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Early Exposure to Certain Antibiotics Might Promote Autism Spectrum Disorders (ASDs)—Related Dysbiosis in European Countries, Supporting Experimental Observations

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Abstract: Autism spectrum disorders (ASDs) are complex illnesses of largely unknown etiology with lifelong neurodevelopmental consequences. The global prevalence of autism has increased twentyfold to thirtyfold since the earliest epidemiologic observations were reported in the late 1960s and early 1970s. Recent reports agree on the association of ASD with the modification of the microbiome (dysbiosis), which raises the possible role of external factors. Our study aimed to identify antibiotic classes that might be associated with the prevalence of ASD in 30 European countries through the possible modification of the intestinal microbiome. Statistical comparison was made between the average yearly consumption of different antibiotic classes (1997-2020) and the number of individuals living with ASD estimated for 2023/100000 population in 30 European countries, and the results were statistically analyzed. Tetracycline (J01A) showed significant positive (promoting) association with the prevalence of ASD (Pearson r : 0.373, p : 0.043. OR: 1.312, CI95%: 0.995-1.791, p : 0.065) and narrow-spectrum, beta-lactamase resistant penicillin (J01CF) (Pearson r : 0.524, p : 0.003, OR: 3.240, CI95%: 1.710-8.853, p : 0.004, Kruskal-Wallis p : 0.032, post hoc test p : 0.027). Mild, negative (inhibitory) association was observed with broad-spectrum, beta-lactamase sensitive penicillin (J01CA) (Pearson r : -0.278, p : 0.157, OR: 0.808, CI95%: 0.649-0.957, p : 0.028) and narrow-spectrum, beta-lactamase-sensitive penicillin (J01CE) (OR: 0.725, CI95%: 0.543-0.885, p : 0.009). Our findings strongly support the animal experiments when penicillin V-exposed newborn mice developed "autism-like" behavior.

Keywords: autism spectrum disorder (ASD); microbiome; antibiotics; penicillin; tetracycline; gut-brain axis (GBA)

1. Introduction

Autism spectrum disorder (ASD), which was first described by Karner (1942), is a neurodevelopmental disease characterized by deficits in social communication and the presence of restricted interests and repetitive behaviors [1]. Its recent, pandemic-like spread is alarming, and it has become a public health issue. According to the Centers for Disease Control and Prevention (CDC) report in 2018, the ASD prevalence among children by 8 years has been estimated as 16.8/ 1000 (1:56)

with more boys (26.6/ 1000 boys) than girls (6.6/ 1000 girls), which means a 150% increase from 2000 to 2014 [2]. The World's prevalence of ASD has increased several-fold since the first epidemiological observations in the past 50 years and has reached the level of 1-2% of all children by 2000. The reasons for the increase could not be clearly stated, but researchers agree that better diagnostic criteria and some external factors might play a role [2].

The pandemic of autism is rapidly spreading, and by now, 1 in 56 children has a chance to develop autism. We are still unable to identify any suitable reason for this ailment, despite several associations with different conditions being suspected as potential causative agents [3, 4]. Genetic mechanisms are considered for approximately 10–20% of ASD cases [5, 6]. It was observed that vancomycin treatment alleviated the symptoms of ASD, suggesting a role for certain clostridial bacteria [7-9]. The abundance of *Clostridium bolte* in autism prompted speculation about the use of developing a vaccine against this pathogen [10]. *Desulfovibrio* was also suspected as an etiological agent in autism [11]. Several publications support the role of intestinal bacteria [12, 13], transvaginal ultrasonography [14], conjugate vaccines [15], and the accumulation of insulin-like growth factors [16] as causative agents. Other reasons leading to autism were also discussed in the literature [17]. A few of the latest proposal includes the role of electromagnetic frequency and radiofrequency radiation exposures (EMF/RFR) [18]. Tylenol/Augmentin and pesticide combined are also suspected [19]. New publications suggest that the peripheral immune system plays an important role in normal neuronal function [20-22].

ASD comprises a collection of neuro-developmental disorders, starting in early childhood and characterized by pervasive behavioral deficits and social interaction [23]. The International Classification of Diseases and the Diagnostic Statistical Manual state that the ASD group includes childhood autism, atypical autism, Asperger's syndrome, disintegrative disorders, and Rett syndrome. ASD is considered genetically and phenotypically as a heterogeneous group of symptoms with various severity, symptomatology, and outcome [24]. All manifestations are considered serious, devastating manifestations regarding the outcome and co-morbidity, which bear down on the affected families, society, and the health care systems alike. The overlap between autism and childhood obesity indicates that autistic patients are obese as well. Recent publications in increasing numbers indicate the role of microbiome alteration (dysbiosis) in autism. Experiments proved that fecal samples from autistic children, when injected into germ-free mice, produce similar symptoms [25, 26].

In ASD cases, the balance between the pro-inflammatory bacteria (clostridia and *desulfovibrio*) and the anti-inflammatory bacteria (bifidobacteria) was destabilized even before the symptoms of ASD occurred. This imbalance results in the so-called "leaky gut" syndrome, and through the more porous epithelial membrane, toxins produced by the microbes can easily enter the circulation, which may affect the brain and the development of ASD [27, 28]. The alteration of the intestinal microbial taxa in ASD cases is repeatedly reported as the reduction of Firmicutes and an increase in the number of Bacteroidetes [29].

Other investigators observed the relative abundance of other bacteria, like clostridia, *Caloramator*, *Alistipes*, *Sarcina*, *Akkermansia*, *Lactobacillus*, enterobacteriaceae, and sutterellaceae [30-34], and the reduction in the abundance of Bifidobacteria, *Desulfovibrio*, *Coprococcus*, veillonellaceae, and prevotellaare species [8]. The pathogenetic process of ASD might be associated with the overgrowth or the reduction of different species, like *Clostridium*, which develops particularly with the use of antibiotics [11, 35-39].

The presence of an altered microbiome (dysbiosis) in ASD is well documented, and its possible role in the development of ASD has also been reported (see above references). It has been hypothesized that antibiotic exposure to the microbiome might produce "ASD-promoting" dysbiosis in the mother or the early modification of the gut microbiome in infancy, which might facilitate the development of ASD through the action of different mediator molecules involving the gut-brain axis (GBA) [40].

In utero exposure to the anticonvulsant valproic acid (VPA) in mice leads to developmental and behavioral deficits in offspring that are similar to ASD [41]. The role of two microbiota-derived host metabolites, p-cresol sulfate and 4-ethylphenyl sulfate, has been reported to contribute to the development of ASD in patients and animal experiments alike. They are produced by certain microbial taxa, such as p-cresol and 4-ethylphenol, which are generated through aromatic acid fermentation by several commensal bacteria, most frequently by the *Clostridioides* genus, which is often detected in the microbiome of ASD patients. The metabolites, after entering the bloodstream, cross the blood-brain barrier and enter the central nervous system, probably through the microglial cells. They affect neuroinflammation and microglial phagocytosis [42].

Epidemiological studies indicated that early-life antibiotic exposure can augment the risk of neurodevelopmental disorders later in life [43, 44].

We aimed to identify the use of antibiotic classes that might show a statistical association with the prevalence of ASD in 30 European countries, probably initiating ASD-related dysbiosis and hence triggering the development of ASD.

2. Hypothesis

The average consumption of systemic antibiotics in the community (1997-2020) varies between 30.474 DID (Defined Daily Dose/ 1000 Inhabitants/ Day) in Greece, and 9.218 DID in the Netherlands (Table 1.), which indicates an over 3-fold difference in the consumption. Similarly, the relative share of the different antibiotic classes used in the European countries of the total antibiotic consumption in the community (J01) is very different. The highest relative share of tetracycline (J01CA) consumption is recorded in the UK (26.259%), while the lowest is in Spain (2.407%), indicating an over 10-fold difference. Similarly, the highest consumption share of beta-lactamase-resistant penicillin (J01F) is the highest in Sweden (9.970%), and the lowest is in Greece (0.013%), which indicates a 766.9-fold difference. Different classes of antibiotics might induce different dysbiosis and trigger the production of diverse molecular products, which are leaking into the circulation and might “augment” or “inhibit” the development of ASD. In this context, “augmentation” means that higher consumption of a given class of antibiotics might statistically positively correlate with the prevalence of a certain disease (e.g., ASD), and “inhibition” means that the relatively higher consumption of a certain class of antibiotics is related to lower prevalence.

We have aimed to identify antibiotic classes that might “augment” or “inhibit” the development of ASD-related dysbiosis and hence the prevalence of ASD.

3. Results

Tetracycline (J01A) usage indicated a significant positive (promoting) association with the prevalence of ASD (Pearson r : 0.373, p : 0.043. OR: 1.312, CI95%: 0.995-1.791, p : 0.065) and narrow-spectrum, beta-lactamase resistant penicillin (J01CF) (Pearson r : 0.524, p : 0.003, OR: 3.240, CI95%: 1.710-8.853, p : 0.004, Kruskal-Wallis p : 0.032). Mild, negative (inhibitory) association was observed with broad-spectrum, beta-lactamase sensitive penicillin (J01CA) (Pearson r : -0.278, p : 0.157, OR: 0.808, CI95%: 0.649-0.957, p : 0.028) and narrow-spectrum, beta-lactamase-sensitive penicillin (J01CE) (OR: 0.725, CI95%: 0.543-0.885, p : 0.009). Our comparative analyses indicated a significant, positive correlation between the consumption of tetracycline and the prevalence of ASD (Fig. 1-2). The highest rate of ASD and the highest tetracycline consumption, together with the second highest consumption of narrow-spectrum, beta-lactamase-resistant penicillin (J01CF), is reported from the UK. Similarly, Sweden records the highest consumption of narrow-spectrum, beta-lactamase-resistant penicillin (J01CF), the fifth place on the tetracycline consumption rank order, and second place on the ASD prevalence list (Table 1).

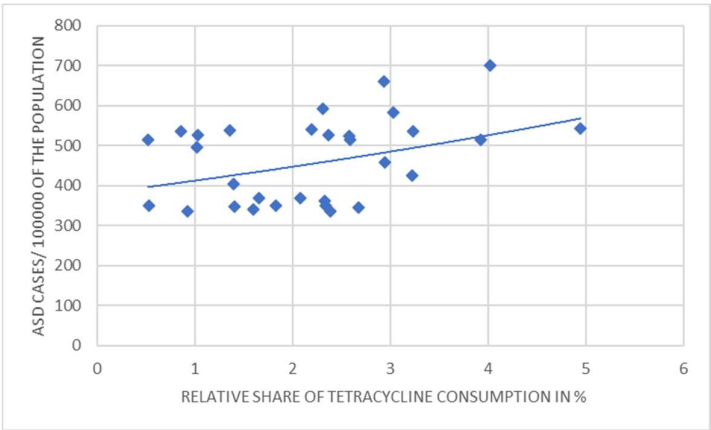


Figure 1. A significant positive association was observed between ASD prevalence and the consumption of tetracycline.

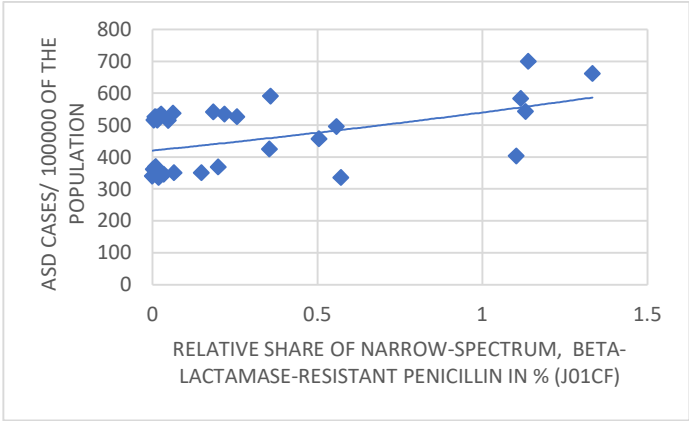


Figure 2. A significant, positive association was observed between ASD prevalence and the consumption of narrow-spectrum, beta-lactamase-resistant penicillin (J01CF).

Table 1. Average antibiotic consumption in 30 European countries between 1997-2020 expressed as the relative share (%) of the total consumption of systemic antibiotics in the community (expressed in DID) and the rank order of ASD prevalence/100000 inhabitants. Significant correlations are marked with a yellow filling color. Non-significant correlations are marked with a blue filling color.

Glossary for ATC groups: J01: antibiotics for systemic use, J01A: tetracycline, J01C: penicillin group, J01CA: broad spectrum, beta-lactamase sensitive penicillin, J01CE: narrow spectrum, beta-lactamase sensitive penicillin, J01CF: narrow spectrum, beta lactamase-resistant penicillin, J01CR: broad spectrum, beta-lactamase resistant penicillin, J01D: cephalosporin, J01F: macrolides, lincoseamid and streptogramin, J01M: quinolone

Average antibiotic consumption 1997-2020	J01 (DID)	J01A%	J01C%	J01CA%	J01CE%	J01CF%	J01CR%	J01D%	J01F%	J01M%	People li with A/ 2023/100 populat
Countries											
UK	15.294	26.259	38.257	21.296	4.878	7.447	4.799	3.570	17.177	3.492	700.070
Sweden	13.370	21.960	47.771	7.831	28.579	9.970	1.361	2.094	5.460	6.171	661.850
Netherlands	9.218	24.995	32.046	13.734	3.873	3.884	10.545	0.966	15.003	9.069	591.540
Ireland	18.256	16.608	45.021	15.425	5.094	6.113	18.482	8.600	18.695	4.700	583.690
Iceland	19.148	25.804	48.146	17.715	12.774	5.907	11.657	2.961	8.147	4.162	543.420
Luxembourg	22.410	9.768	35.582	13.788	0.393	0.826	20.607	18.237	17.840	10.768	541.660
Malta	18.465	7.344	33.929	2.908	0.498	0.336	30.176	21.847	20.422	11.373	537.950
Cyprus	27.338	11.800	34.582	11.405	0.369	0.095	22.803	21.666	11.435	16.911	535.350
Spain	17.697	4.826	51.794	20.405	0.571	1.232	29.011	11.804	14.341	13.330	535.140
Belgium	21.427	11.051	40.720	17.716	0.425	1.195	21.366	11.140	14.958	10.295	526.130
Austria	11.683	8.816	36.386	7.010	8.097	0.068	21.176	13.396	26.329	11.221	526.020
Germany	12.610	20.412	27.113	16.122	8.438	0.111	2.395	15.131	18.184	9.556	525.310
Italy	21.524	2.407	42.241	16.303	0.060	0.070	25.832	13.171	21.957	14.277	516.090
Greece	30.474	8.470	28.152	13.415	1.447	0.013	13.185	23.571	26.337	8.814	515.700
Finland	16.157	24.293	28.217	14.539	9.371	0.297	4.023	12.960	9.123	4.945	514.680
Portugal	18.094	5.610	43.307	11.087	0.155	3.078	29.004	12.756	17.569	13.795	496.610
Norway	15.046	19.567	40.635	12.754	24.465	3.356	0.060	0.957	10.415	3.110	457.900
France	24.478	13.171	46.666	27.796	0.735	1.446	16.615	11.512	16.697	7.856	425.410
Denmark	14.068	9.937	62.425	19.420	32.521	7.840	2.687	0.213	14.437	3.234	403.840
Estonia	10.947	18.928	33.808	22.125	2.412	0.091	9.199	8.148	17.786	7.582	369.490
Lithuania	16.714	9.908	52.315	30.531	13.312	1.191	7.341	7.784	10.452	5.881	369.320
Latvia	10.653	21.862	37.924	25.505	0.901	0.019	11.508	5.107	13.470	9.218	362.530
Slovenia	12.873	4.109	55.543	16.686	15.397	1.150	22.380	4.350	19.125	9.827	351.370
Czech Rep.	14.685	15.887	37.719	9.227	12.782	0.443	15.131	10.555	20.620	7.014	350.850
Slovakia	21.098	8.660	39.241	10.579	13.366	0.081	15.253	17.082	21.955	8.731	349.180
Croatia	17.911	7.839	42.326	13.774	5.622	0.190	22.746	17.710	14.745	8.352	346.790
Poland	18.773	14.228	32.952	19.459	2.243	0.117	11.144	12.332	17.823	6.797	345.130
Hungary	14.711	10.842	35.817	9.972	4.310	0.000	21.508	14.696	20.760	12.528	340.310
Romania	22.897	4.044	46.840	17.745	3.035	2.494	23.925	18.697	12.517	12.901	335.890
Bulgaria	17.828	13.383	36.740	22.655	5.346	0.107	8.750	14.988	14.522	11.168	335.580
RESULTS											
Pearson R	0.039	0.373	-0.146	-0.278	-0.032	0.524	-0.078	-0.157	-0.121	-0.089	
Pearson p	0.839	0.043	0.442	0.137	0.865	0.003	0.682	0.408	0.523	0.640	
OR	1.077	1.312	1.131	0.808	0.725	3.240	0.892	1.073	1.063	1.296	
CI95%	0.922 - 1.259	0.995 - 1.791	0.934 - 1.395	0.649 - 0.957	0.543 - 0.885	1.710 - 8.852	0.747 - 1.029	0.859 - 1.354	0.868 - 1.308	0.954 - 1.811	
p	0.348	0.065	0.221	0.028	0.009	0.004	0.154	0.534	0.554	0.107	
Kruskal-Wallis p	0.679	0.342	0.606	0.668	0.278	0.032	0.481	0.733	0.447	0.903	
Post hoc											
*Group 1-2	0.775	0.636	0.684	0.951	0.818	0.165	0.493	0.924	0.448	0.924	
*Group 1-3	0.684	0.332	0.924	0.636	0.191	0.027	0.972	0.73	0.636	0.988	
*Group 2-3	0.997	0.857	0.684	0.857	0.775	0.857	0.636	0.893	0.951	0.924	

*Group 1 contains the countries with the 10 highest prevalence of ASD, Group 2 is the middle, and Group 3 includes the countries with the 10 lowest prevalence of ASD

4. Discussion

An increasing trend of neurodevelopmental diseases, including ASD, attention-deficit/hyperactivity disorder (ADHD), and learning disabilities, has been reported in the past few decades. Early life is crucial for the appropriate neurodevelopmental processes, but external/internal agents might interfere with it. It has been reported that the incidence of ASD increased significantly over time, especially among toddlers and preschool children, but also in older age groups [45]. Recent publications (cited above) agree on the principal role of certain gut bacteria in the development of autism, particularly of clostridia species. This theory was supported by the fact that vancomycin ameliorated the symptoms of autism. Still, it returned after the cessation of antibiotic treatment, probably indicating that after temporarily suppressing the clostridial species, the appropriate balance of the gut flora was not restored [46]. A large cohort study including over half a million newborn babies reported that among those who were exposed to prenatal (maternal) antibiotics (29.8%), ASD

developed in 1.5% of the observed children, which indicated a slightly higher risk for developing ASD in the perinatal antibiotic exposure group, particularly in the first and second trimester [47]. The intrauterine development of the brain is a critical period and is particularly sensitive to external stimuli coming from the surrounding environment, which interfere with the normal development of the central nervous system (CNS). The alteration of the prenatal environment, particularly the exposure to maternal drug intake, like antibiotics, might trigger neurodevelopmental changes, which lead to the development of neurodevelopmental disorders (NDDs). To prevent the most frequent streptococcal infections in pregnancy, antibiotics, particularly penicillin, are prescribed as prophylactic, which might interfere with the intrauterine development of the CNS [48]. The role of the microbial taxa composition populating the intestine and acting through the gut-brain axis (GBA) is not fully elucidated and probably includes several factors and mechanisms. The microbial colonization of the fetal/newborn intestine mainly originates from the mother. Antibiotics prescribed as prevention for maternal infections are mostly penicillin against suspected streptococcal infections, which might change the composition of the maternal microbial flora along with the milk composition when antibiotics are used during lactation and influence the microbial flora of the newborns [49]. Epidemiological studies indicated the association between maternal infections during pregnancy and the probability of developing NDDs in the offspring [50]. Infections of the newborn treated with antibiotics can induce long-lasting modification of brain development, probably through the GBA [51, 52]. The previous use of antibiotics in children with NDDs was also reported [53]. Antibiotic consumption in childhood and adolescence showed association with mental disturbances in adults, particularly schizophrenia and affective disorders, which may be mediated by effects of infections/inflammation on the brain, triggered by the alterations of the microbial taxa, genetics, or other factors [54]. It has been reported that children receiving antibiotics in their first year of life had more behavioral difficulties and more symptoms of depression at follow-up [55]. Similarly, ADHD risk increase was observed in those exposed to 4 or more antibiotic courses or a duration longer than 3 weeks in their first year of life [56]. Twin studies from the Netherlands and Sweden indicated that the use of antibiotics during the early life of 0 and 2 years increased the risk of ADHD and ASD; however, the importance of the familial environment and the genetic influence in the etiology of NDDs also must be considered [57]. A major meta-analysis on the association between early exposure to antibiotics and ASD did not indicate any significant increase in ASD in the antibiotic exposure group [58], but they did not separate the antibiotic classes. It might be concluded that antibiotics, as major disruptors of the microbiome, can change the composition and the abundance of different microbial taxa and, hence, the composition of the microbiome-derived molecules as well. The altered molecules, acting through the GBA, interfere with the normal neurodevelopmental mechanisms, ending up in neurodevelopmental abnormalities, including ASD. Several reports indicate that the gut microbiota plays a crucial role in the bidirectional communication between the gut and the brain suggesting that the gut microbial taxa may influence neural development, and neurotransmission, which affect behavior, contributing to the pathogenesis and/or progression of many neurodevelopmental, neuropsychiatric, and neurological conditions [59]. A recent review of the literature (2021) summarizes the role of the microbiome in different diseases like depression, anxiety, schizophrenia, autism spectrum disorders, Parkinson's disease, migraine, and epilepsy [59]. Alteration of the microbiome is followed by the appropriate response from the CNS. Signals, arising from the microbiome as mediator molecules produced by different microbial taxa, are transmitted via the afferent spinal and vagal sensory nerves, and they include polysynaptic entrances to higher areas of the CNS, such as the hypothalamus and limbic forebrain (60). In an experimental model, when germ-free mice have been colonized with a microbiome from ASD individuals, the hallmarks of autistic behaviors developed, and the brains of mice colonized with ASD microbiota display alternative splicing of ASD-relevant genes. It appears that the gut microbiota regulates behaviors in mice via the production of neuroactive metabolites, suggesting that gut-brain connections contribute to the pathophysiology of ASD [61].

The ASD-related altered microbiome, when transferred to mice (fecal microbial transfer, FMT), is capable of inducing ASD-like behavior changes in the experimental animals [62]. Similarly, fecal microbial transfer (FMT) has dramatically ameliorated the symptoms of ASD [62-65]. A study by Kang et al. applied extensive FMT treatment to 18 autistic children of 7-16 years who were diagnosed by the Autism Diagnostic Interview-Revised (ADI-R) with moderate to severe gastrointestinal problems. After maintaining the FMT treatment for 8 weeks, they observed significant amelioration of the ASD-related symptoms, including gastroenteric symptoms as well [66]. Sporadic observations indicated that extended antibiotic treatment could ameliorate the symptoms of autism, probably suppressing the proliferation of ASD-related microbial taxa. According to a report, five autistic children who were suffering from Lyme disease were treated with amoxicillin and azithromycin, and the symptoms of ASD were ameliorated [67]. A case report indicated the rapid improvement of ASD after introducing vancomycin treatment, which returned after stopping the use of vancomycin [68]. In an animal model (male Wistar rats), ASD-like behavioral symptoms had been induced either by using an antibiotic cocktail or valproic acid (VPA). They observed that by adding a probiotic mixture to the antibiotic-treated group and the VPA-treated group, the symptoms of ASD-like behavior attenuated both the antibiotics and the VPA-generated antisocial behavioral symptoms [69]. The initial development and maturation of the neonatal microbiome are largely determined by maternal-offspring exchanges of microbiota, which indicates that a dysbiotic maternal microbiome might also populate the newborn intestine [70]. Several researchers consider the importance of biochemical changes observed early in the newborn, such as the depressed level of insulin-like growth factor-1 (IGF-1) in the neurodevelopmental period, in the development of ASD. This observation leads to early diagnosis and probable intervention in the prevention of dysconnectivity. According to Steinmann [71], the process of developing ASD could be preventable even before irreversible psychosocial changes develop. It might be concluded that the biochemical changes observed in ASD are probably triggered by the altered microbiome induced by certain antibiotics. Reports on the association between autism and antibiotics are controversial. Some observations indicate a specific association between prenatal exposure to antibiotics and autism [72], while others found some beneficial effects of perinatal antibiotic use [73]. Controversy might arise from the fact that different antibiotic classes induce different dysbiosis, which either promotes or inhibits the development of ASD-related dysbiosis. In animal experiments, when newborn mice were exposed to low-dose penicillin, profound changes in the intestinal microbiota were observed, which might be implicated in the perturbation of neurodevelopmental and neuropsychiatric pathways. Significant effects were observed in different areas of the brain (frontal cortex, amygdala gene, etc.). Linkage was observed between the specific microbial taxa and the early-life expression of particularly affected genes [74]. The results derived from animal models [75] strongly support our observations, as we have detected the possible association between the consumption of the narrow-spectrum, beta-lactamase-resistant penicillin (J01CF) and the prevalence of autism, together with tetracycline (J01A), which might induce developmental difficulties in the brain, resulting in ASD. Group B Streptococcus (GBS) remains the most common cause of neonatal early-onset sepsis among term infants and a significant cause of late-onset sepsis among both term and preterm infants, and penicillin is frequently used for prophylaxis [76]. As far as pregnant women, newborn babies, and infants are not taking any tetracycline compound because of its contraindication, it could be suspected that tetracycline might have arrived from environmental antibiotic pollution or the women were exposed to tetracycline before being pregnant, which triggered the ASD-related dysbiosis. Tetracycline is widely used in humans and agriculture, mixed with animal fodder. More than 70% of the drug is excreted in an active form in the urine and feces and pollutes the environment. Due to its overuse and weak ability to degrade, it has become a serious threat to the environment. Tetracycline can accumulate in the food chain, and it can influence the composition of the human microbial taxa [77]. It might be concluded that higher consumption could result in higher environmental pollution. Several reports indicate that the risk of developing ASD is associated with ongoing inflammatory changes during pregnancy, which are followed by the alteration of the gut flora with the capability to induce T17 cells and promote the risk

for neurodevelopmental syndromes in the offspring of pregnant mothers with infections or auto-inflammatory disorders [78-82]. The gut microbiota permanently interacts with the immune system. Children suffering from ASD experience GI symptoms, an imbalance observed in a propensity to impaired gut barrier function, which may contribute to their symptoms and clinical outcome [83]. Several researchers have observed immune dysregulation in ASD cases by modifying circulating and brain cytokines, chemokines, and several other inflammatory molecules. The abnormal distribution of different leukocyte subtypes, like inflammatory cytokines (i.e., interferon (IFN)- γ , IL-1 β , IL-6, IL-12, tumor necrosis factor (TNF)- α), has been observed, which might be associated with the symptoms and severity of the ASD [84]. The microbiome's bacterial composition influences the nerve cells' development from the fetus to adulthood as it affects neural stem cell differentiation, the migration of immature neurons, axons, dendritic growth, and synapse formation [85]. The observation of the altered microbiome in ASD patients has prompted researchers to focus attention on reconstructing "normal" microbiomes with diet, antibiotics, prebiotics, probiotics, and FMT as therapeutic approaches [86]. Those attempts with prebiotics, probiotics, and symbiotics have yielded some results in ameliorating the symptoms of ASD by modifying the gut flora and the metabolic activity of children with ASD. Probiotic treatment increases the abundance of *Lactobacillus*, and prebiotic treatment augments the relative abundance of *Bifidobacterium*, but decreases the abundance of *Lachnospirillum*. According to reports, changes in microbial metabolism were observed with the increasing short-chain fatty acid and reduced ammonium level [86]. According to another meta-analysis of the literature dealing with the possible beneficial effects of probiotics, the beneficial effects of probiotics are not confirmed. Combining prebiotics and symbiotic treatment has demonstrated some benefits for behavioral abnormalities, but the most promising intervention is FMT, even if the reports are scarce [87, 88]. A prominent autism stool metagenomics analysis, including stool samples from 247 children (99 with autism), found limited direct autism associations. Only *Romboutsia timonensis* was the specific taxa associated with autism diagnosis. This Australian metagenomics study reported negligible direct associations between ASD diagnosis and the gut microbiome. It indicated that ASD-related restricted interests are associated with a less-diverse diet, which results in reduced microbial taxonomic diversity and looser stool consistency. The study concluded that microbiome differences in ASD may reflect dietary preferences that are related to diagnostic features, and the microbial diversity itself could not be considered a driving force in the development of ASD [89]. Fecal samples from autistic children showed a higher abundance of *Roseburia* and *Candida* genera, and lower abundance of *Dialister*, *Bilophila*, *Veillonella*, *Streptococcus*, *Coprococcus*, and *Prevotella* genera (90).

5. Materials and Methods

To evaluate the above hypothesis, antibiotic consumption pattern databases were compared to the autism (ASD) prevalence in countries with people living with autism/ 100000 population in 30 European countries estimated for 2023 (<https://www.theworlddrinking.com/statistics/178/global-comparison-of-autism-rates/>). (Accessed on 30. Jan. 2025)

Average yearly antibiotic consumption was calculated from the publicly available antibiotic databases of 30 European countries (ECDC yearly reports) for 1997-2020 (accessed: 12 01 2023).

<https://www.ecdc.europa.eu/en/antimicrobial-consumption/database/qualityindicators>

Based on the Anatomical Therapeutic Chemical classification system (ATC), the average yearly consumption of the total systemic antibiotics (J01) has been calculated and expressed in Defined Daily Dose/1000 Inhabitants/ Day (DID). The consumption of major antibiotic classes covering 94% of antibiotic use in the community at ATC levels two and three was calculated as a relative share of the total amount of systemic antibiotics (J01) and expressed in percentage (%). Antibiotic classes included: tetracycline (J01A), penicillin (J01C), broad-spectrum, beta-lactamase sensitive penicillin (J01CA), narrow spectrum, beta-lactamase sensitive penicillin (J01CE), narrow spectrum, beta-lactamase resistant penicillin (J01CF), broad-spectrum, beta-lactamase resistant combination

penicillin (J01CR), cephalosporin (J01D), macrolide and lincosamides, streptogramins (J01F), and quinolone (J01M) expressed in percentage of the total amount.

Statistics: Pearson's correlation was used to estimate the correlation between antibiotic consumption and the prevalence of ASD. A significant correlation (positive/negative) was considered when p values were ≤ 0.05 . A non-significant correlation was estimated when the p -values fell between 0.051 and 0.09. Positive (supportive) and negative (inhibiting) significant correlations were considered and evaluated. Logistic regression analysis was performed to determine the odds ratio (OR, CI 95%) for each antibiotic class. To confirm our results, we have performed the more advanced Kruskal-Wallis test (non-parametric version of ANOVA analysis) to determine the statistically significant differences among the three groups formed by country ranking. Group one contained the countries with the 10 highest ranking ASD prevalence, the second group contained the middle 10 countries and the third group contained the 10 countries with the lowest prevalence of ASD. Post hoc multiple testing was performed comparing the three groups of country ranking. Statistical results were recorded and featured in Table 1. Scatter diagrams were plotted to demonstrate the association (positive/negative) between the prevalence of autism and the average usage of narrow-spectrum, beta-lactamase resistant penicillin (J01CF) and tetracycline (J01A), (Figure 1-2).

6. Conclusions

There is a considerable body of knowledge regarding the association between ASD and early exposure (maternal/fetal, newborn) to antibiotics. Human observational studies [91] and animal experiments [92] indicated that narrow-spectrum penicillin produces perturbation of the microbiome with the reduction of *Lactobacillus* and the proliferation of non-*E. coli* gram-negative species. Our comparative analyses of antibiotic consumption in 30 European countries indicated that certain classes of antibiotics, like tetracycline (J01A) and methicillin-resistant narrow-spectrum penicillin (J01CF), might alter the microbiome in newborns or the maternal microbiome during pregnancy, which favors the development of ASD. Animal models (mice) indicated that applying a low dose of Penicillin V induced a more robust behavioral abnormalities response than a cocktail of broad-spectrum antibiotics [93]. Tetracycline reduces the abundance of *Bacteroides fragilis*, bifidobacteria, lactobacilli, and enterococcus spp., and increases the abundance of lactose-fermenting enterobacteriaceae. A Swedish population study [94] indicated that penicillin is the most frequently prescribed antibiotic for pregnant women and infants. A significant association was observed between the prevalence of autism and antibiotic exposure during pregnancy and early childhood. This observation is in accord with the high consumption ratio of penicillin and the high prevalence of autism [95]. A large meta-analysis of the literature [96] on the association of maternal antibiotic exposure and the attention-deficit/hyperactivity disorder of children indicated a significant relationship. Still, no similar association was observed with post-partum antibiotic intake. The authors emphasize the heterogeneity of the publications included in the study. Other large cohort studies did not observe any association between antibiotic use and autism [97], although they did not separate the use of different classes of antibiotics and the relationship with the development of autism. Because several external exposures are considered possible factors in the development of ASD, including dysbiosis [98], convincing evidence emerged in the relevant literature supporting the crucial role of an altered microbiome [99], which might develop as a result of antibiotic exposure. Reports using FMT in ASD patients have found that oral administration of lyophilized FMT is an effective and safe treatment for children with ASD. FMT could improve GI, ASD related symptoms, and sleep disturbances, and alter the gut bacterial and fungal microbiota composition in children with ASD. A more extensive, updated publication on the burden of ASD is presented in a recent publication [100].

Limitations of our study: Our results could not be interpreted at the individual level; only statistical correlations and concordance could be established. However, a firm statistical correlation between tetracycline and penicillin consumption and ASD was estimated. As shown in the scatter

diagrams, this might indicate a possible association between maternal tetracycline/penicillin (dysbiosis) consumption and autism.

Strength of our study: To our knowledge, our study is the first one comparing the consumption of different antibiotic classes and the prevalence of ASD in European countries. The results of our comparative analysis are in full accord with previous reports and animal experiments indicating the possible role of certain antibiotics, even low-dose penicillin applied to intrauterine, or newborn mice, in the development of autism-like behavior.

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