

Review

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Review

A Systematic Review of Marine Habitat Mapping in the Central-Eastern Atlantic Archipelagos: Methodologies, Current Trends, and Knowledge Gaps

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Abstract: Mapping marine habitats is fundamental for biodiversity conservation and ecosystem-based management in oceanic regions facing increasing anthropogenic and climatic pressures. Currently, with evolving Marine Spatial Planning processes in different countries, habitat mapping is a keystone element for conservation policy. This systematic review (PRISMA guidelines) examined 69 peer-reviewed studies across the Central-Eastern Atlantic archipelagos of Azores, Madeira, Canary Islands and Cabo Verde, including Mid-Atlantic Ridge sectors. We identified knowledge-gaps, methodological trends, and key challenges in habitat mapping, emphasizing the integration of cartographic, ecological, and technological perspectives. The analysis revealed a diversification of mapping methodologies; however, the lack of standardized protocols, heterogeneous datasets, and limited ground-truthing undermine the production of high-resolution bionomic maps across the region. Additionally, we found substantial regional disparities in mapping efforts, technological access, and habitat representation. Azores showed the highest taxonomic richness (393 species), dominated by acoustic surveys for deep-sea corals. Madeira advanced in remote mapping of rhodoliths and hydrozoans; studies in Canary Islands focused mainly on shallow macrophyte beds via direct observation whereas Cabo Verde remains underrepresented, with sporadic studies on corals under abiotic-stress. This review also highlights the need for harmonized frameworks, investment in technological infrastructure, and regional cooperation to improve data interoperability and modeling accuracy.

Keywords: *habitat mapping; coastal ecosystems; remote sensing; rhodolith beds; macrophyte beds; corals*

1. Introduction

Continental shelves provide a diverse network of habitats that support numerous species [1–3] and hold significant ecological, economic, and social values. Their importance is especially demonstrated through key ecosystem services, such as blue carbon sequestration [4–6], habitat formation [7–9], or preservation of fishing grounds and cultural activities [10–14]. These ecosystem services arise from various key habitats underpinned by “ecosystem engineers”, which not only provide resources and services, but also modify local conditions by creating, maintaining, or transforming habitats [15–17]. The complexity of these habitats, related to their three-dimensionality, diversity, and arrangement of physical elements (rocks, crevices, organisms, etc.), largely determines the increased diversity and density of species [18–22]. Consequently, habitat complexity is considered decisive in the structure and function of biological communities [23,24].

Key habitats include macrophyte beds and coral reefs [25–29]. Macrophyte beds encompass various habitats (e.g., macroalgal forests, rhodolith and seagrass seabeds), which function as submerged “forests”, offering protection and food for invertebrates and fish [30–32]. These habitats are known for their high primary production, biodiversity increases through “facilitation cascades” [14,16,33–35] and their role in carbon sequestration [3,16,17,36–38]. Particularly, rhodolith beds, consist of free-living, calcified red algae forming mobile nodules, creating unique habitats with high structural complexity over soft bare substrates. Seagrass meadows can also play a crucial role in sediment retention, juvenile fish habitat provision, and carbon fixation (e.g., blue carbon) [16,17,36]. Coral reefs, found in both shallow and deep waters [39–43], form biogenic structures that provide shelter and breeding grounds for invertebrates and fish, further enhancing biodiversity [16,17,44].

The high added value of these habitats, coupled with increasing pressures and threats, highlights the need to protect marine biodiversity while preserving essential ecosystem services [25,45–49]. Contemporary environmental management methodologies are shifting toward an ecosystem-based approach [50–53] that integrates all interactions related to the functioning of marine ecosystems -including anthropogenic- rather than addressing them in isolation [54–56]. The implementation of this marine spatial management framework requires understanding multiple factors, such as ecosystem functioning and former inter- and intraspecific interactions [57–60]. In this context, accurate data on the distribution and extent of marine habitats is essential [61–64]. Effective management and conservation strategies rely on baseline geographic information, including habitat distribution patterns, connectivity, associated biodiversity [46,65–67], and the impacts of different pressures [3,68].

Habitat mapping is a crucial first step in the sustainable management of the marine environments, a field that remains largely unexplored due to financial, logistical, and technical challenges [3,46,56]. Traditionally, habitat assessment relied on direct sampling and observations [69,70]. However, advancements in remote sensing technologies, such as acoustic, satellite, and optical methods [30,50,63,86,87], allow for the characterization of physical and biological features over large areas with reduced logistical and economic efforts [73–78]. Consequently, habitat mapping underpins essential baseline data for effective coastal management and conservation policies, though limited ecological knowledge of many marine habitats continues to hinder management effectiveness [29,59,66,69,79].

The Lusitanian biogeographical Province [80], known for its rich biodiversity, faces significant environmental pressures from climate change, tourism overdevelopment, coastal exploitation, and invasive species [81,82]. Specially, coastal economic and urban development intensifies anthropogenic pressures, as a large portion of the global population resides and/or works in these areas [83–87]. These pressures are even more pronounced in insular territories, where coastal zones constitute a substantial part of the total land area [83,88], such as the case of the Central-Eastern Atlantic volcanic archipelagos of the Azores, Madeira, Canary Islands, and Cabo Verde [44,89,90]. As insular systems, they exhibit high isolation shaping their marine biota [90,91]. These unique characteristics make them highly relevant in terms of their biogeography, ecology, evolution, and conservation biology, as they serve as natural laboratories for studying evolutionary and ecological processes [89,92,93]. However, their vulnerability underscores the importance of studying these islands to ensure their unique marine ecosystems, which requires detailed knowledge of marine habitat distribution and characteristics, highlighting the importance of advancements in seabed mapping [63,89,94–96].

This study aims to provide a comprehensive review of the current knowledge and gaps in mapping key habitats across the Central-Eastern Atlantic volcanic archipelagos. To achieve this, available methodologies mapping tools and methodologies were cataloged, their chronological use examined, and the challenges associated with their application across different habitats and depths analyzed. The review also identified knowledge gaps, such as understudied geographical areas and key habitats—both in terms of habitat type and taxonomy—highlighting regions with data scarcity that could hinder conservation efforts. Finally, recommendations were proposed to strengthen

research initiatives and enhance the development of useful habitat mapping methodologies. In this sense, international cooperation will foster more effective decision-making processes in the framework of evolving Marine Spatial Planning activities, where marine conservation sector deserves a more intense focus.

2. Materials and Methods

This systematic review was conducted following the guidelines of the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA), to ensure transparency, reproducibility, and comprehensiveness at each stage of the process, from sourcing and selection to the analysis of the selected articles [97,98].

2.1. Study Area

The study focuses on the four volcanic archipelagos located in the Central-Eastern Atlantic Ocean: Azores, Madeira, Canary Islands, and Cabo Verde. A section of the Mid-Atlantic Ridge within the Azores exclusive economic zone (EEZ) was also considered, as it plays a crucial role in ocean current dynamics and ecological connectivity among the four archipelagos (Figure 1; [99]).

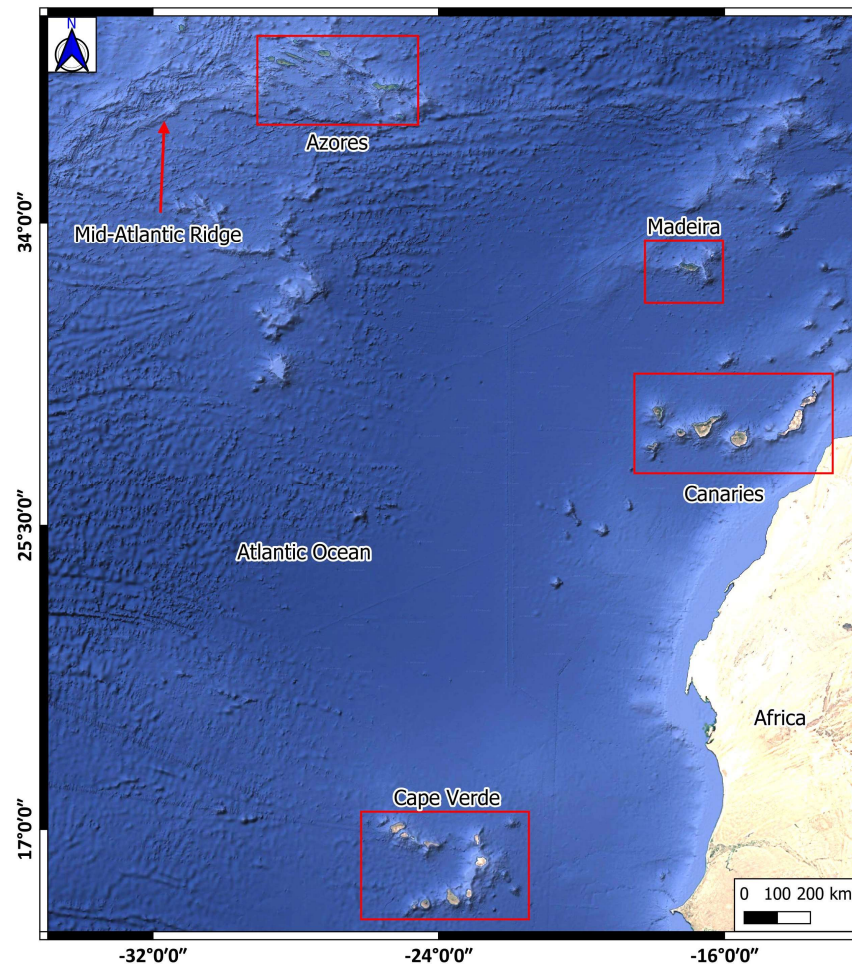


Figure 1. Location of the study area in the Central-Eastern Atlantic Ocean, showing the study areas: (a) Azores and the adjacent Mid-Atlantic Ridge, (b) Madeira archipelago, (c) Canary Islands, and (d) Cabo Verde.

The marine study area covers the so-called European Macaronesian region [100]; the term Macaronesia (composes by two Greek words: *makarion* [fortunate] and *neosi* [islands], have being used in the biogeographic literature to include the four archipelagos located in the Central-East North

Atlantic Ocean: Azores, Madeira (including the Selvagens Isles), the Canary Islands and Cabo Verde. These oceanic archipelagos have a volcanic origin, as geological demonstrations of hotspots off Northwestern Africa with large depths found between adjacent islands. The biogeographic position of these European archipelagos encompasses a large macroclimatic gradient among them of almost 2500 km of latitudinal extension, from hyperoceanic temperate conditions in the Azores to Mediterranean to subtropical ecosystems in the case of Madeira, Selvagens, and the Canaries [101,102]. Their marine environments are mainly connected through the Canary Current, a branch of the Gulf Stream in the Eastern Atlantic moving down from the Azores through Madeira to the Canary Islands and further south to the Cape Verde Islands [103]. Recent biogeographical studies have shown a more isolated biogeographical position in the case of the Azores, whereas the marine biota of the three central archipelagos: Madeira, Selvagens and the Canary Islands showed stronger affinities among them [104,105]; as consequence, these three archipelagos defined a new biogeographical unit (= *Webbnesia*) and the marine biota of the southernmost archipelago of Cabo Verde is considered as part of the West African Transition province [106].

Aside from their common volcanic origin, these archipelagos exhibit a high degree of endemism in the terrestrial realm, underscoring their worldwide biogeographic significance [99,107], and including marked differences in both terrestrial and coastal habitats. The Azores, Madeira and Canary Islands, located at more northerly latitudes, display temperate and subtropical oceanic climates alongside rugged coastlines featuring rocky reefs that support a range of benthic communities. Their coasts range from volcanic cliffs to sandy beaches and emergent rock outcrops, which foster highly diverse marine habitats encompassing seagrass beds, macroalgal assemblages on rocky reefs and cold-water coral reefs (CWC) [99,108,109]. Conversely, Cabo Verde is situated in a dry subtropical zone, characterized by a warmer and more arid climate, where marine ecosystems are dominated by sedimentary seabeds (sandy and muddy), followed by rocky and coral reefs, and whose biota differs in composition from that of the more northerly islands [110].

2.2. Data Sources and Search Strategies

The literature collection was conducted in two primary databases, SciVerse Scopus and ISI Web of Science, with Google Scholar as a supplementary source. These platforms were selected for their extensive coverage and ability to capture relevant publications on cartography and mapping in the different oceanic archipelagos of the Central-Eastern Atlantic Ocean.

The search strategy was carefully designed and executed in December 2024. A Boolean sequence of search terms was developed, using synonyms and related terms aligned with the main objective of the study. As shown in Table 1, each column represented a set of similar terms combined using the “OR” logic, while different columns were linked using the “AND” logic. Wildcard characters (*) were applied to account for variations in term usage (e.g., “canar*” retrieves for canaries, canary, or canarias). The complete search formula is available in the supplementary material. In the Web of Science, the TS= operator was used in the search equation, while in Scopus, a similar logic was applied by adapting the TS= to TITLE-ABS-KEY=. In both, filters were applied to exclude non-relevant subject areas, such as Medicine, Arts, and Physical-Chemical-Mathematical Sciences.

Table 1. Search terms for the systematic review.

Area	Target		Habitat / Species
Macaronesia	Acoustic	Cartography	Zostera
Webbnesia	Aerial	Coastal Ecosystem Services	Aquatic Vegetation
Azores	Backscatter	Conservation	Black Coral
Açores	Bathymetric	Distribution	Coral
Madeira	Drone	Geospatial Data	Eelgrass
Wild Island	GIS ⁽¹⁾	Habitat	Forest

Islas Salvajes	Hydroacoustic	Habitat Connectivity	Garden
Ilhas Selvagens	Hydroacoustic Survey	Habitat Mapping	Halodule
Desertas Islands	LIDAR ⁽²⁾	Imaging	Halophila
Islas Desertas	Multispectral Imaging	Map	Maerl
Canary	Radar	Mapping	MAF ⁽⁵⁾
Canarias	Remote Sensing	Marine Environmental Assessment	Nodule
Canaries	Satellite	Marine Flora Conservation	Rhodolith
Canary Islands	Scanning	Marine Flora Mapping	Seagrass
Islas Canarias	Side Scan Sonar	Marine Habitat Protection	Seaweed
Cabo Verde	Sonar	Marine Protected Areas	
Cape Verde	Sonic	MBES	
	Sound	Monitoring	
	Spatial Mapping	MPA ⁽⁴⁾	
	SSS ⁽³⁾	Seabed Mapping	
	Telemetry	Spatial Analysis	
	Thermal Imaging	Multibeam	

(1) Geographic Information Systems; (2) Light Detection and Ranging; (3) Side Scan Sonar; (4) Marine Protected Area; (5) Marine Animal Forest.

2.3. Inclusion and Exclusion Criteria

To ensure the quality and relevance of the selected studies, rigorous “inclusion” and “exclusion” criteria were established. Studies addressing cartography and providing spatial data were included, even if explicit maps were absent. Reviews containing maps or spatial data, as well as studies focused on one or more Central-Eastern Atlantic archipelagos or the Mid-Atlantic Ridge, when linked to the concerned archipelagos, were also considered. No language or date restrictions were applied, but only peer-reviewed journal articles were included. Conversely, studies that only reported species presence/absence without spatial data, articles focused solely on methodologies without data or maps, and grey literature or non-peer-reviewed works were excluded.

2.4. Screening and Selection Process of Articles

The screening process followed the PRISMA guidelines, and it was conducted in two phases (Figure 2). First, the results were imported into Rayyan software [111], where duplicates were automatically removed and manually verified. At least two authors independently screened titles and abstracts to exclude non-relevant articles [90]. In the second phase, the preselected articles underwent a thorough review to confirm their relevance, with discrepancies resolved by a third reviewer.

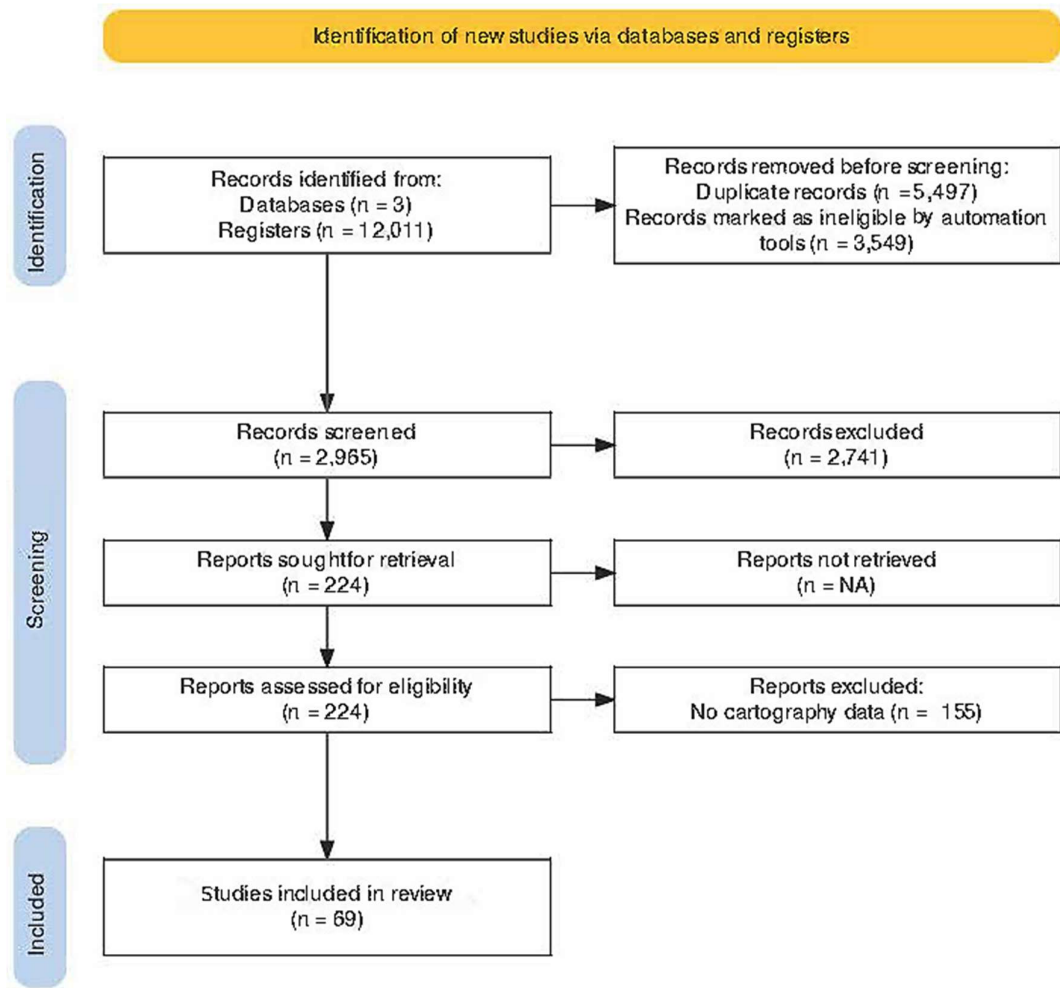


Figure 2. PRISMA flow diagram that shows the bibliography search process (created with Ryyan app [111]).

2.5. Data Extraction and Coding

The information from each article was first consolidated into an Excel template, creating two separate databases (Table 2): one for studies and another for species observations. The first database compiled methodological and contextual features, including mapping techniques, study area, research period, and other relevant factors. The second database focuses on taxonomic details, tracking the species recorded, the number of studies in which they appeared, and their presence or absence in each archipelago.

Table 2. Summary of data sources extracted from the literature review.

i. DATA RELATED TO STUDIES	
Parameter	Option / Detail
Mapping techniques	i. Direct (SCUBA, <i>in-situ</i> observations, etc.)
	ii. Remote (ROVs, submersibles, etc.)
	iii. Aerial (drones, airborne sensors, etc.)
	iv. Satellite (MODIS, etc.)
	v. Acoustic (side scan sonar, multi-frequency sonar, etc.)
	vi. Sampling (dredges, corers, nets, CTD Rosette, traps etc.)

	vii. Bibliographic studies
Study area	Archipelago id (Azores, Madeira, Canary Islands & Cabo Verde)
Year of the study	Year in which the research was conducted
Main objective	i. Exploratory (generation of original data) ii. Monitoring (tracking variables over time) iii. Mapping (mapping of habitats and distributions) iv. Bibliographic review (analysis of existing literature)
Postprocessed maps	Type (Habitat and/or geological map)
Habitat type	i. Macrophyte beds ii. Rhodolith beds iii. Coral reefs or gardens
Depth	Maximum depth range recorded
Substrate type	Hard / Soft / Mixed
Data modelling	Use of advanced statistical algorithms
Cartographic data analysis	i. Geographic Information Systems (GIS) ii. Bathymetric or acoustic analysis iii. Image analysis and computer vision iv. Statistics and modeling (including ML) v. Taxonomic or morphological approaches
Mapped zones area	Reported area (when available)

ii. DATA RELATED TO OBSERVED SPECIES

Parameter	Option / Detail
Taxonomy	Revised taxonomic classification in WORMS and AlgaeBase
Number of studies	i. Number of studies in which the species appears ii. Number of archipelagos in which its presence is documented
Zone	Archipelago id (Azores, Madeira, Canary Islands & Cabo Verde)

2.6. Methodological Quality Assessment and Data Analysis

Methodological quality was assessed using an ad hoc strategy that evaluated the clarity of the technique descriptions, transparency in identifying limitations, and the relevance of the methodologies employed. Three of the authors conducted the literature review, resolving any discrepancies through consensus [90].

The data analysis incorporates qualitative descriptions and graphical representations to evaluate the temporal distribution and application of mapping techniques. Statistical tools, such as SPSS [112] and PAST [113], were used for descriptive analyses and to assess correlations between different variables. Additionally, methods and patterns of habitat representation were examined to identify optimal combinations for mapping complex ecosystems. Studies with a high risk of bias were excluded to enhance the validity of the conclusions. In the graphical and tabular summaries, individual publications may be represented multiple times, as a single article can report multiple

mapping methodologies and/or assess multiple habitat types across archipelagos. Consequently, cumulative frequencies and percentage distributions in the figures do not sum to 100%, reflecting the overlapping nature of methodological and habitat classifications rather than data omission.

3. Results

3.1. Research Effort and Objectives

From a total of 12,011 articles identified across three databases, the initial screening narrowed the sample to 2,965, of which 224 were analyzed in detail. The review focused on 69 studies, providing key insights into the mapping of marine ecosystem-engineers in the concerned archipelagos. Additionally, data on 506 species was collected and compiled into a separate database as part of the taxonomic analysis derived from the reviewed articles.

Geographical and temporal patterns revealed that 30 studies were recorded in the Canary Islands, 29 in Azores, 8 in Madeira, and 3 in Cabo Verde. Research effort has exhibited temporal and spatial variations, with differences in intensity across each archipelago (Figure 3). Azores experienced a peak in research activity between the late 1990s and early 2000s, followed by a resurgence and stabilization between 2005 and 2015, with a relative increase from 2020 onwards. In Madeira, research progressively increased starting in 2020. In the Canary Islands, research levels remained relatively constant since the early 2010s, with more intense activity peaks observed from 2020 onwards, persisting to the present day. Finally, in Cabo Verde, all recorded studies were conducted between 2023 and 2024. Regarding the study objectives, 41% (28 studies) focused on mapping, 38% (26 studies) on explorations, and 21% (15 studies) on monitoring (Table A1).

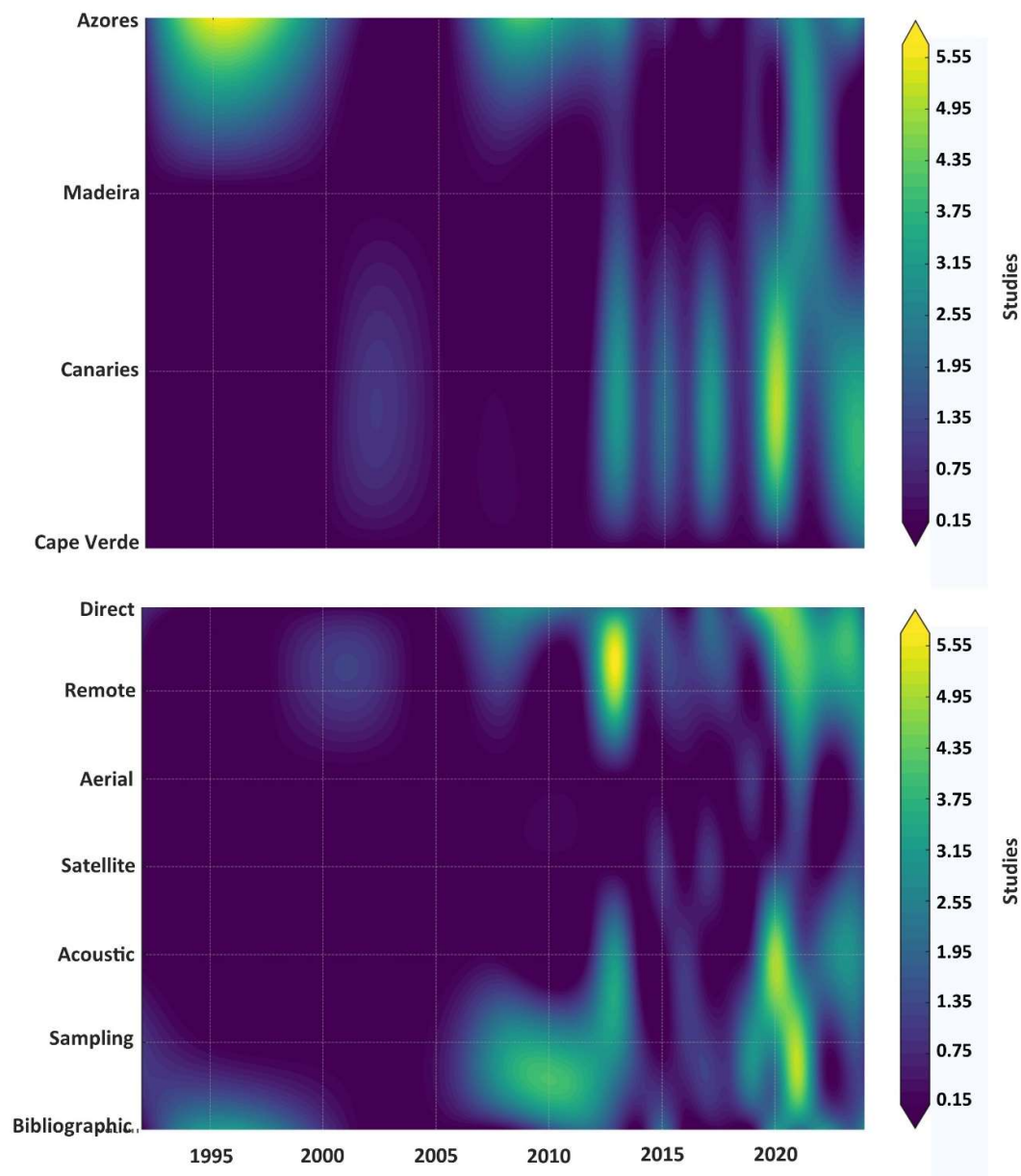


Figure 3. Temporal changes in the number of studies by region (top) and by type of sampling method (bottom) in the context of marine research in Central-Eastern Atlantic archipelagos. The color intensity represents the density of surveys over time (1990-2025), with higher values in yellow and lower values in purple.

3.2. Methodologies and Habitats Analysis

The analyzed studies covered three main types of marine habitats: macrophyte beds, rhodolith beds, and coral reefs. Various methodologies were employed, including direct, remote, acoustic, satellite and/or aerial observation, sampling, and bibliographic review.

The methodologies have exhibit different usage patterns over time (Figure 3). Direct observation has been documented since the beginning, with higher recurrence during 2020 and 2021. Remote observation first appeared in 2000 and showed notable occurrences in 2013 and 2021. Aerial methodologies emerged in 2019, with additional occurrences in 2021 and 2024. Satellite observation first appeared in 2015 and was documented in isolated studies until 2024. Acoustics was not reported until 2012 and became more prevalent from 2020 onwards. Sampling has been used intermittently

since 1992, with increased application during the 2010s. Finally, bibliographic reviews were initially recorded in the early 2000s and reappeared consistently from 2020 to 2024.

Comparative analysis among (Figure 4) and within (Figure 5) archipelagos revealed similar variations in the application of observation and data collection methodologies. In Azores, remote observation was the predominant method, followed to a lesser extent by direct observation and sampling, with similar usage. Acoustic methods, aerial techniques, and bibliographic review were employed less frequently. In contrast, although all methodologies—except remote observation—were relatively more common in the Canary Islands, direct observation emerged as the primary method, complemented by sampling. The use of the remaining methodologies (remote observation, acoustic methods, and bibliographic review) was identical, whereas satellite observation was incorporated in a moderate and exclusive manner in the Canary Islands. In Madeira, direct observation, acoustic methods, and sampling constituted the most employed techniques, whereas remote sensing, aerial observation, and bibliographic review were applied to a lesser extent. The index of aerial observation usage was similar to that used in Canary Islands and Azores. Finally, in Cabo Verde, the only methodologies implemented were remote observation—which proved to be the most used—followed by direct observation and acoustic methods, with equivalent usage. Bibliographic review was the only additional methodology recorded in all studies conducted in Cabo Verde.

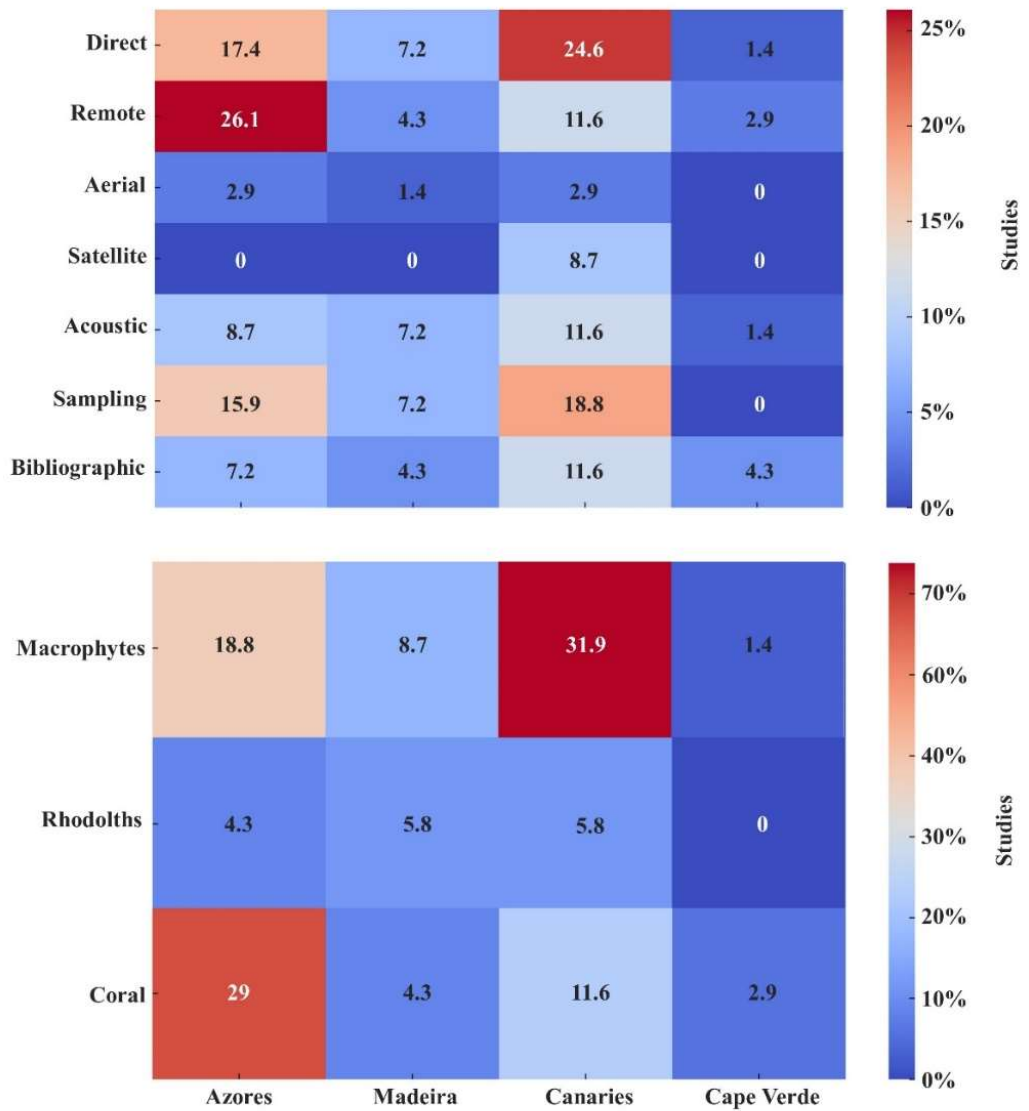


Figure 4. Interregional differences in terms of (top) the types of sampling methods used and (bottom) the habitats analyzed. Individual studies may be counted in multiple categories (methodologies and/or habitats), so cumulative percentages do not sum to 100%.

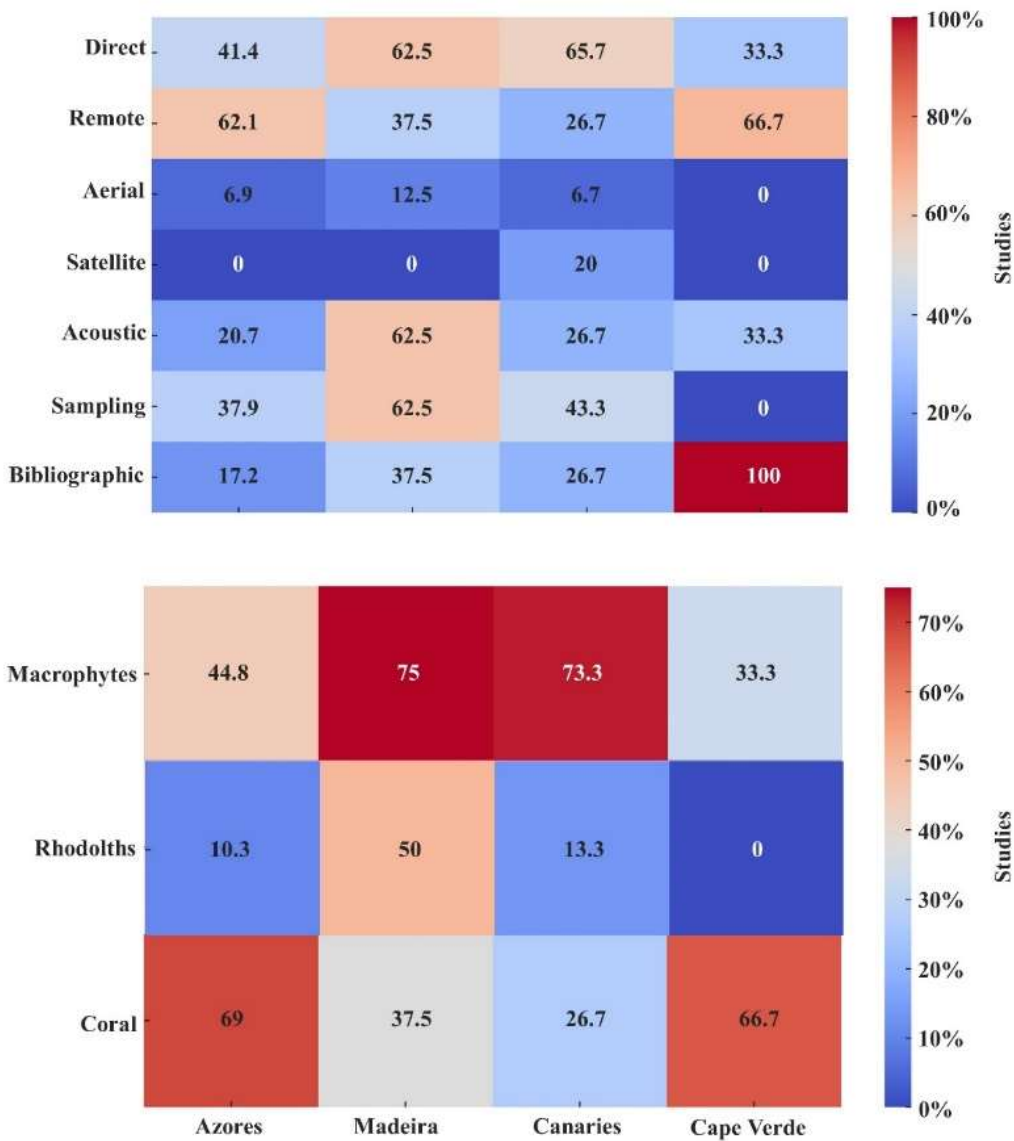


Figure 5. Intraregional differences in (top) the types of sampling methods used and (bottom) the habitats analyzed. Individual studies may be counted in multiple categories (methodologies and/or habitats), so cumulative percentages do not sum to 100%.

3.3. Biodiversity Analysis

The greatest research effort was carried out on macroalgal beds (42 studies), followed by coral reefs (33 studies) and, to a lesser extent, by rhodolith beds (11 studies). Similarly, habitats comparisons revealed differences in the research efforts on these habitats among (Figure 4) and within (Figure 5) the archipelagos, where Azores exhibited the higher percentage of studies related to coral reefs. Conversely, the Canary Islands focused its research efforts toward macroalgal beds, followed by Azores and Madeira. Rhodolith beds had a similar percentage of studies across all the archipelagos, except for Cabo Verde, where no studies targeting this habitat were observed.

Our literature review recorded a total of 505 species, with the highest diversity observed in the Azores (393 species), followed by the Canary Islands (131), Madeira (69), and Cabo Verde (10) (Figure 6, 7; Table A1). The Azores concentrated the highest percentage of cnidarian species, exhibiting a similar diversity in both Hexacorallia and Octocorallia classes, accounting for ca. 86.4% of the

cnidarian species observed. In particular, the Mid-Atlantic Ridge predominantly documented cnidarians (ca. 99%: Table A1). Likewise, the Azores included the greatest diversity of marine plants (Figure 7; Table A2), with the exception of Tracheophyta, which were not represented. Rhodophyta were predominant, followed by Chlorophyta and a similar percentage of Ochrophyta (Figure 7; Table A2). In the other archipelagos, a similar pattern was observed regarding cnidarians and macroalgae. However, Madeira documented a similarity in the diversity of cnidarians between Hexacorallia and Hydrozoa, presenting a lower representation of Octocorallia and a slight representation of Tracheophyta. The Canary Islands exhibited the highest diversity of Tracheophyta (Figure 7; Table A2). With respect to cnidarians in the Canary Islands, a higher percentage of Hexacorallia was observed, followed by Octocorallia and Hydrozoa. Finally, in Cabo Verde, the diversity was concentrated in cnidarians, with a greater diversity of Octocorallia than of Hexacorallia, and no macroalgae were documented, except for Tracheophyta (Figure 7; Table A2).

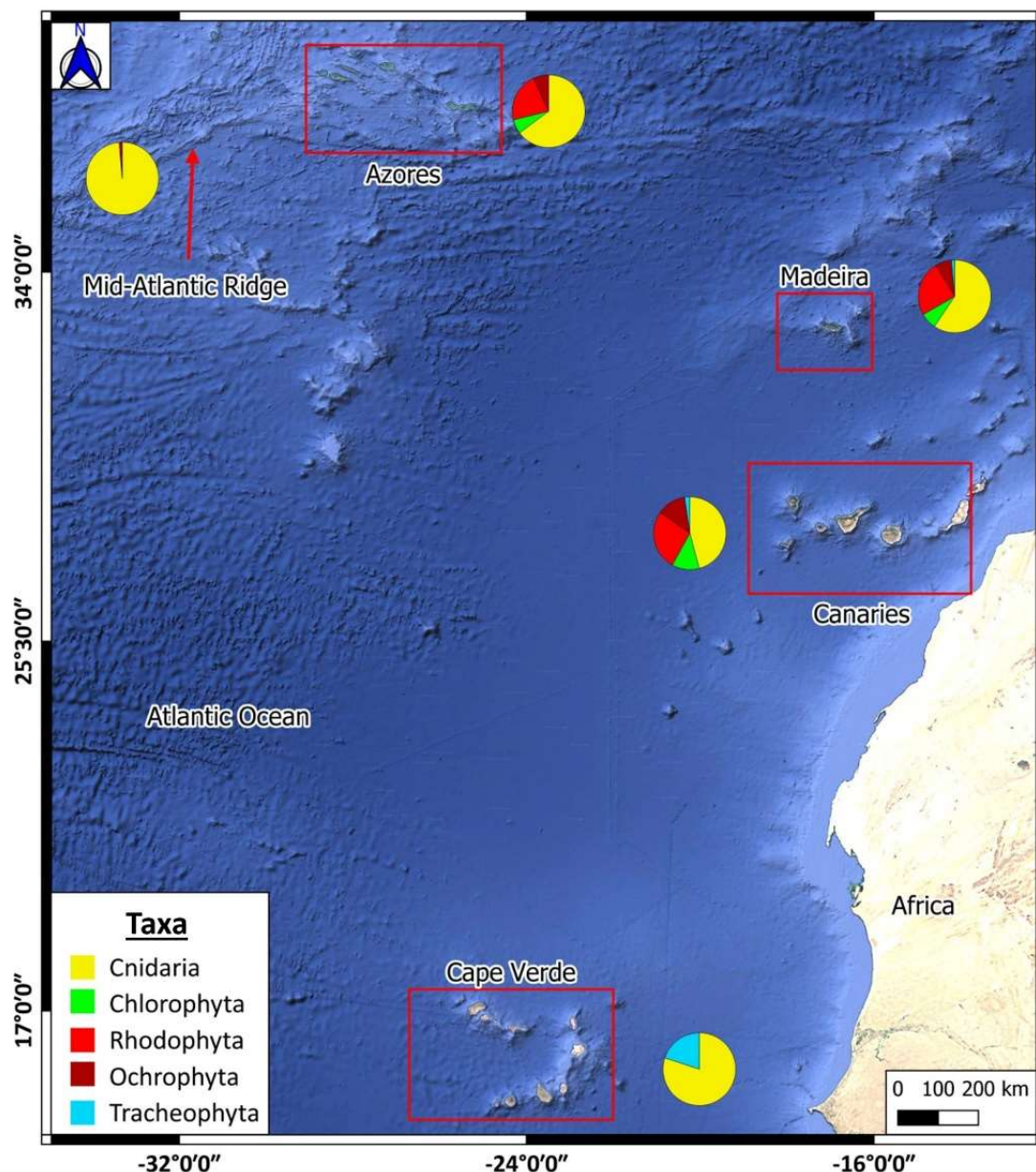


Figure 6. Spatial distribution of the study regions in Central-Eastern Atlantic archipelagos and the Mid-Atlantic Ridge, with proportional representation of the main taxonomic categories registered in each region.

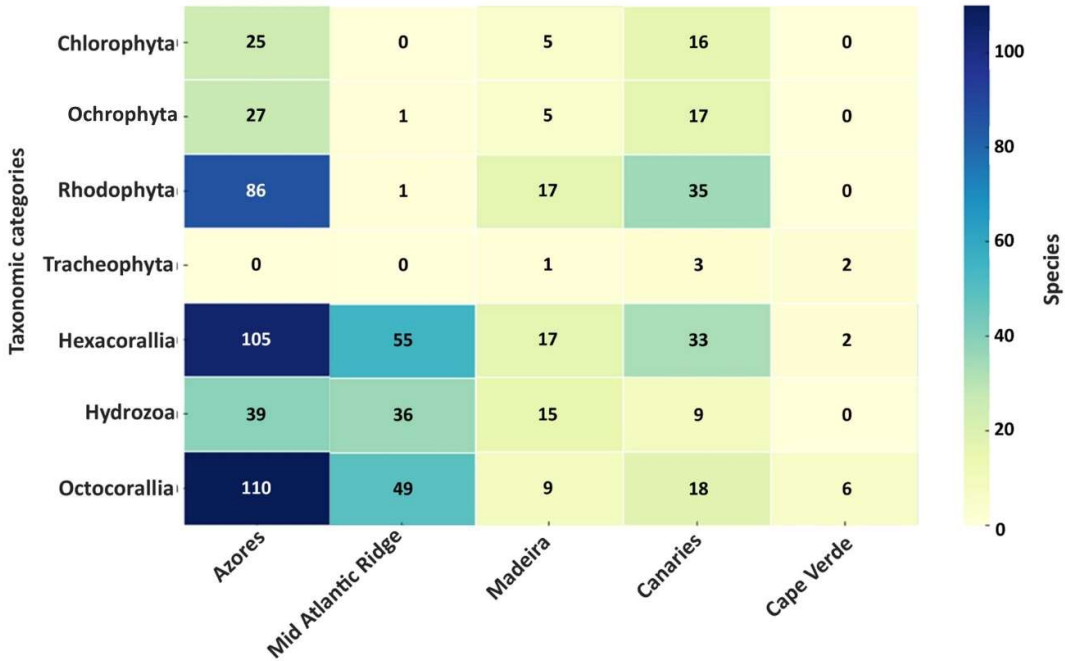


Figure 7. Species richness by taxonomic category and region in the Central-Eastern Atlantic archipelagos and the Mid-Atlantic Ridge. The matrix shows the number of species recorded per group.

Regarding to singular species (Table A2), it was evidenced that the greatest research effort was focused on seagrass meadows, specifically on *Cymodocea nodosa* (Ucria) Ascherson, 1870, which was the subject of 14 studies. Subsequently, the effort was directed toward brown macroalgae, in particular *Dictyota spp* (J.V. Lamouroux, 1809) and *Padina pavonica* (Linnaeus, Thivy, 1960), which were the subject of 12 and 10 studies, respectively. Among corals, the species that were studied most intensively were the octocoral *Viminella flagellum* (Johnson, 1863) and two hexacorals, *Desmophyllum pertusum* (Linnaeus, 1758) and *Antipathella wollastoni* (Gray, 1857), each appearing in 8 studies.

Methodological approaches to studying marine benthic communities varied according to habitat and region (Figure 8). Throughout the geographical range considered, macroalgae were studied mainly by direct observation and sampling, while remote sensing and acoustic methods were used less frequently. In rhodolith beds, direct observation (7 cases) and acoustic methods (5 cases) were the most common approaches. Corals, which were found at depths of up to 4,000 m, were investigated primarily by remote sensing (25 cases) and acoustic methods (13 cases), although sampling and direct observation were also applied. Acoustic methodologies, in particular, were used more frequently in the Canary Islands (8 cases) and Azores (6 cases). Overall, the Azores and the Canary Islands represented the majority of the studies, demonstrating a diverse methodological approach.

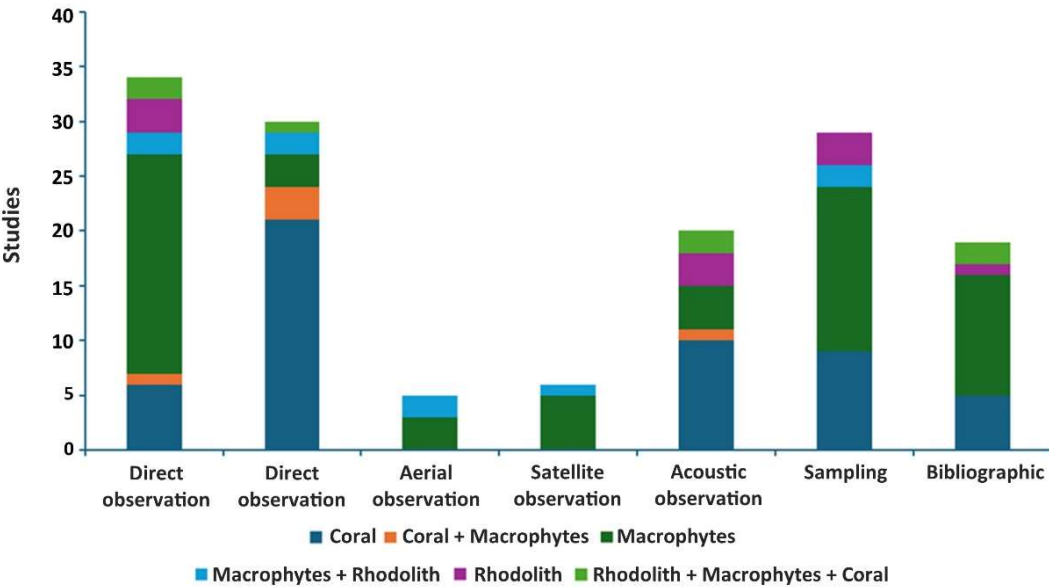


Figure 8. Number of studies by type of sampling method, broken down according to the combination of areas and habitats analyzed.

3.4. Postprocessing Data Analyses

Of the total studies analyzed, 62% incorporated some form of mapping, with 25% generating geological cartography. The remaining studies provided cartographic data as coverage or visual percentages, supplementing spatial analyses. Computational modeling appeared in ca. 49% of the studies, showing a ca. 750% increase between 2010 and 2020, followed by an additional 100% rise between 2020 and 2024 (Table A1).

This growth coincided with the integration of Geographic Information Systems (GIS), described in ca. 47% of the studies, combined often with image analysis tools (18 studies) or remote sensing data (15 studies). GIS was primarily used to predict coral and macrophyte beds distributions based on remote, acoustic, and direct observation data (Figure 9).

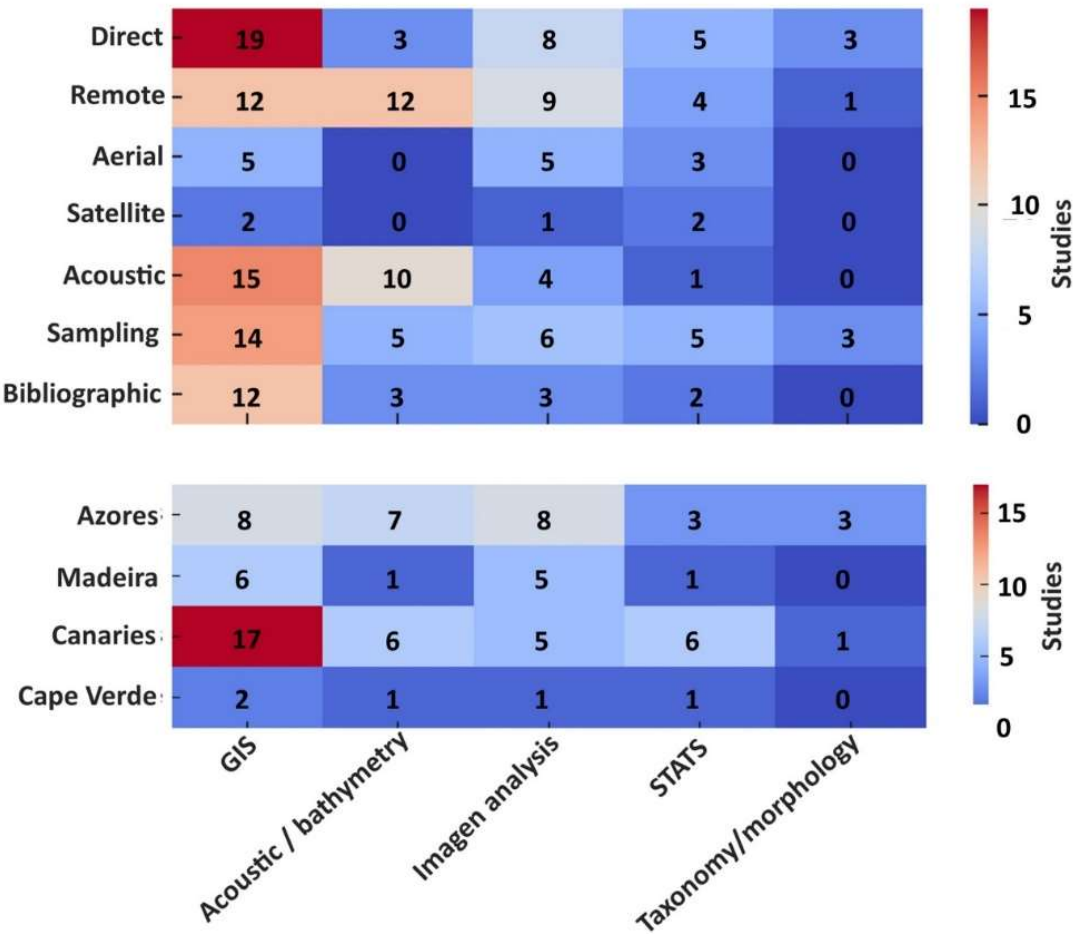


Figure 9. List of post-processing analyses, including Geographic Information Systems (GIS), acoustic data analysis, image analysis, statistical analysis (STATS) and taxonomic/morphological analysis, with: (top) the types of sampling methods used; and (bottom) the archipelagos analyzed.

The Azores and Canary Islands showed a strong preference for GIS, while Madeira relied more on taxonomic and morphological tools. Rhodolith beds were linked to GIS and acoustic/bathymetric analysis, macrophytes to GIS and statistical analysis, and corals to GIS and image analysis (Figure 9).

4. Discussion

Although marine research in some of the Central-Eastern Atlantic archipelagos dates back to the 19th century, these studies have predominantly focused on taxonomy and species cataloguing, relegating habitat mapping to a secondary position [114]. During the last 30 years, research efforts have been concentrated, particularly in the Azores and the Canary Islands archipelagos, while in Cabo Verde, such efforts are scarce or nearly non-existent (Figure 10).

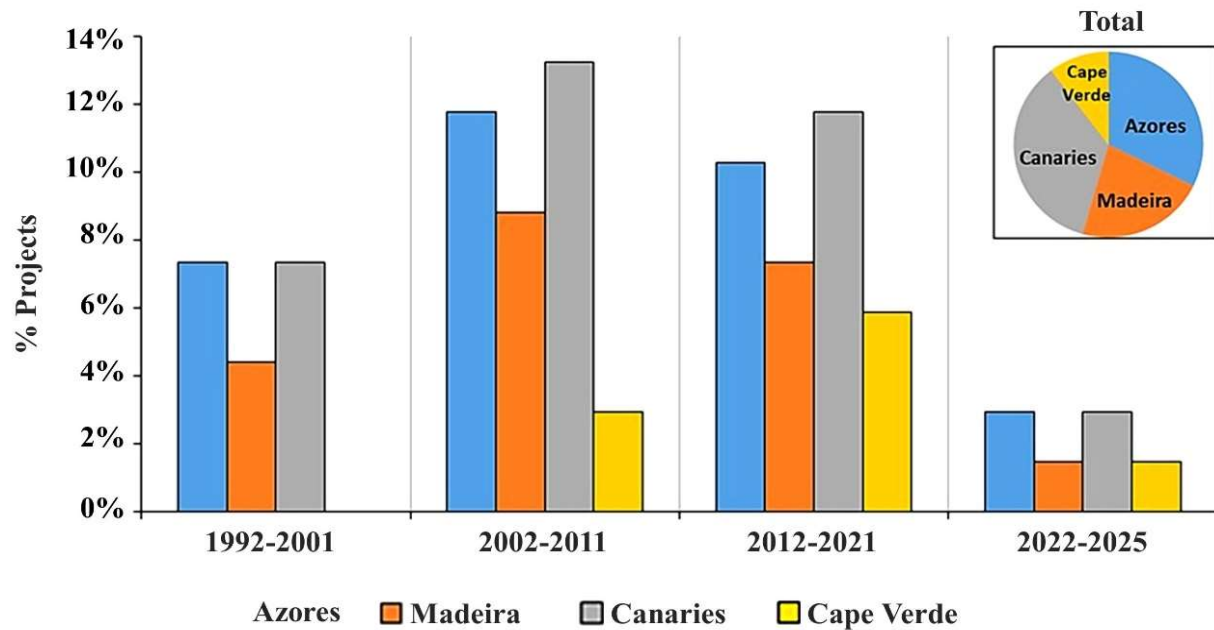


Figure 10. Percentage of European-funded projects by Archipelago. Sources: https://cinea.ec.europa.eu/programmes/life_en; <https://cordis.europa.eu/>; <https://interreg.eu/>; <https://fundacion-biodiversidad.es/>; <https://www.biodiversa.eu/>; <https://plocan.eu/>; <https://www.mac-interreg.org/>.

Transnational cooperation projects focus on monitoring, conservation, cataloguing, and mapping of marine biotopes dominated by habitat-forming megafauna, considered Vulnerable Marine Ecosystems (VMEs), through projects such as MISTICSEAS, MIMAR, POPCORN, and MOVE ON (<https://misticseas3.com/es/>; <https://www.mare-centre.pt/en/proj/mimar>; <https://www.ecoaqua.eu/es/habitats-popcorn.html>; <https://www.moveon-project.eu/homepage/>). However, updated information on the current progress in comprehensive seabed mapping remains limited, a situation comparable to studies conducted on the Mid-Atlantic Ridge, where much accumulated knowledge from multiple multinational expeditions since 1954 remains unpublished in indexed scientific journals, primarily existing as technical reports or grey literature [115,116].

4.1. Methodologies and Technological Advances

4.1.1. Direct and Remote Observation Techniques

The most cost-effective methodology for characterizing marine habitats is direct observation, generally complemented by in situ sampling, as it provides detailed descriptions and generates thematic maps based on biota distribution, facilitating the identification of spatial patterns and ecological processes [96,117–120].

Although direct observation of keystone species and habitats has been systematically employed over time, it is now complemented by emerging remote technologies, such as optical remote observation, or its combination with acoustic and satellite methods, enabling three-dimensional analysis (horizontal and vertical dimensions) [96,121,122]. Currently, marine habitats can be effectively mapped using various stationery and mobile cameras, including remotely operated vehicles (ROVs) and autonomous underwater vehicles (AUVs), providing high-resolution imagery for species identification with expert analysis [123,124]. These tools can explore challenging areas, enabling long-term monitoring and broad data collection [125–128]. However, they face limitations in covering very extensive areas and dealing with the “canopy effect,” which obscures deeper layers, necessitating complementary mapping methods [120,129]. Specifically, in the Canary Islands, camera-based methods — either standalone or combined with other methods — has proven

particularly effective for subtidal habitats and deep-water ecological studies, more so than for intertidal zones [120,130]. Furthermore, this approach aligns well with other methodologies, such as the MNCR biotope method employed in the Azores, facilitating the identification and monitoring of structural changes within algal communities [131].

4.1.2. Acoustic Methodologies

Technological advancements have transformed the way information is acquired and interpreted [132,133]. The development of acoustic technology in the 1940s revolutionized seabed exploration, enabling the interpretation of seabed textures over extensive areas and the creation of more realistic images [134,135]. Over time, technological improvements have significantly enhanced data acquisition and image resolution by refining acoustic signal properties and advancing digital tools for acoustic data analysis [135,136]. Among the various acoustic systems, wide-beam or multibeam systems (Side-Scan Sonar or SSS) [134,135,137], single-beam echo sounders or fish-finders [135,138], and multibeam bathymetric systems (MBES) [135,139] are widely utilized. Within the concerned oceanic archipelagos, this review highlighted SSS and MBES as the most versatile acoustic technologies employed by regional marine research, particularly for habitat characterization along the Mid-Atlantic Ridge [14,56,140,141].

Our review suggests that SSS was particularly effective in identifying substrate textures, highly efficient in habitats with sedimentary and rocky bottoms, as well as detecting coral reefs and seagrass beds in the Azores and Canary Islands [14,141,142]. Conversely, MBES was more precise in measuring bathymetry and seabed relief, making it more suitable for characterizing deep-water habitats, such as coral reefs along the Mid-Atlantic Ridge [56,143–145]. Furthermore, advancements in MBES post-processing have facilitated the implementation of new Multi-Detection (MD) techniques capable of estimating the presence of CWC gardens in the Azores and Canary Islands (e.g., black corals, [46,56,146], undetectable directly via SSS or MBES due to their proteinaceous composition. Such techniques lay the groundwork for enhancing future distribution maps of other key habitats [146].

4.1.3. Aerial and Satellite Methodologies

Aerial remote sensing is transforming marine mapping by providing high-resolution data on habitat coverage and coastal morphological changes using optical, thermal, and hyperspectral sensors [147]. Its capacity for operating at high spatial resolution makes it ideal for monitoring coastal ecosystems and intertidal zones, facilitating vegetation cover analysis [4,147–150]. Aerial platforms equipped with LIDAR (Light Detection and Ranging) enable precise bathymetric mapping in shallow waters [96], generating detailed digital models of sandy bottoms and reefs [4,148–151]. Specifically, in the Azores, Madeira and the Canary Islands, drones (Unmanned Aircraft Systems or UAS) have effectively monitored shallow waters and algal blooms; however, highly heterogeneous habitats present methodological challenges due to environmental and meteorological variables limiting their spatio-temporal applicability [149–151]. Additionally, they cannot accurately differentiate benthic communities without ground-truthing support [149,150]. To overcome these limitations, Monteiro et al. (2021) proposes an integrated approach, by combining in situ data—specific benthic composition—with physiographic mapping derived from UAS imagery.

Satellite platforms (Sentinel, Landsat, WorldView) employ multispectral and hyperspectral sensors, enabling large-scale monitoring of shallow marine ecosystems [152–158]. They offer extensive temporal and spatial coverage, providing characterizable images alongside abiotic and oceanographic variables such as temperature, pH, or chlorophyll [147,159–165]. Despite their effectiveness in marine habitat mapping, satellite studies have been conducted almost exclusively in the Canary Islands, frequently adopting a single-methodological approach and facing limitations due to reliance on reflectance and/or spectral profiles without integrating in situ biological data [149,164]. For example, multitemporal satellite image analysis was applied to evaluate the impact of the Granadilla port construction on the Special Conservation Area (ZEC) of “Sebadales del Sur” in

Tenerife (Canary Islands) [166]. Cosme De Esteban et al. (2023) underscored the importance of ground-truthing regarding satellite approach, as for example comparisons carried out in Príncipe Island (Gulf of Guinea) that revealed discrepancies in delineating marine habitats of high ecological value, such as rhodolith beds, where the acoustic methodology supported by direct observations was more precise compared to satellite methods. Furthermore, Eugenio et al. (2023) highlights the efficacy of these multitemporal approaches integrating bathymetric and benthic studies to detect variations through qualitative and quantitative analysis, producing high-resolution maps.

Globally, aerial methods provide higher spatial resolution and greater flexibility for selecting optimal environmental conditions, whereas satellite imagery analysis reduces exploratory sampling needs, improving efficiency in time, cost, and reducing sampling bias [149–151]. Despite proven efficacy, aerial and satellite remote sensing methodologies remain underrepresented, predominantly applied in the Canary Islands and the Azores, where usage has increased since 2021. In the Canary Islands, image processing and automatic classification algorithms based on artificial intelligence and deep learning have achieved up to ca. 96% effectiveness in discriminating species, such as the genera *Cymodocea* and *Gongolaria* [151], highlighting the increased analytical precision made possible by these advancements; however, clearly defined training areas and ground-truthing validation remain essential [160,164,167].

4.1.4. Data Postprocessing

Once cartographic data is collected, integrating them into a unified reference framework becomes essential. This integration is facilitated by geographic information systems (GIS), which aid in spatial organization and analysis [168–170]. GIS effectively consolidate datasets that differ in spatial, temporal, and thematic scales, thereby addressing informational heterogeneity [171,172]. Literature demonstrates extensive use of specialized software for image analysis [114,120] and GIS programs (e.g., ESRI's ArcGIS or QGIS) for the integration of georeferenced vector and raster data [4,115,148,173]. Additionally, analytical interfaces have been optimized through additional tools that facilitate visualization and map overlays, allowing the combination of habitat information and physicochemical variables. Such integration is pivotal for conducting statistical analyses and creating predictive maps through modeling [141,142].

Following database integration, habitat mapping advances by modeling habitat conditions, emphasizing the biological integrity index and anthropogenic stress factors [72,174].

Identifying new biotopes and extensive data collection on taxonomic diversity significantly enhances predictive statistical modeling at multiple spatial scales [175,176]. This modeling facilitates an integrated assessment of biodiversity and generates predictive maps incorporating anthropogenic risks and impacts [177–180]. According to Braga-Henriques et al. (2013), accurate modeling and data analysis require standardized data formats. However, heterogenous data collection from different periods, campaigns, and multinational organizations complicates direct standardization, requiring the use of probability mapping or prior data refinement according to established standards [114]. Refinement involves standardizing classifications and terminologies to ensure coherent mapping and effective environmental data storage [114]. Consequently, the hierarchical EUNIS habitat classification system, extensively applied across Europe, can support large-scale habitat mapping by integrating comprehensive ecological knowledge and spatial data on seabed characteristics [166]. Accordingly, in the Canary Islands, a study provided the first comprehensive spatial assessment of benthic ecosystem service supply (until 50 m depth; [181]), generating maps through a flexible, updatable methodological approach based on scientific literature, directly applicable to marine spatial planning.

One of the initial steps in Mapping and Assessment of Ecosystems and their Services (MAES) involves ecological niche and food web characterization using specialized models and software (e.g., ECOPATH, ECOSIM, ENFA, or MAXENT). These tools are crucial for analyzing energy flows and predicting species distribution changes due to environmental disturbances and anthropogenic activities [96,147]. Their application has been pivotal in predictive analyses of species distribution,

such as CWC, in studies conducted in the Azores and Cabo Verde [176,182]. When information is limited or historically incomplete, implementing ensemble models has proven effective. In Cabo Verde and the Canary Islands, combining these models with AI and deep learning classification techniques, predicted (i) new CWC presence areas on seamounts [182], (ii) identified the potential distribution of algae (both essential and invasive), and (iii) assessed seagrass meadow conditions following port infrastructure construction [130,183]. In the Azores, integrating data on anthropogenic activities (e.g., fisheries) and protected areas has facilitated evaluations of interactions between key habitats, such as macroalgae and CWC, and resources [129,142,176]. This information provides critical insights into ecosystem functioning, niche interactions and influencing variables, contributing to the development of distribution maps that inform spatial planning and conservation strategies of these essential habitats [4,142,162,176]. Finally, in the Canary Islands, visual analyses and the first ecological model of an MPA have demonstrated the potential of this conservation tool and the responses of communities to natural events [162,184], thereby facilitating adjustments in MPA design and management policies. Collectively, these data support improvements in design and management policies to ensure the conservation and recovery of ecosystems [4,162,184].

4.2. *Habitat Mapping*

Marine habitat maps are developed using a variety of methodologies that integrate multiple sources of data (see section 4.1). Biological data (e.g., species and community distributions), geophysical variables (e.g., bathymetry, geomorphology, physicochemical parameters), and habitat-related factors (e.g., ecosystem services and local and global threats) collectively determine the precision, spatial coverage, quality, and overall utility of the final habitat map.

4.2.1. Methodologies

Marine habitat mapping remains operationally challenging and costly despite technological advancements since the late 1990s, which have enhanced spatial coverage and data accuracy [114,185]. While studies often rely on either direct imaging or remote observation methods individually [149,186], integrating multiple methodologies significantly enhance accuracy by offsetting each technique's inherent limitations [120]. For instance, in the Canary Islands and Azores, studies relying exclusively on remote sensing techniques effectively describe facies and community structure. However, these approaches can be limited by the “canopy effect”, where dense algae cover or CWC gardens may obscure underlying strata, hindering the observation of all present taxa [120]. Furthermore, although most benthic areas provide valuable data, not all provide sufficient information to produce comprehensive habitat mapping. Therefore, it is essential to clearly define the types of thematic maps that can be generated from such data [31,144]. No single approach exists for their development, given the multidisciplinary complexity and the variability associated with the specific objectives of each project. Consequently, multidisciplinary campaigns are conducted—such as the one carried out in the Azores by Somoza et al. (2020)—in which both biological and abiotic data (e.g., geophysical, hydrographic, geological, oceanographic, etc.) were collected, enabling the description of new hard and soft coral gardens and the analysis of the factors that determined their location and distribution.

4.2.2. Biotic-Abiotic Data

Marine habitat maps integrate multiple data sources, including biological (e.g., species and community distributions) and geophysical variables (e.g., bathymetry, geomorphology and physicochemical parameters). According to Brown et al. (2011), habitat maps can be classified based on mapped variables, distinguishing between biological and abiotic data [45,187,188]. Adopting a hierarchical, multiscale approach at species, community, and ecoregion levels enhances maps comprehensiveness [123,189,190]. Additionally, discovering new biotopes and detecting structural or distributional changes through repeat mapping or predictive modelling improve habitat-

classification accuracy [149,162,167,183]. However, maps focused solely on geophysical features remain incomplete without integrating biological information [169,191–193]. In these volcanic archipelagos, geological studies commonly regard biological analyses as secondary, especially in deep-water habitats such as the Mid-Atlantic Ridge and hydrothermal areas of the Azores [141,143,185,194]. Since 2013, however, multidisciplinary research —particularly in the Azores and the Canary Islands— has significantly expanded, driven by technological innovations, advanced methodologies, integrated management plans, and improved infrastructure and funding availability (Figure 10; [178]).

Other mapping investigations treat spatial information as complementary or secondary to the primary objective, which is to collect biotic or abiotic data from marine habitats or facies. For example, research on hydrothermal habitats, in metal-rich areas—such as those hosting *Rimicaris* shrimp populations along the Mid-Atlantic Ridge (Azores)—enable the generation of more accurate predictive models for species distributions [186]. Conversely, investigations that specifically map and characterize valuable habitats, such as the seagrass meadows of *C. nodosa* in the Canary Islands, facilitate the prediction of dynamics in related communities, including fishery populations [164,195].

4.2.3. Habitat-Related Factors

Marine habitats are increasingly threatened by global and local threats (e.g., climate change, overfishing, pollution and coastal development), underscoring the need for up-to-date habitat mapping to manage these ecosystems effectively [149]. In the concerned oceanic archipelagos, several studies have already used direct and remote-sensing techniques to detect and map local threats (e.g., marine litter, trawling, long line fisheries, etc.), showing how cumulative stressors erode functional diversity and community structure over time, particularly in morphologically complex taxa such as CWC [136,140,185,196]. Additionally, marine litter data has been used in advanced predictive modelling to pinpoint plastic accumulation hotspots and compare them with potential key habitat areas [197]. Documenting spatial links between habitats and both historical and contemporary local threats is therefore essential for evidence-based conservation and management [116,130,148,164]. Thus, Martín-García et al. (2022) established cause and effect relationships between human and environmental drivers that influence the distribution of brown macroalgae (e.g., *Gongolaria abies-marina*) in the western Canary waters. Equally, studies performed in Cabo Verde on abiotic factors (e.g., topography and hydrodynamics) are essential for characterizing habitats and supporting predictive models. This approach has effectively identified food supply mechanisms for suspension feeders and potential areas for key species such as the coral *Enalllopsammia rostrata* [184].

Understanding how the distribution and persistence of key habitats have changed in response to local threats is also critical for contextualizing recent observations. In Madeira, a southward regression of *Laminaria ochroleuca* has been documented [140], along with a decline in several Sargassaceae species [130]. Nonetheless, high resolution imagery of adult *L. ochroleuca* specimens (ca. 5–7 years) suggests that the archipelago may serve as a climate refuge against ocean warming [140]. Meanwhile, the first exhaustive mapping of rhodolith beds in Madeira revealed a widespread distribution across the archipelago, extending to depths of ~100 m—considerably deeper than those typically reported in the northeastern Atlantic—and comparable to the Azores [175]. Finally, regarding seagrass beds, *Cymodocea nodosa* and *Avrainvillea canariensis* patches have decreased in southern areas within a Marine Protected Area (Parque Natural Marino de Cabo Girão) [114,173,198]. In the Azores, regressions have also been observed in several species of the Sargassaceae family [130]. However, while the mesophotic niche occupied by algae in the Azores benefits from some protection against natural and anthropogenic stressors, it remains vulnerable to climate change, with predictions indicating a decrease of between 23% and 85% in the thermal niche of *L. ochroleuca* by 2100 [199]. Finally, in the Canary Islands, intensified maritime traffic favors invasive species introduction, notably populations of scleractinian corals (*Savalia savaglia*, *Tubastraea* sp., *Oculina* sp., *Culicia* sp.), necessitating continuous monitoring of these species [136,200]. Brown macroalgae forest of *Gongolaria abies-marina* transitioned from extensive to fragmented forests in the upper sublittoral

zone, almost disappearing in areas such as Canary Islands [130,148,151]. This coincides temporally with the increase in marine heatwaves (MHW), which displace species toward cooler waters [201], and with increased coastal urbanization. These activities can also negatively impact seagrass seabeds, causing reduction or disappearance of *C. nodosa* meadows and rhodolith beds, frequently replaced by invasive species (*Caulerpa racemosa*, *Caulerpa prolifera*) accompanied by cyanobacterial proliferations (*Lyngbya spp.*) [163,195,196,202].

4.2.4. Final Habitat Map in the Central-Eastern Atlantic Archipelagos

Most conservation projects are undertaken within MPAs, whose design has progressively shifted—driven by technological advances and the adoption of ecosystem-based policies—from a focus on single emblematic species to an integrated view of ecosystem functioning. Consistent, high resolution habitat mapping is therefore indispensable for redefining existing MPAs, proposing new ones, and ensuring comparability at the regional scale [3,56,140,203]. For example, Cosme De Esteban et al. (2024) recommended revising an Azorean MPA originally delimited to protect a seabird colony; the absence of a marine component resulted in a buffer zone dominated by sedimentary substrates and excluded reef habitats characterized by cold water corals. Likewise, cartographic and taxonomic surveys along the Mid Atlantic Ridge—sites initially designated as Natura 2000 Sites of Community Importance for cetacean conservation—have revealed extensive mesophotic and deep-water habitats of high ecological value, warranting an expansion of the protection boundaries [140].

4.3. Challenges and Future Implications

Globally, the review of the current status of marine habitat mapping in the oceanic and volcanic archipelagos highlighted two fundamental and interdependent methodological stages: exploration and monitoring. The exploratory stage aims to address initial questions about habitat presence, distribution, and ecological importance, laying the groundwork for subsequent, detailed studies. Clear examples from the Azores and the Canary Islands demonstrate how initial exploration identified essential habitats, such as cold-water coral gardens and rhodolith beds, which might otherwise have been overlooked in marine protected area planning. Conversely, the monitoring stage assesses temporal habitat changes and evaluates the effectiveness of implemented conservation measures, as exemplified in Azores, Madeira and the Canary Islands, where inadequate monitoring has limited effective management of key habitats.

A comprehensive analysis of methodologies and outcomes revealed significant progress in advanced technological applications, particularly in the Azores and Canary Islands, where a wide array of sophisticated tools—such as remote sensing, acoustic, and satellite sensors (e.g., Side-Scan Sonar, MBES)—have been successfully implemented. However, this contrasts sharply with Madeira and especially Cabo Verde, archipelagos constrained by limited technological infrastructure and marine research funding. This regional disparity directly impacts the quality and comprehensiveness of the bionomic maps produced, hindering the necessary interregional comparisons and integrations required for effective integrated management. To address this, regional initiatives promoting technological cooperation and specialized training of local researchers are crucial, as partially achieved by the Azores and Canary Islands through European-funded programs.

Furthermore, integrating and standardizing data generated from diverse methodological techniques remained a significant challenge. The involvement of various stakeholders employing different methodologies resulted in heterogeneous databases, complicating their coherent integration into thematic maps. Studies conducted in the different archipelagos, notably in the Azores, emphasized the imperative need for standardized reference frameworks, systematic field validation (ground-truthing), and robust data validation protocols. Adopting these measures will enhance data reliability and accuracy thereby strengthening predictive modeling capabilities essential for effective marine ecosystem management.

Advanced techniques such as computational modeling, GIS systems, and artificial intelligence have also revolutionized marine mapping in the region. Nevertheless, these technological

innovations introduced specific challenges related to processing, analyzing, and interpreting large volumes of data, especially in contexts with limited technological infrastructure. In areas such as Cabo Verde, where technological adoption remains constrained, it is essential to strengthen analytical capabilities and provide specialized training in spatial data processing. Such improvements are crucial for accurate data interpretation and scientifically informed decision-making.

Effective and sustainable management of marine habitats in the Central-Eastern Atlantic archipelagos require an integrated approach that combines biological and geophysical data to produce detailed bionomic maps. In the Azores, the initial lack of such integration resulted in the exclusion of keystone habitats, including CWC, from established MPAs. Therefore, regular updates and continuous refinement of habitat maps, supported by GIS tools and predictive modeling, are essential for accurate ecological assessments of anthropogenic pressures, ensuring the effectiveness of ecosystem-based conservation strategies.

At present time, stronger scientific cooperation about marine habitat mapping will foster more effective decision-making processes in the framework of evolving Marine Spatial Planning activities, where marine conservation sector usually is not so much represented and deserves a more intense focus.

In conclusion, overcoming the challenges identified in this review requires coordinated efforts integrating methodological standardization, investment in advanced technologies, and robust interregional collaboration. The successful implementation of these strategies will substantially enhance the precision of marine habitat mapping in the region, providing a solid foundation for the effective management and conservation on marine ecosystems in face of current and future environmental and anthropogenic pressures.

Supplementary Materials: Searching Formula (TITLE-ABS-KEY if searching with SCOPUS; and TS= if searching with Web of Science): TITLE-ABS-KEY // TS= ((“Macarones” OR “Webbnesia” OR “Azore” OR “Açore” OR “Madeira” OR “Wild Island” OR “Islas Salvajes” OR “Ilhas Salvagens” OR “Desertas Island” OR “Islas Desertas” OR “Canar” OR “Canary Island” OR “Islas Canarias” OR “Cabo Verde” OR “Cape Verde”) AND (“Acoustic” OR “Aerial” OR “Backscatter” OR “Bathymetr” OR “Drone” OR “GIS” OR “Hydroacoustic” OR “Hydroacoustic Survey” OR “LIDAR” OR “Multispectral Imag” OR “Radar” OR “Remote Sensing” OR “Satellite” OR “Scan” OR “Side Scan Sonar” OR “Sonar” OR “Sonic” OR “Sound” OR “Spatial Mapping” OR “SSS” OR “Telemetr” OR “Thermal Imag” OR “Cartograph” OR “Coastal Ecosystem Service” OR “Conservation” OR “Distribut” OR “Geospatial Data” OR “Habitat” OR “Habitat Connectiv” OR “Habitat Mapping” OR “Imaging” OR “Map” OR “Mapping” OR “Marine Environmental Assessment” OR “Marine Flora Conservation” OR “Marine Flora Mapping” OR “Marine Habitat Protection” OR “Marine Protected Area” OR “MBES” OR “Monitoring” OR “MPA” OR “Seabed Mapping” OR “Spatial Analys” OR “Multibeam”) AND (“Zostera” OR “Aquatic Vegetation” OR “Black Coral” OR “Coral” OR “Eelgrass” OR “Forest” OR “Garden” OR “Halodule” OR “Halophila” OR “Maerl” OR “MAF” OR “Nodule” OR “Rhodolith” OR “Seagrass” OR “Seaweed”))

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Appendix A

Table A1. Table of variables analyzed from the reviewed articles.

Pa per	Refer ence	Ye ar	Zone	Main objecti ve	Habitats			Hab itat map	Geolo gical map	De pth (m)	Subst rate	Area size (ha)
					Macrop hytes	Rhod olith beds	Co ral					
1	[175]	2021	Madeira	exploratory	-	x	-	x	-	35	mixed	2,368
2	[198]	2022	Madeira	monitoring	x	-	-	x	-	20	soft	-
3	[131]	2000	Azores	monitoring	x	-	-	-	-	40	hard	-
4	[186]	2000	Azores	exploratory	-	-	x	-	-	3800	hard	-
5	[203]	2008	Azores	monitoring	x	-	-	x	-	30	hard	400
6	[136]	2015	Canary Islands	exploratory	-	-	x	-	-	600	hard	-
7	[140]	2022	Madeira	exploratory	x	x	x	-	-	990	mixed	1,55
8	[165]	2017	Canary Islands	mapping	x	-	-	x	-	20	mixed	-
9	[158]	2022	Azores	exploratory	-	-	x	-	-	210	mixed	0,019
10	[166]	2015	Azores	mapping	x	x	x	x	-	n/a	mixed	167692,2
11	[146]	2023	Canary Islands	mapping	-	-	x	x	-	110	mixed	48,075
12	[116]	2012	Azores	exploratory	-	-	x	-	-	3,3	mixed	-
13	[129]	2017	Azores, Canary Islands	exploratory	x	-	x	-	-	734	mixed	-
14	[204]	1992	Azores	exploratory	x	-	-	x	-	5,5	hard	-

15	[185]	2013	Madeira	mappi ng	-	-	x	x	x	2660	soft	56000
16	[143]	2020	Azores	explor atory	-	-	x	-	x	1700	hard	0,045
17	[196]	2014	Canary Islands	monit oring	x	-	-	-	-	15	soft	393,9
18	[195]	2014	Azores	monit oring	-	-	x	-	x	1092	hard	-
19	[205]	2021	Azores	explor atory	-	-	x	-	x	595	hard	-
20	[206]	2013	Azores	explor atory	-	-	x	-	x	1097	mixe d	2,378
21	[176]	2013	Azores	explor atory	-	-	x	x	-	1500	hard	-
22	[15]	2019	Canary Islands	monit oring	-	x	-	x	-	50	mixe d	0,72
23	[164]	2021	Canary Islands	monit oring	x	-	-	-	-	75	soft	-
24	[14]	2020	Canary Islands	monit oring	-	x	-	-	-	40	soft	-
25	[163]	2024	Canary Islands	monit oring	x	x	-	x	-	25	mixe d	2150
26	[182]	2024	Cabo Verde	mappi ng	-	-	x	x	x	2100	hard	118800
27	[130]	2022	Canary Islands	monit oring	x	-	-	-	-	n/a	hard	-
28	[114]	2020	Madeira	mappi ng	x	x	x	x	-	50	mixe d	240
29	[56]	2024	Azores	mappi ng	x	-	x	x	-	60	mixe d	394,08
30	[207]	2018	Canary Islands	mappi ng	x	-	-	x	-	50	mixe d	9138
31	[202]	2013	Canary Islands	explor atory	x	-	-	-	-	19	mixe d	-
32	[208]	2019	Madeira	explor atory	x	-	-	-	-	2,6	mixe d	-
33	[209]	2006	Azores	explor atory	x	-	-	x	-	20	hard	-
34	[200]	2019	Canary Islands	explor atory	-	-	x	x	-	35	hard	0,018
35	[148]	2024	Canary Islands	monit oring	x	-	-	x	-	50	hard	-

36	[173]	20 21	Madeira	monit oring	x	-	-	-	-	12	soft	0,009
37	[4]	20 21	Azores	mappi ng	x	-	-	x	-	15	mixe d	-
38	[210]	20 12	Canary Islands	explor atory	x	-	-	-	-	46	mixe d	-
39	[211]	20 02	Canary Islands	mappi ng	x	-	-	x	-	10	mixe d	-
40	[212]	20 23	Canary Islands	mappi ng	x	-	-	x	-	n/a	soft	-
41	[213]	20 12	Azores	mappi ng	-	-	x	-	x	135 0	hard	-
42	[214]	20 17	Canary Islands	monit oring	x	-	-	x	-	20	hard	7,4
43	[215]	20 20	Canary Islands	mappi ng	x	-	-	x	-	40	mixe d	746
44	[162]	20 20	Canary Islands	mappi ng	x	-	-	x	-	40	hard	746
45	[194]	20 16	Canary Islands	mappi ng	-	-	x	-	x	250 0	mixe d	3673 88
46	[144]	20 20	Azores	explor atory	-	-	x	x	x	400 0	mixe d	-
47	[216]	20 08	Azores	explor atory	-	-	x	-	x	297 7	mixe d	-
48	[161]	20 15	Canary Islands	mappi ng	x	-	-	x	-	25	mixe d	-
49	[197]	20 23	Azores	mappi ng	-	-	x	x	-	238 7	mixe d	154
50	[142]	20 12	Azores	mappi ng	x	-	-	x	x	80	mixe d	5375 0
51	[217]	20 23	Azores	mappi ng	-	-	x	x	x	270 0	hard	-
52	[199]	20 21	Azores	explor atory	x	x	-	x	-	85	mixe d	-
53	[218]	20 23	Cabo Verde	explor atory	x	-	-	x	-	5	soft	0,62
54	[219]	20 23	Azores	explor atory	-	-	x	x	-	200 0	mixe d	2200 000
55	[220]	20 22	Canary Islands	mappi ng	x	-	-	x	-	n/a	mixe d	-
56	[221]	20 08	Azores	monit oring	x	x	x	-	-	30	mixe d	-

[illegible]

5	[203]	2008	x	-	-	-	-	x	-	-	-	-	-	-	x
6	[136]	2015	x	x	-	-	-	-	x	-	x	-	-	-	-
7	[140]	2022	-	x	-	-	x	-	-	-	-	-	x	-	-
8	[165]	2017	-	-	-	x	-	-	-	x	-	-	-	-	-
9	[158]	2022	-	x	-	-	-	-	-	-	-	-	x	-	-
10	[166]	2015	-	-	-	-	-	-	x	x	x	x	-	-	-
11	[146]	2023	x	x	-	-	x	-	-	x	x	x	-	-	-
12	[116]	2012	-	x	-	-	-	-	x	-	-	-	-	-	-
13	[129]	2017	x	x	-	-	-	-	-	-	-	-	x	-	-
14	[204]	1992	x	-	-	-	-	x	-	-	-	-	-	-	-
15	[185]	2013	-	x	-	-	x	x	-	x	x	x	-	-	-
16	[143]	2020	-	x	-	-	x	-	-	x	-	-	x	-	-
17	[196]	2014	x	-	-	-	-	x	-	x	-	-	-	-	-
18	[195]	2014	x	x	-	-	-	-	-	x	x	-	-	-	-
19	[205]	2021	-	x	-	-	-	-	-	-	-	-	-	-	-
20	[206]	2013	-	x	-	-	-	-	-	-	-	-	x	-	-
21	[176]	2013	x	x	-	-	-	x	x	-	-	-	-	-	-
22	[15]	2019	x	-	-	-	x	x	-	-	x	-	-	-	-
23	[164]	2021	-	-	-	x	-	-	x	x	x	-	-	-	-
24	[14]	2020	x	-	-	-	x	x	-	-	x	-	-	-	-
25	[163]	2024	-	-	-	x	-	-	-	x	-	-	-	x	-

26	[182]	20 24	-	x	-	-	x	-	x	x	-	-	x	x	-
27	[130]	20 22	x	-	-	-	-	-	x	x	x	-	-	-	-
28	[114]	20 20	x	-	-	-	x	-	x	-	x	-	-	-	-
29	[56]	20 24	-	x	-	-	x	-	-	-	x	x	-	-	-
30	[207]	20 18	x	x	-	-	-	x	-	x	-	-	-	x	-
31	[202]	20 13	x	-	-	-	-	x	-	-	-	-	-	-	x
32	[208]	20 19	x	-	-	-	-	x	-	-	-	-	x	-	-
33	[209]	20 06	x	-	-	-	-	x	-	-	-	-	-	x	-
34	[200]	20 19	x	-	-	-	-	x	-	-	-	-	-	-	-
35	[148]	20 24	-	-	x	-	-	-	x	x	x	-	x	-	-
36	[173]	20 21	x	-	-	-	-	x	-	-	x	-	x	-	-
37	[4]	20 21	x	-	x	-	-	x	-	x	x	-	x	x	-
38	[210]	20 12	x	-	-	-	-	x	-	-	-	-	-	-	-
39	[211]	20 02	-	x	-	-	-	-	-	x	-	-	x	-	-
40	[212]	20 23	x	-	-	-	x	-	x	x	x	-	-	-	-
41	[213]	20 12	-	-	-	-	-	x	-	-	-	-	-	-	-
42	[214]	20 17	x	-	-	-	-	x	-	-	x	-	-	-	-
43	[215]	20 20	-	-	-	x	-	-	x	x	-	-	-	-	-
44	[162]	20 20	x	-	-	-	-	x	x	x	-	-	-	-	-
45	[194]	20 16	-	x	-	-	x	x	-	x	x	x	-	-	-
46	[144]	20 20	-	x	-	-	x	-	-	x	-	x	-	-	-

47	[216]	2008	-	x	-	-	-	-	x	-	-	-	-	-	-	x
48	[161]	2015	-	-	-	x	-	-	-	-	-	-	-	-	x	-
49	[197]	2023	-	x	-	-	-	-	-	-	x	-	x	-	-	-
50	[142]	2012	x	x	-	-	x	x	x	x	x	x	-	-	-	-
51	[217]	2023	-	x	-	-	x	-	-	-	x	-	x	-	-	-
52	[199]	2021	x	x	-	-	-	-	x	-	-	-	-	-	-	-
53	[218]	2023	x	-	-	-	-	-	-	x	-	x	-	-	-	-
54	[219]	2023	-	-	-	-	-	-	x	-	x	x	-	-	-	-
55	[220]	2022	-	-	-	-	-	-	-	x	x	-	-	-	-	x
56	[221]	2008	x	-	-	-	-	-	-	-	-	-	-	-	-	x
57	[46]	2020	x	-	-	-	-	x	x	-	-	-	x	x	-	-
58	[201]	2021	-	x	-	-	-	x	x	-	-	x	x	x	-	-
59	[141]	2013	-	x	-	-	-	x	-	-	-	-	x	-	-	-
60	[183]	2020	x	-	-	-	-	-	-	-	-	-	x	-	-	-
61	[184]	2013	-	x	-	-	-	x	-	-	-	x	x	x	-	x
62	[150]	2019	x	-	x	-	-	-	-	-	-	x	x	-	x	-
63	[167]	2022	-	x	-	-	-	-	-	-	-	-	-	x	x	-
64	[115]	2024	-	x	-	-	-	-	-	x	-	-	x	x	-	-
65	[222]	2013	-	-	-	-	-	x	-	-	-	x	x	x	-	-
66	[160]	2023	-	-	-	x	-	-	-	-	-	x	x	-	x	-
67	[151]	2024	x	-	x	-	-	-	x	-	-	x	x	-	x	x

68	[149]	20	-	x	x	-	-	-	-	x	x	-	x	x	-
		21													
69	[120]	20	x	-	-	-	-	x	-	x	-	-	x	x	-
		08													

Table A2. Table of species observed and mapped in the reviewed papers.

<i>Species</i>	<i>Order</i>	<i>Family</i>	St ud ie s	A zo re s	M ad eir a	Can a ry Is lan ds	Cab o Ver de	Atlant ic Mid- Ridge
<i>Phyllum Cnidaria</i>								
<i>Subphyllum Anthozoa</i>								
Anthozoa			1	x	-	-	-	-
<i>Class Hexacorallia</i>								
<i>Elatopathes aff. abietina (Pourtalès, 1874)</i>	Antipath aria	Aphanipat hidae	1	x	-	-	-	-
Actiniaria	Actiniari a		4	x	-	-	x	x
<i>Anemonia viridis (Forsskål, 1775)</i>	Actiniari a	Actiniidae	1	-	x	-	-	-
<i>Actinoscyphia aurelia (Stephenson, 1918)</i>	Actiniari a	Actinoscy phiidae	1	-	-	x	-	-
<i>Parasicyonis ingolfi Carlgren, 1942</i>	Actiniari a	Actinostoli dae	1	x	-	-	-	x
<i>Parasicyonis Carlgren, 1921</i>	Actiniari a	Actinostoli dae	1	x	-	-	-	x
<i>Telmatactis cricoides (Duchassaing, 1850)</i>	Actiniari a	Andvakiid ae	1	-	x	-	-	-
Antipatharia	Antipath aria		3	x	-	-	-	x
<i>Antipathes furcata Gray, 1857</i>	Antipath aria	Antipathid ae	1	-	-	x	-	-
<i>Antipathes grayi (Roule, 1902)</i>	Antipath aria	Antipathid ae	1	x	-	-	-	-
<i>Antipathes virgata Esper, 1798</i>	Antipath aria	Antipathid ae	1	x	-	-	-	-
<i>Antipathes Pallas, 1766</i>	Antipath aria	Antipathid ae	2	x	x	-	-	-
<i>Stichopathes flagellum Roule, 1902</i>	Antipath aria	Antipathid ae	1	x	-	-	-	-
<i>Stichopathes gracilis (Gray, 1857)</i>	Antipath aria	Antipathid ae	1	x	-	x	-	x

<i>Stichopathes gravieri</i> Molodtsova, 2006	Antipatharia	Antipathidae	2	x	-	-	-	x
<i>Stichopathes richardi</i> Roule, 1902	Antipatharia	Antipathidae	1	x	-	-	-	-
<i>Stichopathes</i> Brook, 1889	Antipatharia	Antipathidae	4	x	x	x	-	x
<i>Elatopathes</i> Opresko, 2004	Antipatharia	Aphanipathidae	1	x	-	-	-	x
<i>Aphanostichopathes dissimilis</i> (Roule, 1902)	Antipatharia	Aphanipathidae	1	x	-	-	-	-
<i>Distichopathes</i> Opresko, 2004	Antipatharia	Aphanipathidae	2	x	x	-	-	-
<i>Phanopathes erinaceus</i> (Roule, 1905)	Antipatharia	Aphanipathidae	2	x	-	-	-	-
<i>Heteropathes</i> Opresko, 2011	Antipatharia	Cladopathidae	1	x	-	-	-	x
<i>Leiopathes expansa</i> Johnson, 1899	Antipatharia	Leiopathidae	2	x	-	-	-	x
<i>Leiopathes glaberrima</i> (Esper, 1792)	Antipatharia	Leiopathidae	3	x	-	x	-	-
<i>Leiopathes grimaldii</i> Roule, 1902	Antipatharia	Leiopathidae	1	x	-	-	-	-
<i>Leiopathes</i> Haime, 1849	Antipatharia	Leiopathidae	1	x	-	x	-	x
<i>Leiopathes</i> Haime, 1849	Antipatharia	Leiopathidae	4	x	-	x	-	x
<i>Antipathella subpinnata</i> (Ellis & Solander, 1786)	Antipatharia	Myriopathidae	4	x	-	x	-	x
<i>Antipathella wollastoni</i> (Gray, 1857)	Antipatharia	Myriopathidae	8	x	-	x	-	x
<i>Antipathella</i> Brook, 1889	Antipatharia	Myriopathidae	3	x	-	x	-	x
<i>Tanacetipathes squamosa</i> (Koch, 1886)	Antipatharia	Myriopathidae	1	x	-	-	-	-
<i>Tanacetipathes</i> Opresko, 2001	Antipatharia	Myriopathidae	4	x	x	-	-	x
<i>Bathypathes patula</i> Brook, 1889	Antipatharia	Schizopathidae	1	x	-	-	-	-
<i>Bathypathes</i> Brook, 1889	Antipatharia	Schizopathidae	3	x	-	x	-	x
<i>Stauropathes punctata</i> (Roule, 1905)	Antipatharia	Schizopathidae	1	x	-	-	-	-

<i>Stauropathes arctica</i> (Lütken, 1871)	Antipatharia	Schizopathidae	1	x	-	-	-	x
<i>Parantipathes hirondelle</i> Molodtsova, 2006	Antipatharia	Schizopathidae	2	x	-	-	-	-
<i>Parantipathes</i> Brook, 1889	Antipatharia	Schizopathidae	2	x	x	-	-	x
Ceriantharia	Ceriantharia		1	x	-	-	-	-
<i>Pachycerianthus</i> Roule, 1904	Ceriantharia	Cerianthidae	1	-	x	-	-	-
Scleractinia	Scleractinia		2	x	-	-	-	x
<i>Coenocyathus cylindricus</i> Milne Edwards & Haime, 1848	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Concentrotheca laevigata</i> (Pourtalès, 1871)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Dasmosmilia lymani</i> (Pourtalès, 1871)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Dasmosmilia variegata</i> (Pourtalès, 1871)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Desmophyllum dianthus</i> (Esper, 1794)	Scleractinia	Caryophylliidae	5	x	-	-	-	x
<i>Desmophyllum pertusum</i> (Linnaeus, 1758)	Scleractinia	Caryophylliidae	8	x	-	x	-	x
<i>Desmophyllum</i> Ehrenberg, 1834	Scleractinia	Caryophylliidae	2	x	-	x	-	x
<i>Pourtalosmilia anthophyllites</i> (Ellis & Solander, 1786)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Premocyathus cornuformis</i> (Pourtalès, 1868)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Anomocora fecunda</i> (Pourtalès, 1871)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Paracyathus pulchellus</i> (Philippi, 1842)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Phyllangia americana</i> Milne Edwards & Haime, 1849	Scleractinia	Caryophylliidae	1	-	-	x	-	-
<i>Polycyathus muelleriae</i> (Abel, 1959)	Scleractinia	Caryophylliidae	1	-	-	x	-	-
<i>Polycyathus senegalensis</i> Chevalier, 1966	Scleractinia	Caryophylliidae	1	-	-	x	-	-
<i>Aulocyathus atlanticus</i> Zibrowius, 1980	Scleractinia	Caryophylliidae	2	x	x	-	-	x

<i>Caryophyllia (Caryophyllia) abyssorum</i> Duncan, 1873	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Caryophyllia (Caryophyllia) alberti</i> Zibrowius, 1980	Scleractinia	Caryophylliidae	3	x	-	-	-	x
<i>Caryophyllia (Caryophyllia) atlantica</i> (Duncan, 1873)	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Caryophyllia (Caryophyllia) calveri</i> Duncan, 1873	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Caryophyllia (Caryophyllia) cyathus</i> (Ellis & Solander, 1786)	Scleractinia	Caryophylliidae	2	x	-	-	-	x
<i>Caryophyllia (Caryophyllia) foresti</i> Zibrowius, 1980	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Caryophyllia (Caryophyllia) inornata</i> (Duncan, 1878)	Scleractinia	Caryophylliidae	2	x	-	x	-	-
<i>Caryophyllia (Caryophyllia) sarsiae</i> Zibrowius, 1974	Scleractinia	Caryophylliidae	2	x	-	-	-	x
<i>Caryophyllia (Caryophyllia) smithii</i> Stokes & Broderip, 1828	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Caryophyllia (Caryophyllia) Lamarck</i> , 1801	Scleractinia	Caryophylliidae	5	x	-	x	-	x
<i>Caryophylliidae</i> Dana, 1846	Scleractinia	Caryophylliidae	2	x	-	-	-	x
<i>Solenosmilia variabilis</i> Duncan, 1873	Scleractinia	Caryophylliidae	5	x	-	x	-	x
<i>Solenosmilia</i> Duncan, 1873	Scleractinia	Caryophylliidae	1	x	-	-	-	x
<i>Tethocyathus variabilis</i> Cairns, 1979	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Trochocyathus (Trochocyathus) spinosocostatus</i> Zibrowius, 1980	Scleractinia	Caryophylliidae	1	x	-	-	-	-
<i>Deltocyathus eccentricus</i> Cairns, 1979	Scleractinia	Deltocyathidae	2	x	x	-	-	x
<i>Deltocyathus italicus</i> (Michelotti, 1838)	Scleractinia	Deltocyathidae	1	x	-	-	-	-
<i>Deltocyathus moseleyi</i> Cairns, 1979	Scleractinia	Deltocyathidae	2	x	x	-	-	x
<i>Dendrophyllia alternata</i> Pourtales, 1880	Scleractinia	Dendrophylliidae	2	x	-	x	-	x
<i>Dendrophyllia cornigera</i> (Lamarck, 1816)	Scleractinia	Dendrophylliidae	4	x	x	x	-	x
<i>Dendrophyllia ramea</i> (Linnaeus, 1758)	Scleractinia	Dendrophylliidae	4	x	x	x	-	x

<i>Dendrophyllia de Blainville, 1830</i>	Scleractinia	Dendrophylliidae	2	x	-	-	-	x
<i>Enallopsammia pusilla (Alcock, 1902)</i>	Scleractinia	Dendrophylliidae	1	x	-	-	-	-
<i>Enallopsammia rostrata (Pourtalès, 1878)</i>	Scleractinia	Dendrophylliidae	4	x	-	-	x	x
<i>Enallopsammia Michelloti, 1871</i>	Scleractinia	Dendrophylliidae	1	x	-	-	-	x
<i>Leptopsammia formosa (Gravier, 1915)</i>	Scleractinia	Dendrophylliidae	2	x	-	-	-	-
<i>Balanophyllia (Balanophyllia) cellulosa Duncan, 1873</i>	Scleractinia	Dendrophylliidae	2	x	-	-	-	x
<i>Tubastraea coccinea Lesson, 1830</i>	Scleractinia	Dendrophylliidae	1	-	-	x	-	-
<i>Tubastraea tagusensis Wells, 1982</i>	Scleractinia	Dendrophylliidae	1	-	-	x	-	-
<i>Tubastraea Lesson, 1830</i>	Scleractinia	Dendrophylliidae	1	-	-	x	-	-
<i>Javania cailleti (Duchassaing & Michelotti, 1864)</i>	Scleractinia	Flabellidae	1	x	-	-	-	-
<i>Javania pseudoalabastra Zibrowius, 1974</i>	Scleractinia	Flabellidae	1	x	-	-	-	-
<i>Flabellum (Flabellum) chunii Marenzeller, 1904</i>	Scleractinia	Flabellidae	1	x	-	-	-	-
<i>Flabellum (Ulocyathus) alabastrum Moseley, 1876</i>	Scleractinia	Flabellidae	2	x	-	-	-	x
<i>Flabellum (Ulocyathus) angulare Moseley, 1876</i>	Scleractinia	Flabellidae	2	x	-	-	-	x
<i>Flabellum (Ulocyathus) macandrewi Gray, 1849</i>	Scleractinia	Flabellidae	2	x	-	-	-	x
<i>Flabellum Lesson, 1831</i>	Scleractinia	Flabellidae	3	x	-	-	-	x
<i>Fungiacyathus (Bathyactis) crispus (Portalès, 1871)</i>	Scleractinia	Fungiacyathidae	2	x	-	-	-	x
<i>Fungiacyathus (Bathyactis) marenzelleri (Vaughan, 1906)</i>	Scleractinia	Fungiacyathidae	1	x	-	-	-	-
<i>Fungiacyathus (Bathyactis) symmetricus (Portalès, 1871)</i>	Scleractinia	Fungiacyathidae	1	x	-	-	-	-
<i>Fungiacyathus (Fungiacyathus) fragilis Sars, 1872</i>	Scleractinia	Fungiacyathidae	3	x	-	-	-	x
<i>Guynia annulata Duncan, 1872</i>	Scleractinia	Guyniidae	1	x	-	-	-	-

<i>Madrepora oculata</i> Linnaeus, 1758	Scleractinia	Madreporidae	8	x	x	x	-	x
<i>Oculina patagonica</i> de Angelis D'Ossat, 1908	Scleractinia	Oculinidae	1	-	-	x	-	-
<i>Oculina</i> Lamarck, 1816	Scleractinia	Oculinidae	1	-	-	x	-	-
<i>Madracis pharensis</i> (Heller, 1868)	Scleractinia	Pocilloporidae	1	x	-	-	-	-
<i>Madracis profunda</i> Zibrowius, 1980	Scleractinia	Pocilloporidae	1	x	-	-	-	-
<i>Madracis</i> Milne Edwards & Haime, 1849	Scleractinia	Pocilloporidae	1	-	x	-	-	-
<i>Culicia tenella</i> Dana, 1846	Scleractinia	Rhizangiidae	1	-	-	x	-	-
<i>Culicia</i> Dana, 1846	Scleractinia	Rhizangiidae	1	-	-	x	-	-
<i>Schizocyathidae</i> Stolarski, 2000	Scleractinia	Schizocyathidae	1	x	-	-	-	x
<i>Schizocyathus fissilis</i> Pourtalès, 1874	Scleractinia	Schizocyathidae	1	x	-	-	-	-
<i>Stenocyathus vermiformis</i> (Portalès, 1868)	Scleractinia	Stenocyathidae	3	x	x	-	-	x
<i>Stephanocyathus</i> (<i>Odontocyathus</i>) <i>nobilis</i> (Moseley, 1876)	Scleractinia	Stephanocyathidae	1	x	-	-	-	-
<i>Stephanocyathus</i> (<i>Stephanocyathus</i>) <i>crassus</i> (Jourdan, 1895)	Scleractinia	Stephanocyathidae	1	x	-	-	-	-
<i>Stephanocyathus</i> (<i>Stephanocyathus</i>) <i>diadema</i> (Moseley, 1876)	Scleractinia	Stephanocyathidae	1	x	-	-	-	-
<i>Stephanocyathus</i> (<i>Stephanocyathus</i>) <i>moseleyanus</i> (Sclater, 1886)	Scleractinia	Stephanocyathidae	2	x	-	-	-	x
<i>Vaughanella concinna</i> Gravier, 1915	Scleractinia	Stephanocyathidae	1	x	-	-	-	-
<i>Peponocyathus stimpsonii</i> (Portalès, 1871)	Scleractinia	Turbinolidae	1	x	-	-	-	-
<i>Sphenotrochus</i> (<i>Sphenotrochus</i>) <i>andrewianus</i> Milne Edwards & Haime, 1848	Scleractinia	Turbinolidae	1	x	-	-	-	-
<i>Thrypticotrochus</i> Cairns, 1989	Scleractinia	Turbinolidae	1	-	-	-	-	x
<i>Peponocyathus folliculus</i> (Portalès, 1868)	Scleractinia	Turbinolidae	2	x	x	-	-	x

Zoantharia	Zoanthar ia		1	x	-	-	-	x
<i>Epizoanthus martinsae</i> Carreiro-Silva, <i>Ocaña, Stanković, Sampaio, Porteiro, Fabri & Stefanni, 2017</i>	Zoanthar ia	Epizoanthi dae	1	x	-	-	-	-
<i>Parazoanthus</i> Haddon & Shackleton, 1891	Zoanthar ia	Parazoant hidae	1	x	-	x	-	x
<i>Savalia savaglia</i> (Bertoloni, 1819)	Zoanthar ia	Parazoant hidae	1	-	-	x	-	-
<i>Palythoa canariensis</i> Haddon & Duerden, 1896	Zoanthar ia	Sphenopid ae	1	-	-	x	-	-
<i>Zoanthidae</i> Rafinesque, 1815	Zoanthar ia	Zoanthida e	2	x	-	-	-	x
Class Octocorallia								
Octocorallia			5	x	-	x	x	x
<i>Rolandia coralloides</i> de Lacaze Duthiers, 1900	Alcyonac ea	Clavularii dae	1	x	-	-	-	-
Malacalcyonacea	Malacalc yonacea		1	x	-	-	-	-
<i>Acanthogorgia armata</i> Verrill, 1878	Malacalc yonacea	Acanthogo rgiidae	3	x	-	-	-	x
<i>Acanthogorgia aspera</i> Pourtalès, 1867	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Acanthogorgia hirsuta</i> Gray, 1857	Malacalc yonacea	Acanthogo rgiidae	3	x	-	x	-	x
<i>Acanthogorgia muricata</i> Verrill, 1883	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Acanthogorgia pico</i> Grasshoff, 1973	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Acanthogorgia</i> sp. Gray, 1857	Malacalc yonacea	Acanthogo rgiidae	4	x	-	x	-	x
<i>Bebryce mollis</i> Philippi, 1842	Malacalc yonacea	Acanthogo rgiidae	3	x	x	-	-	x
<i>Dentomuricea meteor</i> Grasshoff, 1977	Malacalc yonacea	Acanthogo rgiidae	7	x	x	x	-	x
<i>Dentomuricea</i> Grasshoff, 1977	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	x
<i>Muriceides lepida</i> Carpine & Grasshoff, 1975	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Muriceides sceptrum</i> (Studer, 1891)	Malacalc yonacea	Acanthogo rgiidae	2	x	-	-	-	x

<i>Placogorgia becena</i> Grasshoff, 1977	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Placogorgia coronata</i> Carpine & Grasshoff, 1975	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Placogorgia intermedia</i> (Thomson, 1927)	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Villogorgia bebrycoides</i> (von Koch, 1887)	Malacalc yonacea	Acanthogo rgiidae	2	x	-	-	-	x
<i>Gersemia clavata</i> (Danielssen, 1887)	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Azoriella bayeri</i> (López-González & Gili, 2001)	Malacalc yonacea	Cerveridae	1	x	-	-	-	-
<i>Clavularia arctica</i> (Sars, 1860)	Malacalc yonacea	Clavularii dae	1	x	-	-	-	-
<i>Clavularia armata</i> Thomson, 1927	Malacalc yonacea	Clavularii dae	1	x	-	-	-	-
<i>Clavularia charcoti</i> (Tixier-Durivault & d'Hondt, 1974)	Malacalc yonacea	Clavularii dae	1	x	-	-	-	-
<i>Clavularia elongata</i> Wright & Studer, 1889	Malacalc yonacea	Clavularii dae	1	x	-	-	-	-
<i>Clavularia marioni</i> von Koch, 1890	Malacalc yonacea	Clavularii dae	1	x	-	-	-	-
<i>Clavularia tenuis</i> Tixier-Durivault & d'Hondt, 1974	Malacalc yonacea	Clavularii dae	1	x	-	-	-	-
<i>Clavularia</i> Blainville, 1830	Malacalc yonacea	Clavularii dae	1	x	-	-	-	x
<i>Schizophytum echinatum</i> Studer, 1891	Malacalc yonacea	Clavularii dae	2	x	-	-	-	x
<i>Eunicella verrucosa</i> (Pallas, 1766)	Malacalc yonacea	Eunicellid ae	1	-	x	-	-	-
<i>Eunicella</i> Verrill, 1869	Malacalc yonacea	Eunicellid ae	1	-	x	-	-	-
<i>Pseudotelestula humilis</i> (Thomson, 1927)	Malacalc yonacea	Incrustatid ae	1	x	-	-	-	-
<i>Thesea rigida</i> (Thomson, 1927)	Malacalc yonacea	Malacalcy onacea incertae sedis	1	x	-	-	-	-
<i>Nephtheidae</i> Gray, 1862	Malacalc yonacea	Nephtheid ae	1	-	x	-	-	-
<i>Scleronephthya macrospina</i> Thomson, 1927	Malacalc yonacea	Nephtheid ae	1	x	-	-	-	-

<i>Plexauridae</i> Gray, 1859	Malacalc yonacea	Plexaurida e	3	x	-	-	-	x
<i>Dacrygorgia modesta</i> (Verrill, 1883)	Malacalc yonacea	Pterogorgi idae	1	x	-	-	-	-
<i>Sarcophyton</i> Lesson, 1834	Malacalc yonacea	Sarcophyti dae	1	-	x	-	-	-
<i>Bathytelesto rigida</i> (Wright & Studer, 1889)	Malacalc yonacea	Tubiporid ae	1	x	-	-	-	-
<i>Scyphopodium ingolfi</i> (Madsen, 1944)	Malacalc yonacea	Tubiporid ae	1	x	-	-	-	-
<i>Scleranthelia rugosa</i> (Pourtalès, 1867)	Octocora llia incertae sedis	Octocoralli a incertae sedis	1	x	-	-	-	-
Scleralcyonacea	Scleralcy onacea		2	x	-	-	x	x
<i>Chelidonisis aurantiaca</i> Studer, 1890	Scleralcy onacea	Chelidonis ididae	2	x	-	-	-	-
<i>Chrysogorgia agassizii</i> (Verrill, 1883)	Scleralcy onacea	Chrysogor giidae	3	x	-	-	-	x
<i>Chrysogorgia fewkesii</i> Verrill, 1883	Scleralcy onacea	Chrysogor giidae	1	x	-	-	-	-
<i>Chrysogorgia quadruplex</i> Thomson, 1927	Scleralcy onacea	Chrysogor giidae	1	x	-	-	-	-
<i>Chrysogorgia Duchassaing & Michelotti</i> , 1864	Scleralcy onacea	Chrysogor giidae	3	x	-	x	-	x
<i>Parachrysogorgia squamata</i> (Verrill, 1883)	Scleralcy onacea	Chrysogor giidae	1	x	-	-	-	-
<i>Radicipes gracilis</i> (Verrill, 1884)	Scleralcy onacea	Chrysogor giidae	2	x	-	-	-	-
<i>Chrysogorgia elegans</i> (Verrill, 1883)	Scleralcy onacea	Chrysogor giidae	1	x	-	-	-	-
<i>Pleurocorallium johnsoni</i> (Gray, 1860)	Scleralcy onacea	Coralliidae	3	x	-	-	-	x
<i>Coralliidae</i> Lamouroux, 1812	Scleralcy onacea	Coralliidae	4	x	-	-	-	x
<i>Cornularia cornucopiae</i> (Pallas, 1766)	Scleralcy onacea	Cornularii dae	1	x	-	-	-	-
<i>Viminella flagellum</i> (Johnson, 1863)	Scleralcy onacea	Ellisellidae	8	x	x	x	-	x
<i>Acanella arbuscula</i> (Johnson, 1862)	Scleralcy onacea	Keratoisidi dae	5	x	-	x	x	x

<i>Calyptrophora trilepis</i> (Pourtalès, 1868)	Scleralcy onacea	Primnoida e	1	x	-	x	-	x
<i>Candidella imbricata</i> (Johnson, 1862)	Scleralcy onacea	Primnoida e	5	x	-	-	-	x
<i>Narella bellissima</i> (Kükenthal, 1915)	Scleralcy onacea	Primnoida e	4	x	-	-	-	x
<i>Narella versluysi</i> (Hickson, 1909)	Scleralcy onacea	Primnoida e	3	x	-	-	-	x
<i>Paracalyptrophora josephinae</i> (Lindström, 1877)	Scleralcy onacea	Primnoida e	4	x	-	-	-	x
<i>Primnoidae</i> Milne Edwards, 1857	Scleralcy onacea	Primnoida e	3	x	-	-	-	x
<i>Thouarella</i> (<i>Euthouarella</i>) <i>grasshoffi</i> Cairns, 2006	Scleralcy onacea	Primnoida e	1	x	-	-	-	-
<i>Thouarella</i> (<i>Euthouarella</i>) <i>hilgendorfi</i> (Studer, 1879)	Scleralcy onacea	Primnoida e	1	x	-	-	-	x
<i>Thouarella</i> (<i>Thouarella</i>) <i>variabilis</i> Wright & Studer, 1889	Scleralcy onacea	Primnoida e	1	x	-	-	-	-
<i>Thouarella</i> Gray, 1870	Scleralcy onacea	Primnoida e	2	x	-	-	-	x
<i>Sarcodictyon catenatum</i> Forbes in Johnston, 1847	Scleralcy onacea	Sarcodicty onidae	2	x	-	-	-	x
<i>Telestula batoni</i> Weinberg, 1990	Scleralcy onacea	Sarcodicty onidae	1	x	-	-	-	-
<i>Telestula kuekenthali</i> Weinberg, 1990	Scleralcy onacea	Sarcodicty onidae	1	x	-	-	-	-
<i>Telestula tubaria</i> Wright & Studer, 1889	Scleralcy onacea	Sarcodicty onidae	1	x	-	-	-	-
<i>Scleroptilum grandiflorum</i> Kölliker, 1880	Scleralcy onacea	Scleroptili dae	1	x	-	-	-	x
<i>Titanideum obscurum</i> Thomson, 1927	Scleralcy onacea	Spongiode rmidae	1	x	-	-	-	-
<i>Paramuricea candida</i> Grasshoff, 1977	Malacalc yonacea	Acanthogo rgiidae	1	x	-	-	-	-
<i>Paramuricea grayi</i> (Johnson, 1861)	Malacalc yonacea	Acanthogo rgiidae	1	-	x	-	-	-
<i>Paramuricea</i> Kölliker, 1865	Malacalc yonacea	Acanthogo rgiidae	5	x	-	x	x	x
<i>Alcyoniidae</i> Lamouroux, 1812	Malacalc yonacea	Alcyoniida e	3	x	-	-	-	x
<i>Alcyonium bocagei</i> (Saville Kent, 1870)	Malacalc yonacea	Alcyoniida e	2	x	-	-	-	x

<i>Alcyonium burmedju</i> Sampaio, Stokvis & van Ofwegen, 2016	Malacalc yonacea	Alcyoniida e	2	x	-	-	-	x
<i>Alcyonium maristenebrosi</i> (Stiasny, 1937)	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Alcyonium palmatum</i> Pallas, 1766	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Alcyonium profundum</i> Stokvis & van Ofwegen, 2006	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Alcyonium</i> Linnaeus, 1758	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	x
<i>Anthothela grandiflora</i> (Sars, 1856)	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Bellonella tenuis</i> Tixier-Durivault & d'Hondt, 1974	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Bellonella variabilis</i> (Studer, 1891)	Malacalc yonacea	Alcyoniida e	1	x	-	-	-	-
<i>Lateothela grandiflora</i> (Tixier-Durivault & d'Hondt, 1974)	Malacalc yonacea	Alcyoniida e	2	x	-	-	-	-
<i>Isididae</i> Lamouroux, 1812	Malacalc yonacea	Isididae	2	x	-	-	-	x
<i>Paralcyonium spinulosum</i> (Delle Chiaje, 1822)	Malacalc yonacea	Paralcyoni idae	1	x	-	-	-	-
<i>Swiftia dubia</i> (Thomson, 1929)	Malacalc yonacea	Plexaurida e	2	x	-	-	-	x
<i>Swiftia Duchassaing & Michelotti, 1864</i>	Malacalc yonacea	Plexaurida e	2	x	-	x	-	-
<i>Pennatuloides</i> Ehrenberg, 1834	Scleralcy onacea		2	x	-	-	-	x
<i>Anthoptilum</i> Kölliker, 1880	Scleralcy onacea	Anthoptili dae	1	x	-	-	-	x
<i>Iridogorgia fontinalis</i> Watling, 2007	Scleralcy onacea	Chrysogor giidae	1	x	-	-	-	-
<i>Iridogorgia pourtalesii</i> Verrill, 1883	Scleralcy onacea	Chrysogor giidae	1	x	-	-	-	-
<i>Iridogorgia</i> Verrill, 1883	Scleralcy onacea	Chrysogor giidae	2	x	-	-	x	x
<i>Metallogorgia melanotrichos</i> (Wright & Studer, 1889)	Scleralcy onacea	Chrysogor giidae	2	x	-	x	-	-
<i>Metallogorgia</i> Versluys, 1902	Scleralcy onacea	Chrysogor giidae	1	-	-	-	x	-
<i>Corallium</i> Cuvier, 1798	Scleralcy onacea	Coralliidae	2	x	-	x	-	x

<i>Hemicorallium niobe</i> (Bayer, 1964)	Scleralcy onacea	Coralliidae	3	x	-	x	-	-
<i>Hemicorallium tricolor</i> (Johnson, 1899)	Scleralcy onacea	Coralliidae	2	x	-	x	-	-
<i>Paragorgia arborea</i> (Linnaeus, 1758)	Scleralcy onacea	Coralliidae	1	x	-	-	-	x
<i>Paragorgia johnsoni</i> Gray, 1862	Scleralcy onacea	Coralliidae	4	x	-	-	-	x
<i>Anthomastus canariensis</i> Wright & Studer, 1889	Scleralcy onacea	Coralliidae	1	x	-	-	-	-
<i>Anthomastus grandiflorus</i> Verrill, 1878	Scleralcy onacea	Coralliidae	1	x	-	-	-	-
<i>Anthomastus</i> Verrill, 1878	Scleralcy onacea	Coralliidae	4	x	-	x	-	x
<i>Pseudoanthomastus agaricus</i> (Studer, 1890)	Scleralcy onacea	Coralliidae	2	x	-	-	-	x
<i>Pseudoanthomastus Tixier-Durivault & d'Hondt, 1974</i>	Scleralcy onacea	Coralliidae	1	x	-	-	-	-
<i>Nicella granifera</i> (Köl liker, 1865)	Scleralcy onacea	Ellisellidae	1	x	-	-	-	-
<i>Gyrophyllum hirondellei</i> Studer, 1891	Scleralcy onacea	Gyrophylli dae	1	x	-	-	-	-
<i>Isidella longiflora</i> (Verrill, 1883)	Scleralcy onacea	Keratoisidi dae	1	x	-	-	-	-
<i>Lepidisis cyanae</i> Grasshoff, 1986	Scleralcy onacea	Keratoisidi dae	1	x	-	-	-	-
<i>Lepidisis</i> Verrill, 1883	Scleralcy onacea	Keratoisidi dae	1	x	-	-	-	-
<i>Keratoisididae</i> Gray, 1870	Scleralcy onacea	Keratoisidi dae	1	x	-	-	-	x
<i>Keratoisis grayi</i> Wright, 1869	Scleralcy onacea	Keratoisidi dae	1	x	-	-	-	-
<i>Keratoisis</i> Wright, 1869	Scleralcy onacea	Keratoisidi dae	3	x	-	x	-	x
<i>Pennatula</i> Linnaeus, 1758	Scleralcy onacea	Pennatulid ae	1	-	-	x	-	-
<i>Callogorgia verticillata</i> (Pallas, 1766)	Scleralcy onacea	Primnoida e	4	x	-	x	-	x
<i>Paracalyptrophora</i> Kinoshita, 1908	Scleralcy onacea	Primnoida e	1	x	-	-	-	x
<i>Umbellula</i> Cuvier, [1797]	Scleralcy onacea	Umbelluli dae	1	x	-	-	-	x

<i>Veretillum cynomorium</i> (Pallas, 1766)	Scleralcyonacea	Veretillidae	1	-	x	-	-	-
Subphyllum Medusozoa								
Class Hydrozoa								
Hydrozoa			1	x	-	-	-	x
<i>Candelabrum phrygium</i> (Fabricius, 1780)	Anthoathecata	Candelabridae	1	x	-	-	-	x
<i>Candelabrum serpentarii</i> Segonzac & Vervoort, 1995	Anthoathecata	Candelabridae	1	x	-	-	-	x
<i>Eudendrium</i> Ehrenberg, 1834	Anthoathecata	Eudendriidae	1	x	-	-	-	x
<i>Pennaria disticha</i> Goldfuss, 1820	Anthoathecata	Pennariidae	1	-	x	-	-	-
<i>Errina atlantica</i> Hickson, 1912	Anthoathecata	Stylasteridae	2	x	-	-	-	x
<i>Errina dabneyi</i> (Pourtalès, 1871)	Anthoathecata	Stylasteridae	5	x	-	x	-	x
<i>Errina</i> Gray, 1835	Anthoathecata	Stylasteridae	2	x	x	-	-	x
<i>Stenohelia</i> Kent, 1870	Anthoathecata	Stylasteridae	2	x	x	-	-	x
<i>Stylaster</i> Gray, 1831	Anthoathecata	Stylasteridae	1	-	x	-	-	x
<i>Stylasteridae</i> Gray, 1847	Anthoathecata	Stylasteridae	5	x	-	-	-	x
<i>Pliobothrus symmetricus</i> Pourtalès, 1868	Anthoathecata	Stylasteridae	2	x	-	-	-	x
<i>Crypthelia affinis</i> Moseley, 1879	Anthoathecata	Stylasteridae	1	x	-	-	-	-
<i>Crypthelia medioatlantica</i> Zibrowius & Cairns, 1992	Anthoathecata	Stylasteridae	1	x	-	-	-	-
<i>Crypthelia tenuiseptata</i> Cairns, 1986	Anthoathecata	Stylasteridae	1	x	-	-	-	-
<i>Crypthelia vascomarquesi</i> Zibrowius & Cairns, 1992	Anthoathecata	Stylasteridae	1	x	-	-	-	-
<i>Crypthelia</i> Milne Edwards & Haime, 1849	Anthoathecata	Stylasteridae	2	x	-	-	-	x
<i>Lepidopora eburnea</i> (Calvet, 1903)	Anthoathecata	Stylasteridae	1	x	-	-	-	-
<i>Lepidopora</i> Pourtalès, 1871	Anthoathecata	Stylasteridae	1	-	x	-	-	x

<i>Ectopleura crocea</i> (Agassiz, 1862)	Anthoat hecata	Tubulariid ae	1	-	x	-	-	-
<i>Macrorhynchia philippina</i> Kirchenpauer, 1872	Leptothe cata	Aglaophen iidae	2	-	x	-	-	-
<i>Macrorhynchia</i> Kirchenpauer, 1872	Leptothe cata	Aglaophen iidae	1	-	x	-	-	-
<i>Aglaophenia lophocarpa</i> Allman, 1877	Leptothe cata	Aglaophen iidae	1	x	-	-	-	x
<i>Aglaophenia pluma</i> (Linnaeus, 1758)	Leptothe cata	Aglaophen iidae	1	-	x	-	-	-
<i>Aglaophenia</i> Lamouroux, 1812	Leptothe cata	Aglaophen iidae	3	x	x	x	-	x
<i>Lytocarpia myriophyllum</i> (Linnaeus, 1758)	Leptothe cata	Aglaophen iidae	5	x	-	x	-	x
<i>Obelia geniculata</i> (Linnaeus, 1758)	Leptothe cata	Campanul ariidae	1	x	-	-	-	-
<i>Halecium</i> Oken, 1815	Leptothe cata	Haleciidae	1	x	-	-	-	x
<i>Polyplumaria flabellata</i> Sars, 1874	Leptothe cata	Halopterid idae	5	x	-	x	-	x
<i>Polyplumaria</i> Sars, 1874	Leptothe cata	Halopterid idae	1	x	-	-	-	x
<i>Antennella secundaria</i> (Gmelin, 1791)	Leptothe cata	Halopterid idae	1	x	-	-	-	x
<i>Antennella</i> Allman, 1877	Leptothe cata	Halopterid idae	1	-	x	-	-	-
<i>Kirchenpaueria halecioides</i> (Alder, 1859)	Leptothe cata	Kirchenpa ueriidae	1	-	x	-	-	-
<i>Filellum serratum</i> (Clarke, 1879)	Leptothe cata	Lafoeidae	1	x	-	-	-	x
<i>Grammaria abietina</i> (Sars, 1851)	Leptothe cata	Lafoeidae	1	x	-	-	-	x
<i>Acryptolaria conferta</i> (Allman, 1877)	Leptothe cata	Lafoeidae	1	x	-	-	-	x
<i>Acryptolaria crassicaulis</i> (Allman, 1888)	Leptothe cata	Lafoeidae	1	x	-	-	-	x
<i>Acryptolaria</i> Norman, 1875	Leptothe cata	Lafoeidae	1	x	-	x	-	x
<i>Nemertesia antennina</i> (Linnaeus, 1758)	Leptothe cata	Plumularii dae	3	x	-	x	-	x
<i>Nemertesia ramosa</i> (Lamarck, 1816)	Leptothe cata	Plumularii dae	1	-	x	-	-	-

<i>Nemertesia Lamouroux, 1812</i>	Leptotheca	Plumulariidae	1	x	-	-	-	x
<i>Plumulariidae McCrady, 1859</i>	Leptotheca	Plumulariidae	1	x	-	x	-	x
<i>Sertularella gayi (Lamouroux, 1821)</i>	Leptotheca	Sertularellidae	1	x	-	-	-	x
<i>Sertularella Gray, 1848</i>	Leptotheca	Sertularellidae	1	-	x	-	-	x
<i>Diphasia alata (Hincks, 1855)</i>	Leptotheca	Sertulariidae	1	x	-	x	-	x
<i>Diphasia margareta (Hassall, 1841)</i>	Leptotheca	Sertulariidae	1	x	-	-	-	x
<i>Diphasia Agassiz, 1862</i>	Leptotheca	Sertulariidae	2	x	-	-	-	x
<i>Dynamena Lamouroux, 1812</i>	Leptotheca	Sertulariidae	1	-	x	-	-	-
<i>Cryptolaria pectinata (Allman, 1888)</i>	Leptotheca	Zygophylacidae	1	x	-	-	-	x
<i>Cryptolaria Busk, 1857</i>	Leptotheca	Zygophylacidae	1	x	-	x	-	x
<i>Zygophylax biarmata Billard, 1905</i>	Leptotheca	Zygophylacidae	1	x	-	-	-	x
Phyllum Chlorophyta								
Subphyllum Chlorophytina								
Class Ulvophyceae								
<i>Bryopsis J.V.Lamouroux, 1809</i>	Bryopsidales	Bryopsidaceae	1	-	x	-	-	-
<i>Caulerpa cylindracea Sonder, 1845</i>	Bryopsidales	Caulerpaceae	1	-	-	x	-	-
<i>Caulerpa mexicana Sonder ex Kützinger, 1849</i>	Bryopsidales	Caulerpaceae	1	-	-	x	-	-
<i>Caulerpa prolifera (Forsskål) J.V.Lamouroux, 1809</i>	Bryopsidales	Caulerpaceae	7	-	x	x	-	-
<i>Caulerpa racemosa (Forsskål) J.Agardh, 1873</i>	Bryopsidales	Caulerpaceae	1	-	-	x	-	-
<i>Caulerpa webbiana f. disticha Vickers, 1896</i>	Bryopsidales	Caulerpaceae	1	-	-	x	-	-
<i>Caulerpa webbiana Montagne, 1837</i>	Bryopsidales	Caulerpaceae	1	-	x	-	-	-
<i>Caulerpa J.V.Lamouroux, 1809</i>	Bryopsidales	Caulerpaceae	4	-	-	x	-	-

<i>Codium adhaerens</i> C.Agardh, 1822	Bryopsid ales	Codiaceae	4	x	-	-	-	-
<i>Codium elisabethiae</i> O.C.Schmidt, 1929	Bryopsid ales	Codiaceae	2	x	-	-	-	-
<i>Codium fragile</i> subsp. <i>fragile</i> (Suringar) Hariot, 1889	Bryopsid ales	Codiaceae	1	x	-	-	-	-
<i>Codium fragile</i> (Suringar) Hariot, 1889	Bryopsid ales	Codiaceae	1	x	-	-	-	-
<i>Avrainvillea canariensis</i> A.Gepp & E.S.Gepp, 1911	Bryopsid ales	Dichotomo siphonacea e	1	-	x	-	-	-
<i>Halimeda incrassata</i> (J.Ellis) J.V.Lamouroux, 1816	Bryopsid ales	Halimedac eae	1	-	-	x	-	-
<i>Anadyomene stellata</i> (Wulfen) C.Agardh, 1823	Cladoph orales	Anadyome naceae	1	-	-	x	-	-
<i>Microdictyon calodictyon</i> (Montagne) Kützing, 1849	Cladoph orales	Anadyome naceae	1	-	-	x	-	-
<i>Microdictyon</i> Decaisne, 1841	Cladoph orales	Anadyome naceae	1	-	-	x	-	-
<i>Cladophoropsis membranacea</i> Bang ex C.Agardh) Børgesen, 1905	Cladoph orales	Boodleace ae	1	x	-	-	-	-
<i>Chaetomorpha aerea</i> (Dillwyn) Kützing, 1849	Cladoph orales	Cladophor aceae	2	x	-	-	-	-
<i>Chaetomorpha pachynema</i> (Montagne) Kützing, 1847	Cladoph orales	Cladophor aceae	2	x	-	-	-	-
<i>Chaetomorpha</i> Kützing, 1845	Cladoph orales	Cladophor aceae	2	x	-	x	-	-
<i>Pseudorhizoclonium africanum</i> (Kützing) Boedeker, 2016	Cladoph orales	Cladophor aceae	1	x	-	-	-	-
<i>Cladophora albida</i> (Nees) Kützing, 1843	Cladoph orales	Cladophor aceae	1	x	-	-	-	-
<i>Cladophora coelothrix</i> Kützing, 1843	Cladoph orales	Cladophor aceae	1	x	-	-	-	-
<i>Cladophora prolifera</i> (Roth) Kützing, 1843	Cladoph orales	Cladophor aceae	3	x	-	-	-	-
<i>Cladophora</i> Kützing, 1843	Cladoph orales	Cladophor aceae	3	x	-	x	-	-
<i>Valonia utricularis</i> (Roth) C.Agardh, 1823	Cladoph orales	Valoniacea e	1	x	-	-	-	-
<i>Valonia</i> C.Agardh, 1823	Cladoph orales	Valoniacea e	1	x	-	-	-	-

<i>Cymopolia barbata</i> (Linnaeus) J.V.Lamouroux, 1816	Dasyclad ales	Dasyclada ceae	1	-	-	x	-	-
<i>Dasycladus vermicularis</i> (Scopoli) Krasser, 1898	Dasyclad ales	Dasyclada ceae	1	-	x	-	-	-
<i>Dasycladus</i> C.Agardh, 1828	Dasyclad ales	Dasyclada ceae	1	-	-	x	-	-
<i>Parvocaulis parvulus</i> (Solms-Laubach) S.Berger, Fettweiss, Gleissberg, Liddle, U.Richter, Sawitzky & Zuccarello, 2003	Dasyclad ales	Polyphysa ceae	2	-	-	x	-	-
<i>Blidingia minima</i> (Nägeli ex Kützing) Kylin, 1947	Ulvales	Kornmann iaceae	1	x	-	-	-	-
<i>Blidingia</i> Kylin, 1947	Ulvales	Kornmann iaceae	2	x	-	-	-	-
<i>Ulva clathrata</i> (Roth) C.Agardh, 1811	Ulvales	Ulvaceae	1	x	-	-	-	-
<i>Ulva compressa</i> Linnaeus, 1753	Ulvales	Ulvaceae	2	x	-	-	-	-
<i>Ulva intestinalis</i> Linnaeus, 1753	Ulvales	Ulvaceae	1	x	-	-	-	-
<i>Ulva linza</i> Linnaeus, 1753	Ulvales	Ulvaceae	1	x	-	-	-	-
<i>Ulva rigida</i> C.Agardh, 1823	Ulvales	Ulvaceae	3	x	-	-	-	-
<i>Ulva torta</i> (Mertens) Trevisan, 1842	Ulvales	Ulvaceae	1	x	-	-	-	-
<i>Ulva</i> Linnaeus, 1753	Ulvales	Ulvaceae	3	x	-	-	-	-
<i>Ulvaceae</i> J.V. Lamouroux ex Dumortier, 1822	Ulvales	Ulvaceae	1	x	-	-	-	-
Class Chlorophyceae								
<i>Pseudotetraspora marina</i> Wille, 1906	Chlamyd omonada les	Palmellops idaceae	1	-	-	x	-	-
Phyllum Rhodophyta								
Subphyllum Eurhodophytina								
Class Bangiophyceae								
<i>Porphyra umbilicalis</i> Kützing, 1843	Bangiiale s	Bangiaceae	1	x	-	-	-	-
<i>Porphyra</i> C.Agardh, 1824	Bangiiale s	Bangiaceae	1	x	-	-	-	-
<i>Bangia atropurpurea</i> (Mertens ex Roth) C.Agardh, 1824	Bangiiale s	Bangiaceae	1	x	-	-	-	-
Class Florideophyceae								
<i>Asparagopsis armata</i> f. <i>rufolanosa</i> Harvey, 1856	Bonnem aisioniale s	Bonnemais oniaceae	1	x	-	-	-	-
<i>Asparagopsis armata</i> Harvey, 1855	Bonnem aisioniale s	Bonnemais oniaceae	4	x	-	-	-	-

<i>Asparagopsis taxiformis</i> (Delile) Trevisan de Saint-Léon, 1845	Bonnemaisoniales	Bonnemaisoniaceae	7	x	x	x	-	-
<i>Asparagopsis</i> Montagne, 1840	Bonnemaisoniales	Bonnemaisoniaceae	4	x	-	-	-	-
Ceramieales	Ceramiales		1	-	-	x	-	-
<i>Chondracanthus teedei</i> (Mertens ex Roth) Kützinger, 1843	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Aglaothamnion</i> Feldmann-Mazoyer, 1941	Ceramiales	Callithamniaceae	1	x	-	-	-	-
<i>Callithamnion corymbosum</i> (Smith) Lyngbye, 1819	Ceramiales	Callithamniaceae	1	x	-	-	-	-
<i>Gaillona hookeri</i> (Dillwyn) Athanasiadis, 2016	Ceramiales	Callithamniaceae	1	x	-	-	-	-
<i>Spyridia filamentosa</i> (Wulfen) Harvey, 1833	Ceramiales	Callithamniaceae	2	-	-	x	-	-
<i>Spyridia hypnoides</i> (Bory de Saint-Vincent) Papenfuss, 1968	Ceramiales	Callithamniaceae	1	-	-	x	-	-
<i>Antithamnion</i> Nägeli, 1847	Ceramiales	Ceramiaceae	1	x	-	-	-	-
<i>Centroceras clavulatum</i> (C.Agardh) Montagne, 1846	Ceramiales	Ceramiaceae	2	x	-	x	-	-
<i>Ceramieae</i> (Dumortier) Schmitz, 1889	Ceramiales	Ceramiaceae	1	x	-	-	-	-
<i>Ceramium ciliatum</i> (J.Ellis) Ducluzeau, 1806	Ceramiales	Ceramiaceae	2	x	-	-	-	-
<i>Ceramium diaphanum</i> (Lightfoot) Roth, 1806	Ceramiales	Ceramiaceae	2	x	-	x	-	-
<i>Ceramium echionotum</i> J.Agardh, 1844	Ceramiales	Ceramiaceae	2	x	-	-	-	-
<i>Ceramium gaditanum</i> (Clemente) Cremades, 1990	Ceramiales	Ceramiaceae	1	x	-	-	-	-
<i>Ceramium virgatum</i> Roth, 1797	Ceramiales	Ceramiaceae	2	x	-	-	-	-
<i>Ceramium</i> Roth, 1797	Ceramiales	Ceramiaceae	2	-	x	-	-	-
<i>Gayliella mazoyerae</i> T.O.Cho, Fredericq & Hommersand, 2008	Ceramiales	Ceramiaceae	1	-	-	x	-	-
<i>Stirkia codii</i> (H.Richards) Barros-Barreto & Maggs, 2023	Ceramiales	Ceramiaceae	1	-	-	x	-	-

<i>Acrosorium venulosum</i> (Zanardini) Kylin, 1924	Ceramiales	Delesseriaceae	1	x	-	-	-	-
<i>Acrosorium</i> Zanardini ex Kützinger, 1869	Ceramiales	Delesseriaceae	1	x	-	-	-	-
<i>Cottoniella filamentosa</i> (M.A.Howe) Børgesen, 1920	Ceramiales	Delesseriaceae	3	-	-	x	-	-
<i>Dasya pedicellata</i> (C.Agardh) C.Agardh, 1824	Ceramiales	Delesseriaceae	1	-	-	x	-	-
<i>Dasya</i> C.Agardh, 1824	Ceramiales	Delesseriaceae	1	x	-	-	-	-
<i>Delesseriaceae</i> Bory, 1828	Ceramiales	Delesseriaceae	1	x	-	-	-	-
<i>Hypoglossum hypoglossoides</i> (Stackhouse) Collins & Hervey, 1917	Ceramiales	Delesseriaceae	1	x	-	-	-	-
<i>Chondria capillaris</i> (Hudson) M.J.Wynne, 1991	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Chondria coerulescens</i> (J.Agardh) Sauvageau, 1897	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Chondria dasyphylla</i> (Woodward) C.Agardh, 1817	Ceramiales	Rhodomelaceae	2	x	-	x	-	-
<i>Chondria</i> C.Agardh, 1817	Ceramiales	Rhodomelaceae	1	-	x	-	-	-
<i>Laurencia viridis</i> Gil-Rodríguez & Haroun, 1992	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Laurencia</i> J.V.Lamouroux, 1813	Ceramiales	Rhodomelaceae	5	x	-	x	-	-
<i>Lophocladia trichoclados</i> (C.Agardh) F.Schmitz, 1893	Ceramiales	Rhodomelaceae	2	-	-	x	-	-
<i>Lophosiphonia</i> Falkenberg, 1897	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Polysiphonia atlantica</i> Kapraun & J.N.Norris, 1982	Ceramiales	Rhodomelaceae	2	x	-	x	-	-
<i>Polysiphonia flexella</i> (C.Agardh) J.Agardh, 1842	Ceramiales	Rhodomelaceae	1	-	-	x	-	-
<i>Polysiphonia havanensis</i> Montagne, 1837	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Polysiphonia opaca</i> (C.Agardh) Moris & De Notaris, 1839	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Polysiphonia stricta</i> (Mertens ex Dillwyn) Greville, 1824	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Polysiphonia</i> Greville, 1823	Ceramiales	Rhodomelaceae	4	x	-	x	-	-

<i>Symphycladia marchantioides</i> (Harvey) Falkenberg, 1897	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Carradoriella denudata</i> (Dillwyn) Savoie & G.W.Saunders, 2019	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Halopithys incurva</i> (Hudson) Batters, 1902	Ceramiales	Rhodomelaceae	1	-	-	x	-	-
<i>Herposiphonia secunda</i> (C.Agardh) Ambronn, 1880	Ceramiales	Rhodomelaceae	1	-	-	x	-	-
<i>Herposiphonia</i> Nägeli, 1846	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Osmundea pinnatifida</i> (Hudson) Stackhouse, 1809	Ceramiales	Rhodomelaceae	3	x	-	-	-	-
<i>Osmundea</i> Stackhouse, 1809	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Pterosiphonia</i> Falkenberg, 1897	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Vertebrata fucoides</i> (Hudson) Kuntze, 1891	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Vertebrata subulifera</i> (C.Agardh) Kuntze, 1891	Ceramiales	Rhodomelaceae	1	-	-	x	-	-
<i>Vertebrata tripinnata</i> (Harvey) Kuntze, 1891	Ceramiales	Rhodomelaceae	1	x	-	-	-	-
<i>Plumaria</i> F.Schmitz, 1896	Ceramiales	Wrangeliaceae	1	x	-	-	-	x
<i>Wrangelia penicillata</i> (C.Agardh) C.Agardh, 1828	Ceramiales	Wrangeliaceae	1	-	-	x	-	-
<i>Bornetia</i> Thuret, 1855	Ceramiales	Wrangeliaceae	1	-	x	-	-	-
Articulated Corallinaceae	Corallinales		3	x	-	x	-	-
Rhodolith	Corallinales		4	x	-	x	-	-
<i>Corallina officinalis</i> Linnaeus, 1758	Corallinales	Corallinaceae	1	x	-	-	-	-
<i>Corallina</i> Linnaeus, 1758	Corallinales	Corallinaceae	3	x	-	-	-	-
<i>Ellisolandia elongata</i> (J.Ellis & Solander) K.R.Hind & G.W.Saunders, 2013	Corallinales	Corallinaceae	2	x	-	-	-	-
<i>Jania adhaerens</i> J.V.Lamouroux, 1816	Corallinales	Corallinaceae	2	x	-	x	-	-
<i>Jania capillacea</i> Harvey, 1853	Corallinales	Corallinaceae	1	x	-	-	-	-

<i>Jania longifurca</i> Zanardini, 1844	Corallina les	Corallinac eae	2	x	-	-	-	-
<i>Jania rubens</i> (Linnaeus) J.V.Lamouroux, 1816	Corallina les	Corallinac eae	3	x	-	x	-	-
<i>Jania virgata</i> (Zanardini) Montagne, 1846	Corallina les	Corallinac eae	1	x	-	-	-	-
<i>Jania</i> J.V.Lamouroux, 1812	Corallina les	Corallinac eae	5	x	-	-	-	-
<i>Lithophyllum crouaniorum</i> Foslie, 1899	Corallina les	Lithophyll aceae	1	-	-	-	-	-
<i>Lithophyllum hibernicum</i> Foslie, 1906	Corallina les	Lithophyll aceae	1	-	-	-	-	-
<i>Lithophyllum incrustans</i> Philippi, 1837	Corallina les	Lithophyll aceae	4	-	x	x	-	-
<i>Lithophyllum</i> Philippi, 1837	Corallina les	Lithophyll aceae	1	-	x	-	-	-
<i>Tenarea tortuosa</i> (Esper) Me.Lemoine, 1910	Corallina les	Lithophyll aceae	2	x	-	-	-	-
<i>Amphiroa</i> J.V.Lamouroux, 1812	Corallina les	Lithophyll aceae	1	-	x	-	-	-
<i>Neogoniolithon brassica-florida</i> (Harvey) Setchell & L.R.Mason, 1943	Corallina les	Spongitida ceae	1	-	-	-	-	-
<i>Gelidiella</i> Feldmann & G.Hamel, 1934	Gelidiale s	Gelidiellac eae	1	x	-	-	-	-
<i>Gelidium microdon</i> Kützing, 1849	Gelidiale s	Gelidiellac eae	3	x	-	-	-	-
<i>Gelidium pusillum</i> (Stackhouse) Le Jolis, 1863	Gelidiale s	Gelidiellac eae	2	x	-	-	-	-
<i>Gelidium spinosum</i> (S.G.Gmelin) P.C.Silva, 1996	Gelidiale s	Gelidiellac eae	3	x	-	-	-	-
<i>Millerella pannosa</i> (Feldmann) G.H.Boo & L.Le Gall, 2016	Gelidiale s	Gelidiellac eae	1	-	-	x	-	-
<i>Pterocladia capillacea</i> (S.G.Gmelin) Santelices & Hommersand, 1997	Gelidiale s	Pterocladi aceae	5	x	-	-	-	-
<i>Catenella caespitosa</i> (Withering) L.M.Irvine, 1976	Gigartin ales	Caulacant haceae	2	x	-	-	-	-
<i>Caulacanthus ustulatus</i> (Turner) Kützing, 1843	Gigartin ales	Caulacant haceae	1	x	-	-	-	-
<i>Calliblepharis</i> Kützing, 1843	Gigartin ales	Cystocloni aceae	1	-	x	-	-	-
<i>Hypnea musciformis</i> (Wulfen) J.V.Lamouroux, 1813	Gigartin ales	Cystocloni aceae	1	x	-	-	-	-

<i>Hypnea spinella</i> (C.Agardh) Kützing, 1847	Gigartinales	Cystocloniaceae	1	-	-	x	-	-
<i>Hypnea</i> J.V.Lamouroux, 1813	Gigartinales	Cystocloniaceae	1	-	-	x	-	-
<i>Dudresnaya</i> P.L.Crouan & H.M.Crouan, 1835	Gigartinales	Dumontiaceae	1	-	x	-	-	-
<i>Halarachnion ligulatum</i> (Woodward) Kützing, 1843	Gigartinales	Furcellariaceae	1	-	-	x	-	-
<i>Chondracanthus acicularis</i> (Roth) Fredericq, 1993	Gigartinales	Gigartiniaceae	3	x	-	-	-	-
<i>Gigartina pistillata</i> (S.G.Gmelin) Stackhouse, 1809	Gigartinales	Gigartiniaceae	1	x	-	-	-	-
<i>Callophyllis</i> Kützing, 1843	Gigartinales	Kallymeniaceae	1	-	x	-	-	-
<i>Kallymenia reniformis</i> (Turner) J.Agardh, 1842	Gigartinales	Kallymeniaceae	1	-	x	-	-	-
<i>Erythrodermis traillii</i> (Holmes ex Batters) Guiry & Garbary, 1990	Gigartinales	Phyllophoraceae	1	x	-	-	-	-
<i>Gymnogongrus griffithsia</i> (Turner) C.Martius, 1833	Gigartinales	Phyllophoraceae	1	x	-	-	-	-
<i>Phyllophora gelidioides</i> P.L.Crouan & H.M.Crouan ex Karsakoff, 1896	Gigartinales	Phyllophoraceae	1	x	-	-	-	-
<i>Sphaerococcus coronopifolius</i> Stackhouse, 1797	Gigartinales	Sphaerococcaceae	2	x	-	-	-	-
<i>Gracilaria</i> Greville, 1830	Gracilariiales	Gracilariaceae	2	-	x	x	-	-
<i>Dermocorynus dichotomus</i> (J.Agardh) Gargiulo, M.Morabito & Manghisi, 2013	Halymeniales	Grateloupiaceae	1	x	-	-	-	-
<i>Lithothamnion corallioides</i> (P.Crouan & H.Crouan) P.Crouan & H.Crouan, 1867	Hapalidiales	Hapalidiaceae	3	-	-	x	-	-
<i>Lithothamnion</i> Heydrich, 1897	Hapalidiales	Hapalidiaceae	1	-	x	-	-	-
<i>Phymatolithon calcareum</i> (Pallas) W.H.Adey & D.L.McKibbin ex Woelkerling & L.M.Irvine, 1986	Hapalidiales	Hapalidiaceae	3	-	-	x	-	-
<i>Phymatolithon lusitanicum</i> V.Peña, 2015	Hapalidiales	Hapalidiaceae	1	-	-	-	-	-
<i>Phymatolithon</i> Foslie, 1898	Hapalidiales	Hapalidiaceae	1	-	x	-	-	-
<i>Mesophyllum lichenoides</i> (J.Ellis) Me.Lemoine, 1928	Hapalidiales	Mesophyllumaceae	1	x	-	-	-	-

<i>Mesophyllum sphaericum</i> V.Pena, Bárbara, W.H.Adey, Riosmena-Rodrigues & H.G.Choi, 2011	Hapalidiales	Mesophyllumaceae	1	-	-	-	-	-
<i>Mesophyllum</i> Me.Lemoine, 1928	Hapalidiales	Mesophyllumaceae	1	-	x	-	-	-
<i>Hildenbrandia rubra</i> (Sommerfelt) Meneghini, 1841	Hildenbrandiales	Hildenbrandiaceae	1	x	-	-	-	-
<i>Hildenbrandia</i> Nardo, 1834	Hildenbrandiales	Hildenbrandiaceae	4	x	-	-	-	-
<i>Liagora</i> J.V.Lamouroux, 1812	Nemaliales	Liagoraceae	1	-	x	-	-	-
<i>Nemalion elminthoides</i> (Velley) Batters, 1902	Nemaliales	Nemaliaceae	1	x	-	-	-	-
<i>Scinaia Bivona-Bernardi</i> , 1822	Nemaliales	Scinaiaceae	1	-	-	x	-	-
<i>Platoma Schousboe</i> ex F.Schmitz, 1894	Nemastomatales	Schizymeniaceae	1	x	-	-	-	-
<i>Schizymenia dubyi</i> (Chauvin ex Duby) J.Agardh, 1851	Nemastomatales	Schizymeniaceae	1	x	-	-	-	-
<i>Palmaria palmata</i> (Linnaeus) F.Weber & D.Mohr, 1805	Palmariales	Palmariaceae	1	x	-	-	-	-
<i>Peyssonnelia rubra</i> (Greville) J.Agardh, 1851	Peyssonneliales	Peyssonneliaceae	1	x	-	-	-	-
<i>Peyssonnelia Decaisne</i> , 1841	Peyssonneliales	Peyssonneliaceae	4	x	-	x	-	-
<i>Plocamium cartilagineum</i> (Linnaeus) P.S.Dixon, 1967	Plocamiales	Plocamiaceae	2	x	-	-	-	-
<i>Champia parvula</i> (C.Agardh) Harvey, 1853	Rhodymeniales	Champiaceae	1	-	-	x	-	-
<i>Champia Desvaux</i> , 1809	Rhodymeniales	Champiaceae	1	-	x	-	-	-
<i>Gastroclonium ovatum</i> (Hudson) Papenfuss, 1944	Rhodymeniales	Champiaceae	1	x	-	-	-	-
<i>Gastroclonium reflexum</i> (Chauvin) Kützinger, 1849	Rhodymeniales	Champiaceae	1	x	-	-	-	-
<i>Lomentaria articulata</i> (Hudson) Lyngbye, 1819	Rhodymeniales	Lomentariaceae	2	x	-	-	-	-
<i>Rhodymenia holmesii</i> Ardissonne, 1893	Rhodymeniales	Rhodymeniaceae	2	x	-	-	-	-
<i>Rhodymenia pseudopalmata</i> (J.V.Lamouroux) P.C.Silva, 1952	Rhodymeniales	Rhodymeniaceae	1	x	-	-	-	-

Phyllum Ochrophyta

Class Dictyochophyceae

<i>Padina pavonica</i> (Linnaeus) Thivy, 1960	Dictyoch ales	Dictyochac eae	10	x	-	x	-	-
<i>Padina</i> Adanson, 1763	Dictyoch ales	Dictyochac eae	1	-	-	x	-	-

Class Phaeophyceae

<i>Lobophora variegata</i> (J.V.Lamouroux) Womersley ex E.C.Oliveira, 1977	Dictyotal es	Dictyotace ae	4	-	-	x	-	-
<i>Lobophora</i> J.Agardh, 1894	Dictyotal es	Dictyotace ae	4	-	x	x	-	-
<i>Stypopodium zonale</i> (J.V.Lamouroux) Papenfuss, 1940	Dictyotal es	Dictyotace ae	1	-	x	-	-	-
<i>Canistrocarpus cervicornis</i> (Kützinger) De Paula & De Clerck, 2006	Dictyotal es	Dictyotace ae	1	-	-	x	-	-
<i>Dictyopteris polypodioides</i> (A.P.De Candolle) J.V.Lamouroux, 1809	Dictyotal es	Dictyotace ae	1	x	-	-	-	-
<i>Dictyota bartayresiana</i> J.V.Lamouroux, 1809	Dictyotal es	Dictyotace ae	1	x	-	-	-	-
<i>Dictyota dichotoma</i> (Hudson) J.V.Lamouroux, 1809	Dictyotal es	Dictyotace ae	5	x	-	x	-	-
<i>Dictyota fasciola</i> (Roth) J.V.Lamouroux, 1809	Dictyotal es	Dictyotace ae	1	-	-	x	-	-
<i>Dictyota</i> J.V.Lamouroux, 1809	Dictyotal es	Dictyotace ae	12	x	-	x	-	-
<i>Zonaria tournefortii</i> (J.V.Lamouroux) Montagne, 1846	Dictyotal es	Dictyotace ae	6	x	-	-	-	-
<i>Leathesia marina</i> (Lyngbye) Decaisne, 1842	Ectocarp ales	Chordaria ceae	1	x	-	-	-	-
<i>Hydroclathrus clathratus</i> (C.Agardh) M.Howe, 1920	Ectocarp ales	Scytosipho naceae	3	x	-	x	-	-
<i>Colpomenia sinuosa</i> (Mertens ex Roth) Derbès & Solier, 1851	Ectocarp ales	Scytosipho naceae	5	x	-	x	-	-
<i>Petalonia binghamiae</i> (J.Agardh) K.L.Vinogradova, 1973	Ectocarp ales	Scytosipho naceae	1	x	-	-	-	-
<i>Fucus spiralis</i> Linnaeus, 1753	Fucales	Fucaceae	3	x	-	-	-	-
<i>Gongolaria abies-marina</i> (S.G.Gmelin) Kuntze 1891	Fucales	Sargassace ae	2	x	-	x	-	-
<i>Sargassum furcatum</i> Kützinger, 1843	Fucales	Sargassace ae	1	-	-	x	-	-
<i>Sargassum</i> C.Agardh, 1820	Fucales	Sargassace ae	4	x	-	x	-	-

<i>Cystoseira</i> C.Agardh, 1820	Fucales	Sargassaceae	2	x	-	-	-	-
<i>Ericaria selaginoides</i> (Linnaeus) Molinari & Guiry, 2020	Fucales	Sargassaceae	1	x	-	-	-	-
<i>Treptacantha abies-marina</i> (S.G.Gmelin) Kützing, 1843	Fucales	Sargassaceae	4	x	-	x	-	-
<i>Laminaria ochroleuca</i> Bachelot de la Pylaie, 1824	Laminariales	Laminariaceae	4	x	-	x	-	x
<i>Nemoderma tingitanum</i> Schousboe ex Bornet, 1892	Nemodermatales	Nemodermataceae	2	x	-	-	-	-
<i>Nemoderma</i> Schousboe ex Bornet, 1892	Nemodermatales	Nemodermataceae	1	-	x	-	-	-
<i>Ralfsia</i> Berkeley, 1843	Ralfsiales	Ralfsiaceae	1	x	-	-	-	-
<i>Cladostephus spongiosus</i> (Hudson) C.Agardh, 1817	Sphacelariales	Cladostephaceae	1	x	-	-	-	-
<i>Sphacelaria</i> Lyngbye, 1818	Sphacelariales	Sphacelariaceae	1	x	-	-	-	-
<i>Halopteris filicina</i> (Grateloup) Kützing, 1843	Sphacelariales	Stypocaulaceae	9	x	-	x	-	-
<i>Halopteris scoparia</i> (Linnaeus) Sauvageau, 1904	Sphacelariales	Stypocaulaceae	5	x	-	x	-	-
<i>Halopteris</i> Kützing, 1843	Sphacelariales	Stypocaulaceae	3	x	x	-	-	-
<i>Sporochnus pedunculatus</i> (Hudson) C.Agardh, 1817	Sporochnales	Sporochnaaceae	2	-	x	-	-	-
<i>Carpomitra costata</i> (Stackhouse) Batters, 1902	Sporochnales	Sporochnaaceae	1	x	-	-	-	-
<i>Cutleria multifida</i> (Turner) Greville, 1830	Tilopteridales	Cutleriaceae	1	x	-	-	-	-
<i>Phyllariopsis brevipes</i> (C.Agardh) E.C.Henry & G.R.South, 1987	Tilopteridales	Phyllariaceae	1	x	-	-	-	-

Phyllum Tracheophyta
Subphyllum Spermatophytina
Class Magnoliopsida

Alismatales	Alismatales		1	-	-	x	-	-
<i>Cymodocea nodosa</i> (Ucria) Ascherson, 1870	Alismatales	Cymodoceaceae	14	-	x	x	-	-
<i>Halodule wrightii</i> Ascherson, 1868	Alismatales	Cymodoceaceae	1	-	-	-	x	-

<i>Halophila decipiens</i> Ostenfeld, 1902	Alismata les	Hydrochar itaceae	1	-	-	x	-	-
<i>Ruppia maritima</i> Linnaeus, 1753	Alismata les	Ruppiacea e	1	-	-	-	x	-

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