

Effect of 3-Nitrooxypropanol Combined with Different Feed Additives on Growth Performance, Carcass Traits, Enteric Methane Emissions, and Physiological Response in Feedlot Beef Cattle Fed a High-Concentrate Finishing Diet

[William Luiz de Souza](#)*, Maria Betânia Niehues, [Abmael da Silva Cardoso](#), Victor Valério de Carvalho, [Alexandre Perdigão](#), Tiago Sabella Acedo, [Diogo Fleury Azevedo Costa](#), [Luis Fernando Monteiro Tamassia](#), Maik Kindermann, [Ricardo Andrade Reis](#)

Posted Date: 26 September 2024

doi: 10.20944/preprints202409.2127.v1

Keywords: 3-nitrooxypropanol; beef nutrition; beef production; Bos indicus; carbon footprint; greenhouse gas; intensive systems; sustainable intensification; technology adoption



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article

Effect of 3-Nitrooxypropanol Combined with Different Feed Additives on Growth Performance, Carcass Traits, Enteric Methane Emissions, and Physiological Response in Feedlot Beef Cattle Fed a High-Concentrate Finishing Diet

William Luiz de Souza ^{1,2,*}, Maria Betânia Niehues ³, Abmael da Silva Cardoso ⁴, Victor Valério de Carvalho ⁵, Alexandre Perdigão ⁵, Tiago Sabella Acedo ⁵, Diogo Fleury Azevedo Costa ², Luis Fernando Monteiro Tamassia ⁶, Maik Kindermann ⁶ and Ricardo Andrade Reis ¹

¹ Department of Animal Science, Faculty of Agricultural and Veterinary Sciences, São Paulo State University, 14884-900, Jaboticabal, SP, Brazil

² Institute for Future Farming Systems, Central Queensland University, 4701, Rockhampton, QLD, Australia

³ School of Veterinary Medicine and Animal Science, São Paulo State University, 18618-681, Botucatu, SP, Brazil

⁴ Department of Plant and Agroecosystems Science, University of Wisconsin, Madison, 53706, WI, USA

⁵ DSM Nutritional Products, Innovation & Applied Science, 01452-001, São Paulo, SP, Brazil

⁶ DSM Nutritional Products, 4303, Kaiseraugst, Aargau, Switzerland

* Correspondence: williamluizdesouzaa@gmail.com

Simple Summary: The beef industry faces significant challenges in increasing red protein production, given the growing population with projections to be approximately 28% higher by 2050. Additionally, there are increasing government and public pressures for all industries to mitigate greenhouse gases, which include methane. The livestock industry contributes approximately 14.5% of greenhouse gas emissions attributed to human activities. This study investigated the combination of the methane inhibitor 3-nitrooxypropanol with sodium monensin or a feed additive package consisting of essential oils, 25-Hydroxy-Vitamin-D3 an active metabolite of vitamin D3, and carbo-amino-phospho-chelated minerals such as chromium and zinc. The results showed that 3-nitrooxypropanol, regardless of the combination used, reduced daily enteric methane emissions by over 38%, in addition to increasing feed conversion by over 6% when combined with sodium monensin compared to the treatment without 3-nitrooxypropanol. Furthermore, the combination of 3-nitrooxypropanol with the additive package showed better conditions in growth performance, carcass traits, meat quality, blood parameters, and nutrient intake and digestibility. This offers the meat industry more nutritional strategies to meet the growing demand for protein while reducing environmental impacts.

Abstract: The objective was to evaluate the effect of 3-nitrooxypropanol (3-NOP) in combination with different feed additives on growth performance, carcass traits, meat quality, enteric methane (CH₄) emissions, nutrient intake and digestibility, and blood parameters in feedlot beef cattle. In experiment (Exp.) 1, one hundred sixty-eight Nellore bulls (initial bodyweight (BW) 410 ± 8 kg) were allocated to 24 pens in a completely randomized block design. In Exp. 2, thirty Nellore bulls (Initial BW 410 ± 3 kg) were allocated to a collective pen in a completely randomized design. Three treatments were applied: Control (CTL): Sodium monensin (26 mg/kg of dry matter, DM), M3NOP: CTL with 3-NOP (100 mg/kg DM) and Combo: 3-NOP (100 mg/kg DM) with essential oils (100 mg/kg DM), 25-Hydroxy-Vitamin-D3 (0.10 mg/kg DM), organic chromium (4 mg/kg DM), and zinc (60 mg/kg DM). In Exp 1, bulls in Combo had greater (P<0.01) dry matter intake (DMI) at d 28 compared with CTL and M3NOP. During d 0 to 102, bulls final BW and average daily gain (ADG) were greater (P≤0.03) for Combo compared with CTL. Bulls in Combo and M3NOP had better (P<0.01) feed conversion (FC) and feed efficiency (FE) compared with CTL. Hot carcass weight (HCW), carcass ADG and carcass yield were greater (P≤0.05) for bulls from Combo compared with CTL and M3NOP. Bulls in Combo had greater (P=0.01) dressing compared with M3NOP. Combo bulls had better (P=0.02) biological efficiency compared with CTL. Bulls in Combo had lower (P<0.01) carcass pH compared to CTL and M3NOP. In Exp. 2, bulls in Combo had

greater ($P=0.04$) DMI at d 28 compared with CTL and had greater ($P<0.01$) DMI at d 102 compared with CTL and M3NOP. Bulls in Combo had greater ($P=0.04$) HCW compared with CTL and M3NOP and carcass ADG was greater ($P=0.04$) for bulls Combo compared with M3NOP. Bulls in Combo and M3NOP had lower ($P<0.01$) CH_4 production (38.8%, g/d), yield (41.1%, g/kg DMI), intensity (40.8%, g/kg carcass ADG) and increased ($P<0.01$) H_2 emissions (291%, g/d) compared with CTL. Combo bulls had lower ($P<0.01$) blood glucose and insulin, and higher nutrient intake and digestibility ($P\leq 0.05$) compared with CTL and M3NOP. Combining 3-NOP with different feed additives improved FC, FE, and reduced enteric CH_4 emissions. Combo treatment improved growth performance, carcass traits, nutrient intake and digestibility, and improved glucose and insulin responses in feedlot beef cattle on a high-concentrate finishing diet.

Keywords: 3-nitrooxypropanol; beef nutrition; beef production; *Bos indicus*; carbon footprint; greenhouse gas; intensive systems; sustainable intensification; technology adoption

1. Introduction

Nowadays, the beef industry faces major challenges to increase productivity and improve product quality—carcass and meat—efficiently and sustainably to meet growing global demand [1,2]. A projection made by the Food and Agriculture Organization of the United Nations [3] estimates a population growth of approximately 28% and a total demand for meat of approximately 75% higher by 2050, in millions of tons. Of this total, 25% is comprised by beef [4]. Alongside the growing demand for meat, concerns related to greenhouse gas emissions arise [5]. The livestock industry contributes approximately 14.5% of greenhouse gas emissions from human activities, with 40% of these emissions coming from methane originated from ruminants [6], of which 90% comes from ruminal fermentation [7].

Towards more sustainable production, some management and nutritional strategies should be implemented to increase the productivity of beef cattle throughout their life. As example are strategies to conserve forage, the use of supplements, amongst others to minimize the effects of seasonal fluctuations in forage production inherent to the climate, by incorporating nutritional strategies to assure the continuous supply of nutrients to the beef cattle [8,9,10]. One of these alternatives is to finish beef cattle in feedlots during periods of low forage availability, to establish continuous growth performance and, consequently, increase carcass gains [2], in addition to promoting the reduction of CH_4 emissions by yield and intensity, along with the reduction of age at slaughter. In this sense, it has been observed that pathways, including enteric methane production can reduce efficiency due to energy loss during its formation [11,12], causing a reduction of up to 12% in the intake of gross energy, and potentially reducing feed efficiency [13].

To mitigate energy losses in addition to management techniques and promote continuous growth in beef cattle throughout their productive life, it is also possible to use feed additives with specific characteristics to reduce enteric CH_4 emissions, such as 3-nitrooxypropanol (3-NOP), which directly inhibits methanogenesis in beef cattle [14,15]. Its mode of action involves the binding of the molecule to the enzyme methyl-coenzyme M reductase, thus inhibiting the formation of enteric CH_4 , with no negative impact on non-methanogenic bacteria [16]. Reductions in enteric CH_4 emissions with the inclusion of 3-NOP in the diet of finished cattle can reach up to 90% [17]. However, the need to increase productivity is growing due to the extra cost of the feed additive [18,19]. Therefore, the combination of 3-NOP with other feed additives, like essential oil blends (thymol, eugenol, limonene, and vanillin) [20], 25-Hydroxy-Vitamin-D3, an active metabolite of vitamin D3 [21], and minerals like chromium and zinc amino chelates [22,23], may be beneficial to improve performance and carcass traits. In addition to some, such as chelated chromium and zinc, promoting beneficial changes in blood parameters [24,25], as well as essential oil [26,20] and 3-NOP, in some cases [27,28], promotes an increase in the digestibility of nutrients present in the diet.

Given this, the combination of 3-NOP with other additives would potentially promote beneficial effects on the system, increase productivity, reduce the carbon footprint per unit of dry matter intake and per unit of carcass produced and mitigate the daily production of enteric CH_4 emissions. We

hypothesized that 3-NOP would promote reductions in enteric CH₄ emissions regardless of the combination, in addition to the fact that the combination of 3-NOP with different additives would result in increases in growth performance based on feed conversion and feed efficiency, including increases in carcass gains and yields and in nutrient intake and digestibility in the high concentrate finishing diet. The objective of the current work was to evaluate the effect of 3-NOP in combination with different feed additives on growth performance, carcass traits, meat quality, enteric methane (CH₄) emissions, nutrient intake and digestibility, and blood parameters in lot fed beef cattle on a high-concentrate finishing diet.

2. Materials and Methods

The experimental methods were approved by the Ethics Committee on Animal Use of DSM Nutritional Products SA, (protocol numbers BR 220121 and BR 211215), following the guidelines of the Animal Research Ethics Committee of São Paulo State University. The experiments were conducted at the Center for Innovation and Applied Science in Ruminants of DSM Nutritional Products, located in Rio Brilhante, Mato Grosso do Sul, Brazil.

2.1. Animals, Experimental Design and Treatments (Exp. 1 and 2)

In Exp 1, one hundred and sixty-eight Nellore bulls (*Bos indicus*), 16 ± 1 month old, initial bodyweight (BW) of 410 ± 8 kg, were randomly allocated to 24 pens, using a block design based on the initial body weight after 16 h fasting. There were three treatments, with 8 replicates (pens) and 7 bulls in each pen. Each pen had an area of 120 m² (17 m²/animal), drinkers with high-flow floats (30 linear cm/animal) and collective feeders (70 linear cm/animal). In Exp. 2, thirty Nellore bulls, 16 ± 1 month-old and initial BW of 410 ± 3 kg, were randomly distributed in a 480 m² collective pen (16 m²/animal). A completely randomized design was used, with 10 replicates in each treatment (the bulls were the experimental unit). The collective pen was equipped with a high-flow float drinker (30 cm linear/animal), 6 electronic feeders (Sistema Intergado®, Minas Gerais, Brazil), and 2 electronic feeders per treatment.

The treatments were the same for Exp. 1 and 2: Control (CTL): Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM; M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: Synthetic 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM; and Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); Butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monopropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelated zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil).

2.2. Management, Animal Feeding and Diet Analyses (Exp. 1 and 2).

The animal reception program included weighing, deworming, and vaccination following the annual prophylactic schedule. Diets were formulated using the RLM system (Ração de Lucro Máximo, version 3.2, ESALQ, São Paulo, Brazil) (Table 1).

Table 1. Ingredients and nutritional profile of diets (dry matter basis) in Exp. 1 and 2.

Item	Diet Adaptation 1			Diet Adaptation 2			Diet Finishing		
	CTL	M3NOP	Combo	CT	M3NO	Comb	CTL	M3NO	Comb
	1	2	3	L	P	o	CTL	P	o
Ingredients, % DM									
Sugarcane bagasse, %	30.0	30.0	30.0	20.0	20.0	20.0	10.0	10.0	10.0
Soybean meal, %	11.0	11.0	11.0	6.35	6.35	6.35	1.70	1.70	1.70
Ground corn, %	50	50	50	59.5	59.5	59.5	69.0	69.0	69.0
Cottonseed, %	5.00	5.00	5.00	10.0	10.0	10.0	15.0	15.0	15.0
Urea, %	1.00	1.00	1.00	1.15	1.15	1.15	1.30	1.30	1.30
Mineral Supplement ⁴ , %	3.00	3.00	3.00	3.00	3.00	3.00	3.00	3.00	3.00
Sodium monensin, mg/kg DM	26.0	26.0	-	26.0	26.0	-	26.0	26.0	-
Essential oils, mg/kg DM	-	-	100	-	-	100	-	-	100
25-Hydroxy-Vitamin-D3, mg/kg DM	-	-	0.10	-	-	0.10	-	-	0.10
3-nitrooxypropanol, mg/kg DM	-	100	100	-	100	100	-	100	100
Chemical composition (DM-basis)									
Dry matter, %	89.5	89.5	89.5	90.3	90.3	90.3	90.5	90.5	90.5
Organic matter, % DM	84.6	84.6	84.6	85.2	85.2	85.2	85.3	85.3	85.3
Crude protein, % DM	16.2	16.2	16.2	15.4	15.4	15.4	14.8	14.8	14.8
Mineral matter, % DM	4.93	4.93	4.93	5.05	5.05	5.05	5.24	5.24	5.24
Total Digestible Nutrients, % DM	69.8	69.8	69.8	74.2	74.2	74.2	81.1	81.1	81.1
Neutral Detergent Fiber, % DM	32.2	32.2	32.2	27.8	27.8	27.8	26.5	26.5	26.5
Acid Detergent Fiber, % DM	22.8	22.8	22.8	19.3	19.3	19.3	18.4	18.4	18.4
PeNDF ⁵ , % DM	27.2	27.2	27.2	22.2	22.2	22.2	17.2	17.2	17.2
Starch, % DM	35.5	35.5	35.5	42.2	42.2	42.2	49.0	49.0	49.0
Vitamin D3, UI/kg DM	510	4510	510	4510	510	510	4510		
Chromium, mg/kg DM	0.20	0.20	0.60	0.20	0.20	0.60	0.20	0.20	0.60
Zinc, mg/kg DM	60.0	60.0	120	60.0	60.0	120	60.0 0	60.00	120
ME ⁶ , Mcal/kg DM	2.52	2.52	2.52	2.68	2.68	2.68	2.93	2.93	2.93

Abbreviations: DM - dry matter; ME - metabolizable energy; Mcal – megacalorie. ¹ CTL: Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM. ² M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM. ³ Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monopropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg

DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-Vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelates zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil). ⁴ Mineral and vitamin supplement containing (per kg DM): 150 g Calcium; 16 g Phosphorus; 36 g Sulfur; 20 g Magnesium; 34 g Potassium; 56 g Sodium; 8 mg Cobalt; 540 mg Copper; 6.7 mg Chromium; 27.5 mg Iodine; 1070 mg Manganese; 6.70 mg Selenium; 2000 mg Zinc; 168,000 IU Vitamin A; 17,000 IU Vitamin D3; 1740 IU Vitamin E; 90 mg Biotin; 1.35×10^{11} CFU *Saccharomyces cerevisiae*. Manufactured by DSM Nutritional Products, São Paulo, Brazil. ⁵ peNDF - physically effective neutral detergent fiber: Lammers et al. [33], the peNDF in the diet, based on DM, was determined by the fraction kept on the 1.17 mm sieve.

The adaptation to the finishing diet involved a 14-day protocol with two diets offered over seven days each. Once in the finishing diet, bulls were fed daily at 8:00 am. Diet and residues were collected twice per week in both experiments. These samples were subsequently dried in a forced air circulation oven regulated at 55 °C for 72 h. After were ground using a sieve of 1 mm and then subsequent chemical analysis. The DM (AOAC 934.01), organic matter (OM; calculated: DM – Mineral matter (MM)), ether extract (EE; AOAC 920.39), MM (AOAC 942.05) and nitrogen content (AOAC official method 954.01) was converted to crude protein (CP) using a factor of 6.25, as proposed by AOAC [34]. Neutral detergent fiber (NDF), and acid detergent fiber (ADF) were determined with the methodology of Van Soest et al. [35], using a fiber analyzer (Ankon Technologies, New York, USA). Starch was determined using the method described by Hall [36]. The total digestible nutrients (TDN) of the dietary components were calculated according to the table values and the digestible energy (DE) using the equations proposed by the NRC [37]. The metabolizable energy (ME) was calculated with NASEM [38].

2.3. Growth Performance and Carcass Traits (Exps 1 and 2)

In Exp 1, the DMI was determined by calculating the difference of amount of the diet offered and the leftover collected daily from each pen on the following day. This value was then divided by bulls in the pen. Bulls were under *ad libitum* intake conditions, with a 3% residue allowance. In Exp 2, DMI was measured individually for each animal using the electronic feeder system with scales (System Intergado®, Intergado Ltd., Minas Gerais, Brazil) in addition to the sum of individual pellet DMI for each animal from the Greenfeed unit system (C-lock Inc., Rapid City, SD, USA). Diets were provided *ad libitum* and daily adjustments were made based on the amount of feed refused, with a 5% leftover allowance.

Nellore bulls were weighed individually after a 16 h fasting period on d 0, 28 and 102 of the experiment. Average daily gain (ADG) was computed using data acquired from weight assessments. Furthermore, calculations were conducted for feed conversion (FC; ratio of DMI/ADG, kg/kg) and feed efficiency (FE; ratio of ADG/DMI, kg/kg).

In the end of the exp. 1 and 2 on d 102, A certified technician collected and analyzed ultrasound images using the BIA/DGT Brazil software. Measurements of *Longissimus* muscle (LM) area, rib fat thickness (RFT), marbling, and rump fat thickness were obtained via ultrasound (ALOKA SSD 100 VET®, Aloka, Japan) utilizing a 17-cm linear probe operating at a frequency of 3.5MHz. After measuring the carcass traits with ultrasound, the bulls were transported to a slaughterhouse (Campo Grande, Mato Grosso do Sul, Brazil), located 165 km from the experiments site. The slaughter procedures adhered to the standards established by the slaughterhouse following prevailing regulations governing the slaughter of cattle. The hot carcass weight (HCW) at the time of slaughter was determined. The initial HCW was established based on a yield derived from the reference slaughter conducted at the beginning of the experiments, resulting in the following equation: Initial HCW = Initial BW * 0.5226 for further calculations. The carcass ADG in kg/day was calculated as: (final HCW - initial HCW) / 102 days. Dressing % was calculated as: (final HCW / final BW) * 100.

Carcass yield percentage was determined by: $((\text{final HCW} - \text{initial HCW}) / (\text{final BW} - \text{initial BW})) * 100$. Biological efficiency (kg DM / 15 kg HCW) was calculated by: $(\text{total DMI intake over 102 days}) / ((\text{final HCW} - \text{initial HCW}) * 15 \text{ kg HCW})$.

2.4. Meat Quality (Exp. 1)

Two bulls were selected per pen for meat quality assessment, totaling sixteen per treatment. Carcass pH was measured 24 h after slaughter in a cold room at 4 °C, in the region of the LM (12th rib), using a pH meter (model HI 99163, Hanna Instruments, São Paulo, Brazil). Samples for quality assessments were collected from the LM between the 11 - 13th rib, stored in vacuum-sealed bags and frozen. Meat color analyses (L^* , a^* , b^*) were performed at 24 and 72 h. The steaks were thawed in a refrigerator at 6 °C for a duration of 24 h and exposed to oxygen for 30 min, measurements at four different points on the sample were conducted using a portable spectrophotometer (model CM2500d, Konica Minolta, São Paulo, Brazil) with D65 illumination and a 10° standard observer angle, calibrated with black and white standards [39].

The analyses of shear force and cooking losses, 3 steaks (2.54 cm thick) were selected, thawed under refrigeration at 6 °C for a duration of 24 h, with the initial weight of the sample being determined. A thermometer was used at the geometric center of each steak, and the steaks were heated in an industrial electric oven (model F130/L, Flecha de Ouro Ind. e Com. Ltda., São Paulo, Brazil) set to a target temperature of 170 °C until they reached an internal temperature of 40 °C. After turning the steaks, they were cooked until the internal temperature reached 71 °C. The steaks were then cooled to room temperature, and the difference between the final and initial weight of each steak was calculated to determine the cooking loss. The steaks were subsequently stored in refrigeration at 6 °C for a duration of 24 h and later analyzed for shear force. The analysis was performed using five cylinders with a diameter of 1.27 cm to remove fragments of the steaks along the muscle fiber direction and analyzed using a texturometer (TMS-PRO, Food Technology Corporation, Virginia, USA). For calculation purposes, the values of the five cylinders were used to obtain the shear force value expressed in Newtons (N). All procedures for these analyses were described by the AMSA [40].

Myoglobin determination was performed using a steak (1.27 cm thick), which the connective tissue and external fat were removed, and the meat was reduced to cubes and quickly frozen in liquid nitrogen. Myoglobin extraction and quantification were performed following the methodology established by Warris [41] and adapted by Hunt et al. [42]. The concentration of myoglobin extracted was quantified by measuring the absorbance at 433 nm. The calculations were performed using a molar extinction coefficient of 1.14×10^5 M/cm, the molecular weight of myoglobin, and the corresponding dilution factor. Total lipids were determined in the meat samples in triplicate, following the methodology of Bligh and Dyer [43]. The methodology of Sorensen and Jorgensen [https://i.mdpi.cn/team/training/lesson_series/56/details], [44] for the analysis of lipid oxidation, five grams of meat from each sample were used, in triplicate, and homogenized for 1 minute with 15 mL of trichloroacetic acid, with subsequent absorbance readings at 530 and 632 nm. A standard curve containing 5 points was created using the tetraethoxypropane solution of known concentration, thus obtaining the malonaldehyde (MDA) concentrations through the standard curve equation, expressed in mg MDA/kg of meat.

2.5. Blood Variables (Exp 2)

Blood samples were collected from 30 bulls on d 0, 28, and 102 of feedlot period. Samples were obtained via venipuncture of the jugular vein using appropriate tubes (BD Vacutainer®, São Paulo, Brazil), collecting approximately 10 mL of blood per sample. Following collection, Blood samples were immediately transferred to ice and subsequently centrifuged at 3,000 rpm for 15 min. at 4 °C. After, serum was promptly transferred to a polypropylene container and stored at -20 °C. Biochemical markers assessed in the blood included glucose, urea, albumin, creatinine, total proteins, cholesterol, triglycerides, aspartate aminotransferase (AST), and gamma-glutamyl transferase (GGT). These markers were quantified using commercial Bioclin® kits (Bioclin®, Minas Gerais, Belo Horizonte, Brazil) and absorbance measurements were conducted with a spectrophotometer (SBA

200, CELM, São Caetano do Sul, Brazil), according to the manufacturer's instructions. Serum levels of insulin and Insulin-like Growth Factor 1 (IGF-1) were measured using the Immulite 1000® commercial kit (Diagnostic Products Corporation, California, USA).

2.6. Digestibility of Nutrients (Exp 2)

Diet and faeces collections were conducted at three specific time points on d 28, 56, and 84 of the feedlot periods, covering a continuous period of five days, to evaluate the overall digestibility of DM, organic matter (OM), CP, EE, NDF, ADF, and starch. Diet samples and leftovers were collected once daily. These samples were identified and stored at -20°C. Faecal samples were collected three times daily (08:00 am, 01:00 and 05:00 pm) directly from the ground in the pens, immediately after the bulls defecated. Faecal samples were placed on ice and stored at -20°C shortly after collection. Diet, leftover, and fecal samples were dried at 55 °C for a duration of 48 h in a forced-air oven and ground using a Wiley mill (Model 4 Wiley, Thomas Scientific, New Jersey, USA) to pass through both 2 mm and 1 mm sieves, and after grinding, the samples were sent for composition analysis, where indigestible neutral detergent fiber (iNDF) was used as an internal marker [45,46]. Apparent nutrient digestibility was then calculated using the formula: $\text{nutrient digestibility (\%)} = 100 - (100 * [\text{nutrient content in faeces} * \text{iNDF content in the feed}] / [\text{nutrient content in faeces} * \text{iNDF content in faeces}])$.

2.7. Greenhouse Gas Emissions (Exp. 2)

Measurement data for enteric CH₄ and hydrogen (H₂) emissions were collected from 30 individual bulls during the total feedlot period of 102 days for 24 h. Enteric CH₄ emissions were determined using the Greenfeed unit (C-lock Inc., South Dakota, USA), following the methods described by Della Rosa et al. [47]. Through this system, it was possible to measure the emissions of gases from individual bulls, in which they voluntarily approached the equipment and received a small amount of additive-free pelleted feed with the composition of 88.5% DM, 17.0% CP, 3.00% EE, 1.20% calcium and 0.60% phosphorus. Average daily emissions were estimated by aggregating data from several visits throughout the observation period. The equipment was programmed to produce 6 drops of concentrate pellets per visit and with a minimum interval of 4 h between each visit. For each feeding period, a maximum of five drops of feed containing an average of 35.5 g of pellets were allowed, with an interval of 35 seconds between drops. After the measurements, daily enteric CH₄ emissions were calculated in g/d and expressed as yield based on DMI (g CH₄/kg DMI), as intensity based on carcass ADG (g CH₄/kg carcass ADG), and as enteric H₂ emissions in g/d.

2.8. Statistical Analysis (Exps 1 and 2)

In both experiments, statistical analyses were conducted using SAS software version 9.4 (SAS Institute, North Carolina, USA). The Cramér-von Mises normality test was applied to all data, followed by ANOVA, and the Tukey test was employed for mean comparisons. In Exp. 1, the experimental units were the pens (n=24; 7 bulls/pen). The experimental design was randomized blocks, using the initial fasting BW of the bulls for block allocation. The treatments were: Control (CTL, n=56 bulls), M3NOP (n=56 bulls) and Combo (n=56 bulls), and the analysis included data on growth performance, carcass traits and meat quality. In Exp. 2, a completely randomized design was used, where individual bulls in the CTL (n=10 bulls), M3NOP (n=10 bulls), and Combo (n=10 bulls) treatments were considered as experimental units, and treatments were considered as fixed effects. The analysis included data on growth performance, carcass traits, greenhouse gas measurements, blood parameters, and nutrient intake and digestibility. Blood parameters, nutrient intake, and digestibility data were analyzed as repeated measurements. Due to potential inter-animal variability, blood parameter measurements on d 0 were tested and, when significant, used as covariates for measurements on d 28 and d 102, to mitigate the impact of inter-animal variability on the analysis and allowing a more accurate assessment of treatment effects repeated measurements. The Bayesian Information Criterion was employed to select the best covariance structure. For all statistical results, a significance level of $P \leq 0.05$ was considered.

3. Results

3.1. Growth Performance, Carcass Traits and Meat Quality (Exp. 1)

Effects of CTL, M3NOP and Combo treatments were detected ($P \leq 0.05$) on growth performance, carcass traits and meat quality (Table 2). Treatment Combo showed an increase in DMI of 5.56% ($P < 0.01$) compared to the average of CTL and M3NOP on d 28. On d 102, BW and ADG increased by 13 kg ($P = 0.03$) and 0.140 kg/d ($P = 0.01$), respectively, for Combo treatment compared to CTL, with no difference for M3NOP. Combo and M3NOP treatments showed an improvement in FC of 5.87% ($P < 0.01$) and an FE of 5.58% ($P < 0.01$) compared to CTL. No differences were detected ($P \geq 0.07$) among treatments for initial BW, BW at d 28, ADG at d 28, FC at d 28, FE at d 28 and DMI at d 102 (Table 2).

Combo treatment showed an increase of 13 kg in final HCW ($P < 0.01$), carcass ADG of 0.113 kg/d in carcass ADG ($P < 0.01$) and 3.42% in carcass yield ($P = 0.05$) compared to the means of CTL and M3NOP treatments. Combo treatment was 1.75% higher ($P = 0.01$) compared to M3NOP, with no difference for CTL treatment. Biological efficiency showed a reduction of 12 kg of total DM to produce 15 kg of HCW ($P = 0.02$) for the Combo treatment compared to CTL, with no difference for the M3NOP treatment. No differences ($P \geq 0.21$) were observed between treatments for initial HCW, LM area, marbling, 12th rib fat thickness and rump fat thickness (Table 2). The Combo treatment reduced by 1.52% ($P < 0.01$) the carcass pH value when compared to the average of the CTL and M3NOP treatments. No difference ($P \geq 0.17$) between treatments for shear force, cooking loss, meat color (Chroma) at 24 and 72 h, myoglobin, total lipids and lipid oxidation (Table 2).

Table 2. Effect of combining 3-nitrooxypropanol with different feed additives on growth performance, carcass traits, and meat quality of feedlot Nellore bulls fed a high-concentrate finishing diet in Exp. 1.

Item	CTL ¹	M3NOP ²	Combo ³	SEM ⁴	P-value
Growth performance					
Initial bodyweight, kg	410	410	410	8.17	0.83
d 28 dry matter intake, kg/d	11.2 ^b	10.9 ^b	11.7 ^a	0.22	<0.01
d 28 average daily gain, kg/d	1.816	1.795	1.918	0.06	0.34
d 28 feed conversion, kg/kg	6.23	6.11	6.14	0.21	0.91
d 28 feed efficiency, kg/kg	0.162	0.164	0.163	0.01	0.93
d 28 bodyweight, kg	462	460	464	8.18	0.46
d 102 dry matter Intake, kg/d	12.1	11.8	12.4	0.26	0.07
d 102 average daily gain, kg/d	1.733 ^b	1.808 ^{ab}	1.873 ^a	0.03	0.01
d 102 feed conversion, kg/kg	6.98 ^a	6.54 ^b	6.60 ^b	0.13	<0.01
d 102 feed efficiency, kg/kg	0.144 ^a	0.154 ^b	0.152 ^b	0.01	<0.01
Final bodyweight, kg	588 ^b	594 ^{ab}	601 ^a	8.63	0.03
Carcass traits					
Initial hot carcass weight, kg	214	213	213	4.47	0.57
Final hot carcass weight, kg	332 ^b	334 ^b	345 ^a	5.29	<0.01
Carcass average daily gain, kg/d	1.159 ^b	1.188 ^b	1.287 ^a	0.02	<0.01
Dressing, %	56.5 ^{ab}	56.3 ^b	57.3 ^a	0.23	0.01
Carcass yield, %	67.0 ^b	65.7 ^b	68.7 ^a	0.76	0.05
Biological Efficiency, DM kg/15 kg HCW ⁶	156 ^b	150 ^{ab}	144 ^a	3.29	0.02
LM ⁵ area, cm ²	89.2	88.7	91.1	1.80	0.31
Marbling	2.38	2.57	2.40	0.09	0.21

12th-rib-fat, mm	5.52	5.47	5.87	0.18	0.21
Rump fat thickness, mm	8.11	8.26	8.24	0.27	0.87
Meat quality					
pH	5.92 ^b	5.92 ^b	5.83 ^a	0.01	<0.01
Shear force, N	65.9	64.6	65.1	3.11	0.95
Cooking loss, %	27.4	28.0	27.5	0.50	0.69
Chroma L* (24h)	39.6	39.7	38.9	0.85	0.77
Chroma a* (24h)	21.6	21.2	21.2	0.54	0.84
Chroma b* (24h)	14.8	14.9	14.2	0.51	0.57
Chroma L* (72h)	42.7	41.8	41.9	0.92	0.73
Chroma a* (72h)	22.7	22.0	22.4	0.63	0.76
Chroma b* (72h)	17.0	16.6	16.6	0.61	0.85
Myoglobin, mg/g	4.22	4.20	4.15	0.04	0.35
Total lipids, %	1.72	1.90	1.76	0.09	0.12
Lipid oxidation, mgMDA/kg meat	1.05	1.03	1.05	0.01	0.15

Abbreviations: SEM - standard error of the mean; d - day; DM - dry matter; LM - *Longissimus* Muscle; HCW - hot carcass weight; N - Newton; MDA - malonaldehyde. ¹ CTL: Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM. ² M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM. ³ Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monopropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-Vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelates zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil). Superscripts ^{a, b} signify difference with $P \leq 0.05$, as determined by the Tukey test.

3.2. Growth Performance and Carcass Traits (Exp. 2)

Effects of CTL, M3NOP and Combo treatments were detected ($P \leq 0.04$) on growth performance and carcass traits (Table 3). Over d 28, bulls in the Combo treatment showed an increase in DMI of 5.51% ($P = 0.04$) compared to CTL, with no difference for M3NOP. However, on d 102, bulls in the Combo treatment showed an increase in DMI of 8.20% ($P < 0.01$) compared to CTL and M3NOP. No differences ($P \geq 0.19$) between treatments were observed for initial BW, BW at d 28, ADG at d 28, FC at d 28, FE at d 28, BW at d 102, ADG at d 102, FC at d 102 and FE at d 102 (Table 3).

Combo treatment showed an increase in final HCW of 14 kg ($P < 0.01$) and carcass ADG of 0.118 kg/d ($P < 0.01$) compared with CTL, no difference for M3NOP. No difference was observed ($P \geq 0.11$) between treatments for initial BW, carcass yield, biological efficiency, LM area, marbling, 12th-rib fat thickness, and rump fat thickness (Table 3).

Table 3. Effect of combining 3-nitrooxypropanol with different feed additives on growth performance and carcass traits of feedlot Nellore bulls fed a high-concentrate finishing diet in Exp. 2.

Item	CTL ¹	M3NOP ²	Combo ³	SEM ⁴	P-value
Growth performance					
Initial bodyweight, kg	410	410	410	3.23	0.93
d 28 dry matter intake, kg/d	11.0 ^b	11.3 ^{ab}	11.8 ^a	0.20	0.04
d 28 average daily gain, kg/d	1.500	1.536	1.750	0.10	0.21
d 28 feed conversion, kg/kg	8.30	7.71	7.61	0.50	0.58
d 28 feed efficiency, kg/kg	0.135	0.138	0.149	0.008	0.45
d 28 bodyweight, kg	452	453	459	2.77	0.19
d 102 dry matter Intake, kg/d	11.8 ^b	11.7 ^b	12.8 ^a	0.23	<0.01
d 102 average daily gain, kg/d	1.814	1.765	1.892	0.05	0.36
d 102 feed conversion, kg/kg	6.57	6.61	6.78	0.16	0.61
d 102 feed efficiency, kg/kg	0.153	0.152	0.148	0.004	0.61
Final bodyweight, kg	595	590	603	5.99	0.35
Carcass traits					
Initial hot carcass weight, kg	212	213	213	3.38	0.93
Final hot carcass weight, kg	333 ^b	331 ^b	346 ^a	3.89	0.04
Carcass average daily gain, kg/d	1.186 ^b	1.157 ^b	1.304 ^a	0.04	0.04
Dressing, %	56.1	56.0	57.3	0.47	0.23
Carcass yield, %	65.2	65.3	68.9	1.63	0.34
Biological efficiency, DM kg/15 kg HCW	151	153	149	5.05	0.85
LM area, cm ²	88.2	86.6	88.8	2.58	0.82
Marbling	2.35	2.41	1.93	0.17	0.11
12 th -rib fat, mm	5.37	6.01	5.62	0.28	0.24
Rump fat thickness, mm	7.67	8.41	7.88	0.34	0.30

Abbreviations: SEM - standard error of the mean; d - day; DM - dry matter; LM - *Longissimus* Muscle; HCW - hot carcass weight. ¹ CTL: Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM.²M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM.³Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monopropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-Vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelates zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil).Superscripts ^a, ^b signify difference with P ≤ 0.05, as determined by the Tukey test.

3.3. Greenhouse Gas Measurements

An effect ($P<0.01$) was detected between treatments for greenhouse gas measurements (Table 4). M3NOP and Combo treatments resulted, on average, in reductions of 38.8% in daily CH₄ production ($P<0.01$), 41.1% in CH₄ yield ($P<0.01$), and 40.8% in CH₄ intensity ($P<0.01$) compared to the CTL treatment. In addition, an increase in H₂ emission by 291% ($P<0.01$) was observed for the Combo and M3NOP treatments compared with CTL. No differences ($P\geq0.44$) between the treatments on drops as total per day and amount in kilograms, time in minutes per day (Table 4).

Table 4. Effect of combining 3-nitrooxypropanol with different feed additives on greenhouse gas emissions of feedlot Nellore bulls fed a high-concentrate finishing diet in Exp. 2.

Item	CTL ¹	M3NOP ²	Combo ³	SEM ⁴	P-value
GreenFeed data					
Drop, total/d	16.1	15.4	14.8	1.62	0.74
Time, minutes/visit	5.10	5.06	5.09	0.07	0.60
Drop, kg	0.56	0.54	0.52	0.06	0.54
CH ₄ production, g/d	141 ^a	87.0 ^b	85.6 ^b	6.67	<0.01
H ₂ g/d	1.23 ^b	4.59 ^a	5.04 ^a	0.22	<0.01
CH ₄ yield, g/kg DMI	12.0 ^a	7.45 ^b	6.69 ^b	0.50	<0.01
CH ₄ intensity, g/kg Carcass ADG	119 ^a	75.2 ^b	65.6 ^b	5.43	<0.01

Abbreviations: ADG – average daily gain; SEM - standard error of the mean; d - day; DMI - dry matter intake; H - hydrogen; CH₄ - methane. ¹CTL: Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM. ²M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM. ³Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monopropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-Vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelates zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil). Superscripts ^{a, b} signify difference with $P\leq0.05$, as determined by the Tukey test.

3.4. Blood Variables (Exp. 2)

Effects on blood parameters were observed, significantly influenced by treatments ($P\leq0.05$) and by the treatment \times time interaction ($P\leq0.01$) (Table 5). Serum insulin and glucose levels were affected by the treatment \times time interaction ($P\leq0.01$), with all treatments showing increased levels on d 28 and 102. The Combo treatment exhibited reduced serum insulin and glucose levels compared to the average levels observed in the CTL and M3NOP treatments. A treatment \times time interaction ($P=0.01$) was also noted for serum albumin, with lower concentrations on d 102 for the Combo treatment compared to the average levels of the CTL and M3NOP treatments. The Combo treatment showed a reduction in AST by 9.04% ($P=0.04$) and cholesterol by 12.4% compared to the mean of the CTL and M3NOP treatments. No differences were observed for treatment ($P\geq0.09$) or treatment \times time interaction ($P\geq0.17$) for variables like IGF-1, total protein, creatinine, GGT, and triglycerides.

Table 5. Effect of combining 3-nitrooxypropanol with different feed additives on blood parameters of feedlot Nellore bulls fed a high-concentrate finishing diet in Exp. 2.

Item	CT	M3N	Com	CT	M3N	Com	CT	M3N	Com	SE	P-value			
	L ¹	OP ²	bo ³	L	OP	bo	L	OP	bo		M ⁴	T ⁵	Ti me	T x Time
	d 0			d 28			d 102							
Blood parameters,														
Insulin, uIU/ml	7.1 8	6.68	6.99	10. 3 ^a	11.2 ^a	9.28 ^b	16. 0 ^a	16.1 ^a	13.9 ^b	0.20	<0. 01	<0.0 1	<0.01	
IGF-1, ng/ml	313	331	339	427	412	437	450	458	455	15.7	0.5 9	<0.0 1	0.78	
Glucose, mg/dl	75. 9	80.4	83.1	99. 5 ^a	103 ^a	89.2 ^b	113 a	107 ^{ab}	95.8 ^b	3.72	0.0 5	<0.0 1	0.01	
Albumin, g/dl	3.0 6	3.18	3.34	3.6	3.55	3.35	3.8	3.85	3.67	0.08	0.5 1	<0.0 1	0.01	
Total protein, g/dl	6.7 9	6.79	6.80	6.6 0	6.69	6.63	6.6 0	6.90	6.79	0.07	0.0 9	0.01	0.48	
Creatinine, mg/dl	1.7 5	1.75	1.73	1.5	1.54	1.56	2.0 0	2.04	2.06	0.05	0.6 7	<0.0 1	0.93	
AST ⁶ , u/l	68. 8	67.3	69.6	78. 0	80.0	64.6	82. 0	93.0	79.2	4.05	0.0 4	<0.0 1	0.11	
GGT ⁷ , u/l	18. 8	19.1	19.6	24. 0	28.5	28.6	32. 0	31.1	28.0	1.86	0.6 5	<0.0 1	0.19	
Cholesterol, mg/dl	86. 7	87.3	86.3	126	132	107	165	175	145	6.04	<0. 01	<0.0 1	0.17	
Triglycerides, mg/dl	46. 3	41.7	41.6	43. 0	44.7	42.1	46. 0	46.2	46.5	2.21	0.6 4	0.15	0.59	
Urea, mg/dl	48. 0	48.2	46.6	46. 0	47.7	47.1	36. 0	39.3	41.6	1.54	0.2 8	<0.0 1	0.31	

Abbreviations: SEM - standard error of the mean; d - day; DM - dry matter; T – treatments; AST - aspartate aminotransferase; GGT - gamma-glutamyl transferase; IGF-1 - Insulin-like Growth Factor 1. ¹CTL: Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM. ²M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM. ³Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monopropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-Vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelates zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich

Nutritional Products Ltd., São Paulo, Brazil). Superscripts ^{a, b} signify difference with $P \leq 0.05$, as determined by the Tukey test.

3.5. Intake and Digestibility of Nutrients

Effects of treatments were detected ($P \leq 0.05$) on nutrient intake (Table 6). Combo treatment an increase in DMI of 7.95% ($P=0.03$), OM of 8.08% ($P=0.03$), CP of 7.68% ($P=0.02$), ADF of 8.04% ($P=0.02$), NDF of 7.27% ($P=0.04$), and starch of 7.33% ($P=0.05$) was observed, except for EE ($P=0.18$) compared with treatments CTL and M3NOP. No difference detected ($P \geq 0.85$) for the interaction treatment x time on nutrient intake, including DM, OM, CP, EE, ADF, NDF, and starch (table 6).

The interaction between treatment x time detected an effect ($P = 0.04$) on NDF digestibility, in which the Combo treatment increased from d 0 to d 102 when compared to the CTL and M3NOP treatments, which reduced and maintained the results, respectively (Table 6). No difference ($P \geq 0.07$) for the treatment x time interaction on DM, OM, CP, EE, ADF, and starch. However, treatments were detected an effect ($P \leq 0.05$), in which the Combo treatment increased the digestibility of the nutrients by 1.23% for CP ($P = 0.05$), 4.25% for EE ($P = 0.01$), and 0.88% for starch ($P = 0.03$). No difference ($P \geq 0.08$) for the treatment in the variables DM, OM, and ADF (Table 6).

Table 6. Effect of combining 3-nitrooxypropanol with different feed additives on nutrients intake and digestibility of feedlot Nellore bulls fed a high-concentrate finishing diet in Exp. 2.

Item	CT	M3N	Com	CT	M3N	Com	CT	M3N	Com	SE	P-value			
	L ¹	OP ²	bo ³	L	OP	bo	L	OP	bo		M ⁴	T ⁵	Ti me	T x Time
	d 28			d 56			d 84							
Intake, kg/d														
Dry matter	11.6	11.7	12.4	11.9	12.0	13.1	11.7	11.5	12.5	0.47	0.03	0.46	0.98	
Organic matter	11.0	11.1	11.8	11.3	11.4	12.4	11.1	10.9	11.9	0.47	0.03	0.49	0.98	
Crude protein	1.71	1.73	1.83	1.77	1.77	1.93	1.74	1.70	1.85	0.07	0.02	0.42	0.97	
Ether extract	0.58	0.59	0.63	0.65	0.65	0.70	0.59	0.57	0.59	0.03	0.18	<0.01	0.85	
Acid detergent fiber	2.17	2.21	2.30	2.13	2.16	2.37	2.34	2.30	2.52	0.09	0.02	0.02	0.96	
Neutral detergent fiber	3.07	3.08	3.24	3.09	3.15	3.40	3.13	3.05	3.32	0.12	0.04	0.70	0.96	
Starch	6.07	6.12	6.45	6.24	6.26	6.80	6.01	5.88	6.38	0.24	0.05	0.26	0.99	
Digestibility, g/kg dry matter														
Dry matter	765	770	763	749	747	753	732	749	756	5.24	0.08	<0.01	0.07	
Organic matter	779	787	778	775	769	775	756	771	772	4.68	0.25	<0.01	0.09	
Crude protein	783	785	793	778	769	779	769	782	790	5.59	0.05	0.04	0.29	

Ether extract	748	749	762	738	712	766	69	698	740	14.4	0.0	<0.	0.61
							8			3	1	01	
Acid detergent fiber	435	470	435	390	369	411	43	441	480	16.3	0.2	<0.	0.13
							7			0	8	01	
Neutral detergent fiber	579	572	582	569 _{ab}	544 ^b	589 ^a	55	571 ^{ab}	593 ^a	8.04	<0.	0.2	0.04
							9 ^b				01	9	
Starch	932	934	933	910	913	923	90	908	920	4.06	0.0	<0.	0.48
							6				3	01	

Abbreviations: SEM - standard error of the mean; d - day; DM - dry matter; T – treatments. ¹CTL: Sodium monensin (Rumensin 200®; main composition: sodium monensin 20%, microtracer 0.06%, and excipients q.s.p. 100%; Elanco Ltd., São Paulo, Brazil) at 26 mg/kg DM. ²M3NOP: Sodium monensin at 26 mg/kg DM and 3-NOP (Bovaer®; main composition: 3-NOP (10.5%), propylene glycol (35.2%), and silicic acid (54.3%) [29]; DSM-Firmenich Nutritional Products Ltd., Kaiseraugst, Switzerland) at 100 mg/kg DM. ³Combo: 3-NOP at 100 mg/kg DM, essential oils (Crina® Ruminants; main composition: in 1 g of product - natural or nature-identical aromatic substances (195 mg), including thymol (25%–35%), guaiacol (10%–15%), eugenol (5%–10%), vanillin (10%–20%), salicylaldehyde (5%–10%), and limonene (20%–35%); silicon dioxide (130 mg); lecithins (8 mg); butylated hydroxytoluene (5 mg); organic carriers (662 mg: palm oil, carob flour, starch, monpropylene glycol, stearic acid, and calcium sulfate) [30, 31]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 100 mg/kg DM; 25-Hydroxy-Vitamin-D3 (Hy-D®; main composition: 25-Hydroxy-Vitamin-D3 (103–105%), water (4%), other sterols (0.9–1.7%), and erythrosine (<0.08 mg/kg) [32]; DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil) at 0.10 mg/kg DM (corresponding to 4,000 IU vitamin D3/kg DM); carbo-amino-phospho-chelates chromium at 4 mg/kg DM (main composition: chromium (5%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil); and carbo-amino-phospho-chelates zinc at 60 mg/kg DM (main composition: zinc (24%) and excipients q.s.p. (100%); Tortuga minerals®, DSM-Firmenich Nutritional Products Ltd., São Paulo, Brazil). Superscripts ^a, ^b signify difference with P ≤ 0.05, as determined by the Tukey test.

4. Discussion

Our hypothesis was confirmed that adding 3-NOP, regardless of the combination, would mitigate enteric CH₄ emissions and that the combination of 3-NOP with different feed additives would increase growth performance, FC and FE, in addition to promoting carcass gains and yields with further improvements in the digestibility of nutrients in a high concentrate finishing diet. The use of methane inhibitors may lead to high hydrogen in the rumen, hence why it would be important to consider the use of other feed ingredients to ameliorate rumen health and improve overall performance. Recent research evaluated the possibility of combining different additives to promote growth performance and have improvements in carcass traits [26,20,48,49].

The M3NOP and Combo treatments resulted in significant reductions in CH₄ emissions, both in terms of absolute production, yield, and intensity, compared to the CTL treatment. Researchers are in agreement that the addition of 3-NOP at different doses and conditions to ruminant feed points to its efficiency in reducing CH₄ emissions in beef cattle [50,51,52,28,53,17,54], dairy cows [55,56,57,58] and sheep [59]. The latter authors found that 3-NOP is able reduce enteric CH₄ emissions in different metrics, like daily production, DMI, energy intake, carcass ADG, or in milk production, although not the focus of this research. Mitigation of enteric CH₄ emissions in beef cattle can reach up to 90% with the use of 3-NOP [18]. However, a global review on the use of 3-NOP by Yu et al. [5] suggests that reductions in enteric CH₄ emissions tend to approach 30% in daily production in beef cattle, which corroborates with our findings of 38.8% reduction in daily production. Yu et al. [5] also emphasized that oscillations in enteric CH₄ mitigation depend mainly on the composition of the diet (e.g. % NDF) and the doses of 3-NOP used in the studies. These reductions are attributed to the high molecular specificity of 3-NOP in binding to the enzyme methyl-coenzyme M reductase, inhibiting it. This enzyme is responsible for catalyzing the formation of methane in the last stage of methanogenesis in archaea [16], resulting in the availability of CO₂ and H₂ to the ruminal environment. In this study, a

291% increase in enteric H₂ emissions was found. These findings are complemented by other literature, which also observed reductions in enteric CH₄ emissions and increase in H₂ emissions [27,52,5]. Zhang et al. [28] highlight the importance of experiments investigating the combinations of 3-NOP with other additives, aiming to identify those that, when implemented together, provide better results. Although our findings are not different between the combinations of 3-NOP with different strategies (M3NOP and Combo).

Exp. 1 and 2 are complementary in elucidating the effects of combining 3-NOP with different feed additives. We highlight the improvement in FC of 5.87% and FE of 5.50% in the M3NOP and Combo treatments when compared to the CTL treatment in Exp. 1. This increase in FC and FE may be related to several factors linked to the multiple additives present in the Combo treatment. However, the combination of 3-NOP with sodium monensin in the M3NOP treatment showed better FC and FE compared to CTL, which may have enhanced the effect observed in the Combo treatment. Vyas et al. [60], the addition of 3-NOP promoted an improvement in feed efficiency of 5% in the growing phase in feedlot and of 3% in the finishing phase of beef cattle. We cannot rule out a possible mechanism of hydrogen utilization that stops participating in the formation of CH₄, with part being emitted via H₂ and part as dissolved hydrogen ([H]) in the ruminal content. According to Ungerfeld et al. [61], this [H] can be directed to sinks such as the propionate pathway, consequently driving improvements in feed efficiency and beef cattle growth performance, the increase in rumen [H] concentration shows a negative correlation with acetate levels and a positive correlation with propionate levels [62,63].

As for the higher DMI in Combo bulls, this occurred due to the inclusion of sodium monensin in the CTL and M3NOP treatments, in accordance with Salzar et al. [64] observed higher DMI in dairy calves that received essential oils compared to diets supplemented with sodium monensin. Corroborating our findings, a meta-analysis carried out by Duffield et al. [65] reported a 3% reduction in DMI when a dose of sodium monensin of 28.1 mg/kg DM was supplemented in the diet of beef cattle. Gadberry et al. [66] also performed a meta-analysis with data from heifers and beef cows and observed a DMI reduction effect of around 4.30% at daily doses ranging from 125 to 200 mg/d per animal. The literature is in line with Exp. 1, in which, on d 28, we observed a lower DMI of 5.56% on average for the CTL and 3-NOP treatments (both containing sodium monensin) compared to Combo. In Exp. 2, on d 28, a 6.78% lower DMI was observed for CTL compared to Combo and, on d 102, an 8.20% reduction for the average of CTL and 3-NOP treatments compared to Combo. These findings in Exp. 1 and 2, despite the alignment observed in the literature on treatments containing sodium monensin in the reduction of DMI, were slightly higher, probably due to the stimulation of DMI by essential oils or other components contained in Combo, and the reduction observed with the addition of sodium monensin to the diet of beef cattle. Consequently, higher BW and ADG were observed in the Combo treatment in Exp. 1, due to the metrics of FC, FE and higher DMI in relation to the other treatments. The literature demonstrates increases in BW and ADG when essential oils [26,20,67] and chromium [68,69,70,71] are included in beef cattle diets. The complementarity of the results in Exp. 1 and 2 in growth performance is also observed in carcass traits, with the Combo treatment being superior to the others. In the CTL and M3NOP treatments, which contain sodium monensin, no effect was observed in stimulating carcass deposition or improving intake and efficiency characteristics [65]. In contrast, Combo contained in its composition several elements with proven capacity to increase carcass gains and yields. However, the essential oil (containing thymol, eugenol, limonene and vanillin) included in the diet of feedlot beef cattle results in carcass traits like those presented by animals that received sodium monensin [72, 26], highlighting the essential oil as a viable alternative for use in ruminants. Another component of Combo is 25-Hydroxy-Vitamin-D3, which, when included at a dose of 1 mg/animal/day in the finishing diet of Nellore cattle, increased yield by 0.96% [73] and an increase of 4.2 kg in HCW [74]. In addition, the inclusion of 25-Hydroxy-Vitamin-D3 in finishing diets for feedlot beef cattle increased the expression of the Insulin-like growth factor (IGF)-1, IGF-2, mammalian target of rapamycin (mTOR), and myostatin (MSTN) genes, genes related to muscle anabolism, protein turnover, and regulation of muscle growth [75].

One challenge for researchers using complex mixtures is that many may include proprietary formulations. It can be difficult to use analytical techniques like gas or liquid chromatography-mass spectrometry. These methods may not be feasible, suitable, or may involve lengthy sample preparations and separations. Additionally, techniques like colorimetric assays require the development of specific methods for each compound. Batley et al [76] suggested the use of fourier transform infrared-attenuated total reflectance spectroscopy to efficiently identify compounds by generating unique spectra in a faster and more cost-effective manner. Another element presents in the Combo treatment that may have contributed to better carcass traits was chromium, when included in feedlot diets for beef cattle, an increase in HCW was observed [69, 22, 70, 23]. However, there are studies in the literature indicating that the addition of chromium in finishing diets for beef cattle, compared to the control group, did not show an increase in HCW [77,78,79,80,25]. This same inconsistency is also reported in zinc administration, with some studies showing an increase in carcass traits [81], while others show no effect in relation to the control treatment [78,22,23]. However, the intentional use of a mixture containing chromium and zinc increases HCW, as observed in our Exp. 1 and 2, with a consequent increase in gain and carcass yield characteristics. In view of this, Budde et al. [22], studying the effect of including chromium and zinc in combination in feedlot cattle diets, compared it with animals that received only zinc and observed an increase of 8.5 kg in HCW. These findings corroborate our results in HCW found in Exp. 1 and 2, where there was an increase of 12 and 14 kg, respectively. These superior results are probably the effect of the combination of additives that allowed better results in terms of carcass. Combo improved the carcass pH range 24 h post-mortem, as found by Matthews et al. [82], who reported lower pH values at 24 h for pigs supplemented during finishing with chromium chelated with the amino acid methionine. In addition, there is the possibility that the chromium mechanism acts as a stress-mitigating element, reducing cortisol secretion [83], which could mitigate the stress of transportation to the slaughterhouse, which interferes with carcass pH values [84].

In Exp. 2, all treatments showed an increase in serum glucose and insulin concentrations over time. However, the Combo treatment, as described in the results, showed lower glucose and insulin concentrations compared to the CTL and M3NOP treatments from d 28 to d 102. This lower concentration suggests lower insulin resistance, which may promote greater DMI during the feedlot phase and increased carcass traits such as HCW, a result found and previously discussed in the Combo treatment in Exp. 1 and 2. Therefore, there is evidence that minerals, such as chromium and zinc, contained in the Combo treatment, are effective in improving glucose tolerance and reducing insulin resistance in beef cattle [85,86,87,25], increasing insulin binding and the number of insulin receptor receptors and enzymes, promoting insulin sensitivity through β -cells and facilitating its internalization [88, 89], positively regulating gluconeogenesis [25]. Insulin reduces gluconeogenesis by suppressing the secretion of glucagon from the pancreas [90].

AST and cholesterol parameters were compared to reference values [91,92] and demonstrated normal concentrations. However, the Combo treatment, as illustrated in the results, presented lower AST and cholesterol concentrations. The literature highlights potential health benefits linked to minerals like chromium and zinc in terms of AST parameters related to liver health [93], corroborating our findings in Exp. 2 for the Combo treatment. Soumar et al. [24] included chromium and zinc in the diet of lambs before transport and observed that in all treatments there were increases in AST concentration after transport, but the supplemented animals had lower AST concentrations compared to the CTL treatment, indicating an enhancement in the health indicator. This suggests that, possibly, the inclusion of chromium and zinc in the Combo treatment improves the health of bulls in feedlot situations under a high concentrate diet. The serum albumin concentration in the Combo treatment remained unchanged on d 28 in relation to the initial concentration, while in the CTL and M3NOP treatments there was an increase. In addition, albumin in the Combo treatment increased on d 102, although the concentrations remained lower than those observed in the CTL and M3NOP treatments, corroborating Hassan et al. [94] showed lower albumin concentrations in rats supplemented with the combination of chromium and zinc in the diet, associated with reduced serum AST concentrations. Additionally, regarding cholesterol concentrations, we observed that minerals

such as chromium and zinc can also influence lipid metabolism, reducing cholesterol levels [93, 94]. Soumar et al. [24] found that the inclusion of chromium and zinc led to a reduction in cholesterol concentrations in animals. Experiments with rats show that minerals like chromium and zinc can promote the reduction of serum levels of AST, albumin and total cholesterol, in addition to effects on insulin sensitivity and resistance [94,95].

As previously discussed, bulls in the Combo group had higher DMI compared to CTL and M3NOP, which in turn resulted in higher intake of OM, CP, NDF, ADF and starch (Table 6). This condition, depending on digestibility, can favor growth performance and, consequently, improve carcass traits. Meschiatti et al. [26] showed that treatment containing essential oils, compared to sodium monensin in feedlot beef cattle, resulted in higher intake of DM, CP, NDF and starch. Furthermore, Toseti et al. [20] demonstrated that the inclusion of a combination of essential oils together with alpha amylase in diets of finishing beef cattle increased the intake of DM, OM, CP, NDF, ADF, EE and starch compared to the inclusion of sodium monensin. Nutrient digestibility was higher in bulls that received Combo, specifically for CP, EE, NDF, and starch. One possible explanation is the action of the essential oil present in this treatment, which can positively affect nutrient digestibility. Toseti et al. [20] demonstrated greater digestibility of CP and EE in finishing beef cattle. Li et al. [96] reported that the inclusion of essential oil in an in vitro experiment resulted in increased NDF digestibility. The presence of 3-NOP may have enhanced the improved effects on digestibility values observed in Combo. Several studies in the literature show positive results of 3-NOP in ruminants, including increased digestibility of CP [11,27,28], NDF [56,97] and starch [28] in the diet. Although 3-NOP is present in the M3NOP treatment and does not show significant differences compared to the CTL treatment, it may be related to the similarity in DMI between the CTL and M3NOP treatments. Moreover, sodium monensin has the main effect of reducing DMI in beef cattle. [65,66].

5. Conclusions

The addition of 3-NOP to our experimental diets, both with sodium monensin or with a feed additive package containing essential oil, 25-Hydroxy-Vitamin-D3 and carbo-amino-phospho-chelates minerals such as chromium and zinc, proved to be an effective mitigator of enteric CH₄ emissions, with an average reduction of 38.8%, promoting sustainability within the beef cattle feedlot operation. Furthermore, including 3-NOP with sodium monensin improved feed conversion and feed efficiency compared to exclusive inclusion of sodium monensin in high-concentrate diets. The combination of 3-NOP with a feed additive package containing essential oil, 25-Hydroxy-Vitamin-D3 and carbo-amino-phospho-chelates minerals such as chromium and zinc, positively affected several parameters, such as growth performance and carcass traits, resulting in improved intake and digestibility of nutrients, accompanied by changes in the responses to serum glucose, insulin, albumin and cholesterol, indicating improved glucose tolerance and reduced insulin resistance. Furthermore, improved aspartate aminotransferase parameters were observed, a crucial indicator of liver health in finishing beef cattle under high-concentrate diets. It is suggested to investigate the effects of 3-NOP in different diets and its interaction with other feed additives, as well as its impact on health, growth performance, and carcass traits in different phases of cattle production, to broaden the understanding of best practices to maximize efficiency and sustainability in beef cattle production.

Author Contributions: Conceptualization: W.L.S., V.V.C., L.F.M.T., M.K., T.S.A., R.A.R.; Methodology: W.L.S., V.V.C., L.F.M.T., M.K., A.P., A.S.C.; Validation: W.L.S., M.B.N., R.A.R.; Formal analysis: W.L.S., A.S.C., M.B.N.; Investigation: W.L.S.; M.B.N., A.P., R.A.R.; Resources: R.A.R., V.V.C.; Data curation: W.L.S., A.S.C., M.B.N.; Writing—original draft preparation: W.L.S.; Writing—review and editing: W.L.S., V.V.C., L.F.M.T., M.K., A.S.C., D.F.A.C., R.A.R.; Visualization: W.L.S., M.B.N., A.P., R.A.R.; Supervision: W.L.S., V.V.C., L.F.M.T., M.K., R.A.R.; Project administration: R.A.R.; Funding acquisition: R.A.R. All authors have read and agreed to the published version of the manuscript.

Funding: This project was supported by the Department of Innovation and Applied Science, DSM-firmenich. First author's scholarship funded by Coordination for the Improvement of Higher Education Personnel (CAPES 88887.610201/2021-00).

Institutional Review Board Statement: The experimental procedures were approved by the DSM Nutritional Products SA Ethics Committee on Animal Use (numbers BR 220121 and BR 211215), in accordance with the São Paulo State University's Animal Research Ethics Committee guidelines.

Informed Consent Statement: Not applicable.

Data Availability Statement: All data supporting the reported results are available and can be requested directly from the corresponding author.

Acknowledgments: This study is part of the PhD. thesis of the first author. We would like to express our gratitude to the UnespFor team at São Paulo State University, Jaboticabal campus; the Graduate Program in Animal Science at São Paulo State University, Faculty of Agricultural and Veterinary Sciences, Jaboticabal campus, where the first author conducted his PhD research; DSM-Firmenich for sponsoring the experiment; The staff from Fazenda Caçadinha for their support during the experimental work; and the Coordination for the Improvement of Higher Education Personnel for funding the PhD scholarship.

Conflicts of Interest: The authors have read the journal's guideline and have the following competing interests: the co-authors Victor Valério de Carvalho, Alexandre Perdigão, Tiago Sabella Acedo, Luis Fernando Monteiro Tamassia and Maik Kindermann are employees of DSM-Firmenich. The other authors have no competing interests.

References

- Ornaghi, M.G.; Guerrero, A.; Vital, A.C.P.; De Souza, K.A.; Passetti, R.A.C.; Mottin, C.; De Araújo Castilho, R.; Sañudo, C.; Do Prado, I.N. Improvements in the quality of meat from beef cattle fed natural additives. *Meat Sci.* **2020**, *163*, 108059. <https://doi.org/10.1016/j.meatsci.2020.108059>.
- Greenwood, P.L. Review: An overview of beef production from pasture and feedlot globally, as demand for beef and the need for sustainable practices increase. *Animal* **2021**, *15*, 100295. <https://doi.org/10.1016/j.animal.2021.100295>.
- Alexandratos, N.; Bruinsma, J. *World Agriculture: Towards 2030/2050*; ESA Working Paper No. 12-03; FAO: Rome, Italy, 2012. <https://doi.org/10.22004/ag.econ.288998>.
- Cooke, R.F.; Daigle, C.L.; Moriel, P.; Smith, S.B.; Tedeschi, L.O.; Vendramini, J.M.B. Cattle adapted to tropical and subtropical environments: social, nutritional, and carcass quality considerations. *J. Anim. Sci.* **2020**. <https://doi.org/10.1093/jas/skaa014>.
- Yu, G.; Beauchemin, K.A.; Dong, R. A review of 3-nitrooxypropanol for enteric methane mitigation from ruminant livestock. *Animals* **2021**, *11*, 3540. <https://doi.org/10.3390/ani11123540>.
- Gerber, P.J.; Hristov, A.N.; Henderson, B.; Makkar, H.; Oh, J.; Lee, C.; Meinen, R.; Montes, F.; Ott, T.; Firkins, J.; Dell Al Rotz, C.; et al. Technical options for the mitigation of direct methane and nitrous oxide emissions from livestock – A review. *Anim.* **2013**, *7* (Suppl. 2), 220–234. <https://doi.org/10.1017/S1751731113000876>.
- Nibedita, S.; Swati, P.; Pattnaik, M.; Mohapatra, S. Methane emission and strategies for mitigation in livestock. In *Environmental and Agricultural Microbiology: Applications for Sustainability*; pp. 257–274. <https://doi.org/10.1002/9781119525899.ch12>.
- Detmann, E.; Paulino, M.F.; Valadares Filho, S.C.; Huhtanen, P. Nutritional aspects applied to grazing cattle in the tropics: A review based on Brazilian results. *Semina: Ciênc. Agrár.* **2014**, *35*, 2829–2854. <https://doi.org/10.5433/1679-0359.2014v35n4Suplp2829>.
- Cardoso, A.S.; Barbero, R.P.; Romanzini, E.P.; Teobaldo, R.W.; Ongaratto, F.; Fernandes, M.H.M.R.; Ruggieri, A.C.; Reis, R.A. Intensification: A key strategy to achieve great animal and environmental beef cattle production sustainability in Brachiaria grasslands. *Sustainability* **2020**, *12*, 6656. <https://doi.org/10.3390/su12166656>.
- Sousa, L.M.; de Souza, W.L.; Oliveira, K.A.; Cidrini, I.A.; Moriel, P.; Nogueira, H.C.R.; Ferreira, I.M.; Ramirez-Zamudio, G.D.; Oliveira, I.M.d.; Prados, L.F.; et al. Effect of Different Herbage Allowances from Mid to Late Gestation on Nellore Cow Performance and Female Offspring Growth until Weaning. *Animals* **2024**, *14*, 163. <https://doi.org/10.3390/ani14010163>.
- Hristov, A.; Oh, J.; Giallongo, F.; Frederick, T.; Harper, M.; Weeks, H.; Branco, A.; Moate, P.; Deighton, M.; Williams, S. An inhibitor persistently decreased enteric methane emission from dairy cows with no negative effect on milk production. *Proc. Natl. Acad. Sci. U.S.A.* **2015**, *112*, 10663–10668. <https://doi.org/10.1073/pnas.1504124112>.
- Min, B.R.; Lee, S.; Jung, H.; Miller, D.N.; Chen, R. Enteric methane emissions and animal performance in dairy and beef cattle production: Strategies, opportunities, and impact of reducing emissions. *Animals* **2022**, *12*, 948. <https://doi.org/10.3390/ani12080948>.
- Johnson, K.A.; Johnson, D.E. Methane emissions from cattle. *J. Anim. Sci.* **1995**, *73*, 2483–2492. <https://doi.org/10.2527/1995.7382483x>.

14. Honan, M.; Feng, X.; Tricarico, J.M.; Kebreab, E. Feed additives as a strategic approach to reduce enteric methane production in cattle: Modes of action, effectiveness and safety. *Anim. Prod. Sci.* **2021**, *61*, 15. <https://doi.org/10.1071/AN20295>.
15. Kelly, L.; Kebreab, E. Recent advances in feed additives with the potential to mitigate enteric methane emissions from ruminant livestock. *J. Soil Water Conserv.* **2023**, *78*, 111–123. <https://doi.org/10.2489/jswc.2023.00070>.
16. Duin, E.; Wagner, T.; Shima, S.; Prakash, D.; Cronin, B.; Yáñez-Ruiz, D.; Duval, S.; Rumbeli, R.; Stemmler, R.; Thauer, R. Mode of action uncovered for the specific reduction of methane emissions from ruminants by the small molecule 3-nitrooxypropanol. *Proc. Natl. Acad. Sci. U.S.A.* **2016**, *113*, 6172–6177. <https://doi.org/10.1073/pnas.1600298113>.
17. Almeida, A.K.; Cowley, F.; McMeniman, J.P.; Karagiannis, A.; Walker, N.; Tamassia, L.F.; McGrath, J.J.; Hegarty, R.S. Effect of 3-Nitrooxypropanol on Enteric Methane Emissions of Feedlot Cattle Fed with a Tempered Barley-Based Diet with Canola Oil. *J. Anim. Sci.* **2023**, *101*. <https://doi.org/10.1093/jas/skad237>.
18. Beauchemin, K.A.; Ungerfeld, E.M.; Eckard, R.J.; Wang, M. Fifty Years of Research on Rumen Methanogenesis: Lessons Learned and Future Challenges for Mitigation. *Animal* **2020**, *14*. <https://doi.org/10.1017/S1751731119003100>.
19. Souza, W.L.d.; Romanzini, E.P.; Delevatti, L.M.; Leite, R.G.; Bernardes, P.A.; Cardoso, A.S.; Reis, R.A.; Malheiros, E.B. Economic evaluation of nitrogen fertilization levels in beef cattle production: Implications for sustainable tropical pasture management. *Agriculture* **2023**, *13*, 2233. <https://doi.org/10.3390/agriculture13122233>.
20. Toseti, L.B.; Goulart, R.S.; Gouvêa, V.N.; Acedo, T.S.; Vasconcellos, G.S.F.M.; Pires, A.V. Effects of a blend of essential oils and exogenous α -amylase in diets containing different roughage sources for finishing beef cattle. *Anim. Feed Sci. Technol.* **2020**, *269*, 114643. <https://doi.org/10.1016/j.anifeedsci.2020.114643>.
21. Estrada-Angulo, A.; Mendoza-Cortéz, D.A.; Ramos-Méndez, J.L.; Arteaga-Wences, Y.; Uriás-Estrada, J.D.; Castro-Pérez, B.I.; Ríos-Rincón, F.G.; Rodríguez-Gaxiola, M.A.; Barreras, A.; Zinn, R.A. Comparing blend of essential oils plus 25-hydroxy-vit-D3 versus monensin plus virginiamycin combination in finishing feedlot cattle: Growth performance, dietary energetics, and carcass traits. *Animals* **2022**, *12*, 1715. <https://doi.org/10.3390/ani12131715>.
22. Budde, A.M.; Sellins, K.; Karen, E.L.; Wagner, J.J.; Heldt, J.S.; Spears, J.W.; Engle, T.E. Effect of Zinc Source and Concentration and Chromium Supplementation on Performance and Carcass Traits in Feedlot Steers. *J. Anim. Sci.* **2019**, *97*, 1286–1295. <https://doi.org/10.1093/jas/skz016>.
23. Hallmark, H.D.; Zervoudakis, J.T.; Torrecilhas, J.A.; Hatamoto-Zervoudakis, L.K.; Toller, H.; Guimaraes, O.; Engle, T.E. PSII-19 Effect of zinc and chromium supplementation on performance and carcass traits in feedlot steers. *J. Anim. Sci.* **2020**, *98*, 400–401. <https://doi.org/10.1093/jas/skaa278.703>.
24. Soumar, S.K.; Hozhabri, F.; Moeini, M.M.; Nikousefat, Z. Impacts of feeding zinc-methionine or chromium-methionine on performance, antioxidant status and physiological responses to transportation stress on lambs. *Anim. Prod. Sci.* **2020**, *60*, 796–805. <https://doi.org/10.1071/AN18070>.
25. Trojan, S.J.; Hergenreder, J.E.; Canterbury, L.G.; Leonhard, J.T.; Clark, W.D.; Beckett, J.L.; Long, J.M. The effects of chromium propionate supplementation to yearling steers in a commercial feedyard on growth performance, carcass traits, and health. *Transl. Anim. Sci.* **2023**, *7*. <https://doi.org/10.1093/tas/txad078>.
26. Meschiatti, M.A.; Gouvêa, V.N.; Pellarin, L.A.; Batalha, C.D.; Biehl, M.V.; Acedo, T.S.; Dórea, J.R.; Tamassia, L.F.; Owens, F.N.; Santos, F.A. Feeding the Combination of Essential Oils and Exogenous Alpha Amylase Increases Performance and Carcass Production of Finishing Beef Cattle. *J. Anim. Sci.* **2019**, *97*, 456–471. <https://doi.org/10.1093/jas/sky415>.
27. Melgar, A.; Harper, M.T.; Oh, J.; Giallongo, F.; Young, M.E.; Ott, T.L.; Duval, S.; Hristov, A.N. Effects of 3-Nitrooxypropanol on Rumen Fermentation, Lactational Performance, and Resumption of Ovarian Cyclicity in Dairy Cows. *J. Dairy Sci.* **2020**, *103*, 410–432. <https://doi.org/10.3168/jds.2019-17085>.
28. Zhang, X.; Smith, M.; Gruninger, R.; Kung, L.; Vyas, D.; McGinn, S.; Kindermann, M.; Wang, M.; Tan, Z.; Beauchemin, K. Combined Effects of 3-Nitrooxypropanol and Canola Oil Supplementation on Methane Emissions, Rumen Fermentation and Biohydrogenation, and Total Tract Digestibility in Beef Cattle. *J. Anim. Sci.* **2021**, *99*. <https://doi.org/10.1093/jas/skab081>.
29. EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP); Bampidis, V.; Azimonti, G.; Bastos, M.D.L.; Christensen, H.; Dusemund, B.; Durjava, M.; Kouba, M.; López-Alonso, M.; López Puente, S.; Marcon, F. Safety and efficacy of a feed additive consisting of 25-hydroxycholecalciferol monohydrate produced with *Saccharomyces cerevisiae* CBS 146008 for all ruminants (DSM Nutritional Products Sp. zoo). *EFSA J.* **2023**, *21*, e08169. <https://doi.org/10.2903/j.efsa.2023.8169>.
30. Rossi, J., Crina S.A., 1996. Additives for animal nutrition and technique for their preparation. *U.S. Patent 5,558,889*.
31. DSM Nutritional Products, 2016. Product information: Crina Protect Composition. *DSM Nutritional Products*, Parsippany, NJ.

32. EFSA Panel on Additives and Products or Substances used in Animal Feed (FEEDAP), Bampidis, V., Azimonti, G., Bastos, M.D.L., Christensen, H., Dusemund, B., Durjava, M., Kouba, M., López-Alonso, M., López Puente, S., Marcon, F., 2023. Safety and efficacy of a feed additive consisting of 25-hydroxycholecalciferol monohydrate produced with *Saccharomyces cerevisiae* CBS 146008 for all ruminants (DSM Nutritional Products Sp. zoo). *EFSA Journal*, 21(8), e08169. <https://doi.org/10.2903/j.efsa.2023.8169>.
33. Lammers, B.P.; Buckmaster, D.R.; Heinrichs, A.J. A Simple Method for the Analysis of Particle Sizes of Forage and Total Mixed Rations. *J. Dairy Sci.* **1996**, 79, 922–928. [https://doi.org/10.3168/jds.S0022-0302\(96\)76442-1](https://doi.org/10.3168/jds.S0022-0302(96)76442-1).
34. Association of Official Analytical Chemists (AOAC). *Official Methods of Analysis of AOAC International*, 18th ed.; AOAC International: Gaithersburg, MD, USA, 2006.
35. Van Soest, P.J.; Robertson, J.B.; Lewis, B.A. Methods for Dietary Fiber, Neutral Detergent Fiber, and Nonstarch Polysaccharides in Relation to Animal Nutrition. *J. Dairy Sci.* **1991**, 74, 3583–3597. [https://doi.org/10.3168/jds.S0022-0302\(91\)78551-2](https://doi.org/10.3168/jds.S0022-0302(91)78551-2).
36. Hall, M.B. Determination of Starch, Including Maltooligosaccharides, in Animal Feeds: Comparison of Methods and a Method Recommended for AOAC Collaborative Study. *J. AOAC Int.* **2009**, 92, 42–49. <https://doi.org/10.1093/jaoac/92.1.42>.
37. NRC. *Nutrient Requirements of Dairy Cattle*, 7th rev. ed.; National Academies Press: Washington, DC, USA, 2001.
38. NASEM. *Nutrient Requirements of Beef Cattle*, 8th ed.; National Academies Press: Washington, DC, USA, 2016.
39. CIE. Recommendations on Uniform Color Spaces-Color Equations, Psychometric Color Terms. In Proceedings of the Commission Internationale de l'Eclairage; CIE: Paris, France, 1986.
40. AMSA-American Meat Science Association. Research Guidelines for Cookery, Sensory Evaluation, and Instrumental Tenderness Measurements of Fresh Meat. *Proceedings of the American Meat Science Association*; AMSA: Chicago, IL, USA, 1995. Available online: https://meatscience.org/docs/default-source/publications-resources/research-guide/amsa-research-guidelines-for-cookery-and-evaluation-1-02.pdf?sfvrsn=4c6b8eb3_2 (accessed on 3 September 2024).
41. Warriss, P.D. The Extraction of Haem Pigments from Fresh Meat. *J. Food Technol.* **1979**, 14, 75–80. <https://doi.org/10.1111/j.1365-2621.1979.tb00849.x>.
42. Hunt, M.C.; Sørheim, O.; Slinde, E. Color and Heat Denaturation of Myoglobin Forms in Ground Beef. *J. Food Sci.* **1999**, 64, 847–851. <https://doi.org/10.1111/j.1365-2621.1999.tb15925.x>.
43. Bligh, E.G.; Dyer, W.J. A Rapid Method of Total Lipid Extraction and Purification. *Can. J. Biochem. Physiol.* **1959**, 37, 911–917. <https://doi.org/10.1139/o59-099>.
44. Sørensen, G.; Storgaard Jørgensen, S. A Critical Examination of Some Experimental Variables in the 2-Thiobarbituric Acid (TBA) Test for Lipid Oxidation in Meat Products. *Eur. Food Res. Technol.* **1996**, 202, 205–210.
45. Cole, N.A.; McCuiston, K.; Greene, L.W.; McCollum, F.T. Effects of Concentration and Source of Wet Distiller's Grains on Digestibility of Steam-Flaked Corn-Based Diets Fed to Finishing Steers. *Prof. Anim. Sci.* **2011**, 27, 302–311. [https://doi.org/10.15232/S1080-7446\(15\)30493-9](https://doi.org/10.15232/S1080-7446(15)30493-9).
46. Krizsan, S.J.; Huhtanen, P. Effect of Diet Composition and Incubation Time on Feed Indigestible Neutral Detergent Fiber Concentration in Dairy Cows. *J. Dairy Sci.* **2013**, 96, 1715–1726. <https://doi.org/10.3168/jds.2012-5752>.
47. Della Rosa, M.M.; Jonker, A.; Waghorn, G.C. A Review of Technical Variations and Protocols Used to Measure Methane Emissions from Ruminants Using Respiration Chambers, SF6 Tracer Technique, and GreenFeed, to Facilitate Global Integration of Published Data. *Anim. Feed Sci. Technol.* **2021**, 279, 115018. <https://doi.org/10.1016/j.anifeedsci.2021.115018>.
48. Webb, M.J.; Block, J.J.; Harty, A.A.; Salverson, R.R.; Daly, R.F.; Jaeger, J.R.; Underwood, K.R.; Funston, D.P.; Pendell, C.A.; Rotz, C.A.; et al. Cattle and Carcass Performance, and Life Cycle Assessment of Production Systems Utilizing Additive Combinations of Growth Promotant Technologies. *Transl. Anim. Sci.* **2020**, 4, 1–15. <https://doi.org/10.1093/tas/txaa216>.
49. Carvalho, V.V. de; Souza, W.L.L.; Perdigão, A.; Niehues, M.B.; Matos, I.E.; Ribeiro, M.M.; Acedo, T.S.; Tamassia, L.; Walker, N.; Kindermann, M.; Cardoso, A. Silva.; Reis, R. Combination of Feed Additives to Reduce Methane Emissions and Increase Performance by Feedlot Cattle. *J. Anim. Sci.* **2023**, 101 (Supplement_3), 275–276. <https://doi.org/10.1093/jas/skad281.330>.
50. Vyas, D.; McGinn, S.; Duval, S.; Kindermann, M.; Beauchemin, K. Effects of Sustained Reduction of Enteric Methane Emissions with Dietary Supplementation of 3-Nitrooxypropanol on Growth Performance of Growing and Finishing Beef Cattle. *J. Anim. Sci.* **2016**, 94, 2024–2034. <https://doi.org/10.2527/jas.2015-0268>.
51. Martinez-Fernandez, G.; Duval, S.; Kindermann, M.; Schirra, H.J.; Denman, S.E.; McSweeney, C.S. 3-NOP vs. Halogenated Compound: Methane Production, Ruminant Fermentation and Microbial Community Response in Forage Fed Cattle. *Front. Microbiol.* **2018**, 9, 1582. <https://doi.org/10.3389/fmicb.2018.01582>.

52. Alemu, A.; Shreck, A.; Booker, C.; McGinn, S.; Pekrul, L.; Kindermann, M.; Beauchemin, K. Use of 3-Nitrooxypropanol in a Commercial Feedlot to Decrease Enteric Methane Emissions from Cattle Fed a Corn-Based Finishing Diet. *J. Anim. Sci.* **2021**, *99*. <https://doi.org/10.1093/jas/skaa394>.
53. Araújo, T.L.; Rabelo, C.H.; Cardoso, A.S.; Carvalho, V.V.; Acedo, T.S.; Tamassia, L.F.; Vasconcelos, G.S.; Duval, S.M.; Kindermann, M.; Gouvêa, V.N.; Fernandes, M.H. Feeding 3-Nitrooxypropanol Reduces Methane Emissions by Feedlot Cattle Under Tropical Conditions. *J. Anim. Sci.* **2023**, *101*. <https://doi.org/10.1093/jas/skad225>.
54. Pedrini, C.A.; Machado, F.S.; Fernandes, A.R.M.; Cônsolo, N.R.B.; Ocampos, F.M.M.; Colnago, L.A.; Perdigão, A.; de Carvalho, V.V.; Acedo, T.S.; Tamassia, L.F.M.; et al. Performance, Meat Quality and Meat Metabolomics Outcomes: Efficacy of 3-Nitrooxypropanol in Feedlot Beef Cattle Diets. *Animals* **2024**, *14*, 2576. <https://doi.org/10.3390/ani14172576>.
55. Haisan, J.; Sun, Y.; Guan, L.; Beauchemin, K.; Iwaasa, A.; Duval, S.; Barreda, D.; Oba, M. The Effects of Feeding 3-Nitrooxypropanol on Methane Emissions and Productivity of Holstein Cows in Mid Lactation. *J. Dairy Sci.* **2014**, *97*, 3110–3119. <https://doi.org/10.3168/jds.2013-7834>.
56. Haisan, J.; Sun, Y.; Guan, L.; Beauchemin, K.A.; Iwaasa, A.; Duval, S.; Kindermann, M.; Barreda, D.R.; Oba, M. The Effects of Feeding 3-Nitrooxypropanol at Two Doses on Milk Production, Rumen Fermentation, Plasma Metabolites, Nutrient Digestibility, and Methane Emissions in Lactating Holstein Cows. *Anim. Prod. Sci.* **2017**, *57*, 282–289. <https://doi.org/10.1071/AN15219>.
57. Melgar, A.; Lage, C.; Nedelkov, K.; Räisänen, S.; Stefenoni, H.; Fetter, M.; Chen, X.; Oh, J.; Duval, S.; Kindermann, M.; et al. Enteric Methane Emission, Milk Production, and Composition of Dairy Cows Fed 3-Nitrooxypropanol. *J. Dairy Sci.* **2021**, *104*, 357–366.
58. Schilde, M.; von Soosten, D.; Hüther, L.; Kersten, S.; Meyer, U.; Zeyner, A.; Dänicke, S. Dose–Response Effects of 3-Nitrooxypropanol Combined with Low- and High-Concentrate Feed Proportions in the Dairy Cow Ration on Fermentation Parameters in a Rumen Simulation Technique. *Animals* **2021**, *11*, 1784. <https://doi.org/10.3390/ani11061784>.
59. Martínez-Fernández, G.; Abecia, L.; Arco, A.; Cantalapiedra-Hijar, G.; Martín-García, A.; Molina-Alcaide, E.; Kindermann, M.; Duval, S.; Yáñez-Ruiz, D. Effects of Ethyl-3-Nitrooxy Propionate and 3-Nitrooxypropanol on Ruminal Fermentation, Microbial Abundance, and Methane Emissions in Sheep. *J. Dairy Sci.* **2014**, *97*, 3790–3799. <https://doi.org/10.3168/jds.2013-7398>.
60. Vyas, D.; Alemu, A.W.; McGinn, S.M.; Duval, S.M.; Kindermann, M.; Beauchemin, K.A. The Combined Effects of Supplementing Monensin and 3-Nitrooxypropanol on Methane Emissions, Growth Rate, and Feed Conversion Efficiency in Beef Cattle Fed High-Forage and High-Grain Diets. *J. Anim. Sci.* **2018**, *96*, 2923–2938. <https://doi.org/10.1093/jas/sky174>.
61. Ungerfeld, E.M. Metabolic Hydrogen Flows in Rumen Fermentation: Principles and Possibilities of Interventions. *Front. Microbiol.* **2020**, *11*, 589. <https://doi.org/10.3389/fmicb.2020.00589>.
62. Wang, M.; Wang, R.; Zhang, X.; Ungerfeld, E.M.; Long, D.; Mao, H.; Jiao, J.; Beauchemin, K.A.; Tan, Z. Molecular Hydrogen Generated by Elemental Magnesium Supplementation Alters Rumen Fermentation and Microbiota in Goats. *Br. J. Nutr.* **2017**, *118*, 401–410. <https://doi.org/10.1017/S0007114517002161>.
63. Wang, R.; Wang, M.; Ungerfeld, E.M.; Zhang, X.M.; Long, D.L.; Mao, H.X.; Deng, J.P.; Bannink, A.; Tan, Z.L. Nitrate Improves Ammonia Incorporation into Rumen Microbial Protein in Lactating Dairy Cows Fed a Low-Protein Diet. *J. Dairy Sci.* **2018**, *101*, 9789–9799. <https://doi.org/10.3168/jds.2018-14904>.
64. Salzar, L.F.; Nero, L.A.; Campos-Galvão, M.E.; Cortinhas, C.S.; Acedo, T.S.; Tamassia, L.F.; Busato, K.C.; Morais, V.C.; Rotta, P.P.; Silva, A.L.; Marcondes, M.I. Effect of Selected Feed Additives to Improve Growth and Health of Dairy Calves. *PLoS ONE* **2019**, *14*, e0216066. <https://doi.org/10.1371/journal.pone.0216066>.
65. Duffield, T.F.; Merrill, J.K.; Bagg, R.N. Meta-Analysis of the Effects of Monensin in Beef Cattle on Feed Efficiency, Body Weight Gain, and Dry Matter Intake. *J. Anim. Sci.* **2012**, *90*, 4583–4592. <https://doi.org/10.2527/jas.2011-5018>.
66. Gadberry, S.; Beck, P.; Moore, M.; White, F.; Linneen, S.; Lalman, D. Meta-analysis of the effects of monensin on performance of beef replacement heifers and beef cows. *Transl. Anim. Sci.* **2022**, *6*. <https://doi.org/10.1093/tas/txac086>.
67. Silva, T.I.; Souza, J.M.; Acedo, T.S.; Carvalho, V.V.; Perdigão, A.; Silva, L.A.; Silvestre, A.M.; Niehues, M.B.; Schleifer, W.F.; Casali, D.M.; Martins, C.L. Feedlot performance, rumen and cecum morphometrics of Nelore cattle fed increasing levels of diet starch containing a blend of essential oils and amylase or monensin. *Front. Vet. Sci.* **2023**, *10*, 1090097. <https://doi.org/10.3389/fvets.2023.1090097>.
68. Barajas, R.; Cervantes, B.J.; Velazquez, E.A.; Romo, J.A.; Juarez, F.; Rojas, P.J. Cr Methionine Supplementation on Feedlot Performance and Carcass Traits of Bulls: I. Results During the Cool Season in the Northwest of Mexico. *Proc. West. Sect. Am. Soc. Anim. Sci.* **2008**, *59*, 383–386.
69. Barajas, R.; Cervantes, B.J.; Velazquez, E.A.; Romo, J.A.; Juarez, F.; Rojas, P.J.; Peña, F.R. Chromium Methionine Supplementation on Feedlot Performance and Carcass Traits of Bulls: II. Results Including

- through Hot and Humid Season in the Northwest of Mexico. *Proc. West. Sect. Am. Soc. Anim. Sci.* **2008**, *59*, 374–375.
70. Bernhard, B.C.; Burdick, N.C.; Rounds, W.; Rathmann, R.J.; Carroll, J.A.; Finck, D.N.; Jennings, M.A.; Young, T.R.; Johnson, B.J. Chromium Supplementation Alters the Performance and Health of Feedlot Cattle During the Receiving Period and Enhances Their Metabolic Response to a Lipopolysaccharide Challenge. *J. Anim. Sci.* **2012**, *90*, 3879–3888. <https://doi.org/10.2527/jas.2011-4981>.
 71. Baggerman, J.O.; Smith, Z.K.; Thompson, A.J.; Kim, J.; Hergenreder, J.E.; Rounds, W.; Johnson, B.J. Chromium Propionate Supplementation Alters Animal Growth Performance, Carcass Traits, and Skeletal Muscle Properties in Feedlot Steers. *Transl. Anim. Sci.* **2020**, *4*. <https://doi.org/10.1093/tas/txaa146>.
 72. Meyer, N.F.; Erickson, G.E.; Klopfenstein, T.J.; Greenquist, M.A.; Luebbe, M.K.; Williams, P.; Engstrom, M.A. Effect of essential oils, tylosin, and monensin on finishing steer performance, carcass traits, liver abscesses, ruminal fermentation, and digestibility. *J. Anim. Sci.* **2009**, *87*, 2346–2354. <https://doi.org/10.2527/jas.2008-1493>.
 73. Acedo, T.S.; Gouvêa, V.N.; Vasconcellos, G.M.; Arrigoni, M.; Martins, C.L.; Millen, D.D.; Muller, L.R.; Melo, G.F.; Rizzieri, R.A.; Costa, C.F. Effect of 25-Hydroxy-Vitamin-D3 on Feedlot Cattle. *J. Anim. Sci.* **2018**, *96*(Suppl. 3), 447–448. <https://doi.org/10.1093/jas/sky404.976>.
 74. Carvalho, V.V.; Perdigão, A. Supplementation of 25-hydroxy-vitamin-D3 and increased vitamin E as a strategy to increase carcass weight of feedlot beef cattle. *J. Anim. Sci.* **2019**, *97*(Suppl. 3), 440. <https://doi.org/10.1093/jas/skz258.871>.
 75. Martins, T.E.; Acedo, T.S.; Gouvêa, V.N.; Vasconcellos, G.M.; Arrigoni, M.; Martins, C.L.; Millen, D.D.; Pai, M.D.; Perdigão, A.; Melo, G.F. Effects of 25-hydroxycholecalciferol supplementation on gene expression of feedlot cattle. *J. Anim. Sci.* **2020**, *98*, 302–303. <https://doi.org/10.1093/jas/skaa278.542>.
 76. Batley, R.J.; Romanzini, E.P.; Johnson, J.B.; de Souza, W.L.; Naiker, M.; Trotter, M.G.; Quigley, S.P.; de Souza Congio, G.F.; Costa, D.F.A. Rapid Screening of Methane-Reducing Compounds for Deployment via Water with a Commercial Livestock Supplement Using In Vitro and FTIR-ATR Analyses. *Methane* **2024**, *3*, 437–455. <https://doi.org/10.3390/methane3030025>
 77. Bohrer, B.M.; Edenburn, B.M.; Boler, D.D.; Dilger, A.C.; Felix, T.L. Effect of Feeding Ractopamine Hydrochloride (Optaflexx) with or without Supplemental Zinc and Chromium Propionate on Growth Performance, Carcass Traits, and Meat Quality of Finishing Steers. *J. Anim. Sci.* **2014**, *92*, 3988–3996. <https://doi.org/10.2527/jas.2014-7824>.
 78. Edenburn, B.M.; Kneeskern, S.G.; Bohrer, B.M.; Rounds, W.; Boler, D.D.; Dilger, A.C.; Felix, T.L. Effects of supplementing zinc or chromium to finishing steers fed ractopamine hydrochloride on growth performance, carcass traits, and meat quality. *J. Anim. Sci.* **2016**, *94*, 771–779. <https://doi.org/10.2527/jas.2015-9979>.
 79. Kneeskern, S.G.; Dilger, A.C.; Loerch, S.C.; Shike, D.W.; Felix, T.L. Effects of chromium supplementation to feedlot steers on growth performance, insulin sensitivity, and carcass traits. *J. Anim. Sci.* **2016**, *94*, 217–226. <https://doi.org/10.2527/jas.2015-9517>.
 80. Van Bibber-Krueger, C.L.; Axman, J.E.; Gonzalez, J.M.; Vahl, C.I.; Drouillard, J.S. Effects of a yeast combined with chromium propionate on growth performance and carcass quality of finishing steers. *J. Anim. Sci.* **2016**, *94*, 3003–3011. <https://doi.org/10.2527/jas.2016-0454>.
 81. Spears, J.W.; Kegley, E.B. Effect of zinc source (zinc oxide vs zinc proteate) and level on performance, carcass characteristics, and immune response of growing and finishing steers. *J. Anim. Sci.* **2002**, *80*, 2747–2752. <https://doi.org/10.1093/ansci/80.10.2747>.
 82. Matthews, J.O.; Guzik, A.C.; LeMieux, F.M.; Southern, L.L.; Bidner, T.D. Effect of chromium propionate on growth, carcass traits, and pork quality of growing-finishing pigs. *J. Anim. Sci.* **2005**, *83*, 858–862. <https://doi.org/10.2527/2005.834858x>.
 83. Tian, Y.Y.; Gong, L.M.; Xue, J.X.; Cao, J.; Zhang, L.Y. Effects of graded levels of chromium methionine on performance, carcass traits, meat quality, fatty acid profiles of fat, tissue chromium concentrations, and antioxidant status in growing-finishing pigs. *Biol. Trace Elem. Res.* **2015**, *168*, 110–121. <https://doi.org/10.1007/s12011-015-0352-1>.
 84. Hultgren, J.; Segerkvist, K.A.; Berg, C.; Karlsson, A.H.; Öhgren, C.; Algers, B. Preslaughter stress and beef quality in relation to slaughter transport of cattle. *Livest. Sci.* **2022**, *264*, 105073. <https://doi.org/10.1016/j.livsci.2022.105073>.
 85. Kegley, E.B.; Spears, J.W.; Brown, T.T., Jr. Effect of shipping and chromium supplementation on performance, immune response, and disease resistance of steers. *J. Anim. Sci.* **1997**, *75*, 1956–1964. <https://doi.org/10.2527/1997.7571956x>.
 86. Stahlhut, H.S.; Whisnant, C.S.; Lloyd, K.E.; Baird, E.J.; Legleiter, L.R.; Hansen, S.L.; Spears, J.W. Effect of chromium supplementation and copper status on glucose and lipid metabolism in Angus and Simmental beef cows. *Anim. Feed Sci. Technol.* **2005**, *128*, 253–265. <https://doi.org/10.1016/j.anifeeds.2005.11.002>.

87. Spears, J.W.; Whisnant, C.S.; Huntington, G.B.; Lloyd, K.E.; Fry, R.S.; Krafka, K.; Lamptey, A.; Hyda, J. Chromium propionate enhances insulin sensitivity in growing cattle. *J. Dairy Sci.* **2012**, *95*, 2037–2045. <https://doi.org/10.3168/jds.2011-4845>.
88. Anderson, R.A. Nutritional Factors Influencing the Glucose/Insulin System: Chromium. *J. Am. Coll. Nutr.* **1997**, *16*, 404–410. <https://doi.org/10.1080/07315724.1997.10718705>.
89. Yao, X.; Liu, R.; Li, X.; Li, Y.; Zhang, Z.; Huang, S.; Ge, Y.; Chen, X.; Yang, X. Zinc, selenium, and chromium co-supplementation improves insulin resistance by preventing hepatic endoplasmic reticulum stress in diet-induced gestational diabetes rats. *J. Nutr. Biochem.* **2021**, *96*, 108810. <https://doi.org/10.1016/j.jnutbio.2021.108810>.
90. Greenbaum, C.J.; Havel, P.J.; Taborsky, G.J.; Klaff, L.J. Intra-islet insulin permits glucose to directly suppress pancreatic A cell function. *J. Clin. Invest.* **1997**, *88*, 767–773. <https://doi.org/10.1172/JCI115375>.
91. Hoffmann, W.E.; Solter, P.F. Diagnostic enzymology of domestic animals. In *Clinical Biochemistry of Domestic Animals*, 6th ed.; Kaneko, J.J.; Harvey, J.W.; Bruss, M.L., Eds.; Elsevier: San Diego, CA, USA, 2008; pp. 351–378.
92. Lawrence, P.; Kenny, D.A.; Earley, B.; Crews, D.H., Jr.; McGee, M. Grass silage intake, rumen and blood variables, ultrasonic and body measurements, feeding behavior, and activity in pregnant beef heifers differing in phenotypic residual feed intake. *J. Anim. Sci.* **2011**, *89*, 3248–3261. <https://doi.org/10.2527/jas.2010-3774>.
93. Anderson, T.J.; Meredith, I.T.; Yeung, A.C.; Frei, B.; Selwyn, A.P.; Ganz, P. The Effect of Cholesterol-Lowering and Antioxidant Therapy on Endothelium-Dependent Coronary Vasomotion. *N. Engl. J. Med.* **1995**, *332*, 488–493. <https://doi.org/10.1056/NEJM199502233320802>.
94. Hassan, R.M.; Elsayed, M.; Kholief, T.E.; Hassanen, N.H.M.; Gafer, J.A.; Attia, Y.A. Mitigating effect of single or combined administration of nanoparticles of zinc oxide, chromium oxide, and selenium on genotoxicity and metabolic insult in fructose/streptozotocin diabetic rat model. *Environ. Sci. Pollut. Res.* **2021**, *28*, 48517–48534. <https://doi.org/10.1007/s11356-021-14089-w>.
95. Abou Zaid, O.A.R.; El-sonbaty, S.M.; Afifi, M.W.F. The Biochemical Effect of Chromium Nanoparticles Administration on Adiponectin Secretion, Oxidative Stress, and Metabolic Disorders in Streptozotocin-Induced Diabetic Rats. *Benha Vet. Med. J.* **2015**, *28*, 266–275. <https://doi.org/10.21608/bvmj.2015.32661>.
96. Li, Y.L.; Li, C.; Beauchemin, K.A.; Yang, W.Z. Effects of a commercial blend of essential oils and monensin in a high-grain diet containing wheat distillers' grains on in vitro fermentation. *Can. J. Anim. Sci.* **2013**, *93*, 387–398. <https://doi.org/10.4141/cjas2013-028>.
97. Van Gastelen, S.; Dijkstra, J.; Binnendijk, G.; Duval, S.M.; Heck, J.M.L.; Kindermann, M.; Zandstra, T.; Bannink, A. 3-Nitrooxypropanol decreases methane emissions and increases hydrogen emissions of early lactation dairy cows, with associated changes in nutrient digestibility and energy metabolism. *J. Dairy Sci.* **2020**, *103*, 8074–8093. <https://doi.org/10.3168/jds.2019-17936>.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.