Experimental Models of Irritable Bowel Syndrome and the Role of the Enteric Neurotransmission

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Abstract

Irritable bowel syndrome (IBS) is one of the most common gastrointestinal diseases in humans. It is characterized by visceral pain and/or discomfort, hypersensitivity and abnormal motor responses along with change in gut habits. Although the etiopathogenesis of IBS is only partially understood, a main role has been attributed to psychosocial stress of different origin. Animals models such as neonatal maternal separation, water avoidance stress and wrap restraint stress have been developed as psychosocial stressors in the attempt to reproduce the IBS symptomatology and identify the cellular mechanisms responsible for the disease. The study of these models has led to the production of drugs potentially useful for IBS treatment. This review intends to give an overview on the results obtained with the animal models; to emphasize the role of the enteric nervous system in IBS appearance and evolution and as a possible target of drug therapies.

Keywords:

corticotropin releasing factor, irritable bowel syndrome (IBS), maternal separation (MS), neurotransmitters, pain, psychosocial stress, visceral hyperalgesia, water avoidance stress (WAS), wrap restrain stress (WRS).

Introduction

Irritable bowel syndrome (IBS) is a highly prevalent (12-20% in western countries) and multifactorial gastrointestinal disorder (1) whose symptoms are mainly referred to the large bowel. Clinically the IBS is considered a chronic disease characterized by the alternation of remission periods and periods of symptoms exacerbation (2). Several pathophysiological mechanisms may contribute to IBS manifestations; current concepts attribute an important role to visceral hypersensitivity. In colonic mucosal biopsies of IBS patients it has been shown an increase in activated lymphocytes, mast cells, neutrophilic (3,4) and eosinophilic granulocytes (5) suggesting that inflammation might play a role in IBS hyperalgesia. However, in randomized, double-blind, placebo-controlled trials, the administration of antiinflammatory drugs while decreased the infiltration of immune cells, including mast cells, in the mucosal biopsies, had no significant effect on abdominal pain, bloating or bowel habits, thus leading to reconsider the role of inflammation in hyperalgesia in IBS (6,7). Indeed, although IBS is commonly listed as a functional disorder, the presence of mucosal barrier alterations, low-grade inflammation, brain-gut axis dysfunction and dysbiosis represent evidences for an organic origin of the disease (8) The etiology of IBS is still poorly known, however, psychosocial stress is the most widely acknowledged risk factor for the development and/or relapse of IBS (9,10). Notably, up to 80% IBS patients experience comorbid behavioral disorders, such as anxiety or depression (11).

In summary, it is possible that hyperalgesia and psychosocial stressors are able to aliment each other in a vicious circuit responsible for the chronic progression of the disease. The possibility to interrupt this circuit is a challenge for researchers and clinicians.

Psychosocial stress belongs to the exteroceptive stressors (12,13) and are limbic-sensitive and dependent upon the forebrain cognitive circuits mediating the endocrine and autonomic responses to these types of stressors. In the attempt to improve the knowledges of the mechanisms underlining IBS, animal models reproducing psychosocial stressing conditions able to induce some of the IBS main signs, notably the hyperalgesia and pain (14, 15, 16), have been developed. The choice of one or another of these tests is not random since each of them present diverse advantages and limits that have to be weight before initiating the experimental procedure.

Presently we will review three of the most used psychosocial stress models: the wrap restrain stress (WRS), an acute stress and two models of chronic stress: the maternal separation (MS) and the water avoidance stress (WAS). In parallel we will focused our attention on those data, obtained by us and other researchers, that highlight the role played by the enteric nervous system in the genesis of IBS symptomatology and the available experimental or clinical pharmacological therapies affecting, directly or indirectly, the enteric neurotransmission (Fig.1).

Animal models of IBS

The wrap restrain stress (WRS)

The WRS was introduced in the scientific literature more than 30 years ago as the model for human IBS (17). Since then, further investigations have partially revised this hypothesis and currently no animal model is considered able to wholly mimic the human disease. The WRS model is commonly applied once (acute test) and consists of a forced immobilization of the animal lasting at least for 2 hours. The efficacy of this test is confirmed by the development of an immediate hyperalgesia, quantifiable in colon-rectal distention (CRD) number (18), the inhibition of small intestinal transit, the stimulation of large intestinal activity and increased fecal excretion (17). We recently demonstrated that rats underwent to the WRS presented a low-grade mucosal inflammation with a significant increase in mast cells and eosinophylic granulocytes (19,20) that overlapped that described in colonic biopsies of IBS (21). Moreover, these animals showed important changes in the glial cells, in inhibitory and excitatory neurotransmitters and receptors that were interpreted as responsible for the dysmotility and hypersensitivity present in IBS patients. Therefore, it is reasonable to conclude that WRS represents a suitable model for reproducing, at least in part, the main symptoms present in IBS (i.e. hypersensitivity and dysmotility), and to affirm that WRS shares the *face-validity*, one of the criteria proposed by Meyer and Collins, (22) to validate an animal model. It is matter of debate whether WRS is adequate to investigate potential drugs (16). However, WRS misses the most characterizing clinical condition of IBS such as the chronic course. In other words, it does not show good construct-validity (22), In agreement, when WRS is applied repetitively it triggers adaptive responses (16, 23) in opposite to what happens in patients affected by IBS.

Chronic stressors

Beyond the chronic course, accurate anamneses of IBS patients have attested a significant association between IBS and childhood traumas of different origin or the presence of repeated stress situations in the adulthood (24). Thus, the pre-existence/recurrence of adverse conditions characterize and correlate with the development and maintenance of IBS (8). These clinical data have moved the researchers to look for animal models carrying chronic rather than acute stressors. Two main model of chronic stress have been proposed to mimic childhood trauma or repeated stress conditions in the adulthood, the maternal separation (MS) and the water avoidance stress (WAS), respectively. These two behavioral tests are distinct not only for the period of application but also for the intensity being the latter considered a mild chronic stress compared to MS. Nevertheless, they share several features, are able to induce a symptomatology similar to that of IBS and reveal sex-differences in sensitivity.

The Maternal Separation (MS)

The stress consists in removing the puppies from the mother for 3 h per day during the first two weeks of life. Since maternal care affects the hypothalamic-pituitary-adrenal (HPA) axis and the cognitive and emotional functions (25), the MS causes stable changes in the central nervous system of these animals (26) At the level of large intestine, MS promotes, in the adult animals, the development of a condition characterized by visceral hypersensitivity to colorectal distension (CRD) (27-29) and colonic mast cell hyperplasia often concentrates close to nerve endings (21), that are two typical signs of IBS (4,30,31). Moreover, these animals manifest an hypersensitivity to acute psychosocial stress application i.e. the WAS applied for 1 h (32).

Interestingly, the experimental data demonstrated a greater sensitivity to MS of females compared to males (16). Notably, IBS incidence is twofold greater in women than in men and this is true independently of the triggering factors (8). On the other hand, the potency of this stressor is such that the insurgence of depressive symptoms rather than anxiety is likely (33). In summary, the MS model possesses either *face*- or *construct*- (consistency with the hypothetical pathogenesis) *validity* (22).

The water avoidance stress (WAS)

Several reports agree that daily chronic stress predicts the severity of the IBS symptoms (34-36). In the light of these observations, rodent models characterized by alternating exposure to stress have been settled and WAS was considered one of the most effective psychological stressor carrying this property. Literature data showed that, in male Wistar rats, a strain highly responsiveness, chronic application of WAS for 10 days provoked colonic hyperalgesia, increased colonic motility and caused local inflammation. Moreover, the colonic signs persisted for the following 20 days (37). The main limit of this model regards its susceptibility to several factors such as animal strain, sex and environmental conditions that could affect the results (8,16). To synthesize, the WAS model carries both *face-* and *construct-validity* (22).

Less known and still matter of investigation is the possibility that the two models of chronic stress presently reviewed possess also the *predictive validity* i.e. how well the models are responsive to treatment showing some efficacy in alleviating visceral pain to predict triggered treatment. Noteworthy, WAS shows some peculiarities, i.e. the absence of a tolerance after repeated applications and the maintenance of hyperalgesia and hypermotility after the stress is removed, that make it particularly suitable to investigate the ability of drugs to prevent or attenuate the appearance and evolution of the colonic alterations induced by stress, and also to evaluate long lasting beneficial effects of the drug after its withdrawal.

THE ENTERIC NEUROTRANSMISSION AS A POTENTIAL TARGET OF IBS TREATMENT

It has been suggested that pain signals originate in intrinsic primary afferent neurons and are transmitted by extrinsic primary afferent neurons (38). Afferent nerve endings are endowed with a variety of pro- and antinociceptive ion channels and receptors; the balance between pain sensing and suppressing signals finally determines their activation status (38). Important neurotransmitters involved in visceral sensation are CRF, 5-HT and neurokinins; channels mediating activation of afferent nerves are e.g. transient receptor potential (TRP) ion channels, that act as molecular detector of thermal and chemical stimuli that activate sensory neurons to produce acute or persistent pain. Due to recent development of drugs approved for IBS, guanylate cyclase (GC) receptors and chloride channels have received much attention.

Cortocotropin releasing factor (CRF)

Psychological stress at any point of the lifetime can bring to permanent alterations of several systems such as the descending pain modulatory system, the immune system, the gut microbiota. Preclinical evidence has accumulated over the years suggesting that stress-related alterations of colonic motor and visceral functions are primarily mediated by the activation of brain CRFr/CRF1 signaling pathway, while CRF2 receptor activation may exert a counteracting influence (39-41). CRFr are located in effector neurons of the hypothalamus, the amygdala, the cingulate cortex and the locus caeruleus complex. Central injection of CRF can reproduce behavioral and physiological reactions similar to those seen in response to acute psychological stress (42,43), and inhibition of CRF-mediated responses by antagonists (43,44) or in knockout animals results in a decrease of the animals' answer to stress (45,46). However, recent experimental studies point to an equally important contribution of the gut CRFr1-2/CRF signaling to the GI stress response (20,47). In the GI tract the CRFr are expressed by enteric neurons (Fig.2) and mucosal cells (19,20,48-52) and an upregulation of these receptors has been reported in the colon of male Wistar rats underwent to delayed stress-induced visceral hyperalgesia (37,44) and WRS (19,20). Although CRF has been the subject of thirty three years of efforts in research by academia and pharmaceutical industry that resulted in several thousand papers and patents, yet little progress has been made to identify and market diagnostic or therapeutic CRF peptides and small molecule ligand and no modulators of this peptide has reached the pharmaceutical market for IBS so far (53).

The serotonin or 5-hydroxytriptamine (5-HT)

Several lines of evidence indicate that altered 5-HT signaling in the central nervous system and in the gut contributes to the pathophysiology in IBS (54). Further, patients with IBS-diarrhea (IBS-D) have increased postprandial plasma 5-HT, while those with IBS-costipation (IBS-C) have reduced postprandial 5-HT levels (55). 5-HT released from enterochromaffin cells regulates sensory, motor and secretory functions of the digestive system through the interaction with different receptor subtypes. A recent experimental work (56) has shown that 5-HT3r antagonist alosetron and the CRF1r antagonist E2508 reduced defecation and visceral pain in WRS-subjected rats. Nevertheless, the anti-serotoninergic drugs that have been introduced on the market such as tegaserod, a 5-HT-4 agonist, and the same alosetron, seem to be effective only

in a limited part of the IBS patients and suffer from the evidence of side effects (ischemic colitis, diarrhea, cardiovascular ones) that has let FDA to impose limitation of their widest use (https://www.alosetronrems.com/AlosetronUI/rems/pdf/letterforhealthcareproviders.pdf; https://www.fda.gov/Drugs/DrugSafety/ucm103223.htm). Newer 5-HT4 agonists such as prucalopride, naronapride, velusetrag, and YKP10811 have been found active only in chronic idiopathic constipation. Further studies are needed in order to develop an optimal treatment in the field of serotonin.

Tachykinins

NK2r antagonists have been proven effective in animal models of visceral hyperalgesia induced by inflammation (57,58) or WAS (59). Nepadutant (58), ibodutant (57) or saredutant (59) were found able to inhibit rectal hypersensitive responses in rats, guinea-pigs or gerbils pretreated with TNBS or previously subjected to restraint, suggesting that the NK2r has a role in mediating visceral allodynia/hyperalgesia. This role was further supported by the studies of Birder et al. (60), who showed that the increased expression of either c-fos and c-jun protooncogene markers in spinal cord and dorsal root ganglia neurons of rats pretreated with TNBS was prevented by nepadutant, and by Laird et al., (61), who found nepadutant capable of preventing the hypersensitivity of single spinal cord neurons responding to colorectal distension or pelvic nerve stimulation in rats pretreated with intracolonic acetic acid. Laird et al., (62) have also shown that intracolonic instillation of acetic acid or capsaicin fails to produce either acute (cardiovascular) responses or primary hyperalgesia in NK1r-knockout mice, thus suggesting a role for even this latter receptor, as mediator of visceral hyperalgesia. The NK1r antagonist aprepitant effectively reduced defecation and CD as well increase in adrenocorticotropin in gerbils subjected to WRS (59). Conversely, a recent study suggests that the SP system might play little role in the development of visceral hyperalgesia in the neonatal maternal separation (MS) rat model (63). We found a decrease in myenteric NK1immunoreactive (IR) neurons and decrease in Substance P-IR nerve fibres in the muscle wall after WRS (19).

Also the NK3r (located at peripheral or/and central level) could play a role in visceral hyperalgesia, due to the reported effectiveness of either intraperitoneal (64) or intrathecal (65) administration of the NK3r selective antagonist SR 142801 in

reducing reflex abdominal contractions elicited by various nociceptive stimuli. Gender and species differences are reported in the modulating effect on pain of tachykinin antagonists (57,66,67)

In spite of these promising experimental results, none of these NKr antagonists has reached the market due to limitation of their effect in humans and presence of side effects found in the development clinical phases.

Calcium channels

It has been reported that T-type calcium channels encoded by the Ca(V)3.2 isoform are expressed in colonic nociceptive primary afferent neurons and that they contribute to the exaggerated pain perception in a butyrate-mediated rodent model of IBS (68). These preliminary results render T-type antagonists as potential candidates for the development of novel therapeutic agents in the treatment of IBS. A modulation of Ttype calcium channels has been reported for the antispasmodic otilonium bromide (OB) (69). OB is a quaternary ammonium derivative poorly absorbed from the GI tract, that concentrates mainly in the large intestine acting locally as an L-type calcium channel blocker, an antimuscarinic and a NK2r antagonist (70). Probably thanks to this multi-target activity, OB effectively reduced pain and improved defecation alterations in placebo-controlled trials in IBS patients (71). Conversely, OB tested in experimental models of visceral sensitivity, at least in WRS, was less active as compared to the other agents previously reviewed (20). Another antispasmodic such as pinaverium bromide endowed with L-type calcium channel blocker and antimuscarinic activity improved motility disorders and consequently reduced stool problems in IBS patients.

Others receptors and channels

Since irritation, immune challenge and inflammation releases both tachykinin and CGRP from extrinsic afferents and from intrinsic neurons inside the gut, these peptides may either increase the peripheral sensory gain of extrinsic afferents or contribute to the afferent transmission of algetic signal within the central nervous system. As gut extrinsic afferents do not possess receptors for CGRP, it is probable that peptide-induced sensitization or excitation of sensory neurons is an indirect consequence of peptide action on gastrointestinal effectors (38). In experimental models of colorectal distension-induced visceral pain, CGRP has been shown to play

a role (72). In rats underwent to MS it has been found that the high number of mast cells associated with an increase in CGRP-IR (27); similarly, an increase in CGRP expression and a higher number of mast cells were described in the mucosa and submucosa of rats underwent to WRS (19). Noteworthy, in rectal biopsy from patients with well characterized rectal hypersensitivity a marked increase in CGRP was found (73).

The increase in the vanilloid receptor TRPV1 significantly correlated with the decrease in heat and distension sensory threshold confirming that both afferent and intrinsic fibers take part in the change of signaling pathways of visceral hypersensitivity. Further, WAS could induce the upregulation of TRPV1 and TRPA1 in the colonic afferent DRG indicating these receptors as candidate molecules in stress-induced hyperalgesia in rats (74). An antispasmodic largely used in USA for IBS such as peppermint oil, whose active component is menthol, has recently been found antinociceptive thanks to its capability to activate temperature sensing ion channel TRPM8 (75).

It should be noted that while modulators of vanilloid receptors are under preclinical development, activators of guanylate cyclase (GC) receptor has produced a drug successfully introduced in the market for IBS with predominant constipation (IBS-C) such as linactotide. Linaclotide has shown potent anti-nociceptive effects in several mechanistically different rodent models of visceral hypersensitivity (WRS, WAS, inflammatory conditions; see Fig.3) both in Wistar rats and GC-null mice (76). It activates the GC-C expressed on mucosal epithelial cells, resulting in the production and release of cGMP. This extracellular cGMP acts on and inhibits nociceptors, thereby reducing nociception (77). In this area another GC activator such as plecanatide will be reviewed by the FDA for approval in IBS-C indication in the first quarter of 2018 (78).

Another drug approved for IBS-C is lubiprostone, a chloride channel activator which acts locally on the apical membrane of epithelial cells in the gastrointestinal lumen. As a derivative of prostaglandin E1, lubiprostone selectively combines with prostaglandin receptor to activate chloride channel 2 leading to chloride-rich fluid secretion. Increasing fluid secretion softens stools and improves intestinal motility, which finally contributes to alleviate constipation symptoms of IBS-C (78).

Tenapanor is a small-molecule inhibitor of the gastrointestinal sodium/hydrogen exchanger NHE₃, a protein present in the apical side of enterocytes, which results in

increased intraluminal sodium and water excretion and is in active development for IBS-C (79)

Translocator protein 18 kDa (TSPO) is a five-domain transmembrane protein that is highly expressed in steroid-producing tissues, including the glial cells within the brain. ONO-2952 is a novel and selective inhibitor of TSPO that reduces stress-induced defectaion and visceral hyperalgesia in rat models (80).

Other class drug challenged in IBS are those acting on opioid or GABA receptors. Among opioid agonists loperamide and diphenoxylate, are anti-diarrheal agents that have been used in IBS for many years but their efficacy is already discussed and not completely proven while the recent one eluxadoline has been licensed in USA for IBS-D with some limitations due to its side effects (https://www.fda.gov/DrugS/DrugSafety/DrugSafetyPodcasts/ucm547907.htm),

GABAergic agents are alpha2β ligands that generally bind potently to an auxiliary protein associated with voltage-gated calcium channels, reducing depolarization-induced calcium influx at nerve terminals. This reduces the release of several excitatory neurotransmitters, including glutamate, noradrenaline, substance P, and calcitonin gene-related peptide (CGRP), which are involved in pain mechanisms. Gabapentin and pregabalin did not show a good efficacy/risk ratio in clinical trials.

Conclusions

Despite the limits of the IBS experimental models as compared to the complex picture characterizing the human manifestations, these models have allowed to confirm the consistent role of psychosocial stressors in the genesis and time-course of the gut symptomatology. The models analyzed (WRS, WAS, MS) produce visceral hypersensitivity that is one the main sign of IBS. A lot of neurotransmitters seem to be involved in this nociception and modulators of them are generally active in reduce experimental pain. Due to complexity of this phenomenon only few of these latter molecules have been found active in clinical studies and this has hampered the development of new drugs for IBS.

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References

- 1. Canavan, C.; West, J.; Card, T. The epidemiology of irritable bowel syndrome. *Clin. Epidemiol.* **2014**, 6, 71–80.
- 2. Evangelista, S. Benefits from long-term treatment in irritable bowel syndrome. *Gastroenterol. Res Pract.* **2012**, 2012: 936-960.
- 3. Chadwick, V.S.; Chen, W.; Shu, D.; Paulus, B.; Bethwaite, P.; Tie, A.; Wilson, I. Activation of the mucosal immune system in irritable bowel syndrome. *Gastroenterology* **2002**, 122, 1778–1783.
- 4. O'Sullivan, M.; Clayton, N.; Breslin, N.P.; Harman, I.; Bountra, C.; McLaren, A.; O'Morain, C.A. Increased mast cells in the irritable bowel syndrome. *Neurogastroenterol*. Motil. **2000**, 12, 449–457.
- 5. Walker, M.M.; Warwick, A.; Ung, C.; Talley, N.J. The role of eosinophils and mast cells in intestinal functional disease. *Curr. Gastroenterol. Rep.* **2011**, 13, 323-330.
- 6. Tuteja, A.K.; Fang, J.C.; Al-Suqi, M.; Stoddard, G.J.; Hale, D.C. Double-blind placebo-controlled study of mesalamine in post-infective irritable bowel syndrome-a pilot study. *Scand. J. Gastroenterol.* **2012**, 47, 1159-1164.
- 7. Sarna, S.K. Lessons Learnt from Post-Infectious IBS. *Front, Physiol*, **2011**, 23, 2-49.
- 8. Moloney, R.D.; O'Mahony, S.M.; Dinan, T.G.; Cryan, J.F. Stress-induced visceral pain: toward animal models of irritable-bowel syndrome and associated comorbidities. *Front. Psychiatry* **2015**, 6,15-45.
- 9. Mayer, E.A.; Naliboff, B.D.; Chang, L.; Coutinho, S.V. Stress and the Gastrointestinal Tract V. Stress and irritable bowel syndrome *Am. J. Physiol. Gastrointest. Liver Physiol.* **2001**, 280, G519–G524.
- Rosenberger, C.; Elsenbruch, S.; Scholle, A.; de Greiff, G.A.; Schedlowski, M.; Forsting, M.; Gizewski, E.R. Effects of psychological stress on the cerebral processing of visceral stimuli in healthy women. *Neurogastroenterol*. *Motil.* 2009, 21, 740-e45.
- 11. Fond, G.; Loundou, A.; Hamdani, N.; Boukouaci, W.; Dargel, A.; Oliveira, J.; Roger, M.; Tamouza, R.; Leboyer, M.; Boyer, L. Anxiety and depression comorbidities in irritable bowel syndrome (IBS): a systematic review and meta-analysis. *Eur. Arch. Psychiatry Clin. Neurosci.* **2014**, 264, 651-660.

- 12. Herman, J.P.; Cullinan, W.E. Neurocircuitry of stress: central control of the hypothalamo-pituitary-adrenocortical axis. *Trends Neurosci.* **1997**, 20, 78–84.
- 13. Sawchenko, P.E.; Li, H.Y.; Ericsson, A. Circuits and mechanisms governing hypothalamic responses to stress: a tale of two paradigms. *Prog. Brain Res.* **2000**, 122, 61–78.
- 14. Bouin, M.; Plourde, V.; Boivin, M.; Riberdy, M.; Lupien, F.; Laganière, M.; Verrier, P.; Poitras, P. Rectal distention testing in patients with irritable bowel syndrome: sensitivity, specificity, and predictive values of pain sensory thresholds. *Gastroenterology* **2002**, 122, 1771–1777.
- 15. Elsenbruch, S. Abdominal pain in irritable bowel syndrome: a review of putative psychological, neural and neuro-immune mechanisms. *Brain Behav. Immun.* **2011**, 25, 386–394.
- 16. Larauche, M.; Mulak, A.; Tachè Y. Stress and visceral pain: from animal model to clinical therapies. *Experimental Neurology* **2012**, 233, 49-67.
- 17. Williams, C.L.; Villar, R.G.; Peterson, J.M.; Burks, T.F. Stress-induced changes in intestinal transit in the rat: a model for irritable bowel syndrome. *Gastroenterology* **1988**, 94, 611–621.
- 18. Gue, M.; Del Rio-Lacheze, C.; Eutamene, H.; Theodorou, V.; Fioramonti, J.; Bueno, L. Stress-induced visceral hypersensitivity to rectal distension in rats: role of CRF and mast cells. *Neurogastroenterol. Motil.* **1997,** 9, 271–279.
- 19. Traini, C.; Evangelista, S.; Girod, V.; Faussone-Pellegrini, M.S.; Vannucchi, M.G. Changes of excitatory and inhibitory neurotransmitters in the colon of rats underwent to the wrap partial restraint stress (WRS). *Neurogastroenterol. Motil.* **2016**, 28,1172-1185.
- 20. Traini, C.; Evangelista, S.; Girod, V.; Faussone-Pellegrini, M.S.; Vannucchi, M.G. Repeated otilonium bromide administration prevents neurotransmitter changes in colon of rats underwent to wrap restraint stress. *J. Cell. Mol. Med.* **2017**, 21, 735-745.
- 21. Bashashati, M.; Moossavi, S.; Cremon, C.; Barbaro, M.R.; Moraveji, S.; Talmon, G.; Rezaei, N.; Hughes, P.A.; Bian Z.X.; Choi C.H.; et al. Colonic immune cells in irritable bowel syndrome: A systematic review and meta-analysis. *Neurogastroenterol. Motil.* **2017**, doi: 10.1111/nmo.13192.
- 22. Mayer, E.A.; Collins, S.M. Evolving pathophysiologic models of functional gastrointestinal disorders. *Gastroenterology* **2002**, 122, 2032–2048.
- 23. Bouwknecht, J.A. Behavioral studies on anxiety and depression in a drug discovery environment: keys to a successful future. *Eur. J. Pharmacol.* **2015**, 753, 158-176.

- 24. Mayer, E.A.; Labus, J.S.; Tillisch, K.; Cole, S.W.; Baldi, P. Towards a systems view of IBS. Nat Rev *Gastroenterol. Hepatol.* **2015**, 12, 592-605.
- 25. Fish, E.W.; Shahrokh, D.; Bagot, R.; Caldji, C.; Bredy, T.; Szyf, M.; Meaney, M.J. Epigenetic programming of stress responses through variations in maternal care. *Ann. N. Y. Acad. Sci.* **2004**, 1036, 167–180.
- 26. Szyf, M.; Weaver, I.; Meaney, M. Maternal care, the epigenome and phenotypic differences in behavior. *Reprod. Toxicol.* **2007**, 24, 9–19.
- 27. Barreau, F.; Salvador-Cartier, C.; Houdeau, E.; Bueno, L.; Fioramonti, J. Long term alterations of colonic nerve mast cell interactions induced by neonatal maternal deprivation in rats. *Gut* **2008**, 57, 582–590.
- 28. Welting, O.; Van Den Wijngaard, R.M.; De Jonge, W.J.; Holman, R.; Boeckxstaens, G.E. Assessment of visceral sensitivity using radio telemetry in a rat model of maternal separation. *Neurogastroenterol. Motil.* **2005**, 17, 838–845.
- 29. Rosztoczy, A.; Fioramonti, J.; Jarmay, K.; Barreau, F.; Wittmann, T.; Bueno, L. Influence of sex and experimental protocol on the effect of maternal deprivation on rectal sensitivity to distension in the adult rat. *Neurogastroenterol. Motil.* **2003**, 15, 679–686.
- 30. Whitehead, W.E.; Palsson, O.S. Is rectal pain sensitivity a biological marker for irritable bowel syndrome: psychological influences on pain perception. *Gastroenterology* **1998**, 115, 1263–1271.
- 31. O'Sullivan, M.; Clayton, N.; Breslin, N.P.; Harman, I.; Bountra, C.; McLaren, A.; O'Morain, C.A. Increased mast cells in the irritable bowel syndrome. *Neurogastroenterol. Motil.* **2000**, 12. 449–457.
- 32. Barreau, F.; Cartier, C.; Ferrier, L.; Fioramonti, J.; Buéno, L. Nerve growth factor mediates alterations of colonic sensitivity and mucosal barrier induced by neonatal stress in rats. *Gastroenterology* **2004**, 127, 524–534.
- 33. O'Mahony, S.M.; Hyland, N.P.; Dinan, T.G.; Cryan, J.F. Maternal separation as a model of brain-gut axis dysfunction. *Psychopharmacology (Berl)* **2011**, 214, 71–88.
- 34. Elsenbruch, S.; Rosenberger, C.; Bingel, U.; Forsting, M.; Schedlowski, M.; Gizewski, E.R. Patients with irritable bowel syndrome have altered emotional modulation of neural responses to visceral stimuli. *Gastroenterology* **2010**, 139, 1310–1319.
- 35. Elsenbruch, S.; Rosenberger, C.; Enck, P.; Forsting, M.; Schedlowski, M.; Gizewski, E.R. Affective disturbances modulate the neural processing of visceral pain stimuli in irritable bowel syndrome: an fMRI study. *Gut* **2010**, 59, 489–495.

- 36. Lackner, J.M.; Brasel, A.M.; Quigley, B.M.; Keefer, L.; Krasner, S.S.; Powell, C.; Katz, L.A.; Sitrin, M.D. The ties that bind: perceived social support, stress, and IBS in severely affected patients. *Neurogastroenterol. Motil.* **2010**, 22, 893–900.
- 37. Bradesi, S.; Schwetz, I.; Ennes, H.S.; Lamy, C.M.; Ohning, G.; Fanselow, M.; Pothoulakis, C.; McRoberts, J.A.; Mayer, E.A. Repeated exposure to water avoidance stress in rats: a new model for sustained visceral hyperalgesia. *Am. J. Physiol. Gatrointest. Liver Physiol.* **2005**, 289, G42-53.
- 38. Holzer, P.; Holzer-Petsche, U. Pharmacology of inflammatory pain: local alteration in receptors and mediators. *Dig. Dis.* **2009**, 27, 24-30.
- 39. Million, M.; Wang, L.; Wang, Y.; Adelson, D.W.; Yuan P.Q.; Maillot, C.; Coutinho, S.V.; McRoberts, J.A.; Bayati, A.; Mattsson, H.; et al. CRF2 receptor activation prevents colorectal distension induced visceral pain and spinal ERK1/2 phosphorylation in rats. *Gut* **2006**, 55, 172-181.
- 40. Fukudo, S. Role of corticotropin-releasing hormone in irritable bowel syndrome and intestinal inflammation. *J. Gastroenterol.* **2007**, 42 (Suppl. 17), 48-51.
- 41. Taché, Y.; Brunnhuber, S. From Hans Selye's discovery of biological stress to the identification of corticotropin-releasing factor signaling pathways: implication in stress-related functional bowel diseases. *Ann. N. Y. Acad. Sci.* **2008**, 1148, 29-41.
- 42. Owens, M.J.; Nemeroff, C.B. The role of corticotropin-releasing factor in the pathophysiology of affective and anxiety disorders: laboratory and clinical studies. *Ciba Found. Symp.* **1993**, 172, 296–316.
- 43. Taché, Y.; Martinez, V.; Million, M.; Rivier, J. Corticotropin releasing factor and the brain-gut motor response to stress. *Can. J. Gastroenterol.* **1999**, 13 (Suppl. A), 18A–25A.
- 44. Larauche, M.; Bradesi, S.; Million, M.; McLean, P.; Taché, Y.; Mayer, E.A.; McRoberts, J.A. Corticotropin-releasing factor type 1 receptors mediate the visceral hyperalgesia induced by repeated psychological stress in rats. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2008**, 294, G1033-1040
- 45. Smith, G.W.; Aubry, J.M.; Dellu, F.; Contarino, A.; Bilezikjian, L.M.; Gold, L.H.; Chen, R.; Marchuk, Y.; Hauser, C.; Bentley, C.A.; et al. Corticotropin releasing factor receptor 1-deficient mice display decreased anxiety, impaired stress response, and aberrant neuroendocrine development. *Neuron* **1998**, 20, 1093–1102.
- 46. Timpl, P.; Spanagel, R.; Sillaber, I.; Kresse, A.; Reul, J.M.; Stalla, G.K.; Blanquet, V.; Steckler, T.; Holsboer, F.; Wurst, W. Impaired stress response and reduced anxiety in mice lacking a functional corticotropin-releasing

- hormone receptor. Nat. Genet. 1998, 19, 162–166.
- 47. Larauche, M.; Kiank, C.; Tachè, Y. Corticotropin releasing factor signaling in colon and ileum: regulation by stress and pathophysiological implications. *J. Physiol. Pharmacol.* **2009**, 60 (Suppl. 7), 33-46.
- 48. Muramatsu, Y.; Fukushima, K.; Iino, K.; Totsune, K.; Takahashi, K.; Suzuki, T.; Hirasawa, G.; Takeyama, J.; Ito, M.; Nose, M.; et al. Urocortin and corticotropin-releasing factor receptor expression in the human colonic mucosa. *Peptides* **2000**, 21, 1799-1809.
- 49. Chatzaki, E.; Crowe, P.D.; Wang, L.; Million, M.; Taché, Y.; Grigoriadis, D.E. CRF receptor type 1 and 2 expression and anatomical distribution in the rat colon. *J. Neurochem.* **2004**, 90, 309–316.
- 50. Liu, S.; Gao, X.; Gao, N.; Wang, X.; Fang, X.; Hu, H.Z.; Wang, G.D.; Xia, Y.; Wood, J.D. Expression of type 1 corticotropin-releasing factor receptor in the guinea pig enteric nervous system. *J. Comp. Neurol.* **2005**, 481, 284-298.
- 51. Yuan, P.Q.; Million, M.; Wu, S.V.; Rivier, J.; Tachè Y. Peripheral corticotropin releasing factor (CRF) and a novel CRF1 receptor agonist, stressin1-A activate CRF1 receptor expressing cholinergic and nitrergic myenteric neurons selectively in the colon of conscious rats. *Neurogastroenterol. Motil.* **2007**, 19, 923–936.
- 52. Yuan, P.Q.; Wu, S.V.; Elliott, J.; Anton, P.A.; Chatzaki, E.; Million, M.; Taché, Y. Expression of corticotropin releasing factor receptor type 1 (CRF1) in the human gastrointestinal tract and upregulation in the colonic mucosa in patients with ulcerative colitis. *Peptides* **2012**, 38, 62-69.
- 53. Rivier, J.E. Prospective Clinical Applications of CRF Peptide Antagonists. *Curr. Mol. Pharmacol.* **2017**, 10, 264-269.
- 54. Stasi, C.; Bellini, M.; Bassotti, G.; Blandizzi, C.; Milani, S. Serotonin receptors and their role in the pathophysiology and therapy of irritable bowel syndrome. *Tech. Coloproctol.* **2014**, 18, 613-621.
- 55. Atkinson, W.; Lockhart, S.; Whorwell, P.J.; Keevil, B.; Houghton, L.A. Altered 5-hydroxytryptamine signaling in patients with constipation- and diarrhea-predominant irritable bowel syndrome. *Gastroenterology* **2006**, 130, 34–43.
- 56. Taguschi, R.; Shikata, K.; Furuya, Y.; Hirakawa, T.; Ino, M.; Shin, K.; Shibata, H. Selective corticotropin-releasing factor 1 receptor antagonist E2508 reduces restraint stress-induced defecation and visceral pain in rat models. *Psychoneuroendocrinology* **2017**, 75. 110-115.
- 57. Bellucci, F.; Buéno, L.; Bugianesi, R.; Crea, A.; D'Aranno, V.; Meini, S.; Santicioli, P.; Tramontana, M.; Maggi, C.A. Gender-related differential effect

- of tachykinin NK2 receptor-mediated visceral hyperalgesia in guinea pig colon. *Br. J. Pharmacol.* **2016**, 173, 1329-1338.
- 58. Toulouse, M.; Coelho, A.M.; Fioramonti, J.; Lecci, A.; Maggi, C.A.; Bueno, L. Role of tachykinin NK₂ receptors in normal and altered rectal sensitivity in rats. *Br. J. Pharmacol.* **2000**, 129, 193-199.
- 59. Kakol-Palm, D.; Brusberg, M.; Sand, E.; Larsson, H.; Martinez, V.; Johansson, A.; von Mentzer, B.; Påhlman, I.; Lindström, E. Role of tachykinin NK(1) and NK(2) receptors in colonic sensitivity and stress-induced defecation in gerbils. *Eur. J. Pharmacol.* **2008**, 582, 123-131.
- 60. Birder, L.A.; Kiss, S.; De Groat, W.C.; Lecci, A.; Maggi, C.A. Effect of MEN 11420, an NK2 tachykinin antagonist, on immediate-early gene expression following TNBS induced colitis in the rat. *J. Pharmacol. Exp. Ther.* **2003**, 304, 272-276.
- 61. Laird, J.M.; Olivar, T.; Lopez-Garcia, J.A.; Maggi, C.A.; Cervero, F. Responses of rat spinal neurons to distension of inflamed colon: role of tachykinin NK₂ receptors. *Neuropharmacol.* **2001**, 40, 696-701.
- 62. Laird, J.M.; Olivar, T.; Roza, C.; De Felipe, C.; Hunt, S.P.; Cervero, F. Deficits in visceral pain and hyperalgesia of mice with a disruption of the tachykinin NK₁ receptor gene. *Neuroscience* **2000**, 98, 345-352.
- 63. Teng, W.; Chen, H.; Guo, F.; Du, X.; Fu, X.; Fang, Y.; Zhang, H.; Fang, M.; Ding, M. Expression and distribution of SP and its NK1 receptor in the braingut axis in neonatal maternally separated rat model with visceral hypersensitivity. *Genet. Mol. Res.* **2016**, 15, doi: 10.4238/gmr.15038999.
- 64. Julia, V.; Su, X.; Bueno, L.; Gebhart, G.F. Role of neurokinin 3 receptors on responses to colorectal distension in the rat: electrophysiological and behavioral studies. *Gastroenterology* **1999**, 116, 1124-1131.
- 65. Kamp, E.H.; Beck, D.R.; Gebhart, G.F. Combinations of neurokinin receptor antagonists reduce visceral hyperalgesia. *J. Pharmacol. Exp. Ther.* **2001**, 299, 105-113.
- 66. Bradesi, S.; Eutamene, H.; Fioramonti, J.; Bueno, L. Acute restraint stress activates functional NK1 receptor in the colon of female rats: involvement of steroids. *Gut* **2002,**50, 349-354.
- 67. Gaudreau, G.A.; Plourde, V. Role of tachykinin NK1, NK2 and NK3 receptors in the modulation of visceral hypersensitivity in the rat. *Neurosci. Lett.* **2003**, 351, 59-62.
- 68. Marger, F.; Gelot, A.; Alloui, A.; Matricon, J.; Ferrer, J.F.; Barrère, C.; Pizzoccaro, A.; Muller, E.; Nargeot, J.; Snutch, T.P.; et al. T-type calcium channels contribute to colonic hypersensitivity in a rat model of irritable bowel syndrome. *Proc. Natl. Acad. Sci. U. S. A.* **2011,** 108, 11266-11273

- 69. Strege, P.R.; Sha, L.; Beyder, A.; Bernard, C.E.; Perez-Reyes, E.; Evangelista, S.; Gibbons, S.J.; Szurszewski, J.H.; Farrugia, G. T-type Ca(2+) channel modulation by otilonium bromide. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2010**, 298, G706-713
- 70. Martínez-Cutillas, M.; Gil, V.; Gallego, D.; Mañé, N.; Martín, M.T.; Jiménez, M. Mechanisms of action of otilonium bromide (OB) in human cultured smooth muscle cells and rat colonic strips. *Neurogastroenterol. Motil.* **2013**, 25, e803-812.
- 71. Clavé, P.; Acalovschi, M.; Triantafillidis, J.K.; Uspensky, Y.P.; Kalayci, C.; Shee, V.; Tack, J.; OBIS Study Investigators. Randomised clinical trial: otilonium bromide improves frequency of abdominal pain, severity of distention and time to relapse in patients with irritable bowel syndrome. *Aliment. Pharmacol. Ther.* **2011**, 34, 432-442.
- 72. Plourde, V.; St-Pierre, S.; Quirion, R. Calcitonin gene-related peptide in viscerosensitive response to colorectal distension in rats. *Am. J. Physiol.* **1997**, 273, G191-196.
- 73. Chan, C.L.H.; Facer, P.; Davis, J.B.; Smith, G.D.; Egerton, J.; Williams, N.S.; Anand, P. Sensory fibres expressing capsaicin receptor TRPV1 in patients with rectal hypersensitivity and faecal urgency. *Lancet* **2003**, 361, 385-391.
- 74. Yu, Y.B.; Yang, J.; Zuo, X.L.; Gao, L.J.; Wang, P.; Li, Y.Q. Transient receptor potential vanilloid-1 (TRPV1) and ankyrin-1 (TRPA1) participate in visceral hyperalgesia in chronic water avoidance stress rat model. *Neurochem. Res.* **2010**, 35, 797-803.
- 75. Liu, B.; Fan, L.; Balakrishna, S.; Sui, A.; Morris, J.B.; Jordt, S.E. TRPM8 is the principal mediator of menthol-induced analgesia of acute and inflammatory pain. *Pain* **2013**, 154, 2169–2177.
- 76. Eutamene, H.; Bradesi, S.; Larauche, M.; Theodorou, V.; Beaufrand, C.; Ohning, G.; Fioramonti, J.; Cohen, M.; Bryant, A.P.; Kurtz, C.; et al. Guanylate cyclase C-mediated antinociceptive effects of linaclotide in rodent models of visceral pain. *Neurogastroenterol. Motil.* 2010, 22, 312-e84.
- 77. Castro, J.; Harrington, A.M.; Hughes, P.A.; Martin, C.M.; Ge, P.; Shea, C.M.; Jin, H.; Jacobson, S.; Hannig, G.; Mann, E.; et al. Linaclotide inhibits colonic nociceptors and relieves abdominal pain via guanylate cyclase-C and extracellular cyclic guanosine 3',5'-monophosphate. *Gastroenterology* **2013**,145, 1334-1346.e1-11.
- 78. Camilleri, M.; Ford, A.C. Pharmacotherapy for irritable bowel syndrome. *J. Clin. Med.* **2017**, 6, pii: E101. doi: 10.3390/jcm6110101.
- 79. Chey, W.D.; Lembo, A.J.; Rosenbaum, D.P. Tenapanor treatment of patients with constipation-predominant irritable bowel syndrome: A phase 2,

- randomized, placebo-controlled efficacy and safety trial. *Am. J. Gastroenterol.* **2017,** 112, 763–774.
- 80. Mitsui, K.; Niwa, T.; Kawahara, Y.; Morimoto, N.; Ohmoto, K.; Kato, M.; Yamaura, Y.; Yoshimoto, N.; Suna, H.; Katsumata, S. Anti-stress effects of ONO-2952, a novel translocator protein 18 kDa antagonist, in rats. *Neuropharmacology* **2015**, 99, 51–66.

Legend to the Figures

- Fig. 1: IBS pharmacotherapy, Modified from Camilleri and Ford (78).
- Fig. 2: **A**: CRFr1 and **B**: CRFr2 -immunoreactivity in the myenteric and submucous plexuses of rat colon. Bar=40μm.
- Fig 3: A: Effect of oral administration of linaclotide (0.3, 3 or 30 microg/kg) or vehicle on acute restraint stress-induced colorectal hypersensitivity to colorectal distension (CRD). **B**: Effect of acute water avoidance stress on electromyografic response (EMG) to CRD. An acute session of WAS-induced increased EMG response 24 h later. Linaclotide at 3 microg/kg p.o. completely abolished stress-induced hyperalgesia. Modified from Eutamene et al (76).

Figure 1

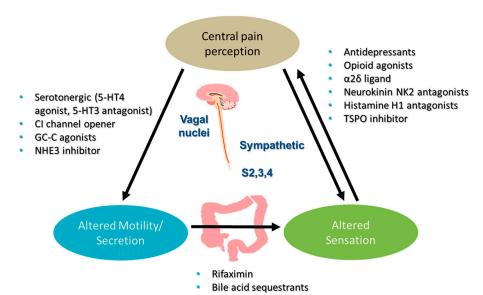


Figure 2

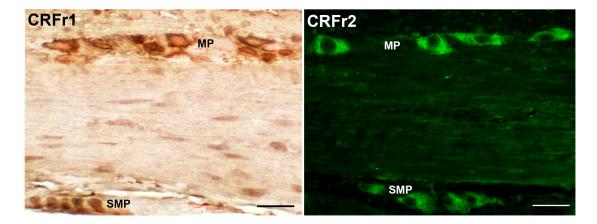


Figure 3

