

Review

Not peer-reviewed version

A Review on the Biorefinery Approach and Marketing Strategy of Leafy Biomass

Nushrat Yeasmen*, Md. Hafizur Rahman Bhuiyan, Valerie Orsat

Posted Date: 28 December 2023

doi: 10.20944/preprints202312.1969.v1

Keywords: Antioxidant; biomass; biorefinery; by-products; forestry biomass; phenolic compounds.



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Remiero

A Review on the Biorefinery Approach and Marketing Strategy of Leafy Biomass

Nushrat Yeasmen 1,*, Md. Hafizur Rahman Bhuiyan 1 and Valérie Orsat 1

- Department of Bioresource Engineering, McGill University, Sainte-Anne-de-Bellevue, Quebec, H9X 3V9, Canada
- * Correspondence: nushrat.yeasmen@mail.mcgill.ca

Abstract: In terms of sustainability, there is a pressing need to evaluate agricultural and forestry leafy biomass that has no current economic value or may pose a threat to the environment. In this aspect, leafy biomass, which represents around 5% of the total tree, can be used for purposes that would be more profitable and ecofriendly. To this end, biorefinery of leafy biomass in the way of extracting valuable health compounds such as phenolics would ensure a valuable societal health ingredient besides alleviating waste disposal problem. However, during the biorefinery process, the primary challenges start with ensuring the feedstock to produce phenol rich leaves extract followed by their isolation and commercialization of this leaves extract. This current review aims to detail the types of biomasses, biorefinery approach of leafy biomass towards the phenolic extraction, leaves commercialization followed by the marketing strategy of the application of phenol rich leaves extract. The outcome of this overview will serve researchers and relevant industries in understanding the process for feedstock collection and the suitable sectors to apply leafy biomass derived healthy compounds.

Keywords: Antioxidant; biomass; biorefinery; by-products; forestry biomass; phenolic compounds

1. Introduction

Energy production, biochemical waste minimization, availability of nutritious food, and climate change are some of the major concerns worldwide. To feed the increasing world population, on the one hand advanced agricultural practices and postharvest techniques are inevitable, while on the other hand, these increased food, feed, and energy crop productions has resulted in a huge amount of agricultural wastes and by-products [1]. In other words, the production stage represents 24–30% of global agricultural by-products, while post-harvest stage accounts for 20% [2,3]. The other sector of interest is forestry which also produces large amounts of biomass derived from harvestable yield. In total, the global annual generation of biomass waste is in the order of 140 Gt, and this presents significant management problems for industry in terms of cost and environmental effects induced by their disposal [4]. However, with the increase in environmental awareness, there is growing interest in circular economy where these underutilized biomasses can be a rich and cheapest source of several bioactive components viz. polyphenols, carotenoids, flavonoids, anthocyanins, and vitamins. Sustainability and circularity of processes are key words for industrial sector encompassing the reduction of postharvest losses, valorization of by-products, and extraction of bioactive compounds [5]. This review aims to highlight the types of biomasses, biorefinery approach of leafy biomass towards the phenolic extraction, leaves commercialization followed by the marketing strategy of leaves extract application.

2. Biomass

Biomass can be derived from two sectors namely agricultural biomass and forestry biomass. Agricultural biomass can be categorized as (1) agricultural harvest generated by-products, (2) postharvest by-products, and (3) food-processing by-products and wastes. Agricultural biomass including crop stalks, leaves, hays, straws, twigs, and roots (agricultural harvest generated by-products); fruit peels, rinds, pulps, bran, husks (postharvest by-products); pomace, seed/nut shells

(food-processing by-products and wastes), among others, are normally discarded or burned (Table 1) [1,3]. Burning organic material emits nitrogen oxides, carbon dioxide, carbon monoxide, sulfur dioxide, lead, mercury, and other hazardous air pollutants which contributes to climate change, being responsible for an estimated 8% of global greenhouse gas emissions [2,4]. It is estimated that 140 Billion tons of biomass are generated each year from agricultural by-products in liquid, solid and gaseous form [5]. On the other hand, forestry biomass includes residues such as stumps, branches, barks, stems, and leaves and processing waste such as logs and sawdust which accounts for around 4.6 Billion tons annually of which 60% goes to energy generation, 20% to industrial 'round wood' and the remaining 20% being primary production loss that remains in-field to decay [6]. Forestry biomass used in power plants to produce energy causes huge increase in carbon emissions compared to coal and other fossil fuels. However, biomasses are underutilized source of material as such there has been little activity focusing on a comparatively 'low-carbon' route for their valorization [4].

Fruit/vegetab **Biomass** References le Carrot 30-40% (pomace) Kaur et al. [7] **Tomato** 3-7% (seed and peel) Schieber et al. [8] Apple 11% (seed core and pulp) Ayala-Zavala et al. [9] Banana ≤30% (peel) Schieber, Stintzing and Carle [8] Guava 10-15% (seed and peel) Selvamuthukumaran and Shi [3] Ayala-Zavala, Rosas-Domínguez, 11% (peel), 13.5% (seed), 17.9% Mango Vega-Vega and González-Aguilar (unusable pulp) [9] Orange 66% (peel) Li et al. [10] Ayala-Zavala, Rosas-Domínguez, 8.5% (peels), 6.5% (seeds), 32.1% Vega-Vega and González-Aguilar Papaya (unusable pulp) [9] 13.5% (peels), 14.5% (pulp), 9.1% (core), Selvamuthukumaran and Shi [3] Pineapple 14.9% (top)

Table 1. Biomasses generated from fruit and vegetable processing industry.

Regarding biorefinery aspect, several reports have shown that agricultural and forestry biomass can be a potential source of bioactive compounds such as polyphenols which are reported to show several biological activities (anti-allergenic, anti-inflammatory, anti-artherogenic, cardioprotective, anti-microbial, anti-thrombotic, and antioxidant effects). Having said that agricultural and forestry biomass could be a good choice as sustainable and renewable feedstock for the functional, nutraceuticals, and pharmaceutical industries [11-19]. To this end, extraction of bioactive compounds using green extraction methods as a 'low-carbon' route can be very efficient knowing their uprising market demand. For example, based on Grand View Research (2019), the global polyphenols market size was already worth USD 1.68 Billion in 2022 and a Compound annual growth rate (CAGR) of 7.4% is expected to register from 2023 to 2030. According to them, the functional food and beverage industries are the largest sectors of the global market share (> 70%) of polyphenols followed by dietary supplements, cosmetics, animal feed, and others [20]. Hence, biomass processing through green extraction technology would offer double benefit (i) extraction of high value compounds that have market demand, and (ii) reduction of carbon emissions, that is caused by conventional biomassto-energy conversion plant. In this way, use of biomass as a source of extraction materials will not only benefit the agri-food economy but also would have positive environmental impact.

3. Promising biomass as a part of bio-refinery approach

Leaves could be a promising source to exploit valuable compounds such as phenolics. Different phenolic compounds have been separated from different leaves, for instance, chlorogenic acids and anthocyanin from blueberry leaves [21]; 5-O-galloylquinic acid and 5-O-galloylquinic acid from

mastic tree leaves [22]; phenolic acid derivatives and flavonoids from moringa leaves [23]; oleuropein from olive leaves [24]; isoquercetin, catechin, hydroquinin, gallic acid, tannic acid, and rutin from cashew leaves [25], among others. Other biomass of agricultural origin, in specific, biomass form postharvest by-products, and food-processing by-products and wastes, are not homogenous and may come from different fields with differing cultivation procedures, conditions and processing; consequently, a standardization process is necessary. Moreover, agricultural food by-products are characterized by their biological instability due to their high microbial load that can affect product safety and naturally accelerate microbial degradation [5]. Another point is, most of the agricultural biomass coming from processing industries undergo several processing treatments such as heating, which might have negative impact on the stability of phenolic compounds. On contrary, leaves can be a potential source of phenolics and the justification behind choosing leaves among other biomasses of agricultural and forestry origin can be given as such,

- leaves are a sustainable source where overexploitation is believed not to be an issue as leaves can be easily grown back by the plant [26,27].
- leaves can be used as fresh, therefore, can preserve the quality of phenolic compounds.
- leaves can be a cheapest source for the extraction of phenolic compounds.

Note that, leaves from one source might be rich in some selective phenolic compounds, while other leaves may contain higher amount of other phenolic compounds, as phenolic compounds possess a broad category of chemical compounds [28]. To date, more than 10,000 phenolic compounds have been identified in plants [29]. In this scenario, the combination/blend of several leaves extract would offer higher benefit. However, before commercialization of leaves' phenolic compounds (either single/ blend of leaves), thorough toxicological study would be the mandatory step, as leaves may contain toxic substance or potentially toxic substance (that comes into the extract during extraction) [30–32]. Moreover, toxicological study on blend of extract (which is separately safe) need to be performed as well, as there might be any synergistic event that could develop new toxic substance, even though they were not present before the mixing or blending of the leaves / leaves' extracts.

4. Leaves commercialization

Availability of raw material is the prerequisite of any industrial production process that takes part in a circular economy. Likewise, the regular supply of leaves (as raw/semi-processed) throughout the year should be ensured. However, leaves are identified as perishable commodity (moisture content up to 80% w.b.) and hence proper storage is required until they are used for required purposes, for example, extraction, etc. [33]. High water containing commodities are highly perishable as they are prone to microbial growth and enzymatic activity. Therefore, perishable commodities are preferably stored at lower temperatures to increase shelf life and retain quality. For prolonged storage and maintenance of the quality of biological samples, storage at -20°C or lower is preferred. However, storage of fresh biomaterial consumes space and is more expensive in terms of transportation (due to the greater mass from the high-water content). To this context, drying can reduce the space requirements and make the transportation of the dried commodity easier because it decreases the mass of the commodity to be transported, along with increasing its shelf life [34].

Drying is a process to remove moisture from fresh commodities and reduce its water activity, which inhibits microbial growth and minimizes deteriorative biochemical reactions. It also reduces the weight and volume of the sample thereby reducing storage and transportation costs as already mentioned. In addition, drying can modify the physical microstructure of plant tissues, which leads to increased extraction yields [35]. For example, freeze drying causes significant changes in the microstructure of the final dried product making it more porous so that solvents can easily penetrate the sample and thus extract more phytochemicals [36–39]. Several drying methods have been practiced with leaves aiming for extraction of phenolic compounds such as freeze drying [40–47], microwave drying [34,40,42,44,48–50], oven drying [41,44,49], spray drying [51], sun drying [40,42,44], vacuum drying [40], shade drying [43], among others.

3

4

As seen above, for sample preparation for phenolic content analysis, freeze drying has often been applied, however, it is a relatively expensive drying method. For economical commercial applications, the most practical method is hot air drying, however, application of higher temperatures can lead to degradation of the phenolic components [40]. Considering these aspects, it is necessary to identify the most suitable drying method and conditions for a specific type of plant sample. Next to drying, the grinding of dried leaves followed by sieving and storing in controlled environment (temperature, relative humidity, light, oxygen, etc.) would offer rapid industrial processing of maple leaves as their raw material.

To give an example of how leaves can be commercialized throughout the year, here the species Acer saccharum is considered as a representative. In Canada, sugar maple leaves calendar year starts in late spring (May) and ends in late fall (November) where they fall to ground to prepare for the winter. Having said that, supply of maple leaves within these months (May-November) can be easily secured. Collecting of leaves in peak season followed by their preservation might also ensure the availability of maple leaves in rest of the months (December-April). Maple leaves can be obtained from both cities and forests. In this aspect, existing maple farms utilized for maple sap collection for the production of maple syrup, could be a good choice to have homogeneous leaves (i.e., maple leaves only). Currently there are 5340 maple farms exist in Canada, with 4776 maple farms originating from Quebec, 391 from Ontario, 111 from New Brunswick, 48 from Nova Scotia, and 14 maple farms from rest [52]. As these farms are under regular maintenance, collecting leaves from these farms besides forests and cities would be most convenient. Maple leaves could be collected from tree/surrounding ground, depending on the time of the life cycle of the tree in a calendar year. Afterwards, the masscollected leaves would be sorted out (based on shape/density/color/etc.) to separate the targeted leaves from unwanted substances i.e., from damaged and unhealthy leaves, rocks, debris, etc. The sorted leaves could be blown by cold air to remove dirt/debris followed by washing them in running tap water. Then the selected healthy leaves will undergo drying process and followed by grinding and sieving of dried powder. The dried powder will be packed in a good quality container (that would not allow interference from surrounding environment to the leaves powder), and the packed powder will be stored in controlled environment until its industrial processing.

5. Marketing strategy of phenol rich leaves extract

Phenol rich leaves extract obtained through green extraction can be utilized in different sectors based on their functionalities. Knowing that there is already an increasing market demand for phenolic compounds. For instance, Belwal et al. [53] reported that the nutraceutical market share would increase through phenol derived biological materials. In this aspect, the demand for phenolic compounds in global markets was expected to reach USD 873.3 million in 2018 based on the study conducted by Transparency Market Research [54-56]. Therefore, it can be assumed that leaves extracts have a better future to dominate the phenolic market as well as relevant industrial sectors. However, marketing strategy of leaves phenolics could be linked with their several biotechnological activities [28]. Generally, the most analyzed property of phenolics is their antioxidant activities, which have been often associated with the prevention, modulation, or treatment of significant diseases and health disorders in humans, including neurodegenerative and cardiovascular disorders, atherosclerosis, and diabetes [57]. Phenolic compounds are considered as antioxidants due to the donation of a hydrogen atom and/or an electron to free radicals, causing the break of chain reaction of oxidation. The antioxidant effect depends on the number and position of the hydroxyl groups [27]. Other important bioactivities for maintaining good health have also been associated with these compounds, e.g., anti-inflammatory, antimicrobial, and anti-proliferative activities, and these biological activities have aroused interest in the use of these molecules in the formulation of nutraceutical products [28]. In addition to the pharmacological interest in these compounds, their biological activities (antioxidant, antimicrobial) could also be explored in other industry sectors, such as food industries, cosmetic industries, and packaging industries. To be precise, depending on the biological activities showed by phenolic compounds, they can be marketed to the focused industries. For example, phenolic compounds can be used in nutraceutical industries as therapeutic agents; in

food industry as food additives (in the form of preservatives- antioxidant and antimicrobial actions; food colorant) and functional ingredients; in packaging industries as active packaging (antioxidant and antimicrobial action) and smart packaging (color indicator for deterioration); in cosmetic industries as antimicrobial, antioxidant, tyrosinase inhibitor, UV protector [28].

5.1. Nutraceutical industries

"A nutraceutical is a product isolated or purified from foods that is generally sold in medicinal/supplemental forms not usually associated with food. A nutraceutical is demonstrated to have a physiological benefit or provide protection against certain chronic diseases" [58]. However, a major challenge of the current nutraceutical sector is ensuring bioavailability for obtaining the desired effect. To define, bioavailability is the fraction of a nutrient that has been digested and absorbed and is available for the metabolic functions of the organisms [59]. Bioavailability can be increased by increasing the fraction/quantity of these compounds or the quality of the extract. In this aspect phenolic nanoparticles can be more effective. Yehia et al. [60] produced phenolic nanoparticles from eggplant skin through dispersion with the application of ultrasounds and obtained higher antimicrobial activity. An enhanced delivery system using nanocarriers can contribute towards increasing the efficiency of the antioxidant [57]. Selivanova and Terekhov [61] has presented the increased bioavailability of flavonoids achieved through the application of crystal engineering which is "an applied field of supramolecular chemistry including design and synthesis of new crystalline compounds with specified physicochemical properties".

5.2. Cosmetic industries

The defensive properties of the phenolic compounds have been useful in other applications. Phenolic acids are being used as natural antioxidants in the cosmetic industry, as they possess defensive properties against growth and evolution during the pathological conditions associated with oxidative stress [57]. Ma et al. [62] evaluated the use of glucitol-core containing gallotannins (GA) of red maple leaves extract in cosmetic applications and found that GA was able to: (1) reduce the levels of reactive oxygen species, (2) down-regulate the expression of MITF, TYR, TRP-1, and TRP-2 gene levels in a time- dependent manner, and (3) significantly reduce protein expression of the TRP-2 gene. Olive leaves extracts had a strong potential as an antiaging ingredient and cosmetic/food preservative exhibiting antioxidant activity, capacity to inhibit elastase (82.5%), collagenase (98.7%) and tyrosinase (50%) [63].

5.3. Food industries

5.3.1. Functional food

According to Health Canada, "A *functional food* is similar in appearance to, or may be, a conventional food, which is consumed as part of a usual diet, and is demonstrated to have physiological benefits and/or reduce the risk of chronic disease beyond basic nutritional functions" [58]. To date, as a source of phenolic compounds, the application of rosemary extract, anthocyanin, steviol glycosides as food additives has been approved by European Union and therefore they are practiced on a variety of food products [64–67]. Due to their strong antioxidant properties, olive leave extracts were seen in pork patties, sunflower oil, palm oil, and olive oil applications [68], *Gingko biloba* leaves extract in pork meat [69], green tea phenolic extracts have been seen in bread applications [70].

5.3.2. Preservatives

Phenolic compounds with strong antioxidant properties can also be used as preservatives. According to the United States Food and Drug Administration (USFDA) Code of Federal Regulation, "antioxidants are the substances participates in physiological, biochemical, or cellular processes that inactivate free radicals or prevent free radical-initiated chemical reactions". Apart from the consumers increasing demand for "clean label" natural ingredients, the raising safety concerns about

5

6

synthetic antioxidants (e.g., BHA, BHT, PG, and TBHQ) also necessitate the search for effective antioxidants from natural sources [71]. Phenolic compounds are classified as primary antioxidants that delay or inhibit the initiation step or interrupt the propagation step of oxidation by scavenging the free radicals. The antioxidant potential of phenolic compounds however mainly depends on the number and arrangement of the hydroxyl groups (-OH) in the molecules of interest [72]. The phenolic extracts obtained from fermented rice bran were applied in pizza doughs stored at room temperature, where the shelf life increased by 10 days [73]. In a different study by Christ-Ribeiro et al. [74], the efficiency of antifungal effects of phenolic extracts obtained from fermented rice bran and Spirulina sp. have been found to be 2.5- and 1.5-folds, respectively, compared with calcium propionate. Ab Rashid et al. [75] employed microencapsulated forms of anthocyanins from Clitoria ternatea as a biopreservative and coloring agent for baked food products, where encapsulation enhanced the stability of the compounds for pH fluctuations and applicability. Moreover, green tea extracts have been used in pork sausages which in turn ensured shelf life extension [76].

5.3.3. Packaging industries

Phenolic compounds have also been proposed as part of the composition of membranes and films packaging material, contributing towards the enhancement of the shelf life of beef [77,78]. Catechin and quercetin as phenolic compounds were used in ethylene-vinyl alcohol copolymer (EVOH) film for the packaging of oxygen sensitive foods, which resulted in the improvement of food stability [79]. Attributing to the sensitivity of anthocyanins to the change in pH, these compounds were utilized for monitoring pH variations on the development of chitosan and starch-based intelligent films embedded with anthocyanins [80]. Gelatin/ polyvinyl alcohol based colorimetric indicator film with mulberry anthocyanins was applied for monitoring fish freshness, where visible color changes were observed with the release of volatile nitrogenous com- pounds over the storage period [81]. Da Rosa et al. [82] developed of biodegradable films with improved antioxidant properties based on the addition of carrageenan containing olive leaf extract for food packaging applications.

5.3.4. Potential application of leaves extracts in food applications

Facing restrictions to the use of artificial food additives, the scientific community has been working on the development of natural alternatives. Moreover, owing to consumers' preferences and demands for functional foods, bioactive phytochemicals from leaves extracts can be progressively applied as the ingredients to improve quality traits, nutritional and therapeutic properties. A list of potential food applications fortified with different plant extracts as a source of phenolic compounds is given in Table 2. Based on the provided food applications fortified with different plant extracts, it can be hypothesized that application of leaves extracts might also be possible to those food as leaves can also possess similar classes of phenolic compounds. For example, with reference to the approval of rosemary extract by the EU, the main phenolic compound isolated from rosemary leaves extracts include coumaric acid, caffeic acid, gallocatechin, luteolin, and isorhamnetin [83,84]. These phenolic compounds have also been isolated in the extract of leaves from stevia, moringa, sea buckthorn, and pistacia [56].

PCs Product Effects References Source Matricaria recutita, Caleja et al. Yogurt Antioxidant activity Phenolic Foeniculum [85] extracts vulgare. Sheep meat Litchi chinensis Antioxidant activity Das et al. [86] nuggets

Table 2. Application of phenolic compounds (PCs) in different foods.

6. Conclusion and future directions

Leafy biomass coming from either forest or agricultural practices or from food industries, is rich in many beneficial components. Therefore, in stead of leaving them to become the treat to the environment, biorefineries combining the profitable extraction and production of various nutraceuticals, pharmaceuticals, or natural antioxidant ingredients will not only decrease the environmental pollution but also help the economy by creating new sources of income. However, future research on the life cycle assessment of the biorefinery process of leaves toward the extraction of phenolic compounds is recommended to find out the most cost effective and eco-friendly method is recommended. In addition, research is required on the controlled delivery of the leaves' phenolic compounds both in in vitro and in vivo conditions.

properties

Author contributions: Nushrat Yeasmen: Conceptualization, Methodology, Investigation, Formal analysis, Writing – original draft, Writing – review & editing. **Md. Hafizur Rahman Bhuiyan:** Writing – review & editing. **Valérie Orsat:** Resources, Supervision, Writing – review & editing.

Funding statement: Authors would like to appreciate the funding support of IDB–McGill Scholarship, Natural Science and Engineering Research Council (NSERC), Canada, and Fonds de Recherche du Québec Nature et technologies (FRQnt), Government of Quebec, Canada.

Data availability: Data will be made available on request.

Conflict of interest: The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

1. W. Routray, V. Orsat, Plant by-products and food industry waste: A source of nutraceuticals and biopolymers, in Food bioconversion. 2017, Elsevier. p. 279-315.

- 2. M.V. Vilariño, C. Franco, C. Quarrington, *Food loss and waste reduction as an integral part of a circular economy.* Frontiers in environmental science, 2017. 5: p. 21.
- 3. M. Selvamuthukumaran, J. Shi, *Recent advances in extraction of antioxidants from plant by-products processing industries.* Food Quality and Safety, 2017. 1(1): p. 61-81.
- 4. N. Tripathi, C.D. Hills, R.S. Singh, C.J. Atkinson, *Biomass waste utilisation in low-carbon products: harnessing a major potential resource*. NPJ climate and atmospheric science, 2019. 2(1): p. 1-10.
- 5. Pagano, L. Campone, R. Celano, A.L. Piccinelli, L. Rastrelli, *Green non-conventional techniques for the extraction of polyphenols from agricultural food by-products: a review.* Journal of Chromatography A, 2021. 1651: p. 462295.
- 6. UNEP, *Technologies for Converting Waste Agricultural Biomass to Energy*. 2013, United Nations Environmental Programme, Division of Technology, Industry and Economics, International Environmental Technology Centre: Osaka, Japan. p. 1-229.
- 7. G.J. Kaur, V. Orsat, A. Singh, *Sustainable usage of carrot discards in food processing*. International Journal of Sustainable Development & World Ecology, 2022. 29(1): p. 18-26.
- 8. Schieber, F.C. Stintzing, R. Carle, *By-products of plant food processing as a source of functional compounds—recent developments*. Trends in Food Science & Technology, 2001. 12(11): p. 401-413.
- 9. J. Ayala-Zavala, C. Rosas-Domínguez, V. Vega-Vega, G. González-Aguilar, *Antioxidant enrichment and antimicrobial protection of fresh-cut fruits using their own byproducts: Looking for integral exploitation.* Journal of food science, 2010. 75(8): p. R175-R181.
- 10. B. Li, B. Smith, M.M. Hossain, *Extraction of phenolics from citrus peels: II. Enzyme-assisted extraction method.* Separation and Purification Technology, 2006. 48(2): p. 189-196.
- 11. C.M. Ajila, S.K. Brar, M. Verma, R.D. Tyagi, S. Godbout, J.R. Valero, *Extraction and analysis of polyphenols: recent trends*. Crit Rev Biotechnol, 2011. 31(3): p. 227-49.
- 12. J. Madureira, L. Barros, S. Cabo Verde, F.M.A. Margaça, C. Santos-Buelga, I.C.F.R. Ferreira, *Ionizing Radiation Technologies to Increase the Extraction of Bioactive Compounds from Agro-Industrial Residues: A Review.*Journal of Agricultural and Food Chemistry, 2020. 68(40): p. 11054-11067.
- 13. J. Giacometti, D.B. Kovacevic, P. Putnik, D. Gabric, T. Bilusic, G. Kresic, V. Stulic, F.J. Barba, F. Chemat, G. Barbosa-Canovas, A.R. Jambrak, *Extraction of bioactive compounds and essential oils from mediterranean herbs by conventional and green innovative techniques: A review.* Food Research International, 2018. 113: p. 245-262.
- 14. Y.X. Wang, H. Zhang, Advances in the extraction, purification, structural-property relationships and bioactive molecular mechanism of Flammulina velutipes polysaccharides: A review. International Journal of Biological Macromolecules, 2021. 167: p. 528-538.
- 15. J.K. Patra, G. Das, S. Lee, S.S. Kang, H.S. Shin, Selected commercial plants: A review of extraction and isolation of bioactive compounds and their pharmacological market value. Trends in Food Science & Technology, 2018. 82: p. 89-109.
- 16. J.X. Zhang, C.T. Wen, H.H. Zhang, Y.Q. Duan, H.L. Ma, Recent advances in the extraction of bioactive compounds with subcritical water: A review. Trends in Food Science & Technology, 2020. 95: p. 183-195.
- 17. Gligor, A. Mocan, C. Moldovan, M. Locatelli, G. Crisan, I. Ferreira, *Enzyme-assisted extractions of polyphenols A comprehensive review*. Trends in Food Science & Technology, 2019. 88: p. 302-315.
- 18. M. Banozic, J. Babic, S. Jokic, *Recent advances in extraction of bioactive compounds from tobacco industrial waste-a review*. Industrial Crops and Products, 2020. 144.
- 19. Z.Y. Wang, S.Y. Li, S.H. Ge, S.L. Lin, *Review of Distribution, Extraction Methods, and Health Benefits of Bound Phenolics in Food Plants*. Journal of Agricultural and Food Chemistry, 2020. 68(11): p. 3330-3343.
- 20. GrandViewResearch, Polyphenols Market Size, Share & Trends Analysis Report By Product (Grape Seed, Green Tea, Cocoa), By Application (Beverages, Food, Feed, Dietary Supplements, Cosmetics), And Segment Forecasts, 2023 2030. 2023: California, United States. p. 125.
- 21. W. Routray, V. Orsat, *MAE of phenolic compounds from blueberry leaves and comparison with other extraction methods.* Industrial Crops and Products, 2014. 58: p. 36-45.
- 22. S. Dragović, V. Dragović-Uzelac, S. Pedisić, Z. Čošić, M. Friščić, I.E. Garofulić, Z. Zorić, *The mastic tree* (Pistacia lentiscus l.) leaves as source of BACs: Effect of growing location, phenological stage and extraction solvent on phenolic content. Food Technology and Biotechnology, 2020. 58(3): p. 303-313.
- 23. C. Rodriguez-Perez, R. Quirantes-Pine, A. Fernandez-Gutierrez, A. Segura-Carretero, *Optimization of extraction method to obtain a phenolic compounds-rich extract from Moringa oleifera Lam leaves*. Industrial Crops and Products, 2015. 66: p. 246-254.

- 24. P. Sucharitha, S.V. Satyanarayana, K.B. Reddy, Pretreatment and Optimization of Processing Conditions for Extraction of Oleuropein from Olive Leaves using Central Composite Design. Pharmacognosy Research, 2019. 11(2): p. 178-187.
- 25. L. Chotphruethipong, S. Benjakul, K. Kijroongrojana, Optimization of extraction of antioxidative phenolic compounds from cashew (Anacardium occidentale L.) leaves using response surface methodology. Journal of Food Biochemistry, 2017. 41(4).
- 26. H.K. Kala, R. Mehta, K.K. Sen, R. Tandey, V. Mandal, *Critical analysis of research trends and issues in microwave assisted extraction of phenolics: Have we really done enough.* Trac-Trends in Analytical Chemistry, 2016. 85: p. 140-152.
- 27. N. Yeasmen, V. Orsat, Advanced separation of phytochemical and bioactive attributes from various morphological components of sugar maple (Acer saccharum) tree. Food Bioscience, 2023. 56: p. 103260.
- 28. B.R. Albuquerque, S.A. Heleno, M.B.P. Oliveira, L. Barros, I.C. Ferreira, *Phenolic compounds: Current industrial applications, limitations and future challenges.* Food & Function, 2021. 12(1): p. 14-29.
- 29. L.P. Mallmann, B. Tischer, M. Vizzotto, E. Rodrigues, V. Manfroi, Comprehensive identification and quantification of unexploited phenolic compounds from red and yellow araçá (Psidium cattleianum Sabine) by LC-DAD-ESI-MS/MS. Food Research International, 2020. 131: p. 108978.
- 30. Ogori, *Plant toxins*. Ame J Biomed Sci Res, 2019. 4(3): p. 173-175.
- 31. S. Patel, M.K. Nag, S. Daharwal, M.R. Singh, D. Singh, *Plant toxins: an overview*. Research Journal of Pharmacology and Pharmacodynamics, 2013. 5(5): p. 283-288.
- 32. M. Wink, Mode of action and toxicology of plant toxins and poisonous plants. Julius-Kühn-Archiv, 2010(421): p. 93.
- 33. X. Jin, C. Shi, C.Y. Yu, T. Yamada, E.J. Sacks, *Determination of Leaf Water Content by Visible and Near-Infrared Spectrometry and Multivariate Calibration in Miscanthus*. Frontiers in Plant Science, 2017. 8.
- 34. W. Routray, V. Orsat, Y. Gariepy, Effect of Different Drying Methods on the Microwave Extraction of Phenolic Components and Antioxidant Activity of Highbush Blueberry Leaves. Drying Technology, 2014. 32(16): p. 1888-1904.
- 35. M. Saifullah, R. McCullum, A. McCluskey, Q. Vuong, Effects of different drying methods on extractable phenolic compounds and antioxidant properties from lemon myrtle dried leaves. Heliyon, 2019. 5(12): p. e03044.
- 36. N. Harnkarnsujarit, K. Kawai, M. Watanabe, T. Suzuki, Effects of freezing on microstructure and rehydration properties of freeze-dried soybean curd. Journal of Food Engineering, 2016. 184: p. 10-20.
- 37. V.P. Oikonomopoulou, M.K. Krokida, V.T. Karathanos, The influence of freeze drying conditions on microstructural changes of food products. Procedia Food Science, 2011. 1: p. 647-654.
- 38. N. Yeasmen, V. Orsat, *Maximization of the recovery of phenolic compounds from sugar maple leaves*. Biomass Conversion and Biorefinery, 2022: p. 1-16.
- 39. N. Yeasmen, V. Orsat, *Phenolic mapping and associated antioxidant activities through the annual growth cycle of sugar maple leaves.* Food Chemistry, 2023: p. 136882.
- 40. M. Saifullah, R. McCullum, A. McCluskey, Q. Vuong, Effects of different drying methods on extractable phenolic compounds and antioxidant properties from lemon myrtle dried leaves. Heliyon, 2019. 5(12).
- 41. L.M. Garcia, C. Ceccanti, C. Negro, L. De Bellis, L. Incrocci, A. Pardossi, L. Guidi, *Effect of Drying Methods on Phenolic Compounds and Antioxidant Activity of Urtica dioica L. Leaves.* Horticulturae, 2021. 7(1).
- 42. M. Saifullah, R. McCullum, A. McCluskey, V.V. Quan, Effect of drying techniques and operating conditions on the retention of color, phenolics, and antioxidant properties in dried lemon scented tea tree (Leptospermum petersonii) leaves. Journal of Food Processing and Preservation, 2021. 45(3).
- 43. Q.M. Cui, Y.D. Wang, W.B. Zhou, S.Y. He, M.L. Yang, Q.W. Xue, Y.F. Wang, T.R. Zhao, J.X. Cao, A. Khan, G.G. Cheng, *Phenolic composition, antioxidant and cytoprotective effects of aqueous-methanol extract from Anneslea fragrans leaves as affected by drying methods.* International Journal of Food Science and Technology, 2021. 56(9): p. 4807-4819.
- 44. F.M. Mashitoa, T. Shoko, J.L. Shai, R.M. Slabbert, Y. Sultanbawa, D. Sivakumar, *Influence of Different Types of Drying Methods on Color Properties, Phenolic Metabolites and Bioactivities of Pumpkin Leaves of var. Butternut squash (Cucurbita moschata Duchesne ex Poir)*. Frontiers in Nutrition, 2021. 8.
- 45. K. Ravichai, R. Muangrat, Effect of different coating materials on freeze-drying encapsulation of bioactive compounds from fermented tea leaf wastewater. Journal of Food Processing and Preservation, 2019. 43(10).

- 46. M.H. Ahmad-Qasem, E. Barrajon-Catalan, V. Micol, A. Mulet, J.V. Garcia-Perez, Influence of freezing and dehydration of olive leaves (var. Serrana) on extract composition and antioxidant potential. Food Research International, 2013. 50(1): p. 189-196.
- 47. N. Hidar, A. Noufid, A. Mourjan, E. El Adnany, S. Mghazli, M. Mouhib, A. Jaouad, M. Mahrouz, Effect of Preservation Methods on Physicochemical Quality, Phenolic Content, and Antioxidant Activity of Stevia Leaves. Journal of Food Quality, 2021. 2021.
- 48. K. Mouhoubi, L. Boulekbache-Makhlouf, K. Madani, A. Palatzidi, J. Perez-Jimenez, I. Mateos-Aparicio, A. Garcia-Alonso, *Phenolic compounds and antioxidant activity are differentially affected by drying processes in celery, coriander and parsley leaves.* International Journal of Food Science and Technology, 2022.
- 49. Snoussi, I. Essaidi, H.B. Koubaier, H. Zrelli, I. Alsafari, T. Zivoslav, J. Mihailovic, M. Khan, A. El Omri, T.C. Velickovic, N. Bouzouita, *Drying methodology effect on the phenolic content, antioxidant activity of Myrtus communis L. leaves ethanol extracts and soybean oil oxidative stability.* Bmc Chemistry, 2021. 15(1).
- 50. K. Mouhoubi, L. Boulekbache-Makhlouf, W. Mehaba, H. Himed-Idir, K. Madani, Convective and microwave drying of coriander leaves: Kinetics characteristics and modeling, phenolic contents, antioxidant activity, and principal component analysis. Journal of Food Process Engineering, 2022. 45(1).
- 51. Q.R. Hu, P.W. Lai, F. Chen, Y.F. Yu, B. Zhang, H.Y. Li, R. Liu, Y.W. Fan, Z.Y. Deng, Whole mulberry leaves as a promising functional food: From the alteration of phenolic compounds during spray drying and in vitro digestion. Journal of Food Science, 2022. 87(3): p. 1230-1243.
- 52. AAFC, Statistical Overview of the Canadian Maple Industry 2020, A.a.A.-F.C. Crops and Horticulture and Division, Editor. 2021, Government of Canada: Canada. p. 1-16.
- 53. T. Belwal, A. Pandey, I.D. Bhatt, R.S. Rawal, Z.S. Luo, Trends of polyphenolics and anthocyanins accumulation along ripening stages of wild edible fruits of Indian Himalayan region. Scientific Reports, 2019. 9.
- 54. K. Ameer, H.M. Shahbaz, J.H. Kwon, *Green Extraction Methods for Polyphenols from Plant Matrices and Their Byproducts: A Review*. Comprehensive Reviews in Food Science and Food Safety, 2017. 16(2): p. 295-315.
- 55. TransparencyMarketResearch. Polyphenols Market by Product (Grape seed, Green tea, Apple and Others), by Application (Functional beverages, Functional food, Dietary supplements and Others) Global Industry Analysis, Size, Share, Growth, Trends and Forecast, 2012 2018. 2016 [cited 2021 25 February]; Available from: https://www.transparencymarketresearch.com/polyphenol-market.html.
- 56. N. Yeasmen, V. Orsat, *Green extraction and characterization of leaves phenolic compounds: a comprehensive review.* Critical Reviews in Food Science and Nutrition, 2021: p. 1-39.
- 57. W. Routray, B. Jena, V. Orsat, *Recent advances in extraction, isolation, characterization, and applications of phenolic compounds*. Studies in Natural Products Chemistry, 2022. 72: p. 29-55.
- 58. HealthCanada, *Nutraceuticals/Functional Foods and Health Claims On Foods*, H. Canada, Editor. 2002, Government of Canada: Canada. p. 1-29.
- 59. D. Bosscher, M. Van Caillie-Bertrand, R. Van Cauwenbergh, H. Deelstra, *Availabilities of calcium, iron, and zinc from dairy infant formulas is affected by soluble dietary fibers and modified starch fractions.* Nutrition, 2003. 19(7-8): p. 641-645.
- 60. H.M. Yehia, M.F.S. El-Din, H.S.M. Ali, M.S. Alamri, W.A. Al-Megrin, M.F. Elkhadragy, M.A.G. Awad, *Synthesis of black eggplant (Solanum melongena) skin antioxidant nanoparticles*. 2019, Google Patents.
- 61. Selivanova, R. Terekhov, *Crystal engineering as a scientific basis for modification of physicochemical properties of bioflavonoids*. Russian Chemical Bulletin, 2019. 68(12): p. 2155-2162.
- 62. H. Ma, J.L. Xu, N. DaSilva, L. Wang, Z.X. Wei, L.R. Guo, S.L. Johnson, W. Lu, J. Xu, Q. Gu, N.P. Seeram, Cosmetic applications of glucitol-core containing gallotannins from a proprietary phenolic-enriched red maple (Acer rubrum) leaves extract: inhibition of melanogenesis via down-regulation of tyrosinase and melanogenic gene expression in B16F10 melanoma cells. Archives of Dermatological Research, 2017. 309(4): p. 265-274.
- 63. A.L. Oliveira, S. Gondim, R. Gómez-García, T. Ribeiro, M. Pintado, *Olive leaf phenolic extract from two Portuguese cultivars–bioactivities for potential food and cosmetic application.* Journal of Environmental Chemical Engineering, 2021. 9(5): p. 106175.
- 64. EU, Regulation (EC) No 1333/2008 of the European Parliament and of the Council of 16 December 2008 on food additives. Official Journal of the European Union, 2008. L354: p. 16-33.
- 65. EU, Commission Regulation (EU) No 1129/2011 of 11 November 2011 amending Annex II to Regulation (EC) No 1333/2008 of the European Parliament and of the Council by establishing a Union list of food additives. Official Journal of the European Union, 2011. L295: p. 1-211.

- 66. EU, Commission Regulation (EU) No 1131/2011 of 11 November 2011 amending Annex II to Regulation (EC) No 1333/2008 of the European Parliament and of the Council with regard to steviol glycosides. Official Journal of the European Union, 2011. L295: p. 205-211.
- 67. A.I. Kalogianni, T. Lazou, I. Bossis, A.I. Gelasakis, Natural Phenolic Compounds for the Control of Oxidation, Bacterial Spoilage, and Foodborne Pathogens in Meat. Foods, 2020. 9(6).
- 68. S. Martillanes, J. Rocha-Pimienta, M. Cabrera-Bañegil, D. Martín-Vertedor, J. Delgado-Adámez, Application of Phenolic Compounds for Food Preservation: Food Additive and Active Packaging, in Phenolic Compounds Biological Activity, M. Soto-Hernandez, M. Palma-Tenango, M.D.R. Garcia-Mateos, Editors. 2017, IntechOpen.
- 69. S.C. Lourenço, M. Moldão-Martins, V.D. Alves, *Antioxidants of Natural Plant Origins: From Sources to Food Industry Applications*. Molecules (Basel, Switzerland), 2019. 24(22): p. 4132.
- 70. D. Pasrija, P.N. Ezhilarasi, D. Indrani, C. Anandharamakrishnan, *Microencapsulation of green tea polyphenols and its effect on incorporated bread quality*. LWT Food Science and Technology, 2015. 64(1): p. 289-296.
- 71. F. Shahidi, Y. Zhong, Measurement of antioxidant activity. Journal of Functional Foods, 2015. 18: p. 757-781.
- 72. F. Shahidi, P. Ambigaipalan, *Phenolics and polyphenolics in foods, beverages and spices: Antioxidant activity and health effects A review.* Journal of Functional Foods, 2015. 18: p. 820-897.
- 73. Christ-Ribeiro, C.C. Bretanha, G.G. Luz, M.M. de Souza, E. Badiale-Furlong, < b> Antifungal compounds extracted from rice bran fermentation applied to bakery product conservation. Acta Scientiarum. Technology, 2017. 39(3): p. 263-268.
- 74. Christ-Ribeiro, C. Graça, L. Kupski, E. Badiale-Furlong, L. de Souza-Soares, *Cytotoxicity, antifungal and anti mycotoxins effects of phenolic compounds from fermented rice bran and Spirulina sp.* Process Biochemistry, 2019. 80: p. 190-196.
- 75. S. Ab Rashid, W.Y. Tong, C.R. Leong, N.M. Abdul Ghazali, M.A. Taher, N. Ahmad, W.-N. Tan, S.H. Teo, *Anthocyanin microcapsule from Clitoria ternatea: Potential bio-preservative and blue colorant for baked food products.* Arabian Journal for Science and Engineering, 2021. 46(1): p. 65-72.
- 76. U. Siripatrawan, S. Noipha, *Active film from chitosan incorporating green tea extract for shelf life extension of pork sausages*. Food Hydrocolloids, 2012. 27(1): p. 102-108.
- 77. T. Mehdizadeh, H. Tajik, A.M. Langroodi, R. Molaei, A. Mahmoudian, *Chitosan-starch film containing pomegranate peel extract and Thymus kotschyanus essential oil can prolong the shelf life of beef.* Meat science, 2020. 163: p. 108073.
- 78. M. Yaghoubi, K. Alirezalu, S.M. Mazloomi, M. Marcinkowska-Lesiak, S. Azadmard-Damirchi, S.H. Peighambardoust, J. Hesari, A. Rastgoo, Y. Phimolsiripol, A.M. Khaneghah, Enhancing beef sausage packaging with calcium alginate active film infused with nisin and ε-polylysine nanoparticles and beetroot extract. LWT, 2023: p. 115665.
- 79. López-de-Dicastillo, J.M. Alonso, R. Catalá, R. Gavara, P. Hernández-Muñoz, *Improving the Antioxidant Protection of Packaged Food by Incorporating Natural Flavonoids into Ethylene–Vinyl Alcohol Copolymer (EVOH) Films.* Journal of Agricultural and Food Chemistry, 2010. 58(20): p. 10958-10964.
- 80. S. Bilgiç, E. Söğüt, A.C. Seydim, *Chitosan and starch based intelligent films with anthocyanins from eggplant to monitor pH variations*. Turkish Journal of Agriculture-Food Science and Technology, 2019. 7(sp1): p. 61-66.
- 81. P. Zeng, X. Chen, Y.-R. Qin, Y.-H. Zhang, X.-P. Wang, J.-Y. Wang, Z.-X. Ning, Q.-J. Ruan, Y.-S. Zhang, Preparation and characterization of a novel colorimetric indicator film based on gelatin/polyvinyl alcohol incorporating mulberry anthocyanin extracts for monitoring fish freshness. Food Research International, 2019. 126: p. 108604.
- 82. G.S. da Rosa, S.K. Vanga, Y. Gariepy, V. Raghavan, Development of Biodegradable Films with Improved Antioxidant Properties Based on the Addition of Carrageenan Containing Olive Leaf Extract for Food Packaging Applications. Journal of Polymers and the Environment, 2020. 28(1): p. 123-130.
- 83. N. Mulinacci, M. Innocenti, M. Bellumori, C. Giaccherini, V. Martini, M. Michelozzi, *Storage method, drying processes and extraction procedures strongly affect the phenolic fraction of rosemary leaves: An HPLC/DAD/MS study.* Talanta, 2011. 85(1): p. 167-176.
- 84. K. Tzima, N.P. Brunton, J.G. Lyng, D. Frontuto, D.K. Rai, *The effect of Pulsed Electric Field as a pre-treatment step in Ultrasound Assisted Extraction of phenolic compounds from fresh rosemary and thyme by-products.* Innovative Food Science & Emerging Technologies, 2021. 69.

- 85. Caleja, L. Barros, A.L. Antonio, M. Carocho, M.B.P. Oliveira, I.C. Ferreira, Fortification of yogurts with different antioxidant preservatives: A comparative study between natural and synthetic additives. Food chemistry, 2016. 210: p. 262-268.
- 86. A.K. Das, V. Rajkumar, P.K. Nanda, P. Chauhan, S.R. Pradhan, S. Biswas, *Antioxidant efficacy of litchi (Litchi chinensis Sonn.) pericarp extract in sheep meat nuggets*. Antioxidants, 2016. 5(2): p. 16.
- 87. Ribeiro, C. Caleja, L. Barros, C. Santos-Buelga, M.F. Barreiro, I.C. Ferreira, Rosemary extracts in functional foods: Extraction, chemical characterization and incorporation of free and microencapsulated forms in cottage cheese. Food & function, 2016. 7(5): p. 2185-2196.
- 88. M. Çam, N.C. İçyer, F. Erdoğan, *Pomegranate peel phenolics: Microencapsulation, storage stability and potential ingredient for functional food development.* LWT-Food Science and Technology, 2014. 55(1): p. 117-123.
- 89. D.S. Pillai, P. Prabhasankar, B. Jena, C. Anandharamakrishnan, Microencapsulation of Garcinia cowa fruit extract and effect of its use on pasta process and quality. International Journal of Food Properties, 2012. 15(3): p. 590-604.
- 90. E.M.F. Lima, M.C.M. Madalão, W.C. dos Santos, P.C. Bernardes, S.H. Saraiva, P.I. Silva, *Spray-dried microcapsules of anthocyanin-rich extracts from Euterpe edulis M. as an alternative for maintaining color and bioactive compounds in dairy beverages*. Journal of Food Science and Technology, 2019. 56(9): p. 4147-4157.
- 91. Y. Aguilera, L. Mojica, M. Rebollo-Hernanz, M. Berhow, E.G. De Mejía, M.A. Martín-Cabrejas, *Black bean coats: New source of anthocyanins stabilized by β-cyclodextrin copigmentation in a sport beverage.* Food chemistry, 2016. 212: p. 561-570.
- 92. J.C. Baldin, E.C. Michelin, Y.J. Polizer, I. Rodrigues, S.H.S. de Godoy, R.P. Fregonesi, M.A. Pires, L.T. Carvalho, C.S. Favaro-Trindade, C.G. de Lima, *Microencapsulated jabuticaba (Myrciaria cauliflora) extract added to fresh sausage as natural dye with antioxidant and antimicrobial activity.* Meat Science, 2016. 118: p. 15-21.
- 93. T.L. Swer, K. Chauhan, C. Mukhim, K. Bashir, A. Kumar, *Application of anthocyanins extracted from Sohiong* (*Prunus nepalensis L.*) in food processing. Lwt, 2019. 114: p. 108360.
- 94. Rashidinejad, E.J. Birch, D. Sun-Waterhouse, D.W. Everett, *Effect of liposomal encapsulation on the recovery and antioxidant properties of green tea catechins incorporated into a hard low-fat cheese following* in vitro *simulated gastrointestinal digestion*. Food and bioproducts processing, 2016. 100: p. 238-245.
- 95. Ajila, M. Aalami, K. Leelavathi, U.P. Rao, *Mango peel powder: A potential source of antioxidant and dietary fiber in macaroni preparations.* Innovative Food Science & Emerging Technologies, 2010. 11(1): p. 219-224.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.