

Article

Not peer-reviewed version

---

# Recent Advances in Biofilm Cycle by Bibliography Analysis

---

[Yuanzhao Ding](#) \*

Posted Date: 8 January 2025

doi: 10.20944/preprints202501.0368.v1

Keywords: Biofilm Cycle; Bibliography; VOSviewer; Big Data; Machine Learning



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

*Article*

# Recent Advances in Biofilm Cycle by Bibliography Analysis

Yuanzhao Ding

School of Geography and the Environment, University of Oxford, South Parks Road, Oxford, OX1 3QY, United Kingdom. ORCID: 0000-0003-0116-3648. armstrongding@163.com

**Abstract:** Biofilms are communities of microorganisms attached to surfaces, embedded within self-produced extracellular polymeric substances (EPS). They play a dual role in human and environmental contexts. Beneficial biofilms are widely utilized in bioreactors for applications such as energy production and pollutant immobilization, while harmful biofilms are associated with severe infections, including lung infections and urinary tract infections. Understanding the biofilm cycle, encompassing attachment, maturation, and dispersal phases, is critical for optimizing beneficial biofilms and mitigating harmful ones. For example, identifying the optimal late exponential or early maturation phases can enhance biofilm utility, while predicting dispersal timing can inform strategies for disrupting harmful biofilms, safeguarding human health. This study employs a bibliometric approach to analyze the biofilm cycle, extracting core keywords, leading organizations, and active countries/regions. By synthesizing data from the Web of Science, this analysis highlights key trends and knowledge gaps in the field, providing a comprehensive overview of biofilm cycle dynamics. The study also identifies future opportunities for integrating big data and machine learning, which have the potential to revolutionize biofilm research. These technologies could enable precise predictions of biofilm behavior, fostering advancements in healthcare, environmental protection, and industrial processes, and ensuring more effective biofilm management strategies.

**Keywords:** biofilm cycle; bibliography; VOSviewer; big data; machine learning

## 1. Introduction

Biofilms are complex communities of microorganisms that attach to surfaces and are encased in self-produced extracellular polymeric substances (EPS) [1,2]. This structure provides a protective and stable environment for the microorganisms, enabling them to thrive under various conditions [3,4]. The study of biofilms is critical due to their dual nature: beneficial biofilms can be harnessed for environmentally friendly applications like bioreactors for energy production [5] or pollutant immobilization [6], while harmful biofilms can lead to serious medical and environmental problems [7,8]. For instance, pathogenic biofilms are a significant cause of infections, including lung infections [9] and urinary tract infections [10], presenting challenges in healthcare and sanitation.

Understanding the biofilm cycle is central to both utilizing beneficial biofilms [5] and mitigating harmful ones [9,10]. The biofilm cycle encompasses various phases, including the attachment phase, maturation phase, and dispersal phase [11]. Each phase involves specific physiological and structural changes within the biofilm community. For example, knowing the duration of the attachment phase can help optimize the conditions for beneficial biofilm growth [12], while understanding the maturation phase timeline aids in exploiting biofilms at their most productive stage, such as during the late exponential or early maturation phase [13]. Similarly, insights into the dispersal phase can inform strategies to disrupt harmful biofilms before they become more resilient or infectious, helping to protect human health by anticipating critical periods in biofilm development.

This study employs a bibliographic approach [14,15] to analyze the biofilm cycle comprehensively, identifying core keywords, leading organizations, and influential

countries/regions in this field. By examining the literature, the research maps out the dominant trends and patterns in biofilm cycle studies [16,17], providing a clearer understanding of how these microbial communities develop and behave. The analysis not only summarizes the current knowledge base but also highlights critical gaps and opportunities for innovation. Through this systematic evaluation, the study aims to deepen our understanding of biofilm dynamics and provide actionable insights for both researchers and practitioners.

## 2. Materials and Methods

The method followed previous studies with slightly modifications [18,19]. This study conducted a bibliometric analysis focusing on the term "Biofilm Cycle" by searching the Web of Science database [20,21] on December 6, 2024. The search yielded 4,385 results, reflecting the significant attention this topic has garnered in recent years. To ensure manageable and insightful analysis, we selected the default 1,000 most relevant articles for further investigation using VOSviewer (Version 1.6.20) [22,23]. This subset represents a comprehensive but concise dataset that aligns with the study's aim of identifying dominant trends, key contributors, and geographical focus in biofilm research. The decision to limit the dataset to 1,000 records ensures that the analysis remains focused and visually interpretable while retaining enough data for meaningful insights.

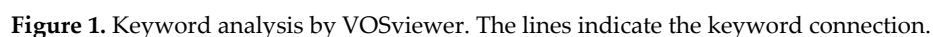
For the keyword co-occurrence analysis, we set the minimum occurrence threshold at "5," which means only keywords that appeared at least five times across the selected articles were included. This threshold was chosen to strike a balance between comprehensiveness and clarity, allowing us to highlight prominent themes without cluttering the visualization with less relevant or infrequently occurring terms. This approach revealed recurring topics and concepts in biofilm research, emphasizing their importance in understanding biofilm cycles. Too low a threshold would have overwhelmed the visualization with minor terms, while a higher threshold risked overlooking potentially critical but less frequent keywords.

In the organizational collaboration analysis, we applied a similar logic, selecting organizations with at least five published documents. This criterion was designed to highlight institutions with consistent contributions to biofilm research while filtering out organizations with minimal involvement. The resulting analysis revealed leading research institutions driving advancements in this field and their collaborative networks. Such insights are crucial for identifying centers of excellence and potential partnerships in biofilm studies, offering a roadmap for researchers seeking collaborators or understanding institutional priorities.

For the country and regional analysis, a threshold of ten documents was established to focus on countries with substantial contributions to the field. This choice enabled the identification of regions with robust research activity while minimizing the noise from countries with only marginal contributions. The analysis highlighted global trends in biofilm research, showing a strong concentration of activity in developed nations with well-funded scientific programs. It also provided a comparative perspective on regional expertise and resources in biofilm studies, offering valuable context for future international collaborations and funding strategies. These thresholds and methodological decisions collectively ensured that our analysis remained both manageable and impactful.

## 3. Results

Figure 1 highlights the results of the keyword analysis, offering a comprehensive view of the research focus within the biofilm cycle domain. A significant group of keywords pertains to microbial strains, including "Escherichia coli," "Pseudomonas aeruginosa," and "Staphylococcus aureus." These bacteria are prominent in biofilm studies due to their roles in both beneficial and harmful biofilm activities. For example, *Escherichia coli* is commonly associated with biofilm formation in industrial and medical contexts, while *Pseudomonas aeruginosa* is a well-known pathogen that thrives in biofilm states, complicating infection treatment. The frequent appearance of these



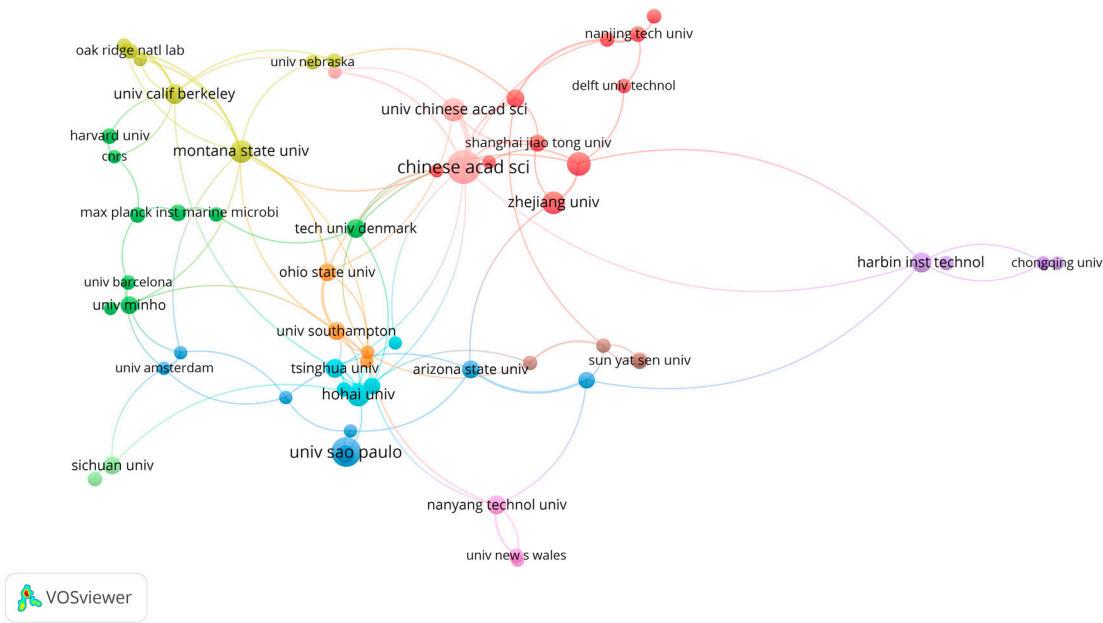
A third cluster of keywords relates to pollutants, including “copper,” “nitrite,” “nitrate,” and “phosphorus.” These terms reflect a growing focus on biofilms’ interactions with contaminants, particularly their ability to immobilize, transform, or degrade these substances. For instance, biofilms can sequester heavy metals like copper or reduce nitrates and nitrites in water systems, offering potential solutions for pollution mitigation. The presence of phosphorus as a keyword suggests interest in biofilms’ role in nutrient dynamics, especially in preventing eutrophication. This cluster highlights the applied significance of biofilm research, particularly in environmental remediation and industrial wastewater treatment.

Lastly, several keywords pertain to biological processes fundamental to the biofilm cycle, including “adhesion,” “infections,” “dispersal,” and “resistance.” These terms represent key phases and characteristics of biofilm formation and development. For instance, adhesion marks the initial attachment phase, which is critical for biofilm establishment, while dispersal reflects the phase where cells leave the biofilm, spreading to new locations. Keywords like “infections” and “resistance” are particularly relevant in medical contexts, highlighting the challenges biofilms pose in healthcare, such as their role in persistent infections and their heightened resistance to antibiotics. Together, these process-related keywords emphasize the complexity of biofilm behavior and the necessity of understanding these dynamics for both harnessing biofilm benefits and mitigating their risks.

Figure 2 illustrates the primary research organizations contributing to the field of biofilm cycle studies, with several institutions occupying central positions in the network. At the core are renowned organizations such as the Chinese Academy of Sciences, Technical University of Denmark,



Ohio State University, Montana State University, University of California, Berkeley, Oak Ridge National Laboratory, Harvard University, Tsinghua University, and Nanyang Technological University. These institutions are distinguished by their extensive contributions to biofilm research, encompassing areas like microbial behavior, biofilm formation, and environmental applications. Their central positioning reflects their critical role in driving innovation, disseminating knowledge, and shaping the field's research trajectory.



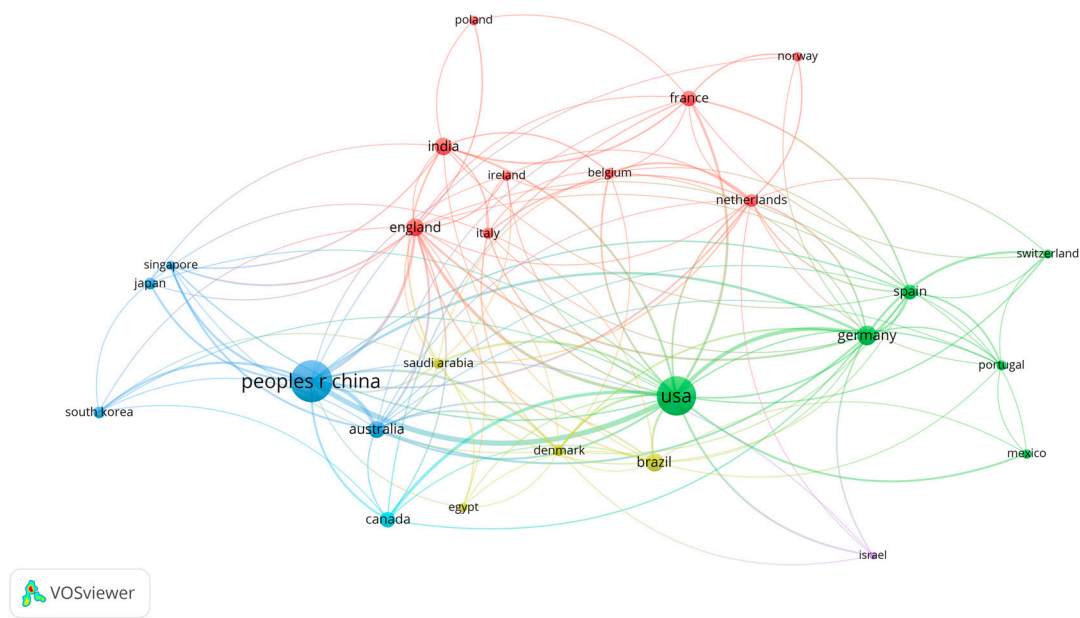
**Figure 2.** Organizational analysis by VOSviewer. The lines indicate the research collaboration.

Beyond these core institutions, numerous other organizations also play significant roles in advancing biofilm cycle research. These include Nanjing Tech University, Delft University of Technology, Shanghai Jiao Tong University, Zhejiang University, Harbin Institute of Technology, Chongqing University, Sun Yat-sen University, Arizona State University, University of New South Wales, University of São Paulo, Hohai University, University of Southampton, Sichuan University, University of Amsterdam, University of Minho, University of Barcelona, Max Planck Institute for Marine Microbiology, and CNRS. These organizations contribute through diverse research efforts, from investigating biofilm mechanics to exploring its applications in environmental and medical sciences. Their geographic distribution also highlights the global interest and collaborative nature of this field, with contributions spanning continents.

Collaboration among these institutions is a defining feature of the biofilm cycle research landscape. By leveraging shared expertise, resources, and datasets, these organizations have collectively advanced our understanding of biofilm dynamics. The partnerships between institutions, both domestic and international, have facilitated groundbreaking discoveries, fostered interdisciplinary research, and accelerated the development of practical applications. The network depicted in Figure 2 underscores the importance of such collaborations in addressing complex scientific questions, demonstrating how collective efforts can propel the biofilm cycle research field forward. This interconnected framework not only enhances scientific productivity but also encourages innovation through diverse perspectives and shared objectives.

Figure 3 highlights the major countries and regions contributing to the field of biofilm cycle research, emphasizing the global nature of scientific collaboration in this domain. At the forefront are China and the United States, which dominate as the two most influential countries in biofilm research. These nations lead in terms of publication volume, research funding, and the establishment of cutting-edge facilities. Their contributions span a wide array of topics, from fundamental studies

on biofilm formation and dispersal phases to practical applications in healthcare, environmental remediation, and industrial processes. China’s rapid rise in research output, supported by initiatives like the Chinese Academy of Sciences, and the United States’ strong academic and industrial partnerships, have solidified their positions as the primary drivers of innovation in this field.



**Figure 3.** Country/region analysis by VOSviewer. The lines indicate the research collaboration.

Beyond China and the United States, several other countries have established themselves as significant contributors to biofilm cycle research. In Asia, nations such as South Korea, Japan, and Singapore have made substantial advancements, often focusing on technological innovations and interdisciplinary research approaches. Similarly, Canada and Australia have contributed extensively, particularly in environmental and medical biofilm studies. In Europe, countries like the United Kingdom, Germany, France, Italy, Belgium, Denmark, Ireland, Portugal, and Spain have long-standing traditions in microbiology and biotechnology, enriching the field through diverse research perspectives. Emerging players like Saudi Arabia, Egypt, India, Brazil, and Mexico reflect a growing global interest, bringing unique regional challenges and solutions to biofilm research.

International collaboration among these nations is a hallmark of the biofilm cycle research field, fostering the exchange of knowledge, methodologies, and resources. For instance, partnerships between developed and emerging economies facilitate capacity building and enable access to diverse datasets, enhancing the scope and applicability of biofilm studies. Cross-border initiatives have resulted in breakthroughs in understanding biofilm-related health challenges and environmental issues, demonstrating the value of a collective global effort. Figure 3 underscores the importance of such collaborations, illustrating how nations, despite their geographical and cultural differences, work together to advance the understanding and application of biofilm cycles, ultimately addressing critical global challenges.

## 4. Discussion

### 4.1. Analysis of the Biofilm Cycle Stages

Biofilm formation is a complex and dynamic process that occurs in five distinct stages: 1) Bacteria Land on Surface, 2) Irreversible Attachment, 3) Proliferation, 4) Maturation, and 5) Dispersal [24]. These stages represent the lifecycle of biofilm formation, starting with the initial contact of bacteria

with a surface and culminating in the release of bacterial cells into the surrounding environment. Bibliographic analysis has been instrumental in understanding the various factors that influence each stage, including microbial interactions, environmental conditions, and the role of extracellular matrix (ECM) components [25].

In the first stage, Bacteria Land on Surface, free-floating bacteria encounter and adhere to a surface [26,27]. This is a reversible attachment, where the bacteria are loosely bound to the surface through weak interactions such as van der Waals forces and electrostatic interactions. The choice of surface material, surface roughness, and environmental factors such as nutrient availability significantly influence this initial phase. Studies have shown that surfaces with hydrophobic properties tend to attract more bacterial adhesion, especially for certain pathogenic strains.

The second stage, Irreversible Attachment, is marked by stronger adhesion of bacteria to the surface [28,29]. This stage is characterized by the production of surface-associated molecules, such as pili and fimbriae, that anchor the bacteria more firmly. Once bacteria irreversibly attach, they begin to produce extracellular polymeric substances (EPS), a key feature of biofilm formation. EPS consists of polysaccharides, proteins, and nucleic acids that contribute to the stability and integrity of the biofilm. This stage is critical because it sets the foundation for further biofilm growth and maturation.

Proliferation, the third stage, involves the growth and division of the attached bacterial cells, leading to an increase in biomass [30,31]. As the bacterial population expands, the biofilm structure becomes more complex. The growing bacteria interact with each other, forming microcolonies that are encased in the EPS matrix. These microcolonies are not homogenous, and gradients of nutrients, oxygen, and waste products often exist within the biofilm, leading to a heterogeneous distribution of microbial activity. This stage is essential for the development of biofilm architecture and the establishment of microbial communities with diverse functions.

The Maturation stage sees the biofilm reaching its full structural complexity [32,33]. The biofilm becomes more organized, with distinct layers and channels formed for nutrient and waste exchange. This stage is characterized by the production of a robust extracellular matrix that provides the biofilm with resistance to antimicrobial agents and physical stress. Maturation enhances the biofilm’s ability to persist in various environments, making it more resilient to external challenges, such as antibiotics and immune responses. At this stage, biofilms can exhibit a high degree of resistance to treatment, which poses challenges in medical and industrial contexts.

Finally, in the Dispersal stage, bacteria are released from the biofilm into the surrounding environment, either as single cells or as clusters [34,35]. This is a crucial stage for biofilm propagation, as it enables bacteria to colonize new surfaces and expand the biofilm. Dispersal can be triggered by environmental changes, such as nutrient depletion, or by the biofilm reaching a certain size. The release of bacteria allows the biofilm to spread, leading to the colonization of new niches. This stage is also associated with the formation of new biofilms in different locations, contributing to the persistence of bacterial populations.

**Table 1.** Summary of biofilm cycle stages.

Stages	Main Characteristics	References
1.Bacteria Land on Surface	- Bacteria touch the surface. - Weak, temporary bonds form (e.g., physical forces).	[26,27]
2.Irreversible Attachment	- Bacteria stick firmly using structures like pili or slime (EPS). - Hard to remove.	[28,29]
3.Proliferation	- Bacteria grow and produce slime to form a shield. - They communicate and work together.	[30,31]
4. Maturation	- Biofilm gets thicker and more complex. - High resistance to antibiotics and harsh conditions.	[32,33]

5. Dispersal	<div>- Parts of the biofilm break off. - Bacteria spread to new places to start over.</div>	[34,35]
--------------	---	---------

4.2. Beneficial Biofilm in Biofilm Reactors and Harmful Biofilm in Infections

Biofilms are highly dynamic microbial communities that develop through distinct stages, with the late Proliferation phase and early Maturation phase being particularly important in determining their potential and strength [33,36]. During the late Proliferation phase, the microbial population within the biofilm increases significantly, and the EPS that form the biofilm's matrix begin to solidify. This creates a highly organized structure that is resistant to external environmental stressors, such as antibiotics and immune responses. The early Maturation phase marks the start of biofilm's structural development, where cells continue to organize into complex, multi-layered structures that contribute to the biofilm's increased resilience. These phases enhance the biofilm's stability and resistance, making it difficult to remove and giving it its maximum potential strength. Studies have shown that biofilms formed during these stages are particularly effective in both beneficial and harmful applications, as their growth and development lead to a more resilient and long-lasting structure

In terms of beneficial applications, biofilms have been utilized in various fields with impressive results. For instance, biofilms play an important role in environmental remediation by immobilizing heavy metals in water treatment processes [6]. Microorganisms within the biofilm matrix are able to break down and detoxify harmful substances, improving the quality of water and reducing environmental pollution. Biofilms are also integral to microbial fuel cells [5,37], where they facilitate electricity generation by harnessing the metabolic activity of microorganisms to convert organic matter into energy [38]. This provides a sustainable energy solution with potential applications in waste treatment and energy production. Additionally, biofilms are being explored in the field of construction, particularly in self-healing concrete [39]. In this application, bacteria embedded in the biofilm matrix react to cracks in the concrete by precipitating calcium carbonate, which fills the cracks and helps to prolong the lifespan of the material [40,41]. These advancements demonstrate how biofilms can be harnessed for sustainable practices across diverse industries.

However, biofilms can also have significant negative impacts, particularly when pathogenic microorganisms form biofilms on medical devices or within the human body. For example, *Pseudomonas aeruginosa* forms biofilms in the lungs of individuals with cystic fibrosis, leading to chronic lung infections that are difficult to treat [42,43]. Similarly, *Staphylococcus aureus* is a common pathogen in urinary tract infections [44,45], and *Escherichia coli* biofilms can cause severe gastrointestinal diseases such as diarrhea [46,47]. The resilience of these pathogenic biofilms makes them particularly challenging to eradicate, requiring advanced treatments. To prevent the formation of harmful biofilms, researchers have developed various strategies, such as incorporating titanium dioxide (TiO<sub>2</sub>) into surface coatings like concrete and medical devices [48,49]. TiO<sub>2</sub> has antimicrobial properties that prevent bacterial adhesion and biofilm formation, thus reducing the risk of infections [50,51]. This strategy is particularly effective in reducing the impact of biofilms in healthcare settings and promoting hygiene in environments where bacterial contamination is a concern.

4.3. Future Direction: Big Data and Machine Learning on Biofilm Cycle

The future of biofilm cycle research holds immense promise, especially with the integration of big data and machine learning technologies [52,53]. These tools offer powerful capabilities for analyzing and predicting complex biological processes, such as biofilm formation and behavior. In recent years, big data and machine learning have already made significant strides in fields such as facial recognition [54,55], autonomous driving [56,57], species distribution prediction [58], and educational outcome forecasting [59]. Their success in these domains illustrates the potential for similar breakthroughs in biofilm research. Machine learning algorithms can analyze vast datasets to uncover patterns and correlations that would be difficult or impossible to detect through traditional methods. As biofilm research advances, leveraging these technologies could lead to more precise and



comprehensive models of biofilm formation, maturation, and detachment processes, helping to improve disease prevention and treatment strategies.

One of the key innovations that big data and machine learning [60] can bring to biofilm research is the development of large-scale databases that track various factors influencing biofilm dynamics. Such a database could include data on bacterial species, surface materials to which bacteria attach, nutrient media composition, biofilm growth duration, and detachment times. By compiling vast amounts of experimental and observational data, researchers can create a comprehensive framework for understanding the multiple variables that govern biofilm behavior. This database would provide a solid foundation for developing predictive models that could forecast the progression of biofilm-related phenomena under different conditions. For instance, machine learning models could analyze this data to predict when a specific biofilm might become most virulent, such as in the case of human infections, enabling timely interventions and personalized medical treatments.

Building upon the foundation of large databases, the use of neural networks [61,62] and other machine learning techniques could further refine predictions and diagnostics in biofilm cycle research. By training neural networks on extensive datasets, researchers could develop models capable of accurately predicting biofilm formation stages, identifying risk factors for infection, and determining when biofilms reach their most dangerous state. For example, a trained neural network model could assess factors such as bacterial species, patient health conditions, and environmental variables to predict when biofilm formation would peak, giving healthcare providers a better understanding of when patients might be at their most vulnerable to biofilm-related infections. This predictive capability would be especially valuable in clinical settings, where timely detection and intervention are critical to patient outcomes. By combining big data with machine learning, the potential to revolutionize biofilm research and enhance both clinical and environmental management is vast.

## 5. Conclusions

In conclusion, this study utilized bibliographic analysis to explore key aspects of the biofilm cycle, including prominent keywords, organizations, and countries/regions involved in the research. The five stages of biofilm development—bacteria landing on surfaces, irreversible attachment, proliferation, maturation, and dispersal—were summarized from essential literature. The study also highlighted biofilm's significant industrial applications and its potential health risks. Looking ahead, the integration of big data and machine learning is presented as a promising opportunity for enhancing biofilm cycle research, offering new methods for prediction and management in both industrial and medical contexts.

## References

1. Flemming, H.-C.; Wingender, J. The biofilm matrix. *Nature reviews microbiology* **2010**, *8*, 623-633.
2. Xiao, R.; Zheng, Y. Overview of microalgal extracellular polymeric substances (EPS) and their applications. *Biotechnology advances* **2016**, *34*, 1225-1244.
3. Siddharth, T.; Sridhar, P.; Vinila, V.; Tyagi, R.D. Environmental applications of microbial extracellular polymeric substance (EPS): a review. *Journal of Environmental Management* **2021**, *287*, 112307.
4. Flemming, H.-C.; Neu, T.R.; Wozniak, D.J. The EPS matrix: the "house of biofilm cells". *Journal of bacteriology* **2007**, *189*, 7945-7947.
5. Zhao, C.-e.; Chen, J.; Ding, Y.; Wang, V.B.; Bao, B.; Kjelleberg, S.; Cao, B.; Loo, S.C.J.; Wang, L.; Huang, W. Chemically functionalized conjugated oligoelectrolyte nanoparticles for enhancement of current generation in microbial fuel cells. *ACS Applied Materials & Interfaces* **2015**, *7*, 14501-14505.
6. Ding, Y.; Peng, N.; Du, Y.; Ji, L.; Cao, B. Disruption of putrescine biosynthesis in *Shewanella oneidensis* enhances biofilm cohesiveness and performance in Cr (VI) immobilization. *Applied and environmental microbiology* **2014**, *80*, 1498-1506.
7. Costerton, J.W.; Stewart, P.S. Battling biofilms. *Scientific American* **2001**, *285*, 74-81.

8. Rather, M.A.; Gupta, K.; Bardhan, P.; Borah, M.; Sarkar, A.; Eldiehy, K.S.H.; Bhuyan, S.; Mandal, M. Microbial biofilm: A matter of grave concern for human health and food industry. *Journal of Basic Microbiology* **2021**, *61*, 380-395.
9. Maurice, N.M.; Bedi, B.; Sadikot, R.T. Pseudomonas aeruginosa biofilms: host response and clinical implications in lung infections. *American journal of respiratory cell and molecular biology* **2018**, *58*, 428-439.
10. Trautner, B.W.; Darouiche, R.O. Role of biofilm in catheter-associated urinary tract infection. *American journal of infection control* **2004**, *32*, 177-183.
11. Sauer, K.; Stoodley, P.; Goeres, D.M.; Hall-Stoodley, L.; Burmølle, M.; Stewart, P.S.; Bjarnsholt, T. The biofilm life cycle: expanding the conceptual model of biofilm formation. *Nature Reviews Microbiology* **2022**, *20*, 608-620.
12. Teughels, W.; Van Assche, N.; Sliepen, I.; Quirynen, M. Effect of material characteristics and/or surface topography on biofilm development. *Clinical oral implants research* **2006**, *17*, 68-81.
13. Lappin-Scott, H.M.; Bass, C. Biofilm formation: attachment, growth, and detachment of microbes from surfaces. *American journal of infection control* **2001**, *29*, 250-251.
14. Chen, S.; Ding, Y. From bibliography to understanding: water microbiology and human health. *Journal of Water and Health* **2024**, *22*, 1911-1921.
15. Chen, S.; Ding, Y. A bibliography study of Shewanella oneidensis biofilm. *FEMS Microbiology Ecology* **2023**, *99*, fiad124.
16. Ma, R.; Hu, X.; Zhang, X.; Wang, W.; Sun, J.; Su, Z.; Zhu, C. Strategies to prevent, curb and eliminate biofilm formation based on the characteristics of various periods in one biofilm life cycle. *Frontiers in Cellular and Infection Microbiology* **2022**, *12*, 1003033.
17. Rice, S.A.; Tan, C.H.; Mikkelsen, P.J.; Kung, V.; Woo, J.; Tay, M.; Hauser, A.; McDougald, D.; Webb, J.S.; Kjelleberg, S. The biofilm life cycle and virulence of Pseudomonas aeruginosa are dependent on a filamentous prophage. *The ISME journal* **2009**, *3*, 271-282.
18. Chen, S.; Ding, Y. Tackling heavy metal pollution: evaluating governance models and frameworks. *Sustainability* **2023**, *15*, 15863.
19. Chen, S.; Ding, Y. Bibliographic Insights into Biofilm Engineering. *Acta Microbiologica Hellenica* **2024**, *69*, 3-13.
20. Zhu, J.; Liu, W. A tale of two databases: the use of Web of Science and Scopus in academic papers. *Scientometrics* **2020**, *123*, 321-335.
21. Mongeon, P.; Paul-Hus, A. The journal coverage of Web of Science and Scopus: a comparative analysis. *Scientometrics* **2016**, *106*, 213-228.
22. Van Eck, N.J.; Waltman, L. Citation-based clustering of publications using CitNetExplorer and VOSviewer. *Scientometrics* **2017**, *111*, 1053-1070.
23. Bukar, U.A.; Sayeed, M.S.; Razak, S.F.A.; Yogarayan, S.; Amodu, O.A.; Mahmood, R.A.R. A method for analyzing text using VOSviewer. *MethodsX* **2023**, *11*, 102339.
24. Hall-Stoodley, L.; Stoodley, P. Biofilm formation and dispersal and the transmission of human pathogens. *Trends in microbiology* **2005**, *13*, 7-10.
25. Toyofuku, M.; Roschitzki, B.; Riedel, K.; Eberl, L. Identification of proteins associated with the Pseudomonas aeruginosa biofilm extracellular matrix. *Journal of proteome research* **2012**, *11*, 4906-4915.
26. Palmer, J.; Flint, S.; Brooks, J. Bacterial cell attachment, the beginning of a biofilm. *Journal of Industrial Microbiology and Biotechnology* **2007**, *34*, 577-588.
27. Feng, G.; Cheng, Y.; Wang, S.-Y.; Borca-Tasciuc, D.A.; Worobo, R.W.; Moraru, C.I. Bacterial attachment and biofilm formation on surfaces are reduced by small-diameter nanoscale pores: how small is small enough? *npj Biofilms and Microbiomes* **2015**, *1*, 1-9.
28. Hinsa, S.M.; Espinosa-Urgel, M.; Ramos, J.L.; O'Toole, G.A. Transition from reversible to irreversible attachment during biofilm formation by Pseudomonas fluorescens WCS365 requires an ABC transporter and a large secreted protein. *Molecular microbiology* **2003**, *49*, 905-918.
29. Caiazza, N.C.; O'Toole, G.A. SadB is required for the transition from reversible to irreversible attachment during biofilm formation by Pseudomonas aeruginosa PA14. **2004**.

30. Krsmanovic, M.; Biswas, D.; Ali, H.; Kumar, A.; Ghosh, R.; Dickerson, A.K. Hydrodynamics and surface properties influence biofilm proliferation. *Advances in Colloid and Interface Science* **2021**, *288*, 102336.
31. He, W.; Liu, H.; Wang, Z.; Tay, F.R.; Shen, Y. The dynamics of bacterial proliferation, viability, and extracellular polymeric substances in oral biofilm development. *Journal of Dentistry* **2024**, *143*, 104882.
32. Otto, M. Staphylococcal infections: mechanisms of biofilm maturation and detachment as critical determinants of pathogenicity. *Annual review of medicine* **2013**, *64*, 175-188.
33. Reisner, A.; Haagensen, J.A.J.; Schembri, M.A.; Zechner, E.L.; Molin, S. Development and maturation of *Escherichia coli* K-12 biofilms. *Molecular microbiology* **2003**, *48*, 933-946.
34. Kaplan, J.Á. Biofilm dispersal: mechanisms, clinical implications, and potential therapeutic uses. *Journal of dental research* **2010**, *89*, 205-218.
35. Rumbaugh, K.P.; Sauer, K. Biofilm dispersion. *Nature Reviews Microbiology* **2020**, *18*, 571-586.
36. Kierek-Pearson, K.; Karatan, E. Biofilm development in bacteria. *Advances in applied microbiology* **2005**, *57*, 79-111.
37. Zhao, C.e.; Wu, J.; Ding, Y.; Wang, V.B.; Zhang, Y.; Kjelleberg, S.; Loo, J.S.C.; Cao, B.; Zhang, Q. Hybrid conducting biofilm with built-in bacteria for high-performance microbial fuel cells. *ChemElectroChem* **2015**, *2*, 654-658.
38. Yang, Y.; Ding, Y.; Hu, Y.; Cao, B.; Rice, S.A.; Kjelleberg, S.; Song, H. Enhancing bidirectional electron transfer of *Shewanella oneidensis* by a synthetic flavin pathway. *ACS synthetic biology* **2015**, *4*, 815-823.
39. Zhang, Z.; Ding, Y.; Qian, S. Influence of bacterial incorporation on mechanical properties of engineered cementitious composites (ECC). *Construction and Building Materials* **2019**, *196*, 195-203.
40. Zhang, Z.; Weng, Y.; Ding, Y.; Qian, S. Use of genetically modified bacteria to repair cracks in concrete. *Materials* **2019**, *12*, 3912.
41. Zhang, Z.; Liu, D.; Ding, Y.; Wang, S. Mechanical performance of strain-hardening cementitious composites (SHCC) with bacterial addition. *Journal of Infrastructure Preservation and Resilience* **2022**, *3*, 3.
42. Reynolds, D.; Kollef, M. The epidemiology and pathogenesis and treatment of *Pseudomonas aeruginosa* infections: an update. *Drugs* **2021**, *81*, 2117-2131.
43. Bisht, K.; Baishya, J.; Wakeman, C.A. *Pseudomonas aeruginosa* polymicrobial interactions during lung infection. *Current opinion in microbiology* **2020**, *53*, 1-8.
44. Saenkharn-Huntsinger, P.; Hyre, A.N.; Hanson, B.S.; Donati, G.L.; Adams, L.G.; Ryan, C.; Londoño, A.; Moustafa, A.M.; Planet, P.J.; Subashchandrabose, S. Copper resistance promotes fitness of methicillin-resistant *Staphylococcus aureus* during urinary tract infection. *Mbio* **2021**, *12*, 10-1128.
45. Selim, S.; Faried, O.A.; Almuhayawi, M.S.; Saleh, F.M.; Sharaf, M.; El Nahhas, N.; Warrad, M. Incidence of vancomycin-resistant *Staphylococcus aureus* strains among patients with urinary tract infections. *Antibiotics* **2022**, *11*, 408.
46. Robins-Browne, R.M.; Hartland, E.L. *Escherichia coli* as a cause of diarrhea. *Journal of gastroenterology and hepatology* **2002**, *17*, 467-475.
47. Cabrera-Sosa, L.; Ochoa, T.J. *Escherichia coli* diarrhea. In *Hunter's Tropical Medicine and Emerging Infectious Diseases*; Elsevier: 2020; pp. 481-485.
48. Hamdany, A.H.; Ding, Y.; Qian, S. Graphene-Based TiO<sub>2</sub> Cement Composites to Enhance the Antibacterial Effect of Self-Disinfecting Surfaces. *Catalysts* **2023**, *13*, 1313.
49. Hamdany, A.H.; Ding, Y.; Qian, S. Visible light antibacterial potential of graphene-TiO<sub>2</sub> cementitious composites for self-sterilization surface. *Journal of Sustainable Cement-Based Materials* **2023**, *12*, 972-982.
50. Hamdany, A.H.; Ding, Y.; Qian, S. Cementitious Composite Materials for Self-Sterilization Surfaces. *ACI Materials Journal* **2022**, *119*, 197-210.
51. Hamdany, A.H.; Ding, Y.; Qian, S. Mechanical and antibacterial behavior of photocatalytic lightweight engineered cementitious composites. *Journal of Materials in Civil Engineering* **2021**, *33*, 04021262.
52. Zhou, L.; Pan, S.; Wang, J.; Vasilakos, A.V. Machine learning on big data: Opportunities and challenges. *Neurocomputing* **2017**, *237*, 350-361.
53. L'heureux, A.; Grolinger, K.; Elyamany, H.F.; Capretz, M.A.M. Machine learning with big data: Challenges and approaches. *Ieee Access* **2017**, *5*, 7776-7797.

54. Almeida, D.; Shmarko, K.; Lomas, E. The ethics of facial recognition technologies, surveillance, and accountability in an age of artificial intelligence: a comparative analysis of US, EU, and UK regulatory frameworks. *AI and Ethics* **2022**, *2*, 377-387.
55. Kaur, P.; Krishan, K.; Sharma, S.K.; Kanchan, T. Facial-recognition algorithms: A literature review. *Medicine, Science and the Law* **2020**, *60*, 131-139.
56. Atakishiyev, S.; Salameh, M.; Yao, H.; Goebel, R. Explainable artificial intelligence for autonomous driving: A comprehensive overview and field guide for future research directions. *IEEE Access* **2024**.
57. Ning, H.; Yin, R.; Ullah, A.; Shi, F. A survey on hybrid human-artificial intelligence for autonomous driving. *IEEE Transactions on Intelligent Transportation Systems* **2021**, *23*, 6011-6026.
58. Chen, S.; Ding, Y. Machine learning and its applications in studying the geographical distribution of ants. *Diversity* **2022**, *14*, 706.
59. Chen, S.; Ding, Y. A machine learning approach to predicting academic performance in Pennsylvania's schools. *Social Sciences* **2023**, *12*, 118.
60. Yang, C.-T.; Kristiani, E.; Leong, Y.K.; Chang, J.-S. Big data and machine learning driven bioprocessing—recent trends and critical analysis. *Bioresource technology* **2023**, *372*, 128625.
61. Kaveh, A. Applications of artificial neural networks and machine learning in civil engineering. *Studies in computational intelligence* **2024**, *1168*, 472.
62. Yang, X.; Ye, J.; Wang, X. Factorizing knowledge in neural networks. 2022; pp. 73-91.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.