

Article

Not peer-reviewed version

Antitumor Activity by an Anti-CD44 Variant 9 Monoclonal Antibody in Gastric and Colorectal Cancer Xenograft Models

Mayuki Tawara , [Hiroyuki Suzuki](#) ^{*} , [Tomokazu Ohishi](#) , [Mika K. Kaneko](#) , [Yukinari Kato](#) ^{*}

Posted Date: 26 August 2025

doi: 10.20944/preprints202508.1939.v1

Keywords: CD44v9; monoclonal antibody therapy; ADCC; CDC; gastric cancer; colorectal cancer



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Article

Antitumor Activity by an Anti-CD44 Variant 9 Monoclonal Antibody in Gastric and Colorectal Cancer Xenograft Models

Mayuki Tawara ¹, Hiroyuki Suzuki ^{1,*}, Tomokazu Ohishi ², Mika K. Kaneko ¹ and Yukinari Kato ^{1,*}

¹ Department of Antibody Drug Development, Tohoku University Graduate School of Medicine, 2-1 Seiryō-machi, Aoba-ku, Sendai, Miyagi 980-8575, Japan

² Institute of Microbial Chemistry (BIKAKEN), Laboratory of Oncology, Microbial Chemistry Research Foundation, 3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141-0021, Japan

* Correspondence: hiroyuki.suzuki.b4@tohoku.ac.jp (H.S.); yukinari.kato.e6@tohoku.ac.jp (Y.K.); Tel.: +81-22-717-8207 (H.S. & Y.K.)

Abstract

Background: CD44 variants (CD44v) play essential roles in the promotion of tumor metastasis, maintenance of cancer stem cell properties, and resistance to treatments. Therefore, the development of anti-CD44v mAbs is essential for targeting CD44v-positive tumor cells. An anti-CD44v9 mAb, C₄₄Mab-1 (mouse, IgG₁, kappa), was previously established. C₄₄Mab-1 recognizes the CD44v variant exon 9-encoded region and applies to multiple research techniques. **Methods:** A mouse IgG_{2a} version of C₄₄Mab-1 (C₄₄Mab-1-mG_{2a}) was generated to evaluate the *in vitro* and *in vivo* antitumor activities using gastric and colorectal cancer cell lines. **Results:** C₄₄Mab-1-mG_{2a} showed a reactivity to CD44v3–10-overexpressed Chinese hamster ovary-K1 (CHO/CD44v3–10), gastric cancer MKN45, and colorectal cancer COLO205 in flow cytometry. C₄₄Mab-1-mG_{2a} exhibited both antibody-dependent cellular cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) against CHO/CD44v3–10, MKN45, and COLO205. Furthermore, administration of C₄₄Mab-1-mG_{2a} significantly suppressed CHO/CD44v3–10, MKN45, and COLO205 xenograft tumor growth compared with control mouse IgG_{2a}. **Conclusions:** These results indicated that C₄₄Mab-1-mG_{2a}, which possesses ADCC/CDC activities, could be applied to the mAb-based therapy against CD44v9-positive carcinomas.

Keywords: CD44v9; monoclonal antibody therapy; ADCC; CDC; gastric cancer; colorectal cancer

1. Introduction

A type I transmembrane glycoprotein CD44 was initially characterized in the 1980s as a lymphocyte homing receptor [1]. The human CD44 gene is located on chromosome 11p13 and comprises 19 exons. Exons 1–5 encode the conserved extracellular domain, exons 16 and 17 encode the stalk region, exon 18 encodes the transmembrane domain, and exons 19 and 20 encode the intracellular cytoplasmic domain. The isoform encoded by exons 1–5 and 16–20 is referred to as the standard form (CD44s), which is presented in most types of cells. Exons 6–15 are subject to alternative splicing, generating multiple variant (CD44v) isoforms by insertion between the extracellular and stalk regions [2]. CD44 is broadly expressed in lymphocytes, fibroblasts, and smooth muscle cells. In normal epithelial tissues, various CD44v isoforms are detected, with variant 9 (v9) being the most prevalent, followed by v6 and v4 [3]. Aberrant expression of CD44v has been implicated in tumor progression [4]. CD44v8-10 (CD44E) is predominantly expressed in epithelial cells, whereas CD44v3–10, the largest isoform, is mainly found in keratinocytes [5].

Both CD44s and CD44v (pan-CD44) bind to hyaluronic acid (HA) through the conserved extracellular domain, which plays critical roles in cellular homing, migration, adhesion, and proliferation [6]. The variant isoforms of CD44 (CD44v) have been implicated in multiple oncogenic processes, including the promotion of tumor invasion and metastasis [7], acquisition of cancer stem cell (CSC) properties [8], and resistance to chemotherapy and radiotherapy [9]. The v3-encoded region undergoes heparan sulfate modification, enabling high-affinity binding to heparin-binding growth factors, such as fibroblast growth factors. This modification allows the v3 region to function as a co-receptor for receptor tyrosine kinases, thereby enhancing downstream signal transduction [10]. The v6-encoded region is essential for c-MET activation through the formation of a ternary complex with hepatocyte growth factor [11]. In addition, the v8–10-encoded region interacts with and stabilizes the cystine–glutamate transporter (xCT), facilitating cystine uptake and subsequent glutathione synthesis, which mitigates reactive oxygen species (ROS)-induced stress [12]. Regulation of redox homeostasis through CD44v8–10–xCT interaction is associated with poor clinical prognosis [13]. Accordingly, the generation and characterization of monoclonal antibodies (mAbs) that recognize each variant exon are essential for elucidating their distinct biological functions and for the development of CD44-targeted cancer therapies.

CD44 is also recognized as a cell surface marker of cancer stem-like cells (CSCs) in various carcinomas [5]. CD44s or CD44v isoforms-specific mAbs have been employed to isolate CD44-high CSC populations [5]. These CD44-high cells exhibit CSC properties, including drug resistance and robust tumorigenic potential *in vivo* [5]. Consequently, the development of anti-CD44 mAbs capable of selectively recognizing individual variants is crucial for elucidating CSC biology and developing variant-specific cancer therapy.

Multiple isoforms of CD44 have been implicated in malignant progression across diverse tumor types [7], including colorectal cancers [14], pancreatic cancers [15,16], prostate cancers [17], head and neck squamous cell carcinomas [18], breast cancers [19], and gliomas [20,21]. Furthermore, a comprehensive multi-omic analysis of malignant gastric cancers revealed genomic amplifications of established cancer driver genes such as EGFR, ERBB2, MET, FGFR2, and CD44 in gastric cancer with peritoneal metastasis [10]. These cell surface antigens are potential therapeutic targets for mAb-based interventions [11]. While mAb therapies and diagnostic approaches have been developed for the first four antigens, equivalent strategies targeting CD44 remain largely unestablished.

We previously generated highly sensitive and specific mAbs targeting CD44 by immunizing mice with CD44v3–10-overexpressed Chinese hamster ovary-K1 (CHO/CD44v3–10). The critical epitopes recognized by these mAbs were identified through enzyme-linked immunosorbent assay, followed by functional characterization in flow cytometry, western blotting, and immunohistochemistry. Among the established clones, C₄₄Mab-1 (mouse IgG₁, κ) bound to a peptide corresponding to the v9-encoded region. Flow cytometric analysis demonstrated that C₄₄Mab-1 recognized CHO/CD44v3–10 cells as well as colorectal cancer cell lines (COLO201 and COLO205). Furthermore, C₄₄Mab-1 successfully detected endogenous CD44v9 in colorectal cancer tissues by immunohistochemistry [22]. These findings indicate that C₄₄Mab-1 is a valuable tool for detecting CD44v9 across multiple applications.

In this study, we converted C₄₄Mab-1 into a mouse IgG_{2a} type mAb (C₄₄Mab-1-mG_{2a}). We evaluated antibody-dependent cellular cytotoxicity (ADCC), complement-dependent cytotoxicity (CDC), and antitumor effect in gastric and colorectal cancer xenograft models.

2. Results

2.1. Production of Mouse IgG_{2a}-Type Anti-CD44v9 mAb, C₄₄Mab-1-mG_{2a}

Immunizing mice previously developed an anti-CD44v9 mAb, C₄₄Mab-1, with CHO/CD44v3–10. C₄₄Mab-1 recognizes the CD44v variant exon 9-encoded region and shows a high binding affinity against CHO/CD44v3–10 and tumor cells [22]. In this study, the complementarity-determining regions (CDRs) of C₄₄Mab-1 were first determined from the cDNA of C₄₄Mab-1-producing hybridoma. To evaluate the antitumor activity, a mouse IgG_{2a}-type C₄₄Mab-1 (C₄₄Mab-1-mG_{2a}) was

constructed by fusing the V_H and V_L CDRs of C_{44} Mab-1 with the C_H and C_L chains of mouse IgG_{2a} (Figure 1A). As a control mouse IgG_{2a} (mIgG_{2a}), PMab-231 (an anti-tiger podoplanin mAb, mouse IgG_{2a}) was produced as described previously [23]. We confirmed the purity of the recombinant mAbs by SDS-PAGE under reduced conditions (Figure 1B).

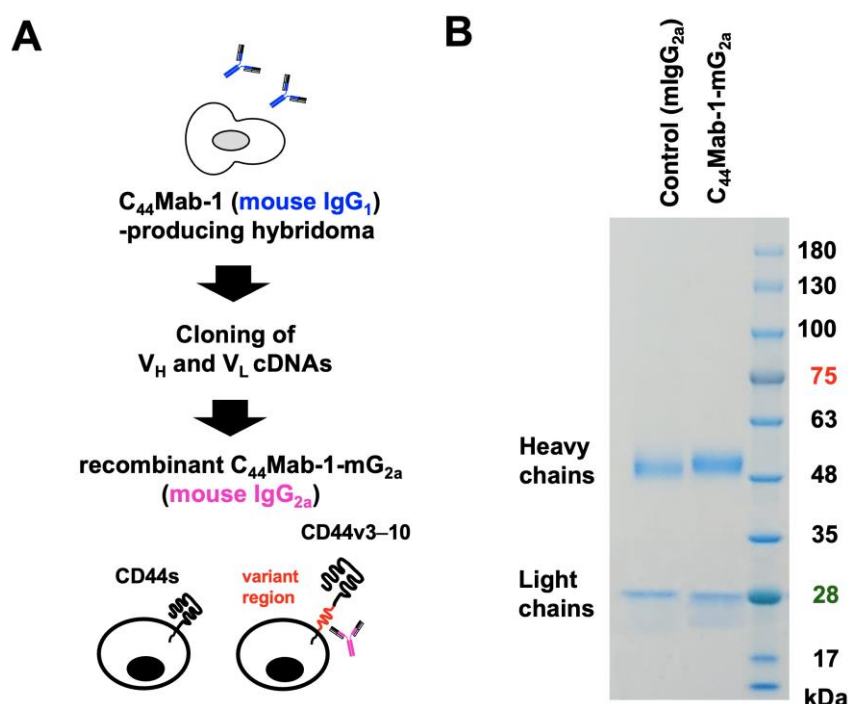


Figure 1. Production of a recombinant anti-CD44v9 mAb, C_{44} Mab-1-mG_{2a}, from C_{44} Mab-1-producing hybridoma. (A) After determination of CDRs of C_{44} Mab-1 (mouse IgG₁), recombinant C_{44} Mab-1-mG_{2a} (mouse IgG_{2a}) was produced and purified. (B) C_{44} Mab-1-mG_{2a} and PMab-231 (control mIgG_{2a}) were treated with sodium dodecyl sulfate sample buffer containing 2-mercaptoethanol. Proteins were separated on a polyacrylamide gel. The gel was stained with Bio-Safe CBB G-250 Stain.

2.2. Flow Cytometry Using C_{44} Mab-1-mG_{2a}

The reactivity of C_{44} Mab-1-mG_{2a} was first confirmed using CHO/CD44v3–10. As shown in Figure 2A, C_{44} Mab-1-mG_{2a} showed dose-dependent reactivity to CHO/CD44v3–10, but not to CHO/CD44s or CHO-K1. We confirmed the expression of CD44s by a pan-CD44 mAb, 5-mG_{2a} (Supplementary Figure 1). Control mIgG_{2a} did not recognize CHO/CD44v3–10 (Figure 2B). Next, the binding affinity was investigated. The dissociation constant value of C_{44} Mab-1-mG_{2a} for CHO/CD44v3–10 was determined to be 4.7×10^{-9} M (Figure 2C). These results indicated that C_{44} Mab-1-mG_{2a} possesses comparable reactivity and affinity with parental mAb, C_{44} Mab-1.

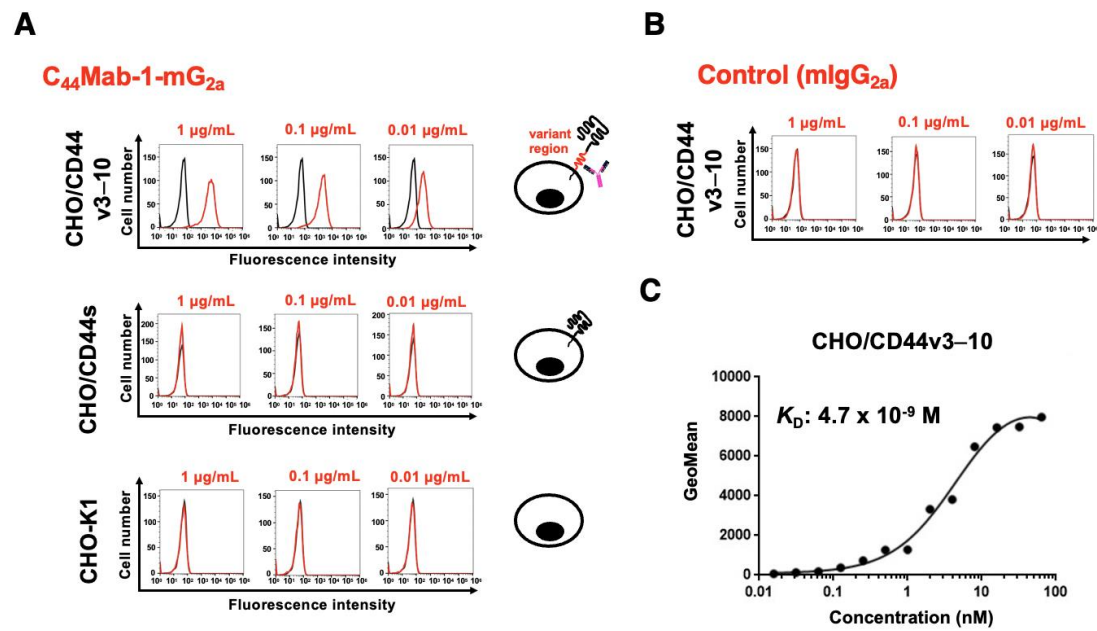


Figure 2. Flow cytometry analysis using C₄₄Mab-1-mG_{2a}. (A) CHO/CD44v3-10, CHO/CD44s, and CHO-K1 were treated with 0.01, 0.1, and 1 µg/mL of C₄₄Mab-1-mG_{2a}. (B) CHO/CD44v3-10 was treated with 0.01, 0.1, and 1 µg/mL of control mIgG_{2a}. Then, the cells were treated with Alexa Fluor 488-conjugated anti-mouse IgG. Fluorescence data were analyzed using the SA3800 Cell Analyzer. (C) CHO/CD44v3-10 were treated with serially diluted C₄₄Mab-1-mG_{2a}, followed by Alexa Fluor 488-conjugated anti-mouse IgG treatment. The fluorescence data were analyzed, and the K_D values were determined.

C₄₄Mab-1 could recognize colorectal cancer cell lines, including COLO205, in flow cytometry [22]. In addition to colorectal cancer, the reactivity of C₄₄Mab-1-mG_{2a} to gastric cancer MKN45 was investigated in flow cytometry. As shown in Figure 3A, C₄₄Mab-1-mG_{2a} exhibited comparable reactivity to MKN45 and COLO205. Control mIgG_{2a} did not recognize MKN45 and COLO205 (Figure 3B). C₄₄Mab-1-mG_{2a} also reacted with other gastric cancer cell lines (LMSU, KatoIII, and NUGC-4, Supplementary Figure 2). These results indicated that C₄₄Mab-1-mG_{2a} retains the reactivity to CD44v9-positive cells.

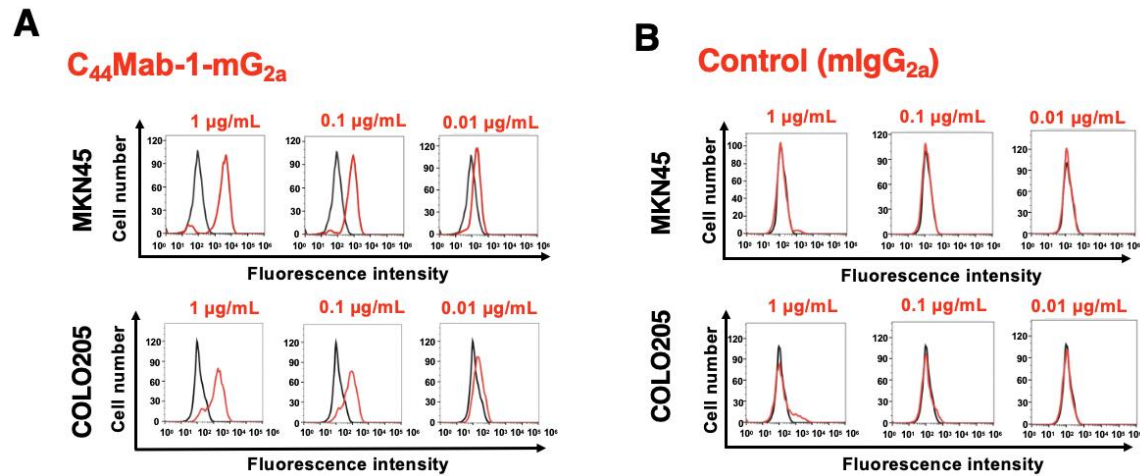


Figure 3. Flow cytometry analysis of C₄₄Mab-1-mG_{2a} to CD44v9-positive cancer cell lines. (A) MKN45 and COLO205 were treated with 0.01, 0.1, and 1 µg/mL of C₄₄Mab-1-mG_{2a}. (B) MKN45 and COLO205 were treated with 0.01, 0.1, and 1 µg/mL of control mIgG_{2a}. Then, the cells were treated with Alexa Fluor 488-conjugated anti-mouse IgG. Fluorescence data were analyzed using the SA3800 Cell Analyzer.

2.3. Immunohistochemistry of Gastric Cancer Using C₄₄Mab-1-mG_{2a}

We previously stained a colorectal cancer tissue array using C₄₄Mab-1 [22]. Therefore, we next investigated the CD44v9 expression in the gastric cancer tissue array (BS01011b) using C₄₄Mab-1-mG_{2a} and an anti-pan CD44 mAb, C₄₄Mab-46-mG_{2a} [24]. C₄₄Mab-1-mG_{2a} exhibited membranous staining in intestinal-type gastric cancer (Figure 4A, left). C₄₄Mab-46-mG_{2a} stained the same type of cancer cells and surrounding stroma cells (Figure 4A, right). In diffuse-type gastric cancer, diffusely spread tumor cells were strongly stained by both C₄₄Mab-1-mG_{2a} and C₄₄Mab-46-mG_{2a} (Figure 4B). Additionally, stromal staining by C₄₄Mab-46 was also observed in the tissue (Figure 4B, right).

We summarized the immunohistochemistry of gastric cancer tissue array in supplementary Table S1; C₄₄Mab-1-mG_{2a} stained 52 out of 72 cases (72%) of gastric cancer but did not stain the stromal tissues. These results indicated that C₄₄Mab-1-mG_{2a} is also helpful to detect CD44v9 in immunohistochemistry of formalin-fixed paraffin-embedded gastric cancers.

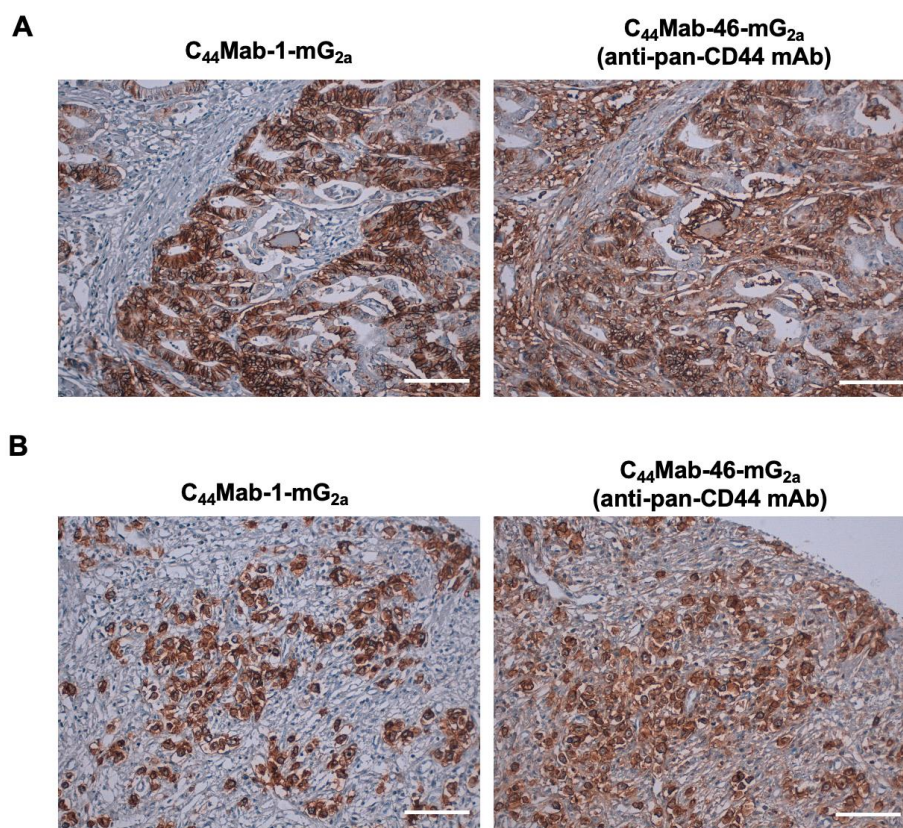


Figure 4. Immunohistochemistry of gastric cancer using C₄₄Mab-1-mG_{2a}. Immunohistochemical analysis using C₄₄Mab-1-mG_{2a} and C₄₄Mab-46-mG_{2a} (a pan-CD44 mAb) against gastric cancer tissue array (BS01011b). After antigen retrieval, serial sections of the tissue arrays were incubated with 5 µg/mL of C₄₄Mab-1-mG_{2a} or 5 µg/mL of C₄₄Mab-46-mG_{2a}, followed by treatment with the Envision+ kit. The color was developed using 3,3'-diaminobenzidine tetrahydrochloride (DAB), and the sections were counterstained with hematoxylin. Scale bar = 100 µm. Typical staining patterns of intestinal-type gastric cancer (**A**, no. 8) and diffuse-type gastric cancer (**B**, no. 58) were shown.

2.4. ADCC and CDC by C₄₄Mab-1-mG_{2a}

The ADCC caused by C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, MKN45, and COLO205 was investigated. The splenocytes derived from BALB/c nude mice were used as an effector. C₄₄Mab-1-mG_{2a} showed potent ADCC against CHO/CD44v3-10 (37.2% vs. 9.4% cytotoxicity of control mIgG_{2a}, $p < 0.05$, Figure 5A), MKN45 (33.1% vs. 14.6% cytotoxicity of control mIgG_{2a}, $p < 0.05$, Figure 5B), and COLO205 (15.0% vs. 3.9% cytotoxicity of control mIgG_{2a}, $p < 0.05$, Figure 5C).

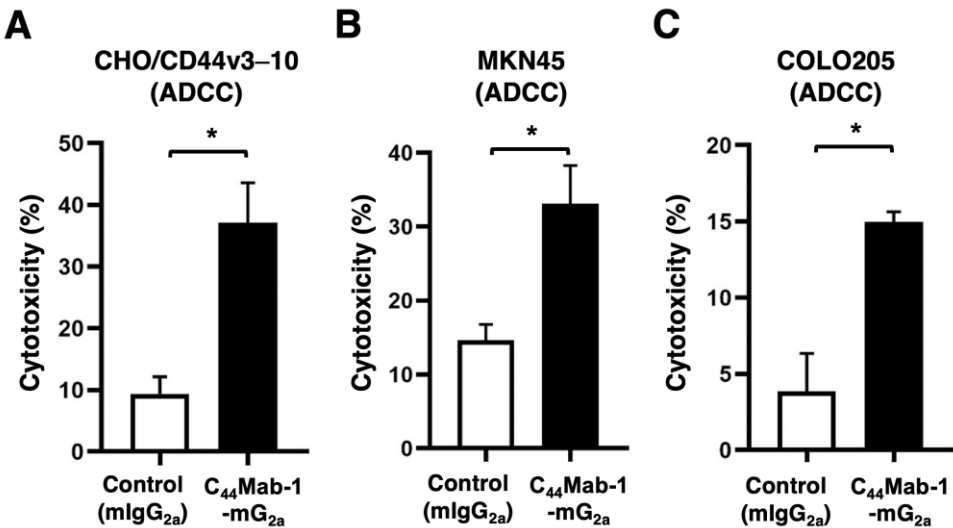


Figure 5. ADCC by C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, MKN45, and COLO205. ADCC induced by C₄₄Mab-1-mG_{2a} or control mouse IgG_{2a} (mIgG_{2a}) against CHO/CD44v3-10 (A), MKN45 (B), and COLO205 (C). Calcein AM-labeled target cells (CHO/CD44v3-10, MKN45, and COLO205) were incubated with the effector splenocytes in the presence of 100 µg/mL of C₄₄Mab-1-mG_{2a} or control mIgG_{2a}. Following a 4.5-hour incubation, the Calcein release into the medium was measured. Values are shown as mean ± SEM. Asterisks indicate statistical significance (* $p < 0.05$; Two-tailed unpaired t test).

The CDC caused by C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, MKN45, and COLO205 was next examined. C₄₄Mab-1-mG_{2a} showed significant CDC against CHO/CD44v3-10 (51.6% vs. 12.1% cytotoxicity of control mIgG_{2a}, $p < 0.05$, Figure 6A), MKN45 (13.3% vs. 3.4% cytotoxicity of control mIgG_{2a}, $p < 0.05$, Figure 6B), and COLO205 (22.5% vs. 13.0% cytotoxicity of control mIgG_{2a}, $p < 0.05$, Figure 6C). These results indicated that C₄₄Mab-1-mG_{2a} exerts antitumor efficacy through activation of effector cells and complements *in vitro*.

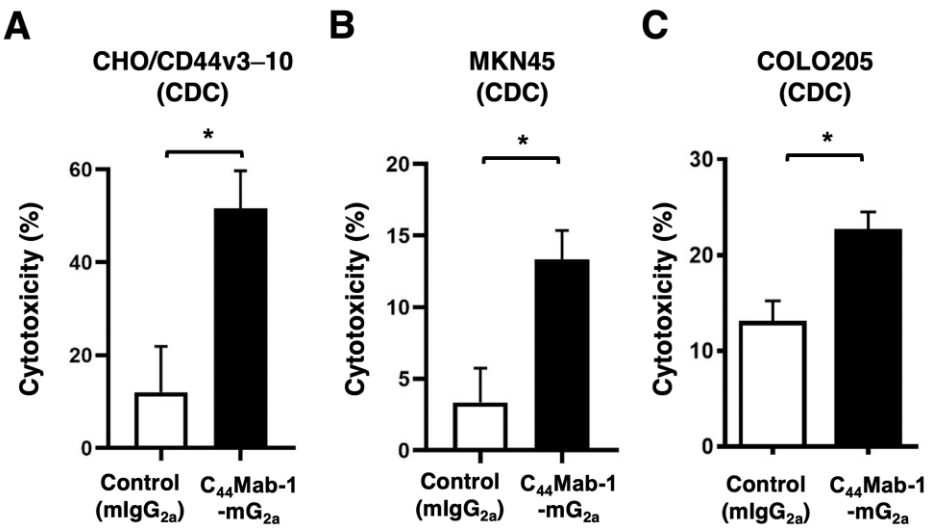


Figure 6. CDC by C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, MKN45, and COLO205. CDC induced by C₄₄Mab-1-mG_{2a} or control mouse IgG_{2a} (mIgG_{2a}) against CHO/CD44v3-10 (A), MKN45 (B), and COLO205 (C). Calcein AM-labeled target cells (CHO/CD44v3-10, MKN45, and COLO205) were incubated with complements and C₄₄Mab-1-mG_{2a} or control mIgG_{2a}. Following incubation for 4.5 h at 37 °C, the Calcein release into the medium was measured. Values are shown as mean ± SEM. Asterisks indicate statistical significance (* $p < 0.05$; Two-tailed unpaired t test).

2.5. Antitumor Effect by C₄₄Mab-1-mG_{2a} Against CHO/CD44v3-10, MKN45, and COLO205 Xenografts

The antitumor effects caused by C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, MKN45, and COLO205 xenografts were evaluated. Following the inoculation of CHO/CD44v3-10, C₄₄Mab-1-mG_{2a} or control mIgG_{2a} was intraperitoneally administered into CHO/CD44v3-10 xenograft-bearing mice on days 7, 14, and 21. The tumor volume was measured on days 7, 10, 14, 17, 21, 23, and 28 after the inoculation. The C₄₄Mab-1-mG_{2a} administration resulted in a potent and significant reduction in CHO/CD44v3-10 xenografts on days 23 ($p < 0.05$) and 28 ($p < 0.01$) compared with that of mIgG_{2a} (Figure 7A). In the cases of MKN45 and COLO205 xenografts, C₄₄Mab-1-mG_{2a} or control mIgG_{2a} was intraperitoneally administered on days 7 and 14. The significant reductions were observed in MKN45 xenografts on days 10 ($p < 0.05$), 14 ($p < 0.01$), and 21 ($p < 0.01$) compared with that of mIgG_{2a} (Figure 7B). The significant reductions were also observed in COLO205 xenografts on days 14 ($p < 0.05$) and 21 ($p < 0.01$) compared with that of mIgG_{2a} (Figure 7C).

Significant decreases in xenograft weight caused by C₄₄Mab-1-mG_{2a} were observed in CHO/CD44v3-10 xenografts (52% reduction; $p < 0.05$; Figure 7D), MKN45 xenografts (24% reduction; $p < 0.05$; Figure 7E), and COLO205 xenografts (23% reduction; $p < 0.05$; Figure 7F). Body weight loss was not observed in the xenograft-bearing mice treated with C₄₄Mab-1-mG_{2a} (Figure 7G, H, and I).

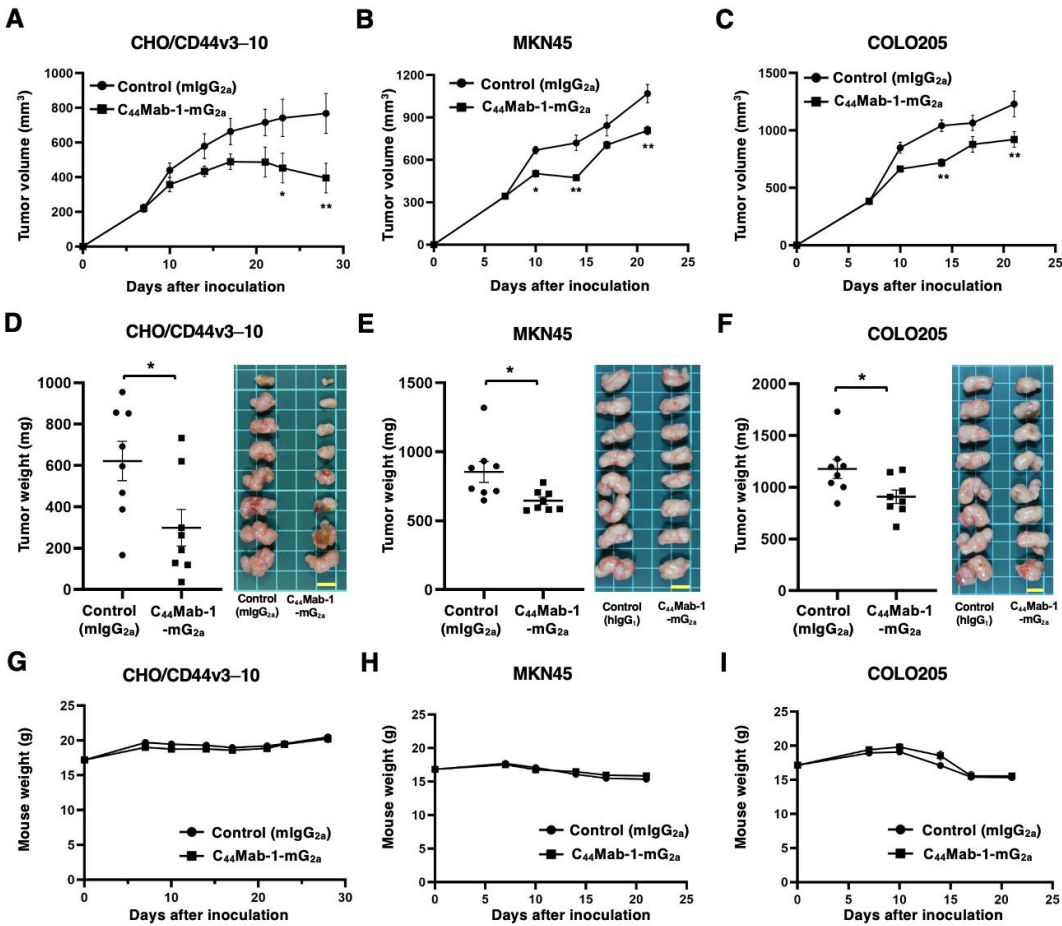


Figure 7. Antitumor activity of C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, MKN45, and COLO205 xenograft. (A-C) CHO/CD44v3-10 (A), MKN45 (B), and COLO205 (C) were subcutaneously injected into BALB/c nude mice (day 0). An amount of 100 µg of C₄₄Mab-1-mG_{2a} or control mouse IgG_{2a} (mIgG_{2a}) was intraperitoneally injected into each mouse on day 8. Additional antibodies were injected on day 14 (MKN45 and COLO205) or 14 and 21 (CHO/CD44v3-10). The tumor volume is represented as the mean ± SEM. ** $p < 0.01$, * $p < 0.05$ (ANOVA with Sidak's multiple comparisons test). (D-F) The mice treated with the mAbs were euthanized on day 21 (MKN45 and COLO205) or 28 (CHO/CD44v3-10). The xenograft weights were measured. Values are presented as the mean ± SEM. ** $p < 0.01$ (Two-tailed unpaired t test). (G-I) Body weights of xenograft-bearing mice treated with the mAbs. There is no statistical difference.

3. Discussion

Given the critical role of CD44 in cancer metastasis and therapeutic resistance, several strategies targeting CD44 have been developed for the treatment of diverse malignancies, including head and neck, breast, gynecological, and ovarian cancers [25]. However, clinical trials evaluating the safety and efficacy of these interventions have demonstrated limited success. RG7356, an anti-pan CD44 mAb, demonstrated an acceptable safety profile. However, the trial was discontinued due to the absence of a clinically significant or dose-dependent response [26]. Clinical evaluation of an antibody–drug conjugate (ADC) comprising an anti-CD44v6 mAb (bivatuzumab–mertansine) was performed, but development was halted because of severe cutaneous toxicity [27,28]. In this study, a mouse IgG_{2a} version of anti-CD44v9 mAb, C₄₄Mab-1-mG_{2a}, was developed (Figure 1). C₄₄Mab-1-mG_{2a} showed a high reactivity to CD44v9-positive gastric and colorectal cancer cell lines (Figure 3) and exhibited the *in vitro* (Figure 5 and 6) and *in vivo* antitumor activities (Figure 7) in their xenograft models. These findings underscore the need for a more comprehensive understanding of CD44 biology to enhance therapeutic efficacy while minimizing side effects.

In a gastric cancer cell line, the predominant CD44v transcripts—CD44v3,8–10, CD44v6–10, CD44v8–10, and CD44v3,8—were identified [29]. C₄₄Mab-1-mG_{2a} recognizes almost all products derived from these transcripts, enabling detection of a broad spectrum of CD44v-expressing gastric cancer cells. Among these variants, CD44v8–10 has been shown to play a critical role in regulating ROS defense and in promoting gastric cancer progression [12]. The CD44v8–10 stabilizes xCT, an essential component of the cellular antioxidant defense system, thereby allowing cancer stem cells to mitigate oxidative stress, confer ferroptosis resistance, and maintain their tumorigenic capacity [30]. Accordingly, an anti-CD44v9 mAb, clone RV3, has predominantly been used for immunohistochemistry. Previous studies have demonstrated that CD44v9 serves as a predictive marker for gastric cancer recurrence [31] and as a biomarker for patient selection and therapeutic efficacy of the xCT inhibitor sulfasalazine [32]. Further studies using C₄₄Mab-1-mG_{2a} are warranted to clarify the association between CD44v9 expression and clinicopathological features. In addition, C₄₄Mab-1-mG_{2a} detected both intestinal-type gastric cancer and diffuse-type gastric cancer in immunohistochemistry (Figure 4). Future investigations should determine whether CD44v9 expression is enriched in specific molecular subtypes of gastric cancer [33]. In gastric cancer with peritoneal metastasis, CD44 was identified as an amplified cancer driver gene [10]. It is worthwhile to evaluate the effect of C₄₄Mab-1-mG_{2a} in a peritoneal metastasis model of gastric cancer.

CD44 is reported to be a target gene of Wnt/ β -catenin in a mouse intestinal tumor model [34]. However, the mechanism of the v9 inclusion during colorectal cancer development remains to be determined. Large-scale genomic analyses have revealed that colorectal cancers were classified into four subtypes: canonical, mesenchymal, metabolic, and microsatellite instability immune types [35]. The relationship between CD44v9 and the subtypes should be investigated in the future. We previously examined the CD44v9 expression on colorectal cancer tissues by immunohistochemistry and found that the CD44v9 was expressed on the basolateral surface of colorectal cancers [22]. The basolateral CD44 expression was previously reported and co-localized with HA [36], Annexin II [37], and Claudin-7-EpCAM complex [38]. Therefore, the basolateral expression of CD44 may serve as an adhesion-mediated signal transduction, which contributes to colorectal cancer tumorigenesis.

Since bivatuzumab–mertansine showed toxicity to normal skin epithelium [27,28], CD44v6-targeted strategies have been further developed to chimeric antigen receptor T (CAR-T) cell therapy. CD44v6 CAR-T cells exhibited antitumor activity against primary human multiple myeloma and acute myeloid leukemia [39]. Moreover, CD44v6 CAR-T cells effectively suppressed xenograft tumor growth in models of lung and ovarian carcinomas [40], highlighting their potential for application in solid tumor therapy. Although CD44v9 expression is minimally detected in normal colonic epithelium by C₄₄Mab-1, it is observed in other normal tissues, such as squamous epithelium [22]. Therefore, to enable the therapeutic application of C₄₄Mab-1, additional studies are required to mitigate potential toxicities in these tissues.

To reduce the reactivity and toxicities in normal tissues, cancer-specific mAbs (CasMabs) for various antigens have been developed. More than three hundred anti-human epidermal growth factor receptor 2 (HER2) mAb clones have been established by immunization of mice with cancer cell-expressed HER2. Among them, H₂CasMab-2 (H₂Mab-250) was screened for cancer-specific reactivity using flow cytometry. H₂CasMab-2 specifically recognized HER2 expressed on breast cancer cells but not on normal epithelial cells from lung bronchus, kidney proximal tubule, colon, and mammary gland. Epitope analysis revealed the mode of cancer-specific recognition [41]. Furthermore, chimeric antigen receptor (CAR)-T cell therapy using a single chain variable fragment of H₂CasMab-2 has been developed and evaluated in a phase I clinical trial (NCT06241456) for patients with HER2-positive advanced solid tumors [42]. Although we additionally obtained several clones of anti-CD44v9 mAbs, further clones are required for the CD44v9 CasMab screening. C₄₄Mab-1-mG_{2a} would serve as a reference mAb when the CD44v9 CasMabs are established in the future.

4. Materials and Methods

4.1. Cell Lines

CHO-K1 and COLO205 (colorectal cancer) cell lines were obtained from the American Type Culture Collection (Manassas, VA, USA). MKN45 and NUGC-4 (human gastric cancer) cell lines were obtained from the Japanese Collection of Research Bioresources (Osaka, Japan). KatoIII (human gastric cancer) was obtained from the Cell Resource Center for Biomedical Research Institute of Development, Aging, and Cancer at Tohoku University (Miyagi, Japan). CHO/CD44v3-10 was established previously [43].

MKN45 and NUGC-4 were maintained in Dulbecco's Modified Eagle's Medium (DMEM; Nacalai Tesque, Inc., Kyoto, Japan). CHO/CD44v3-10, CHO-K1, KatoIII, and COLO205 were cultured in Roswell Park Memorial Institute (RPMI) 1640 medium (Nacalai Tesque, Inc.). All culture media were supplemented with 10% heat-inactivated fetal bovine serum (FBS; Thermo Fisher Scientific Inc., Waltham, MA, USA), 100 U/mL penicillin, 100 µg/mL streptomycin, and 0.25 µg/mL amphotericin B (Nacalai Tesque, Inc.). Cells were incubated at 37 °C in a humidified atmosphere containing 5% CO₂ and 95% air.

4.2. Recombinant mAb Production

A control mIgG_{2a} mAb, PMAb-231 (mouse IgG_{2a}, κ, an anti-tiger podoplanin mAb) was previously produced [23]. An anti-CD44v9 mAb, C₄₄Mab-1 (mouse IgG₁, κ) was established previously [22]. To create the mouse IgG_{2a} version (C₄₄Mab-1-mG_{2a}), the V_H cDNA of C₄₄Mab-1 and the C_H of mouse IgG_{2a} were cloned into the pCAG-Neo vector [FUJIFILM Wako Pure Chemical Corporation (Wako), Osaka, Japan]. Similarly, the V_L cDNA of C₄₄Mab-1 and the C_L of the mouse kappa chain were cloned into the pCAG-Ble vector (Wako). The antibody expression vectors were transfected into BINDS-09 (fucosyltransferase 8-knockout ExpiCHO-S) using the ExpiCHO Expression System (Thermo Fisher Scientific Inc.) to produce C₄₄Mab-1-mG_{2a} and PMAb-231. All antibodies were purified using Ab-Capcher (ProteNova Co., Ltd., Kagawa, Japan).

4.3. Flow Cytometry

Cells were detached by brief incubation with a solution containing 1 mM ethylenediaminetetraacetic acid (EDTA; Nacalai Tesque, Inc.) and 0.25% trypsin. Following centrifugation and washing with phosphate-buffered saline (PBS) containing 0.1% bovine serum albumin (BSA) as a blocking buffer, the cells were incubated with primary mAbs for 30 minutes at 4 °C. Subsequently, cells were treated with Alexa Fluor 488-conjugated anti-mouse IgG secondary antibody (1:2000; Cell Signaling Technology, Inc., Danvers, MA). Fluorescence signals were acquired using an SA3800 Cell Analyzer (Sony Corp., Tokyo, Japan).

4.4. Immunohistochemical Analysis

Formalin-fixed paraffin-embedded tissue arrays of gastric cancer (BS01011b) were purchased from US Biomax Inc. (Rockville, MD, USA). The tissue arrays were autoclaved in citrate buffer (pH 6.0; Nichirei Biosciences, Inc., Tokyo, Japan) for 20 min. The blocking was performed using SuperBlock T20 (Thermo Fisher Scientific, Inc.). The sections were incubated with 5 µg/mL of C₄₄Mab-1-mG_{2a} and 5 µg/mL of an anti-pan-CD44 mAb, C₄₄Mab-46-mG_{2a} [44]. Then, the tissue arrays were treated with the EnVision+ Kit for mouse (Agilent Technologies Inc.). The chromogenic reaction was performed using 3,3'-diaminobenzidine tetrahydrochloride (DAB; Agilent Technologies Inc.). Hematoxylin (FUJIFILM Wako Pure Chemical Corporation) was used for counterstaining. A Leica DMD108 (Leica Microsystems GmbH, Wetzlar, Germany) was used to obtain images.

4.5. ADCC by C₄₄Mab-1-mG_{2a}

The animal experiments were conducted in accordance with the institutional regulations and ethical guidelines to minimize pain and distress, and were approved by the Institutional Committee for Animal Experiments of the Institute of Microbial Chemistry (Numazu, Japan; approval number: 2025-029). Five-week-old female BALB/c nude mice were purchased from Japan SLC Inc. (Shizuoka, Japan). The splenocytes (designated as effector cells) were prepared as described previously [45]. The ADCC activity of C₄₄Mab-1-mG_{2a} was investigated as follows. Calcein AM-labeled target cells (CHO/CD44v3-10, MKN45, and COLO205) were co-incubated with the effector cells at an effector-to-target (E:T) ratio of 50:1 in the presence of 100 µg/mL of C₄₄Mab-1-mG_{2a} (n=3) or control mIgG_{2a} (n=3). Following a 4.5-hour incubation, the Calcein release into the medium was measured using a microplate reader (Power Scan HT; BioTek Instruments, Inc., Winooski, VT, USA).

Cytotoxicity was calculated as a percentage of lysis using the following formula: % lysis = $(E - S)/(M - S) \times 100$, where E represents the fluorescence intensity from co-cultures of effector and target cells, S denotes the spontaneous fluorescence from target cells alone, and M corresponds to the maximum fluorescence obtained after complete lysis using a buffer containing 10 mM Tris-HCl (pH 7.4), 10 mM EDTA, and 0.5% Triton X-100. Data are presented as mean ± standard error of the mean (SEM). Statistical significance was evaluated using a two-tailed unpaired t-test.

4.6. CDC by C₄₄Mab-1-mG_{2a}

The Calcein AM-labeled target cells (CHO/CD44v3-10, MKN45, and COLO205) were plated and mixed with rabbit complement (final dilution 10%, Low-Tox-M Rabbit Complement; Cedarlane Laboratories, Hornby, ON, Canada) and 100 µg/mL of C₄₄Mab-1-mG_{2a} (n=3) or control mIgG_{2a} (n=3). Following incubation for 4.5 h at 37 °C, the Calcein release into the medium was measured, as described above.

4.7. Antitumor Activity of C₄₄Mab-1-mG_{2a}

The Institutional Committee approved the animal experiment for Animal Experiments of the Institute of Microbial Chemistry (approval number: 2025-011). Throughout the study, mice were housed under specific pathogen-free conditions with an 11-hour light/13-hour dark cycle and provided with food and water *ad libitum*. Body weight was measured twice weekly, and general health status was assessed three times per week. Humane endpoints were predefined as a body weight loss exceeding 25% of the initial weight and/or a tumor volume exceeding 3,000 mm³.

Female BALB/c nude mice (4 weeks old) were obtained from Japan SLC, Inc. Tumor cells (0.3 mL of a 1.33×10^8 cells/mL suspension in DMEM) were mixed with 0.5 mL of BD Matrigel Matrix Growth Factor Reduced (BD Biosciences, San Jose, CA, USA). A 100 µL aliquot of the mixture, containing 5×10^6 cells, was subcutaneously injected into the left flank of each mouse (day 0). To evaluate the antitumor activity of C₄₄Mab-1-mG_{2a} against CHO/CD44v3-10, 100 µg of C₄₄Mab-1-mG_{2a} (n=8) or control mIgG_{2a} (n=8) diluted in 100 µL of PBS was administered intraperitoneally to tumor-bearing mice on day 7 post-inoculation. The mAbs were further administered on days 14 and 21. Mice were euthanized on day 28 following tumor cell implantation. To evaluate the antitumor activity of

C₄₄Mab-1-mG_{2a} against MKN45 and COLO205, 100 µg of C₄₄Mab-1-mG_{2a} (n=8) or control mIgG_{2a} (n=8) diluted in 100 µL of PBS was administered intraperitoneally to tumor-bearing mice on day 7 post-inoculation. A second dose was administered on day 14. Mice were euthanized on day 21 following tumor cell implantation.

Tumor size was measured, and volume was calculated using the formula: $\text{volume} = W^2 \times L / 2$, where W represents the short diameter and L the long diameter. Data are presented as the mean \pm standard error of the mean (SEM). Statistical analysis was performed using one-way ANOVA followed by Sidak's post hoc test. A p-value < 0.05 was considered statistically significant.

Supplementary Materials: The following supporting information can be downloaded at: <https://www.mdpi.com/article/doi/s1>. Figure S1: Flow cytometry analysis using 5-mG_{2a}, an anti-pan-CD44 mAb.; Figure S2: Flow cytometry analysis of C₄₄Mab-1-mG_{2a} to CD44v9-positive gastric cancer cell lines.; Table S1: Immunohistochemical analysis using C₄₄Mab-46-mG_{2a} and C₄₄Mab-1-mG_{2a} against gastric cancer tissue array (BS01011b).

Author Contributions: Conceptualization, M.K.K. and Y.K.; methodology, M.K.K. and T.O.; validation, H.S. and Y.K.; investigation, M.T., T.O., and H.S.; data curation, M.T. and H.S.; writing—original draft preparation, M.T. and H.S.; writing—review and editing, Y.K.; project administration, Y.K.; funding acquisition, Y.K. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported in part by the Japan Agency for Medical Research and Development (AMED) under Grant Numbers: JP25am0521010 (to Y.K.), JP25ama121008 (to Y.K.), JP25ama221339 (to Y.K.), and JP25bm1123027 (to Y.K.), and by the Japan Society for the Promotion of Science (JSPS) Grants-in-Aid for Scientific Research (KAKENHI) grant no—25K10553 (to Y.K.).

Institutional Review Board Statement: Animal experiments were approved by the Institutional Committee for Experiments of the Institute of Microbial Chemistry (approval no. 2025-011, approval date 22 April, and approval no. 2025-029, approval date 22 April).

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available in the article and Supplementary Material.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Stamenkovic, I.; Amiot, M.; Pesando, J.M.; Seed, B. A lymphocyte molecule implicated in lymph node homing is a member of the cartilage link protein family. *Cell* 1989;56(6): 1057-1062.
2. Ponta, H.; Sherman, L.; Herrlich, P.A. CD44: from adhesion molecules to signalling regulators. *Nat Rev Mol Cell Biol* 2003;4(1): 33-45.
3. Mackay, C.R.; Terpe, H.J.; Stauder, R.; et al. Expression and modulation of CD44 variant isoforms in humans. *J Cell Biol* 1994;124(1-2): 71-82.
4. Naor, D.; Nedvetzki, S.; Golan, I.; Melnik, L.; Faitelson, Y. CD44 in cancer. *Crit Rev Clin Lab Sci* 2002;39(6): 527-579.
5. Zöller, M. CD44: can a cancer-initiating cell profit from an abundantly expressed molecule? *Nat Rev Cancer* 2011;11(4): 254-267.
6. Slevin, M.; Krupinski, J.; Gaffney, J.; et al. Hyaluronan-mediated angiogenesis in vascular disease: uncovering RHAMM and CD44 receptor signaling pathways. *Matrix Biol* 2007;26(1): 58-68.
7. Hassn Mesrati, M.; Syafruddin, S.E.; Mohtar, M.A.; Syahir, A. CD44: A Multifunctional Mediator of Cancer Progression. *Biomolecules* 2021;11(12).
8. Guo, Q.; Yang, C.; Gao, F. The state of CD44 activation in cancer progression and therapeutic targeting. *Febs j* 2021.
9. Morath, I.; Hartmann, T.N.; Orian-Rousseau, V. CD44: More than a mere stem cell marker. *Int J Biochem Cell Biol* 2016;81(Pt A): 166-173.

10. Bennett, K.L.; Jackson, D.G.; Simon, J.C.; et al. CD44 isoforms containing exon V3 are responsible for the presentation of heparin-binding growth factor. *J Cell Biol* 1995;128(4): 687-698.
11. Orian-Rousseau, V.; Chen, L.; Sleeman, J.P.; Herrlich, P.; Ponta, H. CD44 is required for two consecutive steps in HGF/c-Met signaling. *Genes Dev* 2002;16(23): 3074-3086.
12. Ishimoto, T.; Nagano, O.; Yae, T.; et al. CD44 variant regulates redox status in cancer cells by stabilizing the xCT subunit of system xc(-) and thereby promotes tumor growth. *Cancer Cell* 2011;19(3): 387-400.
13. Kagami, T.; Yamade, M.; Suzuki, T.; et al. High expression level of CD44v8-10 in cancer stem-like cells is associated with poor prognosis in esophageal squamous cell carcinoma patients treated with chemoradiotherapy. *Oncotarget* 2018;9(79): 34876-34888.
14. Wang, Z.; Tang, Y.; Xie, L.; et al. The Prognostic and Clinical Value of CD44 in Colorectal Cancer: A Meta-Analysis. *Front Oncol* 2019;9: 309.
15. Durko, L.; Wlodarski, W.; Stasikowska-Kanicka, O.; et al. Expression and Clinical Significance of Cancer Stem Cell Markers CD24, CD44, and CD133 in Pancreatic Ductal Adenocarcinoma and Chronic Pancreatitis. *Dis Markers* 2017;2017: 3276806.
16. Gzil, A.; Zarebska, I.; Bursiewicz, W.; et al. Markers of pancreatic cancer stem cells and their clinical and therapeutic implications. *Mol Biol Rep* 2019;46(6): 6629-6645.
17. Li, W.; Qian, L.; Lin, J.; et al. CD44 regulates prostate cancer proliferation, invasion and migration via PDK1 and PFKFB4. *Oncotarget* 2017;8(39): 65143-65151.
18. Ludwig, N.; Szczepanski, M.J.; Gluszko, A.; et al. CD44(+) tumor cells promote early angiogenesis in head and neck squamous cell carcinoma. *Cancer Lett* 2019;467: 85-95.
19. Liu, X.; Taftaf, R.; Kawaguchi, M.; et al. Homophilic CD44 Interactions Mediate Tumor Cell Aggregation and Polyclonal Metastasis in Patient-Derived Breast Cancer Models. *Cancer Discov* 2019;9(1): 96-113.
20. Hassn Mesrati, M.; Behrooz, A.B.; A, Y.A.; Syahir, A. Understanding Glioblastoma Biomarkers: Knocking a Mountain with a Hammer. *Cells* 2020;9(5).
21. Wolf, K.J.; Shukla, P.; Springer, K.; et al. A mode of cell adhesion and migration facilitated by CD44-dependent microtentacles. *Proc Natl Acad Sci U S A* 2020;117(21): 11432-11443.
22. Tawara, M.; Suzuki, H.; Goto, N.; et al. A Novel Anti-CD44 Variant 9 Monoclonal Antibody C44Mab-1 was Developed for Immunohistochemical Analyses Against Colorectal Cancers *Curr. Issues Mol. Biol.* 2023;45(4): 3658-3673.
23. Kaneko, M.K.; Suzuki, H.; Ohishi, T.; et al. A Cancer-Specific Monoclonal Antibody against HER2 Exerts Antitumor Activities in Human Breast Cancer Xenograft Models. *Int J Mol Sci* 2024;25(3).
24. Goto, N.; Suzuki, H.; Tanaka, T.; et al. Development of a Novel Anti-CD44 Monoclonal Antibody for Multiple Applications against Esophageal Squamous Cell Carcinomas. *Int J Mol Sci* 2022;23(10).
25. Orian-Rousseau, V.; Ponta, H. Perspectives of CD44 targeting therapies. *Arch Toxicol* 2015;89(1): 3-14.
26. Menke-van der Houven van Oordt, C.W.; Gomez-Roca, C.; van Herpen, C.; et al. First-in-human phase I clinical trial of RG7356, an anti-CD44 humanized antibody, in patients with advanced, CD44-expressing solid tumors. *Oncotarget* 2016;7(48): 80046-80058.
27. Riechelmann, H.; Sauter, A.; Golze, W.; et al. Phase I trial with the CD44v6-targeting immunoconjugate bivatuzumab mertansine in head and neck squamous cell carcinoma. *Oral Oncol* 2008;44(9): 823-829.
28. Tjink, B.M.; Buter, J.; de Bree, R.; et al. A phase I dose escalation study with anti-CD44v6 bivatuzumab mertansine in patients with incurable squamous cell carcinoma of the head and neck or esophagus. *Clin Cancer Res* 2006;12(20 Pt 1): 6064-6072.
29. Qiu, S.; Iimori, M.; Eda, K.; et al. CD44v3,8-10 is essential for Slug-dependent vimentin gene expression to acquire TGF- β 1-induced tumor cell motility. *Cancer Sci* 2022;113(8): 2654-2667.
30. Ando, T.; Yamasaki, J.; Saya, H.; Nagano, O. CD44: a key regulator of iron metabolism, redox balance, and therapeutic resistance in cancer stem cells. *Stem Cells* 2025;43(6).
31. Hirata, K.; Suzuki, H.; Imaeda, H.; et al. CD44 variant 9 expression in primary early gastric cancer as a predictive marker for recurrence. *Br J Cancer* 2013;109(2): 379-386.
32. Shitara, K.; Doi, T.; Nagano, O.; et al. Dose-escalation study for the targeting of CD44v(+) cancer stem cells by sulfasalazine in patients with advanced gastric cancer (EPOC1205). *Gastric Cancer* 2017;20(2): 341-349.

33. TheCancerGenomeAtlasResearchNetwork. Comprehensive molecular characterization of gastric adenocarcinoma. *Nature* 2014;513(7517): 202-209.
34. Wielenga, V.J.; Smits, R.; Korinek, V.; et al. Expression of CD44 in Apc and Tcf mutant mice implies regulation by the WNT pathway. *Am J Pathol* 1999;154(2): 515-523.
35. Guinney, J.; Dienstmann, R.; Wang, X.; et al. The consensus molecular subtypes of colorectal cancer. *Nat Med* 2015;21(11): 1350-1356.
36. Green, S.J.; Tarone, G.; Underhill, C.B. Distribution of hyaluronate and hyaluronate receptors in the adult lung. *J Cell Sci* 1988;90 (Pt 1): 145-156.
37. Oliifrenko, S.; Paiha, K.; Harder, T.; et al. Analysis of CD44-containing lipid rafts: Recruitment of annexin II and stabilization by the actin cytoskeleton. *J Cell Biol* 1999;146(4): 843-854.
38. Kuhn, S.; Koch, M.; Nübel, T.; et al. A complex of EpCAM, claudin-7, CD44 variant isoforms, and tetraspanins promotes colorectal cancer progression. *Mol Cancer Res* 2007;5(6): 553-567.
39. Casucci, M.; Nicolis di Robilant, B.; Falcone, L.; et al. CD44v6-targeted T cells mediate potent antitumor effects against acute myeloid leukemia and multiple myeloma. *Blood* 2013;122(20): 3461-3472.
40. Porcellini, S.; Asperti, C.; Corna, S.; et al. CAR T Cells Redirected to CD44v6 Control Tumor Growth in Lung and Ovary Adenocarcinoma Bearing Mice. *Front Immunol* 2020;11: 99.
41. Kaneko, M.K.; Suzuki, H.; Kato, Y. Establishment of a Novel Cancer-Specific Anti-HER2 Monoclonal Antibody H(2)Mab-250/H(2)CasMab-2 for Breast Cancers. *Monoclon Antib Immunodiagn Immunother* 2024;43(2): 35-43.
42. Hosking, M.P.; Shirinbak, S.; Omilusik, K.; et al. Preferential tumor targeting of HER2 by iPSC-derived CAR T cells engineered to overcome multiple barriers to solid tumor efficacy. *Cell Stem Cell* 2025;32(7): 1087-1101.e1084.
43. Kudo, Y.; Suzuki, H.; Tanaka, T.; Kaneko, M.K.; Kato, Y. Development of a Novel Anti-CD44 Variant 5 Monoclonal Antibody C(44)Mab-3 for Multiple Applications against Pancreatic Carcinomas. *Antibodies (Basel)* 2023;12(2).
44. Ishikawa, K.; Suzuki, H.; Ohishi, T.; et al. Antitumor activities of anti-CD44 monoclonal antibodies in mouse xenograft models of esophageal cancer. *Oncol Rep* 2024;52(5).
45. Li, G.; Suzuki, H.; Ohishi, T.; et al. Antitumor activities of a defucosylated anti-EpCAM monoclonal antibody in colorectal carcinoma xenograft models. *Int J Mol Med* 2023;51(2).

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.