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Article

Driving Forces in the Formation of Biocondensates of Highly Charged Proteins: A Thermodynamic Analysis of Binary Complex Formation

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Abstract: A thermodynamic analysis of the binary complex formation of the highly positively charged linker histone H1 and the highly negatively charged chaperone, prothymosin α (ProT α) is given. ProT α and H1 have large opposite net charges (−44 and +53, respectively) and form complexes at physiological salt concentrations with high affinity as shown by Chowdhury et al. (*Proc Natl Acad Sci U S A* **2023**, *120* (41), e2304036120). Data obtained by Chowdhury et al. for binary complex formation are analyzed by a thermodynamic model that is based on counterion condensation modulated by hydration effects. The analysis demonstrates that the release of the counterions mainly bound to ProT α is the main driving force, effects related to water release play no role within the limits of error. A strongly negative ΔC_p (= −0.87 kJ/(K mol)) is found which is due to the loss of conformational degrees of freedom.

Keywords: biocondensate; complex formation; counterion release; hydration

Introduction

When positively charged polyelectrolytes interact with negatively charged polyelectrolytes in aqueous solution, complex formation will result. [1,2] These “complex coacervates” have been the subject of intensive research and surveys of the older literature may be found in various reviews. [3,4] Recently, this problem has found renewed interest because of the formation of biocondensates in living cells. [5–9] Here coacervates are formed within cells by interaction of anionic with cationic proteins. [10,11] The new interest in coacervates has led to a multitude of studies and details may be found in recent reviews. [7,8,12] Up to now, it seems to be understood that charge-charge interaction may play a major role in the formation of biocondensates. [7,13–16] Simulations have led to a better understanding of the driving forces for biocondensate formation in charged systems. [17–20] Here the question arises whether complex formation is driven by enthalpic or by entropic factors. Thus, the release of counterions [21,22] balancing the charge of macroions is expected to contribute to the gain of free energy during complex formation. In an important paper, Ou and Muthukumar [17] performed a comprehensive study of polyelectrolyte complexation by Langevin dynamics and found a significant enthalpic contribution at low to medium charge density. Only at sufficiently high charge density of the polyelectrolytes as expressed through the linear charge density (Γ in ref.[17]), the entropic part of the free energy of binding prevails. This finding has recently been criticized by Chen and Wang [23] who showed that a strong entropic contribution follows from the temperature dependence of the dielectric constant of water. They state that this electrostatic entropy which is due to the reorganization of water dipoles is the main driving force for complex formation rather than the entropic contribution due to counterion release. Counterion release, on the other hand, has been identified as main driving force for a number of systems for quite some time. [21,22,24–32] Moreover, the release of counterions has been observed directly by NMR-techniques. [33–35] The work of Wang and coworkers [23], on the other hand, has underscored the importance of temperature as one of the decisive variables.

It is important to note that accurate data on the complex formation of flexible polyelectrolytes of opposite charge had already been obtained by Mascotti and Lohman many years ago. [26–29] In

that work the interaction of single stranded RNA with oligolysines was analyzed by applying fluorescence techniques. The binding constant K_b was measured at different concentrations of added salt c_s at different temperatures. Thus, a full thermodynamic analysis could be done leading to the result that the release of condensed counterions during complex formation is the main driving force for binding. A quantitative analysis of the data in terms of the counterion release model [21,36] was presented: Counterions are condensed to the highly charged polyelectrolyte and the fraction of counterions thus bound to the macroion could be quantitatively modeled in terms of Manning's theory. [37] Central to this approach is the definition of the charge parameter ξ :

$$\xi = \frac{\lambda_B}{b} \quad (1)$$

where b denotes the distance between the charge along the chain of the polyelectrolyte whereas λ_B is the Bjerrum-length ($\lambda_B = e^2/4\pi\epsilon_0\epsilon kT$; e : elementary charge, ϵ_0 : permittivity of vacuum, ϵ : dielectric constant, k : Boltzmann constant, T : temperature). If $\xi > 1$, a fraction $1 - 1/\xi$ of counterions is condensed onto the polyelectrolyte chain. A part of these condensed counterions is released when the polyelectrolyte forms a complex with an oppositely charged protein. The gain of entropy provides a strong driving force for binding and the model predicts that $\log K_b$ should scale with $\log c_s$. These predictions of theory were met with gratifying agreement when compared to the experimental data of various single stranded RNAs with oligolysines varying in length. [26–29] Small deviations could be traced back and described quantitatively by effects due to hydration. [26–29] Hence, these thorough investigations strongly suggested that counterion release is the main driving force for the first step in biocondensate formation, namely the binary interaction of a cationic with an anionic polyelectrolyte. This finding is in full agreement with a more recent study by Priftis et al. using isothermal titration calorimetry. [13]

The counterion release model can also explain the formation of complexes of long linear or branched polyelectrolytes with proteins.[22] Hence, natural polyelectrolytes as e.g. DNA or Heparin can interact with patches of positive charge on the surface of the protein which leads to formation of a binary complex. Again, the $\log K_b$ is found for many systems to scale linearly with $\log c_s$ as predicted by the counterion release model. [22,30,38–41] It should be noted that these considerations are directly supported by simulations on model systems [42,43] and by simulations using coarse-grained proteins interacting with polyelectrolytes. [44,45] Hydration effects that lead to slightly nonlinear plots of $\log K_b$ vs. $\log c_s$ can be incorporated using the fact that the complex formation is usually done at low concentrations of polyelectrolyte and protein. [21,28] Thus, the activity of water is bound to the activity of the salt ions by the Gibbs-Duhem relation. [46] Changes in hydration lead to a contribution that scales linearly with salt concentration and can be modeled [47] in terms of a parameter Δw . [48–52] A closed expression could be given that comprises both the effect of counterion release as well as of hydration. [53] The model is based on the solute-partitioning model of Record and coworkers [54,55] which provides a quantitative treatment of Hofmeister-effects. An application of this model [53] to the interaction of Heparin with lysozyme was recently presented and demonstrated that a comprehensive thermodynamic analysis of complex formation in solution can distinguish between the effects of counterion release and hydration. [41]

An important experimental contribution to the analysis of biocondensates has been made by the Schuler group who demonstrated that fluorescence techniques can be used for the study of complex formation down to lowest concentrations. [11,56,57] Thus, Chowdhury et al. presented a comprehensive study of the formation of biocondensates using the IDPs prothymosin α (ProT α) carrying 44 negative charges and linker histone H1 having 53 cationic charges. [57] The formation of complexes could be studied on the level of single molecules which leads to the unambiguous determination of binding constant of binary and ternary complexes. In this way the binding constant of binary complex formation can be obtained without the interference of a concomitant phase separation which would set in when working at higher concentrations. [13]

Here the data of Chowdhury et al.[57] for the first step in biocondensate formation, namely formation of a binary complex, will be evaluated. Binary complex formation is characterized by dissociation constants in the nanomolar region and thus provides a strong driving force for biocondensate formation. The present analysis will be done in terms of a purely thermodynamic model developed recently for the analysis of complex formation between proteins and polyelectrolytes. [53] Only two assumptions are necessary for this model: The effect of counterion

release scales with the log of the salt concentration whereas hydration effects scale linearly with salt concentration. The latter dependence is well-established [21] and used successfully to analyze Hofmeister effects on proteins in solution. [54] It is important to note that the counterion release model employed here is based on experimental studies on the interaction of highly charged polyelectrolytes with their counter- and co-ions. [58–62] Hence, the decomposition of the thermodynamic data employed here is based on a firm experimental basis. The present analysis is therefore capable of identifying the main driving forces for the interaction of highly charged IDPs in solution and the parameters derived here provide a firm basis for a more detailed statistical-mechanical model.

Thermodynamic Analysis

All studies done so far on complexation rely on the mass action law and the strength of binding can be expressed in terms of a measured binding constant K_b which is related to the free energy of binding $\Delta G_b(T, c_s)$ by

$$\Delta G_b(T, c_s) = -RT \ln K_b \quad (2)$$

where the measured binding constant K_b is defined through

$$K_b = \frac{[PEP]}{[PEa][PEc]} \quad (3)$$

where $[PEP]$, $[PEa]$, and $[PEc]$ denote the concentrations of the complex, the anionic and the cationic polyelectrolyte, respectively. Going along the lines devised by Record et al.[21,36], the derivative of K_b follows as[53]

$$\frac{d \ln K_b}{d \ln a_{\pm}} = \Delta n_{ci} - \frac{pm}{55.6} \Delta w + \frac{d \ln \frac{\gamma_{PEP}}{\gamma_{PEa} \gamma_{PEc}}}{d \ln a_{\pm}} \quad (4)$$

Here a_{\pm} is the mean activity of the salt ions and $p = 2$ for a monovalent salt with molality m . The first term on the right-hand side is the net number of anions and cations which either released or taken up during complexation. The second term is due to the Gibbs-Duhem relation and the hydration parameter Δw measures the impact of the water molecules released or taken up when the complex is formed. The third term contains the activity coefficients of the complex and of both reaction partners. For the linear polyelectrolytes under consideration here, this term takes care of the Debye-Hückel contribution of the free counterions. Since this term is being considered properly (see below the discussion of eq.(5)), the activity a_{\pm} can be replaced by the salt concentration c_s in the system with full generality. Hence, all subsequent evaluations can be done using the concentrations of the components.

Evidently, Δn_{ci} constitutes the leading term in eq.(4) since the hydration term will only give an appreciable contribution if the molality m of the added salt is high. As a central step of the counterion release model, Δn_{ci} can now be written as [21,36]

$$\Delta n_{ci} \cong Z \left(1 - \frac{1}{2\xi}\right) \quad (5)$$

Here Z denotes the number of charges involved in the binding of the complex. The expression in the bracket stems from the part $1-1/\xi$ of the condensed counterions plus at term $(2\xi)^{-1}$ that is due to the Debye-Hückel contribution of the ions (see eq.(4)).[21,36] For sufficiently high charged polyelectrolytes, $\Delta n_{ci} \sim Z$ and with this the number of released counterions. In this way, Δn_{ci} becomes a stoichiometric coefficient and the release of the counterions can be described by a simple mass action law:

$$K_{th} = \frac{[PEP][M^+]^{\Delta n_{ci}}}{[PEa][PEc]} = K_b [M^+]^{\Delta n_{ci}} \cong \frac{[PEP][M^+]^Z}{[PEa][PEc]} \quad (3a)$$

where K_{th} is the thermodynamic binding constant and $[M^+]$ denotes the activity of the counterions.[21] Since the concentration of the polyelectrolyte is very small, $[M^+]$ can be equated to the salt concentration c_s in the system. Insertion of eq.(3a) into eq.(2) then leads to the logarithmic dependence of $\Delta G_b(T, c_s)$ on c_s . This argument shows that the counterion release model can be traced back in good approximation to the mass action law when the charge density ξ is high enough.

Eq.(5) thus connects the Manning theory and with this general knowledge on the colligative properties of polyelectrolytes to the problem of complex formation under consideration here. Hence, the general validity of eq.(5) can be discussed on the basis of earlier experimental results on polyelectrolytes in solution:

i) For small concentrations of the polyelectrolyte, the charge parameter ξ does not depend on the concentration of added salt c_s . This is a basic assumption of the Manning theory and well-supported by experimental evidence already discussed by Manning. [37] The osmotic pressure measured in a system of a polyelectrolyte and added salt is given in very good approximation by the osmotic contributions of the free counterions of the polyelectrolyte and the salt ions [37] (additivity rule; see also the discussion by Alexandrowicz [58] and by Blaul et al. [61]). It is thus evident that the condensed counterions behave much in the way of a chemical bound species that does not contribute to the osmotic pressure in the system. The released counterions, on the other hand, do contribute to the osmotic pressure and the mass action law applies for them (cf. eq.(3a)). Thus, the logarithmic dependence of the free energy of binding on salt concentration derives directly from this fact, we only need to take into account the effect of counterion condensation. As a consequence, Δn_{ci} does not depend on salt concentration and eq.(4) can be integrated.

ii) The charge parameter $\xi \sim (\epsilon T)^{-1}$ due to the definition of the Bjerrum length (see eq.(1)). Therefore the decrease of the dielectric constant ϵ with temperature is mostly compensated and the change of ξ in the usual range of experimental temperatures (5 – 50°C) is very small. Thus, in excellent approximation, Δn_{ci} does not depend on temperature either. This agrees very well with a great number of experimental observations on e.g. the interaction of DNA with various proteins [38] or on the interaction of highly charged systems with proteins in general. [22,44,53] Thus, a given system can be characterized by a single value of Δn_{ci} independent of temperature. By virtue of argument i), Δn_{ci} is independent of salt concentration as well and presents therefore the central parameter of this analysis.

Given these fact, is evident that eq.(4) can be integrated and rendered in presence of monovalent salt ions [21,28,53]

$$\Delta G_b(T, c_s) = RT \Delta n_{ci} \ln c_s - RT 0.036 \Delta w c_s + \Delta G_{res} \quad (6)$$

The quantity ΔG_{res} describes the remaining part of $\Delta G_b(T, c_s)$ at a suitably chosen reference state.[53] Going along these lines, a closed expression that combines both the effects of counterion release and hydration can be developed. [53] Central to its derivation is the fact that Δn_{ci} does not depend on temperature.

$$\Delta G_b(T, c_s) = RT \Delta n_{ci} \ln c_s + \Delta H_0 - T \Delta S_0 + \left(\Delta c_{p,0} + c_s \frac{d\Delta c_p}{dc_s} \right) \left[T - T_0 - T \ln \left(\frac{T}{T_0} \right) \right] \quad (7)$$

The change of the specific heat Δc_p that is another central parameter (cf. the discussion of the specific heat in ref. [63]) of this analysis and comprises two terms: First, the intrinsic $\Delta c_{p,0}$ which takes into account the changes of Δc_p due to the gain or loss of degrees of freedom during binding. Second, a term due to hydration that scales with salt concentration c_s . Previous work devoted to complex formation of rodlike polyelectrolytes and rigid proteins showed that $\Delta c_{p,0}$ is negligible in these systems.[53] Here we deal with highly flexible and disordered protein and $\Delta c_{p,0}$ is expected to be of appreciable magnitude (cf. also the discussion in ref. [64]).

A new characteristic temperature T_0 has been introduced here which describes the dependence of hydration on temperature through [53,64]

$$\Delta w = \frac{\frac{d\Delta c_p}{dc_s}}{0.036R} \left(\ln \frac{T}{T_0} + \frac{T_0}{T} - 1 \right) \quad (8)$$

For a positive coefficient $\frac{d\Delta c_p}{dc_s}$ the effect of hydration increases the magnitude of ΔG_b for temperatures above and below T_0 . The parameters ΔH_0 and ΔS_0 denote the enthalpic and entropic contributions to the free binding at contact.[53,65] The residual free energy ΔG_{res} follows as [53,65]

$$\Delta G_{res} = \Delta H_0 - T_0 \Delta S_0 \quad (9)$$

It is interesting to note that eq.(7) resembles the well-known generalized van't Hoff expression [66]

$$\Delta G_b(T) = \Delta H_{b,ref} - T\Delta S_{b,ref} + \Delta c_p \left[(T - T_{ref}) - T \ln \left(\frac{T}{T_{ref}} \right) \right] \quad (10)$$

which can be re-written by use of T_s as the reference temperature at which $\Delta S_b = 0$ [53,67]

$$\Delta G_b(T) = \Delta H_b(T_s) + \Delta c_p \left[(T - T_s) - T \ln \left(\frac{T}{T_s} \right) \right] \quad (11)$$

where $\Delta H_b(T_s)$ denotes the enthalpy of binding at $T_{ref} = T_s$. [67] The characteristic temperature T_0 defined through eq.(7) equals T_s if the term due to counterion release is vanishing. In this way, T_0 becomes a parameter that measures the influence of hydration on complex formation.

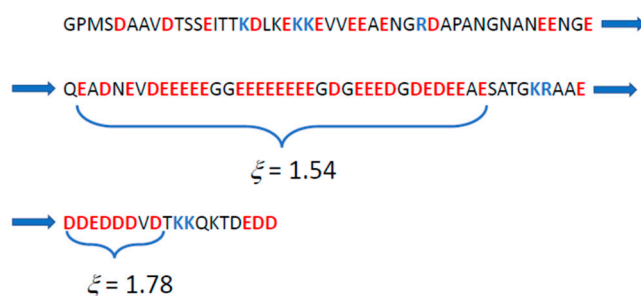
The foregoing considerations suggest to analyze the experimental data in two steps: First, the dependence of ΔG_b on c_s can be determined by eq.(6) neglecting the parameter Δw . In this way a good estimate of the parameters Δn_{ci} and ΔG_{res} can be obtained. Subsequently, the dependence of ΔG_b on T can be analyzed in terms of eq.(11) to obtain an estimate of Δc_p . Both steps proceed in a fully model-free fashion. In a second step Equation (7) can be used to analyze $\Delta G_b(T, c_s)$ for all data at once. [41,53,67] In this way the thermodynamic information embodied in the present set of data can be assessed in a secure fashion.

Results and Discussion

The investigations of Chowdhury et al. [57] leads to the binding constants of the binary complex formation between the anionic IDP ProTα and the cationic IDP H1. Figure 1 displays the respective sequences of amino acids of both IDPs. In case of ProTα a highly inhomogeneous distribution is seen, that is, there is a long sequence consisting only of glutamic acid and aspartic acid, only interrupted by one or at most two uncharged amino acids. If we approximate the mean length of an amino acid by 0.35 nm, such an interruption of a charge sequence is smaller than the Debye-length which for high salt concentrations is still of the order of 0.8 nm and concomitantly larger for a smaller ionic strength. Hence, both sequences indicated in Figure 1a) can be treated as a single highly charged line that can be characterized by a local charge parameter $\xi = 1.54$. (cf. eq.(1)). For these sequences the percentage of condensed counterions can be estimated to be 70% according to eq.(5). Thus, Δn_{ci} (eq.(5)) is expected to be ca. 19, i.e., 19 counterions are condensed and do not contribute to the osmotic pressure in the system. For the smaller sequence, $\xi = 1.78$ which is followed by $\Delta n_{ci} \sim 5$. Thus, the arguments expounded in the section thermodynamic analysis can be fully applied here.

a)

Unlabeled ProTα



b)



Figure 1. a) Sequence of amino acids for the unlabeled anionic IDP ProTα, and b) sequence of amino acids of the unlabeled cationic IDP H1. Negatively charged amino acids are labeled in red whereas positively charged amino acids are labeled blue. Highly charged sequences in ProTα (Figure 1a)) are characterized by the charge parameter ξ (cf. eq.(1)).

The increase of the strength of charge-charge interaction with increasing “blockiness” is well-borne out also from recent simulations. [7,31,68] It should be noted, however, that the polyelectrolyte must exceed a certain length so that counterion condensation can take place. These effects related to the termini of the polyelectrolyte chains have been considered in detail by Manning [69] and seem to describe the experimental data obtained for short polyelectrolytes very well. [70] Thus, the effective length of a polyelectrolyte in which counterion condensation may take place follows by subtracting the Debye-length of each end. Hence, Δn_{ci} (eq.(5)) is expected to be slightly smaller than the above estimate of 19. It should be kept in mind, however, that Δn_{ci} contains not only the release counterions of the anionic polyelectrolyte but also a number of the counterions of the cationic polyelectrolyte (see below).

It is interesting to compare this sequence of charged amino acids to the one found for the cationic IDP H1 (see Figure 1b). Here it is obvious that there is no long virtually uninterrupted series of cationic amino acids, the charges are far more evenly distributed than it is found for ProTα (Figure 1a). Typically, doublets or at most triplets of lysine are separated by one or two uncharged and hydrophobic amino acids. Such a sequences has found earlier in proteins that interact strongly with Heparin (Cardin-Weintraub sequences [71,72]; cf. also ref. [73]). Thus, these sequences can interact closely with highly charged anionic polyelectrolytes. However, the correlation of the counterions to the macroion will be smaller for H1 than in case of ProTα. This can be estimated from the fact that the charge parameter ξ (cf. eq.(1)) as calculated for the entire chain is only 0.58.

Evidently, the interaction of ProTα with H1 can be compared to the well-studied case of globular proteins binding to highly charged polyelectrolytes: [74] A highly charged negative polyelectrolyte as e.g. DNA or heparin having a large number of condensed counterion interact with small cationic patches localized on the surface of a protein (“Heparin binding site”; cf. ef. [73]). The number of positive charges of H1, however, is very high and the question arises why H1 exhibits no toxic interaction with cell organelles. [75] Since a long time it is known that the charge density of a cationic polyelectrolyte is largely determining its toxicity. [76] This feature has been corroborated by more recent investigations and surveys of the problem. [75,77] One may speculate that the smaller correlation of the counterions may mitigate the intrinsic toxicity of H1 by preventing its unspecific interaction with cell membranes.

In the following, the binding constants obtained by Chowdhury et al. [57] for the binary complexes ProTα/H1 will analyzed in terms of eq.(6) and (10) as described above. Figure 2 displays the free energies of binary complex formation of ProTα/H1 as the function of salt concentration for three temperatures. The lines in this semilogarithmic diagram run parallel to each other. Similar findings have been made for many other systems [22,44,53] which directly demonstrate that Δn_{ci} does not depend on temperature. From the data measured at 295K this parameter follows as $\Delta n_{ci} = 18.2$.

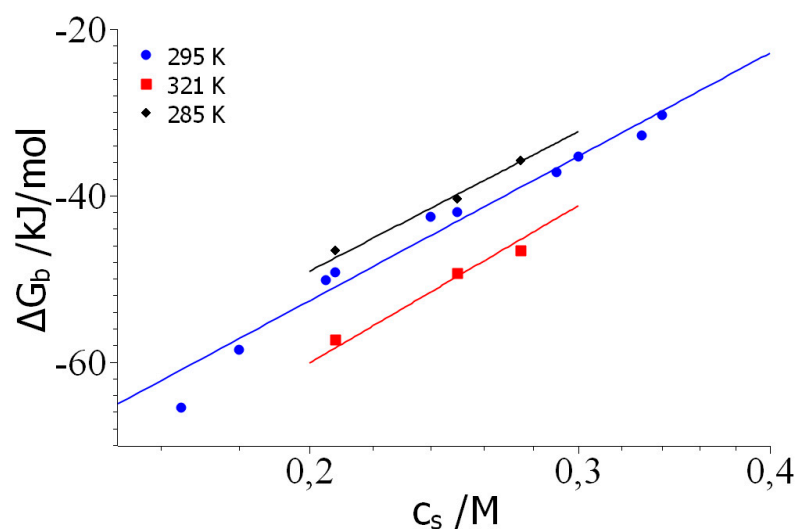


Figure 2. Semilogarithmic plot of the free energies of binary complex formation at a temperature of 285 K (black diamonds), 295 K (blue circles) and for 321 K (red quadrangles). Data taken from Figure 4a of Chowdhury et al..[57] The solid lines show the fits according to eq.(7).

To the author's best knowledge, this is the highest value ever measured for this parameter. It points to a very strong effect of counterion release which is ultimately responsible for the huge binding constants measured at lowest salt concentration for this system. Indeed, free energies of binding with a magnitude larger than 60 kJ/mol are enormous given the fact that complex formation of polyelectrolytes with proteins is usually characterized by values between 30 and 40 kJ/mol. [22,74]

The results shown in Figure 2B of ref. [57] show two additional points of interest: 1. Experiments using different monovalent ions demonstrate the independence of the counterion release effect on the type of ions used in the experiments. This observation points to the fact that here we deal with purely electrostatic interaction that do not involve any specific interaction of the macroion with the monovalent counterions. [53] This finding points to the absence of hydration effects and the analysis of the dependence of the free energy of binding on temperature will fully corroborate this result (see the discussion of Figure 3 below). 2. Figure 2B of ref. [57] displays data relating to divalent ions. In principle, such data should allow us to differentiate between the release of cations and anions: If only monovalent cations condensed to ProTα would be released, replacement of Na⁺-ions by Mg²⁺-ions would lead to a slope in plot of $\ln K_D$ vs. $\ln a_+$ in Figure 2B that is half of the slope found for monovalent salt ions. [21] The slope seen in Figure 2B of ref.[57] both for experiments with Mg²⁺-ions as well as in presence of SO₄²⁻-ions suggests that ca. 13 ions are released. This finding could be explained by assuming that an equal number of positive and negative ions are released, which means that the number of released counterions will be reduced for salts like MgCl₂ or K₂SO₄ by a factor of 0.75 as already noted by Chowdhury et al. [57]. The experimental accuracy of the data referring to divalent ions is smaller than the data referring to monovalent ions, however, and the comparison of theory and experiment is only semi-quantitative in this point. Experiments using mixtures of Mg²⁺- and Na⁺-ions would be very helpful to decide this point. [78]

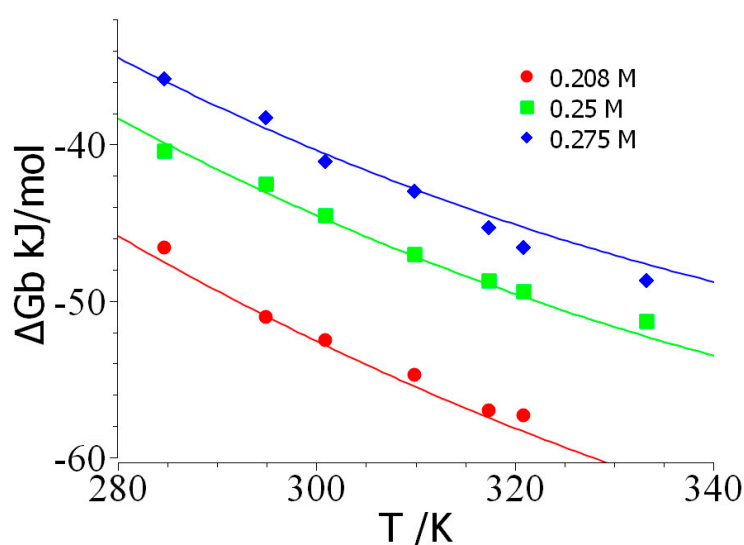


Figure 3. Analysis of the dependence of the free energy of binding ΔG_b on temperature. The data have been taken from Figure 4E of Chowdhury et al.[57]. The solid lines mark the fit of the data by eq.(7).

The dependence of the free energy of binary complex formation of ProTα/H1 temperature is shown in Figure 3. First, it is important to note that the curvature of these plots directly point to a negative specific heat of binding Δc_p . A quantitative analysis of this point by eq.(11) shows that $\Delta c_p \sim -0.8$ kJ/(K mol) for salt concentrations of 0.25 and 0.275 M whereas a value of -2 kJ/(K mol) follows for 0.208 M. The problems of this analysis are at hand, however: Application of eq.(10) ultimately amounts to a numerical differentiation of ΔG_b and small experimental errors will lead to huge errors in Δc_p . Nevertheless, this analysis clearly reveals that Δc_p is negative.

In the second step, an in-depth analysis of the data displayed in Figure 2 and 3 can now be done with the aid of eq.(7). Here the entire set of data is analyzed at once using the MathLab routine *cftool*, that is, all 31 data points $\Delta G_b(T, c_s)$ at fitted by a single set of parameters.[41,53,67] At first, the hydration parameter $d\Delta c_p/dc_s$ is neglected and the data are fitted for a chosen value of the characteristic temperature T_0 . For $T_0 = 290$ K we obtain the following set of parameters: $\Delta n_{ci} = 17.5$; $\Delta H_0 = 48.1$ kJ/mol; $\Delta S_0 = 0.107$ kJ/(K mol), and $\Delta c_{p,0} = -0.87$. The solid lines in Figure 2 and 3 display the fit of all data by this set of parameters. The data set would be fully compatible with $d\Delta c_p/dc_s \cong 1$ kJ/(K mol M) which points to $\Delta w \sim +100$, that is, to a release of water molecules at room temperature. However, the fit with neglect of this parameter shown by the solid lines in Figure 2 and 3 is fully sufficient. This is in full agreement with the fact that no ion-specific effects are seen (see above) which would point to Hofmeister effects due to hydration.

The fit of in total 4 parameters may appear questionable. However, Δn_{ci} is more or less identical with the value obtained directly from the plot in Figure 2 and $\Delta c_{p,0}$ agrees with the value estimated from the above analysis by eq.(10). So we are left with the two remaining parameters ΔH_0 and ΔS_0 that lead to $\Delta G_{res} = 16.8$ kJ/mol according to eq.(9). Linear extrapolation of the plot shown in Figure 2 to $c_s = 1$ M leads to $\Delta G_{res} = 18.5$ kJ/mol which is close by. This extrapolation works under the assumption that $\Delta w = 0$ which seems to be fully supported by the above finding that $d\Delta c_p/dc_s$ can be neglected in good approximation.

The strong positive enthalpy $\Delta H_0 = 48$ kJ/mol compares favorably to the values of 37 – 58 kJ/mol measured directly by calorimetry. [57] In general, the interpretation of the measured enthalpies, however, should proceed with caution as already remarked by Ou and Muthukumar. [17] The enthalpy measured in an ITC-experiment may contain many contributions that are difficult to calculate and which may cancel out each other. In particular, it should be noted, that the enthalpy ΔH_{ITC} measured directly by ITC must not be confounded with the enthalpy of complex formation ΔH_B . It is well-known that ΔH_{ITC} may contain additional contributions from linked equilibria as e.g. protonation during complex formation. Thus, Baker and Murphy analyzed the binding enthalpies for biomolecular complex formation by comparison with the enthalpies of buffer ionization. [79,80]

They could demonstrate that the linked equilibrium of buffer ionization can furnish a marked contribution to the observed enthalpy. Extrapolated to zero heat of buffer ionization, the measured binding enthalpies may even change their sign. The same observation was made Hileman et al. [81] or by La et al. more recently in a series of carefully conducted ITC-experiments. [82] In principle, ΔH_B can be derived from the measured ΔH_{ITC} by extrapolation to a vanishing heat of buffer ionization. It also can be obtained through analysis of the experimental data with the aid of eq.(7) giving ΔH_0 as the enthalpy at $T = T_0$ or by eq.(11) which yields $\Delta H_b(T_s)$. Since the evaluation via eq.(7) used the entire set of data, the resulting ΔH_0 is certainly the more accurate result. The resulting $\Delta H_0 = 48$ kJ/mol hence can be taken as a reliable result which indicates that complex formation is accompanied by a strongly positive enthalpic contribution partially balanced by a marked positive entropic term $T_0\Delta S_0 = 31$ kJ/mol. A similar observation has been made by Priftis et al. in their studies of complex coacervation of synthetic polyelectrolytes.[13] A possible explanation of these findings may be sought in the disturbance of the water structure by the released counterions which have formerly condensed onto the negative polyelectrolyte. A cleavage of hydrogen bonds hence will lead to a loss of enthalpy together with a concomitant raise of entropy because of the induced disorder. In general, a strongly positive enthalpic contribution ΔH_0 together with a markedly positive entropy ΔS_0 points to a disordering of water molecules upon formation of the complex. A similar situation has been discussed by Netz and coworkers when considering the hydration repulsion between biomembranes. [83,84] Here too it was found that at close distance a strong enthalpic repulsion results which is not compensated by a concomitant entropic term. In addition to this, water polarization effects may enhance these effects. [83,84] The strong positive enthalpy may have its origin in terms related to the change of the structure of the water phase during complex formation.

Summarizing the above analysis, it is evident that the counterion release term with $\Delta n_{ci} = 17.5$ is dominating ΔG_b . Thus, for the physiological salt concentration of 0.15 M, the first term on the right-hand side of eq.(7) amounts to -80 kJ/mol at 290 K which demonstrates that counterion release is by far the strongest driving force for binary complex formation. At this point it is interesting to discuss the entropic contributions in eq.(7) again and calculate explicitly the contribution to entropy from the dependence of the dielectric constant ϵ on temperature. [23] If the part of ΔG_b due to the release of counterions is defined by

$$\Delta G_{b,CR}(T, c_s) = RT \Delta n_{ci} \ln c_s \quad (12)$$

the respective entropy deriving from this part is (cf. eq.(5))

$$-\Delta S_{b,CR} = \left(\frac{\partial \Delta G_{b,CR}}{\partial T} \right) = R \ln c_s \left(\Delta n_{ci} + \frac{T}{2\xi} \frac{d \ln \lambda_B}{d \ln T} \right) \quad (13)$$

In order to calculate the dependence of Bjerrum length λ_B on temperature, we use the expression furnished for $\epsilon(T)$ by Malmberg and Maryott. [85] From these data $d \ln \lambda_B / d \ln T$ follows as 0.3 for a temperature of 300 K. With a charge parameter $\xi = 1.5$ of ProT α , the correction to $\Delta S_{b,CR}$ given by the second term in the bracket is 0.1 which is much smaller than $\Delta n_{ci} = 17.5$. This result shows that there is no significant part of the entropy which is due to the dependence of the dielectric constant on temperature. It also demonstrates that the neglect of the dependence of Δn_{ci} on temperature (cf. the discussion of eq.(5) and (6) above) is fully justified.

It rests to discuss the change of the specific heat Δc_p . Strong effects from hydration can be ruled out since no ion-specific effects are observed for the present system which would lead to a term depending on salt concentration (cf. the discussion of this problem in ref. [41]). The strongly negative $\Delta c_{p,0} = -0.87$ kJ/(K mol) must therefore be traced back to the loss of conformational degrees of freedom of both binding partners during complex formation.

Conclusions

A phenomenological thermodynamic analysis of the binary complex formation of the highly positively charged linker histone H1 and the highly negatively charged chaperone, Prothymosin α (ProT α) has been presented. The analysis is fully based on a model-free phenomenological approach that is in full accord with the well-known colligative properties of polyelectrolytes, no additional assumptions need to be invoked. We find that the release of counterions as expressed through the parameter Δn_{ci} (eq.(6) and (7)) is the main driving force for complex formation. In addition to this, a

strong additional entropic term ΔS_0 (eq.(7)) supports binding whereas the enthalpic term ΔH_0 (eq.(7)) derived from the above analysis is positive. Moreover, hydration effects as expressed through the term Δv in eq.(6) were found to be small. Finally, the negative $\Delta C_{p,0} = -0.87$ kJ/(K mol) points to a loss of conformational degrees of freedom of the IDPs in the complex as expected. The entire analysis shows that complex formation of the IDPs in solution may well be compared to the much-studied problem of proteins interacting with highly charged polyelectrolytes. [22,74]

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