

Review

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Review

The Extremophiles: A Biotechnological Wonder Child and an Evolutionary Relic!

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Simple Summary: Extremophiles are remarkable organisms capable of growing and developing in extreme environments such as volcanic areas, polar regions, deep seas, salt and acidic lakes, deserts, and even space. These organisms therefore play an important role in understanding the limits of life and expanding our knowledge of biology. By studying extremophiles, we can unlock the secrets of their enzymes, proteins, and molecular mechanisms, leading to potential applications in biotechnological and sustainable processes. Furthermore, extremophiles are increasingly being harnessed in biotechnology to produce biofunctional molecules and biopharmaceuticals within the ecosystem-friendly circular economy.

Abstract: This review provides an overview of terrestrial extremophiles, highlighting their adaptive strengths and strategies for coping with environmental challenges through the use of specialized proteins. It also explores why their unique lifestyle and ability to adapt to extreme conditions have become a major focus of research, as well as the main benefits and advancements in the study of these organisms in recent decades. The review aims to present an objective summary of the knowledge acquired and its translation into applied science and biotechnological applications.

Keywords: eukaryotic and prokaryotic extremophile; extremozymes; extreme pressure; anti-freezing proteins; cryoprotection; extreme biotechnology

1. Introduction

Extremophilic organisms, which thrive in environments characterized by extreme conditions, have redefined our understanding of life's resilience and adaptability [1]. Their presence in harsh settings, such as hot springs, deep-sea hydrothermal vents, and hypersaline bodies of water, challenges traditional beliefs about the limits of biological activity and broadens our perspective on potential habitats supporting life on Earth and beyond. (see Table 1) [2,3].

Extremophiles are crucial to our comprehension of adaptive evolution and pivotal in tracing the origins of life on our planet, as their habitats closely resemble early Earth's conditions [4]. From an evolutionary standpoint, studies on extremophiles have revealed that some of these organisms cluster near the universal ancestor on the tree of life. Hyperthermophiles, in particular, appear to be closely related to the origin of all life on Earth, making extremophiles crucial for understanding life's origins [5]. Hence, the study of these organisms provides valuable insights into the environmental conditions and life forms that may have existed during the early stages of Earth's history [1]. Additionally, it suggests that their molecular building blocks—such as proteins, fatty acids, and smaller molecules—must have unique adaptive properties, distinct from those of other organisms.

In addition, their significance extends to astrobiology. The ability of life to adapt and survive in harsh terrestrial conditions suggests the possibility of analogous extremophilic life forms existing on other planets, moons, or even in environments beyond our solar system [6]. Thus, investigating

Earth's extremophiles can shed light on the potential for life beyond Earth. This research has profound implications for identifying possible habitats for life elsewhere in the universe and for developing strategies for exploration and potential colonization of such environments [6–8].

Table 1. Description of extreme conditions for organisms and the biologic stress burden.

Type of extreme condition	Description	Biological consequences
Alkaline or acidic environment	Natural habitats above pH 9 like alkaline/soda lakes, limestone caves and some hot springs, or under pH 5 like volcanic lakes, acidic wetlands, streams and soil and mine drainage, which are persistently, or with regular frequency or for short periods extremely acidic or basic.	Protein denaturation, cell membrane damage, enzyme inactivation, disruption of internal pH balance and altered metabolic processes
Cold	Habitats periodically or consistently below -17 °C either persistently, or with regular frequency or for short periods like mountains, polar sites, and deep ocean.	Cell membrane damage, intracellular ice formation, dehydration, enzymatic inhibition and cellular damage, cold adaptation
Hot	Broadly conceived habitats periodically or constantly in excess of 40 °C either persistently, or with regular frequency or for protracted periods. Volcanic regions and geothermal streams	Dehydration, cell membrane damage, protein denaturation, DNA denaturation, enzyme deactivation and disruption of biological processes.
Hypersaline	Environments with salt concentrations greater than that of seawater, that is, >3.5% like salt lakes and mines.	Osmotic stress, shrinkage of the cell, desiccation, enzyme inactivation
High pressure	Habitats under extreme hydrostatic pressure like aquatic regions more than 2000 meter in the ocean and deep lakes.	Cellular compression, enzyme inactivation, membrane disruption, possible DNA and protein denaturation and cellular adaptation
Radiation	Background radiation beyond the natural average of annual exposure at about 2.4 mSv or 240 mrem.	DNA damage, cell death, carcinogenesis, cell dysfunction and cell cycle arrest
Absence of water	Habitats without free water whether persistently, or with regular frequency or for short periods. Includes hot and cold desert environments, and some endolithic habitats	Dehydration, protein denaturation, cell dysfunction, impaired cellular communication and functions, metabolic inactivity, growth arrest and death.
Absence of light	Unreachable regions for sun light like deep ocean environments and habitats such as caves	Reduced energy production, retarded biological rhythms, dependency to alternative energy source, reduced biodiversity and increased adaptation and specialization
Absence of oxygen	Habitats without free oxygen – whether persistently, or with regular frequency, or for protracted periods. Includes habitats in deeper sediments.	Oxygen deprivation, cellular damage and death. Development of semi- or full anaerobe metabolism.
Absence of nutrients	Areas on earth that lack an abundance of nutrients such as the vast ocean, desert and high country.	nutrient deficiency and energy depletion, organ dysfunction and damages leading to death, or growth arrest
Human made extreme environment	Anthropogenic affected habitats. Including waste depots, mine tailings, oil influenced habitats and pollution by heavy metals or organic compounds.	severe cellular and tissue damage

The study of extremophiles began in the late 19th century with the discovery of microorganisms in hot springs [1]. However, extremophile research gained significant momentum in the latter half of the 20th century [9], propelled by advancements in genetic sequencing and DNA analysis techniques that facilitated a more in-depth exploration of these organisms [10]. Unique proteins and enzymes from extremophiles have since found applications in molecular biology. Extremophilic enzymes—

extremozymes—characterized by their high stability and functionality under extreme conditions, prove valuable for *in vitro* molecular processes requiring high temperatures or other challenging conditions [11]. The discovery of thermoresistant enzymes from extremophiles, for instance, has been instrumental in the development of fundamental techniques of DNA analytics [12].

Extremophiles span both prokaryotic and eukaryotic domains of life, encompassing both uni- and multicellular organisms [13]. The main distinction between prokaryotic and eukaryotic extremophiles lies in their cellular structure and complexity, which influence the type and severity of habitats they can inhabit [1,4]. Prokaryotes, including bacteria and archaea, represent the most common and diverse group of extremophiles. These organisms thrive in a vast array of extreme environments that are inhospitable to humans, such as hot springs, deep-sea hydrothermal vents, and acidic and alkaline regions, which collectively account for approximately 75% of our planet [14]. Prokaryotes’ prevalence in such environments is largely attributed to their simpler cellular structure, genetic flexibility, and adaptability to a wide range of conditions [15] (see Table 2). Taking into consideration that proteins constitute 60 to 70% of any cell’s composition, they must play a crucial role in empowering extremophiles to withstand harsh environmental conditions.

Table 2. Classification of extremophilic organisms due to environmental conditions.

Type of extremophile	Type of environment	Taxonomic families of extremophiles
Psychrophile	Cold environment	Mostly bacteria, archaea and Eukaryotes (algae)
Therrmophile and acidophilic thermophile	Hot or hot acidic environment	Mostly bacteria, archaea and rarely Eukaryotes (fungi and algae)
Halophile or osmophile	High salt/high sugar	Mostly bacteria, archaea and rarely Eukaryotes (fungi and algae)
Acidophile	Acidic environment	Mostly bacteria, archaea and Eukaryotes (algae)
Alkaliphile	Alkaline environment	Mostly bacteria, archaea and Eukaryotes (black fungi and algae)
Barophile or Piezophile	High-pressure environment	Mostly bacteria, archaea and rarely Eukaryotes (single cell protists, deep-sea fish and invertebrates)
Xerophile	Dry environment	Mostly bacteria, archaea and Eukaryotes (fungi)
Radiotolerant	High level of radiating environment	Mostly bacteria, archaea and Eukaryotes (fungi)
Endolithic extremophile	Rocky environment	Mostly bacteria, archaea and Eukaryotes (black fungi, lichens and algae)

Extremophiles have evolved various survival strategies and present a unique ability to influence their environments through nutrient cycling and energy production, thereby contributing to enhanced habitability and biological diversity [16]. As a consequence, these habitats become more hospitable to organisms with lower tolerance and adaptation capabilities, including humans [17,18]. For example, the psychrophilic algae *Chlamydomonas nivalis* survives in extreme cold environments by producing anti-freeze proteins (AFPs), enabling it to maintain photosynthetic activity at sub-zero temperatures [19]. Similarly, the prokaryotic bacterium *Psychrobacter sp.*, isolated from Antarctic ice, is fully adapted to survive in very low temperatures [20]. Both organisms contribute to their ecosystems by providing organic nutrients and oxygen, which are beneficial for heterotrophic organisms.

Eukaryotes, characterized by a true nucleus and membrane-bound organelles within their cells, include several multicellular organisms typically found in moderate or stable environments. However, certain extremophilic eukaryotes exhibit unique adaptations, allowing them to survive in conditions lethal to most other organisms [21]. While multicellular eukaryotic extremophiles – such as those living near deep-sea hydrothermal vents – are relatively recent in evolutionary terms, having originated within the last 100 million years, extremophilic bacteria and archaea trace their ancient origins back several billion years, possibly to the early history of life on Earth [13]. Examples of

extremophilic eukaryotes include the thermophilic fungus *Thermomyces lanuginosus*, which produces heat-resistant enzymes for survival in high temperatures – i.e., reaching up to 80 degrees Celsius [22] and the halophilic algae, *Dunaliella salina* [23], found in highly saline environments such as salt pans and salt lakes. This unicellular organism has evolved specialized mechanisms to cope with the high salt concentrations, including the production of compatible solutes that help maintain cellular osmotic balance and protect against dehydration.

Extremophiles, with their remarkable ability to cope with harsh conditions, represent a reservoir of genetic diversity, offering genes and proteins essential for adaptation. For billions of years, they have significantly expanded the range of environments habitable by life, resulting in profound ecological and global implications, which, looking ahead, also include the potential applications of their macromolecules and compounds [24]. For example, enzymes and AFPs that facilitate biological processes at very low temperatures show promising applications in the food and biotech industries. Proteases, amylases, and lipases, exhibiting optimal activities at cold temperatures, are of enormous interest for establishing novel bioprocessing strategies [20,25]. Due to their versatile properties, extremozymes are widely used in wastewater treatment, bioremediation and biodegradation as part of a green chemistry approach that involves the use of various hydrolases such as esterases, catalases, peroxidases, lipases, laccases, etc [26]. Similarly, hydrolases derived from both bacterial and fungal extremophiles are also effective in the biodegradation of plastic materials [27,28]. Moreover, enzymes from extremophiles, particularly acidophiles and thermophiles, play a role in biomining - an eco-friendly process used for the extraction of metals, also known as bioleaching [29,30]. Considering their diverse applications, including the bioremediation and biodegradation of toxic compounds and waste material, extremophilic microorganisms serve as sustainable sources of novel biomolecules and biotechnological tools that can significantly contribute to a bio-based economy. In this context, extremophiles serve as valuable evolutionary relics that can illuminate our path through the complex train of ecological challenges posed by climate change and human-based interventions and activities such as industrial energy and goods production [31].

Scientific inquiry into extremophiles is still in its early stages. This review article aims to provide a concise overview of the repertoire of extremophilic organisms and the extremoproteins that play a central role in extremotolerance and adaptation, particularly with an eye toward their biotechnological applications. The goal is to explore the potential of this largely untapped consortium of organisms to improve technological processes with a keen awareness of the need to preserve biodiversity and pursue eco-friendly interventions, both on Earth and beyond.

2. Extreme Conditions Triggering the Adaptation

Extreme environments play a significant role in driving the evolution and diversity of extremophiles (Table 1). Extremophiles are organisms that thrive in conditions considered extreme by human standards, such as high or low temperatures, high salinity, extreme pressure, high acidity or alkalinity, and high radiation levels. The mechanisms through which these environments promote the evolution and diversity of extremophiles can be illustrated by the following examples:

Selective Pressure: Extreme conditions impose intense selective pressure, ensuring that only organisms with specific adaptive traits can survive and reproduce. Over time, these pressures can lead to the development of unique characteristics that enable survival in harsh environments. A well-known example is *Taq* Polymerase, a thermostable enzyme that remains functional at high temperatures, crucial for DNA replication in the extreme conditions of hot springs where *Thermus aquaticus* is found. This enzyme guarantees the survival of the organism and has also revolutionized molecular biology [32]. Its discovery significantly simplified the Polymerase Chain Reaction (PCR) process, by enabling the combination of the DNA denaturation and amplification step [33]. Since the advent of *Taq* polymerase, other thermostable polymerases with improved properties, such as higher fidelity and greater processivity, have been discovered. These include enzymes like Pfu polymerase, isolated from the hyperthermophilic archaeon *Pyrococcus furiosus*, that thrives in extremely hot environments, such as hydrothermal vents and various others [34].

Genetic Mutations and Horizontal Gene Transfer: Harsh environments can increase mutation rates, leading to the introduction of new genetic variations [35]. Additionally, horizontal gene transfer (HGT) enables extremophiles to acquire genes from other organisms, potentially conferring them advantageous traits [36,37]. HGT is especially common in prokaryotes, such as bacteria and archaea, which are frequently found in extreme environments [38]. A notable example is the radiophilic bacterium *Deinococcus radiodurans*, which has developed PprA, a DNA protection protein aiding the DNA repair and protection against radiation-induced damage, contributing to the organism's extraordinary radiation resistance [39].

DNA protection proteins have several biotechnological applications, leveraging their ability to protect DNA from damage, degradation, or environmental stresses. These proteins are utilized across various fields of biotechnology and medicine including 1) *Gene Cloning and Expression Systems to stabilize plasmids and other vectors*, preserving the integrity of recombinant DNA during manipulation and expression in host cells [40]; 2) *DNA Preservation in biobanking and forensic science*, these proteins are employed to enhance the stability and longevity of DNA samples [41]; 3) *Gene Therapy by DNA protection proteins*, that play a critical role in gene therapy ap by safeguarding therapeutic genes from degradation once introduced into target cells; This ensures the efficacy and safety of gene delivery systems [42], and 4) *Synthetic Biology* by proteins, which stabilize synthetic genetic circuits and engineered pathways, thereby improving the reliability and performance of engineered organisms [43,44].

Metabolic Diversification: Extremophiles exhibit diverse metabolic pathways that enable them to utilize unconventional energy sources and tolerate extreme conditions [45]. For instance, some extremophiles can metabolize inorganic compounds, such as sulfur, to obtain energy [46,47], demonstrating metabolic flexibility that supports their colonization of extreme environments [48]. Enzymes like sulfide quinone oxidoreductase (SQR) are the game player in the oxidation of sulfur compounds, enabling acidophilic and thermophilic organisms like *Sulfolobus acidocaldarius* to derive energy from inorganic sulfur sources in hot, acidic environments [49]. SQR enzymes have various biotechnological applications: they can be used in bioremediation processes to detoxify and convert sulfur compounds in polluted environments, aiding in the cleanup of contaminated sites by facilitating the reduction of sulfur-containing compounds [50]. Further, SQR enzymes can be utilized in bioenergy production processes, particularly in converting sulfur compounds into biofuels or other energy carriers [51]. Their role in electron transfer and redox reactions is crucial for efficient energy conversion. SQR enzymes are valuable in various industrial processes due to their ability to catalyze redox reactions involving quinones and sulfur compounds. This includes applications in the synthesis of chemicals [52]. Different oxidoreductases are being explored for their potential in biocatalysis, particularly in the synthesis of complex organic molecules. Their ability to perform specific redox reactions makes them valuable in synthetic chemistry.

Symbiotic Relationships: Extremophiles sometimes engage in symbiotic relationships, where two or more species collaborate to support each other's survival in extreme conditions. These interactions can drive co-evolution, resulting in further diversification of extremophiles. Proteins play a critical role in facilitating these symbiotic relationships, enabling extremophiles to thrive in harsh environments by interacting beneficially with other organisms. One example is Nitrogenases, enzyme essential in the symbiosis between extremophiles such as diazotrophic bacteria and plants or algae. Nitrogenase is essential for fixing atmospheric nitrogen into a usable form for the host. This is crucial in extreme environments where nitrogen availability is limited [53]. In agricultural biotechnology, nitrogenases are used to enhance nitrogen fixation in crops, reducing the reliance on synthetic nitrogen fertilizers. Approaches include engineering plants or microorganisms to express nitrogenase or developing microbial inoculants that harbor nitrogen-fixing bacteria, thus improving soil fertility and crop yields [54,55].

Another example are Peroxidases and Superoxide Dismutases, which help extremophiles manage oxidative stress. In symbiotic relationships, particularly in environments with high radiation or reactive oxygen species, extremophiles often have enhanced peroxidase and superoxide dismutase

activities; these enzymes play a protective role for both symbionts [56]. Both enzymes have also been used in biotechnological applications in the field of bioremediation to degrade environmental pollutants including dyes and organic contaminants; bio-electrochemistry to develop biosensors for detecting environmental toxins; biomedical applications and biopharmaceuticals ([57]. These examples illustrate the diverse and impactful roles of peroxidases and superoxide dismutases from extremophiles in various biotechnological fields.

Isolation and Speciation: Extreme environments often serve as isolated niches, limiting gene flow between populations and promoting speciation. Geographic and ecological isolation can result in the evolution of distinct species adapted to specific extreme conditions [58]. Within this context, heat shock proteins (HSPs), such as HSP70 and HSP90, have evolved as crucial adaptive mechanisms. These proteins protect cells from thermal stress by assisting in protein folding and preventing aggregation. In extremophiles, HSPs help them survive extreme temperatures [59]. Both HSPs are widely used to improve the expression and stability of recombinant proteins in various host systems [60]. Their chaperone activity ensures the proper folding of proteins, which is crucial for producing functional proteins in biotechnological applications [61].

Stress Response Mechanisms: Extremophiles often possess specialized stress response mechanisms that protect them from the damage caused by extreme conditions. For example, extremophiles in high-radiation environments may have efficient DNA repair systems or protective pigments.

Interestingly, in the most cases, a few proteins are sufficient to guarantee the survival and thriving of extremophilic organisms in extreme habitat [62]. This might be because one or two dominant stress factors such as salt concentration, radiation, heat, or others often characterize extreme environments. These factors can frequently be neutralized by the biofunctionality of a single extremoprotein, allowing the cell or organism to remain viable. Therefore, extreme habitats can be seen as crucial drivers of evolution and adaptation.

3. Extreme Habitat as Key Decision Maker

Extremophiles are primarily classified based on the specific extreme conditions prevalent in their habitats, rather than the type of organism (Table 2).

While prokaryotic organisms can be found in nearly any extreme environment, it is important to note that the study of eukaryotic extremophiles is an evolving field. Ongoing discoveries in this area continue to broaden our understanding of life's adaptability to extreme conditions [6].

Extremophilic eukaryotes can be either unicellular (unicellular) or multicellular [63] encompassing a diverse array of organisms. These include single-celled entities, such as extremophilic protists and fungi, as well as multicellular species, including extremophilic algae, extremotolerant fungi, fishes, insects, and even mammals. In the following section, we delve into prominent and representative higher organisms that have evolved in response to particular environmental categories.

In the forefront of coping with hyperthermic conditions, unicellular eukaryotes thriving in extreme environments are not uncommon, and include certain species of archaea and protists. Thermophilic protists, for instance, exhibit remarkable adaptability to extremely high-temperature environments, such as hot springs [64]. Furthermore, certain unicellular extremophilic fungi and algae are capable of surviving and growing in highly acidic conditions, like those found in acid mine drainage sites [65]. Examples include the acidophilic algae *Euglena mutabilis* and various acidophilic fungi [66]. In general, eukaryotes have a lower temperature tolerance compared to prokaryotes or archaea, with the highest reported temperature being 62°C, and most being unable to grow above 50°C [1].

Among extremophilic eukaryotes, algae species hold particular significance. They exhibit autotrophic or mixotrophic behavior, displaying high diversity in metabolism and habitats and various producing organic material in water or soil [67]. Originating from endosymbiosis between a cyanobacteria and an ancient eukaryotic organism over a billion years ago, algae paved the way for

oxygen production and chloroplasts establishment [68]. Red and green algae, which evolved from this ancestral eukaryotic algae, along with a range of other extremotolerant organisms, play a remarkable ecological and evolutionary importance [69]. Furthermore, unicellular algae serve as excellent model systems for experimental simulations and applied studies in laboratory settings [70]. Unicellular algal cultures react by producing relatively homogeneous populations compared to other cell types under stable environmental conditions, such as light or nutrient concentration, etc.. Their rapid growth—particularly compared to plant cells—makes them well-suited for large-scale bioproduct formation, including pigments, nutraceuticals and even biofuel [71].

The thermoacidophilic red algae *Galdieria sulphuraria* thrives in highly acidic environments (pH 0) and temperatures up to 56°C. This organism employs a combination of photosynthetic and heterotrophic growth, utilizing various carbon sources while exhibiting tolerance to high levels of activated oxygen and heavy metals. Depending on the strain, *G. sulphuraria* possesses a compact genome (13.1 to 16.0 Mb) distributed across 72 to 73 chromosomes, suggesting a robust mechanism for adaptability through a large number of chromosomes relative to its genome size [72]. The exceptional adaptability and resilience of *G. sulphuraria* underscores the potential of unicellular eukaryotes to address environmental challenges such as Acid Mine Drainage (AMD), where mining activities lead to major hydrological and geochemical issues and reduced biodiversity [73]. AMDs can contaminate soil, surfaces and ground sediments by high concentrations of sulfates and heavy metals in water. Unicellular eukaryotes, including algae such as *Chlamydomonas* or *Euglena*, alongside protozoa and multicellular protists, therefore demonstrate biotechnological potential for environmental remediation [74].

The reduction of biodiversity and selection pressure are not only potential outcomes of human-caused ecological changes. Climate change-induced abiotic alterations, such as those leading to frost, heat and increased salinity shock, can also impact the community composition of unicellular eukaryotes (plankton) in hydro-ecosystems [75]. Experimental simulations conducted across different sea regions worldwide, e.g. warm and cold seas, revealed dynamic changes in plankton community composition in response to heat, consistent with findings from previous studies [76]. While increased tolerance to salinity could not reconfirm previous assumptions about lowered biodiversity [77], it appears that high diversity within the local community composition of plankton is only facilitating the enrichment of high salinity tolerant members, provided they are present in the population [75], rather than being driven by adaptability alone. Noteworthy, increased temperature or salinity have served as two long-standing stressors over millions of years of evolution, shaping different local planktic communities. In the case of heavy metals exposure, dynamic tolerance has been observed in the unicellular eukaryote protozoa *Tetrahymena thermophila* to titaniumoxide (TiO₂) particles, as demonstrated by proteomics studies [78]. Various metabolic pathways were found to be significantly altered in response to different TiO₂ concentrations, with no apparent toxic effects even at high amounts of heavy metals. These findings highlight the physiological adaptability within one single species and suggest promising applications for heavy metal pollution sanitation and waste management through the internalization of TiO₂ particles by phagocytic activity [79].

Another notable example of unicellular extremophile is the red alga *C. merolae*, recognized as one of the first fully sequenced algal genome and the one complete eukaryotic genome assembled without gaps [80–82]. Its nuclear genome, in terms of the number of protein-coding genes and RNA genes, is comparable to biotechnologically established yeasts such as *Saccharomyces cerevisiae*. *C. merolae* harbors genes crucial for photosynthesis, chloroplasts biogenesis and regulation, making it a promising host cell candidate for the production of bioproducts like pigments, feed and proteins [83]. This extremophile grows in acidic seawater-nutrient media, enabling nonsterile fermentations with low risk of contamination [83]. Various transformation techniques have been developed or are ongoing under development to enhance DNA introduction into *C. merolae*, driven by the increasing demand for low-cost protein-based pharmaceutical. This includes vaccine production within biomanufacturing platforms, which represents a huge challenge, predominantly for mucosal vaccines [84]. Photosynthetic microalgae such as *Chlamydomonas*, *Dunaliella*, *Chlorella*, *Haematococcus*

and *Spirulina*, are also being actively explored as efficient host cells for recombinant proteins expression, offering economical and biotechnological advantages over other biofactories [85]. Key benefits for their use in biotechnological processes include the authentic post-translational modifications, biosafety (free from human pathogenic viruses or virus-like particles), high growth rates, availability of genetic engineering tools, and high biosynthetic capacities [77]. In summary, research on unicellular extremophilic algae is paving the way for innovative advancements in the competitive field of biotechnological manufacturing of complex molecules. Additionally, it enhances our understanding of systems biology adaptation to extreme environmental conditions driven by climate change.

Among the most renowned multicellular extremophiles are tardigrades, also known as “Water Bears”. These microscopic, water-dwelling animals, exhibit remarkable resilience to extreme conditions, including extreme temperatures, radiation, and desiccation [86]. Despite being multicellular, tardigrades consist of a relatively small number of cells and serve as model organisms akin to *Caenorhabditis elegans* or fruit flies, offering insights into biological mechanisms, and exhibiting an evolutionary library for coping with diverse extreme conditions, applicable in the biotechnological contexts [87]. Recent research highlights the role of three protein families in tardigrades: Cytoplasmic-, Secreted- and Mitochondrial-Abundant Heat Soluble (CAHS, SAHS, and MAHS) proteins. These proteins, collectively referred to as tardigrade disordered proteins (TDPs), are not found in other organisms [88]. Remarkably, TDPs lack a stable three-dimensional structure, which may allow them to adopt different conformations in the absence of water molecules- such as during freezing, desiccation or exposure to irradiation [89].

Additionally, certain species of some extremophilic insects, particularly those from the order *Coleoptera* and the family *Carabidae*, have adapted to survive in extreme environments such as deserts with high temperatures and low moisture levels, as well as near volcanic vents. These insects have evolved various adaptations to conserve water and regulate their body temperature. Extensive monitoring of *Coleoptera* beetles in Kunashir Island of the Kuril archipelago, Russia, has revealed increasing biodiversity over some decades. Although much of this data is still unpublished, ongoing studies are trying to provide a clearer understanding of these organisms and their adaptability processes. Typically, higher animals exhibit the lower tolerance to extreme environments compared to some lower extremophiles like archaea or fungi; however, *Coleoptera sp.* and *Carabidae sp.* demonstrate a remarkable combination of resistance to desiccation, high temperatures and acidic conditions, making them noteworthy among multicellular extremophiles [90].

Higher animals rarely thrive in extreme environments compared to microorganisms, monocytic plants and fungi. However, certain species, such as the African lungfish *Protopterus annectes*, demonstrate remarkable adaptations. This ancient fish, considered a living ancestor of tetrapods, can endure prolonged drought due to specific branchial proteins, Aquapurine 1 and 3. These proteins enable it to withstand desiccation and rehydration stresses [91]. Similarly, Emperor penguins (*Aptenodytes forsteri*) exhibit interesting adaptations. They dive to depths of up to 500 meters, deeper than any other bird, to hunt their food and cope with water pressure by evolving high-affinity hemoglobin and various physiological strategies [92]. Another example of an extremophilic animal is the vampire squid (*Vampyroteuthis infernalis*), which inhabits depth of up to 1000 meter in the deep sea, where no light penetrates and oxygen levels are extremely low, through highly adaptive strategies, including an exceptionally low metabolic rate and a detritivorous trophic strategy [93]. Lowered metabolism appears to be a key survival strategy in extreme environments, whether in the deep sea or hot and arid desert.

In summary, the expression of specific proteins plays a dominant role in support for extremophilic character of organisms, complemented by smart physiological strategies and the production of low molecular weight compounds like non-canonical amino acids or subunits for biodegradable polymers.

4. Future Direction

The field of extremophilic research is poised to advance through several exciting avenues. One key area of focus will be the discovery and characterization of novel extremophiles from previously unexplored extreme environments, such as deep-sea hydrothermal vents, acidic hot springs, and polar ice caps. Emerging genomic and metagenomic technologies are expected to play a crucial role in uncovering the genetic and metabolic pathways that enable these organisms to thrive in such harsh conditions. Additionally, there is growing interest in the biotechnological potential of extremophiles, including the development of extremozymes for industrial applications, bioremediation of contaminated environments, and the synthesis of novel bioactive compounds. Understanding the mechanisms of extremophilic adaptation could also provide valuable insights into the origins of life on Earth and the potential for life on other planets. As interdisciplinary collaborations expand, integrating microbiology, molecular biology, bioinformatics, and environmental science, the field of extremophilic research is set to progress further, offering new perspectives on the resilience and versatility of life while driving innovative biotechnological developments.

5. Conclusions

Extremophilic organisms, which thrive in extreme environments such as hot springs, deep-sea hydrothermal vents, and hypersaline bodies of water, have greatly expanded our understanding of life's resilience and adaptability. Their existence challenges traditional notions about the boundaries of biological activity and broadens the perspective on potential habitats for life on Earth and beyond. Proteins are central to enabling these organisms to survive and function under such harsh conditions, making extremophiles invaluable for studying adaptive evolution and the origins of life on Earth. Studies have shown that some extremophiles, particularly hyperthermophiles, are closely related to the universal ancestor of all life, providing valuable insights into early Earth's conditions and the evolutionary processes that shaped life.

Proteins, such as heat shock proteins and DNA protection proteins, found in these organisms, play a pivotal role in extremophile survival, offering clues about the molecular mechanisms of early life. Additionally, extremophiles have unique proteins and enzymes, known as extremozymes, which remain stable and functional under extreme conditions. These extremozymes have found diverse applications in molecular biology, industrial processes, bioremediation, biodegradation, and biomining. For instance, thermoresistant enzymes from extremophiles have been instrumental in DNA analytics and other biotechnological processes. Proteins such as Taq polymerase, used in PCR, and sulfur-oxidizing enzymes, used in bioremediation, highlight the practical applications of extremophilic proteins. Furthermore, extremophiles often engage in symbiotic relationships and exhibit distinctive metabolic pathways, allowing them to utilize unusual energy sources and thrive in extreme environments. These interactions and metabolic flexibility are often mediated by specialized proteins, such as nitrogenases in nitrogen-fixing bacteria and peroxidases in organisms managing oxidative stress. These proteins contribute to the ability of extremophiles to colonize and influence their habitats.

Extremophiles represent a reservoir of genetic diversity, offering genes and proteins essential for adaptation to extreme conditions. This genetic diversity has positive ecological and global implications, including potential applications in the food and biotech industries, wastewater treatment, and the development of sustainable biotechnological tools. Proteins that enable extremophiles to metabolize unusual energy sources or tolerate extreme conditions are particularly valuable for such innovations.

In conclusion, the study of extremophiles not only deepens our understanding of life's adaptability and evolution but also holds significant potential for biotechnological advancements and the search for extraterrestrial life. At the core of these discoveries are proteins, which provide the molecular tools that empower extremophiles to thrive in some of the most inhospitable environments on the planet.

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References

1. Rothschild, L.J.; Mancinelli, R.L. Life in Extreme Environments. *Nature* **2001**, *409*, 1092–1101, doi:10.1038/35059215.
2. Cockell, C.S. Vacant Habitats in the Universe. *Trends Ecol Evol* **2011**, *26*, 73–80, doi:https://doi.org/10.1016/j.tree.2010.11.004.
3. Pikuta, E. V; Hoover, R.B.; Tang, J. Microbial Extremophiles at the Limits of Life. *Crit Rev Microbiol* **2007**, *33*, 183–209, doi:10.1080/10408410701451948.
4. Cavicchioli, R.; Charlton, T.; Ertan, H.; Mohd Omar, S.; Siddiqui, K.S.; Williams, T.J. Biotechnological Uses of Enzymes from Psychrophiles. *Microb Biotechnol* **2011**, *4*, 449–460, doi:10.1111/j.1751-7915.2011.00258.x.
5. Martínez-Espinoza, R.M. Microorganisms and Their Metabolic Capabilities in the Context of the Biogeochemical Nitrogen Cycle at Extreme Environments. *Int J Mol Sci* **2020**, *21*, doi:10.3390/ijms21124228.
6. Merino, N.; Aronson, H.S.; Bojanova, D.P.; Feyhl-Buska, J.; Wong, M.L.; Zhang, S.; Giovannelli, D. Living at the Extremes: Extremophiles and the Limits of Life in a Planetary Context. *Frontiers in Microbiology* **2019**, *10*.
7. Mastascusa, V.; Romano, I.; Di Donato, P.; Poli, A.; Della Corte, V.; Rotundi, A.; Bussoletti, E.; Quarto, M.; Pugliese, M.; Nicolaus, B. Extremophiles Survival to Simulated Space Conditions: An Astrobiology Model Study. *Origins of Life and Evolution of Biospheres* **2014**, *44*, 231–237, doi:10.1007/s11084-014-9397-y.
8. Kochhar, N.; I.K, K.; Shrivastava, S.; Ghosh, A.; Rawat, V.S.; Sodhi, K.K.; Kumar, M. Perspectives on the Microorganism of Extreme Environments and Their Applications. *Curr Res Microb Sci* **2022**, *3*, 100134, doi:https://doi.org/10.1016/j.crmicr.2022.100134.
9. Dalmaso, G.Z.; Ferreira, D.; Vermelho, A.B. Marine Extremophiles: A Source of Hydrolases for Biotechnological Applications. *Mar Drugs* **2015**, *13*, 1925–1965.
10. Colman, D.R.; Poudel, S.; Hamilton, T.L.; Havig, J.R.; Selensky, M.J.; Shock, E.L.; Boyd, E.S. Geobiological Feedbacks and the Evolution of Thermoacidophiles. *ISME J* **2018**, *12*, 225–236, doi:10.1038/ismej.2017.162.
11. Mesbah, N.M. Industrial Biotechnology Based on Enzymes From Extreme Environments . *Frontiers in Bioengineering and Biotechnology* **2022**, *10*.
12. Ishino, S.; Ishino, Y. DNA Polymerases as Useful Reagents for Biotechnology – the History of Developmental Research in the Field . *Frontiers in Microbiology* **2014**, *5*.
13. Rampelotto, P.H. Extremophiles and Extreme Environments. *Life* **2013**, *3*, 482–485.
14. Coker, J.A. Recent Advances in Understanding Extremophiles. *F1000Res* **2019**, *8*, doi:10.12688/f1000research.20765.1.
15. Dimitriu, T.; Szczelkun, M.D.; Westra, E.R. Evolutionary Ecology and Interplay of Prokaryotic Innate and Adaptive Immune Systems. *Current Biology* **2020**, *30*, R1189–R1202.

16. Singh, P.; Jain, K.; Desai, C.; Tiwari, O.; Madamvar, D. Microbial Community Dynamics of Extremophiles/Extreme Environment. In: 2019; pp. 323–332 ISBN 978-0-12-814849-5.
17. Grant, P.R.; Grant, B.R.; Huey, R.B.; Johnson, M.T.J.; Knoll, A.H.; Schmitt, J. Evolution Caused by Extreme Events. *Philos Trans R Soc Lond B Biol Sci* **2017**, *372*, doi:10.1098/rstb.2016.0146.
18. Chevin, L.-M.; Hoffmann, A.A. Evolution of Phenotypic Plasticity in Extreme Environments. *Philos Trans R Soc Lond B Biol Sci* **2017**, *372*, doi:10.1098/rstb.2016.0138.
19. Peng, Z.; Liu, G.; Huang, K. Cold Adaptation Mechanisms of a Snow Alga *Chlamydomonas Nivalis* During Temperature Fluctuations. *Front Microbiol* **2020**, *11*, 611080, doi:10.3389/fmicb.2020.611080.
20. Perfumo, A.; Freiherr von Sass, G.J.; Nordmann, E.-L.; Budisa, N.; Wagner, D. Discovery and Characterization of a New Cold-Active Protease From an Extremophilic Bacterium via Comparative Genome Analysis and in Vitro Expression. *Front Microbiol* **2020**, *11*, 881, doi:10.3389/fmicb.2020.00881.
21. Berlemont, R.; Gerday, C. 1.16 - Extremophiles. In: Moo-Young, M.B.T.-C.B. (Third E., Ed.; Pergamon: Oxford, 2011; pp. 203–216 ISBN 978-0-444-64047-5.
22. Maheshwari, R.; Bharadwaj, G.; Bhat, M.K. Thermophilic Fungi: Their Physiology and Enzymes. *Microbiol Mol Biol Rev* **2000**, *64*, 461–488, doi:10.1128/MMBR.64.3.461-488.2000.
23. Santos, F.; Antón, J. Extremophiles: Hypersaline Environments. In: Schmidt, T.M.B.T.-E. of M. (Fourth E., Ed.; Academic Press: Oxford, 2019; pp. 270–275 ISBN 978-0-12-811737-8.
24. Blaser, M.J.; Cardon, Z.G.; Cho, M.K.; Dangl, J.L.; Donohue, T.J.; Green, J.L.; Knight, R.; Maxon, M.E.; Northen, T.R.; Pollard, K.S.; et al. Toward a Predictive Understanding of Earth's Microbiomes to Address 21st Century Challenges. *mBio* 2016, *7*.
25. Furhan, J. Adaptation, Production, and Biotechnological Potential of Cold-Adapted Proteases from Psychrophiles and Psychrotrophs: Recent Overview. *J Genet Eng Biotechnol* **2020**, *18*, 36, doi:10.1186/s43141-020-00053-7.
26. Gunjal, A.; Waghmode, M.; Annasaheb Magar Mahavidyalaya, S.; Patil, N.; Aparna, G.; Meghmala, W.; Neha, P.) *March (2021) Res*; 2021; Vol. 4;.
27. Tesei, D.; Quartinello, F.; Guebitz, G.M.; Ribitsch, D.; Nöbauer, K.; Razzazi-Fazeli, E.; Sterflinger, K. Shotgun Proteomics Reveals Putative Polyesterses in the Secretome of the Rock-Inhabiting Fungus *Knufia Chersonesos*. *Sci Rep* **2020**, *10*, 1–15, doi:10.1038/s41598-020-66256-7.
28. Atanasova, N.; Stoitsova, S.; Paunova-krasteva, T.; Kambourova, M. Plastic Degradation by Extremophilic Bacteria. *Int J Mol Sci* 2021, *22*.
29. Gumulya, Y.; Boxall, N.J.; Khaleque, H.N.; Santala, V.; Carlson, R.P.; Kaksonen, A.H. In a Quest for Engineering Acidophiles for Biomining Applications: Challenges and Opportunities. *Genes (Basel)* 2018, *9*.
30. CA, J. Biomining of Metals: How to Access and Exploit Natural Resource Sustainably. *Microb Biotechnol* **2010**, *10*, 1191–1193.
31. Anwar, U.B.; Zwar, I.P.; de Souza, A.O. Biomolecules Produced by Extremophiles Microorganisms and Recent Discoveries. *New and Future Developments in Microbial Biotechnology and Bioengineering: Microbial Biomolecules: Properties, Relevance, and Their Translational Applications* **2020**, 247–270, doi:10.1016/B978-0-444-64301-8.00012-3.
32. Stetter, K.O. Extremophiles and Their Adaptation to Hot Environments. *FEBS Lett* **1999**, *452*, 22–25, doi:https://doi.org/10.1016/S0014-5793(99)00663-8.
33. Chien, A.; Edgar, D.B.; Trela, J.M. Deoxyribonucleic Acid Polymerase from the Extreme Thermophile *Thermus Aquaticus*. *J Bacteriol* **1976**, *127*, 1550–1557, doi:10.1128/jb.127.3.1550-1557.1976.

34. Takagi, M.; Nishioka, M.; Kakihara, H.; Kitabayashi, M.; Inoue, H.; Kawakami, B.; Oka, M.; Imanaka, T. Characterization of DNA Polymerase from *Pyrococcus* Sp. Strain KOD1 and Its Application to PCR. *Appl Environ Microbiol* **1997**, *63*, 4504–4510, doi:10.1128/aem.63.11.4504-4510.1997.
35. Gifford, D.R.; Bhattacharyya, A.; Geim, A.; Marshall, E.; Krašovec, R.; Knight, C.G. Environmental and Genetic Influence on the Rate and Spectrum of Spontaneous Mutations in *Escherichia Coli*. *Microbiology (Reading)* **2024**, *170*, doi:10.1099/mic.0.001452.
36. Marie, V.; Lin, J. Cannibalistic Viruses in the Aquatic Environment: Role of Virophages in Manipulating Microbial Communities. *International Journal of Environmental Science and Technology* **2016**, *13*, 2097–2104, doi:10.1007/s13762-016-1027-y.
37. Kobras, C.M.; Falush, D. Adapting for Life in the Extreme. *Elife* **2019**, *8*, doi:10.7554/eLife.48999.
38. Polz, M.F.; Alm, E.J.; Hanage, W.P. Horizontal Gene Transfer and the Evolution of Bacterial and Archaeal Population Structure. *Trends Genet* **2013**, *29*, 170–175, doi:10.1016/j.tig.2012.12.006.
39. Daly, M.J. A New Perspective on Radiation Resistance Based on *Deinococcus Radiodurans*. *Nat Rev Microbiol* **2009**, *7*, 237–245.
40. Martínez-Puente, D.H.; Pérez-Trujillo, J.J.; Zavala-Flores, L.M.; García-García, A.; Villanueva-Olivo, A.; Rodríguez-Rocha, H.; Valdés, J.; Saucedo-Cárdenas, O.; Montes de Oca-Luna, R.; Loera-Arias, M. de J. Plasmid DNA for Therapeutic Applications in Cancer. *Pharmaceutics* **2022**, *14*, doi:10.3390/pharmaceutics14091861.
41. Sguazzi, G.; Fasani, G.; Renò, F.; Gino, S. Biobanks: Archives or Resources? Their Secondary Use for Forensic Purposes—A Systematic Review. *Forensic Sciences* **2024**, *4*, 42–61.
42. Uherek, C.; Wels, W. DNA-Carrier Proteins for Targeted Gene Delivery. *Adv Drug Deliv Rev* **2000**, *44*, 153–166, doi:https://doi.org/10.1016/S0169-409X(00)00092-2.
43. Benner, S.A.; Sismour, A.M. Synthetic Biology. *Nat Rev Genet* **2005**, *6*, 533–543, doi:10.1038/nrg1637.
44. Foo, J.L.; Ching, C.B.; Chang, M.W.; Leong, S.S.J. The Imminent Role of Protein Engineering in Synthetic Biology. *Biotechnol Adv* **2012**, *30*, 541–549, doi:https://doi.org/10.1016/j.biotechadv.2011.09.008.
45. Kour, D.; Rana, K.L.; Kaur, T.; Singh, B.; Chauhan, V.S.; Kumar, A.; Rastegari, A.A.; Yadav, N.; Yadav, A.N.; Gupta, V.K. Extremophiles for Hydrolytic Enzymes Productions: Biodiversity and Potential Biotechnological Applications. In *Bioprocessing for Biomolecules Production*; 2019; pp. 321–372 ISBN 9781119434436.
46. Seckbach, J.; Chela-Flores, J. Extremophiles and Chemotrophs as Contributors to Astrobiological Signatures on Europa: A Review of Biomarkers of Sulfate-Reducers and Other Microorganisms. In *Proceedings of the SPIE Optical Engineering + Applications*; 2007.
47. Mangold, S.; Valdés, J.H.; Holmes, D.S.; Dopson, M. Sulfur Metabolism in the Extreme Acidophile *Acidithiobacillus caldus*. *Front Microbiol* **2011**, *2*.
48. Marlow, J.J.; Steele, J.A.; Case, D.H.; Connon, S.A.; Levin, L.A.; Orphan, V.J. Microbial Abundance and Diversity Patterns Associated with Sediments and Carbonates from the Methane Seep Environments of Hydrate Ridge, OR. *Front Mar Sci* **2014**, *1*.
49. Witt, M.; Pozzi, R.; Diesch, S.; Hädicke, O.; Grammel, H. New Light on Ancient Enzymes – in Vitro CO₂ Fixation by Pyruvate Synthase of *Desulfovibrio Africanus* and *Sulfolobus acidocaldarius*. *FEBS J* **2019**, 286.
50. Narayanan, M.; Ali, S.S.; El-sheekh, M. A Comprehensive Review on the Potential of Microbial Enzymes in Multipollutant Bioremediation: Mechanisms, Challenges, and Future Prospects. *J Environ Manage* **2023**, *334*, 117532.

51. Haque, S.; Singh, R.; Harakeh, S.M.; Teklemariam, A.D.; Alharthy, S.A.; Tripathi, S.C.; Singh, R.P.; Hassan, A.A.; Srivastava, N.; Gupta, V.K. Enzymes Based Biocatalysis for the Treatment of Organic Pollutants and Bioenergy Production. *Curr Opin Green Sustain Chem* **2022**.
52. May, S.W. Applications of Oxidoreductases. *Curr Opin Biotechnol* **1999**, *10*, 370–375, doi:10.1016/S0958-1669(99)80067-6.
53. Burns, R.C.; Hardy, R.W. Nitrogen Fixation in Bacteria and Higher Plants. *Mol Biol Biochem Biophys* **1975**, *1*–189, doi:10.1007/978-3-642-80926-2.
54. Neemisha; Kumar, A.; Sharma, P.; Kaur, A.; Sharma, S.; Jain, R. Harnessing Rhizobacteria to Fulfil Inter-Linked Nutrient Dependency on Soil and Alleviate Stresses in Plants. *J Appl Microbiol* **2022**, *133*, 2694–2716, doi:https://doi.org/10.1111/jam.15649.
55. Timofeeva, A.M.; Galyamova, M.R.; Sedykh, S.E. Plant Growth-Promoting Soil Bacteria: Nitrogen Fixation, Phosphate Solubilization, Siderophore Production, and Other Biological Activities. *Plants (Basel)* **2023**, *12*, doi:10.3390/plants12244074.
56. Steimbrück, B.A.; Sartorio, M.G.; Cortez, N.; Albanesi, D.; Lisa, M.; Repizo, G.D. The Distinctive Roles Played by the Superoxide Dismutases of the Extremophile *Acinetobacter* sp. Ver3. *Sci Rep* **2022**, *12*.
57. Tanwir, K.; Amna; Javed, M.T.; Shahid, M.; Akram, M.S.; Ali, Q. Chapter 32 - Antioxidant Defense Systems in Bioremediation of Organic Pollutants. In *Handbook of Bioremediation*; Hasanuzzaman, M., Prasad, M.N.V., Eds.; Academic Press, 2021; pp. 505–521 ISBN 978-0-12-819382-2.
58. Cabej, N.R. 18 - Species and Allopatric Speciation. In *Epigenetic Principles of Evolution*; Cabej, N.R., Ed.; Elsevier: London, 2012; pp. 707–723 ISBN 978-0-12-415831-3.
59. Laksanalamai, P.; Robb, F.T. Small Heat Shock Proteins from Extremophiles: A Review. *Extremophiles* **2004**, *8*, 1–11, doi:10.1007/s00792-003-0362-3.
60. Assenberg, R.; Wan, P.T.; Geisse, S.; Mayr, L.M. Advances in Recombinant Protein Expression for Use in Pharmaceutical Research. *Curr Opin Struct Biol* **2013**, *23*, 393–402, doi:https://doi.org/10.1016/j.sbi.2013.03.008.
61. Saibil, H. Chaperone Machines for Protein Folding, Unfolding and Disaggregation. *Nat Rev Mol Cell Biol* **2013**, *14*, 630–642, doi:10.1038/nrm3658.
62. De Champdoré, M.; Staiano, M.; Rossi, M.; D'Auria, S. Proteins from Extremophiles as Stable Tools for Advanced Biotechnological Applications of High Social Interest. *J R Soc Interface* **2007**, *4*, 183–191.
63. Weber, A.P.M.; Horst, R.J.; Barbier, G.G.; Oesterhelt, C. Metabolism and Metabolomics of Eukaryotes Living under Extreme Conditions. *Int Rev Cytol* **2007**, *256*, 1–34, doi:10.1016/S0074-7696(07)56001-8.
64. Rappaport, H.B.; Oliverio, A.M. Extreme Environments Offer an Unprecedented Opportunity to Understand Microbial Eukaryotic Ecology, Evolution, and Genome Biology. *Nat Commun* **2023**, *14*, 4959, doi:10.1038/s41467-023-40657-4.
65. Aguilera, A. Eukaryotic Organisms in Extreme Acidic Environments, the Río Tinto Case. *Life (Basel)* **2013**, *3*, 363–374, doi:10.3390/life3030363.
66. Nancucheo, I.; Barrie Johnson, D. Acidophilic Algae Isolated from Mine-Impacted Environments and Their Roles in Sustaining Heterotrophic Acidophiles. *Front Microbiol* **2012**, *3*, 325, doi:10.3389/fmicb.2012.00325.
67. Lin, S. Algae. By Linda E Graham and Lee W Wilcox. Upper Saddle River (New Jersey): Prentice Hall. Xvi + 700 p; Ill.; Taxonomic and Subject Indexes. ISBN: 0-13-660333-5. 2000. *Q Rev Biol* **2002**, *77*, 70–71, doi:10.1086/343625.
68. McFadden, G.I. Origin and Evolution of Plastids and Photosynthesis in Eukaryotes. *Cold Spring Harb Perspect Biol* **2014**, *6*, a016105–a016105, doi:10.1101/cshperspect.a016105.

69. Elias, M.; Archibald, J.M. Sizing up the Genomic Footprint of Endosymbiosis. *Bioessays* **2009**, *31*, 1273–1279, doi:10.1002/bies.200900117.
70. Ana A. Ramos [Review] The Unicellular Green Alga *Dunaliella salina* Teod. as a Model for Abiotic Stress Tolerance: Genetic Advances and Future Perspectives. *Algae* **2011**, *26*, 3–20, doi:10.4490/algae.2011.26.1.003.
71. Khan, M.I.; Shin, J.H.; Kim, J.D. The Promising Future of Microalgae: Current Status, Challenges, and Optimization of a Sustainable and Renewable Industry for Biofuels, Feed, and Other Products. *Microb Cell Fact* **2018**, *17*, 36, doi:10.1186/s12934-018-0879-x.
72. Downing, J.M.; Lock, S.C.L.; Iovinella, M.; Davey, J.W.; Mackinder, L.C.M.; Chong, J.P.J.; Ashton, P.D.; Feichtinger, G.A.; James, S.; Jeffares, D.C.; et al. Comparisons between Complete Genomes of the Eukaryotic Extremophile *Galdieria Sulphuraria* Reveals Complex Nuclear Chromosomal Structures. *bioRxiv* **2022**.
73. Clapcott, J.E.; Goodwin, E.O.; Harding, J.S. Identifying Catchment-Scale Predictors of Coal Mining Impacts on New Zealand Stream Communities. *Environ Manage* **2016**, *57*, 711–721, doi:10.1007/s00267-015-0627-5.
74. Hogsden, K.L.; Harding, J.S. Consequences of Acid Mine Drainage for the Structure and Function of Benthic Stream Communities: A Review. *Freshwater Science* **2012**, *31*, 108–120, doi:10.1899/11-091.1.
75. Stefanidou, N.; Genitsaris, S.; Lopez-bautista, J.M.; Sommer, U.; Moustaka-Gouni, M. Unicellular Eukaryotic Community Response to Temperature and Salinity Variation in Mesocosm Experiments. *Front Microbiol* **2018**, *9*.
76. Moustaka-Gouni, M.; Kormas, K.A.; Scotti, M.; Vardaka, E.; Sommer, U. Warming and Acidification Effects on Planktonic Heterotrophic Pico- and Nanoflagellates in a Mesocosm Experiment. *Protist* **2016**, *167*, 389–410, doi:10.1016/j.protis.2016.06.004.
77. Loreau, M.; Naeem, S.; Inchausti, P.; Bengtsson, J.; Grime, J.P.; Hector, A.; Hooper, D.U.; Huston, M.A.; Raffaelli, D.; Schmid, B.; et al. Biodiversity and Ecosystem Functioning: Current Knowledge and Future Challenges. *Science* **2001**, *294*, 804–808, doi:10.1126/science.1064088.
78. Rajapakse, K.; Drobne, D.; Kastelec, D.; Kogej, K.; Makovec, D.; Gallampois, C.; Amelina, H.; Danielsson, G.; Fanedl, L.; Marin\vsek-Logar, R.; et al. Proteomic Analyses of Early Response of Unicellular Eukaryotic Microorganism *Tetrahymena Thermophila* Exposed to TiO₂ Particles. *Nanotoxicology* **2016**, *10*, 542–556.
79. Yang, W.W.; Wang, Y.; Huang, B.; Wang, N.X.; Wei, Z.B.; Luo, J.; Miao, A.J.; Yang, L.Y. TiO₂ Nanoparticles Act as a Carrier of Cd Bioaccumulation in the Ciliate *Tetrahymena Thermophila*. *Environ Sci Technol* **2014**, *48*, 7568–7575, doi:10.1021/es500694t.
80. Merola, A.; Castaldo, R.; Luca, P. De; Gambardella, R.; Musacchio, A.; Taddei, R. Revision of *Cyanidium Caldarium*. Three Species of Acidophilic Algae. *Plant Biosyst* **1981**, *115*, 189–195.
81. Matsuzaki, M.; Misumi, O.; Shin-I, T.; Maruyama, S.; Takahara, M.; Miyagishima, S.-Y.; Mori, T.; Nishida, K.; Yagisawa, F.; Nishida, K.; et al. Genome Sequence of the Ultrasmall Unicellular Red Alga *Cyanidioschyzon Merolae* 10D. *Nature* **2004**, *428*, 653–657, doi:10.1038/nature02398.
82. Nozaki, H.; Takano, H.; Misumi, O.; Terasawa, K.; Matsuzaki, M.; Maruyama, S.; Nishida, K.; Yagisawa, F.; Yoshida, Y.; Fujiwara, T.; et al. A 100%-Complete Sequence Reveals Unusually Simple Genomic Features in the Hot-Spring Red Alga *Cyanidioschyzon Merolae*. *BMC Biol* **2007**, *5*, 28, doi:10.1186/1741-7007-5-28.
83. Hirooka, S.; Tomita, R.; Fujiwara, T.; Ohnuma, M.; Kuroiwa, H.; Kuroiwa, T.; Miyagishima, S. Efficient Open Cultivation of Cyanidialean Red Algae in Acidified Seawater. *Sci Rep* **2020**, *10*, 13794, doi:10.1038/s41598-020-70398-z.
84. Castellanos-Huerta, I.; Gómez-Verduzco, G.; Tellez-Isaias, G.; Ayora-Talavera, G.; Bañuelos-Hernández, B.; Petrone-García, V.M.; Fernández-Siurob, I.; Garcia-Casillas, L.A.; Velázquez-Juárez, G. *Dunaliella Salina* as a Potential Biofactory for Antigens and Vehicle for Mucosal Application. *Processes* **2022**, *10*.

85. Raja, R.; Hemaiswarya, S.; Rengasamy, R. Exploitation of *Dunaliella* for Beta-Carotene Production. *Appl Microbiol Biotechnol* **2007**, *74*, 517–523, doi:10.1007/s00253-006-0777-8.
86. Goldstein, B. Tardigrades and Their Emergence as Model Organisms. *Curr Top Dev Biol* **2022**, *147*, 173–198.
87. Goldstein, B.; Blaxter, M. Tardigrades. *Current Biology* **2002**, *12*, R475, doi:https://doi.org/10.1016/S0960-9822(02)00959-4.
88. Hesgrove, C.; Boothby, T.C. The Biology of Tardigrade Disordered Proteins in Extreme Stress Tolerance. *Cell Communication and Signaling* **2020**, *18*, 178, doi:10.1186/s12964-020-00670-2.
89. Yamaguchi, A.; Tanaka, S.; Yamaguchi, S.; Kuwahara, H.; Takamura, C.; Imajoh-Ohmi, S.; Horikawa, D.D.; Toyoda, A.; Katayama, T.; Arakawa, K.; et al. Two Novel Heat-Soluble Protein Families Abundantly Expressed in an Anhydrobiotic Tardigrade. *PLoS One* **2012**, *7*, e44209, doi:10.1371/journal.pone.0044209.
90. Makarov, K.V.; Sundukov, Yu.N.; Matalin, A. V Ground Beetles (Coleoptera, Carabidae) in Fumarole Fields of Kunashir Island, Kuril Archipelago, Russia. *Acta Zoologica Academiae Scientiarum Hungaricae* **2020**.
91. Chng, Y.R.; Ong, J.L.Y.; Ching, B.; Chen, X.L.; Hiong, K.C.; Wong, W.P.; Chew, S.F.; Lam, S.H.; Ip, Y.K. Molecular Characterization of Aquaporin 1 and Aquaporin 3 from the Gills of the African Lungfish, *Protopterus Annectens*, and Changes in Their Branchial mRNA Expression Levels and Protein Abundance during Three Phases of Aestivation. *Front Physiol* **2016**, *7*.
92. Meir, J.U.; Ponganis, P.J. High-Affinity Hemoglobin and Blood Oxygen Saturation in Diving Emperor Penguins. *J Exp Biol* **2009**, *212*, 3330–3338, doi:10.1242/jeb.033761.
93. Golikov, A. V.; Ceia, F.R.; Sabirov, R.M.; Ablett, J.D.; Gleadall, I.G.; Gudmundsson, G.H.; Hoving, H.J.T.; Judkins, H.; Palsson, J.; Reid, A.L.; et al. The First Global Deep-Sea Stable Isotope Assessment Reveals the Unique Trophic Ecology of Vampire Squid *Vampyroteuthis Infernalis* (Cephalopoda). *Sci Rep* **2019**, *9*.

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