

Review

Not peer-reviewed version

# From Mutation to Metabolism: Toxins, Mitochondria, and Integrative Orthomolecular Cancer Therapy (IOCT) – Implications for ASCVD and T2DM

#### Richard Cheng

Posted Date: 15 October 2025

doi: 10.20944/preprints202510.1142.v1

Keywords: cancer; mitochondria; oxidative stress; metabolic therapy; orthomolecular; IOCT; ketogenic diet; intravenous vitamin C; TPIM; adaptive trials



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

# From Mutation to Metabolism: Toxins, Mitochondria, and Integrative Orthomolecular Cancer Therapy (IOCT) — Implications for ASCVD and T2DM

#### Richard Z. Cheng 1,2,3

- <sup>1</sup> Cheng Integrative Health Center, Columbia, SC, USA; richzc@gmail.com
- <sup>2</sup> Cheng Health Consulting, Ltd., Shanghai, China
- <sup>3</sup> Orthomolecular Medicine News Service (OMNS), USA

#### **Abstract**

Conventional oncology remains mutation-centric despite modest survival gains and substantial toxicity. Converging evidence indicates that environmental and dietary toxins, together with micronutrient insufficiency, act upstream to damage mitochondria-precipitating redox collapse, impaired oxidative phosphorylation, metabolic inflexibility, genomic instability, and therapy resistance. These lesions not only initiate and sustain malignancy but also underpin a broader chronic-disease continuum that includes atherosclerotic cardiovascular disease (ASCVD) and type 2 diabetes mellitus (T2DM). Building on prior root-cause analyses, we introduce the Triple-Principle **Intervention Model (TPIM)**—Safety first, Effectiveness via titration-to-target, and Affordability—as a conservative clinical filter for Integrative Orthomolecular Medicine. We then operationalize Integrative Orthomolecular Cancer Therapy (IOCT) through a systems-based program combining restricted ketogenic nutrition and fasting; high-dose intravenous vitamin C; targeted micronutrient repletion (e.g., niacin/NAD+, D3/K2, Mg, omega-3); mitochondrial redox modulation (e.g., PBMT/"Red-Blue" with methylene blue); immune-inflammation regulation; bioidentical hormone optimization; and circadian/lifestyle realignment. To evaluate such multi-modal, adaptive care, we propose the Triple-Principle Adaptive RCT (TP-ARC)—a next-generation clinical trial framework that prioritizes clinical outcomes over isolated pharmacologic effects. Unlike conventional drug-centric RCTs designed to test single agents under rigid inclusion criteria, TP-ARC assesses the whole-person therapeutic program and its impact on quality of life, functional capacity, and survival. Biomarker titration serves as a secondary, supportive tool rather than the primary endpoint. By adapting interventions to fit each patient-rather than forcing patients to fit protocols-TP-ARC bridges realworld practice and research, embodying a pragmatic, ethical, and patient-centered evolution of clinical science. This mitochondrial-metabolism-focused integrative framework reframes cancer as a preventable and modifiable systems disorder, enabling ethical and economical bedside translation while providing mechanistic continuity with ASCVD and T2DM to guide future cross-disciplinary trials.

**Keywords:** cancer; mitochondria; oxidative stress; metabolic therapy; orthomolecular; IOCT; ketogenic diet; intravenous vitamin C; TPIM; adaptive trials

#### **Series Note**

This is Part III of *From Mutation to Metabolism*. Part I mapped upstream drivers of cancer[1]; Part II detailed environmental/dietary toxins as initiators of mitochondrial dysfunction[2]. This paper (Part III) integrates these insights into a mitochondria-centered oncology model (IOCT) filtered by TPIM[3] and shows how the same mitochondrial lesions explain parallel risk and progression patterns in chronic diseases such as ASCVD and T2DM.

#### 1. Introduction

More than fifty years after the launch of the "War on Cancer," survival gains for most advanced malignancies remain modest, despite an exponential rise in molecular data, targeted drugs, and treatment cost. The prevailing paradigm continues to view **genetic mutations** as cancer's primary cause and mutation-targeted therapy as its logical cure; however, accumulating metabolic and mitochondrial evidence challenges the sufficiency of this model [4–10]. This framework cannot explain the wide variability of clinical outcomes, the common metabolic features shared by genetically diverse tumors, or the overlap between cancer and other chronic diseases such as cardiovascular disease and type 2 diabetes mellitus (T2DM) [4–9,11–16].

Converging data across oncology, cardiometabolism, and aging biology indicate that mitochondrial dysfunction—rather than random nuclear mutations—is the earliest and unifying lesion linking environmental, nutritional, and metabolic stressors into a single integrative pathophysiologic axis—the very foundation of the Integrative Orthomolecular Medicine (IOM) model [4,9,10,17–21]. Environmental and dietary toxins, together with widespread micronutrient insufficiency, impair mitochondrial respiration, collapse redox balance, and erode metabolic flexibility [1,2,22–31]. These insults propagate upward as genomic instability, chronic inflammation, insulin resistance, and impaired apoptosis, creating the biochemical terrain from which malignant transformation arises [4,10,17–21].

In this paper—the third installment of the *From Mutation to Metabolism* series [1,2]—we extend prior root-cause analyses to position **mitochondrial injury as a central driver of cancer** and the mechanistic bridge linking cancer with atherosclerotic cardiovascular disease (ASCVD) and T2DM. We introduce a mitochondria-centered clinical framework, **Integrative Orthomolecular Cancer Therapy (IOCT)**, filtered by the **Triple-Principle Intervention Model (TPIM)** emphasizing **safety**, **titration-to-target effectiveness**, and **affordability**[3]. While this article focuses on cancer as the lead indication, the same mechanistic lens clarifies how toxin- and nutrient-driven mitochondrial collapse underlies multiple chronic diseases, offering a unified, preventive, and therapeutic strategy within the broader **Integrative Orthomolecular Medicine (IOM)** framework [1,2,11–16].

#### 2. From Root Causes to Mitochondria: Cancer as the Lead Indication

Cancer seldom arises from a single insult. Instead, multiple upstream drivers—environmental, dietary, metabolic, and psychosocial—converge on mitochondrial dysfunction. Mitochondria are both power plants and sentinels of cellular integrity, orchestrating ATP generation, redox balance, apoptosis, and biosynthetic flux; when their function is compromised, the result is not merely energetic failure but a cascade of downstream disturbances: excess reactive oxygen species (ROS), redox collapse, genomic instability, and loss of growth control [4,9,10,17–21].

Our prior analyses (Parts I and II) mapped ten categories of upstream drivers that precede malignant transformation—ranging from **toxin exposure** and **nutrient deficiency** to **hormonal**, **inflammatory**, **and lifestyle stressors** [1,2]. Although clinically diverse, these factors share a single biochemical consequence: erosion of mitochondrial resilience [2,22–33]. The same injury patterns recur in other chronic diseases, notably **ASCVD** and **T2DM**, which are likewise characterized by oxidative stress, metabolic inflexibility, and impaired apoptosis [11–16,34–40].

#### 2.1. Mechanistic Convergence

Upstream Driver	Mitochondria	Cancer	(Lead	ASCVD Parallel	T2DM
	l Lesion	Example)			Parallel
Environmental &	ETC	Warburg	shift,	Endothelial	β-cell
industrial toxins	inhibition;	genomic		mitochondrial	oxidative
(heavy metals,	↑ROS;	instability,		ROS → ↓NO	stress ->
		therapy		bioavailability;	impaired



	(DNIA	[2, 4, 0, 22]	1	A TED / A D.D.
pesticides, air	mtDNA	resistance[2,4,9,22	vascular	ATP/ADP
pollutants)	damage	-25]	calcification[11–	signaling;
			16]	insulin
				secretory
				failure[26,34–
				40]
High-carbohydrate /	Metabolic	Tumor glucose	Dyslipidemia;	Insulin
ultra-processed diet;	inflexibility;	dependence;	foam-cell	resistance;
hyperinsulinemia	sustained	anabolic	inflammation;	hepatic
	glycolysis;	signaling[4–9,41–	adverse	steatosis;
	lipotoxicity	43]	cardiometabolic	adverse
			profiles[11,12,14	metabolomic
			-16]	signatures[34–
				40]
Micronutrient	Redox	Impaired	†Oxidative	↓Mitochondria
insufficiency (C, D3,	collapse;	apoptosis;	burden;	l biogenesis;
K2, Mg, Se, Niacin)	↓OXPHOS	immune	endothelial	reduced
	enzyme	escape[10,26–33]	dysfunction[26-	insulin
	activity		33]	sensitivity[26-
				33]
Chronic	Cytokine-	Pro-tumor	Plaque	Systemic low-
infection/inflammatio	driven	microenvironmen	inflammation;	grade
n; stress hormones	ROS/RNS;	t; resistance[10,17-	immune	inflammation;
	mtDNA	19,21,36]	activation[11–	β-cell
	editing		16]	fatigue[34–40]
Iatrogenic & lifestyle	Accumulated	Therapy	Accelerated	Circadian
stressors (drugs,	oxidative	resistance; stem-	vascular aging;	insulin
radiation, inactivity,	load;	cell	metabolic	dysregulation;
poor sleep)	circadian	survival[10,17–	syndrome	insulin
	disruption	19,21,36]	features[11–16]	resistance[34–
				40]

Interpretation. Across these categories, mitochondrial lesions precede structural disease. The same ETC inhibition, redox imbalance, and impaired apoptosis that initiate tumorigenesis also accelerate endothelial injury and  $\beta$ -cell failure. Thus, mitochondrial injury is a systems-level failure mode, explaining why cancer, ASCVD, and T2DM frequently co-cluster in the same metabolic terrain[2,4–9,11–16,34–43].

#### 2.2. Conceptual Shift

Traditional oncology interprets these observations downstream—mutations as cause, mitochondrial changes as effect. The reverse appears more consistent with biochemical evidence: **toxins and nutrient deficits first disable mitochondria**, with genomic instability emerging secondarily [2,4,9,10,17–19,21–25,36]. This inversion reframes carcinogenesis as a **bioenergetic and** 

**redox disorder** within a broader chronic-disease spectrum [4–9,11–16,35–43]. Consequently, interventions that **restore mitochondrial efficiency**—through source control of toxins, orthomolecular repletion, metabolic flexibility, and redox modulation—are not merely supportive but **causally corrective** within oncology's remit [1,2,4–9,41–43]. In the next section, we formalize this therapeutic logic through the **Triple-Principle Intervention Model (TPIM)**, which functions as the ethical and clinical filter for **Integrative Orthomolecular Cancer Therapy (IOCT)**.

# 3. The Triple-Principle Intervention Model (TPIM)

Conventional hierarchies that privilege regulatory status and single-agent RCTs risk excluding safe, biologically plausible, and accessible strategies for patients with complex metabolic terrain [44]. TPIM is introduced here (and in Parts I–II of this series) as a conservative clinical filter that prioritizes **Safety first, Effectiveness via titration-to-target**, and **Affordability/accessibility** to operationalize **IOCT** within real-world oncology [1,2].

#### 3.1. Safety First

Cancer patients already face substantial toxicity from standard care; any added intervention must demonstrate a favorable safety profile and incorporate routine monitoring.

- Low-toxicity metabolic and orthomolecular modalities. Ketogenic/metabolic therapy (KMT) and related approaches show acceptable tolerability in contemporary reviews and case-series across tumor types [41–43,45]. High-dose intravenous vitamin C (HDIVC) has extensive clinical experience with documented safety signals and modern pharmacologic understanding; adverse-event reviews and clinical protocols emphasize key contraindications (e.g., G6PD deficiency, significant renal impairment) and practical safeguards [30,31,46–49].
- Safety monitoring (labs/biomarkers). Standard hepatobiliary panels (ALT/AST, bilirubin,
  ALP, bile-acid profiles where indicated) remain foundational for detecting treatment-related
  liver stress [50–52]. The GGT + ferritin pair can function as an early, low-cost screen for
  oxidative/toxic burden and metabolic risk, supporting risk stratification and follow-up [53–55].
- Program integration. Within IOCT, safety checkpoints are staged before dose-escalation of diet, fasting, HDIVC, niacin, and other agents, with reassessment after meaningful titration windows [30,31,41–43,46,47,49].

## 3.2. Effectiveness via Titration-to-Target

Rather than one-size-fits-all protocols, TPIM defines effectiveness as **restoration of biological function** measurable through validated or biologically coherent biomarkers, with dose and program intensity **titrated to explicit targets**.

- Metabolic/ketogenic monitoring. The Glucose–Ketone Index (GKI) provides a practical, continuous readout of the host–tumor metabolic gradient and is suitable for longitudinal titration in metabolic oncology programs [45,56–58].
- Terrain correction through diet quality. Evidence linking ultra-processed foods (UPFs) with adverse cardiometabolic profiles and cancer risk underscores diet quality as a titratable lever within IOCT [11–16], with metabolomic and insulin-resistance signatures (e.g., TyG, TG/HDL) supporting monitoring [34–40].
- Orthomolecular repletion. Mechanistic and translational literature supports targeted correction
  of micronutrient insufficiencies (ascorbate, D3/K2, Mg, Se, niacin/NAD+) to stabilize redox,
  support OXPHOS, and improve therapy tolerance [10,26–33]; HDIVC delivers pharmacologic
  ascorbate levels with tumor-selective effects and host redox support [33,47–49]. (Note: HDIVC



can transiently interfere with point-of-care glucose meters; confirm glucose with laboratory assays when needed.)

#### Mitochondrial Repair and Restoration as the Ultimate Target

Within TPIM, effectiveness is ultimately defined as restoration of mitochondrial integrity and function—the biochemical common denominator of recovery. Mitochondrial repair can be tracked through improvements in redox balance (\$\psi\$ GGT + ferritin), OXPHOS efficiency (\$\psi\$ ATP/ADP ratio, lactate \$\psi\$), and metabolic flexibility (optimized GKI). Orthomolecular and biophysical interventions—including HDIVC, niacin-driven NAD+ repletion, magnesium and CoQ10 support, omega-3 membrane stabilization, and Red–Blue PBMT with methylene blue—synergistically enhance electron-transport integrity and antioxidant recycling. When titrated-to-target, these measures not only reverse tumor-associated mitochondrial injury but also normalize cardiometabolic and endocrine terrain, providing a measurable endpoint for IOCT programs.

## Titration-to-target exemplars used in IOCT (programmatic thresholds):

- **GKI:** <1 for intensive phases; <3 for maintenance/metabolic control[45,56–59].
- **Redox/toxic burden:** trending **GGT + ferritin** toward low-normal ranges, with source control when elevated[53–55,60].
- **Insulin resistance:** fasting insulin/HOMA-IR, **TyG**, and **TG/HDL** tracked to predefined clinic thresholds during diet/fasting cycles[34–40].
- Micronutrient sufficiency: correction guided by serial labs for ascorbate status (where available), 25(OH)D, Mg, Se, and lipids while titrating niacin and omega-3s[10,26–33].

*Note*: Targets are operational within the IOCT program (author framework) and are justified by mechanism and feasibility; they are not presented as guideline mandates.

#### 3.3. Affordability and Accessibility

For integrative programs to scale across diverse settings and sustain adherence, components must be **practical and cost-conscious**. Many IOCT levers—dietary restructuring, vitamin D3, magnesium, niacin, and protocolized HDIVC—are comparatively inexpensive and broadly deployable in community settings[1,2,32]. Affordability therefore functions as an **ethical design constraint** within TPIM, improving reach and persistence of care.

TPIM as a Clinical Filter for IOCT (decision funnel)

Root causes  $\rightarrow$  TPIM filter (Safety, Titration-to-Target Effectiveness, Affordability)  $\rightarrow$  IOCT protocol modules.

Any modality that meets all three TPIM conditions—and aligns with patient goals—qualifies for inclusion in an IOCT plan; safety labs and biomarker targets guide intensity and sequencing.

# 4. Clinical Application: Integrative Orthomolecular Cancer Therapy (IOCT)

Effective cancer care within the Integrative Orthomolecular Medicine (IOM) framework requires systematic evaluation and intervention across all 10 categories of root-level drivers[1]. The therapeutic aim extends beyond tumor suppression: while controlling tumor growth, the clinician seeks to restore systemic balance, reduce metabolic and oxidative stress, and rebuild host resilience and quality of life.



#### 4.1. Environmental & Occupational Toxins- The Dominant Driver of Global Mortality

In the modern era, toxin exposure has emerged as a leading global health threat. The 2017 Lancet Commission on Pollution and Health reported that pollution is responsible for **over 9 million deaths annually**, or 16% of all deaths worldwide—more than war, smoking, and malnutrition combined[61,62]. These exposures are not isolated to industrial zones; they are pervasive across air, water, soil, and consumer products, silently eroding metabolic resilience and driving carcinogenesis.

Cancer represents one of the most visible clinical endpoints of this toxic overload. Heavy metals (arsenic, cadmium, mercury, lead) damage DNA and mitochondria; pesticides and herbicides disrupt endocrine and immune function; plastics and phthalates act as estrogenic mimics; airborne particulates induce chronic oxidative stress; and persistent organic pollutants accumulate in fatty tissues, altering gene expression and promoting malignant transformation. From an orthomolecular perspective, toxins accelerate nutrient depletion, glutathione exhaustion, and redox collapse—precisely the vulnerabilities that set the stage for cancer[22–25].

Recent evidence[2] underscores that toxins drive not only cancer, but also heart disease and type 2 diabetes, through a common mechanism: mitochondrial damage. Industrial chemicals (benzene, formaldehyde, phthalates), pesticides (glyphosate, atrazine), heavy metals (lead, mercury, cadmium), air pollutants (PM2.5), and even everyday household chemicals (parabens, triclosan) all impair mitochondrial DNA integrity, disrupt the electron transport chain, and increase reactive oxygen species (ROS). The resulting oxidative stress and redox collapse compromise ATP production and push cells toward glycolysis, a shift that promotes carcinogenesis, atherosclerosis, and insulin resistance. In cardiovascular disease, such toxic-driven mitochondrial injury accelerates endothelial dysfunction and arterial stiffness; in type 2 diabetes, it damages pancreatic beta cells and disrupts insulin signaling. In all three conditions, the unifying feature is that toxins erode metabolic resilience at the mitochondrial level, creating a fertile ground for chronic disease. This convergence strengthens the argument that cancer should not be studied in isolation but as part of a broader toxin-induced chronic disease continuum.

#### 4.1.1. Assessment

Effective toxin management begins with **multi-level evaluation**:

- Laboratory testing:
- Hair and mineral analysis (chronic heavy metal burden)[63,64].
- Serum and urine panels for lead, cadmium, mercury, arsenic[65,66].
- Organic pollutant screens (PCBs, dioxins, pesticides)[67–69].
- Functional biomarkers:
- GGT + ferritin pair as early markers of oxidative stress and toxic overload[53– 55,60].
- ALT, AST, bilirubin, and alkaline phosphatase for hepatic burden[50–52].
- Organic Acids Test (OAT) evaluates mitochondrial intermediates, oxidative stress markers, and detox-related metabolites (e.g., pyroglutamate, orotate), providing a systems view of redox and energy balance[70–72].



- Comprehensive detox-function profile includes phase I/II conjugation capacity (sulfation, glucuronidation, methylation pathways) and glutathione status to assess liver detoxification efficiency[70,73–79].
- Intestinal permeability ("leaky gut") and microbiome balance tests identify gut-derived endotoxin translocation and dysbiosis that perpetuate systemic inflammation and toxin recirculation[79–82].
- **Imaging:** Coronary artery calcium (CAC) and low dose CT scans for vascular calcification, often reflecting cumulative toxic stress[83–85].
- Clinical history: Occupational exposures (mining, welding, agriculture, manufacturing), residential proximity to industrial/agricultural zones, and lifestyle habits (plastic use, processed foods, contaminated water).

# 4.1.2. Mechanistic Impact on Carcinogenesis

- Mitochondrial injury: Heavy metals and pesticides inhibit key enzymes in oxidative phosphorylation, forcing reliance on glycolysis (the Warburg effect)[2,86].
- DNA damage: Arsenic, cadmium, and air pollutants induce DNA adducts and chromosomal instability[2,87,88].
- Endocrine disruption: Phthalates, bisphenol A (BPA), and pesticides mimic or block hormone signaling, driving hormone-dependent cancers (breast, prostate, endometrial)[2,89–94].
- **Chronic inflammation:** Particulate matter (PM2.5) and diesel exhaust activate NF-κB and IL-6 pathways, fueling systemic inflammation and tumor progression[95–97].
- Nutrient depletion: Many toxins increase turnover of antioxidants (e.g., vitamin C, vitamin E, glutathione) and micronutrients (selenium, zinc, magnesium), compounding deficiencies already common in cancer patients[98–102].

# 4.1.3. Management - Detoxification & Resilience Building

Within the TPIM framework, management of toxin burden is **twofold: remove the source**, and **restore resilience**. Detoxification therapy in cancer treatment encompasses various approaches aimed at reducing toxicity and improving treatment outcomes. <u>Savchenko et al. (2022)</u> demonstrated that enterosorbent-based detoxification therapy effectively restored immune function in cancer patients, normalizing cellular immunity parameters and relieving inflammatory responses compared to standard therapy alone[103]. However, established alternative detoxification regimens like the Gerson therapy, which involves dietary modifications and coffee enemas, lack scientific substantiation and showed no evidence of clinical benefit in controlled studies[104]. Additionally, fasting-induced metabolic changes may enhance chemotherapy efficacy by protecting normal cells while potentially reducing multidrug resistance in cancer cells through alterations in glucose, IGF-I, and other protein levels[105]. These findings suggest that evidence-based detoxification approaches may complement conventional cancer therapy.

## • Source elimination:

- Clean air (filtration), clean water (reverse osmosis or distilled), toxin-free diet (organic, pesticide-free, low in ultra-processed foods)[11–16].
- o Reduction of plastic exposure, safe household and personal care products[25].
- Biochemical detoxification:



- Liver Detox protocol supports hepatic phase I/II detoxification pathways, enhances bile flow, and promotes elimination[24,50–52].
- Targeted chelation therapy (EDTA, DMSA, DMPS) for confirmed heavy metal overload [24,64].
- Orthomolecular cofactors: Vitamin C, magnesium, selenium, alpha-lipoic acid, and N-acetylcysteine to replenish glutathione and enhance redox balance[10,33,48].
- Mitochondrial repair and antioxidant defense:
- o High-dose vitamin C (oral/IV) to counteract oxidative burden[33,46–49].
- o CoQ10 and carnitine for mitochondrial stabilization[10,18].
- o Omega-3 fatty acids to counter inflammatory signaling[26–33].

#### 4.1.4. Clinical Imperative

The pervasive, cumulative, and synergistic effects of environmental toxins make them arguably the most urgent and modifiable driver of cancer and chronic disease today. Unlike genetic predispositions, toxin exposures are preventable and reversible—provided they are systematically identified and addressed. Orthomolecular strategies not only facilitate detoxification but also restore the biochemical defenses required to neutralize ongoing exposures.

In this sense, tackling environmental and occupational toxins represents the **frontline of integrative cancer prevention and management**: before cellular mutations arise, before metabolic collapse occurs, and before irreversible malignant transformation takes hold.

#### 4.2. Dietary & Metabolic Stressors

Cancer cells exhibit profound metabolic inflexibility, relying predominantly on glycolysis for ATP production even in the presence of oxygen—a phenomenon known as the **Warburg effect**. This dependency creates a therapeutic opportunity: by restricting glucose availability and promoting ketone utilization, one can selectively stress tumor cells while enhancing host metabolic resilience[4–8].

#### 4.2.1. Restricted Ketogenic Diet and Intermittent Fasting

A **restricted ketogenic diet (KD)** provides adequate protein and high-quality fats while minimizing carbohydrate intake, thereby lowering circulating glucose and insulin. This shifts systemic metabolism toward ketosis, depriving tumor cells of their primary fuel source while simultaneously enhancing mitochondrial efficiency in normal tissues. **Intermittent fasting** or prolonged fasting intervals further reinforce these effects by lowering insulin-like growth factor 1 (IGF-1), improving autophagy, and reducing inflammatory signaling. Together, these strategies remodel systemic metabolism to favor host survival over tumor growth[5,9,41–43].

# Continuous Monitoring

To ensure effectiveness and adherence, **continuous glucose monitoring (CGM)** and periodic measurement of **blood ketones** are employed. These tools allow clinicians and patients to track the **Glucose Ketone Index (GKI)** in real time, providing an objective biomarker of metabolic control[45,56,57,59]. Within the TPIM framework, interventions are **titrated-to-target**, with a therapeutic goal of achieving GKI values consistent with metabolic stress on tumors (<1 in intensive protocols, <3 in maintenance)[45,58,59].

# Clinical Goals

The primary aim of metabolic therapies is to **reduce tumor glycolysis**, thereby limiting growthpromoting substrates and weakening malignant cells. A secondary but equally important aim is to **improve host metabolic resilience** by lowering insulin resistance, enhancing mitochondrial function,



and stabilizing redox balance. These improvements not only contribute to tumor control but also reduce the systemic toxicity of conventional therapies, mitigate cachexia, and improve quality of life.

Within an integrative orthomolecular model, metabolic therapies serve as the **foundation of cancer management**, upon which micronutrient, redox, immune, and lifestyle interventions can be layered.

#### 4.2.2. Ultra-Processed Foods (UPFs): A Dual Driver of Metabolic Stress and Toxic Exposure

While ketogenic diets, fasting, and metabolic monitoring provide powerful tools to exploit the tumor's glycolytic dependency, these strategies are constantly undermined by the pervasive global consumption of ultra-processed foods (UPFs). UPFs are not simply "empty calories"; they represent a toxic dietary system that simultaneously drives metabolic stress and introduces carcinogenic exposures[2]. Their elimination, therefore, is not optional but a non-negotiable intervention within integrative metabolic cancer management.

#### Defining UPFs

UPFs, as defined by the NOVA classification, are industrially manufactured formulations composed of refined starches, added sugars, seed oils, emulsifiers, preservatives, colorants, and flavor enhancers[11]. They can constitute the majority of calories in some high-income populations and are rapidly expanding into middle- and low-income nations, mirroring the global rise in obesity, diabetes, and cancer incidence[12–16].

#### Metabolic Impact

- Hyperglycemia and hyperinsulinemia: Refined starches and sugars create repeated glucose surges, feeding glycolysis and promoting chronic insulin signaling that favors tumor growth and increases overall cancer risk, while fructose—abundant in UPFs—specifically drives pancreatic cancer proliferation through the non-oxidative pentose phosphate pathway for nucleic acid synthesis[34–37].
- Metabolic inflexibility: UPFs suppress fatty acid oxidation and ketone production, locking
  metabolism into glycolytic dependence—the same pathway exploited by tumor cells (Warburg
  effect)[38–40].
- Obesity and inflammation: UPFs drive visceral adiposity, low-grade inflammation, and altered
  adipokine signaling, all of which accelerate tumorigenesis[26,27].
- Micronutrient dilution: Despite their caloric load, UPFs are deficient in protective nutrients such as vitamin D, vitamin C, magnesium, selenium, and omega-3 fatty acids. This compounds orthomolecular insufficiency and weakens mitochondrial and immune defenses.

#### Toxicological Burden

UPFs function not only as metabolic disruptors but also as **vectors of toxins**:

- Pesticide residues in grains, oils, and processed produce bioaccumulate with chronic intake.
- Endocrine disruptors (phthalates, BPA) leach from plastics and packaging.
- Chemical additives (emulsifiers, preservatives, sweeteners) disrupt gut microbiota, increase intestinal permeability, and promote chronic inflammation.
- Oxidized seed oils and AGEs (advanced glycation end-products) act as oxidative stressors, damaging DNA and proteins, and generating mutagenic byproducts.



#### Clinical Assessment

- Dietary history: Quantify daily caloric proportion derived from packaged, ready-to-eat, or fast foods.
- Biomarkers: Elevated fasting insulin, HOMA-IR, TG/HDL ratio, and GGT often reflect UPF-induced metabolic stress and hepatic burden.
- Functional measures: Microbiome imbalance and gut permeability testing can reveal downstream effects of UPF consumption.

#### Management - Elimination as Core Therapy

- Strict elimination of UPFs: Central to dietary intervention; without this step, neither ketogenic strategies nor micronutrient repletion can achieve full efficacy.
- Replacement with nutrient-dense whole foods: Pasture-raised animal proteins, low-carb
  vegetables, wild-caught fish, and orthomolecular supplementation.
- Ketogenic and fasting integration: Rebuilds metabolic flexibility, reverses insulin resistance, and restores mitochondrial resilience.
- Gut restoration: Anti-inflammatory diets, probiotics, and antioxidant-rich whole foods to repair intestinal barrier and microbiota disrupted by UPFs.

# Clinical Imperative

UPFs represent a **modern dietary toxin** at the intersection of metabolic and environmental cancer drivers. They simultaneously:

- 1. Provide the substrates that fuel tumor glycolysis.
- 2. Deliver a **toxicological payload** of pesticides, endocrine disruptors, and synthetic additives.
- 3. Displace the very nutrients needed to maintain redox stability, mitochondrial function, and immune surveillance.

For these reasons, the elimination of UPFs is not a lifestyle preference but a therapeutic mandate. Within the Integrative Orthomolecular Medicine (IOM) framework, the fight against cancer begins by cutting off this dual source of metabolic stress and toxic exposure, thereby restoring the biological terrain necessary for recovery and resilience

#### 4.3. Micronutrient Deficiencies – The Orthomolecular Foundation of Cancer Management

Among all root drivers of cancer, **micronutrient deficiency is uniquely fundamental**, because it directly compromises mitochondrial energy metabolism, antioxidant defenses, DNA repair, immune surveillance, and cellular differentiation. Whereas toxins and infections act as external stressors, and genetic/epigenetic factors shape susceptibility, it is nutrient deficiency that deprives cells of the essential biochemical "currency" to withstand and repair damage. Orthomolecular medicine, by definition, seeks to restore optimal concentrations of these molecules, thereby strengthening the body's capacity to resist, contain, and reverse malignant transformation.

#### 4.3.1. Assessment of Micronutrient Status

A comprehensive orthomolecular assessment includes:

- Core serum markers: 25(OH)D (Vitamin D3), plasma ascorbate (Vitamin C), serum magnesium, selenium, zinc, iodine, lipid profile with omega-3 index.
- Functional assays: Organic acids test for mitochondrial intermediates, oxidative stress panels (glutathione status, F2-isoprostanes, 8-OHdG), redox balance (GGT + ferritin pair).



Clinical context: History of long-term medication use (statins, metformin, PPIs, chemotherapy),
dietary recall (low intake of fresh animal foods, fish, and nutrient-dense vegetables), and
symptom clusters (fatigue, depression, immune suppression, bone fragility).

Deficiencies are nearly universal among cancer patients. Vitamin D insufficiency (<30 ng/mL) has been documented in 60–80% of cancer cohorts, low vitamin C status is common even in developed nations, and magnesium intake rarely exceeds two-thirds of the RDA, far below therapeutic needs.

#### 4.3.2. Mechanistic Rationale

- Vitamin C (ascorbate): Master antioxidant and electron donor; protects DNA from oxidative
  mutagenesis, supports collagen matrix integrity, and at pharmacologic IV doses generates
  hydrogen peroxide selectively cytotoxic to tumor cells.
- Vitamin D3: Regulates over 2,000 genes, including those controlling cell cycle, apoptosis, and immune function; deficiency promotes immune escape and unchecked proliferation.
- Vitamin K2 (MK-7): Directs calcium away from soft tissues and into bone, reducing vascular
  calcification and tumor-associated microcalcifications; synergistic with D3.
- Magnesium: Essential for >300 enzymes, stabilizes ATP, DNA repair enzymes, and mitochondrial oxidative phosphorylation; deficiency accelerates genomic instability.
- Niacin (Vitamin B3): Precursor of NAD+, central to mitochondrial respiration and DNA repair (PARP enzymes); high-dose niacin restores redox capacity and improves lipid and metabolic profiles.
- Selenium: Critical for glutathione peroxidase and thioredoxin reductase, neutralizing
  peroxides and supporting immune function; selenium deficiency is linked to higher cancer
  incidence.
- Omega-3 fatty acids (EPA/DHA): Compete with pro-inflammatory omega-6 fats, produce resolvins/protectins, stabilize membranes, and modulate oncogenic signaling pathways.
- **Zinc and Iodine:** Cofactors for DNA repair, thyroid metabolism, and antioxidant defense; deficiencies impair immune surveillance and endocrine regulation.

These micronutrients act not in isolation but as a **network of redox and metabolic cofactors**. Insufficiency in even one node destabilizes the system, compounding the effects of toxins, infections, and metabolic stressors.

#### 4.3.3. Management: Orthomolecular Repletion

Guided by TPIM's principles of safety, titration-to-target, and affordability, micronutrient repletion is prioritized for every cancer patient:

- Vitamin C: 5–15 g/day orally (in divided doses), plus high-dose intravenous vitamin C (HDIVC) up to 1,500 mg/kg body weight, administered 2–3 times per week in advanced cases[30,46–49,106]. Dosing should be titrated by bowel tolerance and plasma ascorbate monitoring, with screening for G6PD deficiency, renal impairment, and oxalate risk prior to initiation. Clinicians should also note that HDIVC can transiently interfere with point-of-care glucose meters, occasionally producing falsely elevated readings; confirm with laboratory glucose assays when clinically relevant.
- Vitamin D3: 5,000–30,000 IU/day, carefully titrated to maintain serum 25(OH)D levels between 50–100 ng/mL for the general population and up to 100–150 ng/mL in patients with autoimmune comorbidities. Such dosing should be conducted only under clinician supervision with

regular monitoring of serum calcium, renal function, and parathyroid hormone. Avoid calcium cosupplementation and pair with magnesium and vitamin K<sub>2</sub> for optimal balance and safety [102,107–111].

- Vitamin K2: Plays a key role in activating matrix Gla protein and osteocalcin, thereby helping
  prevent vascular calcification and supporting bone integrity—especially important in patients
  with ASCVD or concomitant bone loss. A dose as high as 45 mg/day has been suggested in clinical
  studies of osteoporosis, though the optimal formulation and dosing for cardiovascular protection remain
  under investigation[112–119].
- Magnesium: 400–800 mg/day (elemental; glycinate/threonate/citrate forms; titrate to bowel tolerance; adjust for CKD).
- Niacin (IR): 500–3,000 mg/day, titrated upward for lipid normalization, NAD+ replenishment, and mitochondrial support; manage flush with gradual escalation.
- **Selenium:** 200–400 μg/day (selenomethionine or sodium selenite).
- Omega-3 (EPA/DHA): 2–4 g/day, titrated to achieve Omega-3 index ≥8%.
- Zinc and Iodine: Individualized dosing, guided by labs and thyroid status.

Regular lab monitoring ensures safety, detects imbalances (e.g., selenium excess), and validates titration-to-target (e.g., Vit D3, Omega-3 index, redox biomarkers).

#### 4.3.4. High-Dose Intravenous Vitamin C (HDIVC): Hallmark of Orthomolecular Oncology

While broad micronutrient sufficiency forms the biological baseline, **HDIVC represents the flagship therapeutic tool of orthomolecular oncology**. By bypassing intestinal absorption limits, IV dosing achieves plasma levels 100–500 times higher than oral intake, unlocking unique pharmacologic effects.

# **Historical Context**

HDIVC has a long and distinguished history within orthomolecular medicine. Linus Pauling, Ph.D.—two-time Nobel laureate—and Scottish surgeon Ewan Cameron first advanced the use of intravenous vitamin C for cancer in the 1970s, reporting extended survival and improved quality of life in advanced cancer patients[28]. Their pioneering work was followed by decades of contributions from orthomolecular physicians including Abram Hoffer[29], Hugh Riordan[30], Ronald Hunninghake[31] and Thomas Levy[32], who refined dosing strategies, safety protocols, and mechanistic understanding.

Building on this foundation, modern investigators such as Mark Levine (NIH) and Qi Chen (University of Kansas) have clarified the pharmacokinetic differences between oral and intravenous ascorbate, demonstrating the tumor-selective cytotoxicity of HDIVC[33]. More recently, Baghli et al. (2024), in an ISOM consensus paper, highlighted targeting the mitochondrial–stem cell connection through a hybrid orthomolecular protocol[120]. In parallel, the Society for International Metabolic Oncology (SIMO), under the leadership of Thomas N. Seyfried, assembled an international consortium to outline a clinical research framework for ketogenic metabolic therapy in glioblastoma, including HDIVC and mitochondrial support[45].

# • Mechanisms:

- Generates extracellular hydrogen peroxide cytotoxic to cancer cells deficient in catalase.
- o Enhances mitochondrial respiration and redox stability in normal tissues.
- o Promotes collagen synthesis, reinforcing tissue barriers to invasion.
- Reduces inflammation and chemotherapy toxicity.
- Dosing: Typically 25–100 g per infusion, 2–3 times weekly; in advanced cases up to 1,500 mg/kg bodyweight[106].

- Safety: Decades of clinical use demonstrate an excellent profile, with contraindications limited to G6PD deficiency, renal insufficiency, and active oxalate nephropathy.
- Evidence: Preclinical studies consistently show tumor-selective toxicity[46,121]. Early-phase
  clinical trials report improved quality of life, reduced chemotherapy side effects, and signals of
  survival benefit[47–49].
  - HDIVC thus embodies the **Triple-Principle Intervention Model**:
- Safe (low toxicity, decades of use),
- Effective (selectively stresses tumor metabolism while protecting host),
- Accessible (low cost relative to targeted drugs).

#### 4.3.5. Clinical Integration

Micronutrient repletion is not an adjunct but a **cornerstone therapy**:

- It **corrects the biochemical insufficiency** that renders cells vulnerable to carcinogenesis.
- It enhances host resilience against chemotherapy and radiation toxicity.
- It **potentiates metabolic and immune therapies**, creating synergistic benefits.

In the IOM framework, orthomolecular sufficiency is the **biological baseline**—without it, no other intervention can reach its full potential. Cancer, therefore, must be understood not just as a disease of mutation and metabolism, but as a **disease of biochemical insufficiency compounded by toxic overload**.

- 4.4. Chronic Infections & Immune Dysregulation
- Assessment: Viral (HBV, HCV, EBV, HPV), bacterial, fungal or biofilm-related pathogens;
   labs—hsCRP, ESR, cytokines, autoantibodies.
- Management: Eradicate active infections; strengthen terrain with Vit D, Vit C, zinc, selenium; low-dose naltrexone (LDN); immune cell therapies (NK/T-cell, dendritic) where feasible.
- 4.5. Hormonal Imbalance & Endocrine Disruption
- Assessment: Thyroid (TSH, fT3, fT4, antibodies, rT3), sex hormones (E2, P, T, DHEA, SHBG), adrenal function (cortisol rhythm via saliva), melatonin.
- Management: BHRT (bioidentical hormones for thyroid, adrenal, sex hormones), circadian rhythm alignment (light/dark therapy, sleep hygiene), melatonin (10–100 mg).
- 4.6. Lifestyle & Behavioral Risk Factors
- Assessment: Exercise level, sleep quality, alcohol use, smoking, screen time, stress load.
- Management: Outdoor exercise with sunlight exposure, progressive strength training, sleep
  optimization, alcohol/smoking cessation, structured breathing and relaxation practices.
- 4.7. Psychosocial & Emotional Stress
- Assessment: Chronic stress, trauma history, depression, anxiety, HRV monitoring.



- Management: Mindfulness, cognitive behavioral therapy, resilience training, community/social support, adaptogens (ashwagandha, rhodiola), omega-3s, high-dose B vitamins.
- 4.8. Developmental & Early-Life Programming
- Assessment: Birth history, early malnutrition, toxin/vaccine exposures, childhood illnesses, methylation/epigenetic testing if available.
- Management: Lifelong orthomolecular nutrition, detox support, mitochondrial repair (CoQ10, carnitine, NAD precursors), regenerative therapies (stem cells, PBMT).
- 4.9. Genetic & Epigenetic Susceptibility
- Assessment: Family history of cancer, germline mutations, somatic drivers (NGS panels), methylation profiles.
- Management: Personalized nutrition (e.g., methyl donors in MTHFR), fasting, red-blue PBMT therapy, polyphenols as epigenetic modulators, exercise and hormesis.
- 4.10. Iatrogenic Injury
- Assessment: History of chemotherapy, radiation, chronic drug use (statins, PPIs, metformin, steroids), nutrient-depletion side effects.
- Management: Corrective orthomolecular support (e.g., CoQ10 with statins, B12 with metformin, Mg with diuretics), integrative oncology protocols to minimize side effects, patient education on medication risks.
- 4.11. Restoration of Mitochondrial Function: Reversing Malignancy

Mitochondrial dysfunction is characteristic of many cancers—driving metabolic rewiring, apoptosis resistance, oxidative stress imbalance, and therapy resistance. Restoring mitochondrial function (reactivating oxidative phosphorylation, or OXPHOS) can reduce malignancy, promote apoptosis, and increase treatment sensitivity, effectively reversing key malignant traits. Cancer cells typically display hyperpolarized mitochondria, altered calcium flux, reduced ROS signaling, and impaired apoptosis; restoration of function reactivates OXPHOS, normalizes redox signaling, and resensitizes cells to programmed cell death[17]. Because electron-transport integrity and mitochondrial DNA (mtDNA) stability are critical for tumor progression, loss of mitochondrial function can act as a metabolic checkpoint; conversely, restoring mitochondrial quality can constrain tumor growth[18,19]. Cancer stem cells also rely heavily on OXPHOS, so targeting mitochondrial metabolism in these cells reduces survival and aggressiveness, with downstream effects on differentiation state, metastatic potential, and immune interactions[10,20,21].

Mechanistic and translational exemplars reinforce this therapeutic angle. Dichloroacetate (DCA) inhibits pyruvate dehydrogenase kinase, shifts metabolism from glycolysis toward mitochondrial oxidation, and can reduce tumor growth while restoring apoptosis in therapy-resistant models[17]. In a direct test of causality, transplantation of normal mitochondria from mammary epithelial cells into human breast cancer cells inhibited proliferation dose-dependently and increased sensitivity to doxorubicin, Abraxane, and carboplatin[122]. Likewise, cancer cells lacking mtDNA showed delayed tumor growth until host mtDNA was acquired to restore respiration and tumorigenic potential[123]. More broadly, modulating mitochondrial respiration can induce

proliferation arrest, differentiation, or death, underscoring the tractability of mitochondrial manipulation as an anticancer strategy[124].

Beyond metabolic switches, **novel modalities** target mitochondrial structure–function coupling. **Mitochondrial transplantation** has shown promise in drug-resistant settings (e.g., TNBC) by restoring bioenergetics and improving mitochondrial dynamics (fusion via MFN1/OPA1), reorganizing the network to enhance metabolism and apoptotic responsiveness, and thereby reducing tumor growth while improving chemotherapy sensitivity[17–19]. Emerging **mitochondriatargeting organic sensitizers (MTOSs)** disrupt mitochondrial homeostasis (membrane potential, ROS generation, ETC integrity), induce regulated cell-death programs, and promote release of DAMPs to activate antitumor immunity—though clinical translation will require better delivery and biocompatibility within heterogeneous tumors[20]. Collectively, **mitochondrial restoration and modulation** (via metabolic reprogramming, organelle transfer, and targeted sensitizers) provide convergent routes to reverse malignant phenotypes and overcome resistance[10,17–21].

#### Red-Blue Therapy in IOCT.

Red–Blue Therapy (near infrared light + methylene blue) exemplifies how integrative orthomolecular cancer therapy incorporates **biophysical and biochemical modalities** into a unified mitochondrial-restoration framework. Within PBMT, two wavelengths are particularly well studied: **660 nm red light**, which penetrates a few millimeters and optimally activates **cytochrome c oxidase** (**Complex IV**) in superficial tissues, and **850 nm near-infrared light**, which penetrates deeper into muscle, brain, and thoracic organs to stimulate mitochondrial respiration and microcirculatory dynamics. Together, these bands span the "optical window" for tissue penetration, with evidence of enhanced **ATP generation**, **mitochondrial membrane potential recovery**, **and hormetic ROS modulation**.

**Methylene blue (MB)** complements these effects by acting as a **redox shuttle** within the electron transport chain, bypassing Complex I and III bottlenecks, restoring NAD+/NADH ratios, and limiting electron leak that drives oxidative stress. MB also broadens light absorption, functioning as a **photosensitizer**, so PBMT at 660 nm and 850 nm enhances MB's cycling between reduced and oxidized forms—yielding synergistic improvements in OXPHOS and redox stability.

#### Clinical implications.

- Supportive oncology: PBMT at 660/850 nm is guideline-endorsed for oral mucositis and has
  demonstrated benefit for neuropathic pain, wound healing, and fatigue in cancer patients, per
  MASCC/ISOO clinical practice guidelines[125].
- Neurocognitive and fatigue support: MB at low doses (e.g., <1-2 mg/kg has shown
  mitochondrial and cognitive benefits in neurodegeneration and may reduce "chemo-brain" or
  treatment-related fatigue when combined with PBMT[126–132].</li>
- Integration with IOCT: Red-Blue Therapy can be combined with HDIVC, ketogenic metabolic therapy, niacin, CoQ10, magnesium, and lifestyle-based mitochondrial conditioning to consolidate OXPHOS resilience, terrain optimization, and therapy tolerance.

# TPIM alignment.

- Safety: Both PBMT (when delivered at appropriate fluence and with eye protection) and low-dose MB have favorable long-term safety records, with well-defined contraindications (e.g., serotonergic medications, G6PD deficiency).
- Effectiveness: Improvements in fatigue, neuropathy, cognitive function, and mitochondrial biomarkers are measurable and titratable.

 Affordability: PBMT devices emitting 660/850 nm wavelengths are increasingly accessible, and MB is inexpensive at therapeutic doses compared to conventional biologics.

In sum, mitochondrial restoration represents a unifying therapeutic axis in Integrative Orthomolecular Cancer Therapy (IOCT). Normalizing mitochondrial function decreases cancer cell malignancy by reinstating apoptosis, re-establishing metabolic control, and enhancing therapy sensitivity [10,17–21,122–124,133]. Within this framework, **Red–Blue Therapy**—the combination of 660 nm red and 850 nm near-infrared light with **methylene blue**—extends the IOCT mitochondrial toolkit by safely enhancing OXPHOS, stabilizing redox signaling, and improving host resilience. Early translational and supportive-care data justify its structured evaluation in adaptive clinical trials and its integration alongside HDIVC and metabolic therapy as part of a mitochondria-centered cancer-terrain strategy.

#### 4.12. Integrated Care Model

Cancer patients rarely have a single dominant driver; **multiple drivers interact synergistically**. A patient-specific IOM plan requires:

- 1. **Mapping all 10 categories** via labs, imaging, and history.
- 2. **Prioritizing interventions** based on safety, impact, and feasibility.
- 3. **Titrating therapies-to-target** (e.g., GKI, Vit D3 levels).
- 4. Continuous monitoring with labs, imaging, and symptom feedback.

This structured approach transforms cancer care from a **tumor-centered war** into a **host-centered restoration strategy**, integrating metabolic, nutritional, and regenerative therapies to support remission and long-term resilience.

# 5. Research and Trial Implications: TP-ARC

The **Triple-Principle Adaptive RCT (TP-ARC)** expands TPIM into a research model. It differs from conventional trials by:

- Testing **programs**, not isolated agents
- Allowing titration and personalization
- Embedding safety, effectiveness, and affordability metrics
- Using real-world populations with comorbidities
- Measuring survival, quality of life, and cost rather than short-term surrogates

TP-ARC thus provides a reproducible yet pragmatic pathway to validate integrative cancer management strategies.

# 6. Discussion

By combining **root cause analysis** with **triple-principle intervention**, cancer is reframed as a **preventable and modifiable systems failure**. While genetics and advanced therapies have their place, the majority of upstream drivers remain environmental, nutritional, and metabolic—domains amenable to intervention.

Integrative metabolic therapy offers both **tumor control** and **host protection**, minimizing toxicity while maximizing resilience. The TPIM framework ensures that interventions remain safe,

titrated to effect, and broadly accessible. The TP-ARC research design ensures that such strategies can be tested rigorously and scaled globally.

#### 7. Conclusions

Cancer care must evolve beyond late-stage, mutation-focused rescue toward upstream, mitochondria-centered prevention and management. The combination of root cause analysis, triple-principle intervention, and adaptive clinical trials offers a blueprint for transforming oncology into a discipline grounded in safety, effectiveness, and sustainability.

**Author Contributions:** R.Z.C. conceptualized the study, conducted the literature review, and drafted and finalized the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable.

Conflicts of Interest: R.Z.C. is the founder of Cheng Integrative Health Center (USA) and Cheng Health Consulting Ltd. (Shanghai), and serves as Editor-in-Chief of the Orthomolecular Medicine News Service (OMNS). He also develops nutraceutical formulations, including several nutraceutical formulations (e.g., ImmuneBoost, Yutrix, Liver Detox, Whole Cell Nutrition). The author declares that these interests did not influence the content or conclusions of this manuscript. The author's product lines include formulations overlapping with interventions discussed (e.g., detox and micronutrient formulas).

Intellectual Property Statement: The Triple-Principle Intervention Model (TPIM)™ is a proprietary clinical framework developed by R.Z.C. and has been filed for trademark registration with the United States Patent and Trademark Office (USPTO).

**Conceptual Continuity Statement:** The **Triple-Principle Intervention Model (TPIM)**™ evolved from the author's earlier *Toolkit Approach* framework first described in the *Orthomolecular Medicine News Service* article "A Toolkit for COVID-19 Management and Future Epidemics"[3].

# References

- Cheng, R.Z. From Mutation to Metabolism: Root Cause Analysis of Cancer's Initiating Drivers. 2025. DOI: 10.20944/preprints202509.0903.v1; Available online: https://www.preprints.org/manuscript/202509.0903/v1.
- Cheng, R.Z. From Mutation to Metabolism: Environmental and Dietary Toxins as Upstream Drivers of Mitochondrial Dysfunction and Chronic Disease. 2025. DOI: 10.20944/preprints202509.1767.v1; Available online: https://www.preprints.org/manuscript/202509.1767/v1.
- Cheng, R.Z. A Paradigm Shift in Epidemic and Chronic Disease Management. Orthomolecular Medicine News Service, 2024. v20, n23. Available online: https://orthomolecular.org/resources/omns/v20n23.shtml (accessed 13 March 2025).
- Seyfried, T.N.; Flores, R.E.; Poff, A.M.; D'Agostino, D.P. Cancer as a Metabolic Disease: Implications for Novel Therapeutics. *Carcinogenesis* 2014, 35, (3), 515–527. DOI: 10.1093/carcin/bgt480.
- Seyfried, T.N. Cancer as a Metabolic Disease: On the Origin, Management, and Prevention of Cancer; Wiley, 2012.
   Available online: https://www.wiley.com/en-us/Cancer+as+a+Metabolic+Disease%3A+On+the+Origin%2C+Management%2C+and+Prevention+of+Cancer-p-9780470584927.
- Zhao, Y.; Liu, H.; Riker, A.I.; Fodstad, O.; Ledoux, S.P.; Wilson, G.L.; Tan, M. Emerging Metabolic Targets in Cancer Therapy. Front Biosci (Landmark Ed) 2011, 16, (5), 1844–1860. DOI: 10.2741/3826.

- 7. Pan, J.G.; Mak, T.W. Metabolic Targeting as an Anticancer Strategy: Dawn of a New Era? *Sci STKE* 2007, 2007, (381), pe14. DOI: 10.1126/stke.3812007pe14.
- 8. Dang, C.V.; Hamaker, M.; Sun, P.; Le, A.; Gao, P. Therapeutic Targeting of Cancer Cell Metabolism. *J Mol Med (Berl)* 2011, 89, (3), 205–212. DOI: 10.1007/s00109-011-0730-x.
- 9. Seyfried, T.N. Cancer as a Mitochondrial Metabolic Disease. *Front Cell Dev Biol* 2015, 3, 43. DOI: 10.3389/fcell.2015.00043.
- 10. Porporato, P.E.; Filigheddu, N.; Pedro, J.M.B.-S.; Kroemer, G.; Galluzzi, L. Mitochondrial Metabolism and Cancer. *Cell Res* 2018, 28, (3), 265–280. DOI: 10.1038/cr.2017.155.
- 11. Monteiro, C.A.; Cannon, G.; Moubarac, J.-C.; Levy, R.B.; Louzada, M.L.C.; Jaime, P.C. The UN Decade of Nutrition, the NOVA Food Classification and the Trouble with Ultra-Processing. *Public Health Nutr* 2018, 21, (1), 5–17. DOI: 10.1017/S1368980017000234.
- Baker, P.; Machado, P.; Santos, T.; Sievert, K.; Backholer, K.; Hadjikakou, M.; Russell, C.; Huse, O.; Bell, C.; Scrinis, G.; Worsley, A.; Friel, S.; Lawrence, M. Ultra-Processed Foods and the Nutrition Transition: Global, Regional and National Trends, Food Systems Transformations and Political Economy Drivers. *Obes Rev* 2020, 21, (12), e13126. DOI: 10.1111/obr.13126.
- Martini, D.; Godos, J.; Bonaccio, M.; Vitaglione, P.; Grosso, G. Ultra-Processed Foods and Nutritional Dietary Profile: A Meta-Analysis of Nationally Representative Samples. *Nutrients* 2021, 13, (10), 3390. DOI: 10.3390/nu13103390.
- 14. Monteiro, C.A.; Moubarac, J.-C.; Cannon, G.; Ng, S.W.; Popkin, B. Ultra-Processed Products Are Becoming Dominant in the Global Food System. *Obes Rev* 2013, 14 *Suppl* 2, 21–28. DOI: 10.1111/obr.12107.
- 15. Martínez Steele, E.; Juul, F.; Neri, D.; Rauber, F.; Monteiro, C.A. Dietary Share of Ultra-Processed Foods and Metabolic Syndrome in the US Adult Population. *Prev Med* 2019, 125, 40–48. DOI: 10.1016/j.ypmed.2019.05.004.
- Anastasiou, I.A.; Kounatidis, D.; Vallianou, N.G.; Skourtis, A.; Dimitriou, K.; Tzivaki, I.; Tsioulos, G.; Rigatou, A.; Karampela, I.; Dalamaga, M. Beneath the Surface: The Emerging Role of Ultra-Processed Foods in Obesity-Related Cancer. *Curr Oncol Rep* 2025, 27, (4), 390–414. DOI: 10.1007/s11912-025-01654-6.
- 17. Bhat, T.A.; Kumar, S.; Chaudhary, A.K.; Yadav, N.; Chandra, D. Restoration of Mitochondria Function as a Target for Cancer Therapy. *Drug Discov Today* 2015, 20, (5), 635–643. DOI: 10.1016/j.drudis.2015.03.001.
- 18. Vasan, K.; Werner, M.; Chandel, N.S. Mitochondrial Metabolism as a Target for Cancer Therapy. *Cell Metab* 2020, 32, (3), 341–352. DOI: 10.1016/j.cmet.2020.06.019.
- 19. Miao, X.; Jiang, P.; Wang, Z.; Kong, W.; Feng, L. Mitochondrial Transplantation: A Novel Therapeutic Approach for Treating Diseases. *MedComm* (2020) 2025, 6, (6), e70253. DOI: 10.1002/mco2.70253.
- 20. Liu, Y.; Sun, Y.; Guo, Y.; Shi, X.; Chen, X.; Feng, W.; Wu, L.-L.; Zhang, J.; Yu, S.; Wang, Y.; Shi, Y. An Overview: The Diversified Role of Mitochondria in Cancer Metabolism. *Int J Biol Sci* 2023, *19*, (3), 897–915. DOI: 10.7150/ijbs.81609.
- Li, Z.; Zhang, W.; Guo, S.; Qi, G.; Huang, J.; Gao, J.; Zhao, J.; Kang, L.; Li, Q. A Review of Advances in Mitochondrial Research in Cancer. Cancer Control 2024, 31, 10732748241299072. DOI: 10.1177/10732748241299072.
- 22. Henkler, F.; Brinkmann, J.; Luch, A. The Role of Oxidative Stress in Carcinogenesis Induced by Metals and Xenobiotics. *Cancers (Basel)* 2010, 2, (2), 376–396. DOI: 10.3390/cancers2020376.
- 23. Talha; Zahoor, U.; Azhar, S.; Yasin, A.; Saleem, S.; Yousaf, S.; Nawaz, I.; Khanzada, A.; Aslam, M.A. Oxidative Stress Pathways in Cancer: Lessons Learned from Heavy Metal Exposures: Oxidative Stress and Heavy Metals. *Journal of Health and Rehabilitation Research* 2024, 4, (3), 1–6. DOI: 10.61919/jhrr.v4i3.1703; Available online: https://jhrlmc.com/index.php/home/article/view/1703.
- 24. Patrick, L. Toxic Metals and Antioxidants: Part II. The Role of Antioxidants in Arsenic and Cadmium Toxicity. *Altern Med Rev* 2003, *8*, (2), 106–128.
- 25. Koedrith, P.; Seo, Y.R. Advances in Carcinogenic Metal Toxicity and Potential Molecular Markers. *Int J Mol Sci* 2011, 12, (12), 9576–9595. DOI: 10.3390/ijms12129576.
- Konieczna, J.; Morey, M.; Abete, I.; Bes-Rastrollo, M.; Ruiz-Canela, M.; Vioque, J.; Gonzalez-Palacios, S.; Daimiel, L.; Salas-Salvadó, J.; Fiol, M.; Martín, V.; Estruch, R.; Vidal, J.; Martínez-González, M.A.; Canudas, S.; Jover, A.J.; Fernández-Villa, T.; Casas, R.; Olbeyra, R.; Buil-Cosiales, P.; Babio, N.; Schröder, H.; Martínez,

- J.A.; Romaguera, D.; PREDIMED-Plus investigators Contribution of Ultra-Processed Foods in Visceral Fat Deposition and Other Adiposity Indicators: Prospective Analysis Nested in the PREDIMED-Plus Trial. *Clin Nutr* 2021, 40, (6), 4290–4300. DOI: 10.1016/j.clnu.2021.01.019.
- 27. Donohoe, C.L.; Doyle, S.L.; Reynolds, J.V. Visceral Adiposity, Insulin Resistance and Cancer Risk. *Diabetol Metab Syndr* 2011, *3*, 12. DOI: 10.1186/1758-5996-3-12.
- Cameron, E.; Pauling, L. Ascorbic Acid and the Glycosaminoglycans. An Orthomolecular Approach to Cancer and Other Diseases. *Oncology* 1973, 27, (2), 181–192. DOI: 10.1159/000224733.
- Hoffer, A. Orthomolecular Treatment of Cancer. In *Nutrients in Cancer Prevention and Treatment*eds.: K.N. Prasad, L. Santamaria, R.M. Williams, Humana Press, Totowa, NJ, 1995, pp. 373–391. DOI: 10.1007/978-1-4612-0237-0\_23; Available online: http://link.springer.com/10.1007/978-1-4612-0237-0\_23.
- Riordan, H.D.; Hunninghake, R.B.; Riordan, N.H.; Jackson, J.J.; Meng, X.; Taylor, P.; Casciari, J.J.; González, M.J.; Miranda-Massari, J.R.; Mora, E.M.; Rosario, N.; Rivera, A. Intravenous Ascorbic Acid: Protocol for Its Application and Use. P R Health Sci J 2003, 22, (3), 287–290.
- 31. Riordan, H.D.; Riordan, N.H.; Jackson, J.A.; Casciari, J.J.; Hunninghake, R.; González, M.J.; Mora, E.M.; Miranda-Massari, J.R.; Rosario, N.; Rivera, A. Intravenous Vitamin C as a Chemotherapy Agent: A Report on Clinical Cases. *P R Health Sci J* 2004, 23, (2), 115–118.
- 32. Levy, T.E. *A Guide to the Optimal Administration of Vitamin C*; MedFox Publishing, 2022. Available online: https://orthomolecular.org/resources/omns/Thomas-Levy-Guide-To-The-Optimal-Administration-of-Vitamin-C.pdf?
- Chen, Q.; Espey, M.G.; Krishna, M.C.; Mitchell, J.B.; Corpe, C.P.; Buettner, G.R.; Shacter, E.; Levine, M. Pharmacologic Ascorbic Acid Concentrations Selectively Kill Cancer Cells: Action as a pro-Drug to Deliver Hydrogen Peroxide to Tissues. *Proc. Natl. Acad. Sci. U.S.A.* 2005, 102, (38), 13604–13609. DOI: 10.1073/pnas.0506390102.
- 34. Isaksen, I.M.; Dankel, S.N. Ultra-Processed Food Consumption and Cancer Risk: A Systematic Review and Meta-Analysis. *Clin Nutr* 2023, 42, (6), 919–928. DOI: 10.1016/j.clnu.2023.03.018.
- Venkateswaran, V.; Haddad, A.Q.; Fleshner, N.E.; Fan, R.; Sugar, L.M.; Nam, R.; Klotz, L.H.; Pollak, M. Association of Diet-Induced Hyperinsulinemia with Accelerated Growth of Prostate Cancer (LNCaP) Xenografts. J Natl Cancer Inst 2007, 99, (23), 1793–1800. DOI: 10.1093/jnci/djm231.
- Liu, H.; Huang, D.; McArthur, D.L.; Boros, L.G.; Nissen, N.; Heaney, A.P. Fructose Induces Transketolase
  Flux to Promote Pancreatic Cancer Growth. Cancer Res 2010, 70, (15), 6368–6376. DOI: 10.1158/00085472.CAN-09-4615.
- 37. Hirabayashi, S.; Baranski, T.J.; Cagan, R.L. Transformed Drosophila Cells Evade Diet-Mediated Insulin Resistance through Wingless Signaling. *Cell* 2013, *154*, (3), 664–675. DOI: 10.1016/j.cell.2013.06.030.
- Handakas, E.; Chang, K.; Khandpur, N.; Vamos, E.P.; Millett, C.; Sassi, F.; Vineis, P.; Robinson, O. Metabolic Profiles of Ultra-Processed Food Consumption and Their Role in Obesity Risk in British Children. Clin Nutr 2022, 41, (11), 2537–2548. DOI: 10.1016/j.clnu.2022.09.002.
- 39. O'Connor, L.E.; Hall, K.D.; Herrick, K.A.; Reedy, J.; Chung, S.T.; Stagliano, M.; Courville, A.B.; Sinha, R.; Freedman, N.D.; Hong, H.G.; Albert, P.S.; Loftfield, E. Metabolomic Profiling of an Ultraprocessed Dietary Pattern in a Domiciled Randomized Controlled Crossover Feeding Trial. *J Nutr* 2023, 153, (8), 2181–2192. DOI: 10.1016/j.tjnut.2023.06.003.
- Harlan, T.S.; Gow, R.V.; Kornstädt, A.; Alderson, P.W.; Lustig, R.H. The Metabolic Matrix: Re-Engineering Ultraprocessed Foods to Feed the Gut, Protect the Liver, and Support the Brain. Front Nutr 2023, 10, 1098453. DOI: 10.3389/fnut.2023.1098453.
- Barrea, L.; Caprio, M.; Tuccinardi, D.; Moriconi, E.; Di Renzo, L.; Muscogiuri, G.; Colao, A.; Savastano, S.;
   Obesity Programs of nutrition, Education, Research and Assessment (OPERA) group Could Ketogenic Diet
   "Starve" Cancer? Emerging Evidence. Crit Rev Food Sci Nutr 2022, 62, (7), 1800–1821. DOI:
   10.1080/10408398.2020.1847030.
- 42. Weber, D.D.; Aminzadeh-Gohari, S.; Tulipan, J.; Catalano, L.; Feichtinger, R.G.; Kofler, B. Ketogenic Diet in the Treatment of Cancer Where Do We Stand? *Mol Metab* 2020, 33, 102–121. DOI: 10.1016/j.molmet.2019.06.026.

- 43. Woolf, E.C.; Scheck, A.C. The Ketogenic Diet for the Treatment of Malignant Glioma. *J Lipid Res* 2015, *56*, (1), 5–10. DOI: 10.1194/jlr.R046797.
- 44. Cheng, R.Z. RCTs: The Foundational Flaw of Modern Medicine? Orthomolecular Medicine News Service. Available online: https://orthomolecular.org/resources/omns/index.shtml.
- 45. Duraj, T.; Kalamian, M.; Zuccoli, G.; Maroon, J.C.; D'Agostino, D.P.; Scheck, A.C.; Poff, A.; Winter, S.F.; Hu, J.; Klement, R.J.; Hickson, A.; Lee, D.C.; Cooper, I.; Kofler, B.; Schwartz, K.A.; Phillips, M.C.L.; Champ, C.E.; Zupec-Kania, B.; Tan-Shalaby, J.; Serfaty, F.M.; Omene, E.; Arismendi-Morillo, G.; Kiebish, M.; Cheng, R.; El-Sakka, A.M.; Pflueger, A.; Mathews, E.H.; Worden, D.; Shi, H.; Cincione, R.I.; Spinosa, J.P.; Slocum, A.K.; Iyikesici, M.S.; Yanagisawa, A.; Pilkington, G.J.; Chaffee, A.; Abdel-Hadi, W.; Elsamman, A.K.; Klein, P.; Hagihara, K.; Clemens, Z.; Yu, G.W.; Evangeliou, A.E.; Nathan, J.K.; Smith, K.; Fortin, D.; Dietrich, J.; Mukherjee, P.; Seyfried, T.N. Clinical Research Framework Proposal for Ketogenic Metabolic Therapy in Glioblastoma. BMC Med 2024, 22, (1), 578. DOI: 10.1186/s12916-024-03775-4.
- 46. Padayatty, S.J.; Sun, A.Y.; Chen, Q.; Espey, M.G.; Drisko, J.; Levine, M. Vitamin C: Intravenous Use by Complementary and Alternative Medicine Practitioners and Adverse Effects. *PLoS One* 2010, *5*, (7), e11414. DOI: 10.1371/journal.pone.0011414.
- 47. Giansanti, M.; Karimi, T.; Faraoni, I.; Graziani, G. High-Dose Vitamin C: Preclinical Evidence for Tailoring Treatment in Cancer Patients. *Cancers* (*Basel*) 2021, *13*, (6), 1428. DOI: 10.3390/cancers13061428.
- 48. Böttger, F.; Vallés-Martí, A.; Cahn, L.; Jimenez, C.R. High-Dose Intravenous Vitamin C, a Promising Multi-Targeting Agent in the Treatment of Cancer. *J Exp Clin Cancer Res* 2021, 40, (1), 343. DOI: 10.1186/s13046-021-02134-v.
- 49. Ngo, B.; Van Riper, J.M.; Cantley, L.C.; Yun, J. Targeting Cancer Vulnerabilities with High-Dose Vitamin C. *Nat Rev Cancer* 2019, *19*, (5), 271–282. DOI: 10.1038/s41568-019-0135-7.
- 50. Ennulat, D.; Magid-Slav, M.; Rehm, S.; Tatsuoka, K.S. Diagnostic Performance of Traditional Hepatobiliary Biomarkers of Drug-Induced Liver Injury in the Rat. *Toxicol Sci* 2010, 116, (2), 397–412. DOI: 10.1093/toxsci/kfq144.
- 51. Kwo, P.Y.; Cohen, S.M.; Lim, J.K. ACG Clinical Guideline: Evaluation of Abnormal Liver Chemistries. *Am J Gastroenterol* 2017, 112, (1), 18–35. DOI: 10.1038/ajg.2016.517.
- 52. Luo, L.; Schomaker, S.; Houle, C.; Aubrecht, J.; Colangelo, J.L. Evaluation of Serum Bile Acid Profiles as Biomarkers of Liver Injury in Rodents. *Toxicol Sci* 2014, 137, (1), 12–25. DOI: 10.1093/toxsci/kft221.
- 53. Mohapatra, E.; Priya, R.; Nanda, R.; Patel, S. Serum GGT and Serum Ferritin as Early Markers for Metabolic Syndrome. *J Family Med Prim Care* 2020, *9*, (7), 3458–3463. DOI: 10.4103/jfmpc.jfmpc\_570\_20.
- 54. Wei, D.; Chen, T.; Li, J.; Gao, Y.; Ren, Y.; Zhang, X.; Yu, H.; Tian, H. Association of Serum Gamma-Glutamyl Transferase and Ferritin with the Metabolic Syndrome. *J Diabetes Res* 2015, 2015, 741731. DOI: 10.1155/2015/741731.
- 55. Koenig, G.; Seneff, S. Gamma-Glutamyltransferase: A Predictive Biomarker of Cellular Antioxidant Inadequacy and Disease Risk. *Dis Markers* 2015, 2015, 818570. DOI: 10.1155/2015/818570.
- Meidenbauer, J.J.; Mukherjee, P.; Seyfried, T.N. The Glucose Ketone Index Calculator: A Simple Tool to Monitor Therapeutic Efficacy for Metabolic Management of Brain Cancer. *Nutrition & metabolism* 2015, 12,
   DOI: 10.1186/s12986-015-0009-2; Available online: http://www.ncbi.nlm.nih.gov/pubmed/25798181 http://www.pubmedcentral.nih.gov/articlerender.fcgi?artid=PMC4367849.
- 57. Akgoc, Z.; Mukherjee, P.; Seyfried, T.N. The Glucose Ketone Index Predicts Overall Survival and Metastasis of Mouse Tumor Cells to Visceral Organs and Brain. *Longhua Chinese Medicine* 2022, 5, (0). DOI: 10.21037/lcm-21-43; Available online: https://lcm.amegroups.org/article/view/8366.
- Seyfried, T.N.; Mukherjee, P.; Iyikesici, M.S.; Slocum, A.; Kalamian, M.; Spinosa, J.-P.; Chinopoulos, C. Consideration of Ketogenic Metabolic Therapy as a Complementary or Alternative Approach for Managing Breast Cancer. Front Nutr 2020, 7, 21. DOI: 10.3389/fnut.2020.00021.
- 59. Phillips, M.C.L.; Thotathil, Z.; Dass, P.H.; Ziad, F.; Moon, B.G. Ketogenic Metabolic Therapy in Conjunction with Standard Treatment for Glioblastoma: A Case Report. *Oncol Lett* 2024, 27, (5), 230. DOI: 10.3892/ol.2024.14363.
- 60. Rashmi, B.M.; Pattanshetty, S.; Bhagyajyoti; Savadi, B.V.; Prashanth, G. Association of Serum Ferritin and Gamma-Glutamyl Transferase Levels with Metabolic Syndrome and Insulin Resistance. *Dentistry and*

- Medical Research 2023, 11, (1), 26. DOI: 10.4103/dmr.dmr\_28\_23; Available online https://journals.lww.com/dmrs/fulltext/2023/11010/association\_of\_serum\_ferritin\_and\_gamma\_glutamyl. 6.aspx.
- 61. Fuller, R.; Landrigan, P.J.; Balakrishnan, K.; Bathan, G.; Bose-O'Reilly, S.; Brauer, M.; Caravanos, J.; Chiles, T.; Cohen, A.; Corra, L.; Cropper, M.; Ferraro, G.; Hanna, J.; Hanrahan, D.; Hu, H.; Hunter, D.; Janata, G.; Kupka, R.; Lanphear, B.; Lichtveld, M.; Martin, K.; Mustapha, A.; Sanchez-Triana, E.; Sandilya, K.; Schaefli, L.; Shaw, J.; Seddon, J.; Suk, W.; Téllez-Rojo, M.M.; Yan, C. Pollution and Health: A Progress Update. *Lancet Planet Health* 2022, 6, (6), e535–e547. DOI: 10.1016/S2542-5196(22)00090-0.
- 62. Landrigan, P.J.; Fuller, R.; Acosta, N.J.R.; Adeyi, O.; Arnold, R.; Basu, N.N.; Baldé, A.B.; Bertollini, R.; Bose-O'Reilly, S.; Boufford, J.I.; Breysse, P.N.; Chiles, T.; Mahidol, C.; Coll-Seck, A.M.; Cropper, M.L.; Fobil, J.; Fuster, V.; Greenstone, M.; Haines, A.; Hanrahan, D.; Hunter, D.; Khare, M.; Krupnick, A.; Lanphear, B.; Lohani, B.; Martin, K.; Mathiasen, K.V.; McTeer, M.A.; Murray, C.J.L.; Ndahimananjara, J.D.; Perera, F.; Potočnik, J.; Preker, A.S.; Ramesh, J.; Rockström, J.; Salinas, C.; Samson, L.D.; Sandilya, K.; Sly, P.D.; Smith, K.R.; Steiner, A.; Stewart, R.B.; Suk, W.A.; Schayck, O.C.P. van; Yadama, G.N.; Yumkella, K.; Zhong, M. The Lancet Commission on Pollution and Health. *Lancet* 2018, 391, (10119), 462–512. DOI: 10.1016/S0140-6736(17)32345-0.
- 63. Baron, P.; Schweinsberg, F. [A study of the literature on the concentrations of arsenic, lead, cadmium and mercury in body fluids and tissues to define normal values and detection of overload. 1. Description of analytical methods and arsenic]. Zentralbl Bakteriol Mikrobiol Hyg B Umwelthyg Krankenhaushyg Arbeitshyg Prav Med 1988, 186, (4), 289–310.
- 64. Filipoiu, D.C.; Bungau, S.G.; Endres, L.; Negru, P.A.; Bungau, A.F.; Pasca, B.; Radu, A.-F.; Tarce, A.G.; Bogdan, M.A.; Behl, T.; Nechifor, A.C.; Hassan, S.S.U.; Tit, D.M. Characterization of the Toxicological Impact of Heavy Metals on Human Health in Conjunction with Modern Analytical Methods. *Toxics* 2022, 10, (12), 716. DOI: 10.3390/toxics10120716.
- 65. Keil, D.E.; Berger-Ritchie, J.; McMillin, G.A. Testing for Toxic Elements: A Focus on Arsenic, Cadmium, Lead, and Mercury.; Available online: https://dx.doi.org/10.1309/LMYKGU05BEPE7IAW.
- 66. Esteban, M.; Castaño, A. Non-Invasive Matrices in Human Biomonitoring: A Review. *Environ Int* 2009, 35, (2), 438–449. DOI: 10.1016/j.envint.2008.09.003.
- Dorgan, J.F.; Brock, J.W.; Rothman, N.; Needham, L.L.; Miller, R.; Stephenson, H.E.; Schussler, N.; Taylor,
   P.R. Serum Organochlorine Pesticides and PCBs and Breast Cancer Risk: Results from a Prospective Analysis (USA). *Cancer Causes Control* 1999, 10, (1), 1–11. DOI: 10.1023/a:1008824131727.
- 68. Porta, M.; Bosch de Basea, M.; Benavides, F.G.; López, T.; Fernandez, E.; Marco, E.; Alguacil, J.; Grimalt, J.O.; Puigdomènech, E.; PANKRAS II Study Group Differences in Serum Concentrations of Organochlorine Compounds by Occupational Social Class in Pancreatic Cancer. *Environ Res* 2008, 108, (3), 370–379. DOI: 10.1016/j.envres.2008.06.010.
- 69. Sévère, S.; Marchand, P.; Guiffard, I.; Morio, F.; Venisseau, A.; Veyrand, B.; Le Bizec, B.; Antignac, J.-P.; Abadie, J. Pollutants in Pet Dogs: A Model for Environmental Links to Breast Cancer. *Springerplus* 2015, 4, 27. DOI: 10.1186/s40064-015-0790-4.
- Lord, R.S.; Bralley, J.A. Clinical Applications of Urinary Organic Acids. Part I: Detoxification Markers. *Altern Med Rev* 2008, 13, (3), 205–215.
- Kałużna-Czaplińska, J. Noninvasive Urinary Organic Acids Test to Assess Biochemical and Nutritional Individuality in Autistic Children. Clin Biochem 2011, 44, (8–9), 686–691. DOI: 10.1016/j.clinbiochem.2011.01.015.
- 72. Pieczenik, S.R.; Neustadt, J. Mitochondrial Dysfunction and Molecular Pathways of Disease. *Exp Mol Pathol* 2007, *83*, (1), 84–92. DOI: 10.1016/j.yexmp.2006.09.008.
- Zamek-Gliszczynski, M.J.; Hoffmaster, K.A.; Nezasa, K.; Tallman, M.N.; Brouwer, K.L.R. Integration of Hepatic Drug Transporters and Phase II Metabolizing Enzymes: Mechanisms of Hepatic Excretion of Sulfate, Glucuronide, and Glutathione Metabolites. *Eur J Pharm Sci* 2006, 27, (5), 447–486. DOI: 10.1016/j.ejps.2005.12.007.
- 74. Jancova, P.; Anzenbacher, P.; Anzenbacherova, E. Phase II Drug Metabolizing Enzymes. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Repub* 2010, 154, (2), 103–116. DOI: 10.5507/bp.2010.017.

- 75. Grant, D.M. Detoxification Pathways in the Liver. *J Inherit Metab Dis* 1991, 14, (4), 421–430. DOI: 10.1007/BF01797915.
- Geenen, S.; Preez, F.B. du; Snoep, J.L.; Foster, A.J.; Sarda, S.; Kenna, J.G.; Wilson, I.D.; Westerhoff, H.V. Glutathione Metabolism Modeling: A Mechanism for Liver Drug-Robustness and a New Biomarker Strategy. *Biochim Biophys Acta* 2013, 1830, (10), 4943–4959. DOI: 10.1016/j.bbagen.2013.04.014.
- 77. Bock, K.W.; Lilienblum, W.; Fischer, G.; Schirmer, G.; Bock-Henning, B.S. The Role of Conjugation Reactions in Detoxication. *Arch Toxicol* 1987, 60, (1–3), 22–29. DOI: 10.1007/BF00296941.
- 78. Forman, H.J.; Zhang, H.; Rinna, A. Glutathione: Overview of Its Protective Roles, Measurement, and Biosynthesis. *Mol Aspects Med* 2009, 30, (1–2), 1–12. DOI: 10.1016/j.mam.2008.08.006.
- 79. Silva, M.; Carvalho, M. da G. Detoxification Enzymes: Cellular Metabolism and Susceptibility to Various Diseases. *Rev Assoc Med Bras* (1992) 2018, 64, (4), 307–310. DOI: 10.1590/1806-9282.64.04.307.
- Simeonova, D.; Ivanovska, M.; Murdjeva, M.; Carvalho, A.F.; Maes, M. Recognizing the Leaky Gut as a Trans-Diagnostic Target for Neuroimmune Disorders Using Clinical Chemistry and Molecular Immunology Assays. Curr Top Med Chem 2018, 18, (19), 1641–1655. DOI: 10.2174/1568026618666181115100610.
- 81. Fukui, H. Endotoxin and Other Microbial Translocation Markers in the Blood: A Clue to Understand Leaky Gut Syndrome. *Cellular & Molecular Medicine: Open access* 2016, 3, (2). DOI: 10.21767/2573-5365.100023; Available online: https://cellular-molecular-medicine.imedpub.com/abstract/endotoxin-and-other-microbial-translocation-markers-in-the-blood-a-clue-to-understand-leaky-gut-syndrome-17990.html.
- 82. Fasano, A. Zonulin and Its Regulation of Intestinal Barrier Function: The Biological Door to Inflammation, Autoimmunity, and Cancer. *Physiol Rev* 2011, *91*, (1), 151–175. DOI: 10.1152/physrev.00003.2008.
- 83. Shemesh, J.; Henschke, C.I.; Shaham, D.; Yip, R.; Farooqi, A.O.; Cham, M.D.; McCauley, D.I.; Chen, M.; Smith, J.P.; Libby, D.M.; Pasmantier, M.W.; Yankelevitz, D.F. Ordinal Scoring of Coronary Artery Calcifications on Low-Dose CT Scans of the Chest Is Predictive of Death from Cardiovascular Disease. *Radiology* 2010, 257, (2), 541–548. DOI: 10.1148/radiol.10100383.
- 84. Greenland, P.; Blaha, M.J.; Budoff, M.J.; Erbel, R.; Watson, K.E. Coronary Calcium Score and Cardiovascular Risk. *J Am Coll Cardiol* 2018, 72, (4), 434–447. DOI: 10.1016/j.jacc.2018.05.027.
- 85. Onnis, C.; Virmani, R.; Kawai, K.; Nardi, V.; Lerman, A.; Cademartiri, F.; Scicolone, R.; Boi, A.; Congiu, T.; Faa, G.; Libby, P.; Saba, L. Coronary Artery Calcification: Current Concepts and Clinical Implications. *Circulation* 2024, 149, (3), 251–266. DOI: 10.1161/CIRCULATIONAHA.123.065657.
- 86. Vander Heiden, M.G.; Cantley, L.C.; Thompson, C.B. Understanding the Warburg Effect: The Metabolic Requirements of Cell Proliferation. *Science* 2009, 324, (5930), 1029–1033. DOI: 10.1126/science.1160809.
- 87. Whyatt, R.M.; Santella, R.M.; Jedrychowski, W.; Garte, S.J.; Bell, D.A.; Ottman, R.; Gladek-Yarborough, A.; Cosma, G.; Young, T.L.; Cooper, T.B.; Randall, M.C.; Manchester, D.K.; Perera, F.P. Relationship between Ambient Air Pollution and DNA Damage in Polish Mothers and Newborns. *Environ Health Perspect* 1998, 106 Suppl 3, (Suppl 3), 821–826. DOI: 10.1289/ehp.98106821.
- 88. Knudsen, L.E.; Norppa, H.; Gamborg, M.O.; Nielsen, P.S.; Okkels, H.; Soll-Johanning, H.; Raffn, E.; Järventaus, H.; Autrup, H. Chromosomal Aberrations in Humans Induced by Urban Air Pollution: Influence of DNA Repair and Polymorphisms of Glutathione S-Transferase M1 and N-Acetyltransferase 2. *Cancer Epidemiol Biomarkers Prev* 1999, 8, (4 Pt 1), 303–310.
- 89. Wan, M.L.Y.; Co, V.A.; El-Nezami, H. Endocrine Disrupting Chemicals and Breast Cancer: A Systematic Review of Epidemiological Studies. *Crit Rev Food Sci Nutr* 2022, 62, (24), 6549–6576. DOI: 10.1080/10408398.2021.1903382.
- 90. Gore, A.C.; Chappell, V.A.; Fenton, S.E.; Flaws, J.A.; Nadal, A.; Prins, G.S.; Toppari, J.; Zoeller, R.T. Executive Summary to EDC-2: The Endocrine Society's Second Scientific Statement on Endocrine-Disrupting Chemicals. *Endocr Rev* 2015, *36*, (6), 593–602. DOI: 10.1210/er.2015-1093.
- 91. Schug, T.T.; Janesick, A.; Blumberg, B.; Heindel, J.J. Endocrine Disrupting Chemicals and Disease Susceptibility. *J Steroid Biochem Mol Biol* 2011, 127, (3–5), 204–215. DOI: 10.1016/j.jsbmb.2011.08.007.
- 92. Gao, H.; Yang, B.-J.; Li, N.; Feng, L.-M.; Shi, X.-Y.; Zhao, W.-H.; Liu, S.-J. Bisphenol A and Hormone-Associated Cancers: Current Progress and Perspectives. *Medicine (Baltimore)* 2015, 94, (1), e211. DOI: 10.1097/MD.0000000000000011.

- 93. Di Donato, M.; Cernera, G.; Giovannelli, P.; Galasso, G.; Bilancio, A.; Migliaccio, A.; Castoria, G. Recent Advances on Bisphenol-A and Endocrine Disruptor Effects on Human Prostate Cancer. *Mol Cell Endocrinol* 2017, 457, 35–42. DOI: 10.1016/j.mce.2017.02.045.
- 94. Giulivo, M.; Lopez de Alda, M.; Capri, E.; Barceló, D. Human Exposure to Endocrine Disrupting Compounds: Their Role in Reproductive Systems, Metabolic Syndrome and Breast Cancer. A Review. *Environ Res* 2016, 151, 251–264. DOI: 10.1016/j.envres.2016.07.011.
- 95. Li, R.; Zhou, R.; Zhang, J. Function of PM2.5 in the Pathogenesis of Lung Cancer and Chronic Airway Inflammatory Diseases. *Oncol Lett* 2018, 15, (5), 7506–7514. DOI: 10.3892/ol.2018.8355.
- 96. Karin, M. NF-kappaB as a Critical Link between Inflammation and Cancer. *Cold Spring Harb Perspect Biol* 2009, *1*, (5), a000141. DOI: 10.1101/cshperspect.a000141.
- 97. Philip, M.; Rowley, D.A.; Schreiber, H. Inflammation as a Tumor Promoter in Cancer Induction. *Semin Cancer Biol* 2004, 14, (6), 433–439. DOI: 10.1016/j.semcancer.2004.06.006.
- 98. Mayland, C.; Allen, K.R.; Degg, T.J.; Bennet, M. Micronutrient Concentrations in Patients with Malignant Disease: Effect of the Inflammatory Response. *Ann Clin Biochem* 2004, 41, (Pt 2), 138–141. DOI: 10.1258/000456304322880032.
- 99. Luo, M.; Fernandez-Estivariz, C.; Jones, D.P.; Accardi, C.R.; Alteheld, B.; Bazargan, N.; Hao, L.; Griffith, D.P.; Blumberg, J.B.; Galloway, J.R.; Ziegler, T.R. Depletion of Plasma Antioxidants in Surgical Intensive Care Unit Patients Requiring Parenteral Feeding: Effects of Parenteral Nutrition with or without Alanyl-Glutamine Dipeptide Supplementation. *Nutrition* 2008, 24, (1), 37–44. DOI: 10.1016/j.nut.2007.10.004.
- 100. Fritz, H.; Kennedy, D.; Fergusson, D.; Fernandes, R.; Cooley, K.; Seely, A.; Sagar, S.; Wong, R.; Seely, D. Selenium and Lung Cancer: A Systematic Review and Meta Analysis. *PLoS One* 2011, 6, (11), e26259. DOI: 10.1371/journal.pone.0026259.
- 101. Steele, E.; Liu, D.; Omer, E. Managing Micronutrient Deficiencies in High-Risk Patients: No Small Feat! *Curr Nutr Rep* 2024, 13, (3), 668–678. DOI: 10.1007/s13668-024-00552-w.
- 102. Grant, W.B.; Wimalawansa, S.J.; Pludowski, P.; Cheng, R.Z. Vitamin D: Evidence-Based Health Benefits and Recommendations for Population Guidelines. *Nutrients* 2025, 17, (2), 277. DOI: 10.3390/nu17020277; Available online: https://www.mdpi.com/2072-6643/17/2/277.
- 103. Savchenko, A.A.; A, C.A.; Kasparov, E.V.; B, K.Э.; Borisov, S.A.; A, Б.С.; Masterova, A.A.; A, M.A.; Borisov, A.G.; Геннадьевич, Б.А. Evaluation of the effectiveness of detoxification in the immunorehabilitation of cancer patients. *Cytokines and inflammation* 2022, *19*, (1–4), 61–68. DOI: 10.17816/CI2022221-4-11; Available online: https://doi.org/10.17816/CI2022221-4-11.
- 104. Cassileth, B. Gerson Regimen. Oncology (Williston Park) 2010, 24, (2), 201.
- 105. Lee, C.; Raffaghello, L.; Longo, V.D. Starvation, Detoxification, and Multidrug Resistance in Cancer Therapy. *Drug Resist Updat* 2012, 15, (1–2), 114–122. DOI: 10.1016/j.drup.2012.01.004.
- 106. NCI, N. Intravenous Vitamin C (PDQ®) NCI. 2025 nciglobal,ncienterprise; Available online: https://www.cancer.gov/about-cancer/treatment/cam/hp/vitamin-c-pdq.
- 107. Takacs, I.; Bakos, B.; Nemeth, Z.; Toth, B.E.; Szili, B.; Lakatos, P. Controlled Randomized Open Label Clinical Study Comparing the Safety and Efficacy of Loading Schedules in Vitamin D Deficient Patients. *J Steroid Biochem Mol Biol* 2023, 231, 106330. DOI: 10.1016/j.jsbmb.2023.106330.
- 108. Amon, U.; Yaguboglu, R.; Ennis, M.; Holick, M.F.; Amon, J. Safety Data in Patients with Autoimmune Diseases during Treatment with High Doses of Vitamin D3 According to the "Coimbra Protocol." *Nutrients* 2022, *14*, (8), 1575. DOI: 10.3390/nu14081575.
- 109. McCullough, P.J.; Lehrer, D.S.; Amend, J. Daily Oral Dosing of Vitamin D3 Using 5000 TO 50,000 International Units a Day in Long-Term Hospitalized Patients: Insights from a Seven Year Experience. J Steroid Biochem Mol Biol 2019, 189, 228–239. DOI: 10.1016/j.jsbmb.2018.12.010.
- 110. Grant, W.B.; Boucher, B.J.; Cheng, R.Z.; Pludowski, P.; Wimalawansa, S.J. Vitamin D and Cardiovascular Health: A Narrative Review of Risk Reduction Evidence. *Nutrients* 2025, 17, (13), 2102. DOI: 10.3390/nu17132102.
- 111. Lemke, D.; Klement, R.J.; Schweiger, F.; Schweiger, B.; Spitz, J. Vitamin D Resistance as a Possible Cause of Autoimmune Diseases: A Hypothesis Confirmed by a Therapeutic High-Dose Vitamin D Protocol. Front Immunol 2021, 12, 655739. DOI: 10.3389/fimmu.2021.655739.

- 112. Hariri, E.; Kassis, N.; Iskandar, J.-P.; Schurgers, L.J.; Saad, A.; Abdelfattah, O.; Bansal, A.; Isogai, T.; Harb, S.C.; Kapadia, S. Vitamin K2-a Neglected Player in Cardiovascular Health: A Narrative Review. *Open Heart* 2021, 8, (2), e001715. DOI: 10.1136/openhrt-2021-001715.
- 113. DiNicolantonio, J.J.; Bhutani, J.; O'Keefe, J.H. The Health Benefits of Vitamin K. *Open Heart* 2015, 2, (1), e000300. DOI: 10.1136/openhrt-2015-000300.
- 114. Vik, H. Highlighting The Substantial Body Of Evidence Confirming The Importance Of Vitamin K2 As A Cardio-Support Nutrient, And How The Right K2 Makes All The Difference. *Integr Med (Encinitas)* 2019, 18, (6), 24–28.
- 115. Shiraki, M.; Shiraki, Y.; Aoki, C.; Miura, M. Vitamin K2 (Menatetrenone) Effectively Prevents Fractures and Sustains Lumbar Bone Mineral Density in Osteoporosis. *J Bone Miner Res* 2000, 15, (3), 515–521. DOI: 10.1359/jbmr.2000.15.3.515.
- 116. Knapen, M.H.J.; Schurgers, L.J.; Vermeer, C. Vitamin K2 Supplementation Improves Hip Bone Geometry and Bone Strength Indices in Postmenopausal Women. *Osteoporos Int* 2007, 18, (7), 963–972. DOI: 10.1007/s00198-007-0337-9.
- 117. Bunyaratavej, N.; Penkitti, P.; Kittimanon, N.; Boonsangsom, P.; Bonjongsat, A.; Yunoi, S. Efficacy and Safety of Menatetrenone-4 Postmenopausal Thai Women. *J Med Assoc Thai* 2001, 84 Suppl 2, S553-559.
- 118. Villa, J.K.D.; Diaz, M.A.N.; Pizziolo, V.R.; Martino, H.S.D. Effect of Vitamin K in Bone Metabolism and Vascular Calcification: A Review of Mechanisms of Action and Evidences. *Crit Rev Food Sci Nutr* 2017, 57, (18), 3959–3970. DOI: 10.1080/10408398.2016.1211616.
- 119. Iwamoto, J. Vitamin K2 Therapy for Postmenopausal Osteoporosis. *Nutrients* 2014, *6*, (5), 1971–1980. DOI: 10.3390/nu6051971; Available online: https://pmc.ncbi.nlm.nih.gov/articles/PMC4042573/.
- 120. Baghli, I.; Makis, W.; Marik, P.E.; Gonzalez, M.J.; Grant, W.B.; Hunninghake, R.; Levy, T.E.; Lim, H.; Cheng, R.Z.; Bondarenko, I.; Bousquet, P.; Ortiz, R.; Mary, M.; D'Agostino, D.P.; Martinez, P. Targeting the Mitochondrial-Stem Cell Connection in Cancer Treatment: A Hybrid Orthomolecular Protocol. *Journal of Orthomolecular Medicine* 2024, 39, (3).; Available online: https://isom.ca/wp-content/uploads/2024/09/Targeting-the-Mitochondrial-Stem-Cell-Connection-in-Cancer-Treatment-JOM-39.3.pdf.
- 121. Schoenfeld, J.D.; Sibenaller, Z.A.; Mapuskar, K.A.; Wagner, B.A.; Cramer-Morales, K.L.; Furqan, M.; Sandhu, S.; Carlisle, T.L.; Smith, M.C.; Abu Hejleh, T.; Berg, D.J.; Zhang, J.; Keech, J.; Parekh, K.R.; Bhatia, S.; Monga, V.; Bodeker, K.L.; Ahmann, L.; Vollstedt, S.; Brown, H.; Shanahan Kauffman, E.P.; Schall, M.E.; Hohl, R.J.; Clamon, G.H.; Greenlee, J.D.; Howard, M.A.; Schultz, M.K.; Smith, B.J.; Riley, D.P.; Domann, F.E.; Cullen, J.J.; Buettner, G.R.; Buatti, J.M.; Spitz, D.R.; Allen, B.G. O2-- and H2O2-Mediated Disruption of Fe Metabolism Causes the Differential Susceptibility of NSCLC and GBM Cancer Cells to Pharmacological Ascorbate. *Cancer Cell* 2017, 31, (4), 487-500.e8. DOI: 10.1016/j.ccell.2017.02.018.
- 122. Elliott, R.L.; Jiang, X.P.; Head, J.F. Mitochondria Organelle Transplantation: Introduction of Normal Epithelial Mitochondria into Human Cancer Cells Inhibits Proliferation and Increases Drug Sensitivity. *Breast Cancer Res Treat* 2012, 136, (2), 347–354. DOI: 10.1007/s10549-012-2283-2.
- 123. Tan, A.S.; Baty, J.W.; Dong, L.-F.; Bezawork-Geleta, A.; Endaya, B.; Goodwin, J.; Bajzikova, M.; Kovarova, J.; Peterka, M.; Yan, B.; Pesdar, E.A.; Sobol, M.; Filimonenko, A.; Stuart, S.; Vondrusova, M.; Kluckova, K.; Sachaphibulkij, K.; Rohlena, J.; Hozak, P.; Truksa, J.; Eccles, D.; Haupt, L.M.; Griffiths, L.R.; Neuzil, J.; Berridge, M.V. Mitochondrial Genome Acquisition Restores Respiratory Function and Tumorigenic Potential of Cancer Cells without Mitochondrial DNA. *Cell Metab* 2015, 21, (1), 81–94. DOI: 10.1016/j.cmet.2014.12.003.
- 124. Scatena, R. Mitochondria and Cancer: A Growing Role in Apoptosis, Cancer Cell Metabolism and Dedifferentiation. *Adv Exp Med Biol* 2012, 942, 287–308. DOI: 10.1007/978-94-007-2869-1\_13.
- 125. Zadik, Y.; Arany, P.R.; Fregnani, E.R.; Bossi, P.; Antunes, H.S.; Bensadoun, R.-J.; Gueiros, L.A.; Majorana, A.; Nair, R.G.; Ranna, V.; Tissing, W.J.E.; Vaddi, A.; Lubart, R.; Migliorati, C.A.; Lalla, R.V.; Cheng, K.K.F.; Elad, S.; Mucositis Study Group of the Multinational Association of Supportive Care in Cancer/International Society of Oral Oncology (MASCC/ISOO) Systematic Review of Photobiomodulation for the Management of Oral Mucositis in Cancer Patients and Clinical Practice Guidelines. Support Care Cancer 2019, 27, (10), 3969–3983. DOI: 10.1007/s00520-019-04890-2.



- 126. Callaway, N.L.; Riha, P.D.; Bruchey, A.K.; Munshi, Z.; Gonzalez-Lima, F. Methylene Blue Improves Brain Oxidative Metabolism and Memory Retention in Rats. *Pharmacol Biochem Behav* 2004, 77, (1), 175–181. DOI: 10.1016/j.pbb.2003.10.007.
- 127. Riha, P.D.; Bruchey, A.K.; Echevarria, D.J.; Gonzalez-Lima, F. Memory Facilitation by Methylene Blue: Dose-Dependent Effect on Behavior and Brain Oxygen Consumption. *Eur J Pharmacol* 2005, 511, (2–3), 151–158. DOI: 10.1016/j.ejphar.2005.02.001.
- 128. Rojas, J.C.; Bruchey, A.K.; Gonzalez-Lima, F. Neurometabolic Mechanisms for Memory Enhancement and Neuroprotection of Methylene Blue. *Prog Neurobiol* 2012, 96, (1), 32–45. DOI: 10.1016/j.pneurobio.2011.10.007.
- 129. Tucker, D.; Lu, Y.; Zhang, Q. From Mitochondrial Function to Neuroprotection-an Emerging Role for Methylene Blue. *Mol Neurobiol* 2018, 55, (6), 5137–5153. DOI: 10.1007/s12035-017-0712-2.
- 130. Gonzalez-Lima, F.; Barksdale, B.R.; Rojas, J.C. Mitochondrial Respiration as a Target for Neuroprotection and Cognitive Enhancement. *Biochem Pharmacol* 2014, 88, (4), 584–593. DOI: 10.1016/j.bcp.2013.11.010.
- 131. Gonzalez-Lima, F.; Auchter, A. Protection against Neurodegeneration with Low-Dose Methylene Blue and near-Infrared Light. *Front Cell Neurosci* 2015, *9*, 179. DOI: 10.3389/fncel.2015.00179.
- 132. Donegan, S. Novel Treatment for the Management of Ifosfamide Neurotoxicity: Rationale for the Use of Methylene Blue. *J Oncol Pharm Pract* 2001, 6, (4), 153–165. DOI: 10.1177/107815520100600404; Available online: https://doi.org/10.1177/107815520100600404.
- 133. Jiang, H.; Tiche, S.J.; He, C.J.; Liu, J.; Bian, F.; Jedoui, M.; Forgo, B.; Islam, M.T.; Zhao, M.; Emengo, P.; He, B.; Li, Y.; Li, A.M.; Truong, A.T.; Ho, J.; Simmermaker, C.; Yang, Y.; Zhou, M.-N.; Hu, Z.; Svensson, K.J.; Cuthbertson, D.J.; Hazard, F.K.; Xing, L.; Shimada, H.; Chiu, B.; Ye, J. Restoring Mitochondrial Quantity and Quality to Reverse the Warburg Effect and Drive Neuroblastoma Differentiation. *Proc Natl Acad Sci U S A* 2025, *122*, (36), e2502483122. DOI: 10.1073/pnas.2502483122.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.