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Article

Effects of Lincomycin and Faecal Bacterial Stress on Intestinal Metabolite Composition of Weaned Tibetan Piglets

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Abstract: To investigate the effects of feeding the Lincomycin and faecal bacterial on weaning stress in Tibetan piglets, we divided the weaned Tibetan piglets at 30 days of age into four groups. We measured growth performance, blood physiological indexes, and metabolomics. The results showed that the Ant, Tib, and Fec groups significantly reduced the ratio of diarrhoea to feed/weight (F/G) and increased the average daily gain (ADG) compared to the Nor group ($p < 0.01$). The Nor group had significantly lower leukocyte counts, hemoglobin levels, and erythrocyte counts compared to the other three groups on 21d ($p < 0.05$). These physiological indexes tended to stabilize on 42d. We found that there were beneficial metabolites and metabolic pathways for gastrointestinal function. Specifically, the porphyrin metabolic pathway was elevated in the Ant group, and the tryptophan metabolic pathway was significantly elevated in the Tib and Fec groups compared to the Nor group ($p < 0.05$). In conclusion, adding faecal bacterial supernatant and fifteen-flavour black pills of Tibetan medicine to the feed reduced the rate of diarrhoea and improved the growth performance of piglets. Moreover, it had an effect on microorganisms and their metabolites and pathways in the gastrointestinal tract of the animals, which might be the main reason for influencing the diarrhoea rate of weaned Tibetan piglets and the growth and development of the piglets. This study provides a new approach for anti-stress applications in weaned Tibetan piglets and the development of substitute anti-products.

Keywords: Weaned piglets; metabonomics; Tibetan medicine; diarrhea; growth performance

1. Introduction

One of the most stressful events in the lifecycle of pigs is the weaning of piglets from the sow ^[1]. During the weaning process, piglets are exposed to multiple stressors, such as transportation, handling, transitioning from highly palatable milk to a less palatable dry feed, adjusting to a new environment, and establishing social hierarchy ^[2]. These stressors, along with the physiological and anatomical adaptations in the digestive tract, can lead to digestive dysfunctions and impaired immune systems, ultimately resulting in reduced growth and health in weaned nursery pigs ^[2]. In addition to optimizing the piglet transfer process, implementing scientific weaning techniques, reducing stress, and paying attention to temperature, humidity control, and strict disinfection of the barn after weaning ^[3], the addition of antibiotics to the feed is a common method to prevent and control diarrhea in piglets. However, this practice can lead to the development of antibiotic-resistant bacteria, which can be transferred from animals to humans, posing a threat to human health. Consequently, the development of efficient antibiotic replacement products has become the focus of research.

Fecal microbiota transplantation (FMT) is a technique in which the fecal microbiota from a healthy individual is transplanted into the gut of a recipient individual [4,5]. FMT has garnered attention as an important therapy for intestinal diseases, including inflammatory bowel disease and irritable bowel syndrome [6-8]. This technique has the ability to restore the intestinal microbiota, making it a promising approach for researchers [9-11]. Additionally, natural herbal medicines have also gained attention due to their low side effects, multi-targeting, and multi-pathway advantages [12]. The Fifteen Black Herbal Pills of Tibetan Medicine, a proprietary Chinese medicine preparation, is composed of fifteen pure natural mineral and botanical medicines. These pills are commonly used to treat chronic gastroenteritis, indigestion, loss of appetite, vomiting, and diarrhea [12]. The metabolites produced by different feed additives significantly impact the structure of the intestinal flora and play a crucial role in stress-induced diarrhea in weaned piglets [13]. The interaction between the metabolism of the intestinal flora and the nutrient metabolism of the host contributes to the maintenance of the intestinal tract and the overall health of the host [13]. Metabolites provide detailed information about the biological endpoints as downstream components of gene regulatory networks and protein interaction networks [14].

Currently, several studies have demonstrated the regulatory effects of probiotics, microecological agents, antimicrobial peptides on intestinal flora. These effects include inhibiting the growth of harmful microorganisms, promoting feed intake and growth of pigs, and improving immune function [15,16]. Such characteristics make them promising alternatives to antibiotics. Fecal transplantation has the ability to reshape the composition of the intestinal flora. In this regard, studies have shown that transplanting fecal microbiota from healthy donors to recipients with intestinal flora disorders can stabilize the intestinal microbiota and improve intestinal barrier function [17-19]. Research has also shown that different pig breeds can transfer their intestinal characteristics to germ-free mice through the transplantation of pig fecal fluids [20]. Moreover, recent findings indicate that the Fifteen-flavour Black Pill of Tibetan Medicine has the potential to protect the gastric mucosa, improve gastric muscle movement, promote gastric emptying, and accelerate small intestine propulsion. Apart from these mechanisms, an increase in blood flow to the gastric mucosa of gastritis patients has been observed, along with improved microcirculation and enhanced absorption of proliferative lesions. However, the impact of the Fifteen-flavour Black Pill of Tibetan Medicine on the intestinal flora remains unexplored. Faecal metabolites, which are end products of cellular and intestinal flora metabolism, offer a comprehensive reflection of the intestinal flora as well as the absorption and digestion of nutrients in the digestive tract. They not only elaborate on the composition of the intestinal microbiota, but also serve as biomarkers to study the relationship between intestinal microbial metabolism and host phenotype [21]. Post-weaning presents a major challenge for pigs as they are exposed to pathogenic bacteria and experience nutritional, environmental, and immunological stresses [22]. Enhancing the health condition of piglets is not only cost-effective but also advantageous for sustainable pork production. Therefore, we propose the utilization of metabolomics technology to analyze the effects of feeding four experimental groups of weaned Tibetan piglets without antibiotics and with the addition of fecal bacterial supernatant, the Fifteen-flavour Black Pill of Tibetan Medicine, and lincomycin, respectively.

This analysis aims to investigate the impact of feeding fecal bacterial supernatant and the Fifteen-flavour Black Pill of Tibetan Medicine on the growth performance, blood physiological indexes, and metabolites of weaned Tibetan piglets. Ultimately, this study aims to contribute to the comprehensive understanding and practical implementation of feed additives in livestock production. Consequently, it intends to provide a theoretical basis for the utilization of feed additives in animal husbandry.

2. Materials and Methods

2.1. Animals, design and diets

Twenty-eight weaned Tibetan pigs, aged 30 days with an initial body weight (BW) of 5.4 ± 0.2 kg, were allocated into four dietary treatments using a randomized complete block design. The trial

consisted of four treatment groups: the control group (Nor1/2 group), the lincomycin group (Ant1/2 group), the faecal bacterial supernatant group (Fec1/2 group), and the Tibetan medicine fifteen-flavour black pill group (Tib1/2 group). The Fifteen-flavour Black Pill of Tibetan medicine consists of 15 kinds of natural minerals and plant medicine, such as cold water stone, salt (fried), rhododendron, nutmeg, Tibetan mu tong, coriander fruit, sand, bright salt, purple, sand, wood incense, piper, black pepper and dried ginger. In the control group (Nor), piglets were fed a basal diet, while the Fec group, the Tib group, and the Ant group were fed the same basal diet but supplemented with 2000 mL/kg of faecal bacterial supernatant, 1000 mg/kg of Tibetan medicine fifteen-flavour black pill, and 1000 mg/kg of lincomycin hydrochloride (Schedule 1).

Schedule 1. Feed formulation and nutrient composition of diets.

| Items | Proportion/% |
|------------------------|--------------|
| Corn | 45.00 |
| Wheat bran | 35.00 |
| Soybean mea | 16.00 |
| Additive premixes 2624 | 4.00 |
| DE (Mcal/kg) | 2.8487 |
| CP (%) | 15.89 |
| CF(%) | 3.94 |
| Ca (%) | 0.70 |
| TP (%) | 0.66 |
| NPP (%) | 0.30 |
| Lys (%) | 0.79 |
| Met+Cys(%) | 0.53 |
| Thr (%) | 0.58 |
| Trp (%) | 0.21 |

Note: (1) The premix provides per kg of concentrate supplement: vitamin A 10 000 IU, vitamin D32 550 IU, vitamin E 20 IU, biotin 0.06 mg, copper sulphate 22 mg, iron sulphate 94 mg, manganese sulphate 80 mg, zinc sulphate 88 mg, potassium iodide 0.75 mg, sodium selenite 0.50 mg, calcium 0.35 %, phosphorus 0.125 %, and sodium chloride 0.80 %. Sodium chloride 0.80 %. (2) The digestible energy in the table is the calculated value, and the other nutrients are measured values.

For the nutrient compositions and experimental diets, refer to Table 1. All experimental diets were formulated to meet or exceed the nutrient requirements recommended by NRC [23]. The pigs were fed these diets for a duration of 42d, which were divided into two phases: phase 1 (0d to 21d) and phase 2 (21d to 42d).

Table 1. Effect of different feed additions on growth performance of weaned piglets 0-42d after the start of the trial (n=7).

| Groups | Initial weight (kg) | Diarrhoea rate in piglets (%) | Average weight (kg) | daily gain (ADG, kg) | Average feed intake (ADFI, kg) | daily feed-to-weight ratio (F/G) | finaly weight (kg) |
|--------|---------------------|-------------------------------|-------------------------|----------------------|--------------------------------|----------------------------------|--------------------|
| | | | kg) | (%) | (|) | kg) |
| | | | kg) | kg) | kg) | kg) | kg) |
| Nor | 5.42±0.06 | 10.5%±0.01 ^A | 0.037±0.00 ^C | 0.414±0.25 | 11.19±0.11 ^A | 6.97±0.47 ^A | |
| Ant | 5.43±0.07 | 7.8%±0.00 ^B | 0.083±0.00 ^A | 0.451±0.00 | 5.43±0.36 ^B | 8.93±0.09 ^B | |
| Tib | 5.40±0.03 | 8.2%±0.00 ^B | 0.089±0.00 ^A | 0.449±0.03 | 5.04±0.04 ^B | 9.13±0.35 ^B | |
| Fec | 5.48±0.09 | 8.2%±0.00 ^B | 0.06±0.00 ^B | 0.413±0.02 | 6.88±0.18 ^C | 8.00±0.10 ^C | |

Note: Data in the same column labelled with no letter or with the same letter on the shoulder indicate a non-significant difference ($P > 0.05$), a different lowercase letter indicates a significant difference ($P > 0.05$), and a different uppercase letter indicates a highly significant difference ($P < 0.01$).

2.2. Growth Performance

Individual body weight (BW) was recorded at the beginning and end of each phase (0d and 42d) to track average daily gain (ADG), average daily feed intake (ADFI), and feed-to-weight ratio (F/G) throughout the experiment.

2.3. Sample collection and processing

Fresh fecal samples were collected using rectal massage techniques from the pigs on 0d, 21d, and 42d of the experiment. The collected fecal samples were freeze-dried, ground (60 Hz, 30 s), and 50 mg of the sample was weighed into a centrifuge tube. Then, 700 μ L of the extraction solution (methanol-water, 3:1 by volume, pre-cooled at -40 °C, containing the internal standard) was added. The mixture was vortexed for 30 s, homogenized for 4 min at 35 Hz, sonicated in an ice-water bath for 5 min, homogenized and sonicated three more times. After that, the samples were repeated in the homogenizer at 4 °C overnight. Subsequently, the samples were centrifuged at 4 °C, 12000 rpm (centrifugal force 13800 \times g, radius 8.6 cm) for 15 min, and the supernatant was carefully filtered through a 0.22 μ m microporous membrane. The supernatant was then diluted 5-fold with the extraction solution and vortexed for 30 s. A QC sample was prepared by taking 40 μ L of each sample and stored at -80 °C until analysis.

Blood samples were collected from the anterior vena cava of the piglets while in a fasting state on 0d, 21d, and 42d of the study period. The blood samples were added to an EDTA vacuum tube (purple top) for subsequent analysis. Protein, glucose, white blood cell (WBC), red blood cell (RBC), and lymphocyte concentrations in the whole blood samples were analyzed using an automatic blood analyzer (BC-2600Vet Veterinary Fully-Automated Blood Cell Analyzer, Wuxi Xuanye Science and Technology Development Co).

2.4. LC-MS analysis

In this experiment, the target compounds were separated through chromatography using a Waters UPLC liquid chromatography column and an EXIONLC system (SCIEX) ultra-high performance liquid chromatograph (UPLC). The mass spectrometry data acquisition and quantitative analysis of target compounds were performed using SCIEX Analyst WorkStation Software (Version 1.6.3). The raw mass spectra were converted to TXT format using MSconverter software. Peak extraction and annotation were conducted using a self-developed R package and a self-constructed database. This was followed by performing orthogonal partial least squares discriminant analysis (OPLS-DA). The model validity was evaluated using the R^2Y and Q^2 parameters. Differential metabolites were identified based on the variable importance in the projection (VIP) of the OPLS-DA analysis between the two groups. A VIP value greater than 1 was used as a screening criterion, combined with the P value of the univariate analysis. Two ionization modes, positive ion mode (POS) and negative ion mode (NEG), were employed for metabolite detection in this experiment to improve metabolite coverage and detection effectiveness. The subsequent metabolite data analysis will include OPLS-DA of both the positive and negative ion mode data.

2.5. Mass spectrometer parameters

LC-MS/MS analysis was conducted using the SCIEX 6500 QTRAP+ triple quadrupole mass spectrometer, which was equipped with an IonDriveTurbo VESI ion source. The experiment was carried out in the multiple reaction monitoring (MRM) mode. The ion source parameters were set as: IonSpray Voltage: +5500/-4500 V, Curtain Gas Pressure: 35 psi, Temperature: 400 °C, Ion Source Gas 1 Pressure: 60 psi, Ion Source Gas 2 Pressure: 60 psi, and DP: \pm 100 V. The identical ion source parameters were utilized for the study.

2.6. Preprocessing and quality control of raw sequencing data

The constructed sequencing libraries were sequenced on the Illumina NovaSeq high-throughput sequencing platform using the whole-genome birdshot method. In order to assess the stability of the analytical system and identify variables that exhibited considerable variation during the analysis process, all samples were analyzed as Quality Control (QC) samples. A QC sample was included every 8-10 samples during instrumental analysis.

2.7. Statistical analysis

We performed Duncan's multiple comparisons using SPSS 18.0 software. Growth data and blood physiological indexes of weaned Tibetan piglets were analyzed using one-way ANOVA to determine differences between treatment groups. Differences with $p < 0.05$ were considered significant, differences with $p < 0.01$ were considered highly significant, and differences with $p > 0.05$ were considered not significant.

3. Results

3.1. Growth performance

Effects on growth performance parameters from 0d to 42d are presented in Table 1. The incidence of diarrhea was significantly lower ($P < 0.01$) in the Ant, Tib, and Fec groups compared to the Nor group. When comparing the average daily gain (ADG) among the four groups, the Ant, Tib, and Fec groups had significantly higher ADG values ($P < 0.01$) than the Nor group. Additionally, the Ant and Tib groups showed significantly higher ADG values ($P < 0.05$) than the Fec group. However, there was no significant difference ($P > 0.05$) observed in the comparison of average daily feed intake (ADFI) among the four groups. Comparing the feed conversion ratio (F/G) among the four groups, the Nor group had the highest value and it was significantly higher ($P < 0.01$) than the three other groups (Ant, Tib, and Fec). Furthermore, the Fec group had significantly higher F/G values ($P < 0.01$)

than the Ant and Tib groups, with no significant difference between the Ant and Tib groups ($P > 0.05$).

3.2. Comparison of blood physiological indicators

Comparison of blood physiological indices examined in this experiment under the same rearing conditions and at the same time of the experiment on 21d (Table 2), the leukocyte count in the Tib1 and Fec1 groups was significantly higher than that in the Nor1 group ($P < 0.01$). Hemoglobin (HGB) levels were the lowest in the Nor1 group and significantly higher in the Ant1, Tib1, and Fec1 groups compared to the Nor1 group ($P < 0.05$). The analysis of blood physiological indices in weaned piglets on 42d of the experiment yielded no significant differences in any of the blood physiological indices ($P > 0.05$), including platelet count, which did not reach statistical significance.

Table 2. Haematopoietic index data on 21d after the start of the trial (n=7).

| Norm | Unit | Nor1 | Ant1 | Tib1 | Fec1 |
|------|---------------------|--------------------------|--------------------------|--------------------------|--------------------------|
| WBC | 10 ⁹ /L | 19.67±0.43 ^a | 28.03±0.57 ^b | 27.83±0.22 ^b | 29.23±0.93 ^b |
| RBC | 10 ¹² /L | 7.96±0.20 ^a | 8.66±0.18 ^b | 8.12±0.31 ^b | 8.65±0.20 ^b |
| HGB | g/L | 133.00±0.58 ^a | 148.67±1.45 ^b | 147.33±8.09 ^b | 145.33±0.88 ^b |
| HCT | % | 42.90±0.46 | 44.80±1.16 | 42.90±1.30 | 44.67±0.20 |
| MCV | fL | 54.73±0.73 | 54.77±0.24 | 55.43±0.07 | 53.77±0.20 |
| MCH | Pg | 17.56±0.30 | 17.57±0.07 | 17.77±0.09 | 17.40±0.06 |
| MCHC | g/L | 322.08±1.92 | 322.67±1.76 | 322.00±2.52 | 321.67±1.73 |
| RDW | % | 16.23±0.46 | 16.83±1.30 | 16.10±0.10 | 17.30±0.50 |
| MPV | fL | 7.93±0.27 | 7.40±0.35 | 7.70±0.06 | 7.47±0.23 |
| PDW | % | 16.63±0.15 | 16.63±0.13 | 16.50±0.15 | 16.50±0.10 |
| PCT | % | 0.32±0.04 | 0.25±0.01 | 0.30±0.01 | 0.30±0.05 |

Note: Peer data labelled with no letter or with the same letter on the shoulder indicate a non-significant difference ($p > 0.05$), while different lower case letters indicate a significant difference ($p < 0.05$).

3.3. Quality control

When conducting metabolomics research using mass spectrometry, quality control (QC) samples are typically utilized to ensure the reliability and high quality of the obtained metabolomics data. Although the QC samples are theoretically identical, there are usually systematic errors introduced during the processes of sample extraction, detection, and analysis. These errors lead to discrepancies between the QC samples, with smaller differences indicating higher method stability

and better data quality. The visualization of the PCA analysis graph for this experiment reveals a dense distribution of the QC samples, indicating the reliability of the data (Figure 1AB). The starting group of the pre-test is referred to as the STA group.

3.4. Multivariate statistical analysis

R^2X , R^2Y , and Q^2 are commonly used metrics for this purpose. In general, R^2Y and Q^2 values greater than 0.5 indicate the reliability of the model. As presented in Table 3, the OPLS-DA model established in this experiment exhibits R^2Y and Q^2 values exceeding 0.5, thus signifying the stability, reliability, and statistical significance of the model.

Table 3. OPLS-DA model.

| Group | $R^2X(\text{cum})$ | $R^2Y(\text{cum})$ | $Q^2(\text{cum})$ |
|------------|--------------------|--------------------|-------------------|
| Nor1vsAnt1 | 0.669 | 0.994 | 0.955 |
| Nor1vsTib1 | 0.722 | 0.995 | 0.964 |
| Nor1vsFec1 | 0.482 | 0.983 | 0.871 |
| Ant1vsTib1 | 0.601 | 0.978 | 0.767 |
| Ant1vsFec1 | 0.597 | 0.998 | 0.955 |
| Tib1vsFec1 | 0.587 | 0.996 | 0.910 |
| Nor2vsAnt2 | 0.572 | 0.967 | 0.569 |
| Nor2vsTib2 | 0.578 | 0.976 | 0.518 |
| Nor2vsFec2 | 0.533 | 0.981 | 0.549 |
| Ant2vsTib2 | 0.668 | 0.979 | 0.700 |
| Ant2vsFec2 | 0.721 | 0.990 | 0.799 |
| Tib2vsFec2 | 0.714 | 0.902 | 0.744 |

3.5. Screening for differential metabolites

Differential metabolites were identified based on three parameters: VIP, FC, and P-value. To be considered differential metabolites, the VIP value had to be greater than 1 and the P-value had to be less than 0.01. Furthermore, specific differential metabolites were selected based on the principle of P-value less than 0.01, VIP value greater than 1, and FC value greater than 1 or less than 1. These criteria were determined using the VIP values obtained from statistical analyses and OPLS-DA.

At the 21d of the experiment for the comparisons “Nor1vs.Ant1”, “Nor1vs.Tib1”, and “Nor1vs.Fec1”, a total of 41, 46, and 22 differential metabolites were identified, with 15, 19, and 9

being up-regulated and 26, 27, and 13 being down-regulated (Table 4-1, Table 4-2, and Table 4-3). At the 42d of the trial, in comparison to the control groups “Nor2 vs. Ant2”, “Nor2 vs. Tib2”, and “Nor2 vs. Fec2”, there were respectively 9, 5, and 17 up-regulated metabolites, and 5 and 12 down-regulated metabolites (Table 4-4, Table 4-5, and Table 4-6). Specifically, 2, 5, and 12 metabolites were up-regulated, while 7, 0, and 5 were down-regulated.

All samples and relevant data underwent distance matrix calculation, and hierarchical clustering was performed to generate a dendrogram illustrating the similarity between samples. The sample dendograms revealed significant metabolic differences between different feed additives in both positive and negative ion modes (Figure 1CD).

Table 4. -1 Differential metabolites of Nor1vs. Ant1.

| Metabolites | VIP | P value | FC | Trend |
|----------------------------------|---|---------|-------|-------|
| Stearamide | 2.22 | 0.009 | 7.76 | up |
| Choline | 3.11 | 0.001 | 7.18 | up |
| 1-Stearoylglycerol | 1.12 | 0.005 | 6.94 | up |
| trans-Petroselinic Acid | 3.19 | 0.008 | 5.51 | up |
| L-(-)-Malic acid | 1.76 | 0.007 | 3.89 | up |
| 4-Nitrophenyl glucopyranoside | 2-acetamido-2-deoxy- α -D- 5.89 | 0.002 | 2.44 | up |
| 4-Hydroxybenzoic acid | 5.36 | 0.001 | 2.24 | up |
| Dihydrozeatin | 3.52 | 0.002 | 1.90 | up |
| Panaxtriol | 1.34 | 0.009 | 1.60 | up |
| Stearic Acid | 1.72 | 0.004 | 1.36 | up |
| 2,8-Quinolinediol | 1.89 | 0.008 | 15.31 | down |
| trans-3-Hexenoic acid | 1.40 | 0.009 | 10.96 | down |
| Methionine | 1.61 | 0.004 | 10.36 | down |
| Lithocholic Acid | 1.54 | 0.000 | 8.28 | down |
| 3-Hydroxy-3-methylbutanoic acid | 4.68 | 0.000 | 6.86 | down |

| | | | | |
|-----------------------------|------|-------|------|------|
| N-Acetyl-D-alloisoleucine | 3.95 | 0.003 | 4.10 | down |
| N-Acetyl-L-leucine | 3.75 | 0.000 | 3.50 | down |
| 4-Hydroxybutyric acid (GHB) | 2.33 | 0.007 | 3.35 | down |
| Isoquinoline | 2.30 | 0.008 | 2.90 | down |
| Indole-3-acetic acid | 2.75 | 0.008 | 2.78 | down |

Table 4. -2 Differential metabolites of Nor1 vs. Tib1.

| Metabolites | VIP | P value | FC | Trend |
|--|------|---------|-------|-------|
| Bullatine A | 8.14 | 0.000 | 1.39 | up |
| Prostaglandin E2-1-glyceryl ester | 6.19 | 0.009 | 6.80 | up |
| 4-Nitrophenyl 2-acetamido-2-deoxy- α -D-glucopyranoside | 5.68 | 0.005 | 2.30 | up |
| (+)-12(13)-DiHOME | 3.51 | 0.007 | 4.29 | up |
| 19(R)-hydroxy Prostaglandin A2 | 2.83 | 0.000 | 1.14 | up |
| PC (20:3e/22:5) | 2.82 | 0.000 | 1.11 | up |
| Choline | 2.68 | 0.006 | 6.13 | up |
| 3-Methylglutaric acid | 2.43 | 0.000 | 1.95 | up |
| N1-(2-piperidinophenyl)-3,4,5-trimethoxybenzamide | 2.24 | 0.001 | 1.09 | up |
| 2-Oxindole | 1.83 | 0.009 | 5.53 | up |
| 4-Hydroxybenzaldehyde | 1.22 | 0.004 | 4.38 | down |
| L-Phenylalanine | 1.24 | 0.008 | 12.15 | down |

| | | | | |
|---|------|-------|-------|------|
| Methionine | 1.31 | 0.002 | 10.13 | down |
| Norleucine | 1.82 | 0.003 | 1.04 | down |
| DL-Tryptophan | 1.86 | 0.007 | 8.47 | down |
| Indole-3-acrylic acid | 1.87 | 0.007 | 8.61 | down |
| N1-[4-hydroxy-6-(methoxymethyl)pyrimidin-2-yl]acetamide | 1.88 | 0.003 | 2.09 | down |
| N-Isobutyrylglycine | 2.01 | 0.002 | 1.46 | down |
| 5-amino-1-phenyl-1H-pyrazole-4-carbohydrazide | 2.06 | 0.009 | 1.80 | down |
| (2-oxo-2,3-dihydro-1H-indol-3-yl)acetic acid | 2.11 | 0.000 | 1.39 | down |

Table 4. -3 Differential metabolites of Nor1 vs. Fec1.

| Metabolites | VIP | P value | FC | Trend |
|---------------------------|------|---------|------|-------|
| FAHFA (22:4/3:0) | 2.94 | 0.005 | 4.34 | up |
| gamma-Glutamylleucine | 1.14 | 0.001 | 2.82 | up |
| N-Acetylhistamine | 1.53 | 0.001 | 2.70 | up |
| 2-Methylnicotinamide | 1.55 | 0.001 | 2.70 | up |
| (+/-)12(13)-DiHOME | 1.51 | 0.005 | 2.64 | up |
| Lactitol | 4.87 | 0.000 | 1.28 | up |
| MGDG (19:2/18:4) | 1.25 | 0.003 | 1.17 | up |
| L-Argininosuccinate | 1.86 | 0.000 | 1.16 | up |
| 3-Hydroxyanthranilic acid | 2.89 | 0.007 | 1.08 | up |

| | | | | |
|--|------|-------|-------|------|
| 2,8-Quinolinediol | 5.42 | 0.000 | 26.55 | down |
| L-Tyrosine | 2.03 | 0.005 | 14.97 | down |
| N-Benzylformamide | 2.12 | 0.002 | 11.26 | down |
| 4-Hydroxybenzaldehyde | 2.52 | 0.000 | 7.87 | down |
| 5-Hydroxyindole-3-acetic acid | 2.30 | 0.009 | 5.10 | down |
| 5-Hydroxyindole | 4.39 | 0.001 | 4.39 | down |
| Dopamine | 3.50 | 0.000 | 2.55 | down |
| (2-oxo-2,3-dihydro-1H-indol-3-yl)acetic acid | 2.08 | 0.000 | 1.99 | down |
| N1-(2,3-dihydro-1,4-benzodioxin-6-yl)acetamide | 2.40 | 0.003 | 1.94 | down |
| Glucuronic acid-3,6-lactone | 2.20 | 0.003 | 1.85 | down |

Table 4. -4 Differential metabolites of Nor2 vs. Ant2.

| Metabolites | VIP | P value | FC | Trend |
|---------------------------|------|---------|-------|-------|
| ERH | 5.95 | 0.001 | 1.28 | up |
| 3,3-Dimethylglutaric acid | 1.11 | 0.000 | 1.72 | up |
| Phenethyl isothiocyanate | 0.71 | 0.005 | 1.08 | down |
| Ethylmalonic acid | 1.03 | 0.009 | 2.23 | down |
| D-(-)-Lyxose | 1.19 | 0.001 | 1.70 | down |
| N6-Acetyl-L-lysine | 1.78 | 0.000 | 5.17 | down |
| (+/-)12(13)-DiHOME | 2.00 | 0.008 | 13.11 | down |
| 11-Epiprostaglandin E1 | 2.04 | 0.001 | 5.08 | down |
| Dehydrodiisoeugenol | 2.89 | 0.004 | 1.03 | down |

Table 4. -5 Differential metabolites of Nor2 vs. Tib2.

| Metabolites | VIP | P value | P | | Trend |
|---|------|---------|------|--|-------|
| | | | FC | | |
| Bullatine A | 8.00 | 0.000 | 2.72 | | up |
| Piperine | 5.44 | 0.001 | 1.07 | | up |
| D-Ribulose 1,5-bisphosphate | 2.91 | 0.006 | 2.23 | | up |
| 3-(3-bromo-4-hydroxy-5-methoxyphenyl)-2-cyanoacrylamide | 2.78 | 0.007 | 3.70 | | up |
| Cyanidin | 1.51 | 0.005 | 1.08 | | up |

Table 4. -6 Differential metabolites of Nor2 vs. Fec2.

| Metabolites | VIP | P value | FC | Trend |
|--|------|---------|-------|-------|
| N-(3-chloro-2-methylphenyl)-N'-(3-methoxypropyl)thiourea | 6.17 | 0.000 | 1.17 | up |
| Lactitol | 5.32 | 0.002 | 1.88 | up |
| 3-Iodo-L-tyrosine | 4.47 | 0.042 | 2.14 | up |
| L-Threonic acid-1,4-lactone | 3.36 | 0.048 | 3.79 | up |
| D-Ribulose 1,5-bisphosphate | 3.28 | 0.005 | 1.90 | up |
| (+/-)-CP 47,497-C7-Hydroxy metabolite | 3.23 | 0.044 | 2.77 | up |
| 3-(3-bromo-4-hydroxy-5-methoxyphenyl)-2-cyanoacrylamide | 2.74 | 0.030 | 2.51 | up |
| Cyanidin | 2.08 | 0.001 | 1.09 | up |
| Acetophenone | 1.15 | 0.034 | 23.03 | up |

| | | | | |
|--|------|-------|------|------|
| FAHFA (8:0/18:3) | 1.14 | 0.043 | 1.41 | up |
| 5(S),15(S)-DiHETE | 1.23 | 0.039 | 1.59 | down |
| 3-(propan-2-yl)-octahydropyrrolo [1,2-a]pyrazine-1,4-dione | 1.41 | 0.004 | 1.18 | down |
| Ribitol | 1.57 | 0.000 | 1.05 | down |
| Uric acid | 3.70 | 0.047 | 5.29 | down |

3.6. KEGG analysis of differential metabolites

Pathway analysis bubble plots were generated to illustrate the metabolic pathways involved in different feed additions for the Nor and experimental groups. At the 21d, the Ant1 group screened a total of eight metabolic pathways, including Bile secretion, Biosynthesis of alkaloids derived from the shikimate pathway, Taste transduction, Synaptic vesicle cycle, Secondary bile acid biosynthesis, Gap junctions (Figure 1F). Comparatively, the Tib1 group screened a total of 11 metabolic pathways compared to the Nor1 group (Figure 1G), which included Purine metabolism, Tryptophan metabolism, Chromogranulin metabolism, and ABC transporters. The Fec1 group screened 10 metabolic pathways (Figure 1E), including Tryptophan metabolism, Melanogenesis, Prolactin signaling pathway, Tyrosine signaling pathway, and Gap junction pathway. At the end of the experiment, the Ant2 group exhibited 10 differential metabolic pathways compared to the Nor2 group (Figure 1I), including Porphyrin metabolism, Aminoacyl-tRNA biosynthesis, and amino acid biosynthesis. The Tib2 group showed five differential pathways (Figure 1J), including Tryptophan metabolism, Melanogenesis, Prolactin signaling pathway, and Tyrosine metabolism. Furthermore, Figure 1E displayed five differential pathways, with the major ones being Phenylalanine metabolism, Glyoxylate and dicarboxylate metabolism, Lysine degradation. The Fec2 group exhibited three metabolic pathways (Figure 1H), with the major ones being Tyrosine metabolism, Microbial metabolism in diverse environments, and Phenylalanine metabolism.

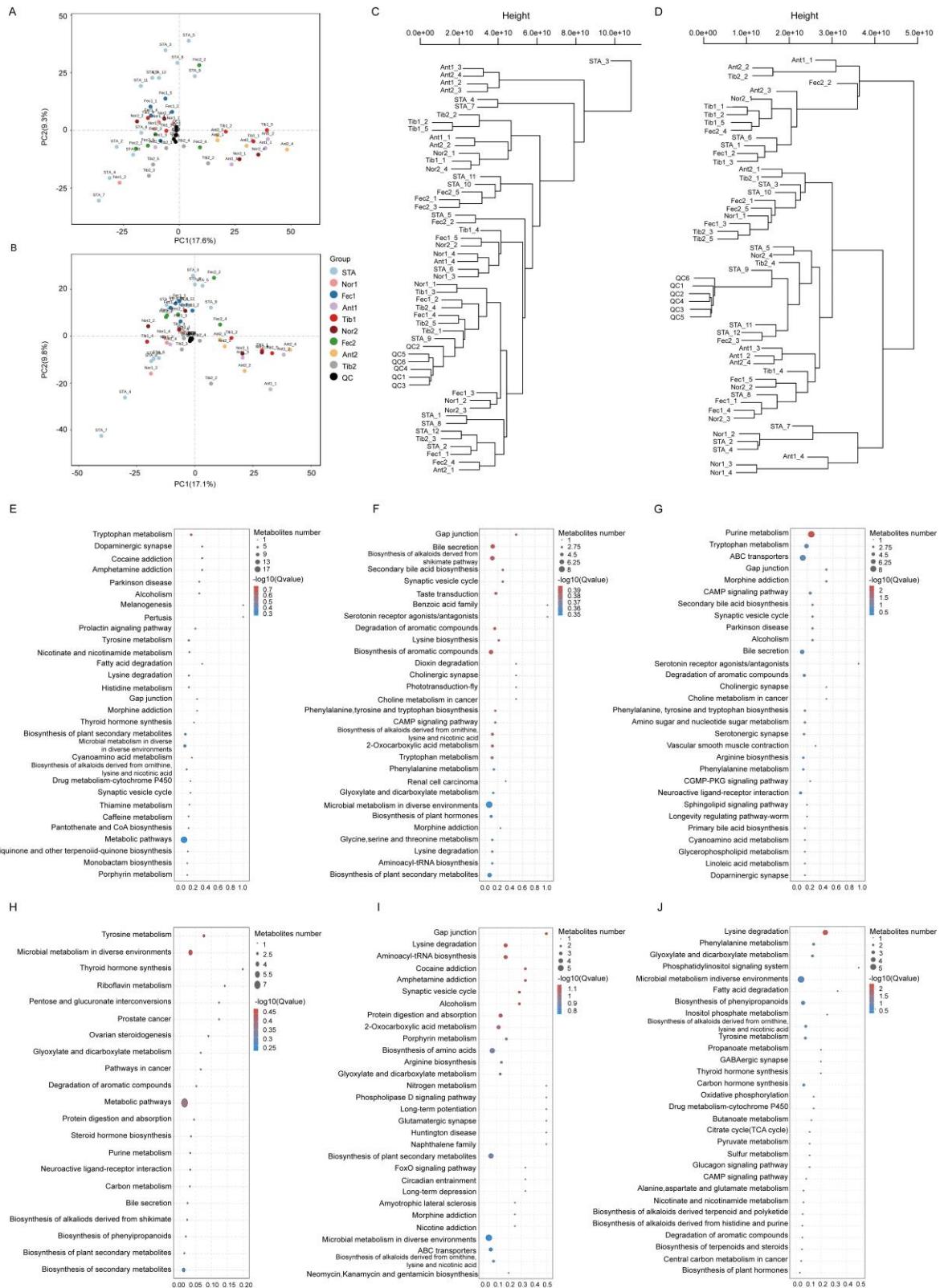


Figure 1. (A) PCA score plot for the ten groups analyzed in the positive ion mode. (B) PCA score plot for the ten groups analyzed in the negative ion mode. (C) Sample dendrogram of the 10 groups analyzed in positive ion mode. (D) Sample dendrogram of the 10 groups analyzed in negative ion mode. (E-J) KEGG significance bubble plot.

3.7. Metabolite KEGG enrichment circle plot analysis

The top 20 enriched pathways were selected to draw KEGG enrichment circle maps, and it can be seen from Figure 2A that benzoic acid family (ko07110) and serotonin receptor agonist (ko07211) were the most significant difference maps in Nor1 and Ant1 groups. As can be seen from Figure 2B fatty acid degradation (ko00071) was the most significant difference plot in Nor2 and Ant2 groups. As can be seen from Figure 2C oocyte meiosis (ko04114) and progesterone-mediated oocyte maturation (ko04914) were the most significant difference maps in Nor1 and Fec1 groups. From Figure 2D, shows that fatty acid degradation (ko00071) and α -linolenic acid metabolism (ko00592) were the most significant difference maps in Nor2 and Fec2 groups. The fatty acid degradation pathway can generate ATP as well as acetyl coenzyme A, which is one of the main sources of energy for the body, and improve the metabolism of the weaned piglets, which in turn promotes growth and development. From Figure 2E, it can be seen that serotonin receptor agonist/antagonist (ko07211) is the most significant difference between Nor1 and Tib1 groups. It has been shown that 95% of serotonin is located in the intestinal tract of animals, and serotonin receptor agonist can regulate the release of serotonin, which is importantly related to the weaned piglets to maintain a good sleep and appetite. From Figure 2F, it can be seen that Nor2 and Tib2 groups, naphthalene (ko07114) and phosphatidylinositol signalling system (ko04070) are the most significant differences between the graphs. Phosphatidylinositol signalling system can regulate HDAC3 activity in intestinal epithelial cells, promoting the repair of intestinal barrier disruption and intestinal homeostasis, thus reducing the diarrhoea rate of piglets.

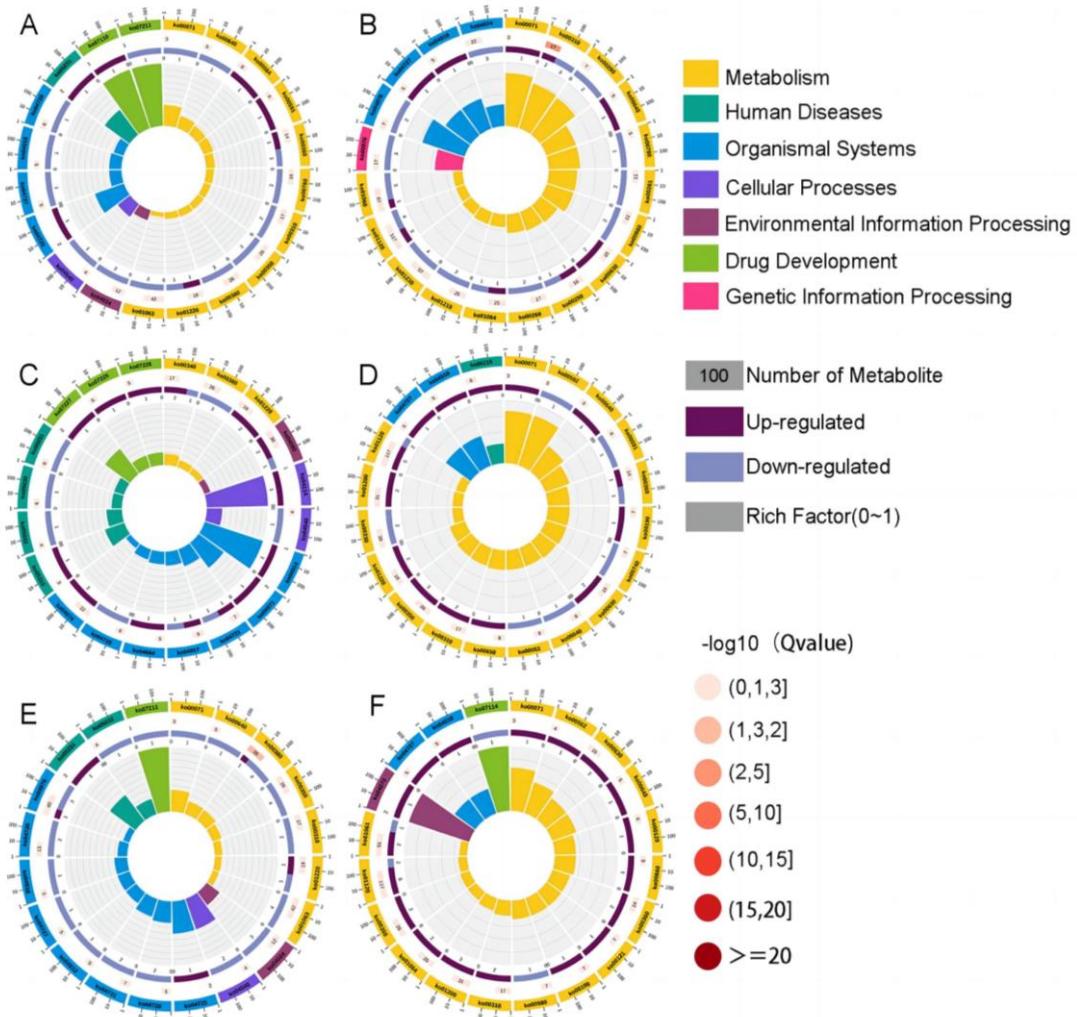


Figure 2. (A-F)Metabolite KEGG enrichment circle plot analysis.

4. Discussion

Based on metabolomics, this experiment utilized multivariate statistical analysis to observe and analyze the effects of adding lincomycin, Fifteen-flavour Black Pill of Tibetan Medicine, and Fecal Bacterial Supernatant to the feed on the growth performance, blood physiological indexes, and metabolites of weaned Tibetan piglets using LC-MC technology. The results indicated that the 15-flavour black pill group and the fecal bacterial supernatant group had more significant effects on the fecal metabolism of weaned piglets. The differential metabolite enrichment analysis revealed that the Tibetan medicine group was involved in the widest range of metabolic pathways on 21d of the experiment.

Faecal microbial transplantation (FMT) is widely used to alter and remodel the gut microbiota of animals, promoting growth and development [23,24]. Oral administration of fecal microbial suspensions to adult gilts has been shown to significantly increase their average daily weight gain and reduce the rate of diarrhea in their piglets [25]. Ren et al. [26] conducted an experimental study on the growth and intestinal health of neonatal piglets, showing that FMT led to greater weight gain and body fat deposition in piglets through the introduction of fecal microorganisms in the colon. Hu et al. [27] demonstrated that transplantation of fecal microbial fluids from healthy adult Jinhua pigs to newborn piglets of "Duchangda" piglets resulted in a significant increase in the average daily gain (ADG) of the piglets. These findings closely align with the results of the present experiment where the feeding of fecal bacterial supernatant significantly reduced the diarrhea rate of weaned piglets and moderately improved their growth performance. Blood, as a crucial component of the animal circulatory system, can objectively reflect the physiology and metabolism of the organism. Changes in the number of leukocytes can indicate the level of cellular immunity [28,29]. The Tib group of weaned piglets exhibited significantly higher leukocyte counts than the Nor group on the 21st day of the test ($P < 0.01$). Furthermore, higher hemoglobin levels contribute to enhanced oxygen transport, immune ability, and metabolic function [30-32]. Clinically, higher hemoglobin levels have been associated with improved body resistance [33]. In this experiment, the Tib group had significantly higher hemoglobin (HGB) levels than the Nor group ($P < 0.05$), which indicates an enhancement in the immune ability of weaned piglets.

In addition to the fecal bacteria refeeding method, the use of natural Chinese herbs to prevent and control piglet diarrhea has gained increasing attention [34-36]. Traditional Chinese herbal medicines, including Tibetan medicines, have been found to be effective in treating bacterial infectious diseases [37]. In this study, the addition of the Tibetan medicine Fifteen Black Herb Pills to the diet reduced the frequency of diarrhea in weaned Tibetan piglets compared to the Nor group. Previous studies have also reported positive effects of dietary supplementation with herbal extracts on the frequency of diarrhea in weaned piglets [38,39]. For instance, Xia et al. [40] used the traditional Tibetan medicine Anzhi Jinhua San in mice and found it effective in treating dyspepsia, anorexia, cold, and diarrhea. Additionally, clinical studies have demonstrated the effectiveness of the Tibetan medicine Fifteen-flavour Black Herbal Pill in treating pain-oriented intermittent diarrhea patients, and in this experiment, it was added to the basal diets of weaned piglets for the first time. The results showed a significant reduction in the diarrhea rate of weaned piglets compared to the Nor group during the 21d and 42d test periods, with no significant difference in growth performance between the Tibetan medicine group and the lincomycin group. Moreover, the Tib group exhibited extremely significantly higher white blood cell counts than the Nor group ($P < 0.01$) and significantly higher hemoglobin (HGB) counts than the Nor group ($P < 0.05$) on 21d of the test. These findings suggest that the Fifteen-flavour black pill of Tibetan medicine has the potential to reduce the diarrhea rate of Tibetan piglets, improve their immune ability, and promote their growth.

Weaning stress disrupts the intestinal flora, leading to a decrease in the growth performance and immunity of the pig [41,42]. The addition of lincomycin, Tibetan medicine's fifteen-flavour black pill, and faecal bacterial supernatant can enhance the growth performance and immunity of weaned piglets, although the effect on intestinal flora still requires further investigation. Therefore, this experiment investigated the effects of different feed additives on the microbial metabolites of intestinal flora. It was found that the Fec group, compared to the Nor group, exhibited up-regulated

differential metabolites including lactitol, L-arginine succinate, and 3-hydroxy-o-cyanoanthranilic acid (3-HAA). 3-HAA, a tryptophan metabolite produced through the indoleamine-2,3-dioxygenase (IDO) pathway, plays a vital role in the regulation of the immune system [43]. Moreover, 3-HAA inhibits cytokine production by Th1 and Th2 cells through the enhancement of heme oxygenase-1 (HO-1) [44] and the expression of inducible nitric oxide synthase (iNOS) [45]. Additionally, the regulatory mechanism of 3-HAA has been shown to inhibit the production of inflammatory mediators by macrophages through the LPS-induced activation of the PI3K/Akt signaling pathway [46]. This experiment found that the Fec group modulated the immune system of weaned piglets by up-regulating the differential metabolite 3-HAA, resulting in an increase in the haemoglobin content of piglets' blood, inhibition of cytokine secretion by T cells, and alteration of macrophage activity. It also exhibited anti-inflammatory activity, leading to a decrease in the diarrhoea rate compared to the Nor group.

According to the differential metabolite KEGG analysis, the Tib, Fec, and Nor groups were found to exhibit tryptophan metabolism, which is the main metabolic pathway in the gastrointestinal tract of animals [47]. Dietary tryptophan is primarily acted upon by gut microbes, producing indole and derivatives. Indole and its derivatives play a protective role for bacteria against antibiotics by regulating flagellar synthesis and expression of virulence factors [48-50]. They also act as agonists of the Aryl Hydrocarbon Receptor (AhR), participating in the regulation of gut microbial load, structure, intestinal endothelium, lymphocyte immune tolerance, and other physiological processes [51]. Bansal et al. [52] demonstrated that indole promotes the expression of genes related to the mucosal barrier and mucus secretion. Moreover, reduced expression of AhR receptors in the intestinal mucosa of Crohn's disease patients was found, worsening symptoms of colitis due to a lack of tryptophan in the diet. Jennis et al. [53] observed increased intestinal mucosal permeability in mice that were fed a high-fat diet for 22 consecutive weeks. However, this increase was restored after treatment with indole-3-acetic acid, along with a decrease in circulating lipopolysaccharide (LPS) in the blood. In this experiment, the addition of Tibetan medicine's fifteen-flavour black pill and faecal bacterial supernatant regulated the intestinal microflora through the tryptophan metabolic pathway, thereby increasing the bacteriostatic activity of the intestine compared to the Nor group. The AhR signaling generated by the metabolism of tryptophan along the pathway of the intestinal flora was essential for the intestinal stability of the weaned piglets [54,55]. It helped to maintain the intestinal mucosal barrier function and reduce the diarrhoea rate of the weaned piglets.

In conclusion, this experiment systematically observed the effects of Tibetan medicine's fifteen-flavour black pill and the supernatant of faecal bacteria on the intestinal metabolites of weaned Tibetan piglets for the first time. The findings confirmed that refeeding with Tibetan medicine's fifteen-flavour black pill and faecal bacterial supernatant can significantly regulate the intestinal metabolism of weaned Tibetan piglets. The therapeutic efficacy of these additives in clinics may be related to the regulation of the organism's metabolic pathways, particularly the metabolic pathway of tryptophan.

5. Conclusions

In summary, our findings suggest that the anti-feeding of faecal bacterial supernatant and the fifteen-flavour black pill of Tibetan medicine improve the growth performance of weaned piglets and certain blood physiological indexes, such as white blood cells and haemoglobin. We also speculate that these improvements are due to the influence on microbial metabolites like 3-hydroxy-octylaminobenzoic acid (3-HAA) and metabolic pathways like tryptophan metabolism. These effects may lead to a reduction in diarrhoea rates, enhancement of immune ability, and increased growth performance of weaned Tibetan piglets. Moreover, our results indicate that the fifteen-flavour black pill of Tibetan medicine and the anti-feeding of faecal bacterial supernatant exhibit a certain substitutive effect. This reveals a potential for the substitution products to be scientifically evaluated and selected, bringing real benefits to the relevant industry's development.

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References

1. Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets[J]. *J Anim Sci Biotechnol*, 2013, 4(1):19.
2. Spring S, Premathilake H, Bradway C et al. Effect of very low-protein diets supplemented with branched-chain amino acids on energy balance, plasma metabolomics and fecal microbiome of pigs[J]. *Sci Rep*, 2020, 10(1):15859.
3. Canibe N, Hojberg O, Kongsted H et al. Review on Preventive Measures to Reduce Post-Weaning Diarrhoea in Piglets[J]. *Animals (Basel)*, 2022, 12(19):2585.
4. Lin C, Wan J, Su Y et al. Effects of Early Intervention with Maternal Fecal Microbiota and Antibiotics on the Gut Microbiota and Metabolite Profiles of Piglets[J]. *Metabolites*, 2018, 8(4):89.
5. Youngster I, Sauk J, Pindar C et al. Fecal microbiota transplant for relapsing *Clostridium difficile* infection using a frozen inoculum from unrelated donors: a randomized, open-label, controlled pilot study[J]. *Clin Infect Dis*, 2014, 58(11):1515-1522.
6. Borody TJ, Khoruts A. Fecal microbiota transplantation and emerging applications[J]. *Nat Rev Gastroenterol Hepatol*, 2011, 9(2):88-96.
7. Heath RD, Cockerell C, Mankoo R et al. Fecal microbiota transplantation and its potential therapeutic uses in gastrointestinal disorders[J]. *North Clin Istanb*, 2018, 5(1):79-88.
8. Pigneur B, Sokol H. Fecal microbiota transplantation in inflammatory bowel disease: the quest for the holy grail[J]. *Mucosal Immunol*, 2016, 9(6):1360-1365.
9. Cheng S, Ma X, Geng S et al. Fecal Microbiota Transplantation Beneficially Regulates Intestinal Mucosal Autophagy and Alleviates Gut Barrier Injury[J]. *mSystems*, 2018, 3(5):e00137-00118.
10. Hu J, Ma L, Nie Y et al. A Microbiota-Derived Bacteriocin Targets the Host to Confer Diarrhea Resistance in Early-Weaned Piglets[J]. *Cell Host Microbe*, 2018, 24(6):817-832 e818.
11. Xiang Q, Wu X, Pan Y et al. Early-Life Intervention Using Fecal Microbiota Combined with Probiotics Promotes Gut Microbiota Maturation, Regulates Immune System Development, and Alleviates Weaning Stress in Piglets[J]. *Int J Mol Sci*, 2020, 21(2):503.
12. Wang H, Xu R, Zhang H et al. Swine gut microbiota and its interaction with host nutrient metabolism[J]. *Anim Nutr*, 2020, 6(4):410-420.
13. Kurbatov I, Dolgalev G, Arzumanian V et al. The Knowns and Unknowns in Protein-Metabolite Interactions[J]. *Int J Mol Sci*, 2023, 24(4):4155.
14. Zhu LR, Li SS, Zheng WQ et al. Targeted modulation of gut microbiota by traditional Chinese medicine and natural products for liver disease therapy[J]. *Front Immunol*, 2023, 14:1086078.
15. Gresse R, Chaucheyras-Durand F, Fleury MA et al. Gut Microbiota Dysbiosis in Postweaning Piglets: Understanding the Keys to Health[J]. *Trends Microbiol*, 2017, 25(10):851-873.
16. Wang T, Teng K, Liu Y et al. Lactobacillus plantarum PFM 105 Promotes Intestinal Development Through Modulation of Gut Microbiota in Weaning Piglets[J]. *Front Microbiol*, 2019, 10:90.
17. Hu J, Chen L, Tang Y et al. Standardized Preparation for Fecal Microbiota Transplantation in Pigs[J]. *Front Microbiol*, 2018, 9:1328.
18. Nowland TL, Tork VA, Low WY et al. A single faecal microbiota transplantation altered the microbiota of weaned pigs[J]. *Life*, 2020, 10(9):203.
19. Liu T, Guo Y, Lu C et al. Effect of Different Pig Fecal Microbiota Transplantation on Mice Intestinal Function and Microbiota Changes During Cold Exposure[J]. *Front Vet Sci*, 2022, 9:805815.
20. Diao H, Yan HL, Xiao Y et al. Intestinal microbiota could transfer host Gut characteristics from pigs to mice[J]. *BMC microbiology*, 2016, 16(1):238.

21. Zierer J, Jackson MA, Kastenmuller G et al. The fecal metabolome as a functional readout of the gut microbiome[J]. *Nat Genet*, 2018, 50(6):790-795.
22. Jayaraman B, Nyachoti CM. Husbandry practices and gut health outcomes in weaned piglets: A review[J]. *Anim Nutr*, 2017, 3(3):205-211.
23. Li M, Li F, Lu Z et al. Effects of TiO(2) nanoparticles on intestinal microbial composition of silkworm, *Bombyx mori*[J]. *Sci Total Environ*, 2020, 704:135273.
24. Barnes D, Ng K, Smits S et al. Competitively Selected Donor Fecal Microbiota Transplantation: Butyrate Concentration and Diversity as Measures of Donor Quality[J]. *J Pediatr Gastroenterol Nutr*, 2018, 67(2):185-187.
25. Rahman R, Fouhse JM, Prisnee TL et al. Comparing the impact of mixed-culture microbial communities and fecal transplant on the intestinal microbiota and metabolome of weaned piglets[J]. *FEMS Microbiol Ecol*, 2023, 99(7).
26. Qi R, Zhang Z, Wang J et al. Introduction of Colonic and Fecal Microbiota From an Adult Pig Differently Affects the Growth, Gut Health, Intestinal Microbiota and Blood Metabolome of Newborn Piglets[J]. *Front Microbiol*, 2021, 12:623673.
27. Hu L, Geng S, Li Y et al. Exogenous Fecal Microbiota Transplantation from Local Adult Pigs to Crossbred Newborn Piglets[J]. *Front Microbiol*, 2017, 8:2663.
28. Mathew DJ, Lucy MC, R DG. Interleukins, interferons, and establishment of pregnancy in pigs[J]. *Reproduction*, 2016, 151(6):R111-122.
29. Quinn JG, Tansey EA, Johnson CD et al. Blood: tests used to assess the physiological and immunological properties of blood[J]. *Adv Physiol Educ*, 2016, 40(2):165-175.
30. Moraes LA, Piqueras L, Bishop-Bailey D. Peroxisome proliferator-activated receptors and inflammation[J]. *Pharmacol Ther*, 2006, 110(3):371-385.
31. Yang R, Gao G, Liu T et al. Enhanced ability of hemoglobin to carry oxygen by salidroside[J]. *Electrochemistry Communications*, 2007, 9(1):94-96.
32. Gabler NK, Spurlock ME. Integrating the immune system with the regulation of growth and efficiency[J]. *J Anim Sci*, 2008, 86(14 Suppl):E64-74.
33. Smuda K, Gienger J, Hönicke P et al. Function of hemoglobin-based oxygen carriers: Determination of methemoglobin content by spectral extinction measurements[J]. *International Journal of Molecular Sciences*, 2021, 22(4):1753.
34. Hu TY, Ju JM, Mo LH et al. Anti-inflammation action of xanthones from *Swertia chirayita* by regulating COX-2/NF-κB/MAPKs/Akt signaling pathways in RAW 264.7 macrophage cells[J]. *Phytomedicine*, 2019, 55:214-221.
35. Liu CS, Liang X, Wei XH et al. Gegen Qinlian Decoction Treats Diarrhea in Piglets by Modulating Gut Microbiota and Short-Chain Fatty Acids[J]. *Front Microbiol*, 2019, 10:825.
36. Zhao J, Zhang G, Zhou X et al. Effect of Dandelion root extract on growth performance, immune function and bacterial community in weaned pigs[J]. *Food and Agricultural Immunology*, 2019, 30(1):95-111.
37. Wang M, Huang H, Hu Y et al. Effects of dietary supplementation with herbal extract mixture on growth performance, organ weight and intestinal morphology in weaning piglets[J]. *J Anim Physiol Anim Nutr (Berl)*, 2020, 104(5):1462-1470.
38. Xia Z, Zhang Y, Li C et al. Traditional Tibetan medicine Anzhijinhua San attenuates ovalbumin-induced diarrhea by regulating the serotonin signaling system in mice[J]. *J Ethnopharmacol*, 2019, 236:484-494.
39. Chen G, Li Z, Liu S et al. Fermented chinese herbal medicine promoted growth performance, intestinal health, and regulated bacterial microbiota of weaned piglets[J]. *Animals*, 2023, 13(3):476.
40. Świtała M, Kołacz R, Bodak-Koszałka E et al. Haematological and biochemical parameters of blood and immune response of runt weaners[J]. *Journal of Animal and Feed Sciences*, 1998, 7:405-413.
41. Xu Q, Cheng M, Jiang R et al. Effects of dietary supplement with a Chinese herbal mixture on growth performance, antioxidant capacity, and gut microbiota in weaned pigs[J]. *Front Vet Sci*, 2022, 9:971647.
42. Mahmud MR, Jian C, Uddin MK et al. Impact of Intestinal Microbiota on Growth Performance of Suckling and Weaned Piglets[J]. *Microbiol Spectr*, 2023, 11(3):e0374422.
43. Lee K, Kwak JH, Pyo S. Inhibition of LPS-induced inflammatory mediators by 3-hydroxyanthranilic acid in macrophages through suppression of PI3K/NF-κB signaling pathways[J]. *Food Funct*, 2016, 7(7):3073-3082.

44. Yoon SJ, Kim SJ, Lee SM. Overexpression of HO-1 Contributes to Sepsis-Induced Immunosuppression by Modulating the Th1/Th2 Balance and Regulatory T-Cell Function[J]. *J Infect Dis*, 2017, 215(10):1608-1618.
45. Yan Y, Zhang GX, Gran B et al. IDO upregulates regulatory T cells via tryptophan catabolite and suppresses encephalitogenic T cell responses in experimental autoimmune encephalomyelitis[J]. *J Immunol*, 2010, 185(10):5953-5961.
46. Xia TT, Hu R, Shao CJ et al. Stanniocalcin-1 secreted by human umbilical mesenchymal stem cells regulates interleukin-10 expression via the PI3K/AKT/mTOR pathway in alveolar macrophages[J]. *Cytokine*, 2023, 162:156114.
47. Grifka-Walk HM, Jenkins BR, Kominsky DJ. Amino Acid Trp: The Far Out Impacts of Host and Commensal Tryptophan Metabolism[J]. *Front Immunol*, 2021, 12:653208.
48. Lu Y, Chong J, Shen S et al. TrpNet: Understanding Tryptophan Metabolism across Gut Microbiome[J]. *Metabolites*, 2021, 12(1).
49. Hyland NP, Cavanaugh CR, Hornby PJ. Emerging effects of tryptophan pathway metabolites and intestinal microbiota on metabolism and intestinal function[J]. *Amino Acids*, 2022, 54(1):57-70.
50. Konopelski P, Ufnal M. Indoles - Gut Bacteria Metabolites of Tryptophan with Pharmacotherapeutic Potential[J]. *Curr Drug Metab*, 2018, 19(10):883-890.
51. Wang YD, Chen WD, Yu D et al. The G-protein-coupled bile acid receptor, Gpbar1 (TGR5), negatively regulates hepatic inflammatory response through antagonizing nuclear factor kappa light-chain enhancer of activated B cells (NF- κ B) in mice[J]. *Hepatology*, 2011, 54(4):1421-1432.
52. Keitel V, Donner M, Winandy S et al. Expression and function of the bile acid receptor TGR5 in Kupffer cells[J]. *Biochem Biophys Res Commun*, 2008, 372(1):78-84.
53. Potthoff MJ, Boney-Montoya J, Choi M et al. FGF15/19 regulates hepatic glucose metabolism by inhibiting the CREB-PGC-1alpha pathway[J]. *Cell Metab*, 2011, 13(6):729-738.
54. Liang H, Dai Z, Liu N et al. Dietary L-Tryptophan Modulates the Structural and Functional Composition of the Intestinal Microbiome in Weaned Piglets[J]. *Front Microbiol*, 2018, 9:1736.
55. Liu JR, Miao H, Deng DQ et al. Gut microbiota-derived tryptophan metabolism mediates renal fibrosis by aryl hydrocarbon receptor signaling activation[J]. *Cell Mol Life Sci*, 2021, 78(3):909-922.

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