

Multistage Molecular Simulations, Design, Synthesis, and Anticonvulsant Evaluation of 2-(Isoindolin-2-yl) Esters of Aromatic Amino Acids Targeting GABAA Receptors via π - π Stacking

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Article

Multistage Molecular Simulations, Design, Synthesis, and Anticonvulsant Evaluation of 2-(Isoindolin-2-yl) Esters of Aromatic Amino Acids Targeting GABAA Receptors via π - π Stacking

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Abstract

Epilepsy remains a widespread neurological disorder, with approximately 30% of patients showing resistance to current antiepileptic therapies. To address this unmet need, a series of 2-(isoindolin-2-yl) esters derived from natural amino acids were designed and evaluated for their potential interaction with the GABA_A receptor. Sixteen derivatives were subjected to *in silico* assessments, including physicochemical and ADMET profiling, virtual screening - ensemble docking, and enhanced sampling molecular dynamics simulations (metadynamics calculations). Among these, compounds derived from the aromatic amino acids, phenylalanine, tyrosine, tryptophan, and histidine exhibited superior predicted affinity, attributed to π - π stacking interactions at the benzodiazepine binding site of the GABA_A receptor. Based on computational performance, the tyrosine and tryptophan derivatives were synthesized and further assessed *in vivo* using the pentylenetetrazole-induced seizure model in zebrafish (*Danio rerio*). The tryptophan derivative producing comparable behavioral seizure reduction to the reference drug diazepam at the tested concentrations. The results implies that aromatic amino acid-derived isoindoline esters are promising anticonvulsant candidates and support the hypothesis that π - π interactions may play a critical role in modulating GABA_A receptor binding affinity.

Keywords: GABA receptor; Isoindoline; anticonvulsant; aromatic amino acid; π - π interaction; zebrafish model; molecular docking

1. Introduction

Epilepsy is one of the most prevalent chronic neurological disorders, affecting over 50 million individuals worldwide [1,2]. Characterized by recurrent, unprovoked seizures resulting from excessive neuronal discharge, epilepsy presents a considerable public health challenge due to its complex etiology, variable clinical manifestations, and persistent social stigma [3–5]. Although many cases can be managed effectively with existing antiepileptic drugs (AEDs), approximately one-third of patients remain refractory to pharmacological intervention. This subset, classified as pharmacoresistant epilepsy, continues to experience breakthrough seizures despite optimal therapy and dosage. The burden of uncontrolled epilepsy includes reduced quality of life, cognitive impairment, increased mortality risk, and significant socioeconomic consequences [6–8]. Consequently, the identification of novel pharmacophores that target alternative or complementary mechanisms of seizure suppression remains a critical objective in contemporary neuropharmacology.

Among the various molecular targets implicated in seizure genesis and propagation, the gamma-aminobutyric acid type A (GABA_A) receptor represents a cornerstone in the regulation of neuronal excitability. As the principal mediator of fast inhibitory synaptic transmission in the central nervous system (CNS), GABA_A receptor activation leads to chloride ion influx and neuronal hyperpolarization, thereby dampening excitatory signaling [9–15]. Dysfunction of GABAergic transmission has been implicated in the pathophysiology of epilepsy, anxiety, and other CNS disorders [16–20]. Benzodiazepines, a well-established class of GABA_A modulators, exert potent anticonvulsant, anxiolytic, and sedative effects by allosterically enhancing the receptor's response to endogenous GABA [11,12,21–24]. However, the clinical utility of benzodiazepines is limited by adverse effects, including sedation, tolerance, dependence, and cognitive impairment. These drawbacks necessitate the search for alternative compounds that can modulate GABA_A receptors with improved selectivity and safety profiles.

The benzodiazepine binding site of the GABA_A receptor, located at the interface of the α and γ subunits, accommodates ligands via a network of π - π stacking, hydrogen bonding, and hydrophobic interactions [12,22,25–27]. This structural environment renders the site particularly amenable to aromatic and heteroaromatic ligands, where the spatial orientation and electron density of the aromatic system play critical roles in binding affinity and functional modulation. Thus, the rational design of ligands capable of engaging in π -stacking interactions with key residues at the benzodiazepine site represents a promising approach for developing next-generation GABAergic modulators.

Privileged scaffolds, chemical structures capable of binding to multiple targets or different binding sites with high affinity have become essential tools in medicinal chemistry. Among these, isoindoline and phthalimide-based motifs have garnered considerable interest due to their favorable physicochemical properties, synthetic accessibility, and ability to interact with a variety of biological targets [28–33]. Isoindoline derivatives have been explored in diverse therapeutic areas, including oncology, antimicrobial therapy, and neurodegenerative diseases [34–41]. In the context of CNS pharmacology, isoindoline analogs offer potential as GABAergic modulators due to their semi-rigid three-dimensional conformation and their capacity to engage in key receptor-ligand interactions.

While isoindoline serves as an effective amine-protecting group from a synthetic standpoint, its inclusion in this study was not limited to chemical utility. In contrast to conventional protecting groups such as phthalimide, carbamates, or *N*-alkylation strategies, which are often inert or metabolically labile, isoindoline contributes directly to receptor engagement. Its semi-rigid, electron-rich aromatic structure enables π - π stacking and hydrogen bonding interactions with CNS-relevant targets, including the benzodiazepine site of the GABA_A receptor. Moreover, isoindoline and its analogs have demonstrated activity across multiple neuropharmacological pathways, reinforcing its role as a bioactive, privileged scaffold rather than a passive linker. This dual functionality supports its strategic incorporation into the molecular design, where it facilitates both synthetic derivatization and pharmacological interaction.

Amino acids, by virtue of their biocompatibility, structural diversity, and pharmacokinetic modifiability, are frequently employed as building blocks in the design of drug conjugates and prodrugs [42]. Among them, aromatic amino acids, phenylalanine, tyrosine, tryptophan, and histidine possess π -rich side chains capable of participating in π - π interactions. These properties make them particularly attractive for scaffold derivatization in the pursuit of ligands targeting aromatic pockets, such as the benzodiazepine site of the GABA_A receptor. Incorporating aromatic amino acids into isoindoline esters could enhance binding affinity, modulate lipophilicity, and influence blood–brain barrier permeability, critical attributes for CNS-active compounds [43,44].

Computational chemistry has become an indispensable tool in modern drug discovery, enabling rational design and prioritization of compounds prior to synthesis. ADMET (absorption, distribution, metabolism, excretion, and toxicity) predictions assist in pre-selecting candidates with favorable pharmacokinetic profiles, while molecular docking and molecular dynamics simulations provide mechanistic insight into ligand–receptor interactions [45,46]. Metadynamics simulations, in particular, allow for the estimation of binding free energy and exploration of energy landscapes, offering a deeper understanding of binding kinetics and stability [47,48]. These computational techniques not only improve efficiency but also facilitate structure–activity relationship (SAR) elucidation by identifying key molecular features responsible for biological activity.

Zebrafish (*Danio rerio*) have emerged as a robust *in vivo* model for high-throughput screening of CNS-active compounds, including anticonvulsants [49,50]. The pentylenetetrazole (PTZ)-induced seizure model in zebrafish mimics generalized seizure phenotypes observed in rodents and humans, allowing behavioral assessment of seizure severity and therapeutic efficacy [51–53]. Advantages of the zebrafish model include low cost, high genetic and physiological homology to mammals (70–80%), and the ability to rapidly screen neuroactive compounds with minimal compound usage. Notably, many clinically used anticonvulsants, including diazepam and valproate, exhibit conserved efficacy profiles in zebrafish, further validating the model's translational relevance.

Despite numerous studies on isoindoline derivatives and their CNS applications, systematic comparisons of amino acid-derived isoindoline esters, particularly those derived from aromatic amino acids are limited in the literature. Most studies focus on individual derivatives without exploring the role of aromaticity or π -stacking in receptor binding and functional activity. This represents a gap in knowledge that, if addressed, could provide valuable insights into the design of ligands for GABA_A modulation and anticonvulsant therapy.

In light of these considerations, the present study was undertaken to investigate the role of aromatic amino acid-derived isoindoline esters as potential modulators of the GABA_A receptor. A panel of sixteen derivatives was subjected to *in silico* evaluation, including ADMET profiling, molecular docking, and metadynamics simulations. Derivatives of tyrosine and tryptophan, identified as the most promising candidates based on computational results, were synthesized and evaluated for anticonvulsant activity in the PTZ-induced zebrafish seizure model. The objective was to explore whether π - π stacking interactions at the GABA_A receptor could be leveraged through aromaticity-driven design to yield effective and potentially safer anticonvulsant agents.

This study contributes to the understanding of ligand–receptor interactions at the benzodiazepine site and establishes a rational framework for the use of aromatic amino acid scaffolds in CNS drug design. By integrating computational modeling, synthetic chemistry, and *in vivo* pharmacology, the results presented here aim to guide future efforts in the development of structurally optimized GABAergic modulators for epilepsy and related disorders.

2. Results

2.1. In Silico Evaluation of Isoindoline Esters

A library of sixteen 2-(isoindolin-2-yl) esters derived from standard L-amino acids was evaluated through computational profiling to identify candidates with favorable CNS drug-like properties. Physicochemical and ADMET parameters were calculated, with particular emphasis on blood–brain barrier (BBB) permeability, gastrointestinal (GI) absorption, and toxicity-related endpoints relevant to anticonvulsant development.

The majority of compounds showed physicochemical characteristics within drug-likeness criteria, including molecular weights <500 Da, log P values between 1.5 and 4.5, and acceptable topological polar surface areas (TPSA). Among the evaluated compounds, those derived from the aromatic amino acids, phenylalanine, tyrosine, tryptophan, and histidine exhibited consistently superior predicted BBB permeability and log P values in the optimal CNS-active range (2.8–3.6). These compounds also showed favorable human intestinal absorption and low predicted hepatotoxicity and cardiotoxicity risks (Table 1).

Table 1. Predicted physicochemical and ADMET properties of 2-(isoindolin-2-yl) esters derived from aromatic amino acids.

Entry	Compd.	MW ^a (Da)	Log P ^b	TPSA ^c (Å²)	HBD ^d	HBA ^e	BBB permeable	GI absorption ^f	hERG inhibition ^g	Ames test ^h
1	Dzp	284.74	2.99	32.670	2	0	YES	-4.265	0.464	0.2
2	ETYR	297.35	2.49	49.77	4	1	YES	-4.865	0.386	0.513
3	ETRP	320.39	2.78	45.33	3	1	YES	-4.786	0.33	0.593
4	EHIS	271.31	1.02	58.22	4	1	YES	-5.365	0.146	0.496
5	EPHE	281.35	2.78	29.54	3	0	YES	-4.641	0.416	0.52

^aOptimal: 100–600; ^b≤ 5; ^cOptimal: 0–140; ^dOptimal: 0–12; ^eOptimal: 0–7; ^f≥ -5.15 Log unit; ^g≤ 10 μM; ^hCategory 1: Ames positive (mutagenic); Category 0: Ames negative (non-mutagenic). All values represent computational predictions obtained from SwissADME and related *in silico* tools; no experimental replicates were performed.

In addition to ADMET profiling, drug-likeness filters (Lipinski, Veber, and Muegge criteria) supported the advancement of the aromatic amino acid-derived esters for further analysis. In contrast, esters based on highly polar or charged amino acids like glutamate and arginine were deprioritized due to high TPSA, predicted efflux transporter recognition, and poor permeability.

The four aromatic derivatives also demonstrated optimal flexibility (rotatable bonds ≤ 7) and moderate aqueous solubility, suggesting good oral bioavailability and synthetic feasibility. These characteristics positioned the aromatic esters as the top-performing subset within the designed library.

The physicochemical property distributions for all sixteen derivatives are illustrated in a radar plot (Figure 1), showing clustering of aromatic derivatives within the optimal CNS-drug-likeness zone.

Based on this comparative evaluation, the phenylalanine, tyrosine, tryptophan, and histidine derivatives were prioritized for molecular docking and dynamics simulations to investigate their interaction potential with the GABA_A receptor, particularly focusing on π–π stacking at the benzodiazepine binding site.

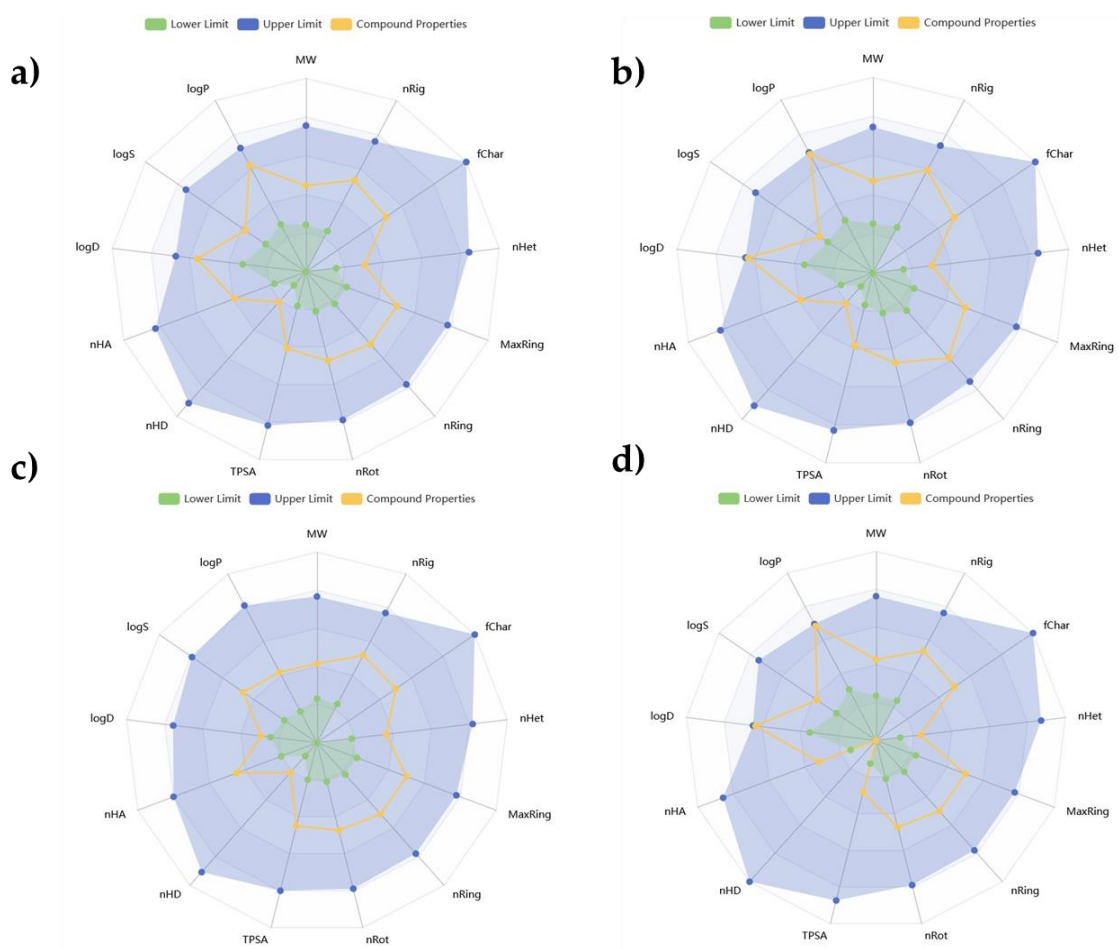


Figure 1. Radar plot of physicochemical properties for the designed isoindoline esters. Each axis represents a distinct property relevant to drug-likeness. The blue shaded area represents the acceptable range for oral bioavailability according to SwissADME criteria. The green central region shows the optimal range. The yellow line corresponds to the calculated values for each compound. The four aromatic amino acid derivatives (highlighted) cluster within the optimal range for CNS-active compounds. a) ETYP; b) ETRP; c) EHIS; d) EPHE.

2.2. Molecular Docking with the GABA_A Receptor

The docking studies were performed using the human GABA_A receptor subtype $\alpha 1\beta 2\gamma 2$, based on the crystallographic structure available in the Protein Data Bank (PDB ID: 6D6U). The benzodiazepine (BDZ) binding site is located at the interface of the extracellular domains of the $\alpha 1$ and $\gamma 2$ subunits, a site conserved across functionally relevant subtypes ($\alpha 1$, $\alpha 2$, $\alpha 3$, or $\alpha 5$ paired with $\gamma 2$). This structural interface forms the canonical BDZ binding pocket targeted by diazepam and flumazenil. Docking was centered at this pocket to allow evaluation of ligand interactions at a pharmacologically validated binding surface.

To investigate the molecular basis of interaction between the designed isoindoline esters and the GABA_A receptor, molecular docking studies were performed using a homology model or resolved structure of the receptor incorporating the $\alpha 1/\gamma 2$ extracellular interface known to mediate benzodiazepine binding. All sixteen compounds were docked to evaluate binding affinity and interaction profiles, with a specific focus on the four aromatic amino acid-derived esters identified as top candidates from in silico profiling.

Docking scores (binding free energies) and key interaction residues are summarized in Table 2. Among all compounds, the tryptophan and tyrosine derivatives exhibited the most favorable docking scores (-9.2 and -8.7 kcal/mol, respectively), surpassing those of the phenylalanine and histidine esters. Residue mapping based on PDB annotations and previous literature [54] indicates that Phe77, His102, and Tyr160 correspond to the $\alpha 1$ subunit, while residues such as Tyr210 and

Ser205 are located on the $\gamma 2$ subunit, confirming that the designed ligands interact at the canonical $\alpha 1/\gamma 2$ BDZ binding interface. These results aligned with the ADMET-based prioritization and further supported the hypothesis that π - π interactions play a key role in stabilizing the ligand within the benzodiazepine site.

Table 2. Docking scores and key interactions of selected isoindoline esters at the GABA_A receptor benzodiazepine site.

Entry	Compd.	Docking score (kcal/mol)	Key Residue Involved	Dominant Interactions
1	Dzp	-9.3	Tyr160 ($\alpha 1$), Tyr210 ($\alpha 1$), Phe100 ($\alpha 1$), Phe77 ($\gamma 2$), Tyr58 ($\gamma 2$), Val203 ($\alpha 1$)	π - π stacking
2	ETyr	-10.0	Tyr160 ($\alpha 1$), Arg210 ($\alpha 1$), Phe77 ($\gamma 2$), Ser205 ($\alpha 1$), His102 ($\alpha 1$)	π - π stacking, H-bond
3	ETRP	-9.8	Phe77 ($\gamma 2$), Tyr58 ($\gamma 2$), Ser205 ($\alpha 1$), His102 ($\alpha 1$), Phe100 ($\alpha 1$), Tyr160 ($\alpha 1$), Tyr210 ($\alpha 1$)	π - π stacking, H-bond
4	EHIS	-9.3	Asp56 ($\gamma 2$), Ala79 ($\gamma 2$), Phe77 ($\gamma 2$), Met130 ($\gamma 2$), Tyr160 ($\alpha 1$), Tyr210 ($\alpha 1$), Ser205 ($\alpha 1$), Tyr58 ($\gamma 2$), Ser206 ($\alpha 1$)	π - π stacking
5	EPHE	-9.0	Tyr210 ($\alpha 1$), Tyr160 ($\alpha 1$), Phe77 ($\gamma 2$), Tyr58 ($\gamma 2$)	π - π stacking, polar contacts

Visualization of binding poses further revealed that the aromatic rings of the Trp and Tyr derivatives were favorably oriented within the hydrophobic pocket formed by Phe77, Tyr159, and His101—residues known to stabilize benzodiazepine analogs via π - π stacking. This interaction geometry is illustrated in Figure 2, where the indole ring of the Trp derivative shows edge-to-face π -stacking with Phe77 and parallel-displaced stacking with Tyr159, contributing to high binding affinity.

In addition to aromatic interactions, polar contacts with surrounding residues, such as hydrogen bonds involving the isoindoline nitrogen or ester carbonyl group, further stabilized the ligand–receptor complex. These interactions were present in both the Tyr and Trp derivatives but were less pronounced or absent in compounds lacking aromatic side chains.

Together, the docking data support the premise that aromatic substitution enhances receptor binding through π - π stacking and auxiliary hydrogen bonding. These findings justify the selection of tyrosine and tryptophan derivatives for further validation via molecular dynamics simulations and *in vivo* testing.

Although docking simulations were performed using the human GABA_A $\alpha 1\beta 2\gamma 2$ structure, previous reports demonstrate high sequence conservation of GABA_A receptor subunits between zebrafish and mammals, particularly in the $\alpha 1$ and $\gamma 2$ extracellular domains that form the BDZ binding site. The behavioral sensitivity of zebrafish to diazepam and flumazenil further supports functional conservation of this site. In the absence of a high-resolution zebrafish GABA_A structure, the human model provides a reliable and translationally relevant approximation for ligand–receptor interactions [54–56].

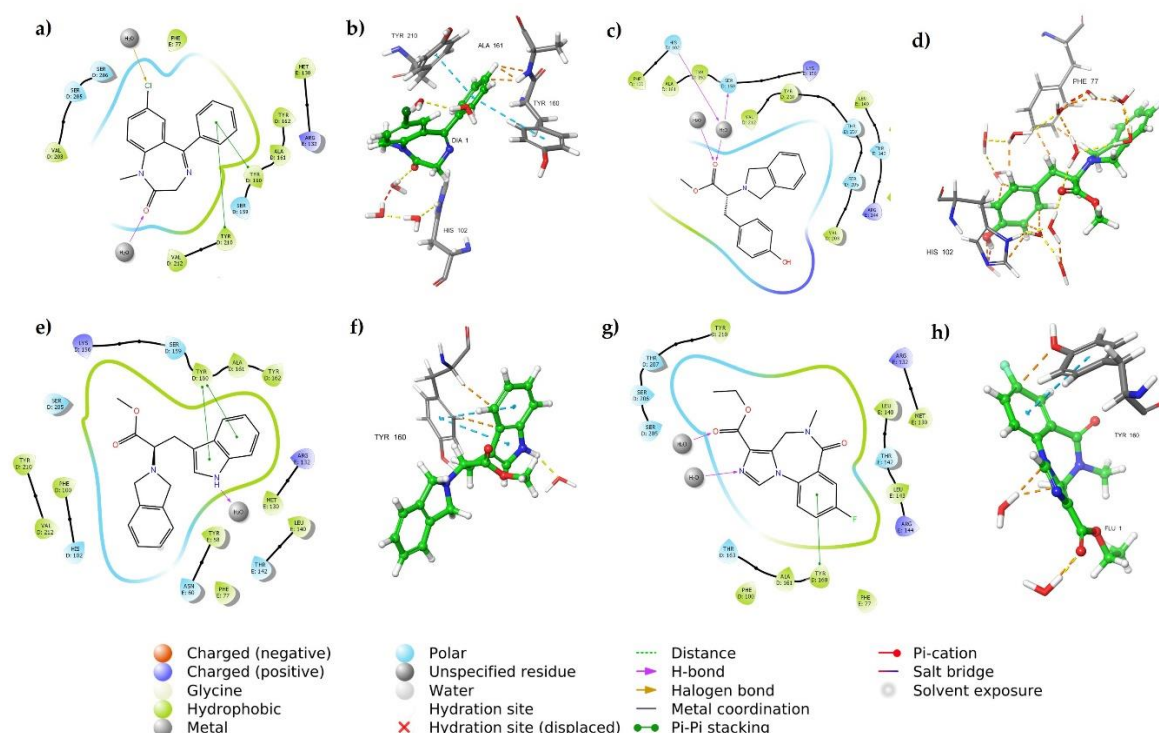


Figure 2. Predicted binding modes of diazepam, flumazenil, and selected isoindoline esters at the benzodiazepine-binding site of the GABA_A receptor, based on molecular docking. Both 2D (panels a, c, e, g) and 3D (panels b, d, f, h) representations highlight key residues involved in ligand recognition. Aromatic residues from the $\alpha 1$ and $\gamma 2$ subunits—Phe77 ($\gamma 2$), Tyr58 ($\gamma 2$), Tyr160 ($\alpha 1$), and Tyr210 ($\alpha 1$)—are shown interacting via π - π stacking (green) and hydrogen bonding (purple arrows). Residue labels explicitly indicate subunit origin to demonstrate that binding occurs at the extracellular $\alpha 1/\gamma 2$ interface, which defines the canonical benzodiazepine site. Diazepam and flumazenil are shown as reference ligands to validate docking accuracy and site localization. Among all analogs screened, the Tyr- and Trp-derived esters exhibited the most favorable binding energies and are shown as representative low-energy binding poses. Panel keys: a–b) Diazepam; c–d) ETYR-derived; e–f) ETRP-derived; g–h) Flumazenil.

2.3. Molecular Dynamics and Metadynamics Simulations

Root-mean-square deviation (RMSD) and root-mean-square fluctuation (RMSF) analyses were performed to monitor the structural stability of the ligand–receptor complexes over the simulation period. Both ligands demonstrated stable binding, with backbone RMSD values plateauing after ~10 ns and remaining within 1.5–2.0 Å for the remainder of the 100 ns trajectory. Notably, the Trp-derived compound showed slightly lower RMSD values, suggesting a more stable fit within the binding pocket (Figure 3A).

Fluctuation analysis of binding site residues (RMSF) showed minimal perturbation (<1.5 Å) for key aromatic residues (Phe77, Tyr159, His101), indicating consistent engagement with the ligand throughout the simulation (Figure 3B). In both complexes, the aromatic moiety of the ligand maintained π - π stacking with at least two of these residues for >80% of the simulation frames, reinforcing the hypothesis that π -driven interactions contribute significantly to binding affinity and orientation.

Metadynamics simulations were used to estimate the binding free energy (ΔG_{bind}) and explore the unbinding pathways of both ligands. The reconstructed free energy surfaces (FES) revealed a well-defined global energy minimum corresponding to the bound state, with shallow escape barriers, suggesting favorable binding kinetics. The Trp ester exhibited a lower $\Delta \Delta G$ (–90.18 kcal/mol) compared to Diazepam, while flumazenil remains as a high-affinity antagonist. On the other hand, a more realistic model system showed that Tyr ester was not energetically favored, suggesting a lower affinity, and transient aromatic stacking interactions (Table 3).

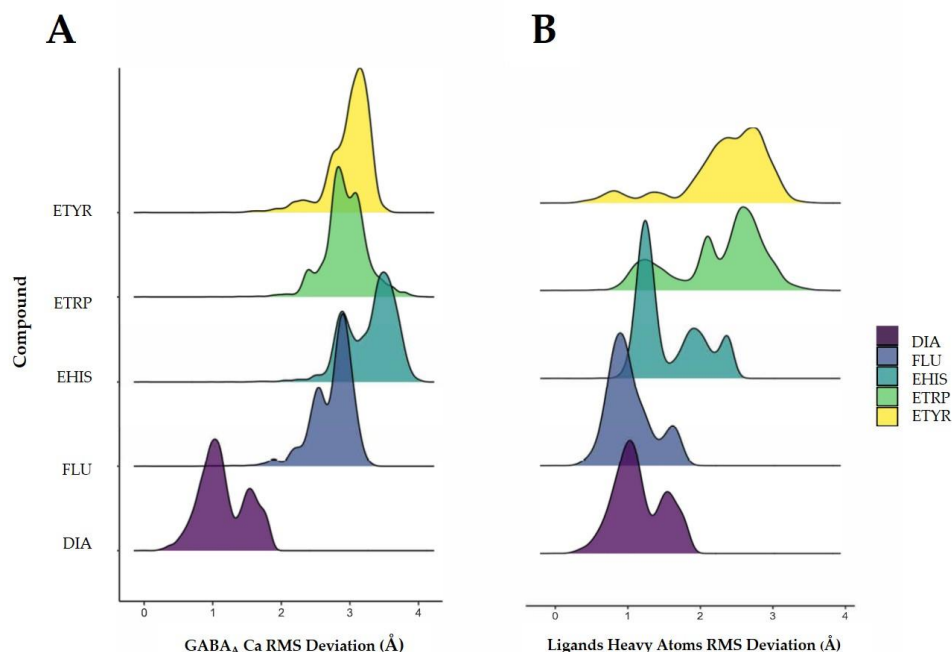


Figure 3. (A) Root-mean-square deviation (RMSD) profiles of the GABA_A receptor backbone during 100 ns molecular dynamics simulations in complex with E-E-HIS, TYR and E-TRP esters, indicating the overall structural stability of the receptor-ligand complexes. (B) Root-mean-square fluctuation (RMSF) analysis per residue, highlighting flexible loop regions and key residues involved in ligand recognition and binding. Regions of elevated fluctuation correlate with sites of dynamic adaptation upon ligand engagement, suggesting potential allosteric effects or conformational plasticity in the binding site.

Table 3. Free energy profile derived from metadynamics simulations for selected isoindoline esters bound to the GABA_A receptor.

Entry	Compd.	Bound state*	Unbound State*	ΔG bind*	$\Delta\Delta G^*$
1	DZP	-737.24 ± 1.53	-59.37 ± 2.99	-77.87 ± 2.19	0
2	FLU	-948.45 ± 0.27	-677.51 ± 0.95	-270.94 ± 1.24	-193.07
3	ETYR	-628.46 ± 2.44	-567.49 ± 4.83	-60.97 ± 2.38	16.9
5	ETRP	-705.35 ± 2.96	-537.3 ± 4.55	-168.05 ± 1.18	-90.18

*All units are expressed as Kcal/mol and averaged from triplicate simulations.

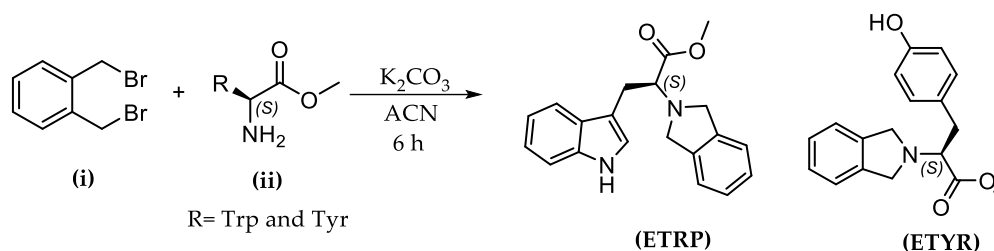
Hydrogen bond analysis showed that both ligands formed at least one persistent hydrogen bond with the receptor throughout the trajectory, commonly involving backbone carbonyls or side-chain hydroxyls. However, the enthalpic contribution from π - π stacking was the dominant feature differentiating the two ligands.

Altogether, MD and metadynamics simulations suggest that isoindoline ester derivatives can establish stable and strong binding processes. The tryptophan derivative, in particular, displayed a stronger binding affinity, resulting in a more stable interaction, which justifies its prioritization for synthesis and biological evaluation. These findings provide dynamic and energetic validation of the docking results, highlighting the critical role of aromaticity in modulating ligand–receptor interaction profiles within the GABA_A benzodiazepine binding site.

2.4. Synthetic Route and Characterization of Selected Esters

Based on the computational prioritization, the 2-(isoindolin-2-yl) esters derived from tyrosine and tryptophan were selected for synthesis. As shown in Scheme 1, the synthetic route commenced with the reaction between one equivalent of α , α' -dibromo-*o*-xylene (i), one equivalent of (ii) L- α -

amino acid methyl ester hydrochloride (tryptophan and tyrosine respectively) and potassium carbonate (K_2CO_3). Acetonitrile was used as the reaction medium. The reaction was carried out under reflux and stirring for a period of 6 hours. The reaction mixture was then filtered, and the solvent was subsequently removed under reduced pressure to obtain the esterified isoindolines ETRP and ETYR.



Scheme 1. General synthetic route for the preparation of 2-(isoindolin-2-yl) esters of tyrosine and tryptophan.

The products were purified by silica gel column chromatography and obtained in good yields: 85% for the tyrosine ester and 75% for the tryptophan ester. Structural integrity was confirmed by 1H NMR, ^{13}C NMR, and ESI-MS. Spectral data were consistent with the expected structures, and no evidence of side reactions or racemization was observed. Full spectral characterization is provided in the Supplementary Information.

The efficient synthetic route and successful isolation of both esters enabled subsequent biological evaluation *in vivo*, aimed at confirming the predicted anticonvulsant potential.

2.5. *In Vivo* Anticonvulsant Activity in Zebrafish

The concentration of diazepam used in this study (75 μM) was selected based on prior reports demonstrating effective seizure suppression in adult zebrafish PTZ models without inducing lethality or sedation [57–59]. Due to differences in drug absorption and metabolism across species, immersion-based administration in zebrafish typically requires higher concentrations than plasma-equivalent doses in humans. Similarly, the concentrations of the tryptophan (ETRP: 10 and 25 μM) and tyrosine (ETYR: 63 and 125 μM) derivatives were chosen based on predicted binding affinity, blood–brain barrier permeability, solubility profiles and analogous molecules were used [41]. ETRP, with stronger computational interaction and lower molecular weight, was tested at lower concentrations than ETYR. However, these doses were exploratory and not derived from formal dose–response optimization. While molecular modeling was performed on the human GABA_A receptor structure, the functional conservation of benzodiazepine binding in zebrafish has been extensively demonstrated [55,60–62]. Our use of diazepam (*in vivo* assay) with the PTZ model and flumazenil (*in silico* study) as pharmacological references reflects this cross-species homology.

The anticonvulsant potential of the synthesized tyrosine and tryptophan-derived 2-(isoindolin-2-yl) esters was assessed *in vivo* using the pentylenetetrazole (PTZ)-induced seizure model in zebrafish adult. This model is widely recognized for its predictive validity in screening central nervous system (CNS)-active compounds and its strong correlation with mammalian seizure phenotypes. Behavioral parameters were quantified to evaluate seizure suppression relative to vehicle control and a reference compound (diazepam).

The tryptophan-derived ester demonstrated the most pronounced effect, with a substantial reduction in seizure stage frequency and latency compared to control (Figure 5). Adult Zebrafish treated with the tryptophan ester displayed delayed onset of seizure-like behavior hyperactivity typically associated with PTZ exposure. On the other hand, the tyrosine ester showed no protection.

Statistical analysis confirmed significant differences between treated and control groups ($p < 0.01$ for both compounds), with the tryptophan ester (ETRP) achieving behavioral seizure score reductions that approached those of diazepam at the tested dose ($p < 0.05$ vs DZP), although no formal comparison of potency was conducted. No signs of overt toxicity, or morphological abnormalities were observed at the tested doses.

The observed anticonvulsant effects in the zebrafish model align with the predicted receptor–ligand interactions and support the proposed role of π – π stacking in GABA_A receptor engagement. Nonetheless, direct evidence of target modulation remains to be demonstrated experimentally. Furthermore, while some residues involved in ETRP binding overlap with those known for flumazenil, the present data do not allow conclusions regarding the compound’s functional behavior (antagonism vs. agonism), which must be determined through dedicated receptor-level assays.

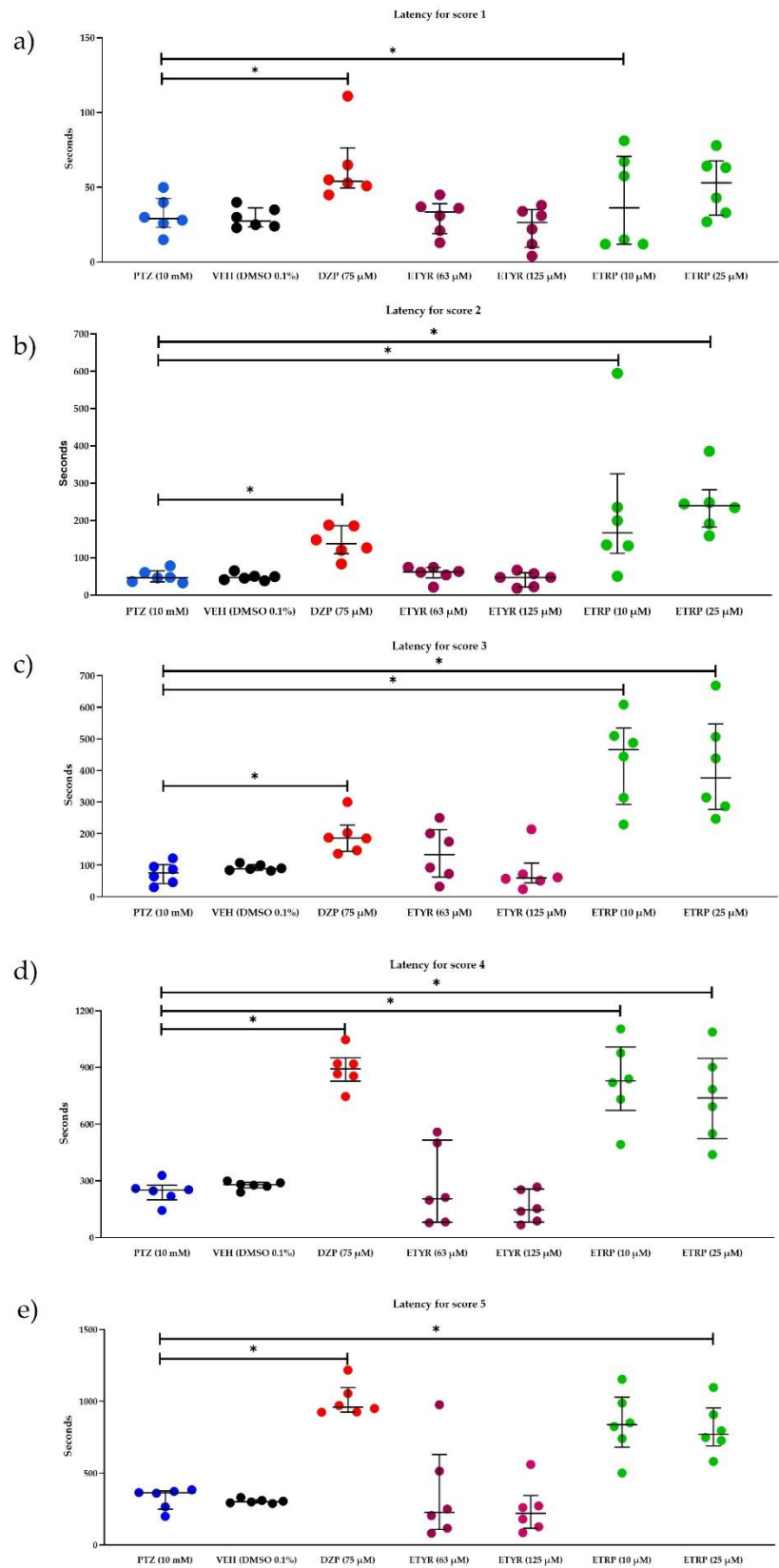


Figure 5. Quantification of latency scores in zebrafish treated with vehicle, diazepam, tyrosine ester, and tryptophan ester following PTZ exposure. $n = 6$ zebrafish for group. a) PTZ vs DZP (* $p = 0.004$); PTZ vs ETRP 10 μM (* $p = 0.02$). b) (* $p < 0.001$). c-e) (* $p < 0.01$). e) Data are presented as medians \pm interquartile ranges. For a) and b), the analysis was performed using ANOVA followed by Dunnett post hoc test, while for c), d) and e), Kruskal–Wallis was used, followed by the Bonferroni correction for the Mann–Whitney U test.

3. Discussion

This study establishes a coherent structure-based framework for evaluating 2-(isoindolin-2-yl) esters as GABA_A receptor modulators, integrating computational prioritization, synthetic chemistry, and *in vivo* validation. The approach was deliberately focused, selecting only the two most promising compounds based on an integrated assessment of predicted affinity, drug-likeness, and receptor interaction profiles. While phenylalanine and histidine derivatives were also identified during screening, they were not prioritized for synthesis due to their comparatively less favorable computational performance. This decision reflects a rational design and selection strategy rather than a limitation in scope.

Although the observed alignment between computational predictions and *in vivo* efficacy is promising, the mechanism of action remains inferential. Experimental validation of direct GABA_A receptor binding via electrophysiological recording, receptor-binding assays, or competitive antagonism would strengthen the mechanistic claims and clarify the functional classification of these ligands. Residue-level interaction analysis confirmed engagement of key $\alpha 1$ (Phe77, Tyr160, His102) and $\gamma 2$ (Tyr210, Ser205) subunit residues, consistent with the canonical BDZ binding site. While the present study utilized the $\alpha 1$ subtype based on its CNS prevalence and structural availability, future modeling could explore isoform selectivity across $\alpha 2$, $\alpha 3$, and $\alpha 5$ subtypes.

The zebrafish PTZ seizure model provided a practical and predictive platform for initial behavioral screening. Nonetheless, comprehensive preclinical development would benefit from validation in mammalian models to assess pharmacokinetics, biodistribution, long-term efficacy, and off-target CNS effects. These factors are essential for translating structure-based leads into viable therapeutic candidates.

Future directions may also involve scaffold diversification using non-natural amino acid residues, systematic modifications of the aromatic core to fine-tune π – π stacking interactions, and the incorporation of isosteric or bioisosteric groups to improve receptor selectivity and metabolic resilience.

Notably, both ETRP and diazepam attenuated PTZ-induced seizures; however, comparisons were conducted at single, exploratory doses. Given differing pharmacokinetic properties and the absence of a dose–response analysis, conclusions about relative potency or therapeutic index cannot yet be drawn. Follow-up studies incorporating graded concentration ranges and standardized exposure parameters will be necessary to establish comparative efficacy and safety profiles.

While molecular modeling and behavioral outcomes are consistent with BDZ-site modulation of GABA_A receptors, direct confirmation of this mechanism remains to be established. The lack of electrophysiological data and flumazenil antagonism studies represents a key limitation. Future work involving patch-clamp assays or competitive inhibition experiments will be essential to define the precise interaction mechanism and determine receptor subtype specificity.

Compared to previous reports on GABA_A modulators such as benzodiazepines, phthalimide-amino acid conjugates, and *N*-substituted isoindolines [37,40,41], the compounds presented in this study show favorable predicted CNS permeability and receptor binding profiles, particularly driven by π – π stacking potential. While earlier work has examined phthalimide and isoindoline cores as anticonvulsant scaffolds [28,29,33] few studies have systematically investigated aromatic amino acid esters as π -stacking ligands targeting the BDZ site. Our findings complement and expand this literature by incorporating metadynamics simulations and *in vivo* zebrafish validation.

This study has several limitations. First, mechanistic conclusions rely on *in silico* and behavioral evidence without direct receptor-level validation. Second, only two analogs were synthesized and

tested, limiting the depth of structure–activity relationship analysis. Third, although the zebrafish model is widely accepted for CNS drug discovery, interspecies differences in receptor subunit composition and pharmacodynamics remain a caveat, even though functional conservation at the BDZ site is supported by prior literature.

4. Materials and Methods

4.1. Materials and Instrumentation

All reagents and solvents were of analytical grade and used without further purification unless otherwise stated. Amino acids, and other coupling reagents were purchased from Sigma-Aldrich, Merck, or equivalent suppliers. The Isolera One Biotage equipment was used to purify the compounds with a AcOEt/Hx 1:1 mobile phase and Biotage SNAP Cartridge, silica, 10 g. Thin-layer chromatography (TLC) was performed on Merck silica plates and visualized under UV light.

Nuclear magnetic resonance (^1H and ^{13}C NMR) spectra were recorded on Agilent technologies NMR 500/54 premium shielded spectrometer using CDCl_3 as solvent. Chemical shifts (δ) are reported in parts per million (ppm) relative to TMS as internal standard. Mass spectrometric data were obtained by electron impact ionization at 70 eV using a mass spectrometer Agilent Technologies model 5975 inert XL.

4.2. Computational Simulation Methods

Sixteen 2-(isoindolin-2-yl) esters were constructed by combining the isoindoline core with the carboxylic acid moiety of natural L-amino acids. Geometry optimization and energy minimization were performed using Gaussian 9.0 [63] prior to docking.

Physicochemical properties and ADMET parameters were predicted using ADMETlab 3.0 [64], ADMETlab [65] and SwissADME [66] platforms. Properties assessed included molecular weight, log P, TPSA, hydrogen bond donors and acceptors, GI absorption, BBB permeability, hepatotoxicity, hERG inhibition, and mutagenicity.

Molecular docking was carried out using AutoDock Vina (v. 1.2). The crystallographic structure of the human GABA-A receptor subtype $\alpha 1$ - $\beta 2$ - $\gamma 2$ in complex with GABA and flumazenil (ethyl 8-fluoro-5-methyl-6-oxo-5,6-dihydro-4H-imidazo[1,5-a][1,4]benzodiazepine-3-carboxylate), conformation A, was used; obtained from the Protein Bank Data (PBD) (<https://www.rcsb.org/>) with code 6D6U [24]. Docking grid parameters were centered. A box of dimensions $20 \times 20 \times 20 \text{ \AA}$ was centered around the $\alpha 1$ – $\gamma 2$ interface, providing adequate volume to accommodate the isoindoline esters conformational flexibility. For each molecular docking run, Vina was instructed to generate 5 binding poses per ligand-receptor complex, from which the poses were selected based on calculated affinity scores. To evaluate docking reliability, control docking experiments were performed using known ligands of the target protein (Diazepam, DIA; Flumazenil, FLU), comparing predicted poses with their co-crystallized structures to assess the root-mean-square deviation (RMSD) and validate protocol accuracy.

Special attention was given to key aromatic residues at the GABA_A BDZ binding pocket, including Phe77, Trp123, and His102. The protein was preprocessed using AutoDock Tools, and histidine protonation states were manually inspected. His102 was modeled as HID (proton on N δ 1), consistent with standard extracellular pH conditions and to preserve hydrogen bonding capability. Moreover, PROPKA3 results suggest a protonated state of this residue as HID (89 % confidence). The aromatic side chains of Phe and Trp were retained as rigid during docking, and π – π stacking was assessed through centroid-to-centroid distance analysis post-docking using visual inspection and LigPlot+ analysis.

Protein-ligand complexes were recovered from Autodock Vina docked complexes, and the simulations were conducted using the Maestro v2024-3 version 13.8.135, MMshare Version 6.4.135, Release 2023-4, and Desmond Multisim v4.0.0 interoperability tools package under the OPLS3 force field. The membrane-peptide complexes were built with the CHARMM-GUI server (<https://charmm->

gui.org, accessed on November 5 2024), with a lipid composition (Outer/inner leaflet ratio) as described: SM (sphingomyelin) 10/10 ; PC (1,2-dioleoyl-sn-glycero- 3-30 phosphocoline) 60/60; PE (1,2-dioleoyl-sn-glycero- 3-30 phosphoethanolamine 65/65; PS (1,2-dioleoyl-sn-glycero- 3-15 phospho-L-serine) 10/10 and, CHL (Cholesterol) 5/5 [67].

GABA_A residue protonation states were set at neutral pH with pKa predictions from *PROPK3*, while the system charges were neutralized at 0.15 M of NaCl and KCl. The cell membrane models total area was set to 2100 Å² (x and y axis extended until 40 Å), and solvated with the three-point TIP3P water model in an periodic box of 15 Å edge distance.

All MD simulations were carried out under periodic boundary conditions. A two-stage energy minimization approximation was applied: 5000 steps of steepest descent followed by 5000 steps of conjugate gradient, with positional restraints (10 kcal·mol⁻¹·Å⁻²) on solute heavy atoms. Subsequently, the system was gradually heated from 0 to 310 K over 100 ps under constant volume conditions, followed by 500 ps of equilibration at a constant temperature of 310 K with relaxation times of 1 ps, using the Langevin thermostat. The Berendsen barostat was applied to control the pressure at 1 atm with relaxation times of 2 ps, and positional restraints were gradually reduced during equilibration. Production runs were conducted for 100 ns under NPT conditions, applying a 2 fs time step with constraints on bonds involving hydrogen atoms. Long-range electrostatics were calculated with a 12 Å cutoff for non-bonded interactions.

Trajectory analysis was conducted using the Simulation Event Module of the Desmond-Maestro interoperability tools; alpha carbon root-mean-square deviation (RMSD) was calculated along complete trajectories to assess structural flexibility.

The free energy landscape governing the binding/unbinding the GABA_A-derivatives complexes was evaluated with the well-tempered metadynamics technique (wt-MetaD); an advanced enhanced sampling technique capable of overcoming the timescale limitations inherent in classical molecular dynamics (MD) simulations [68]. The metadynamics simulations were performed using Desmond v2022-1 (Schrödinger LLC), with post-analysis conducted via the Metadynamics Analysis Plugin in Maestro.

The distance between the centers of mass (COM) of the BZD binding site (initial COM distance ≈ 15 Å) and the center of mass of each derivative was set as a collective variable (CV) to describe the binding process. This CV was selected based on its physical relevance to the dissociation process, consistent with prior studies involving protein complexes [69].

Gaussian potentials were applied at a frequency of 1 ps, with an initial height (ω) of 0.03 Kcal·mol⁻¹ and a width (σ) of 0.5 Å. The bias factor was set to 0.03 Kcal·mol⁻¹ (ω) and 0.5 Å/ps (σ) according to the wt-MetaD protocol, enabling gradual flattening of the energy landscape while retaining resolution near the transition states. The metadynamics simulations continued until a maximum COM distance of 15 Å was reached or complete ligand dissociation was observed. To ensure convergence and reproducibility, three independent replicas of 100 ns each were performed per ligand derivative. The time step was set to 1 fs, and all simulations were conducted under NPT conditions at 310 K and 1 atm, utilizing the Martyna-Tobias-Klein barostat and the Nosé-Hoover thermostat.

The potential of mean force (PMF) profiles were extracted by reconstructing the bias potentials using time-dependent free energy estimators. Convergence was validated by observing plateau behavior in the accumulated bias potential and the PMF curves across replicas. The accuracy of free energy estimates was further supported by bootstrapping the final PMF over trajectory windows, ensuring robustness in the resulting energy barriers.

4.3. Synthesis of 2-(Isoindolin-2-yl) Esters

General Procedure

α , α' -dibromo-*o*-xylene with L-amino acid methyl ester and of potassium carbonate on of acetonitrile was added and refluxed for 6 h. Reaction mixture was filtered and concentrated. The solvent was removed under reduced pressure, and the product was purified by chromatography.

Synthesis of methyl (S)-(1*H*-indol-3-yl)-2-(isoindolin-2-yl) propanoate (ETRP):

Brown solid; yield: 75%. ^1H NMR (500 MHz, CDCl_3 -d): δ 7.20 (s, 4H), 7.09 – 7.04 (m, 2H), 6.71 – 6.65 (m, 2H), 4.29 – 4.21 (m, 3H), 4.18 – 4.10 (m, 2H), 3.75 (dd, J = 8.7, 6.6 Hz, 1H), 3.61 (s, 3H), 3.12 – 3.03 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3 -d): δ 173.09, 139.35, 136.13, 127.42, 126.85, 123.12, 122.69, 122.43, 121.95, 119.35, 118.54, 111.51, 111.24, 77.34, 77.08, 76.83, 65.97, 55.68, 51.44, 27.00; HRMS (ESI): m/z $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{20}\text{H}_{20}\text{N}_2\text{O}_2$: 320.1525, found 320.1531; error = +1.9 ppm.

Synthesis methyl (S)-3-(4-hydroxyphenyl)-2-(isoindolin-2-yl) propanoate (ETYP):

Grey solid; yield: 85%. ^1H NMR (500 MHz, CDCl_3 -d): δ 7.20 (s, 4H), 7.09 – 7.04 (m, 2H), 6.71 – 6.65 (m, 2H), 4.29 – 4.21 (m, 3H), 4.18 – 4.10 (m, 2H), 3.75 (dd, J = 8.7, 6.6 Hz, 1H), 3.61 (s, 3H), 3.12 – 3.03 (m, 2H). ^{13}C NMR (126 MHz, CDCl_3 -d): δ 172.78, 154.50, 139.11, 130.10, 126.83, 122.36, 115.40, 77.28, 77.02, 76.77, 67.08, 55.57, 51.43, 36.56; HRMS (ESI): m/z $[\text{M}+\text{H}]^+$ calculated for $\text{C}_{18}\text{H}_{19}\text{NO}_3$: 297.1365, found 297.1354; error = -3.7 ppm.

4.4. Zebrafish PTZ-Induced Seizure Assay

The anticonvulsant potential of isoindoline was evaluated in adult zebrafish (*Danio rerio*) using a pentylenetetrazol (PTZ)-induced seizure model. Zebrafish were pre-treated by immersion in tanks containing different concentrations for 30 minutes before seizure induction. Diazepam at 75 μM was used as pharmacological control. Diazepam at 75 μM was used as pharmacological control. This concentration was selected based on previous zebrafish studies where immersion doses of 50–100 μM reliably induced seizure protection without sedative or toxic effects [57–59,70]. The immersion model typically requires higher compound concentrations than those found in human plasma due to differences in drug absorption, exposure duration, and metabolic clearance. In this study, the isoindoline ester of tyrosine (ETYP) was tested at 63 and 125 μM , while the isoindoline ester of tryptophan (ETRP) was tested at 10 and 25 μM . Dimethyl sulfoxide (DMSO) at 0.1% was used as the vehicle control. Notably, there is currently no experimental evidence supporting the antiepileptic activity of these compounds. Therefore, to establish initial testing concentrations, we referred to the work of Campos-Rodríguez et al. [41], who evaluated related isoindoline analogs in zebrafish larvae. Based on that precedent, the concentrations of ETYP (63 and 125 μM) and ETRP (10 and 25 μM) were selected for use in our assays.

Following pre-treatment, zebrafish were transferred to a 10 mM PTZ solution (1 L) to induce convulsions. Behavioral responses were recorded and classified into five distinct seizure stages [58]:

Stage 1: Time to present increased swimming activity and high frequency of opercular movement.

Stage 2: Time to present burst swimming, left and right movements, and erratic movements.

Stage 3: Time to present circling movements.

Stage 4: Time to present clonic seizure-like behavior (abnormal whole-body rhythmic muscular contraction).

Stage 5: Time to present fall to the bottom of the tank, tonic seizure-like behavior (sinking to the bottom of the tank, loss of body posture, and principally by rigid extension of the body).

The five stages are described in time in seconds.

4.5. Graphical and Statistical Analysis

All data analyses were performed using IBM SPSS Statistics 29 and GraphPad Prism version 10. For seizure latency in the zebrafish model, normality was tested using the Shapiro–Wilk test, and

homogeneity of variance was assessed with Levene's test. If assumptions of normality and homoscedasticity were met, parametric tests such as one-way ANOVA followed by Dunnett as a post hoc test were used for multiple comparisons using a control group. In cases where data did not meet normality assumptions, non-parametric alternatives, such as the Kruskal–Wallis test with Bonferroni correction for the Mann–Whitney U test used as post hoc correction, were applied. Statistical significance was set at $p < 0.05$ for all analyses. For the construction of the comparison graphs, due to the small sample size and high variability, a boxplot was chosen for the best representation of the data, where the median and the 25th and 75th percentiles are represented, since the mean and standard deviation could be biased.

5. Conclusions

This study demonstrates the successful design, synthesis, and *in vivo* validation of 2-(isoindolin-2-yl) esters derived from aromatic amino acids as potential anticonvulsant agents. The evidence presented suggests modulation of the GABA_A receptor, likely via interaction with the benzodiazepine binding site, based on computational modeling and pharmacological similarity to diazepam. Computational evaluation of sixteen derivatives identified tyrosine and tryptophan esters as the most promising candidates based on predicted pharmacokinetics, binding affinity, and interaction profiles. Molecular docking and dynamics simulations highlighted the critical role of π – π stacking interactions in stabilizing ligand binding at the benzodiazepine site.

Synthesis and characterization of the top-ranked compounds were followed by pharmacological assessment in a zebrafish PTZ-induced seizure model, where tryptophan derivative significantly reduced seizure severity. These findings confirm the predictive value of the computational approach and support the use of aromatic amino acid scaffolds in the development of CNS-active compounds. The work provides a mechanistically grounded framework for the rational design of GABAergic modulators and establishes isoindoline esters as a promising scaffold for further optimization in the context of pharmacoresistant epilepsy.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

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References

1. WHO Epilepsia. <https://www.who.int/es/news-room/fact-sheets/detail/epilepsy> (20 december 2024),
2. Chen, Z.; Brodie, M. J.; Ding, D.; Kwan, P., Editorial: Epidemiology of epilepsy and seizures. *Front Epidemiol* **2023**, 3, 1273163.
3. Beniczky, S.; Trinka, E.; Wirrell, E.; Abdulla, F.; Al Baradie, R.; Alonso Vanegas, M.; Auvin, S.; Singh, M. B.; Blumenfeld, H.; Bogacz Fressola, A.; Caraballo, R.; Carreno, M.; Cendes, F.; Charway, A.; Cook, M.; Craiu, D.; Ezeala-Adikaibe, B.; Frauscher, B.; French, J.; Gule, M. V.; Higurashi, N.; Ikeda, A.; Jansen, F. E.; Jobst, B.; Kahane, P.; Kishk, N.; Khoo, C. S.; Vinayan, K. P.; Lagae, L.; Lim, K. S.; Lizcano, A.; McGonigal, A.; Perez-Gosiengfiao, K. T.; Ryvlin, P.; Specchio, N.; Sperling, M. R.; Stefan, H.; Tatum, W.; Tripathi, M.; Yacubian, E. M.; Wiebe, S.; Wilmschurst, J.; Zhou, D.; Cross, J. H., Updated classification of epileptic seizures: Position paper of the International League Against Epilepsy. *Epilepsia* **2025**.
4. Hu, T.; Zhang, J.; Wang, J.; Sha, L.; Xia, Y.; Ortyl, T. C.; Tian, X.; Chen, L., Advances in Epilepsy: Mechanisms, Clinical Trials, and Drug Therapies. *Journal of Medicinal Chemistry* **2023**, 66, (7), 4434-4467.
5. McCallan, N.; Davidson, S.; Ng, K. Y.; Biglarbeigi, P.; Finlay, D.; Lan, B. L.; McLaughlin, J., Epileptic multi-seizure type classification using electroencephalogram signals from the Temple University Hospital Seizure Corpus: A review. *Expert Systems with Applications* **2023**, 234, 121040.
6. Tomson, T.; Zelano, J.; Dang, Y. L.; Perucca, P., The pharmacological treatment of epilepsy in adults. *Epileptic Disorders* **2023**, 25, (5), 649-669.
7. Mesraoua, B.; Brigo, F.; Lattanzi, S.; Abou-Khalil, B.; Al Hail, H.; Asadi-Pooya, A. A., Drug-resistant epilepsy: Definition, pathophysiology, and management. *Journal of the Neurological Sciences* **2023**, 452, 120766.
8. Wang, Y.; Chen, Z., An update for epilepsy research and antiepileptic drug development: Toward precise circuit therapy. *Pharmacol Ther* **2019**, 201, 77-93.
9. Bryson, A.; Reid, C.; Petrou, S., Fundamental Neurochemistry Review: GABA(A) receptor neurotransmission and epilepsy: Principles, disease mechanisms and pharmacotherapy. *J Neurochem* **2023**, 165, (1), 6-28.
10. Perucca, E.; Bialer, M.; White, H. S., New GABA-Targeting Therapies for the Treatment of Seizures and Epilepsy: I. Role of GABA as a Modulator of Seizure Activity and Recently Approved Medications Acting on the GABA System. *CNS Drugs* **2023**, 37, (9), 755-779.
11. Masiulis, S.; Desai, R.; Uchanski, T.; Serna Martin, I.; Lavery, D.; Karia, D.; Malinauskas, T.; Zivanov, J.; Pardon, E.; Kotecha, A.; Steyaert, J.; Miller, K. W.; Aricescu, A. R., GABA(A) receptor signalling mechanisms revealed by structural pharmacology. *Nature* **2019**, 565, (7740), 454-459.
12. Lavery, D.; Desai, R.; Uchański, T.; Masiulis, S.; Stec, W. J.; Malinauskas, T.; Zivanov, J.; Pardon, E.; Steyaert, J.; Miller, K. W.; Aricescu, A. R., Cryo-EM structure of the human $\alpha 1\beta 3\gamma 2$ GABAA receptor in a lipid bilayer. *Nature* **2019**, 565, (7740), 516-520.
13. Dossi, E.; Huberfeld, G., GABAergic circuits drive focal seizures. *Neurobiol Dis* **2023**, 180, 106102.
14. Zhao, P.; Ding, X.; Li, L.; Jiang, G., A review of cell-type specific circuit mechanisms underlying epilepsy. *Acta Epileptol* **2024**, 6, (1), 18.
15. Kobayashi, K.; Endoh, F.; Ohmori, I.; Akiyama, T., Action of antiepileptic drugs on neurons. *Brain Dev* **2020**, 42, (1), 2-5.
16. Nimgampalle, M.; Chakravarthy, H.; Sharma, S.; Shree, S.; Bhat, A. R.; Pradeepkiran, J. A.; Devanathan, V., Neurotransmitter systems in the etiology of major neurological disorders: Emerging insights and therapeutic implications. *Ageing Res Rev* **2023**, 89, 101994.

17. Arora, I.; Mal, P.; Arora, P.; Paul, A.; Kumar, M., GABAergic implications in anxiety and related disorders. *Biochem Biophys Res Commun* **2024**, 724, 150218.
18. Teleanu, R. I.; Niculescu, A. G.; Roza, E.; Vladacenco, O.; Grumezescu, A. M.; Teleanu, D. M., Neurotransmitters-Key Factors in Neurological and Neurodegenerative Disorders of the Central Nervous System. *Int J Mol Sci* **2022**, 23, (11).
19. Tang, X.; Jaenisch, R.; Sur, M., The role of GABAergic signalling in neurodevelopmental disorders. *Nat Rev Neurosci* **2021**, 22, (5), 290-307.
20. Maguire, J., Mechanisms of Psychiatric Comorbidities in Epilepsy. *Curr Top Behav Neurosci* **2022**, 55, 107-144.
21. Sills, G. J.; Rogawski, M. A., Mechanisms of action of currently used antiseizure drugs. *Neuropharmacology* **2020**, 168, 107966.
22. Jansen, M., An in-depth structural view of a GABAA brain receptor. *Nature* **2019**, 565, (7740), 436-438.
23. Zhu, S.; Sridhar, A.; Teng, J.; Howard, R. J.; Lindahl, E.; Hibbs, R. E., Structural and dynamic mechanisms of GABA(A) receptor modulators with opposing activities. *Nat Commun* **2022**, 13, (1), 4582.
24. Zhu, S.; Noviello, C. M.; Teng, J.; Walsh, R. M., Jr.; Kim, J. J.; Hibbs, R. E., Structure of a human synaptic GABA(A) receptor. *Nature* **2018**, 559, (7712), 67-72.
25. Maramai, S.; Benckekroun, M.; Ward, S. E.; Atack, J. R., Subtype Selective γ -Aminobutyric Acid Type A Receptor (GABAAR) Modulators Acting at the Benzodiazepine Binding Site: An Update. *Journal of Medicinal Chemistry* **2020**, 63, (7), 3425-3446.
26. Golani, L. K.; Yeunus Mian, M.; Ahmed, T.; Pandey, K. P.; Mondal, P.; Sharmin, D.; Rezvanian, S.; Witkin, J. M.; Cook, J. M., Rationalizing the binding and alpha subtype selectivity of synthesized imidazodiazepines and benzodiazepines at GABAA receptors by using molecular docking studies. *Bioorg Med Chem Lett* **2022**, 62, 128637.
27. Goldschen-Ohm, M. P., Benzodiazepine Modulation of GABA(A) Receptors: A Mechanistic Perspective. *Biomolecules* **2022**, 12, (12).
28. de Oliveira, M.; Viana, D. C. F.; Silva, A. A.; Pereira, M. C.; Duarte, F. S.; Pitta, M. G. R.; Pitta, I. R.; Pitta, M. G. R., Synthesis of novel thiazolidinic-phthalimide derivatives evaluated as new multi-target antiepileptic agents. *Bioorg Chem* **2022**, 119, 105548.
29. Chelucci, R. C.; Chiquetto, R.; Chiba, D. E.; Scarim, C. B.; Chin, C. M.; Dos Santos, J. L., Isoindoline-1,3-dione Derivatives as Prototypes for Anticonvulsant Drug Discovery. *Med Chem* **2025**.
30. Jha, M.; Youssef, D.; Sheehy, H.; Jha, A., Synthesis and Pharmacology of Clinical Drugs Containing Isoindoline Heterocycle Core. *Organics* **2025**, 6, (1).
31. Aronica, L.; Albano, G., Cyclization Reactions for the Synthesis of Phthalans and Isoindolines-. *Synthesis* **2018**, 50, (06), 1209-1227.
32. Starosotnikov, A. M.; Bastrakov, M. A., Cycloaddition reactions in the synthesis of isoindolines (microreview). *Chemistry of Heterocyclic Compounds* **2018**, 53, (11), 1181-1183.
33. Fernandes, G. F. S.; Lopes, J. R.; Dos Santos, J. L.; Scarim, C. B., Phthalimide as a versatile pharmacophore scaffold: Unlocking its diverse biological activities. *Drug Dev Res* **2023**, 84, (7), 1346-1375.
34. Aliabadi, A.; Gholamine, B.; Karimi, T., Synthesis and antiseizure evaluation of isoindoline-1,3-dione derivatives in mice. *Medicinal Chemistry Research* **2014**, 23, (6), 2736-2743.
35. Abdel-Hafez, A. A.-M., Synthesis and anticonvulsant evaluation of N-substituted-isoindolinedione derivatives. *Archives of Pharmacol Research* **2004**, 27, (5), 495-501.

36. Iman, M.; Saadabadi, A.; Davood, A.; Shafaroodi, H.; Nikbakht, A.; Ansari, A.; Abedini, M., Docking, Synthesis and Anticonvulsant Activity of N-substituted Isoindoline-1,3-dione. *Iran J Pharm Res* **2017**, *16*, (2), e125027.
37. Asadollahi, A.; Asadi, M.; Hosseini, F. S.; Ekhtiari, Z.; Biglar, M.; Amanlou, M., Synthesis, molecular docking, and antiepileptic activity of novel phthalimide derivatives bearing amino acid conjugated anilines. *Research in Pharmaceutical Sciences* **2019**, *14*, (6), 534-543.
38. Mancilla Percino, T.; Guzmán Ramírez, J. E.; Mera Jiménez, E.; Trejo Muñoz, C. R., Synthesis, characterization of novel isoindolyl- and bis-isoindolylphenylboronic anhydrides. Antiproliferative activity on glioblastoma cells and microglial cells assays of boron and isoindolines compounds. *Journal of Organometallic Chemistry* **2019**, *891*, 35-43.
39. Mancilla Percino, T.; Hernández Rodríguez, M.; Mera Jiménez, E., Synthesis of New Isoindolines Derived from L-A-Amino Acids and their Selectivity on Cancer Cell Lines. *ChemistrySelect* **2024**, *9*, (4).
40. Campos-Rodríguez, C.; Trujillo-Ferrara, J. G.; Alvarez-Guerra, A.; Vargas, I. M. C.; Cuevas-Hernandez, R. I.; Andrade-Jorge, E.; Zamudio, S.; Juan, E. R., Neuropharmacological Screening of Chiral and Non-chiral Phthalimide-Containing Compounds in Mice: in vivo and in silico Experiments. *Med Chem* **2019**, *15*, (1), 102-118.
41. Campos-Rodríguez, C.; Fredrick, E.; Ramirez-San Juan, E.; Olsson, R., Enantiomeric N-substituted phthalimides with excitatory amino acids protect zebrafish larvae against PTZ-induced seizures. *Eur J Pharmacol* **2020**, 888.
42. Özbek, O.; Gürdere, M. B., A review on the synthesis and applications of molecules as anticonvulsant drug agent candidates. *Medicinal Chemistry Research* **2020**, *29*, (9), 1553-1578.
43. Ahuja, P.; Husain, A.; Siddiqui, N., Essential amino acid incorporated GABA-phthalimide derivatives: synthesis and anticonvulsant evaluation. *Medicinal Chemistry Research* **2014**, *23*, (9), 4085-4098.
44. Banerjee, P. S.; Sharma, P. K., New antiepileptic agents: structure-activity relationships. *Medicinal Chemistry Research* **2011**, *21*, (7), 1491-1508.
45. Hollingsworth, S. A.; Dror, R. O., Molecular dynamics simulation for all. *Neuron* **2018**, *99*, (6), 1129-1143.
46. De Vivo, M.; Masetti, M.; Bottegoni, G.; Cavalli, A., Role of Molecular Dynamics and Related Methods in Drug Discovery. *J Med Chem* **2016**, *59*, (9), 4035-61.
47. Barducci, A.; Bonomi, M.; Parrinello, M., Metadynamics. *WIREs Computational Molecular Science* **2011**, *1*, (5), 826-843.
48. Cavalli, A.; Spitaleri, A.; Saladino, G.; Gervasio, F. L., Investigating drug-target association and dissociation mechanisms using metadynamics-based algorithms. *Acc Chem Res* **2015**, *48*, (2), 277-85.
49. D'Amora, M.; Galgani, A.; Marchese, M.; Tantussi, F.; Faraguna, U.; De Angelis, F.; Giorgi, F. S., Zebrafish as an Innovative Tool for Epilepsy Modeling: State of the Art and Potential Future Directions. *International Journal of Molecular Sciences* **2023**, *24*, (9), 7702.
50. Lopez-Rosas, C. A.; Gonzalez-Perianez, S.; Pawar, T. J.; Zurutuza-Lormendez, J. I.; Ramos-Morales, F. R.; Olivares-Romero, J. L.; Saavedra Velez, M. V.; Hernandez-Rosas, F., Anticonvulsant Potential and Toxicological Profile of *Verbesina persicifolia* Leaf Extracts: Evaluation in Zebrafish Seizure and *Artemia salina* Toxicity Models. *Plants (Basel)* **2025**, *14*, (7).
51. Samokhina, E.; Samokhin, A., Neuropathological profile of the pentylenetetrazol (PTZ) kindling model. *Int J Neurosci* **2018**, *128*, (11), 1086-1096.
52. Murugan, R.; Ramya Ranjan Nayak, S. P.; Haridevamuthu, B.; Priya, D.; Chitra, V.; Almutairi, B. O.; Arokiyaraj, S.; Saravanan, M.; Kathiravan, M. K.; Arockiaraj, J., Neuroprotective potential of pyrazole

- benzenesulfonamide derivative T1 in targeted intervention against PTZ-induced epilepsy-like condition in vivo zebrafish model. *Int Immunopharmacol* **2024**, 131, 111859.
53. Ciubotaru, A. D.; Leferman, C.-E.; Ignat, B.-E.; Knieling, A.; Esanu, I. M.; Salaru, D. L.; Foia, L. G.; Minea, B.; Hritcu, L. D.; Dimitriu, C. D.; Stoica, L.; Ciureanu, I.-A.; Ciobica, A. S.; Neamtu, A.; Stoica, B. A.; Ghiciuc, C. M., Behavioral and Biochemical Insights into the Therapeutic Potential of Mitocurcumin in a Zebrafish–Pentylentetrazole (PTZ) Epilepsy Model. *Pharmaceuticals* **2025**, 18, (3).
 54. Kim, J. J.; Gharpure, A.; Teng, J.; Zhuang, Y.; Howard, R. J.; Zhu, S.; Noviello, C. M.; Walsh, R. M., Jr.; Lindahl, E.; Hibbs, R. E., Shared structural mechanisms of general anaesthetics and benzodiazepines. *Nature* **2020**, 585, (7824), 303-308.
 55. Sadamitsu, K.; Shigemitsu, L.; Suzuki, M.; Ito, D.; Kashima, M.; Hirata, H., Characterization of zebrafish GABA(A) receptor subunits. *Sci Rep* **2021**, 11, (1), 6242.
 56. Monesson-Olson, B.; McClain, J. J.; Case, A. E.; Dorman, H. E.; Turkewitz, D. R.; Steiner, A. B.; Downes, G. B., Expression of the eight GABAA receptor alpha subunits in the developing zebrafish central nervous system. *PLoS One* **2018**, 13, (4), e0196083.
 57. Chitolina, R.; Reis, C. G.; Stahlhofer-Buss, T.; Linazzi, A.; Benvenuti, R.; Marcon, M.; Herrmann, A. P.; Piato, A., Effects of N-acetylcysteine and acetyl-L-carnitine on acute PTZ-induced seizures in larval and adult zebrafish. *Pharmacol Rep* **2023**, 75, (6), 1544-1555.
 58. Mussulini, B. H.; Leite, C. E.; Zenki, K. C.; Moro, L.; Baggio, S.; Rico, E. P.; Rosemberg, D. B.; Dias, R. D.; Souza, T. M.; Calcagnotto, M. E.; Campos, M. M.; Battastini, A. M.; de Oliveira, D. L., Seizures induced by pentylentetrazole in the adult zebrafish: a detailed behavioral characterization. *PLoS One* **2013**, 8, (1), e54515.
 59. Fontana, B. D.; Ziani, P. R.; Canzian, J.; Mezzomo, N. J.; Muller, T. E.; Dos Santos, M. M.; Loro, V. L.; Barbosa, N. V.; Mello, C. F.; Rosemberg, D. B., Taurine Protects from Pentylentetrazole-Induced Behavioral and Neurochemical Changes in Zebrafish. *Mol Neurobiol* **2019**, 56, (1), 583-594.
 60. Verma, R.; Raj Choudhary, P.; Kumar Nirmal, N.; Syed, F.; Verma, R., Neurotransmitter systems in zebrafish model as a target for neurobehavioural studies. *Materials Today: Proceedings* **2022**, 69, 1565-1580.
 61. Horzmann, K. A.; Freeman, J. L., Zebrafish Get Connected: Investigating Neurotransmission Targets and Alterations in Chemical Toxicity. *Toxics* **2016**, 4, (3).
 62. Monesson-Olson, B.; McClain, J. J.; Case, A. E.; Dorman, H. E.; Turkewitz, D. R.; Steiner, A. B.; Downes, G. B., Expression of the eight GABAA receptor alpha subunits in the developing zebrafish central nervous system. *PLoS One* **2018**, 13, (4).
 63. M. J. Frisch, G. W. T., H. B. Schlegel, G. E. Scuseria, M. A. Robb;; J. R. Cheeseman, G. S., V. Barone, G. A. Petersson, H. Nakatsuji, X. Li;; M. Caricato, A. M., J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci;; H. P. Hratchian, J. V. O., A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young;; F. Ding, F. L., F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson;; D. Ranasinghe, V. G. Z., J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada;; M. Ehara, K. T., R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda;; O. Kitao, H. N., T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta;; F. Ogliaro, M. B., J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov;; T. Keith, R. K., J. Normand, K. Raghavachari, A. Rendell, J. C. Burant;; S. S. Iyengar, J. T., M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi;; J. W. Ochterski, R. L. M., K. Morokuma, O. Farkas, J. B. Foresman, and; D. J. Fox *Gaussian 09, Revision A.1*, Gaussian, Inc: 2009.
 64. Xiong, G.; Wu, Z.; Yi, J.; Fu, L.; Yang, Z.; Hsieh, C.; Yin, M.; Zeng, X.; Wu, C.; Lu, A.; Chen, X.; Hou, T.; Cao, D., ADMETlab 2.0: an integrated online platform for accurate and comprehensive predictions of ADMET properties. *Nucleic Acids Research* **2021**, 49, (W1), W5-W14.

65. Dong, J.; Wang, N. N.; Yao, Z. J.; Zhang, L.; Cheng, Y.; Ouyang, D.; Lu, A. P.; Cao, D. S., ADMETlab: a platform for systematic ADMET evaluation based on a comprehensively collected ADMET database. *J Cheminform* **2018**, 10, (1), 29.
66. Daina, A.; Michielin, O.; Zoete, V., SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Scientific Reports* **2017**, 7, (1), 42717.
67. Rivel, T.; Ramseyer, C.; Yesylevskyy, S., The asymmetry of plasma membranes and their cholesterol content influence the uptake of cisplatin. *Sci Rep* **2019**, 9, (1), 5627.
68. Barducci, A.; Bussi, G.; Parrinello, M., Well-tempered metadynamics: a smoothly converging and tunable free-energy method. *Phys Rev Lett* **2008**, 100, (2), 020603.
69. Tribello, G. A.; Bonomi, M.; Branduardi, D.; Camilloni, C.; Bussi, G., PLUMED 2: New feathers for an old bird. *Computer Physics Communications* **2014**, 185, (2), 604-613.
70. Pereira, A. C. M.; Cunha, A. A.; Carvalho, H. d. O.; Ferreira, I. M.; Carvalho, J. C. T., Oral diazepam suppresses pentylenetetrazole-induced seizure-like behavior in adult zebrafish: A tool for nonclinical studies. *Journal of Applied Pharmaceutical Science* **2023**.

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