

---

# From Polyphenols to Prodrugs: Bridging the Blood–Brain Barrier with Nanomedicine and Neurotherapeutics

---

[Masaru Tanaka](#)<sup>\*,†</sup>, Adriano Cressoni Araujo, [Vitor Engrácia Valenti](#), Elen Landgraf Guiguer, Vitor Cavallari Strozze Catharin, Cristiano Machado Gualhardi, Eliana de Souza Bastos Mazuqueli Pereira, Ricardo de Alvares Goulart, Rafael Santos de Argolo Haber, Atonelly Cassio Alves de Carvalho, [Sandra Maria Barbalho](#)<sup>\*,†</sup>

Posted Date: 5 February 2026

doi: 10.20944/preprints202601.2275.v2

Keywords: central nervous system diseases (CNS); blood-brain barrier (BBB); drug delivery systems; nanomedicine; phytochemicals; nanoparticles; prodrugs; drug administration; intranasal; ultrasonic therapy; transferrin receptor



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a [Creative Commons CC BY 4.0 license](#), which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

# From Polyphenols to Prodrugs: Bridging the Blood–Brain Barrier with Nanomedicine and Neurotherapeutics

Masaru Tanaka <sup>1,\*†</sup>, Adriano Cressoni Araujo <sup>2,3</sup>, Vítor Engrácia Valenti <sup>4</sup>, Elen Landgraf Guiguer <sup>2,3</sup>, Vitor Cavallari Strozze Catharin <sup>2</sup>, Cristiano Machado Gualhardi <sup>2</sup>, Eliana de Souza Bastos Mazuqueli Pereira <sup>3</sup>, Ricardo de Alvares Goulart <sup>3</sup>, Rafael Santos de Argolo Haber <sup>2</sup>, Atonelly Cassio Alves de Carvalho <sup>3</sup> and Sandra Maria Barbalho <sup>2,3,5,\*†</sup>

<sup>1</sup> Danube Neuroscience Research Laboratory, HUN-REN-SZTE Neuroscience Research Group, Hungarian Research Network, University of Szeged (HUN-REN-SZTE), H-6725 Szeged, Hungary

<sup>2</sup> Department of Biochemistry and Pharmacology, School of Medicine, Faculdade de Medicina de Marília – UNIMAR

<sup>3</sup> Graduate Program in Structural and Functional Interactions in Rehabilitation, School of Medicine, Universidade de Marília (UNIMAR), Marília 17525-902, SP, Brazil

<sup>4</sup> Systematic Reviews and Meta-Analyses Center, School of Philosophy and Sciences, São Paulo State University, Marília 17525-900, SP, Brazil

<sup>5</sup> Research Coordination – UNIMAR Charity Hospital, Faculdade de Medicina de Marília – UNIMAR

\* Correspondence: tanaka.masaru.1@med.u-szeged.hu (M.T.); smbarbalho@gmail.com (S.M.B.); Tel.: +36-62-342-847 (M.T.); +55 14 99655-3190 (S.M.B.)

† These authors contributed equally to this work.

## Abstract

Central nervous system disorders drive disability, yet many neuroactive candidates fail because the brain is a hard compartment to dose. Plant derived molecules spanning polyphenols, alkaloids, terpenoids, and cannabinoids are attractive because their pleiotropic actions can engage oxidative stress, neuroinflammation, and circuit dysfunction. In practice, the blood-brain barrier (BBB) restricts most native phytochemicals through tight-junction selectivity, rapid metabolism, low solubility, and transporter-mediated efflux. Key gaps include poor standardization of exposure metrics, limited human relevant BBB models, and few head-to-head studies that compare delivery platforms on the same payload and outcome. This review tackles the mismatch between mechanistic promise and reliable brain exposure that stalls translation. The objectives are to link phytochemical liabilities to enabling strategies in nanomedicine, alternative routes, and transporter-targeted prodrugs, and to propose decision-grade endpoints for translation. We synthesize evidence on BBB transport logic, nanocarrier families, targeting ligands, intranasal delivery, focused ultrasound mediated opening, and prodrug approaches that hijack influx transporters, while foregrounding safety and chemistry, manufacturing, and controls (CMC) constraints. Here we highlight that effective neurotherapeutics emerge when chemistry, carrier, route, and measurement are co designed rather than optimized in isolation. This framework can guide platform selection, de-risk first in-human studies, and sharpen trial endpoints. More broadly, it offers a transferable playbook for barrier-limited drug development across neurology, psychiatry, and oncology.

**Keywords:** central nervous system diseases (CNS); blood-brain barrier (BBB); drug delivery systems; nanomedicine; phytochemicals; nanoparticles; prodrugs; drug administration; intranasal; ultrasonic therapy; transferrin receptor

## 1. Introduction

### 1.1. Clinical Burden and Therapeutic Gap

Central nervous system (CNS) disorders such as depression, dementia, and chronic pain remain among the leading causes of global morbidity, disability, and economic burden [1,2]. Despite decades of intensive research, therapeutic outcomes remain unsatisfactory, with high relapse rates in major depressive disorder, limited disease-modifying options for dementia, and inadequate pain control across populations [2,3]. These shortcomings are amplified by the fact that most CNS-active drugs show poor penetration across the blood–brain barrier (BBB), resulting in suboptimal central exposure and attenuated efficacy [1,4]. Even when new compounds demonstrate preclinical promise, attrition rates during clinical translation remain staggering, with failure rates in neuropsychiatric drug development exceeding those in nearly all other therapeutic domains [5,6]. The net result is a widening therapeutic gap that leaves millions of patients reliant on outdated, partially effective, or poorly tolerated interventions [2,5–7].

This persistent impasse has renewed attention toward alternative sources of therapeutic innovation. Plant-derived molecules, particularly those rooted in neuroactive amino acid metabolism such as tryptophan, offer a compelling avenue [8–11]. These compounds are celebrated for their structural diversity, multitarget activity, and evolutionary compatibility with human physiology, making them attractive candidates for modulating complex CNS pathologies [8,10,12]. Yet, enthusiasm is tempered by major barriers. Many phytochemicals exhibit low bioavailability, poor stability, and unpredictable BBB permeability, which compromise their therapeutic impact [8,12–14]. Recent advances in nanotechnology, ranging from functionalized nanoparticles to receptor-assisted carriers, seek to overcome these pharmacokinetic and delivery hurdles, but their clinical translation is still in its infancy [8,12–14]. Against this backdrop, revisiting plant-derived tryptophan and its metabolic derivatives provides a unique opportunity to bridge neurobiology and psychiatry, while also testing the integration of phytochemistry with advanced delivery platforms to transform depression management [8–10,13,15].

### 1.2. Blood-Brain Barrier (BBB) as a Bottleneck for Phytochemicals

The BBB stands as the central checkpoint governing molecular access to the brain, designed to maintain homeostasis while excluding xenobiotics and potential toxins [12,16]. Its architecture is highly specialized: endothelial cells form continuous tight junctions that restrict paracellular flux, while efflux transporters such as P-glycoprotein (P-gp), breast cancer resistance protein, and multidrug resistance-associated proteins actively pump out diverse substrates [16]. This dual protection ensures neural integrity but also severely limits drug delivery [17]. For therapeutic compounds to penetrate effectively, they must navigate a gauntlet of physicochemical constraints. Molecules with poor aqueous solubility, inappropriate lipophilicity, or rapid metabolic breakdown are particularly disadvantaged, leading to negligible CNS exposure despite robust systemic availability [17,18].

Phytochemicals exemplify this paradox. Polyphenols such as resveratrol and curcumin exhibit potent antioxidant and neuroprotective activities *in vitro*, yet their hydrophilicity and metabolic instability limit their brain levels to trace amounts [12,14,19–24]. Alkaloids, though often more lipophilic, encounter substantial efflux clearance, which nullifies their apparent permeability advantage [12]. Terpenoids and cannabinoids, despite their lipophilic structures, which favor passive diffusion, are hindered by rapid first-pass metabolism and limited bioavailability, yielding inconsistent central effects [14,19,25]. Studies consistently report that only a minority of native phytochemicals achieve detectable brain penetration, and even fewer reach concentrations required for therapeutic modulation of neurotransmission or neuroinflammation [12,26,27]. The recurring outcome is a stark disconnect between preclinical promise and clinical translation [19,28,29]. These limitations underscore the need for innovative delivery strategies that can re-engineer phytochemicals to evade efflux, improve stability, and optimize solubility [12,14,25,28]. Without such

advances, the native molecular forms of these plant-derived agents remain ill-suited for reliable CNS targeting and fall short of their therapeutic potential in depression and related disorders [30,31].

### 1.3. Scope and Organizing Framework

This review is not a catalog of every BBB nanotechnology reported to date. Instead, we use an organizing framework that starts with payload liabilities, such as poor stability, rapid clearance, limited permeability, or off-target exposure, and then maps these constraints onto enabling strategies, from ligand-targeted carriers and intranasal systems to transporter-leveraging prodrugs and selected physical modulation [32–34]. We then judge platforms by pharmacological endpoints that matter for CNS translation: quantifiable brain exposure, target engagement, and a safety margin compatible with real world dosing [32,35–37].

Accordingly, we largely exclude systemic nanomedicine programs without explicit CNS intent, purely diagnostic nanomaterials, and highly speculative constructs lacking a plausible CMC and regulatory path. With that scope set, the next step is to ground these choices in the biological rules of the barrier itself, because delivery design only works when it respects architecture, transport routes, and disease-driven heterogeneity.

## 2. The Blood–Brain Barrier (BBB): Architecture, Transport, Heterogeneity

### 2.1. Neurovascular Unit Architecture

The neurovascular unit (NVU) forms the structural and functional foundation of the BBB, integrating multiple cellular and extracellular components into a finely tuned system that maintains CNS homeostasis [38–40]. Far from being a passive wall, the NVU is a dynamic interface whose architecture underpins both the protective selectivity of the barrier and its vulnerability in disease states [39,41,42].

At the core of this architecture are brain microvascular endothelial cells, which adopt a highly specialized phenotype distinct from systemic endothelia [39,43]. They exhibit extremely low rates of transcytosis and lack fenestrations, thereby minimizing nonspecific permeability [43]. Their intercellular contacts are enriched with tight junction proteins such as claudins, occludin, and ZO-1, creating an electrically resistant barrier that restricts paracellular diffusion while still permitting finely regulated transport of essential metabolites [44–46]. Adherens and gap junctions provide additional stability and communication, allowing endothelial cells to operate as a synchronized sheet rather than as isolated units [45,47].

Closely apposed to the endothelial layer, pericytes are embedded within the basement membrane and act as guardians of barrier integrity [42,48]. They regulate angiogenesis, modulate permeability, and secrete trophic factors such as angiopoietin I and vitronectin, which sustain endothelial survival and limit inflammatory activation [49]. Astrocytic endfeet ensheath nearly the entire capillary surface, releasing mediators, including vascular endothelial growth factor (VEGF) and glial-derived neurotrophic factor (GDNF), while their aquaporin-4 channels orchestrate water and ion balance, which are crucial for neuronal signaling [38,43,50,51].

The basement membrane itself, composed of extracellular matrix proteins secreted by both endothelial cells and astrocytes, provides not only structural stability but also biochemical cues that regulate cellular behavior and cross-talk within the NVU [39,52]. Central to barrier impermeability are tight and adherens junctions, which act as molecular rivets sealing adjacent endothelial cells [44,45]. These complexes are remarkably plastic, responding to oxidative stress, inflammation, and neurodegenerative insults by loosening or disassembling, thereby amplifying barrier leakiness [44,46,48]. Altogether, the NVU's intricate cellular and extracellular architecture forms the scaffold upon which selective transport processes are built, ensuring both the protection and the metabolic supply of the CNS [38–40].

## 2.2. Transport Pathways and Efflux

The BBB operates under the constant tension of permitting the entry of essential nutrients while simultaneously excluding xenobiotics and potentially harmful agents. This balancing act defines its role as both protector and barrier, a duality that complicates CNS drug development [4,53,54]. Only a limited fraction of compounds traverse the BBB by passive diffusion, largely restricted to small, lipophilic molecules with low molecular weight [4,53,55]. Even for lipophilic drugs, passage is often curtailed by additional regulatory mechanisms that actively limit nonspecific entry [53,56].

To sustain brain metabolism, the BBB relies heavily on carrier-mediated transport [4,55]. Prominent examples include GLUT1 for glucose, LAT1 for large neutral amino acids, and monocarboxylate transporters (MCTs) for lactate and other energy substrates [56,57]. These carriers not only ensure nutrient delivery but also provide entry routes for select phytochemicals, though their activity is sensitive to pathological states and drug interactions [56,57]. For larger molecules, receptor-mediated transcytosis represents a critical pathway, with transferrin and insulin receptors serving as canonical examples [58,59]. Advances in nanomedicine are increasingly focused on exploiting these receptors to deliver therapeutic payloads across the barrier in a controlled manner [53,54,60]. Opposing these influx mechanisms are efflux pumps, the most formidable being P-gp, breast cancer resistance protein (BCRP), and multidrug resistance-associated proteins (MRPs) [53,56]. These ATP-binding cassette (ABC) transporters expel a vast array of xenobiotics and pharmacological compounds, shaping drug distribution within the brain [53,56]. Their expression is dynamic, influenced by neuronal activity, circadian rhythms, aging, and disease states, while interactions with polyphenols and phytochemicals may either inhibit or stimulate efflux, thereby altering drug bioavailability [4]. Ultimately, BBB transport functions as both a sentinel and a bottleneck, safeguarding the CNS but at the same time restricting the therapeutic reach of many promising neuroactive agents [53,54,61] (Table 1, Figure 1).

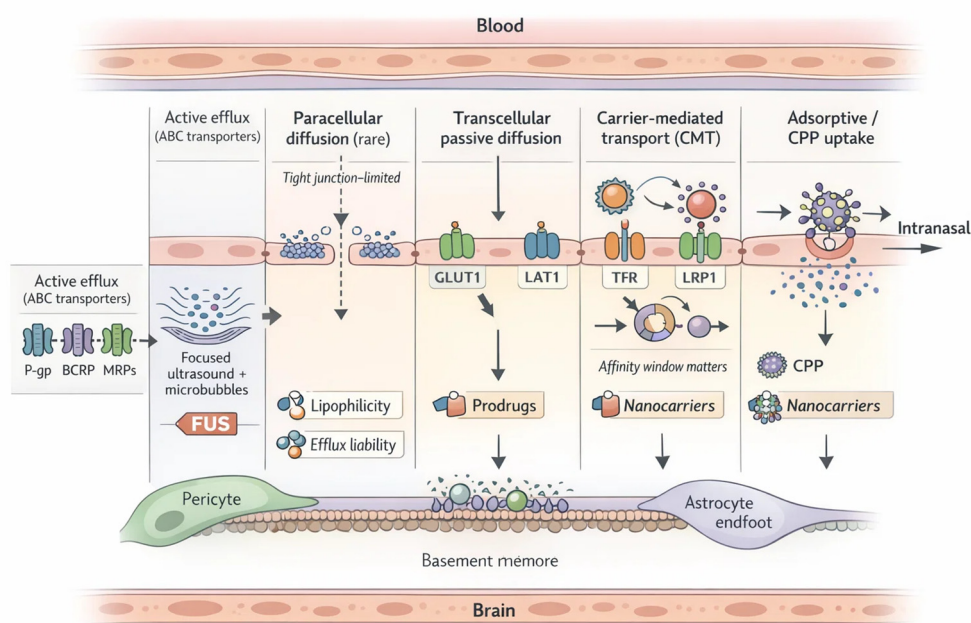
**Table 1.** Blood-brain barrier (BBB) transport machineries and implications for phytochemicals. This table summarizes the dominant BBB transport routes relevant to plant-derived neurotherapeutics and links each route to pragmatic design levers. Pathways are framed as decision levers: passive diffusion is constrained by physicochemical liabilities common in polyphenols; carrier-mediated transport and receptor-mediated transcytosis provide engineered influx opportunities; adsorptive and cell-penetrating peptide strategies can amplify uptake but trade specificity for risk; and efflux pumps (P-gp, BCRP, MRPs) often set the ceiling for unbound brain exposure even when in vitro permeability appears promising.

Pathway	Molecular prerequisites	Exemplars	Impact on phytochemicals	Engineering lever(s)	References
Paracellular diffusion (tight junction limited)	Effectively negligible at an intact BBB; requires transient junction loosening or pathological leak	Small hydrophiles in disease-associated states	Native polyphenols remain largely excluded; leak is disease- and region-dependent and poorly controllable	Localized opening approaches (e.g., focused ultrasound with microbubbles); avoid programs that depend on nonspecific leak	[62–64]
Transcellular passive diffusion	Small size, low polarity, limited H-bonding; favorable	CNS-permeable small molecules; selected alkaloids	Many phytochemicals exceed polarity and H-bonding windows;	Prodrug or soft-drug design; tune logD and polar surface area; stabilize against first-pass	[65–67]

	lipophilicity; minimal efflux liability		metabolism and efflux can negate apparent permeability	metabolism; solubility enabling formulations	
Carrier-mediated transport (CMT)	Structural mimicry of endogenous nutrients; transporter affinity plus adequate chemical stability	GLUT1 (glucose), LAT1 (large neutral amino acids), MCTs (monocarboxylates)	Provides an influx handle for polar phytochemicals, but competition with endogenous substrates and species differences can limit delivery	Transporter-hijacking prodrugs (amino acid, glucose, monocarboxylate promoieties); Km/Vmax-aware design; brain-selective cleavage	[68–70]
Receptor-mediated transcytosis (RMT)	Ligand engagement within a productive affinity window; excessive avidity increases sequestration and lysosomal routing	Transferrin receptor, insulin receptor, LRP1 (targeting designs)	Enables macromolecular and nanoparticle shuttling, but ligand density and valency control release into brain parenchyma	Ligand-decorated nanocarriers; optimize affinity and ligand density; cleavable linkers; designs that favor recycling over degradation	[53,71,72]
Adsorptive-mediated transcytosis and CPP uptake	Net positive charge and/or CPP motifs; electrostatic interactions with endothelial glycocalyx	Tat, penetratin, RVG-derived peptides (as CPP/targeting motifs)	High uptake can trade specificity for off-target accumulation and cytotoxicity; "more cationic" is not always better	Charge-switchable coatings; stimulus-unmasking CPPs; cap surface charge; combine with targeting ligands to improve selectivity	[53,67,71]
Active efflux (ABC transporters)	Substrate recognition by ATP-driven pumps; efflux can	P-gp, BCRP, MRPs	A key barrier for many polyphenols; inhibition or induction can shift CNS	Efflux-evading prodrugs; corona control and stealth coatings; carrier strategies that reduce free substrate	[65–67]

dominate	exposure	at the luminal
even when	unpredictably	membrane; early
passive	across age,	efflux liability
permeability	disease, and	screening
is favorable	comedication	

ABC ATP-binding cassette; BBB, blood–brain barrier; BCRP, breast cancer resistance protein; CMT, carrier-mediated transport; CNS, central nervous system; CPP, cell-penetrating peptide; GLUT1, glucose transporter 1; Km, Michaelis constant; LAT1, L-type amino acid transporter 1; LRP1, low-density lipoprotein receptor–related protein 1; MCTs, monocarboxylate transporters; MRPs, multidrug resistance–associated proteins; P-gp, P-glycoprotein; RVG, rabies virus glycoprotein; RMT, receptor-mediated transcytosis; Vmax, maximum transport rate.



**Figure 1.** Blood-brain barrier (BBB) transport routes and where delivery platforms intervene: passive limits, engineered influx, and efflux ceilings. The schematic summarizes dominant BBB transport routes relevant to phytochemicals and highlights where delivery platforms intervene. At an intact BBB, paracellular diffusion across tight junctions is negligible and disease-dependent; controlled opening is best treated as a localized modulation strategy (e.g., focused ultrasound (FUS) with microbubbles) rather than a baseline assumption. Transcellular passive diffusion depends on size, polarity, hydrogen bonding, and lipophilicity, but apparent permeability is often capped by metabolism and active efflux. ABC transporters—P-gp, BCRP, MRPs—frequently set the ceiling for unbound brain exposure, motivating efflux-evading prodrugs, stealth/corona control for nanocarriers, and early efflux-liability screening. For polar phytochemicals, carrier-mediated transport (CMT) via GLUT1, LAT1, and MCT1 enables engineered influx using nutrient-mimetic promoieties, requiring Km/Vmax-aware design and brain-selective cleavage. For larger cargos, receptor-mediated transcytosis (RMT) via Tfr and LRP1 supports ligand-decorated nanocarriers, but demands an affinity “sweet spot” to avoid endothelial sequestration and lysosomal routing. Adsorptive/CPP uptake can boost entry yet trades specificity for off-target risk; charge-switchable or stimulus-unmasked coatings can mitigate this. Intranasal delivery provides a complementary bypass for suitable payloads. ABC, ATP-binding cassette; BBB, blood–brain barrier; BCRP, breast cancer resistance protein; CMT, carrier-mediated transport; CPP, cell-penetrating peptide; FUS, focused ultrasound; GLUT1, glucose transporter 1; Km, Michaelis constant; LAT1, L-type amino acid transporter 1; LRP1, low-density lipoprotein receptor–related protein 1; MCT1, monocarboxylate transporter 1; MRPs, multidrug resistance–associated proteins; P-gp, P-glycoprotein; RMT, receptor-mediated transcytosis; Tfr, transferrin receptor; Vmax, maximum transport rate.

### 2.3. Disease- and Age-Driven Heterogeneity

The BBB is not a uniform structure but displays striking regional heterogeneity that shapes vulnerability and therapeutic access [73–75]. The hippocampus, for example, exhibits earlier and more pronounced permeability changes compared to the cortex, while the choroid plexus contains fenestrated vasculature that facilitates selective exchange with cerebrospinal fluid [73,75–78]. Such regional differences are further accentuated by pathological states: aging, neurodegeneration, and systemic inflammation remodel barrier integrity, producing spatially distinct patterns of leakage and dysfunction [75,79–82]. These dynamic alterations contribute to selective regional susceptibility in disorders such as Alzheimer’s and Parkinson’s disease [75,79,81,83]. Recognizing and integrating BBB heterogeneity is therefore critical for the rational design of nanomedicine and targeted neurotherapeutics [4,53,84–86].

Ultimately, BBB transport functions as both a sentinel and a bottleneck, safeguarding the CNS but at the same time restricting the therapeutic reach of many promising neuroactive agents [4,53,84–86]. Building on this transport landscape, the subsequent section turns to phytochemicals, profiling their distinct liabilities and highlighting how these molecular features dictate both formulation choices and prodrug design in the pursuit of effective neurotherapeutics [13,14,87,88].

## 3. Phytochemicals as Neurotherapeutics: Classes, Liabilities, MOAs

### 3.1. Polyphenols (*Resveratrol, Quercetin, Curcumin*)

Polyphenols, particularly resveratrol, quercetin, and curcumin, represent the most extensively studied class of neuroprotective phytochemicals, attracting attention due to their pleiotropic activities and broad preclinical support in models of neurodegenerative disease [89–95]. Their neurotherapeutic potential rests on a complex repertoire of mechanisms that extend beyond simple radical scavenging [92,93,95,96]. Resveratrol activates SIRT1 signaling, promoting mitochondrial biogenesis and synaptic resilience, while quercetin modulates AMPK and Nrf2 pathways to counter oxidative stress and restore redox balance [94,96–98]. Curcumin has been shown to suppress NF- $\kappa$ B and Toll-like receptor signaling, thereby dampening neuroinflammatory cascades and protecting neuronal networks [30,65,93,96]. Collectively, these pathways converge to enhance neuronal survival, preserve cognitive function, and mitigate disease-associated cellular stress [89,93–95].

Despite these appealing mechanisms, polyphenols suffer from profound pharmacokinetic limitations [99–102]. All three undergo extensive first-pass metabolism through uridine 5'-diphospho-glucuronosyltransferase (UGT) and sulfotransferase (SULT) pathways, producing conjugated metabolites with limited biological activity [92,101–103]. The predominance of glucuronides and sulfates in circulation sharply reduces the availability of free aglycones that are more pharmacologically active [92,99,101–103]. Moreover, oral bioavailability is poor, with plasma concentrations of parent compounds often remaining below therapeutic thresholds even at high dietary intake [99–101,104]. Such metabolic liabilities have fueled a parallel interest in prodrug approaches and nanoformulations aimed at preserving active moieties for CNS delivery [93,95,100,105].

Even when absorbed, polyphenol penetration into the brain is further constrained by efflux transporters at the BBB [94,95]. P-gp and breast cancer resistance protein (BCRP) actively restrict their accumulation in brain parenchyma, while a “permeability paradox” emerges from the discrepancy between promising *in vitro* BBB transport studies and the much lower exposures seen *in vivo* [32,94,95,106]. This discordance reflects not only transporter activity but also systemic metabolism and protein binding, which together limit CNS bioavailability [95,99–102]. Nevertheless, polyphenols continue to serve as reference scaffolds in neurotherapeutic research, inspiring innovative strategies to overcome BBB constraints while maintaining their broad pharmacodynamic advantages [93–95,100,105].

### 3.2. Alkaloids (*Berberine and Galantamine*)

Alkaloids represent a chemically diverse class of CNS-active molecules with deep roots in both ethnopharmacology and modern clinical medicine [107,108]. Among them, berberine has emerged as a compelling yet pharmacokinetically problematic candidate [109,110]. It interacts strongly with organic cation transporters and is a recognized substrate of P-gp, factors that severely limit its absorption and systemic distribution [109–111]. Berberine undergoes rapid first-pass metabolism, exhibits low oral bioavailability, and suffers from pronounced metabolic instability, resulting in extremely poor CNS exposure despite promising neuroprotective and anti-inflammatory effects demonstrated in cellular and animal models [109,110,112]. These challenges have spurred interest in nanoparticle formulations and prodrug strategies designed to bypass efflux transport and enhance brain uptake [112–117].

By contrast, galantamine provides an example of a plant-derived alkaloid that has successfully transitioned into clinical practice as an approved therapy for Alzheimer's disease [107,108,118,119]. Acting as a selective acetylcholinesterase inhibitor, it improves cholinergic transmission and demonstrates measurable cognitive benefits [108,118,119]. Interestingly, its central activity is not strictly proportional to plasma exposure, as galantamine crosses the BBB primarily through passive diffusion with potential contributions from carrier-mediated processes [65,120–122]. This selective permeability enables therapeutic CNS engagement even at moderate systemic concentrations, underscoring the importance of pharmacodynamic targeting in addition to pharmacokinetics [65,120,121].

Together, berberine and galantamine exemplify the so-called exposure–signal paradox at the BBB, where strong CNS effects can be achieved despite restricted or unpredictable drug penetration [65,120–122].

### 3.3. Terpenoids and Cannabinoids (*CBD/THC, Ginkgolides*)

Lipophilic terpenoids and cannabinoids such as cannabidiol (CBD),  $\Delta^9$ -tetrahydrocannabinol (THC), and ginkgolides display paradoxical behavior at the BBB, where their high hydrophobicity does not consistently translate into effective CNS delivery [123–125]. CBD and THC are both highly lipophilic molecules, yet their brain penetration is actively curtailed by efflux pumps including P-gp and BCRP, which lower their effective concentrations in neural tissue [125–127]. Despite these restrictions, clinical and preclinical evidence demonstrates robust antiepileptic, anxiolytic, and analgesic activity, leading to regulatory approval of CBD for severe childhood epilepsies and THC formulations for spasticity and pain management in multiple sclerosis [128–131]. Their mechanisms are diverse, encompassing CB1 and CB2 receptor modulation, serotonergic signaling through 5-HT1A receptors, and anti-inflammatory as well as antioxidant actions, though their oral bioavailability remains low and interindividual variability in CNS exposure is considerable [127,132–134].

Ginkgolides, diterpenoid lactones derived from *Ginkgo biloba*, present a different profile, achieving moderate penetration into the CNS. Their primary mechanism of action involves antagonism of platelet-activating factor, a pathway linked to neuroinflammation and ischemic injury [135,136]. Preclinical findings suggest neuroprotective and anti-inflammatory potential, yet clinical trials have yielded mixed results, with benefits often modest and outcomes limited by poor BBB permeability and variable bioavailability [135,136]. The discrepancy between mechanistic promise and inconsistent clinical performance reflects the difficulty of translating terpenoid pharmacology into effective CNS therapeutics.

Together, cannabinoids and ginkgolides illustrate the so-called lipophilicity trap, in which excessive hydrophobicity, combined with efflux and metabolic instability, can paradoxically hinder brain delivery rather than facilitate it [123–126]. This paradox underscores the need for nuanced drug design and advanced delivery systems when considering terpenoids as neurotherapeutic candidates [133,134,137].

### 3.4. Formulation-Relevant Liabilities and Structure–Activity Relationship (SAR) Flags

Beyond class-specific mechanisms, the physicochemical properties of phytochemicals largely dictate their ability to cross the BBB and achieve therapeutic relevance [65,138,139]. Optimal penetration is typically observed in compounds with a logD between 1 and 3, balanced pK<sub>a</sub> values that minimize ionization at physiological pH, a hydrogen bond donor count of two or fewer, and a hydrogen bond acceptor count not exceeding five [138,140,141]. Molecular flexibility is equally important, with fewer than ten rotatable bonds generally favoring permeability and sustained CNS exposure [138,142]. These criteria extend Lipinski's Rule of Five into the realm of CNS drug-likeness and provide practical benchmarks for evaluating natural products [138,140,141].

Conversely, several red flags frequently emerge among phytochemicals. Excessive polarity or a topological polar surface area greater than 90 Å<sup>2</sup> strongly predicts poor CNS penetration [138,140,142]. Similarly, a high density of hydrogen bond donors, often in the form of phenolic hydroxyl groups, correlates with both poor permeability and metabolic vulnerability through glucuronidation or sulfation [65,138]. These metabolic soft spots, common in polyphenols and terpenoids, reduce bioavailability and amplify efflux transporter recognition [143–145].

Early recognition of these liabilities through structure–activity relationship analysis and computational screening is therefore essential [138,146,147]. Such insights can guide the rational design of prodrugs and nanoformulations, improving bioavailability and transforming suboptimal scaffolds into viable neurotherapeutic candidates [4,139,148] (Table 2).

**Table 2.** Translational map from phytochemical class to key delivery liabilities and practical enabling strategies for central nervous system (CNS) development across blood-brain barrier (BBB) constrained programs. Phytochemical scaffolds share recurring developability bottlenecks at the BBB, yet the dominant liability differs by class. The table links representative compounds discussed in the manuscript to the most common physicochemical and biopharmaceutical constraints, then pairs each class with a preferred enabling strategy that is compatible with scale-up and safety screening.

Class	Representative compounds	Primary liabilities	Preferred enabling strategy	Notes (e.g., stability, taste, ionization)	References
Polyphenols (flavonoids, stilbenes, curcuminoids)	Resveratrol; quercetin; curcumin	Low exposure in vitro; extensive metabolism (UGT/SULT); efflux liability (P-gp/BCRP); often high polarity or poor solubility; chemical instability (oxidation hydrolysis).	Prodrug transporter-hijacking phase II metabolism nanoencapsulation (polymeric NPs, liposomes, SLNs) to protect the scaffold and modulate release; or BBB adjuncts when justified.	or Phenolic acids are weak acids with context-dependent ionization; many are light and pH sensitive; bitter or astringent taste can limit oral dosing and adherence.	[149–151]
Alkaloids	Berberine; galantamine	Ionization strong	Salt selection plus lipid-based carriers	Often strongly bitter; typical basic	[110,116,149]

		transporter or polymeric pKa yields cationic interactions can micelles; efflux fraction at cap CNS entry; bypass via prodrug physiological pH; variable oral or nanocarrier galantamine bioavailability; shielding; leverage illustrates that efflux driven high target potency pharmacodynamic exposure with lower systemic targeting can partly variability; class exposure through offset limited brain dependent safety controlled release or partitioning. margins and alternative routes. CYP interactions.	
Terpenoids (mono-, sesqui-, diterpenes)	Ginkgolides; pinene; linalool	High lipophilicity with low aqueous solubility; volatility for some monoterpenes; oxidative degradation; high protein binding and rapid metabolism leading to variable CNS exposure.	Self-emulsifying systems, nanoemulsions, cyclodextrin for inclusion, or lipid nanoparticles to raise apparent solubility and stabilize the high protein payload; intranasal formulations for rapid onset when appropriate.
Cannabinoids	Cannabidiol (CBD); THC	Formulation-limited absorption and marked individual variability; extensive hepatic metabolism; drug-drug interactions; psychoactivity and regulatory constraints for THC; long tissue	Lipid vehicles, nanoemulsions, polymeric carriers, or controlled release depots; route optimization (oromucosal, intranasal) and dose fractionation to reduce peak related adverse effects while maintaining exposure. Light and oxygen sensitive; taste can be limiting; largely neutral but highly lipophilic; legal status and labeling requirements can shape trial design. [149,151,152] [153–155]

---

residence due to  
lipophilicity.

---

BBB, blood–brain barrier; BCRP, breast cancer resistance protein; CBD, cannabidiol; CNS, central nervous system; CYP, cytochrome P450; NPs, nanoparticles; pKa, acid dissociation constant; P-gp, P-glycoprotein; SLNs, solid lipid nanoparticles; SULT, sulfotransferases; THC,  $\Delta^9$ -tetrahydrocannabinol; UGT, UDP-glucuronosyltransferases.

## 4. Nanomedicine Platforms for Blood-Brain Barrier (BBB) Delivery

### 4.1. Polymeric Nanoparticles (PLGA, PEG-PLGA, and Chitosan)

Polymeric nanoparticles have emerged as highly versatile carriers for CNS delivery, with poly(lactic-co-glycolic acid) (PLGA) considered the gold standard due to its biocompatibility, biodegradability, and track record of clinical translation [156–158]. The incorporation of polyethylene glycol (PEG) into PLGA scaffolds provides “stealth” properties by shielding the carrier from opsonization and prolonging its circulation half-life, thereby substantially enhancing the probability of crossing the BBB [159–161]. Beyond their pharmacokinetic advantages, PLGA and PEG-PLGA matrices are particularly well suited for encapsulating hydrophobic phytochemicals, thereby improving aqueous solubility and enabling sustained release profiles that minimize burst effects while maintaining therapeutic concentrations within neural tissue [157,160,162]. Several studies have highlighted that surface modifications, ranging from peptide ligands such as Angiopep-2 to functional protein corona interactions, can further refine nanoparticle selectivity for BBB transport and neuronal uptake [33,163–165].

Chitosan-based systems represent a complementary and increasingly significant strategy, exploiting their intrinsic cationic nature and mucoadhesive capacity [159,166]. When used either as a surface coating or as a hybrid scaffold with PLGA, chitosan enables tight interaction with mucosal surfaces and facilitates paracellular transport [159,166,167]. This property is particularly advantageous for intranasal administration, as demonstrated by formulations where PLGA nanoparticles embedded within chitosan microparticles achieved enhanced uptake across the olfactory mucosa and direct delivery to the brain [159,167,168]. Intranasal chitosan–PLGA carriers have been successfully applied to deliver repurposed chemotherapeutics such as gemcitabine for glioblastoma therapy, achieving tumor-selective release while bypassing systemic clearance [158,159,165].

Quantitative pharmacokinetic assessments underscore the translational potential of these approaches [156,161,167]. Enrichment analyses of brain-to-plasma distribution indicate that optimized PLGA- and chitosan-based delivery systems can increase cerebral accumulation by several fold relative to unformulated compounds. These findings support the use of polymeric nanomedicine not merely as a means to enhance phytochemical bioavailability, but as a deliberate strategy to harness transport mechanisms for targeted CNS therapy [157,160,161,167].

### 4.2. Lipid Carriers (Liposomes, Solid Lipid Nanoparticles (SLNs), Nanoemulsions)

Lipid-based nanocarriers have become central to brain-targeted delivery, with liposomes representing the archetypal bilayer system [169,170]. Their amphiphilic structure allows for simultaneous encapsulation of hydrophilic and hydrophobic compounds, while PEGylated liposomes confer stealth properties that prolong circulation time and enhance BBB penetration [169,171,172]. This versatility has been leveraged in multiple preclinical models, where PEGylated formulations not only improved stability but also demonstrated controlled biodistribution within brain parenchyma [169,171,173]. The bilayer’s modularity also facilitates functionalization with targeting ligands, thereby adding a level of precision that polymeric systems often struggle to replicate [169,173].

Solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs) have advanced the field by addressing limitations of traditional liposomes, particularly with respect to stability and drug-loading capacity [155,169,174]. SLNs, composed of solid lipids at body temperature, offer biocompatibility and controlled release but are often restricted by lower payload efficiency [155,175]. In contrast, NLCs incorporate both solid and liquid lipids into their matrix, thereby providing greater drug accommodation and reducing the risk of expulsion during storage [151,155,176]. Comparative studies consistently highlight the superior stability and performance of NLCs over SLNs, particularly for long-term formulations aimed at chronic neurodegenerative diseases [151,176,177].

Nanoemulsions extend the potential of lipid systems by enabling rapid and direct intranasal delivery to the brain [176,178]. Their small droplet size promotes fast absorption through the olfactory epithelium, a pathway particularly attractive for bypassing systemic metabolism [176,178,179]. However, reproducibility in manufacturing, along with challenges in preventing aggregation and ensuring shelf-life stability, remains an unresolved hurdle [178,180,181]. As recent *in vivo* studies emphasize, the promise of nanoemulsions lies in their high uptake efficiency, yet their clinical translation will depend on improved standardization and stabilization strategies [176,178,181].

#### 4.3. Dendrimers and Micelles

Dendrimers represent one of the most structurally sophisticated nanocarriers for CNS therapy [182,183]. Their branched, tree-like architecture provides internal cavities for drug encapsulation and a multivalent surface for ligand attachment, making them particularly effective in tuning brain-specific targeting [182–184] critical challenge, however, lies in balancing ligand density: while higher densities can enhance receptor-mediated transcytosis, excessive functionalization often increases steric hindrance or cytotoxicity [182,184,185]. Studies with polyamidoamine (PAMAM) and carbosilane dendrimers demonstrate that size and surface charge strongly dictate BBB penetration, with mixed-surface or PEGylated variants offering improved biocompatibility and reduced clearance [186–189]. Such design nuances highlight the delicate trade-off between maximizing efficacy and minimizing off-target toxicity *in vivo* [182,184,187].

Micelles, by contrast, exploit their amphiphilic organization to solubilize poorly water-soluble compounds, a feature particularly advantageous for phytochemicals and nucleic acids [190,191]. Their self-assembly into nanosized aggregates enables responsiveness to physiological triggers, such as acidic pH or enzyme activity, resulting in controlled drug release within the brain microenvironment [192–194]. Optimization of micelle size and zeta potential has been shown to extend circulation while facilitating BBB passage without rapid renal clearance [186,190,192]. Recent developments in cation-free micelles for siRNA delivery illustrate how fine-tuned charge control can reduce cytotoxicity while preserving efficient uptake in glioblastoma models [190,195]. Together, dendrimers and micelles provide complementary strategies: dendrimers excel at multivalent, targeted interactions, while micelles offer dynamic, responsive platforms for solubilization and triggered release [186,191].

#### 4.4. Inorganic/Carbon Nanostructures

Carbon- and inorganic-based nanostructures have carved a distinct niche in BBB research due to their dual roles as carriers and imaging agents [196–198]. Carbon dots and quantum dots, for example, exhibit intrinsic fluorescence, enabling simultaneous drug delivery and real-time tracking [196,199–201]. This built-in diagnostic capacity underpins their promise for theranostic applications, particularly in neurodegenerative disease models where monitoring biodistribution is critical [196,202–204]. Magnetic nanoparticles extend this paradigm further by enabling magnetically guided delivery, offering external control over accumulation in targeted brain regions [198,203,205]. Such approaches not only enhance precision but also reduce systemic exposure, positioning these nanostructures as powerful candidates for next-generation CNS therapeutics [197,198,203].

Despite these advantages, their translation faces significant barriers [199,206]. The long-term biocompatibility of quantum dots and carbon nanodots remains uncertain, with concerns over

oxidative stress, protein corona formation, and potential accumulation in neural tissues [196,199,206,207]. Magnetic nanoparticles, while effective in guidance and imaging, also raise questions about clearance and toxicity with repeated use [198,203,205,206]. Animal studies have demonstrated promising biodistribution patterns, yet discrepancies in chronic safety outcomes highlight the need for rigorous toxicological evaluation before clinical adoption [199,206,208,209]. In this context, the field is actively exploring polymer-coated and functionalized variants to mitigate oxidative damage while preserving the diagnostic and therapeutic potential [198,203,205,206]. Together, inorganic and carbon nanostructures embody both the allure and caution of theranostic nanomedicine: they provide unparalleled control and visibility, but demand equally careful assessment of their long-term biological footprint [197,199,206].

#### 4.5. Hybrid/Biodegradable and Protein Corona Control

Hybrid nanomedicine platforms are increasingly recognized as promising strategies for crossing the BBB because they combine complementary features of polymeric, lipid, and inorganic scaffolds [67,210,211]. Polymeric–lipid hybrids, for example, integrate the structural stability of polymers with the biocompatibility and drug-loading flexibility of lipid layers, while inorganic–organic hybrids offer imaging capabilities alongside controlled drug release [67,212,213]. A recurring theme in these designs is the fine-tuning of surface charge: adjusting zeta potential can minimize opsonization and prolong circulation, yet excessive neutralization may compromise cellular uptake [67,149,214]. Dual-targeting systems, such as polyanionic polymalic acid nanodrugs conjugated with Angiopep-2, illustrate how surface chemistry can be leveraged to maintain stability while enabling efficient receptor-mediated transport into the brain [214–216].

A critical determinant of *in vivo* performance lies in protein corona formation, which reshapes nanoparticle identity immediately upon systemic entry [217]. Far from being an inert byproduct, the corona can hinder transcytosis, reduce tumor selectivity, or alternatively, be engineered to guide delivery [217]. Strategies to regulate this interface include pre-coating with tunable surfactants, exploiting biomimetic exosome-mimetic shells, or even deliberately co-opting serum proteins to enhance stealth [217]. Such approaches demonstrate that corona engineering is not merely defensive but can be actively harnessed to improve BBB passage and targeting precision.

At the same time, clinical translation of these platforms hinges on biodegradability and safety [67,149,210]. Polyanhydride-based carriers and bioinspired protein–polymer nanocapsules exemplify progress toward fully degradable designs, yet challenges remain regarding reproducibility, long-term safety, and scale-up [67,149]. As recent reviews emphasize, the success of hybrid systems will depend not only on their multifunctional design but also on overcoming regulatory hurdles by proving that corona control and biodegradability can coexist without compromising efficacy [67,210,211].

#### 4.6. Literature Snapshot

Comparative evaluations of nanomedicine platforms highlight how material choice shapes brain delivery outcomes [61,218,219]. Head-to-head studies reveal that PLGA nanoparticles carrying flavonoids often outperform liposomes in terms of controlled release and systemic stability, while liposomes enable faster brain penetration but can be prone to leakage and reduced retention [220–222]. Similarly, dendrimers and micelles demonstrate complementary advantages when loaded with peptide cargos: dendrimers benefit from multivalency that enhances receptor-mediated uptake, whereas micelles provide stimulus-responsive release and better solubilization of hydrophobic payloads [223–225]. Endpoints such as BBB permeability ratios, neurobehavioral outcomes in stroke or neurodegeneration models, and toxicity remain the unifying benchmarks across these comparisons [61,218,219].

The collective evidence makes one principle clear: there is no universal best nanomedicine platform [218,219,222]. The optimal choice is highly context-driven, defined by the therapeutic payload, disease model, and the balance between efficacy and safety [61,218,219]. Surface chemistry

and biological targeting often make or break brain delivery—hence a focused look at ligands, valency, and stimuli [221,223,224] (Table 3).

**Table 3.** Nanocarrier platforms for blood-brain barrier (BBB)-constrained phytochemicals: design levers, translational performance, and CMC/GMP-critical considerations. This table offers a decision-oriented snapshot of major nanocarrier platforms for brain delivery, linking each to core/shell materials, surface strategies (PEG, receptor ligands, corona control), loading modes, and release logic. Release is categorized as constitutive (diffusion/erosion) or stimulus-enabled (pH/redox/enzymes; external triggers such as magnetic fields or focused ultrasound). Key trade-offs include leakage versus retention (liposomes), payload expulsion (SLNs), reproducibility/stability limits (nanoemulsions), multivalency versus toxicity (dendrimers/ CPP-like surfaces), long-term safety uncertainty (inorganic systems), and batch heterogeneity/regulatory issues (exosomes). CMC/GMP notes emphasize QbD priorities: size/PDI, zeta potential, encapsulation efficiency, release kinetics, sterility/endotoxin, and scalable manufacture.

Platform	Core/shell materials	Size/PDI	Surface (PEG/ligand)	Loading	Release trigger	Pros/Cons	Example payload	Notes (CMC/GMP)	References
PolymERIC nanoparticles (PLGA, PEG-PLGA; chitosan-PLGA; chitosan-n-coated/hybrid/s)	PLGA or PEG-PLGA matrix; optional chitosan coating or chitosan microparticle embedding	Tunable; low PDI targeted as a core CQA	PEG “stealth”; Angio pep-2 and other RMT ligands; corona-Engineering approaches	Encapsulation of hydrophobic phytochemicals; co-loading feasible; protected from metabolism	Diffusion + polymer erosion; can add pH/redox/enzyme-responsive elements	Pros: biodegradable, sustained release, strong stability. Cons: MPS uptake; burst-release risk if not tuned; process sensitivity	Resveratrol; curcumin; flavonoids; intranasal chitosan-PLGA example: gemcitabine	QbD/QT (size, PDI, zeta potential, encapsulation efficiency, release kinetics); sterility/endotoxin control; scale-up reproducibility	[61,20,226]
Liposomes (including phospholipid bilayer ± control)	Phospholipid bilayer	Tunable; control	PEGylation for	Hydrophilic cargo in hydrophobic	Constitutive leakage	Pros: versatile	Resveratrol; curcumin	Composition and lipid	[221,222,227]

ing	cholesterol; aqueous core + hydrophobic bilayer domain	needed to limit leakage and maintain uniformity	circulation; modular and functionalization for BBB targeting	bilayer; encapsulation possible	co-/partitioning; can be engineered for thermosensitive or pH-triggered release	loading; rapid brain access in some models. Cons: leakage and reduced retention; stability/shelf-life constraints	in; peptide cargos (as discussed in platform comparisons)	raw-material controls; filtration - compatibility sterility where feasible; stability/lyophilization program s to preserve CQAs	
Solid lipid nanoparticles (SLNs) and nanostructured lipid carriers (NLCs)	SLN: solid lipid matrix. NLC: mixed solid lipids + drug accommodation	Typically robust colloids when optimized; low PDI targeted as a translation enabler	Surface-stabilized; PEG and ligands can be incorporated as needed	Best for lipophilic phytochemicals; NLCs improve loading vs SLNs	Diffusion from lipid matrix; reorganization influences kinetics	Pros: biocompatible, controlled release, improved stability. Cons: SLNs limited loading; storage driven drug	Polyphenols; terpenoids; chronic neurodegeneration-oriented formulations	Control lipid polymorphism, surfactant system, and storage conditions; monitor size/PDI, encapsulation efficiency, leakage/expulsion during	[228 – 230]

							expuls ion risk reduce d in NLCs	stability testing		
Nanoemulsions / nanoe mulgels (often intranasal)	Oil-in-water droplets with surfactant s/co-surfactant s; optional mucoadhesive gel phase	Small droplet size emphasized for uptake; distribution width is a key CQA	Mucosal polymers (e.g., chitosan coatings)	Solubilization of hydrophobic polyphenols; formulation-driven payload stabilization	of and	Rapid absorption via olfactory epithelium; residence-time extension via in situ gels (thermo/ion-responsive) and aggregation/helf-life challenges	Pros: fast nose-to-brain access; bypass first-pass metabolism. Cons: manufacturing under lipid vehicle stability and aggregation/helf-life challenges	Curcumin; resveratrol; terpenoids; cannabinoids; class examples referenced under lipid vehicle stability and aggregation/helf-life challenges	CMC focuses on droplet size distribution, rheology (for sprays/plume/metered dosing, preservative compatibility, and long-term stability)	[227, 231, 232]
Dendrimers (e.g., PAMAM, carbosilane)	Monodisperse branched polymers; generation-defined architecture; optional PEGylation/mixed surfaces	Defined by generation; surface charge is a dominant performance/safety dial	Multivalent ligand attachment (RMT or disease-homing); PEG/mixed	Internal encapsulation and/or conjugation; suitable for peptides and nucleic acids	cavity and covalent for and	Cleavable linkers (pH/redox/enzyme) for conjugation; diffusivity for encapsulation; small	Pros: programmable multivalency and targeting density. Cons: cationic	Peptide cargos; nucleic acids (siRNA-class examples)	Tight control of generation, residual monomers/solvents, surface substitution ratio; sterility/	[223 – 225]

			-		molecul	c		endotoxi	
			surfac		es	toxicit		n;	
			e			y risk		charge-	
			design			and		related	
			s to			cleara		immuno	
			reduc			nce		toxicity	
			e			trade-		screenin	
			toxicit			offs		g	
			y			with			
						high			
						functio			
						nalizat			
						ion			
Polym	Amphiphi	Nanosi	PEG	Solubilizes poorly	Physiol	Pros:	Hydro	Define	[220,
eric	lic block	zed	coron	water-soluble	ogic	strong	phobic	CMC/C	224,
micelle	copolymer	aggreg	a	phytochemicals;	triggers	solubil	phytoc	QA set	225]
s	s that self-	ates;	comm	can carry nucleic	(acidic	ization	hemica	includin	
	assemble	stabilit	on;	acids with tailored	pH,	;	ls;	g	
	(hydropho	y to	charg	chemistry	enzyme	trigger	siRNA	size/PDI	
	bic core +	dilutio	e		s) that	able	deliver	, critical	
	hydrophili	n is a	tuning		destabil	release	y	micelle	
	c corona)	critical	(inclu		ize	. Cons:	exampl	concentr	
		attribut	ding		micelles	disass	es	ation,	
		e	cation		and	embly	discuss	loading,	
			-free		release	risk in	ed in	and	
			design		cargo	vivo;	gliobla	release;	
			s) to			formul	stoma	assess	
			reduc			ation	context	serum	
			e			sensiti		stability	
			cytoto			vity to		and	
			xicity			concen		storage	
						tration		(lyophil	
						and		ization)	
						serum		effects	
						interac			
						tions			
Inorga	Carbon	Engine	Funci	Conjugation/adsor	Externa	Pros:	Theran	Extende	[218,
nic/car	dots or	ered	onal	ption of small	l	trackin	ostic	d	219,
bon	quantum-	nanosc	coatin	molecules;	triggers	g +	small	toxicolo	226]
nanostr	dot-like	ale;	gs	intrinsic imaging	(magne	deliver	molecu	gy and	
uctures	cores;	surface	(poly	signal enables	tic y;	les;	clearanc		
(carbon	magnetic	chemis	mer/P	theranostics	fields/li	control	imagin	e	
dots/q	nanopartic	try	EG)		ght)	lable	g-	characte	

uantu	les;	domin	and		and	guidan	enable	rization;	
m dots;	polymer/P	ates	ligand		surface	ce.	d CNS	strict	
magnet	EG	biodist	s for		chemist	Cons:	deliver	control	
ic	coatings	ributio	targeti		ry-	long-	y	of	
cores)	for	n and	ng;		depend	term	constru	surface	
	biocompat	clearan	magn		ent	safety/	cts	chemistr	
	ibility	ce	etic		release	retenti		y and	
			guida			on		impuriti	
			nce as			uncert		es;	
			an			ainty;		justificat	
			extern			oxidati		ion of	
			al			on and		long-	
			contro			accum		term	
			l layer			ulation		tissue	
						concer		persiste	
						ns		nce risk	
Hybrid	Polymer-	Tunabl	Dual-	Multi-cargo	Layer	Pros:	Angiop	PAT-	[218,
/biomi	lipid	e;	targeti	capability;	can	integra	ep-2-	enabled	233,
metic	hybrids;	multi-	ng	integrate	prodrug	tes	conjug	monitori	234]
system	inorganic-	compo	(e.g.,	chemistry	with	stabilit	ated	ng; tight	
s and	organic	nent	Angio	carrier features		y,	nanodr	incomin	
corona-	hybrids;	system	pep-2			targeti	ug	g	
control	membrane	s	+			ng,	exampl	material	
led	-coated	amplif	CPP/T			(pH/red	and	specs;	
design	nanopartic	y	AT);			ox/enzy	trigger	dual-	control
s	les;	sensiti	coron			me) or	able	ligand	strategy
	polymalic	vity to	a			external	release	lipid	for
	acid	process	engin			(ultraso	. Cons:	nanoca	surface
	nanodrug	drift	eering			und/lig	CMC	rriers	function
	s		via			ht)	compl	in	alization
			pre-			triggers	exity;	gliobla	and
			coatin				regula	stoma	corona;
			g or				tory	models	batch-to-
			biomi				ambig		batch
			metic				uity		fidelity
			shells				for		prioritiz
							multif		ed
							unctio		
							nal		
							hybrid		
							s		

Biogenic vesicles (mammalian exosomes; plant-derived EVs; synthetic d EVs; synthetic mimetic vesicles)	Natural EV bilayers (mammalian plant-derived or synthetic mimetics (polymers/membrane-coated NPs)	Often heterogenous; purity and population definition are key	Innate tropism plus optimal ligand decoration; mimetics allow controlled ligand presentation	Electroporation/incubation/extrusion of small molecules, nucleic acids; PDEVs may carry intrinsic bioactives	Cellular uptake/ endosomal trafficking; release depends on uptake and intracellular routing	Pros: high biocompatibility; potent initial tropism; PDEVs are abundant and low immunogenicity; BBB mechanisms for PDEVs less defined	Small molecules, proteins, nucleic acids; antioxidant/anti-inflammatory cargos in neuroinflammation models	Standardization, isolation, potency assays; define acceptable heterogeneity; scale-up and regulatory classification; BBB mechanisms for PDEVs less defined	[219, 225, 235]
--	---	--	--	--	---	---	--	--	-----------------

BBB, blood–brain barrier; CMC, chemistry, manufacturing, and controls; CNS, central nervous system; CPP, cell-penetrating peptide; CQA, critical quality attribute; EVs, extracellular vesicles; NLCs, nanostructured lipid carriers; NPs, nanoparticles; PAMAM, poly(amidoamine); PAT, process analytical technology; PDEVs, plant-derived extracellular vesicles; PEG, polyethylene glycol; PLGA, poly(lactic-co-glycolic acid); QbD, quality by design; QTPP, quality target product profile; RMT, receptor-mediated transcytosis; siRNA, small interfering RNA; SLNs, solid lipid nanoparticles; TAT, trans-activator of transcription.

## 5. Targeting and Stimuli Strategies

### 5.1. Receptor-Mediated Transcytosis (RMT) Ligands

Receptor-mediated transcytosis (RMT) has emerged as the cornerstone of BBB targeting, providing a rational route for therapeutic entry into the CNS [72]. Among the most extensively characterized receptors, the transferrin receptor (TfR), low-density lipoprotein receptor-related protein 1 (LRP1), insulin receptor (IR), and LDL receptor (LDLR) have each been exploited to shuttle biologics, peptides, and nanocarriers across endothelial cells [58,236]. Ligands such as Angiopep-2, apolipoprotein E (ApoE)-mimetics, and engineered transferrin derivatives illustrate how endogenous trafficking machinery can be co-opted without major structural perturbations to the BBB [58,236]. This strategy has been validated in multiple preclinical models and increasingly in human-relevant systems, including iPSC-derived BBB platforms that closely recapitulate receptor dynamics [237,238].

The success of RMT hinges on a finely tuned balance between ligand affinity and avidity [72,239]. Excessively strong binding may lead to receptor saturation or sequestration in lysosomes, while weak interactions risk premature dissociation and suboptimal delivery [70,239]. Mathematical modeling and linker engineering have refined our understanding of these trade-offs, highlighting an “affinity window” that permits recycling and productive transcytosis [239,240]. Yet, competition with endogenous ligands imposes a physiological ceiling effect, particularly for receptors like TfR and IR that are critical for nutrient homeostasis [240,241]. This ceiling necessitates dosing strategies and ligand modifications that preserve BBB transport without displacing natural substrates.

Translational progress has been most visible in the diversification of ligand classes. Antibody fragments and bispecific constructs with optimized linkers now outperform earlier monoclonals in preclinical BBB assays [32,242,243]. In parallel, short peptides, nanobodies, and aptamers provide modular alternatives that reduce immunogenicity while retaining high receptor specificity [70,244,245]. Collectively, these innovations illustrate how the conceptual framework of RMT is being transformed into clinically viable neurotherapeutic strategies, setting the stage for next-generation delivery systems tailored to the diseased brain microenvironment.

### 5.2. Adsorptive and Cell-Penetrating Peptide (CPP) Strategies

Adsorptive-mediated transcytosis and cell-penetrating peptides (CPPs) represent alternative strategies to receptor-based delivery for crossing the BBB [71,246,247]. Their principle rests on cationic surfaces that engage in electrostatic interactions with the negatively charged glycocalyx of endothelial cells, thereby initiating uptake [246,248]. Classic CPPs such as Tat, penetratin, and rabies virus glycoprotein (RVG) have been widely studied and consistently demonstrate high internalization efficiency across a range of in vitro and in vivo models [247,249,250]. This strong uptake capacity has made them attractive tools for brain-directed delivery of proteins, nucleic acids, and nanocarriers [246,247,250]. However, the very same non-specificity that enables broad penetration also increases the risk of cytotoxicity and off-target accumulation in peripheral tissues, posing major challenges for clinical translation [251–253].

To overcome these limitations, newer designs exploit reversible or conditional activation of cationic charges [254,255]. Charge-switchable coatings that remain neutral in circulation but expose CPP activity in acidic or enzymatically active microenvironments offer a way to enhance specificity while reducing systemic toxicity [254,255]. Stimulus-responsive CPPs integrated into nanogels or liposomes can be selectively unveiled in glioma or inflamed brain regions, thereby combining the high uptake efficiency of adsorptive strategies with a more targeted therapeutic profile [254,256,257]. These innovations suggest that adsorptive and CPP approaches, once criticized for their lack of selectivity, may reemerge as valuable complements to receptor-mediated systems when combined with smart design principles [247,251,254].

### 5.3. Multivalent/Dual-Targeting Designs

Multivalent and dual-targeting strategies build on the idea that no single ligand can fully capture the complexity of BBB transport and disease-specific recognition [258–260]. By combining receptor-mediated transcytosis ligands with disease-associated epitopes, such as amyloid-binding motifs in Alzheimer's disease or glioma-homing peptides, researchers aim to achieve both efficient barrier penetration and selective accumulation in pathological tissue [241,259,261]. This layered approach provides synergy, since one ligand optimizes endothelial uptake while the second guides delivery toward neuronal or tumor targets [261–263]. The result is not simply additive transport but enhanced fidelity of targeting, often translating into improved therapeutic outcomes in preclinical models [261,262,264].

Nonetheless, designing dual-decorated nanocarriers requires careful calibration [260,264,265]. Steric hindrance between ligands, suboptimal linker lengths, or excessive surface density can compromise binding efficiency and even trigger accelerated clearance [264,265]. Several studies illustrate both promise and pitfalls [259,261,262]. In Alzheimer's disease models, nanoparticles bearing transferrin and neuron-targeting Tet1 peptides showed superior cognitive rescue compared with single-ligand systems [259,261]. In glioblastoma, lipid nanocarriers co-functionalized with Angiopep-2 and TAT achieved deeper tumor penetration and survival benefits [259,261]. These examples highlight how multivalent strategies, when optimized, can balance BBB entry with precision delivery, positioning them as one of the most forward-looking directions in neurotherapeutics [32] [32,70,258].

### 5.4. Stimuli-Responsive Systems

Stimuli-responsive systems harness both endogenous and exogenous cues to achieve precise control over drug delivery across the BBB [33,266,267]. Internal triggers such as acidic pH gradients, redox imbalances, and overexpressed enzymes in the tumor microenvironment have been successfully integrated into nanocarriers to enable controlled and site-specific release [266,268,269]. pH-sensitive polymers, disulfide-cleavable linkers, and enzyme-activated coatings exemplify this strategy, ensuring that therapeutic cargo remains stable in circulation yet becomes rapidly available once inside diseased brain regions [186,269,270]. These approaches not only enhance local efficacy but also reduce systemic exposure, thereby addressing one of the central challenges of neurotherapeutics [220,267].

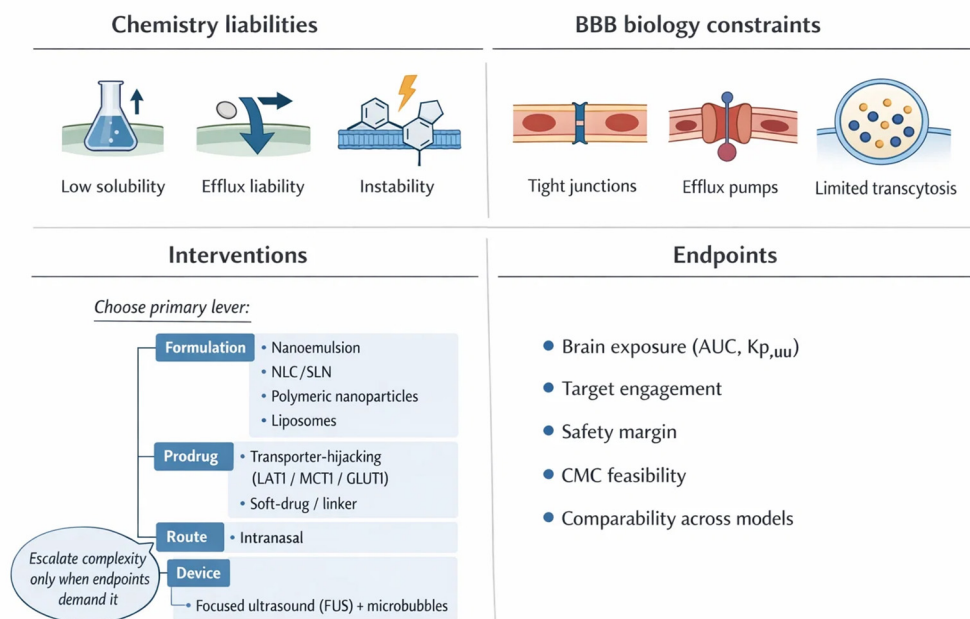
External stimuli offer an additional dimension of spatiotemporal precision [220,267,268]. Magnetic fields, focused ultrasound, and light-based activation provide reversible and non-invasive triggers that can be synchronized with drug administration [267,271,272]. Such methods have been paired with polymeric and lipid nanocarriers to achieve on-demand release and deep penetration into glioblastoma tissue [4,267,272]. However, questions of safety, reproducibility, and clinical feasibility remain unresolved, particularly for modalities requiring specialized equipment or prolonged exposure [220,273,274]. Balancing innovation with practicality is crucial as these systems move [220] toward translation [220,266,275]. Beyond targeting, alternative routes and device-enabled openings can bypass or transiently relax the barrier [4,54]. (Table 4, Figure 2).

**Table 4.** Receptor-mediated transcytosis (RMT) ligands for BBB delivery: binding/valency design levers, species caveats, and reported brain-exposure deltas. This table summarizes RMT ligand classes by target receptor, binding strength ( $K_d$ ), and valency/ligand density—key determinants of productive transcytosis versus endothelial sequestration and lysosomal routing. Species caveats flag translation pitfalls, including endogenous ligand competition (e.g., transferrin/insulin), regional and disease-dependent receptor expression, and limited rodent–human cross-reactivity.  $\Delta$  brain exposure records fold-changes (brain area under the curve (AUC),  $K_p$ ,brain, or preferably  $K_p$ ,uu) versus matched non-targeted controls under comparable dosing and sampling windows. “NR” indicates values not specified in the current draft and to be completed during final reference curation.

Ligand/target	$K_d$ range	Valency	Species caveats	$\Delta$ brain exposure (fold)	Notes	References
Transferrin (Tf) → TfR	NR in manuscript; productive “sweet spot” emphasized (avoid very high avidity)	Mono- to multivalent (ligand density-dependent)	Endogenous Tf receptor expression varies by region/disease; rodent–human differences	NR; enter as fold vs non-targeted control (specify metric: AUC, $K_p$ ,brain, $K_p$ ,uu)	Canonical BBB shuttle; density govern recycling vs lysosomal routing; can be paired with parenchymal motifs	[276,277]
Anti-TfR antibodies / fragments / bispecific shuttles → TfR	NR in manuscript; affinity and epitope selection critical	Often monovalent/low-avidity formats preferred; bispecific designs common	Epitope-specific species cross-reactivity; high affinity can increase trapping; saturation effects	NR; report with dosing window and comparator	Design goal is efficient transcytosis with minimal TfR downregulation and reduced endothelial retention	[278–280]
Angiopep-2 → LRP1	NR in manuscript; ligand density and avidity tuning highlighted	Typically multivalent on nanocarriers; density optimized to avoid sequestration	LRP1 expression/content dependence; human relevance must be confirmed; tumor vs healthy BBB differences	NR; report relative to non-targeted carrier	Widely used peptide shuttle for nanoparticles and conjugates; can support glioblastoma-directed constructs	[281–283]
ApoE-mimetic	NR in manuscript	Mono- or multivalent;	Strong endogenous	NR; specify endpoint	Leverages lipoprotein	[284,285]

peptides → LDLR (±LRP1)	pt; affinity window and release kinetics emphasiz ed	avidity increases uptake but can increase trapping	ApoE/LDL competition; lipid-state effects; species differences in lipoprotein biology	(brain/plasma ratio, AUC)	trafficking; cleavable linkers and controlled valency can aid parenchymal release
Insulin engineered IR ligands / anti-IR formats → IR	NR in manuscri pt; avoid receptor saturation	Low-avidity designs generally favored	Physiological ceiling and safety constraints (glucose homeostasis); high endogenous competition; species differences	NR; report alongside safety/tolerabil ity	Attractive but [240] constrained by homeostatic receptor function; format and dosing are decisive
Aptamers / alternative binders (e.g., TfR- or LRP1- binding) → RMT receptors	NR in manuscri pt; receptor- specific values to be inserted	Usually monovalent; multimerizati on possible	Cross-reactivity and epitope mapping required; stability in plasma and nuclease resistance differ by species	NR; populate with harmonized assay definitions	Modular [276,286] alternatives to peptides/antibod ies; can reduce immunogenicity but require robust CMC characterization

ApoE, apolipoprotein E; AUC, area under the curve; BBB, blood–brain barrier; CMC, chemistry, manufacturing, and controls; IR, insulin receptor;  $K_d$ , dissociation constant;  $K_{p,brain}$ , brain-to-plasma partition coefficient;  $K_{p,uu}$ , unbound brain-to-plasma partition coefficient; LDL, low-density lipoprotein; LDLR, low-density lipoprotein receptor; LRP1, low-density lipoprotein receptor–related protein 1; NR, not reported; RMT, receptor-mediated transcytosis; Tf, transferrin; TfR, transferrin receptor.



**Figure 2.** Decision tree for selecting CNS delivery strategies by payload liabilities and clinical context. The schematic provides a pragmatic decision tree to match a CNS payload to the most efficient delivery strategy. The workflow begins with dominant liabilities: (i) solubility/dissolution limitation, (ii) efflux liability (e.g., P-gp/BCRP/MRPs), and (iii) chemical or metabolic instability. Solubility-driven failure routes first to formulation solutions (solubilizing vehicles, lipid systems, polymeric nanoparticles, mucoadhesive/intranasal formats when appropriate). Predominant efflux liability routes to prodrug design (efflux-evading promoieties or transporter-hijacking approaches such as GLUT1/LAT1/MCT1) and/or carrier shielding/targeting (stealth/corona control; RMT ligands such as TfR/LRP1). Instability-driven failure routes to protective encapsulation or stability-optimized prodrugs with controlled release. Clinical context modifiers (need for rapid onset, diffuse vs focal pathology, and tolerability constraints) direct selection of route (e.g., intranasal bypass) or focused ultrasound (FUS) + microbubbles for localized BBB modulation. Each terminal node specifies decision-grade endpoints: brain exposure (AUC,  $K_{p,brain}$ ,  $K_{p,uu}$ ), target engagement, safety margin, and CMC scalability. AUC, area under the curve; BBB, blood–brain barrier; BCRP, breast cancer resistance protein; CMC, chemistry, manufacturing, and controls; CNS, central nervous system; FUS, focused ultrasound; GLUT1, glucose transporter 1;  $K_{p,brain}$ , brain-to-plasma partition coefficient;  $K_{p,uu}$ , unbound brain-to-plasma partition coefficient; LAT1, L-type amino acid transporter 1; MCT1, monocarboxylate transporter 1; MRPs, multidrug resistance–associated proteins; P-gp, P-glycoprotein; RMT, receptor-mediated transcytosis; TfR, transferrin receptor; LRP1, low-density lipoprotein receptor–related protein 1.

## 6. Alternative Routes and Device-Enabled Blood-Brain Barrier (BBB) Opening

### 6.1. Intranasal Nose-to-Brain

The intranasal route exploits the unique anatomical connectivity between the nasal cavity and the brain through the olfactory epithelium and the branches of the trigeminal nerve [176,287]. These pathways enable both intra- and extra-neuronal transport, providing rapid and direct access to the CNS while bypassing systemic circulation and hepatic first-pass metabolism [176,288]. Such direct trafficking has been demonstrated for a wide range of small molecules, peptides, and nanocarrier systems, reinforcing the potential of this route for delivering neuroprotective phytochemicals and engineered prodrugs [176,289,290].

Formulation science has been central to enhancing this delivery mode [287,289]. Mucoadhesive in situ gels, often thermo- or ion-responsive, prolong nasal residence time and counteract mucociliary clearance, while nanoemulsions improve solubility and stability of hydrophobic polyphenols such as

curcumin or resveratrol [176,291,292]. Nanoparticulate systems—ranging from lipid-based carriers to chitosan-modified polymeric nanoparticles—further allow surface functionalization for improved permeability and targeted release [149,290,293]. Prodrug strategies that exploit enzymatic conversion within the nasal mucosa are being increasingly explored to improve bioavailability and sustain brain exposure [287].

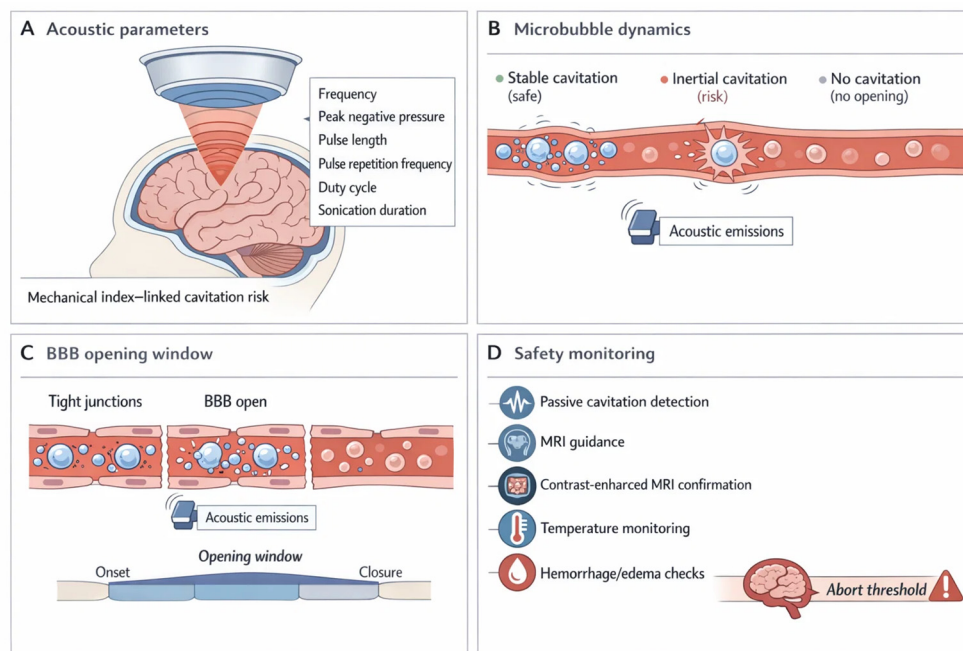
Despite this promise, translational hurdles remain [289,294]. Anatomical variability, short retention time, and interindividual differences in nasal airflow complicate dosing precision and reproducibility in humans [176,294]. Ergonomic device design, accurate metered dosing, and integration with pharmacokinetic modeling will be essential for clinical translation [176,294]. Large-scale human trials, coupled with regulatory harmonization, are still required before intranasal nanoformulations can be considered reliable delivery systems for neurotherapeutics [287,289,295].

## 6.2. Focused Ultrasound + Microbubbles (FUS)

Focused ultrasound combined with circulating microbubbles has emerged as one of the most precise approaches to transiently opening the BBB [296,297]. The mechanism relies on acoustic cavitation, in which microbubbles oscillate in response to ultrasound exposure, producing localized shear stress on the vascular endothelium [296,298,299]. This process induces mechanoporation and loosening of tight junctions, thereby increasing paracellular permeability in a controlled and reversible manner [296,298,299]. Importantly, both stable and inertial cavitation contribute to permeability enhancement, yet parameters must be carefully tuned to avoid endothelial damage or hemorrhage [298,300,301]. The reversible nature of the opening distinguishes FUS from chemical osmotic methods, as the barrier typically restores within hours [296].

A major strength of this technology lies in its spatiotemporal precision [296,302,303]. MRI-guided or neuronavigation-based systems allow targeting of submillimeter brain regions, enabling localized drug accumulation with minimal off-target exposure [302,304,305]. Real-time cavitation monitoring, coupled with feedback-controlled ultrasound delivery, provides essential safety guardrails, reducing risks of edema, neuroinflammation, or microvascular injury [300,306,307]. Longitudinal studies in both primates and humans confirm the feasibility of repeated sessions without significant adverse cognitive effects, although vigilance for subtle inflammatory responses remains necessary [298,308,309].

Clinical translation is already well under way [302]. Phase I trials in gliomas, Alzheimer's disease, and Parkinson's dementia consistently report tolerability and transient BBB disruption, with imaging confirming enhanced delivery of chemotherapeutics, antibodies, and nanoparticles [298,302,310]. While no trials have yet evaluated phytochemicals or natural prodrugs directly, the compatibility of FUS with nanocarriers and controlled-release systems makes such applications plausible [297,299,310]. Integrating polyphenol-based therapeutics into FUS platforms could represent a novel frontier for noninvasive neuroprotection and disease modification [299,310] (Table 3).



**Figure 3.** Focused ultrasound (FUS) delivery paradigm: acoustic control, microbubble cavitation, transient blood–brain barrier (BBB) opening, and safety monitoring. The schematic summarizes the focused ultrasound (FUS)–microbubble paradigm for transient, spatially targeted BBB modulation to enable CNS delivery. (A) Acoustic parameters (frequency, peak negative pressure, pulse length, pulse repetition frequency, duty cycle, and sonication duration) set the cavitation regime and therefore the balance between efficacy and risk[311–313] 10.3171/2017.11.focus17627. 10.1016/j.ultsonch.2018.12.031. 10.1016/j.ultsonch.2016.09.016. (B) Microbubble dynamics span a spectrum from no response (insufficient opening) to stable cavitation (desired oscillation that increases permeability) and inertial cavitation (collapse associated with vascular damage risk). These regimes are inferred in real time using acoustic emissions and passive cavitation detection to support parameter adjustment and stop rules[307,314,315] 10.1038/s41598-020-66994-8. 10.1016/j.ultsonch.2023.106291. 10.1109/tbme.2018.2882337. (C) BBB opening window is depicted as a reversible permeability increase (minutes–hours) during which small molecules, prodrugs, and nanocarriers can cross the endothelium; closure restores barrier integrity[316–318] 10.1371/journal.pone.0125911. 10.1038/s41598-018-36340-0. 10.3171/2022.9.jns221565. (D) Safety monitoring includes image-guided targeting (MRI), confirmation of opening (contrast-enhanced MRI), surveillance for edema or microhemorrhage, and temperature/physiological checks where relevant. Together, the framework links controllable acoustic inputs to microbubble behavior, delivery timing, and verification steps required for reproducible, trial-grade BBB opening[307,316,319] 10.1073/pnas.2103280118. 10.1038/s41598-020-66994-8. 10.1371/journal.pone.0125911. BBB, blood–brain barrier; CNS, central nervous system; FUS, focused ultrasound; MRI, magnetic resonance imaging.

### 6.3. Chemical/Osmotic Opening and Convection-Enhanced Delivery (CED)

Classical BBB-disruption strategies such as intra-arterial mannitol, DMSO co-solvent effects, and bradykinin analogs can transiently loosen tight junctions, yet their clinical utility has waned [273,320]. The reasons are consistent across reviews: non-selective permeability increases, variable magnitude and duration of opening, and procedure-related risks, including seizures, edema, or ischemic events [321,322]. Lack of spatiotemporal control and systemic toxicities further erode risk–benefit in longitudinal care [320]. For phytochemicals and natural prodrugs, which often require sustained or repeated exposure, these invasive and poorly tunable methods are a poor fit; cumulative toxicity and patient burden compound the translational gap [320,323].

CED preserves a niche when focal, high-dose deposition is essential, as in gliomas or diffuse midline lesions, particularly with implantable ports and image-guided catheters, despite technical complexity and heterogeneity of distribution [324–326]. When chemistry can carry the payload,

prodrugs simplify the problem, if transporter hijacking and cleavage are tuned correctly [327,328] (Table 5).

**Table 5.** Alternative routes and device-enabled blood-brain barrier (BBB) modulation: evidence and risk-mitigation matrix for improving central nervous system (CNS) exposure. This table summarizes alternative routes and device-enabled strategies to bypass or locally modulate the BBB when systemic nanocarriers or prodrugs are insufficient. Evidence level (preclinical to clinical use) is paired with key translational risks. Intranasal delivery enables olfactory/trigeminal access but is limited by variability and dose volume. Focused ultrasound (FUS) with microbubbles provides precise, reversible BBB opening requiring cavitation-aware control and imaging verification. Osmotic/chemical disruption increases permeability at the cost of safety and control. Convection-enhanced delivery (CED) enables focal infusion but is invasive. External-field approaches remain exploratory.

Modality	Mechanism	Evidence level	Clinical status	Advantages	Risks/mitigations	References
Intranasal nose-to-brain (sprays, gels, nanoemulsions)	Direct transport along olfactory and trigeminal pathways; reduced first-pass metabolism; mucoadhesion prolongs residence	Preclinical strong; early clinical emerging (context-dependent)	Used clinically for some CNS-active molecules; delivery platforms under evaluation for neurodegeneration/oncology	Noninvasive; rapid onset potential; bypasses systemic dilution for suitable payloads; compatible with solubility-enabling formulations	High inter-individual variability (anatomy, mucociliary clearance); limited dose volume; nasal irritation—mitigate with device optimization, deposition mapping, mucoadhesive/in situ gels, and PK endpoints (AUC, $K_{p,uu}$ when feasible)	[4,155,329]
Focused ultrasound (FUS) + microbubbles	Pulsed acoustic exposure drives stable cavitation-mediated mechanoporation and transient tight-	Robust preclinical; multiple early clinical studies	Clinical translation under way (MRI-guided protocols in neuro-oncology and neurodegeneration)	Spatially targeted, reversible opening; compatible with diverse payloads (small molecules, prodrugs,	Hemorrhage/edema risk with inertial cavitation; off-target opening—mitigate via cavitation monitoring (acoustic emissions/PCD), conservative	[148,330,331]

	junction/transport changes enabling local BBB opening			nanocarriers, biologics); enables region-specific dosing	parameter sets, MRI guidance, contrast-enhanced confirmation, and predefined abort thresholds	
Osmotic BBB disruption (intra-arterial mannitol)	Hyperosmolar shrinkage of endothelial cells transiently widens tight junctions and increases permeability	Established concept; variable evidence by indication and protocol	Applied in select centers/indications; invasive and less commonly used than device-guided opening	Can increase delivery of otherwise excluded agents; compatible with intra-arterial administration	Poor spatial control; seizure/edema/hemorrhage risk; procedure-related risks—mitigate with stringent patient selection, hemodynamic monitoring, imaging surveillance, and avoidance of programs relying on nonspecific leak	[148,330,332]
Chemical permeability modulation (selected permeabilizers/co-solvents)	Transiently alters membrane integrity, tight-junction signaling, or transporter function to raise permeability	Limited to mixed; often preclinical or adjunctive	Not routine for broad delivery; cautiously adjuncts in narrow settings	Potentially simple to implement; can be paired with systemic dosing when local devices are unavailable	Nonspecific barrier disruption and systemic toxicity; unpredictable PK and inflammation—mitigate with minimal-effective exposure, local delivery where possible, tight safety biomarkers, and preference for controllable modalities	[148,332,333]
Convection-enhanced	Pressure-driven	Strong preclinical/c	Clinical use and trials in neuro-	High local dose;	Invasive; catheter placement errors,	[326,334,335]

delivery (CED)	interstitial infusion via intracranial catheter achieves high local concentrations independent of BBB transport	clinical experience in focal indications	oncology and focal CNS procedure-dependent	bypasses targets; systemic barriers; controllable infusion profiles; suitable for macromolecules and particles	reflux/backflow, heterogeneous distribution, infection—mitigate with image-guided planning, real-time distribution tracking, optimized cannula design, and sterility controls	
External-field targeting/trigging (magnetic guidance; remote release)	Magnetic gradients concentrate magnetically responsive carriers; external fields can trigger release from stimuli-responsive constructs	Primarily preclinical; exploratory translation	Investigational; requires specialized hardware and long-term data	Adds spatiotemporal control without barrier-opening; can pair with imaging-enabled carriers for tracking	Uncertain long-term retention/clearance; heating and off-target accumulation; device standardization gaps—mitigate with biocompatible coatings, rigorous dosimetry, biodistribution/clearance studies, and conservative escalation	[4,148]

auc, area under the curve; BBB, blood–brain barrier; ced, convection-enhanced delivery; cns, central nervous system; fus, focused ultrasound; mri, magnetic resonance imaging; pcd, passive cavitation detection; pk, pharmacokinetics; kp,uu, unbound brain-to-plasma partition coefficient.

## 7. Prodrugs and Transporter Hijacking

### 7.1. LAT1/MCT1/GLUT1-Targeted Prodrugs

Among the influx transporters that shape small-molecule entry into the brain, the large neutral amino acid carrier LAT1 has emerged as the most exploited in prodrug design [336–338]. LAT1 is highly expressed on the luminal side of brain capillaries and recognizes aromatic and branched-chain amino acids as substrates [53,338,339]. By conjugating drugs with phenylalanine, tyrosine, or related promoieties, it is possible to achieve carrier-mediated uptake that circumvents passive BBB limitations [336,337,340]. LAT1-linked prodrugs of valproic acid, ferulic acid, and NSAIDs have shown superior brain penetration, and the Xiong 2021 dataset provides compelling evidence that

conjugated neurotherapeutics not only cross the BBB but also accumulate within neurons, astrocytes, and microglia, confirming cellular specificity of uptake [341–343].

Beyond LAT1, other solute carriers are beginning to attract attention [56,338] Monocarboxylate transporter 1 (MCT1) recognizes lactate and pyruvate analogues, providing a scaffold for monocarboxylate-linked prodrugs, whereas GLUT1, the primary glucose transporter, can be hijacked via glucose conjugation [53,56,338]. Proof-of-principle studies demonstrate that indomethacin and ketoprofen conjugated to glucose traverse the BBB in rodents, though kinetic competition with endogenous glucose poses significant challenges [53,343]. These strategies illustrate the expanding toolkit for tailoring prodrug chemistry to align with the substrate repertoire of BBB carriers [53,338].

Transporter hijacking, however, is not without risk [336,344]. Kinetic constraints such as  $K_m$  and  $V_{max}$  dictate the efficiency of uptake, and saturation by high-affinity endogenous substrates can diminish drug delivery [53,337]. Moreover, transporter expression varies across species, complicating preclinical-to-clinical translation [56,338]. LAT1 prodrugs are generally selective, off-target interactions and potential saturation effects remain critical safety considerations [341,345,346]. The challenge now lies in fine-tuning conjugate chemistry to balance affinity, stability, and enzymatic cleavability in the brain while minimizing systemic exposure [339,340,342].

### 7.2. Lipidization, Soft Drugs, Self-Immolative Linkers

Lipidization remains one of the oldest yet most versatile strategies for enhancing drug penetration into the brain [347–349]. By appending lipophilic chains or glyceride motifs, polar APIs can acquire sufficient passive diffusion across endothelial membranes, provided the modifications are designed for efficient cleavage once in the CNS [347–349]. This balance between increased lipophilicity and metabolic lability is critical: too stable and the parent drug may not be released; too labile and systemic hydrolysis prevents brain delivery [347,349]. The approach has been applied successfully to small neuroactive agents, though reproducibility across species remains a central design challenge [348,350].

In parallel, soft drug concepts introduce the inverse logic: compounds are deliberately engineered for predictable inactivation outside the CNS, ensuring that only a fraction escapes rapid metabolism and reaches the brain [347,349]. Self-immolative linkers add yet another layer of sophistication, exploiting pH gradients, enzyme expression, or redox triggers to launch controlled cleavage cascades [351–353]. Modern designs favor traceless release, often with dual stimuli or cascade amplification to achieve brain-first activation while avoiding premature systemic leakage [351,352,354]. The guiding rule across these platforms is to harmonize stability, trigger sensitivity, and cleavage kinetics so that release occurs only under CNS-relevant conditions, minimizing off-target toxicity while maximizing therapeutic gain [351,352,354].

### 7.3. Solubility Boosters (Cyclodextrins, Co-Crystals, Ion Pairing)

Cyclodextrins have been widely used to improve aqueous solubility through inclusion complexes that sequester hydrophobic moieties within their cyclic cavities [142,355,356]. This strategy can significantly enhance systemic exposure and oral bioavailability, yet it offers little direct benefit for BBB permeation, as the bulky complexes rarely cross endothelial tight junctions intact [355,357,358]. Their role is therefore supportive: enabling consistent systemic levels that may feed into other brain-targeted strategies rather than acting as genuine CNS delivery enhancers [356,359,360].

Co-crystals and ion pairing occupy a more dynamic niche [361]. Co-crystals modify dissolution rates and solubility without altering the pharmacodynamic profile of the parent drug, creating opportunities for predictable exposure kinetics [361–363]. Ion pairing, in contrast, transiently adjusts lipophilicity by associating ionizable drugs with counterions, thereby improving membrane partitioning and yielding short-lived permeability gains [364–366]. The central distinction is crucial: while all three approaches may improve systemic bioavailability, only certain ion-pairing strategies directly modulate BBB permeability [364–366]. Recognizing this separation between systemic

solubility enhancers and true BBB permeability modulators is essential when positioning such methods within prodrug pipelines [365–367].

#### 7.4. Nano-Prodrug Conjugates

Nano-prodrug conjugates represent a convergence of nanomedicine and classical prodrug chemistry [368–370]. In these systems, nanocarriers such as polymers, liposomes, or albumin-binding constructs are covalently linked to prodrug moieties, creating assemblies that combine carrier stability with controlled release [368,370,371]. Examples include polymer-drug conjugates that self-assemble into micelles or nanoparticles, and liposome-prodrug hybrids that integrate covalently modified drugs into bilayer structures [227,368,372]. Activation is then triggered by tumor- or CNS-relevant stimuli such as redox gradients, pH shifts, or enzyme cleavage, ensuring spatially restricted release [269,372,373].

The rationale for this complexity is strongest when dealing with drugs that have narrow therapeutic windows or poor solubility, where conventional formulations risk systemic toxicity or inadequate exposure [370,374]. By embedding prodrug chemistry within nanocarriers, it becomes possible to synchronize delivery, minimize premature release, and improve therapeutic indices [368,374,375]. Yet translation remains challenging [375–377]. Manufacturing reproducibility, batch-to-batch stability, and regulatory pathways for hybrid entities blur the lines between drug and device, complicating approvals [376–378]. Scalability and quality control of multifunctional prodrug nanocarriers are further hurdles that limit current clinical penetration despite compelling preclinical evidence [376,377,379]. Delivery vectors and prodrugs must be vetted in models that actually predict human exposure—next we align models with decision-grade endpoints [376,377] (Table 6).

**Table 6.** Prodrug design playbook for BBB delivery: transporter-hijacking promoieties, brain-selective cleavage logic, and translation risks. This table condenses transporter-hijacking prodrug strategies into a practical design checklist for BBB delivery. It centers on key influx carriers (LAT1, GLUT1, MCT1), outlining common promoieties, linker chemistries, and brain-selective cleavage triggers that mitigate polarity, efflux, and instability while limiting premature systemic activation. Exposure gain is reported as fold-change versus parent or non-targeted controls using harmonized endpoints (brain AUC,  $K_p$ ,brain, preferably  $K_p$ ,uu), with “NR” for values not specified. Off-target risks include substrate competition, peripheral uptake, species differences, and unintended metabolite activity; notes emphasize  $K_m/V_{max}$ -aware design, linker stability, and assay/QC needs.

Transporter	Promoiety/linker	Cleavage trigger	Exposure gain	Off-target risks	Notes	References
LAT1 (large neutral amino acid transporter)	L-amino acid promoieties (e.g., phenylalanine/leucine/tyrosine analogs); amide, or carbamate linkers; optional self-immolative spacers	Brain-enriched esterases/peptidases; linker-enabled self-immolation after enzymatic trigger	NR (populated with fold-change in brain AUC, $K_p$ ,brain or $K_p$ ,uu)	Competition with endogenous amino acids; saturation at high dose; peripheral uptake (gut, kidney); rodent-human affinity/epitope differences	Prefer moderate flux over trapping; design should be $K_m/V_{max}$ -aware; verify brain-selective cleavage and low systemic conversion; include efflux	[380–382]

					liability screening for released parent	
GLUT1 (glucose transporter )	Glucose or glucosyl-like promoieties; O- or C-linked glycosides; carbonate/carbamate/ester linkers for release	Glycosidase-assisted unmasking (where applicable) and/or esterase-triggered cleavage of linkers; self-immolative release modules	NR (report with matched control and dosing window)	High peripheral distribution (erythrocytes/endothelium); competition with glucose; risk of rapid systemic cleavage; potential metabolic liabilities	Aim for productive transport without excessive binding; validate stability in plasma and nasal/intestinal matrices; monitor impact on glucose handling only where pharmacologically plausible	[380,383,384]
MCT1 (monocarboxylate transporter )	Monocarboxylate promoieties (e.g., lactate/pyruvate/acetate-like); ester linkers; soft-drug variants to tune logD	Carboxylesterase-mediated cleavage; pH/enzyme-sensitive linkers can bias release toward brain compartment	NR (capture as fold-change in brain exposure and unbound fraction when available)	Peripheral uptake (muscle, liver); competition with endogenous monocarboxylates; acidosis-related confounding in sensitive species differences in transporter expression	Useful for polar acids/phenolics; quantify competition effects under physiological substrate levels; include brain-selective cleavage validation and metabolite profiling	[385–387]
Multiple SLCs (exploratory / case-by-case)	Nutrient-mimetic fragments matched to a selected transporter's substrate modular linkers	Enzyme-labile trigger + self-immolative release	NR (insert when transporter, affinity)	Uncertain selectivity; target tissue uptake; unpredictable metabolism;	Use only with strong transporter evidence (expression at BBB + uptake)	[380,388,389]

	(esters/amides/carbamates)	(design-dependent)	, and PK endpoints are specified)	model-to-human translation risk	assays); pair with orthogonal confirmation (inhibitors/knockdown, saturability, competitive substrates)
Efflux-evading (non-transporter -hijacking) prodrugs	Mask donors/acceptors; increase modestly; promoieties reduce recognition by P-gp/BCRP; soft-drug linkers	H-bond logD that avoid premature conversion)	Systemic brain esterases (must be tuned to avoid premature conversion) or NR (report brain exposure and safety vs parent)	Premature systemic activation; altered distribution and toxicity; metabolite formation; drug-drug interactions	Useful when [389–391] are not practical; requires early efflux screening (P-gp/BCRP) and rigorous metabolite ID; prioritize K <sub>p,uu</sub> as decision endpoint

AUC, area under the curve; BBB, blood–brain barrier; BCRP, breast cancer resistance protein; GLUT1, glucose transporter 1; km, Michaelis constant; kp,brain, brain-to-plasma partition coefficient; kp,uu, unbound brain-to-plasma partition coefficient; LAT1, large neutral amino acid transporter 1; logD, distribution coefficient; MCT1, monocarboxylate transporter 1; nr, not reported; P-gp, P-glycoprotein; pk, pharmacokinetics; SLCs, solute carrier transporters; v<sub>max</sub>, maximum transport rate.

## 8. Biogenic and Exosome-Mimetic Vesicles

### 8.1. Mammalian Exosomes

Mammalian exosomes have attracted intense interest as endogenous delivery vehicles, given their origin from neuronal, immune, and stem-cell lineages [392,393]. Neuron-derived vesicles display inherent neurotropism, while macrophage or dendritic cell exosomes often retain immunological signaling capabilities that can be leveraged for targeted delivery [393,394]. Stem cell-derived vesicles, particularly those from mesenchymal sources, exhibit regenerative properties and have been applied in models of neuroinflammation and tissue repair [395,396]. This natural diversity provides a menu of options for CNS-directed therapy, with the vesicle's parent cell type influencing both tropism and therapeutic payload [395,397].

Several methods exist for incorporating cargo into exosomes [394,398]. Electroporation transiently disrupts vesicle membranes to load nucleic acids, while passive incubation exploits lipid bilayer partitioning [394,399]. Sonication and extrusion, though less subtle, can increase loading efficiency for small molecules and proteins [399,400]. More sophisticated approaches combine chemical conjugation or ligand decoration to engineer selective homing properties, extending beyond the vesicle's innate targeting profile [398,401].

Despite these advantages, translational obstacles remain formidable [402,403]. Batch-to-batch variability complicates reproducibility, and large-scale production has yet to reach regulatory-grade consistency [392,402,403]. Issues of heterogeneity, yield, and purification standards pose barriers to clinical adoption, while classification of exosomes as biologics, devices, or drug–biologic hybrids remains unresolved [404,405]. Thus, mammalian exosomes stand at the intersection of promise and challenge, offering unmatched biocompatibility but demanding rigorous solutions in scalability and regulation before they can function as reliable neurotherapeutic vectors [392,393].

### 8.2. Plant-Derived Extracellular Vesicles

Plant-derived extracellular vesicles (PDEVs) are emerging as abundant, low-cost nanocarriers harvested from edible sources such as ginger, grape, and citrus [406–408]. Their natural stability, low immunogenicity, and tolerance to gastrointestinal conditions make them particularly attractive for oral or intranasal administration, routes that remain challenging for mammalian exosomes [406,409,410]. PDEVs also carry intrinsic bioactive metabolites, adding antioxidant and anti-inflammatory potential to their delivery role [408,411,412].

Despite these advantages, several limitations temper enthusiasm [406,407]. Vesicle heterogeneity across plant species and even between batches complicates reproducibility, while the mechanisms by which PDEVs interact with or traverse the BBB remain poorly defined [406,412,413]. Preclinical studies demonstrate promising antioxidant and anti-inflammatory effects in models of neuroinflammation and oxidative stress, yet translation into predictable CNS uptake remains uncertain [414–416]. Thus, PDEVs occupy a unique space: safe, scalable, and bioactive, but requiring deeper mechanistic insight before they can be positioned as reliable neurotherapeutic vectors [407,413,417].

### 8.3. Synthetic Mimetics

Synthetic exosome-mimetic vesicles are designed to replicate the communication and delivery roles of natural exosomes while sidestepping their limitations of yield and heterogeneity [418,419]. Strategies include polymersomes with controllable membrane chemistry, membrane-coated nanoparticles that borrow cellular surface markers, and hybrid designs that combine synthetic scaffolds with natural membrane fragments [420–422]. Such constructs excel in tunability and scalability, making them better suited for standardized manufacturing compared to their mammalian counterparts [418,419,423].

Yet these advantages come with trade-offs [421,422]. Replacing native membranes often diminishes biocompatibility cues that exosomes naturally provide, raising concerns about immune activation and altered clearance [421–423]. Nonetheless, synthetic platforms allow for reproducible incorporation of targeting ligands or exosomal motifs, offering a level of precision that natural vesicles rarely achieve [418–420]. Early applications highlight their promise in oncology, regenerative medicine, and CNS targeting [421,424,425]. We now connect models to the endpoints that drive go/no-go decisions.

## 9. Translational Models and Decision-Enabling Endpoints

### 9.1. In Vitro Models

In vitro BBB models remain indispensable as early decision tools in neurotherapeutic development [426,427]. Classic Transwell systems with endothelial monolayers offer simplicity and throughput but often fail to reproduce the restrictive tight junctions of the human BBB [428,429]. Adding astrocytes or pericytes in co-culture improves fidelity, as astrocytic signals reinforce junctional protein expression and better align transendothelial electrical resistance (TEER) values with physiological ranges [427,430,431]. These refinements help distinguish passive permeability from transporter-mediated flux, although limitations in dynamic responses persist [428,432].

Human induced pluripotent stem cell (iPSC)-derived BBB organoids introduce greater biological relevance by capturing species-specific expression of transporters and efflux pumps [427,433]. However, the lack of standardized differentiation protocols results in variable permeability and metabolic profiles across laboratories [427,434]. Microfluidic BBB-on-chip systems address some of these issues by incorporating shear stress, nutrient gradients, and continuous flow, recapitulating the hemodynamic conditions that shape BBB integrity [428,432,435]. These dynamic constructs offer a closer physiological context but are more technically demanding and costly to implement [432,435].

Benchmarking against in vivo data remains essential [428,435]. Metrics such as apparent permeability ( $P_{app}$ ) and TEER are routinely compared to animal and human datasets, yet over- or underestimation of drug transport is common [429,436]. The dataset reported by Aday and colleagues (2016) illustrates how carefully calibrated microfluidic models can achieve closer alignment with in vivo permeability coefficients [428,435]. Despite progress, no single in vitro system fully resolves the trade-off between scalability and predictive accuracy, underscoring the need for model selection tailored to the specific decision point in development [426,432].

### 9.2. *In Vivo Models and Species Differences*

Rodent models remain the workhorse of preclinical neurotherapeutics, offering high-throughput screening, ease of genetic manipulation, and well-characterized disease models [437]. Yet their BBB exhibits greater paracellular leakiness than that of primates, which can overestimate drug penetrance [438,439]. This divergence partly explains why promising rodent data often fail to translate into clinical success [437,440]. Moreover, rodents display transporter expression patterns that differ in both abundance and substrate specificity compared to humans, adding further complexity to predictions of central exposure [440–442].

Non-human primates provide the closest approximation of human BBB integrity and regional perfusion characteristics [441,443]. Their barrier tightness, transporter repertoire, and cerebrovascular physiology more closely align with human data, making them critical for late-stage validation [439,444]. However, cost, ethical concerns, and limited availability restrict their widespread use [444]. Adding further complication, disease states reshape barrier permeability: ischemic stroke disrupts endothelial junctions, Alzheimer's disease alters transporter activity, and glioblastoma induces localized leakiness that changes drug distribution [344,445,446].

These species and disease-dependent differences emphasize the translational gap between model systems and patients [437,438]. Cross-species network analyses and computational integration strategies are increasingly used to bridge this gap, but the fundamental challenge remains: no single in vivo model fully captures the nuances of human BBB physiology [429,447]. Strategic selection and careful benchmarking are therefore essential to guide go/no-go decisions in CNS drug development [429,439].

### 9.3. *Quantitative PK Endpoints*

Quantitative pharmacokinetic endpoints are central to linking drug exposure with CNS activity [448]. The unbound fraction in brain tissue ( $f_{u,brain}$ ) defines the pharmacologically active pool, while the brain-to-plasma partition coefficient ( $K_{p,brain}$ ) describes overall distribution across compartments [449]. A more precise index is  $K_{p,uu,brain}$ , the ratio of unbound brain to unbound plasma concentrations, which reflects true equilibrium between compartments and better predicts central efficacy [449,450]. These parameters guide whether a compound achieves sufficient free concentrations at its target site or is limited by efflux transporters and protein binding [451,452].

Measuring these endpoints remains technically demanding [448]. Microdialysis enables direct sampling of interstitial fluid, offering dynamic readouts of unbound concentrations, but it is invasive and limited to specialized settings [453]. Homogenate binding assays, in contrast, are more accessible but prone to overestimation due to disrupted tissue architecture [450,454]. Cerebrospinal fluid is often used as a surrogate for interstitial concentrations, yet differences in turnover and compartmentalization mean CSF rarely mirrors brain extracellular fluid with high fidelity [455,456].

Regulators increasingly emphasize integration of such quantitative PK endpoints with pharmacodynamic measures, particularly through physiologically based pharmacokinetic and PK–PD models, to inform dose selection and reduce translational uncertainty [456–458]. By anchoring drug development in  $f_{u,brain}$ ,  $K_{p,brain}$ , and  $K_{p,uu,brain}$ , researchers can more confidently bridge preclinical data with human predictions and make decision-grade assessments of CNS penetration [450,459].

#### 9.4. Imaging and Biomarker Readouts

Imaging has become a cornerstone in evaluating how drugs and nanocarriers navigate the BBB [460,461]. Positron emission tomography (PET) tracers provide sensitive, quantitative assessments of permeability, while magnetic resonance imaging (MRI) with contrast agents captures dynamic leakage and regional perfusion in vivo [460,462]. Recent work extends these approaches to track nanocarrier fate over time, revealing how size, charge, and surface chemistry influence deposition within target regions [461,463]. Such dynamic readouts are invaluable not only for confirming delivery but also for ruling out vascular compromise or off-target accumulation that could cloud efficacy signals [464].

In parallel, biomarker development is beginning to complement and extend imaging [465]. Neuroinflammation markers such as GFAP or ICAM-1 flag astrocytic and endothelial responses, while circulating exosomal signatures hint at brain-specific injury or remodeling processes [466,467]. The true translational power lies in linking imaging and biomarkers simultaneously to efficacy—drug exposure within the intended region—and safety, including the detection of edema or inflammatory activation [468,469]. Emerging strategies combine multimodal imaging with panels of fluid biomarkers, offering a near real-time window into drug delivery, target engagement, and tissue response [465,470,471]. This convergence is setting the stage for decision frameworks that go beyond single endpoints and instead integrate orthogonal readouts to guide go/no-go calls with greater confidence [468,472]. What has actually reached patients? We summarize clinical traction and why certain bets are moving first [460] (Table 7).

**Table 7.** Models-to-endpoints crosswalk for blood-brain barrier (BBB) delivery: throughput, predictive scope, artifacts, and best-fit development decisions. This crosswalk links commonly used BBB/central nervous system (CNS) delivery models discussed in the manuscript to the endpoints they most reliably support. For each model, throughput is contrasted with predictive scope (barrier integrity, transporter effects, regional delivery, or human translation), and key artifacts that bias interpretation are flagged. The Best-fit decision column indicates where each approach is most informative in a development workflow—from early screening to go/no-go based on decision-grade brain exposure (e.g.,  $K_{p,uu}$ ) and target engagement.

Model	Throughput	What it predicts	Key artifacts	Best-fit decision	References
Transwell (endothelial mono-/co-culture; static TEER/permeability)	High	Relative permeability and gross barrier integrity; early ranking of formulations/prodrugs; qualitative efflux effects (context-dependent)	Static conditions; nonphysiologic shear; variable tight junction maturation; transporter expression drift; adsorption to plastics	Early screen and rank-order; eliminate non-starters before costly models	[473–475]

iPSC-derived BBB endothelium (Transwell)	Medium	More human-relevant tight junctions/transporters; better prediction of human-like permeability windows	Differentiation variability; batch effects; incomplete neurovascular unit (NVU) signaling unless co-cultured	Mid-stage confirmation of BBB-relevant transport and efflux liability	[476–478]
iPSC organoids / spheroids (NVU-like)	Medium–low	3D cell–cell interactions, and trends; neuroinflammation-compatible testing	uptake and penetration limited perfusion; measurement standardization gaps	Mechanism prioritization and safety/uptake profiling; compare targeting vs non-targeting designs	[479–481]
Microfluidic on-chip (flow/shear; NVU co-culture)	Low–medium	Dynamic barrier responses under flow; transporter-mediated flux; inflammation-dependent permeability shifts	Device-to-device variability; bubble/absorption effects; complex operation; limited throughput	Late preclinical de-risking for mechanism and context dependence (inflammation, disease cues)	[482–484]
Rodent in vivo PK (brain + plasma; brain/plasma ratios)	Medium	System-level exposure, metabolism, distribution; signal of CNS delivery improvement	Species differences in BBB properties and transporters; confounding by vascular space and binding; anesthesia effects	Go/no-go based on integrated exposure; prioritize candidates for quantitative endpoints (K <sub>p,uu</sub> )	[473,477,485]
Rodent microdialysis (ISF sampling)	Low	Unbound interstitial exposure and course; preclinical readout to	Invasive; recovery calibration; regional restriction;	Decision-grade confirmation of CNS penetration (K <sub>p,uu</sub> -like)	[473,477,485]

			target-site pharmacology	limited specialized setups	to inference) and PK/PD linkage	
CSF sampling (preclinical/clinical)	Medium	Surrogate trends when ISF is unavailable; supports translational sampling designs	exposure when ISF is unavailable; supports translational sampling designs	CSF ≠ ISF; compartmental delays; protein binding differences; disease-state confounding	Clinical feasibility planning; supportive evidence alongside imaging or modeling	[473,477,485]
PET imaging (labeled payload or marker)	Low	Whole-brain/spatial distribution; target engagement surrogates; longitudinal kinetics in vivo	Whole-brain/spatial distribution; target engagement surrogates; longitudinal kinetics in vivo	Radiolabel alters properties; metabolite signal; resolution limits; tracer-specific assumptions	Translation-facing biodistribution and engagement readouts; de-risk regional delivery claims	[478,486,487]
Non-human primate (NHP) studies	Very low	Closest approximation to human BBB transport and delivery paradigm	Closest approximation to human BBB transport and delivery paradigm	Cost/ethics; small n; limited disease modeling; procedural constraints	Preclinical-to-clinical bridge for top candidates and delivery devices/targeting ligands	[473,477,485]
Mechanistic / BBB models (incl. efflux and binding)	High (in silico)	Scenario testing; dose-to-exposure translation; integrates binding, efflux, and tissue partitioning	Scenario testing; dose-to-exposure translation; integrates binding, efflux, and tissue partitioning	Parameter uncertainty; requires high-quality data; misspecification risk	Study design, endpoint selection, and translation planning; interpret CSF/ISF and imaging outputs	[473,477,485]

BBB, blood–brain barrier; CNS, central nervous system; CSF, cerebrospinal fluid; ISF, interstitial fluid; iPSC, induced pluripotent stem cell; K<sub>p,uu</sub>, unbound brain-to-plasma partition coefficient; NHP, non-human primate; NVU, neurovascular unit; PBPK, physiologically based pharmacokinetic; PD, pharmacodynamics; PET, positron emission tomography; PK, pharmacokinetics; TEER, transendothelial electrical resistance.

## 10. Clinical Landscape and Case Snapshots

### 10.1. Neuro-Oncology

Focused ultrasound (FUS) with microbubbles is the most advanced clinical strategy for transiently opening the BBB in neuro-oncology [488–490]. By generating localized acoustic cavitation, FUS temporarily loosens tight junctions, permitting chemotherapeutics to achieve higher intratumoral concentrations than with systemic dosing alone [488–491]. Early-phase trials in glioblastoma and brain metastases report encouraging safety signals, with most adverse events being transient edema or headaches rather than irreversible damage [489,491]. Imaging-confirmed increases in drug penetration, paired with pharmacokinetic analyses, have strengthened confidence that this approach is technically feasible and biologically impactful [489,491].

Parallel efforts explore ligand-targeted nanocarriers for glioblastoma, including transferrin- and integrin-directed liposomes, which are designed to selectively home to tumor vasculature or infiltrating glioma cells [492–494]. These systems aim not only to improve local accumulation but also to minimize systemic exposure [492,493]. Key endpoints now extend beyond radiographic progression-free survival to include intratumoral drug levels, pharmacodynamic signatures, and radiomic biomarkers that track response heterogeneity [274,489,490]. Yet regulatory progress remains uneven. Enrollment in neuro-oncology trials frequently lags behind projections, with disparities in infrastructure and patient access limiting study completion rates [317,495]. As adaptive designs and external control datasets gain traction, the field is moving toward more flexible, inclusive trial frameworks capable of sustaining momentum in a disease space with urgent unmet needs [317,490,495].

### 10.2. Neurodegeneration

Alzheimer's disease trials have tested diverse delivery routes, with intranasal insulin standing out for its ability to bypass systemic metabolism and provide direct brain access [496,497]. Peptide- and polyphenol-loaded carriers are also under investigation to stabilize bioactive molecules while enhancing their penetration into hippocampal and cortical regions [498,499]. Although some studies report cognitive benefits and favorable biomarker shifts, variability in patient populations and endpoint sensitivity continues to limit clear conclusions [500–502]. Reliance on radiographic and cognitive scales alone often underestimates subtle, early effects, underscoring the need for multimodal biomarker panels [500,501,503].

In Parkinson's disease, dopamine prodrugs and nanoparticle-based formulations represent strategies to extend half-life and reduce peripheral toxicity while restoring striatal dopamine tone [157,504,505]. Several trials show encouraging motor improvements, yet variability in absorption and BBB transport remains a barrier to consistency [504–506]. Lessons from both Alzheimer's and Parkinson's pipelines converge on the importance of robust biomarkers, sensitive endpoints, and trial designs that accommodate disease heterogeneity [500,501,503]. Without these refinements, even promising therapeutic concepts risk falling short in translation [500,503,507].

### 10.3. Psychiatric and Pain Indications

Early translational studies in psychiatry and pain have focused on phytochemicals such as curcumin and resveratrol, as well as terpenoids such as pinene and linalool, which show preclinical promise for mood regulation and analgesia [30,508,509]. Yet progress into robust clinical validation remains limited [30,508]. Subjective endpoints and the strong influence of placebo responses complicate signal detection, while modest funding and heterogeneous trial designs further slow momentum [510,511]. Current exploratory efforts in depression and chronic pain increasingly employ advanced delivery systems and biomarker-informed approaches, but most remain proof-of-concept [512–515]. The field illustrates both opportunity and fragility in translating natural compounds into psychiatric and pain therapeutics [30,508,516,517].

#### 10.4. Snapshot

First-in-human studies with natural compounds illustrate both opportunity and limitation across therapeutic domains. Oncology has the deepest record, with plant-derived chemotherapeutics and semi-synthetic derivatives advancing into late-stage trials [518,519]. Yet many candidates stall due to safety uncertainties, inconsistent batch quality, or regulatory concerns related to chemistry and manufacturing controls [520]. In neurodegeneration, compounds such as curcumin, resveratrol, and quercetin have entered clinical testing, often showing bioactivity but hampered by poor bioavailability and heterogeneous outcomes [21,521,522]. Psychiatric indications remain the least mature, with strong preclinical rationale but scarce head-to-head trials against approved therapies [523,524].

This comparative landscape highlights clear gaps for phytochemicals: translation remains fragmented, efficacy signals are often modest, and reproducibility suffers without rigorous manufacturing standards [519,520]. The field now recognizes that scientific novelty alone is insufficient. Translation hinges on safety, manufacturability, and regulatory clarity—complex systems demand disciplined CMC [524] (Table 8).

**Table 8.** Clinical snapshot of central nervous system (CNS) delivery strategies in the manuscript: indication-by-modality overview, endpoints, and exposure evidence. This table provides a high-level clinical snapshot of the delivery modalities discussed in the manuscript, organized by representative CNS indications. For each entry, the development phase and primary clinical endpoint are summarized alongside the type of exposure evidence available (e.g., imaging-verified BBB opening, CSF/PK surrogates, or decision-grade brain exposure metrics when reported). “NR” denotes details not specified in the current draft and can be completed during final reference curation.

Indication	Modality	Phase	Primary endpoint	Exposure evidence	Status	References
Neurodegeneration (Alzheimer’s / Parkinson’s)	Focused ultrasound (FUS) + microbubbles	Phase I (as noted in manuscript)	Safety/tolerability; imaging-confirmed BBB opening	MRI guidance + contrast-enhanced confirmation; BBB resealing within hours (NR details)	Early clinical translation under way	[296,525,526]
Glioblastoma / focal CNS tumors	RMT-targeted nanocarriers (e.g., TfR/LRP1 ligands; Angiopep-2-type designs)	Preclinical → early clinical (NR)	Tumor response / progression metrics (NR)	Biodistribution/brain uptake signals; comparator vs non-targeted carrier (NR)	Investigational; target/form at-dependent	[215,527,528]
Glioblastoma / focal CNS tumors	Convection-enhanced delivery (CED)	Clinical use/trials (NR)	Local control and safety (procedure-specific)	High local concentration by direct interstitial infusion; distribution tracking (NR)	Procedure-dependent; used in specialized settings	[324,325,529]

Depression / neuropsychiatric disorders	Intranasal nose-to-brain formulations (sprays, gels, nanoemulsions)	Preclinical → early clinical signals (NR)	Symptom scales and tolerability (NR)	PK/PD signals; CSF or surrogate exposure where available (NR)	Emerging; high variability and formulation -sensitive	[329,530,531]
Broad indications (adjunct permeability strategies)	CNS Osmotic disruption (intra-arterial mannitol)	BBB Selective clinical application (NR)	Feasibility/safety; delivery enhancement (NR)	Increased permeability by protocol; exposure quantification variable (NR)	Invasive; limited use due to safety/control trade-offs	[321,532,533]
Exploratory device-enabled targeting	External-field targeting/trigging (magnetic guidance; remote release)	Preclinical	Proof-of-concept delivery and safety	Tracking-enabled carriers; biodistribution and clearance studies (NR)	Exploratory; hardware and long-term safety gaps	[296,534,535]

BBB, blood–brain barrier; CED, convection-enhanced delivery; CNS, central nervous system; CSF, cerebrospinal fluid; FUS, focused ultrasound; LRP1, low-density lipoprotein receptor–related protein 1; MRI, magnetic resonance imaging; NR, not reported; PD, pharmacodynamics; PK, pharmacokinetics; RMT, receptor-mediated transcytosis; TfR, transferrin receptor.

## 11. Material Safety and Immunogenicity

### 11.1. Hemolysis, Complement Activation, Microglial Responses

Early material safety screening hinges on blood compatibility, because initial interactions with blood components often dictate downstream immune trajectories [536,537]. Hemolysis is not a benign artifact but an active trigger of innate immunity [538,539]. Cell-free heme and heme-bearing microvesicles directly activate the complement cascade, driving C3 cleavage, leukocyte activation, and cytokine release, thereby linking red blood cell damage to acute inflammatory toxicity and organ injury [538–540]. As a result, hemolysis assays are most informative when paired with measurements of complement split products and early cytokines in serum or whole-blood systems [536,540,541]. Across preclinical and clinical contexts, rising immune complexes, C3a generation, or depletion of C3 and C4 consistently correlate with infusion reactions and dose-limiting hypersensitivity, particularly during dose escalation [538,541,542].

For CNS-targeted materials, microglia and astrocytes represent a distinct and highly sensitive safety axis [111,543,544]. Complement opsonization can promote microglial uptake that is either neuroprotective or deleterious, depending on persistence and inflammatory tone [543,544]. Astrocyte-derived complement components, together with IL-1, TNF, and IL-6 signaling, shape microglial activation states and synaptic integrity [544–546]. Here, physicochemical parameters act as immune dials rather than binary switches [536]. Smaller size, higher dose, and increased positive surface charge enhance uptake and cytokine release, while excessive activation pushes glia toward chronic inflammatory phenotypes [546–548]. Early integration of these variables helps distinguish immunologically silent designs from those primed to provoke neuroimmune risk [547–549].

### 11.2. Hemocompatibility and Neuroinflammation Assays

Standardized hemocompatibility testing remains the first safety filter for blood-contacting and intravascular materials [550–552]. In vitro panels aligned with ISO 10993-4 typically assess platelet adhesion and aggregation, the intrinsic and extrinsic coagulation pathways, and complement activation, using through thrombin generation, aPTT, platelet surface markers, and C3a or C5b-9 formation [550,552,553]. Sequential whole-blood and platelet-rich plasma assays increasingly capture the cascade from protein adsorption to thrombogenicity and cytokine release, allowing mechanistic interpretation rather than binary pass-fail outcomes [550,554].

Neuroinflammation assays extend this logic into the CNS space. Human iPSC-derived microglia and astrocytes, cocultures, and emerging organoid systems enable multiplex cytokine profiling, complement C3 readouts, and neurotoxicity markers under controlled stimuli [555–557]. Critically, aligning these outputs with clinically validated biomarkers such as GFAP, IL-6, or TNF strengthens the bridge between in vitro signals and patient-level neuroinflammatory risk [558–561].

### 11.3. Biodistribution and Clearance

Biodistribution and clearance represent a central determinant of both efficacy and long-term safety for material-based therapeutics [562,563]. Following systemic administration, the majority of nanoscale materials are rapidly sequestered by the mononuclear phagocyte system, with liver and spleen often capturing most of the injected dose [564]. Uptake by Kupffer cells, splenic macrophages, and sinusoidal endothelium can markedly reduce target tissue exposure while establishing persistent intracellular reservoirs [564,565]. Such retention may remain clinically silent, yet it raises concerns under repeated dosing and complicates the interpretation of chronic toxicity risk [564,566].

Design choices strongly bias this balance between persistence and elimination [562,567]. Ultrasmall or biodegradable architectures favor renal or hepatobiliary clearance, shortening organ residence while preserving therapeutic exposure [563,568]. In contrast, larger or rigid constructs tend toward lysosomal trapping [562,569]. De-risking strategies increasingly combine biodegradable scaffolds, surface chemistry optimization, and dose fractionation to limit cumulative burden without sacrificing pharmacological performance [570–572].

### 11.4. CMC/GMP and Critical Quality Attributes (CQAs)

Critical quality attributes anchor the translation of complex materials from bench to clinic [573,574]. Across quality by design frameworks, particle size, polydispersity, zeta potential, encapsulation efficiency, and release kinetics consistently emerge as core CQAs because they integrate manufacturability with exposure and immunogenicity risk [573,575]. Multivariate and machine learning driven designs show that modest shifts in process parameters can propagate into meaningful changes in these attributes, with downstream effects on stability and biological performance [576–578] 10.3390/ijms262010238. Release profiles, often first order or diffusion controlled, are increasingly treated as quantitative CQAs rather than descriptive outcomes [573,579].

For CNS administered products, sterility and endotoxin control are nonnegotiable [580,581]. Endotoxin thresholds are substantially lower than for systemic routes, reflecting heightened neuroinflammatory sensitivity [582,583]. Routine lot release; therefore, couples validated BET or rFC assays with conservative specifications aligned with intrathecal exposure [582,584]. Stability programs add another layer of complexity [585,586]. Aggregation, content leakage, and loss of redispersibility during storage or lyophilization can silently erode CQAs unless cryoprotectants and freezing protocols are optimized [586,587].

GMP alignment ultimately depends on reproducibility [588,589]. Batch to batch fidelity in physicochemical attributes, sterility, and potency transforms CMC data from descriptive characterization into a predictive safety framework [589,590]

### 11.5. Regulatory Expectations

Regulatory agencies approach nanomedicines through a risk-based, case-by-case lens that reflects their structural diversity and evolving biology [591,592]. Both the FDA and the EMA emphasize nanomedicine-specific risks that extend beyond those of conventional small molecules, including altered biodistribution, immune activation, and long-term tissue persistence [591,593]. Guidance increasingly calls for deeper physicochemical characterization, nano-relevant immunotoxicity testing, and justification when standard ICH assays lack sensitivity [594,595]. For complex biological nanoparticle hybrids such as lipid nanoparticles, polymer conjugates, or gene delivery systems, regulators treat products as non-biological complex drugs, limiting assumptions of equivalence and requiring product-specific clinical evidence [591]

Bridging preclinical data to first-in-human studies relies on standardized safety frameworks that integrate in vitro and ex vivo human blood assays with human blood, and targeted in vivo assays [594,596]. For CNS indications, expectations tighten further [597]. BBB interactions, neuroinflammation risk, and irreversible outcomes demand a transparent risk-benefit narrative grounded in mechanistic data rather than exposure alone [595,597]. Data science tightens design loops and right-sizes risk before first dose in humans [598,599].

## 12. Data Science, Modeling, Artificial Intelligence (AI)-Guided Design

### 12.1. BBB Permeability Prediction and Polypharmacology

Data science increasingly reframes CNS design from intuition to prediction [600,601] QSAR and machine learning models trained on large BBB datasets now capture both quantitative logBB and categorical permeability with accuracy that supports early triage [600,601]. Beyond simple lipophilicity, modern models incorporate nonlinear descriptors and explicitly account for transporter effects, with P-gp emerging as a dominant determinant of CNS variability [147,602]. This is especially relevant for polyphenols, where favorable passive diffusion can be offset by strong efflux liability [147]. Transporter-aware modeling, combined with PBPK frameworks, allows permeability to be interpreted as a balance of influx and clearance rather than a static property [602,603].

AI-guided polypharmacology further expands this view [604,605]. Network-level profiling distinguishes harmful off target promiscuity from coordinated multi target engagement, enabling rational exploitation of pleiotropic mechanisms that are often intrinsic to natural products and CNS therapeutics [606,607].

### 12.2. Multi-Objective Formulation Optimization

Multi objective optimization reframes formulation design as a data driven negotiation between competing constraints [608]. Machine learning models trained on design of experiment data now predict how size, zeta potential, and drug loading jointly shape potency, exposure, stability, and manufacturability [609,610]. Rather than chasing a single optimum, Bayesian and evolutionary algorithms explore Pareto fronts, revealing trade-offs that are invisible to one-factor-at-a-time approaches [608,611]. In practice, this enables probabilistic design spaces where acceptable formulations are defined by balanced desirability rather than maximal performance [609]. Such frameworks accelerate iteration, reduce experimental burden, and align early formulation choices with downstream safety and GMP feasibility [609,610].

### 12.3. PBPK/PKPD and Digital Twins

Physiologically based pharmacokinetic modeling has become a cornerstone for forecasting CNS exposure and BBB penetration in silico [603,612]. Modern CNS PBPK platforms resolve regional brain compartments, passive permeability, and active efflux, allowing human predictions to be extrapolated from limited preclinical or in vitro data [612,613]. Coupling these frameworks to PK PD models refines dose response by linking brain time courses to target engagement and effect kinetics,

enabling virtual dose fractionation before first exposure [602]. Digital twin concepts extend this logic further [614]. By integrating PBPK, machine learning derived BBB parameters, and virtual populations, individualized predictions of permeability and response become feasible [612,614]. We synthesize the major gaps and convert them into concrete, testable strategies.

### 13. Research Gaps and Concrete Strategies

#### 13.1. Standardized Human-Relevant PK Endpoints

A clear gap is the lack of standardized, human-relevant CNS PK endpoints that translate cleanly from animals to early clinical trials. Across conceptual surveys and candidate-selection frameworks, **K<sub>p,uu,brain</sub>** repeatedly emerges as the most defensible common currency because it captures BBB transport and binding within an unbound metric. Yet human K<sub>p,uu,brain</sub> data remain sparse, and many programs still rely on total brain concentrations or non-comparable surrogates. A concrete strategy is universal adoption of K<sub>p,uu,brain</sub>, paired with PBPK-informed target-site exposure ratios that connect unbound concentrations to in vivo IC<sub>50</sub>-class benchmarks and pharmacodynamic effect.

Methodologically, the field needs harmonization of how K<sub>p,uu,brain</sub> and related endpoints are measured in humans. Combined PET plus microdialysis can convert imaging signal into unbound interstitial exposure, while mechanistic PBPK platforms can reconcile compartmental and spatial heterogeneity when CSF is unreliable. Standardized PET endpoints for exposure and engagement, together with aligned CSF metrics such as AUC, C<sub>max</sub>, and C<sub>trough</sub> normalized to potency, would make datasets interoperable. Regulatory pressure could then drive CNS drug development toward quantifiable, auditable endpoints, replacing subjective “brain penetration” claims with decision-grade measures.

#### 13.2. Humanized BBB Models with Disease Fidelity

Humanized BBB models still fall short when they trade biological realism for convenience. A priority gap is scaling iPSC derived BBB organoids, spheroids, and self assembled microvessels into **flow conditioned** systems that reproduce shear, polarization, and transport kinetics seen in vivo. Microfluidic BBB-on-a-chip platforms and perfusable 3D microvessels now achieve low paracellular permeability and strong junctional phenotypes, yet protocol and donor variability remain major sources of noise [615].

Concrete strategy: build disease fidelity into the neurovascular unit, not just the endothelium. Co culture designs that **incorporate pericytes, astrocytes, neurons, and microglia**, ideally from patient or isogenic iPSC backgrounds, capture inter-individual differences in maturation, transporter function, immune cell trafficking, and barrier breakdown [616]. The field then needs rigorous validation against human in vivo benchmarks, such as PET permeability proxies, CSF-to-plasma relationships, and clinical drug exposure patterns, so these models stop being “pretty biology” and start reducing translational attrition.

#### 13.3. Prodrug Translation Playbook

A translational prodrug playbook has to start with transporter rigor, not transporter “positive” checkboxes. For carrier mediated designs, uptake should be quantified with **K<sub>m</sub> and V<sub>max</sub>** under physiologic substrate conditions, then stress tested for competition with endogenous ligands and likely co medications [337]. Pharmacoproteomic transporter expression can anchor these kinetics to realistic barrier capacity, while time course uptake modeling helps separate true transported substrates from high affinity binders that never meaningfully cross.

Next, de-risk activation and safety in parallel. **Cleavage mapping** should quantify where and how fast the promoiety is removed across plasma, liver, brain microvessels, parenchyma, and disease relevant compartments to enforce brain first activation and avoid premature systemic unmasking. Off target profiling can be expanded beyond cell lines using tissue thermal proteome profiling or ABPP style probes across organ panels [617]. Go/no go rules then become tangible: require a brain

unbound exposure gain, a defined brain to plasma activation ratio, no dominant peripheral off target signals, and a pharmacodynamic effect that tracks brain exposure in 3D GBM models or organotypic brain slices.

#### 13.4. Long-Term Safety and Immunogenicity Registries

Long-term safety remains a blind spot when CNS trials end at symptom curves rather than at biology and latency. A concrete strategy is post-trial registries that follow participants for years, capturing delayed toxicities, immune responses, and neuroinflammation through linked EHR and claims data, structured adverse event reports, and longitudinal fluid or imaging biomarkers such as GFAP, YKL-40, sTREM2, or neuroinflammatory PET [618]. These registries should interlock with pharmacovigilance databases via standardized, FAIR data models and privacy preserving linkage, so signals can be detected, replicated, and risk managed across systems. Biologics and cell or gene therapies offer the template: mandated long follow up, harmonized reporting, and global registries that turn rare late events into quantifiable risk [618].

#### 13.5. Manufacturability and QC for Complex Carriers

A central translational gap for complex nanocarriers is that manufacturability and QC often lag behind formulation ingenuity. QbD should be treated as the organizing logic, starting with a clear QTPP and mapping CQAs to CMAs and CPPs so that a justified design space and control strategy survive scale-up [573]. Yet non-linear formulation process couplings and raw material drift still drive lot-to-lot variability, especially for surface functionalized systems where small chemistry changes reshape size, charge, corona, and bioactivity. Concrete fixes include PAT-enabled real-time monitoring (inline or online size sensing, turbidity, spectroscopy, multivariate analytics), semi-continuous or continuous lines, and tighter incoming material specifications plus stage-gated in-process controls to secure reproducible release quality [619].

#### 13.6. Clinical Trial Design Upgrades

Clinical trials for BBB therapeutics need design upgrades that treat BBB heterogeneity as a core covariate rather than background noise. Adaptive platform, basket, and window-of-opportunity approaches can rapidly prune futile delivery strategies while learning which BBB modulation, timing, and dosing actually shift brain exposure [620]. Pair this with enrichment: stratify participants by BBB integrity or permeability status using DCE MRI, PET based uptake metrics, or fluid markers reflecting barrier leakage and clearance kinetics.

Endpoints should prove target engagement, not just clinical change [621]. Imaging derived cerebral PK, longitudinal PD imaging, and permeability limited PBPK models can define exposure response relationships and justify go or no-go decisions. To satisfy regulators for high-cost, high-complexity products, adaptations and estimands must be pre specified, bias controlled, and CMC and companion diagnostics aligned early. Finally, we outline what will likely materialize soon and what needs deeper tech maturation,

## 14. Roadmap: Near-Term vs Long-Term

### 14.1. Near-Term (2–4 Years)

In the near term (2 to 4 years), the most “deployable” polyphenol programs will likely be ligand targeted PLGA nanoparticles and liposomes carrying resveratrol and curcumin, chosen because their safety narratives are mature while formulation science can add real value. Practical targets include transferrin or RVG-style ligands for BBB facing delivery, plus dual loading to exploit complementary redox and anti-inflammatory pharmacology. The translation gate is not efficacy hype, it is reproducible particle size, drug loading, and stability under scalable unit operations [162].

Intranasal mucoadhesive nanoemulsions and nanoemulgels are even closer to early-phase readiness because dosing ergonomics can be engineered into a sprayable, residence-time-extending

product. Chitosan-coated or thermotriggered in situ gel formats already map nicely onto trial-friendly endpoints: nasal tolerability, systemic exposure, and nose-to-brain PK surrogates such as regional brain concentrations in imaging-rich substudies or CSF exposure when justified [178]. Here, the CMC story must be crisp: droplet size distribution, rheology, spray plume metrics, and preservative compatibility.

LAT1 anchored prodrugs are the “biology first” option. The appeal is a validated transporter with design rules for aromatic promoieties and linker choices, enabling higher brain exposure with lower peripheral burden [336,381]. Near term success will come from leveraging known promoieties scaffolds, building a screening cascade that confirms LAT1 affinity, bioconversion kinetics, and intra brain distribution, then anchoring dose selection to target engagement readouts in neurons and glia.

Focused ultrasound-assisted regional delivery fits the same horizon when paired with drugs that already have a clinical path, such as chemotherapy for glioma margins or neuroprotectives with clean systemic safety profiles. It offers a controllable exposure window, but only if trials pre-specify imaging-based BBB opening, local PK confirmation, and safety monitoring that regulators recognize [622]. Across all these tracks, feasibility wins: scalable manufacturing, release tests that predict performance, and endpoints that prove delivery plus mechanism, not just symptomatic change [623] (Figure 4).

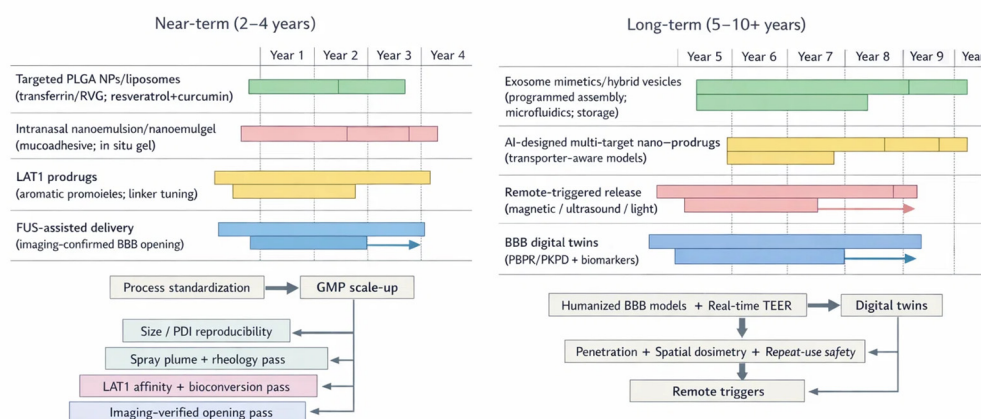
#### 14.2. Longer-Term (5–10+ Years)

Looking 5 to 10 years out, exosome mimetics and hybrid vesicles could become the “biomimetic workhorses” of brain delivery, but only if GMP-scale-up stops being artisanal. The roadmap points to programmed assembly, extrusion, and liposome fusion approaches, followed by process intensification via microfluidics and bottom-up manufacturing to reduce heterogeneity while improving yield [624]. Cost control will hinge on standardized membrane sourcing, robust cargo-loading metrics, and shelf-stable storage protocols.

In parallel, AI-designed multi target nano prodrugs may unlock rational polypharmacology, not by adding more ligands, but by learning which combinations actually cooperate at the BBB. The key upgrade is transporter awareness in silico: prediction stacks that integrate passive permeability, efflux risk, and carrier or promoieties interactions with uptake transporters [625]. Regulatory credibility will depend on curated datasets, auditable models, and prospective validation rather than retrospective fits.

Remote triggered release platforms promise precision with fewer systemic side effects. Magnetic fields [626].

Finally, patient specific BBB digital twins could connect PBPK and PKPD to biomarker-informed adaptation, turning trial dosing into a learning loop [625]. That vision depends on humanized BBB models with functional readouts, such as real-time TEER, longitudinal safety registries for complex nanomedicines, and early alignment with regulators on what counts as validated exposure and engagement evidence [627] (Figure 4).



**Figure 4.** Roadmap timeline for brain delivery platforms: near-term deployables versus long-term maturation, with validation gates and dependencies. Gantt-style bands summarize expected maturation windows for CNS delivery strategies and the dependencies that determine whether programs advance from feasibility to translation. Near-term (2–4 years) tracks emphasize “deployable” options with the clearest CMC and endpoint pathways: ligand-targeted PLGA nanoparticles/liposomes (e.g., transferrin or RVG; resveratrol/curcumin, including dual loading) gated by reproducible size/PDI, loading, and stability under scalable unit operations; intranasal mucoadhesive nanoemulsions/nanoemulgels (chitosan or thermo/in situ gels) gated by droplet-size distribution, rheology, spray plume metrics, preservative compatibility, and tolerability plus PK surrogates (CSF or imaging-rich substudies where justified); LAT1-anchored prodrugs gated by LAT1 affinity, bioconversion kinetics, intrabrain distribution, and target engagement in neurons/glia; and focused ultrasound–assisted regional delivery gated by imaging-confirmed BBB opening, local PK confirmation, and regulator-recognized safety monitoring. Longer-term (5–10+ years) bands capture exosome mimetics/hybrid vesicles (GMP scale-up and heterogeneity control), AI-designed multi-target nano-prodrugs (auditable datasets and prospective validation), remote-triggered release (dosimetry and repeat-use safety standards), and BBB “digital twins” integrating PBPK/PKPD with real-time TEER and registries. AI, artificial intelligence; BBB, blood–brain barrier; CMC, chemistry, manufacturing, and controls; CNS, central nervous system; CSF, cerebrospinal fluid; GMP, good manufacturing practice; LAT1, large neutral amino acid transporter 1; PBPK, physiologically based pharmacokinetic; PDI, polydispersity index; PK, pharmacokinetics; PKPD, pharmacokinetic–pharmacodynamic; PLGA, poly(lactic-co-glycolic acid); RVG, rabies virus glycoprotein; TEER, transendothelial electrical resistance.

## 15. Clinical Applications and Translational Implications

The concepts synthesized in this study have direct clinical relevance for the development of next-generation neurotherapeutics targeting disorders with high unmet medical need, including neurodegenerative diseases, neuropsychiatric conditions, epilepsy, and brain tumors [628]. By systematically linking the physicochemical limitations of plant-derived compounds to rational delivery solutions—such as nanocarriers, transporter-targeted prodrugs, intranasal administration, and device-enabled BBB modulation—this framework provides actionable guidance for improving CNS drug exposure where conventional pharmacotherapy has failed [4]. In clinical contexts characterized by multifactorial pathophysiology, such as Alzheimer’s disease, Parkinson’s disease, depression, and chronic pain, phytochemicals with pleiotropic anti-inflammatory, antioxidant, and

neuromodulatory actions may offer therapeutic advantages if reliable brain delivery can be achieved[629].

From a translational standpoint, the study supports a shift away from empiric compound selection toward delivery-first clinical development, in which candidate molecules are paired early with route, carrier, or prodrug strategies to achieve decision-grade CNS exposure [67]. Clinically, this approach may enable dose reduction, improved safety margins, and more predictable pharmacokinetics, particularly in vulnerable populations such as older adults or patients receiving polypharmacy [630]. Furthermore, the discussed platforms—especially intranasal delivery and focused ultrasound-mediated BBB opening—offer opportunities for region-specific or noninvasive treatment paradigms, which are increasingly relevant in precision neurology and psychiatry [631]. Collectively, these insights inform the design of early-phase clinical trials, guide biomarker and endpoint selection, and support regulatory-aligned translation of plant-derived neurotherapeutics from bench to bedside [632].

## 15. Conclusion and Translational Implications

RMT targeting via. Intranasal delivery enables potent, stable payloads to exploit the nose-to-brain pathways [155]. Focused ultrasound with microbubbles provides a reversible, local window of BBB opening [633]. Exosome mimetics and nano prodrugs stay promising but remain unproven. As authors, we argue that translation will speed up once Kp,uu,brain centric workflows become the common language across teams. The take home message is simple: treat delivery as a coupled system where chemistry, carrier, route, and measurement are co designed. Start with a liability map for each phytochemical, then choose the simplest strategy that can raise unbound brain exposure while meeting safety and chemistry, manufacturing, and controls constraints. Report fu,brain, unbound plasma fractions, and Kp,uu,brain, then connect them to target engagement, imaging, and functional outcomes. Future research should deliver head-to-head platform comparisons on identical payloads, validate human-relevant BBB models against in vivo benchmarks, and build PBPK guided dose projections that survive species shifts. Methodologically, the field also needs harmonized critical quality attributes and longitudinal safety panels that capture complement activation, microglial priming, and vascular repair. Done right, these principles can accelerate barrier-limited therapeutics far beyond phytochemicals.

**Author Contributions:** Conceptualization, M.T. and S.M.B.; methodology, A.C.A., E.L.G., V.E.V., R.S.A.H., R.A.G.; investigation; writing—original draft preparation, M.T., V.C.S.C., C.M.G., E.S.B.M.P., A.C.A.C.; writing—review and editing, M.T., V.E.V., S.M.B; visualization, M.T.; supervision, M.T. and S.M.B.; project administration, M.T. and S.M.B.; funding acquisition, M.T. and S.M.B. All authors have read and agreed to the published version of the manuscript.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** No new data were created or analyzed in this study.

**Acknowledgments:** The authors have reviewed and edited the output and take full responsibility for the content of this publication.

**Conflicts of Interest:** The authors declare no conflicts of interest.

## Abbreviations

The following abbreviations are used in this manuscript:

ABC	ATP-binding cassette
ApoE	apolipoprotein E

AUC	area under the curve
BBB	blood-brain barrier
BCRP	breast cancer resistance protein
CBD	cannabidiol
CED	convection-enhanced delivery
CMC	chemistry, manufacturing, and controls
CMT	carrier-mediated transport
CNS	central nervous system
CPP	cell-penetrating peptide
CQA	critical quality attribute
CSF	cerebrospinal fluid
CYP	cytochrome P450
DOI	digital object identifier
EVS	extracellular vesicles
FUS	focused ultrasound
GDNF	glial-derived neurotrophic factor
GLUT1	glucose transporter 1
GMP	good manufacturing practice
IPSC	induced pluripotent stem cell
IR	insulin receptor
ISF	interstitial fluid
KM	michaelis constant
KP	uu, unbound brain-to-plasma partition coefficient
KP,BRAIN	brain-to-plasma partition coefficient
KP,UU	unbound brain-to-plasma partition coefficient
K_D	dissociation constant
K_P,BRAIN	brain-to-plasma partition coefficient
K_P,UU	unbound brain-to-plasma partition coefficient
LAT1	large neutral amino acid transporter 1
LD	linear dichroism
LDL	low-density lipoprotein
LDLR	low-density lipoprotein receptor
LOGD	distribution coefficient
LRP1	low-density lipoprotein receptor-related protein 1
MCT1	monocarboxylate transporter 1
MCTS	monocarboxylate transporters
MRI	magnetic resonance imaging
MRPS	multidrug resistance-associated proteins
NHP	non-human primate
NLCS	nanostructured lipid carriers
NPS	nanoparticles
NR	not reported

NVU	neurovascular unit
P-gp	P-glycoprotein
PAMAM	poly(amidoamine)
PAT	process analytical technology
PBPK	physiologically based pharmacokinetic
PCD	passive cavitation detection
PD	pharmacodynamics
PDEVS	plant-derived extracellular vesicles
PDI	polydispersity index
PEG	polyethylene glycol
PET	positron emission tomography
PK	pharmacokinetics
PKA	acid dissociation constant
PKPD	pharmacokinetic-pharmacodynamic
PLGA	poly(lactic-co-glycolic acid)
QBD	quality by design
QTPP	quality target product profile
RESVERATROL/CURCUMIN	including dual loading) gated by reproducible size/PDI, loading, and stability under scalable unit operations
RMT	receptor-mediated transcytosis
RVG	rabies virus glycoprotein
SIRNA	small interfering RNA
SLCS	solute carrier transporters
SLNS	solid lipid nanoparticles
SULT	sulfotransferases
TAT	trans-activator of transcription
TEER	transendothelial electrical resistance
TF	transferrin
TFR	transferrin receptor
THC	$\Delta^9$ -tetrahydrocannabinol
UGT	UDP-glucuronosyltransferases
VEGF	vascular endothelial growth factor
VMAX	maximum transport rate

## References

1. Global, regional, and national burden of disorders affecting the nervous system, 1990-2021: a systematic analysis for the Global Burden of Disease Study 2021. *Lancet Neurol* **2024**, *23*, 344-381, doi:10.1016/s1474-4422(24)00038-3.
2. Arias, D.; Saxena, S.; Verguet, S. Quantifying the global burden of mental disorders and their economic value. *EclinicalMedicine* **2022**, *54*, 101675, doi:10.1016/j.eclinm.2022.101675.
3. Tanaka, M. Parkinson's Disease: Bridging Gaps, Building Biomarkers, and Reimagining Clinical Translation. *Cells* **2025**, *14*, doi:10.3390/cells14151161.
4. Wu, D.; Chen, Q.; Chen, X.; Han, F.; Chen, Z.; Wang, Y. The blood-brain barrier: structure, regulation, and drug delivery. *Signal Transduct Target Ther* **2023**, *8*, 217, doi:10.1038/s41392-023-01481-w.

5. Gribkoff, V.K.; Kaczmarek, L.K. The need for new approaches in CNS drug discovery: Why drugs have failed, and what can be done to improve outcomes. *Neuropharmacology* **2017**, *120*, 11-19, doi:10.1016/j.neuropharm.2016.03.021.
6. Howe, J.R.t.; Bear, M.F.; Golshani, P.; Klann, E.; Lipton, S.A.; Mucke, L.; Sahin, M.; Silva, A.J. The mouse as a model for neuropsychiatric drug development. *Curr Biol* **2018**, *28*, R909-r914, doi:10.1016/j.cub.2018.07.046.
7. Tanaka, M. From Monoamines to Systems Psychiatry: Rewiring Depression Science and Care (1960s–2025). *Biomedicines* **2025**, *14*, 35.
8. Naoi, M.; Wu, Y.; Maruyama, W.; Shamoto-Nagai, M. Phytochemicals Modulate Biosynthesis and Function of Serotonin, Dopamine, and Norepinephrine for Treatment of Monoamine Neurotransmission-Related Psychiatric Diseases. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26072916.
9. Chen, X.; Xu, D.; Yu, J.; Song, X.J.; Li, X.; Cui, Y.L. Tryptophan Metabolism Disorder-Triggered Diseases, Mechanisms, and Therapeutic Strategies: A Scientometric Review. *Nutrients* **2024**, *16*, doi:10.3390/nu16193380.
10. Huang, Y.; Zhao, M.; Chen, X.; Zhang, R.; Le, A.; Hong, M.; Zhang, Y.; Jia, L.; Zang, W.; Jiang, C.; et al. Tryptophan Metabolism in Central Nervous System Diseases: Pathophysiology and Potential Therapeutic Strategies. *Aging Dis* **2023**, *14*, 858-878, doi:10.14336/ad.2022.0916.
11. Szabó, Á.; Galla, Z.; Spekker, E.; Martos, D.; Szűcs, M.; Fejes-Szabó, A.; Fehér, Á.; Takeda, K.; Ozaki, K.; Inoue, H.; et al. Behavioral Balance in Tryptophan Turmoil: Regional Metabolic Rewiring in Kynurenine Aminotransferase II Knockout Mice. *Cells* **2025**, *14*, doi:10.3390/cells14211711.
12. Liu, Y.; Chen, Z.; Li, A.; Liu, R.; Yang, H.; Xia, X. The Phytochemical Potential for Brain Disease Therapy and the Possible Nanodelivery Solutions for Brain Access. *Front Oncol* **2022**, *12*, 936054, doi:10.3389/fonc.2022.936054.
13. Mishra, K.; Rana, R.; Tripathi, S.; Siddiqui, S.; Yadav, P.K.; Yadav, P.N.; Chourasia, M.K. Recent Advancements in Nanocarrier-assisted Brain Delivery of Phytochemicals Against Neurological Diseases. *Neurochem Res* **2023**, *48*, 2936-2968, doi:10.1007/s11064-023-03955-3.
14. Rassu, G.; Sorrenti, M.; Catenacci, L.; Pavan, B.; Ferraro, L.; Gavini, E.; Bonferoni, M.C.; Giunchedi, P.; Dalpiaz, A. Conjugation, Prodrug, and Co-Administration Strategies in Support of Nanotechnologies to Improve the Therapeutic Efficacy of Phytochemicals in the Central Nervous System. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15061578.
15. Chehadi, A.C.; Pereira de Lima, E.; Detregiachi, C.R.P.; Santos de Argollo Haber, R.; Catharin, V.; Fornari Laurindo, L.; Engracia Valenti, V.; Machado Galhardi, C.; Tanaka, M.; Maria Barbalho, S. Harnessing Dietary Tryptophan: Bridging the Gap Between Neurobiology and Psychiatry in Depression Management. *Int J Mol Sci* **2026**, *27*, doi:10.3390/ijms27010465.
16. Endo, H.; Ogasawara, M.; Tega, Y.; Kubo, Y.; Hosoya, K.I.; Akanuma, S.I. Upregulation of P-Glycoprotein and Breast Cancer Resistance Protein Activity in Newly Developed in Vitro Rat Blood-Brain Barrier Spheroids Using Advanced Glycation End-Products. *Biol Pharm Bull* **2024**, *47*, 1893-1903, doi:10.1248/bpb.b24-00481.
17. Strazielle, N.; Ghersi-Egea, J.F. Factors affecting delivery of antiviral drugs to the brain. *Rev Med Virol* **2005**, *15*, 105-133, doi:10.1002/rmv.454.
18. Waterhouse, R.N. Determination of lipophilicity and its use as a predictor of blood-brain barrier penetration of molecular imaging agents. *Mol Imaging Biol* **2003**, *5*, 376-389, doi:10.1016/j.mibio.2003.09.014.
19. Ayaz, M.; Mosa, O.F.; Nawaz, A.; Hamdoon, A.A.E.; Elkhalfifa, M.E.M.; Sadiq, A.; Ullah, F.; Ahmed, A.; Kabra, A.; Khan, H.; et al. Neuroprotective potentials of Lead phytochemicals against Alzheimer's disease with focus on oxidative stress-mediated signaling pathways: Pharmacokinetic challenges, target specificity, clinical trials and future perspectives. *Phytomedicine* **2024**, *124*, 155272, doi:10.1016/j.phymed.2023.155272.
20. Lehoczki, A.; Fekete, M.; Jarecsny, T.; Zábó, V.; Szappanos, Á.; Csípó, T.; Lipécz, Á.; Major, D.; Fazekas-Pongor, V.; Varga, P.; et al. The Neuroprotective Role of Curcumin: From Molecular Pathways to Clinical Translation-A Narrative Review. *Nutrients* **2025**, *17*, doi:10.3390/nu17172884.

21. Nunes, Y.C.; Mendes, N.M.; Pereira de Lima, E.; Chehadi, A.C.; Lamas, C.B.; Haber, J.F.S.; Dos Santos Bueno, M.; Araújo, A.C.; Catharin, V.C.S.; Detregiachi, C.R.P.; et al. Curcumin: A Golden Approach to Healthy Aging: A Systematic Review of the Evidence. *Nutrients* **2024**, *16*, doi:10.3390/nu16162721.
22. Laurindo, L.F.; de Carvalho, G.M.; de Oliveira Zanuso, B.; Figueira, M.E.; Direito, R.; de Alvares Goulart, R.; Buglio, D.S.; Barbalho, S.M. Curcumin-Based Nanomedicines in the Treatment of Inflammatory and Immunomodulated Diseases: An Evidence-Based Comprehensive Review. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15010229.
23. Marton, L.T.; Barbalho, S.M.; Sloan, K.P.; Sloan, L.A.; Goulart, R.A.; Araújo, A.C.; Bechara, M.D. Curcumin, autoimmune and inflammatory diseases: going beyond conventional therapy - a systematic review. *Crit Rev Food Sci Nutr* **2022**, *62*, 2140-2157, doi:10.1080/10408398.2020.1850417.
24. Matias, J.N.; Achete, G.; Campanari, G.; Guiguer, É, L.; Araújo, A.C.; Buglio, D.S.; Barbalho, S.M. A systematic review of the antidepressant effects of curcumin: Beyond monoamines theory. *Aust N Z J Psychiatry* **2021**, *55*, 451-462, doi:10.1177/0004867421998795.
25. Kaspute, G.; Ramanavicius, A.; Prentice, U. Natural drug delivery systems for the treatment of neurodegenerative diseases. *Mol Biol Rep* **2025**, *52*, 217, doi:10.1007/s11033-025-10286-9.
26. Sánchez-Martínez, J.D.; Valdés, A.; Gallego, R.; Suárez-Montenegro, Z.J.; Alarcón, M.; Ibañez, E.; Alvarez-Rivera, G.; Cifuentes, A. Blood-Brain Barrier Permeability Study of Potential Neuroprotective Compounds Recovered From Plants and Agri-Food by-Products. *Front Nutr* **2022**, *9*, 924596, doi:10.3389/fnut.2022.924596.
27. Shimazu, R.; Anada, M.; Miyaguchi, A.; Nomi, Y.; Matsumoto, H. Evaluation of Blood-Brain Barrier Permeability of Polyphenols, Anthocyanins, and Their Metabolites. *J Agric Food Chem* **2021**, *69*, 11676-11686, doi:10.1021/acs.jafc.1c02898.
28. Musa, I.; Rotaru-Zavaleanu, A.D.; Sfredel, V.; Aldea, M.; Gresita, A.; Glavan, D.G. Post-Stroke Recovery: A Review of Hydrogel-Based Phytochemical Delivery Systems. *Gels* **2025**, *11*, doi:10.3390/gels11040260.
29. Tanaka, M.; Vécsei, L. From Microbial Switches to Metabolic Sensors: Rewiring the Gut-Brain Kynurenine Circuit. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13082020.
30. Figueiredo Godoy, A.C.; Frota, F.F.; Araújo, L.P.; Valenti, V.E.; Pereira, E.; Detregiachi, C.R.P.; Galhardi, C.M.; Caracio, F.C.; Haber, R.S.A.; Fornari Laurindo, L.; et al. Neuroinflammation and Natural Antidepressants: Balancing Fire with Flora. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13051129.
31. Karad, V.; Gupta, G.L. Phytochemicals encouraging neurotrophic pathways: brain-derived neurotrophic factors as molecular targets in depression. *Naunyn Schmiedebergs Arch Pharmacol* **2025**, *398*, 15075-15094, doi:10.1007/s00210-025-04298-2.
32. Yang, H.M. Overcoming the Blood-Brain Barrier: Advanced Strategies in Targeted Drug Delivery for Neurodegenerative Diseases. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17081041.
33. Zha, S.; Liu, H.; Li, H.; Li, H.; Wong, K.L.; All, A.H. Functionalized Nanomaterials Capable of Crossing the Blood-Brain Barrier. *ACS Nano* **2024**, *18*, 1820-1845, doi:10.1021/acsnano.3c10674.
34. Ali, S.; Ali, S.A.; Kumar, M.; Jahan, I.; Hak, J. Emerging Strategies for Targeted Drug Delivery across the Blood-Brain Barrier in Neurological Disorder. *Current Pharmaceutical Research* **2025**, 1-14.
35. Zou, L.; Chien, H.C.; Pade, D.; Li, Y.; Nguyen, M.; Bhamidipati, R.K.; Wang, Z.; Enogieru, O.J.; Wahlstrom, J. Considerations in K(p,uu,brain)-based Strategy for Selecting CNS-targeted Drug Candidates with Sufficient Target Coverage and Substantial Pharmacodynamic Effect. *Aaps j* **2025**, *27*, 52, doi:10.1208/s12248-025-01035-8.
36. Gonen, O.M.; Porter, T.; Wang, B.; Xue, F.; Ma, Y.; Song, L.; Sun, P.; Fan, W.; Shen, Y. Safety, Pharmacokinetics and Target Engagement of a Novel Brain Penetrant RIPK1 Inhibitor (SIR9900) in Healthy Adults and Elderly Participants. *Clin Transl Sci* **2025**, *18*, e70151, doi:10.1111/cts.70151.
37. Tanaka, M.; Szatmári, I.; Vécsei, L. Quinoline Quest: Kynurenine Acid Strategies for Next-Generation Therapeutics via Rational Drug Design. *Pharmaceutics (Basel)* **2025**, *18*, doi:10.3390/ph18050607.
38. Gong, Y.; Wu, M.; Huang, Y.; He, X.; Yuan, J.; Dang, B. Research developments in the neurovascular unit and the blood-brain barrier (Review). *Biomed Rep* **2025**, *22*, 88, doi:10.3892/br.2025.1966.
39. McConnell, H.L.; Mishra, A. Cells of the Blood-Brain Barrier: An Overview of the Neurovascular Unit in Health and Disease. *Methods Mol Biol* **2022**, *2492*, 3-24, doi:10.1007/978-1-0716-2289-6\_1.

40. Schaeffer, S.; Iadecola, C. Revisiting the neurovascular unit. *Nat Neurosci* **2021**, *24*, 1198-1209, doi:10.1038/s41593-021-00904-7.
41. Gong, Y.; Wu, M.; Huang, Y.; He, X.; Yuan, J.; Dang, B. Research developments in the neurovascular unit and the blood-brain barrier. *Biomedical Reports* **2025**, *22*, 88.
42. Divecha, Y.A.; Rampes, S.; Tromp, S.; Boyanova, S.T.; Fleckney, A.; Fidanboyulu, M.; Thomas, S.A. The microcirculation, the blood-brain barrier, and the neurovascular unit in health and Alzheimer disease: The aberrant pericyte is a central player. *Pharmacol Rev* **2025**, *77*, 100052, doi:10.1016/j.pharmr.2025.100052.
43. Kugler, E.C.; Greenwood, J.; MacDonald, R.B. The "Neuro-Glial-Vascular" Unit: The Role of Glia in Neurovascular Unit Formation and Dysfunction. *Front Cell Dev Biol* **2021**, *9*, 732820, doi:10.3389/fcell.2021.732820.
44. Sasson, E.; Anzi, S.; Bell, B.; Yakovian, O.; Zorsky, M.; Deutsch, U.; Engelhardt, B.; Sherman, E.; Vatine, G.; Dzikowski, R. Nano-scale architecture of blood-brain barrier tight-junctions. *Elife* **2021**, *10*, e63253.
45. Alluri, H.; Peddaboina, C.S.; Tharakan, B. Evaluation of Tight Junction Integrity in Brain Endothelial Cells Using Confocal Microscopy. *Methods Mol Biol* **2024**, *2711*, 257-262, doi:10.1007/978-1-0716-3429-5\_21.
46. Gowrikumar, S.; Tarudji, A.; McDonald, B.Z.; Balusa, S.S.; Kievit, F.M.; Dhawan, P. Claudin-1 impairs blood-brain barrier by downregulating endothelial junctional proteins in traumatic brain injury. *Tissue Barriers* **2025**, *13*, 2470482, doi:10.1080/21688370.2025.2470482.
47. Maiuolo, J.; Gliozzi, M.; Musolino, V.; Scicchitano, M.; Carresi, C.; Scarano, F.; Bosco, F.; Nucera, S.; Ruga, S.; Zito, M.C.; et al. The "Frail" Brain Blood Barrier in Neurodegenerative Diseases: Role of Early Disruption of Endothelial Cell-to-Cell Connections. *Int J Mol Sci* **2018**, *19*, doi:10.3390/ijms19092693.
48. Potjewyd, G.; Moxon, S.; Wang, T.; Domingos, M.; Hooper, N.M. Tissue Engineering 3D Neurovascular Units: A Biomaterials and Bioprinting Perspective. *Trends Biotechnol* **2018**, *36*, 457-472, doi:10.1016/j.tibtech.2018.01.003.
49. Mãe, M.A.; He, L.; Nordling, S.; Vazquez-Liebanas, E.; Nahar, K.; Jung, B.; Li, X.; Tan, B.C.; Chin Foo, J.; Cazenave-Gassiot, A.; et al. Single-Cell Analysis of Blood-Brain Barrier Response to Pericyte Loss. *Circ Res* **2021**, *128*, e46-e62, doi:10.1161/circresaha.120.317473.
50. Guérit, S.; Fidan, E.; Macas, J.; Czupalla, C.J.; Figueiredo, R.; Vijikumar, A.; Yalcin, B.H.; Thom, S.; Winter, P.; Gerhardt, H.; et al. Astrocyte-derived Wnt growth factors are required for endothelial blood-brain barrier maintenance. *Prog Neurobiol* **2021**, *199*, 101937, doi:10.1016/j.pneurobio.2020.101937.
51. Tanaka, M. Neurogenesis and Neuroinflammation in Dialogue: Mapping Gaps, Modulating Microglia, Rewiring Aging. *Cells* **2026**, *15*, doi:10.3390/cells15010078.
52. Reed, M.J.; Damodarasamy, M.; Banks, W.A. The extracellular matrix of the blood-brain barrier: structural and functional roles in health, aging, and Alzheimer's disease. *Tissue Barriers* **2019**, *7*, 1651157, doi:10.1080/21688370.2019.1651157.
53. Ding, L.; Kshirsagar, P.; Agrawal, P.; Murry, D.J. Crossing the Blood-Brain Barrier: Innovations in Receptor- and Transporter-Mediated Transcytosis Strategies. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17060706.
54. Terstappen, G.C.; Meyer, A.H.; Bell, R.D.; Zhang, W. Strategies for delivering therapeutics across the blood-brain barrier. *Nat Rev Drug Discov* **2021**, *20*, 362-383, doi:10.1038/s41573-021-00139-y.
55. Pardridge, W.M. Transport of small molecules through the blood-brain barrier: biology and methodology. *Advanced drug delivery reviews* **1995**, *15*, 5-36.
56. Bao, X.; Wu, J.; Xie, Y.; Kim, S.; Michelhaugh, S.; Jiang, J.; Mittal, S.; Sanai, N.; Li, J. Protein Expression and Functional Relevance of Efflux and Uptake Drug Transporters at the Blood-Brain Barrier of Human Brain and Glioblastoma. *Clin Pharmacol Ther* **2020**, *107*, 1116-1127, doi:10.1002/cpt.1710.
57. Latif, S.; Kang, Y.S. Blood-Brain Barrier Solute Carrier Transporters and Motor Neuron Disease. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14102167.
58. Tashima, T. Smart Strategies for Therapeutic Agent Delivery into Brain across the Blood-Brain Barrier Using Receptor-Mediated Transcytosis. *Chem Pharm Bull (Tokyo)* **2020**, *68*, 316-325, doi:10.1248/cpb.c19-00854.
59. Stanimirovic, D.B.; Sandhu, J.K.; Costain, W.J. Emerging Technologies for Delivery of Biotherapeutics and Gene Therapy Across the Blood-Brain Barrier. *BioDrugs* **2018**, *32*, 547-559, doi:10.1007/s40259-018-0309-y.

60. Kucharz, K.; Kristensen, K.; Johnsen, K.B.; Lund, M.A.; Lønstrup, M.; Moos, T.; Andresen, T.L.; Lauritzen, M.J. Post-capillary venules are the key locus for transcytosis-mediated brain delivery of therapeutic nanoparticles. *Nat Commun* **2021**, *12*, 4121, doi:10.1038/s41467-021-24323-1.
61. Hersh, A.M.; Alomari, S.; Tyler, B.M. Crossing the Blood-Brain Barrier: Advances in Nanoparticle Technology for Drug Delivery in Neuro-Oncology. *Int J Mol Sci* **2022**, *23*, doi:10.3390/ijms23084153.
62. Lochhead, J.J.; Yang, J.; Ronaldson, P.T.; Davis, T.P. Structure, Function, and Regulation of the Blood-Brain Barrier Tight Junction in Central Nervous System Disorders. *Front Physiol* **2020**, *11*, 914, doi:10.3389/fphys.2020.00914.
63. Keaney, J.; Walsh, D.M.; O'Malley, T.; Hudson, N.; Crosbie, D.E.; Loftus, T.; Sheehan, F.; McDaid, J.; Humphries, M.M.; Callanan, J.J.; et al. Autoregulated paracellular clearance of amyloid- $\beta$  across the blood-brain barrier. *Sci Adv* **2015**, *1*, e1500472, doi:10.1126/sciadv.1500472.
64. Nozohouri, E.; Noorani, B.; Patel, D.; Ahn, Y.; Zoubi, S.; Bickel, U. Assessing blood-brain barrier (BBB) integrity in an Alzheimer's disease mouse model: is the BBB globally or locally disrupted? *Fluids Barriers CNS* **2025**, *22*, 79, doi:10.1186/s12987-025-00685-2.
65. Cornelissen, F.M.G.; Markert, G.; Deutsch, G.; Antonara, M.; Faaij, N.; Bartelink, I.; Noske, D.; Vandertop, W.P.; Bender, A.; Westerman, B.A. Explaining Blood-Brain Barrier Permeability of Small Molecules by Integrated Analysis of Different Transport Mechanisms. *J Med Chem* **2023**, *66*, 7253-7267, doi:10.1021/acs.jmedchem.2c01824.
66. Khalil, A.; Barras, A.; Boukherroub, R.; Tseng, C.L.; Devos, D.; Burnouf, T.; Neuhaus, W.; Szunerits, S. Enhancing paracellular and transcellular permeability using nanotechnological approaches for the treatment of brain and retinal diseases. *Nanoscale Horiz* **2023**, *9*, 14-43, doi:10.1039/d3nh00306j.
67. Nájera-Maldonado, L.; Parra-González, M.; Peralta-Cuevas, E.; Gutierrez-Onofre, A.J.; Garcia-Atutxa, I.; Villanueva-Flores, F. Cracking the Blood-Brain Barrier Code: Rational Nanomaterial Design for Next-Generation Neurological Therapies. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17091169.
68. Smith, Q.R. Carrier-mediated transport to enhance drug delivery to brain. In Proceedings of the International Congress Series, 2005; pp. 63-74.
69. Özgür, B.; Puris, E.; Brachner, A.; Appelt-Menzel, A.; Oerter, S.; Balzer, V.; Holst, M.R.; Christiansen, R.F.; Hyldig, K.; Buckley, S.T.; et al. Characterization of an iPSC-based barrier model for blood-brain barrier investigations using the SBAD0201 stem cell line. *Fluids Barriers CNS* **2023**, *20*, 96, doi:10.1186/s12987-023-00501-9.
70. Li, Y.; Liu, R.; Zhao, Z. Targeting Brain Drug Delivery with Macromolecules Through Receptor-Mediated Transcytosis. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17010109.
71. Hervé, F.; Ghinea, N.; Scherrmann, J.M. CNS delivery via adsorptive transcytosis. *Aaps j* **2008**, *10*, 455-472, doi:10.1208/s12248-008-9055-2.
72. Baghirov, H. Receptor-mediated transcytosis of macromolecules across the blood-brain barrier. *Expert Opin Drug Deliv* **2023**, *20*, 1699-1711, doi:10.1080/17425247.2023.2255138.
73. Pfau, S.J.; Langen, U.H.; Fisher, T.M.; Prakash, I.; Nagpurwala, F.; Lozoya, R.A.; Lee, W.A.; Wu, Z.; Gu, C. Characteristics of blood-brain barrier heterogeneity between brain regions revealed by profiling vascular and perivascular cells. *Nat Neurosci* **2024**, *27*, 1892-1903, doi:10.1038/s41593-024-01743-y.
74. Noumbissi, M.E.; Galasso, B.; Stins, M.F. Brain vascular heterogeneity: implications for disease pathogenesis and design of in vitro blood-brain barrier models. *Fluids Barriers CNS* **2018**, *15*, 12, doi:10.1186/s12987-018-0097-2.
75. Beltran-Velasco, A.I.; Clemente-Suárez, V.J. Impact of Peripheral Inflammation on Blood-Brain Barrier Dysfunction and Its Role in Neurodegenerative Diseases. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26062440.
76. Wilhelm, I.; Nyúl-Tóth, Á.; Suciú, M.; Hermenean, A.; Krizbai, I.A. Heterogeneity of the blood-brain barrier. *Tissue Barriers* **2016**, *4*, e1143544, doi:10.1080/21688370.2016.1143544.
77. Ha, I.H.; Lim, C.; Kim, Y.; Moon, Y.; Han, S.H.; Moon, W.J. Regional Differences in Blood-Brain Barrier Permeability in Cognitively Normal Elderly Subjects: A Dynamic Contrast-Enhanced MRI-Based Study. *Korean J Radiol* **2021**, *22*, 1152-1162, doi:10.3348/kjr.2020.0816.

78. Schaffenrath, J.; Huang, S.F.; Wyss, T.; Delorenzi, M.; Keller, A. Characterization of the blood-brain barrier in genetically diverse laboratory mouse strains. *Fluids Barriers CNS* **2021**, *18*, 34, doi:10.1186/s12987-021-00269-w.
79. Kurz, C.; Walker, L.; Rauchmann, B.S.; Perneczky, R. Dysfunction of the blood-brain barrier in Alzheimer's disease: Evidence from human studies. *Neuropathol Appl Neurobiol* **2022**, *48*, e12782, doi:10.1111/nan.12782.
80. Bravo-Ferrer, I.; Gaasdal-Bech, K.; Colvin, C.; Vaughan, H.J.; Moss, J.; Williams, A.; Díaz Castro, B. Multiregional blood-brain barrier phenotyping identifies the prefrontal cortex as the most vulnerable region to ageing in mice. *Brain Commun* **2025**, *7*, fcaf332, doi:10.1093/braincomms/fcaf332.
81. Chen, T.; Dai, Y.; Hu, C.; Lin, Z.; Wang, S.; Yang, J.; Zeng, L.; Li, S.; Li, W. Cellular and molecular mechanisms of the blood-brain barrier dysfunction in neurodegenerative diseases. *Fluids Barriers CNS* **2024**, *21*, 60, doi:10.1186/s12987-024-00557-1.
82. de Lima, E.P.; Tanaka, M.; Lamas, C.B.; Quesada, K.; Detregiachi, C.R.P.; Araújo, A.C.; Guiguer, E.L.; Catharin, V.; de Castro, M.V.M.; Junior, E.B.; et al. Vascular Impairment, Muscle Atrophy, and Cognitive Decline: Critical Age-Related Conditions. *Biomedicines* **2024**, *12*, doi:10.3390/biomedicines12092096.
83. Bhattacharya, T.; Soares, G.; Chopra, H.; Rahman, M.M.; Hasan, Z.; Swain, S.S.; Cavalu, S. Applications of Phyto-Nanotechnology for the Treatment of Neurodegenerative Disorders. *Materials (Basel)* **2022**, *15*, doi:10.3390/ma15030804.
84. Kirit, E.; Gokce, C.; Altun, B.; Yilmazer, A. Nanotherapeutic Strategies for Overcoming the Blood-Brain Barrier: Applications in Disease Modeling and Drug Delivery. *ACS Omega* **2025**, *10*, 32606-32625, doi:10.1021/acsomega.5c02206.
85. Yuan, S.; Hu, D.; Gao, D.; Butch, C.J.; Wang, Y.; Zheng, H.; Sheng, Z. Recent advances of engineering cell membranes for nanomedicine delivery across the blood-brain barrier. *J Nanobiotechnology* **2025**, *23*, 493, doi:10.1186/s12951-025-03572-y.
86. Pedder, J.H.; Sonabend, A.M.; Cearns, M.D.; Michael, B.D.; Zakaria, R.; Heimberger, A.B.; Jenkinson, M.D.; Dickens, D. Crossing the blood-brain barrier: emerging therapeutic strategies for neurological disease. *Lancet Neurol* **2025**, *24*, 246-260, doi:10.1016/s1474-4422(24)00476-9.
87. Dirir, A.M.; Ali, A.; Hachem, M. Recent Advancements in Lipid Nanoparticles-Based Phytoactives Delivery Systems for Neurodegenerative Diseases. *Int J Nanomedicine* **2025**, *20*, 10279-10300, doi:10.2147/ijn.S537566.
88. Kamath, A.P.; Nayak, P.G.; John, J.; Mutalik, S.; Balaraman, A.K.; Krishnadas, N. Revolutionizing neurotherapeutics: Nanocarriers unveiling the potential of phytochemicals in Alzheimer's disease. *Neuropharmacology* **2024**, *259*, 110096, doi:10.1016/j.neuropharm.2024.110096.
89. de Lima, E.P.; Laurindo, L.F.; Catharin, V.C.S.; Direito, R.; Tanaka, M.; Jasmin Santos German, I.; Lamas, C.B.; Guiguer, E.L.; Araújo, A.C.; Fiorini, A.M.R.; et al. Polyphenols, Alkaloids, and Terpenoids Against Neurodegeneration: Evaluating the Neuroprotective Effects of Phytocompounds Through a Comprehensive Review of the Current Evidence. *Metabolites* **2025**, *15*, doi:10.3390/metabo15020124.
90. Laurindo, L.F.; Santos, A.; Carvalho, A.C.A.; Bechara, M.D.; Guiguer, E.L.; Goulart, R.A.; Vargas Sinatora, R.; Araújo, A.C.; Barbalho, S.M. Phytochemicals and Regulation of NF-κB in Inflammatory Bowel Diseases: An Overview of In Vitro and In Vivo Effects. *Metabolites* **2023**, *13*, doi:10.3390/metabo13010096.
91. Buglio, D.S.; Marton, L.T.; Laurindo, L.F.; Guiguer, E.L.; Araújo, A.C.; Buchaim, R.L.; Goulart, R.A.; Rubira, C.J.; Barbalho, S.M. The Role of Resveratrol in Mild Cognitive Impairment and Alzheimer's Disease: A Systematic Review. *J Med Food* **2022**, *25*, 797-806, doi:10.1089/jmf.2021.0084.
92. Ciupei, D.; Colișar, A.; Leopold, L.; Stănilă, A.; Diaconeasa, Z.M. Polyphenols: From Classification to Therapeutic Potential and Bioavailability. *Foods* **2024**, *13*, doi:10.3390/foods13244131.
93. Yang, B.; Dong, Y.; Wang, F.; Zhang, Y. Nanoformulations to Enhance the Bioavailability and Physiological Functions of Polyphenols. *Molecules* **2020**, *25*, doi:10.3390/molecules25204613.
94. Grabska-Kobyłeczka, I.; Szpakowski, P.; Król, A.; Książek-Winiarek, D.; Kobyłeczki, A.; Głabiński, A.; Nowak, D. Polyphenols and Their Impact on the Prevention of Neurodegenerative Diseases and Development. *Nutrients* **2023**, *15*, doi:10.3390/nu15153454.
95. Arias-Sánchez, R.A.; Torner, L.; Fenton Navarro, B. Polyphenols and Neurodegenerative Diseases: Potential Effects and Mechanisms of Neuroprotection. *Molecules* **2023**, *28*, doi:10.3390/molecules28145415.

96. Mamun, A.A.; Shao, C.; Geng, P.; Wang, S.; Xiao, J. Polyphenols Targeting NF- $\kappa$ B Pathway in Neurological Disorders: What We Know So Far? *Int J Biol Sci* **2024**, *20*, 1332-1355, doi:10.7150/ijbs.90982.
97. Kaluza, M.; Ksiazek-Winiarek, D.; Szpakowski, P.; Czapkowska, J.; Fijalkowska, J.; Glabinski, A. Polyphenols in the Central Nervous System: Cellular Effects and Liposomal Delivery Approaches. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26136477.
98. Biswas, P.; Dey, D.; Biswas, P.K.; Rahaman, T.I.; Saha, S.; Parvez, A.; Khan, D.A.; Lily, N.J.; Saha, K.; Sohel, M.; et al. A Comprehensive Analysis and Anti-Cancer Activities of Quercetin in ROS-Mediated Cancer and Cancer Stem Cells. *Int J Mol Sci* **2022**, *23*, doi:10.3390/ijms231911746.
99. Teng, H.; Chen, L. Polyphenols and bioavailability: an update. *Crit Rev Food Sci Nutr* **2019**, *59*, 2040-2051, doi:10.1080/10408398.2018.1437023.
100. Williamson, G. Bioavailability of Food Polyphenols: Current State of Knowledge. *Annu Rev Food Sci Technol* **2025**, *16*, 315-332, doi:10.1146/annurev-food-060721-023817.
101. Manach, C.; Williamson, G.; Morand, C.; Scalbert, A.; Rémésy, C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am J Clin Nutr* **2005**, *81*, 230s-242s, doi:10.1093/ajcn/81.1.230S.
102. Hu, M.; Wu, B.; Liu, Z. Bioavailability of Polyphenols and Flavonoids in the Era of Precision Medicine. *Mol Pharm* **2017**, *14*, 2861-2863, doi:10.1021/acs.molpharmaceut.7b00545.
103. Liu, Z.; Hu, M. Natural polyphenol disposition via coupled metabolic pathways. *Expert Opin Drug Metab Toxicol* **2007**, *3*, 389-406, doi:10.1517/17425255.3.3.389.
104. Kapetanovic, I.M.; Muzzio, M.; Huang, Z.; Thompson, T.N.; McCormick, D.L. Pharmacokinetics, oral bioavailability, and metabolic profile of resveratrol and its dimethylether analog, pterostilbene, in rats. *Cancer Chemother Pharmacol* **2011**, *68*, 593-601, doi:10.1007/s00280-010-1525-4.
105. Lee, D.H.; Kim, J.W.; Cong, R.; Park, J.S.; Nguyen, C.H.B.; Park, K.; Kang, K.; Shim, S.M. Exploring absorption indices for a variety of polyphenols through Caco-2 cell model: insights from permeability studies and principal component analysis. *J Sci Food Agric* **2025**, *105*, 6243-6253, doi:10.1002/jsfa.14339.
106. Martos, D.; Lőrinczi, B.; Szatmári, I.; Vécsei, L.; Tanaka, M. Decoupling Behavioral Domains via Kynurenic Acid Analog Optimization: Implications for Schizophrenia and Parkinson's Disease Therapeutics. *Cells* **2025**, *14*, doi:10.3390/cells14130973.
107. Dos Santos, T.C.; Gomes, T.M.; Pinto, B.A.S.; Camara, A.L.; Paes, A.M.A. Naturally Occurring Acetylcholinesterase Inhibitors and Their Potential Use for Alzheimer's Disease Therapy. *Front Pharmacol* **2018**, *9*, 1192, doi:10.3389/fphar.2018.01192.
108. Kabir, M.T.; Uddin, M.S.; Begum, M.M.; Thangapandiyan, S.; Rahman, M.S.; Aleya, L.; Mathew, B.; Ahmed, M.; Barreto, G.E.; Ashraf, G.M. Cholinesterase Inhibitors for Alzheimer's Disease: Multitargeting Strategy Based on Anti-Alzheimer's Drugs Repositioning. *Curr Pharm Des* **2019**, *25*, 3519-3535, doi:10.2174/1381612825666191008103141.
109. Murakami, T.; Bodor, E.; Bodor, N. Approaching strategy to increase the oral bioavailability of berberine, a quaternary ammonium isoquinoline alkaloid: Part 1. Physicochemical and pharmacokinetic properties. *Expert Opin Drug Metab Toxicol* **2023**, *19*, 129-137, doi:10.1080/17425255.2023.2203857.
110. Abo El-Enin, H.A.; Elkomy, M.H.; Naguib, I.A.; Ahmed, M.F.; Alsaidan, O.A.; Alsalahat, I.; Ghoneim, M.M.; Eid, H.M. Lipid Nanocarriers Overlaid with Chitosan for Brain Delivery of Berberine via the Nasal Route. *Pharmaceuticals (Basel)* **2022**, *15*, doi:10.3390/ph15030281.
111. Kwon, M.; Lim, D.Y.; Lee, C.H.; Jeon, J.H.; Choi, M.K.; Song, I.S. Enhanced Intestinal Absorption and Pharmacokinetic Modulation of Berberine and Its Metabolites through the Inhibition of P-Glycoprotein and Intestinal Metabolism in Rats Using a Berberine Mixed Micelle Formulation. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12090882.
112. El-Nahas, A.E.; Elbedaiwy, H.M.; Masoud, I.M.; Aly, R.G.; Helmy, M.W.; El-Kamel, A.H. Berberine-loaded zein/hyaluronic acid composite nanoparticles for efficient brain uptake to alleviate neuro-degeneration in the pilocarpine model of epilepsy. *Eur J Pharm Biopharm* **2023**, *188*, 182-200, doi:10.1016/j.ejpb.2023.04.008.
113. Xiong, W.; Sang, W.; Linghu, K.G.; Zhong, Z.F.; Cheang, W.S.; Li, J.; Hu, Y.J.; Yu, H.; Wang, Y.T. Dual-functional Brij-S20-modified nanocrystal formulation enhances the intestinal transport and oral bioavailability of berberine. *Int J Nanomedicine* **2018**, *13*, 3781-3793, doi:10.2147/ijn.S163763.

114. Shen, C.C.; Yang, M.Y.; Hsieh, W.Y.; Tsay, G.J.; Yang, Y.C.; Huang, Y.F.; Liu, S.Y.; Lai, C.M.; Lee, C.H.; Tang, C.M.; et al. Berberine's Impact on Apoptosis, Proliferation, Uptake Efficiency, and Nanoparticle-Based Therapy in DBTRG Cells. *ACS Nanosci Au* **2025**, *5*, 165-183, doi:10.1021/acsnanoscienceau.5c00004.
115. Elsheikh, M.A.; Elnaggar, Y.S.R.; Hamdy, D.A.; Abdallah, O.Y. Novel cremochylomicrons for improved oral bioavailability of the antineoplastic phytochemistry berberine chloride: Optimization and pharmacokinetics. *Int J Pharm* **2018**, *535*, 316-324, doi:10.1016/j.ijpharm.2017.11.023.
116. Bian, X.; Guo, Q.; Yau, L.F.; Yang, L.; Wang, X.; Zhao, S.; Wu, S.; Qin, X.; Jiang, Z.H.; Li, C. Berberine-inspired ionizable lipid for self-structure stabilization and brain targeting delivery of nucleic acid therapeutics. *Nat Commun* **2025**, *16*, 2368, doi:10.1038/s41467-025-57488-0.
117. Saleh, S.R.; Abd-Elmegied, A.; Aly Madhy, S.; Khatlab, S.N.; Sheta, E.; Elnozahy, F.Y.; Mehanna, R.A.; Ghareeb, D.A.; Abd-Elmonem, N.M. Brain-targeted Tet-1 peptide-PLGA nanoparticles for berberine delivery against STZ-induced Alzheimer's disease in a rat model: Alleviation of hippocampal synaptic dysfunction, Tau pathology, and amyloidogenesis. *Int J Pharm* **2024**, *658*, 124218, doi:10.1016/j.ijpharm.2024.124218.
118. Marucci, G.; Buccioni, M.; Ben, D.D.; Lambertucci, C.; Volpini, R.; Amenta, F. Efficacy of acetylcholinesterase inhibitors in Alzheimer's disease. *Neuropharmacology* **2021**, *190*, 108352, doi:10.1016/j.neuropharm.2020.108352.
119. Moss, D.E. Improving Anti-Neurodegenerative Benefits of Acetylcholinesterase Inhibitors in Alzheimer's Disease: Are Irreversible Inhibitors the Future? *Int J Mol Sci* **2020**, *21*, doi:10.3390/ijms21103438.
120. Ebert, A.; Goss, K.U. Blood-brain barrier permeability revisited: Predicting intrinsic passive BBB permeability using the Solubility-diffusion model. *Eur J Pharm Sci* **2025**, *215*, 107354, doi:10.1016/j.ejps.2025.107354.
121. Fong, C.W. Permeability of the Blood-Brain Barrier: Molecular Mechanism of Transport of Drugs and Physiologically Important Compounds. *J Membr Biol* **2015**, *248*, 651-669, doi:10.1007/s00232-015-9778-9.
122. Vilar, S.; Sobarzo-Sanchez, E.; Santana, L.; Uriarte, E. Ligand and Structure-based Modeling of Passive Diffusion through the Blood-Brain Barrier. *Curr Med Chem* **2018**, *25*, 1073-1089, doi:10.2174/0929867324666171106163742.
123. Calapai, F.; Cardia, L.; Sorbara, E.E.; Navarra, M.; Gangemi, S.; Calapai, G.; Mannucci, C. Cannabinoids, Blood-Brain Barrier, and Brain Disposition. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12030265.
124. Basavarajappa, B.S.; Subbanna, S. Unveiling the Potential of Phytocannabinoids: Exploring Marijuana's Lesser-Known Constituents for Neurological Disorders. *Biomolecules* **2024**, *14*, doi:10.3390/biom14101296.
125. Chatterjee, S.; Deshpande, A.A.; Shen, H. Recent advances in the in vitro and in vivo methods to assess impact of P-glycoprotein and breast cancer resistance protein transporters in central nervous system drug disposition. *Biopharm Drug Dispos* **2023**, *44*, 7-25, doi:10.1002/bdd.2345.
126. Patel, N.C.; Feng, B.; Hou, X.; West, M.A.; Trapa, P.E.; Sciabola, S.; Verhoest, P.; Liras, J.L.; Maurer, T.S.; Wager, T.T. Harnessing Preclinical Data as a Predictive Tool for Human Brain Tissue Targeting. *ACS Chem Neurosci* **2021**, *12*, 1007-1017, doi:10.1021/acchemneuro.0c00807.
127. Śmiarowska, M.; Białocka, M.; Machoy-Mokrzyńska, A. Cannabis and cannabinoids: pharmacology and therapeutic potential. *Neurol Neurochir Pol* **2022**, *56*, 4-13, doi:10.5603/PJNNS.a2022.0015.
128. Friedman, D.; French, J.A.; Maccarrone, M. Safety, efficacy, and mechanisms of action of cannabinoids in neurological disorders. *Lancet Neurol* **2019**, *18*, 504-512, doi:10.1016/s1474-4422(19)30032-8.
129. Cristino, L.; Bisogno, T.; Di Marzo, V. Cannabinoids and the expanded endocannabinoid system in neurological disorders. *Nat Rev Neurol* **2020**, *16*, 9-29, doi:10.1038/s41582-019-0284-z.
130. Blebea, N.M.; Pricopie, A.I.; Vlad, R.A.; Hancu, G. Phytocannabinoids: Exploring Pharmacological Profiles and Their Impact on Therapeutic Use. *Int J Mol Sci* **2024**, *25*, doi:10.3390/ijms25084204.
131. Pagano, C.; Navarra, G.; Coppola, L.; Avilia, G.; Bifulco, M.; Laezza, C. Cannabinoids: Therapeutic Use in Clinical Practice. *Int J Mol Sci* **2022**, *23*, doi:10.3390/ijms23063344.
132. Kopustinskiene, D.M.; Masteikova, R.; Lazauskas, R.; Bernatoniene, J. Cannabis sativa L. Bioactive Compounds and Their Protective Role in Oxidative Stress and Inflammation. *Antioxidants (Basel)* **2022**, *11*, doi:10.3390/antiox11040660.

133. Grifoni, L.; Landucci, E.; Pieraccini, G.; Mazzantini, C.; Bergonzi, M.C.; Pellegrini-Giampietro, D.E.; Bilia, A.R. Development and Blood-Brain Barrier Penetration of Nanovesicles Loaded with Cannabidiol. *Pharmaceuticals (Basel)* **2025**, *18*, doi:10.3390/ph18020160.
134. Al-Khazaleh, A.K.; Zhou, X.; Bhuyan, D.J.; Münch, G.W.; Al-Dalabeeh, E.A.; Jaye, K.; Chang, D. The Neurotherapeutic Arsenal in Cannabis sativa: Insights into Anti-Neuroinflammatory and Neuroprotective Activity and Potential Entourage Effects. *Molecules* **2024**, *29*, doi:10.3390/molecules29020410.
135. Li, X.; Huang, L.; Liu, G.; Fan, W.; Li, B.; Liu, R.; Wang, Z.; Fan, Q.; Xiao, W.; Li, Y.; et al. Ginkgo diterpene lactones inhibit cerebral ischemia/reperfusion induced inflammatory response in astrocytes via TLR4/NF- $\kappa$ B pathway in rats. *J Ethnopharmacol* **2020**, *249*, 112365, doi:10.1016/j.jep.2019.112365.
136. Yu, T.; Wei, Z.; Wang, J.; Song, C.; Huang, W.; Zhang, P.; Shi, J.; Zhang, R.; Jiang, M.; Wang, D.; et al. Ginkgo biloba Extract GBE50 ameliorates cerebrovascular dysfunction and cognitive impairment in a mouse model of Alzheimer's disease. *Phytomedicine* **2025**, *141*, 156646, doi:10.1016/j.phymed.2025.156646.
137. Upadhyay, G.; Fihurka, O.; Patel, P.; Sanchez-Ramos, J. Quantitation of Cannabidiol (CBD) in brain regions and plasma following intranasal administration of a CBD nanoformulation. *J Cannabis Res* **2025**, *7*, 63, doi:10.1186/s42238-025-00308-5.
138. Dehnostel, F.O.; Dixit, V.A.; Preissner, R.; Banerjee, P. Non-animal models for blood-brain barrier permeability evaluation of drug-like compounds. *Sci Rep* **2024**, *14*, 8908, doi:10.1038/s41598-024-59734-9.
139. Mohan Kumar, D.; Talwar, P. Neurotherapeutics across blood-brain barrier: screening of BBB-permeable and CNS-active molecules for neurodegenerative disease. *Front Pharmacol* **2025**, *16*, 1616144, doi:10.3389/fphar.2025.1616144.
140. Etukudo, E.M.; Usman, I.M.; Ovosun, A.; Ojiakor, V.O.; Makena, W.; Owembabazi, E.; Aja, P.M.; Mutume Nzanzu Vivalya, B.; Archibong, V.B.; Anyanwu, E. Exploring the Neuroprotective Potentials of Flavonoid Metabolites in Syzygium aromaticum: A Review with in-silico Insight to Therapeutic Potential. *J Exp Pharmacol* **2025**, *17*, 587-611, doi:10.2147/jep.S536765.
141. Meng, F.; Xi, Y.; Huang, J.; Ayers, P.W. A curated diverse molecular database of blood-brain barrier permeability with chemical descriptors. *Sci Data* **2021**, *8*, 289, doi:10.1038/s41597-021-01069-5.
142. Janicka, M.; Sztanke, M.; Sztanke, K. Modeling the Blood-Brain Barrier Permeability of Potential Heterocyclic Drugs via Biomimetic IAM Chromatography Technique Combined with QSAR Methodology. *Molecules* **2024**, *29*, doi:10.3390/molecules29020287.
143. Kocsis, A.E.; Kucsápszky, N.; Santa-Maria, A.R.; Hunyadi, A.; Deli, M.A.; Walter, F.R. Much More than Nutrients: The Protective Effects of Nutraceuticals on the Blood-Brain Barrier in Diseases. *Nutrients* **2025**, *17*, doi:10.3390/nu17050766.
144. Isabel, U.-V.; Belén, A.d.I.R.M.; Elena, G.-B. A new frontier in neuropharmacology: Recent progress in natural products research for blood-brain barrier crossing. *Current Research in Biotechnology* **2024**, *8*, 100235.
145. Kato, R.; Zhang, L.; Kinatukara, N.; Huang, R.; Asthana, A.; Weber, C.; Xia, M.; Xu, X.; Shah, P. Investigating blood-brain barrier penetration and neurotoxicity of natural products for central nervous system drug development. *Sci Rep* **2025**, *15*, 7431, doi:10.1038/s41598-025-90888-2.
146. Kumar, V.; Banerjee, A.; Roy, K. Breaking the Barriers: Machine-Learning-Based c-RASAR Approach for Accurate Blood-Brain Barrier Permeability Prediction. *J Chem Inf Model* **2024**, *64*, 4298-4309, doi:10.1021/acs.jcim.4c00433.
147. Spielvogel, C.P.; Schindler, N.; Schröder, C.; Stellnberger, S.L.; Wadsak, W.; Mitterhauser, M.; Papp, L.; Hacker, M.; Pichler, V.; Vraka, C. Enhancing Blood-Brain Barrier Penetration Prediction by Machine Learning-Based Integration of Novel and Existing, In Silico and Experimental Molecular Parameters from a Standardized Database. *J Chem Inf Model* **2025**, *65*, 2773-2784, doi:10.1021/acs.jcim.4c02212.
148. Liu, S.; Jin, X.; Ge, Y.; Dong, J.; Liu, X.; Pei, X.; Wang, P.; Wang, B.; Chang, Y.; Yu, X.A. Advances in brain-targeted delivery strategies and natural product-mediated enhancement of blood-brain barrier permeability. *J Nanobiotechnology* **2025**, *23*, 382, doi:10.1186/s12951-025-03415-w.
149. Maher, R.; Moreno-Borrillo, A.; Jindal, D.; Mai, B.T.; Ruiz-Hernandez, E.; Harkin, A. Intranasal Polymeric and Lipid-Based Nanocarriers for CNS Drug Delivery. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15030746.

150. Ilić, T.; Đoković, J.B.; Nikolić, I.; Mitrović, J.R.; Pantelić, I.; Savić, S.D.; Savić, M.M. Parenteral Lipid-Based Nanoparticles for CNS Disorders: Integrating Various Facets of Preclinical Evaluation towards More Effective Clinical Translation. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15020443.
151. Nguyen, T.T.; Maeng, H.J. Pharmacokinetics and Pharmacodynamics of Intranasal Solid Lipid Nanoparticles and Nanostructured Lipid Carriers for Nose-to-Brain Delivery. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14030572.
152. Zhang, Y.; Guo, Z.; Zhang, H.; Wei, H.; Wang, T.; Du, S.; Li, P. Transnasal PLGA Nanoparticles with Terpene Permeation Enhancers: Membrane Remodeling and Tight Junction Modulation for Enhanced Brain Drug Delivery. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26083861.
153. Reddy, T.S.; Zomer, R.; Mantri, N. Nanoformulations as a strategy to overcome the delivery limitations of cannabinoids. *Phytother Res* **2023**, *37*, 1526-1538, doi:10.1002/ptr.7742.
154. Pires, P.C.; Rodrigues, M.; Alves, G.; Santos, A.O. Strategies to Improve Drug Strength in Nasal Preparations for Brain Delivery of Low Aqueous Solubility Drugs. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14030588.
155. Koo, J.; Lim, C.; Oh, K.T. Recent Advances in Intranasal Administration for Brain-Targeting Delivery: A Comprehensive Review of Lipid-Based Nanoparticles and Stimuli-Responsive Gel Formulations. *Int J Nanomedicine* **2024**, *19*, 1767-1807, doi:10.2147/ijn.S439181.
156. Akpınar Adscheid, S.; Rojas-Rodríguez, M.; Abdel-Hafez, S.M.; Pavone, F.S.; Schneider, M.; Türeli, A.E.; Calamai, M.; Günday-Türeli, N. Scalable Manufacturing Method for Model Protein-Loaded PLGA Nanoparticles: Biocompatibility, Trafficking and Release Properties. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17010087.
157. Lababidi, J.M.; Azzazy, H.M.E. Revamping Parkinson's disease therapy using PLGA-based drug delivery systems. *NPJ Parkinsons Dis* **2025**, *11*, 248, doi:10.1038/s41531-025-01081-1.
158. Yang, J.; Zeng, H.; Luo, Y.; Chen, Y.; Wang, M.; Wu, C.; Hu, P. Recent Applications of PLGA in Drug Delivery Systems. *Polymers (Basel)* **2024**, *16*, doi:10.3390/polym16182606.
159. Sheffey, V.V.; Siew, E.B.; Tanner, E.E.L.; Eniola-Adefeso, O. PLGA's Plight and the Role of Stealth Surface Modification Strategies in Its Use for Intravenous Particulate Drug Delivery. *Adv Healthc Mater* **2022**, *11*, e2101536, doi:10.1002/adhm.202101536.
160. Kesharwani, P.; Kumar, V.; Goh, K.W.; Gupta, G.; Alsayari, A.; Wahab, S.; Sahebkar, A. PEGylated PLGA nanoparticles: unlocking advanced strategies for cancer therapy. *Mol Cancer* **2025**, *24*, 205, doi:10.1186/s12943-025-02410-x.
161. Sánchez-López, E.; Ettcheto, M.; Egea, M.A.; Espina, M.; Cano, A.; Calpena, A.C.; Camins, A.; Carmona, N.; Silva, A.M.; Souto, E.B.; et al. Memantine loaded PLGA PEGylated nanoparticles for Alzheimer's disease: in vitro and in vivo characterization. *J Nanobiotechnology* **2018**, *16*, 32, doi:10.1186/s12951-018-0356-z.
162. De Soricellis, C.; Amante, C.; Russo, P.; Aquino, R.P.; Del Gaudio, P. Prilling as an Effective Tool for Manufacturing Submicrometric and Nanometric PLGA Particles for Controlled Drug Delivery to Wounds: Stability and Curcumin Release. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17010129.
163. Hoyos-Ceballos, G.P.; Ruozi, B.; Ottonelli, I.; Da Ros, F.; Vandelli, M.A.; Forni, F.; Daini, E.; Vilella, A.; Zoli, M.; Tosi, G.; et al. PLGA-PEG-ANG-2 Nanoparticles for Blood-Brain Barrier Crossing: Proof-of-Concept Study. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12010072.
164. Zhang, W.; Refaat, A.; Li, H.; Zhu, D.; Tong, Z.; Nicolazzo, J.A.; Peng, B.; Bai, H.; Esser, L.; Voelcker, N.H. Optimizing Angiopep-2 Density on Polymeric Nanoparticles for Enhanced Blood-Brain Barrier Penetration and Glioblastoma Targeting: Insights From In Vitro and In Vivo Experiments. *Advanced Functional Materials* **2025**, 2425165.
165. Chen, Z.A.; Wu, C.H.; Wu, S.H.; Huang, C.Y.; Mou, C.Y.; Wei, K.C.; Yen, Y.; Chien, I.T.; Runa, S.; Chen, Y.P.; et al. Receptor Ligand-Free Mesoporous Silica Nanoparticles: A Streamlined Strategy for Targeted Drug Delivery across the Blood-Brain Barrier. *ACS Nano* **2024**, *18*, 12716-12736, doi:10.1021/acsnano.3c08993.

166. Ojeda-Hernández, D.D.; Canales-Aguirre, A.A.; Matias-Guiu, J.; Gomez-Pinedo, U.; Mateos-Díaz, J.C. Potential of Chitosan and Its Derivatives for Biomedical Applications in the Central Nervous System. *Front Bioeng Biotechnol* **2020**, *8*, 389, doi:10.3389/fbioe.2020.00389.
167. Meng, Q.; Wang, A.; Hua, H.; Jiang, Y.; Wang, Y.; Mu, H.; Wu, Z.; Sun, K. Intranasal delivery of Huperzine A to the brain using lactoferrin-conjugated N-trimethylated chitosan surface-modified PLGA nanoparticles for treatment of Alzheimer's disease. *Int J Nanomedicine* **2018**, *13*, 705-718, doi:10.2147/ijn.S151474.
168. O'Donnell, A.; Moollan, A.; Baneham, S.; Ozgul, M.; Pabari, R.M.; Cox, D.; Kirby, B.P.; Ramtoola, Z. Intranasal and intravenous administration of octa-arginine modified poly(lactic-co-glycolic acid) nanoparticles facilitates central nervous system delivery of loperamide. *J Pharm Pharmacol* **2015**, *67*, 525-536, doi:10.1111/jphp.12347.
169. Jang, Y.J.; Kang, S.J.; Park, H.S.; Lee, D.H.; Kim, J.H.; Kim, J.E.; Kim, D.I.; Chung, C.H.; Yoon, J.K.; Bhang, S.H. Drug delivery strategies with lipid-based nanoparticles for Alzheimer's disease treatment. *J Nanobiotechnology* **2025**, *23*, 99, doi:10.1186/s12951-025-03109-3.
170. Aslam, M.; Javed, M.N.; Deeb, H.H.; Nicola, M.K.; Mirza, M.A.; Alam, M.S.; Akhtar, M.H.; Waziri, A. Lipid Nanocarriers for Neurotherapeutics: Introduction, Challenges, Blood-brain Barrier, and Promises of Delivery Approaches. *CNS Neurol Disord Drug Targets* **2022**, *21*, 952-965, doi:10.2174/1871527320666210706104240.
171. Chen, L.; Zhang, Z.; M, R.S.; Owens, T.; R, M.H.K.; Wu, C. PEGylated liposomes via ATRP for brain drug delivery. *J Liposome Res* **2025**, *35*, 283-289, doi:10.1080/08982104.2025.2485428.
172. Du, Q.; Liu, Y.; Fan, M.; Wei, S.; Ismail, M.; Zheng, M. PEG length effect of peptide-functional liposome for blood brain barrier (BBB) penetration and brain targeting. *J Control Release* **2024**, *372*, 85-94, doi:10.1016/j.jconrel.2024.06.005.
173. Zuberi, A.; Rehman, U.; Gupta, G.; Ghazwani, M.; Hani, U.; Kesharwani, P. Smart liposomal systems for brain cancer: Technological innovations in drug delivery. *Colloids Surf B Biointerfaces* **2025**, *255*, 114904, doi:10.1016/j.colsurfb.2025.114904.
174. Tapeinos, C.; Battaglini, M.; Ciofani, G. Advances in the design of solid lipid nanoparticles and nanostructured lipid carriers for targeting brain diseases. *J Control Release* **2017**, *264*, 306-332, doi:10.1016/j.jconrel.2017.08.033.
175. Mehrdadi, S. Drug Delivery of Solid Lipid Nanoparticles (SLNs) and Nanostructured Lipid Carriers (NLCs) to Target Brain Tumors. *Adv Pharm Bull* **2023**, *13*, 512-520, doi:10.34172/apb.2023.062.
176. Costa, C.P.; Moreira, J.N.; Sousa Lobo, J.M.; Silva, A.C. Intranasal delivery of nanostructured lipid carriers, solid lipid nanoparticles and nanoemulsions: A current overview of in vivo studies. *Acta Pharm Sin B* **2021**, *11*, 925-940, doi:10.1016/j.apsb.2021.02.012.
177. Zheng, Y.; Cui, L.; Lu, H.; Liu, Z.; Zhai, Z.; Wang, H.; Shao, L.; Lu, Z.; Song, X.; Zhang, Y. Nose to Brain: Exploring the Progress of Intranasal Delivery of Solid Lipid Nanoparticles and Nanostructured Lipid Carriers. *Int J Nanomedicine* **2024**, *19*, 12343-12368, doi:10.2147/ijn.S497480.
178. Bonferoni, M.C.; Rossi, S.; Sandri, G.; Ferrari, F.; Gavini, E.; Rassa, G.; Giunchedi, P. Nanoemulsions for "Nose-to-Brain" Drug Delivery. *Pharmaceutics* **2019**, *11*, doi:10.3390/pharmaceutics11020084.
179. Nguyen, T.T.; Duong, V.A. Advancements in Nanocarrier Systems for Nose-to-Brain Drug Delivery. *Pharmaceutics (Basel)* **2025**, *18*, doi:10.3390/ph18050615.
180. Bahadur, S.; Pardhi, D.M.; Rautio, J.; Rosenholm, J.M.; Pathak, K. Intranasal Nanoemulsions for Direct Nose-to-Brain Delivery of Actives for CNS Disorders. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12121230.
181. Chatterjee, B.; Gorain, B.; Mohananaidu, K.; Sengupta, P.; Mandal, U.K.; Choudhury, H. Targeted drug delivery to the brain via intranasal nanoemulsion: Available proof of concept and existing challenges. *Int J Pharm* **2019**, *565*, 258-268, doi:10.1016/j.ijpharm.2019.05.032.
182. Zhu, Y.; Liu, C.; Pang, Z. Dendrimer-Based Drug Delivery Systems for Brain Targeting. *Biomolecules* **2019**, *9*, doi:10.3390/biom9120790.
183. Li, H.; Zha, S.; Li, H.; Liu, H.; Wong, K.L.; All, A.H. Polymeric Dendrimers as Nanocarrier Vectors for Neurotheranostics. *Small* **2022**, *18*, e2203629, doi:10.1002/smll.202203629.

184. Janaszewska, A.; Lazniewska, J.; Trzepiński, P.; Marcinkowska, M.; Klajnert-Maculewicz, B. Cytotoxicity of Dendrimers. *Biomolecules* **2019**, *9*, doi:10.3390/biom9080330.
185. Bharadwaj, P.; Roullin, V.G.; Chain, J.L. Crossing the blood-brain barrier: advances in dendrimer-based nanocarriers for central nervous system delivery. *Nanoscale* **2025**, *17*, 23202-23227, doi:10.1039/d5nr02548f.
186. Senanayake, D.; Yapa, P.; Dabare, S.; Munaweera, I. Precision targeting of the CNS: recent progress in brain-directed nanodrug delivery. *RSC Adv* **2025**, *15*, 25910-25928, doi:10.1039/d5ra03578c.
187. Zawadzki, S.; Martín-Serrano, Á.; Okła, E.; Kędzierska, M.; Garcia-Gallego, S.; López, P.O.; de la Mata, F.J.; Michlewska, S.; Makowski, T.; Ionov, M.; et al. Synthesis and biophysical evaluation of carbosilane dendrimers as therapeutic siRNA carriers. *Sci Rep* **2024**, *14*, 1615, doi:10.1038/s41598-024-51238-w.
188. Santos, S.D.; Xavier, M.; Leite, D.M.; Moreira, D.A.; Custódio, B.; Torrado, M.; Castro, R.; Leiro, V.; Rodrigues, J.; Tomás, H.; et al. PAMAM dendrimers: blood-brain barrier transport and neuronal uptake after focal brain ischemia. *J Control Release* **2018**, *291*, 65-79, doi:10.1016/j.jconrel.2018.10.006.
189. Yan, X.; Chen, Q. Polyamidoamine Dendrimers: Brain-Targeted Drug Delivery Systems in Glioma Therapy. *Polymers (Basel)* **2024**, *16*, doi:10.3390/polym16142022.
190. Jiang, T.; Qiao, Y.; Ruan, W.; Zhang, D.; Yang, Q.; Wang, G.; Chen, Q.; Zhu, F.; Yin, J.; Zou, Y.; et al. Cation-Free siRNA Micelles as Effective Drug Delivery Platform and Potent RNAi Nanomedicines for Glioblastoma Therapy. *Adv Mater* **2021**, *33*, e2104779, doi:10.1002/adma.202104779.
191. Bhagat, N.; Nalawala, Z.; Patel, J.; Das, D.; Baldha, R.; Sarolia, J.; Rathod, S. Self-Assembled systems for Nose-to-Brain delivery of Temozolamide (TMZ) in brain tumor therapy. *Int J Pharm* **2025**, *675*, 125540, doi:10.1016/j.ijpharm.2025.125540.
192. Kadekar, S.; Nawale, G.N.; Rangasami, V.K.; Le Joncour, V.; Laakkonen, P.; Hilborn, J.; Varghese, O.P.; Oommen, O.P. Redox responsive Pluronic micelle mediated delivery of functional siRNA: a modular nano-assembly for targeted delivery. *Biomater Sci* **2021**, *9*, 3939-3944, doi:10.1039/d1bm00428j.
193. Wei, H.X.; Liu, M.H.; Wang, T.Y.; Shih, M.H.; Yu, J.; Yeh, Y.C. Fabrication of pH- and Ultrasound-Responsive Polymeric Micelles: The Effect of Amphiphilic Block Copolymers with Different Hydrophilic/Hydrophobic Block Ratios for Self-Assembly and Controlled Drug Release. *Biomacromolecules* **2025**, *26*, 2116-2130, doi:10.1021/acs.biomac.4c01202.
194. Convertine, A.J.; Diab, C.; Prieve, M.; Paschal, A.; Hoffman, A.S.; Johnson, P.H.; Stayton, P.S. pH-responsive polymeric micelle carriers for siRNA drugs. *Biomacromolecules* **2010**, *11*, 2904-2911, doi:10.1021/bm100652w.
195. Huang, X.; Li, J.; Li, G.; Ni, B.; Liang, Z.; Chen, H.; Xu, C.; Zhou, J.; Huang, J.; Deng, S. Cation-free siRNA-cored nanocapsules for tumor-targeted RNAi therapy. *Acta Biomater* **2023**, *161*, 226-237, doi:10.1016/j.actbio.2023.03.001.
196. Pechnikova, N.A.; Domvri, K.; Porpodis, K.; Istomina, M.S.; Iaremenko, A.V.; Yaremenko, A.V. Carbon quantum dots in biomedical applications: advances, challenges, and future prospects. *Aggregate* **2025**, *6*, e707.
197. Perini, G.; Palmieri, V.; Ciasca, G.; De Spirito, M.; Papi, M. Unravelling the Potential of Graphene Quantum Dots in Biomedicine and Neuroscience. *Int J Mol Sci* **2020**, *21*, doi:10.3390/ijms21103712.
198. Qiao, R.; Fu, C.; Forgham, H.; Javed, I.; Huang, X.; Zhu, J.; Whittaker, A.K.; Davis, T.P. Magnetic iron oxide nanoparticles for brain imaging and drug delivery. *Adv Drug Deliv Rev* **2023**, *197*, 114822, doi:10.1016/j.addr.2023.114822.
199. Truskewycz, A.; Yin, H.; Halberg, N.; Lai, D.T.H.; Ball, A.S.; Truong, V.K.; Rybicka, A.M.; Cole, I. Carbon Dot Therapeutic Platforms: Administration, Distribution, Metabolism, Excretion, Toxicity, and Therapeutic Potential. *Small* **2022**, *18*, e2106342, doi:10.1002/smll.202106342.
200. Molaei, M.J. Carbon quantum dots and their biomedical and therapeutic applications: a review. *RSC Adv* **2019**, *9*, 6460-6481, doi:10.1039/c8ra08088g.
201. Das, S.; Mondal, S.; Ghosh, D. Carbon quantum dots in bioimaging and biomedicines. *Front Bioeng Biotechnol* **2023**, *11*, 1333752, doi:10.3389/fbioe.2023.1333752.
202. Araújo, C.; Rodrigues, R.O.; Bañobre-López, M.; Silva, A.M.T.; Ribeiro, R.S. Carbon Dots as a Fluorescent Nanosystem for Crossing the Blood-Brain Barrier with Plausible Application in Neurological Diseases. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17040477.

203. Chaparro, C.I.P.; Simões, B.T.; Borges, J.P.; Castanho, M.; Soares, P.I.P.; Neves, V. A Promising Approach: Magnetic Nanosystems for Alzheimer's Disease Theranostics. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15092316.
204. Mohebichamkhorami, F.; Faizi, M.; Mahmoudifard, M.; Hajikarim-Hamedani, A.; Mohseni, S.S.; Heidari, A.; Ghane, Y.; Khoramjouy, M.; Khayati, M.; Ghasemi, R.; et al. Microfluidic Synthesis of Ultrasmall Chitosan/Graphene Quantum Dots Particles for Intranasal Delivery in Alzheimer's Disease Treatment. *Small* **2023**, *19*, e2207626, doi:10.1002/smll.202207626.
205. Chauhan, M.; Basu, S.M.; Qasim, M.; Giri, J. Polypropylene sulphide coating on magnetic nanoparticles as a novel platform for excellent biocompatible, stimuli-responsive smart magnetic nanocarriers for cancer therapeutics. *Nanoscale* **2023**, *15*, 7384-7402, doi:10.1039/d2nr05218k.
206. Raja, I.S.; Song, S.J.; Kang, M.S.; Lee, Y.B.; Kim, B.; Hong, S.W.; Jeong, S.J.; Lee, J.C.; Han, D.W. Toxicity of Zero- and One-Dimensional Carbon Nanomaterials. *Nanomaterials (Basel)* **2019**, *9*, doi:10.3390/nano9091214.
207. Sharma, M.; Kumar, C.; Arya, S.K.; Puri, S.; Khatri, M. Neurological effects of carbon quantum dots on zebrafish: A review. *Neuroscience* **2024**, *560*, 334-346, doi:10.1016/j.neuroscience.2024.10.016.
208. Jacobsen, N.R.; Møller, P.; Clausen, P.A.; Saber, A.T.; Micheletti, C.; Jensen, K.A.; Wallin, H.; Vogel, U. Biodistribution of Carbon Nanotubes in Animal Models. *Basic Clin Pharmacol Toxicol* **2017**, *121 Suppl 3*, 30-43, doi:10.1111/bcpt.12705.
209. Galassi, T.V.; Antman-Passig, M.; Yaari, Z.; Jessurun, J.; Schwartz, R.E.; Heller, D.A. Long-term in vivo biocompatibility of single-walled carbon nanotubes. *PLoS One* **2020**, *15*, e0226791, doi:10.1371/journal.pone.0226791.
210. Clementino, A.R.; Pellegrini, G.; Banella, S.; Colombo, G.; Cantù, L.; Sonvico, F.; Del Favero, E. Structure and Fate of Nanoparticles Designed for the Nasal Delivery of Poorly Soluble Drugs. *Mol Pharm* **2021**, *18*, 3132-3146, doi:10.1021/acs.molpharmaceut.1c00366.
211. Haro-Martínez, E.; Muscolino, E.; Moral, N.; Duran, J.; Fornaguera, C. Crossing the blood-brain barrier: nanoparticle-based strategies for neurodegenerative disease therapy. *Drug Deliv Transl Res* **2025**, doi:10.1007/s13346-025-01887-9.
212. Omar, S.H.; Osman, R.; Mamdouh, W.; Abdel-Bar, H.M.; Awad, G.A.S. Bioinspired lipid-polysaccharide modified hybrid nanoparticles as a brain-targeted highly loaded carrier for a hydrophilic drug. *Int J Biol Macromol* **2020**, *165*, 483-494, doi:10.1016/j.ijbiomac.2020.09.170.
213. Ishak, R.A.H.; Mostafa, N.M.; Kamel, A.O. Stealth lipid polymer hybrid nanoparticles loaded with rutin for effective brain delivery - comparative study with the gold standard (Tween 80): optimization, characterization and biodistribution. *Drug Deliv* **2017**, *24*, 1874-1890, doi:10.1080/10717544.2017.1410263.
214. Yuan, T.; Gao, L.; Zhan, W.; Dini, D. Effect of Particle Size and Surface Charge on Nanoparticles Diffusion in the Brain White Matter. *Pharm Res* **2022**, *39*, 767-781, doi:10.1007/s11095-022-03222-0.
215. Israel, L.L.; Galstyan, A.; Cox, A.; Shatalova, E.S.; Sun, T.; Rashid, M.H.; Grodzinski, Z.; Chiechi, A.; Fuchs, D.T.; Patil, R.; et al. Signature Effects of Vector-Guided Systemic Nano Bioconjugate Delivery Across Blood-Brain Barrier of Normal, Alzheimer's, and Tumor Mouse Models. *ACS Nano* **2022**, *16*, 11815-11832, doi:10.1021/acs.nano.1c10034.
216. Sharma, S.; Lee, D.; Maity, S.; Singh, P.; Chadokiya, J.; Mohaghegh, N.; Hassani, A.; Kim, H.; Gangarade, A.; Ljubimova, J.Y.; et al. Antibody-Free Immunopeptide Nanoconjugates for Brain-Targeted Drug Delivery in Glioblastoma Multiforme. *Bioconjug Chem* **2025**, *36*, 2132-2144, doi:10.1021/acs.bioconjugchem.5c00168.
217. Kim, W.; Ly, N.K.; He, Y.; Li, Y.; Yuan, Z.; Yeo, Y. Protein corona: Friend or foe? Co-opting serum proteins for nanoparticle delivery. *Adv Drug Deliv Rev* **2023**, *192*, 114635, doi:10.1016/j.addr.2022.114635.
218. Kashapov, R.; Ibragimova, A.; Pavlov, R.; Gabdrakhmanov, D.; Kashapova, N.; Buriylova, E.; Zakharova, L.; Sinyashin, O. Nanocarriers for Biomedicine: From Lipid Formulations to Inorganic and Hybrid Nanoparticles. *Int J Mol Sci* **2021**, *22*, doi:10.3390/ijms22137055.
219. Alshawwa, S.Z.; Kassem, A.A.; Farid, R.M.; Mostafa, S.K.; Labib, G.S. Nanocarrier Drug Delivery Systems: Characterization, Limitations, Future Perspectives and Implementation of Artificial Intelligence. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14040883.

220. Beach, M.A.; Nayanathara, U.; Gao, Y.; Zhang, C.; Xiong, Y.; Wang, Y.; Such, G.K. Polymeric Nanoparticles for Drug Delivery. *Chem Rev* **2024**, *124*, 5505-5616, doi:10.1021/acs.chemrev.3c00705.
221. Jiang, Y.; Li, W.; Wang, Z.; Lu, J. Lipid-Based Nanotechnology: Liposome. *Pharmaceutics* **2023**, *16*, doi:10.3390/pharmaceutics16010034.
222. Roszkowski, S.; Durczynska, Z. Advantages and limitations of nanostructures for biomedical applications. *Adv Clin Exp Med* **2025**, *34*, 447-456, doi:10.17219/acem/186846.
223. Kesharwani, P.; Gothwal, A.; Iyer, A.K.; Jain, K.; Chourasia, M.K.; Gupta, U. Dendrimer nanohybrid carrier systems: an expanding horizon for targeted drug and gene delivery. *Drug Discov Today* **2018**, *23*, 300-314, doi:10.1016/j.drudis.2017.06.009.
224. Das, S.S.; Bharadwaj, P.; Bilal, M.; Barani, M.; Rahdar, A.; Taboada, P.; Bungau, S.; Kyzas, G.Z. Stimuli-Responsive Polymeric Nanocarriers for Drug Delivery, Imaging, and Theragnosis. *Polymers (Basel)* **2020**, *12*, doi:10.3390/polym12061397.
225. Wakaskar, R.R. General overview of lipid-polymer hybrid nanoparticles, dendrimers, micelles, liposomes, spongosomes and cubosomes. *J Drug Target* **2018**, *26*, 311-318, doi:10.1080/1061186x.2017.1367006.
226. Yanar, F.; Carugo, D.; Zhang, X. Hybrid Nanoplatfoms Comprising Organic Nanocompartments Encapsulating Inorganic Nanoparticles for Enhanced Drug Delivery and Bioimaging Applications. *Molecules* **2023**, *28*, doi:10.3390/molecules28155694.
227. Mehta, M.; Bui, T.A.; Yang, X.; Aksoy, Y.; Goldys, E.M.; Deng, W. Lipid-Based Nanoparticles for Drug/Gene Delivery: An Overview of the Production Techniques and Difficulties Encountered in Their Industrial Development. *ACS Mater Au* **2023**, *3*, 600-619, doi:10.1021/acsmaterialsau.3c00032.
228. Viegas, C.; Patrício, A.B.; Prata, J.M.; Nadhman, A.; Chintamaneni, P.K.; Fonte, P. Solid Lipid Nanoparticles vs. Nanostructured Lipid Carriers: A Comparative Review. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15061593.
229. Scioli Montoto, S.; Muraca, G.; Ruiz, M.E. Solid Lipid Nanoparticles for Drug Delivery: Pharmacological and Biopharmaceutical Aspects. *Front Mol Biosci* **2020**, *7*, 587997, doi:10.3389/fmolb.2020.587997.
230. Duan, Y.; Dhar, A.; Patel, C.; Khimani, M.; Neogi, S.; Sharma, P.; Siva Kumar, N.; Vekariya, R.L. A brief review on solid lipid nanoparticles: part and parcel of contemporary drug delivery systems. *RSC Adv* **2020**, *10*, 26777-26791, doi:10.1039/d0ra03491f.
231. Ranjbar, S.; Emamjomeh, A.; Sharifi, F.; Zarepour, A.; Aghaabbasi, K.; Dehshahri, A.; Sepahvand, A.M.; Zarrabi, A.; Beyzaei, H.; Zahedi, M.M.; et al. Lipid-Based Delivery Systems for Flavonoids and Flavonolignans: Liposomes, Nanoemulsions, and Solid Lipid Nanoparticles. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15071944.
232. Dhiman, N.; Awasthi, R.; Sharma, B.; Kharkwal, H.; Kulkarni, G.T. Lipid Nanoparticles as Carriers for Bioactive Delivery. *Front Chem* **2021**, *9*, 580118, doi:10.3389/fchem.2021.580118.
233. Hassan, A.A.A.; Ramadan, E.; Kristó, K.; Regdon, G., Jr.; Sovány, T. Lipid-Polymer Hybrid Nanoparticles as a Smart Drug Delivery System for Peptide/Protein Delivery. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17060797.
234. Sivadasan, D.; Sultan, M.H.; Madkhali, O.; Almoshari, Y.; Thangavel, N. Polymeric Lipid Hybrid Nanoparticles (PLNs) as Emerging Drug Delivery Platform-A Comprehensive Review of Their Properties, Preparation Methods, and Therapeutic Applications. *Pharmaceutics* **2021**, *13*, doi:10.3390/pharmaceutics13081291.
235. Seyyedi-Mansour, S.; Carpena, M.; Barciela, P.; Perez-Vazquez, A.; Assadpour, E.; Prieto, M.A.; Jafari, S.M. Lipid-based nanocarriers loaded with bioactive compounds in active food packaging: Fabrication, characterization, and applications. *Adv Colloid Interface Sci* **2025**, *340*, 103457, doi:10.1016/j.cis.2025.103457.
236. Zhang, W.; Liu, Q.Y.; Haqqani, A.S.; Leclerc, S.; Liu, Z.; Fauteux, F.; Baumann, E.; Delaney, C.E.; Ly, D.; Star, A.T.; et al. Differential expression of receptors mediating receptor-mediated transcytosis (RMT) in brain microvessels, brain parenchyma and peripheral tissues of the mouse and the human. *Fluids Barriers CNS* **2020**, *17*, 47, doi:10.1186/s12987-020-00209-0.
237. Piantino, M.; Louis, F.; Shigemoto-Mogami, Y.; Kitamura, K.; Sato, K.; Yamaguchi, T.; Kawabata, K.; Yamamoto, S.; Iwasaki, S.; Hirabayashi, H.; et al. Brain microvascular endothelial cells derived from human

- induced pluripotent stem cells as in vitro model for assessing blood-brain barrier transferrin receptor-mediated transcytosis. *Mater Today Bio* **2022**, *14*, 100232, doi:10.1016/j.mtbio.2022.100232.
238. Katt, M.E.; Waters, E.A.; Gastfriend, B.D.; Herrin, B.R.; Cooper, M.D.; Shusta, E.V. Identification of Variable Lymphocyte Receptors That Target the Human Blood-Brain Barrier. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17091179.
239. Kuhn, P.; Petralla, S.; Dabbagh, F.; Pegoretti, V.; Muranyi, W.; Ishikawa, H.; Schroten, H.; Fischer, R.; Frenzel, A.; Schirrmann, T.; et al. A pH-sensitive binding modality allows successful transferrin receptor-mediated transcytosis of a bivalent antibody across brain barriers. *MAbs* **2025**, *17*, 2563758, doi:10.1080/19420862.2025.2563758.
240. Pardridge, W.M.; Chou, T. Mathematical Models of Blood-Brain Barrier Transport of Monoclonal Antibodies Targeting the Transferrin Receptor and the Insulin Receptor. *Pharmaceutics (Basel)* **2021**, *14*, doi:10.3390/ph14060535.
241. Shen, X.; Li, H.; Zhang, B.; Li, Y.; Zhu, Z. Targeting Transferrin Receptor 1 for Enhancing Drug Delivery Through the Blood-Brain Barrier for Alzheimer's Disease. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26199793.
242. Morrison, J.I.; Petrovic, A.; Metzendorf, N.G.; Rofo, F.; Yilmaz, C.U.; Stenler, S.; Laudon, H.; Hultqvist, G. Standardized Preclinical In Vitro Blood-Brain Barrier Mouse Assay Validates Endocytosis-Dependent Antibody Transcytosis Using Transferrin-Receptor-Mediated Pathways. *Mol Pharm* **2023**, *20*, 1564-1576, doi:10.1021/acs.molpharmaceut.2c00768.
243. Houry, N.; Pizzo, M.E.; Discenza, C.B.; Joy, D.; Tatarakis, D.; Todorov, M.I.; Negwer, M.; Ha, C.; De Melo, G.L.; Sarrafha, L.; et al. Fc-engineered large molecules targeting blood-brain barrier transferrin receptor and CD98hc have distinct central nervous system and peripheral biodistribution. *Nat Commun* **2025**, *16*, 1822, doi:10.1038/s41467-025-57108-x.
244. Ruiz-López, E.; Schuhmacher, A.J. Transportation of Single-Domain Antibodies through the Blood-Brain Barrier. *Biomolecules* **2021**, *11*, doi:10.3390/biom11081131.
245. Qiu, B.; Pompe, S.; Xenaki, K.T.; Di Maggio, A.; Moreno, C.B.; van Bergen En Henegouwen, P.M.P.; Mastrobattista, E.; Oliveira, S.; Caiazzo, M. Receptor-mediated transcytosis of nanobodies targeting the heparin-binding EGF-like growth factor in human blood-brain barrier models. *J Control Release* **2025**, *383*, 113852, doi:10.1016/j.jconrel.2025.113852.
246. Szeckó, A.; Mészáros, M.; Simões, B.; Cavaco, M.; Chaparro, C.; Porkoláb, G.; Castanho, M.; Deli, M.A.; Neves, V.; Veszelka, S. PEPH3-modified nanocarriers for delivery of therapeutics across the blood-brain barrier. *Fluids Barriers CNS* **2025**, *22*, 31, doi:10.1186/s12987-025-00641-0.
247. Ghorai, S.M.; Deep, A.; Magoo, D.; Gupta, C.; Gupta, N. Cell-Penetrating and Targeted Peptides Delivery Systems as Potential Pharmaceutical Carriers for Enhanced Delivery across the Blood-Brain Barrier (BBB). *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15071999.
248. Fu, L.; Bridges, C.A.; Kim, H.N.; Ding, C.; Bao Hou, N.C.; Yeow, J.; Fok, S.; Macmillan, A.; Sterling, J.D.; Baker, S.M.; et al. Cationic Polysaccharides Bind to the Endothelial Cell Surface Extracellular Matrix Involving Heparan Sulfate. *Biomacromolecules* **2024**, *25*, 3850-3862, doi:10.1021/acs.biomac.4c00477.
249. Reveret, L.; Leclerc, M.; Morin, F.; Émond, V.; Calon, F. Pharmacokinetics, biodistribution and toxicology of novel cell-penetrating peptides. *Sci Rep* **2023**, *13*, 11081, doi:10.1038/s41598-023-37280-0.
250. Habault, J.; Poyet, J.L. Recent Advances in Cell Penetrating Peptide-Based Anticancer Therapies. *Molecules* **2019**, *24*, doi:10.3390/molecules24050927.
251. Nam, S.H.; Park, J.; Koo, H. Recent advances in selective and targeted drug/gene delivery systems using cell-penetrating peptides. *Arch Pharm Res* **2023**, *46*, 18-34, doi:10.1007/s12272-022-01425-y.
252. Polderdijk, S.G.I.; Limzerwala, J.F.; Spiess, C. Plasma membrane damage limits cytoplasmic delivery by conventional cell penetrating peptides. *PLoS One* **2024**, *19*, e0305848, doi:10.1371/journal.pone.0305848.
253. Ghaemi, B.; Tanwar, S.; Singh, A.; Arifin, D.R.; McMahan, M.T.; Barman, I.; Bulte, J.W.M. Cell-Penetrating and Enzyme-Responsive Peptides for Targeted Cancer Therapy: Role of Arginine Residue Length on Cell Penetration and In Vivo Systemic Toxicity. *ACS Appl Mater Interfaces* **2024**, *16*, 11159-11171, doi:10.1021/acsami.3c14908.

254. Mendes, M.; Nunes, S.; Cova, T.; Branco, F.; Dyrks, M.; Kokscho, B.; Vale, N.; Sousa, J.; Pais, A.; Vitorino, C. Charge-switchable cell-penetrating peptides for rerouting nanoparticles to glioblastoma treatment. *Colloids Surf B Biointerfaces* **2024**, *241*, 113983, doi:10.1016/j.colsurfb.2024.113983.
255. Tang, B.; Zaro, J.L.; Shen, Y.; Chen, Q.; Yu, Y.; Sun, P.; Wang, Y.; Shen, W.C.; Tu, J.; Sun, C. Acid-sensitive hybrid polymeric micelles containing a reversibly activatable cell-penetrating peptide for tumor-specific cytoplasm targeting. *J Control Release* **2018**, *279*, 147-156, doi:10.1016/j.jconrel.2018.04.016.
256. Du, J.J.; Zhang, R.Y.; Jiang, S.; Xiao, S.; Liu, Y.; Niu, Y.; Zhao, W.X.; Wang, D.; Ma, X. Applications of cell penetrating peptide-based drug delivery system in immunotherapy. *Front Immunol* **2025**, *16*, 1540192, doi:10.3389/fimmu.2025.1540192.
257. Shi, D.; Mi, G.; Shen, Y.; Webster, T.J. Glioma-targeted dual functionalized thermosensitive Ferri-liposomes for drug delivery through an in vitro blood-brain barrier. *Nanoscale* **2019**, *11*, 15057-15071, doi:10.1039/c9nr03931g.
258. Porro, G.; Basile, M.; Xie, Z.; Tuveri, G.M.; Battaglia, G.; Lopes, C.D.F. A new era in brain drug delivery: Integrating multivalency and computational optimisation for blood-brain barrier permeation. *Adv Drug Deliv Rev* **2025**, *224*, 115637, doi:10.1016/j.addr.2025.115637.
259. Moreira, R.; Nóbrega, C.; de Almeida, L.P.; Mendonça, L. Brain-targeted drug delivery - nanovesicles directed to specific brain cells by brain-targeting ligands. *J Nanobiotechnology* **2024**, *22*, 260, doi:10.1186/s12951-024-02511-7.
260. Luo, F.; Zhong, T.; Chen, Y.; Guo, Q.; Tao, L.; Shen, X.; Fan, Y.; Wu, X. Dual-Ligand Synergistic Targeting Anti-Tumor Nanoplatfoms with Cascade-Responsive Drug Release. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15072014.
261. Guo, Q.; Xu, S.; Yang, P.; Wang, P.; Lu, S.; Sheng, D.; Qian, K.; Cao, J.; Lu, W.; Zhang, Q. A dual-ligand fusion peptide improves the brain-neuron targeting of nanocarriers in Alzheimer's disease mice. *J Control Release* **2020**, *320*, 347-362, doi:10.1016/j.jconrel.2020.01.039.
262. Chen, C.; Duan, Z.; Yuan, Y.; Li, R.; Pang, L.; Liang, J.; Xu, X.; Wang, J. Peptide-22 and Cyclic RGD Functionalized Liposomes for Glioma Targeting Drug Delivery Overcoming BBB and BBTB. *ACS Appl Mater Interfaces* **2017**, *9*, 5864-5873, doi:10.1021/acsami.6b15831.
263. Zhang, Y.; Zhai, M.; Chen, Z.; Han, X.; Yu, F.; Li, Z.; Xie, X.; Han, C.; Yu, L.; Yang, Y.; et al. Dual-modified liposome codelivery of doxorubicin and vincristine improve targeting and therapeutic efficacy of glioma. *Drug Deliv* **2017**, *24*, 1045-1055, doi:10.1080/10717544.2017.1344334.
264. Ling, C.; Chen, L.; Liang, J.; Li, X.; Wei, H.; Yu, C.; Wang, J. Neutrophil/monocyte-targeted dual-ligands modified liposomes delivering puerarin for ischemia stroke treatment. *Mater Today Bio* **2025**, *33*, 102077, doi:10.1016/j.mtbio.2025.102077.
265. Zhu, M.; Liu, Q.; Chen, Z.; Liu, J.; Zhang, Z.; Tian, J.; Wang, X.; Yang, X.; Chen, Q.; Huang, X.; et al. Rational Design of Dual-Targeted Nanomedicines for Enhanced Vascular Permeability in Low-Permeability Tumors. *ACS Nano* **2025**, *19*, 3424-3438, doi:10.1021/acsnano.4c12808.
266. Jiao, Y.; Yang, L.; Wang, R.; Song, G.; Fu, J.; Wang, J.; Gao, N.; Wang, H. Drug Delivery Across the Blood-Brain Barrier: A New Strategy for the Treatment of Neurological Diseases. *Pharmaceutics* **2024**, *16*, doi:10.3390/pharmaceutics16121611.
267. Toader, C.; Dumitru, A.V.; Eva, L.; Serban, M.; Covache-Busuioc, R.A.; Ciurea, A.V. Nanoparticle Strategies for Treating CNS Disorders: A Comprehensive Review of Drug Delivery and Theranostic Applications. *Int J Mol Sci* **2024**, *25*, doi:10.3390/ijms252413302.
268. Luo, S.; Lv, Z.; Yang, Q.; Chang, R.; Wu, J. Research Progress on Stimulus-Responsive Polymer Nanocarriers for Cancer Treatment. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15071928.
269. Deirram, N.; Zhang, C.; Kermaniyan, S.S.; Johnston, A.P.R.; Such, G.K. pH-Responsive Polymer Nanoparticles for Drug Delivery. *Macromol Rapid Commun* **2019**, *40*, e1800917, doi:10.1002/marc.201800917.
270. Meng, X.; Shen, Y.; Zhao, H.; Lu, X.; Wang, Z.; Zhao, Y. Redox-manipulating nanocarriers for anticancer drug delivery: a systematic review. *J Nanobiotechnology* **2024**, *22*, 587, doi:10.1186/s12951-024-02859-w.
271. Hersh, A.M.; Bhimreddy, M.; Weber-Levine, C.; Jiang, K.; Alomari, S.; Theodore, N.; Manbachi, A.; Tyler, B.M. Applications of Focused Ultrasound for the Treatment of Glioblastoma: A New Frontier. *Cancers (Basel)* **2022**, *14*, doi:10.3390/cancers14194920.

272. He, C.; Wu, Z.; Zhuang, M.; Li, X.; Xue, S.; Xu, S.; Xu, J.; Wu, Z.; Lu, M. Focused ultrasound-mediated blood-brain barrier opening combined with magnetic targeting cytomembrane based biomimetic microbubbles for glioblastoma therapy. *J Nanobiotechnology* **2023**, *21*, 297, doi:10.1186/s12951-023-02074-z.
273. Teleanu, R.I.; Preda, M.D.; Niculescu, A.G.; Vladăncenco, O.; Radu, C.I.; Grumezescu, A.M.; Teleanu, D.M. Current Strategies to Enhance Delivery of Drugs across the Blood-Brain Barrier. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14050987.
274. Sonabend, A.M.; Gould, A.; Amidei, C.; Ward, R.; Schmidt, K.A.; Zhang, D.Y.; Gomez, C.; Bebawy, J.F.; Liu, B.P.; Bouchoux, G.; et al. Repeated blood-brain barrier opening with an implantable ultrasound device for delivery of albumin-bound paclitaxel in patients with recurrent glioblastoma: a phase 1 trial. *Lancet Oncol* **2023**, *24*, 509-522, doi:10.1016/s1470-2045(23)00112-2.
275. Manzari, M.T.; Shamay, Y.; Kiguchi, H.; Rosen, N.; Scaltriti, M.; Heller, D.A. Targeted drug delivery strategies for precision medicines. *Nat Rev Mater* **2021**, *6*, 351-370, doi:10.1038/s41578-020-00269-6.
276. Johnsen, K.B.; Burkhart, A.; Thomsen, L.B.; Andresen, T.L.; Moos, T. Targeting the transferrin receptor for brain drug delivery. *Prog Neurobiol* **2019**, *181*, 101665, doi:10.1016/j.pneurobio.2019.101665.
277. Thomsen, M.S.; Johnsen, K.B.; Kucharz, K.; Lauritzen, M.; Moos, T. Blood-Brain Barrier Transport of Transferrin Receptor-Targeted Nanoparticles. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14102237.
278. Bien-Ly, N.; Yu, Y.J.; Bumbaca, D.; Elstrott, J.; Boswell, C.A.; Zhang, Y.; Luk, W.; Lu, Y.; Dennis, M.S.; Weimer, R.M.; et al. Transferrin receptor (TfR) trafficking determines brain uptake of TfR antibody affinity variants. *J Exp Med* **2014**, *211*, 233-244, doi:10.1084/jem.20131660.
279. Arguello, A.; Mahon, C.S.; Calvert, M.E.K.; Chan, D.; Dugas, J.C.; Pizzo, M.E.; Thomsen, E.R.; Chau, R.; Damo, L.A.; Duque, J.; et al. Molecular architecture determines brain delivery of a transferrin receptor-targeted lysosomal enzyme. *J Exp Med* **2022**, *219*, doi:10.1084/jem.20211057.
280. Bonvicini, G.; Singh, S.; Sandersjö, L.; Sehlin, D.; Syvänen, S.; Andersson, K.G. The effects of dose, valency, and affinity on TfR-mediated brain delivery in vivo. *Fluids Barriers CNS* **2025**, *22*, 36, doi:10.1186/s12987-025-00643-y.
281. di Polidoro, A.C.; Cafarchio, A.; Vecchione, D.; Donato, P.; De Nola, F.; Torino, E. Revealing Angiopoep-2/LRP1 Molecular Interaction for Optimal Delivery to Glioblastoma (GBM). *Molecules* **2022**, *27*, doi:10.3390/molecules27196696.
282. Khan, N.U.; Ni, J.; Ju, X.; Miao, T.; Chen, H.; Han, L. Escape from abluminal LRP1-mediated clearance for boosted nanoparticle brain delivery and brain metastasis treatment. *Acta Pharm Sin B* **2021**, *11*, 1341-1354, doi:10.1016/j.apsb.2020.10.015.
283. Kafa, H.; Wang, J.T.; Rubio, N.; Klippstein, R.; Costa, P.M.; Hassan, H.A.; Sosabowski, J.K.; Bansal, S.S.; Preston, J.E.; Abbott, N.J.; et al. Translocation of LRP1 targeted carbon nanotubes of different diameters across the blood-brain barrier in vitro and in vivo. *J Control Release* **2016**, *225*, 217-229, doi:10.1016/j.jconrel.2016.01.031.
284. Kato, N.; Yamada, S.; Suzuki, R.; Iida, Y.; Matsumoto, M.; Fumoto, S.; Arima, H.; Mukai, H.; Kawakami, S. Development of an apolipoprotein E mimetic peptide-lipid conjugate for efficient brain delivery of liposomes. *Drug Deliv* **2023**, *30*, 2173333, doi:10.1080/10717544.2023.2173333.
285. Benitez Amaro, A.; Solanelles Curco, A.; Garcia, E.; Julve, J.; Rives, J.; Benitez, S.; Llorente Cortes, V. Apolipoprotein and LRP1-Based Peptides as New Therapeutic Tools in Atherosclerosis. *J Clin Med* **2021**, *10*, doi:10.3390/jcm10163571.
286. Hjelm, L.C.; Lindberg, H.; Ståhl, S.; Löfblom, J. Affibody Molecules Intended for Receptor-Mediated Transcytosis via the Transferrin Receptor. *Pharmaceutics (Basel)* **2023**, *16*, doi:10.3390/ph16070956.
287. Crowe, T.P.; Hsu, W.H. Evaluation of Recent Intranasal Drug Delivery Systems to the Central Nervous System. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14030629.
288. Crowe, T.P.; Greenlee, M.H.W.; Kanthasamy, A.G.; Hsu, W.H. Mechanism of intranasal drug delivery directly to the brain. *Life Sci* **2018**, *195*, 44-52, doi:10.1016/j.lfs.2017.12.025.
289. Islam, S.U.; Shehzad, A.; Ahmed, M.B.; Lee, Y.S. Intranasal Delivery of Nanoformulations: A Potential Way of Treatment for Neurological Disorders. *Molecules* **2020**, *25*, doi:10.3390/molecules25081929.

290. Chen, Y.; Zhang, C.; Huang, Y.; Ma, Y.; Song, Q.; Chen, H.; Jiang, G.; Gao, X. Intranasal drug delivery: The interaction between nanoparticles and the nose-to-brain pathway. *Adv Drug Deliv Rev* **2024**, *207*, 115196, doi:10.1016/j.addr.2024.115196.
291. Agrawal, M.; Saraf, S.; Saraf, S.; Dubey, S.K.; Puri, A.; Gupta, U.; Kesharwani, P.; Ravichandiran, V.; Kumar, P.; Naidu, V.G.M.; et al. Stimuli-responsive In situ gelling system for nose-to-brain drug delivery. *J Control Release* **2020**, *327*, 235-265, doi:10.1016/j.jconrel.2020.07.044.
292. Vigani, B.; Rossi, S.; Sandri, G.; Bonferoni, M.C.; Caramella, C.M.; Ferrari, F. Recent Advances in the Development of In Situ Gelling Drug Delivery Systems for Non-Parenteral Administration Routes. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12090859.
293. Xu, K.; Duan, S.; Wang, W.; Ouyang, Q.; Qin, F.; Guo, P.; Hou, J.; He, Z.; Wei, W.; Qin, M. Nose-to-brain delivery of nanotherapeutics: Transport mechanisms and applications. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* **2024**, *16*, e1956, doi:10.1002/wnan.1956.
294. Lofts, A.; Abu-Hijleh, F.; Rigg, N.; Mishra, R.K.; Hoare, T. Using the Intranasal Route to Administer Drugs to Treat Neurological and Psychiatric Illnesses: Rationale, Successes, and Future Needs. *CNS Drugs* **2022**, *36*, 739-770, doi:10.1007/s40263-022-00930-4.
295. Huang, Q.; Chen, Y.; Zhang, W.; Xia, X.; Li, H.; Qin, M.; Gao, H. Nanotechnology for enhanced nose-to-brain drug delivery in treating neurological diseases. *J Control Release* **2024**, *366*, 519-534, doi:10.1016/j.jconrel.2023.12.054.
296. Gorick, C.M.; Breza, V.R.; Nowak, K.M.; Cheng, V.W.T.; Fisher, D.G.; Debski, A.C.; Hoch, M.R.; Demir, Z.E.F.; Tran, N.M.; Schwartz, M.R.; et al. Applications of focused ultrasound-mediated blood-brain barrier opening. *Adv Drug Deliv Rev* **2022**, *191*, 114583, doi:10.1016/j.addr.2022.114583.
297. Chen, K.T.; Wei, K.C.; Liu, H.L. Theranostic Strategy of Focused Ultrasound Induced Blood-Brain Barrier Opening for CNS Disease Treatment. *Front Pharmacol* **2019**, *10*, 86, doi:10.3389/fphar.2019.00086.
298. Zhu, H.; Allwin, C.; Bassous, M.G.; Pouliopoulos, A.N. Focused ultrasound-mediated enhancement of blood-brain barrier permeability for brain tumor treatment: a systematic review of clinical trials. *J Neurooncol* **2024**, *170*, 235-252, doi:10.1007/s11060-024-04795-z.
299. Gosselet, F.; Loiola, R.A.; Roig, A.; Rosell, A.; Culot, M. Central nervous system delivery of molecules across the blood-brain barrier. *Neurochem Int* **2021**, *144*, 104952, doi:10.1016/j.neuint.2020.104952.
300. Sun, T.; Samiotaki, G.; Wang, S.; Acosta, C.; Chen, C.C.; Konofagou, E.E. Acoustic cavitation-based monitoring of the reversibility and permeability of ultrasound-induced blood-brain barrier opening. *Phys Med Biol* **2015**, *60*, 9079-9094, doi:10.1088/0031-9155/60/23/9079.
301. Xie, Y.; Hu, J.; Lei, W.; Qian, S. Prediction of vascular injury by cavitation microbubbles in a focused ultrasound field. *Ultrason Sonochem* **2022**, *88*, 106103, doi:10.1016/j.ultsonch.2022.106103.
302. Meng, Y.; Kalia, L.V.; Kalia, S.K.; Hamani, C.; Huang, Y.; Hynynen, K.; Lipsman, N.; Davidson, B. Current Progress in Magnetic Resonance-Guided Focused Ultrasound to Facilitate Drug Delivery across the Blood-Brain Barrier. *Pharmaceutics* **2024**, *16*, doi:10.3390/pharmaceutics16060719.
303. Timbie, K.F.; Mead, B.P.; Price, R.J. Drug and gene delivery across the blood-brain barrier with focused ultrasound. *J Control Release* **2015**, *219*, 61-75, doi:10.1016/j.jconrel.2015.08.059.
304. Burgess, A.; Shah, K.; Hough, O.; Hynynen, K. Focused ultrasound-mediated drug delivery through the blood-brain barrier. *Expert Rev Neurother* **2015**, *15*, 477-491, doi:10.1586/14737175.2015.1028369.
305. Wu, S.Y.; Aurup, C.; Sanchez, C.S.; Grondin, J.; Zheng, W.; Kamimura, H.; Ferrera, V.P.; Konofagou, E.E. Efficient Blood-Brain Barrier Opening in Primates with Neuronavigation-Guided Ultrasound and Real-Time Acoustic Mapping. *Sci Rep* **2018**, *8*, 7978, doi:10.1038/s41598-018-25904-9.
306. Sun, T.; Zhang, Y.; Power, C.; Alexander, P.M.; Sutton, J.T.; Aryal, M.; Vykhodtseva, N.; Miller, E.L.; McDannold, N.J. Closed-loop control of targeted ultrasound drug delivery across the blood-brain/tumor barriers in a rat glioma model. *Proc Natl Acad Sci U S A* **2017**, *114*, E10281-e10290, doi:10.1073/pnas.1713328114.
307. Novell, A.; Kamimura, H.A.S.; Cafarelli, A.; Gerstenmayer, M.; Flament, J.; Valette, J.; Agou, P.; Conti, A.; Selingue, E.; Aron Badin, R.; et al. A new safety index based on intrapulse monitoring of ultra-harmonic cavitation during ultrasound-induced blood-brain barrier opening procedures. *Sci Rep* **2020**, *10*, 10088, doi:10.1038/s41598-020-66994-8.

308. Bae, S.; Liu, K.; Pouliopoulos, A.N.; Ji, R.; Jiménez-Gambín, S.; Yousefian, O.; Kline-Schoder, A.R.; Batts, A.J.; Tsitsos, F.N.; Kokossis, D.; et al. Transcranial blood-brain barrier opening in Alzheimer's disease patients using a portable focused ultrasound system with real-time 2-D cavitation mapping. *Theranostics* **2024**, *14*, 4519-4535, doi:10.7150/thno.94206.
309. Todd, N.; Angolano, C.; Ferran, C.; Devor, A.; Borsook, D.; McDannold, N. Secondary effects on brain physiology caused by focused ultrasound-mediated disruption of the blood-brain barrier. *J Control Release* **2020**, *324*, 450-459, doi:10.1016/j.jconrel.2020.05.040.
310. Wang, J.; Li, Z.; Pan, M.; Fiaz, M.; Hao, Y.; Yan, Y.; Sun, L.; Yan, F. Ultrasound-mediated blood-brain barrier opening: An effective drug delivery system for theranostics of brain diseases. *Adv Drug Deliv Rev* **2022**, *190*, 114539, doi:10.1016/j.addr.2022.114539.
311. Shin, J.; Kong, C.; Cho, J.S.; Lee, J.; Koh, C.S.; Yoon, M.S.; Na, Y.C.; Chang, W.S.; Chang, J.W. Focused ultrasound-mediated noninvasive blood-brain barrier modulation: preclinical examination of efficacy and safety in various sonication parameters. *Neurosurg Focus* **2018**, *44*, E15, doi:10.3171/2017.11.Focus17627.
312. Cheng, M.; Li, F.; Han, T.; Yu, A.C.H.; Qin, P. Effects of ultrasound pulse parameters on cavitation properties of flowing microbubbles under physiologically relevant conditions. *Ultrason Sonochem* **2019**, *52*, 512-521, doi:10.1016/j.ultsonch.2018.12.031.
313. Lin, Y.; Lin, L.; Cheng, M.; Jin, L.; Du, L.; Han, T.; Xu, L.; Yu, A.C.H.; Qin, P. Effect of acoustic parameters on the cavitation behavior of SonoVue microbubbles induced by pulsed ultrasound. *Ultrason Sonochem* **2017**, *35*, 176-184, doi:10.1016/j.ultsonch.2016.09.016.
314. Zhao, X.; Wright, A.; Goertz, D.E. An optical and acoustic investigation of microbubble cavitation in small channels under therapeutic ultrasound conditions. *Ultrason Sonochem* **2023**, *93*, 106291, doi:10.1016/j.ultsonch.2023.106291.
315. Patel, A.; Schoen, S.J., Jr.; Arvanitis, C.D. Closed Loop Spatial and Temporal Control of Cavitation Activity with Passive Acoustic Mapping. *IEEE Trans Biomed Eng* **2018**, doi:10.1109/tbme.2018.2882337.
316. Downs, M.E.; Buch, A.; Sierra, C.; Karakatsani, M.E.; Teichert, T.; Chen, S.; Konofagou, E.E.; Ferrera, V.P. Long-Term Safety of Repeated Blood-Brain Barrier Opening via Focused Ultrasound with Microbubbles in Non-Human Primates Performing a Cognitive Task. *PLoS One* **2015**, *10*, e0125911, doi:10.1371/journal.pone.0125911.
317. Mainprize, T.; Lipsman, N.; Huang, Y.; Meng, Y.; Bethune, A.; Ironside, S.; Heyn, C.; Alkins, R.; Trudeau, M.; Sahgal, A.; et al. Blood-Brain Barrier Opening in Primary Brain Tumors with Non-invasive MR-Guided Focused Ultrasound: A Clinical Safety and Feasibility Study. *Sci Rep* **2019**, *9*, 321, doi:10.1038/s41598-018-36340-0.
318. Rezai, A.R.; Ranjan, M.; Haut, M.W.; Carpenter, J.; D'Haese, P.F.; Mehta, R.I.; Najib, U.; Wang, P.; Claassen, D.O.; Chazen, J.L.; et al. Focused ultrasound-mediated blood-brain barrier opening in Alzheimer's disease: long-term safety, imaging, and cognitive outcomes. *J Neurosurg* **2023**, *139*, 275-283, doi:10.3171/2022.9.Jns221565.
319. Anastasiadis, P.; Gandhi, D.; Guo, Y.; Ahmed, A.K.; Bentzen, S.M.; Arvanitis, C.; Woodworth, G.F. Localized blood-brain barrier opening in infiltrating gliomas with MRI-guided acoustic emissions-controlled focused ultrasound. *Proc Natl Acad Sci U S A* **2021**, *118*, doi:10.1073/pnas.2103280118.
320. Pinkiewicz, M.; Zaczynski, A.; Walecki, J.; Zawadzki, M. Beyond the Walls of Troy: A Scoping Review on Pharmacological Strategies to Enhance Drug Delivery Across the Blood-Brain Barrier and Blood-Tumor Barrier. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26157050.
321. Priest, R.; Ambady, P.; Neweult, E. ACTR-23. safety of intra-arterial chemotherapy with osmotic opening of the blood-brain barrier. *Neuro-Oncology* **2018**, *20*, vi16.
322. Ferreira, C.; Ferreira, M.Y.; Cardoso, L.J.C.; Scarramal, J.P.L.; Nogueira, A.; Wong, T.; Bokil, S.; Singh, F.; Massimo, S.; Albers, O.; et al. Safety and efficacy of selective and superselective intra-arterial cerebral infusion with blood-brain barrier disruption for glioma: a systematic review and meta-analysis. *J Neurointerv Surg* **2025**, doi:10.1136/jnis-2025-024057.
323. Lei, K.; Zhou, L.; Dan, M.; Yang, F.; Jian, T.; Xin, J.; Yu, Z.; Wang, Y. Trojan Horse Delivery Strategies of Natural Medicine Monomers: Challenges and Limitations in Improving Brain Targeting. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17030280.

324. D'Amico, R.S.; Aghi, M.K.; Vogelbaum, M.A.; Bruce, J.N. Convection-enhanced drug delivery for glioblastoma: a review. *J Neurooncol* **2021**, *151*, 415-427, doi:10.1007/s11060-020-03408-9.
325. Jahangiri, A.; Chin, A.T.; Flanigan, P.M.; Chen, R.; Bankiewicz, K.; Aghi, M.K. Convection-enhanced delivery in glioblastoma: a review of preclinical and clinical studies. *J Neurosurg* **2017**, *126*, 191-200, doi:10.3171/2016.1.Jns151591.
326. Mehta, A.M.; Sonabend, A.M.; Bruce, J.N. Convection-Enhanced Delivery. *Neurotherapeutics* **2017**, *14*, 358-371, doi:10.1007/s13311-017-0520-4.
327. Huttunen, K.M. Improving drug delivery to the brain: the prodrug approach. *Expert Opin Drug Deliv* **2024**, *21*, 683-693, doi:10.1080/17425247.2024.2355180.
328. Puris, E.; Fricker, G.; Gynther, M. Targeting Transporters for Drug Delivery to the Brain: Can We Do Better? *Pharm Res* **2022**, *39*, 1415-1455, doi:10.1007/s11095-022-03241-x.
329. Papakyriakopoulou, P.; Valsami, G. The nasal route for nose-to-brain drug delivery: advanced nasal formulations for CNS disorders. *Expert Opin Drug Deliv* **2025**, *22*, 823-839, doi:10.1080/17425247.2025.2489553.
330. Chu, C.; Jablonska, A.; Gao, Y.; Lan, X.; Lesniak, W.G.; Liang, Y.; Liu, G.; Li, S.; Magnus, T.; Pearl, M.; et al. Hyperosmolar blood-brain barrier opening using intra-arterial injection of hyperosmotic mannitol in mice under real-time MRI guidance. *Nat Protoc* **2022**, *17*, 76-94, doi:10.1038/s41596-021-00634-x.
331. Virtanen, P.S.; Ortiz, K.J.; Patel, A.; Blocher, W.A., 3rd; Richardson, A.M. Blood-Brain Barrier Disruption for the Treatment of Primary Brain Tumors: Advances in the Past Half-Decade. *Curr Oncol Rep* **2024**, *26*, 236-249, doi:10.1007/s11912-024-01497-7.
332. Chen, T.; Wang, W.; Ramos, N.M.; Schonthal, A. EXTH-66. NEO100 TRANSIENTLY OPENS UP THE BLOOD BRAIN BARRIER VIA TIGHT JUNCTION INHIBITION. *Neuro-Oncology* **2020**, *22*, ii101.
333. Trevisani, M.; Berselli, A.; Alberini, G.; Centonze, E.; Vercellino, S.; Cartocci, V.; Millo, E.; Ciobanu, D.Z.; Braccia, C.; Armirotti, A.; et al. A claudin5-binding peptide enhances the permeability of the blood-brain barrier in vitro. *Sci Adv* **2025**, *11*, eadq2616, doi:10.1126/sciadv.adq2616.
334. Parvar, S.J.; Wong, C.I.; Lewis, A.; Szychoth, E.; Morris, C.J.; Shorthouse, D.; Dziemidowicz, K. Convection-enhanced delivery for brain malignancies: Technical parameters, formulation strategies and clinical perspectives. *Adv Drug Deliv Rev* **2025**, *224*, 115657, doi:10.1016/j.addr.2025.115657.
335. Lonser, R.R.; Sarntinoranont, M.; Morrison, P.F.; Oldfield, E.H. Convection-enhanced delivery to the central nervous system. *J Neurosurg* **2015**, *122*, 697-706, doi:10.3171/2014.10.Jns14229.
336. Puris, E.; Gynther, M.; Auriola, S.; Huttunen, K.M. L-Type amino acid transporter 1 as a target for drug delivery. *Pharm Res* **2020**, *37*, 88, doi:10.1007/s11095-020-02826-8.
337. Chien, H.C.; Colas, C.; Finke, K.; Springer, S.; Stoner, L.; Zur, A.A.; Venteicher, B.; Campbell, J.; Hall, C.; Flint, A.; et al. Reevaluating the Substrate Specificity of the L-Type Amino Acid Transporter (LAT1). *J Med Chem* **2018**, *61*, 7358-7373, doi:10.1021/acs.jmedchem.8b01007.
338. Parvez, M.M.; Sadighi, A.; Ahn, Y.; Keller, S.F.; Enoru, J.O. Uptake Transporters at the Blood-Brain Barrier and Their Role in Brain Drug Disposition. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15102473.
339. Bahrami, K.; Järvinen, J.; Laitinen, T.; Reinisalo, M.; Honkakoski, P.; Poso, A.; Huttunen, K.M.; Rautio, J. Structural Features Affecting the Interactions and Transportability of LAT1-Targeted Phenylalanine Drug Conjugates. *Mol Pharm* **2023**, *20*, 206-218, doi:10.1021/acs.molpharmaceut.2c00594.
340. Hügele, A.; Löffler, S.; Molina, B.H.; Guillon, M.; Montaser, A.B.; Auriola, S.; Huttunen, K.M. Aminopeptidase B can bioconvert L-type amino acid transporter 1 (LAT1)-utilizing amide prodrugs in the brain. *Front Pharmacol* **2022**, *13*, 1034964, doi:10.3389/fphar.2022.1034964.
341. Huttunen, J.; Peltokangas, S.; Gynther, M.; Natunen, T.; Hiltunen, M.; Auriola, S.; Ruponen, M.; Vellonen, K.S.; Huttunen, K.M. L-Type Amino Acid Transporter 1 (LAT1/Lat1)-Utilizing Prodrugs Can Improve the Delivery of Drugs into Neurons, Astrocytes and Microglia. *Sci Rep* **2019**, *9*, 12860, doi:10.1038/s41598-019-49009-z.
342. Montaser, A.B.; Järvinen, J.; Löffler, S.; Huttunen, J.; Auriola, S.; Lehtonen, M.; Jalkanen, A.; Huttunen, K.M. L-Type Amino Acid Transporter 1 Enables the Efficient Brain Delivery of Small-Sized Prodrug across the Blood-Brain Barrier and into Human and Mouse Brain Parenchymal Cells. *ACS Chem Neurosci* **2020**, *11*, 4301-4315, doi:10.1021/acschemneuro.0c00564.

343. Tampio, J.; Huttunen, J.; Montaser, A.; Huttunen, K.M. Targeting of Perforin Inhibitor into the Brain Parenchyma Via a Prodrug Approach Can Decrease Oxidative Stress and Neuroinflammation and Improve Cell Survival. *Mol Neurobiol* **2020**, *57*, 4563-4577, doi:10.1007/s12035-020-02045-7.
344. Al Rihani, S.B.; Darakjian, L.I.; Deodhar, M.; Dow, P.; Turgeon, J.; Michaud, V. Disease-Induced Modulation of Drug Transporters at the Blood-Brain Barrier Level. *Int J Mol Sci* **2021**, *22*, doi:10.3390/ijms22073742.
345. Huttunen, J.; Gynther, M.; Vellonen, K.S.; Huttunen, K.M. L-Type amino acid transporter 1 (LAT1)-utilizing prodrugs are carrier-selective despite having low affinity for organic anion transporting polypeptides (OATPs). *Int J Pharm* **2019**, *571*, 118714, doi:10.1016/j.ijpharm.2019.118714.
346. Huttunen, J.; Gynther, M.; Vellonen, K.-S.; Huttunen, K.M. L-Type amino acid transporter 1 (LAT1)-utilizing prodrugs are carrier-selective despite having low affinity for organic anion transporting polypeptides (OATPs). *International Journal of Pharmaceutics* **2019**, *571*, 118714.
347. van de Waterbeemd, H.; Smith, D.A.; Jones, B.C. Lipophilicity in PK design: methyl, ethyl, futile. *J Comput Aided Mol Des* **2001**, *15*, 273-286, doi:10.1023/a:1008192010023.
348. Patel, M.M.; Patel, B.M. Crossing the Blood-Brain Barrier: Recent Advances in Drug Delivery to the Brain. *CNS Drugs* **2017**, *31*, 109-133, doi:10.1007/s40263-016-0405-9.
349. Sethi, B.; Kumar, V.; Mahato, K.; Coulter, D.W.; Mahato, R.I. Recent advances in drug delivery and targeting to the brain. *J Control Release* **2022**, *350*, 668-687, doi:10.1016/j.jconrel.2022.08.051.
350. Juhairiyah, F.; de Lange, E.C.M. Understanding Drug Delivery to the Brain Using Liposome-Based Strategies: Studies that Provide Mechanistic Insights Are Essential. *Aaps j* **2021**, *23*, 114, doi:10.1208/s12248-021-00648-z.
351. Gonzaga, R.V.; do Nascimento, L.A.; Santos, S.S.; Machado Sanches, B.A.; Giarolla, J.; Ferreira, E.I. Perspectives About Self-Immolative Drug Delivery Systems. *J Pharm Sci* **2020**, *109*, 3262-3281, doi:10.1016/j.xphs.2020.08.014.
352. Zhang, X.; Wang, S.; Cheng, G.; Yu, P.; Chang, J.; Chen, X. Cascade Drug-Release Strategy for Enhanced Anticancer Therapy. *Matter* **2021**, *4*, 26-53, doi:10.1016/j.matt.2020.10.002.
353. Muñoz-Sánchez, S.; Gong, J.; de la Mata, F.J.; Gillies, E.R.; García-Gallego, S. Functional Self-Immolative Hydrogels with Dendritic Cross-Linkers for Controlled Drug Delivery. *Chem Mater* **2025**, *37*, 5814-5824, doi:10.1021/acs.chemmater.5c01006.
354. van der Westhuyzen, A.E.; Hodson, L.E.; Pashikanti, G.; Fritzeimer, R.; Yeung, S.B.; Mancía, A.; Lian, D.R.; Tholath, P.J.; Paez, A.C.; Chavan, L.N.; et al. Rotamer-Controlled Self-Immolative Linkers Enable Tunable Release of Neurosteroid Oxime Prodrugs. *ACS Med Chem Lett* **2025**, *16*, 2022-2031, doi:10.1021/acsmchemlett.5c00452.
355. Wang, H.; Zheng, C.; Tian, F.; Xiao, Z.; Sun, Z.; Lu, L.; Dai, W.; Zhang, Q.; Mei, X. Improving the Dissolution Rate and Bioavailability of Curcumin via Co-Crystallization. *Pharmaceutics (Basel)* **2024**, *17*, doi:10.3390/ph17040489.
356. Nicolaescu, O.E.; Belu, I.; Mocanu, A.G.; Manda, V.C.; Rău, G.; Pîrvu, A.S.; Ionescu, C.; Ciulu-Costinescu, F.; Popescu, M.; Cioâlțeu, M.V. Cyclodextrins: Enhancing Drug Delivery, Solubility and Bioavailability for Modern Therapeutics. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17030288.
357. Lanevskij, K.; Japertas, P.; Didziapetris, R.; Petrauskas, A. Ionization-specific prediction of blood-brain permeability. *J Pharm Sci* **2009**, *98*, 122-134, doi:10.1002/jps.21405.
358. Xing, Y.; Meng, B.; Chen, Q. Cyclodextrin-Containing Drug Delivery Systems and Their Applications in Neurodegenerative Disorders. *Int J Mol Sci* **2024**, *25*, doi:10.3390/ijms251910834.
359. Văruț, R.M.; Popescu, A.I.S.; Gaman, S.; Niculescu, C.E.; Niculescu, A.; Dop, D.; Stepan, M.D.; Ionovici, N.; Singer, C.E.; Popescu, C. Cyclodextrin-Based Drug Delivery Systems for Depression: Improving Antidepressant Bioavailability and Targeted Central Nervous System Delivery. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17030355.
360. Thiberville, L.; Faivre, V.; Sizun, C.; Dehouck, M.P.; Landry, C.; Baati, R.; Tsapis, N. Cyclodextrin-based formulations for delivering broad-spectrum nerve agent antidote to the central nervous system: stability, physicochemical characterization and application in a human blood-brain barrier model. *Int J Pharm* **2025**, *674*, 125505, doi:10.1016/j.ijpharm.2025.125505.

361. Guo, M.; Sun, X.; Chen, J.; Cai, T. Pharmaceutical cocrystals: A review of preparations, physicochemical properties and applications. *Acta Pharm Sin B* **2021**, *11*, 2537-2564, doi:10.1016/j.apsb.2021.03.030.
362. Spiridon, I.; Anghel, N. Cyclodextrins as Multifunctional Platforms in Drug Delivery and Beyond: Structural Features, Functional Applications, and Future Trends. *Molecules* **2025**, *30*, doi:10.3390/molecules30143044.
363. Duan, C.; Liu, W.; Tao, Y.; Liang, F.; Chen, Y.; Xiao, X.; Zhang, G.; Chen, Y.; Hao, C. Two Novel Palbociclib-Resorcinol and Palbociclib-Orcinol Cocrystals with Enhanced Solubility and Dissolution Rate. *Pharmaceutics* **2021**, *14*, doi:10.3390/pharmaceutics14010023.
364. Peterson, B.; Weyers, M.; Steenekamp, J.H.; Steyn, J.D.; Gouws, C.; Hamman, J.H. Drug Bioavailability Enhancing Agents of Natural Origin (Bioenhancers) that Modulate Drug Membrane Permeation and Pre-Systemic Metabolism. *Pharmaceutics* **2019**, *11*, doi:10.3390/pharmaceutics11010033.
365. Zhang, Q.L.; Fu, B.M.; Zhang, Z.J. Borneol, a novel agent that improves central nervous system drug delivery by enhancing blood-brain barrier permeability. *Drug Deliv* **2017**, *24*, 1037-1044, doi:10.1080/10717544.2017.1346002.
366. Neaz, S.; Alam, M.M.; Imran, A.B. Advancements in cyclodextrin-based controlled drug delivery: Insights into pharmacokinetic and pharmacodynamic profiles. *Heliyon* **2024**, *10*, e39917, doi:10.1016/j.heliyon.2024.e39917.
367. de Souza, M.M.; Gini, A.L.R.; Moura, J.A.; Scarim, C.B.; Chin, C.M.; Dos Santos, J.L. Prodrug Approach as a Strategy to Enhance Drug Permeability. *Pharmaceutics (Basel)* **2025**, *18*, doi:10.3390/ph18030297.
368. Dong, X.; Brahma, R.K.; Fang, C.; Yao, S.Q. Stimulus-responsive self-assembled prodrugs in cancer therapy. *Chem Sci* **2022**, *13*, 4239-4269, doi:10.1039/d2sc01003h.
369. Kopeček, J.; Yang, J. Polymer nanomedicines. *Adv Drug Deliv Rev* **2020**, *156*, 40-64, doi:10.1016/j.addr.2020.07.020.
370. Guo, H.; Mi, P. Polymer-drug and polymer-protein conjugated nanocarriers: Design, drug delivery, imaging, therapy, and clinical applications. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* **2024**, *16*, e1988, doi:10.1002/wnan.1988.
371. Jiang, Y.; Stenzel, M. Drug Delivery Vehicles Based on Albumin-Polymer Conjugates. *Macromol Biosci* **2016**, *16*, 791-802, doi:10.1002/mabi.201500453.
372. Mi, P. Stimuli-responsive nanocarriers for drug delivery, tumor imaging, therapy and theranostics. *Theranostics* **2020**, *10*, 4557-4588, doi:10.7150/thno.38069.
373. Lee, Y.; Thompson, D.H. Stimuli-responsive liposomes for drug delivery. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* **2017**, *9*, doi:10.1002/wnan.1450.
374. Wu, X.; Shu, Y.; Zheng, Y.; Zhang, P.; Cong, H.; Zou, Y.; Cai, H.; Zha, Z. Recent Advances in Nanomedicine: Cutting-Edge Research on Nano-PROTAC Delivery Systems for Cancer Therapy. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17081037.
375. Li, X.; Peng, X.; Zoulikha, M.; Bofo, G.F.; Magar, K.T.; Ju, Y.; He, W. Multifunctional nanoparticle-mediated combining therapy for human diseases. *Signal Transduct Target Ther* **2024**, *9*, 1, doi:10.1038/s41392-023-01668-1.
376. Đorđević, S.; Gonzalez, M.M.; Conejos-Sánchez, I.; Carreira, B.; Pozzi, S.; Acúrcio, R.C.; Satchi-Fainaro, R.; Florindo, H.F.; Vicent, M.J. Current hurdles to the translation of nanomedicines from bench to the clinic. *Drug Deliv Transl Res* **2022**, *12*, 500-525, doi:10.1007/s13346-021-01024-2.
377. Ahmad, A.; Imran, M.; Sharma, N. Precision Nanotoxicology in Drug Development: Current Trends and Challenges in Safety and Toxicity Implications of Customized Multifunctional Nanocarriers for Drug-Delivery Applications. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14112463.
378. Buya, A.B.; Mahlangu, P.; Witika, B.A. From lab to industrial development of lipid nanocarriers using quality by design approach. *Int J Pharm X* **2024**, *8*, 100266, doi:10.1016/j.ijpx.2024.100266.
379. Sammasagi, S.S.; Sutar, K.P.; Hooli, S. Scale-up and quality control challenges in the industrial manufacturing of nanoformulations: current trends and future perspectives. *IJSAT-International Journal on Science and Technology* **2025**, *16*.
380. Gyimesi, G.; Hediger, M.A. Transporter-Mediated Drug Delivery. *Molecules* **2023**, *28*, doi:10.3390/molecules28031151.

381. Puris, E.; Gynther, M.; Huttunen, J.; Petsalo, A.; Huttunen, K.M. L-type amino acid transporter 1 utilizing prodrugs: How to achieve effective brain delivery and low systemic exposure of drugs. *J Control Release* **2017**, *261*, 93-104, doi:10.1016/j.jconrel.2017.06.023.
382. Montaser, A.; Lehtonen, M.; Gynther, M.; Huttunen, K.M. L-Type Amino Acid Transporter 1-Utilizing Prodrugs of Ketoprofen Can Efficiently Reduce Brain Prostaglandin Levels. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12040344.
383. Patching, S.G. Glucose Transporters at the Blood-Brain Barrier: Function, Regulation and Gateways for Drug Delivery. *Mol Neurobiol* **2017**, *54*, 1046-1077, doi:10.1007/s12035-015-9672-6.
384. Suzuki, K.; Miura, Y.; Mochida, Y.; Miyazaki, T.; Toh, K.; Anraku, Y.; Melo, V.; Liu, X.; Ishii, T.; Nagano, O.; et al. Glucose transporter 1-mediated vascular translocation of nanomedicines enhances accumulation and efficacy in solid tumors. *J Control Release* **2019**, *301*, 28-41, doi:10.1016/j.jconrel.2019.02.021.
385. Vijay, N.; Morris, M.E. Role of monocarboxylate transporters in drug delivery to the brain. *Curr Pharm Des* **2014**, *20*, 1487-1498, doi:10.2174/13816128113199990462.
386. Wang, G.; Zhao, L.; Jiang, Q.; Sun, Y.; Zhao, D.; Sun, M.; He, Z.; Sun, J.; Wang, Y. Intestinal OCTN2- and MCT1-targeted drug delivery to improve oral bioavailability. *Asian J Pharm Sci* **2020**, *15*, 158-173, doi:10.1016/j.ajps.2020.02.002.
387. Sun, Y.; Zhao, D.; Wang, G.; Jiang, Q.; Guo, M.; Kan, Q.; He, Z.; Sun, J. A novel oral prodrug-targeting transporter MCT 1: 5-fluorouracil-dicarboxylate monoester conjugates. *Asian J Pharm Sci* **2019**, *14*, 631-639, doi:10.1016/j.ajps.2019.04.001.
388. Hu, C.; Tao, L.; Cao, X.; Chen, L. The solute carrier transporters and the brain: Physiological and pharmacological implications. *Asian J Pharm Sci* **2020**, *15*, 131-144, doi:10.1016/j.ajps.2019.09.002.
389. Huttunen, J.; Adla, S.K.; Markowicz-Piasecka, M.; Huttunen, K.M. Increased/Targeted Brain (Pro)Drug Delivery via Utilization of Solute Carriers (SLCs). *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14061234.
390. Begley, D.J. ABC transporters and the blood-brain barrier. *Curr Pharm Des* **2004**, *10*, 1295-1312, doi:10.2174/1381612043384844.
391. Colclough, N.; Alluri, R.V.; Tucker, J.W.; Gozalpour, E.; Li, D.; Du, H.; Li, W.; Harlfinger, S.; O'Neill, D.J.; Sproat, G.G.; et al. Utilizing a Dual Human Transporter MDCKII-MDR1-BCRP Cell Line to Assess Efflux at the Blood Brain Barrier. *Drug Metab Dispos* **2024**, *52*, 95-105, doi:10.1124/dmd.123.001476.
392. Tian, J.; Han, Z.; Song, D.; Peng, Y.; Xiong, M.; Chen, Z.; Duan, S.; Zhang, L. Engineered Exosome for Drug Delivery: Recent Development and Clinical Applications. *Int J Nanomedicine* **2023**, *18*, 7923-7940, doi:10.2147/ijn.S444582.
393. Koh, H.B.; Kim, H.J.; Kang, S.W.; Yoo, T.H. Exosome-Based Drug Delivery: Translation from Bench to Clinic. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15082042.
394. Zeng, H.; Guo, S.; Ren, X.; Wu, Z.; Liu, S.; Yao, X. Current Strategies for Exosome Cargo Loading and Targeting Delivery. *Cells* **2023**, *12*, doi:10.3390/cells12101416.
395. Hade, M.D.; Suire, C.N.; Suo, Z. Mesenchymal Stem Cell-Derived Exosomes: Applications in Regenerative Medicine. *Cells* **2021**, *10*, doi:10.3390/cells10081959.
396. Ge, Y.; Wu, J.; Zhang, L.; Huang, N.; Luo, Y. A New Strategy for the Regulation of Neuroinflammation: Exosomes Derived from Mesenchymal Stem Cells. *Cell Mol Neurobiol* **2024**, *44*, 24, doi:10.1007/s10571-024-01460-x.
397. Zubair, M.; Abouelnazar, F.A.; Iqbal, M.A.; Pan, J.; Zheng, X.; Chen, T.; Shen, W.; Yin, J.; Yan, Y.; Liu, P.; et al. Mesenchymal stem cell-derived exosomes as a plausible immunomodulatory therapeutic tool for inflammatory diseases. *Front Cell Dev Biol* **2025**, *13*, 1563427, doi:10.3389/fcell.2025.1563427.
398. Fu, S.; Wang, Y.; Xia, X.; Zheng, J.C. Exosome engineering: Current progress in cargo loading and targeted delivery. *NanoImpact* **2020**, *20*, 100261.
399. Xi, X.M.; Xia, S.J.; Lu, R. Drug loading techniques for exosome-based drug delivery systems. *Pharmazie* **2021**, *76*, 61-67, doi:10.1691/ph.2021.0128.
400. Chen, C.; Sun, M.; Wang, J.; Su, L.; Lin, J.; Yan, X. Active cargo loading into extracellular vesicles: Highlights the heterogeneous encapsulation behaviour. *J Extracell Vesicles* **2021**, *10*, e12163, doi:10.1002/jev.2.12163.

401. Ahmed, W.; Mushtaq, A.; Ali, S.; Khan, N.; Liang, Y.; Duan, L. Engineering Approaches for Exosome Cargo Loading and Targeted Delivery: Biological versus Chemical Perspectives. *ACS Biomater Sci Eng* **2024**, *10*, 5960-5976, doi:10.1021/acsbomaterials.4c00856.
402. Kimiz-Gebologlu, I.; Oncel, S.S. Exosomes: Large-scale production, isolation, drug loading efficiency, and biodistribution and uptake. *J Control Release* **2022**, *347*, 533-543, doi:10.1016/j.jconrel.2022.05.027.
403. Ahn, S.H.; Ryu, S.W.; Choi, H.; You, S.; Park, J.; Choi, C. Manufacturing Therapeutic Exosomes: from Bench to Industry. *Mol Cells* **2022**, *45*, 284-290, doi:10.14348/molcells.2022.2033.
404. Wang, C.K.; Tsai, T.H.; Lee, C.H. Regulation of exosomes as biologic medicines: Regulatory challenges faced in exosome development and manufacturing processes. *Clin Transl Sci* **2024**, *17*, e13904, doi:10.1111/cts.13904.
405. Verma, N.; Arora, S. Navigating the Global Regulatory Landscape for Exosome-Based Therapeutics: Challenges, Strategies, and Future Directions. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17080990.
406. Lian, M.Q.; Chng, W.H.; Liang, J.; Yeo, H.Q.; Lee, C.K.; Belaid, M.; Tollemeto, M.; Wacker, M.G.; Czarny, B.; Pastorin, G. Plant-derived extracellular vesicles: Recent advancements and current challenges on their use for biomedical applications. *J Extracell Vesicles* **2022**, *11*, e12283, doi:10.1002/jev2.12283.
407. Lo, K.J.; Wang, M.H.; Ho, C.T.; Pan, M.H. Plant-Derived Extracellular Vesicles: A New Revolutionization of Modern Healthy Diets and Biomedical Applications. *J Agric Food Chem* **2024**, *72*, 2853-2878, doi:10.1021/acs.jafc.3c06867.
408. Kim, M.; Jang, H.; Kim, W.; Kim, D.; Park, J.H. Therapeutic Applications of Plant-Derived Extracellular Vesicles as Antioxidants for Oxidative Stress-Related Diseases. *Antioxidants (Basel)* **2023**, *12*, doi:10.3390/antiox12061286.
409. Karamanidou, T.; Tsouknidas, A. Plant-Derived Extracellular Vesicles as Therapeutic Nanocarriers. *Int J Mol Sci* **2021**, *23*, doi:10.3390/ijms23010191.
410. Alzahrani, F.A.; Khan, M.I.; Kameli, N.; Alsahafi, E.; Riza, Y.M. Plant-Derived Extracellular Vesicles and Their Exciting Potential as the Future of Next-Generation Drug Delivery. *Biomolecules* **2023**, *13*, doi:10.3390/biom13050839.
411. Shao, M.; Jin, X.; Chen, S.; Yang, N.; Feng, G. Plant-derived extracellular vesicles -a novel clinical anti-inflammatory drug carrier worthy of investigation. *Biomed Pharmacother* **2023**, *169*, 115904, doi:10.1016/j.biopha.2023.115904.
412. Calzoni, E.; Bertoldi, A.; Cusumano, G.; Buratta, S.; Urbanelli, L.; Emiliani, C. Plant-derived extracellular vesicles: natural nanocarriers for biotechnological drugs. *Processes* **2024**, *12*, 2938.
413. Zuo, Y.; Zhang, J.; Sun, B.; Wang, X.; Wang, R.; Tian, S.; Miao, M. A New Perspective on Regenerative Medicine: Plant-Derived Extracellular Vesicles. *Biomolecules* **2025**, *15*, doi:10.3390/biom15081095.
414. Zhu, Y.; Zhao, J.; Ding, H.; Qiu, M.; Xue, L.; Ge, D.; Wen, G.; Ren, H.; Li, P.; Wang, J. Applications of plant-derived extracellular vesicles in medicine. *MedComm (2020)* **2024**, *5*, e741, doi:10.1002/mco2.741.
415. Huang, J.; Chen, L.; Li, W.; Chang, C.J. Anti-inflammatory and antioxidative effects of *Perilla frutescens*-derived extracellular vesicles: Insights from Zebrafish models. *Mol Immunol* **2025**, *182*, 126-138, doi:10.1016/j.molimm.2025.04.008.
416. Lai, W.Y.; Chuang, C.W.; Huang, Y.C.; Huang, C.J. Therapeutic potential of plant-derived small extracellular vesicles in sepsis: A network meta-analysis. *Pharmacol Res* **2025**, *217*, 107795, doi:10.1016/j.phrs.2025.107795.
417. Cui, L.; Perini, G.; Minopoli, A.; Palmieri, V.; De Spirito, M.; Papi, M. Plant-derived extracellular vesicles as a natural drug delivery platform for glioblastoma therapy: A dual role in preserving endothelial integrity while modulating the tumor microenvironment. *Int J Pharm X* **2025**, *10*, 100349, doi:10.1016/j.ijpx.2025.100349.
418. Xu, X.; Xu, L.; Wen, C.; Xia, J.; Zhang, Y.; Liang, Y. Programming assembly of biomimetic exosomes: An emerging theranostic nanomedicine platform. *Mater Today Bio* **2023**, *22*, 100760, doi:10.1016/j.mtbio.2023.100760.
419. Lu, M.; Huang, Y. Bioinspired exosome-like therapeutics and delivery nanoplatfoms. *Biomaterials* **2020**, *242*, 119925, doi:10.1016/j.biomaterials.2020.119925.

420. Chen, Y.; Douanne, N.; Wu, T.; Kaur, I.; Tsering, T.; Erzingatzian, A.; Nadeau, A.; Juncker, D.; Nerguizian, V.; Burnier, J.V. Leveraging nature's nanocarriers: Translating insights from extracellular vesicles to biomimetic synthetic vesicles for biomedical applications. *Sci Adv* **2025**, *11*, eads5249, doi:10.1126/sciadv.ads5249.
421. Poinot, V.; Pizzinat, N.; Ong-Meang, V. Engineered and Mimicked Extracellular Nanovesicles for Therapeutic Delivery. *Nanomaterials (Basel)* **2024**, *14*, doi:10.3390/nano14070639.
422. Antimisariar, S.G.; Mourtas, S.; Marazioti, A. Exosomes and Exosome-Inspired Vesicles for Targeted Drug Delivery. *Pharmaceutics* **2018**, *10*, doi:10.3390/pharmaceutics10040218.
423. Kooijmans, S.A.; Vader, P.; van Dommelen, S.M.; van Solinge, W.W.; Schiffelers, R.M. Exosome mimetics: a novel class of drug delivery systems. *Int J Nanomedicine* **2012**, *7*, 1525-1541, doi:10.2147/ijn.S29661.
424. Rayamajhi, S.; Nguyen, T.D.T.; Marasini, R.; Aryal, S. Macrophage-derived exosome-mimetic hybrid vesicles for tumor targeted drug delivery. *Acta Biomater* **2019**, *94*, 482-494, doi:10.1016/j.actbio.2019.05.054.
425. Ailuno, G.; Baldassari, S.; Lai, F.; Florio, T.; Caviglioli, G. Exosomes and Extracellular Vesicles as Emerging Theranostic Platforms in Cancer Research. *Cells* **2020**, *9*, doi:10.3390/cells9122569.
426. Jagtiani, E.; Yeolekar, M.; Naik, S.; Patravale, V. In vitro blood brain barrier models: An overview. *J Control Release* **2022**, *343*, 13-30, doi:10.1016/j.jconrel.2022.01.011.
427. Sivandzade, F.; Cucullo, L. In-vitro blood-brain barrier modeling: A review of modern and fast-advancing technologies. *J Cereb Blood Flow Metab* **2018**, *38*, 1667-1681, doi:10.1177/0271678x18788769.
428. Wang, Y.I.; Abaci, H.E.; Shuler, M.L. Microfluidic blood-brain barrier model provides in vivo-like barrier properties for drug permeability screening. *Biotechnol Bioeng* **2017**, *114*, 184-194, doi:10.1002/bit.26045.
429. Thomsen, M.S.; Humle, N.; Hede, E.; Moos, T.; Burkhart, A.; Thomsen, L.B. The blood-brain barrier studied in vitro across species. *PLoS One* **2021**, *16*, e0236770, doi:10.1371/journal.pone.0236770.
430. Bernard-Patrzynski, F.; Lécuyer, M.A.; Puscas, I.; Boukhatem, I.; Charabati, M.; Bourbonnière, L.; Ramassamy, C.; Leclair, G.; Prat, A.; Roullin, V.G. Isolation of endothelial cells, pericytes and astrocytes from mouse brain. *PLoS One* **2019**, *14*, e0226302, doi:10.1371/journal.pone.0226302.
431. Park, J.S.; Choe, K.; Khan, A.; Jo, M.H.; Park, H.Y.; Kang, M.H.; Park, T.J.; Kim, M.O. Establishing Co-Culture Blood-Brain Barrier Models for Different Neurodegeneration Conditions to Understand Its Effect on BBB Integrity. *Int J Mol Sci* **2023**, *24*, doi:10.3390/ijms24065283.
432. Jiang, L.; Li, S.; Zheng, J.; Li, Y.; Huang, H. Recent Progress in Microfluidic Models of the Blood-Brain Barrier. *Micromachines (Basel)* **2019**, *10*, doi:10.3390/mi10060375.
433. Goldeman, C.; Andersen, M.; Al-Robai, A.; Buchholtz, T.; Svane, N.; Ozgür, B.; Holst, B.; Shusta, E.; Hall, V.J.; Saaby, L.; et al. Human induced pluripotent stem cells (BIONi010-C) generate tight cell monolayers with blood-brain barrier traits and functional expression of large neutral amino acid transporter 1 (SLC7A5). *Eur J Pharm Sci* **2021**, *156*, 105577, doi:10.1016/j.ejps.2020.105577.
434. Singh, N.R.; Gromnicova, R.; Brachner, A.; Kraev, I.; Romero, I.A.; Neuhaus, W.; Male, D. A hydrogel model of the human blood-brain barrier using differentiated stem cells. *PLoS One* **2023**, *18*, e0283954, doi:10.1371/journal.pone.0283954.
435. Campisi, M.; Shin, Y.; Osaki, T.; Hajal, C.; Chiono, V.; Kamm, R.D. 3D self-organized microvascular model of the human blood-brain barrier with endothelial cells, pericytes and astrocytes. *Biomaterials* **2018**, *180*, 117-129, doi:10.1016/j.biomaterials.2018.07.014.
436. Srinivasan, B.; Kolli, A.R.; Esch, M.B.; Abaci, H.E.; Shuler, M.L.; Hickman, J.J. TEER measurement techniques for in vitro barrier model systems. *J Lab Autom* **2015**, *20*, 107-126, doi:10.1177/2211068214561025.
437. Andjelkovic, A.V.; Stamatovic, S.M.; Phillips, C.M.; Martinez-Revollar, G.; Keep, R.F. Modeling blood-brain barrier pathology in cerebrovascular disease in vitro: current and future paradigms. *Fluids Barriers CNS* **2020**, *17*, 44, doi:10.1186/s12987-020-00202-7.
438. O'Brown, N.M.; Pfau, S.J.; Gu, C. Bridging barriers: a comparative look at the blood-brain barrier across organisms. *Genes Dev* **2018**, *32*, 466-478, doi:10.1101/gad.309823.117.
439. Watanabe, D.; Nakagawa, S.; Morofuji, Y.; Tóth, A.E.; Vastag, M.; Aruga, J.; Niwa, M.; Deli, M.A. Characterization of a Primate Blood-Brain Barrier Co-Culture Model Prepared from Primary Brain Endothelial Cells, Pericytes and Astrocytes. *Pharmaceutics* **2021**, *13*, doi:10.3390/pharmaceutics13091484.

440. Zhang, W.; Liu, Q.Y.; Haqqani, A.S.; Liu, Z.; Sodja, C.; Leclerc, S.; Baumann, E.; Delaney, C.E.; Brunette, E.; Stanimirovic, D.B. Differential Expression of ABC Transporter Genes in Brain Vessels vs. Peripheral Tissues and Vessels from Human, Mouse and Rat. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15051563.
441. Hoshi, Y.; Uchida, Y.; Tachikawa, M.; Inoue, T.; Ohtsuki, S.; Terasaki, T. Quantitative atlas of blood-brain barrier transporters, receptors, and tight junction proteins in rats and common marmoset. *J Pharm Sci* **2013**, *102*, 3343-3355, doi:10.1002/jps.23575.
442. Hussner, J.; Foletti, A.; Seibert, I.; Fuchs, A.; Schuler, E.; Malagnino, V.; Grube, M.; Meyer Zu Schwabedissen, H.E. Differences in transport function of the human and rat orthologue of the Organic Anion Transporting Polypeptide 2B1 (OATP2B1). *Drug Metab Pharmacokinet* **2021**, *41*, 100418, doi:10.1016/j.dmpk.2021.100418.
443. Strazielle, N.; Blondel, S.; Confais, J.; El Khoury, R.; Contamin, H.; Ghersi-Egea, J.F. Molecular determinants of neuroprotection in blood-brain interfaces of the cynomolgus monkey. *Front Pharmacol* **2025**, *16*, 1523819, doi:10.3389/fphar.2025.1523819.
444. Chaves, C.; Do, T.M.; Cegarra, C.; Roudières, V.; Tolou, S.; Thill, G.; Rocher, C.; Didier, M.; Lesuisse, D. Non-Human Primate Blood-Brain Barrier and In Vitro Brain Endothelium: From Transcriptome to the Establishment of a New Model. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12100967.
445. Plotnikov, M.B.; Anishchenko, A.M.; Khlebnikov, A.I.; Schepetkin, I.A. Regulation of Blood-Brain Barrier Permeability via JNK Signaling Pathway: Mechanisms and Potential Therapeutic Strategies for Ischemic Stroke, Alzheimer's Disease and Brain Tumors. *Molecules* **2025**, *30*, doi:10.3390/molecules30112353.
446. Vazana, U.; Veksler, R.; Pell, G.S.; Prager, O.; Fassler, M.; Chassidim, Y.; Roth, Y.; Shahar, H.; Zangen, A.; Raccach, R.; et al. Glutamate-Mediated Blood-Brain Barrier Opening: Implications for Neuroprotection and Drug Delivery. *J Neurosci* **2016**, *36*, 7727-7739, doi:10.1523/jneurosci.0587-16.2016.
447. Brynildsen, J.K.; Rajan, K.; Henderson, M.X.; Bassett, D.S. Network models to enhance the translational impact of cross-species studies. *Nat Rev Neurosci* **2023**, *24*, 575-588, doi:10.1038/s41583-023-00720-x.
448. de Lange, E.C.M.; van den Brink, W.; Yamamoto, Y.; de Witte, W.E.A.; Wong, Y.C. Novel CNS drug discovery and development approach: model-based integration to predict neuro-pharmacokinetics and pharmacodynamics. *Expert Opin Drug Discov* **2017**, *12*, 1207-1218, doi:10.1080/17460441.2017.1380623.
449. Vallianatou, T.; Tsopelas, F.; Tsantili-Kakoulidou, A. Prediction Models for Brain Distribution of Drugs Based on Biomimetic Chromatographic Data. *Molecules* **2022**, *27*, doi:10.3390/molecules27123668.
450. Loryan, I.; Reichel, A.; Feng, B.; Bundgaard, C.; Shaffer, C.; Kalvass, C.; Bednarczyk, D.; Morrison, D.; Lesuisse, D.; Hoppe, E.; et al. Unbound Brain-to-Plasma Partition Coefficient,  $K_{(p,uu,brain)}$ -a Game Changing Parameter for CNS Drug Discovery and Development. *Pharm Res* **2022**, *39*, 1321-1341, doi:10.1007/s11095-022-03246-6.
451. Kalvass, J.C.; Maurer, T.S.; Pollack, G.M. Use of plasma and brain unbound fractions to assess the extent of brain distribution of 34 drugs: comparison of unbound concentration ratios to in vivo p-glycoprotein efflux ratios. *Drug Metab Dispos* **2007**, *35*, 660-666, doi:10.1124/dmd.106.012294.
452. Li, J.; Jiang, J.; Wu, J.; Bao, X.; Sanai, N. Physiologically Based Pharmacokinetic Modeling of Central Nervous System Pharmacokinetics of CDK4/6 Inhibitors to Guide Selection of Drug and Dosing Regimen for Brain Cancer Treatment. *Clin Pharmacol Ther* **2021**, *109*, 494-506, doi:10.1002/cpt.2021.
453. Sundheimer, J.K.; Benzel, J.; Longuespée, R.; Burhenne, J.; Pfister, S.M.; Maaß, K.K.; Sauter, M.; Pajtler, K.W. Experimental Insights and Recommendations for Successfully Performing Cerebral Microdialysis With Hydrophobic Drug Candidates. *Clinical and translational science* **2025**, *18*, e70226.
454. Fridén, M.; Gupta, A.; Antonsson, M.; Bredberg, U.; Hammarlund-Udenaes, M. In vitro methods for estimating unbound drug concentrations in the brain interstitial and intracellular fluids. *Drug Metab Dispos* **2007**, *35*, 1711-1719, doi:10.1124/dmd.107.015222.
455. Liu, X.; Van Natta, K.; Yeo, H.; Vilenski, O.; Weller, P.E.; Worboys, P.D.; Monshouwer, M. Unbound drug concentration in brain homogenate and cerebral spinal fluid at steady state as a surrogate for unbound concentration in brain interstitial fluid. *Drug Metab Dispos* **2009**, *37*, 787-793, doi:10.1124/dmd.108.024125.
456. Yamamoto, Y.; Väitalo, P.A.; Wong, Y.C.; Huntjens, D.R.; Proost, J.H.; Vermeulen, A.; Krauwinkel, W.; Beukers, M.W.; Kokki, H.; Kokki, M.; et al. Prediction of human CNS pharmacokinetics using a

- physiologically-based pharmacokinetic modeling approach. *Eur J Pharm Sci* **2018**, *112*, 168-179, doi:10.1016/j.ejps.2017.11.011.
457. Mu, R.J.; Liu, T.L.; Liu, X.D.; Liu, L. PBPK-PD model for predicting morphine pharmacokinetics, CNS effects and naloxone antagonism in humans. *Acta Pharmacol Sin* **2024**, *45*, 1752-1764, doi:10.1038/s41401-024-01255-2.
458. Wu, K.; Li, X.; Zhou, Z.; Zhao, Y.; Su, M.; Cheng, Z.; Wu, X.; Huang, Z.; Jin, X.; Li, J.; et al. Predicting pharmacodynamic effects through early drug discovery with artificial intelligence-physiologically based pharmacokinetic (AI-PBPK) modelling. *Front Pharmacol* **2024**, *15*, 1330855, doi:10.3389/fphar.2024.1330855.
459. Zou, Y.; Yuan, H.; Guo, Z.; Guo, T.; Fu, Z.; Wang, R.; Xu, D.; Wang, Q.; Wang, T.; Chen, L. Predicting the Brain-To-Plasma Unbound Partition Coefficient of Compounds via Formula-Guided Network. *J Chem Inf Model* **2025**, *65*, 5099-5112, doi:10.1021/acs.jcim.5c00590.
460. Chung, K.J.; Abdelhafez, Y.G.; Spencer, B.A.; Jones, T.; Tran, Q.; Nardo, L.; Chen, M.S., Jr.; Sarkar, S.; Medici, V.; Lyo, V.; et al. Quantitative PET imaging and modeling of molecular blood-brain barrier permeability. *Nat Commun* **2025**, *16*, 3076, doi:10.1038/s41467-025-58356-7.
461. Mishra, V.; Kumari, N.; Vyas, M.; Aljabali, A.A.A.; Chattaraj, A.; Mishra, Y. Advances in multimodal imaging techniques in nanomedicine: enhancing drug delivery precision. *RSC Adv* **2025**, *15*, 27187-27209, doi:10.1039/d5ra03255e.
462. Varatharaj, A.; Liljeroth, M.; Darekar, A.; Larsson, H.B.W.; Galea, I.; Cramer, S.P. Blood-brain barrier permeability measured using dynamic contrast-enhanced magnetic resonance imaging: a validation study. *J Physiol* **2019**, *597*, 699-709, doi:10.1113/jp276887.
463. Mishra, A.; Payne, C.; Carrascal-Miniño, A.; Sunassee, K.; Halbherr, S.; Pouliopoulos, A.N.; de Rosales, R.T. PET imaging for non-invasive monitoring of <sup>89</sup>Zr-Talidox delivery to the brain following focused ultrasound-mediated blood-brain barrier opening. *Journal of Controlled Release* **2025**, *387*, 114183.
464. Hugon, G.; Goutal, S.; Dauba, A.; Breuil, L.; Larrat, B.; Winkeler, A.; Novell, A.; Tournier, N. [(18)F]2-Fluoro-2-deoxy-sorbitol PET Imaging for Quantitative Monitoring of Enhanced Blood-Brain Barrier Permeability Induced by Focused Ultrasound. *Pharmaceutics* **2021**, *13*, doi:10.3390/pharmaceutics13111752.
465. Pa, C.; Shen, S.; Dai, Y.; Wu, M. Bibliometric analysis of neural injury biomarkers in neurodegenerative diseases: research trends and future perspectives. *Front Hum Neurosci* **2025**, *19*, 1614132, doi:10.3389/fnhum.2025.1614132.
466. Gerber, K.S.; Alvarez, G.; Alamian, A.; Behar-Zusman, V.; Downs, C.A. Biomarkers of Neuroinflammation in Traumatic Brain Injury. *Clin Nurs Res* **2022**, *31*, 1203-1218, doi:10.1177/10547738221107081.
467. Mondello, S.; Guedes, V.A.; Lai, C.; Czeiter, E.; Amrein, K.; Kobeissy, F.; Mechref, Y.; Jeromin, A.; Mithani, S.; Martin, C.; et al. Circulating Brain Injury Exosomal Proteins following Moderate-To-Severe Traumatic Brain Injury: Temporal Profile, Outcome Prediction and Therapy Implications. *Cells* **2020**, *9*, doi:10.3390/cells9040977.
468. Lopez, B.G.C.; Kohale, I.N.; Du, Z.; Korsunsky, I.; Abdelmoula, W.M.; Dai, Y.; Stopka, S.A.; Gaglia, G.; Randall, E.C.; Regan, M.S.; et al. Multimodal platform for assessing drug distribution and response in clinical trials. *Neuro Oncol* **2022**, *24*, 64-77, doi:10.1093/neuonc/noab197.
469. Barbalho, S.M.; Leme Boaro, B.; da Silva Camarinha Oliveira, J.; Patočka, J.; Barbalho Lamas, C.; Tanaka, M.; Laurindo, L.F. Molecular Mechanisms Underlying Neuroinflammation Intervention with Medicinal Plants: A Critical and Narrative Review of the Current Literature. *Pharmaceutics (Basel)* **2025**, *18*, doi:10.3390/ph18010133.
470. Tanaka, M.; Battaglia, S.; Liloia, D. Navigating Neurodegeneration: Integrating Biomarkers, Neuroinflammation, and Imaging in Parkinson's, Alzheimer's, and Motor Neuron Disorders. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13051045.
471. Barbalho, S.M.; Laurindo, L.F.; de Oliveira Zanuso, B.; da Silva, R.M.S.; Gallerani Caglioni, L.; Nunes Junqueira de Moraes, V.B.F.; Fornari Laurindo, L.; Dogani Rodrigues, V.; da Silva Camarinha Oliveira, J.; Beluce, M.E.; et al. AdipoRon's Impact on Alzheimer's Disease-A Systematic Review and Meta-Analysis. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26020484.
472. Tanaka, M. From Serendipity to Precision: Integrating AI, Multi-Omics, and Human-Specific Models for Personalized Neuropsychiatric Care. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13010167.

473. Shamul, J.G.; Wang, Z.; Gong, H.; Ou, W.; White, A.M.; Moniz-Garcia, D.P.; Gu, S.; Clyne, A.M.; Quiñones-Hinojosa, A.; He, X. Meta-analysis of the make-up and properties of in vitro models of the healthy and diseased blood-brain barrier. *Nat Biomed Eng* **2025**, *9*, 566-598, doi:10.1038/s41551-024-01250-2.
474. Stone, N.L.; England, T.J.; O'Sullivan, S.E. A Novel Transwell Blood Brain Barrier Model Using Primary Human Cells. *Front Cell Neurosci* **2019**, *13*, 230, doi:10.3389/fncel.2019.00230.
475. Wu, Y.; Chen, Z.; Chen, F.; Su, J.; Han, J.; Liu, S. Using Regular Porous Membrane-Based Blood-Brain Barrier Model to Screen Brain-Targeted Drugs with Nanochannel Electrochemistry. *Anal Chem* **2025**, *97*, 7968-7977, doi:10.1021/acs.analchem.5c00365.
476. Burgio, F.; Gaiser, C.; Brady, K.; Gatta, V.; Class, R.; Schrage, R.; Suter-Dick, L. A Perfused In Vitro Human iPSC-Derived Blood-Brain Barrier Faithfully Mimics Transferrin Receptor-Mediated Transcytosis of Therapeutic Antibodies. *Cell Mol Neurobiol* **2023**, *43*, 4173-4187, doi:10.1007/s10571-023-01404-x.
477. Ohshima, M.; Kamei, S.; Fushimi, H.; Mima, S.; Yamada, T.; Yamamoto, T. Prediction of Drug Permeability Using In Vitro Blood-Brain Barrier Models with Human Induced Pluripotent Stem Cell-Derived Brain Microvascular Endothelial Cells. *Biores Open Access* **2019**, *8*, 200-209, doi:10.1089/biores.2019.0026.
478. Workman, M.J.; Svendsen, C.N. Recent advances in human iPSC-derived models of the blood-brain barrier. *Fluids Barriers CNS* **2020**, *17*, 30, doi:10.1186/s12987-020-00191-7.
479. Mathew-Schmitt, S.; Oerter, S.; Reitenbach, E.; Gätzner, S.; Höchner, A.; Jahnke, H.G.; Piontek, J.; Neuhaus, W.; Brachner, A.; Metzger, M.; et al. Generation of Advanced Blood-Brain Barrier Spheroids Using Human-Induced Pluripotent Stem Cell-Derived Brain Capillary Endothelial-Like Cells. *Adv Biol (Weinh)* **2025**, *9*, e2400442, doi:10.1002/adbi.202400442.
480. Cho, C.F.; Wolfe, J.M.; Fadzen, C.M.; Calligaris, D.; Hornburg, K.; Chiocca, E.A.; Agar, N.Y.R.; Pentelute, B.L.; Lawler, S.E. Blood-brain-barrier spheroids as an in vitro screening platform for brain-penetrating agents. *Nat Commun* **2017**, *8*, 15623, doi:10.1038/ncomms15623.
481. Logan, S.; Arzua, T.; Canfield, S.G.; Seminary, E.R.; Sison, S.L.; Ebert, A.D.; Bai, X. Studying Human Neurological Disorders Using Induced Pluripotent Stem Cells: From 2D Monolayer to 3D Organoid and Blood Brain Barrier Models. *Compr Physiol* **2019**, *9*, 565-611, doi:10.1002/cphy.c180025.
482. Hajal, C.; Offeddu, G.S.; Shin, Y.; Zhang, S.; Morozova, O.; Hickman, D.; Knutson, C.G.; Kamm, R.D. Engineered human blood-brain barrier microfluidic model for vascular permeability analyses. *Nat Protoc* **2022**, *17*, 95-128, doi:10.1038/s41596-021-00635-w.
483. Oddo, A.; Peng, B.; Tong, Z.; Wei, Y.; Tong, W.Y.; Thissen, H.; Voelcker, N.H. Advances in Microfluidic Blood-Brain Barrier (BBB) Models. *Trends Biotechnol* **2019**, *37*, 1295-1314, doi:10.1016/j.tibtech.2019.04.006.
484. Vetter, J.; Palagi, I.; Waisman, A.; Blaeser, A. Recent advances in blood-brain barrier-on-a-chip models. *Acta Biomater* **2025**, *197*, 1-28, doi:10.1016/j.actbio.2025.03.041.
485. Chaulagain, B.; Gothwal, A.; Lamptey, R.N.L.; Trivedi, R.; Mahanta, A.K.; Layek, B.; Singh, J. Experimental Models of In Vitro Blood-Brain Barrier for CNS Drug Delivery: An Evolutionary Perspective. *Int J Mol Sci* **2023**, *24*, doi:10.3390/ijms24032710.
486. Park, T.E.; Mustafaoglu, N.; Herland, A.; Hasselkus, R.; Mannix, R.; FitzGerald, E.A.; Prantil-Baun, R.; Watters, A.; Henry, O.; Benz, M.; et al. Hypoxia-enhanced Blood-Brain Barrier Chip recapitulates human barrier function and shuttling of drugs and antibodies. *Nat Commun* **2019**, *10*, 2621, doi:10.1038/s41467-019-10588-0.
487. Katt, M.E.; Shusta, E.V. In vitro Models of the Blood-Brain Barrier: Building in physiological complexity. *Curr Opin Chem Eng* **2020**, *30*, 42-52, doi:10.1016/j.coche.2020.07.002.
488. Filieri, S.; Miciaccia, M.; Armenise, D.; Baldelli, O.M.; Litorri, A.; Ferorelli, S.; Sardanelli, A.M.; Perrone, M.G.; Scilimati, A. Can Focused Ultrasound Overcome the Failure of Chemotherapy in Treating Pediatric Diffuse Intrinsic Pontine Glioma Due to a Blood-Brain Barrier Obstacle? *Pharmaceuticals (Basel)* **2025**, *18*, doi:10.3390/ph18040525.
489. Tian, H.; Zhang, T.; Qin, S.; Huang, Z.; Zhou, L.; Shi, J.; Nice, E.C.; Xie, N.; Huang, C.; Shen, Z. Enhancing the therapeutic efficacy of nanoparticles for cancer treatment using versatile targeted strategies. *J Hematol Oncol* **2022**, *15*, 132, doi:10.1186/s13045-022-01320-5.

490. Rahman, R.; Ventz, S.; McDunn, J.; Louv, B.; Reyes-Rivera, I.; Polley, M.C.; Merchant, F.; Abrey, L.E.; Allen, J.E.; Aguilar, L.K.; et al. Leveraging external data in the design and analysis of clinical trials in neuro-oncology. *Lancet Oncol* **2021**, *22*, e456-e465, doi:10.1016/s1470-2045(21)00488-5.
491. Chen, K.T.; Chai, W.Y.; Lin, Y.J.; Lin, C.J.; Chen, P.Y.; Tsai, H.C.; Huang, C.Y.; Kuo, J.S.; Liu, H.L.; Wei, K.C. Neuronavigation-guided focused ultrasound for transcranial blood-brain barrier opening and immunostimulation in brain tumors. *Sci Adv* **2021**, *7*, doi:10.1126/sciadv.abd0772.
492. Liu, Y.; Castro Bravo, K.M.; Liu, J. Targeted liposomal drug delivery: a nanoscience and biophysical perspective. *Nanoscale Horiz* **2021**, *6*, 78-94, doi:10.1039/d0nh00605j.
493. McDannold, N.; Wen, P.Y.; Reardon, D.A.; Fletcher, S.M.; Golby, A.J. Cavitation monitoring, treatment strategy, and acoustic simulations of focused ultrasound blood-brain barrier disruption in patients with glioblastoma. *J Control Release* **2024**, *372*, 194-208, doi:10.1016/j.jconrel.2024.06.036.
494. Kim, Y.; Armstrong, T.S.; Gilbert, M.R.; Celiku, O. A critical analysis of neuro-oncology clinical trials. *Neuro Oncol* **2023**, *25*, 1658-1671, doi:10.1093/neuonc/noad036.
495. Liu, Y.; Wang, Q.; Feng, Z.; Qin, M.; Zhang, Z.; Jiang, J.; Ren, T.; Liu, X.; Jeffrey Brinker, C.; Zhao, Y.; et al. Unlocking tumor barrier: annexin A2-mediated transcytosis boosts drug delivery in pancreatic and breast tumors. *Nat Commun* **2025**, *16*, 6531, doi:10.1038/s41467-025-61434-5.
496. Hallschmid, M. Intranasal Insulin for Alzheimer's Disease. *CNS Drugs* **2021**, *35*, 21-37, doi:10.1007/s40263-020-00781-x.
497. Wong, C.Y.J.; Baldelli, A.; Hoyos, C.M.; Tietz, O.; Ong, H.X.; Traini, D. Insulin Delivery to the Brain via the Nasal Route: Unraveling the Potential for Alzheimer's Disease Therapy. *Drug Deliv Transl Res* **2024**, *14*, 1776-1793, doi:10.1007/s13346-024-01558-1.
498. Chen, L.; Guan, Y.; Wang, S.; Han, X.; Guo, F.; Wang, Y. Engineered nanoplatfoms for brain-targeted co-delivery of phytochemicals in Alzheimer's disease: Rational design, blood-brain barrier penetration, and multi-target therapeutic synergy. *Neurotherapeutics* **2025**, *22*, e00722, doi:10.1016/j.neurot.2025.e00722.
499. Wei, Y.; Xia, X.; Wang, X.; Yang, W.; He, S.; Wang, L.; Chen, Y.; Zhou, Y.; Chen, F.; Li, H.; et al. Enhanced BBB penetration and microglia-targeting nanomodulator for the two-pronged modulation of chronically activated microglia-mediated neuroinflammation in Alzheimer's disease. *Acta Pharm Sin B* **2025**, *15*, 1098-1111, doi:10.1016/j.apsb.2025.01.015.
500. Oosthoek, M.; Vermunt, L.; de Wilde, A.; Bongers, B.; Antwi-Berko, D.; Scheltens, P.; van Bokhoven, P.; Vijverberg, E.G.B.; Teunissen, C.E. Utilization of fluid-based biomarkers as endpoints in disease-modifying clinical trials for Alzheimer's disease: a systematic review. *Alzheimers Res Ther* **2024**, *16*, 93, doi:10.1186/s13195-024-01456-1.
501. Elghanam, Y.; Purja, S.; Kim, E.Y. Biomarkers as Endpoints in Clinical Trials for Alzheimer's Disease. *J Alzheimers Dis* **2024**, *99*, 693-703, doi:10.3233/jad-240008.
502. Pascoal, T.A.; Aguzzoli, C.S.; Lussier, F.Z.; Crivelli, L.; Suemoto, C.K.; Fortea, J.; Rosa-Neto, P.; Zimmer, E.R.; Ferreira, P.C.L.; Bellaver, B. Insights into the use of biomarkers in clinical trials in Alzheimer's disease. *EBioMedicine* **2024**, *108*, 105322, doi:10.1016/j.ebiom.2024.105322.
503. Petersen, R.C.; Graf, A.; Brady, C.; De Santi, S.; Florian, H.; Landen, J.; Pontecorvo, M.; Randolph, C.; Sink, K.; Carrillo, M.; et al. Operationalizing selection criteria for clinical trials in Alzheimer's disease: Biomarker and clinical considerations: Proceedings from the Alzheimer's Association Research Roundtable (AARR) Fall 2021 meeting. *Alzheimers Dement (N Y)* **2025**, *11*, e70038, doi:10.1002/trc2.70038.
504. Monge-Fuentes, V.; Biolchi Mayer, A.; Lima, M.R.; Geraldles, L.R.; Zannotto, L.N.; Moreira, K.G.; Martins, O.P.; Piva, H.L.; Felipe, M.S.S.; Amaral, A.C.; et al. Dopamine-loaded nanoparticle systems circumvent the blood-brain barrier restoring motor function in mouse model for Parkinson's Disease. *Sci Rep* **2021**, *11*, 15185, doi:10.1038/s41598-021-94175-8.
505. Liang, K.; Yang, L.; Kang, J.; Liu, B.; Zhang, D.; Wang, L.; Wang, W.; Wang, Q. Improving treatment for Parkinson's disease: Harnessing photothermal and phagocytosis-driven delivery of levodopa nanocarriers across the blood-brain barrier. *Asian J Pharm Sci* **2024**, *19*, 100963, doi:10.1016/j.ajps.2024.100963.
506. Leta, V.; Klingelhofer, L.; Longardner, K.; Campagnolo, M.; Levent, H.; Aureli, F.; Metta, V.; Bhidayasiri, R.; Chung-Faye, G.; Falup-Pecurariu, C.; et al. Gastrointestinal barriers to levodopa transport and absorption in Parkinson's disease. *Eur J Neurol* **2023**, *30*, 1465-1480, doi:10.1111/ene.15734.

507. Qiu, Y.; Jacobs, D.M.; Messer, K.; Salmon, D.P.; Wellington, C.L.; Stukas, S.; Revta, C.; Brewer, J.B.; Léger, G.C.; Askew, B.; et al. Prognostic value of plasma biomarkers for informing clinical trial design in mild-to-moderate Alzheimer's disease. *Alzheimers Res Ther* **2025**, *17*, 97, doi:10.1186/s13195-025-01745-3.
508. Asgharian, P.; Quispe, C.; Herrera-Bravo, J.; Sabernavaei, M.; Hosseini, K.; Forouhandeh, H.; Ebrahimi, T.; Sharafi-Badr, P.; Tarhriz, V.; Soofiyan, S.R.; et al. Pharmacological effects and therapeutic potential of natural compounds in neuropsychiatric disorders: An update. *Front Pharmacol* **2022**, *13*, 926607, doi:10.3389/fphar.2022.926607.
509. Küpeli Akkol, E.; Tatlı Çankaya, I.; Şeker Karatoprak, G.; Carpar, E.; Sobarzo-Sánchez, E.; Capasso, R. Natural Compounds as Medical Strategies in the Prevention and Treatment of Psychiatric Disorders Seen in Neurological Diseases. *Front Pharmacol* **2021**, *12*, 669638, doi:10.3389/fphar.2021.669638.
510. McIntyre, R.S.; Kwan, A.T.H.; Mansur, R.B.; Oliveira-Maia, A.J.; Teopiz, K.M.; Maletic, V.; Suppes, T.; Stahl, S.M.; Rosenblatt, J.D. Psychedelics for the Treatment of Psychiatric Disorders: Interpreting and Translating Available Evidence and Guidance for Future Research. *Am J Psychiatry* **2025**, *182*, 21-32, doi:10.1176/appi.ajp.20230902.
511. Barakji, J.; Korang, S.K.; Feinberg, J.; Maagaard, M.; Mathiesen, O.; Gluud, C.; Jakobsen, J.C. Cannabinoids versus placebo for pain: A systematic review with meta-analysis and Trial Sequential Analysis. *PLoS One* **2023**, *18*, e0267420, doi:10.1371/journal.pone.0267420.
512. Jiang, P.; Li, J. Recent advances in biomimetic nanodelivery systems for the treatment of depression. *Mater Today Bio* **2025**, *32*, 101781, doi:10.1016/j.mtbio.2025.101781.
513. Davis, K.D.; Aghaepour, N.; Ahn, A.H.; Angst, M.S.; Borsook, D.; Brenton, A.; Burczynski, M.E.; Crean, C.; Edwards, R.; Gaudilliere, B.; et al. Discovery and validation of biomarkers to aid the development of safe and effective pain therapeutics: challenges and opportunities. *Nat Rev Neurol* **2020**, *16*, 381-400, doi:10.1038/s41582-020-0362-2.
514. Tanaka, M. Beyond the boundaries: Transitioning from categorical to dimensional paradigms in mental health diagnostics. *Adv Clin Exp Med* **2024**, *33*, 1295-1301, doi:10.17219/acem/197425.
515. Tanaka, M.; Battaglia, S. Dualistic Dynamics in Neuropsychiatry: From Monoaminergic Modulators to Multiscale Biomarker Maps. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13061456.
516. Tanaka, M.; Szabó, Á.; Vécsei, L. Redefining Roles: A Paradigm Shift in Tryptophan-Kynurenine Metabolism for Innovative Clinical Applications. *Int J Mol Sci* **2024**, *25*, doi:10.3390/ijms252312767.
517. Szabó, Á.; Galla, Z.; Spekker, E.; Szűcs, M.; Martos, D.; Takeda, K.; Ozaki, K.; Inoue, H.; Yamamoto, S.; Toldi, J.; et al. Oxidative and Excitatory Neurotoxic Stresses in CRISPR/Cas9-Induced Kynurenine Aminotransferase Knockout Mice: A Novel Model for Despair-Based Depression and Post-Traumatic Stress Disorder. *Front Biosci (Landmark Ed)* **2025**, *30*, 25706, doi:10.31083/fbl25706.
518. Naeem, A.; Hu, P.; Yang, M.; Zhang, J.; Liu, Y.; Zhu, W.; Zheng, Q. Natural Products as Anticancer Agents: Current Status and Future Perspectives. *Molecules* **2022**, *27*, doi:10.3390/molecules27238367.
519. Chunarkar-Patil, P.; Kaleem, M.; Mishra, R.; Ray, S.; Ahmad, A.; Verma, D.; Bhayye, S.; Dubey, R.; Singh, H.N.; Kumar, S. Anticancer Drug Discovery Based on Natural Products: From Computational Approaches to Clinical Studies. *Biomedicines* **2024**, *12*, doi:10.3390/biomedicines12010201.
520. Choudhari, A.S.; Mandave, P.C.; Deshpande, M.; Ranjekar, P.; Prakash, O. Phytochemicals in Cancer Treatment: From Preclinical Studies to Clinical Practice. *Front Pharmacol* **2019**, *10*, 1614, doi:10.3389/fphar.2019.01614.
521. Wang, G.; Zhou, X.; Pang, X.; Ma, K.; Li, L.; Song, Y.; Hou, D.; Wang, X. Pharmacological effects, molecular mechanisms and strategies to improve bioavailability of curcumin in the treatment of neurodegenerative diseases. *Front Pharmacol* **2025**, *16*, 1625821, doi:10.3389/fphar.2025.1625821.
522. Grabarczyk, M.; Justyńska, W.; Czapkowska, J.; Smolińska, E.; Bielenin, A.; Glabinski, A.; Szpakowski, P. Role of Plant Phytochemicals: Resveratrol, Curcumin, Luteolin and Quercetin in Demyelination, Neurodegeneration, and Epilepsy. *Antioxidants (Basel)* **2024**, *13*, doi:10.3390/antiox13111364.
523. Ahn-Horst, R.Y.; Turner, E.H.; Kesselheim, A.S. Characteristics of Trials Preceding FDA Approval of Novel Psychiatric Drugs. *JAMA Netw Open* **2025**, *8*, e2456588, doi:10.1001/jamanetworkopen.2024.56588.
524. Markowitz, J.C.; Milrod, B.L. Lost in Translation: The Value of Psychiatric Clinical Trials. *J Clin Psychiatry* **2022**, *83*, doi:10.4088/JCP.22com14647.

525. Lipsman, N.; Meng, Y.; Bethune, A.J.; Huang, Y.; Lam, B.; Masellis, M.; Herrmann, N.; Heyn, C.; Aubert, I.; Boutet, A.; et al. Blood-brain barrier opening in Alzheimer's disease using MR-guided focused ultrasound. *Nat Commun* **2018**, *9*, 2336, doi:10.1038/s41467-018-04529-6.
526. Gasca-Salas, C.; Fernández-Rodríguez, B.; Pineda-Pardo, J.A.; Rodríguez-Rojas, R.; Obeso, I.; Hernández-Fernández, F.; Del Álamo, M.; Mata, D.; Guida, P.; Ordás-Bandera, C.; et al. Blood-brain barrier opening with focused ultrasound in Parkinson's disease dementia. *Nat Commun* **2021**, *12*, 779, doi:10.1038/s41467-021-21022-9.
527. Choudhury, H.; Pandey, M.; Chin, P.X.; Phang, Y.L.; Cheah, J.Y.; Ooi, S.C.; Mak, K.K.; Pichika, M.R.; Kesharwani, P.; Hussain, Z.; et al. Transferrin receptors-targeting nanocarriers for efficient targeted delivery and transcytosis of drugs into the brain tumors: a review of recent advancements and emerging trends. *Drug Deliv Transl Res* **2018**, *8*, 1545-1563, doi:10.1007/s13346-018-0552-2.
528. Ramalho, M.J.; Sevin, E.; Gosselet, F.; Lima, J.; Coelho, M.A.N.; Loureiro, J.A.; Pereira, M.C. Receptor-mediated PLGA nanoparticles for glioblastoma multiforme treatment. *Int J Pharm* **2018**, *545*, 84-92, doi:10.1016/j.ijpharm.2018.04.062.
529. Kretsoulas, D.; Damante, M.; Cua, S.; Lonser, R.R. Adjuvant convection-enhanced delivery for the treatment of brain tumors. *J Neurooncol* **2024**, *166*, 243-255, doi:10.1007/s11060-023-04552-8.
530. Upadhyay, R.; Ghosh, P.; Desavathu, M. Advancement in the Nose-to-Brain Drug delivery of FDA-approved drugs for the better management of Depression and Psychiatric disorders. *Int J Pharm* **2024**, *667*, 124866, doi:10.1016/j.ijpharm.2024.124866.
531. Yang, H.; Liu, R.; Huang, J.; Zhao, Y.; Zou, C.; Wu, D.; Zhao, H. Nose-to-brain delivery of biomineralized silk fibroin nanoparticles synergistically extinguish neuroinflammation for efficient depressive-like symptoms alleviation. *Int J Biol Macromol* **2025**, *322*, 146761, doi:10.1016/j.ijbiomac.2025.146761.
532. Zünkeler, B.; Carson, R.E.; Olson, J.; Blasberg, R.G.; DeVroom, H.; Lutz, R.J.; Saris, S.C.; Wright, D.C.; Kammerer, W.; Patronas, N.J.; et al. Quantification and pharmacokinetics of blood-brain barrier disruption in humans. *J Neurosurg* **1996**, *85*, 1056-1065, doi:10.3171/jns.1996.85.6.1056.
533. Burks, S.R.; Kersch, C.N.; Witko, J.A.; Pagel, M.A.; Sundby, M.; Muldoon, L.L.; Neuwelt, E.A.; Frank, J.A. Blood-brain barrier opening by intracarotid artery hyperosmolar mannitol induces sterile inflammatory and innate immune responses. *Proc Natl Acad Sci U S A* **2021**, *118*, doi:10.1073/pnas.2021915118.
534. Wang, W.; Marín-Ramos, N.I.; He, H.; Zeng, S.; Cho, H.Y.; Swenson, S.D.; Zheng, L.; Epstein, A.L.; Schönthal, A.H.; Hofman, F.M.; et al. NEO100 enables brain delivery of blood-brain barrier impermeable therapeutics. *Neuro Oncol* **2021**, *23*, 63-75, doi:10.1093/neuonc/noaa206.
535. Pinkiewicz, M.; Pinkiewicz, M.; Walecki, J.; Zaczynski, A.; Zawadzki, M. Breaking Barriers in Neuro-Oncology: A Scoping Literature Review on Invasive and Non-Invasive Techniques for Blood-Brain Barrier Disruption. *Cancers (Basel)* **2024**, *16*, doi:10.3390/cancers16010236.
536. Mollnes, T.E.; Storm, B.S.; Brekke, O.L.; Nilsson, P.H.; Lambris, J.D. Application of the C3 inhibitor compstatin in a human whole blood model designed for complement research - 20 years of experience and future perspectives. *Semin Immunol* **2022**, *59*, 101604, doi:10.1016/j.smim.2022.101604.
537. Luo, S.; Hu, D.; Wang, M.; Zipfel, P.F.; Hu, Y. Complement in Hemolysis- and Thrombosis- Related Diseases. *Front Immunol* **2020**, *11*, 1212, doi:10.3389/fimmu.2020.01212.
538. Roumenina, L.T.; Bartolucci, P.; Pirenne, F. The role of Complement in Post-Transfusion Hemolysis and Hyperhemolysis Reaction. *Transfus Med Rev* **2019**, *33*, 225-230, doi:10.1016/j.tmr.2019.09.007.
539. Poillerat, V.; Gentinetta, T.; Leon, J.; Wassmer, A.; Edler, M.; Torset, C.; Luo, D.; Tuffin, G.; Roumenina, L.T. Hemopexin as an Inhibitor of Hemolysis-Induced Complement Activation. *Front Immunol* **2020**, *11*, 1684, doi:10.3389/fimmu.2020.01684.
540. Nguyen-Peyre, K.-A.; Meuleman, M.-S.; Bodivit, G.; Kassasseya, C.; Decrouy, X.; Vingert, B.; Bencheikh, L.; Lambris, J.; Pirenne, F.; Roumenina, L.T. Red Blood Cell-Derived Particles Induce Endothelial Damage Via the Alternative Complement Pathway. **2024**.
541. Kang, H.J.; Roh, J.; Lee, H.; Park, E.M.; Lee, H.W.; Lee, J.Y.; Hwang, J.H.; Shim, J.; Choi, K. Complement Activation and Hemolysis in Non-human Primates Following Transfusion of Genetically Modified Pig Red Blood Cells. *Ann Lab Med* **2025**, *45*, 509-519, doi:10.3343/alm.2024.0443.

542. Alharbi, Y.M. Phenotype- and age-associated variations in non-specific agglutinins and complement components (C3 and C5a) in camels: Implications for transfusion compatibility and immune function. *Vet World* **2025**, *18*, 2811-2822, doi:10.14202/vetworld.2025.2811-2822.
543. Singh, D. Astrocytic and microglial cells as the modulators of neuroinflammation in Alzheimer's disease. *J Neuroinflammation* **2022**, *19*, 206, doi:10.1186/s12974-022-02565-0.
544. Müller, L.; Di Benedetto, S. Neuroimmune crosstalk in chronic neuroinflammation: microglial interactions and immune modulation. *Front Cell Neurosci* **2025**, *19*, 1575022, doi:10.3389/fncel.2025.1575022.
545. West, P.K.; McCorkindale, A.N.; Guennewig, B.; Ashhurst, T.M.; Viengkhou, B.; Hayashida, E.; Jung, S.R.; Butovsky, O.; Campbell, I.L.; Hofer, M.J. The cytokines interleukin-6 and interferon- $\alpha$  induce distinct microglia phenotypes. *J Neuroinflammation* **2022**, *19*, 96, doi:10.1186/s12974-022-02441-x.
546. Fornari Laurindo, L.; Aparecido Dias, J.; Cressoni Araújo, A.; Torres Pomini, K.; Machado Galhardi, C.; Rucco Penteado Detregiachi, C.; Santos de Argollo Haber, L.; Donizeti Roque, D.; Dib Bechara, M.; Vialogo Marques de Castro, M.; et al. Immunological dimensions of neuroinflammation and microglial activation: exploring innovative immunomodulatory approaches to mitigate neuroinflammatory progression. *Front Immunol* **2023**, *14*, 1305933, doi:10.3389/fimmu.2023.1305933.
547. Alaei, M.; Koushki, K.; Taebi, K.; Yousefi Taba, M.; Keshavarz Hedayati, S.; Keshavarz Shahbaz, S. Metal nanoparticles in neuroinflammation: impact on microglial dynamics and CNS function. *RSC Adv* **2025**, *15*, 5426-5451, doi:10.1039/d4ra07798a.
548. Zhao, N.; Francis, N.L.; Calvelli, H.R.; Moghe, P.V. Microglia-targeting nanotherapeutics for neurodegenerative diseases. *APL Bioeng* **2020**, *4*, 030902, doi:10.1063/5.0013178.
549. Benita, B.A.; Koss, K.M. Peptide discovery across the spectrum of neuroinflammation; microglia and astrocyte phenotypical targeting, mediation, and mechanistic understanding. *Front Mol Neurosci* **2024**, *17*, 1443985, doi:10.3389/fnmol.2024.1443985.
550. Weber, M.; Steinle, H.; Golombek, S.; Hann, L.; Schlensak, C.; Wendel, H.P.; Avci-Adali, M. Blood-Contacting Biomaterials: In Vitro Evaluation of the Hemocompatibility. *Front Bioeng Biotechnol* **2018**, *6*, 99, doi:10.3389/fbioe.2018.00099.
551. Nalezinková, M. In vitro hemocompatibility testing of medical devices. *Thromb Res* **2020**, *195*, 146-150, doi:10.1016/j.thromres.2020.07.027.
552. van Oeveren, W. Obstacles in haemocompatibility testing. *Scientifica (Cairo)* **2013**, *2013*, 392584, doi:10.1155/2013/392584.
553. Seyfert, U.T.; Biehl, V.; Schenk, J. In vitro hemocompatibility testing of biomaterials according to the ISO 10993-4. *Biomol Eng* **2002**, *19*, 91-96, doi:10.1016/s1389-0344(02)00015-1.
554. Tienda-Vazquez, M.A.; Arredondo, P.; Mejía-Delgadillo, X.; Rodríguez-González, J.A.; Soto-Cajiga, J.A.; Sabath, E.; Lozano, O.; Almanza-Arjona, Y.C. Biological testing unification for hemodialysis membranes evaluation: A step towards standardization. *Biomater Adv* **2025**, *169*, 214165, doi:10.1016/j.bioadv.2024.214165.
555. Tujula, I.; Hyvärinen, T.; Lotila, J.; Rogal, J.; Voulgaris, D.; Sukki, L.; Tornberg, K.; Korpela, K.; Jäntti, H.; Malm, T.; et al. Modeling neuroinflammatory interactions between microglia and astrocytes in a human iPSC-based coculture platform. *Cell Commun Signal* **2025**, *23*, 298, doi:10.1186/s12964-025-02304-x.
556. Guttikonda, S.R.; Sikkema, L.; Tchieu, J.; Saurat, N.; Walsh, R.M.; Harschnitz, O.; Ciceri, G.; Sneeboer, M.; Mazutis, L.; Setty, M.; et al. Fully defined human pluripotent stem cell-derived microglia and tri-culture system model C3 production in Alzheimer's disease. *Nat Neurosci* **2021**, *24*, 343-354, doi:10.1038/s41593-020-00796-z.
557. Yuan, N.Y.; Richards, W.D.; Parham, K.T.; Clark, S.G.; Greuel, K.; Polzin, B.; Smith, S.W.; Lebakken, C.S. Neural organoids incorporating microglia to assess neuroinflammation and toxicities induced by known developmental neurotoxins. *Curr Res Toxicol* **2025**, *9*, 100252, doi:10.1016/j.crttox.2025.100252.
558. Heneka, M.T.; Gauthier, S.; Chandekar, S.A.; Hviid Hahn-Pedersen, J.; Bentsen, M.A.; Zetterberg, H. Neuroinflammatory fluid biomarkers in patients with Alzheimer's disease: a systematic literature review. *Mol Psychiatry* **2025**, *30*, 2783-2798, doi:10.1038/s41380-025-02939-9.
559. Roveta, F.; Bonino, L.; Piella, E.M.; Rainero, I.; Rubino, E. Neuroinflammatory Biomarkers in Alzheimer's Disease: From Pathophysiology to Clinical Implications. *Int J Mol Sci* **2024**, *25*, doi:10.3390/ijms252211941.

560. Battaglia, S.; Tanaka, M. Screen, Sample, Stratify: Biomarkers and Machine Learning Compress Dementia Pathways. *2026*, *14*, 159.
561. Tanaka, M.; Battaglia, S. From Biomarkers to Behavior: Mapping the Neuroimmune Web of Pain, Mood, and Memory. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13092226.
562. Wei, Y.; Quan, L.; Zhou, C.; Zhan, Q. Factors relating to the biodistribution & clearance of nanoparticles & their effects on in vivo application. *Nanomedicine (Lond)* **2018**, *13*, 1495-1512, doi:10.2217/nmm-2018-0040.
563. Dogra, P.; Adolph, N.L.; Wang, Z.; Lin, Y.S.; Butler, K.S.; Durfee, P.N.; Croissant, J.G.; Noureddine, A.; Coker, E.N.; Bearer, E.L.; et al. Establishing the effects of mesoporous silica nanoparticle properties on in vivo disposition using imaging-based pharmacokinetics. *Nat Commun* **2018**, *9*, 4551, doi:10.1038/s41467-018-06730-z.
564. Gustafson, H.H.; Holt-Casper, D.; Grainger, D.W.; Ghandehari, H. Nanoparticle Uptake: The Phagocyte Problem. *Nano Today* **2015**, *10*, 487-510, doi:10.1016/j.nantod.2015.06.006.
565. Ngo, W.; Ahmed, S.; Blackadar, C.; Bussin, B.; Ji, Q.; Mladjenovic, S.M.; Sepahi, Z.; Chan, W.C.W. Why nanoparticles prefer liver macrophage cell uptake in vivo. *Adv Drug Deliv Rev* **2022**, *185*, 114238, doi:10.1016/j.addr.2022.114238.
566. Lérida-Viso, A.; Estepa-Fernández, A.; García-Fernández, A.; Martí-Centelles, V.; Martínez-Mañez, R. Biosafety of mesoporous silica nanoparticles; towards clinical translation. *Adv Drug Deliv Rev* **2023**, *201*, 115049, doi:10.1016/j.addr.2023.115049.
567. Baek, M.J.; Hur, W.; Kashiwagi, S.; Choi, H.S. Design Considerations for Organ-Selective Nanoparticles. *ACS Nano* **2025**, *19*, 14605-14626, doi:10.1021/acsnano.5c00484.
568. Zhu, G.H.; Gray, A.B.C.; Patra, H.K. Nanomedicine: controlling nanoparticle clearance for translational success. *Trends Pharmacol Sci* **2022**, *43*, 709-711, doi:10.1016/j.tips.2022.05.001.
569. Zhang, Y.; Lin, X.; Chen, X.; Fang, W.; Yu, K.; Gu, W.; Wei, Y.; Zheng, H.; Piao, J.; Li, F. Strategies to Regulate the Degradation and Clearance of Mesoporous Silica Nanoparticles: A Review. *Int J Nanomedicine* **2024**, *19*, 5859-5878, doi:10.2147/ijn.S451919.
570. Wang, L.; Quine, S.; Frickenstein, A.N.; Lee, M.; Yang, W.; Sheth, V.M.; Bourlon, M.D.; He, Y.; Lyu, S.; Garcia-Contreras, L.; et al. Exploring and Analyzing the Systemic Delivery Barriers for Nanoparticles. *Adv Funct Mater* **2024**, *34*, doi:10.1002/adfm.202308446.
571. Zelepukin, I.V.; Shevchenko, K.G.; Deyev, S.M. Rediscovery of mononuclear phagocyte system blockade for nanoparticle drug delivery. *Nat Commun* **2024**, *15*, 4366, doi:10.1038/s41467-024-48838-5.
572. Bresinsky, M.; Goepferich, A. Control of biomedical nanoparticle distribution and drug release in vivo by complex particle design strategies. *Eur J Pharm Biopharm* **2025**, *208*, 114634, doi:10.1016/j.ejpb.2025.114634.
573. Hidayat, A.F.; Wardhana, Y.W.; Suwendar, S.; Mohammed, A.F.A.; Mahmoud, S.A.; Elamin, K.M.; Wathoni, N. A Review on QbD-Driven Optimization of Lipid Nanoparticles for Oral Drug Delivery: From Framework to Formulation. *Int J Nanomedicine* **2025**, *20*, 8611-8651, doi:10.2147/ijn.S534137.
574. Yang, S.; Hu, X.; Zhu, J.; Zheng, B.; Bi, W.; Wang, X.; Wu, J.; Mi, Z.; Wu, Y. Aspects and Implementation of Pharmaceutical Quality by Design from Conceptual Frameworks to Industrial Applications. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17050623.
575. Pielenhofer, J.; Meiser, S.L.; Gogoll, K.; Ciciliani, A.M.; Denny, M.; Klak, M.; Lang, B.M.; Staubach, P.; Grabbe, S.; Schild, H.; et al. Quality by Design (QbD) Approach for a Nanoparticulate Imiquimod Formulation as an Investigational Medicinal Product. *Pharmaceutics* **2023**, *15*, doi:10.3390/pharmaceutics15020514.
576. Walsh, I.; Myint, M.; Nguyen-Khuong, T.; Ho, Y.S.; Ng, S.K.; Lakshmanan, M. Harnessing the potential of machine learning for advancing "Quality by Design" in biomanufacturing. *MAbs* **2022**, *14*, 2013593, doi:10.1080/19420862.2021.2013593.
577. Suriyaamporn, P.; Kansom, T.; Pamornpathomkul, B.; Ngawhirunpat, T.; Opanasopit, P.; Ramjan, S. Predictive modeling approach using machine learning-integrated design of experiments in quality by design for optimizing resveratrol-loaded polymeric nanoparticle formulation. *Int J Pharm* **2025**, *683*, 126080, doi:10.1016/j.ijpharm.2025.126080.
578. Tanaka, M. Special Issue "Translating Molecular Psychiatry: From Biomarkers to Personalized Therapies". *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms262010238.

579. Palakurthi, S.S.; Charbe, N.B.; Kapre, S.; Zheng, W.; Thalla, M.; Palaniappan, D.; Lu, D.; Palakurthi, S. Evaluating critical quality attributes and novel drug release testing of difluprednate nanoemulsions. *Int J Pharm* **2025**, *674*, 125431, doi:10.1016/j.ijpharm.2025.125431.
580. Soncin, S.; Lo Cicero, V.; Astori, G.; Soldati, G.; Gola, M.; Sürder, D.; Moccetti, T. A practical approach for the validation of sterility, endotoxin and potency testing of bone marrow mononucleated cells used in cardiac regeneration in compliance with good manufacturing practice. *J Transl Med* **2009**, *7*, 78, doi:10.1186/1479-5876-7-78.
581. Vuilleminot, B.R.; Korte, S.; Wright, T.L.; Adams, E.L.; Boyd, R.B.; Butt, M.T. Safety Evaluation of CNS Administered Biologics-Study Design, Data Interpretation, and Translation to the Clinic. *Toxicol Sci* **2016**, *152*, 3-9, doi:10.1093/toxsci/kfw072.
582. Cooper, J.F.; Latta, K.S.; Smith, D. Automated endotoxin testing program for high-risk-level compounded sterile preparations at an institutional compounding pharmacy. *Am J Health Syst Pharm* **2010**, *67*, 280-286, doi:10.2146/ajhp090290.
583. Cooper, J.F.; Thoma, L.A. Screening extemporaneously compounded intraspinal injections with the bacterial endotoxins test. *Am J Health Syst Pharm* **2002**, *59*, 2426-2433, doi:10.1093/ajhp/59.24.2426.
584. Dubczak, J.; Reid, N.; Tsuchiya, M. Evaluation of limulus amoebocyte lysate and recombinant endotoxin alternative assays for an assessment of endotoxin detection specificity. *Eur J Pharm Sci* **2021**, *159*, 105716, doi:10.1016/j.ejps.2021.105716.
585. Secretan, P.H.; Annereau, M.; Do, B. A Risk-Based Framework for Hospital Compounding: Integrating Degradation Mechanisms and Predictive Toxicology. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17091202.
586. Pallerla, S.; Pires, I.S.; Melo, M.B.; Yun, D.; Wagner, A.; Budai, M.; Kumar, D.; Katinger, D.; Sayeed, E.; Lombardo, A.; et al. Scale-up and cGMP manufacturing of next-generation vaccine adjuvant saponin/MPLA nanoParticles (SMNP). *J Pharm Sci* **2025**, *114*, 103913, doi:10.1016/j.xphs.2025.103913.
587. Cunha, S.; Costa, C.P.; Loureiro, J.A.; Alves, J.; Peixoto, A.F.; Forbes, B.; Sousa Lobo, J.M.; Silva, A.C. Double Optimization of Rivastigmine-Loaded Nanostructured Lipid Carriers (NLC) for Nose-to-Brain Delivery Using the Quality by Design (QbD) Approach: Formulation Variables and Instrumental Parameters. *Pharmaceutics* **2020**, *12*, doi:10.3390/pharmaceutics12070599.
588. Laggner, M.; Gugerell, A.; Bachmann, C.; Hofbauer, H.; Vorstandlechner, V.; Seibold, M.; Gouya Lechner, G.; Peterbauer, A.; Madlener, S.; Demyanets, S. Reproducibility of GMP-compliant production of therapeutic stressed peripheral blood mononuclear cell-derived secretomes, a novel class of biological medicinal products. *Stem cell research & therapy* **2020**, *11*, 9.
589. Sanyal, G.; Särnefält, A.; Kumar, A. Considerations for bioanalytical characterization and batch release of COVID-19 vaccines. *NPJ Vaccines* **2021**, *6*, 53, doi:10.1038/s41541-021-00317-4.
590. Laggner, M.; Gugerell, A.; Bachmann, C.; Hofbauer, H.; Vorstandlechner, V.; Seibold, M.; Gouya Lechner, G.; Peterbauer, A.; Madlener, S.; Demyanets, S.; et al. Reproducibility of GMP-compliant production of therapeutic stressed peripheral blood mononuclear cell-derived secretomes, a novel class of biological medicinal products. *Stem Cell Res Ther* **2020**, *11*, 9, doi:10.1186/s13287-019-1524-2.
591. Csóka, I.; Ismail, R.; Jójárt-Laczkovich, O.; Pallagi, E. Regulatory Considerations, Challenges and Risk-based Approach in Nanomedicine Development. *Curr Med Chem* **2021**, *28*, 7461-7476, doi:10.2174/0929867328666210406115529.
592. Ali, F.; Neha, K.; Parveen, S. Current regulatory landscape of nanomaterials and nanomedicines: A global perspective. *Journal of Drug Delivery Science and Technology* **2023**, *80*, 104118.
593. Dobrovolskaia, M.A. Lessons learned from immunological characterization of nanomaterials at the Nanotechnology Characterization Laboratory. *Front Immunol* **2022**, *13*, 984252, doi:10.3389/fimmu.2022.984252.
594. Giannakou, C.; Park, M.; Bosselaers, I.E.M.; de Jong, W.H.; van der Laan, J.W.; van Loveren, H.; Vandebriel, R.J.; Geertsma, R.E. Nonclinical regulatory immunotoxicity testing of nanomedicinal products: Proposed strategy and possible pitfalls. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* **2020**, *12*, e1633, doi:10.1002/wnan.1633.

595. Hofer, S.; Hofstätter, N.; Punz, B.; Hasenkopf, I.; Johnson, L.; Himly, M. Immunotoxicity of nanomaterials in health and disease: Current challenges and emerging approaches for identifying immune modifiers in susceptible populations. *Wiley Interdiscip Rev Nanomed Nanobiotechnol* **2022**, *14*, e1804, doi:10.1002/wnan.1804.
596. Dobrovolskaia, M.A.; McNeil, S.E. Understanding the correlation between in vitro and in vivo immunotoxicity tests for nanomedicines. *J Control Release* **2013**, *172*, 456-466, doi:10.1016/j.jconrel.2013.05.025.
597. van Gerven, J.; Bonelli, M. Commentary on the EMA Guideline on strategies to identify and mitigate risks for first-in-human and early clinical trials with investigational medicinal products. *Br J Clin Pharmacol* **2018**, *84*, 1401-1409, doi:10.1111/bcp.13550.
598. Musazzi, U.M.; Franzè, S.; Condorelli, F.; Minghetti, P.; Caliceti, P. Feeding Next-Generation Nanomedicines to Europe: Regulatory and Quality Challenges. *Adv Healthc Mater* **2023**, *12*, e2301956, doi:10.1002/adhm.202301956.
599. Zhuang, Y.; Mendes, B.B.; Menon, D.; Oliveira, J.; Chen, X.; Duman, F.D.; Conniot, J.; Mercado, S.; Liu, X.; Zhang, S.Y.; et al. Multiscale Profiling of Nanoscale Metal-Organic Framework Biocompatibility and Immune Interactions. *Adv Healthc Mater* **2025**, *14*, e01809, doi:10.1002/adhm.202501809.
600. Nabi, A.E.; Pouladvand, P.; Liu, L.; Hua, N.; Ayubcha, C. Machine Learning in Drug Development for Neurological Diseases: A Review of Blood Brain Barrier Permeability Prediction Models. *Mol Inform* **2025**, *44*, e202400325, doi:10.1002/minf.202400325.
601. Liu, L.; Zhang, L.; Feng, H.; Li, S.; Liu, M.; Zhao, J.; Liu, H. Prediction of the Blood-Brain Barrier (BBB) Permeability of Chemicals Based on Machine-Learning and Ensemble Methods. *Chem Res Toxicol* **2021**, *34*, 1456-1467, doi:10.1021/acs.chemrestox.0c00343.
602. Gülave, B.; van den Maagdenberg, H.W.; van Boven, L.; van Westen, G.J.P.; de Lange, E.C.M.; van Hasselt, J.G.C. Prediction of the Extent of Blood-Brain Barrier Transport Using Machine Learning and Integration into the LeicNS-PK3.0 Model. *Pharm Res* **2025**, *42*, 281-289, doi:10.1007/s11095-025-03828-0.
603. Mehta, P.; Soliman, A.; Rodriguez-Vera, L.; Schmidt, S.; Muniz, P.; Rodriguez, M.; Forcadell, M.; Gonzalez-Perez, E.; Vozmediano, V. Interspecies Brain PBPK Modeling Platform to Predict Passive Transport through the Blood-Brain Barrier and Assess Target Site Disposition. *Pharmaceutics* **2024**, *16*, doi:10.3390/pharmaceutics16020226.
604. Cichońska, A.; Ravikumar, B.; Rahman, R. AI for targeted polypharmacology: The next frontier in drug discovery. *Curr Opin Struct Biol* **2024**, *84*, 102771, doi:10.1016/j.sbi.2023.102771.
605. Abdelsayed, M. AI-Driven Polypharmacology in Small-Molecule Drug Discovery. *Int J Mol Sci* **2025**, *26*, doi:10.3390/ijms26146996.
606. Noor, F.; Asif, M.; Ashfaq, U.A.; Qasim, M.; Tahir Ul Qamar, M. Machine learning for synergistic network pharmacology: a comprehensive overview. *Brief Bioinform* **2023**, *24*, doi:10.1093/bib/bbad120.
607. Chaudhari, R.; Fong, L.W.; Tan, Z.; Huang, B.; Zhang, S. An up-to-date overview of computational polypharmacology in modern drug discovery. *Expert Opin Drug Discov* **2020**, *15*, 1025-1044, doi:10.1080/17460441.2020.1767063.
608. Lambrinidis, G.; Tsantili-Kakoulidou, A. Multi-objective optimization methods in novel drug design. *Expert Opin Drug Discov* **2021**, *16*, 647-658, doi:10.1080/17460441.2021.1867095.
609. Waibel, I.; Schneider, T.N.; Fischer, F.J.; Dumnoenchanvanit, P.; Kulakova, A.; Nguyen, T.D.; Egebjerg, T.; Bertelsen, S.; Lorenzen, N.; Arosio, P. Bayesian Optimization for Efficient Multiobjective Formulation Development of Biologics. *Mol Pharm* **2025**, *22*, 6636-6645, doi:10.1021/acs.molpharmaceut.5c00591.
610. Li, L.; Back, S.I.; Ma, J.; Guo, Y.; Galeandro-Diamant, T.; Clénet, D. Bayesian optimization and machine learning for vaccine formulation development. *PLoS One* **2025**, *20*, e0324205, doi:10.1371/journal.pone.0324205.
611. Luukkonen, S.; van den Maagdenberg, H.W.; Emmerich, M.T.M.; van Westen, G.J.P. Artificial intelligence in multi-objective drug design. *Curr Opin Struct Biol* **2023**, *79*, 102537, doi:10.1016/j.sbi.2023.102537.
612. Wickramasinghe, C.D.; Kim, S.; Li, J. SpatialCNS-PBPK: An R/Shiny Web-Based Application for Physiologically Based Pharmacokinetic Modeling of Spatial Pharmacokinetics in the Human Central

- Nervous System and Brain Tumors. *CPT Pharmacometrics Syst Pharmacol* **2025**, *14*, 864-880, doi:10.1002/psp4.70026.
613. Bowman, C.; Ma, F.; Mao, J.; Plise, E.; Chen, E.; Liu, L.; Zhang, S.; Chen, Y. Evaluation of bottom-up modeling of the blood-brain barrier to improve brain penetration prediction via physiologically based pharmacokinetic modeling. *Biopharm Drug Dispos* **2023**, *44*, 60-70, doi:10.1002/bdd.2344.
614. Tregub, P.P.; Bystrov, D.A.; Kushnir, I.A.; Korsakova, S.A.; Yurchenko, S.O.; Salmina, A.B. Blood-brain barriers and drug pharmacokinetics: mechanisms and models. *Eur J Pharmacol* **2025**, *1003*, 177872, doi:10.1016/j.ejphar.2025.177872.
615. Linville, R.M.; DeStefano, J.G.; Sklar, M.B.; Xu, Z.; Farrell, A.M.; Bogorad, M.I.; Chu, C.; Walczak, P.; Cheng, L.; Mahairaki, V.; et al. Human iPSC-derived blood-brain barrier microvessels: validation of barrier function and endothelial cell behavior. *Biomaterials* **2019**, *190-191*, 24-37, doi:10.1016/j.biomaterials.2018.10.023.
616. Canfield, S.G.; Stebbins, M.J.; Faubion, M.G.; Gastfriend, B.D.; Palecek, S.P.; Shusta, E.V. An isogenic neurovascular unit model comprised of human induced pluripotent stem cell-derived brain microvascular endothelial cells, pericytes, astrocytes, and neurons. *Fluids Barriers CNS* **2019**, *16*, 25, doi:10.1186/s12987-019-0145-6.
617. Ratcliffe, L.; Vermond, S.; Hill, S.; de Munnik, S.; van Puijvelde, G.; Quist, B.; Pooley, R.; Carr, K.; Brown, N.; Aspinall-O'Dea, M. Abstract LB009: Safety profile assessment for IND-enabling studies: Bispecific antibodies and antibody-drug conjugates (ADCs). *Cancer Research* **2025**, *85*, LB009-LB009.
618. Youssef, E.; Weddle, K.; Zimmerman, L.; Palmer, D. Pharmacovigilance in Cell and Gene Therapy: Evolving Challenges in Risk Management and Long-Term Follow-Up. *Drug Saf* **2026**, *49*, 27-53, doi:10.1007/s40264-025-01596-9.
619. Glader, C.; Jeitler, R.; Wang, Y.; van Tuijn, R.; Grau-Carbonell, A.; Tetyczka, C.; Mesite, S.; Caisse, P.; Khinast, J.; Roblegg, E. Process Analytical Strategies for Size Monitoring: Offline, At-Line, Online, and Inline Methods in a Top-Down Nano-Manufacturing Line. *Pharmaceutics* **2025**, *17*, doi:10.3390/pharmaceutics17060684.
620. Adaptive platform trials: definition, design, conduct and reporting considerations. *Nat Rev Drug Discov* **2019**, *18*, 797-807, doi:10.1038/s41573-019-0034-3.
621. Narsinh, K.H.; Perez, E.; Haddad, A.F.; Young, J.S.; Savastano, L.; Villanueva-Meyer, J.E.; Winkler, E.; de Groot, J. Strategies to Improve Drug Delivery Across the Blood-Brain Barrier for Glioblastoma. *Curr Neurol Neurosci Rep* **2024**, *24*, 123-139, doi:10.1007/s11910-024-01338-x.
622. Zhu, M.; Zhuang, J.; Li, Z.; Liu, Q.; Zhao, R.; Gao, Z.; Midgley, A.C.; Qi, T.; Tian, J.; Zhang, Z.; et al. Machine-learning-assisted single-vessel analysis of nanoparticle permeability in tumour vasculatures. *Nat Nanotechnol* **2023**, *18*, 657-666, doi:10.1038/s41565-023-01323-4.
623. Pan, F.; Xia, Y.; Zhang, B.; Mohammed, A.; Zhao, X. Microfluidic Fabrication of Peptide-Functionalized Poly(lactic-co-glycolic acid) Nanoparticles for Targeted Curcumin Delivery in Breast Cancer. *Langmuir* **2025**, *41*, 19514-19525, doi:10.1021/acs.langmuir.5c02318.
624. Kim, J.; Lee, D.H.; Seo, H.; Lee, H.J.; Kim, G.; Nie, C.; Hong, Y.; Yoon, Y.J.; Lim, J.Y.; Park, S.J.; et al. Microfluidic Generation of Exosome-Mimetic Nanoparticles for Scalable Production and Enhanced Therapeutic Efficacy. *Small* **2025**, *21*, e06162, doi:10.1002/smll.202506162.
625. Fan, S.; Wang, W.; Che, W.; Xu, Y.; Jin, C.; Dong, L.; Xia, Q. Nanomedicines Targeting Metabolic Pathways in the Tumor Microenvironment: Future Perspectives and the Role of AI. *Metabolites* **2025**, *15*, doi:10.3390/metabo15030201.
626. Wang, Y.; Kohane, D.S. External triggering and triggered targeting strategies for drug delivery. *Nature Reviews Materials* **2017**, *2*, 1-14.
627. Sanadgol, N.; Abedi, M.; Hashemzaei, M.; Kamran, Z.; Khalseh, R.; Beyer, C.; Voelz, C. Exosomes as nanocarriers for brain-targeted delivery of therapeutic nucleic acids: advances and challenges. *J Nanobiotechnology* **2025**, *23*, 453, doi:10.1186/s12951-025-03528-2.
628. Schreiner, T.G.; Menéndez-González, M.; Schreiner, O.D.; Ciobanu, R.C. Intrathecal Therapies for Neurodegenerative Diseases: A Review of Current Approaches and the Urgent Need for Advanced Delivery Systems. *Biomedicines* **2025**, *13*, doi:10.3390/biomedicines13092167.

629. Sebghatollahi, Z.; Yogesh, R.; Mahato, N.; Kumar, V.; Mohanta, Y.K.; Baek, K.H.; Mishra, A.K. Signaling Pathways in Oxidative Stress-Induced Neurodegenerative Diseases: A Review of Phytochemical Therapeutic Interventions. *Antioxidants (Basel)* **2025**, *14*, doi:10.3390/antiox14040457.
630. Mehta, R.S.; Kochar, B.D.; Kennelty, K.; Ernst, M.E.; Chan, A.T. Emerging approaches to polypharmacy among older adults. *Nat Aging* **2021**, *1*, 347-356, doi:10.1038/s43587-021-00045-3.
631. de Koning, L.A.; Vazquez-Matias, D.A.; Beaino, W.; Vugts, D.J.; van Dongen, G.; van der Flier, W.M.; Ries, M.; van Vuurden, D.G.; Vijverberg, E.G.B.; van de Giessen, E. Drug delivery strategies to cross the blood-brain barrier in Alzheimer's disease: a comprehensive review on three promising strategies. *J Prev Alzheimers Dis* **2025**, *12*, 100204, doi:10.1016/j.tjpad.2025.100204.
632. Vargas, R.; Martinez-Martinez, N.; Lizano-Barrantes, C.; Pacheco-Molina, J.A.; García-Montoya, E.; Pérez-Lozano, P.; Suñé-Negre, J.M.; Suñé, C.; Suñé-Pou, M. Advancing through the blood-brain barrier: mechanisms, challenges and drug delivery strategies. *Admet dmpk* **2025**, *13*, 2988, doi:10.5599/admet.2988.
633. Durham, P.G.; Butnariu, A.; Alghorazi, R.; Pinton, G.; Krishna, V.; Dayton, P.A. Current clinical investigations of focused ultrasound blood-brain barrier disruption: A review. *Neurotherapeutics* **2024**, *21*, e00352, doi:10.1016/j.neurot.2024.e00352.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.