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Article

Essential Oils as Repellents Against the House Cricket, *Acheta domesticus*

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Simple Summary

The aim of this study was to determine the behavior of house crickets, *Acheta domesticus*, to essential oils. We tested a panel of 27 essential oils and hypothesized that some would act as strong repellents, while others would have weak or no significant effects. In addition to the essential oils tested, we tested two synthetic repellents, which elicited no significant repellent effects. This study is important in contributing to our current understanding of insect repellency and the use of naturally derived plant compounds, in lieu of their synthetic counterparts, to serve as repellents against insect pests.

Abstract

The house cricket *Acheta domesticus* is found globally. It is an agricultural pest causing economic damage to a wide variety of crops including cereal seedlings, vegetable crops, fruit plants, and stored grains. Additionally, crickets act as mechanical vectors of pathogens by harboring bacteria, fungi, viruses and toxins causing foodborne illnesses. They can contaminate stored grains, packaged foods, or animal feed due to deposition of their feces, lowering the quality of the food and creating food safety risks. Synthetic insect repellents, such as pyrethroids and carbamates, have been used previously in integrated pest management practices to control crickets. Though successful as repellents, they have been associated with health and environmental risks and concerns. The use of organic green repellents, such as plant essential oils, may be a viable alternative in pest management practices. In this study, we tested the behavioral effects of 27 plant-based essential oils on the behavioral effects of the house crickets, *Acheta domesticus* in dual choice bioassays. Crickets were introduced into an open arena to allow them unrestricted movement. A transparent plastic bottle containing an essential oil treatment was placed in the arena to allow voluntary entry by crickets. Following a predetermined observation period, the number of crickets that entered the bottle was recorded, and percent entry was calculated as the proportion of individuals inside the bottle relative to the total number in the arena. Analysis of the percentage entry into the bottle allowed for a comparative assessment of repellency of the panel of essential oils that were tested. Essential oils that elicited high levels of entry into the bottle were categorized as having weak or no repellency, while those that produced reduced entry were classified as moderate or strong repellents. This ranking system enabled a clear differentiation among essential oils with respect to impact on cricket behavior. Our results indicated that house crickets responded with a strong repellent behavior to nearly half of the essential oils tested, while four essential oils and two synthetic repellents evoked no significant repellent responses. Four strong repellent essential oils were tested at different concentrations and showed a clear dose-dependent repellent effect. The results suggest that selected essential oils can be useful in the development of more natural “green” insect repellents.

Keywords: olfaction; repellent; behavior; locomotion; essential oils; cricket; Orthoptera; antenna; green repellent; pest

1. Introduction

Crickets are agricultural pests, disease vectors and noise nuisances in household settings. While they are not typically associated with the transmission of diseases as other arthropods such as ticks, mosquitoes, and cockroaches, they still raise significant concern with respect to food contamination and hygiene (i.e. stored grains, flour, etc.), crop, fabric and paper damage, and annoyance attributed to their nocturnal chirping [1]. In addition, cricket home infestations are often indicators of poor sanitation and/or structural inadequacies to home dwellings. More specifically, they can enter garbage cans to feed on trash due to their omnivorous feeding preference and, thereby, potentially contaminate food sources with their fecal matter. Therefore, finding safe pest management practices to control house crickets and other arthropods that carry diseases is critical. While the application of baits and sprays containing synthetic chemicals, such as pyrethroids, carbamates, neonicotinoids, isooxazolines, organophosphates, are options, these applications can pose problems and result in drawbacks, such as toxicity to adults, children, and pets, potential build-up of harmful chemical residues in the environment, and increased chance of resistance to insect populations (e.g., [2–5]). In addition, commonly used household methods to dispose of crickets have proven to be relatively ineffective. These include glue boards, baited cornmeal, diatomaceous earth, vacuum cleaner hose suction, sprays, and baits laced with synthetic chemicals [6]. Considering this, there is a chemical and ecological need to find plant-derived alternatives, i.e., natural, “green” repellents, as candidates in integrated pest management.

House crickets, like other insects, use their chemical senses (smell and taste) for orientation, food selection, and mate finding. To detect olfactory environmental cues, they possess a pair of antennae bearing antennal sensory organs or sensilla. These sensilla contain receptor neurons which encode and process olfactory stimuli. Such stimuli can trigger behaviors, such as orientation toward food and mating partners or avoidance of predators. In this study, the objective was to test a panel of essential oils on house cricket behavior.

Essential oils are lipophilic, volatile, organic, “green” (i.e., environmentally friendly) compounds representing mixtures of volatile secondary metabolites synthesized by aromatic plants [7]. Plants use these compounds to protect themselves against predators, such as insects and pathogens, and they can also serve as signaling compounds. These compounds include terpenes, terpenoids, aldehydes, phenols, etc. and can be extracted from plants using several methods (i.e., steam distillation, cold pressing, solvent extraction) [8]. In traditional medicine, they have been used as fragrances, antiseptics, and remedies. Essential oils are comprised of hundreds of compounds and variability of their content arises from both intrinsic and extrinsic factors [9]. Intrinsic factors include the plant’s genotype, whereas extrinsic factors are governed by e.g., seasonality, water availability and soil composition [10]. Other contributing factors may include temperature, humidity, light intensity, soil composition, etc. [11]. Essential oils are comprised of hydrocarbon molecules and can be classified as terpenes, alcohols, esters, aldehydes, ketones, phenols, oxygenated compounds, monoterpene alcohols, sesquiterpene alcohols, aldehydes, esters, lactones, coumarins, ethers, and oxides [7]. It is noted, however, that the most active compounds fall into two chemical groups: terpenoids (monoterpenoids and sesquiterpenoids) and phenylpropanoids [7,12].

Volatile molecules emanating from essential oils can be detected by olfactory receptor cells housed within olfactory sensory organs (or sensilla) on insect antennae [13]. Such molecules may be responsible for regulating insect behavior. In the case of essential oils, recent evidence has demonstrated that they can serve as olfactory repellents. Essential oils can disrupt the insects’ olfactory-guided locomotory behavior, making it difficult for insects to detect humans, as well as offer antimicrobial activity, as they contain active compounds like terpenes, phenols, and aldehydes which inhibit a broad range of microorganisms, including bacteria, fungi, and viruses [14–17].

Insect repellents differ from insecticides in their mode of action as they do not kill insects, they deter them approaching, landing, or establishing residence [18]. This has important relevance to cricket management, as the primary goal is to protect household materials and prevent food contamination, rather than eradication of insect populations. Due to their volatile nature, their odors

do not persist on a long-term basis in the environment, thereby requiring frequent reapplication [19], in addition to exploring methods to advance the formulation to ensure stabilization and prolonged action. In practice, this may require techniques to microencapsulate, nano emulsify, and the use of polymer-based slow-release methods to optimize their efficacy [20], in addition to developing more portable applications (e.g., sprays) tailored to cricket control.

2. Materials and Methods

Adult house crickets (Timberline, Marion, IL) were housed in a controlled environment maintained at $22^{\circ}\text{C} \pm 2^{\circ}\text{C}$ with a 16:8 light-dark cycle. Crickets were provided *ad libitum* access to cricket food and Easy Water (Timberline, Marion, IL) prior to experimentation. Female and male crickets were randomly selected. Each cricket was naïve to the specific odorant prior to testing and was discarded following its use to prevent any potential bias from prior exposure. Testing occurred in large plastic containers (Figure 1). The open arena (i.e., container) measured 42.9 cm x 29.2 cm x 14.9 cm (Sterilite, Townsend, MA) and was filled with 350 g of unscented paper bedding material (Kaytee Products, Inc., Chilton, WI). Plastic water bottles (500 ml) were cut in half, and the top half was inverted into the bottom half and secured with clear tape. This design prevented the crickets from escaping during testing. Filter paper disks (2.4 cm diameter circles) (Whatman/Cytiva, Marlborough, MA) served as carriers for the selected essential oils (E.O.s) (test substance) which were dissolved with pure ethyl alcohol (200 proof; Pharmco Products Inc., Brookfield, CT). For control experiments, only pure ethyl alcohol was applied to the filter paper. Experimental bioassays for control and test groups were conducted in separate locations to ensure no cross contamination of the odorant-containing bottles and control bottles. For experiments, 10 crickets were introduced into each container which maintained consistency of sample size. For each essential oil tested, the experiment was replicated at least 10 times (with some essential oils, more than 10 replicates were made) with 10 crickets per container ($n = 100$ crickets).

Orientation towards or away from the odorant source was assessed to determine the effects of the odorant on cricket behavior and to calculate the percentage of crickets that entered the bottle or remained in the open arena of the container. The degree of repellency associated with a particular essential oil was determined as the number of crickets entering the bottle (i.e., a lower number of crickets entering a bottle was deemed to indicate higher repellency of the essential oil). An evaluation of the strength of repellency for all the essential oils tested was carried out by assigning three ranges of percent entry: 0 - 25% (strong repellent), >25 - 50% entry (medium repellent), and >50% - <70% entry (weak repellent). Statistical analysis of data was performed using a table of cross-categorized frequency data for a Chi Square Test of Association (VassarStats Website for Statistical Computation, <http://vassarstats.net/>). Yates values were reported and corrected for continuity, and probability estimates were non-directional. Statistical significance was considered for p-values smaller than 0.05.

Essential oils (100% pure and organic) were purchased from Cliganic, Walnut, CA (peppermint, rosemary, tea tree, lemon grass, lavender, frankincense, orange), Anjou, Fremont, CA (sage, cinnamon, bergamot, lemon, grapefruit, patchouli, geranium, cypress, ylang ylang, palmarosa), Aromappeal, Holbrook, NY (basil, citronella, lemon eucalyptus, clove, eucalyptus), Now, Bloomingdale, IL (wintergreen), Plant Therapy, Twin Falls, ID (juniper berry), Aura Cacia, Urbana, IA (sweet fennel), Edens Garden, San Clements, CA (catnip), and Sedbuwza, Hagerstown, MD (coffee). The essential oils were pipetted onto a filter paper and placed at the bottom of each bottle. Four minutes was allowed to elapse prior to placing the bottles into the arena. The bottle (containing the test odorant or the control) was positioned at random locations in the containers to prevent bias. To compare essential oil repellency with repellent effects of known synthetic repellents, IR3535 and DEET were tested with the same bioassay design. All bioassays were run in the evening, as crickets are most active in the evenings due to their nocturnal lifestyle, for 6 hours to ensure sufficient observation time for behavioral responses. Upon completion of each experiment, all crickets and bottles used were disposed of to prevent potential cross-contamination.



Figure 1. Top-down view of the behavioral bioassay showing the inverted bottle, crickets, bedding, and plastic arena (lid removed). Ten crickets were allowed to explore freely in the arena during the experiment. The inverted bottle contained either an essential oil (test) or alcohol (control).

3. Results

3.1. Essential Oils as Repellent Compounds

In this study, we tested the effects of 27 essential oils (i.e., ylang ylang, sweet fennel, frankincense, juniper berry, cypress, wintergreen, geranium, cinnamon, sage, peppermint, basil, rosemary, lavender, catnip, patchouli, tea tree, lemon eucalyptus, clove, eucalyptus, citronella, lemongrass, palmarosa, coffee, bergamot, lemon, grapefruit, orange). They belonged to 12 different plant families (i.e., Annonaceae, Apiaceae, Burseraceae, Cupressaceae, Ericaceae, Geraniaceae, Lauraceae, Lamiaceae, Myrtaceae, Poaceae, Rubiaceae, and Rutaceae) (Table 1) and fell within three main chemical groups: monoterpenes/monoterpenoids, sesquiterpenes/sesquiterpenoids, and diterpenes/diterpenoids (Table 2).

These essential oils served as possible repellents of house crickets, *Acheta domesticus*, using a dual choice olfactory bioassay design (Figure 1). Test data was compared with control results. During control experiments, when no essential oil was present on the filter paper, 69.8% of the crickets in the container entered the inverted bottle. This served as the baseline to assess repellency of the tested essential oils. The strength of repellency was categorized by assigning essential oils to one of three ranges of percent entry: 0 - 25% entry (strong repellent), >25 - 50% entry (medium repellent), and >50% - <70% entry (weak repellent) (Figures 2–5). Four of the 27 essential oils tested, geranium, cypress, ylang ylang, and palmarosa, elicited no repellent effect. Yates values were reported and corrected for continuity, and probability estimates were non-directional. Statistical significance was considered for p-values smaller than 0.05 and Yates values were reported for the following essential oils: sage (Yates, 26.79, $p < 0.0001$), peppermint (Yates, 210.87, $p < 0.0001$), wintergreen (Yates, 103.06, $p < 0.0001$), basil (Yates, 90.49, $p < 0.0001$), rosemary (Yates, 148.24, $p < 0.0001$), cinnamon (Yates, 141.16, $p < 0.0001$), tea tree (Yates, 129.24, $p < 0.0001$), bergamot (Yates, 97.39, $p < 0.0001$), citronella (Yates, 94.66, $p < 0.0001$), juniper berry (Yates, 78.53, $p < 0.0001$), lemon grass (Yates, 128.79, $p < 0.0001$), lemon eucalyptus (Yates, 128.75, $p < 0.0001$), lavender (Yates, 78.04, $p < 0.0001$), lemon (Yates, 92.12, $p < 0.0001$), catnip (Yates, 99.71, $p < 0.0001$), clove (Yates, 46.8, $p < 0.0001$), grapefruit (Yates, 39.64, $p < 0.0001$), frankincense (Yates, 34.8, $p < 0.0001$), eucalyptus (Yates, 27.92, $p < 0.0001$), orange (Yates, 17.58, $p < 0.0001$), coffee (Yates, 13.06, $p < 0.0003$), patchouli (Yates, 7.09, $p < 0.0077$), sweet fennel (Yates, 6.06, $p < 0.0138$), geranium (Yates, 3.71, $p < 0.05408$), cypress (Yates, 1.11, $p < 0.2921$), ylang ylang (Yates, 0.28, $p < 0.5967$), palmarosa (Yates, 0.01, $p < 0.9203$). All the essential oils in Figure 2 were statistically significant from the control and elicited repellency, except for geranium, cypress, ylang ylang, and palmarosa. Percent entry values are discussed for each group of essential oils (i.e.,

strong, medium, weak repellency), in sections 2.2-2.4. Importantly, when known synthetic repellents, such as IR3535 and DEET were tested, they elicited no significant repellency (percent entry, 59%, Yates, 3.01, $p < 0.0827$ and percent entry, 68%, Yates, 0.03, $p < 0.8625$), respectively.

Table 1. Essential oils tested in this study belonging to 12 plant families.

Plant Family	Oils Included
Annonaceae	ylang ylang
Apiaceae	sweet fennel
Burseraceae	frankincense
Cupressaceae	juniper berry, cypress
Ericaceae	wintergreen
Geraniaceae	geranium
Lauraceae	cinnamon
Lamiaceae	sage, peppermint, basil, rosemary, lavender, catnip, patchouli
Myrtaceae	tea tree, lemon eucalyptus, clove, eucalyptus
Poaceae	citronella, lemongrass, palmarosa
Rubiaceae	coffee
Rutaceae	bergamot, lemon, grapefruit, orange

Note: Resources used [21].

Table 2. Composition of essential oils tested.

Oil / Plant	Monoterpenoids	Sesquiterpenoids	Dominant Functional Families (terpenoid or non-terpenoid)
Basil	✓	minor	Monoterpenoid alcohols (linalool), phenylpropanoids (estragole, eugenol)
Bergamot	✓	–	Monoterpenoid esters (linalyl acetate), monoterpenoid alcohols (linalool), coumarins (nonvolatile)
Catnip	✓ (iridoids)	–	Iridoids (monoterpenoid lactones, nepetalactone)
Cinnamon	–	–	Phenylpropanoid aldehydes (cinnamaldehyde)
Citronella	✓	minor	Monoterpenoid aldehydes (citronellal), monoterpenoid alcohols (citronellol, geraniol), monoterpenoid esters
Clove	–	✓	Phenylpropanoid phenols (eugenol), phenylpropanoid esters (eugenyl acetate), sesquiterpenes (β-caryophyllene)
Coffee	minor	–	Diterpenes (cafestol, kahweol), phenolic acids
Cypress	✓	✓	Monoterpenes (α-pinene), sesquiterpenoid alcohols (cedrol)
Eucalyptus	✓	–	Monoterpenoid oxides (1,8-cineole)
Frankincense	✓	✓	Monoterpenes + sesquiterpenes; resin contains diterpenes (boswellic acids)
Geranium	✓	minor	Monoterpenoid alcohols (citronellol, geraniol), monoterpenoid esters
Grapefruit	✓	minor	Monoterpenes (limonene), sesquiterpenoid ketones (nootkatone), monoterpenoid aldehydes
Juniper berry	✓	✓	Monoterpenes (α-pinene), sesquiterpenes (β-caryophyllene)
Lavender	✓	–	Monoterpenoid esters (linalyl acetate), monoterpenoid alcohols (linalool)
Lemon	✓	–	Monoterpenes (limonene), monoterpenoid aldehydes (citral, small %)
Lemon eucalyptus	✓	–	Monoterpenoid aldehydes (citronellal), monoterpenoid alcohols (citronellol)
Lemongrass	✓	minor	Monoterpenoid aldehydes (citral), monoterpenoid alcohols (geraniol)
Orange	✓	–	Monoterpenes (limonene)
Palmarosa	✓	–	Monoterpenoid alcohols (geraniol), monoterpenoid esters
Patchouli	–	✓	Sesquiterpenoid alcohols (patchoulol)
Peppermint	✓	–	Monoterpenoid alcohols (menthol), monoterpenoid ketones (menthone), monoterpenoid esters
Rosemary	✓	minor	Monoterpenoid oxides (1,8-cineole), monoterpenoid ketones (camphor), monoterpenes (α-pinene)
Sage	✓	–	Monoterpenoid ketones (thujone, camphor)
Sweet fennel	–	–	Phenylpropanoid ethers (anethole)
Tea tree	✓	minor	Monoterpenoid alcohols (terpinen-4-ol), monoterpenes, minor sesquiterpenes
Wintergreen	–	–	Phenylpropanoid esters / phenolic acid esters (methyl salicylate)
Ylang ylang	✓	✓	Monoterpenoid alcohols (linalool), sesquiterpenes (β-caryophyllene), esters (benzyl acetate)

Note: Resource used [22].

The essential oils tested in this study were comprised of terpenes with non-oxygenated hydrocarbons built from isoprene units (C₅H₈). They were further differentiated as monoterpenes (C₁₀ structures bearing two isoprene units; most common and highly volatile), sesquiterpenes (C₁₅ structures bearing three isoprene units; less volatile) and diterpenes (C₂₀ structures bearing four isoprene units; less common, less volatile, and more resin-like). Terpenoid families included monoterpenes, sesquiterpenes, and diterpenes (bearing a hydrocarbon skeleton and oxygen-containing functional groups) which included alcohols (-OH), aldehydes (-CHO), ketones (C=O), esters (-COO-), phenols (aromatic ring with an -OH group on the benzene ring), and ethers (R-O-R). Non-terpenoid families included phenylpropanoids (C₆H₅-CH₂-CH₂-R), and coumarins (benzopyrone compounds). Functional groups, like lactones (cyclic esters) and oxides (cyclic ethers) straddled both terpenoid and non-terpenoid classifications.

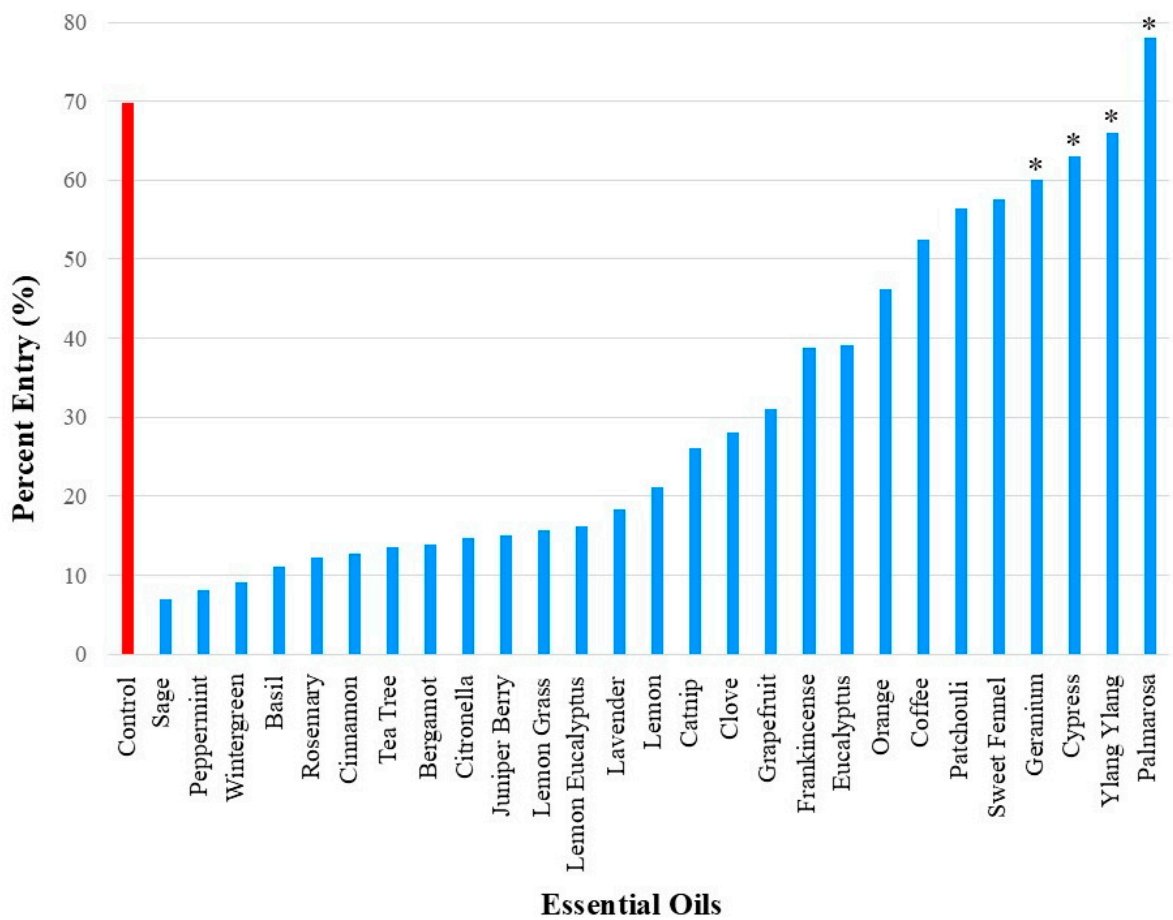


Figure 2. Overall results showing the degree of repellency elicited by 27 essential oils. Repellency was expressed as percent entry into a plastic bottle containing a test essential oil. Ethanol was used for all control experiments. Percent entry for test compounds was compared with that for the control compound (69.8%). Asterisks indicate essential oils that did not elicit significant repellency.

3.2. Essential Oils Eliciting Strong Repellent Responses

In terms of percent entry into bottles with test compounds, we found 14 essential oils that elicited between 7 and 21.1% entry. In ascending order, the strongest repellency using a 50-μg dose was found for the following essential oils (Figure 3): sage (7.00%), peppermint (8.06%), wintergreen (9.09%), basil (11.0%), rosemary (12.2%), cinnamon (12.7%), tea tree (13.5%), bergamot (13.8%), citronella (14.6%), juniper berry (15.0%), lemon grass (15.0%), lemon eucalyptus (16.3%), lavender (18.3%), and lemon (21.1%). These essential oils were categorized into five main odor groups (Table 3).

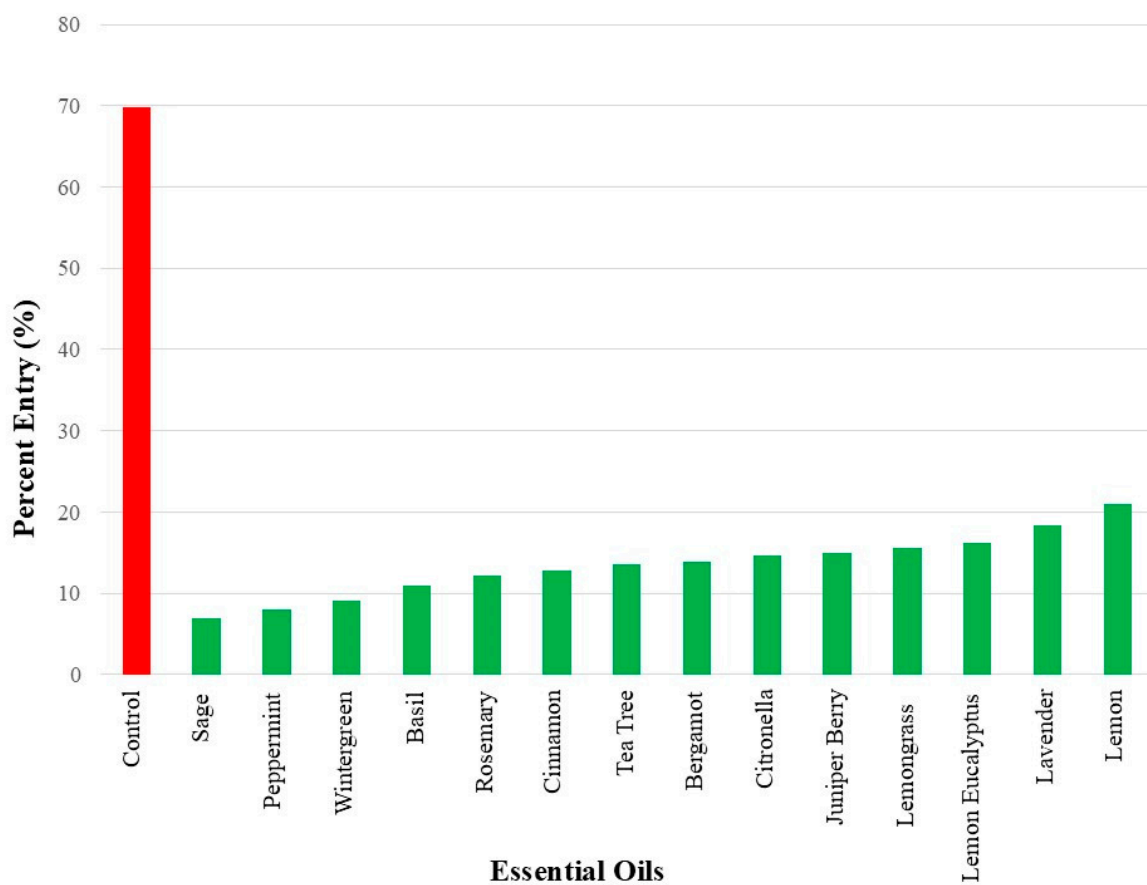


Figure 3. Repellency results of the essential oils exhibiting the strongest repellency (i.e., percent entry range: 0 - 25%) using a 50-μg dose are shown. Repellency was expressed as percent entry into a plastic bottle containing the test compound (i.e., essential oil). Ethanol was used for all control experiments. Percent entry for test compounds was compared with that for the control compound (69.8%). All essential oils in this figure elicited significant repellency.

Table 3. Categorization of 14 essential oils exhibiting the strongest repellency organized into five main odor groups.

Odor category	Essential oils
Camphor-like	Basil, Rosemary, Tea tree, Sage, Lemon eucalyptus
Minty	Peppermint, Wintergreen
Floral (sweet)	Lavender, Bergamot
Citrus	Lemon, Lemongrass, Citronella
Spicy (woody)	Cinnamon
Pine-like	Juniper berry

Note: Resource used [22,23].

3.3. Essential Oils Eliciting Medium Repellent Responses

The essential oils that elicited medium repellency using a 50-μg dose in ascending order were: catnip (26.0%), clove (26.0%), grapefruit (31.0%), frankincense (38.8%), eucalyptus (39.2%), and orange oil (46.2.%) (Figure 4).

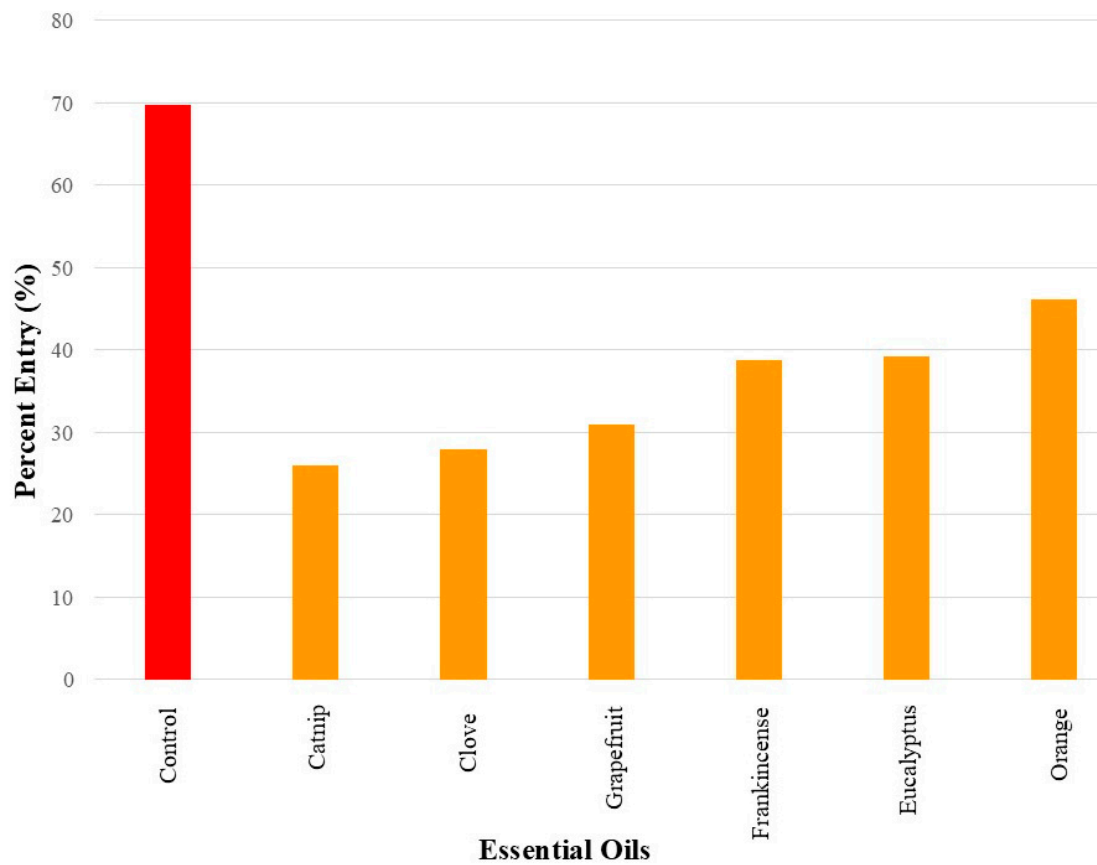


Figure 4. Results of repellency tests showing the essential oils that exhibited medium repellency (i.e., percent entry range > 25 - 50%) using a 50-μg dose. Repellency was expressed as percent entry into a plastic bottle containing the test compound (i.e., essential oil). Ethanol was used for all control experiments. Percent entry for test compounds was compared with that for the control compound (69.8%). All essential oils in this figure elicited significant repellency.

3.4. Essential Oils Eliciting Weak Repellent Responses

The essential oils that caused weakest repellency using a 50-μg dose in ascending order were: coffee (52.5%), patchouli 56.5%), and sweet fennel (57.5%). Geranium (60%), cypress (63%), ylang ylang (66%), and palmarosa (78%) did not elicit significant repellency (Figure 5).

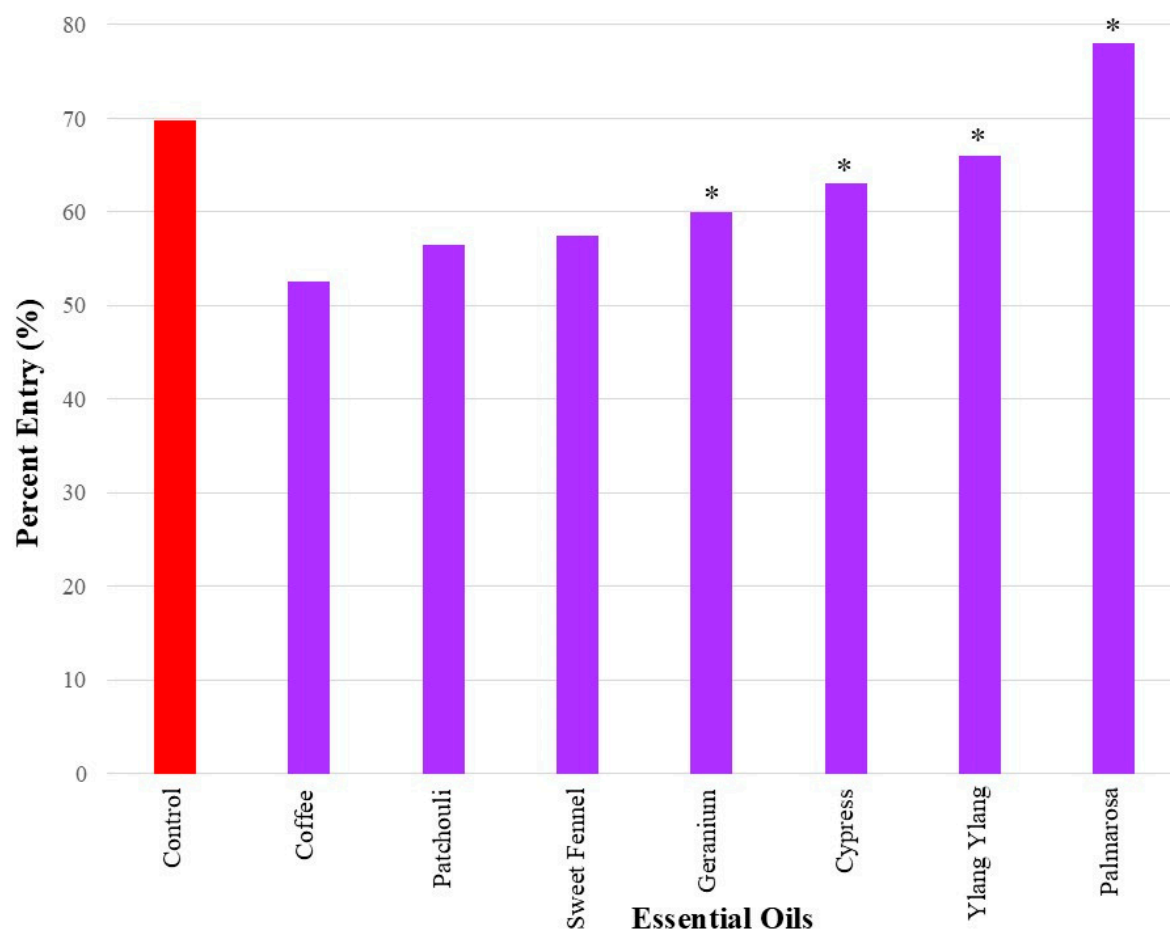


Figure 5. Results of repellency tests showing the essential oils that exhibited weak repellency (i.e., percent entry range >50% - <70%) using a 50- μ g dose. Only coffee, patchouli, and sweet fennel elicited significant repellency. Four compounds did not elicit significant repellency, namely geranium, cypress, ylang ylang, palmarosa indicated by the asterisks. Repellency was expressed as percent entry into a plastic bottle containing the test compound (i.e., essential oil). Ethanol was used for all control experiments. Percent entry for test compounds was compared with that for the control compound (69.8%).

3.5. Dose Response Curves

From those essential oils eliciting strong repellency, four were selected (i.e., peppermint, rosemary, cinnamon, and lemongrass) to determine their dose-response dynamics at four different concentrations (0.05 μ g, 0.5 μ g, 5 μ g, and 50 μ g) (Figure 6). In all cases, we observed a decrease in the mean number of entries into the test bottle with increasing concentration. Differently shaped sigmoid curves were noted. The RT (repellency threshold) concentration (i.e., lowest concentration at which repellency was significantly different from that of the control) was determined for each of the four essential oils. In the case of both peppermint and lemongrass, the RT was 0.05 μ g, whereas for rosemary and cinnamon this value was 0.5 μ g. We estimated the RD₅₀ concentration (concentration at 50% of the population of insects in the test group were repelled). As the control compound elicited 69.8% (~70%), and not 100% entry into control bottles, the concentration at which 35% (i.e., 50% of control entries) was used as the RD₅₀ value for test compounds. Since, on average, 7 out of 10 insects entered the bottle in control experiments, the RD₅₀ value was set to 3.5 number of entries (see y axes in Figure 6). In the case of lemon grass, peppermint, cinnamon, and rosemary, these RD₅₀ values were found to be approximately 1.1 μ g, 2.3 μ g, 6.8 μ g, and 10 μ g, respectively.

The data presented in the four dose-response curves suggested three main findings: (a) peppermint and lemongrass were the most repellent compounds; (b) while lemongrass appeared to have a slightly lower RD₅₀ concentration than peppermint (1.1 μ g versus 2.3 μ g, respectively), it was

evident that more crickets were initially repelled by peppermint at the highest dose tested (i.e., 50 μ g) and (c) rosemary was the least potent compound tested among the four potent repellents.

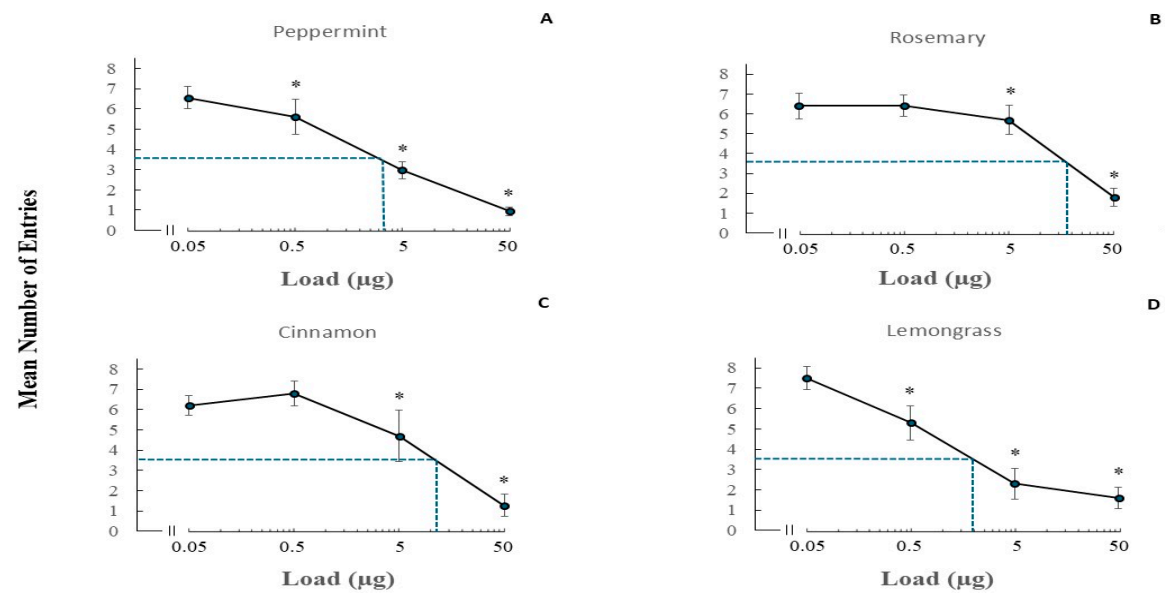


Figure 6. Dose-response curves for four essential oils eliciting strong repellency, namely A) peppermint, B) rosemary, C) cinnamon, and D) lemongrass at 0.05 μ g 0.5 μ g, 5 μ g, and 50 μ g. Each test compound was applied to a filter paper disk which was then placed in an empty bottle. Control experiments were run in a similar manner by applying alcohol to a filter paper disk. The hatched line indicates the RD₅₀ concentration (concentration which represented 50% of the insect population that was repelled). The RT (repellency threshold) concentrations represents the lowest concentrations at which repellency was significantly different from that of the control.

4. Discussion

Essential oils are plant-produced, bioactive secondary metabolites and thought to be less toxic to beneficial insects and environmentally friendly alternatives to synthetic repellents. They are highly concentrated, volatile, fat-soluble, biodegradable, colorless mixtures of aromatic compounds used for defense and signaling purposes (e.g., deter pests and pathogens, attract pollinators, and communicate with other organisms) [7,9]. Essential oils have been found to demonstrate antimicrobial, antioxidant, and/or insect-repellent activity [14]. Some essential oils also interfere with insect odorant receptors (i.e., “olfactory interference”) and can disrupt navigation, oviposition, and feeding, leading to avoidance of food sources and a reduction in oviposition [15,19] In this study, we determined the potential repellency of 27 essential oils using an olfactory test bioassay against *A. domesticus*. Effectiveness of repellency was determined by the number of crickets entering the plastic bottle containing the test compound. Repellency effectiveness was assigned to three groups those essential oils eliciting strong repellency (0 - 25% entry), medium repellency (>25 - 50% entry), and weak repellency (>50% - <70% entry). Our results demonstrated that 14 of the 27 essential oils tested elicited strong repellency, namely sage, peppermint, wintergreen, basil, rosemary, cinnamon, tea tree, bergamot, citronella, juniper berry, lemongrass, lemon eucalyptus, lavender, and lemon. Medium repellent behavior was observed with catnip, clove, grapefruit, frankincense, eucalyptus, and orange. Weak repellent behavior was observed with coffee, patchouli and sweet fennel. Geranium, cypress, ylang ylang, and palmarosa elicited no significant repellent effect. Similarly, synthetic repellents, IR3535 and DEET, elicited no significant repellency. Dose-response data from four essential oils that elicited strong repellent behavior in this study (i.e., peppermint, rosemary, cinnamon, and lemongrass) indicated that RT and RD₅₀ values were similar for peppermint and lemongrass, as they were for rosemary and cinnamon.

The essential oils found to elicit the strongest repellency (i.e., 0 - 25% entry) were classified as monoterpenes and monoterpenoids (alcohols, esters, aldehydes, oxides, and ketones), phenylpropanoids (esters, aldehydes) and sesquiterpenes (Table 2). These essential oils fell into five main odor categories: camphor-like, minty, floral (sweet), citrus, spicy (woody) and pine-like (Table 3). Five (i.e., basil, rosemary, tea tree, sage, and lemon eucalyptus) of the 14 essential oils that elicited strongest repellency, fell into the category of camphor-smelling, two (i.e., peppermint and wintergreen) with a minty odor, two (lavender and bergamot) with a sweet floral odor, three (lemon, lemongrass, citronella) with a citrus/lemon odor, one (cinnamon) with (woody) odor, and one (juniper berry) with a pine-like odor (Table 3).

Some of the essential oils tested in this study (i.e., ylang ylang, sweet fennel, geranium, lemon eucalyptus, citronella, lemongrass, palmarosa, peppermint, basil, rosemary, lavender, catnip, patchouli, tea tree, clove, cinnamon, sage, juniper berry, cypress wintergreen, frankincense, coffee, and eucalyptus, lemon, orange, grapefruit) are known to exhibit both larvicidal, as well as repellent activity in mosquitoes, blowflies, ticks, and store-product pest species. Studies on various insect and arthropod species have shown in addition that some essential oils serve as natural repellents including citronella and lemongrass (containing citronellal and geraniol, both mosquito repellents), catnip (nepetalactone), peppermint and clove (containing menthol and eugenol), and cinnamon and basil (containing eugenol and cinnamaldehyde) [24–44].

The effect of essential oils on target receptors, enzymes, and channels is key to understanding insecticidal activity and biological mechanisms of action. Examples include inhibiting gamma-aminobutyric acid (GABA), octopamine, and tyramine receptors, changes in acetylcholinesterase activity, interference with glutamate-gated chloride channels, disruption of growth, and effects on cuticular permeability. These effects result in dehydration or enhanced means for transdermal drug delivery. *Alpha*-pinene carvacrol, limonene, menthol, menthone, and 1,8-cineole have been shown to interact with acetylcholinesterase activity, while eugenol and cinnamaldehyde have been shown to interact with octopamine and tyramine receptors. Carvacrol, pulegone, and thymol have been found to boost the inhibitory effect of GABA at its receptors without directly turning the receptor on by themselves. In addition, thymol and menthol were found to inhibit glutamate-gated chloride channels [45–48].

Essential oils exhibit measurable repellent activity across multiple insect species and assay types. While performance varies among oils and formulations, many show comparable short-term efficacy to synthetic repellents and often do so with lower ecological and toxicological risk. As resistance to conventional insecticides continues to rise and demand grows for eco-conscious solutions, essential oils represent a promising avenue for future repellent development. With further refinement of delivery systems and deeper investigation into active constituents, botanically derived repellents can become viable, sustainable alternatives to synthetic chemicals in integrated pest management strategies.

5. Conclusions

In this study, we identified strongly repellent essential oils: sage, peppermint, wintergreen, basil, rosemary, cinnamon, tea tree, bergamot, citronella, juniper berry, lemon grass, lemon eucalyptus, lavender, and lemon. These essential oils fell into five main odor categories: camphor-like, minty, floral (sweet), citrus, spicy (woody) and pine-like. The effectiveness of these essential oil repellents shows their potential as ecofriendly repellents against the house cricket suggesting their suitability for integration into pest management methods. Essential oils can provide meaningful repellent effects. With refined formulation and further study on stability and safety, essential oils could evolve into practical, environmentally conscious alternatives to synthetic repellents. Their future use in integrated pest management could help reduce reliance on synthetic chemicals while maintaining effective protection.

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Abbreviations

The following abbreviations are used in this manuscript:

IR3535	Ethyl 3-(N-butylacetamido)propionate
DEET	N,N-Diethyl-meta-toluamide
RT	Repellency threshold
RD ₅₀ value	Concentration at 50% of the population of insects in the test group were repelled
GABA	Gamma-aminobutyric acid

References

1. Ebeling, W. Urban entomology. Univ. Calif. Div. Agr. Sci. 695 p. 1975.
2. Isman, M. B. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology*, 51, 45–66. 2006. <https://doi.org/10.1146/annurev.ento.51.110104.151146>
3. Snyder, D.E.; Meyer, J.; Zimmermann, A.G.; Qiao, M.; Gissendanner, S.J.; Cruthers, L.R.; Slone, R.L.; Young, D.R. Preliminary 896 Studies on the Effectiveness of the Novel Pulicide, Spinosad, for the Treatment and Control of Fleas on Dogs. *Vet Parasitol* 2007, 897 150, 345–351, doi:10.1016/j.vetpar.2007.09.011.
4. Matsumura, F. *Toxicology of insecticides* (2nd ed.). Springer. 2010.
5. Gupta, A.; Baker, C.; Wang, H.; Targa, N.; Pfefferkorn, A.; Tielemans, E. Target Animal Safety Evaluation of a Novel Topical 909 Combination of Esafoxolaner, Eprinomectin and Praziquantel for Cats. *Parasite* 2021, 28, 18, doi:10.1051/parasite/2021015.
6. Allen, N. How to get rid of crickets in your house. *Better Homes & Gardens*. 2025. <https://www.bhg.com/how-to-get-rid-of-crickets-in-the-house-8610165>
7. Miresmailli, S.; Isman, M.B. Botanical Insecticides Inspired by Plant–Herbivore Chemical Interactions. *Trends Plant Sci* 2014, 19, 940 29–35, doi:10.1016/j.tplants.2013.10.002.
8. Bakkali, F.; Averbek, S.; Averbek, D.; Idaomar, M. Biological Effects of Essential Oils – A Review. *Food and Chemical Toxicology* 942 2008, 46, 446–475, doi:10.1016/j.fct.2007.09.106.
9. De S. O. Batista, L.C.; Cid, Y.P.; De Almeida, A.P.; Prudêncio, E.R.; Riger, C.J.; De Souza, M.A.A.; Coumendouros, K.; Chaves, 944 D.S.A. In Vitro Efficacy of Essential Oils and Extracts of Schinus Molle L. against *Ctenocephalides felis felis*. *Parasitology* 2016, 143, 945 627–638, doi:10.1017/S0031182016000081.
10. dos Santos Cavalcanti, A.; de Souza Alves, M.; da Silva, L.C.P.; dos Santos Patrocínio, D.; Sanches, M.N.; Chaves, D.S. de A.; de 949 Souza, M.A.A. Volatiles Composition and Extraction Kinetics from *Schinus terebinthifolius* and *Schinus molle* Leaves and Fruit. 950 *Revista Brasileira de Farmacognosia* 2015, 25, 356–362, doi:10.1016/j.bjpp. 2015.07.003.
11. Regina, H.; Lima, P.; Kaplan, M.C.; Valéria De Mello Cruz, A. Influence of Abiotic Factors on Terpenoids Production and Variability 952 in the Plants; 2003.

12. Bruneton, J. Pharmacognosy: Phytochemistry, Medicinal Plants; Editions Médicales.; 2008
13. Shields VD, Hildebrand JG. Recent advances in insect olfaction, specifically regarding the morphology and sensory physiology of antennal sensilla of the female sphinx moth *Manduca sexta*. *Microsc Res Tech*. 2001 Dec 1;55(5):307-29. doi: 10.1002/jemt.1180. PMID: 11754510; PMCID: PMC2386875.
14. Wińska K., Mączka W., Łyczko J., Grabarczyk M., Czubaszek A., Szumny A. Essential Oils as Antimicrobial Agents—Myth or Real Alternative? *Molecules*. 2019; 24(11):2130. <https://doi.org/10.3390/molecules24112130>
15. Kamaraj C, Satish Kumar RC, Al-Ghanim KA, Nicoletti M, Sathiyamoorthy V, Sarvesh S, Ragavendran C, Govindarajan M. Novel Essential Oils Blend as a Repellent and Toxic Agent against Disease-Transmitting Mosquitoes. *Toxics*. 2023 Jun 8;11(6):517. doi: 10.3390/toxics11060517. PMID: 37368617; PMCID: PMC10303837.
16. Hazarika H, Krishnatreyya H. Technological Advancements in Mosquito Repellents: Challenges and Opportunities in Plant-Based Repellents. *Acta Parasitol*. 2025 May 28;70(3):117. doi: 10.1007/s11686-025-01054-7. PMID: 40434490.
17. Yadav, Deepak K., Rathee, S. Sharma, V. Patil, U.K. A Comprehensive Review on Insect Repellent Agents: Medicinal Plants and Synthetic Compounds. *Anti-inflammatory & Anti-allergy Agents in Medicinal Chemistry*. 2024 Oct 16. doi:10.2174/0118715230322355240903072704.
18. Debboun, M., Frances, S.P., & Strickman, D. (Eds.). (2006). *Insect Repellents: Principles, Methods, and Uses* (1st ed.). CRC Press. <https://doi.org/10.1201/9781420006650>.
19. Nerio, Luz S., Olivero-Verbel, J., and Stashenko, E. Repellent activity of essential oils: a review. *Bioresour Technol*. 2010 Jan;101(1):372-8. <https://doi.org/10.1016/j.biortech.2009.07.048>
20. Pavela, R. History, Presence and Perspective of Using Plant Extracts as Commercial Botanical Insecticides and Farm Products 974 for Protection against Insects - A Review. *Plant Protection Science* 2016, 52, 229–241, doi: 10.17221/31/2016-PPS
21. AromaWeb 1997-2025. <https://www.aromaweb.com/essential-oils/bergamot-essential-oil.php>
22. Buchbauer, G. (Ed.). (2024). *Handbook of Essential Oils: Science, Technology, and Applications* (3rd ed.). CRC Press. [ISBN: 9780367504021]
23. PubChem <https://pubchem.ncbi.nlm.nih.gov/>
24. Chaiphongpachara T, Laojun S. Comparative efficacy of commercial ylang-ylang (*Cananga odorata*) essential oils from India and Thailand against larval *Aedes aegypti* (L.) (Diptera: Culicidae). *J Adv Vet Anim Res*. 2020 Jun 29;7(3):391-396. doi: 10.5455/javar.2020.g433. PMID: 33005663; PMCID: PMC7521804.
25. Sajid Iqbal, Fawad Ahmed Khan, Abdullah Haris, Raimondas Mozūratīs, Muhammad Binyameen, Muhammad Azeem. Essential oils of four wild plants inhibit the blood seeking behaviour of female *Aedes aegypti*, *Experimental Parasitology*, Volume 244, 2023, 108424, ISSN 0014-4894.
26. Jaenson TG, Pålsson K, Borg-Karlson AK. Evaluation of extracts and oils of mosquito (Diptera: Culicidae) repellent plants from Sweden and Guinea-Bissau. *J Med Entomol*. 2006 Jan;43(1):113-9. doi: 10.1093/jmedent/43.1.113. PMID: 16506457.
27. Haris, A.; Azeem, M.; Abbas, M.G.; Mumtaz, M.; Mozūratīs, R.; Binyameen, M. Prolonged Repellent Activity of Plant Essential Oils against Dengue Vector, *Aedes aegypti*. *Molecules* **2023**, 28, 1351. <https://doi.org/10.3390/molecules28031351>
28. Caballero-Gallardo, K., Olivero-Verbel, J., Stashenko, E., Repellency and toxicity of essential oils from *Cymbopogon martinii*, *Cymbopogon flexuosus* and *Lippia origanoides* cultivated in Colombia against *Tribolium castaneum*. *Journal of Stored Products Research*, Volume 50, 2012, Pages 62-65, ISSN 0022-474X, <https://doi.org/10.1016/j.jspr.2012.05.002>.
29. Kumar S, Wahab N, Warikoo R. Bioefficacy of *Mentha piperita* essential oil against dengue fever mosquito *Aedes aegypti* L. *Asian Pac J Trop Biomed*. 2011 Apr;1(2):85-8. doi: 10.1016/S2221-1691(11)60001-4. PMID: 23569733; PMCID: PMC3609176.
30. Sutthanont N, Sudsawang M, Phanpoowong T, Sriwichai P, Ruangsittichai J, Rotejanaprasert C, Srisawat R. Effectiveness of Herbal Essential Oils as Single and Combined Repellents against *Aedes aegypti*, *Anopheles dirus* and *Culex quinquefasciatus* (Diptera: Culicidae). *Insects*. 2022 Jul 21;13(7):658. doi: 10.3390/insects13070658. PMID: 35886836; PMCID: PMC9322308.

31. Bedini S, Guarino S, Echeverria MC, Flamini G, Ascrizzi R, Loni A, Conti B. *Allium sativum*, *Rosmarinus officinalis*, and *Salvia officinalis* Essential Oils: A Spiced Shield against Blowflies. *Insects*. 2020 Feb 25;11(3):143. doi: 10.3390/insects11030143. PMID: 32106555; PMCID: PMC7143423.
32. Batume, C., Mulongo, I.M., Ludlow, R. et al. Evaluating repellence properties of catnip essential oil against the mosquito species *Aedes aegypti* using a Y-tube olfactometer. *Sci Rep* **14**, 2269 (2024). <https://doi.org/10.1038/s41598-024-52715-y>
33. Gokulakrishnan, J., Kuppusamy, E., Shanmugam, D., Appavu, A., Kaliyamoorthi, K. Pupicidal and repellent activities of *Pogostemon cablin* essential oil chemical compounds against medically important human vector mosquitoes. *Asian Pacific Journal of Tropical Disease*, Volume 3, Issue 1, 2013, Pages 26-31, ISSN 2222-1808, [https://doi.org/10.1016/S2222-1808\(13\)60006-7](https://doi.org/10.1016/S2222-1808(13)60006-7).
34. Sritabutra D, Soonwera M. Repellent activity of herbal essential oils against *Aedes aegypti* (Linn.) and *Culex quinquefasciatus* (Say.). *Asian Pac J Trop Dis*. 2013 Aug;3(4):271–6. doi: 10.1016/S2222-1808(13)60069-9. PMCID: PMC4027311.
35. Luker, H.A., Salas, K.R., Esmaeli, D. et al. Repellent efficacy of 20 essential oils on *Aedes aegypti* mosquitoes and *Ixodes scapularis* ticks in contact-repellency assays. *Sci Rep* **13**, 1705 (2023). <https://doi.org/10.1038/s41598-023-28820-9>
36. Barbosa, L. C. A., Filomeno, C. A. & Teixeira, R. R. Chemical variability and biological activities of *Eucalyptus* spp. essential oils. *Molecules* **21**, 1671. 2016.
37. Carroll, J.F., Tabanca, N., Kramer, M., Elejalde, N.M., Wedge, D.E., Bernier, U.R., Coy, M., Becel, J.J., Demirci, B., Husnu, K., Baser, C., Zhang, J., and Zhang, S. Essential oils of *Cupressus funebris*, *Juniperus communis* and *J. chinensis* (Cupressaceae) as repellents against ticks (Acari: Ixodidae) and mosquitoes (Diptera: Culicidae) and as toxicants against mosquitoes. *J. Vect. Ecol.*, 36 (2): 258-268. 2011.
38. Su XL, Huang ZC, Chen L, Chen DY, Zhao DX, Zeng ZJ. Active Components of 16 Essential Oils and Their Fumigation Effects on *Galleria mellonella* (Lepidoptera: Pyralidae). *Insects*. 2024 Dec 9;15(12):977. doi: 10.3390/insects15120977. PMID: 39769579; PMCID: PMC11677612.
39. Metayi MH, Abd El-Naby SS, El-Habal NA, Fahmy HH, Abdou MS, Ali B, Abdel-Rheim KH, Abdel-Megeed A. Omani Frankincense nanoemulsion formulation efficacy and its latent effects on biological aspects of the spiny bollworm *Earias insulana* (Boisd.). *Front Physiol*. 2022 Oct 6;13:1001136. doi: 10.3389/fphys.2022.1001136. PMID: 36277209; PMCID: PMC9583008.
40. Aditama, Wiwit and F. Sitepu. The effectiveness of arabica coffee (*Coffea arabica* L) grounds on mortality and growth of *Aedes aegypti* larva. (2019).
41. Ahmed HA, Nassrallah AA, Abdel-Raheem MA, Elbehery HH. Lemon peel essential oil and its nano-formulation to control *Agrotis ipsilon* (Lepidoptera: Noctuidae). *Sci Rep*. 2023 Oct 20;13(1):17922. doi: 10.1038/s41598-023-44670-x. PMID: 37863942; PMCID: PMC10589301.
42. Ugwu, Francis Stephen Ogbonna and Chidiebere Divine Chime. "Repellence of *Aedes aegypti* with oils from *Citrus sinensis* and *Citrus paradisi* fruit peels from Nsukka." *Bio-Research* (2023): n. pag.
43. Sarma, R., Adhikari, K., Mahanta, S., Khanikor, B. Insecticidal activities of *Citrus aurantifolia* essential oil against *Aedes aegypti* (Diptera: Culicidae), *Toxicology Reports*, Volume 6, 2019, Pages 1091-1096, ISSN 2214-7500, <https://doi.org/10.1016/j.toxrep.2019.10.009>.
44. Fernando SSST, Jayasooriya RGPT, Samarakoon KW, Wijegunawardana NDAD, Alahakoon SB. Citrus-Based Bio-Insect Repellents-A Review on Historical and Emerging Trends in Utilizing Phytochemicals of Citrus Plants. *J Toxicol*. 2024 Nov 28;2024:6179226. doi: 10.1155/jt/6179226. PMID: 39640379; PMCID: PMC11620817.
45. Corrêa, E.J.A.; Carvalho, F.C.; de Castro Oliveira, J.A.; Bertolucci, S.K.V.; Scotti, M.T.; Silveira, C.H.; Guedes, F.C.; Melo, J.O.F.; 963 de Melo-Minardi, R.C.; de Lima, L.H.F. Elucidating the Molecular Mechanisms of Essential Oils' Insecticidal Action Using a 964 Novel Cheminformatics Protocol. *Sci Rep* 2023, 13, 4598, doi:10.1038/s41598-023-29981-3.
46. Jankowska, M.; Rogalska, J.; Wyszowska, J.; Stankiewicz, M. Molecular Targets for Components of Essential Oils in the Insect 966 Nervous System—A Review. *Molecules* 2017, 23, 34, doi:10.3390/molecules23010034.

47. Liu, X.; Cao, A.; Yan, D.; Ouyang, C.; Wang, Q.; Li, Y. Overview of Mechanisms and Uses of Biopesticides. *Int J Pest Manag* 2021, 968 67, 65–72, doi:10.1080/09670874.2019.1664789.
48. Popescu, I.E.; Gostin, I.N.; Blidar, C.F. An Overview of the Mechanisms of Action and Administration Technologies of the 970 Essential Oils Used as Green Insecticides. *Agri Engineering* 2024, 6, 1195–1217, doi:10.3390/agriengineering6020068.

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