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Case Report

Biologic Response to Crest to Restoration Distance in Subcrestally Placed Implants: A Case Report

Chiyun Won

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Abstract: Background: Subcrestally placed implants (SPI) offer biological and mechanical advantages, but their deeper positioning requires careful management of the peri-implant soft tissue interface, particularly the Crest to Restoration Distance (CRD). This vertical space, extending from the crestal bone to the implant restoration, plays a crucial role in maintaining peri-implant health. If CRD exceeds an optimal range, void formation can occur, increasing the risk of bacterial colonization, peri-implant mucositis, and eventual peri-implantitis. Optimizing CRD, including soft tissue thickness (STT), is essential for preserving peri-implant stability. **Objective:** This study aims to evaluate the influence of CRD optimization in peri-implant health by comparing two subcrestally placed implants in a split-mouth case. It further explores the structural role of peri-implant soft tissue, particularly the Transitional Zone (TZ) and Subcrestal Zone (SZ), in maintaining a stable biologic seal. Materials and Methods: Study Design: A split-mouth analysis involving two subcrestally placed implants (upper left and lower right first molars) in a 57-year-old female patient. 3D Analysis: Cone Beam Computed Tomography (CBCT) was utilized to measure CRD, Depth of Placement (DP), and Crest Restoration Length (CRL). Clinical Evaluation: Periodic assessments included Implant Paper Point Probing (IPPP), Bleeding on Probing (BOP), and other peri-implant health indicators. Intervention: A restoration adjustment was performed to optimize CRD at the test site (upper left implant). Results: Test Site (Upper Left Implant): Before optimization: Excessive CRD, deep probing depths (>3 mm), and clinical signs of mucositis (BOP, redness, swelling). After optimization: Adjusted CRD resolved mucositis symptoms, achieving stable probing depths (<1 mm), no BOP, and healthy peri-implant soft tissue. CBCT confirmed an improved biologic seal. Control Site (Lower Right Implant): Maintained a CRD within the optimal range from the beginning, with consistently healthy peri-implant tissues. No signs of peri-implant disease throughout the observation period. Conclusion: Maintaining an optimal CRD is associated with peri-implant soft tissue stability and may reduce the risk of sulcular epithelial downgrowth and pocket formation, potentially minimizing peri-implant disease. This study underscores the importance of considering 3D peri-implant soft tissue parameters in implant planning, particularly in subcrestally placed implants (SPI). Future research should focus on standardizing CRD guidelines to improve long-term implant outcomes.

Keywords: junctional epithelium; peri-implant soft tissue; implant; subcrestal placement; transitional zone

Introduction

Dental implants are placed in the human body to replicate the function and appearance of natural teeth, offering long-term solutions for missing teeth. Prosthodontic components, such as abutments and crowns, are connected to these implants to restore both functionality and aesthetics [1]. Due to their high predictability, dental implants have become the most recommended and widely practiced treatment option for restoring lost teeth in totally or partially edentulous patients. The discovery of osseointegration—the process by which bone heals around titanium—has been the foundation for the success of modern implant dentistry [2].



There are two critical interfaces between living tissues and artificial components in implant dentistry. The first is the bone-implant interface, known as osseointegration, which is well-established and extensively studied [3]. The second is the peri-implant soft tissue interface, often referred to as the "soft tissue seal" or "mucointegration." Unlike osseointegration, our understanding of this soft tissue interface is still evolving. The term "soft tissue seal" reflects the fact that its characteristics are not yet fully defined or understood [4,5].

When osseointegration lacks adequate protection from a functional soft tissue barrier, external threats—particularly bacterial infiltration—can compromise implant stability and long-term success. This soft tissue barrier plays a pivotal role in preventing peri-implant diseases such as peri-implantitis, which can lead to bone loss and, if untreated, implant failure. Enhancing our understanding of the peri-implant soft tissue interface is essential for improving implant outcomes and developing preventive strategies against peri-implant complications [6].

Despite its importance, there is no unified consensus on the anatomy, histology, physiology, or clinical guidelines for optimizing the soft tissue seal around implants. While interest in this area has grown in recent years, many questions remain unresolved [7–10].

Similar to periodontal diseases—gingivitis and periodontitis—peri-implant diseases include peri-implant mucositis and peri-implantitis. Peri-implant mucositis is limited to the soft tissue, while peri-implantitis involves progressive bone loss [11–13]. These conditions typically begin with disruption of the soft tissue seal, which, if left unchecked, can eventually compromise the osseointegration achieved during the early phase of treatment.

Like periodontal bone loss, peri-implant bone loss can result from bacterial invasion. However, bone loss around implants may also occur due to inadequate soft tissue width, independent of bacterial factors. Linkevicius et al. reported that when vertical soft tissue thickness (STT) at the alveolar crest is 2 mm or less, bone resorption is likely to occur, as the body attempts to establish an adequate biologic width [14]. This adaptive resorption highlights the importance of ensuring a critical soft tissue thickness to maintain peri-implant health and minimize crestal bone loss.

To address the inherent weakness of peri-implant soft tissue in forming a biological seal, various strategies have been explored:

- Optimizing abutment surface characteristics to enhance tissue attachment [15,16].
- Modifying prosthodontic components, such as the emergence profile and platform-switching, to improve soft tissue stability [17,18].
- Adjusting the implant placement depth to strengthen the soft tissue's protective barrier [19].

Among these approaches, subcrestal implant placement (SPI) has gained attention as a method for improving peri-implant soft tissue adaptation. Vervaeke et al. described SPI as a technique that promotes bone remodeling and soft tissue stability, particularly in cases with thin peri-implant mucosa [22,23].

Several studies have demonstrated that subcrestally placed implants provide stable peri-implant conditions and minimize crestal bone loss over time. (Figure 1) Bicon implants, widely used during the 2000s and 2010s, were often placed subcrestally, leading to lasting biological stability [24]. SPIs are commonly used in cases with narrow or sloped ridges, where deeper placement ensures that the implant's rough surface is fully embedded in bone. This approach is also beneficial for achieving a natural emergence profile, especially in molar regions. Despite their clinical success, SPI outcomes in the literature have been inconsistent, possibly due to:

- Lack of standardized clinical guidelines.
- Variability in surgical and prosthodontic techniques.
- Limited data on peri-implant soft tissue adaptation [23–25].

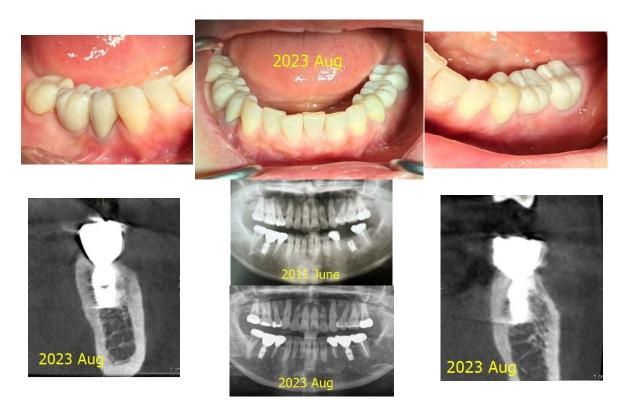


Figure 1. Results after 10 years of subcrestally placed implants (SPI) – Fixtures were placed subcrestally, with X-rays showing no crestal bone loss and clinical photos demonstrating healthy peri-implant soft tissue and a natural emergence profile. This approach reflects a trend seen with Bicon implants during the 2000s to 2010s, where implant fixtures were often placed slightly deeper than those of other systems to support lasting stability.

Most analyses of peri-implant soft tissue have focused on horizontal extensions between the implant restoration and the underlying soft tissue, while vertical soft tissue dimensions have been largely overlooked. The Crest to Restoration Distance (CRD) represents the vertical space between the crestal bone and the implant restoration. This space is occupied by peri-implant soft tissue and is critical for biologic stability.

When CRD is within an optimal range:

- The biologic seal is reinforced, reducing bacterial infiltration.
- Void spaces or pockets are eliminated, lowering the risk of mucositis.
- Soft tissue adaptation is improved, preventing excessive sulcular epithelial downgrowth.
 When CRD is too large
- Soft tissue stability is compromised, leading to pocket formation.
- Bacterial accumulation increases, raising the risk of peri-implant disease.

This case study examines how CRD optimization affects peri-implant soft tissue stability by comparing two implants placed in the same patient:

- Control Implant: Maintained peri-implant health with stable CRD.
- Test Implant: Developed peri-implant mucositis due to excessive CRD.

By modifying CRD through prosthodontic adjustments, this study explores its role in disease prevention and peri-implant soft tissue integrity.

Crest to Restoration Distance (CRD) plays a fundamental role in ensuring peri-implant soft tissue stability, particularly in subcrestally placed implants. This study underscores the importance of CRD optimization as a modifiable factor in implant success and suggests that future research should focus on establishing standardized CRD guidelines to improve clinical protocols and long-term outcomes.

Case Presentation

A 57-year-old woman underwent extraction of her lower right first molar on June 3, 2020, due to a severe periodontal abscess. Later, on September 22, 2020, her upper left first molar was extracted due to a fracture involving both the crown and root. (Figure 2)





Figure 2. Initial radiograph and clinical image of the upper left first molar on the patient's first visit (September 22, 2020). The radiograph illustrates the condition of the tooth and surrounding structures. The clinical image provides a clear view of the crown and root fracture prior to extraction.

On October 29, 2020, an implant (Oneplant, Warrentec, manufactured in Seoul, South Korea) was placed at the upper left first molar site. The procedure included a sinus lift using the transcrestal approach along with bone grafting surgery. (Figure 3)

A second implant (Oneplant, Warrentec) was placed at the lower right first molar site on December 21, 2020.

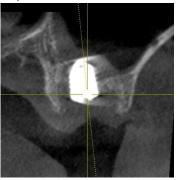




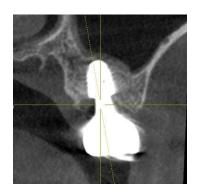
Figure 3. October 29, 2020 – Implant placement at the upper left first molar site with sinus lift and bone grafting surgery.

The second operation to connect the healing abutments on both sides was performed on March 3, 2021. Healing abutments with a diameter of 6.0 mm and a height of 5.0 mm were placed at each site.

The final implant restoration for the lower right molar was completed on March 18, 2021, using a 4.5 mm diameter, 3 mm gingival cuff abutment.

For the upper left molar, the restoration was completed on March 25, 2021, with a 4.5 mm diameter, 4 mm gingival cuff abutment.

Zirconia crowns were cemented onto the prefabricated abutments (manufactured by Warrentec) using the extraoral cementation technique. (Figure 4)



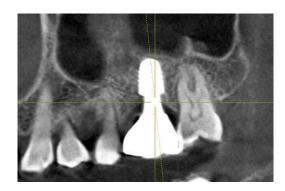


Figure 4. March 25, 2021 – Restoration completed using a 4 mm gingival cuff abutment.

She was provided with instructions on implant care, including thorough cleaning using proximal brushes and water irrigation devices.

Until October 17, 2022, the patient had not exhibited any signs or symptoms of implant-associated diseases, such as peri-implant mucositis or peri-implantitis, with the implant functioning well during routine check-ups for approximately 19 months. However, starting from that date, she reported discomfort in the upper left first molar implant area, including gingival bleeding, swelling, and food impaction.

On clinical examination, the peri-implant soft tissue around the upper left first molar implant exhibited redness and slight swelling; however, radiographic evaluation showed no signs of bone loss. Consequently, the patient was diagnosed with peri-implant mucositis in the upper left first molar implant, while the lower right first molar implant remained healthy.

For the treatment of peri-implant mucositis, the following measures were implemented: reinforcing the patient's oral hygiene around the implant, scheduling more frequent recall appointments for check-ups and saline irrigation around the sulcus, and applying topical antibiotics to the sulcus. However, there was no improvement, and the patient's discomfort due to food impaction persisted. Clinical evaluation revealed that the interproximal contact point was adequately tight, preventing food from becoming lodged between the teeth. The lateral profile of the crown appeared natural, with sufficient space in the embrasure area for cleaning. Despite this, food seemed to accumulate in the sulcus rather than in other areas, such as the interproximal or contact zones. In contrast, the lower right first molar implant remained stable, with no signs or symptoms of peri-implant disease.

For the purpose of analyzing and comparing the peri-implant soft tissue structure and peri-implant bone topography, a CBCT (Cone-Beam Computed Tomography) scan was performed. The analysis specifically focused on the Crest to Restoration Distance (CRD), which represents the vertical distance between the implant restoration and the underlying crestal bone. The results of this analysis are summarized in Table 1. (Table 1)

Table 1. presents the measurements of the Crest to Restoration Distance (CRD), which quantifies the vertical distance between the implant restoration and the crestal bone. The values are divided into central CRD (cCRD) and peripheral CRD (pCRD) to distinguish the dimensional variations along the implant interface. These measurements are taken at both central and peripheral locations, evaluating the mesial (M), distal (D), buccal (B), and lingual (L) aspects around the implants of the upper left first molar and lower first molar. The data were collected from the CBCT scan conducted on October 26, 2021.

	Upper left 1st molar		Lower righ	ıt 1st molar
•	cCRD	pCRD	cCRD	pCRD
M	1.52	1.91	0.36	0.91
D	1.54	1.87	0.78	1.66
В	1.88	2.04	0.3	0.3
L	0.98	2.05	0.6	1.41
Average	1.48	1.97	0.51	1.07

Measurement of Crest to Restoration Distance (CRD)

The Crest to Restoration Distance (CRD) refers to the vertical space between the underlying crestal bone and the overlying implant restoration. To assess the dimensional variations along the implant interface, CRD is measured at two specific locations: centrally (cCRD) and peripherally (pCRD).

The results showed that the CRD for the upper left first molar implant was consistently larger than that for the lower right first molar implant. Specifically, the difference in central CRD (cCRD) measured 0.97 mm (1.48 mm – 0.51 mm), and the difference in peripheral CRD (pCRD) measured 0.90 mm (1.97 mm – 1.07 mm). Since comparable CBCT-based CRD measurements are not available in the existing literature, a clinical study by Won, which analyzed CRD (or Soft Tissue Thickness, STT) in 20 cases of subcrestally placed implants with stable outcomes over more than two years, was used as a reference. This analysis reported an average pCRD of 0.6 mm and an average cCRD of 0.3 mm (Table 2).

Table 2. Summary of CRD measurements in 20 cases of subcrestally placed implants, as analyzed by Won. The average pCRD was 0.6 mm, and the average cCRD was 0.3 mm.

	cCRD				pCR	D		
	cCRD/M	cCRD/D	cCRD/B	cCRD/L	pCRD/M	pCRD/D	pCRD/B	pCRD/L
average	0.4	0.2	0.2	0.2	0.8	0.6	0.4	0.5
	0.3			0.6	1			

The analysis suggested that the increased CRD in the upper left first molar implant may have contributed to the development of peri-implant mucositis. To address this, it was decided to remake the restoration for the upper left implant, aiming to adjust the CRD to the optimal range as indicated by the data. A new restoration was fabricated using a 4.5 mm diameter, 3 mm gingival cuff abutment, and subsequently delivered to the patient (Figure 5).

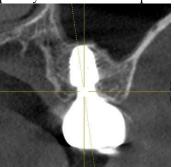




Figure 5. October 31, 2022 The restoration was remade with gingival cuff 3 mm abutment.

Following the restoration remake, the patient reported no further discomfort related to the implant, including gingival bleeding, swelling, or food impaction. Clinical examinations confirmed healthy peri-implant soft tissue, with no signs of radiographic bone resorption. The sulcus was probed using the Implant Paper Point Probe (IPPP), a device specifically designed for probing the peri-implant sulcus and applied with a yield strength of 0.35 N (manufactured by Sure Endo, Seoul, South Korea). The probing depth consistently measured less than 1 mm, with no evidence of bleeding. These findings confirm the successful resolution of peri-implant mucositis.

Figure 6 illustrates the comparison between the two restorations before and after the revision, highlighting differences in their profiles. It also includes a panoramic X-ray taken after the revision procedure and clinical photographs captured on October 31, 2022 following the restoration adjustment. (Figure 6)

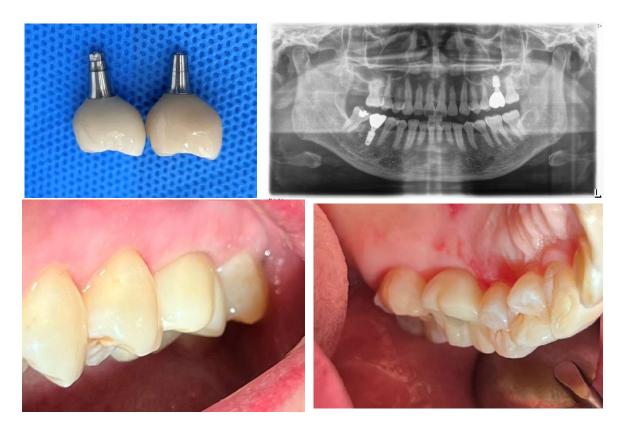


Figure 6. The upper left image presents a comparison of the two restorations before and after the revision, while the upper right image shows a panoramic X-ray taken post-revision. The lower two images display the clinical appearance of the upper left first molar implant restoration following the revision procedure conducted on October 31, 2022.

Radiographic and Clinical Analysis: A Split-Mouth Comparison

These two implant sites serve as a comparison for a split-mouth study. CBCT scans taken in 2021 and 2024 were utilized for a three-dimensional analysis of peri-implant soft tissue and bone structure.

Measurements of CRD changes for both implant sites (control and treatment) were conducted using Cone Beam Computed Tomography (CBCT). The analysis focused on parameters such as Crest to Restoration Distance (CRD), Depth of Placement (DP), and other structural features using CBCT imaging, complemented by clinical assessments. The primary objective was to evaluate how changes in CRD correlate with peri-implant health and disease, particularly in managing peri-implant mucositis.

Radiographic measurements were obtained from panoramic views and CBCT scans using Sirona Dental Systems devices (ORTHOPHOS XG 3D or AXEOS) and software programs (SIDEXIS and GALILEOS Implant Viewer, version 1.9.5605.25519 ID7).

Radiographic Measurements

Crest to Restoration Distance (CRD)

CRD was assessed at two locations:

- Central CRD (cCRD): Measured at the outer edge of the implant fixture diameter, representing
 the vertical distance from the crestal bone to the restoration at the central portion of the implant
 platform.
- Peripheral CRD (pCRD): Measured at the most peripheral part of the crestal bone in the buccolingual direction and at the most coronal point at the midpoint of the mesiodistal crestal bone.

- DP was measured perpendicularly at the same central and peripheral locations as CRD:
- Central DP (cDP): Measured from the implant fixture top (or fixture-abutment connection) to the crestal bone at the outer side of the fixture diameter.
- Peripheral DP (pDP): Measured from the implant fixture top (or fixture-abutment connection)
 to the crestal bone at the most coronal part of the buccal and lingual crestal bone, as well as the
 most coronal or midpoint of the mesial and distal crestal bone between adjacent teeth or
 implants.

Clinical Evaluation

Peri-implant soft tissue health was assessed through the following clinical parameters:

- Implant Paper Point Probing (IPPP): Used to measure probing depths with a standardized method.
- Bleeding on Probing (BOP): Evaluated as an indicator of inflammation.
- Visual Assessment: Conducted to identify redness and swelling as markers of peri-implant tissue health.

Results

The CRG and DP measurements for the upper left first molar implant and the lower right first molar implant were taken in 2021 and 2024. The averages for each measurement were calculated and compared over time. The results are summarized in Tables 3–8. (Table 3, Table 4, Table 5, Table 6, Table 7, Table 8)

A. Control site (the lower right 1st molar) for comparison

a) Changes of CRD

Table 3. Changes in CRD from 2021 to 2024 at the peripheral and central areas for the control site (the lower right first molar implant).

pCRD	2021	2024	changes
M	0.91	0.87	-0.04
D	1.66	1.54	-0.12
В	0.3	0.34	0.04
L	1.41	1.15	-0.26
average	1.07	0.98	-0.10
cCRD	2021	2024	changes
M	0.36	0.38	0.02
D	0.78	0.61	-0.17
В	0.3	0.21	-0.09
L	0.6	0.51	-0.09
average	0.51	0.43	-0.08

b) Changes of Depth of Placement (DP)

Table 4. Changes in Depth of Placement (DP) from 2021 to 2024 at the peripheral and central areas for the control Site (the lower right first molar implant) The DP changes over time indicate variations in crestal bone level, signifying potential bone loss or gain around the implant.

pDP	2021	2024	changes
M	3.78	3.8	0.02
D	3.5	3.56	0.06
В	3.71	3.67	-0.04
L	2.61	3.15	0.54
average	3.4	3.55	0.15

cDP	2021	2024	changes
M	2.79	2.81	0.02
D	2.65	2.65	0
В	2.71	2.73	0.02
L	2.31	2.31	0
average	2.62	2.63	0.01

c) The clinical parameters for the soft tissue state

Table 5. Clinical parameters for soft tissue condition at the lower right first molar implant. This table presents the clinical assessment of soft tissue health around the lower right first molar implant, including Implant Paper Point Probing (IPPP), Bleeding on Probing (BOP), redness, and swelling, demonstrating a healthy soft tissue state.

Depth of Probing	ВОР	redness	swelling
Less than 1 mm	no	no	no

B. Treatment site (the upper left 1st molar) for peri-implant mucositis

a) Changes of CRD

Table 6. Changes in CRD from 2021 to 2024 at the peripheral and central areas for the treatment site (the upper left first molar implant).

pCRD	2021	2024	changes
M	1.91	0.58	-1.33
D	1.87	1.2	-0.67
В	2.04	0.76	-1.28
L	2.05	0.99	-1.06
average	1.97	0.88	-1.09
cCRD	2021	2024	changes
M	1.52	0.62	-0.90
D	1.54	0.5	-1.04
В	1.88	0.58	-1.3
L	0.98	0.5	-0.48
average	1.48	0.55	-0.93

b) Changes of Depth of Placement (DP)

Table 7. Changes in Depth of Placement (DP) from 2021 to 2024 at the peripheral and central areas for the treatment site (the upper left first molar implant) The DP changes over time indicate variations in crestal bone level, signifying potential bone loss or gain around the implant.

pDP	2021	2024	changes
M	6.32	6.53	0.21
D	4.00	4.06	0.06
В	4.31	5.03	0.72
L	1.61	2.79	1.18
average	4.06	4.60	0.54
cDP	2021	2024	changes
M	2.79	2.86	0.07
D	2.66	2.69	0.03
В	2.23	3.57	1.34
L	2.53	2.43	-0.1

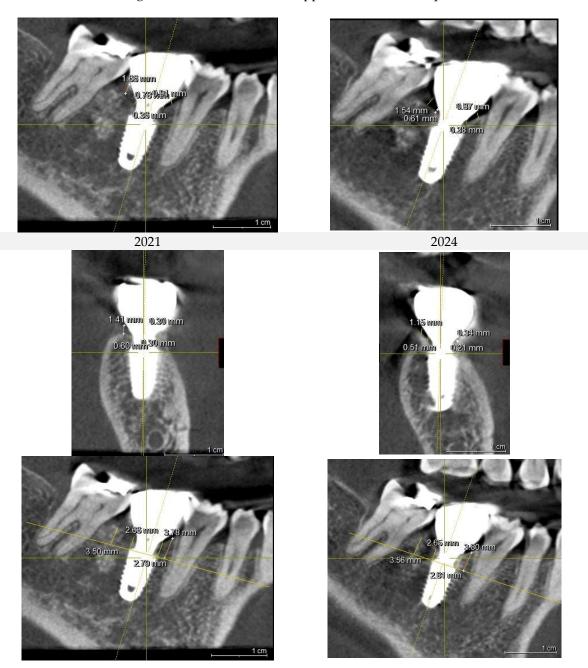
21/2#2@2	2.55	2 80	0.24
average	2.55	2.09	0.34

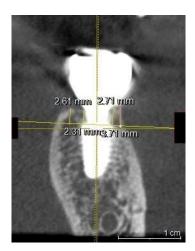
c) The probing depth with IPPP (Implant paper probing point)

Table 8. Clinical parameters for soft tissue condition at the upper left first molar implant. This table presents the clinical assessment of soft tissue health around the upper left first molar implant, including Implant Paper Point Probing (IPPP), Bleeding on Probing (BOP), redness, and swelling, demonstrating a healthy soft tissue state after revision.

Depth of Probing	ВОР	redness	swelling
Less than 1 mm	no	no	no

Figure 7 and Figure 8 illustrate the radiographic changes observed from 2021 to 2024 at the lower right first molar implant site (control site) and the upper left first molar implant site (experimental site), respectively. (Figure 7, Figure 8) The clinical photographs taken in 2024 (Figure 9) depict the final results, showcasing both a stable and natural appearance for both implant sites.





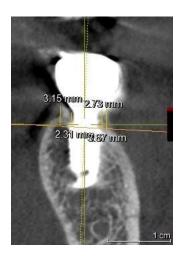
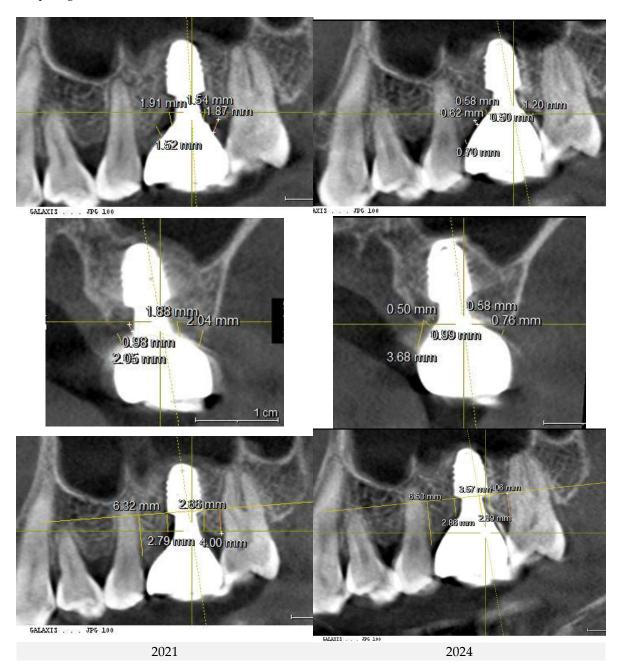
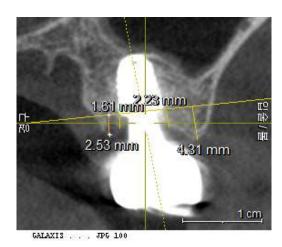


Figure 7. This image illustrates the X-ray measurements conducted on the lower right first molar implant, comparing data from 2021 and 2024.





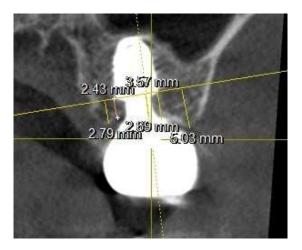


Figure 8. This image illustrates the X-ray measurements conducted on the upper left first molar implant, comparing data from 2021 and 2024..





Figure 9. Clinical images of control and experimental sites post-revision.

This figure presents clinical photographs of the control and experimental implant sites following the revision procedure. Both sites exhibit a natural and healthy appearance, indicating successful soft tissue adaptation and restoration aesthetics, taken at 2024.

Summary of Results

The upper left molar showed notable changes in soft tissue thickness and peri-implant mucositis, while the lower right molar demonstrated consistent CRD. (Table 9)

Table 9. Summary of the comparison for the average changes between the control site (lower right molar) and the treatment site (upper left molar) for CRD.

	Control site	Treated site
The average change in pCRD	-0.10	-1.09
The average change in cCRD	-0.08	-0.93

After revising the prosthodontic components to reduce CRD, the upper left implant showed clinical resolution of mucositis, with no BOP and stable probing depths of less than 1 mm.

The post-revision evaluation provides significant insights into the structural changes in perimplant soft tissue and their impact on implant health. Figure 10 compares X-rays from October 2021 and July 2024, illustrating a marked decrease in the Crest to Restoration Distance (CRD) following the restoration modification. (Figure 10) After the reduction achieved by modifying the implant restoration, the gap can now be regarded as entirely filled by peri-implant soft tissue, without voids. From this observation, *it can be suggested that maintaining the CRD within a critical range is*

essential for achieving a stable biological seal, as it ensures the peri-implant soft tissue provides an effective barrier against external irritants and bacterial infiltration.

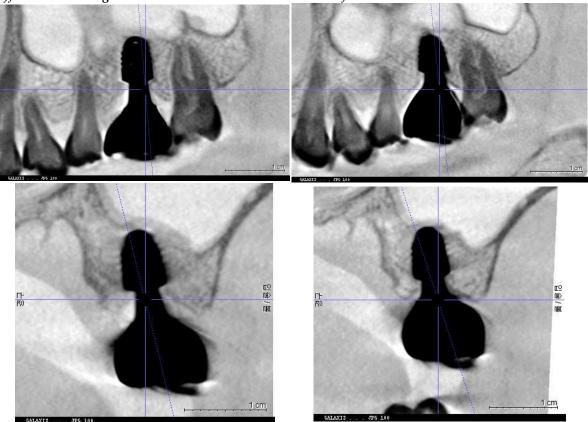


Figure 10. Comparison of X-rays taken in October 2021 and July 2024 after the restoration revision, demonstrating a decrease in the CRD. When the CRD is maintained within the critical range, it can be inferred that the gap space is occupied exclusively by peri-implant soft tissue, without voids, and the GRD can be represented as Soft Tissue Thickness (STT).

These findings emphasize the potential importance of CRD optimization in implant restoration, necessitating histologic evidence supporting its role in promoting a healthy and functional perimplant soft tissue barrier.

Discussions

1. Biological Role of Peri-Implant Soft Tissue

Peri-implant mucositis and peri-implantitis are biological complications with an infectious etiology, primarily caused by the disruption of the soft tissue seal around implants [24]. The peri-implant soft tissue serves a crucial function as a biological barrier, protecting the underlying crestal bone from bacterial infiltration. Linkevicius et al. highlighted the importance of vertical soft tissue thickness in forming a strong biologic seal around implants but did not fully address the three-dimensional architecture of peri-implant soft tissue. This omission raises concerns about the potential for pocket formation, which can lead to bacterial colonization and food accumulation, particularly in subcrestally placed implants (SPI), where deeper positioning can create greater challenges for maintaining a stable soft tissue seal (25, 13).

Bosshardt et al. described the pathogenesis of periodontal pocket formation, emphasizing that the breakdown of the junctional epithelium serves as the initiating event in disease progression. Their study highlighted the critical function of the junctional epithelium as the first line of defense and reinforced the importance of probing as a diagnostic tool [26].

Although acquiring sufficient Supracrestal Tissue Height (STH) is essential for maintaining biologic width and preventing esthetic complications, excessive STH has been linked to pocket formation and an increased risk of peri-implant disease. A balanced STH within the range of 3 mm < STH < 5 mm has been suggested to optimize tissue health [27]. While numerous studies emphasize the vertical component of peri-implant soft tissue as a key determinant of tissue stability, there is a lack of literature addressing the horizontal component of peri-implant soft tissue, which is inevitably generated in SPI cases due to the extended transitional interface between the submucosal portion of the implant restoration and the underlying peri-implant tissue.

This case report suggests that maintaining an optimal CRD plays a pivotal role in stabilizing peri-implant soft tissue architecture within this transitional interface. A properly maintained CRD may limit epithelial downgrowth and prevent deep pocket formation, thus reinforcing the biological seal.

The observed clinical improvement in peri-implant mucositis following CRD optimization suggests a structural transformation in peri-implant soft tissue. Figure 11 presents a schematic representation of this proposed histologic adaptation. Prior to modification, the peri-implant interface exhibited void spaces (pockets) alongside sulcular epithelium and connective tissue layers. Following restoration revision, it is hypothesized that the void was eliminated, leading to the reorganization of peri-implant soft tissue into a more structured zone composed of sulcular epithelium, junctional epithelium, and connective tissue.

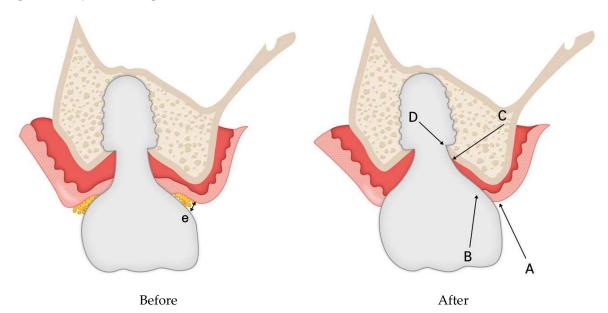


Figure 11. Hypothetical schematic representation of histologic changes in peri-implant soft tissue following restoration modification. Before modification, the Crest to Restoration Distance (CRD) contained voids (pockets), sulcular epithelium, and connective tissue. After modification, it is hypothesized that the CRD was restructured into a stable peri-implant soft tissue barrier, consisting of sulcular epithelium, junctional epithelium, and connective tissue, reinforcing the biological seal and reducing the risk of peri-implant inflammation.

This transformation may contribute to a more effective biologic seal, preventing bacterial infiltration and promoting peri-implant health. However, while these observations are inferred from clinical and radiographic data, further histologic research is required to validate the proposed structural adaptation.

2. Clinical Indicators and Diagnosis of Peri-Implant Disease

Clinical signs of peri-implant mucositis include redness, swelling, bleeding on probing (BOP), suppuration on probing (SOP), and increased probing pocket depth (PPD). In contrast, peri-implantitis is confirmed through radiographic evidence of bone loss. Periodic probing is emphasized

as a critical diagnostic tool for monitoring peri-implant conditions, as increased PPD, accompanied by profuse BOP and SOP, is strongly associated with peri-implantitis [28].

The diagnosis of peri-implant mucositis relies on clinical criteria without requiring histological confirmation. Unlike peri-implantitis, peri-implant mucositis is reversible. Destruction of the junctional epithelium attachment is a hallmark of gingivitis, underscoring the role of hemidesmosomes in periodontal gingiva [26]. However, peri-implant tissues exhibit significant morphological differences from periodontal tissues, particularly weaker hemidesmosomal attachment in the junctional epithelium around implants [29]. This weaker attachment may render peri-implant tissues more susceptible to bacterial infiltration and disease progression, including peri-implantitis.

Despite skepticism regarding the role of hemidesmosomes around implants [10], the resistance observed during probing suggests that peri-implant soft tissue retains some sealing ability. This may be attributed to the functional role of hemidesmosomes, similar to that observed in natural teeth [9,30]. This sealing function highlights the importance of peri-implant soft tissue integrity in maintaining peri-implant health and preventing disease progression.

Numerous reports underscore the pivotal role of probing in maintaining peri-implant soft tissue health. Probing serves not only as an essential diagnostic tool for early detection of peri-implant diseases but also as a preventive measure to mitigate progression toward peri-implant bone destruction. When combined with clinical signs such as erythema, BOP, SOP, and swelling, probing provides a comprehensive assessment of peri-implant tissue health, facilitating timely intervention [28].

3. Probing implants' sulcus with IPPP (Implant Paper Point Probing)

For natural teeth, periodontal sulcus depth is measured using periodontal probes with a limited probing force. Sulcus depth is defined as the distance from the gingival margin to the most apically penetrated portion of the junctional epithelium. In a healthy state, the probe tip remains within the junctional epithelium. However, in the presence of inflammation, the probe penetrates beyond the junctional epithelium into the connective tissue, reflecting the compromised barrier function of the epithelial attachment.

Similarly, in peri-implant tissues, inflammation causes the probe tip to penetrate deeper into the connective tissue, reaching closer to the alveolar bone crest than in natural teeth. The 2017 World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Conditions recommended that peri-implant pocket probing should be performed with a light force (~0.25 N), as healthy peri-implant pocket depths generally remain <5 mm [24].

Controlled probing with a force of 0.2 N has demonstrated similar probe penetration depths in healthy implants and natural teeth, aligning with the dimensions of the epithelial barrier, as observed in studies by Abrahamsson and Soldini [31].

The challenge in clinical examinations lies in applying a standardized probing force. While automated periodontal probes have been proposed to ensure consistent force, their utility for probing implants remains limited due to variations in the curvature and design of implant prosthetic restorations [32].

To overcome these challenges, the author used the Implant Paper Point Probe (IPPP), developed by Suredent in South Korea. The IPPP has a yield strength of 0.25–0.35 N, allowing it to flexibly penetrate the sulcus within this range. If excessive force is applied, the paper points bend rather than damage the tissue, ensuring controlled probing. This approach provides reliable sulcus depth measurements while minimizing the risk of over-penetration, offering an effective alternative to metal probes for implants.

Additionally, the Implant Paper Point Probe (IPPP) offers dual functionality, providing valuable insights into sulcus health by detecting sulcus fluid and bleeding on probing (BOP). These conditions can be directly observed through wetting and discoloration of the paper probe, enhancing diagnostic accuracy.

Some authors have raised concerns about potential tissue damage caused by probing, particularly when the probe penetrates deeper than intended. However, the IPPP method alleviates this concern, as its supple design allows it to bend upon reaching a preset force, preventing damage to the underlying peri-implant soft tissue beyond the appropriate depth.

4. Possible explanation for the results from this study

This case study primarily focused on the impact of Crest to Restoration Distance (CRD) on perimplant soft tissue health. The key finding was that maintaining CRD within an optimal range was associated with peri-implant soft tissue stability, preventing the onset of peri-implant mucositis—a precursor to peri-implantitis.

To explain this observation, three possible mechanisms are proposed:

- 1. Preservation of Hemidesmosomal Attachment
 - o By eliminating pocket formation at the peri-implant interface, the hemidesmosomal attachment of the junctional epithelium remains functional at the entrance.
 - o This prevents sulcular epithelial downgrowth and maintains the integrity of the peri-implant mucosa, reinforcing the soft tissue barrier against bacterial infiltration.
- 2. Hydraulic Pressure from the Connective Tissue
 - With a stable junctional epithelium at the entrance, the connective tissue beneath the implant restoration is able to exert hydraulic pressure against the prosthesis.
 - This pressure generates a protective sealing force, further contributing to the biologic defense mechanism.
- 3. Sustained Structural and Immunologic Integrity
 - Maintaining structural stability ensures that each component of the peri-implant soft tissue performs its specialized role:
 - Sulcular epithelium: High turnover rate providing rapid barrier renewal.
 - Junctional epithelium: Adhesion to the implant surface with immunologic function.
 - Connective tissue: Mechanical support, vascular supply, and immune defense.
 - These roles are well-established in basic histology and can be inferred without direct histological investigation. The structural integrity of peri-implant soft tissue can be assessed through dimensional analysis of each layer, as demonstrated in this case study.

From this case, two key clinical findings support this hypothesis:

- 1. Peri-implant mucositis resolved following the optimization of CRD.
- 2. Peri-implant soft tissue returned to a healthy state, confirmed through Implant Paper Point Probing (IPPP).

These findings suggest that maintaining CRD within an optimal range may help prevent downward migration of the sulcular epithelium, allowing the underlying connective tissue to remain stable without requiring sulcular epithelial coverage. This self-sustained tissue maintains its structural and functional integrity, preventing inflammatory pocket formation and minimizing the risk of disease progression.

Conversely, when CRD exceeds the critical range, peri-implant soft tissue stability is compromised. Without sufficient support, the connective tissue becomes unsustainable, requiring compensatory epithelial coverage. This leads to apical migration of the junctional epithelium, forming deep pockets that facilitate bacterial colonization and inflammation. The resulting inflammatory response, aimed at managing the foreign material, often manifests as soft tissue swelling, further deepening the pocket and increasing susceptibility to peri-implant mucositis and peri-implantitis.

This mechanism closely parallels the pathogenesis of periodontitis in natural teeth, where the junctional epithelium converts into pocket epithelium, marking disease progression [26]. Thus, this study reinforces the importance of CRD as a modifiable factor in maintaining peri-implant health and preventing disease progression.

These results challenge the traditional assumption that subcrestally placed implants (SPIs) are inherently predisposed to deep pocket formation and peri-implant complications. Instead, this study

demonstrates that when CRD is maintained within a critical range, SPIs can achieve stable perimplant soft tissue adaptation—even when a greater peri-implant soft tissue dimension develops beneath the restoration. Conversely, this broader and more voluminous connective tissue may contribute to peri-implant soft tissue health through its intrinsic biological properties, including enhanced vascularization, mechanical support, and immunologic function. This underscores the necessity of meticulous soft tissue management and precision in prosthetic design to ensure long-term implant success.

Furthermore, all cases in this study employed an extraoral cementation technique, effectively eliminating excess cement—a well-documented factor contributing to peri-implant disease. These findings highlight the importance of integrating both soft tissue and prosthetic management strategies to optimize peri-implant health and prevent complications.

5. Histologic Features of the Junctional Epithelium and Connective Tissue

The peri-implant soft tissue consists of two primary components:

- 1. Junctional epithelium
- 2. Connective tissue

These structures work synergistically to form a biologic seal around implants, functionally mimicking the protective roles observed in natural teeth.

Junctional Epithelium

The junctional epithelium serves as a critical defense barrier, forming the first line of protection against microbial colonization and infection. It plays a vital role in adapting gingival tissues to implants while also contributing to immunologic defense mechanisms [9].

Key characteristics of the junctional epithelium:

- High cell turnover: Comprising 15–30 cell layers, with a maximum average thickness of about 0.5 mm.
- Hemidesmosomal adhesion: Hemidesmosomes are present on both the implant-facing and connective tissue-facing surfaces of the epithelium, facilitating adhesion.
- Larger intercellular spaces: Compared to oral epithelium, these spaces allow immune cells to traverse the epithelium, enhancing its defensive function.
- Endocytosis and decomposition: The epithelium has the ability to process and neutralize exogenous factors, contributing to its protective role [34].

Clinically, the junctional epithelium is indistinguishable from connective tissue, but its presence can often be inferred by the absence of bleeding on probing (BOP). Its high cell turnover rate and ability to facilitate immune cell migration further enhance its role as a protective barrier. Therefore, in self-sustained peri-implant soft tissue, the upper portion may include junctional epithelium, even if composed of only a single cell layer, distinguishing it from pathologic sulcular epithelial downgrowth observed in disease progression.

Connective Tissue

Located apical to the junctional epithelium, the peri-implant connective tissue is primarily composed of collagen fibers. In natural teeth, this connective tissue is anchored to cementum via dentogingival fibers, which provide a mechanical barrier against bacterial invasion. However, dental implants lack this intrinsic fibrous attachment.

Instead, the peri-implant connective tissue compensates by forming a functional seal, which is maintained through:

- Expansile forces generated by the inherent elasticity of the connective tissue
- Compressive forces exerted against the implant surface and surrounding structures

These biomechanical interactions ensure stability and integration of the peri-implant soft tissue. (Figure 12) The connective tissue effectively supports the overlying junctional epithelium, playing a

critical role in maintaining peri-implant soft tissue stability and protecting the underlying crestal bone.

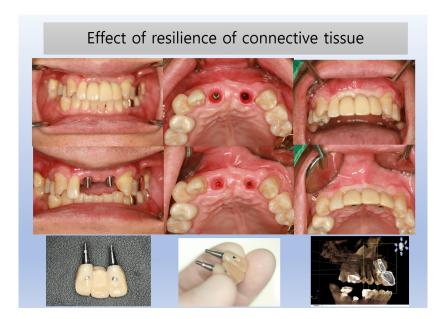


Figure 12. Submucosal Swelling and Hydraulic Pressure in the Connective Tissue.

Figure 12 illustrates a temporary submucosal swelling, representing the resilience and hydraulic positive pressure generated within the connective tissue. This phenomenon occurs during the removal of the prosthesis for temporary repair, highlighting the structural integrity and adaptability of peri-implant connective tissue.

6. New model for biologic width for implants

To structurally understand peri-implant soft tissue, the traditional concept of biologic width, originally developed for natural teeth, has been adapted for implants. This concept was first introduced by Gargiulo et al. [35] and later expanded by Vacek et al. [36], defining biologic width as comprising:

- Gingival sulcus
- Epithelial attachment
- Connective tissue attachment

These studies reported consistent connective tissue measurements of approximately 1.07 mm and 0.77 mm, respectively. Further investigations by Saso Ivanovski explored the supra-alveolar transmucosal architecture, highlighting both similarities and distinct features between teeth and implants [37].

Limitations of Traditional Biologic Width Models in Implant Dentistry

Traditional biologic width models emphasize vertical soft tissue dimensions but largely overlook horizontal and oblique components of peri-implant connective tissue. This is particularly relevant for subcrestally placed implants (SPIs), where the peri-implant soft tissue adapts due to the coronal flaring of implant restorations.

One key adaptation is the formation of an extended soft tissue interface between the crestal bone and the implant restoration, referred to in this study as Crest to Restoration Distance (CRD). Without an adequate understanding of this peri-implant soft tissue, the overall transition zone from the fixture-abutment connection to the coronal gingival margin may be misinterpreted as pathological or mistaken for a socket-like condition.

Previous efforts to standardize peri-implant soft tissue evaluation have largely focused on:

Vertical soft tissue thickness

Crestal bone loss assessments

However, these approaches are limited by their two-dimensional perspective, failing to capture the complex three-dimensional architecture of peri-implant tissues [39]. This oversight underscores the need for a new model tailored to subcrestally placed implants (SPIs) with internal platform-switching connections, where the three-dimensional interplay of soft tissue components plays a critical role in long-term stability and function.

Proposed New Model for Biologic Width in Implants

This study introduces a new biologic width model for implants, categorizing submucosal periimplant soft tissue into three distinct zones: (Figure 13)

1. Sulcus (A-B):

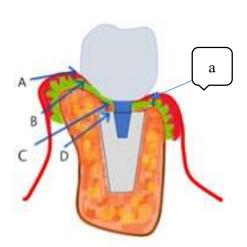
- o Extends from the gingival margin to the beginning of the Transitional Zone.
- Composed of oral epithelium and sulcular epithelium, forming the primary defense barrier against external irritants.

2. Transitional Zone (TZ) (B-C):

- o Located immediately beneath the Sulcus.
- o Composed of elastic connective tissue and may include junctional epithelium.
- Adapts to the implant restoration shape, playing a critical role in stabilizing peri-implant soft tissue.
- Ensures a functional biologic seal, maintaining close contact with the implant surface, essential for resisting bacterial infiltration.

3. Subcrestal Zone (SZ) (C-D):

- o Present in subcrestally placed implants.
- o Consists of inelastic connective tissue in direct contact with the crestal bone.
- Provides mechanical resistance and structural support to the implant, contributing significantly to long-term peri-implant stability.
- The SZ is more likely to be identified when tissue-level implants are placed subcrestally, especially when a matching abutment technique is employed [33].



zone	differentiation		
		Oral	
A-B	Sulcus	(Sulcular)	
		epithelium	
		Junctional	
	Transitional zone (TZ)	epithelium	
B-C		and Elastic	
		connective	
		tissue	
	Subcrestal	Inelastic	
C-D		connective	
	zone	tissue	
a	Traditional biologic width		

Figure 13. provides a schematic drawing that differentiates submucosal peri-implant soft tissue into the three distinct zones:.

1. Sulcus (A-B)

- The outermost layer extending from the gingival margin to the Transitional Zone.
- o Forms the visible interface between the implant restoration and the oral cavity.

2. Transitional Zone (TZ) (B-C)

- o Beneath the Sulcus, transitioning into connective tissue while maintaining some epithelial characteristics.
- Adapts to the contours of the implant restoration, stabilizing the soft tissue and creating a biological seal.
- 3. Subcrestal Zone (SZ) (C-D)
 - Positioned closest to the crestal bone and composed entirely of inelastic connective tissue.
 - Provides foundational support, offering mechanical resistance to external forces.
 - o Plays a crucial role in subcrestally placed implants, ensuring long-term stability.

This schematic model integrates real X-ray images and illustrations, enhancing visualization of peri-implant soft tissue structure. The Transitional Zone (B-C) and Subcrestal Zone (C-D) are emphasized, highlighting their distinct configurations and clinical significance in implant success.

Additionally, this model visually incorporates traditional biologic width (A-E) for comparison.

Differences Between Epi-Crestal Placement and SPI: Considerations for CRD Optimization

The adaptation of peri-implant soft tissue varies depending on implant positioning, prosthetic design, and recipient site characteristics. Understanding these differences underscores the necessity of a comprehensive biologic width model that accounts for both vertical and horizontal peri-implant soft tissue dimensions.

Classification of Implant Placement Approaches (Figure 14)

Type 1: Epi-Crestal Placement with Vertical Biologic Width

When the implant platform is positioned at the crestal bone without horizontal extension of the implant restoration in contact with the underlying mucosa, the peri-implant soft tissue aligns vertically, forming what is traditionally referred to as biologic width. The soft tissue primarily consists of vertically oriented connective tissue and epithelial layers. Because no coronal flaring occurs at the submucosal level, the resulting emergence profile is less natural in appearance.

A slender prosthetic design is typically required to minimize soft tissue compression, and a large CRD is created to facilitate easier hygiene maintenance.

Type 2: Epi-Crestal Placement with Horizontal Extension Generating a Transitional Zone (TZ)

When the implant platform is placed at the crestal bone level with horizontal extension of the implant restoration in contact with the underlying mucosa, the coronal flaring extends the soft tissue attachment laterally, generating a horizontal component within the biologic width. This adaptation stabilizes the peri-implant soft tissue and results in the formation of a Transitional Zone (TZ).

Compared to Type 1, this configuration enhances the natural emergence profile while maintaining biologic function. However, it requires sufficient crestal bone width to support the extended interface between the implant restoration and the soft tissue.

Type 3: Subcrestal Placement (SPI) with Subcrestal Zone (SZ) Formation

In subcrestal placement (SPI), the implant is positioned below the crestal bone level, necessitating peri-implant soft tissue adaptation both vertically and horizontally. This results in the formation of both a Transitional Zone (TZ) and a Subcrestal Zone (SZ), which enhance soft tissue sealing and biologic stability.

SPI generates a more natural-looking emergence profile compared to epi-crestal placements.

CRD must be carefully controlled to prevent excessive soft tissue voids, and optimized CRD (or Soft Tissue Thickness - STT) is critical for long-term biologic stability.

Biological Implications of SPI with CRD Optimization

Enhances vascularization and mechanical support.

- Reduces the risk of epithelial downgrowth and peri-implant mucositis.
- Provides long-term stability, particularly in cases with thin crestal bone, where additional soft tissue volume serves as a protective barrier.

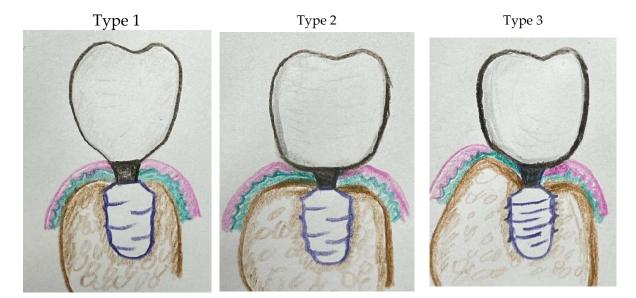


Figure 14. Schematic Models of Peri-Implant Soft Tissue Adaptation Based on Implant Placement Depth and Restoration Design.

This figure illustrates three distinct types of peri-implant soft tissue adaptation, categorized by implant placement depth and the morphology of the implant restoration:

Type 1: Epi-Crestal Placement with Vertical Biologic Width – The implant is positioned at the crestal bone level without horizontal extension of the restoration in contact with the underlying mucosa. This results in a vertically aligned peri-implant soft tissue seal, forming a classical biologic width composed of connective tissue and epithelial layers. A slender restoration design is typically required to reduce soft tissue compression and facilitate hygiene maintenance.

Type 2: Epi-Crestal Placement with Horizontal Extension Forming a Transitional Zone (TZ) – The implant is placed at the crestal bone level with horizontal extension of the implant restoration in contact with the underlying mucosa. This coronal flaring extends the soft tissue attachment laterally, forming a Transitional Zone (TZ) that enhances soft tissue stabilization. This configuration provides a more natural emergence profile than Type 1 while maintaining biologic function, but it requires adequate crestal bone width for optimal support.

Type 3: Subcrestal Placement (SPI) with Subcrestal Zone (SZ) Formation – The implant is positioned below the crestal bone level, promoting peri-implant soft tissue adaptation in both vertical and horizontal dimensions. This leads to the formation of both a Transitional Zone (TZ) and a Subcrestal Zone (SZ), contributing to enhanced soft tissue sealing and biologic stability. Proper CRD management and optimized Soft Tissue Thickness (STT) are critical for maintaining peri-implant health and preventing complications such as peri-implant mucositis.

These models highlight the structural differences in peri-implant soft tissue adaptation and emphasize the importance of CRD optimization in achieving long-term implant stability.

7. Summary of Key Findings

In the presented case of a subcrestally placed implant (SPI) in the upper left first molar region, the peri-implant soft tissue interposed between the crestal bone and the implant restoration emerged as a critical factor in maintaining peri-implant health. Initially, probing depths exceeding 3 mm indicated compromised peri-implant conditions. Following interventions to optimize soft tissue thickness and adjust the CRD within an optimal range, the probing depth improved to less than 1 mm, and this stability was consistently maintained over time.

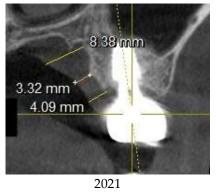
These findings underscore the vital importance of maintaining CRD within a critical range. When CRG exceeds a certain threshold, the soft tissue overlying the crestal bone may transform into oral epithelium, leading to the formation of a pocket prone to bacterial colonization and inflammation. Conversely, by optimizing CRD, epithelial downgrowth was prevented, and the perimplant soft tissue remained stable.

This case highlights the proposed roles of the TZ and SC as structural components essential for peri-implant health. Figure 12 illustrates a temporary submucosal swelling observed during the removal of the prosthesis for repair (Figure 12). This swelling reflects the hydraulic positive pressure and resilience of the connective tissue within the TZ, suggesting a self-sustaining nature that maintains soft tissue integrity and prevents external irritation.

Interestingly, the palatal soft tissue remained unaffected by the implant restoration (Figure 15). This may be attributed to the unique characteristics of palatal tissue, including its abundant blood supply and keratinized nature, which contribute to its resilience and stability despite external modifications [40].

Ultimately, this case highlights the importance of precisely modulating peri-implant soft tissue dimensions. By maintaining an optimal CRD and promoting the formation of TZ and SZ, clinicians can enhance peri-implant health, reduce complications such as inflammation and mucositis, and ensure both functional and esthetic stability. These findings offer valuable insights for integrating advanced peri-implant soft tissue management strategies into clinical practice.

Future research should aim to further elucidate the structural and functional roles of the perimplant soft tissue and establish standardized guidelines for its management.



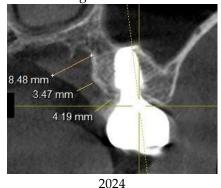


Figure 15. Palatal Mucosal Thickness Over Time.

A comparison of palatal mucosal thickness at three vertical levels (from apical to crestal) between 2021 and 2024. The data demonstrates minimal changes in palatal mucosal thickness over time, suggesting that the palatal soft tissue remains stable despite external influences. This thickness stability may contribute to its strong sealing capability, providing an optimal peri-implant environment in the maxilla.

8. Future Directions

1. Histologic Analysis of Peri-Implant Soft Tissues in SPI

Further histologic studies are required to provide detailed insights into the junctional epithelium and connective tissue adaptations in subcrestally placed implants (SPIs). These studies should analyze how soft tissue interacts with implant surfaces and its role in maintaining peri-implant health. Given the challenges associated with direct histologic validation in human subjects, alternative methodologies, such as advanced imaging techniques and computational modeling, may also be explored to enhance our understanding.

2. Standardization of CRD Measurements for SPI

Large-scale clinical studies are necessary to establish standardized CRD measurement protocols for SPIs. A systematic approach to defining optimal CRD parameters for **Transitional Zone (TZ) and Subcrestal Zone (SZ)** formation would improve clinical assessments and long-term peri-

implant health. By analyzing a broader patient population, future research can refine these parameters and develop evidence-based guidelines for implant placement and prosthetic design.

3. Advancing Prosthodontic Design Based on Bony Architecture

Future prosthodontic designs should transition from empirical approaches to data-driven methodologies tailored to individual bony architecture. The integration of **Cone Beam Computed Tomography (CBCT) and STL file technology** could allow for customized restorations that optimize both esthetic and functional outcomes. Computational simulations and machine learning applications may further enhance precision in predicting peri-implant soft tissue behavior.

4. Establishing a Logical Framework for TZ and SZ

While this case report introduces a novel perspective on peri-implant soft tissue adaptation, its findings are derived from a single case and require validation through larger-scale studies. The proposed concepts of the **Transitional Zone (TZ) and Subcrestal Zone (SZ)** must be tested across various clinical scenarios to determine their reproducibility and clinical relevance. Logical reasoning serves as a fundamental tool in shaping these research directions, forming hypotheses, and refining clinical applications. By systematically integrating empirical data with structured theoretical frameworks, future investigations can strengthen the scientific foundation for peri-implant soft tissue management, ensuring long-term implant success.

A comprehensive research approach that combines histologic, clinical, and theoretical validation will refine peri-implant soft tissue management, ultimately advancing implant dentistry and improving patient outcomes.

Conclusion

This study highlights the critical role of optimizing peri-implant soft tissue characteristics, particularly the Crest to Restoration Distance (CRD), in the successful management of peri-implant mucositis and the long-term stability of subcrestally placed implants (SPIs). Maintaining CRD within an optimal range may help establish a stable biological interface, which could support soft tissue integrity and reduce the risk of epithelial downgrowth and inflammatory pocket formation.

Adjusting prosthodontic design to achieve an optimal CRD was associated with the resolution of peri-implant mucositis, with stable bone levels and probing depths consistently remaining below 1 mm.

The split-mouth case design in this study demonstrated that SPIs, when managed with proper CRD, can promote peri-implant health by:

- Enhancing the peri-implant soft tissue seal
- Maintaining functional stability
- Reducing biologic complications

This study further emphasizes the importance of a three-dimensional approach in analyzing peri-implant soft tissue, integrating key parameters such as CRD, soft tissue thickness (STT), and the proposed Transitional Zone (TZ). These insights provide a comprehensive framework for enhancing implant outcomes through precise soft tissue management strategies.

Furthermore, this research underscores the necessity of customizing prosthodontic designs to match individual patient anatomy, offering a pathway to improved functional and aesthetic outcomes in clinical practice. Future investigations should focus on:

- Establishing standardized CRD thresholds for optimal peri-implant health
- Developing advanced techniques for personalized prosthodontic design

These efforts will broaden the clinical applicability of the findings and pave the way for enhanced long-term stability and health of subcrestally placed implants.

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Ethics Statement: This study is a retrospective clinical report conducted independently at a private dental clinic. As it does not involve any experimental procedures or interventions beyond routine clinical practice, the need for formal ethical approval by any ethical committee was deemed unnecessary. All procedures were performed in accordance with the ethical standards outlined in the Helsinki Declaration and its later amendments or comparable ethical standards. Written informed consent was obtained from all participants for the use of their clinical data and images for research and publication purposes.

Consent for Publication: Written informed consent for the publication of clinical images and case details was obtained from the patient.

Availability of Data and Materials: The data and materials used in this study are available upon reasonable request from the corresponding author.

Clarification: The author, Chiyun Won, DDS, is the sole author of this manuscript. The corresponding author is the same person. The author operates a private dental clinic and is not affiliated with any academic or institutional organizations. This clarification ensures transparency, as standard journal affiliations typically reference well-known institutions.

Conflicts of Interest: The author declares no conflicts of interest.

Abbreviations

Abbreviation	Full Description
SPI	Subcrestally Placed Implant
STT	Soft Tissue Thickness
CRD	Crest to Restoration Distance
cCRD	Central Crest to Restoration Distance
pCRD	Peripheral Crest to Restoration Distance
DP	Depth of Placement
cDP	Central Depth of Placement
pDP	Peripheral Depth of Placement
SZ	Subcrestal Zone
TZ	Transitional Zone
CBCT	Cone Beam Computed Tomography
PPD	Probing Pocket Depth
BOP	Bleeding on Probing
SOP	Suppuration on Probing
IPPP	Implant Paper Point Probing
CAC	Crown-Abutment Complex
3DSTA	3-Dimensional Soft Tissue Analysis
Ep	Epi-Crestal
STL	Standard Tessellation Language (used for 3D modeling and digital impressions)
STH	Supracrestal Tissue Height

References

- 1. Gomez-Meda, R., Esquivel, J., & Blatz, M. B. (2021). "The esthetic biological contour concept for implant restoration emergence profile design." *Journal of Esthetic and Restorative Dentistry*, 33(1), 173–184. https://doi.org/10.1111/jerd.12714.
- 2. Adell, R., Lekholm, U., Rockler, B., & Brånemark, P. I. (1981). "A 15-year study of osseointegrated implants in the treatment of the edentulous jaw." *International Journal of Oral Surgery*, 10(6), 387–416. https://doi.org/10.1016/s0300-9785(81)80077-4.
- 3. Buser, D., Sennerby, L., & De Bruyn, H. (2017). "Modern implant dentistry based on osseointegration: 50 years of progress, current trends, and open questions." *Periodontol* 2000, 73(1), 7–21. https://doi.org/10.1111/prd.12185.

- 4. Kunrath, M. F., & Gerhardt, M. D. N. (2023). "Trans-mucosal platforms for dental implants: Strategies to induce muco-integration and shield peri-implant diseases." *Dental Materials*, 39(9), 846–859. https://doi.org/10.1016/j.dental.2023.07.009.
- 5. Abdallah, M.-N., Badran, Z., Ciobanu, O., Hamdan, N., & Tamimi, F. (2017). "Strategies for optimizing the soft tissue seal around osseointegrated implants." *Advanced Healthcare Materials*, 6(20). https://doi.org/10.1002/adhm.201700549. PMID: 28960892.
- 6. Zheng, Z., Ao, X., Xie, P., Jiang, F., & Chen, W. (2021). "The biological width around implant." *Journal of Prosthodontic Research*, 65(1), 11–18. https://doi.org/10.2186/jpr.JPOR-2019-356.
- 7. Monje, A., González-Martín, O., & Ávila-Ortiz, G. (2023). "Impact of peri-implant soft tissue characteristics on health and esthetics." *Journal of Esthetic and Restorative Dentistry*, 35(1), 183–196. https://doi.org/10.1111/jerd.13003.
- 8. Ávila-Ortiz, G., González-Martín, O., Couso-Queiruga, E., & Wang, H.-L. (2020). "The peri-implant phenotype." *Journal of Periodontology*, 91(3), 283–288. https://doi.org/10.1002/JPER.19-0566.
- 9. Gibbs, S., Roffel, S., Meyer, M., & Gasser, A. (2019). "Biology of soft tissue repair: gingival epithelium in wound healing and attachment to the tooth and abutment surface." *European Cells and Materials*, 38, 63–78. https://doi.org/10.22203/eCM.v038a06.
- Aellos, F., Grauer, J. A., Harder, K. G., Dworan, J. S., Fabbri, G., Cuevas, P. L., Yuan, X., Liu, B., Brunski, J. B., & Helms, J. A. (2024). "Dynamic analyses of a soft tissue-implant interface: Biological responses to immediate versus delayed dental implants." *Journal of Clinical Periodontology*, 51(7), 806–817. https://doi.org/10.1111/jcpe.13980. PMID: 38708491.
- 11. Lang NP, Kinane DF, Lindhe J, Sanz M, Tonetti MS. "Sixth European Workshop on Periodontology of the European Academy of Periodontology at the Charterhouse at Ittingen, Thurgau, Switzerland." *Journal of Clinical Periodontology*, 35(8 Suppl), 1–2. https://doi.org/10.1111/j.1600-051X.2008.01255.x.
- 12. Sanz, M., & Chapple, I. L. (2012). "Clinical research on peri-implant diseases: consensus report of Working Group 4." *Journal of Clinical Periodontology*, 39(Suppl 12), 202–206. https://doi.org/10.1111/j.1600-051X.2011.01837.x.
- 13. Panos N Papapanou et al. (2018). "Periodontitis: Consensus report of workgroup 2 of the 2017 World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Conditions." *Journal of Periodontology*, 89(Suppl 1), S173–S182. https://doi.org/10.1002/JPER.17-0721. PMID: 29926951.
- 14. Linkevicius, T., Apse, P., Grybauskas, S., & Puisys, A. (2009). "The influence of soft tissue thickness on crestal bone changes around implants: a 1-year prospective controlled clinical trial." *International Journal of Oral and Maxillofacial Implants*, 24(4), 712–719. PMID: 19885413.
- 15. Al Rezk, F., Trimpou, G., Lauer, H.-C., Weigl, P., & Krockow, N. (2018). "Response of soft tissue to different abutment materials with different surface topographies: A review of the literature." *General Dentistry*, 66(1), 18–25. PMID: 29303758.
- 16. Jin, S., Yu, Y., Zhang, T., Xie, D., Zheng, Y., Wang, C., Liu, Y., & Xia, D. (2024). "Surface modification strategies to reinforce the soft tissue seal at the transmucosal region of dental implants." *Bioactive Materials*, 42, 404–432. https://doi.org/10.1016/j.bioactmat.2024.08.042. PMID: 39308548. PMCID: PMC11415887.
- 17. González-Martín, O., Lee, E., Weisgold, A., Veltri, M., & Su, H. (2020). "Contour management of implant restorations for optimal emergence profiles: Guidelines for immediate and delayed provisional restorations." *International Journal of Periodontics & Restorative Dentistry*, 40(1), 61–70. https://doi.org/10.11607/prd.4422.
- 18. Rompen, E., Domken, O., Degidi, M., Farias Pontes, A. E., & Piattelli, A. (2006). "The effect of material characteristics, of surface topography and of implant components and connections on soft tissue integration: A literature review." Clinical Oral Implants Research, 17(Suppl 2), 55–67. https://doi.org/10.1111/j.1600-0501.2006.01367.x.
- 19. Hsu, Y.-T., Lin, G.-H., & Wang, H.-L. (2017). "Effects of platform-switching on peri-implant soft and hard tissue outcomes: A systematic review and meta-analysis." *International Journal of Oral and Maxillofacial Implants*, 32(1), e9–e24. https://doi.org/10.11607/jomi.5140. PMID: 28095526.

- 20. Palacios-Garzón, N., Velasco-Ortega, E., & López-López, J. (2019). "Bone loss in implants placed at subcrestal and crestal level: A systematic review and meta-analysis." *Materials (Basel, 12)*(1), 154. https://doi.org/10.3390/ma12010154. PMID: 30621286. PMCID: PMC6337530.
- Baus-Domínguez, M., Oliva-Ferrusola, E., Maza-Solano, S., Ruiz-de-León, G., Serrera-Figallo, M.-Á., Gutiérrez-Perez, J.-L., Torres-Lagares, D., & Macías-García, L. (2024). "Biological response of the periimplant mucosa to different definitive implant rehabilitation materials." *Polymers (Basel)*, 16(11), 1534. https://doi.org/10.3390/polym16111534.
- 22. Esquivel, J., Gomez Meda, R., & Blatz, M. B. (2021). "The impact of 3D implant position on emergence profile design." *The International Journal of Periodontics & Restorative Dentistry*, 41(1), 79–86. https://doi.org/10.11607/prd.5126.
- 23. Vervaeke, S., Dierens, M., Besseler, J., & De Bruyn, H. (2014). "The influence of initial soft tissue thickness on peri-implant bone remodeling." *Clinical Implant Dentistry and Related Research*, 16(2), 238–247. https://doi.org/10.1111/j.1708-8208.2012.00474.x. PMID: 22758656.
- 24. Paolantoni, G., Tatullo, M., Miniello, A., Sammartino, G., & Marenzi, G. (2023). "Influence of crestal and sub-crestal implant position on development of peri-implant diseases: A 5-year retrospective analysis." *Clinical Oral Investigations*, 28(1), 16. https://doi.org/10.1007/s00784-023-05413-4. PMID: 38135770.
- Sun, Y., Yang, J., Chen, K., Li, Z., Chen, Z., & Huang, B. (2023). "Clinical and radiographic results of crestal vs. subcrestal placement of implants in posterior areas: A split-mouth randomized controlled clinical trial." Clinical Implant Dentistry and Related Research, 25(5), 948–959. https://doi.org/10.1111/cid.13230. PMID: 37259774.
- Cristina Valles et al. (2018). "Influence of subcrestal implant placement compared with equicrestal position
 on the peri-implant hard and soft tissues around platform-switched implants: A systematic review and
 meta-analysis." Clinical Oral Investigations, 22(2), 555–570. https://doi.org/10.1007/s00784-017-2301-1. PMID:
 29313133.
- 27. Bosshardt, D. D. (2018). "The periodontal pocket: Pathogenesis, histopathology and consequences." *Periodontology* 2000, 76(1), 43–50. https://doi.org/10.1111/prd.12153.
- 28. Wang, I.-C. I., Barootchi, S., Tavelli, L., & Wang, H.-L. (2021). "The peri-implant phenotype and implant esthetic complications: Contemporary overview." *Journal of Esthetic and Restorative Dentistry*, 33(1), 212–223. https://doi.org/10.1111/jerd.12709. PMID: 33459483.
- 29. Monje, A., Amerio, E., Farina, R., Nart, J., Ramanauskaite, A., Renvert, S., Roccuzzo, A., Salvi, G. E., Schwarz, F., Trombelli, L., & Wang, H.-L. (2021). "Significance of probing for monitoring peri-implant diseases." *International Journal of Oral Implantology (Berlin)*, 14(4), 385–399. PMID: 34726848.
- 30. Berglundh, T., & Lindhe, J. (1996). "Dimension of the periimplant mucosa. Biological width revisited." *Journal of Clinical Periodontology*, 23(10), 971–973. https://doi.org/10.1111/j.1600-051x.1996.tb00520.x.
- 31. Fischer, N. G., & Aparicio, C. (2022). "Junctional epithelium and hemidesmosomes: Tape and rivets for solving the 'percutaneous device dilemma' in dental and other permanent implants." *Bioactive Materials*, 18, 178–198. https://doi.org/10.1016/j.bioactmat.2022.03.019. PMID: 35387164. PMCID: PMC8961425.
- 32. Abrahamsson, I., & Soldini, C. (2006). "Probe penetration in periodontal and peri-implant tissues. An experimental study in the beagle dog." *Clinical Oral Implants Research*, 17(6), 601–605. https://doi.org/10.1111/j.1600-0501.2006.01235.x.
- 33. D. S. Barendregt et al. (2006). "Comparison of two automated periodontal probes and two probes with a conventional readout in periodontal maintenance patients." *Journal of Clinical Periodontology*, 33(4), 276–282. https://doi.org/10.1111/j.1600-051X.2006.00900.x. PMID: 16553636.
- 34. Won, C. (2024). "3-Dimensional Soft Tissue Analysis (3DSTA) of Subcrestally Placed Implants: Relating Biologic Stability with Morphologic Achievements." *Preprints, Version 1*. Received: 15 April 2024; Approved: 17 April 2024.
- 35. Atsuta, I., Ayukawa, Y., Kondo, R., Oshiro, W., Matsuura, Y., Furuhashi, A., Tsukiyama, Y., & Koyano, K. (2016). "Soft tissue sealing around dental implants based on histological interpretation." *Journal of Prosthodontic Research*, 60(1), 3–11. https://doi.org/10.1016/j.jpor.2015.07.001. PMID: 26725967.
- 36. Gargiulo, A. W., & Arrocha, R. (1967). "Histo-clinical evaluation of free gingival grafts." *Periodontics*, 5(6), 285–291. PMID: 4864122.

- 37. Vacek, J. S., Gher, M. E., Assad, D. A., Richardson, A. C., & Giambarresi, L. I. (1994). "The dimensions of the human dentogingival junction." *International Journal of Periodontics & Restorative Dentistry*, 14(2), 154–165. PMID: 7928131.
- 38. Ivanovski, S., & Lee, R. (2018). "Comparison of peri-implant and periodontal marginal soft tissues in health and disease." *Periodontology* 2000, 76(1), 116–130. https://doi.org/10.1111/prd.12150.
- 39. Lin, G.-H., & Madi, I. M. (2019). "Soft-Tissue Conditions Around Dental Implants: A Literature Review." *Implant Dentistry*, 28(2), 138–143. https://doi.org/10.1097/ID.00000000000000871.
- 40. Fuchigami, K., Munakata, M., Kitazume, T., Tachikawa, N., Kasugai, S., & Kuroda, S. (2017). "A diversity of peri-implant mucosal thickness by site." *Clinical Oral Implants Research*, 28(2), 171–176. https://doi.org/10.1111/clr.12778.

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