
Based on UHPLC-Q-Orbitrap HRMS, the Effect of Chemical Composition before and after Fermentation of Gastrodia Tuder Halimasch Powder Was Compared

[Yaning Wu](#) , Hongwei Zhang , [Zhenling Zhang](#) * , [Jianguang Zhu](#) * , Songbo Ma , Yongqi Zhao , Yiming Wang , Jun Yuan , Xing Guo , Yajing Li , Shuai Zhang

Posted Date: 8 September 2023

doi: 10.20944/preprints202309.0606.v1

Keywords: gastrodia tuder halimasch powder; ultra high performance liquid chromatography tan- 34 dem quadrupole electrostatic field orbitrap high resolution mass spectrometry; stoichiometry; fer- 35 mentation processing



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article

Based on UHPLC-Q-Orbitrap HRMS, the Effect of Chemical Composition before and after Fermentation of Gastrodia Tuder Halimasch Powder Was Compared

Yaning Wu ¹, Hongwei Zhang ¹, Zhenling Zhang ^{1,2,3,*}, Jianguang Zhu ^{1,*}, Songbo Ma ⁴, Yongqi Zhao ¹, Yiming Wang ¹, Jun Yuan ¹, Xing Guo ¹, Yajing Li ¹ and Shuai Zhang ¹

¹ Henan University of Chinese Medicine and Zhengzhou 450046

² Henan Engineering Research Center of Traditional Chinese Medicine Characteristic Processing Technology, Zhengzhou 450046

³ Henan Engineering Technology Research Center for Integrated Traditional Chinese Medicine Production, Zhengzhou 450046

⁴ Luoyang Wokang Pharmaceutical Co., Ltd. Luoyang, 471521 China

* Correspondence: zhangzl6758@163.com (Z.Z.); 13503859285@139.com (J.Z.)

Abstract: Objective: To compare the effects of fermentation on the chemical composition of Gastrodia Tuder Halimasch Powder, and to provide a basis for the processing, processing and clinical application of this medicinal material. Methods: UHPLC-Q-Orbitrap HRMS was used to identify compound structure by comparing excimer ions, characteristic fragment ions and reference substances, and referring to relevant literature and database search. The main differential components were screened by orthogonal partial least squares-discriminant analysis (OPLS-DA), and the differential components were quantitatively studied by high performance liquid chromatography, and the differences in the types and contents of chemical components before and after fermentation were compared. Results: A total of 77 compounds were identified or preliminarily derived after fermentation, including 21 terpenes, 18 organic acids, 9 flavonoids, 7 nucleosides, 6 amides, 4 amino acids, 2 pyrrolidone derivatives, 2 sterols, 2 alkaloids and 6 others. Through high performance liquid chromatography, it was found that the content of daidzein genistein and ergosterol was significantly increased after fermentation, and a new component ergosterol was produced. Compared with Gastrodia Tuder Halimasch Powder before fermentation, the content of genistin and other components was significantly reduced, and there may be a process of genistin conversion to genistein. Conclusion: The fermentation process will have a certain influence on the types and contents of the chemical components of Gastrodia Tuder Halimasch Powder, and ergosterol, genistein and daidzein may be the main pharmacodynamic components, which can provide a useful reference for the clinical rational application, quality control and mechanism of action of Gastrodia Tuder Halimasch Powder.

Keywords: gastrodia tuder halimasch powder; ultra high performance liquid chromatography tandem quadrupole electrostatic field orbitrap high resolution mass spectrometry; stoichiometry; fermentation processing

1. Introduction

Gastrodia Tuder Halimasch Powder is a member of the family *Armillaria mellea* (vane. ex. Fr.) Quel. The dried bacteria powder obtained by liquid fermentation and culture, separated and extracted, was included in the 2002 (Chemical Landmark Upgrading National Standard 13 Volumes) [1]. Honey ring bacteria are rich in chemical components, including terpenes, sugars, nucleosides, sterols and other components [2]. Modern pharmacological studies have shown that aqueous extracts and ethanol extracts of mycelium of *Mycomycetes* have the effect of improving insomnia [3]; Ethyl

acetate extract of *Honeycilla* is able to inhibit inflammatory mediators [4]; *Melania* polysaccharides are resistant to Alzheimer's disease and hypoglycemia [5,6]; Sesquiterpene aromatic esters in Honey Ring are antidepressant [7]; Melleolide-like compounds in honey ring bacteria have anti-liver cancer and anti-*Aspergillus* effects [8,9].

After reviewing the literature, soy glycogen can improve ischemic brain injury [10], Cerebral edema [11], Vascular endothelial dysfunction [12,13] and anti-epileptic [14]; Genistein has anti-A β neurotoxicity [15], atherosclerosis [16], Regulates blood sugar [17] and lipids [18]; Ergosterol against Alzheimer's disease [19], diabetes [20], Hepatic steatosis [21] and to neuroprotection [22]; Clinically used for neuroprotection with *Gastrodia Tuder Halimasch Powder* [23], Treatment of tension headaches [24], Lower blood sugar [25] and other effects. Daidzein, genistein and ergosterol may be the main pharmacodynamic components of *Gastrodia Tuder Halimasch Powder*.

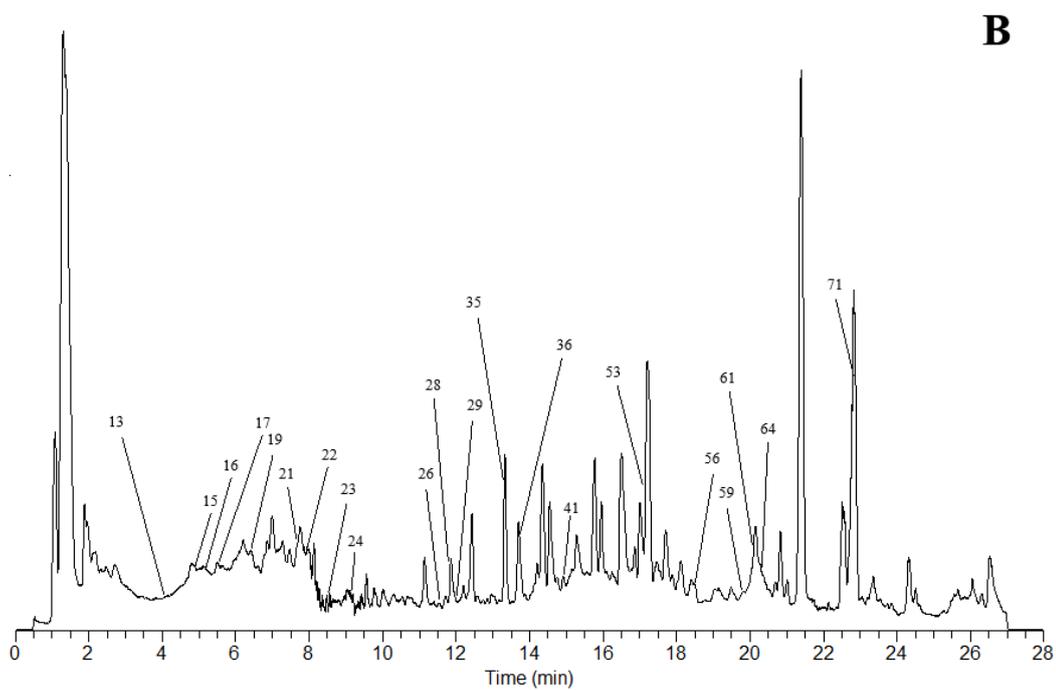
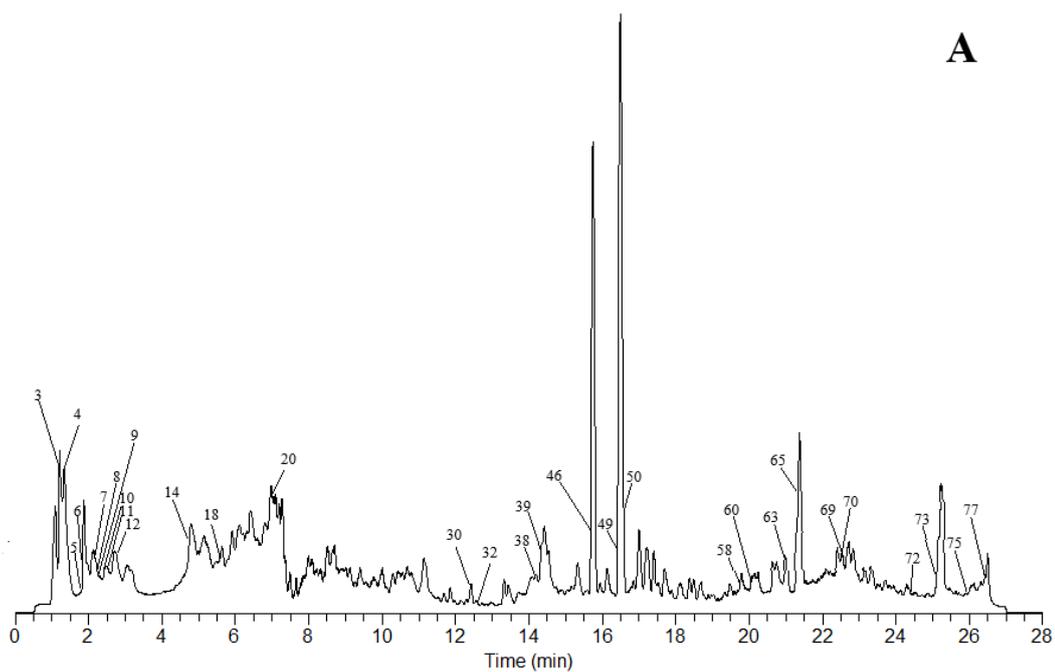
At present, there are few reports on the effect of *Gastrodia Tuder Halimasch Powder* on its chemical composition before and after fermentation, and the chemical components of traditional Chinese medicines are of great significance for elucidating the mechanism of action of traditional Chinese medicines and guiding the safety of clinical medication, which restricts the clinical rational application of *Gastrodia Tuder Halimasch Powder* to a certain extent [26]. Ultra-performance liquid chromatography-quadrupole / electrostatic field orbital well high-resolution mass spectrometry (UHPLC-Q-Orbitrap HRMS) It is mostly used in the analysis of complex compounds in traditional Chinese medicine, and has the characteristics of high resolution, high sensitivity and strong qualitative ability [27]. Therefore, the chemical components of *Gastrodia Tuder Halimasch Powder* before and after fermentation were analyzed based on UHPLC-Q-ORBITRAP HRMS, and the main chemical components were identified according to the chromatographic peak retention time (tR), secondary fragment ions and reference comparison, combined with relevant literature reports; Orthogonal partial least squares-discriminant analysis (OPLS-DA) was used to screen the main differentiating components; The HPLC method was used to determine the multi-component content in *Gastrodia Tuder Halimasch Powder*, and then PCA treatment was carried out to group the *Gastrodia Tuder Halimasch Powder* before and after fermentation, and the SPSS 26.0 software was used to analyze the LSD variance of the data in pairs; According to this, the Origin 8.0 drawing software was used to draw a significant difference marker histogram to compare the difference in the content of components before and after fermentation of *Gastrodia Tuder Halimasch Powder*. The fermentation-based method adopted by *Gastrodia Tuder Halimasch Powder* is of great significance to enhance the clinical efficacy, and provides a reference for exploring the clinical rational application and quality control research of *Gastrodia Tuder Halimasch Powder*.

2. Results and Discussion

2.1. Chemical composition analysis of *Gastrodia Tuder Halimasch Powder*

The ion flow (TIC) of *Armillaria gastrodia* powder before and after fermentation under positive and negative ion modes is shown in Figure 1. Xcalibur 4.5 was used to process the original mass spectrometry data, and chemical formulas were obtained by accurate relative molecular mass and excimer ion peaks of compounds. 77 components were preliminarily qualitatively analyzed by using MS/MS spectra, characteristic fragment ions and other fragment ions, reference reports and database analysis. It included 21 terpenoids, 18 organic acids, 9 flavonoids, 7 nucleosides, 6 amides, 4 amino acids, 2 pyrrolidone derivatives, 2 steroids, 2 alkaloids and 6 other classes, as shown in Table 1.

2.2. Analysis of main components of *Gastrodia Tuder Halimasch Powder*



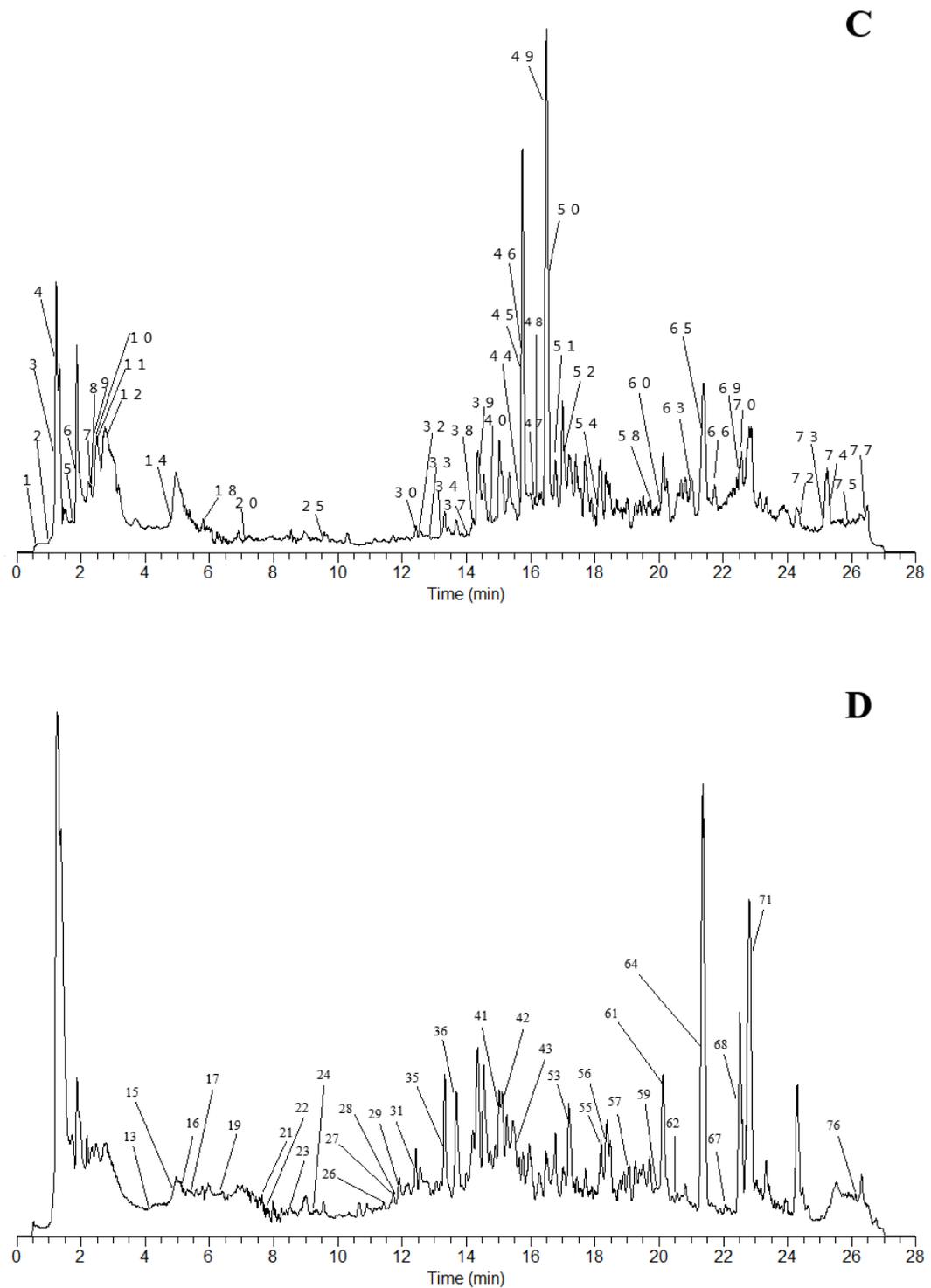


Figure 1. TIC diagram of Gastrodia Tuder Halimasch Powder under positive ion mode (A) and negative ion mode (B) before fermentation and positive ion mode (C) and negative ion mode (D) after fermentation.

Table 1. UHPLC-Q-Exactive Orbitrap HRMS information of chemical components of Gastrodia Tuder Halimasch Powder before and after fermentation.

| Numbering | Class | Compound | Retention Time | Molecular Formula | Measured Value | Calculated Value | Error $\times 10^{-6}$ | Fragment Ions | After Fermentation | Before Fermentation |
|-----------|-------------------------|---------------------|----------------|----------------------|----------------|------------------|------------------------|--|--------------------|---------------------|
| 1 | Other | Benzaldehyde | 0.54 | C_7H_6O | 107.04893 | 107.04914 | -1.96 | 79.05432 | + | - |
| 2 | Amides | Phenacetin | 1.03 | $C_{10}H_{13}NO_2$ | 180.10207 | 180.10191 | 0.89 | 138.09169 | + | - |
| 3 | Alkaloids | Choline | 1.29 | $C_5H_{13}NO$ | 104.10693 | 104.10699 | -0.58 | 60.08082, 58.06518 | + | + |
| 4 | Nucleosides | Cytidine* | 1.38 | $C_9H_{13}N_3O_5$ | 244.09282 | 244.09279 | 0.12 | 112.05049 | + | + |
| 5 | Nucleosides | Uridine* | 1.81 | $C_9H_{12}N_2O_6$ | 245.0768 | 245.07681 | -0.04 | 113.03452 | + | + |
| 6 | Organic acids | Nicotinic acid | 1.86 | $C_6H_5NO_2$ | 124.03936 | 124.0393 | 0.48 | 80.04947, 96.04435, 78.03384 | + | + |
| 7 | Amino acids | L-Tyrosine | 2.22 | $C_9H_{11}NO_3$ | 182.08092 | 182.08117 | -1.37 | 165.05461, 136.07568 | + | + |
| 8 | Nucleosides | Adenosine* | 2.34 | $C_{10}H_{13}N_5O_4$ | 268.10406 | 268.10403 | 0.11 | 136.06178, 119.03522 | + | + |
| 9 | Pyrrolidone derivatives | Piracetam | 2.39 | $C_6H_{10}N_2O_2$ | 143.08144 | 143.0815 | -0.42 | 98.05994, 126.05515, 70.06517 | + | + |
| 10 | Nucleosides | Guanosine* | 2.45 | $C_{10}H_{13}N_5O_5$ | 284.09894 | 284.09894 | 0 | 152.0567 | + | + |
| 11 | Nucleosides | Guanine* | 2.57 | $C_5H_5N_5O$ | 152.05667 | 152.05669 | -0.13 | 135.03012, 110.03482, 109.05079, 128.04536 | + | + |
| 12 | Nucleosides | Uracil* | 2.8 | $C_4H_4N_2O_2$ | 113.03458 | 113.03455 | 0.27 | 96.00798 | + | + |
| 13 | Organic acids | Methylsuccinic acid | 4.1 | $C_5H_8O_4$ | 131.035 | 131.03498 | 0.15 | 87.0451 | + | + |
| 14 | Nucleosides | Thymine | 4.84 | $C_5H_6N_2O_2$ | 127.05011 | 127.0502 | -0.71 | 110.03175 | + | + |
| 15 | Amino acids | D-Phenylalanine | 4.98 | $C_9H_{11}NO_2$ | 164.07172 | 164.0717 | 0.12 | 164.07172 | + | + |

| | | | | | | | | | | |
|----|-------------------------|---------------------------------|-------|----------------------|-------------------|---------------|-----------|---------------------------------|---|---|
| 16 | Organic acids | 3,4-Dihydroxyphenylacetic acid | 5.23 | $C_8H_8O_4$ | [M-] 167.03506 | 167.0 3498 | 0.48 | 123.04531, 122.03760 | + | + |
| 17 | Amino acids | 3-Hydroxy-3-methylglutaric acid | 5.41 | $C_6H_{10}O_5$ | [M-] 161.0455 | 161.0 4555 | - 0.31 | 57.03456, 59.01381, 99.04536, | + | + |
| 18 | Organic acids | Pantothenic acid | 5.79 | $C_9H_{17}NO_5$ | [M+] 220.11797 | 220.1 1795 | 0.09 | 202.10672, 184.09637 | + | + |
| 19 | Organic acids | 3-Hydroxy-3-methylbutanoic acid | 6.43 | $C_5H_{10}O_3$ | [M-] 117.05579 | 117.0 5572 | 0.60 | 71.05026, 115.03962, 99.04516 | + | + |
| 20 | Pyrrolidone derivatives | Levetiracetam | 7.05 | $C_8H_{14}N_2O_2$ | [M+] 171.11284 | 171.1 128 | 0.23 | 126.09127, 89.07092, 72.08067 | + | + |
| 21 | Other | Salicylic acid | 7.62 | $C_7H_6O_3$ | [M-] 137.02437 | 137.0 2442 | - 0.36 | 93.03452 | + | + |
| 22 | Other | 2-Isopropylmalic acid | 7.9 | $C_7H_{12}O_5$ | [M-] 175.06126 | 175.0 612 | 0.34 | 115.04008, 85.06586, 113.06087 | + | + |
| 23 | Organic acids | Terephthalic acid | 8.58 | $C_8H_6O_4$ | [M-] 165.01938 | 165.0 193 | 0.48 | 121.0296 | + | + |
| 24 | Organic acids | benzoic acid | 9.12 | $C_7H_6O_2$ | [M-] 121.02943 | 121.0 295 | - 0.58 | 93.03461 | + | + |
| 25 | Organic acids | 5-Hydroxyindole-3-acetic acid | 9.63 | $C_{10}H_9NO_3$ | [M+] 192.06543 | 192.0 6552 | - 0.47 | 146.06000, 147.06816 | + | - |
| 26 | Flavonoids | Genistin* | 11.53 | $C_{21}H_{20}O_{10}$ | [M-] 431.09836 | 431.0 9837 | - 0.02 | 269.04517 | + | + |
| 27 | Flavonoids | Hispidulin | 11.89 | $C_{16}H_{12}O_6$ | [M-] 299.0563 | 299.0 5611 | 0.64 | 300.05914, 284.03268, 285.03671 | + | - |
| 28 | Flavonoids | Kaempferide | 11.93 | $C_{16}H_{12}O_6$ | [M-] 299.05658 | 299.0 5611 | 0.64 | 284.03271, 285.03638 | + | + |
| 29 | Flavonoids | Naringenin | 12.05 | $C_{15}H_{12}O_5$ | [M-] 271.06149 | 271.0 612 | 1.07 | 151.00380, 119.05028, 107.01356 | + | + |
| 30 | Flavonoids | Daidzein* | 12.4 | $C_{15}H_{10}O_4$ | [M+] 255.06519 | 255.0 6519 | 0 | 227.07022, 199.07542, 137.02338 | + | + |
| 31 | Flavonoids | Fisetin | 12.49 | $C_{15}H_{10}O_6$ | [M-] 285.04059 | 285.0 4046 | 0.46 | 135.00888, 256.03601 | + | - |

[M+H]⁺ in the positive ion mode, and its molecular formula is inferred to be C₂₄H₃₀O₆. The fragment ion 165.05457 [M+H-C₁₅H₂₁O₂-OH]⁺ combined with reference identified it as armillarin^[28]. Compound 50 (t_R=16.66 min) gives the molecular ion peak m/z 397.20105 [M+H]⁺ in the positive ion mode, and its molecular formula is inferred to be C₂₄H₂₈O₅. Fragment ions 232.14107 [M+H-C₉H₉O₃]⁺, 215.14299 [M+H-C₉H₉O₃-OH]⁺, 187.14807 [M+H-C₉H₉O₃-CO]⁺, 185.13257 [M+H-C₉H₉O₃-CO-H₂]⁺, 171.11671 [M+H-C₉H₉O₃-CO-H₂-CH₄]⁺, 165.05460 [M+H-C₁₅H₂₀O₂]⁺ and 131.08546 [M+H-CO-C₄H₈]⁺ were identified as armillaribin by reference^[29]. Compound 51 (t_R=16.85 min) gives the molecular ion peak m/z 431.16205 [M+H]⁺ in the positive ion mode, and its molecular formula is inferred to be C₂₄H₂₇O₅Cl. Fragment ion 215.14302 [M+H-C₉H₈ClO₃-OH]⁺, 199.01566 [M+H-C₁₅H₂₀O₂]⁺, 187.14810 [M+H-C₉H₈ClO₃-CO]⁺ and 171.11707 [M+H-C₁₅H₂₀O₂-CO]⁺. It was identified as armillaricin by reference^[30]. Compound 52 (t_R=17.18 min) gives the molecular ion peak m/z 401.19601 [M+H]⁺ in the positive ion mode, and its molecular formula is inferred to be C₂₃H₂₈O₆, fragment ion 233.15363 [M+H-C₈H₈O₄]⁺. It was identified as melleolide by reference^[29]. Compound 53 (t_R =17.22 min) gives the molecular ion peak m/z 295.22781 in the positive ion mode, and its molecular formula is inferred to be C₁₈H₃₂O₃ and fragment ion 277.21704 [M-H-H₂O]⁺, which is identified as coriolic acid by reference^[31]. Compound 54 (t_R=18.19 min) gives a molecular ion peak m/z 449.17240 [M+H]⁺ in the positive ion mode, and its molecular formula is inferred to be C₂₄H₂₉O₆Cl. The fragment ions m/z 233.15359 [M+H-C₉H₉O₄Cl]⁺ and 199.01558 [M+H-C₁₅H₂₁O₂-OH]⁺ were identified as armillaridin by reference^[28].

2.2.2. Identification of organic acid compounds

According to MS information, a total of 18 organic acid compounds were detected, taking compounds 18, 64 and 68 as examples. Compound 18 (t_R=5.79 min) gives the molecular ion peak m/z 220.11797 in the positive ion mode, and its molecular formula is inferred to be C₉H₁₇NO₅, fragment ion 202.10672 [M+H-H₂O]⁺ and 184.09637 [M+H-2H₂O]⁺. It was identified as Pantothenic acid by reference^[32]. Compound 64 (t_R=21.37min) was given a molecular ion peak m/z 279.23291 in the negative ion mode, and its molecular formula was inferred to be C₁₈H₃₂O₃ and fragment ion 261.22281 [M-H-H₂O]⁻, which was identified as linoleic acid by reference^[33]. Compound 68 (t_R=22.45min) gives a molecular ion peak m/z 255.23296 in the negative ion mode, and its molecular formula is inferred to be C₁₆H₃₂O₂ and fragment ion 237.22198 [M-H-H₂O]⁻, which is identified as palmitic acid by reference^[34].

2.2.3. Identification of flavonoids

According to MS information, a total of 9 flavonoids were detected, taking compounds 26, 30 and 35 as examples. Compound 26 (t_R=11.53min) was given a molecular ion peak m/z 431.09836 in the negative ion mode, and its molecular formula was inferred to be C₂₁H₂₀O₁₀ and fragment ion 269.04517 [M-H-C₆O₅H₁₀]⁻, which was identified as genistin by comparison with reference materials^[35]. Compound 30 (t_R=12.4 min) gives a molecular ion peak m/z 255.0652 in the positive ion mode, and its molecular formula is inferred to be C₁₅H₁₀O₄. Fragment ions 227.07022 [M+H-CO]⁺, 199.07542 [M+H-2CO]⁺ and 137.02338 [M+H-H₂O-C₈H₄]⁺ were identified as daidzein by comparison with reference^[36]. Compound 35 (t_R=13.35min) gives a molecular ion peak m/z 269.04547 in the negative ion mode, and its molecular formula is inferred to be C₁₅H₁₀O₅. Fragment ion 241.05162 [M-H-CO]⁻, 240.04305 [M-H-CHO]⁻, 225.05518 [M-H-CO₂]⁻, 213.05608 [M-H-2CO]⁻ and 197.06015 [M-H-CO₂-CO]⁻. According to the literature^[37] and the comparison of control products, it was identified as genistein.

2.2.4. Identification of nucleoside compounds and amide compounds

According to MS information, 7 nucleoside compounds and 6 amide compounds were detected. Compounds 8, 12, 58, 65 and 73 were taken as examples. Compound 8 (t_R=2.34 min) gives a molecular ion peak m/z 268.10406 in the positive ion mode, and its molecular formula is inferred to be C₁₀H₁₃N₅O₄, fragment ion 136.06178 [M+H-C₅H₉O₄]⁺ and 119.03522 [M+H-C₅H₉O₄-NH₃]⁺. It was

identified as adenosine by comparison with reference^[38]. Compound 12 ($t_R=2.8\text{min}$) was given a molecular ion peak m/z 113.03458 in the positive ion mode, and its molecular formula was inferred to be $C_4H_4N_2O_2$ and fragment ion 96.00798 $[M+H-NH_3]^+$, which was identified as uracil by comparison with reference materials^[39]. Compound 58 ($t_R=19.86\text{min}$) gives a molecular ion peak m/z 280.2638 in the positive ion mode, and its molecular formula is inferred to be $C_{18}H_{33}NO$, fragment ions 263.23645 $[M+H-NH_3]^+$ and 245.22620 $[M+H-NH_3-H_2O]^+$. According to literature^[34], it was identified as linoleamide. Compound 65 ($t_R=21.43\text{min}$) gives a molecular ion peak m/z 282.2792 in the positive ion mode, and its molecular formula is inferred to be $C_{18}H_{35}NO$ and fragment ion 265.25272 $[M+H-NH_3]^+$. According to the literature^[40], it was identified as oleic acid amide. Compound 73 ($t_R=25.11\text{min}$) gives a molecular ion peak m/z 338.342 in the positive ion mode, and its molecular formula is inferred to be $C_{22}H_{43}NO$ and fragment ion 321.31503 $[M+H-NH_3]^+$. Combined with literature^[40], it is identified as erucic amide.

2.2.5. Identification of amino acid compounds and pyrrolidone derivatives

According to MS information, a total of 4 amino acid compounds and 2 pyrrolidone derivatives were detected, taking compounds 7, 9 and 20 as examples. Compound 7 ($t_R = 2.22\text{min}$) gives a molecular ion peak m/z 182.08092 in the positive ion mode, and its molecular formula is inferred to be $C_9H_{11}NO_3$, fragment ion 165.05461 $[M+H-NH_3]^+$ and 136.07568 $[M+H-COOH_2]^+$. It was identified as L-tyrosine^[41]. Compound 9 ($t_R=2.39\text{min}$) gives the molecular ion peak m/z 143.08144 in the positive ion mode, and its molecular formula is inferred to be $C_6H_{10}N_2O_2$. The fragment ions 126.05515 $[M+H-NH_3]^+$, 98.05994 $[M+H-NH_3-CO]^+$ and 70.06517 $[M+H-NH_3-2CO]^+$, combined with references^[42], were identified as Piracetam. Compound 20 ($t_R=7.05\text{min}$) gave a molecular ion peak m/z 171.11284 in the positive ion mode, and its molecular formula was inferred to be $C_8H_{14}N_2O_2$ and fragment ion 126.09127 $[M+H-NH_3-CO]^+$. Combined with literature^[42], it was identified as Levetiracetam.

2.2.6. Identification of steroids and alkaloids

According to MS information, a total of 2 sterols and 2 alkaloids were detected, taking compounds 66 and 77 as examples. Compound 66 ($t_R = 21.77\text{min}$) was given a molecular ion peak m/z 397.34583 in the positive ion mode, and its molecular formula was inferred to be $C_{28}H_{44}O$ and fragment ion 379.33572 $[M+H-CH_2COOH]$. Combined with the reference^[43] and the comparison, it was identified as ergosterol. Compound 77 ($t_R=26.48\text{min}$) was given a molecular ion peak m/z 118.08617 in the positive ion mode, and its molecular formula was inferred to be $C_5H_{11}NO_2$, fragment ions 59.07298 $[M+H-CH_2COOH]$ and 58.06517 $[M+H-CH_3COOH]$. It was identified as betaine^[44].

2.2.7. Other classes

According to MS information, a total of 6 other classes were detected, including 2 sphingolipids, 1 aromatic aldehyde, phenolic, aromatic amine and acid compounds, taking compound 21 as an example. Compound 21 ($t_R = 7.62\text{min}$) gave a molecular ion peak m/z 137.02437 in the negative ion mode, and its molecular formula was inferred to be $C_7H_6O_3$ and fragment ion 93.03452 $[M-H-CO_2]$. Combined with literature^[45], it was identified as salicylic acid.

2.2.8. Orthogonal Partial least squares-Discriminant Analysis (OPLS-DA)

SIMCA 14.1 software was used, based on PCA, and OPLS-DA modeling analysis was shown in Figure 2. The VIP value diagram was used to determine the main difference components of the powder before and after fermentation. The greater the VIP value, the greater the weight value and the stronger the ability to distinguish samples, as shown in Figure 3. Results The R^2X (cum) = 0.947, Q^2 (cum) = 0.998 of the OPLS-DA models of the pre-fermentation and post-fermentation Gastrodia Tudor Halimasch Powder. It is proved that the OPLS-DA model is good and can explain the component difference generally. Projecting importance as a variable (VIP) value >1 shows the difference components: armillarin, Genistein, Linoleic acid, armillaridin, 4',7-Dihydroxyflavanone, 10 α ,13 α -dihydroxyarmillaridin, adenosine, 4'-methoxyarmillasin, 7-(2-aminophenyl)heptanoic acid,

daidzein, 2',5-epoxy-4-dehydroxyarmillaridiene, choline, uracil, 3-Hydroxy-3-methylbutanoic acid, phenacetin, ergosterol, 2-Amino-1,3,4-octadecanetriol, guanine, L-Tyrosine, soyasaponin I, armillaricin, genistin, palmitoleic Acid, nicotinic acid, melleolide, Guanosine, armillaridine, armillaribin, however, due to the limitation of chromatographic conditions, a multi-component determination method was established for the different components of Gastrodia Tuder Halimasch Powder before and after fermentation.

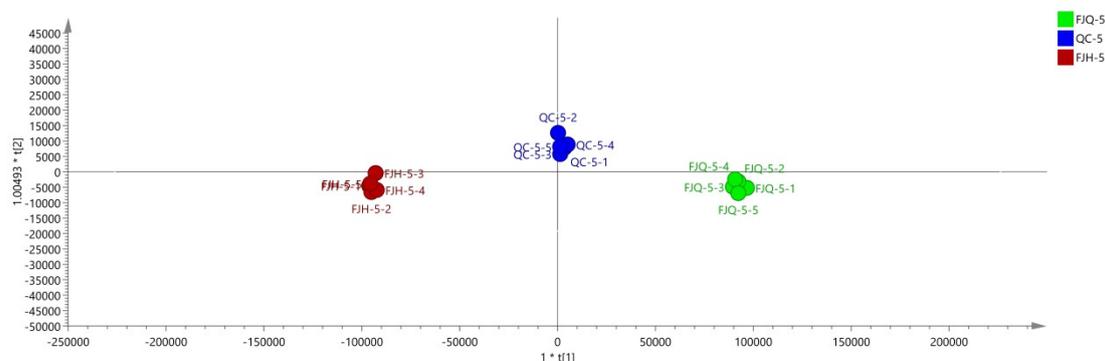


Figure 2. OPLS-DA analysis of Gastrodia Tuder Halimasch Powder (FJY-5) before fermentation and Gastrodia Tuder Halimasch Powder (FJH-5) after fermentation. Note: YL-5-1~5 and FJH-5-1~5 represent 5 parallel experiments.

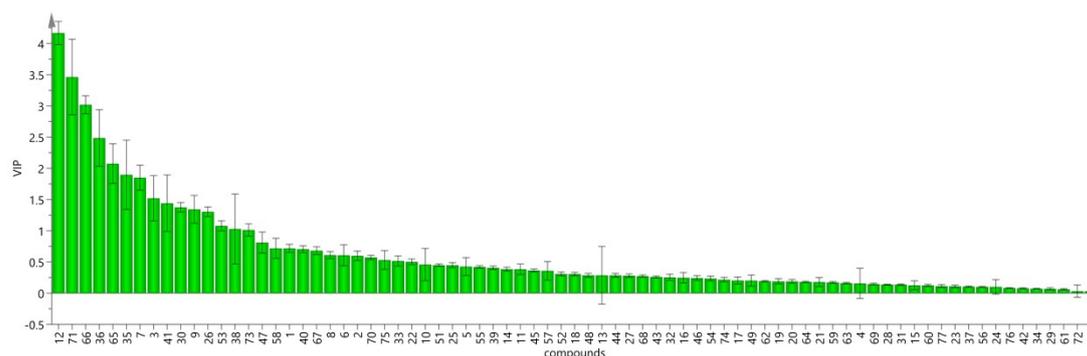


Figure 3. VIP value of Gastrodia Tuder Halimasch Powder before and after fermentation.

2.3. Establishment of a method for the determination of multi-component content of Gastrodia Tuder Halimasch Powder before and after fermentation

2.3.1. Chromatographic conditions

Shimadzu LC-20AD high performance liquid chromatogram (made in Shimadzu, Kyoto, Japan) was performed on Waters Symmetry C18 column (4.6 mm×250 mm, 5 μm) with detection wavelength of 270 nm and flow rate of 1.0 mL•min⁻¹. The column temperature was 30 °C. Gradient elution of methanol (A) -0.1% acetic acid aqueous solution (B) (0~5 min, 40% A; 5~15 min, 40~60% A; 15~30 min, 60~70% A; 30~35 min, 70%~100% A; 35~45 min, 100% A), the sample size was 5 μL. Figure 4.

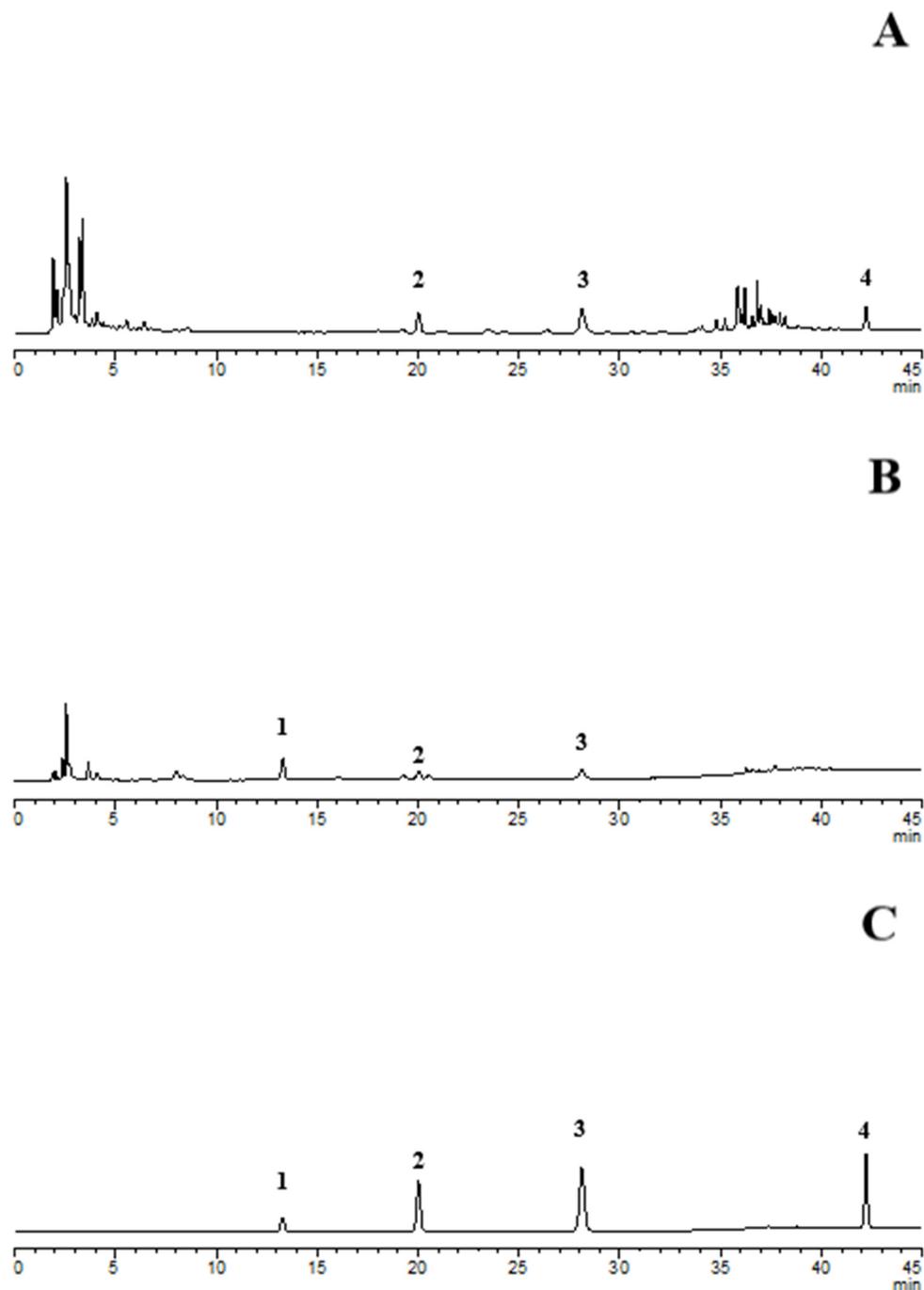


Figure 4. A. Gastrodia Tudor Halimasch Powder after fermentation, B. Gastrodia Tudor Halimasch Powder before fermentation, C-4 mixed label. Note: 1-genistin, 2-daidzein, 3-genistein, 4-ergosterol.

2.3.2. Preparation of reference solution

Take appropriate amount of genistin, daidzein, genistein and ergosterol reference products, weigh them accurately, add methanol to dissolve them, prepare the reference product mother liquor with mass concentrations of 0.1550, 0.6167, 0.4360 and 0.1200 $\text{mg}\cdot\text{mL}^{-1}$, and precisely absorb the appropriate amount of each reference product mother liquor. Mixed reference solution with methanol with mass concentrations of 5.3952, 17.9904, 19.3446 and 47.0368 $\text{ug}\cdot\text{mL}^{-1}$ was filtered by 0.22 μm microporous filter membrane and then prepared for use.

2.3.3. Preparation of test product solution

Take 0.5 g of Gastrodia Tuder Halimasch Powder before and after fermentation, add 5 mL methanol solution, ultrasonic for 1 h, take out, cool to room temperature, make up for weight loss, filter paper, filter through 0.22 μm microporous filter membrane, and then obtain.

2.3.4. Investigation of linear relationship

The mixture of reference solution 0.4, 0.5, 1, 3, 5, 7 μL under item 2.6.2 was precisely absorbed and determined according to the chromatographic conditions under item 2.6.1. The standard curve was drawn with the injection volume as the horizontal coordinate (X) and the peak area as the vertical coordinate (Y), as shown in Table 2.

Table 2. Regression equations, correlation coefficients and linear ranges of the four components.

| Compound | Linear Equation | R ² | Linear Range/ $\mu\text{g}\cdot\text{mL}^{-1}$ |
|------------|--------------------------|----------------|--|
| Genistin | $Y=3682.9592X-745.8583$ | 0.9999 | 2.1581~37.7667 |
| Daidzein | $Y=3519.0934X-1460.4269$ | 0.9999 | 7.1962~125.9328 |
| Genistein | $Y=5724.1012X-3375.2034$ | 0.9999 | 7.7379~135.4125 |
| Ergosterol | $Y=1504.3237X-1830.0128$ | 0.9999 | 18.8147~329.2574 |

2.3.5. Precision test

The same batch of test product solution was taken before and after fermentation, and the relative standard deviation (RSD) of genistin, dzeidin, genistein and ergosterol were calculated as 1.49%, 1.65%, 0.85% and 0.64%, respectively, according to the chromatographic conditions of 3.9.1. The accuracy of the instrument was good.

2.3.6. Stability test

The same batch of test solution was taken before and after fermentation, and the samples were injected at 0, 2, 4, 8, 12 and 24 h after preparation according to chromatographic conditions under item 3.9.1. The RSD of genistin, dzeidin, genistein and ergosterol peak area were 1.56%, 1.63%, 0.49% and 0.67%, respectively. The results showed that the test solution had good stability within 24 h.

2.3.7. Repeatability test

The same batch of Gastrodia Tuder Halimasch Powder was taken before and after fermentation, and 6 sample solutions were prepared in parallel according to the method in item 3.9.3. The average mass fraction of genistin, dzeidin, genistein and ergosterol was calculated as 0.04, 0.14, 0.16 and 0.43 $\text{mg}\cdot\text{g}^{-1}$, respectively, according to the chromatographic conditions in item 3.9.1. RSDs were 1.06%, 1.18%, 1.03% and 2.13%, respectively, indicating good reproducibility of the method.

3.3.8. Sample addition recovery test

The test Gastrodia Tuder Halimasch Powder with known component content before and after fermentation was finely weighed to about 0.25g, 6 parallel parts were added to the control product with the same content as the sample, and the test product solution was prepared according to the method under 3.9.2, and determined according to the chromatographic conditions under 3.9.1. The average recoveries of genistin, daidzein, genistein and ergosterol were 97.50%, 102.97%, 98.65% and 102.87%, and RSD were 0.39%, 0.97%, 0.51% and 0.85%, respectively. See Table 3.

Table 3. Recovery table of 4 components in Gastrodia Tuder Halimasch Powder samples before and after fermentation.

| Compound | Weighing sample (g) | Sample content (mg/g) | Added content (mg/g) | Real measurement (mg/g) | Recovery (%) | Average recovery (%) | RSD value (%) |
|----------|---------------------|-----------------------|----------------------|-------------------------|--------------|----------------------|---------------|
| | 0.2497 | 0.0084 | 0.0083 | 0.0165 | 0.9741 | 97.50% | 0.39% |

| | | | | | | | |
|------------|--------|--------|--------|--------|--------|---------|-------|
| | 0.2503 | 0.0084 | 0.0083 | 0.0165 | 0.9700 | | |
| Genistin | 0.2501 | 0.0084 | 0.0083 | 0.0165 | 0.9753 | | |
| | 0.2498 | 0.0084 | 0.0083 | 0.0165 | 0.9766 | | |
| | 0.2498 | 0.0084 | 0.0083 | 0.0166 | 0.9892 | | |
| | 0.2502 | 0.0084 | 0.0083 | 0.0164 | 0.9649 | | |
| | 0.2501 | 0.0341 | 0.0343 | 0.0692 | 1.0239 | | |
| Daidzein | 0.2503 | 0.0342 | 0.0343 | 0.0685 | 1.0008 | | |
| | 0.2502 | 0.0341 | 0.0343 | 0.0696 | 1.0352 | 102.97% | 0.97% |
| | 0.2501 | 0.0341 | 0.0343 | 0.0690 | 1.0164 | | |
| | 0.2498 | 0.0341 | 0.0343 | 0.0701 | 1.0491 | | |
| | 0.2501 | 0.0341 | 0.0343 | 0.0702 | 1.0527 | | |
| Genistein | 0.2502 | 0.0387 | 0.0389 | 0.0766 | 0.9738 | | |
| | 0.2501 | 0.0387 | 0.0389 | 0.0773 | 0.9915 | | |
| | 0.2502 | 0.0387 | 0.0389 | 0.0767 | 0.9747 | 98.65% | 0.51% |
| | 0.2503 | 0.0388 | 0.0389 | 0.0772 | 0.9874 | | |
| | 0.2502 | 0.0387 | 0.0389 | 0.0773 | 0.9919 | | |
| Ergosterol | 0.2499 | 0.0387 | 0.0389 | 0.0776 | 0.9997 | | |
| | 0.2503 | 0.1081 | 0.1081 | 0.2203 | 1.0384 | | |
| | 0.2501 | 0.1080 | 0.1081 | 0.2192 | 1.0286 | | |
| | 0.2502 | 0.1080 | 0.1081 | 0.2157 | 0.9962 | 102.87% | 0.85% |
| | 0.2501 | 0.1080 | 0.1081 | 0.2199 | 1.0348 | | |
| | 0.2502 | 0.1080 | 0.1081 | 0.2210 | 1.0451 | | |
| | 0.2503 | 0.1081 | 0.1081 | 0.2193 | 1.0290 | | |

2.4. Sample Determination

Prepare Gastrodia Tuder Halimasch Powder before and after fermentation in parallel, 3 parts per batch, prepare the test solution according to the method in item 3.9.3, and determine according to the chromatographic conditions in item 3.9.1, as shown in Table 4. SPSS 26.0 software was used to average the contents of 4 components in each parallel sample, and then PCA was applied to each batch of data. The results showed that the Gastrodia Tuder Halimasch Powder before and after fermentation was divided into 3 groups, the first group before fermentation, the second group after fermentation, and the third group after fermentation, the eighth to the 10th group. SPSS 26.0 software was used to perform LSD variance analysis on the data, and Origin 8.0 was used to draw the histogram of significant difference markers. See Figure 5. The results showed that the contents of daidzein, genistein and ergosterol (not detected before fermentation) increased significantly after fermentation, and the contents of genistin were not detected after fermentation, which may be due to the possible transformation of chemical components during fermentation.

Table 4. Contents of 4 components in Gastrodia Tuder Halimasch Powder samples before and after fermentation.

| Sample | Genistin(mg/g) | Daidzein(mg/g) | Genistein(mg/g) | Ergosterol(mg/g) |
|-----------------------------|----------------|----------------|-----------------|------------------|
| Prefermentation sample | 0.0336 | 0.0184 | 0.0188 | / |
| After fermentation sample 1 | / | 0.1391 | 0.1617 | 0.4653 |
| After fermentation sample 2 | / | 0.1364 | 0.1549 | 0.4318 |
| After fermentation sample 3 | / | 0.1390 | 0.1576 | 0.4371 |
| After fermentation sample 4 | / | 0.1116 | 0.1321 | 0.4392 |
| After fermentation sample 5 | / | 0.1092 | 0.1348 | 0.4359 |
| After fermentation sample 6 | / | 0.1487 | 0.1547 | 0.4368 |
| After fermentation sample 7 | / | 0.1498 | 0.1259 | 0.4472 |
| After fermentation sample 8 | / | 0.1938 | 0.1915 | 0.4248 |
| After fermentation sample 9 | / | 0.1820 | 0.1803 | 0.4342 |

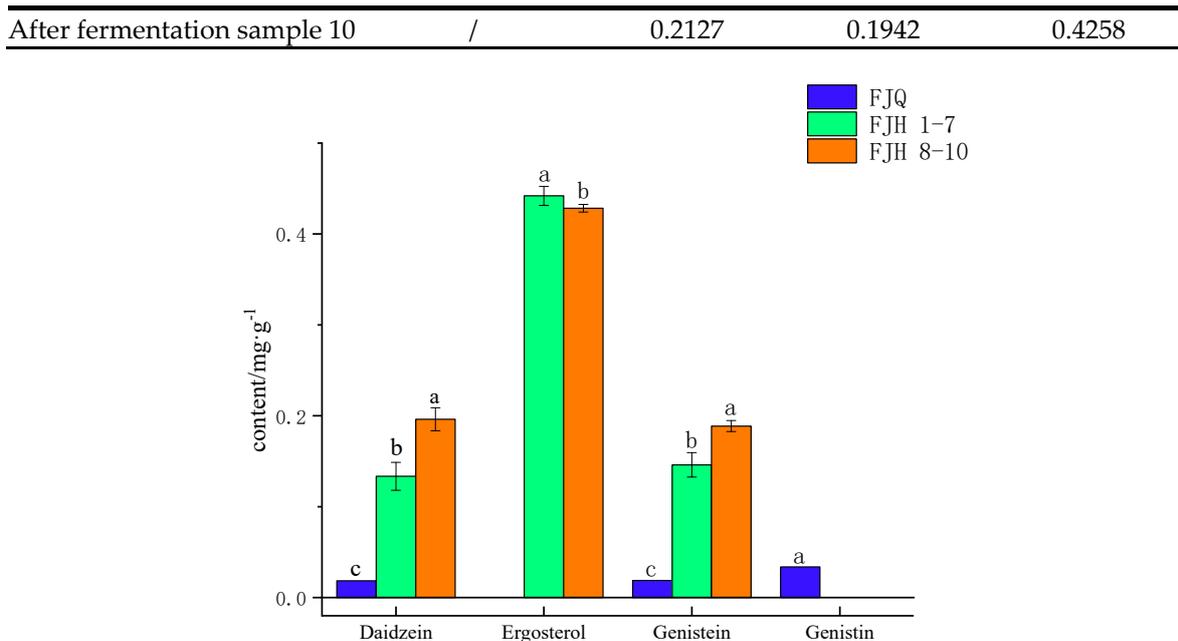


Figure 5. Difference of Gastrodia Tuder Halimasch Powder before and after fermentation.

3. Concluding Remarks

According to the analysis of components of Gastrodia Tuder Halimasch Powder before and after fermentation, due to the large number of chemical components, different compounds have different responses under different modes, so the positive and negative ion modes were respectively scanned, and combined with relevant data such as reference products, PubChem database and references, a total of 77 compounds were speculated and identified. It contains 21 terpenoids, 18 organic acids, 9 flavonoids, 7 nucleosides, 6 amides, 4 amino acids, 2 pyrrolidone derivatives, 2 sterols, 2 alkaloids and 6 other compounds. Combined with VIP>1, the main chemical components of Gastrodia Tuder Halimasch Powder before and after fermentation were further quantitatively analyzed, including genistin, daidzein, genistein and ergosterol. By PCA analysis, the Gastrodia Tuder Halimasch Powder before fermentation, 10 batches of different batches and the Gastrodia Tuder Halimasch Powder after fermentation by the manufacturer were divided into 3 groups, the raw materials were group 1, and the samples 1-7 after fermentation were group 2. After fermentation, samples 8-10 were the third group. Combined with SPSS 26.0 software and LSD analysis of variance, daidzein and genistein were significantly higher than those before fermentation, with daidzein content less than 0.02 mg·g⁻¹ before fermentation and higher than 0.10 mg·g⁻¹ after fermentation. The content of genistein was less than 0.02 mg·g⁻¹ before fermentation and higher than 0.13 mg·g⁻¹ after fermentation. After fermentation, no genistin was detected in Gastrodia Tuder Halimasch Powder, which may be converted to other compounds. Ergosterol was not detected in the Gastrodia Tuder Halimasch Powder before fermentation, which may be due to the formation of a new compound after fermentation. The ergosterol content was higher than 0.42 mg·g⁻¹ after fermentation. This is of great significance for the clinical use of Gastrodia Tuder Halimasch Powder, and this study provides a theoretical basis for the subsequent research on the material basis of Gastrodia Tuder Halimasch Powder. Therefore, ergosterol, genistein and daidzein may be the active ingredients of Gastrodia Tuder Halimasch Powder after fermentation, which can be used as quality control indicators for Gastrodia Tuder Halimasch Powder after fermentation in the future, and provide reference for the quality control of Gastrodia Tuder Halimasch Powder. and the basic research of pharmacodynamic substances.

4. Materials and Methods

4.1. Drugs and reagents

Before fermentation, all the Gastrodia Tuder Halimasch Powder were provided by Luoyang Wokang Pharmaceutical Co., LTD. After fermentation, there were 10 batches of Gastrodia Tuder Halimasch Powder; Luoyang Wokang Pharmaceutical Co., LTD. 7 batches, batch numbers are 220101, 220102, 220103, 220201, 220202, 220203, 220701. Jiangsu Shenhua Pharmaceutical Co., Ltd. 3 batches, batch numbers are 211108, 211109, 211110.

Genistin, genistein, uridine, adenosine reference products (China Institute for Identification of Pharmaceutical and Biological Products, batch number: 111709-200501, 111704-200501, 887-200202, 110879-200202, purity $\geq 98\%$); Ergosterol, ursolic acid, guanosine, uracil (Chengdu Pusi Biotechnology Co., LTD., batch number PS000668, PS000730, PS012671, PS010291, PS020117, purity $\geq 98\%$); Daidzein and cytidine reference products (Shanghai Yuanye Biotechnology Co., LTD., batch number B20227, B20073, purity $\geq 98\%$).

BSA224S-CW type 1/10,000 balance and BT25S type 1/100,000 balance (Sartorius Technology Instrument Co., LTD.); UPT-II-10T ultrapure water device (Chengdu Ultrapure Technology Co., LTD.); KQ-500DV type ultrasonic cleaner (Kunshan Ultrasonic Instrument Co., LTD.), water is Watsons drinking water and laboratory self-made ultra-pure water, acetonitrile, formic acid are mass spectrometry pure, methanol, acetic acid are chromatographic pure, the rest of the reagents are analytical pure.

4.2. Preparation of Gandouling tablets and standard solutions

Precision weigh 0.5g Gastrodia Tuder Halimasch Powder into 100 mL volumetric bottle, add 10 mL 50% methanol, ultrasonic extraction for 1 h, cool, make up for weight loss, filter, and pass 0.22 μ m microporous filter membrane, take 1ml filtrate into 2ml centrifugal tube, put it into high-speed refrigerated centrifuge, then add 500ul methanol to redissolve. Then put it into a high-speed centrifuge at 12000 r \cdot min⁻¹ for 10 min, and take the supernatant to obtain the test product solution.

Accurately weigh the appropriate amount of each reference product, add 50% methanol to prepare the reference product reserve solution with a mass concentration of 1 mg \cdot mL⁻¹, accurately absorb each reference product reserve solution, mix and dilute it into a mixed reference solution with a mass concentration of about 10 μ g \cdot mL⁻¹ for HRMS analysis.

4.3. UHPLC conditions

Using the Ultimate 3000-Orbitrap Exploris 240 LMS (Thermo Fisher Scientific, USA), Hypersil GOLD column (100 \times 2.1mm, 1.9 μ m), The flow rate was 0.3 mL \cdot min⁻¹, the column temperature was 35 $^{\circ}$ C, and the mobile phase was gradient elution with acetonitrile (A) -0.1% formic acid aqueous solution (B) (0~1 min, 2% A; 1~8 min, 2~20% A; 8~14 min, 20~70% A; 14~22 min, 70~95% A; 22~24 min, 95% A; 24~24.5 min, 95%~2% A; 24.5~28 min, 2% A;) The sample size was 2 μ L.

4.4. Mass spectrometry conditions

The scanning range of positive and negative ion detection modes is m/z 100~1200. The positive and negative ion spray voltages are 3.5kV and -3.0kV respectively. The sheath gas flow rate is 25 arb, the auxiliary gas flow rate is 10 arb, and the auxiliary temperature is 350 $^{\circ}$ C. Ion transfer tube temperature 350 $^{\circ}$ C.

4.5. Data processing

Compounds related to Gastrodia Tuder Halimasch Powder were collected by searching PubMed, TCMSP, CNKI and other databases, and the main chemical components were identified according to the chromatographic peak retention time (t_R), secondary fragment ions and reference materials, combined with relevant literature reports.

Author Contributions: Y.W. performed the experiments, analyzed the data, and wrote the first draft of the paper. Z.Z. and J.Z. provided guidance on experimental ideas and structure. S.M. and H.Z. provided experimental materials and suggested experimental methods. H.Z. revised some of the paper content. Y.W., Y.Z., Y.W., J.Y., X.G., L.Y. and S.Z. for data curation. We also thank Shu-Ding Sun for technical support from Public service platform for scientific research of Academy of Chinese Medicine Sciences, Henan University of Chinese Medicine. We also thank Shu-Ding Sun for technical support from Public service platform for scientific research of Academy of Chinese Medicine Sciences, Henan University of Chinese Medicine.

Funding: This research was supported by the Public Welfare Industry Special Project - Construction of Traditional Chinese Medicine Concoction Technology Inheritance Base (Grant No. 38103021-2022) and the 2022 Postgraduate Research Innovation Category Project of Henan University of Traditional Chinese Medicine (Grant No. 2022SHDY009).

Conflicts of Interest: All of the authors have declared that no competing interests exist.

Sample Availability: Samples of the compounds are available from the authors.

References

1. National Pharmacopoeia Commission (Ed.). National Drug Standard Chemical landmark National Standard thirteenth. Beijing: National Pharmacopoeia Commission. 2002:99.
2. Wang R, ZHANG SY, Mu Q. Research progress on chemical constituents and biological activities of *Armillaria*. Chinese herbal medicine. 2016,47(11):1992-1999.
3. Li IC, Lin TW, Lee TY, et al. Oral administration of *Armillaria mellea* mycelia promotes non-rapid eye movement and rapid eye movement sleep in rats. Journal of Fungi. 2021,7(5):371.
4. Geng Y, Zhu S, Cheng P, et al. Bioassay-guided fractionation of ethyl acetate extract from *Armillaria mellea* attenuates inflammatory response in lipopolysaccharide (LPS) stimulated BV-2 microglia. Phytomedicine. 2017,26:55-61.
5. An SS. Study on the pharmacological effects of *Armillaria* polysaccharides on Alzheimer's disease through anti-apoptosis and anti-oxidation. D Jilin University, 2018.
6. Baokui X, Zhang Y. The Prevention and Treatment of Polysaccharide from the Rhizomorph of *Armillaria mellea* on Diabetic Cataract in Rat. Agricultural Science & Technology. 2014,15(7).
7. Zhang T, Du Y, Liu X, et al. Study on antidepressant-like effect of protoilludane sesquiterpenoid aromatic esters from *Armillaria Mellea*. Natural Product Research. 2021,35(6):1042-1045.
8. Li Z, Wang Y, Jiang B, et al. Structure, cytotoxic activity and mechanism of protoilludane sesquiterpene aryl esters from the mycelium of *Armillaria mellea*[J]. Journal of ethnopharmacology. 2016,184:119-127.
9. Dorfer M, Heine D, König S, et al. Melleolides impact fungal translation via elongation factor 2. Organic & biomolecular chemistry. 2019,17(19):4906-4916.
10. Aras A B, Guven M, Akman T, et al. Neuroprotective effects of daidzein on focal cerebral ischemia injury in rats. Neural regeneration research. 2015,10(1):146.
11. Li X, Liu RZ, Lin YF, et al. Protective effect of daidzein on cerebral ischemia-reperfusion injury in rats by inhibiting inflammatory response. When Zhen Chinese medicine. 2014,25:6-8.
12. Cheong S H, Furuhashi K, Ito K, et al. Daidzein promotes glucose uptake through glucose transporter 4 translocation to plasma membrane in L6 myocytes and improves glucose homeostasis in Type 2 diabetic model mice. The Journal of nutritional biochemistry. 2014,25(2):136-143.
13. Park M H, Ju J-W, Kim M, et al. The protective effect of daidzein on high glucose-induced oxidative stress in human umbilical vein endothelial cells. Zeitschrift für Naturforschung C. 2016,71(1-2):21-28.
14. Westmark C J. A hypothesis regarding the molecular mechanism underlying dietary soy-induced effects on seizure propensity. Frontiers in Neurology. 2014,5:169.
15. Li Q, Zheng YJ, Zhao MS, et al. The signaling mechanism of genistein against A β neurotoxicity and its research progress. Chinese Journal of Gerontology. 2017,37:3086-3089.
16. Babu P V A, Si H, Fu Z, et al. Genistein prevents hyperglycemia-induced monocyte adhesion to human aortic endothelial cells through preservation of the cAMP signaling pathway and ameliorates vascular inflammation in obese diabetic mice. The Journal of nutrition. 2012,142(4):724-730.
17. El-Kordy E A, Alshahrani A M. Effect of genistein, a natural soy isoflavone, on pancreatic β -cells of streptozotocin-induced diabetic rats: Histological and immunohistochemical study. Journal of microscopy and ultrastructure. 2015,3(3):108-119.

18. Hu WM, Zhang L, Li LZ, et al. Mechanism of genistein inhibition of 3T3-L1 cell adipogenic differentiation. *Food science*. 2016,37:219-224.
19. Kushairi N, Tarmizi N a K A, Phan C W, et al. Modulation of neuroinflammatory pathways by medicinal mushrooms, with particular relevance to Alzheimer's disease. *Trends in Food Science & Technology*. 2020,104:153-162.
20. Xiong M, Huang Y, Liu Y, et al. Antidiabetic activity of ergosterol from *Pleurotus ostreatus* in KK-Ay mice with spontaneous type 2 diabetes mellitus. *Molecular Nutrition & Food Research*. 2018,62(3):1700444.
21. Rangsinth P, Sharika R, Pattarachotanant N, et al. Potential Beneficial Effects and Pharmacological Properties of Ergosterol, a Common Bioactive Compound in Edible Mushrooms. *Foods*,2023,12(13):2529.
22. Sillapachaiyaporn C, Mongkolpobsin K, Chuchawankul S, et al. Neuroprotective effects of ergosterol against TNF- α -induced HT-22 hippocampal cell injury. *Biomedicine & Pharmacotherapy*. 2022,154:113596.
23. Zhou LS. Observation of therapeutic effect of *Gastrodia Armillaria* tablet on 100 cases of neurasthenia and hypertension. *New Journal of Medicine and Pharmacy*. 1978(10):13.
24. Dai HJ, Meng WW, Wang XQ, et al. Observation of curative effect of compound *Gastrodia Armillaria* tablet in treatment of tension-type headache. *Chinese general clinic*. 2008,24:365-366.
25. Gong HQ. Study on the hypoglycemic effect of *Armillaria polysaccharide* by oral administration and its mechanism. D Northeast Normal University, 2018.
26. Hu WD, Wang SY, Xu AC, et al. Characterization and identification of chemical components of *psoralea* based on UHPLC-Q-TOF-MS technique. *Chinese journal of traditional Chinese Medicine*. 2023,48:2989-2999.
27. Dai SY, Cui YF, Xu J, et al. UHPLC-Q-Exactive Orbitrap MS/MS Comparative analysis of alkaloids in *Aconite*, *Aconite* and *aconite*. *Chinese journal of traditional Chinese Medicine*. 2023,48:126-139.
28. Yang J, Yuwu C, Xiaozhang F, et al. Chemical constituents of *Armillaria mellea* mycelium I. Isolation and characterization of armillararin and armillaridin. *Planta medica*. 1984,50(04):288-290.
29. Yang JS, Cong PZ. Study of sesquiterpene alcohol-aromatic acid esters in mycelia of *Armillaria* by mass spectrometry. *Chemical journal*. 1988(11):1093-1100.
30. Yang J, Chen Y, Feng X, et al. Isolation and structure elucidation of armillaricin1. *Planta medica*, 1989,55(06):564-565.
31. Liang N, Cai P, Wu D, et al. High-speed counter-current chromatography (HSCCC) purification of antifungal hydroxy unsaturated fatty acids from plant-seed oil and *Lactobacillus* cultures. *Journal of agricultural and food chemistry*. 2017,65(51):11229-11236.
32. Serrano-García I, Hurtado-Fernández E, Gonzalez-Fernandez J J, et al. Prolonged on-tree maturation vs. cold storage of Hass avocado fruit: Changes in metabolites of bioactive interest at edible ripeness. *Food Chemistry*. 2022,394:133447.
33. Li J, Wang YW, Zhang Q, et al. Qualitative and quantitative analysis of Xiaoer Jiegan granules based on HPLC-Q-Exactive-MS and HPLC-MS/MS techniques. *Chinese Journal of Hospital Pharmacy*. 2023, 43: 868-876.
34. Dong F, Li ZX, Jia CM, et al. Composition analysis of sesame oil based on UPLC/Q-TOF MS/MS. *Chinese grease*. 2022,47(10):130-136.
35. Fang G, Zhang P, Ye XL, et al. Analysis of isoflavone glycosides and their aglycans by electrospray ion trap mass spectrometry. *Journal of Second Military Medical University*. 2013,34:1108-1115.
36. Chen X, Huang ZF, Liu YH, et al. Identification of metabolites of dangerolone capsules in vivo based on UHPLC-Q/Orbitrap-MS/MS. *Chinese journal of traditional Chinese Medicine*. 2022,47:5052-5063.
37. Zhao WJ, Liang YY, Wang ZJ, et al. The metabolites of genistein in rats were identified by UHPLC-LTQ-Orbitrap mass spectrometry. *Journal of mass spectrometry*. 2019,40:109-122.
38. Qin WH, Yang Y, Li Q, et al. Chemical constituents of *Cordyceps sinensis* from Nepal were analyzed based on UPLC-Q-TOF-MS. *Chinese Journal of New Drugs*. 2019,28:1574-1581.
39. Stentoft C, Vestergaard M, Løvendahl P, et al. Simultaneous quantification of purine and pyrimidine bases, nucleosides and their degradation products in bovine blood plasma by high performance liquid chromatography tandem mass spectrometry. *Journal of Chromatography A*. 2014,1356:197-210.
40. Dabur R, Mittal A. Detection and qualitative analysis of fatty acid amides in the urine of alcoholics using HPLC-QTOF-MS. *Alcohol*,2016,52:71-78.

41. Zheng W, Shi HY, Wang P, et al. The chemical components and blood entry components of Banxia Baizhu Tianma Decoction were analyzed by UPLC-Q-Orbitrap-HRMS technique. *Chinese Journal of Hospital Pharmacy*. 2022,42:2331-2339.
42. Zhang ZT. Study on pharmacokinetics and mass spectrum lysis of piracetam. D Shenyang Pharmaceutical University. 2007.
43. Li SM, Feng Y, Zeng X. Determination of ergosterone and ergosterol in Zhuling granules by HPLC-APCI-MS/MS method. *Journal of Pharmaceutical Analysis*. 2014,34(04):649-653.
44. Wood K V, Bonham C C, Miles D, et al. Characterization of betaines using electrospray MS/MS. *Phytochemistry*. 2002,59(7):759-765.
45. Zhang L, Song S, Chen B, et al. Integration of UHPLC/Q-OrbitrapMS-based metabolomics and activities evaluation to rapidly explore the anti-inflammatory components from lasianthus. *Heliyon*. 2023,9(6).

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.