

Review

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Posted Date: 2 June 2025

doi: 10.20944/preprints202506.0107.v1

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Review

Electrochemical and Optical Biosensors: Innovation in **Electrode Material**

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Abstract: New developments in biosensing technologies have transformed the entire field of diagnostics. It is possible to diagnose health problems via this technology in ways that are more accurate, faster, and less invasive than ever before. The article provides a nice overview of the newest biosensing technologies. It also discusses key advances in methods of detection, fabrication techniques, and materials used. Various detection technologies have been developed. These include biosensors, microfluidic devices and lab on a chip devices. Moreover, the studies take the pulse of progress in areas such as electrochemical and optical biosensors. In particular, indium tin oxide ranks among interesting new materials and holds promise for further advances. Moreover, various 'nano' materials, such as silver nanoparticles, are being studied along with new metalorganic frameworks. By incorporating such advances, this study can make previews more thorough and accurate. This review offers insights that are valuable to both researchers studying biosensors in industry and people practicing as doctors.

Keywords: microfluidic devices; electrochemical biosensors; optical biosensors; indium tin oxide; silver nanoparticles; metalorganic frameworks; silicon materials; nanotechnology; diagnostic tools

1. Introduction

Biosensors turn biological responses into electrical signals. The devices had three main parts. The first part was the bioreceptor. It interacts with the target substance. Examples of bioreceptors include enzymes, antibodies, nucleic acids, and cells. The choice of a bioreceptor depends on the target analyte and the required sensitivity and specificity of the detection system. The second part was the transducer. This changed the biorecognition event into a signal. The signal can be electrical, optical, thermal, or mechanical. The third part was the signal processor. It processes the signal from the transducer and makes it readable. This involved amplifying, filtering, and converting it from analog to digital [1].

Electrochemical biosensors are a common modern chemical technique. Various types exist on the basis of measurement methods. An amperometric biosensor is one that measures the current produced from the oxidation or reduction of a substance, so it is suitable for testing the levels of glucose [2]. A potentiometric biosensor measures potential differences that are related to substance concentration, such as pH meters [3]. A conductometric biosensor detects changes in electrical conductivity caused by changes in the ionic strength or mobility of a substance [1]. An impedimetric biosensor measures changes in impedance after a recognition element has bound with a substance, which is useful for detecting large biomolecules such as proteins [4]. Electrochemical biosensors are vital in diagnostics, environmental monitoring and biomedical research. They provide high specificity and sensitivity because of diverse recognition elements, such as enzymes, antibodies or nucleic acids [2–5].

Optical biosensors use light to detect biological interactions. They offer many options on the basis of different optical methods. Fluorescence-based biosensors measure the concentration of substances by emitting light. SPR biosensors detect changes in the refractive index when substances bind. Optical waveguide-based sensors improve sensitivity by confining light. Colorimetric biosensors are simple and show color changes [1]. Biosensors using Raman technology can detect unique molecular vibrations of specific substances. These methods are especially effective when combined with SERS amplification [6]. These biosensors are crucial in diagnostic and biomedical research. They provide fast, precise, and nonintrusive analysis [7] (Error! Reference source not found.).

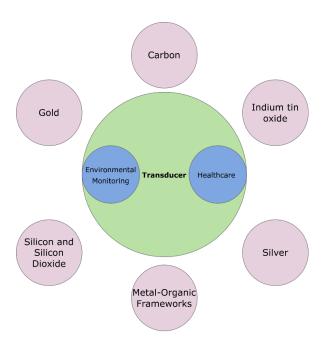


Figure 1. Figure of transducer made with Au electrodes, carbon electrodes, ITO, Ag nanoparticles, MOFs and Si/SiO₂ for health care and environment monitoring applications.

2. Biosensing Technology

This section discusses a variety of disease detection devices. The topics covered include biosensors, lab-on-a-chip devices, imaging technologies, digital PCR, microfluidic devices, assays, blood tests, liquid biopsies, BioMEMS, and genetic testing. These technologies are crucial because they are highly sensitive, fast, and portable. In addition, these methods are noninvasive. They also provide real-time monitoring and personalized treatment plans that are critical for early and accurate disease detection and the management of individual patients.

2.1. Lab On-Chip Devices

LOC technology is used to change the fluid volume includes microfluidic chips that have biological elements built into them that can detect disease biomarkers from fluids. The responses are then converted into measurable signals by a transducer, and its interpretation for the detection unit is used in analysis. The system needs only small sample sizes, which are crucial for the early detection of diseases since low biomarker concentrations are key to accurate diagnosis. In addition, the reduction in size and portability of LOC devices make them ideal for point-of-care testing, especially those suitable in remote areas lacking proper laboratory infrastructure. However, the complex fabrication processes involved in creating LOC systems can increase production costs. In addition, scalability problems exist. These limitations are being addressed by advances in technology, which is expected to reduce costs and improve scalability. Some LOC devices have difficulty detecting more than one analyte at a time; this can complicate the diagnosis of diseases with complex biomarker

profiles. To overcome this, researchers are developing more sophisticated LOC systems that can handle multiple analytes, thereby improving their diagnostic capacity [8].

2.2. Biosensors

This configuration can speed up testing processes, reduce the risk of contamination and increase overall efficiency. A variety of systems include biosensors, which are devices that detect biomarkers. The device consists of a signal processing unit, transducer and biorecognition element. With a biosensor, real-time observation can be realized. Specifically, designed to be the least invasive method of sample collection. Interference from many different unwanted factors can cause the accuracy of a biosensor to decrease. The stability of the biorecognition components gradually decreases with time. This has a direct effect on the reliability of biosensors. To avoid any negative responses, it is important to evaluate the biocompatibility first. For example, a glucose biosensor for diabetes management achieves fast and accurate glucose readings but has problems with interference from other blood components [9]. Error! Reference source not found.

Figure 2. (A) Representation of electrochemical biosensors: (a) amperometric, (b) potentiometric, (c) conductometric, and (d) impedimetric modes.(B) Classification of nanomaterials-based biosensors including nanoparticles, nanorods, nanowires, quantum dots, carbon nanotubes, and dendrimers.(C) Dimensional classification of nanomaterials used in biosensing based on confinement direction and application domains.(D) Optical biosensor schematics: (a)chemiluminescence-based, (b) surface plasmon resonance (SPR)-based, and (c) evanescent wave-based optical biosensor [1].

2.3. Microfluidic Devices

These devices can test for diseases even when material is scarce, and their compactness enables them to be carried around easily. These methods are particularly suitable for emergency diagnosis. All of these microchips, biosensors, pumps, valves, chambers and microchannels are widely used in

this area. Through automation, the efficient screening of large populations and examination of biomarkers in one assay are ensured. Nevertheless, compatibility, universal production standards and intricate fabrication still present major problems. For example, a study developed and demonstrated a portable, efficient microfluidic detector for rapid sepsis testing [10]. This has brought attention to the challenges of standardizing production.

2.4. Assay

An assay is a laboratory test used to look for the presence, quantity, or activity of biomarkers in samples. Assays are the most cost-effective option, especially with many samples. This saves time and money, particularly when working with extensive sample sizes. The protocols for them are standardized, making them suitable for many different environments. However, conventional assays are time-consuming and often require the expertise of skilled personnel, increasing the possibility of human error. Assays can be less portable because of the need for multiple reagents and equipment, and larger sample volumes may limit specific assays. This could limit the number of assays a person can run on any one experiment. For example, a study aknowledge the cost-effectiveness and efficiency of enzyme-linked immunosorbent assays (ELISAs) for the detection of infectious diseases is needed. However, the authors also indicated that a drawback lies in the fact that the process was time-consuming and relied on skilled personnel [11].

2.5. BioMEMS

Biomolecular devices combine biological and electronic parts and are being developed in biological microelectromechanical systems (BioMEMS). These devices, which are smaller than biosensors, are used in real-time data processing and are both useful for controlled drug delivery. Fabricating them demands highly specialized processes, making them expensive. Over time, the stability of materials can compromise the endurance and performance of devices. This creates new concerns with biocompatibility, a frequent reason that companies will have trouble obtaining FDA regulatory approval. Some studies have described the development of a BioMEMS device for controlled drug delivery, including the need for specialized manufacturing techniques and biocompatibility issues [12].

3. Material as Transducer

3.1. Gold (Au) Electrodes

Gold Nanoparticles for Biomolecule Detection

The biosensor uses gold nanoparticles (AuNPs) capped with specific antisense oligonucleotides (ssDNA) targeting the viral nucleocapsid phosphoprotein (N-gene). The sensing probes are immobilized on a paper-based electrochemical platform, creating a nucleic acid testing device suitable for widespread diagnostic use. The readout is recorded using a simple handheld reader, which is practical. The sensor was tested with SARS-CoV-2-infected Vero cells and clinical samples and produced improved output in under 5 minutes. It has a sensitivity of 231 (copies μL^{-1})⁻¹ and a detection limit of 6.9 copies per micro liter without further amplification. Even during viral mutation, ssDNA-conjugated AuNPs targeting two N gene regions allow the sensor to remain functional. This dual-target strategy reduces the chance of false negatives due to viral mutations [13]. Despite advancements in nucleic acid testing, early detection of breast cancer remains a significant challenge. Achieving high specificity and sensitivity in breast cancer diagnostics is critical for improving outcomes. Early detection significantly improves treatment success, with HER2 as a key biomarker. This study introduces an electrochemical immunosensor for highly sensitive detection of HER 2 using a nanodiamond (nanoD) and gold nanoparticle (AuNP) platform. Nanodiamonds were immobilized on a glassy carbon electrode (GCE), followed by AuNP electrodeposition to increase conductivity.

Techniques such as voltammetry, EIS, TEM, XRD, and FESEM confirmed the material deposition and surface morphology. The sensor achieved a detection limit of 0.29 pg mL⁻¹ under optimal conditions, with minimal cross-reactivity [14]. Although the electrochemical immunosensor for HER 2 offers high sensitivity and specificity, its focus is limited. Broader applications are necessary to increase its versatility across different biomarkers. This study presents a method for lysozyme detection using a gold nanoparticle-based biosensor. The biosensor is integrated with decomposition Muller matrix polarimetry to increase accuracy and sensitivity. A DNA aptamer with specificity for lysozyme is used for precise binding in the detection process. Gold nanoparticles enhance biosensor sensitivity through their unique optical properties. The detected signal was processed via a Muller matrix. This focuses on depolarization, linear diattenuation, and depolarization index measurements. The results revealed a linear relationship between the optical parameters and lysozyme concentration. The dynamic range observed was between 0.01 and 500 pM with high precision. The limit of detection (LOD) was determined to be 1.24 fM, confirming the device's sensitivity. These results highlight the potential of biosensors for expanding diagnostic applications. The use of gold nanoparticles greatly improves sensor capabilities[15]. Lysozyme detection is highly sensitive but lacks the ability for realtime biomarker monitoring, which is essential for infectious diseases. This remains a challenge. In this study, a QCM biosensor was developed to detect HBsAg, and antibodies were covalently attached to primary amines for detection. Such improvements are key. Surface modifications were performed using PEI and thiolated PEI, which improved electrode performance and enabled more efficient detection. This optimized the process. RSM optimization revealed no significant difference in immobilization yield between the modified layers used for the biosensor. Testing confirmed success. Surface analysis using FESEM, AFM, FTIR, and CA revealed increased hydrophilicity and roughness, enhancing the biosensor performance. This improved the detection reliability. The QCM biosensor demonstrated a wide dynamic range from 1 to 1 × 10³ ng/mL. The accuracy was very high. This makes it a promising tool for noninvasive and timely monitoring of HBV in human serum samples. Healthcare outcomes have improved greatly [16] (Figure 1).

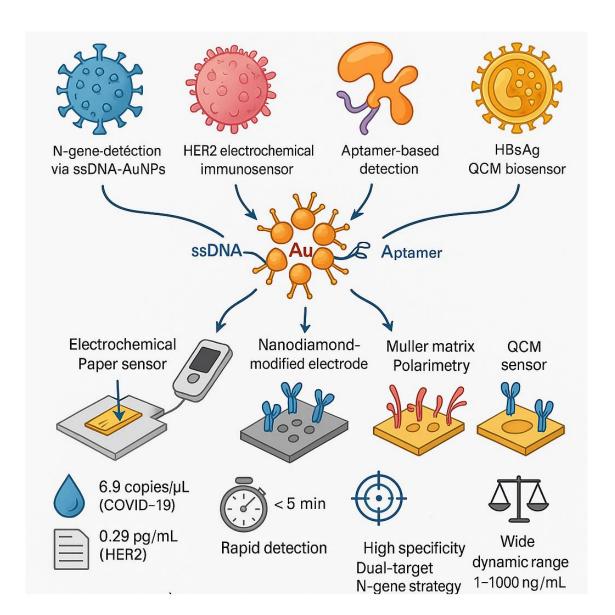


Figure 1. Gold nanoparticle-based biosensing platforms for rapid and sensitive detection of SARS-CoV-2 Ngene, HER2, lysozyme, and HBsAg using electrochemical, optical, and QCM detection methods.

Electrochemical Detection in Biological Monitoring

The research focuses on creating a sensitive and flexible sensor for real-time monitoring of glucose and lactate levels in sweat. A breakthrough in flexibility. The working electrode was enhanced with gold nanopine needles (AuNNs) via electrochemical deposition, effectively amplifying signals and increasing sensor sensitivity. Precision was at every step. For enzyme immobilization, poly(ethylene glycol) diglycidylether (PEGDE) offered better enzyme activity retention than glutaraldehyde did, ensuring long-term sensor performance. Efficiency redefined in biosensing. The biosensor demonstrated excellent catalytic performance with AuNNs, achieving low detection limits of 7 μ mol L⁻¹ for glucose. Unmatched sensitivity was achieved here [17]. Although the AuNN-based sensor is sensitive and flexible, integrating sensors into wearable applications requires materials that can endure stress. The intrinsic power of gold manifests itself through its active influence. Gold is ideal for sensing because of its chemical inertness and broad electrochemical window, making it perfect for biosensing applications. This innovation meets real-world needs. Using a dry-spinning method, stretchable and conductive gold fibers fabricated that maintained conductivity even under mechanical stress. Stretchable and still powerful. These fibers were used to create lactate-sensing working electrodes, reference electrodes, and counter electrodes for integration

into wearable textiles. Wearable tech with impact can be produce. The textile biosensors exhibited high sensitivity, with 19.13 μ A/mM cm² in PBS and 14.6 μ A/mM cm² in artificial sweat. Performance under any strain [18]. However, as biosensing applications expand, there is a growing demand for devices that can withstand stress, offering ultrawide detection ranges. There demand is rising. The development of blood glucose monitoring devices requires greater sensitivity and lower detection limits to enhance biosensor performance and reliability. These advancements have boosted diagnostics. In this study, we fabricated a high-performance flexible enzymatic glucose biosensor by electrochemically depositing dendritic gold nanostructures on a carbon cloth substrate. The addition of gold greatly improved the conductivity. Glucose oxidase was immobilized on the dendritic gold, and enzyme loading was enhanced using the enzyme precipitation coating method. This increases the detection efficiency. Scanning electron microscopy (SEM) was used to characterize the dendritic gold nanostructures in detail to confirm successful deposition and structure. Visualization aids precise measurement. Cyclic voltammetry (CV), chronoamperometry, and electrochemical impedance spectroscopy (EIS) were used to evaluate the overall electrochemical performance of the biosensor. Multiple tests ensure reliability. The biosensor exhibited high sensitivity (72.45 $\mu A~mM^{-1}$ cm⁻²), a low detection limit (6.7 µM), and a broad linear glucose detection range. These results indicate precision. It also demonstrated good selectivity, reproducibility, and stability, with strong accuracy when tested in real serum samples for glucose detection. It is accurate for clinical use. These results suggest that the new biosensor holds great potential for future applications in the biomedical and healthcare sectors. Promising innovations are needed ahead [19]. For comprehensive diagnostics, handling multiple analytes at once is crucial to ensure accurate and reliable results in medical tests. This is vitally important. In another study, a highly sensitive electrochemical sensor was designed for glucose detection using carbon nanotubes (CNTs) grown in situ at low temperatures. On a glass substrate. This setup involved CNTs grown on photolithographically defined gold microelectrode arrays (CNT/Au MEAs) placed on a glass substrate. Each sample is distinct. Selectivity was achieved by modifying CNT/Au MEA with the glucose oxidase (GOx) enzyme immobilized in poly(paraphenylenediamine). It improves sensor precision. The biosensor showed a linear response in the 0.2–27.5 μ M range, with a sensitivity of 168.03 $k\Omega^{-1}$ M $^{-1}$ and low detection limits. The accuracy was confirmed further. It demonstrated excellent anti-interference properties and was validated through HPLC in blood serum samples, ensuring accuracy in real-world applications. Validation through HPLC was previously reported [20]. Overall, this research highlights advancements in biosensor technology, with improved sensitivity, stability, and versatility for medical monitoring

Advanced Nanostructures for Specific Detection

applications. It is useful for real diagnostics.

A biosensor system for isoprocarb detection was developed, utilizing acetylcholinesterase (AChE) inhibition by isoprocarb as the fundamental detection mechanism. Gold nanoparticles modify electrode. A gold nanoparticle-polyaniline-modified graphite pencil electrode (AuNP-PANI-GPE) was employed to detect thiocholine changes in response to isoprocarb. In the process two fabrication steps followed. The electrode was fabricated using cyclic voltammetry: electropolymerization of aniline on a graphite pencil and then electrodeposition of gold nanoparticles. Gold nanoparticles were observed. SEM-EDX revealed that gold nanoparticles ranging from 8–80 nm were successfully deposited on the modified electrode surface. A larger active area formed. Cyclic voltammetry indicated that the active surface area was 0.17019 cm², which was larger than that of both the PANI-GPE and unmodified GPE. Oxidation peak recorded. The oxidation peak of thiocholine at +0.675 V (vs. Ag/AgCl) increased with increasing acetylthiocholine concentration and decreased in the presence of isoprocarb. Linear curve observed. Under optimal conditions, a linear calibration curve for isoprocarb (0.05–1.0 μ M) was generated, demonstrating high accuracy. It has impressive detection limits. The detection and quantification limits were 0.1615 nM

and 0.5382 nM, respectively, with a sensitivity of 1.7771 μ A/ μ M.mm². Also, excellent stability was achieved. The electrode showed strong stability, with an RSD of 4.87% over eight measurements, making it promising for real-world detection. The scope of future applications is vast [21]. To increase the sensitivity and broaden detection beyond isoprocarb, a new folic acid biosensor was developed in which dihydrofolic acid reductase (DHFR) was immobilized. The titanium nanoparticles were characterized. The biosensor utilized a c-MWCNT/TiO2NP-modified gold electrode and was analyzed at various stages using SEM, EIS, FTIR, and cyclic voltammetry. The pH was optimal. The biosensor exhibited a low detection limit of 11.48 nM, a wide linear range (5–50 nM), a sensitivity of 0.42 μ A/nM/cm², and excellent stability. The maximum current was appear at 0.125 V . This method proved effective for folic acid detection in serum samples from pregnant women, showing potential in clinical applications. As well stability was maintained [22]. Overall, advancements from isoprocarb detection to folic acid biosensing have improved biosensor sensitivity, specificity, and application range for biomolecules.

Table 1. Gold-Based Electrodes for Biosensing Applications.

							, 11			
Title	Summary	Targe	Material	Techniq	Dete	Sensiti	Measureme	Key	Applica	R
		t		ue	ction	vity	nt Method	Features	tions	ef
		Analy			Limit					
		te			(LOD					
)					
Gold	Biosensor	SARS	AuNPs,	Paper-	6.9	231	Hand-held	Dual-	SARS-	[1
(Au)	using	-CoV-	ssDNA	based	copie	(copies	reader	target	CoV-2	3]
Electrod	AuNPs	2		electroch	$s/\mu L$	μL–		approac	detectio	
es for	capped	RNA		emical		1)-1		h, no	n	
SARS-	with			platform				amplific		
CoV-2	antisense							ation		
Detectio	ssDNA							needed,		
n	targeting							rapid		
	the viral N-							detectio		
	gene.							n (< 5		
								mins)		
Electroc	Highly	HER	NanoDia	GCE,	0.29	Not	Differential	High	Breast	[1
hemical	sensitive	2	monds,	DPV	pg	specifie	pulse	specificit	cancer	4]
Immuno	detection		AuNPs		mL-1	d	voltammetr	y, low	biomar	
sensor	of HER 2						y (DPV)	cross-	ker	
for HER	using a							reactivit	detectio	
2	NanoDiam							y,	n	
Detectio	ond and							optimal		
n	AuNP							at 35 °C,		
	nanohybri							pH 7.2		
	d platform.									

Gold	Detection	Lysoz	Gold	Muller	1.24	Not	Muller	High	Lysozy	[1
Nanopar	of	yme	nanopart	matrix	fM	specifie	matrix	specificit	me	5]
ticle-	Lysozyme		icles,	polarime		d	calculation	y, broad	detectio	
Based	using gold		DNA	try				dynamic	n	
Biosenso	nanopartic		aptamer					range		
r for	les and							(0.01 to		
Lysozym	decomposi							500 pM)		
e	tion Muller									
Detectio	matrix									
n	polarimetr									
	y.									
Flexible	Real-time	Gluco	AuNNs	Electroch	7	Not	Electrochem	High	Glucose	[1
Sensor	monitorin	se,	Autvivs	emical	μmol	specifie	ical	selectivit	and	[1 7]
for	g of	Lactat		depositio	μπιοτ L-1	d	detection		lactate	7]
Glucose	glucose	e		•	(gluc	u	detection	y, stability,	monitor	
and	and lactate	e		n, PEGDE	ose),			reprodu	ing in	
Lactate	in sweat			TEGDE	54			cibility	sweat	
Monitori	using a				μmol			Cibility	Sweat	
ng	flexible				L-1					
116	chip with				(lacta					
	AuNNs.				te)					
	1101110				tc)					
Stretcha	Fabricatio	Lactat	Gold	Dry-	Not	19.13	Electrochem	High	Wearab	[1
ble Gold	n of	e	fibers	spinning	specif	μA/m	ical	sensitivit	le	8]
Fiber-	stretchable			, three-	ied	M cm ²	detection	y,	lactate	
Based	, strain-			electrode		(PBS),		maintain	monitor	
Lactate	insensitive			system		14.6		S	ing	
Biosenso	gold fibers					μA/m		perform		
rs	for lactate					M cm ²		ance		
	sensing in					(artifici		under		
	wearable					al		100%		
	application					sweat)		strain		
	s.									

Flexible	High-	Gluco	Dendritic	Electroch	6.7	72.45	SEM, CV,	High	Biomed	[1
Enzymat	performan	se	gold	emical	μM	μΑ	chronoamp	sensitivit	ical and	9]
ic	ce glucose		nanostru	depositio		mM-1	erometry,	y, low	health	
Glucose	biosensor		ctures,	n,		cm-2	EIS	detectio	care	
Biosenso	using		carbon	enzyme				n limit,		
r	dendritic		cloth	precipita				wide		
	gold			tion				linear		
	nanostruct			coating				range,		
	ures on							good		
	carbon							selectivit		
	cloth.							y,		
								reprodu		
								cibility,		
								stability		
QCM-	QCM-	HBsA	Anti-	PEI and	3.14	Not	FESEM,	Broad	Non-	[1
Biosenso	biosensor	g	HBsAg	thiolated	ng/m	specifie	AFM, ATR-	dynamic	invasiv	6]
r for	for label-		antibodie	-PEI	L	d	FTIR, CA	range,	e	
HBsAg	free		s, gold	surface			measureme	high	monitor	
Detectio	detection		electrode	modifica			nt	accuracy	ing of	
n	of HBsAg.			tions,				, good	HBV-	
				RSM				selectivit	biomar	
				optimiza				y,	ker	
				tion				stability,		
								regenera		
								bility		
Isoproca	Biosensor	Isopr	Gold	Electro-	0.161	1.7771	SEM-EDX,	High	Real	[2
rb	system	ocarb	nanopart	polymeri	5 nM	μΑ/μ	CV	sensitivit	detectio	1]
Detectio	based on		icles,	zation,		M.mm ²		y,	n of	
n	AChE		polyanili	electro-				excellent	isoproc	
Biosenso	inhibition		ne,	depositio				stability	arb	
r	by		graphite	n						
	isoprocarb		pencil							
	using		electrode							
	AuNPs-									
	PANI-									
	GPE.									

Electroc	Sensor	Gluco	CNTs,	Immobili	0.2	168.03	CV, EIS	S	Good	Multipl	[2
hemical	using	se	gold	zing	μM	kΩ-1			reprodu	e	0]
Sensor	CNTs		microelec	GOx		M-1			cibility,	electroa	
for	grown on		trode	enzyme,					anti-	ctive	
Glucose	Au MEA		arrays	poly (p-					interfere	biomole	
Detectio	for glucose			PDA)					nce,	cules	
n	detection.			matrix					validate	detectio	
									d	n	
									through		
									HPLC		
Folic	Biosensor	Folic	C-	ТЕМ,	11.48	0.42	SEM,	EIS,	Wide	Folic	[2
								,			•
Acid	for folic	Acid	MWCNT	XRD,	nM	μA/nM	FTIR, C	CV	linear	acid	2]
Biosenso	acid		, TiO2	FTIR,		/cm ²			range,	quantifi	
r	detection		nanopart	SEM,					good	cation	
	using		icles,	EIS, CV					storage	in	
	DHFR		gold						stability	serum	
	immobiliz		electrode							samples	
	ed on c-										
	MWCNT/										
	TiO2NPs										
	modified										
	Au										
	electrode.										

3.2. Carbon-Based Electrodes

Electrochemical Sensors for Biological Molecules

A novel screen-printed carbon electrode (SPCE) with multiple working electrodes and a single signal output channel has been developed. This enhances immunosensing applications. The design eliminates counter and reference electrodes, reducing costs and environmental impact by excluding precious metals completely. This lowers waste. The SPCE enables simultaneous detection of various analytes through individually modified working electrodes for enhanced efficiency. Each electrode was performed independently. An independent platinum network acts as a counter electrode, improving the reproducibility and reliability of the SPCE during operation. This ensures accuracy. A hydrogel on a working electrode improves conductivity, significantly enhancing the overall sensitivity of the electrode in use. This increases the conductivity. The SPCE was applied to a multiplexed, label-free amperometric immunosensor for detecting four tumor markers with high precision. The detection limits improved. This method achieved detection limits as low as 5.5 pg mL⁻¹ for SCCA and 2.3 pg mL⁻¹ for NSE. Such low limits improve the performance[23]. Additionally, a nickel ferrite/reduced graphene oxide (NiFe₂O₄/rGO) nanocomposite was synthesized and used to fabricate a bioelectrode on a screen-printed carbon electrode (SPCE). This improved electrochemical biosensing. The uricase/nickel ferrite/reduced graphene oxide/screen-printed carbon electrode (Uricase/NiFe₂O₄/rGO/SPCE) demonstrated enhanced performance, with a linear range from 5 to 900 micromolar (μM) and a detection limit of 21.9 micromolar (μM). The sensor gets excellent stability and selectivity. The biosensor showed repeatability, making it suitable for detecting uric acid (UA)

in real samples with point-of-care potential. It is used for UA disorders [24]. Screen-printed carbon electrodes (SPECs) are commonly utilized in point-of-care testing (POCT) because of their affordability, disposability, and simple design. The surface modification is challenging. Unlike gold or platinum electrodes, modifying the SPCE carbon surface is difficult because of its inherent stability properties. Carboxyl groups introduced. Oxygen plasma (O₂) treatment can enable covalent bonding by introducing carboxyl groups on the SPCE carbon surface. It has improved bonding efficiency. This research studied the effect of oxygen plasma (O₂) treatment using a novel immunosensor with gold nanoparticles (AuNPs) on electrode performance. Four modifications were compared. The modifications tested were oxygen plasma-treated/covalent-bonded antibodies (a), oxygen plasmatreated/physically adsorbed antibodies (b), bare/covalent-bonded antibodies (c), and bare/physically adsorbed antibodies (d). The results showed varying limits. The detection limits were 0.50 nanograms per milliliter (ng/mL), 9.7 nanograms per milliliter (ng/mL), 0.54 nanograms per milliliter (ng/mL), and 1.2 nanograms per milliliter (ng/mL) across the four configurations tested. Higher sensitivity was observed. The oxygen plasma-treated electrodes had an increased number of carboxyl groups, which enhanced antibody adsorption and improved sensor sensitivity and performance. Effective surface modification. Oxygen plasma (O2) treatment was found to significantly improve surface modification, resulting in better antibody binding and sensor performance [25]. Additionally, a glucose biosensor incorporating Prussian blue nanoparticles (PBNPs) and carbon nanotubes was developed using sweat for noninvasive glucose monitoring. GOx was immobilized. Glucose oxidase (GOx) was immobilized by chitosan and encapsulated in Nafion to ensure stability and detection performance. Detection limit was 7 μ M. The GOx/PBNP/MWCNT-COOH sensor demonstrated a low detection limit, high sensitivity, and excellent interference resistance across the sweat glucose range. Suitable for diabetic patients. The sensor, which was applied to screen-printed carbon electrodes (SPECs), maintained stability for two weeks, showing promise in personalized medical detection. Stability achieved was expecting in future [26]. These advancements in SPCE technologies highlight their potential for cost-effective, reliable biosensors in medical diagnostics and environmental monitoring applications. The SPCE is promising (Error! Reference source not found.).

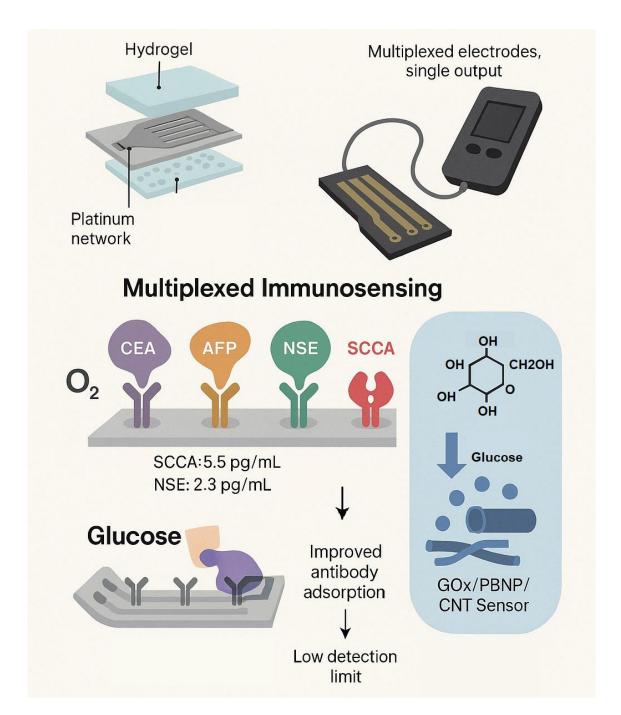


Figure 4. Versatile SPCE platform enabling multiplexed tumor marker detection and noninvasive glucose monitoring with enhanced sensitivity and portability.

Electrochemical and Optical Sensors for Environmental and Pharmaceutical Applications

A highly sensitive electrochemical sensor has been developed for detecting doxorubicin hydrochloride (DOX), a potent antitumor agent. DOX was detected well. The sensor uses a glassy carbon electrode modified with multiwalled carbon nanotubes (MWCNTs) decorated with gold nanoparticles. Nanotubes has enhanced performance. Characterization techniques, such as scanning electron microscopy (SEM), confirmed the successful decoration of the MWCNTs with gold nanoparticles. SEM confirmed the success of the study. Cyclic and linear sweep voltammetry showed efficient catalytic activity for DOX reduction, with increased peak current and reduced overpotential. It has effective catalytic activity. The sensor achieved a linear detection range from 10^{-11} to 10^{-6} M, with a low detection limit of 6.5 pM. It has wide detection range [27].

Single-walled carbon nanotubes (SWCNTs) have shown great promise for near-infrared fluorescence-based biosensing in various applications. This study introduces a reusable sensor. The

drop-cast optical biosensor uses peptide-encapsulated SWCNTs to detect low concentrations of acetic acid in air. It is reusable. SWCNTs, specifically those with (6,5) chirality, exhibit peak fluorescence in the 970–1050 nm range, which is compatible with low-cost silicon-based detectors. It is also stable. Peptide encapsulation enhances stability and sensitivity, allowing the detection of acetic acid vapor at concentrations as low as 0.05%. It is crucial. This advancement is important for cost-effective, realtime SWCNT-based gas phase detection in areas such as wine spoilage monitoring [28]. Along with this a new method has emerged. An enzymatic biosensor for the indirect detection of organophosphates (OPs) was developed that utilizes acetylcholinesterase inhibition for detection. The biosensor used the electrochemical principle. The sensor was built on a screen-printed carbon electrode modified with copper nanowires (CuNWs) and reduced graphene oxide (rGO). The current was measured. The oxidation current was measured through cyclic voltammetry (CV), which is generated by the enzymatic interaction between acetylcholinesterase and acetylthiocholine. Signal reduction happened. The biosensor response showed signal reduction due to AChE inhibition by an organophosphate inhibitor in the test sample. This enhanced the sensitivity. The CuNW/rGO nanocomposite enhanced the signal current and lowered the oxidation potential for chlorpyrifos detection in the test solution. It is highly effective. The detection range was 10–200 µg/L, with a limit of detection of 3.1 µg/L and a quantification limit of 12.5 µg/L. It detects chlorpyrifos [29]. A hydrogen peroxide (H2O2) sensor was developed using a calcium titanate-modified electrode (CaTiO3@SPE). The electrode synthesized via a hydrothermal method was characterized using a XRD, EDX, SEM, and BET, and CaTiO3 had a specific surface area of 57.6 m²/g. The sensor, which was fabricated through drop-casting, exhibited a detection limit of 0.08 µM for H2O2 with good selectivity. Stability and repeatability are also notable features [30]. These advancements highlight the continuous innovation and diversification of sensor technologies, which improve detection capabilities across a range of applications. Error! Reference source not found.

Figure 5. (A) Cyclic voltammograms of ferricyanide and ruthenium complexes on bare and plasma-treated electrodes at varying scan rates.(B) XPS spectra confirming increased surface carboxylation after plasma treatment.(C) Differential pulse voltammograms for IgA detection under four modification strategies: plasma-treated/covalent bonding, plasma-treated/physical adsorption, bare/covalent bonding, and bare/physical adsorption.(D) Electrochemical kinetics analysis showing diffusion current relationships and calculated electroactive surface areas.(E) Electrochemical impedance analysis comparing charge transfer resistance (Rct)

and double-layer capacitance (Cdl) across electrode treatments.(F) Calibration curves for IgA detection demonstrating lowest limit of detection (LOD) and highest sensitivity with plasma-treated electrodes and covalent antibody immobilization.

The biosensor represents a significant improvement in point-of-care uric acid monitoring technology. The working electrode is based on a 3D SACNT array immobilized with uricase through precipitation and crosslinking, which increases the enzyme density and contact area with reactants while preserving the conductivity of the SACNT structure. The biosensor showed impressive sensitivity, measuring 518.8 μA/(mM·cm²), with an operating range of 100-1000 μM and a low detection limit of 1 µM. Dynamic uric acid monitoring was validated in serum samples, with no significant difference compared with an FDA-approved electrochemical analyzer (paired t test, p > 0.05). Its large surface area and electrocatalytic activity indicate its potential for broader point-of-care biomolecule monitoring applications [31]. The SACNT array biosensor demonstrated excellent sensitivity but focused mainly on uric acid detection. To solve this problem, a new biosensor was developed using carbon dots. These carbon dots (CDs) were synthesized from curcumin and dimethylformamide (DMF) through microwave irradiation, resulting in CDD-CDs. The CDD-CDs were functionalized with 3-(aminopropyl)-triethoxysilane (APTES), forming APT-CDs. Laccase was then covalently immobilized onto APT-CDs. This created a novel bioprobe. The CDD-CDs emitted orange fluorescence at 586 nm, APT-CDs emitted green fluorescence at 533 nm, and the bioprobe emitted blue fluorescence at 476 nm. The bioprobe could detect dopamine linearly from 0 to 30 μ M, with a detection limit of 41.2 nM. For the tapered optical fibers, the detection limit improved to 46.4 nM across a range of 0-10 µM. This material showed high biocompatibility and excellent stability. This finding was validated in human serum and cerebrospinal fluid, confirming its clinical potential [32]. These new biosensors represent an innovative step forward in enhancing sensitivity and specificity for biochemical monitoring. They show promise for applications in both clinical diagnostics and environmental analysis.

3.3. Indium Tin Oxide (ITO)

Electrochemical Biosensors Using ITO for Pathogen Detection

This article explores the development of advanced electrochemical biosensors using indium tin oxide (ITO) electrodes for improved sensitivity and detection. Biosensors detect pathogens. These biosensors are crucial for detecting proteins and pathogens with high sensitivity and specificity in medical diagnostics. They are reliable tools. This study presents a electrochemical DNA biosensor for detecting Shigella flexneri via innovative detection techniques. The detection method is label-free. The sensor utilizes ITO electrodes, where a detection probe is attached to poly melamine (P-Mel) and polyglutamic acid (PGA). Probes are crucial in this method. Disuccinimidyl suberate (DSS) was used to prepare the flexible ITO electrode, and anthraquinone-2-sulfonic acid monohydrate sodium salt (AQMS) served as a signal indicator. This increases the sensitivity. This biosensor detects S. flexneri DNA at concentrations ranging from 1×10^{-6} to 1×10^{-21} mol/L, with a detection limit of 7.4×10^{-22} mol/L. The detection limit was acceptable . In real samples, it detects S. flexneri from 8×10¹⁰ to 80 cells/mL, with a detection limit of 10 cells/mL.Here, accuracy is essential [33]. Another study introduced an immunosensor for detecting GM2 activator protein (GM2A) using an ITO substrate with enhanced sensitivity. It detects GM2A. This substrate is enhanced with gold nanoparticles (GNPs) and an amino-functionalized thiophene polymer (P(ThiAmn)) multilayer. Gold enhances detection. To create this biosensor, GNPs are deposited on the substrate, and ThiAmn is electropolymerized to increase the surface area. The area increases attachment. This allows for the attachment of many anti-GM2A biorecognition elements, improving the overall detection capability and reliability in biosensing applications. The attachment is crucial. Electrochemical impedance spectroscopy (EIS) was used to study the specific interaction between anti-GM2A antibodies and GM2A antigens for targeted detection. It measures interactions. Under optimal conditions, GM2A is detected in a linear

concentration range from 0.0185 to 111 pg/mL, with a detection limit (LOD) of 5.8 fg/mL. It is highly sensitive. This biosensor shows good reproducibility, long storage stability, and excellent specificity for GM2A antigens in various applications. The results are reproducible [34]. Additionally, a new platform has been developed for the detection of heat shock protein 70 (HSP70) via advanced techniques. This platform uses ITO. It employs a three-electrode system on a chip, modifying the reference and working electrodes for enhanced detection. HSP70 antibody assembly. The PS-AuNPs@Cys/Au film deposited on ITO glass provides an excellent substrate for antibody attachment, amplifying signals. This enhances detection. Under optimal conditions, the sensor shows a linear range from 0.1 ng/mL to 1000 ng/mL for protein detection. The limit was 25.7 pg/mL. This method detects HSP70 in normal human blood samples and outperforms the commonly used ELISA method for analysis. In this way, HSP70 has been detected [35]. Additionally, a novel label-free impedimetric immunosensor was developed for sensitive and selective analysis of the Aβ42 protein. Rapid detection occurs. This immunosensor uses cost-effective, disposable ITO-PET electrodes modified with 3-glycidoxypropyldimethoxymethylsilane (GPDMMS) for functionality. Functional groups are form. The interaction between anti-Aβ42 and Aβ42 was analyzed via electrochemical impedance spectroscopy (EIS) and cyclic voltammetry (CV). These changes are confirmed. Morphological changes on the electrode surface during each immobilization step were confirmed by scanning electron microscopy (SEM). The sensitivity is high. The immunosensor shows a linear detection range from 1 to 100 pg/mL, with a detection limit of 0.37 pg/mL. This method is highly selective and stable. For the first time, the kinetic behavior of the antibody-antigen complex was analyzed via singlefrequency impedance (SFI). The binding is very well. Its potential for clinical application was confirmed by analyzing Aβ42 levels in human serum, highlighting its diagnostic utility. The patient's diagnosis was confirmed [36]. Overall, these studies highlight the advancements in ITO-based biosensors with enhanced sensitivity, specificity, and versatility for biomedical use (Error! Reference source not found.).

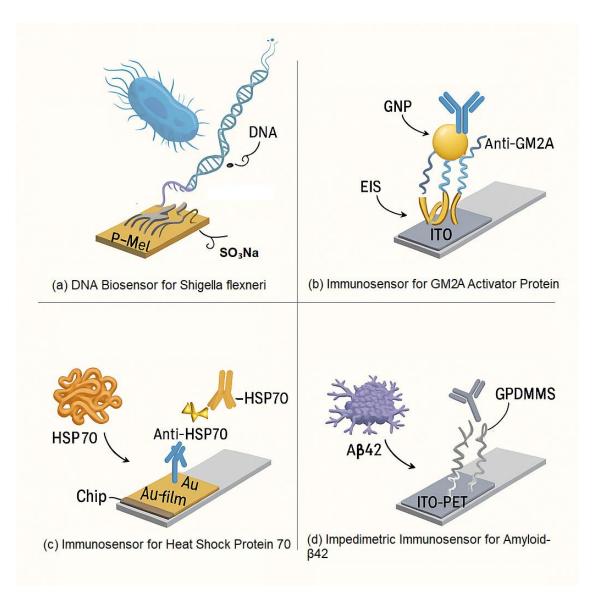


Figure 6. Advanced ITO-based electrochemical biosensors for ultra-sensitive detection of Shigella flexneri, GM2A, HSP70, and A β 42 in clinical diagnostics.

Biosensors Using ITO for Environmental and Health Applications

Recent advancements in biosensor technology have led to innovative devices made with advanced materials, significantly transforming traditional detection methods. This is revolutionary. One such advancement is the creation of a high-tech optical fiber probe designed for remote cysteine detection with precision. The design is compact. Researchers have coated optical fiber surfaces with indium tin oxide, zinc oxide, and cuprous oxide semiconductor nanomaterials for improved performance. It also increases efficiency. This setup replaces bulky macroscopic spatial light systems with a compact optical fiber path, allowing for remote light injection. The light is guided precisely. The probe acts as a working electrode, interacting with the analyte and directing light where needed for photoelectrochemical reactions. This makes the setup more effective. This fiber-optic-based cysteine biosensor has a linear detection range of 0.01-1 µM, indicating that it is promising for biochemical analysis. It has the scope in future for detection devices [37]. Another exciting development is a biosensor for detecting aflatoxin B1 (AFB1) without labels, which uses advanced nanomaterials for accurate detection. The approach used is effective. This sensor uses gold nanobipyramids placed on an indium tin oxide-coated glass substrate modified with a self-assembled APTES film. The modification is strong. These modifications ensure effective AuNBP formation and immobilization of anti-AFB1 antibodies, enabling precise electrochemical biosensing of AFB1. It is

robust. The sensor has a detection limit of 0.1 nM and a recovery rate of 95-100%, demonstrating robustness in real-world applications. The results are excellent [38]. Additionally, a molecularly imprinted electrochemical sensor using Ti3C2Tx MXene has been developed for detecting bilirubin with high selectivity. It is a reliable method. Ti3C2Tx MXene is synthesized through chemical etching and applied to an ITO electrode surface using the drop-casting method. The process is simple. This modification enhances the binding properties, which are essential for molecularly imprinted polymer formation, and offers excellent electrochemical characteristics. It's very effective. The sensor has a linear range of 10 μ M to 90 μ M and a detection limit of 0.197 μ M, with strong reproducibility. The performance is optimal [39]. A new glucose sensor has also been introduced using a biomoleculeassisted method to deposit copper-cobalt bimetallic heteronanostructures. It is groundbreaking. This method results in a sensitive and disposable glucose-sensing electrode, solving instability issues common with transition metal films. Stability was improved. Typically, metal-based films on ITO electrodes are prone to peeling during electrocatalysis, but this innovation efficiently solves that problem. Peeling can be avoided. Researchers have used methionine as a structure-directing agent, leading to a glucose sensor with high sensitivity and selectivity. The results are accurate. This sensor showed a detection limit of 9.1 μ M and a sensitivity of 1418 μ A mM⁻¹ cm⁻², opening new pathways for stable electrodes. This method is highly promising [40].

Advanced Nanomaterial-Based Sensors

The current study developed a nonenzymatic glucose sensor using iodine-doped reduced graphene oxide with a titanium dioxide nanocomposite (I-rGO@TiO2). It was effective. Titanium dioxide nanoparticles and reduced graphene oxide were synthesized via sol-gel and thermal reduction methods to ensure consistency and quality. The analysis confirmed this finding. The IrGO@TiO2 composite was prepared through a hydrothermal technique, combining I-rGO and TiO2 in a 1:4 ratio. The results were clear. XRD and TEM analysis revealed that nanocrystalline anatase titanium was distributed across the I-rGO sheets, confirming the nanostructure of the composite. The findings were satisfactory. The sensor was fabricated by modifying an indium tin oxide electrode with I-rGO@TiO2, resulting in a wide linear detection range. It analyses well. Additionally, the nanocomposite showed positive temperature coefficient behavior with good conductivity at 200°C. Testing was performed. Antibacterial properties were tested against E. coli and Bacillus subtilis, and promising results were obtained in antimicrobial resistance studies. It shows impressive outcomes [41]. Biosensors have gained interest for the diagnosis of infectious diseases, such as tuberculosis (TB), because of their simplicity and point-of-care potential. They are useful. The incorporation of aptamer molecules and nanomaterials offers advantages such as high binding affinity and low immunogenicity, significantly enhancing aptasensor performance. Great improvements were observed. This study used microwave-synthesized copper indium tin sulfide (CITS) nanomaterials combined with the natural biopolymer chitosan for signal amplification. Better sensitivity is achieved. The optical properties of CITS include strong UV absorption, which is characteristic of kesterite semiconductor nanomaterials, confirming successful optical traits. The optical properties can be applied similar devices . X-ray diffraction confirmed the presence of the kesterite phase, with an average crystallite size of 6.188 nm, confirming the desired material phase. It was reliable. The aptasensor's electrochemical properties were enhanced by 77.5%, and aptamer loading improved by 73.7%, increasing the overall performance. The sensor achieves good performance. The aptasensor showed sensitivity to IFN-γ concentrations, with a limit of detection of 6885 fg/mL and a wide linear range. It also have high accuracy. This sensor exhibited excellent stability, selectivity, and the ability to be applied to real-world sample testing with promising results. It achives certain durability [42]. This study integrates nanocomposites and aptamers, achieving high sensitivity and specificity in biosensors for point-of-care diagnostics.

3.4. Silver Nanoparticles

Glucose Detection Using Nanomaterials

Owing to their unique properties, nanomaterials have been extensively explored in the development of sensors, contributing to reliable sensor designs with enhanced sensitivity and specificity. This study proposes a self-powered biosensor. The fluorescent/electrochemical dualmode biosensor uses DNA-templated silver nanoclusters (AgNCs@DNA) for advanced biosensing in glucose detection applications. AgNC@DNA serves as a probe. The study evaluated its efficacy for glucose detection, using the fluorescence emitted by AgNCs@DNA as the readout signal for glucose levels. Glucose oxidase (GOx) generates hydrogen peroxide. The fluorescence emitted by AgNCs@DNA correlates with hydrogen peroxide levels, making it highly effective for monitoring glucose. GOx facilitates detection. This biosensor is highly sensitive. The electrochemical signal uses AgNCs as charge mediators between GOx and the carbon electrode, improving the detection accuracy. The biosensor achieved low limits of detection (LODs) for optical and electrochemical signals, providing excellent sensitivity for glucose monitoring. Specifically, the LODs were 23 µM for optical readouts and 29 µM for electrochemical readouts, confirming the effectiveness of the biosensor [43]. Stretchable transparent electrodes (STEs) based on silver nanowires (AgNWs) have garnered attention for their superior optoelectronic properties in sensor development. Oxidation is a challenge. AgNWs exhibit low oxidation resistance, which limits their durability in devices utilizing stretchable transparent electrodes. A recent study developed core-sheath nanowires. Ag@Au NWs with a dual-headed matchstick morphology and a gold sheath thickness of 2.5 nm were fabricated. These nanowires enabled STEs with an optical transmittance of 78.7%, 13.0% haze, and excellent mechanical properties. The electrodes resist oxidation. The STEs showed a sheet resistance of 13.5 Ω ·sq. ⁻¹ and a tensile strain capacity of 240%, which were achieved through welding. The dense gold sheath provided oxidation resistance. These Ag@Au NW STEs enabled nonenzymatic glucose biosensors, offering a high sensitivity of 967 $\mu A \cdot mM^{-1} \cdot cm^{-2}$. The detection limit was 125 μM . The performance of these materials shows potential [44]. The high level of glucose present in daily nutrition is a significant factor contributing to conditions such as diabetes (diabetes mellitus) and obesity. Glucose levels must be monitored. Determining glucose concentrations in food production processes is crucial for accurate quality control measures. Biosensors enable fast analysis. Biosensors are bioanalytical devices offering cost-effective, simple analysis and providing quick response times for glucose detection in various settings. Silver nanoparticles (AgNPs) are useful. They can be synthesized through green methods and modify electrodes for efficient glucose biosensing applications. This helps research. Agricultural waste provides a sustainable source for synthesizing these nanoparticles. In this study, an amperometric glucose biosensor was created by modifying a carbon paste electrode (CPE) with waste tea-based silver nanoparticles (WT-AgNPs). The glucose oxidase (GOx) enzyme was immobilized. This was accomplished by cross-linking onto a modified carbon paste electrode (MCPE), enabling effective glucose detection mechanisms. Detection relies on hydrogen peroxide (H₂O₂). Hydrogen peroxide reduction occurred at +0.4 V versus silver/silver chloride (Ag/AgCl), providing clear readings. The biosensor demonstrated a linear range between 0.10 and 1.0 micromolar (µM) glucose[45]. It works in juice. This sensor was successfully applied to detect glucose in commercial fruit juice samples. This method showed excellent reproducibility. Low detection limits, high reproducibility, selectivity, and long shelf-life were all demonstrated. This article explores a sensor. The development of a glossy photopaper (GPP)-based screen-printed chemiresistive interdigitate electrode (SPCIDE) for glucose sensing is highlighted. Interdigitate electrodes (IDEs) were printed. Using screen-printing methods, low-sheet resistance electrodes were made with graphene ink on the GPP. Polyaniline (PANI) was applied. Polyaniline was drop-cast onto the interdigitate electrode to form a chemiresistive matrix. Selective detection is key. The glucose oxidase (GOx) enzyme and green-synthesized silver nanoparticles (GS-AgNPs) were immobilized on PANI to increase glucose selectivity. Amperometric testing was performed. The amperometric measurements revealed a strong linear relationship between the current change and glucose concentration. The coefficient of determination (R2) was high. The sensor demonstrated a detection

limit of 198 nanomolar (nM) and a sensitivity of 291.19 microamperes per millimolar per square centimeter (μA mM⁻¹ cm⁻²). SPCIDE has potential. This sensor holds promise for developing affordable and eco-friendly point-of-care (PoC) diagnostic kits. Mass production is viable[46]. The use of inexpensive screen-printing techniques makes the creation of eco-friendly biosensors feasible for large-scale applications.

Detection Using AgNPs and Other Nanoparticles

In this work, silver nanoparticle (AgNP)/reduced graphene oxide (rGO) nanocomposites were electrodeposited on glassy carbon electrodes (GCEs) to develop electrochemical sensors for hydrogen peroxide (H₂O₂) and dopamine (DA) detection. AgNPs were synthesized successfully. These electrochemical sensors were designed for the sensitive detection of H₂O₂ and DA with high performance. GO reduction occurred effectively. The nanocomposites formed on the electrodes were confirmed by scanning electron microscopy (SEM) and electrochemical impedance spectroscopy (EIS). The method was reliable. The AgNP/rGO/GCE exhibited a linear response to H_2O_2 from 5 μM to 620 μ M, with a sensitivity of 49 μ A mM⁻¹cm⁻². The LOD was 3.19 μ A. For DA, the sensor showed a linear range of 1 μM to 276 μM, with a sensitivity of 7.86 μA mM⁻¹cm⁻² and a limit of detection (LOD) of $0.18 \mu M$. The detection was stable. These sensors could simultaneously detect DA and H_2O_2 without interference, maintaining excellent stability over time with eco-friendly fabrication. The sensors worked reliably. This fabrication method offers great potential for the sensitive detection of DA and H_2O_2 with robust and reproducible results. This is a highly effective approach [47]. A study developed an aptamer-based electrochemical sensor (AEC) for detecting STX via silver nanoparticles (AgNPs) modified with an aptamer. Aptamers were attached to the sensor. Under optimized conditions, AECs exhibited a linear response to STX between 0.04 and 0.15 µM, with high sensitivity and accuracy. Regression equation applied. The detection limit was 1 nM, which is below the regulatory limits for STX in seafood, demonstrating excellent sensitivity for practical applications. The performance was excellent. The potassium ferricyanide (K₃Fe(CN)₀)-etched AgNPs served as a signal source, providing a stable ratiometric electrochemical signal for enhanced sensitivity. STX was detected well[48]. In another study, hollow Prussian blue with ultrasmall silver nanoparticles (Ag-HPB) was fabricated via a coating-etching method for sensor development. Silver nanoparticles (Ag NPs) diffuse into Prussian blue (PB). Prussian blue (PB) was coated on silver nanoparticles (Ag NPs), allowing them to diffuse into the Prussian blue framework, enhancing sensor performance. This increased the conductivity. The biosensing platform combines the electrical conductivity of silver nanoparticles (Ag NPs) with the high enzyme loading capacity of the hollow structure. The performance was enhanced. Using glucose oxidase (GOx) and acetylcholinesterase (AChE), the sensor showed a sensitive glucose response of 24.37 $\mu A~mM^{-1}~cm^{-2}$ and detected trichlorfon. It was highly stable. The detection limit for trichlorfon (TCF) is 2.28 pg/mL, and the system is effective for monitoring trichlorfon in apples[49]. Nanocellulose improved the sensitivity. Nanocellulose derived from hemp (HNC), combined with silver nanoparticles (AgNPs), enhances electrochemical sensing, particularly for detecting lactate in wearable sensors. The electrode performance increased. Hemp nanocellulose (HNC) was extracted through alkali treatment and acid hydrolysis, whereas silver nanoparticles (AgNPs) were nucleated via a self-reduction process to form a nanocomposite. This composite improved the conductivity. A 10 weight percentage (wt%) hemp nanocellulose/silver nanoparticle-polyvinyl alcohol (HNC/AgNP-PVA) modification achieved the highest current response with a redox couple [ferricyanide/ferrocyanide (Fe(CN)6]^{3-/4-}]. Lactate was detected precisely. The modified screen-printed graphene electrode detected lactate concentrations in a linear range of 0-25 millimolar (mM), addressing the 12.5 millimolar (mM) cutoff for muscle fatigue [50]. Wearable sensors are promising. In summary, techniques such as electrodeposition, aptamer immobilization, coating etching, and self-reduction offer diverse approaches to achieve sensitive electrochemical sensing (Error! Reference source not found.).

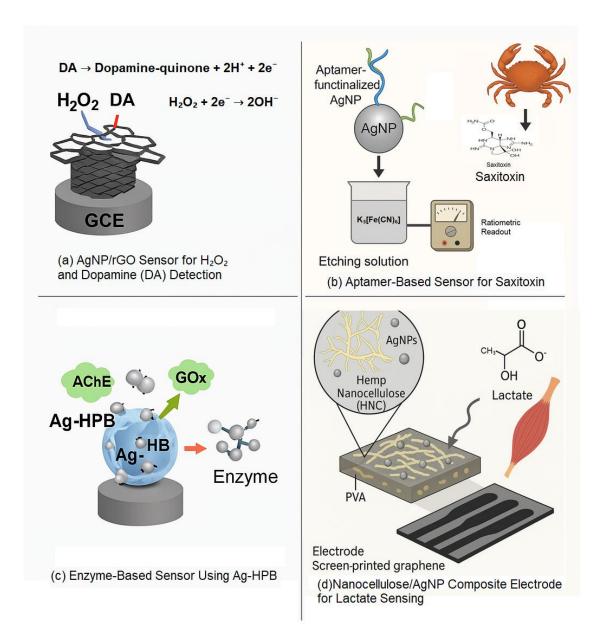


Figure 7. Multifunctional AgNP-based electrochemical sensors for simultaneous detection of H_2O_2 , dopamine, saxitoxin, and lactate with high sensitivity and stability.

Biosensors Using Specific Electrode Materials

Silver/silver chloride (Ag/AgCl) electrodes are widely used in electrochemical biosensing because of their stability, making them reliable reference electrode materials. This stability is essential. However, recent studies have shown that silver/silver chloride (Ag/AgCl) electrodes interact significantly with the alcohol oxidase enzyme, impacting its stability and reducing its effectiveness in biosensing applications. This affects the measurement accuracy. Screen-printed silver/silver chloride (Ag/AgCl) ink was found to negatively affect enzyme stability, as demonstrated by an optical absorbance assay, highlighting a crucial limitation of the material. It reduces enzyme stability. The Ag/AgCl electrode caused a sharp decline in enzyme activity, shortening its duration from one week to just 10 hours in a buffer solution [51]. This accelerates enzyme degradation. Understanding this interaction is essential for improving biosensors by considering alternative materials or strategies to minimize the degradation effect. A potential solution exists. A novel biosensor design features a mini-reactor with lactate oxidase (LOx) integrated with a silver amalgam screen-printed electrode for improved stability. It enhances usability. The mini-reactor, containing mesoporous silica (SBA–15) coated with covalently immobilized lactate oxidase (LOx), allows easy

replacement and extended usability, enhancing performance. This improves durability. Lactate detection is based on monitoring oxygen consumption via four-electron oxygen reduction at -900 mV versus the silver (Ag) pseudo reference electrode. This minimizes interference. With 270 µg of lactate oxidase (LOx) per minireactor, 93.8% of its signal was retained after 350 measurements, and 96.9% of its signal was retained after seven months [52]. This ensures excellent stability. Successfully tested for lactate detection in saliva, wine, and dairy products, it is useful in clinical diagnostics and food quality control applications. A promising solution indeed exists.

Table 2. Silver Nanoparticles in Biosensing.

Title	Summary	Targe	Materia	Techniq	Detectio	Sensitiv	Measur	Key	Applic	R
		t	1	ue	n Limit	ity	ement	Feature	ations	e
		Anal			(LOD)		Method	s		f
		yte								
Impact	Investigates	Alcoh	Ag/AgC	Screen-	N/A	N/A	Optical	Signific	Enzym	[5
of	the interaction	ol	1	printed			absorba	ant	atic	1]
Ag/AgC	between	Oxid	electrod	Ag/AgCl			nce	enzyme	biosens	
1	Ag/AgCl	ase	es	ink,			assay	activity	ors	
Electrod	electrodes and	enzy		optical				degrada		
es on	Alcohol	me		absorban				tion in		
Alcohol	Oxidase			ce assay				presenc		
Oxidase	enzyme							e of		
Stability	stability.							Ag/Ag		
								Cl;		
								enzyme		
								activity		
								halftim		
								e		
								reduced		
								from ~1		
								week to		
								10		
								hours		

Lactate	Design of a	Lacta	Silver	LOx	N/A	L	N/A	Ampero	High	Clinical	[5
Oxidase	biosensor with	te	amalga	mini-				metric	operati	diagno	2]
-Based	LOx-based		m	reactor,				monitor	onal	stics,	
Biosenso	mini-reactor		electrod	mesopor				ing	stability	food	
r with	and silver		e, lactate	ous silica					(93.8%	and	
Silver	amalgam		oxidase	powder,					after	bevera	
Amalga	electrode.		(LOx)	ampero					350	ge	
m				metric					measur	quality	
Electrod				monitori					ements,	control	
e				ng					96.9%		
									after 7		
									months		
); Easy		
									replace		
									ment of		
									mini-		
									reactor		
Dual-	Fluorescent/el	Gluco	DNA-	Fluoresc	23	μΜ	N/A	Fluores	Dual-	Glucos	[4
Mode	ectrochemical	se	templat	ence		ical),	14/11	cence	mode	e	3]
Biosenso	dual-mode	50	ed silver	detectio	29	μM		and	detectio	monito	ر
r Using	biosensor for		nanoclu	n,		ctroc		electroc	n, high	ring in	
DNA-	glucose		sters	electroch		nical)		hemical	sensitiv	body	
Templat	detection.		(AgNCs	emical	nen	псату		richiicai	ity,	fluids	
ed Silver	detection.		@DNA),						small	naids	
Nanoclu			glucose	pathway					size		
			oxidase						SIZE		
sters											
			(GOx)								
Core-	Development	Gluco	Ag@Au	Capillar	125	μΜ	967	Electroc	High	Flexibl	[4
Sheath	of Ag@Au	se	nanowir	y-force-			$\mu A{\cdot}mM$	hemical	optical	e	4]
Ag@Au	NWs for		es	induced			−1·cm−2		transmi	electro	
Nanowi	improved		(NWs)	welding					ttance	nics,	
res for	STEs with								(78.7%),	glucose	
Stretcha	high oxidation								high	biosens	
ble	resistance.								tensile	ors	
Transpa									strain		
rent									(240%),		
Electrod									excellen		
es									t		
									robustn		
									ess		

Green	Amperometric	Gluco	Green	CPE	N/A	N/A	Ampero	Low	Glucos	[4
Synthesi	glucose	se	synthesi	modified			metric	detectio	e	5]
s of	biosensor		zed	with				n limit,	detecti	
Silver	using green		waste	WT-				high	on in	
Nanopa	synthesized		tea-	AgNPs,				reprodu	food,	
rticles	WT-AgNPs.		based	enzyme				cibility,	environ	
for			silver	immobili				good	mental	
Glucose			nanopar	zation				selectivi	applica	
Biosenso			ticles					ty	tions	
rs			(WT-							
			AgNPs),							
			glucose							
			oxidase							
			enzyme							
HNC/A	Uses hemp-	Lacta	HNC,	Alkali	Not	Not	Cyclic	High	Wearab	[5
gNPs-	derived	te	AgNPs,	treatmen	specified	specifie	voltam	current	le	0]
PVA for	nanocellulose		PVA	t, acid		d	metry	respons	lactate	
Lactate	(HNC) with			hydrolys			(CV),	e at 10	sensor	
Detectio	silver			is, self-			Ampero	wt%		
n	nanoparticles			reductio			metry	HNC/A		
	(AgNPs) for			n				gNPs-		
	lactate							PVA,		
	detection.							Linear		
								range:		
								0–25		
								mM		
Ag-HPB	Hollow	Gluco	AgNPs,	Coating-	2.28	24.37	Ampero	High	Trichlo	[4
Nanoco	Prussian blue	se,	Prussia	etching	pg/mL	μΑ	metry	sensitiv	rfon	9]
mposite	with ultra-	Trichl	n blue	method	(trichlorf	mM-1	metry	ity and	monito	2]
s for	small AgNPs	orfon	(PB)	nieulou	on)	cm-2		enzyme	ring in	
	J	011011	(1 D)		OH)			-	_	
Biosensi	for glucose					(glucose		loading, Flexible	apples	
ng	and)				
	trichlorfon							biosensi		
	detection.							ng		
								platfor		
								m		

GPP-	Glossy Photo-	Gluco	Graphe	Screen-	198 nM	291.19	Ampero	Low-	Point-	[4
Based	Paper based	se	ne,	printing,		μΑ	metry	cost,	of-Care	6]
SPCIDE	sensor for		Polyanil	drop-		mM-1		eco-	glucose	
for	glucose		ine	casting		cm-2		friendly	testing	
Glucose	detection.		(PANI),					, Linear		
Sensing			GS-					range:		
			AgNPs,					198 nM		
			Glucose					to 30		
			oxidase					mM		
			(GOx)							
Aptame	Uses aptamer	Saxit	AgNPs,	Aptamer	1 nM	Not	Differen	High	STX	[4
r-Based	and	oxin	Aptame	immobili		specifie	tial	sensitiv	detecti	8]
Electroc	K3Fe(CN)6	(STX)	r,	zation,		d	pulse	ity,	on in	-
hemical	regulated		K3Fe(C	K3Fe(C			voltam	stability	seafood	
Sensor	AgNPs for		N)6	N)6			metry	, and		
for STX	STX detection.			etching			(DPV)	reprodu		
								cibility		
AgNPs/r	Electrochemic	Hydr	AgNPs,	Hydroth	3.19 μΑ	49 μΑ	Ampero	Simulta	Detecti	[4
GO			Reduce	ermal	(H2O2),	mM-1c	•	neous	on of	[4 7]
Nanoco	al sensors using	ogen perox	d	synthesi	0.18 μM	m-2	metry, CV	detectio	H2O2	7]
mposite	AgNPs/rGO	ide	graphen	synthesi s,	(DA)	(H2O2),	CV	n of	and	
s for	nanocomposit	(H2O	e oxide	electrode	(DA)	7.86 μA		H2O2	dopami	
H2O2	es.	2),	(rGO)	position		mM-1c		and	ne	
and DA	es.		(IGO)	position		m-2		DA,	He	
Detectio		Dopa mine				(DA)		Good		
		(DA)				(DA)		stability		
n		(DA)						Stavility		

3.5. Metal-Organic Frameworks

Electrochemical Detection with Aptamer/Enzyme Modification

In parallel with the development of biosensors, metal–organic frameworks (MOFs), such as MIL–53(Al), decorated with Au@Pt nanoparticles and enzymes have led to the development of an advanced detection system for 2019–nCoV–NPs. The combination of horseradish peroxidase (HRP) and G-quadruplex DNAzyme, along with the thiol-modified aptamers N48 and N61 immobilized on a gold electrode (GE), formed a biosensing platform in accordance with increasing demand for sensitive COVID-19 diagnosis. The principle of using Au@Pt/MIL-53(Al) nanocomposites, HRP, and hemin/G-quadruplex DNAzymes to cocatalyze hydroquinone oxidation in the presence of hydrogen peroxide has been recognized as a signal amplification strategy. This system enabled the detection of 2019-nCoV-NPs with a detection limit of 8.33 pg/mL and a linear range of 0.025 to 50 ng/mL, thereby establishing high potential for early COVID-19 diagnosis [53]. However, the complexity and cost of the system pose challenges for wider application. The next development was an electrochemical biosensor system based on magnetic metal–organic frameworks (MMOFs) synthesized via an in situ growth method, which employs aptamer–biotin and streptavidin–horseradish peroxidase for the

detection of spike proteins. Unlike the earlier MOF-based system, which involved a more complex setup, this electrochemical system uses MMOFs modified on a screen-printed electrode and a smaller sample size. The MMOF-based biosensors demonstrated a detection limit of 6 pM for voltammetry and 5.12 pM for impedance spectroscopy in human serum samples. This miniaturization reduces complexity and cost, simultaneously enabling a cost-effective and sensitive detection platform for the SARS-CoV-2 spike protein during point-of-care testing [54]. In parallel with opticle sensors, noninvasive glucose detection offers a pain-free, easy, and low-cost option that can improve home glucose testing for diabetic patients. The enzyme-encapsulated MOF nanomesh, equipped with a uniform shape, regular structure, and excellent catalytic performance with high electrical conductivity, realized the development of a sensitive and stable enzymatic biosensing platform, which demonstrated a sensitivity of 86.86 µA mm⁻¹ cm⁻² and a detection limit of 16.57 µm and remained stable for 16 days. Tests with real saliva samples showed greater than 93% accuracy compared with a commercial glucose assay kit, with a Pearson's r > 0.7 correlation with blood glucose. Noninvasive glucose biosensors have strong potential for improving home glucose testing for diabetic patients, thereby establishing a promising new method for diabetic care technology [55]. The next development involved enhancing the porosity of the nanomaterial for sensitivity issues by attaching porphyrin (H2TMP) to MOF-5/CoNi2S4, improving the detection of the recombinant SARS-CoV-2 spike antigen. Atomic force microscopy (AFM) revealed a surface roughness between 0.54 and 0.80 µm, indicating strong antigen interactions. The synthesized nanomaterials demonstrated a detection limit of 5 nM for the antigen, highlighting their high sensitivity and biocompatibility. Compared with porphyrin, these nanobiosensors offer low cost, safety, and bioactivity, making them promising for future COVID-19 detection platforms [56]. These examples highlight the versatility and effectiveness of using advanced metal-organic frameworks and nanomaterials in biosensors, emphasizing their potential to improve detection sensitivity, selectivity, and performance in various medical applications.

Electrochemical Detection with Metal/Carbon Nanocomposites

In parallel with carbon sensors, enzyme encapsulation methods equipped with ZIF-8, BC, and c-MWCNTs in combination with a self-powered EBFC platform led to the first bisphenol A (BPA) detection system, achieving a detection limit of 1.95×10^{-3} mM, which was developed in accordance with increasing demand for sensitive BPA detection [57]. In principle, a Cu MOF synthesized at room temperature with BTC as a ligand and copper nitrate trihydrate as a precursor has been recognized as a highly sensitive nonenzymatic glucose sensor, which was proposed by researchers in the field. The development of a Cu MOF deposited on a graphite sheet electrode sensing system made glucose detection available with a detection limit of 0.019 mM and sensitivity of 229.4 µA mM-1 cm², thereby establishing a stably growing and highly reproducible detection platform. The next development was the characterization of the Cu MOF structure using FTIR, SEM, EDX, and PXRD, confirming the electrocatalytic activity of the sensor for glucose oxidation [58]. Unlike enzyme-based BPA detection systems, which are equipped with relatively high power requirements, Cu MOF-based glucose sensors are mainly based on electrochemical principles and employ fast response times and excellent stability. In parallel with the development of glucose detection methods, measuring the levels of monoamine neurotransmitters (MNTs), such as dopamine (DA), adrenaline (Adr), norepinephrine (NE), and 5-hydroxytryptamine (5-HT), has become important for understanding MNT-related diseases such as Alzheimer's disease, Parkinson's disease, and depression. A novel electrochemical sensor equipped with a nanocomposite of multiwalled carbon nanotubes (MWCNTs) and an aminefunctionalized Zr(IV) metal-organic framework (UIO-66-NH2) was developed for detecting multiple MNTs, in accordance with increasing demand for monitoring neurotransmitter levels. The use of MWCNTs-UIO-66-NH2 for enhanced sensor performance, owing to its high surface area, low impedance, and excellent electrocatalytic activity, has been recognized as a significant advancement in MNT detection. The development of such a sensor enabled real-time monitoring of DA from PC12

and C6 cells, offering potential for diagnosing MNT-related disorders and establishing a promising platform in electrochemical sensor technology [59]. The next development could involve further integration of MOFs with other nanomaterials to expand the versatility and applicability of the sensor systems. Unlike earlier detection methods, which require complex procedures and bulky equipment, this new sensor system provides high sensitivity, a low detection limit, and real-time monitoring while reducing the cost of analysis and making the process more accessible for medical diagnostics.

Electrochemiluminescence (ECL) and Immunosensor Methods

In parallel with the development of glucose monitoring technologies, a novel sandwich paperbased electrochemiluminescence (ECL) biosensor for detecting HbA1c was introduced, utilizing an advanced nanocomposite as the tracing tag and a specialized immobilization platform for the sensing element. The biosensor was developed via screen-printed electrodes (SPEs) fabricated on a paper substrate, followed by the deposition of a thick gold layer and the electrochemical reduction of aminophenylboronic acid (APBA)-functionalized graphene oxide (GO) to form reduced graphene oxide (rGO)/APBA. This system demonstrated excellent performance by detecting HbA1c within a wide range (2%-18%) and a low detection limit of 0.072%, in accordance with the need for sensitive detection in complex biomarker applications [60]. However, this method involves complex electrode modifications, which are addressed in the subsequent approach. The development of a new sandwich-type electrochemical immunosensor based on Prussian blue (PB) as a signal indicator and functionalized metal-organic framework nanocomposites as signal amplifiers enabled the quantitative analysis of HE4, a crucial biomarker for ovarian cancer diagnosis. Unlike previous systems, this new immunosensor displayed a wide linear range (0.1-80 ng / mL) and a lower detection limit of 0.02 ng/ mL, thereby overcoming the limitations of earlier approaches and advancing the detection capabilities for more complex biomarkers, improving both sensitivity and practicality [61]. Error! Reference source not found.

Table 3. Metal-Organic Frameworks (MOFs) in Biosensors.

Title	Summary	Target Analyte	Materi al	Technique	Detect ion	Sens itivit	Measureme nt Method	Key Feature	Ap pli	Ref
		Analyte	aı		Limit	y	nt Wethou	s	cat	
					(LOD)	,			io	
									ns	
MOF-	Biosensor	2019-	MIL-	Thiol-	8.33	Not	Aptamer-	Wide	Ea	[53]
based	using MIL-	nCoV-	53(Al),	modified	pg/mL	speci	protein-	linear	rly	
Aptase	53(Al) with	NP	Au@Pt	aptamers,		fied	nanoprobe	range	C	
nsor	Au@Pt		nanop	Au@Pt/MIL-			sandwich	(0.025	О	
for	nanoparticle		article	53(Al), HRP,			electrochem	to 50	VI	
COVID	s and		s,	hemin/G-			ical	ng/mL),	D-	
-19	enzymes for		HRP,	quadruplex			detection	high	19	
Detecti	detecting		hemin	DNAzyme				sensitiv	dia	
on	2019-nCoV-		, G-					ity,	gn	
	NP.		quadr					selectivi	osi	
			uplex					ty,	s	
			DNAz					reliabili		
			yme					ty		

MOF-	Enhanced	SARS-	MOF-	Surface	5 nM	Not	Atomic force	High	SA	[56]
5/CoNi	sensitivity	CoV-2	5,	roughness		speci	microscopy,	biocom	RS	
2S4	for detecting	spike	CoNi2	measuremen		fied	MTT assay	patibilit	-	
Nanobi	SARS-CoV-	antigen	S4,	t, cell				y, low	Co	
osenso	2 spike		H2TM	viability				cytotoxi	V-	
r for	antigen		P	assay				city,	2	
SARS-	using MOF-							tunabili	det	
CoV-2	5/CoNi2S4							ty	ect	
Detecti	with								ion	
on	porphyrin.									
ZIF-8	Encapsulati	Bispheno	ZIF-8,	Enzymatic	1.95 x	Not	Biofuel cell-	High	BP	[57]
Encaps	on of laccase	l A (BPA)	BC, c-	biofuel cell	10-3	speci	driven	flexibili	A	
ulated	in ZIF-8		MWC		mM	fied	sensing	ty,	det	
Laccas	combined		NTs,				platform	excellen	ect	
e	with BC and		laccas					t	ion	
Electro	c-MWCNTs		e					mechan		
de for	for detecting							ical		
BPA	BPA.							propert		
Detecti								ies,		
on								conduct		
								ivity		
ECL	Paper-based	HbA1c	Zr-	Electrochem	0.07%	Not	ECL and	Wide	Hb	[72]
Biosen	ECL ECL	110/110	MOF,	iluminescen	0.07 70	speci	cyclic	respons	A1	[, -]
sor for	biosensor		Fe3O4	ce, cyclic		fied	voltammetr	e range	c	
HbA1c	using		, TMC,	voltammetry		iica	у	(2% to	det	
Detecti	nanocompo		AuNC	voltaininetry			measureme	18%),	ect	
on	site tracing		s,				nts	high	ion	
OH	_		APBA				1115	sensitiv	1011	
	tag for HbA1c									
	detection.		functi					ity		
	detection.		functi							
			onaliz							
			ed GO							

Cu	Non-	Glucose	Cu	Electrochem	0.019	229.4	Cyclic	Excepti	Gl	[58]
MOF-	enzymatic		MOF,	ical	mM	μAm	voltammetr	onal	uc	
Based	glucose		BTC,	detection,		M ⁻¹	у,	stability	ose	
Glucos	sensor using		Coppe	cyclic		cm ⁻²	chronoampe	, short	det	
e	Cu MOF for		r	voltammetry			rometry	respons	ect	
Biosen	electrochem		nitrate					e time,	ion	
sor	ical		trihyd					good		
	detection.		rate					repeata		
								bility		
Electro	A	HE4	PB,	Electrochem	0.02	N/A	Electrochem	PB as	Cli	[61]
chemic	sandwich-		TiMO	ical	ng/mL		ical	signal	nic	
al	type		F-	immunosens				indicato	al	
Immun	electrochem		KB@A	or				r,	ov	
osenso	ical		uNPs					TiMOF-	ari	
r for	immunosen							KB@Au	an	
HE4	sor for HE4							NPs for	ca	
	detection							signal	nc	
	using							amplifi	er	
	Prussian							cation	dia	
	blue (PB)								gn	
	and								osi	
	functionaliz								s	
	ed MOF									
	nanocompo									
	sites.									
Non-	An	Glucose	MOF	Enzymatic	16.57	86.86	Electrochem	High	Но	[55]
Invasiv	enzymatic	Glucose	nano	biosensor	μm	μΑ	ical	sensitiv	me	[55]
e	biosensing		mesh,	Dioscrisor	μπ	mm-	icai	ity and	-	
Glucos	platform		enzym			1		stability	ba	
e	using MOF		e			cm-2		Stability	se	
Detecti	nanomesh					CIII Z			d	
on	for sensitive		encaps ulatio						gl	
OII	and stable		n							
	glucose		11						uc ose	
	detection.								tes	
	actection.								tin	
									g	

ECL	An	microRN	Cu	ECL	0.5 fM	N/A	Electrochem	Cu	G	[60]
Biosen	electrochem	A	NCs,	biosensor			iluminescen	NCs/Zn	C	
sor for	iluminescen	(miRNA-	Zn				ce	MOF	pe	
GC	ce (ECL)	421)	MOF					nanosh	rit	
Detecti	biosensor		nanos					eet for	on	
on	for detecting		heet,					high	eal	
	microRNA		Au					quantu	me	
	in GC		NPs/					m yield	tas	
	extracellular		MXen					and	tas	
	vesicles.		e,					stability	is	
			phosp						dia	
			holipi						gn	
			d layer						osi	
									s	
MNT	A platform	Dopamin	MWC	Electrochem	Low	N/A	Electrochem	Synergi	Cli	[59]
Detecti	using	e (DA),	NTs,	ical sensor	detecti	14/21	ical	stic	nic	
on	MWCNTs	Adrenali	UIO-	icai scrisoi	on		icui	effect	al	
with	and UIO-66-	ne (Adr),	66-		limit			betwee	dia	
Electro	NH2	Norepine	NH2		for			n	gn	
chemic	nanocompo	phrine	11112		DA,			MWCN	osi	
al	site for	(NE), 5-			Adr,			Ts and	s	
Sensor	detecting	Hydroxy			NE, 5-			UIO-66-	of	
	multiple	tryptami			НТ			NH2	M	
	monoamine	ne (5-HT)							NT	
	neurotrans	,							_	
	mitters								rel	
	(MNTs).								ate	
	,								d	
									dis	
									or	
									de	
									rs	

MMOF	An MMOF-	SARS-	MMO	Electrochem	6 p	М	N/A	Electrochem	MMOF	Poi	[54]
-Based	based	CoV-2	F,	ical	(volta			ical	for easy	nt-	
Electro	aptasensor	spike	aptam	aptasensor	mmetr				washin	of-	
chemic	for detecting	proteins	er-		y), 5.	.12			g and	car	
al	SARS-CoV-		biotin,		pМ				depositi	e	
Aptase	2 spike		strepta		(imp	e			on, high	tes	
nsor	proteins		vidin-		danc	æ			sensitiv	tin	
for	using		HRP		spect	tr			ity	g	
SARS-	electrochem				oscoj	ру				for	
CoV-2	ical)					SA	
	methods.									RS	
										-	
										Co	
										V-	
										2	

3.6. Silicon and Silicon Dioxide

Electrochemical Etching and Functionalization

Porous silicon (PSi) biosensors equipped with anti-HSP70 antibodies immobilized with APTES and glutaraldehyde constitute the first selective detection system for HSP70, which was developed in accordance with increasing demand for the cost-effective detection of large biomolecules. This principle, which uses the large surface area and optical properties of PSi, has been recognized as a significant advancement in label-free biosensors and was proposed as described above. The development of a mesoporous PSi biosensor system has made label-free detection of large biomolecules available for researchers, thereby establishing a stably growing research and development sector. The next development was in biosensor optimization to reduce pore clogging while maintaining high sensitivity for large biomolecules such as HSP70, with the PSi sensor having a porosity of $68\% \pm 3\%$ and a thickness of 2400 ± 30 nm. Unlike other biosensing systems that face challenges with large-molecule detection, this system achieved a detection range of 3000-500000 ng/ mL and a detection limit of 1290 ± 160 ng / mL. Employing this porous structure and hand-held spectrometers, which provide results within several minutes, requires careful examination of the porosity and surface chemistry via SEM and FTIR techniques. This sensor simultaneously realized high selectivity for HSP70 detection over other proteins, such as IgG. The use of a mesoporous PSi sensor enables sensitive, label-free detection, establishing a promising platform for large biomolecule biosensing [62]. In addition to addressing pore-clogging issues in PSi, a conductive glucose sensor equipped with glucose oxidase immobilized on macroporous silicon has been studied. The macroporous silicon layer, which is fabricated on a p-type silicon substrate through an electrochemical etching process, along with two silver electrodes on the front side, serves as the sensor platform. The functionalization of the surface is carried out by the physisorption of glucose oxidase, which enables glucose detection. The current-voltage characteristics are recorded at different glucose concentrations, revealing an initial current increase up to 1 mM, followed by a decrease as the concentration increases. The sensor response is analyzed by employing conduction mechanisms such as hopping, Poole-Frenkel, trap-assisted, and Fowler-Nordheim tunneling, each prevailing at distinct applied field ranges. This sensor demonstrates excellent performance in terms of sensitivity, response, and repeatability, effectively resolving the issues of selectivity and functionalization [63]. In parallel with the development of early glucose sensors, high-sensitivity glucose sensors face challenges in real-time applications that demand a high degree of specificity. To address this, early diagnosis and prognosis based on the monitoring of breast cancer biomarkers have become increasingly critical. In this study, gold-coated silicon microneedle arrays (Au-Si-MNAs) were employed both as a biomarker extraction platform and as an electrochemical transducer, facilitating selective immunocapture and the quantification of the breast cancer biomarker ErbB2. The device demonstrated a linear response in artificial interstitial fluid within the range of 10-250 ng/mL, with a detection limit of 4.8 ng/mL, which is lower than the levels typically observed in breast cancer patients. As a proof of concept, the immunosensor was able to extract ErbB2 from a phantom gel designed to mimic skin layers, with a linear range from 50 to 250 ng/mL and a detection limit of 25 ng/mL. This unique platform, which integrates direct transdermal biomarker extraction and quantification, offers promising new directions for developing high-performance wearable point-ofcare devices [64]. These studies collectively underscore the potential of advanced fabrication techniques, including electrochemical etching, thermal oxidation, and microneedle arrays, to enhance sensor performance in various applications.

Lithography and Etching Techniques

In parallel with traditional hydrogen peroxide (H2O2) sensing technologies, a field-effect transistor (FET) based on reduced graphene oxide-polypyrrole (rGO/PPy) nanocomposites was developed, which uses lithography and etching techniques to fabricate a SiO2 trench-type structure designed for both holding and sensing H2O2. Morphological and structural analyses of the rGO/PPy

nanocomposites confirmed their bonding, enabling a high-performance sensor with a fast response time of 5 s, high stability, and sensitivity, with a detection limit of 10 pM at an appropriate signal–to–noise ratio (S/N). However, its high selectivity toward H2O2 restricts its applicability to other uses [65]. To address this limitation, researchers have explored 3-D integrated graphene–porous silicon (p-Si) plasmonic waveguide-based nanostructures for DNA hybridization, employing the full-vectorial finite element method. The p-Si waveguide, designed with the Maxwell–Garnett model, is sandwiched between two low-index silicon dioxide layers, incorporating a graphene layer to increase absorption, tunability, and sensitivity. This design achieves extraordinary optical transmission (EOT) through a subwavelength nanoaperture, reducing ohmic losses and improving optical transmission near the infrared region. Parametric analysis of this waveguide achieved a sensitivity of 318.5 nm/RIU, a figure of merit of 3.395/RIU, a quality factor of 17.36, and a detection accuracy of 0.01/nm. The combination of advanced fabrication techniques such as lithography and etching for H2O2 sensors, alongside sophisticated modeling and material layering in the plasmonic waveguide, highlights the critical role of innovative approaches in improving the performance and applicability of biosensors for lab-on-a-chip biological applications [66].

Microcontact and Layer-by-Layer Techniques

An electrochemical capacitance biosensor utilizing silicon nitride substrate a (Si3N4/SiO2/Si[P]/Al) was developed for the sensitive detection of tumor necrosis factor alpha (TNFα) cytokines. The sensor was fabricated via microcontact printing to immobilize the biological recognition element onto the sensor surface, and fluorescence microscopy confirmed successful biomolecule attachment. Additionally, contact angle measurements verified biofunctionalization. TNF- α detection was carried out via Mott–Schottky analysis across a concentration range of 1 pg/mL to 30 pg/mL. The biosensor exhibited high linearity and sensitivity, achieving responses of 4 mV·pM⁻¹ in PBS and 4.4 mV·pM⁻¹ in AS. The LOD values were 0.38 pg/mL in PBS and 1 pg/mL in AS. Selectivity tests revealed minimal interference from substances such as cortisol and interleukin-10 in artificial saliva [67]. A silicon-based, multilayer biosensor was reported for the selective detection of Sphingobium yanoikuyae in nonbeverage alcohols via the use of RNA aptamers as biomolecular recognition elements. The sensor was fabricated by immobilizing RNA aptamers on a silicon substrate and covalently attaching microbes to the silicon surface. This functionalization enabled specific detection of the target bacterium through iridescent color changes caused by the increased thickness of the nanolayers. The detection limit was verified as 2×10^6 CFU/mL via UV-Vis reflectance spectrophotometry, which was supported by analysis via atomic force microscopy and X-ray photoelectron spectroscopy. This approach offers a sensitive and selective method for bacterial detection, confirmed through reproducible iridescent responses [68].

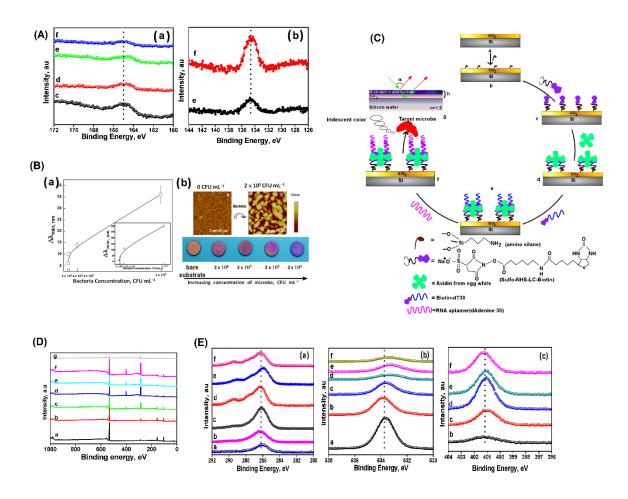


Figure 8. (A) High-resolution XPS spectra of N 1s and Si 2p for successive surface modifications: e- Biotin–dT30 and f- RNA aptamer layers.(B) (a) Binding isotherm showing Δλmin versus Sphingobium yanoikuyae concentration; (b) AFM images and corresponding optical chip color changes before and after microbial binding.(C) Schematic of multilayer surface functionalization: (a) oxidized SiO₂, (b) aminated, (c) biotinylated, (d) avidin-modified, (e) Biotin–dT30, (f) RNA aptamer layer, (g) bacterial binding, and (h) interference-based color generation.(D) XPS survey spectra for each functional layer: (a) SiO₂, (b) amine, (c) biotin, (d) avidin, (e) Biotin–dT30, (f) RNA aptamer, (g) bacteria-bound.(E) High-resolution XPS spectra for (a) C 1s, (b) O 1s, and (c) N 1s across layers (a) SiO₂, (b) amine, (c) biotin, (d) avidin, (e) Biotin–dT30, (f) RNA aptamer, (g) bacteria-bound.

Advanced Coating and Deposition Techniques

Compared with traditional sensors, a photonic crystal (PC)-based biosensor has been reported to demonstrate superior performance in therapeutic applications, making it suitable for use in clinical decision systems because of its enhanced sensitivity and resolution. The biosensor was constructed via the use of photonic crystals made of silicon dioxide (SiO2), gold (Au), and graphene oxide (GO) for protein analysis and immunological evaluation. The design included (i) gold plating to enhance reflectivity, (ii) GO-coated rods to increase plasmonic attraction, and (iii) improvements in biosensing by increasing the absorption of antibodies and antigens. This construction resulted in higher sensitivity and significant spectral changes for accurate detection. For this purpose, simulations based on the finite-difference time-domain (FDTD) method confirmed a spectral shift and transmission amplitude of 100 nm/RIU with a quality factor of 597, demonstrating the sensor's ability for protein analysis and immunological evaluation, which is crucial for clinical decision-making [69]. An FP-based optical fiber sensor has been reported for the detection of ultralow glucose concentrations in liquids. The optical fiber sensor was constructed using (i) phase shifted Bragg-grating lab-on-fiber (PSBG-LOF) technology, (ii) positioning a novel PSBG at the end of a single-mode fiber (SMF), (iii) deposition of 4.5 pairs of siliconoxynitrite (SiON)-doped silicon (Si) thin films via plasma-enhanced

chemical vapor deposition (PECVD), and (iv) inclusion of a silica layer between the two PSBGs. This is the first instance of applying SiON-doped Si in PSBG structures for glucose detection. The sensor demonstrated notable features, such as high sensitivity (14904 nm/RIU), a low detection limit (1.98 × 10-6 RIU), and resistance to temperature variations within the range of 25°C--45°C. These properties highlight its potential for in vivo biosensing applications, where temperature stability is crucial [70]. A porous silicon (PSi) Bragg mirror (Bm)-based grayscale variation method using fluorescence imaging was proposed for quick and easy detection of pesticides, and FP optical fiber sensors are known for their high sensitivity. In this approach, CdSe/ZnS quantum dots (QDs) were employed as fluorescence labels to reduce the fluorescence intensity, with a digital imaging method applied for detection. Specifically, acetamiprid was designed to bind to its specific aptamer, which led to the replacement of the complementary strand aptamers of the QDs, resulting in a weakened fluorescence intensity. The fluorescence image of the PSi Bm surface was captured using an imaging device, and the detection of the acetamiprid pesticide was achieved by calculating the variation in the average grey value of the image. The experimental results revealed that the average grey value variation increased as the pesticide concentration increased, indicating a linear relationship within a defined range. The detection limit for the acetamiprid pesticide using this method was found to be 2.8 nM, suggesting a low-cost, fast, and simple approach for pesticide detection [71]. Error! Reference source not found.

Table 4. Silicon and Silicon Dioxide-Based Biosensors.

Title	Summar y	Target Analyte	Material	Techniq ue	Detect ion Limit	Sensit ivity	Measur ement Method	Key Feature s	Applica tions	Ref
					(LOD)					
Porous	High	Heat	Porous	Electroch	1290 ±	N/A	Fiber	High	Label-	[62]
Silicon	sensitivit	Shock	Silicon	emical	160		Optic	selectivi	free	
Optical	y for	Protein	(PSi)	etching,	ng/mL		Spectro	ty, -10	optical	
Biosenso	detecting	70		thermal			meter,	dB wide	biosensi	
r	HSP70	(HSP70)		oxidation			SEM,	bandwi	ng	
				,			FTIR,	dth,		
				antibody			Contact	porosity		
				functiona			Angle	: 68% ±		
				lization			Measur	3%,		
				with			ement	thicknes		
				APTES				s: 2400 ±		
				and				30 nm		
				glutarald						
				ehyde						

Capacita	High	Tumor	Silicon	Micro-	0.38	4	Mott-	High	Early	[67]
nce	sensitivit	Necrosis	Nitride	contact	pg/mL	$mV{\cdot}p$	Schottk	linearity	detectio	
Electroch	y and	Factor	(Si3N4/Si	printing,	(PBS),	M-1	y	,	n of	
emical	selectivit	Alpha	O2/Si[P]/	fluoresce	1	(PBS),	analysis	selectivi	inflam	
Biosenso	y for	(TNF- α)	Al)	nce	pg/mL	4.4	,	ty	matory	
r for	TNF-α			microsco	(AS)	$mV{\cdot}p$	Fluoresc	against	respons	
TNF-α	cytokines			py,		M-1	ence	Cortisol	es	
Detection				contact		(AS)	Microsc	and		
				angle			opy,	Interleu		
				measure			Contact	kin-10		
				ment			Angle			
							Measur			
							ement			
RNA	Detection	Sphingob	Silicon	Layer-by-	2 × 106	N/A	Visual	Visual	Pathoge	[68]
Aptamer	of	ium	omeon	layer	CFU/	14/11	color	detectio	n	[00]
-Based	specific	yanoikuy		depositio	mL		change,	n,	detectio	
Color	bacteria	ae		n, RNA			UV-Vis	iridesce	n in	
Sensor	with			aptamers			reflecta	nt color	non-	
for	visual			uptumers			nce	changes	beverag	
Pathogen	color						spectro	, UV-Vis	e	
Detection	changes						photom	confirm	alcohols	
Detection	changes						etry,	ation	aiconois	
							AFM,	ation		
							XPS			
							AFS			
Photonic	Responsi	Proteins,	SiO2,	Gold	N/A	100	FDTD	High	Clinical	[69]
Crystal-	ve	antibodie	Gold	plating,		nm/RI	method	Quality	decisio	
Based	biosensor	S,	(Au),	graphene		U		factor:	n	
Biosenso	for	antigens	Graphen	oxide				597,	systems	
r for	protein		e Oxide	coating,				increase		
Protein	analysis		(GO)	FDTD				d		
Analysis	and			method				plasmo		
	immunol							nic		
	ogical							attractio		
	evaluatio							n		
	n									

Fabry-	Detecting	Glucose	Siliconox	Plasma	1.98 ×	14904	Lab-on-	Temper	In-vivo	[70]
Perot	ultralow		ynitrite	enhanced	10-6	nm/RI	Fiber	ature	biosensi	
Optical	glucose		(SiON)	chemical	RIU	U	(LOF)	insensiti	ng	
Fiber	concentr		doped	vapor				ve	applicat	
Sensor	ations		silicon	depositio				(25°C-	ions	
for	with high		(Si)	n				45°C),		
Glucose	sensitivit			(PECVD)				high		
Detection	y			, Phase				sensitivi		
				Shifted				ty		
				Bragg-						
				Grating						
				(PSBG)						
3-D	Studies a	DNA	Graphen	Full-	N/A	318.5	COMS	High	Lab-on-	[66]
Integrate	3-D		e, porous	vectorial		nm/RI	OL	sensitivi	a-chip	
d	integrate		silicon,	finite		U	multiph	ty,	biologic	
Graphen	d		silicon	element			ysics	tunabili	al	
e-Porous	graphene		dioxide	method,			softwar	ty,	applicat	
Silicon	-p-Si			Maxwell			e	reduced	ions	
Plasmoni	plasmoni			Garnett				ohmic		
c	c			model				losses		
Wavegui	wavegui									
de for	de-based									
DNA	nanostru									
Hybridiz	cture for									
ation	DNA									
	hybridiza									
	tion.									
High-	Demonst	Hydroge	Reduced	Lithogra	10:00	N/A	Electric	Fast	Liquid	[65]
Performa	rates a	n	graphene	phy,	PM		al signal	respons	sensing	
nce	high-	peroxide	oxide,	etching			measur	e, high	applicat	
Hydroge	performa	(H2O2)	polypyrr	techniqu			ement	stability	ions	
n	nce		ole, SiO2	es				, high		
Peroxide	H2O2							selectivi		
Sensing	sensing							ty		
FET	FET									
	using									
	rGO/PPy									
	nanocom									
	posites.									

Conducti	Studies	Glucose	Glucose	Electroch	N/A	N/A	Current	Good	Glucose	[63]
ve	glucose		oxidase,	emical			_	respons	sensing	
Glucose	sensor		macro	etching,			voltage	e, high		
Sensor	using		porous	physisor			characte	sensitivi		
Using	glucose		silicon,	ption			ristics	ty,		
Macro	oxidase		silver					excellen		
Porous	immobili		electrode					t		
Silicon	zed on		s					repetiti		
	macro							ve		
	porous							behavio		
	silicon.							r		
Gold-	Uses Au-	Epiderm	Gold,	Gold	4.8	N/A	Electroc	High	Wearabl	[64]
Coated	Si-MNA	al growth	silicon,	coating,	ng/mL		hemical	selectivi	e point-	
Silicon	for	factor	artificial	electroch			measur	ty,	of-care	
Micronee	biomarke	receptor	interstitia	emical			ement	simulta	devices	
dle	r	2 (ErbB2)	l fluid	transduc				neous		
Arrays	extractio			er				extracti		
for Breast	n and							on and		
Cancer	electroch							quantifi		
Biomarke	emical							cation		
r	transduct									
Detection	ion for									
	breast									
	cancer									
	diagnosis									
	•									
Fluoresce	Proposes	Acetami	CdSe/Zn	Digital	2.8 nM	N/A	Fluoresc	Low-	Rapid	[71]
nce	a	prid	S	imaging			ence	cost,	and	
Image-	grayscale	(pesticid	quantum	method			intensit	rapid,	simple	
Based	variation	e)	dots, PSi				y	simple	pesticid	
Pesticide	of		Bragg				measur	detectio	e	
Detection	fluoresce		mirrors,				ement	n	detectio	
Using PSi	nce		aptamers						n	
Bragg	image									
Mirrors	using PSi									
	Bragg									
	mirrors									
	for									
	pesticide									
	detection									

Conclusion

Electrochemical sensors have secured high sensitivity, short response times, and compact designs on the market to make portable diagnostic tools feasible. They are generally affordable and can detect many analytes, but interference and fouling can affect their performance so that they have to be maintained often. By comparison, optical sensors are highly specific and capable of detecting a variety of analytes without interference. However, these methods require sophisticated instruments and are also influenced by environmental factors. When these sensors are chosen, characteristics such as sensitivity, sample type, and methodological convenience are key parameters for evaluating them. Both sensor types have strong and weak points, with research trends toward improving sensitivity, selectivity, and miniaturization. The advancement of nanomaterials and molecularly imprinted polymers offers promise for better performance. Miniaturization is essential to make biosensors in the healthcare field portable, affordable and scalable. Innovations in these areas will extend the applications and improve the performance of biosensor systems in healthcare environmental monitoring.

Author Contributions: D.U.R.: Written original draft; conducted a survey of the literature; prepared the tables; collected the references; methodology; edited and proofread the final manuscript. B.K.Y.: Guidance on review writing; methodology; Supervision. All authors have read and agreed to the published version of the manuscript.

Funding: No funding was provided for the development of this manuscript.

Ethics Approval and Consent to Participate: Not applicable.

Consent for Publication: Not applicable.

Data Availability Statement: All data relevant to this review are included in the text, references, tables, and figures.

Acknowledgments: The authors would like to express sincere gratitude to the Shreenivas Deshpande Library at the Indian Institute of Technology (BHU) Varanasi for providing invaluable resources and support.

Conflicts of Interest: The authors declare that they have no competing interests

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