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Article

Thoracic Fat Pad Biopsy in Cardiac Amyloidosis: Diagnostic Yield in an Afro-Caribbean Population

Short Title: Thoracic Fat Pad Biopsy in Cardiac Amyloidosis

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Highlights

- This study is the first to systematically assess the diagnostic potential of thoracic fat pad biopsy in patients with cardiac amyloidosis (CA).
- Thoracic fat pad biopsy has a limited sensitivity for CA diagnosis, particularly in patients with transthyretin-related (ATTR) CA, with a sensitivity of 31%.
- This study underscores the importance of utilizing established standard diagnostic criteria for CA.

Abstract: Aims: Cardiac amyloidosis (CA) is associated with amyloid infiltration of extra-cardiac tissue, which may occur at early stages of the disease. This study evaluates the diagnostic utility of thoracic fat pad biopsy obtained during pacemaker or ICD implantation as an alternative to standard diagnostic criteria for systemic amyloidosis. **Methods and results:** This exploratory, retrospective study included 27 patients with suspected or diagnosed cardiac amyloidosis (CA) who underwent pacemaker or defibrillator therapy. Of these, 16 patients were confirmed to have CA (15 with technetium-labeled bisphosphonate bone scintigraphy and 1 with protein electrophoresis and echocardiographic findings), while 11 were confirmed to be CA-negative. Thoracic fat pad biopsy demonstrated a specificity of 100% but a sensitivity of only 31%. Among patients with transthyretin (ATTR)-CA, sensitivity remained similarly low at 27%. These results are consistent with prior findings on abdominal fat pad biopsy in ATTR-CA, highlighting the limited diagnostic yield of this method. **Conclusion:** Thoracic fat pad biopsy cannot be recommended as a standard diagnostic tool for CA, particularly in ATTR-CA, due to its poor sensitivity. However, in AL (amyloid light-chain) amyloidosis, this minimally invasive procedure may aid diagnosis without additional invasive interventions.

Keywords: amyloidosis; ATTR; cardiac; scintigraphy; fat pad; biopsy; pacemaker

1. Introduction

Amyloidosis is a severe condition resulting from the systemic accumulation of amyloid protein deposits in various tissues, leading to structural and functional abnormalities. It is primarily associated with cardiac and neurological manifestations, with cardiac and renal forms posing significant threats to health and life. Cardiac amyloidosis (CA) primarily manifests as restrictive cardiomyopathy, which can result in heart failure and arrhythmias. Amyloidosis can arise from various types of amyloid proteins, but in Europe, the most prevalent forms are AL (amyloid light-chain) and ATTR (transthyretin) amyloidosis [1,2].

The hereditary form of ATTR cardiac amyloidosis (ATTRv-CA) disproportionately afflicts the black Afro-Caribbean population, who when compared to mainly whites with wild-type transthyretin amyloidosis (wt-ATTR-CA), a phenotypically similar condition, present with more advanced disease despite genetic testing may permit early identification [3].

Current scientific understanding highlights the typically “late” diagnosis of amyloidosis, emphasizing the challenges of achieving an earlier diagnosis. Numerous scientific publications suggest that systemic manifestations might precede the phenotypic presentation of the traditionally recognized cardiac and neurological forms by several years. Patients with ATTR amyloidosis may experience musculoskeletal symptoms 5 to 15 years prior to signs or symptoms of systemic disease [4–6]. Carpal tunnel syndrome, the most prevalent non-cardiac symptom in patients with ATTR-CA, often appears years before a formal diagnosis of either wt-ATTR-CA or hereditary ATTRv-CA [4]. ATTR amyloid deposits could be identified in various tissues of patients with amyloidosis removed during orthopedic procedures including carpal tunnel syndrome, rotator cuff tears, and lumbar canal stenosis [7].

Identifying early manifestations of CA could potentially shorten the diagnostic delay, enabling early therapeutic intervention [2,8,9]. Among these early signs, conduction disorders or other arrhythmias necessitating the implantation of pacemakers or defibrillators, may be an early manifestation of CA. Early signs of CA, often referred to as “red flag” symptoms (Figure 1), should prompt suspicion of CA and initiate further diagnostic evaluation [1,6,10].

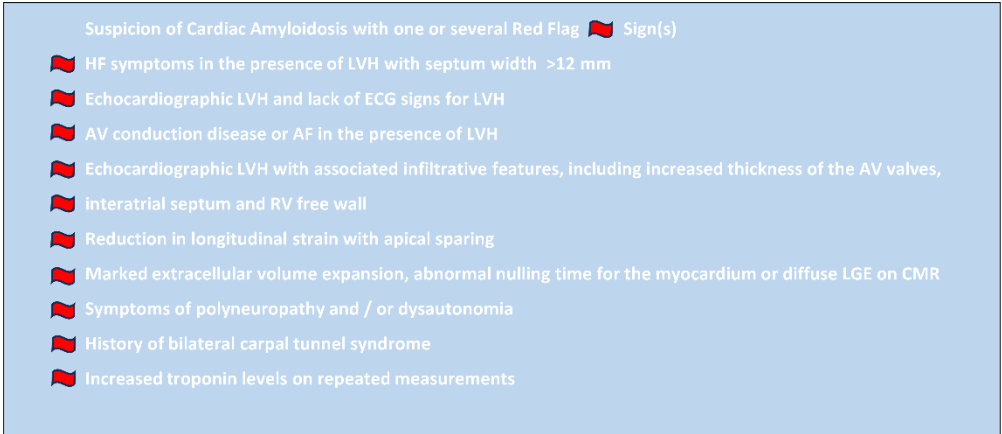


Figure 1. Possible clinical signs that should raise the suspicion of cardiac amyloidosis (CA). HF Heart Failure; LVH Left Ventricular Hypertrophy; AV Atrio-Ventricular; AF Atrial Fibrillation; RV Right Ventricular; LGE Late Gadolinium Enhancement; CMR Cardiac Magnetic Resonance Tomography.

Definitive diagnosis relies on histopathological examination, which involves biopsy procedures that can be particularly challenging and pose risks, especially in cardiac and renal forms of the disease. Historically, a conclusive diagnosis of amyloidosis has been made through a tissue biopsy stained with Congo red [11]. When observed under polarized light, this reveals the characteristic green birefringence of amyloid deposits, confirming the presence of the disease [11]. Histological confirmation is still required in cases where both bone scintigraphy (indicating ATTR-CA) and tests for monoclonal protein (indicating potential AL amyloidosis) show abnormalities or are inconclusive

[11]. It is also reasonable to verify and classify amyloid deposits through immunohistochemistry or mass spectrometry [12]. Mass spectrometry is the currently preferred method for amyloid typing, as immunohistochemistry results are often subtle and can be misinterpreted without significant expertise [12].

Bone scintigraphy, particularly using ^{99m}Tc -technetium-labeled bisphosphonates, has emerged as a valuable diagnostic tool for ATTR-CA [13]. This imaging technique can non-invasively detect myocardial uptake of amyloid deposits, offering high sensitivity and specificity in distinguishing ATTR-CA from other forms such as AL-CA after exclusion of monoclonal gammopathy [13]. ^{99m}Tc -scintigraphy for cardiac amyloidosis imaging involves administering 10 to 25 mCi of radiotracer intravenously, followed by planar and single-photon emission computed tomography (SPECT) imaging, typically performed 1 or 3 hours later. The degree of myocardial tracer uptake on planar imaging can be assessed using both qualitative and quantitative grading systems. Qualitative grading compares heart uptake to rib bone uptake, with grade 0 indicating no uptake, grade 1 representing mild uptake (less than rib, or less than bone in the case of ^{99m}Tc -DPD), grade 2 showing moderate uptake (equal to rib), and grade 3 indicating severe uptake (greater than rib). Quantitative assessments rely on the heart-to-contralateral (H/CL) ratio (for ^{99m}Tc -PYP) or the heart-to-whole body ratio (for ^{99m}Tc -DPD and ^{99m}Tc -HMDP) [13]. The limited availability of bone scintigraphy in less equipped healthcare settings may pose a significant barrier to timely and accurate diagnosis of cardiac ATTR amyloidosis.

Both ventricular and supraventricular arrhythmias are frequently observed in CA, particularly in ATTR-CA, with their occurrence closely linked to disease progression [14]. The prognostic significance of sustained and non-sustained ventricular tachycardias remains uncertain, and there are currently no clear guidelines for the use of implantable cardiac defibrillators (ICD) in these patients [14]. Atrial fibrillation is the most common supraventricular arrhythmia, affecting up to 88% of individuals with ATTR-CA [14]. Anticoagulation should be considered regardless of the CHADsVA score [15]. Conduction abnormalities and bradyarrhythmias are also prevalent in ATTR-CA, with up to 40% of patients requiring pacemaker implantation [14].

The implantation of pacemakers involves creating a pre-pectoral pocket between the superficial skin plane and the deeper muscular plane. One case report described amyloid deposits within the pacemaker pocket of a patient who had already been diagnosed with ATTRv-CA [16].

The potential for amyloid infiltration in the pacemaker or ICD pocket during the early stages of the disease has not been systematically investigated and warrants further study. As conduction abnormalities and other arrhythmias are linked to amyloidosis, the presence of peripheral deposits in the pacemaker or ICD pocket would not be unexpected.

The current study aims to assess the diagnostic utility of thoracic fat pad biopsy obtained from the pacemaker or ICD pocket as a potential alternative biopsy site for patients with suspicion of systemic amyloidosis. This exploratory, hypothesis generating study will compare the sensitivity and specificity of thoracic fat pad biopsy against standard diagnostic approaches for detecting CA, providing insights into the value of histopathological analysis of fat pad samples collected during pacemaker or ICD implantation. By exploring this previously unstudied method, we aim to determine the clinical yield of an accessible diagnostic tool that could facilitate earlier intervention and improve the management of patients with CA.

2. Methods

Patients with suspicion or diagnosis of CA and indication for pacemaker or ICD implantation or generator change, admitted in our electrophysiology (EP) lab between October 2020 and August 2024, were included into this exploratory, observational, retrospective single center study.

Inclusion criteria are displayed in Figure 1. All patients were managed in accordance with the amended Declaration of Helsinki (<https://www.wma.net/what-we-do/medical-ethics/declaration-of-helsinki/>). Written informed consent from the patients or patients' legal guardians/next of kin was not required to participate in this study in accordance with current legislation and institutional

requirements. All patients provided written informed consent prior to the procedure. The study was approved by the Institutional Review Board of the University Hospital of Martinique.

Patient and procedure selection were non-randomized and carried out upon the discretion of the electrophysiologist in charge. All study procedures were performed under local anesthesia by two experienced electrophysiologists (AM, RV) or by a fellow electrophysiologist (CM).

The indication for pacemaker and ICD therapy followed current guidelines [17,18]. Diagnosis of suspected cardiac amyloidosis (Figure 1) was confirmed or excluded with current gold standard diagnostic tools (Figure 2) [19].

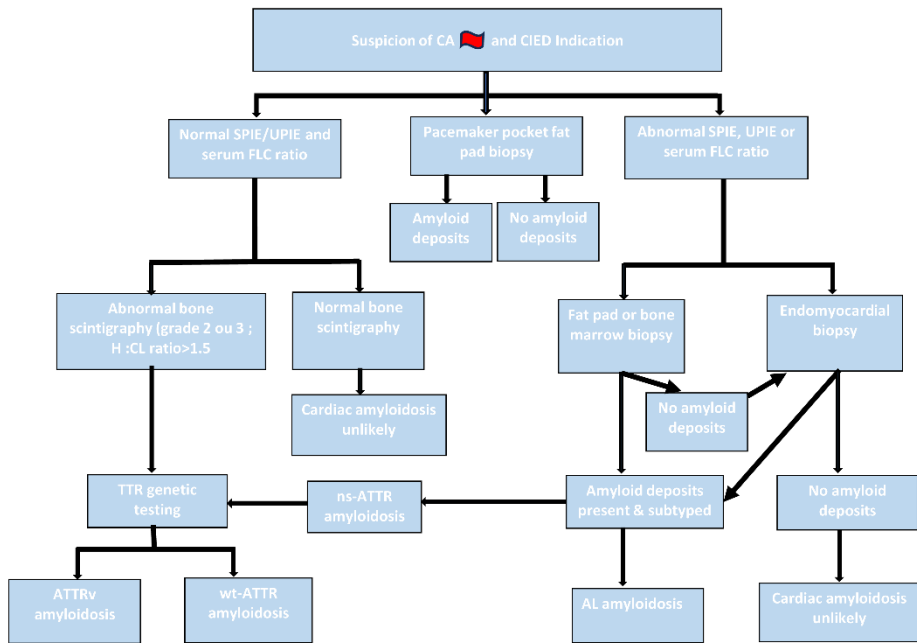


Figure 2. Algorithm for the evaluation of suspected cardiac amyloidosis at the University Hospital Martinique with added thoracic fat pad biopsy during pacemaker implantation: LVH left ventricular hypertrophy; CMR cardiac magnetic resonance; ICD implantable cardioverter defibrillator; FLC free light chain; GI gastrointestinal; SPIE serum protein electrophoresis with immunofixation; UPIE urine protein electrophoresis with immunofixation; H:CL heart to contralateral lung ratio, ns-ATTR non-specified ATTR; ATTRv hereditary (variant) ATTR; wt-ATTR wild type (senile) ATTR. Algorithm modified and adapted after [10,19].

Study data (patient and procedure characteristics) were retrospectively collected. Power calculations were not performed in this exploratory, retrospective study, as it is the first to evaluate this novel fat biopsy site with no prior data to guide sample size estimation. Additionally, limited number of available patients further precluded power calculations.

Statistical analysis was performed with JASP Team (2024, Version 0.19.1). Mean and standard deviations were reported for quantitative variables, while categorical variables were presented as absolute values and percentages. The following tests were accordingly used for group comparisons: t-test, Wilcoxon–Mann–Whitney test, Chi-square test and Fisher’s exact test. Statistical significance was set at $p<0.05$.

3. Results

A total of 27 patients were included in this study. Of these, 16 were diagnosed with CA, while 11 did not have CA based on standard diagnostic criteria (Tables 1 and 2). Among the 16 CA patients, 15 (93.8%) had ATTR-CA and 1 (6.3%) had AL-CA (Tables 1 and 2). Of the ATTR-CA patients, 8 individuals were found to carry a heterozygous mutation for the amyloidogenic allele where isoleucine substitutes for valine at codon position 142 (122 of the mature protein) (p.Val142Ile), leading to the diagnosis of hereditary ATTR-CA (ATTRv-CA). Genetic testing was negative in 3

patients, leading to the diagnosis of wild-type ATTR-CA (wt-ATTR-CA). Genetic testing was not performed in 4 patients with ATTR-CA. As no definitive diagnosis of ATTRv-CA or wt-ATTR-CA could be made, we defined those patients as non-specified ATTR-CA (ns-ATTR-CA) (Tables 1 and 2).

Table 1. Patient characteristics of all patients with definite diagnosis and thoracic fat pad biopsy. AL amyloid light-chain; ATTR transthyretin amyloidosis; CA cardiac amyloidosis; ATTRv-CA hereditary (variant) ATTR-CA; wt-ATTR-CA wild-type ATTR-CA; ns-ATTR-CA non-specified ATTR-CA (without genetic diagnostic); p.Val142Ile heterozygous mutation for the amyloidogenic allele where isoleucine substitutes for valine at codon position 142 (122 of the mature protein); MGUS monoclonal gammopathy of undetermined significance.

Number	Diagnosis	Serum electrophoresis	Salivary glands biopsy	Abdominal fat pad biopsy	Thoracic fat pad biopsy	Bone scintigraphy	Result
1	No CA	Normal	None	None	Negative	Negative	True negative
2	ATTRv-CA (p.Val142Ile)	Normal	None	None	Negative	Positive, Grade 3 uptake	False negative
3	No CA	Normal	None	None	Negative	Negative	True negative
4	ns-ATTR-CA	Normal	None	None	Negative	Positive, Grade 2 uptake	False negative
5	ATTRv-CA (p.Val142Ile)	Normal	Positive	Positive	Positive	Positive, Grade 4 uptake	True positive
6	No CA	Normal	None	Negative	Negative	Negative	True negative
7	No CA	Normal	None	None	Negative	Negative	True negative
8	ns-ATTR-CA	Normal	None	Positive	Positive	Positive, Grade 3 uptake	True positive
9	ATTRv-CA (p.Val142Ile)	Normal	Negative	None	Positive	Positive, Grade 3 uptake	True positive
10	No CA	Normal	None	Negative	Negative	Negative	True negative
11	No CA	Normal	None	None	Negative	Negative	True negative
12	No CA	Normal	None	None	Negative	Negative	True negative

13	ATTRv-CA (p.Val142Ile)	Monoclonal peak (MGUS)	Positive	Negative	Negative	Positive, Grade 3 uptake	False negative
14	No CA	Normal	None	None	Negative	Negative	True negative
15	No CA	Normal	None	Negative	Negative	Negative	True negative
16	No CA	Normal	None	None	Negative	Negative	True negative
17	No CA	Normal	None	None	Negative	Negative	True negative
18	ATTRv-CA (p.Val142Ile)	Normal	None	None	Negative	Positive, Grade 3 uptake	False negative
19	ns-ATTR-CA	Normal	None	None	Negative	Positive Grade 2 uptake	False negative
20	ns-ATTR-CA	Normal	Negative	None	Negative	Positive, Grade 2 uptake	False negative
21	wt-ATTR-CA	Normal	Negative	None	Negative	Positive, Grade 2 uptake	False negative
22	wt-ATTR-CA	Normal	None	None	Negative	Positive, Grade 3 uptake	False negative
23	ATTRv-CA (p.Val142Ile)	Monoclonal peak (MGUS)	Negative	Negative	Negative	Positive, Grade 3 uptake	False negative
24	ATTRv-CA (p.Val142Ile)	Normal	None	None	Positive	Positive, Grade 2 uptake	True positive
25	ATTRv-CA (p.Val142Ile)	Normal	Negative	None	Negative	Positive, Grade 2 uptake	False negative
26	wt-ATTR-CA	Normal	None	None	Negative	Positive, Grade 3 uptake	False negative
27	AL-CA	Monoclonal peak	Positive	Positive	Positive	Negative	True positive

Table 2. Baseline parameters, comparing patients with and without cardiac amyloidosis. Quantitative variables are presented as mean \pm standard deviation, and qualitative variables as absolute number, n (%); level of significance: $p < 0.05$. BMI body mass index; ATTR-CA ATTR cardiac amyloidosis; AV atrio-ventricular; ICD implantable cardioverter defibrillator; LVEF left ventricular ejection fraction; GFR glomerular filtration rate; CKD chronic kidney disease with $GFR < 60$ ml/min; BNP brain natriuretic peptide.

	Amyloidosis (n=16)	No amyloidosis (n=11)	P value
Age (years \pm SD)	76.5 \pm 9	77.1 \pm 10.2	0.870
Male sex n (%)	15 (93.8)	7 (63.6)	0.125
BMI (kg/m ² \pm SD)	25.1 \pm 4.2	28.5 \pm 6.1	0.118
ATTR-CA n (%)	15 (93.8)	0 (0%)	0.001
Baseline heart rate (bpm \pm SD)	59 \pm 25	41 \pm 8	0.026
Atrial fibrillation n (%)	6 (37.5%)	3 (33.3)	0.692
Sick sinus syndrome n (%)	0 (0)	1 (9.1)	0.407
AV conduction disease n (%)	13 (81.3)	10 (90.9)	0.624
Pacemaker n (%)	13 (81.3)	11 (100)	0.248
ICD n (%)	3 (18.8)	0	0.248
QRS width (ms \pm SD)	127 \pm 37	110 \pm 34	0.422
LVEF (% \pm SD)	46 \pm 11	55 \pm 9	0.053
Septum (mm \pm SD)	15 \pm 3	12 \pm 2	0.027
GFR (ml/min \pm SD)	64 \pm 25	73 \pm 34	0.461
CKD n (%)	7 (43.8)	4 (36.4)	1.000
BNP (pg/ml \pm SD)	465.3	574.4	0.676

No complications occurred, related to the intervention. Baseline characteristics were similar between patients with and without CA, with no significant differences in age, sex, BMI, QRS width, LVEF, renal function, or BNP levels. However, patients with CA had higher baseline heart rate, lower LVEF, and greater septal thickness (Table 1). Indication for pacemaker therapy was atrioventricular conduction disease in 13 patients (81.3%) in the amyloidosis group and in 10 patients (90.9%) in the group without amyloidosis. Atrioventricular (AV) conduction disease was defined as second degree AV-block, complete AV-block, alternating bundle branch block, bi-fascicular block or AF with bradyarrhythmia. One patient (9.1%) in the group without amyloidosis was diagnosed with symptomatic sick sinus syndrome. Three patients received an ICD. ICD indications were LVEF of 30% and non-sustained ventricular tachycardia in a 63 year-old patient with wt-ATTR-CA, LVEF of 30% in a 64 year-old patient with ATTRv-CA and LVEF of 28% in a 56 year-old patient with AL-CA.

The histopathological analysis showed a specificity of 100% and a sensitivity of 31.3% in the studied cohort (Table 3). Figures 3–6 provide examples of two true-positive cases, one true-negative and one false-negative case. With no false-positive results, the positive predictive value (PPV) based

on the cohort data was 100%, indicating that all positive biopsy results were true positives for CA. When accounting for the cohort prevalence of 59.3%, the PPV was calculated to be 18.5%, reflecting the diagnostic performance in the context of this specific population. The negative predictive value (NPV) was 50%, indicating that half of the negative biopsies were false negatives (Table 3).

Table 3. Cross table for CA (AL-CA and ATTR-CA): sensitivity 31.3%, specificity 100%, positive predictive value 18.5%, negative predictive value 50%, Fisher’s exact test: P=0.06. CA: Cardiac Amyloidosis.

	CA	No CA
Biopsy positive with evidence of amyloid deposits n (%)	5 (31.3)	0 (0)
Biopsy negative without evidence of amyloid deposits n (%)	11 (68.7)	11 (100)

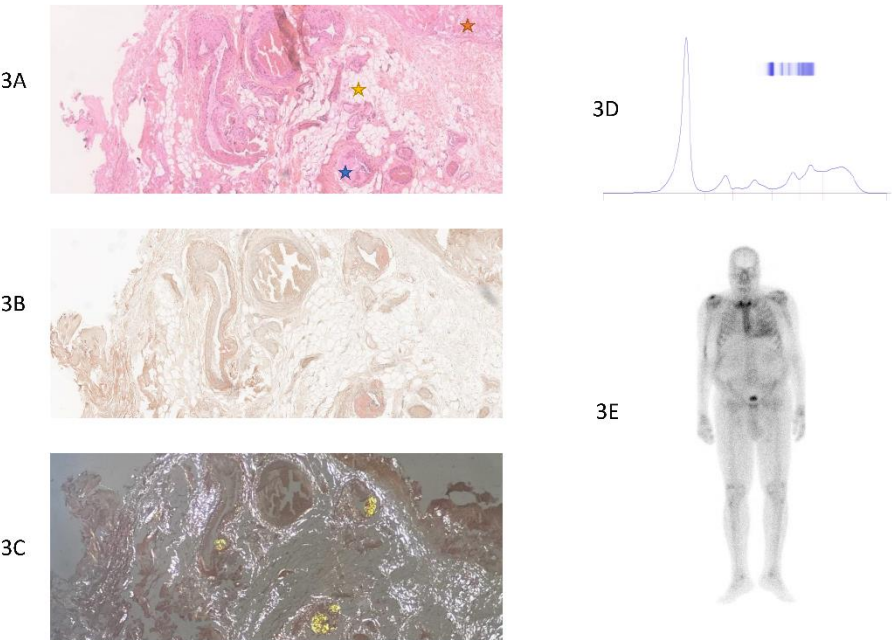


Figure 3. True positive case of a patient with hereditary transthyretin ATTRv (p.Val142Ile) cardiac amyloidosis (patient number 9, Table 1). **3A:** Standard HES staining of thoracic fat pad biopsy with 5x magnification. Fibro-adipose tissue without atypia, showing amyloid deposits in the vascular walls as amorphous eosinophilic extracellular deposits. Blue star: amyloid deposits. Orange star: fibrous tissue. Yellow star: adipose tissue. **3B:** Congo red staining of thoracic fat pad biopsy, 5x magnification. Fibro-adipose tissue without atypia, showing amyloid deposits stained with Congo red. **3C:** Amyloid deposits displaying yellow-green birefringence under polarized light (on Congo red-stained slides). **3D:** Normal protein electrophoresis without monoclonal peak. **3E:** Bone scintigraphy: grade 3 uptake in the cardiac area, suggestive of transthyretin cardiac amyloidosis. Diffuse uptake in soft tissues.

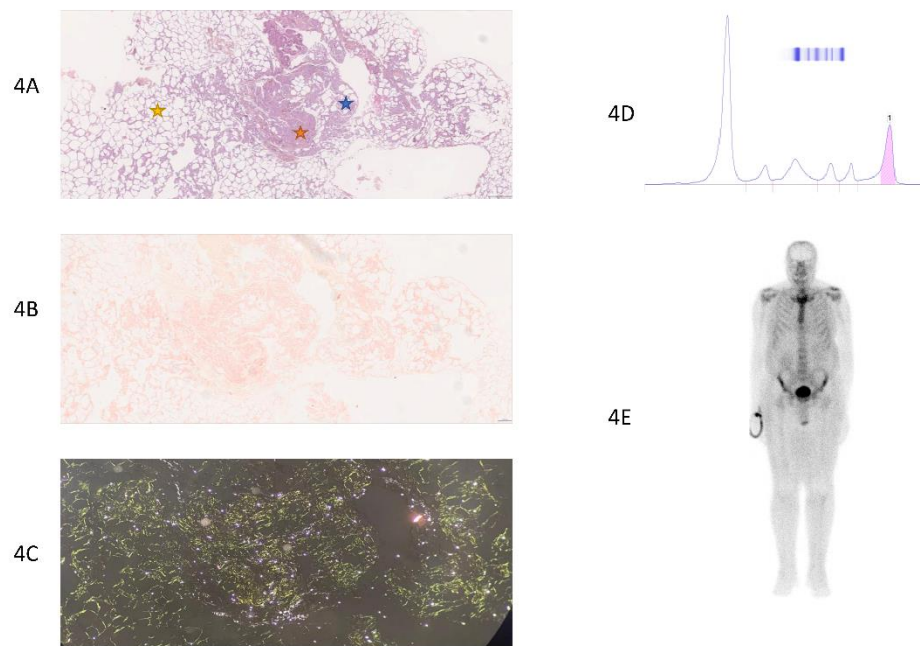


Figure 4. True positive case of a patient with AL cardiac amyloidosis (patient number 27, Table 1). **4A:** Standard HES staining of thoracic fat pad biopsy with 10x magnification. Fibro-adipose tissue without atypia, showing amyloid deposits as amorphous eosinophilic extracellular deposits. Blue star: amyloid deposits. Orange star: fibrous tissue. Yellow star: adipose tissue. **4B:** Congo red staining of thoracic fat pad biopsy, 10x magnification. Fibro-adipose tissue without atypia, showing amyloid deposits stained with Congo red. **4C:** Amyloid deposits displaying yellow-green birefringence under polarized light (on Congo red-stained slides). **4D:** Protein electrophoresis: Monoclonal peak. **4E:** Bone scintigraphy: No uptake observed in the cardiac area, grade 0 uptake. Moderate diffuse uptake in extra-cardiac tissue.

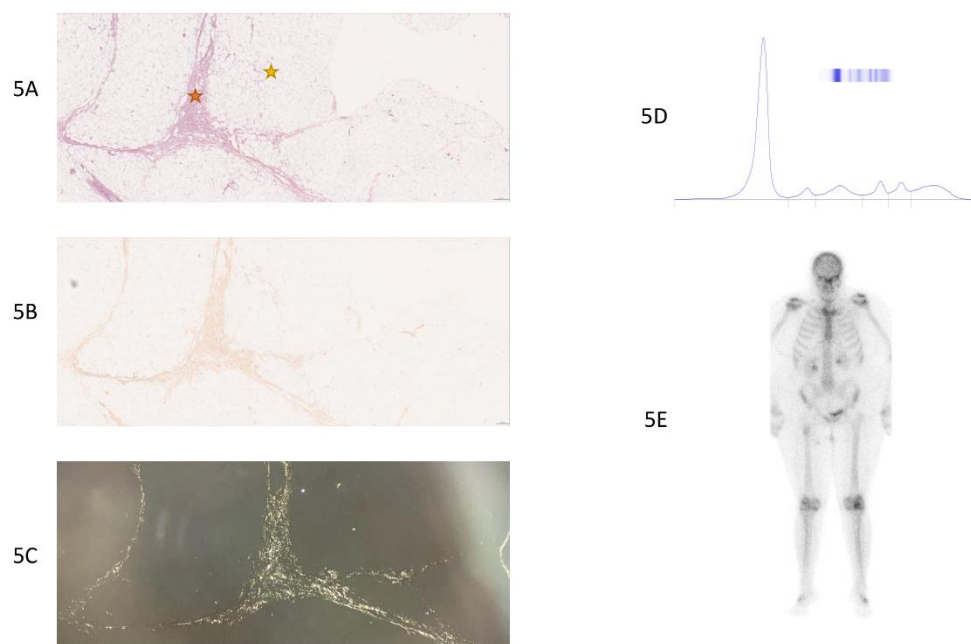


Figure 5. True negative case of a patient without cardiac amyloidosis (patient number 15, Table 1). **5A:** Standard HES staining of thoracic fat pad biopsy, 5x magnification: Fibro-adipose tissue without

amyloid deposits. Orange star: fibrous tissue. Yellow star: adipose tissue. **5B:** Congo red staining, 5x magnification. Fibro-adipose tissue without amyloid deposits. **5C:** Absence of yellow-green birefringence under polarized light. **5D:** Protein electrophoresis: Normal, without monoclonal peak. **5E:** Bone scintigraphy: No uptake in the cardiac area, grade 0. No suspicious uptake foci detected on the whole-body scan. No scintigraphic evidence of transthyretin cardiac amyloidosis.

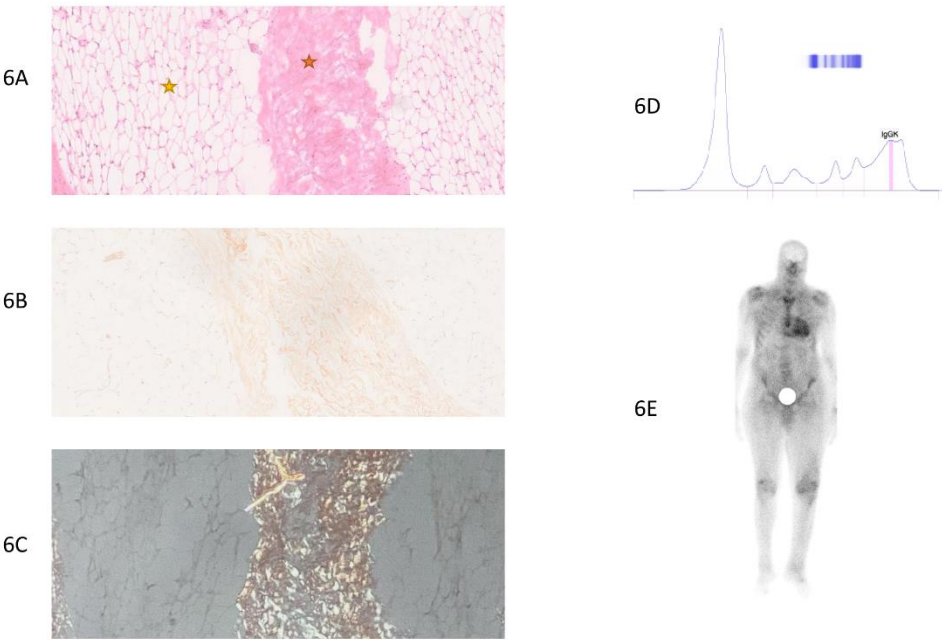


Figure 6. False negative case of a patient with hereditary transthyretin ATTRv (p.Val142Ile) cardiac amyloidosis (patient number 23, Table 1). **6A:** Standard HES staining, 10x magnification. Fibro-adipose tissue without atypia, no amyloid deposits. Orange star: fibrous tissue. Yellow star: adipose tissue. **6B:** Congo red staining, 10x magnification. Adipose tissue without atypia, no amyloid deposits. **6C:** Absence of amyloid deposits (yellow-green) under polarized light (on slide stained with Congo red). **6D:** Protein electrophoresis: Monoclonal peak (IgGκappa). The hematological evaluation, including serum protein electrophoresis, immunofixation, serum free light chain analysis, Bence-Jones protein assessment, clinical evaluation, bone marrow biopsy, and imaging, excluded plasmacytoma and confirmed MGUS (monoclonal gammopathy of undetermined significance). The diagnosis of hereditary ATTRv (p.Val142Ile) cardiac amyloidosis was established through positive bone scintigraphy and supported by genetic testing. **6E:** Bone scintigraphy: Intense uptake in the cardiac area, with intensity greater than the costal framework, grade 3 uptake, suggestive of transthyretin cardiac amyloidosis. Diffuse soft tissue uptake. Degenerative uptake in both knees.

As previous studies have demonstrated a lower sensitivity of abdominal fat pad biopsy in patients with ATTR-CA compared to those with AL-CA, we separately analyzed the sensitivity and specificity for ATTR-CA, excluding the patient with AL-CA. Sensitivity was again found to be poor at 26.7%, with a specificity at 100% (Table 4).

Table 4. Cross table for ATTR-CA: sensitivity 26.7%, specificity 100%, Fisher’s exact test: P=0.113. ATTR-CA: Transthyretin cardiac amyloidosis.

	ATTR-CA	No ATTR-CA
Biopsy positive with evidence of amyloid deposits n (%)	4 (26.7)	0 (0)

Biopsy negative without evidence of amyloid deposits n (%)	11 (73.3)	11 (100)
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In our study, we did not routinely perform biopsies from the abdominal fat pad. Therefore, fat pad biopsies from only eight patients could be compared with the results from thoracic fat pad biopsies. Three patients with evidence of amyloid in the abdominal fat pad biopsy had also evidence of amyloid in the thoracic fat pad biopsy. One of them was diagnosed with ns-ATTR-CA (grade 3 myocardial uptake), one with ATTRv-CA (grade 4 uptake) and one with AL-CA. Five patients without evidence of amyloid in the abdominal fat pad biopsy had also no evidence of amyloid in the thoracic fat pad biopsy ($p=0.018$). Among the five patients without amyloid evidence in their abdominal fat pad biopsies, two had ATTRv-CA with grade 2 and 3 myocardial uptake (false negative cases), respectively, while three showed no evidence of CA (true negative cases) based on standard diagnostic criteria ($p=0.196$). In this group of eight patients with thoracic and abdominal fat pad biopsies we found an excellent correlation with identical results from both anatomical sites ($p=0.018$). In this small group, thoracic and abdominal fat pad biopsies detected only three out of five cases with CA (sensitivity 60%, specificity 100%, $p=0.196$). Seven patients also received a salivary gland biopsy. We found three true positive cases, four false negative cases, when compared to gold standard diagnostic.

4. Discussion

The prevalence of amyloidosis in the general population of Martinique is unknown. However, deducing from the known 3.43% prevalence of the p.Val142Ile variant in African Americans and assuming disease manifestation in 10-15% of carriers, one could estimate a prevalence of manifest ATTRv-CA between 0.3% and 0.5% in Martinique, a Caribbean island where approximately 90% of the population is of African descent [3,20]. The prevalence of manifest amyloidosis in African countries is also unknown but is likely to increase in the future due to the increasing life expectancy [21]. This emphasizes the need for reliable and affordable diagnostic tools.

In patients with clinical amyloidosis (cardiac or neurological), the diagnostic yield of peripheral biopsies varies across studies. Specificity rates are excellent, consistently reaching 100%, while sensitivity rates show more variation. Depending on the etiology and in the most favorable studies, abdominal fat biopsies demonstrate a sensitivity from 58 to 95%, and rectal mucosal biopsies range from 74 to 95% [22–24]. Biopsies of accessory salivary glands exhibit sensitivities between 45 and 61%, and renal biopsies range from 85 to 100% [25–27]. Additionally, some studies report a very high diagnostic yield from endomyocardial biopsies in characteristic cardiac amyloidosis cases [11,28].

In this exploratory analysis, histopathological analysis of thoracic fat pad sample from the pacemaker or ICD pocket could diagnose only five from 16 patients with CA. 11 from in total 16 patients with diagnosis of CA had no evidence of amyloid in the histopathological analysis of the thoracic fat pad biopsy. Thus, the sensitivity of histopathological analysis to diagnose CA was poor with only 31%. Noteworthy, there were no false positive findings.

No correlation was observed between the myocardial uptake in bone scintigraphy and the thoracic fat pad biopsy results. Even though no wt-ATTR-CA was among the cases with positive amyloid finding in the thoracic fat pad sample, no reasonable conclusion can be drawn due to the small sample size.

Although this observation should be interpreted with great caution due to the small number of cases, we found in a group of eight patients with thoracic and abdominal fat pad biopsies an excellent correlation with identical results from both anatomical sites ($p=0.018$).

The limited sensitivity of 31% at thoracic fat pad biopsy in our study with 94% of patients with ATTR-CA is similar to findings of previous studies on abdominal fat pad aspiration. These studies demonstrated that sensitivity of abdominal fat pad aspiration greatly varies depending on the type of amyloid protein. Sensitivity for diagnosis of ATTR amyloidosis using abdominal fat pad aspiration

or biopsy was 15% in wt-ATTR-CA, 39% in ns-ATTR-CA and 45% in ATTRv-CA, whereas sensitivity in patients with AL-CA is up to 84-90% [29,30].

Limitations of this study include the small sample size (n=27), the retrospective single-center design, and the lack of power calculations due to the exploratory nature of the research, which may limit the generalizability of the findings to broader populations.

5. Conclusions

Our initial hypothesis that cardiac amyloidosis (CA) could be reliably diagnosed through histopathological analysis of thoracic fat pad samples obtained from pacemaker or ICD pockets was not confirmed. In this study, involving an Afro-Caribbean population with 94% diagnosed with ATTR-CA, thoracic fat pad biopsy demonstrated a sensitivity of 31% and specificity of 100% compared to protein electrophoresis or technetium-labeled bisphosphonate bone scintigraphy as standard diagnostic tools. Given its low diagnostic yield, histopathological analysis of thoracic fat pad biopsies cannot currently be recommended as a standard diagnostic tool for ATTR-CA. However, it may still hold significant value in diagnosing AL-CA, potentially obviating the need for further invasive procedures. This study underscores the importance of utilizing established standard diagnostic criteria for CA while also highlighting the need for less expensive and more accessible diagnostic methods, especially in resource-limited settings.

Credit statement: Cedric Mvita: data curation, investigation, writing of the original draft; Romain Vergier: conceptualization, investigation, methodology, editing of the original draft; Mathilda Simeon: formal analysis, investigation, editing of the original draft; Nathan Buila Bimbi: supervision; Nathan Malka: conceptualization; Karima Lounaci: data curation, validation; Maria Herrera Bethencourt: data curation, validation; Karim Farid: methodology, supervision; Arnt Kristen: methodology, supervision; Rishika Banydeen: conceptualization, methodology; Astrid Monfort: methodology, supervision; Jocelyn Inamo: methodology, supervision; Andreas Müssigbrodt: conceptualization, data curation, formal analysis, investigation, methodology, visualization, writing of the original draft

Data availability statement: The data that support the findings of this study are available from the corresponding author, (AM), upon reasonable request.

Conflict of interest: None declared

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