Review

The Current Status of Cigarette Smoking and Air Pollution Associated Lung Cancer

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Abstract: Cigarette smoking and air pollution (particulate matter) are recognized as two major etiological factors for lung cancer. Of all the risk factors, cigarette smoking is significantly associated with lung carcinogenesis. The main mechanism lies in the metabolically activated carcinogens (majorly polycyclic aromatic hydrocarbons and nitrosamines), which could covalently bind with DNA molecules and lead to irreversible mutations in pivotal cancer genes, such as *TP53* and *KRAS*. Another major etiological factor for lung cancer is air pollution, which is with complex compositions and ubiquitous in daily life, especially in developing countries as China and India. The latest literatures on lung cancer epidemiology and etiology have been briefly summarized and reviewed in this work

Keywords: lung cancer; cigarette smoking; air pollution; epidemiology; etiology

1. Introduction

Cancer is a leading cause for death worldwide, accounting for nearly 10 million deaths in 2020. Specially, lung cancer is one of the most common types of cancer and imposes a huge health burden for humans (1). The five-year survival of lung cancer patients in the United States and developing countries were only 14% and 8%, respectively (2). There are many verified risk factors responsible for lung carcinogenesis. Etiologically, it is believed that cigarette smoking (CS) and extensive air pollutions are the two major risk factors for lung cancer.

The cultivation of the tobacco plant has a very long history that stretches back to 6,000 BC in the Americas. It was thought that tobacco smoking was primarily performed in religious ceremonies and also used for medical purposes in the ancient indigenous American tribes circa 1 BC. In 1912, Adler proposed that CS might be associated with the growing incidence of lung cancer (3). In 1964, the U.S. Surgeon General definitively announced that CS causes lung cancer and laryngeal cancer (4). It is evidenced that cigarette smoking (CS) not only increases the lung cancer risks. In the Human Early-Life Exposome project containing 1,173 children aged 7 years old, it was found that maternal tobacco smoking during pregnancy and CS exposure in childhood could also influence future health and accelerate biological aging from an early age (5). However, the CS associated lung cancer could largely be prevented through smoking cessation. It is never too late to act, quit smoking as early as possible is of much beneficial to you and your children's future.

For a relatively long period of time, ubiquitous air pollution is posing an urgent challenge to worldwide public health. Ambient air pollution was classified as Group 1 carcinogens for lung cancer by the International Agency for Research on Cancer (IARC) in 2013 (6). Substantial evidence, including mechanistic studies, have demonstrated that the air pollution, especially the outdoor particulate matter (PM), is causatively associated with lung cancer (7). More than one-half of lung cancer deaths attributable to air pollution were projected to East Asian countries, especially China (8). However, air pollution is a manmade disaster attributed to everyone. We are an integral part of the environment, and the

responsibility to curb air pollution and effectively protect the environment lies with all of us.

2. The epidemiology of lung cancer from 1990 to 2020

In the three decades between 1990 and 2020, cancer incidence and mortality continuously grew worldwide (Figure 1A) (1, 2, 9-11). Based on the data from GLOBOCAN, 8.1 million cancer cases occurred in 1990. Thirty years later, the number reached to 19.3 million in 2020. Simultaneously, the number of corresponding cancer related deaths increased from 5.2 million in 1990 to 10 million in 2020 (1, 2). In 2040, the global cancer burden is expected to reach 28.4 million cancer cases, with a 47% increase from 2020 (1). The driving forces behind the curve are complex but reflect aging, social and economic development, and population growth, as well as the changes in the prevalence and distribution of main cancer risk factors, and social and economic development (12, 13).

Lung cancer has been one of the most frequently diagnosed cancers and the leading cause of human cancer death for all humans combined, worldwide (1, 2, 9-11). The incidence and death number of lung cancer for both sexes are still growing (Figure 1B). The incidences of lung cancer were estimated to be 1.04 million (12.8% of the world total) in 1990, 1.61 million (12.7%) in 2008, 1.82 million (13.0%) in 2012, 2.09 million (12.3%) in 2018, and 2.21 million (12.2%) in 2020. The lung cancer associated mortalities were 0.92 million (17.8% of all the cancer caused deaths) in 1990, 1.38 million (18.2%) in 2008, 1.59 million (19.4%) in 2012, 1.76 million (18.6%) in 2018, and 1.80 million (18.2%) in 2020, respectively (1, 2, 9-11). The numbers of new incidences and mortalities for lung cancer continue to increase, whereas the corresponding percentages in all cancer types have peaked and declined since 2012. In 2020, Lung cancer continues to be the leading cause of cancer death for all humans. For both sexes at the national level, lung cancer is the most frequently diagnosed cancer in 37 countries. However, with respect to mortality, lung cancer is the leading cause of cancer death in 90 countries (1). In contrast, the mortality and incidence of lung cancer in the U.S. are dramatically decreased in recent years due to the success of smoking cessation programs (Figure 1C).

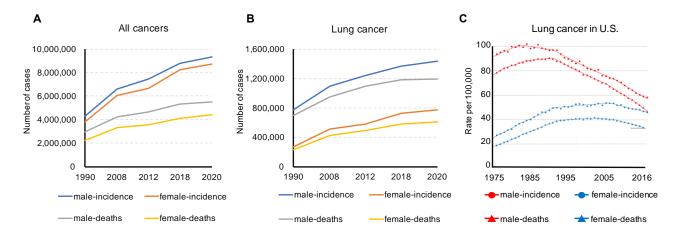


Figure 1. The epidemiology of cancer incidence and mortality from 1990 to 2020 illustrated the growing dynamics in male and female population worldwide. **(A)** The incidence and mortality of all surveyed types of cancer for both sexes are in the process of continuously growing worldwide. **(B)** The incidence and death number of lung cancer for both sexes are still growing. Data source: GLOBOCAN 1990~2020. **(C)** Cancer of the lung and bronchus SEER delay-adjusted incidence and U.S. death rates, 1975-2016, all races, by sex. Source: SEER 9 areas and U.S. mortality files (National Center for Health Statistics, CDC).

2.1. Lung cancer in males

It is estimated that CS accounted for 86% of lung cancer associated mortality in men (2). In 2017, it was estimated that cigarette smoking contributed to 1.19 million lung cancer

deaths (14). For males, lung cancer ranked as the leading cancer type both in terms of incidence and mortality in the extent of 30 years (1, 2, 9-11). In 1990, male lung cancer was 18.0% of the total male cancer cases and 23.4% of the total male cancer deaths (2). In 2020, the corresponding rates for lung cancer incidence and mortality decreased to 15.4% and 21.7%, respectively (Figure 2A~2B). The time trends of lung cancer incidence and mortality generally reflect the past exposure to cigarette smoking in male population and considerable regional variations (2).

For males, from the perspective of global cancer patterns in 2020, lung cancer is the most frequently diagnosed type of cancer in 36 countries, including Russia, China, Korea, Myanmar, Malaysia, Indonesia, Philippines, Kazakhstan, Iraq, Syria, Turkey, Jordan, Ukraine, Poland, Romania, Greece, Algeria, Libya and Morocco (1). In terms of cancer related mortality, lung cancer is the leading cause of cancer death in men in 93 countries (Figure 2C), half of all the 185 surveyed countries, including all the 36 countries with lung cancer as the most frequently diagnosed cancer, and also Canada, United States, Brazil, Bolivia, Argentina, Uruguay, Paraguay, Japan, Australia, New Zealand, Pakistan, Uzbekistan, Turkmenistan, South Africa, and nearly all the European countries except Sweden (1). For instance, the incidence rate in males from Eastern Europe, Eastern Asia, Western Europe, Southern Europe and Northern America is 49.3, 47.2, 43.3, 43.1 and 39.1 per 100,000 people, respectively (1). The highest incidence rates of lung cancer among both men and women were observed in Hungary. The rate in male was as high as 77.4 in 100, 000 in 2018 (11). In 2020, the leading lung cancer incidences in men occurred in Turkey (Western Asia) (1). If the cancer incidence and mortality patterns are viewed via the perspective of the 4-Tier Human Development Index (HDI), for incidences in men, lung cancer ranks first (39 per 100,000) in higher HDI countries, vice versa for lower HDI countries (10.3 per 100,000) (1). The mortality rate for lung cancer is 31.6 per 100,000 in higher HDI countries and 9.4 per 100,000 in lower HDI countries for men (1).

The large variation in regional lung cancer incidences across the world is attributable to the maturity of tobacco epidemic, varied patterns of smoking exposure, including intensity and duration of smoking, type of cigarettes consumed, and degree of inhalation (2). Based on 1990 estimates, the incidence of lung cancer in males in regions of Eastern Europe, North America, Northern Europe, Southern Europe, Western Europe and Australia were 75.85, 69.62, 59.12, 58.81, 54.1, and 47.55 per 100,000 population, respectively (2). In 2008, the incidence of lung cancer in these areas decreased to 57, 48.5, 39.3, 49, 44.7, and 32.4, respectively (9). In 2020, the incidence rates furtherly reduced to 49, 35.7, 33.3, 43.1, 41.7, and 28.1 per 100, 000 population, respectively (1). The motivation behind the decreasing male lung cancer incidence is largely attributed to the decreasing CS epidemic. The United States (US), Northern and Western Europe, and Australia have now passed the peak of the tobacco-related epidemic in the middle of last century, and incidence rates and death rates of lung cancer are declining (2). The decreasing CS prevalence, followed by a peak and decline in lung cancer rates in the same generation, was also observed in developed nations such as the UK, the US, Finland, Australia, New Zealand, the Netherlands, Singapore, and (more recently) Germany (15, 16). After analyzing the Nordic cancer registries of Denmark, Finland, Norway and Sweden from the NORDCAN database, it was found that the approaching of incidence rates for lung and bladder cancer can be expected in the course of the abating smoking epidemic (17).

On the contrary, in Southern and Eastern European countries, the incidence rates of lung cancer continues to increase, where smoking prevalence have not peaked (2, 18-20). Additionally, in many developing countries such as China and Indonesia, the epidemic of tobacco smoking has just been recently established and has either peaked or continues to grow. Therefore, lung cancer rates are likely to increase for the next several decades in these countries (21). Especially in China, the most populous country in the world, with more than 4.5 million new incidences of cancer and 3 million cancer deaths China also tops first in cancer burden worldwide. In which, active smoking is the leading cancer risk in men. It was found that, in 31 Chinese provinces, the highest population-attributable fraction among men was smoking (22). In total, Chinese men now smoke more than a

third of the world's total cigarettes. It is estimated that smoking caused about 20% of all adult male deaths in China during the 2010s. The tobacco-attributed proportion of mortality is still increasing. Although overall adult mortality rates are falling, as the adult population of China grows and the proportion of male deaths due to smoking increases, the annual number of deaths in China that are caused by tobacco will rise from about 1 million in 2010 to 2 million in 2030 and 3 million in 2050, unless there is widespread cessation of CS (23). The above epidemiological data shows that barring intervention measures to accelerate smoking cessation and reduce the number of new adopters are desperately required for the purpose of lung cancer prevention.

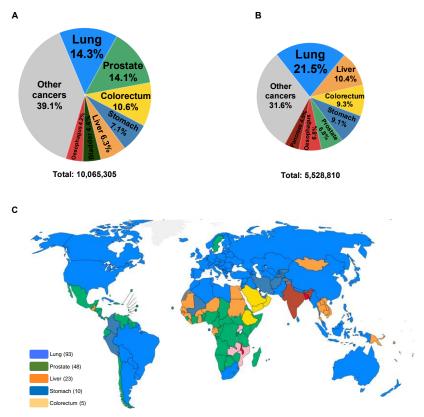


Figure 2. The proportions of top types of cancer in GLOBOCAN 2020. **(A)** shows the estimated percentage of new cancer cases in 2020, worldwide, males, all ages. **(B)** shows the estimated percentage of cancer deaths in 2020, worldwide, males, all ages. **(C)** Globally, the world map shows the top cancer per country, estimated number of deaths in 2020, only males, all ages. For mortality, female lung cancer as the top cancer in one country would be presented in blue color. Lung cancer was estimated as the leading cause of male cancer death in 93 countries showed in the world map. Data source: UNDP. World Health Organization. GLOBOCAN 2020.

2.2. Lung cancer in females

It is estimated that CS accounted for 49% of lung cancer associated mortality in women (2). For females, in the thirty years from 1990 to 2020, the percentage of lung cancer incidence in all cancers increased from 7.0% to 8.8%, and corresponding mortality percentage increased from 10.2% to 13.8% (1, 2, 9-11) (Figure 3A~3B). In 1990, lung cancer ranked fifth in incidence and third in mortality in all cancer types in females (2). The ranking increased to the fourth in incidence and second in mortality in 2008. Afterward, lung cancer consecutively ranked third in incidence and second in mortality in 2012, 2018 and 2020 (1, 10, 11). However, the most marked change is that, in 2020, female breast cancer has now surpassed lung cancer as the leading cause of global cancer incidence, with an estimated 2.3 million new cases, representing 11.7% of all cancer cases (1). Lung cancer incidence in females are of high variety among global regions in 2020. At the national level in females population, lung cancer was the most frequently diagnosed cancer only in

North Korea (1). However, lung cancer was the leading cause of cancer death for females in 25 countries, such as China, Korea, Australia, Canada, the United States, Norway, Sweden, Finland, Iceland, the United Kingdom, Ireland, Denmark, Netherlands, Belgium, Poland, Czech Republic, Hungary, and Slovenia (1) (Figure 3C). The highest incidence rate is in Northern America (30.7 per 100,000), followed by Northern Europe (26.9), Western Europe (25.7), Australia (24.0) and Eastern Asia (21.9) (1). From the perspective of HDI, lung cancer incidence ranks third (18.2 per 100,000) in all cancers in higher HDI countries, vice versa for lower HDI countries (4.2 per 100,000) (1). The corresponding mortality rate for female lung cancer in higher HDI countries is 13.7 per 100,000, and the rate in lower HDI countries is 3.8 per 100,000 (1).

The trends of lung cancer incidence in females are also of high variety, globally. For example, the corresponding female lung cancer incidence in North America, Northern Europe, Southern Europe were 32.91, 20.21, and 7.26 per 100,000 females, respectively in 1990 (2). In 2008, the incidences increased to 35.8, 21.8, and 10.4, respectively (9). In 2020, the rates were 30.1, 26.8, and 16.4 (1). Except Northern America with the continuously decreasing rate of lung cancer incidence from 2008, the rates from Northern Europe and Southern Europe both generally increased from 2008 to 2020 (1, 9-11). Actually, only a few female populations, such as the US and the UK, are showing signs of plateau and decline. In many other countries, the lung cancer incidence rates are increasing in women (24). The trend convergence in the men and women in several European countries is possibly resulted from the sex-specific differences in the distribution of histologic subtypes and smoking prevalence (25).

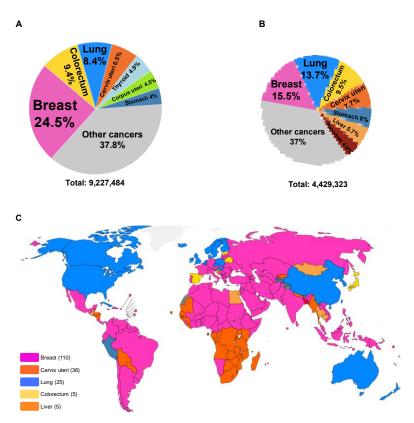


Figure 3. The proportions of top types of cancer in GLOBOCAN 2020. **(A)** shows the estimated percentage of new cancer cases in 2020, worldwide, females, all ages. **(B)** shows the estimated percentage of cancer deaths in 2020, worldwide, females, all ages. **(C)** Globally, the world map shows the top cancer per country, estimated number of deaths in 2020, only females, all ages. For mortality, female lung cancer as the top cancer in one country would be presented in blue color. Lung cancer was estimated as the leading cause of female cancer death in 36 countries showed in the world map. Data source: UNDP. World Health Organization. GLOBOCAN 2020.

2.3. Lung cancer prevention

It is generally agreed that the CS associated lung cancers could be largely prevented by reducing smoking initiation and increasing smoking cessation. A population-based case-control study in France demonstrated that lung cancer risk increased linearly with intensity and duration of tobacco smoking while it decreased with time since cessation, to reach the risk in never-smokers after 20 years of abstinence (26). However, even after diagnosis, smoking cessation could also reduce the risk for lung cancer progression and mortality among the current smokers with early-stage lung cancer (27). Consistently, in a study of 1,134 NSCLC patients with smoking history, the length of smoking period was a significant prognostic indicator for those who underwent curative lung resection (28).

Most lung cancers are preventable by applying proven tobacco control interventions (1). The U.S. CDC launched Comprehensive Tobacco Control Programs in Best-practice, including effectively reducing active smoking, banning smoking in all indoor areas of workplaces and public places to prevent involuntary exposure to secondhand smoke, raising the retail price and taxes rate for tobacco products, restricting tobacco advertising and promotion, especially banning tobacco sales to young people, and enforcing plain packaging and graphic illustration of health warning on the tobacco products (1). In 2003, WHO established the Framework Convention on Tobacco Control to enable international coordinated efforts to curb the tobacco smoking epidemic, and 168 signatories have ratified the agreement after its adoption (29).

Alarmingly, tobacco use among youth is rapidly increasing in many lower-HDI countries. For young smokers, one strategy for tobacco control is printing effective warning images on cigarette products. Which could directly arouse the consciousness of smoking related health risks (Figure 4). Recently, an investigation of 353 Thailand undergraduates found that warning images illustrating patients' sufferings from smoking related cancer and body damages, could generate a significantly higher level of fear than other images in these adolescents. In addition, non-smokers were more sensitive to scary warning images (30).

In U.S., the comprehensive tobacco control programs in many states, such as California and New York, have markedly decreased smoking rates and accelerated the reduction in lung cancer occurrence (31, 32). Although the total number of cigarettes smoked worldwide is decreasing, Asia is still the largest tobacco consuming area, with China and India as the most populous countries in the earlier stage of tobacco epidemic (33). If smoking cessation and initiation prevention could be achieved by immediate actions, the lung cancer incidence and corresponding extraordinary burden experienced in developed countries could be largely attenuated.

The 5-year survival rate for patients with lung cancer in 2018 and 2020 were 18% and 19%, respectively (34, 35). However, the survival rate for lung cancer could be largely improved by early diagnosis. For the purpose of which, there is an increasing realization that most patients who are diagnosed with lung cancer as a result of annual CT lung screening (CTLS) could be cured, and heavy cigarette smokers without screening have a greater risk of lung cancer mortality (36). Consistently, a study with a minimum 10 years' follow-up, demonstrated that lung cancer mortality was significantly lower among those who underwent CTLS than among those without screening (37). Although there are uncertainties about over-diagnosis via CTLS, the meta-analysis suggested that the benefits of CTLS implementation outweigh potential harms, in subjects with CS history, ultimately supporting the systematic implementation of CTLS worldwide (38).

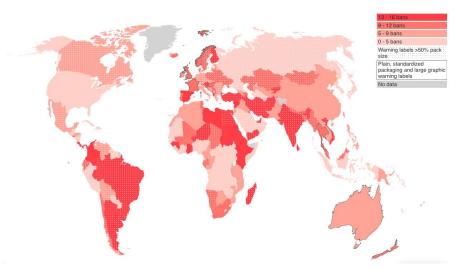


Figure 4. The world map shows the total number of bans on direct and indirect tobacco advertising, 2018. Data source: WHO GTCR 2019.

2.4. Challenges from new forms of cigarette

Recently, new forms of cigarette containing electric devices are raising concerns for potential public health risks. For instance, the heated tobacco products (HTP), have been developed as a less harmful alternative to traditional cigarettes. Although HTP products may impose a reduced risk of respiratory cancer compared to traditional smoking, the actual short-term and long-term impact of HTP on public health is still not fully undertood. It was found that HTP exposure such as I Quit Ordinary Smoking (IQOS) could alter mitochondrial functions and further lead to many adverse health consequences (39).

The prevalence of e-cigarettes has been increasing rapidly, particularly among the adolescents and non-smokers (40). However, the e-cigarette is greatly different from traditional cigarette, same irritant agents and carcinogens could also be found within (41). It was verified that electronic-cigarette smoke (ECS) is capable of inducing lung adenocarcinoma in mice, probably through causing DNA damage and inhibiting DNA repair (42). Similar pathological changes, as well as the mutagenic DNA adducts, were also evidenced in nicotine treated human lung cells (43).

Additionally, given the lag time of several decades extrapolated from tobacco smoking data, the long-term effect of these new forms of cigarette on public health should be concerning. Thus, comprehensive policies and regulations to assess, supervise, and monitor should be closely followed.

3. Mechanisms of CS associated lung cancer

The mechanism underlying the major CS carcinogens associated lung cancer are reviewed and illustrated (Figure 5).

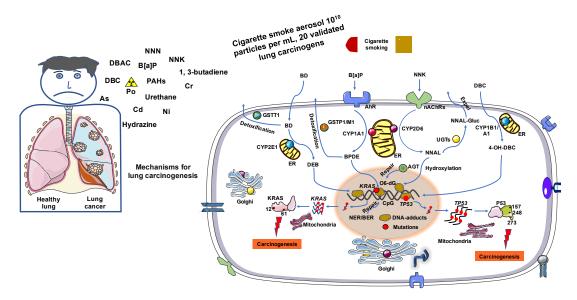


Figure 5. Mechanisms underlying cigarette smoking associated lung cancer. The above graph shows the representatives of major types of components in cigarette, and the mechanisms underlying cigarette smoking associated lung carcinogenesis were integratively illustrated on the cellular level.

3.1. Significant carcinogens in CS

The IARC reported that cigarette smoking could increase the lung cancer risks in all histological types, including adenocarcinoma, squamous-cell carcinoma, small-cell carcinoma, and large-cell carcinoma. In 1826, nicotine was isolated from tobacco for the first time and the investigation of mechanisms underlying cigarette smoking associated lung cancer began one century ago. Tobacco smoke contains a complex mixture of chemicals, with more than 70 validated carcinogens to humans (44). Twenty carcinogens have been demonstrated to convincingly cause lung cancer in laboratory animals (Table 1). Particularly, Nitrosamines (NNK and NNN), polycyclic aromatic hydrocarbons (benzo[a]pyrene) as significant carcinogens are believed to be two major aetiological factors for lung cancer.

Table 1. Lung cancers were induced in experimental animals by the carcinogens presented in CS.

Classifications	Carcinogens in CS	Lung tumorigenicity in rodents
Polycyclic aromatic hydrocarbons	Benzo[a]pyrene	Hamster(45)_1974; Hamster(46)_1978; Rat(47), (48)_1983; Mouse(49)_1995; Mouse(50)_1995;
	Benzo $[b]$ fluoranthene	Rat(47)_1983; Mouse(49)_1995; Mouse(50)_1995; Mouse(51) _1996;
	Benzo[j]fluoranthene	Rat(47)_1983; Mouse(52)_1994;
	Benzo[k]fluoranthene	Rat(47)_1983;
	Dibenzo[a,i]pyrene	Hamster(53)_1974;
	Indeno[1,2,3-cd]pyrene	Rat(47)_1983;
	Dibenz[a,h]anthracene	Mouse(54)_1966; Rat(47)_1983; Mouse(49)_1995; Mouse(50) _1995;
	5-Methylchrysene	Mouse(49)_1995; Mouse(50)_1995;
	Cyclopenta[c,d]pyrene	Mouse(55)_1994; Mouse(49)_1995; Mouse(50)_1995;
Heterocyclics	Dibenz $[a,h]$ acridine	Rat(48)_1983;
	7H-Dibenzo[c,g]carbazole	Hamster(56)_1977; Mouse(57)_1996;
N-Nitrosamines	N-Nitrosodiethylamine	Hamster(58)_1961; Mouse(59)_1992; Mouse(60)_2018;
	NNK	Mouse(61)_1978; Rat(62)_1987; Rat(63)_1997;
Other organics	1,3-Butadiene	Mouse(64)_1985; Mouse(65)_1989; Mouse(66)_1993;
	Urethane(Ethyl carbamate)	Rat(67)_1947; Mouse(68)_2007; Mouse(69)_2015;
Inorganics	Nickel	Rat(70)_1975; Mouse(71) _1984;
	Chromium	Rat(72-74)_1986; Rat(75)_2005;
	Cadmium	Rat(76)_1986; Mouse(77)_1994;
	Polonium-210	Rat(78)_1967; Hamster(45)_1974; Hamster(46)_1978;
	Arsenic	Rat(79)_1977; Human(80)_1980;
	Hydrazine	Mouse(81)_1965; Mouse(82)_1969; Mouse(83)_1976;

3.1.1. Polycyclic aromatic hydrocarbons

A variety of polycyclic aromatic hydrocarbons (PAH), including benzo[a]pyrene, benzo[b,j,k]fluoranthane, dibenzo[a,i]pyrene, indeno[1,2,3-cd]pyrene, dibenz[a,h]anthracene and 5-methylchrysene, could cause lung cancer $in\ vivo\ (84)$. B[a]P was the first and the major carcinogenic precursors detected in CS (85). Millions of people are suffering lung cancer risks of being exposed to B[a]P via CS or consuming B[a]P-contaminated food. It was demonstrated in mice model that the carcinogenic capability of dibenz[a,h]anthracene and 5-methylchrysene with low percentage in CS tended to be stronger than B[a]P (50). However, the carcinogenic mechanisms underlying dibenz[a,h]anthracene may be different from other PAHs, for the frequent Kras mutations induced by PAHs were not shown in dibenz[a,h]anthracene (50). CS was found to enhance the metabolic activation of phenanthrene in humans, which is a representative of PAH (86).

3.1.2. Nitrosamines

4-(methylnitrosamine)-1-(3-pyridyl)-1-butanone (NNK), N'-nitrosonornicotine (NNN), N'-nitrosoanatabine (NAT) and N'-nitrosoanabasine (NAB) belong to a group of

chemicals called tobacco-specific nitrosamines (TSNA), which are found only in tobacco products. Epidemiologically, TSNA concentrations were shown to be higher in every day tobacco users than in intermittent users (87). The IARC has classified NNN and NNK as Group I carcinogen to humans. NNK, a potent carcinogen, can induce lung adenomas in female A/J mice (88), liver cancer and pancreatic cancer in male F344 rats (89). Activated Akt was evidenced in mice lung tumors induced by NNK, probably through the binding of nicotine or NNK with nicotine-acetylcholine receptors (nAChRs) (90). The CSC or NNK exposure was able to neoplatically transform BEAS-2B cells into invasive adenocarcinoma in vitro, which exhibited typical malignant features, such as increased invasiveness, resembling the progressive changes in human lung cancer (91). In mice, lung cancer could be induced by NNK in CS, and lung tumors with more malignant features appeared after combining with LPS (inflammation inducing) compared to NNK treatment alone (92). Interactions between specific chemicals in tobacco smoke were investigated; mice receiving both NNK and aldehydes had more adenomas than only NNK treated mice, which suggested that coexposure to inhaled aldehydes is capable to enhance the carcinogenic effects of NNK (93).

3.1.3. Heterocyclic organics

Heterocyclic organic compounds of lung carcinogenicity presented in tobacco smoke include Dibenz[a,h]acridine (DBAC) and 7H-dibenzo[c,g]carbazole (DBC)(48, 56, 57). In male CD rats treated with DBAC, the resulted genotoxicity was found in the form of DNA adducts using 32P-postlabeling analysis(94). DBC as a skin, liver, and lung carcinogen has both local and systemic effects in mice(95). The DBC-DNA adducts formed from DBC metabolites, including 4-OH-DBC, as the most potent are mostly catalyzed by lung CYP1B1 and CYP1A1(96). Stable DBC-DNA adducts were identified in mouse lung, including two newly discovered DBC-6-N7-Ade and DBC-6-N1-Ade, following a single i.p. of DBC(97). Consistently, seven DBC-DNA adducts were identified in the lung of A/J mice after DBC treatment via i.p. Additionally, K-ras mutations from the induced lung tumor predominantly occurred at codon 61(A-T transversion)(57).

3.1.4. Other carcinogenic organics

Of the many tobacco smoke constituents, 1,3-butadiene (BD) has a high cancer risk index due to its tumorigenic potency and its abundance in cigarette smoke(98, 99). The carcinogenicity of BD has been attributed to the formation of several epoxide metabolites, of which 1,2,3,4-diepoxybutane (DEB) is the most toxic and mutagenic(100). Recently, it is found that the status of GSTT1 genotype significantly influences BD metabolism and acute toxicity (101). Ethyl carbamate also known as urethane is presented in cigarette smoke (102), potent carcinogenicity of urethane to induce lung tumors was demonstrated in mouse and rat, and the process of lung carcinogenesis was partially promoted by the activation of NF-κB (68) (69). NF-κB as a critical effector of inflammatory responses is integral to lung tumorigenesis in urethane induced lung cancer mouse model (68).

3.1.5. Heavy metals

Tobacco products also contain carcinogenic metals in relative large quantities (85). It was shown that the abundance of heavy metal levels in smokers are significantly higher than that in non-smokers, including thallium (Ti), arsenic (As), manganese (Mn), and copper (Cu). Specifically, lead (Pb), cadmium (Cd), and cobalt (Co) are among the most important metals accumulated in smokers' blood (103). Upon nickel exposure, the nickel concentrations in lungs were significantly increased, and no apparent deposition was observed in other tested organs using a rat model (104). It was consistently demonstrated that nickel and cadmium could generate single-strand breaks in rat lung when administered alone (105). Interestingly in NFS mice, lung cancer was induced by cadmium only at a lower dose(77). Malignant and premalignant bronchial lesions in rats were induced in 9 months after a single strontium chromate treatment(75). The chromium-induced

DNA damage and mutations observed in the lung of rodents has been correlated with the potential lung carcinogenicity in humans (106, 107). Arsenic in cigarettes was preliminarily reported in as early as 1896 (108). Arsenic exposure before pregnancy could increase the lung adenoma and bronchiolo-alveolar tumor incidence in the offspring (109). Since arsenic and B[a]P are lung cancer carcinogens both in CS and environment, it was speculated that arsenic and B[a]P combined-exposure have synergistic effect in the increased lung cancer risk observed in arsenic-exposed cigarette smokers (110).

3.2. The metabolic activation and detoxification of tobacco carcinogens

Cytochrome P450s and glutathione-S-transferases (GST) are two major type of enzymes for the metabolic activation and detoxification of the tobacco carcinogens (111, 112). Most carcinogens in cigarette smoke are metabolically activated by cytochrome P450. Subsequent formation of macromolecular adducts are responsible for carcinogenicity (111). Some low molecular weight chemicals such as ethylene oxide, formaldehyde, and acetaldehyde can reactively bind with DNA molecules. The CYP1A1, CYP2D6, and CYP2E1 in the cytochrome P450 gene family are significant enzymes for tobacco carcinogen metabolism. CYP2D6 can metabolically activate NNK (113). CYP2E1 actively metabolizes 1,2-butadiene, N-nitrosodimethylamine, and N-nitrosodiethylamine (114, 115). CS could induce the expression of CYP1A1 which is also known as aryl hydrocarbon hydroxylase (AHH). The catalysis of PAH (B[a]P) into carcinogenic metabolites is also processed by CYP1A1 (116). Further, more CYP1A1 activities promoted by B[a]P is partially attributed to the activation of AhR/Src/ERK axis (117). Consistently, It has been reported that a flavonoid pinocembrin (PCB) is to alleviate B[a]P toxicity via inhibiting DNA adduct formation by attenuating CYP1A1 expression through the suppression of the AhR/Src/ERK pathways (118). PAH detoxification is mainly catalyzed by GSTM1 and GSTP1(119). The detoxification of nitrosamines, such as NNK, is majorly govern by UGTs. First, NNK is metabolized into carcinogenic NNAL. Next, NNAL is glucuronidated and NNAL-Gluc is produced as the detoxified form by UGT. Recently, it was found that variations of the activity of specific UGTs may affect the efficiency of NNAL detoxification (120).

3.3. Genomic aberrances caused by carcinogens and DNA adducts

DNA adducts, formed through covalently binding carcinogen metabolites to human DNA, usually at guanine or adenine, are crucial to the lung carcinogenic process (85). The investigation of "DNA adducts" via a 32P-DNA post-labeling approach was used to elucidate the association between CS carcinogens and lung cancer risk (121). PAHs like B[a]P are metabolically activated and converted to 7,8-diol-9,10-epoxides (BPDE), subsequently forming the BPDE-deoxyguanosine DNA adducts or protein adducts with albumin (122). NNAL is the predominant metabolite of NNK, which are hydroxylated to form the DNA adducts such as methyl adducts (7-methylguanine, O6-methylguanine), and pyridyloxobutyl adducts. The level of these DNA adducts are increased in lung cancer patients compared to in normal cases and are likely to cause lung tumors in rodents (123, 124). NNN could be metabolically activated to produce carcinogenic metabolites via 2'-hydroxylation and 5'-hydroxylation, which could directly alkylate DNA to form adducts (125, 126). 4hydroxy-1-(3-pyridyl)-1-butanone (HPB)-releasing DNA adducts derived from NNK and NNN, are significantly increased in smokers (127, 128). In addition, new DNA adducts from NNN are continuously being discovered (129, 130). Programmed cell death or apoptosis may occur in cells with DNA damage. However, DNA miscoding and genetic mutations would be produced. If these DNA adducts were not efficiently repaired. If a permanent mutation occurs in specific sites of an oncogene or a tumor suppressor gene, and multiple of these events may lead to aberrant genotypes and uncontrolled proliferation, ultimately, to lung cancer.

DNA adducts resulting from PAHs and NNK can directly cause genetic mutations in significant oncogenes and tumor suppressor genes, especially the frequent mutations of G-T and G-A in *KRAS* and *TP53* (131-133). *TP53* mutations with an excess of G-T

transversions have been observed in smoking-associated lung cancers. These G-T transversion hotspots are the sites preferring the formation of PAH adducts and also the position of endogenously methylated CpG dinucleotides (133). BPDE preferentially forms DNA adducts and induces G-T transversion in codon 157, 248, and 273, which are three mutational hot spots in *TP53* (134). In 550 lung tumor samples with *TP53* mutations, 33% of which were classified as G-T transversions, and 26% were G-A transitions, especially at the CpG sites of the deamination of 5-methylcytosine, where the cytosine methylation further enhances guanine alkylation by a variety of carcinogens (135). Collectively, these data suggested that the BPDE-DNA adducts could directly cause mutations in *TP53* (136). G-A transitions induced by the DNA adduct O6-POB-dG in *TP53*, derived from NNK or NNN, could be repaired by O6-methylguanine–DNA alkyltransferase (AGT) (137).

Mutations in codon12 of *KRAS* are prevalent in lung adenocarcinomas and are more common in smokers than in never smokers, indicating that this mutation pattern may be causatively associated with carcinogens in CS (138, 139). Consistently, the same mutation pattern of *Kras* was induced by treating mice with PAHs (140). Particularly, the GGT-GAT mutation predominant in Kras codon12 was shown to be caused by NNK metabolite-DNA adducts (O⁶-methylguanine) (141). CDKN2A, as a tumor-suppressor gene, was hypermethylated in the promoter area was found in 23% lung cancer cases (142). Consistently, the hypermethylated promoter region of Cdkn2a was identified in almost all of the adenocarcinomas induced by NNK, and even detected in precancerous lesions. These genetic lesions were also recapitulated in human SCCs, indicating the causal associations between the silenced Cdkn2a and the carcinogenicity of NNK(143). The mutational inactivation in FHIT identified in lung tumors were found to be associated with the duration of CS (144). These data suggested that carcinogens in CS could induce genetic and epigenetic alterations in tumor-suppressor genes, and make them incapable of efficiently suppressing lung carcinogenesis.

However, these DNA adducts resulting from cigarette smoke could be repaired by host repair mechanisms, such as base excision repair (BER) for oxidized bases and DNA alkylation, and nucleotide excision repair (NER) for PAH resulted DNA lesions (145, 146). In addition to the NER pathway for O⁶–alkylguanines DNA adducts derived from PAH, NNK, N-nitrosodimethylamine, and N-nitrosodiethylamine, the alkyl groups could also be removed from the O⁶-deoxyguanosine by AGT, restored to deoxyguanosine (147, 148).

Moreover, recent studies showed that CS carcinogens not only cause point mutations but also lead to genome instabilities. A large genome-wide study of NSCLC demonstrated that more copy number alterations (CNAs) occurred on different genomic scales in heavy smokers other than in light or non-smokers. The higher level of CNAs may have resulted from CS caused genome instability, which was demonstrated in CSC treated human bronchial epithelial cell (149).

3.4. CS biomarkers for evaluation of lung cancer risks

The DNA adducts, protein adducts, and urinary metabolites produced in the metabolic processing of tobacco carcinogens provide biomarkers for evaluation of lung cancer risks from cigarette smoking. These metabolic products also reflect the carcinogen uptake, metabolic activation and detoxification in people exposed to cigarette smoke (85, 150). For example, the level of DNA-adduct in white blood cells was sensitive in predicting the risks of lung cancer for smokers (151). Cotinine, t,t-Muconic acid, 1-hydroxypyrene, and NNAL and NNAL-Glucuronides (NNAL-Gluc, detoxified metabolite of NNK) are common urinary carcinogen biomarkers for nicotine, benzene, PAH, and NNK, respectively, which are measured for evaluation of environmental tobacco smoke (ETS) exposure (150).

Given the fact that only partial smokers would finally develop lung cancer, which may attributable to the substantial differences in the metabolic response to tobacco carcinogens, with differed capability of catalyze carcinogens into DNA adducts, so these biomarkers could potentially be used for discriminating smokers who can efficiently activate or detoxify tobacco carcinogens. For instance, the ratio of NNAL-Gluc to NNAL was

significantly higher in white smokers than in black smokers, which partially explains the higher lung cancer incidences in black smokers (152). Interestingly, in the normal lung tissue from 63 lung cancer patients, the levels of DNA adducts in females were higher than that in males after adjusted for smoking doses, indicating that women are likely at higher risks for CS associated lung cancer (153).

The Golestan Cohort Study showed that current male smokers with lung cancer had higher average levels of total nicotine equivalents (TNE-2), NNAL, and 3-FLU. On the other hand, lung cancer risk increased with concentrations of TNE-2 and NNN (154). Consistently, the urinary NNAL level was found to be associated with the risk of lung cancer in general population and independent from CS, indicating the experience of potential ETS exposure (155). Collectively, these data indicated that smokers or involuntary smokers, capable of activating NNK or NNN and forming DNA adducts, are likely at higher lung cancer risks.

3.5. Oxidative damage caused by CS

The oxidative damage (OD) induced by CS may be attributed to the reactive oxygen species (ROS, such as nitric oxide, NO) in the gas phase (156), and free radicals (such as the quinones) in particulate matter (157-159). Peroxynitrite, generated from nitric oxide and superoxide anion, causes the DNA single-strand breaks (160). ROS also causes increased DNA adducts such as 8-oxodeoxyguanosine (161). In lung cancer cell lines, CS induces oxidative stress and mitochondrial damage, which further increases glycolytic flux, downregulates FOXO3a, and facilitate EMT and cell migration (162). Additionally, the CS caused OD increases circulating F2-isoprostanes via arachidonic acid peroxidation, which could be improved by supplementation of ascorbic acid (163). In human bronchial epithelial cells, the OD and inflammatory responses induced by CSE could be alleviated by eucalyptol and curcumin (164). CSC could activate the Ca²⁺/PKC signaling in vitro and increase the metastatic nodules in the lung of mice, which could be alleviated via inhibition of Ca²⁺/PKC or decreasing ROS production (165). Extensive hypoxia resulted from CS can trigger endoplasmic reticulum stress (ERS). Subsequently, the unfolded protein response (UPR) caused by ERS would activate cell death pathways and promote lung carcinogenesis (166).

3.6. CS associated inflammation and immunosuppression in lung cancer

The activated NF-κB pathway and increased expression of inflammatory factors occur in response to CS. Therefore, pulmonary inflammatory responses can be induced via AhR activation by CS ligands. However, smoke-induced inflammation can be attenuated by endogenous AhR ligand (167). NF-κB, a critical effector of inflammatory responses, is integral to lung tumorigenesis in the urethane induced lung cancer mouse model (68). DAPK2, as a tumor suppressor gene, can be methylated at N6-adenosine via CS activated NF-kB pathway, which significantly associated with poor prognosis in NSCLC patients, especially in smokers (168). In airway epithelium and lung cancer cells, CS activated MARCKS promotes the expression of pro-inflammatory cytokines, EMT, and stem-like properties. Additionally, CS activated NF-kB signaling pathways can be suppressed via the inhibition of MARCKS phosphorylation (169). Increased levels of IL-17 were accompanied with DNA damage response (DDR) in human lung tissues and in mice after CS exposure, indicating that IL-17 mediated inflammation may promote CS-induced genomic instability. Moreover, IL-17 application can increase DDR and chromosome breakage in human bronchial epithelium cells, which was diminished in IL-17 KO mice after CS exposure (170).

CS causes an influx of inflammatory cells, including Langerhans cells, into the lung tissue and induces extensive pathological changes, such as emphysema and fibrosis. Additionally, this situation can be deteriorated upon increased infiltration of Langerhans cells (171). In mice, alveolar damages were observed, including alveoli enlargement, collagen deposition, mucus production, and higher AIM2 expression in dendric cells and

macrophages recruited in lung after CS exposure (172). A recent study reported that ACE2 was significantly overexpressed in the bronchial and alveolar epithelial cells in smokers, independent of age or gender, indicating that CS related ACE2 overexpression may assist the SARS-CoV-2 infection, and smoking cessation may potentially attenuate the risk of COVID-19 transmission (173). An *in vitro* air-liquid-interface (ALI) human airway tissue model was successfully developed to assess the effects of CS on the function and phenotype of airway epithelial cells in smokers, including xenobiotic metabolism, oxidative stress, and inflammatory responses (174).

The immunological activities of CD8+ T cells against tumor cells could be attenuated by nicotine in CS (175). Furthermore, the density of tumor associated neutrophils increased with lung cancer progression, particularly in NSCLC smokers (176). In mice lung cancer induced by B[a]P, extensive immunosuppressive changes and more cancer stem like cells were observed, including increased expression of TGF beta, CTLA-4, PD-L1, FOXP3 and decreased IL-12, together with increased CD166+ cells and decreased CD83+, CD8+ cells (177). Additionally, a recent study found that B[a]P could induce PD-L1 expression on lung epithelial cells both *in vitro* and *in vivo*, which is mediated by AhR, the B[a]P receptor. Furthermore, AhR inhibitors can synergize with anti-PD-L1 antibody, showing potent anti-tumor effect (178). In NSCLC patients treated with nivolumab and pembrolizumab(anti-PD-1 antibody), longer PFS did not favor never smoking patients, probably due to the low level of mutation heterogeneity (179). Moreover, the response rate of pembrolizumab was nearly two times higher in current or former smokers than in never smokers with NSCLC (180). These findings partially explain why lung cancer patients with a history of smoking respond better to pembrolizumab than non-smoker.

3.7. miRNA in CS associated lung cancer

It was discovered that the levels of miR-532-5p, miR-25-3p, and miR-133a-3p were significantly higher in adenocarcinoma patients than in healthy participants. The miR-133a-3p was independently associated with CS, which was also associated with pulmonary inflammation (181). The immunosuppressive effect of miRNA was observed after CS exposure; upregulated miR-629-5p partially contributed to the exhaustion of CD8+ T cells after nicotine exposure (175). Additionally, the anti-tumor effects of certain miRNAs could be impaired by CS. For example, miR-584-5p, capable of suppressing cancer migration and invasion, was methylated and downregulated after CS application in human bronchial epithelial cells (182).

3.8. Microbiota in smokers

The microbe and lung respiratory host cells form a sophisticated and balanced ecosystem, which can negatively be impacted by CS. In a study of 103 bronchoalveolar lavage fluid samples from lung cancer, the local microbiota was sequenced. The results indicated that lung cancer microbiota possessed higher diversity in squamous cell carcinoma than in adenocarcinoma and was enriched with Proteobacteria (183). The change of microbial diversity was also illustrated in a general population study of 529 Australian adults, which found that CS was associated with diversity loss of the airway bacterial communities, negative effects on abundant taxa, profound alterations to network structure and expansion of Streptococcus spp (184). Particularly for lung squamous cell carcinoma, increased abundance of unique bacterial consortium, such as Acidovorax, was evidenced in CS associated tumors (185). These marked abnormal imbalanced relationships between microbiome and host side in smokers may contribute to lung carcinogenesis.

3.9. Radon

In the general population, environmental exposure to radon is recognized as the second leading cause for lung carcinogenesis. Recently, in a model proposed by the International Commission on Radiological Protection, the radon exposure was found to have a positive correlation with the excess of lung cancer cases (186). From the perspective of CS

status, radon is the leading risk factor for lung cancer in never-smokers (LCINS), especially in radon-prone areas. In these cases, men were at higher risk of lung cancer than women (187). However, an investigation on the interaction between radon and CS indicated that smoking may act as a promoter of radon-initiated cells, and exposure to radon followed by smoking produced a significantly more-than-multiplicative effect (188). CS associated lung carcinogenesis is partly attributed to the cumulative alpha-radiation dose at bronchial bifurcations from indoor radon progeny and radionuclide attached smoke particles produced by tobacco combustion (189). Fractionation in burning cigarettes gives rise to the association of radon progeny with micron particles in mainstream CS, which are selectively deposited in "hot spots" at bifurcations, enriched radon progeny undergo substantial radioactive decay at bifurcations before clearance. Moreover, progressive chemical and radiation damage to the bifurcation epithelium gives rise to prolonged retention of insoluble ²¹⁰Pb-enriched radioactive smoke particles. It was estimated that a carcinogenic alpha-radiation dose of 80-100 rads is delivered to approximately equal to 107 cells of most smokers who die of lung cancer (190).

4. Indoor air pollution, outdoor air pollution

4.1. Indoor air pollution

Indoor air pollution resulting from poor heating ventilation facilities equipped for heating and cooking are main risk factors for Chinese household women. In 2012, lung cancer rates in Chinese women were approximately 20.4 cases per 100,000 (10). In 2018, the incidence of lung cancer in Chinese women increased to 22.8 per 100,000, which is equal to or even higher than the rates among women in some European countries, whereas the smoking prevalence is of substantial differences between the two regions (11). One explanation accounting for this is that the high incidence of lung cancer in Chinese women possibly reflects severe exposure to indoor air pollution directly emitted from the combustion of biomass fuels via unventilated coal-fueled stoves and from cooking oil fumes (19, 191-193). These household pollutants caused by coal burning and biomass fuels have been classified as Group 1 and Group 2A carcinogen for lung cancer by IARC.

4.2. Outdoor air pollution

4.2.1. Particulate matter (PM) in air pollutions

A majority of air pollution is caused by emissions from industry, power generation, transportation, and domestic burning. Together, these sources increase the air particulate matter (PM), which has been shown to have a causal link with lung cancer incidence and mortality (7). In 2013, IARC classified PM and other outdoor air pollutants as Group 1 carcinogens for lung cancer (6). The PM pollution can be evaluated and measured in mass concentration as ug/m3. 10 ug/m3 is the world health-based air-quality guideline from WHO, while the global population-weighted mean annual average PM2.5 was 46 ug/m3 in 2017, almost 5 times higher than the safe value. In India, PM_{2.5} reached as high as 91 ug/m³ in 2017. Particles with diameter more than 10 um would largely be retained at the respiratory barriers in the nose and throat. The PM_{2.5-10} are coarse fraction particles, which are capable of travelling only short distance and then precipitate to the earth. Astonishingly, because of the small size, large amount, and higher surface area-to-mass ratio (194), PM2.5 can travel as far as several thousand kilometers and remain suspended in the air for weeks (195). These properties of PM_{2.5} particles and ultrafine particles (UFPs) greatly increase exposure for humans, and penetrate deeper into the lung tissue (196). After the inhalation of air pollutants, these retained particles can produce long-term effects of inflammation and oxidative stress on both the local and systemic levels (197).

4.2.2. PM associated with lung cancer

Recently, ambient environment pollution has been determined to cause lung cancer (198). In an analysis of 80, 285 participants, PM_{2.5} was shown to have a significant adverse association with total lung cancer incidence (199). The ambient PM_{2.5}, PM₁₀ and NO₂ were

associated with poorer lung cancer survival, particularly in 352,053 lung cancer patients of early stage (200). Moreover, in the ACS CPS-II study, long-term PM_{2.5} exposure was demonstrated to be associated with increased lung cancer mortality in never-smokers, with the exclusion of the potential confounder of cigarette smoking (201). More than half of the lung cancer deaths attributable to ambient fine particles were projected to have been in China and other East Asian countries (8), which include the most polluted regions (Figure 6). Generally, there were combined effects among lung disease history, environmental exposures, and family history toward susceptibility to lung cancer in Chinese non-smokers. Non-smokers who had a family history of lung cancer were at higher risk of lung cancer than non-smokers who did not have lung disease history. Non-smokers with family cancer history may obtain benefits from active treatment of lung disease and removal of environmental exposures (202).

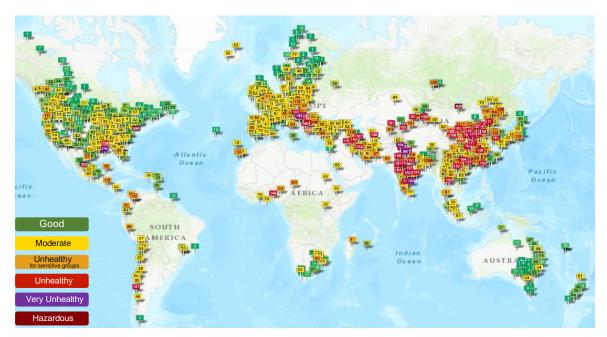


Figure 6. The world's real-time air pollution distribution in term of PM_{2.5} levels on Oct 20, 2021. The air quality index (AQI) is classified into six levels to represent the severity of air pollution, including Good (AQI 0-50), Moderate (AQI 51-100), Unhealthy for Sensitive Groups (AQI 101-150), Unhealthy (AQI 151-200), Very Unhealthy (AQI 201-300) and Hazardous (AQI 300+). The caution statement for "Hazardous" is that everyone should avoid all outdoor exertion. The AQI scale used for indexing the real-time pollution in the map is based on the latest US EPA standard, using the Instant Cast reporting formula.

PM pollution was reported to be associated with many types of lung cancer. The heightened risk of adenocarcinoma over the last four decades is considered as reflecting changes in cigarettes and the delivery of carcinogens. A meta-analysis carried out in North America and Europe reported that PM_{2.5} and PM₁₀ were both found to have a strong adverse association with lung adenocarcinoma. Particularly, the risk for lung cancer incidence or mortality rate increased 8% and 9% per each 10ug/m³ increase in PM2.5 and PM10 concentration, respectively (198, 203). In the participants who reported spending more than one hour per day outdoors, an adverse association of exposure to outdoor pollutions with adenocarcinoma was implicated (204). A significant adverse association of PM2.5 with small cell carcinoma was reported in the Canadian Breast Screening Study with 89,234 women participants (205). Adverse association of ambient PM10 concentration and squamous cell lung carcinoma incidence were concluded after analyzing 4,219 participants (206). Contradictorily, a non-significant association of PM₁₀ concentrations with lung cancer was reported in a large-scale study of 6.5 million participants conducted in Korea, whereas an adverse association of PM10 with lung adenocarcinoma in male smokers was identified in this study (207).

4.2.3. Specific carcinogens in PM

Studies also indicated the associations between lung cancer and the exact carcinogens in PM_{2.5} pollutions. The association of PM_{2.5} pollutants emitted from coal combustion with lung cancer mortality was found, especially Se and S (208). Glutathione-related rather than the ascorbate-related PM_{2.5} was found to be significantly associated with lung cancer mortality (209). In the ESCAPE study of 245, 782 participants, relative risks of lung cancer increase with the elemental components in PM_{2.5} and PM₁₀, such as S and Ni (210). Combustion of fossil fuels, such as coal, account for around 25% of total As and Hg emissions (211, 212), which have close association with several types of cancer including lung cancer. The IARC reported that the PM constituents such as the diesel engine exhaust, nickel, Chromium, Cd and silica dust are potential carcinogens for lung cancer (213). PAHs (214), dioxins (215), sulfur-derivatives (216), and 3-nitrobenzanthrone (217) commonly found in air pollutions are well-defined mutagens and carcinogens. Mutagenic PAHs can interact and bind DNA molecules (218), and the following repair process of these DNA adducts could potentially introduce mutations in the affected DNA sequences (219).

4.2.4. Genetic aberrances associated with PM

Substantial findings have illustrated that the number of mutations in air pollution associated lung cancer were three times higher than those in lung cancer patients from regions with cleaner air (220). Alarmingly, genomic mutations resulting from air borne carcinogens are inheritable. An in vivo mouse study found that higher level of germline mutations could be inherited after being exposed to industrial air pollutants (221). Additionally, ambient air pollution can cause epigenetic alterations, such as the aberrant status of DNA methylation (222-225), which could result in TP53 silencing and chromosome instability (226, 227). TP53 is a significant tumor suppressor gene, its abnormal mutation expression status are closely related with lung carcinogenesis (228). Low-dose PM2.5 is sufficient to induce epigenetic silencing of TP53 in human alveolar epithelial cells (229). Human epithelial cells exposed to PM2.5 are more susceptible to hypomethylation, which can result in upregulation of cancer-related signaling pathways (230). PM2.5 can also promote autophagy and malignancy of lung cells via ROS (231), including transformation of mouse fibroblasts occurs after exposure to ROS (232). Collectively, ROS can stimulate the malignant phenotypes of tumor cells to, such as proliferation, invasiveness, angiogenesis, metastasis, and resistance to apoptosis (233).

4.2.5. Synergistic effect of CS and PM

It is estimated that tobacco smoking contributed to 63.2% of lung cancer deaths, whereas 14.1% of the total lung cancer mortalities was attributable to ambient PM_{2.5} pollutions (14). CS and ambient air pollution are two ubiquitous factors extensively overlapped for a large population. Additionally, evidence has shown that a potential synergistic effect of exposure to CS and ambient air pollution exposure was greater than the sum of the effects from either exposure alone (234). Together, these factors tremendously increase the burden of lung cancer in the areas with high prevalence of cigarette smoking and heavy air pollutions, such as China and India, the most populous Asian countries. However, as the declining trend of lung cancer incidence occurred in many developed countries after the CS epidemic, the air pollution associated lung cancer could also be largely avoided if the etiology factors were effectively eliminated.

5. Conclusion

Genomic aberrations including germline inherited and carcinogen induced, play an important role in tumor initiation, progression, and metastasis. Although CS is the main risk factor for lung cancer, the exposure to potential risk factors of lung cancer is ubiquitous, including exposure to a variety of carcinogens from environment, such as pollutants from second-hand smoke, domestic radon, and fuel combustion. Additionally, complex mechanisms underlie carcinogen associated lung carcinogenesis. However, lung cancer is

one of the most preventable type of cancer that could potentially be largely avoided, if comprehensive policies and measures are taken to control and eliminate lung cancer risk factors.

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References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates
 of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA: a cancer journal for clinicians. 2021;71(3):209-49.
- 2. Parkin DM, Pisani P, Ferlay J. Global cancer statistics. CA: a cancer journal for clinicians. 1999;49(1):33-64, 1.
- 3. Proctor RN. The history of the discovery of the cigarette-lung cancer link: evidentiary traditions, corporate denial, global toll. Tobacco control. 2012;21(2):87-91.
- 4. Brawley OW, Glynn TJ, Khuri FR, Wender RC, Seffrin JR. The first Surgeon General's report on smoking and health: the 50th anniversary. CA: a cancer journal for clinicians. 2014;64(1):5-8.
- 5. de Prado-Bert P, Ruiz-Arenas C, Vives-Usano M, Andrusaityte S, Cadiou S, Carracedo Á, et al. The early-life exposome and epigenetic age acceleration in children. Environment international. 2021;155:106683.
- 6. IARC. Outdoor Air Pollution. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans. Volume 109.2013.
- 7. Turner MC, Andersen ZJ, Baccarelli A, Diver WR, Gapstur SM, Pope CA, 3rd, et al. Outdoor air pollution and cancer: An overview of the current evidence and public health recommendations. CA: a cancer journal for clinicians. 2020.
- 8. IARC. Air Pollution and Cancer. IARC Scientific Pub. No.161. Lyon, France: IARC Press; 2013.
- 9. Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D. Global cancer statistics. CA: a cancer journal for clinicians. 2011;61(2):69-90.
- 10. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA: a cancer journal for clinicians. 2015;65(2):87-108.
- 11. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA: a cancer journal for clinicians. 2018;68(6):394-424.
- 12. Omran AR. The epidemiologic transition. A theory of the epidemiology of population change. The Milbank Memorial Fund quarterly. 1971;49(4):509-38.
- 13. Omer Gersten JW. The cancer transition in Japan since 1951. Demographic Research. 2002;7:36.
- 14. Global, regional, and national comparative risk assessment of 84 behavioural, environmental and occupational, and metabolic risks or clusters of risks for 195 countries and territories, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017. Lancet (London, England). 2018;392(10159):1923-94.
- 15. Parkin DM, Bray FI, Devesa SS. Cancer burden in the year 2000. The global picture. European journal of cancer (Oxford, England: 1990). 2001;37 Suppl 8:S4-66.
- 16. Alonso R, Piñeros M, Laversanne M, Musetti C, Garau M, Barrios E, et al. Lung cancer incidence trends in Uruguay 1990-2014: An age-period-cohort analysis. Cancer epidemiology. 2018;55:17-22.
- 17. Hemminki K, Försti A, Hemminki A, Ljungberg B, Hemminki O. Incidence trends in lung and bladder cancers in the Nordic Countries before and after the smoking epidemic. European journal of cancer prevention: the official journal of the European Cancer Prevention Organisation (ECP). 2021.
- 18. Jemal A, Center MM, DeSantis C, Ward EM. Global patterns of cancer incidence and mortality rates and trends. Cancer epidemiology, biomarkers & prevention: a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology. 2010;19(8):1893-907.
- 19. Lam WK, White NW, Chan-Yeung MM. Lung cancer epidemiology and risk factors in Asia and Africa. The international journal of tuberculosis and lung disease: the official journal of the International Union against Tuberculosis and Lung Disease. 2004;8(9):1045-57.
- 20. Youlden DR, Cramb SM, Baade PD. The International Epidemiology of Lung Cancer: geographical distribution and secular trends. Journal of thoracic oncology: official publication of the International Association for the Study of Lung Cancer. 2008;3(8):819-31.
- 21. Jha P. Avoidable global cancer deaths and total deaths from smoking. Nature reviews Cancer. 2009;9(9):655-64.
- 22. Chen W, Xia C, Zheng R, Zhou M, Lin C, Zeng H, et al. Disparities by province, age, and sex in site-specific cancer burden attributable to 23 potentially modifiable risk factors in China: a comparative risk assessment. The Lancet Global health. 2019;7(2):e257-e69.
- 23. Chen Z, Peto R, Zhou M, Iona A, Smith M, Yang L, et al. Contrasting male and female trends in tobacco-attributed mortality in China: evidence from successive nationwide prospective cohort studies. Lancet (London, England). 2015;386(10002):1447-56.
- 24. Bray FI, Weiderpass E. Lung cancer mortality trends in 36 European countries: secular trends and birth cohort patterns by sex and region 1970-2007. International journal of cancer. 2010;126(6):1454-66.
- 25. Lortet-Tieulent J, Renteria E, Sharp L, Weiderpass E, Comber H, Baas P, et al. Convergence of decreasing male and increasing female incidence rates in major tobacco-related cancers in Europe in 1988-2010. European journal of cancer (Oxford, England: 1990). 2015;51(9):1144-63.

- 26. Rusmaully J, Tvardik N, Martin D, Billmann R, Cénée S, Antoine M, et al. Risk of lung cancer among women in relation to lifetime history of tobacco smoking: a population-based case-control study in France (the WELCA study). BMC cancer. 2021;21(1):711.
- 27. Sheikh M, Mukeriya A, Shangina O, Brennan P, Zaridze D. Postdiagnosis Smoking Cessation and Reduced Risk for Lung Cancer Progression and Mortality: A Prospective Cohort Study. Annals of internal medicine. 2021.
- 28. Takamori S, Shimokawa M, Matsubara T, Haratake N, Toyozawa R, Miura N, et al. Prognostic Impact of Smoking Period in Patients with Surgically Resected Non-small Cell Lung Cancer. Annals of surgical oncology. 2021;28(2):685-94.
- 29. WHO. WHO Framework Convention on Tobacco Control2003. 42 p.
- 30. Chudech S, Janmaimool P. Effectiveness of warning graphic labels on cigarette packs in enhancing late-teenagers' perceived fear of smoking-related harms in Bangkok, Thailand. Journal of public health research. 2021;10(1):1912.
- 31. Jemal A, Thun MJ, Ries LA, Howe HL, Weir HK, Center MM, et al. Annual report to the nation on the status of cancer, 1975–2005, featuring trends in lung cancer, tobacco use, and tobacco control. Journal of the National Cancer Institute. 2008;100(23):1672-94.
- 32. State-specific trends in lung cancer incidence and smoking--United States, 1999-2008. MMWR Morbidity and mortality weekly report. 2011;60(36):1243-7.
- Giovino GA, Mirza SA, Samet JM, Gupta PC, Jarvis MJ, Bhala N, et al. Tobacco use in 3 billion individuals from 16 countries: an analysis of nationally representative cross-sectional household surveys. Lancet (London, England). 2012;380(9842):668-79.
- 34. Wang J, Yu F, Shang Y, Ping Z, Liu L. Thyroid cancer: incidence and mortality trends in China, 2005-2015. Endocrine. 2020;68(1):163-73.
- 35. Kitahara CM, Pfeiffer RM, Sosa JA, Shiels MS. Impact of Overweight and Obesity on US Papillary Thyroid Cancer Incidence Trends (1995-2015). Journal of the National Cancer Institute. 2020;112(8):810-7.
- 36. Henschke CI, Yankelevitz DF, Jirapatnakul A, Yip R, Reccoppa V, Benjamin C, et al. Implementation of low-dose CT screening in two different health care systems: Mount Sinai Healthcare System and Phoenix VA Health Care System. Translational lung cancer research. 2021;10(2):1064-82.
- de Koning HJ, van der Aalst CM, de Jong PA, Scholten ET, Nackaerts K, Heuvelmans MA, et al. Reduced Lung-Cancer Mortality with Volume CT Screening in a Randomized Trial. The New England journal of medicine. 2020;382(6):503-13.
- 38. Passiglia F, Cinquini M, Bertolaccini L, Del Re M, Facchinetti F, Ferrara R, et al. Benefits and Harms of Lung Cancer Screening by Chest Computed Tomography: A Systematic Review and Meta-Analysis. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2021;39(23):2574-85.
- 39. Znyk M, Jurewicz J, Kaleta D. Exposure to Heated Tobacco Products and Adverse Health Effects, a Systematic Review. International journal of environmental research and public health. 2021;18(12).
- 40. Bracken-Clarke D, Kapoor D, Baird AM, Buchanan PJ, Gately K, Cuffe S, et al. Vaping and lung cancer A review of current data and recommendations. Lung cancer (Amsterdam, Netherlands). 2021;153:11-20.
- 41. Notes from the field: electronic cigarette use among middle and high school students United States, 2011-2012. MMWR Morbidity and mortality weekly report. 2013;62(35):729-30.
- 42. Tang MS, Wu XR, Lee HW, Xia Y, Deng FM, Moreira AL, et al. Electronic-cigarette smoke induces lung adenocarcinoma and bladder urothelial hyperplasia in mice. Proceedings of the National Academy of Sciences of the United States of America. 2019;116(43):21727-31.
- 43. Lee HW, Park SH, Weng MW, Wang HT, Huang WC, Lepor H, et al. E-cigarette smoke damages DNA and reduces repair activity in mouse lung, heart, and bladder as well as in human lung and bladder cells. Proceedings of the National Academy of Sciences of the United States of America. 2018;115(7):E1560-e9.
- 44. Hecht SS. Research opportunities related to establishing standards for tobacco products under the Family Smoking Prevention and Tobacco Control Act. Nicotine & tobacco research: official journal of the Society for Research on Nicotine and Tobacco. 2012;14(1):18-28.
- 45. Little JB, O'Toole WF. Respiratory tract tumors in hamsters induced by benzo(a)pyrene and 210Po alpha-radiation. Cancer research. 1974;34(11):3026-39.
- 46. Little JB, McGandy RB, Kennedy AR. Interactions between polonium-210 alpha-radiation, benzo(a)pyrene, and 0.9% NaCl solution instillations in the induction of experimental lung cancer. Cancer research. 1978;38(7):1929-35.
- 47. Deutsch-Wenzel RP, Brune H, Grimmer G, Dettbarn G, Misfeld J. Experimental studies in rat lungs on the carcinogenicity and dose-response relationships of eight frequently occurring environmental polycyclic aromatic hydrocarbons. Journal of the National Cancer Institute. 1983;71(3):539-44.
- 48. Deutsch-Wenzel RP, Brune H, Grimmer G. Experimental studies on the carcinogenicity of five nitrogen containing polycyclic aromatic compounds directly injected into rat lungs. Cancer letters. 1983;20(1):97-101.
- Nesnow S, Ross JA, Stoner GD, Mass MJ. Mechanistic linkage between DNA adducts, mutations in oncogenes and tumorigenesis of carcinogenic environmental polycyclic aromatic hydrocarbons in strain A/J mice. Toxicology. 1995;105(2-3):403-13.
- 50. Ross JA, Nelson GB, Wilson KH, Rabinowitz JR, Galati A, Stoner GD, et al. Adenomas induced by polycyclic aromatic hydrocarbons in strain A/J mouse lung correlate with time-integrated DNA adduct levels. Cancer research. 1995;55(5):1039-44.
- 51. Mass MJ, Abu-Shakra A, Roop BC, Nelson G, Galati AJ, Stoner GD, et al. Benzo[b]fluoranthene: tumorigenicity in strain A/J mouse lungs, DNA adducts and mutations in the Ki-ras oncogene. Carcinogenesis. 1996;17(8):1701-4.

- 52. LaVoie EJ, He ZM, Wu Y, Meschter CL, Weyand EH. Tumorigenic activity of the 4,5- and 9,10-dihydrodiols of benzo[j]fluoranthene and their syn- and anti-diol epoxides in newborn mice. Cancer research. 1994;54(4):962-8.
- 53. Sellakumar A, Shubik P. Carcinogenicity of different polycyclic hydrocarbons in the respiratory tract of hamsters. Journal of the National Cancer Institute. 1974;53(6):1713-9.
- 54. Ianysheva N, Balenko NV. [On experimental lung cancer caused by the introduction of various doses of 1, 2, 5, 6-dibenzanthracene]. Gigiena i sanitariia. 1966;31(7):12-5.
- 55. Nesnow S, Ross JA, Nelson G, Wilson K, Roop BC, Jeffers AJ, et al. Cyclopenta[cd]pyrene-induced tumorigenicity, Ki-ras codon 12 mutations and DNA adducts in strain A/J mouse lung. Carcinogenesis. 1994;15(4):601-6.
- 56. Sellakumar A, Stenbäck F, Rowland J, Shubik P. Tumur induction by 7H-dibenzol[c,g] carbazole in the respiratory tract of Syrian hamsters. Journal of toxicology and environmental health. 1977;3(5-6):935-9.
- 57. Warshawsky D, Talaska G, Jaeger M, Collins T, Galati A, You L, et al. Carcinogenicity, DNA adduct formation and K-ras activation by 7H-dibenzo[c,g]carbazole in strain A/J mouse lung. Carcinogenesis. 1996;17(4):865-71.
- 58. Dontenwill W, Mohr U. [On tracheal and bronchial carcinoma in the golden hamster after treatment with diethylnitrosamine]. Klinische Wochenschrift. 1961;39:493.
- 59. You M, Wang Y, Lineen AM, Gunning WT, Stoner GD, Anderson MW. Mutagenesis of the K-ras protooncogene in mouse lung tumors induced by N-ethyl-N-nitrosourea or N-nitrosodiethylamine. Carcinogenesis. 1992;13(9):1583-6.
- 60. Mervai Z, Egedi K, Kovalszky I, Baghy K. Diethylnitrosamine induces lung adenocarcinoma in FVB/N mouse. BMC cancer. 2018;18(1):157.
- 61. Hecht SS, Chen CB, Hirota N, Ornaf RM, Tso TC, Hoffmann D. Tobacco-specific nitrosamines: formation from nicotine in vitro and during tobacco curing and carcinogenicity in strain A mice. Journal of the National Cancer Institute. 1978;60(4):819-24.
- 62. Hecht SS, Lin D, Castonguay A, Rivenson A. Effects of alpha-deuterium substitution on the tumorigenicity of 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone in F344 rats. Carcinogenesis. 1987;8(2):291-4.
- 63. Staretz ME, Foiles PG, Miglietta LM, Hecht SS. Evidence for an important role of DNA pyridyloxobutylation in rat lung carcinogenesis by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone: effects of dose and phenethyl isothiocyanate. Cancer research. 1997;57(2):259-66.
- 64. Huff JE, Melnick RL, Solleveld HA, Haseman JK, Powers M, Miller RA. Multiple organ carcinogenicity of 1,3-butadiene in B6C3F1 mice after 60 weeks of inhalation exposure. Science (New York, NY). 1985;227(4686):548-9.
- 65. Miller RA, Melnick RL, Boorman GA. Neoplastic lesions induced by 1,3-butadiene in B6C3F1 mice. Experimental pathology. 1989;37(1-4):136-46.
- Melnick RL, Huff JE. 1,3-Butadiene induces cancer in experimental animals at all concentrations from 6.25 to 8000 parts per million. IARC scientific publications. 1993(127):309-22.
- 67. Guyer MF, Claus PE. Tumor of the lung in rats following injections of urethane (ethylcarbamate). Cancer research. 1947;7(6):342-5.
- 68. Stathopoulos GT, Sherrill TP, Cheng DS, Scoggins RM, Han W, Polosukhin VV, et al. Epithelial NF-kappaB activation promotes urethane-induced lung carcinogenesis. Proceedings of the National Academy of Sciences of the United States of America. 2007;104(47):18514-9.
- 69. Westcott PM, Halliwill KD, To MD, Rashid M, Rust AG, Keane TM, et al. The mutational landscapes of genetic and chemical models of Kras-driven lung cancer. Nature. 2015;517(7535):489-92.
- 70. Ottolenghi AD, Haseman JK, Payne WW, Falk HL, MacFarland HN. Inhalation studies of nickel sulfide in pulmonary carcinogenesis of rats. Journal of the National Cancer Institute. 1975;54(5):1165-72.
- 71. Poirier LA, Theiss JC, Arnold LJ, Shimkin MB. Inhibition by magnesium and calcium acetates of lead subacetate- and nickel acetate-induced lung tumors in strain A mice. Cancer research. 1984;44(4):1520-2.
- 72. Levy LS, Martin PA, Bidstrup PL. Investigation of the potential carcinogenicity of a range of chromium containing materials on rat lung. British journal of industrial medicine. 1986;43(4):243-56.
- 73. Glaser U, Hochrainer D, Klöppel H, Oldiges H. Carcinogenicity of sodium dichromate and chromium (VI/III) oxide aerosols inhaled by male Wistar rats. Toxicology. 1986;42(2-3):219-32.
- 74. Levy LS, Venitt S. Carcinogenicity and mutagenicity of chromium compounds: the association between bronchial metaplasia and neoplasia. Carcinogenesis. 1986;7(5):831-5.
- 75. Takahashi Y, Kondo K, Ishikawa S, Uchihara H, Fujino H, Sawada N, et al. Microscopic analysis of the chromium content in the chromium-induced malignant and premalignant bronchial lesions of the rat. Environmental research. 2005;99(2):267-72.
- 76. Oberdörster G. Airborne cadmium and carcinogenesis of the respiratory tract. Scandinavian journal of work, environment & health. 1986;12(6):523-37.
- 77. Waalkes MP, Rehm S. Chronic toxic and carcinogenic effects of cadmium chloride in male DBA/2NCr and NFS/NCr mice: strain-dependent association with tumors of the hematopoietic system, injection site, liver, and lung. Fundamental and applied toxicology: official journal of the Society of Toxicology. 1994;23(1):21-31.
- 78. Yuile CL, Berke HL, Hull T. Lung Cancer Following Polonium-210 Inhalation in Rats. Radiation Research. 1967;31(4):760-74.
- 79. Ishinishi N, Kodama Y, Nobutomo K, Hisanaga A. Preliminary experimental study on carcinogenicity of arsenic trioxide in rat lung. Environmental health perspectives. 1977;19:191-6.
- 80. Some metals and metallic compounds. IARC monographs on the evaluation of the carcinogenic risk of chemicals to humans. 1980;23:1-415.

- 81. Milia U, Di Leo FP. [Tumors of the lung induced with hydrazine hydrate in mice of the BALB-c-Cb-Se substrain]. Lavori dell'Istituto di anatomia e istologia patologica, Universita degli studi di Perugia. 1965;25(3):149-54.
- 82. Toth B. Lung tumor induction and inhibition of breast adenocarcinomas by hydrazine sulfate in mice. Journal of the National Cancer Institute. 1969;42(3):469-75.
- 83. Bhide SV, D'Souza RA, Sawai MM, Ranadive KJ. Lung tumour incidence in mice treated with hydrazine sulphate. International journal of cancer. 1976;18(4):530-5.
- 84. Hecht SS. Tobacco smoke carcinogens and lung cancer. Journal of the National Cancer Institute. 1999;91(14):1194-210.
- 85. Hecht SS. Tobacco carcinogens, their biomarkers and tobacco-induced cancer. Nature reviews Cancer. 2003;3(10):733-44.
- 86. Luo K, Luo X, Cao W, Hochalter JB, Paiano V, Sipe CJ, et al. Cigarette smoking enhances the metabolic activation of the polycyclic aromatic hydrocarbon phenanthrene in humans. Carcinogenesis. 2021;42(4):570-7.
- 87. Xia B, Blount BC, Guillot T, Brosius C, Li Y, Van Bemmel DM, et al. Tobacco-Specific Nitrosamines (NNAL, NNN, NAT, and NAB) Exposures in the US Population Assessment of Tobacco and Health (PATH) Study Wave 1 (2013-2014). Nicotine & tobacco research: official journal of the Society for Research on Nicotine and Tobacco. 2021;23(3):573-83.
- 88. Hecht SS, Morse MA, Amin S, Stoner GD, Jordan KG, Choi CI, et al. Rapid single-dose model for lung tumor induction in A/J mice by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone and the effect of diet. Carcinogenesis. 1989;10(10):1901-4.
- 89. Rivenson A, Hoffmann D, Prokopczyk B, Amin S, Hecht SS. Induction of lung and exocrine pancreas tumors in F344 rats by tobacco-specific and Areca-derived N-nitrosamines. Cancer research. 1988;48(23):6912-7.
- West KA, Brognard J, Clark AS, Linnoila IR, Yang X, Swain SM, et al. Rapid Akt activation by nicotine and a tobacco carcinogen modulates the phenotype of normal human airway epithelial cells. The Journal of clinical investigation. 2003;111(1):81-90.
- 91. Klein-Szanto AJ, Iizasa T, Momiki S, Garcia-Palazzo I, Caamano J, Metcalf R, et al. A tobacco-specific N-nitrosamine or cigarette smoke condensate causes neoplastic transformation of xenotransplanted human bronchial epithelial cells. Proceedings of the National Academy of Sciences of the United States of America. 1992;89(15):6693-7.
- 92. Di ME, Kahkonen B, Liu CH, Di YP. Lung carcinomas induced by NNK and LPS. Methods in cell biology. 2021;163:175-85.
- 93. Peterson LA, Oram MK, Flavin M, Seabloom D, Smith WE, O'Sullivan MG, et al. Coexposure to Inhaled Aldehydes or Carbon Dioxide Enhances the Carcinogenic Properties of the Tobacco-Specific Nitrosamine 4-Methylnitrosamino-1-(3-pyridyl)-1-butanone in the A/J Mouse Lung. Chemical research in toxicology. 2021;34(3):723-32.
- 94. Whong WZ, Stewart JD, Cutler D, Ong T. Induction of in vivo DNA adducts by 4 industrial by-products in the rat-lung-cell system. Mutation research. 1994;312(2):165-72.
- 95. Warshawsky D, Talaska G, Xue W, Schneider J. Comparative carcinogenicity, metabolism, mutagenicity, and DNA binding of 7H-dibenzo[c,g]carbazole and dibenz[a,j]acridine. Critical reviews in toxicology. 1996;26(2):213-49.
- 96. Shertzer HG, Genter MB, Talaska G, Curran CP, Nebert DW, Dalton TP. 7H-dibenzo[c,g]carbazole metabolism by the mouse and human CYP1 family of enzymes. Carcinogenesis. 2007;28(6):1371-8.
- 97. Dowty HV, Xue W, LaDow K, Talaska G, Warshawsky D. One-electron oxidation is not a major route of metabolic activation and DNA binding for the carcinogen 7H-dibenzo[c,g]carbazole in vitro and in mouse liver and lung. Carcinogenesis. 2000;21(5):991-8.
- 98. Brunnemann KD, Kagan MR, Cox JE, Hoffmann D. Analysis of 1,3-butadiene and other selected gas-phase components in cigarette mainstream and sidestream smoke by gas chromatography-mass selective detection. Carcinogenesis. 1990;11(10):1863-8.
- 99. Albertini RJ, Carson ML, Kirman CR, Gargas ML. 1,3-Butadiene: II. Genotoxicity profile. Critical reviews in toxicology. 2010;40 Suppl 1:12-73.
- 100. Walker VE, Degner A, Carter EW, Nicklas JA, Walker DM, Tretyakova N, et al. 1,3-Butadiene metabolite 1,2,3,4 diepoxybutane induces DNA adducts and micronuclei but not t(9;22) translocations in human cells. Chemico-biological interactions. 2019;312:108797.
- 101. Boysen G, Arora R, Degner A, Vevang KR, Chao C, Rodriguez F, et al. Effects of GSTT1 Genotype on the Detoxification of 1,3-Butadiene Derived Diepoxide and Formation of Promutagenic DNA-DNA Cross-Links in Human Hapmap Cell Lines. Chemical research in toxicology. 2021;34(1):119-31.
- 102. Balharry D, Sexton K, BéruBé KA. An in vitro approach to assess the toxicity of inhaled tobacco smoke components: nicotine, cadmium, formaldehyde and urethane. Toxicology. 2008;244(1):66-76.
- 103. Shakeri MT, Nezami H, Nakhaee S, Aaseth J, Mehrpour O. Assessing Heavy Metal Burden Among Cigarette Smokers and Non-smoking Individuals in Iran: Cluster Analysis and Principal Component Analysis. Biological trace element research. 2021.
- 104. Tanaka I, Ishimatsu S, Matsuno K, Kodama Y, Tsuchiya K. Biological half time of deposited nickel oxide aerosol in rat lung by inhalation. Biological trace element research. 1985;8(3):203-10.
- 105. Saplakoğlu U, Işcan M, Işcan M. DNA single-strand breakage in rat lung, liver and kidney after single and combined treatments of nickel and cadmium. Mutation research. 1997;394(1-3):133-40.
- 106. Tsapakos MJ, Hampton TH, Wetterhahn KE. Chromium(VI)-induced DNA lesions and chromium distribution in rat kidney, liver, and lung. Cancer research. 1983;43(12 Pt 1):5662-7.
- 107. Cheng L, Sonntag DM, de Boer J, Dixon K. Chromium(VI)-induced mutagenesis in the lungs of big blue transgenic mice. Journal of environmental pathology, toxicology and oncology: official organ of the International Society for Environmental Toxicology and Cancer. 2000;19(3):239-49.
- 108. Murrell W. Preliminary Report on the Presence of Arsenic in Cigarettes. British medical journal. 1896;2(1854):96.

- 109. Waalkes MP, Qu W, Tokar EJ, Kissling GE, Dixon D. Lung tumors in mice induced by "whole-life" inorganic arsenic exposure at human-relevant doses. Archives of toxicology. 2014;88(8):1619-29.
- 110. Wang Z. Mechanisms of the synergistic lung tumorigenic effect of arsenic and benzo(a)pyrene combined- exposure. Seminars in cancer biology. 2021.
- 111. Reed L, Arlt VM, Phillips DH. The role of cytochrome P450 enzymes in carcinogen activation and detoxication: an in vivo-in vitro paradox. Carcinogenesis. 2018;39(7):851-9.
- 112. Dasari S, Ganjayi MS, Yellanurkonda P, Basha S, Meriga B. Role of glutathione S-transferases in detoxification of a polycyclic aromatic hydrocarbon, methylcholanthrene. Chemico-biological interactions. 2018;294:81-90.
- 113. Crespi CL, Penman BW, Gelboin HV, Gonzalez FJ. A tobacco smoke-derived nitrosamine, 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone, is activated by multiple human cytochrome P450s including the polymorphic human cytochrome P4502D6. Carcinogenesis. 1991;12(7):1197-201.
- 114. Hartman JH, Miller GP, Caro AA, Byrum SD, Orr LM, Mackintosh SG, et al. 1,3-Butadiene-induced mitochondrial dysfunction is correlated with mitochondrial CYP2E1 activity in Collaborative Cross mice. Toxicology. 2017;378:114-24.
- 115. Yamazaki H, Oda Y, Funae Y, Imaoka S, Inui Y, Guengerich FP, et al. Participation of rat liver cytochrome P450 2E1 in the activation of N-nitrosodimethylamine and N-nitrosodiethylamine to products genotoxic in an acetyltransferase-overexpressing Salmonella typhimurium strain (NM2009). Carcinogenesis. 1992;13(6):979-85.
- 116. Gastelum G, Jiang W, Wang L, Zhou G, Borkar R, Putluri N, et al. Polycyclic Aromatic Hydrocarbon-induced Pulmonary Carcinogenesis in Cytochrome P450 (CYP)1A1- and 1A2-Null Mice: Roles of CYP1A1 and CYP1A2. Toxicological sciences: an official journal of the Society of Toxicology. 2020;177(2):347-61.
- 117. Vázquez-Gómez G, Rocha-Zavaleta L, Rodríguez-Sosa M, Petrosyan P, Rubio-Lightbourn J. Benzo[a]pyrene activates an AhR/Src/ERK axis that contributes to CYP1A1 induction and stable DNA adducts formation in lung cells. Toxicology letters. 2018;289:54-62.
- 118. Alzahrani AM, Rajendran P. Pinocembrin attenuates benzo(a)pyrene-induced CYP1A1 expression through multiple pathways: An in vitro and in vivo study. Journal of biochemical and molecular toxicology. 2021;35(4):e22695.
- 119. Butkiewicz D, Grzybowska E, Phillips DH, Hemminki K, Chorazy M. Polymorphisms of the GSTP1 and GSTM1 genes and PAH-DNA adducts in human mononuclear white blood cells. Environmental and molecular mutagenesis. 2000;35(2):99-105.
- 120. Kozlovich S, Chen G, Lazarus P. Stereospecific Metabolism of the Tobacco-Specific Nitrosamine, NNAL. Chemical research in toxicology. 2015;28(11):2112-9.
- 121. Munnia A, Giese RW, Polvani S, Galli A, Cellai F, Peluso MEM. Bulky DNA Adducts, Tobacco Smoking, Genetic Susceptibility, and Lung Cancer Risk. Advances in clinical chemistry. 2017;81:231-77.
- 122. Motwani HV, Westberg E, Lindh C, Abramsson-Zetterberg L, Törnqvist M. Serum albumin adducts, DNA adducts and micronuclei frequency measured in benzo[a]pyrene-exposed mice for estimation of genotoxic potency. Mutation research Genetic toxicology and environmental mutagenesis. 2020;849:503127.
- 123. Hecht SS. Biochemistry, biology, and carcinogenicity of tobacco-specific N-nitrosamines. Chemical research in toxicology. 1998;11(6):559-603.
- 124. Hecht SS, Carmella SG, Foiles PG, Murphy SE, Peterson LA. Tobacco-specific nitrosamine adducts: studies in laboratory animals and humans. Environmental health perspectives. 1993;99:57-63.
- 125. Zarth AT, Upadhyaya P, Yang J, Hecht SS. DNA Adduct Formation from Metabolic 5'-Hydroxylation of the Tobacco-Specific Carcinogen N'-Nitrosonornicotine in Human Enzyme Systems and in Rats. Chemical research in toxicology. 2016;29(3):380-9.
- 126. Zhao L, Balbo S, Wang M, Upadhyaya P, Khariwala SS, Villalta PW, et al. Quantitation of pyridyloxobutyl-DNA adducts in tissues of rats treated chronically with (R)- or (S)-N'-nitrosonornicotine (NNN) in a carcinogenicity study. Chemical research in toxicology. 2013;26(10):1526-35.
- 127. Foiles PG, Akerkar SA, Carmella SG, Kagan M, Stoner GD, Resau JH, et al. Mass spectrometric analysis of tobacco-specific nitrosamine-DNA adducts in smokers and nonsmokers. Chemical research in toxicology. 1991;4(3):364-8.
- 128. Hecht SS, Carmella SG, Foiles PG, Murphy SE. Biomarkers for human uptake and metabolic activation of tobacco-specific nitrosamines. Cancer research. 1994;54(7 Suppl):1912s-7s.
- 129. Li Y, Carlson ES, Zarth AT, Upadhyaya P, Hecht SS. Investigation of 2'-Deoxyadenosine-Derived Adducts Specifically Formed in Rat Liver and Lung DNA by N'-Nitrosonornicotine Metabolism. Chemical research in toxicology. 2021;34(4):1004-15.
- 130. Li Y, Hecht SS. Identification of an N'-Nitrosonornicotine-Specific Deoxyadenosine Adduct in Rat Liver and Lung DNA. Chemical research in toxicology. 2021;34(4):992-1003.
- 131. Ronai ZA, Gradia S, Peterson LA, Hecht SS. G to A transitions and G to T transversions in codon 12 of the Ki-ras oncogene isolated from mouse lung tumors induced by 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone (NNK) and related DNA methylating and pyridyloxobutylating agents. Carcinogenesis. 1993;14(11):2419-22.
- 132. Kawano R, Takeshima Y, Inai K. Effects of K-ras gene mutations in the development of lung lesions induced by 4-(N-methyl-n-nitrosamino)-1-(3-pyridyl)-1-butanone in A/J mice. Japanese journal of cancer research: Gann. 1996;87(1):44-50.
- 133. Pfeifer GP, Denissenko MF, Olivier M, Tretyakova N, Hecht SS, Hainaut P. Tobacco smoke carcinogens, DNA damage and p53 mutations in smoking-associated cancers. Oncogene. 2002;21(48):7435-51.
- 134. Smith LE, Denissenko MF, Bennett WP, Li H, Amin S, Tang M, et al. Targeting of lung cancer mutational hotspots by polycyclic aromatic hydrocarbons. Journal of the National Cancer Institute. 2000;92(10):803-11.
- 135. Chen JX, Zheng Y, West M, Tang MS. Carcinogens preferentially bind at methylated CpG in the p53 mutational hot spots. Cancer research. 1998;58(10):2070-5.

- 136. Denissenko MF, Pao A, Tang M, Pfeifer GP. Preferential formation of benzo[a]pyrene adducts at lung cancer mutational hotspots in P53. Science (New York, NY). 1996;274(5286):430-2.
- 137. Kotandeniya D, Murphy D, Yan S, Park S, Seneviratne U, Koopmeiners JS, et al. Kinetics of O(6)-pyridyloxobutyl-2'-deoxyguanosine repair by human O(6)-alkylguanine DNA alkyltransferase. Biochemistry. 2013;52(23):4075-88.
- 138. Westra WH, Slebos RJ, Offerhaus GJ, Goodman SN, Evers SG, Kensler TW, et al. K-ras oncogene activation in lung adenocarcinomas from former smokers. Evidence that K-ras mutations are an early and irreversible event in the development of adenocarcinoma of the lung. Cancer. 1993;72(2):432-8.
- 139. Menzies GE, Prior IA, Brancale A, Reed SH, Lewis PD. Carcinogen-induced DNA structural distortion differences in the RAS gene isoforms; the importance of local sequence. BMC chemistry. 2021;15(1):51.
- 140. Nesnow S, Ross JA, Mass MJ, Stoner GD. Mechanistic relationships between DNA adducts, oncogene mutations, and lung tumorigenesis in strain A mice. Experimental lung research. 1998;24(4):395-405.
- 141. Belinsky SA, Devereux TR, Maronpot RR, Stoner GD, Anderson MW. Relationship between the formation of promutagenic adducts and the activation of the K-ras protooncogene in lung tumors from A/J mice treated with nitrosamines. Cancer research. 1989;49(19):5305-11.
- 142. Topaloglu O, Hoque MO, Tokumaru Y, Lee J, Ratovitski E, Sidransky D, et al. Detection of promoter hypermethylation of multiple genes in the tumor and bronchoalveolar lavage of patients with lung cancer. Clinical cancer research: an official journal of the American Association for Cancer Research. 2004;10(7):2284-8.
- 143. Belinsky SA, Nikula KJ, Palmisano WA, Michels R, Saccomanno G, Gabrielson E, et al. Aberrant methylation of p16(INK4a) is an early event in lung cancer and a potential biomarker for early diagnosis. Proceedings of the National Academy of Sciences of the United States of America. 1998;95(20):11891-6.
- 144. Nelson HH, Wiencke JK, Gunn L, Wain JC, Christiani DC, Kelsey KT. Chromosome 3p14 alterations in lung cancer: evidence that FHIT exon deletion is a target of tobacco carcinogens and asbestos. Cancer research. 1998;58(9):1804-7.
- 145. Chatterjee N, Walker GC. Mechanisms of DNA damage, repair, and mutagenesis. Environmental and molecular mutagenesis. 2017;58(5):235-63.
- 146. Sancar A. DNA excision repair. Annual review of biochemistry. 1996;65:43-81.
- 147. Cai Y, Geacintov NE, Broyde S. Nucleotide excision repair efficiencies of bulky carcinogen-DNA adducts are governed by a balance between stabilizing and destabilizing interactions. Biochemistry. 2012;51(7):1486-99.
- 148. Pegg AE, Dolan ME, Moschel RC. Structure, function, and inhibition of O6-alkylguanine-DNA alkyltransferase. Progress in nucleic acid research and molecular biology. 1995;51:167-223.
- 149. Huang YT, Lin X, Liu Y, Chirieac LR, McGovern R, Wain J, et al. Cigarette smoking increases copy number alterations in nonsmall-cell lung cancer. Proceedings of the National Academy of Sciences of the United States of America. 2011;108(39):16345-50
- 150. Hecht SS. Human urinary carcinogen metabolites: biomarkers for investigating tobacco and cancer. Carcinogenesis. 2002;23(6):907-22.
- 151. Tang D, Phillips DH, Stampfer M, Mooney LA, Hsu Y, Cho S, et al. Association between carcinogen-DNA adducts in white blood cells and lung cancer risk in the physicians health study. Cancer research. 2001;61(18):6708-12.
- 152. Richie JP, Jr., Carmella SG, Muscat JE, Scott DG, Akerkar SA, Hecht SS. Differences in the urinary metabolites of the tobacco-specific lung carcinogen 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone in black and white smokers. Cancer epidemiology, biomarkers & prevention: a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology. 1997;6(10):783-90.
- 153. Ryberg D, Hewer A, Phillips DH, Haugen A. Different susceptibility to smoking-induced DNA damage among male and female lung cancer patients. Cancer research. 1994;54(22):5801-3.
- 154. Rostron BL, Wang J, Etemadi A, Thakur S, Chang JT, Bhandari D, et al. Associations between Biomarkers of Exposure and Lung Cancer Risk among Exclusive Cigarette Smokers in the Golestan Cohort Study. International journal of environmental research and public health. 2021;18(14).
- 155. Park EY, Lim MK, Park E, Oh JK, Lee DH. Relationship Between Urinary 4-(Methylnitrosamino)-1-(3-Pyridyl)-1-Butanol and Lung Cancer Risk in the General Population: A Community-Based Prospective Cohort Study. Frontiers in oncology. 2021;11:611674.
- 156. Adam T, Mitschke S, Streibel T, Baker RR, Zimmermann R. Quantitative puff-by-puff-resolved characterization of selected toxic compounds in cigarette mainstream smoke. Chemical research in toxicology. 2006;19(4):511-20.
- 157. Yoshie Y, Ohshima H. Synergistic induction of DNA strand breakage by cigarette tar and nitric oxide. Carcinogenesis. 1997;18(7):1359-63.
- 158. Chouchane S, Wooten JB, Tewes FJ, Wittig A, Müller BP, Veltel D, et al. Involvement of semiquinone radicals in the in vitro cytotoxicity of cigarette mainstream smoke. Chemical research in toxicology. 2006;19(12):1602-10.
- 159. Pryor WA, Stone K, Zang LY, Bermúdez E. Fractionation of aqueous cigarette tar extracts: fractions that contain the tar radical cause DNA damage. Chemical research in toxicology. 1998;11(5):441-8.
- 160. Ahmad R, Hussain A, Ahsan H. Peroxynitrite: cellular pathology and implications in autoimmunity. Journal of immunoassay & immunochemistry. 2019;40(2):123-38.
- 161. Rosen JE, Prahalad AK, Williams GM. 8-Oxodeoxyguanosine formation in the DNA of cultured cells after exposure to H2O2 alone or with UVB or UVA irradiation. Photochemistry and photobiology. 1996;64(1):117-22.

- 162. Di Vincenzo S, Sangiorgi C, Ferraro M, Buscetta M, Cipollina C, Pace E. Cigarette smoke extract reduces FOXO3a promoting tumor progression and cell migration in lung cancer. Toxicology. 2021;454:152751.
- 163. Dietrich M, Block G, Hudes M, Morrow JD, Norkus EP, Traber MG, et al. Antioxidant supplementation decreases lipid peroxidation biomarker F(2)-isoprostanes in plasma of smokers. Cancer epidemiology, biomarkers & prevention: a publication of the American Association for Cancer Research, cosponsored by the American Society of Preventive Oncology. 2002;11(1):7-13.
- 164. Reis R, Orak D, Yilmaz D, Cimen H, Sipahi H. Modulation of cigarette smoke extract-induced human bronchial epithelial damage by eucalyptol and curcumin. Human & experimental toxicology. 2021;40(9):1445-62.
- 165. Gopalakrishna R, Chen ZH, Gundimeda U. Tobacco smoke tumor promoters, catechol and hydroquinone, induce oxidative regulation of protein kinase C and influence invasion and metastasis of lung carcinoma cells. Proceedings of the National Academy of Sciences of the United States of America. 1994;91(25):12233-7.
- 166. Bradley KL, Stokes CA, Marciniak SJ, Parker LC, Condliffe AM. Role of unfolded proteins in lung disease. Thorax. 2021;76(1):92-9.
- 167. Rico de Souza A, Traboulsi H, Wang X, Fritz JH, Eidelman DH, Baglole CJ. The Aryl Hydrocarbon Receptor Attenuates Acute Cigarette Smoke-Induced Airway Neutrophilia Independent of the Dioxin Response Element. Frontiers in immunology. 2021:12:630427.
- 168. Jin M, Li G, Liu W, Wu X, Zhu J, Zhao D, et al. Cigarette smoking induces aberrant N(6)-methyladenosine of DAPK2 to promote non-small cell lung cancer progression by activating NF-κB pathway. Cancer letters. 2021;518:214-29.
- 169. Liu J, Chen SJ, Hsu SW, Zhang J, Li JM, Yang DC, et al. MARCKS cooperates with NKAP to activate NF-kB signaling in smokerelated lung cancer. Theranostics. 2021;11(9):4122-36.
- 170. Cao C, Tian B, Geng X, Zhou H, Xu Z, Lai T, et al. IL-17-Mediated Inflammation Promotes Cigarette Smoke-Induced Genomic Instability. Cells. 2021;10(5).
- 171. Masunaga A, Takemura T, Ichiyasu H, Migiyama E, Horio Y, Saeki S, et al. Pathological and clinical relevance of selective recruitment of Langerhans cells in the respiratory bronchioles of smokers. Respiratory investigation. 2021;59(4):513-21.
- 172. Colarusso C, Terlizzi M, Lamort AS, Cerqua I, Roviezzo F, Stathopoulos G, et al. Caspase-11 and AIM2 inflammasome are involved in smoking-induced COPD and lung adenocarcinoma. Oncotarget. 2021;12(11):1057-71.
- 173. Liu A, Zhang X, Li R, Zheng M, Yang S, Dai L, et al. Overexpression of the SARS-CoV-2 receptor ACE2 is induced by cigarette smoke in bronchial and alveolar epithelia. The Journal of pathology. 2021;253(1):17-30.
- 174. Xiong R, Wu Y, Wu Q, Muskhelishvili L, Davis K, Tripathi P, et al. Integration of transcriptome analysis with pathophysiological endpoints to evaluate cigarette smoke toxicity in an in vitro human airway tissue model. Archives of toxicology. 2021;95(5):1739-61
- 175. Cheng CC, Lin HC, Chiang YW, Chang J, Sie ZL, Yang BL, et al. Nicotine exhausts CD8(+) T cells against tumor cells through increasing miR-629-5p to repress IL2RB-mediated granzyme B expression. Cancer immunology, immunotherapy: CII. 2021;70(5):1351-64.
- 176. Aloe C, Wang H, Vlahos R, Irving L, Steinfort D, Bozinovski S. Emerging and multifaceted role of neutrophils in lung cancer. Translational lung cancer research. 2021;10(6):2806-18.
- 177. Salem ML, El-Ashmawy NE, Abd El-Fattah EE, Khedr EG. Immunosuppressive role of Benzo[a]pyrene in induction of lung cancer in mice. Chemico-biological interactions. 2021;333:109330.
- 178. Wang GZ, Zhang L, Zhao XC, Gao SH, Qu LW, Yu H, et al. The Aryl hydrocarbon receptor mediates tobacco-induced PD-L1 expression and is associated with response to immunotherapy. Nature communications. 2019;10(1):1125.
- 179. Borghaei H, Paz-Ares L, Horn L, Spigel DR, Steins M, Ready NE, et al. Nivolumab versus Docetaxel in Advanced Nonsquamous Non-Small-Cell Lung Cancer. The New England journal of medicine. 2015;373(17):1627-39.
- 180. Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. The New England journal of medicine. 2015;372(21):2018-28.
- 181. Ramírez-Salazar EG, Gayosso-Gómez LV, Baez-Saldaña R, Falfán-Valencia R, Pérez-Padilla R, Higuera-Iglesias AL, et al. Cigarette Smoking Alters the Expression of Circulating microRNAs and Its Potential Diagnostic Value in Female Lung Cancer Patients. Biology. 2021;10(8).
- 182. Lee SB, Park YS, Sung JS, Lee JW, Kim B, Kim YH. Tumor Suppressor miR-584-5p Inhibits Migration and Invasion in Smoking Related Non-Small Cell Lung Cancer Cells by Targeting YKT6. Cancers. 2021;13(5).
- 183. Gomes S, Cavadas B, Ferreira JC, Marques PI, Monteiro C, Sucena M, et al. Profiling of lung microbiota discloses differences in adenocarcinoma and squamous cell carcinoma. Scientific reports. 2019;9(1):12838.
- 184. Turek EM, Cox MJ, Hunter M, Hui J, James P, Willis-Owen SAG, et al. Airway microbial communities, smoking and asthma in a general population sample. EBioMedicine. 2021;71:103538.
- 185. Greathouse KL, White JR, Vargas AJ, Bliskovsky VV, Beck JA, von Muhlinen N, et al. Interaction between the microbiome and TP53 in human lung cancer. Genome biology. 2018;19(1):123.
- 186. Ponciano-Rodríguez G, Gaso MI, Armienta MA, Trueta C, Morales I, Alfaro R, et al. Indoor radon exposure and excess of lung cancer mortality: the case of Mexico-an ecological study. Environmental geochemistry and health. 2021;43(1):221-34.
- 187. Cheng ES, Egger S, Hughes S, Weber M, Steinberg J, Rahman B, et al. Systematic review and meta-analysis of residential radon and lung cancer in never-smokers. European respiratory review: an official journal of the European Respiratory Society. 2021;30(159).

- 188. Thomas D, Pogoda J, Langholz B, Mack W. Temporal modifiers of the radon-smoking interaction. Health physics. 1994;66(3):257-62.
- 189. Böhm R, Sedlák A, Bulko M, Holý K. Radon as a Tracer of Lung Changes Induced by Smoking. Risk analysis: an official publication of the Society for Risk Analysis. 2020;40(2):370-84.
- 190. Martell EA. alpha-Radiation dose at bronchial bifurcations of smokers from indoor exposure to radon progeny. Proceedings of the National Academy of Sciences of the United States of America. 1983;80(5):1285-9.
- 191. Personal habits and indoor combustions. Volume 100 E. A review of human carcinogens. IARC monographs on the evaluation of carcinogenic risks to humans. 2012;100(Pt E):1-538.
- 192. Boffetta P, Nyberg F. Contribution of environmental factors to cancer risk. British medical bulletin. 2003;68:71-94.
- 193. Thun MJ, Hannan LM, Adams-Campbell LL, Boffetta P, Buring JE, Feskanich D, et al. Lung cancer occurrence in never-smokers: an analysis of 13 cohorts and 22 cancer registry studies. PLoS medicine. 2008;5(9):e185.
- 194. Leikauf GD, Kim SH, Jang AS. Mechanisms of ultrafine particle-induced respiratory health effects. Experimental & molecular medicine. 2020;52(3):329-37.
- 195. Lall R, Thurston GD. Identifying and quantifying transported vs. local sources of New York City PM2.5 fine particulate matter air pollution. Atmospheric Environment. 2006;40:333-46.
- 196. Thurston G. Outdoor Air Pollution: Sources, Atmospheric Transport, and Human Health Effects. 2016. p. 367-77.
- 197. Brauer M, Avila-Casado C, Fortoul TI, Vedal S, Stevens B, Churg A. Air pollution and retained particles in the lung. Environmental health perspectives. 2001;109(10):1039-43.
- 198. Hamra GB, Guha N, Cohen A, Laden F, Raaschou-Nielsen O, Samet JM, et al. Outdoor particulate matter exposure and lung cancer: a systematic review and meta-analysis. Environmental health perspectives. 2014;122(9):906-11.
- 199. Gharibvand L, Lawrence Beeson W, Shavlik D, Knutsen R, Ghamsary M, Soret S, et al. The association between ambient fine particulate matter and incident adenocarcinoma subtype of lung cancer. Environmental health: a global access science source. 2017;16(1):71.
- 200. Eckel SP, Cockburn M, Shu YH, Deng H, Lurmann FW, Liu L, et al. Air pollution affects lung cancer survival. Thorax. 2016;71(10):891-8.
- 201. Turner MC, Krewski D, Pope CA, 3rd, Chen Y, Gapstur SM, Thun MJ. Long-term ambient fine particulate matter air pollution and lung cancer in a large cohort of never-smokers. American journal of respiratory and critical care medicine. 2011;184(12):1374-81.
- 202. Yu F, Xiao R, Li X, Hu Z, Cai L, He F. Combined effects of lung disease history, environmental exposures, and family history of lung cancer to susceptibility of lung cancer in Chinese non-smokers. Respiratory research. 2021;22(1):210.
- 203. Santos LR, Alves-Correia M, Câmara M, Lélis M, Caldeira C, Brazão ML, et al. Multiple Victims of Carbon Monoxide Poisoning in the Aftermath of a Wildfire: A Case Series. Acta medica portuguesa. 2018;31(3):146-51.
- 204. Gharibvand L, Shavlik D, Ghamsary M, Beeson WL, Soret S, Knutsen R, et al. The Association between Ambient Fine Particulate Air Pollution and Lung Cancer Incidence: Results from the AHSMOG-2 Study. Environmental health perspectives. 2017;125(3):378-84.
- 205. Tomczak A, Miller AB, Weichenthal SA, To T, Wall C, van Donkelaar A, et al. Long-term exposure to fine particulate matter air pollution and the risk of lung cancer among participants of the Canadian National Breast Screening Study. International journal of cancer. 2016;139(9):1958-66.
- 206. Consonni D, Carugno M, De Matteis S, Nordio F, Randi G, Bazzano M, et al. Outdoor particulate matter (PM10) exposure and lung cancer risk in the EAGLE study. PloS one. 2018;13(9):e0203539.
- 207. Moon DH, Kwon SO, Kim SY, Kim WJ. Air Pollution and Incidence of Lung Cancer by Histological Type in Korean Adults: A Korean National Health Insurance Service Health Examinee Cohort Study. International journal of environmental research and public health. 2020;17(3).
- 208. Lippmann M, Chen LC, Gordon T, Ito K, Thurston GD. National Particle Component Toxicity (NPACT) Initiative: integrated epidemiologic and toxicologic studies of the health effects of particulate matter components. Research report (Health Effects Institute). 2013(177):5-13.
- 209. Weichenthal S, Crouse DL, Pinault L, Godri-Pollitt K, Lavigne E, Evans G, et al. Oxidative burden of fine particulate air pollution and risk of cause-specific mortality in the Canadian Census Health and Environment Cohort (CanCHEC). Environmental research. 2016;146:92-9.
- 210. Raaschou-Nielsen O, Beelen R, Wang M, Hoek G, Andersen ZJ, Hoffmann B, et al. Particulate matter air pollution components and risk for lung cancer. Environment international. 2016;87:66-73.
- 211. Matschullat J. Arsenic in the geosphere--a review. The Science of the total environment. 2000;249(1-3):297-312.
- 212. Lim H, Devesa SS, Sosa JA, Check D, Kitahara CM. Trends in Thyroid Cancer Incidence and Mortality in the United States, 1974-2013. Jama. 2017;317(13):1338-48.
- 213. Cogliano VJ, Baan R, Straif K, Grosse Y, Lauby-Secretan B, El Ghissassi F, et al. Preventable exposures associated with human cancers. Journal of the National Cancer Institute. 2011;103(24):1827-39.
- 214. Castaño-Vinyals G, D'Errico A, Malats N, Kogevinas M. Biomarkers of exposure to polycyclic aromatic hydrocarbons from environmental air pollution. Occupational and environmental medicine. 2004;61(4):e12.
- 215. Manisalidis I, Stavropoulou E, Stavropoulos A, Bezirtzoglou E. Environmental and Health Impacts of Air Pollution: A Review. Frontiers in public health. 2020;8:14.

- 216. Yamagishi K, Onuma K, Chiba Y, Yagi S, Aoki S, Sato T, et al. Generation of gaseous sulfur-containing compounds in tumour tissue and suppression of gas diffusion as an antitumour treatment. Gut. 2012;61(4):554-61.
- 217. Arlt VM. 3-Nitrobenzanthrone, a potential human cancer hazard in diesel exhaust and urban air pollution: a review of the evidence. Mutagenesis. 2005;20(6):399-410.
- 218. Moorthy B, Chu C, Carlin DJ. Polycyclic aromatic hydrocarbons: from metabolism to lung cancer. Toxicological sciences: an official journal of the Society of Toxicology. 2015;145(1):5-15.
- 219. Demetriou CA, Vineis P. Carcinogenicity of ambient air pollution: use of biomarkers, lessons learnt and future directions. Journal of thoracic disease. 2015;7(1):67-95.
- 220. Yu XJ, Yang MJ, Zhou B, Wang GZ, Huang YC, Wu LC, et al. Characterization of Somatic Mutations in Air Pollution-Related Lung Cancer. EBioMedicine. 2015;2(6):583-90.
- 221. Somers CM, Yauk CL, White PA, Parfett CL, Quinn JS. Air pollution induces heritable DNA mutations. Proceedings of the National Academy of Sciences of the United States of America. 2002;99(25):15904-7.
- 222. Sanchez-Guerra M, Zheng Y, Osorio-Yanez C, Zhong J, Chervona Y, Wang S, et al. Effects of particulate matter exposure on blood 5-hydroxymethylation: results from the Beijing truck driver air pollution study. Epigenetics. 2015;10(7):633-42.
- 223. Baccarelli A, Wright RO, Bollati V, Tarantini L, Litonjua AA, Suh HH, et al. Rapid DNA methylation changes after exposure to traffic particles. American journal of respiratory and critical care medicine. 2009;179(7):572-8.
- 224. Guo L, Byun HM, Zhong J, Motta V, Barupal J, Zheng Y, et al. Effects of short-term exposure to inhalable particulate matter on DNA methylation of tandem repeats. Environmental and molecular mutagenesis. 2014;55(4):322-35.
- 225. Tarantini L, Bonzini M, Apostoli P, Pegoraro V, Bollati V, Marinelli B, et al. Effects of particulate matter on genomic DNA methylation content and iNOS promoter methylation. Environmental health perspectives. 2009;117(2):217-22.
- 226. Clark SJ, Melki J. DNA methylation and gene silencing in cancer: which is the guilty party? Oncogene. 2002;21(35):5380-7.
- 227. Zhang W, Klinkebiel D, Barger CJ, Pandey S, Guda C, Miller A, et al. Global DNA Hypomethylation in Epithelial Ovarian Cancer: Passive Demethylation and Association with Genomic Instability. Cancers. 2020;12(3).
- 228. Deben C, Van den Bossche J, Van Der Steen N, Lardon F, Wouters A, de Beeck KO, et al. Deep sequencing of the TP53 gene reveals a potential risk allele for non-small cell lung cancer and supports the negative prognostic value of TP53 variants. Tumour biology: the journal of the International Society for Oncodevelopmental Biology and Medicine. 2017;39(2):1010428317694327.
- 229. Zhou W, Tian D, He J, Wang Y, Zhang L, Cui L, et al. Repeated PM2.5 exposure inhibits BEAS-2B cell P53 expression through ROS-Akt-DNMT3B pathway-mediated promoter hypermethylation. Oncotarget. 2016;7(15):20691-703.
- 230. Heßelbach K, Kim GJ, Flemming S, Häupl T, Bonin M, Dornhof R, et al. Disease relevant modifications of the methylome and transcriptome by particulate matter (PM(2.5)) from biomass combustion. Epigenetics. 2017;12(9):779-92.
- 231. Deng X, Feng N, Zheng M, Ye X, Lin H, Yu X, et al. PM(2.5) exposure-induced autophagy is mediated by lncRNA loc146880 which also promotes the migration and invasion of lung cancer cells. Biochimica et biophysica acta General subjects. 2017;1861(2):112-25.
- 232. Zimmerman R, Cerutti P. Active oxygen acts as a promoter of transformation in mouse embryo C3H/10T1/2/C18 fibroblasts. Proceedings of the National Academy of Sciences of the United States of America. 1984;81(7):2085-7.
- 233. Halliwell B. Oxidative stress and cancer: have we moved forward? The Biochemical journal. 2007;401(1):1-11.
- 234. Turner MC, Cohen A, Jerrett M, Gapstur SM, Diver WR, Pope CA, 3rd, et al. Interactions between cigarette smoking and fine particulate matter in the Risk of Lung Cancer Mortality in Cancer Prevention Study II. American journal of epidemiology. 2014;180(12):1145-9.