

Review

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Posted Date: 17 September 2024

doi: 10.20944/preprints202409.1316.v1

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Review

The Utilization of PRAME in the Diagnosis, Prognosis, and Treatment of Melanoma

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Abstract: Melanoma, a deadly form of skin cancer, has seen improved survival rates due to advances in diagnosis and treatment, yet the need for further improvement remains critical. Tumor-associated antigens, such as PRAME (Preferentially Expressed Antigen in Melanoma), offer promising avenues for enhanced diagnostic precision, prognostic assessment, and targeted immunotherapy. PRAME, a cancer-testis antigen, is selectively expressed in various cancers, including melanoma, and plays a key role in promoting tumorigenesis through inhibition of retinoic acid signaling, epithelial-to-mesenchymal transition, and immune evasion. This review explores the diagnostic utility of PRAME in distinguishing melanoma from benign nevi, its prognostic value in aggressive melanoma subtypes, and its potential as a therapeutic target in cancer vaccines and adoptive T-cell therapies. While PRAME-targeted therapies face challenges such as tumor heterogeneity and immune suppression, ongoing research aims to overcome these barriers, offering hope for more effective melanoma treatments.

Keywords: melanoma; PRAME; cell biology; cancer; immunotherapy

1. Introduction

Melanoma, known to develop in part from UV radiation damage of melanocyte DNA or other inherited or acquired mutations, is one of the deadliest forms of skin cancer. The American Cancer Society estimates that in the US, approximately 100,640 new cases of melanoma will be diagnosed in 2024. Of these, 8,290 are expected to die from their cancer (ACS, 2024). Additionally, melanoma incidence rates are consistently rising. The statistics, however, are not all discouraging. Death rates secondary to melanoma have declined by 6% to 7% per year from 2013 to 2017 (ACS, 2024). The reductions in death rates can be explained by the improvements in diagnosis and treatment. Over the past three decades, several paradigm-shifting therapeutics have been developed for the treatment of cancer, including immune checkpoint inhibitors (ICI), BRAF and MEK targeted therapies, and cancer vaccines (Seedor and Orloff, 2022). ICIs, for example, were demonstrated to extend median overall survival from 11.2 months in 2013 to 72.1 months in the CheckMate 067 trial in 2021 (Van Not et al., 2024, Wolchok et al., 2022). Improvements in melanoma screening and rapidity of diagnosis have also contributed to the reduction in death rates. Despite these improvements, the number of deaths remains unacceptably high. Further advancements in melanoma diagnosis and treatment are critical.

A particular area of research that poses promising solutions for diagnosis and treatment of melanoma is the identification of antigen biomarkers. Specifically, investigators are searching for tumor-associated antigens (TAA), proteins expressed by tumor cells, exclusively or at least in higher quantities, as compared to somatic cells. Identification of antigen biomarkers could be utilized in the diagnostic process through the development of immunohistochemistry. These antibodies recognize specific proteins and would provide high utility for pathologists with challenging melanoma cases. Moreover, TAAs would make excellent targets for immune-based therapies. Utilizing T-cells or cancer vaccines to target specific TAAs could provide maximal destruction of tumor cells while minimizing damage to normal tissues. A current TAA being studied in these capacities is PRAME,

or preferentially expressed antigen in melanoma. This paper seeks to explore the utilization of PRAME in different clinical contexts for the management of melanoma, namely diagnostic, prognostic, and therapeutic contexts.

2. The Biology of PRAME and Other Cancer Testis Antigens

2.1. Overview of the PRAME Protein

Preferentially expressed Antigen in Melanoma (PRAME) is a 509 amino acid protein found in the nuclear and cytoplasmic space of several different cell types in the human body. The gene encoding PRAME is found on chromosome 22 (22q11.22), embedded among several genes encoding immunoglobulin proteins (Cassalia et al., 2024). PRAME is a member of the cancer testis antigen (CTA) family. CTAs are proteins typically found in germline cells of the ovaries and testis and are thought to play a role in gametogenesis (Cassalia et al., 2024). These antigens have also been found on various hematologic and solid tumor malignancies. Many CTAs are present on human leukocyte antigen class 1 (HLA-1) molecules in cancer cells, allowing for recognition by T-lymphocytes (Al-Khadairi and Decock, 2019, Cassalia et al., 2024, Fratta et al., 2011). PRAME was first identified by H. Ideka et al. in melanoma cell lines in 1997 while investigating potential therapeutic targets for immunotherapy (Ikeda et al., 1997).

2.2. Biology of Cancer Testis Antigens

Cutting-edge technologies like cancer vaccines and T-cell therapy are exciting due to their utilization of the immune system to find and destroy tumor cells with high selectivity. To continue developing better immunotherapies, investigators must identify immunogenic antigens that are selectively expressed on the HLA-1 molecules of cancer cells and not normal cells. These specific antigens in tumor cells are often called tumor-associated antigens (TAA). In hopes of discovering TAAs, researchers led by Lloyd J Old performed autologous typing experiments with various tumor cells and found a new class of TAAs he called cancer testis antigens (CTAs) (Nin and Deng, 2023). Twenty-five years later, autologous typing techniques were superseded by serological analysis of cDNA expression libraries (SEREX), which led to a rapid expansion of the known CTAs (Fratta et al., 2011, Nin and Deng, 2023). These antigens are called cancer testis antigens because of their expression in both tumor cells and testis germline cells. Today, the CTA family includes over 200 distinct proteins (Gjerstorff et al., 2015, Nin and Deng, 2023). CTAs can be divided into two subclasses by their chromosomal location. One class represents genes found on X-chromosomes, and the other represents genes found on either autosomes or Y-chromosomes (Gjerstorff et al., 2015, Nin and Deng, 2023). In adult somatic tissues, CTA expression is typically restricted to germline cells of the testis, ovaries, and placenta (Fratta et al., 2011, Gjerstorff et al., 2015, Nin and Deng, 2023). However, recent experiments suggest that CTAs such as PRAME are also found in the adrenal and endometrial tissues (Cassalia et al., 2024, Simpson et al., 2005). CTAs are known to be involved in the differentiation and proliferation of germline cells, but the biological mechanisms remain to be fully elucidated (Nin and Deng, 2023, Simpson et al., 2005).

2.3. The Role of PRAME and other Cancer Testis Antigens in Tumorigenesis

Many plausible theories have been developed that attempt to explain the process of somatic cells transitioning to tumor cells. One such theory is that certain silenced genes involved in gametogenesis become activated, resulting in a highly proliferative, apoptotic-resistant, and migratory phenotype implicated in cancer (Nin and Deng, 2023). Given the role of PRAME in gametogenesis, its increased expression is thought to promote tumorigenesis through inhibition of retinoic acid signaling, promotion of epithelial to mesenchymal transition, and promotion of a pro-tumor microenvironment. In testis cells for example, when a gametocyte differentiates into a mature spermatocyte, epigenetic programs methylate promoter regions of DNA that encode proteins involved in gametogenesis. Once the promoter region is methylated, transcription factors can no longer bind, resulting in decreased genetic expression (Moore et al., 2013). In all somatic tissues, excluding the placenta and testis

germline cells, the promoter regions of genes encoding CTAs are hypermethylated. In the germline cells, by contrast, the promoter regions are *hypomethylated*. Interestingly, several studies exploring CTA and PRAME biology in cancer note similar hypomethylation patterns for CTA genes. Somatic tissue with methylated promoter regions can become *hypomethylated*, which increases the expression of CTA product proteins that promote tumor phenotypes. (Cassalia et al., 2024, Schenk et al., 2007). These methylation patterns suggest that epigenetic modifications of CTA genes occur in somatic cells, promoting the transition to cancerous phenotypes (Figure 1).

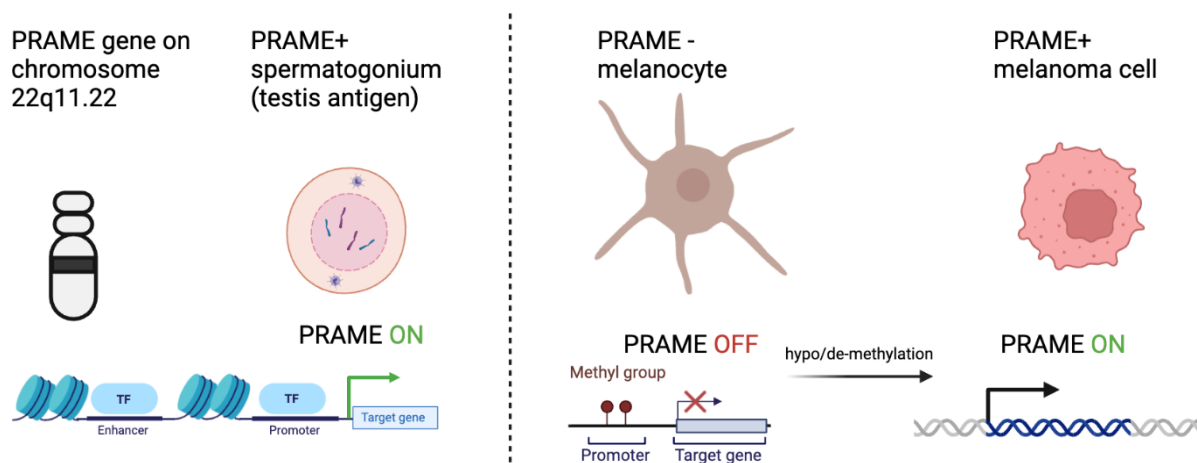


Figure 1. PRAME expression in early progenitor cells and reactivation in melanoma cells. PRAME is located on chromosome 22 and normally expressed in developing gamete cells. PRAME expression is associated with an undifferentiated, proliferative state in cells. Methylation at the promoter region inhibits its expression in benign melanocytes. Via mechanisms not fully yet know, PRAME expression is re-activated in almost all melanoma types, in part due to de-methylation at the PRAME promoter.

The protein PRAME has been identified to promote a cancerous phenotype through its involvement with retinoic acid signaling pathways (Figure 2). Retinoic acid signaling (RAS) is a biochemical pathway involved in the regulation of cell differentiation, proliferation arrest, and apoptosis (Al-Khadairi and Decock, 2019, Szymański et al., 2020). In normal cells, retinoic acid (RA), commonly known as vitamin A, binds to retinoic acid receptor proteins α , β , or γ , which then activate genetic programs involved in cell cycle regulation and growth arrest (Brown, 2023). Normal RAS has anti-tumor effects. Remarkably, activation of RAS through a form of exogenous RA called all-trans retinoic acid (ATRA) has been demonstrated to inhibit the growth and progression of acute promyelocytic leukemia (APL) (Nagai and Ambinder, 2023). Though not entirely established, it is thought that PRAME acts as a dominant repressor of RAS through competitive binding to RA receptors. PRAME prevents RA from binding properly, resulting in the downregulation of target genes involved in growth arrest (Al-Khadairi and Decock, 2019, Cassalia et al., 2024). This is a plausible explanation for the high expression of PRAME in tumor cells with highly proliferative phenotypes.

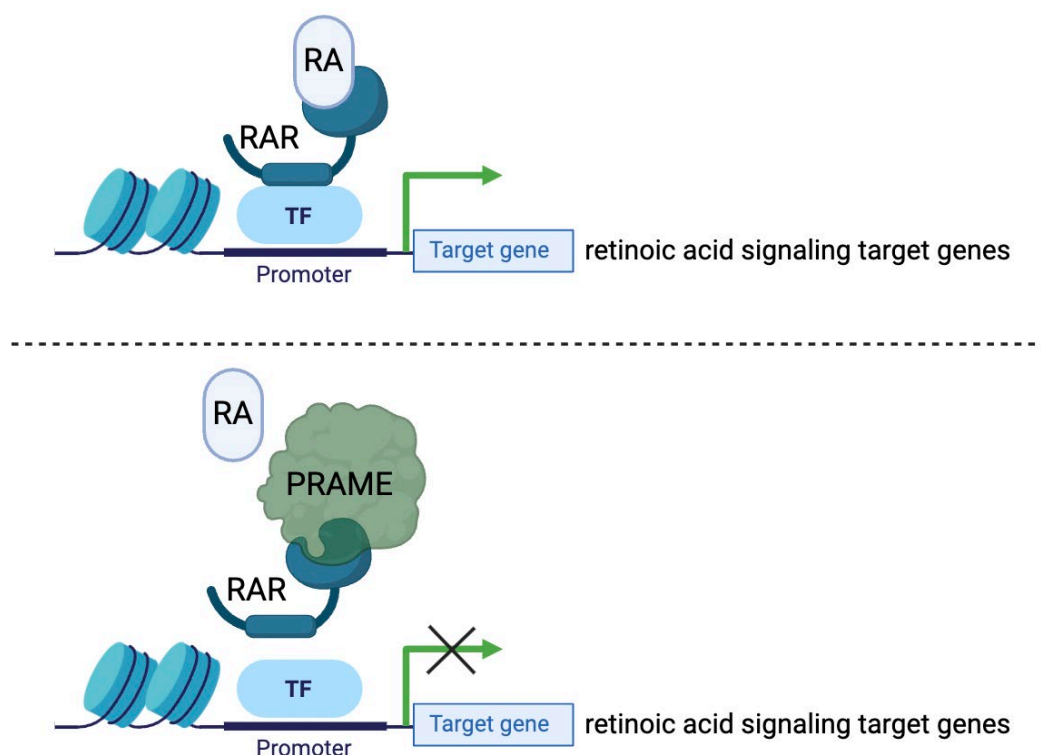


Figure 2. PRAME inhibits binding of retinoic acid (RA)-retinoic acid receptor (RAR), inhibiting target gene expression. RA binds to nuclear hormone receptor RAR to induce expression of retinoic acid signaling genes. PRAME binds to RAR to inhibit the expression of the target genes. Switching OFF of these target genes results in a more proliferative, migratory, and less-differentiated cell state (characteristics that are displayed in melanoma cells).

PRAME, among other CTAs, has been shown to increase the metastatic potential of cancer cells. Several studies have found that tumor cells of metastatic lesions exhibit higher expression levels of CTAs than the cells of the primary tumor site, suggesting they may play a role in cancer metastasis. Researchers have determined one plausible mechanism for this increased metastatic potential. The CTAs PRAME, CT45A1, and MAGEC2 are known to induce epithelial-to-mesenchymal transitions (EMT) in tumor cells. EMT is a process by which somatic cells lose their cell-to-cell adhesion properties and surface polarity, leading to increased cell motility and migratory potential (Al-Khadairi et al., 2019, Nin and Deng, 2023, Shang et al., 2014, Wen et al., 2021). How PRAME induces EMT is not understood, but the mechanism could be similar to the protein CT45A1, which induces EMT in osteosarcoma cells through the β -catenin pathway (Shang et al., 2014, Wen et al., 2021). The third understood role of PRAME in tumorigenesis is in its promotion of a pro-tumor microenvironment by increasing tumor “coldness.” Cold tumors are cancer cells that evade the immune system through down-regulation of antigen-presenting molecules and increase production of immune checkpoint molecules. These “tumor-cold” environments limit the efficacy of immunotherapeutics. PRAME was shown to be involved in this process in a breast cancer study. Investigator A. Naik et al. found that silencing of PRAME genes in MDA-MB-468 breast cancer cells reduced the expression of immune checkpoint molecules and promoted cancer cytotoxicity by T-cells. In the same study, overexpression of PRAME was found to decrease the activation of T-cells and reduce the cytolytic potential (Naik et al., 2021). Further research must be done to better understand the role of PRAME in cancer through its involvement in RAS, EMT promotion, and establishment of a tumor-cold microenvironment.

3. Diagnostic Potential of PRAME in Melanoma

3.1. Current Histopathologic Techniques Used in Melanoma Diagnosis

Melanoma is a cancer of the melanocytes. Many melanomas bear similar features, clinically and pathologically, to benign pigmented or dysplastic lesions, leading to an often-challenging differential diagnosis. Clinically, dermatologists utilize the ABCDE criteria for distinguishing melanomas from benign nevi (Moore and Gasparini, 2011). These criteria include **A**symmetry, **B**order irregularity, **C**olor change, **D**iameter of greater than 6mm, and **E**volution (Duarte et al., 2021, Moore and Gasparini, 2011). If making a confident diagnosis is difficult, clinicians biopsy the lesions for pathology to review. Followed by staining with hematoxylin and eosin (H&E), pathologists look for irregular morphologic features like asymmetry, poor circumscription, cytologic atypia, mitotic activity, and failure of maturation with descent (Moore and Gasparini, 2011). Despite careful analysis of these features, many difficult-to-diagnose cases persist. Diagnosing melanomas with high specificity and sensitivity is essential because incorrect or delayed diagnoses can result in unnecessary treatment or delays in necessary treatment. Immunostaining for a variety of protein markers is often used to increase diagnostic accuracy. Common IHC stains used in the diagnosis and staging of melanomas are S-100, SOX10, Melan-A, and HMB-45 (Pop et al., 2023, Ronchi et al., 2022). S100 is not specific to melanocytes. Sox-10 and Melan-A confirm melanocytic origins, but are not capturing a malignant phenotype. Pathologists may also utilize cytogenic tests like fluorescence in situ hybridization (FISH) or single nucleotide polymorphism array (SNP) to examine the DNA from biopsied tissue samples to identify genetic aberrations (Cazzato et al., 2022; M. W. Moore & Gasparini, 2011). This allows for the identification of genetic aberrations implicated in cancer. Cytogenic testing typically yields more accurate diagnoses; however, there is no perfect test, and final diagnosis relies on clinical, pathologic, and molecular correlation. However, cost and availability of molecular testing often result in less frequent or restricted use of these tests. Because of this, pathologists favor the widely available immunostaining methods for melanoma diagnosis. Continued research is being conducted to identify candidate antigens for IHC. PRAME has become one a strong antigen of interest.

3.2. Utilization of PRAME in Melanoma Diagnosis

Melanocytes are found in a myriad of locations throughout the body, from the lymph nodes to the epidermis. Thus, melanomas are diverse in their location, presentation, and prognosis. The list of melanoma subtypes includes superficial spreading melanoma, lentigo maligna, acral melanoma, desmoplastic melanoma, non-desmoplastic melanoma, nevoid melanoma, uveal melanoma, spitzoid melanoma, and mucosal melanoma (Lezcano et al., 2021, Scolyer et al., 2011). Each of the different melanoma subtypes has a benign lesion counterpart with similar clinical characteristics. Many studies have been performed to explore the frequency of PRAME expression in melanoma compared with benign tumors. The research has shown PRAME to be a highly selective and sensitive marker in melanoma diagnosis. Research by Lezcano et al. found that of 100 lesions confirmed to be metastatic melanoma, 92 were diffusely PRAME positive (92%). Similarly, a sample of 155 primary cutaneous melanomas, excluding desmoplastic melanomas, showed PRAME positivity in 92% of the cases (Lezcano et al., 2018). This data highlights the sensitivity of PRAME staining. In the same study, specificity was measured by PRAME staining of 140 benign cutaneous and nodal melanocytic nevi. Of these nevi, 86.4% were PRAME negative (Lezcano et al., 2018). A similar study performed by Andrea Ronchi et al. utilized immunocytochemistry (ICC) methods to explore PRAME positivity in melanomas. PRAME positivity was observed in 85.4% of 48 cutaneous melanoma metastases. The sensitivities of PRAME were not as significant as S100 (100%), Melan-A (97.9%), SOX10 (100%) and HMB45 (89.6%) (Ronchi et al., 2022). Though PRAME IHC is less sensitive than the other IHC stains marking melanocytes, it may offer greater diagnostic utility by being much more specific for melanoma. PRAME is selectively expressed in various malignancies and limitedly expressed in benign cells. When staining with Melan-A and SOX10, benign and malignant melanocytes are stained, making margin assessment difficult. Staining with PRAME allows for clearer margin assessment and better visualization of malignant melanocytes. For these reasons, PRAME IHC offers unique clinical utility by providing more accurate margin assessment and Breslow thickness; both of

which are important in staging and clinical management of melanoma (Lezcano et al., 2018). Additional studies have evaluated the sensitivity of PRAME IHC compared with other ancillary cytogenetic testing like FISH and SNP-Array. A separate study done by Cecilia Lezcano revealed that out of a cohort of 110 diagnostically challenging melanocytic tumors, there was a concordance of 90% between cytogenetic testing and PRAME IHC as well as a concordance of 92.7% between PRAME IHC and the final diagnosis. High concordance with cytogenetic testing is a testament to the diagnostic utility of PRAME.

As previously mentioned, melanocytes can be present in the lymph nodes. The benign collections of melanocytes in the lymph nodes are called nodal nevi. Nodal nevi are morphologically similar to melanoma lymph node metastases, making the differential diagnosis challenging. Lezcano et al. performed a study evaluating the expression of PRAME in nodal nevi compared to melanoma metastases. They found that of 30 nodal nevi, 0% were PRAME-positive. This data demonstrates remarkable specificity. Furthermore, of the 15 melanoma metastases in the lymph nodes, 100% were PRAME-positive (Lezcano et al., 2020b). Further research should be done to confirm this high sensitivity of PRAME IHC to improve the diagnostic accuracy of lymph node biopsies in melanoma patients. Another challenging class of cases in clinical practice is acral lesions, which are pigmented lesions on the soles, palms, knees, elbows, and nails. Benign acral lesions have similar pathologic and morphologic characteristics under microscopy compared with melanomas, necessitating IHC and ancillary cytogenetic testing to confirm diagnoses. PRAME IHC was found to be positive in 87.1% of malignant acral lesions and faintly expressed or not at all expressed in 82.5% of benign lesions in a study done by Giacomo Santandrea et al. This research suggests a sensitivity of 87.1% and a specificity of 82.5% for PRAME IHC (Santandrea et al., 2022). One of the authors of this manuscript (McBride) found that, in a cohort at the Cleveland Clinic, 100% of acral melanomas (n=10) were positive for PRAME IHC, and, of the benign, dysplastic, and spitz acral nevi 100% (n=20), all were negative for PRAME (McBride et al., 2022). Having an accurate IHC stain like PRAME to aid in the diagnosis of acral lesions is extremely useful and should be utilized more often in clinical practice. PRAME IHC is an exciting tool for differentiating between benign and malignant melanocytic lesions. However, inconsistencies in expression among certain melanoma subtypes can be significant barriers to universal interpretation; PRAME staining must always be taken within the entire context of the neoplasm, along with clinical and, if available, molecular data. Two diagnostically challenging melanocytic proliferations are desmoplastic and spitzoid melanomas. Desmoplastic melanoma, an invasive, neurotropic malignancy, was found to have PRAME positivity in only 35% of cases (Table 1) (Lezcano et al., 2018). However, the gold standard is only the labeled diagnosis of “desmoplastic melanoma” and PRAME expression in the PRAME+ desmoplastic melanomas may be signaling something fundamentally different about these desmoplastic melanoma types. Spitzoid lesions can be difficult to diagnose on H&E alone, and often need additional immunohistochemical and molecular tests as well, especially due to the clinical and pathological similarities between spitz nevi, atypical spitz tumors and spitzoid melanomas (Cassalia et al., 2024). Several studies have identified PRAME IHC to be a useful tool in aiding in the diagnosis of spitzoid melanomas (Googe et al., 2021). Stephen Koh’s lab found PRAME positivity in 82% of spitzoid melanomas and only 20% of benign spitz nevi. Conversely, A. Alomari identified PRAME positivity as a possible diagnostic pitfall in spitzoid melanoma diagnosis due to the cases of diffusely positive benign spitzoid lesions, which are likely just in a proliferative state (Alomari et al., 2021). PRAME correlates highly with the results of molecular testing in melanoma, with high agreement (Lezcano et al., 2020a). The strengths and limitations in sensitivity and specificity of PRAME IHC should continue to be researched. If properly understood, PRAME IHC could yield greater diagnostic utility for future challenging diagnoses. Additionally, the differences in gene expression and prognosis in PRAME+ versus PRAME-melanomas should be studied further. The utility of PRAME extends beyond diagnosis, holding prognostic implications as well as therapeutic potential.

Table 1. PRAME as a Prognostic Marker in Melanoma and Other Cancers.

Cancer Type	Study	PRAME Expression Rate	Prognostic Outcome
Uveal Melanoma (Class 2)	Field et al., 2016	High in Class 2	Increased metastasis risk, worse prognosis
Mucosal Melanoma	Toyama et al., 2019	83.3%	Correlated with worse overall survival
Breast Cancer	Epping et al., 2008	N/A	Associated with decreased overall survival
Sarcomas	Iura et al., 2015	90% in myxoid liposarcoma	PRAME-positive associated with worse prognosis
Leukemia (ALL)	Zhang et al., 2017	N/A	Overexpression correlated with better outcomes in ALL
Hodgkin’s Lymphoma	Xu et al., 2020	N/A	PRAME overexpression associated with drug resistance

3.3. PRAME as a Prognostic Biomarker in Melanoma

PRAME expression has been shown in numerous studies to be highly associated with an unfavorable melanoma prognosis. PRAME, as its name implies, was first identified in melanoma cells and has been demonstrated to increase metastatic potential through repression of RAS, promotion of EMT, and promotion of an immunologically “cold” tumor microenvironment (TME) (Al-Khadairi et al., 2019, Cassalia et al., 2024, Nin and Deng, 2023). These traits all help explain the poorer prognosis observed in melanomas with elevated PRAME expression (Table 1). Most of the prognostic research papers are retrospective studies in uveal melanomas (UM). UMs are malignancies of the melanocytes within the ocular tissue/uveal tissue and can be stratified into two subclasses by gene expression profiling (GEP). Class 1 UMs have low metastatic risk, and Class 2 UMs have high metastatic risk (Field et al., 2016a, Field et al., 2016b, Schefler et al., 2019). A retrospective study performed by Amy Schefler et al. (n=148 patients) involving multiple ocular oncology centers found positive PRAME expression to be associated with increased basal diameter, worse GEP values, and increased metastatic risk (Schefler et al., 2019). A similar analysis by Matthew Field demonstrated higher PRAME positivity in Class 2 UMs compared to Class 1 (Field et al., 2016a). Given that Class 2 UMs have greater metastatic potential, this data holds prognostic implications. In the same study, PRAME-positive UMs were found to metastasize more rapidly than the lesions with minimal PRAME expression (Field et al., 2016b). This corroborates the role of PRAME in promoting the EMT phenotype, whereby cells lose their surface polarity and gain motility (Al-Khadairi et al., 2019). The prognostic effects of PRAME have also been explored in mucosal melanomas. Out of a sample of 29 mucosal melanomas, 20 (83.3%) had high PRAME expression, correlating to a worse prognosis overall (Toyama et al., 2019).

Though these studies suggest a strong correlation between PRAME positivity and poorer prognosis, an analysis by Parra et al. found no significant difference between PRAME expression in primary cutaneous versus metastatic melanoma; this is not surprising, as PRAME levels would be expected to remain high whether or not we are looking at primary melanoma or metastatic cells (Parra et al., 2023). Further research must be conducted to explore the possible prognostic implications of PRAME, in early melanomas, versus late, and in combination with other antigens, in all melanoma subtypes.

3.4. PRAME as a Prognostic Biomarker in Other Cancer Types

PRAME has proven to be a versatile biomarker, correlating to poorer prognosis in tumor cells expressing it. One such cancer class is breast cancer, where several studies have found increased PRAME expression to be associated with worse outcomes. In 2022 alone, 287,850 US women were diagnosed with invasive breast cancer. Of these women, 43,250 died from their disease (Giaquinto et al., 2022). A study analyzing Kaplan-Meier survival curves found that of 295 primary breast cancer lesions, PRAME expression was associated with decreased overall survival and increased rates of metastasis (Epping et al., 2008). The exact mechanism explaining how PRAME promotes these outcomes in breast cancer patients is not fully understood, but some studies suggest it stems from PRAME’s involvement in the promotion of an EMT phenotype (Al-Khadairi et al., 2019). PRAME expression is also associated with decreased survival and poorer outcomes in sarcoma patients. A

sarcoma is a malignancy of the bone and soft tissues, encompassing a broad range of pathologies. Multiple sarcoma subtypes have exhibited CTA expression, including myxoid liposarcomas, synovial sarcomas, chondrosarcomas, and osteosarcomas (Roszik et al., 2017, Tan et al., 2012). A study performed by Roszik et al. found PRAME expression to be associated with lower expression of antigen-presenting protein beta2microglobulin (B2M), providing a biochemical explanation for how PRAME promotes an immunologically cold TME (Roszik et al., 2017). Cancers that downregulate antigen presenting molecules are less recognizable by the immune system, resulting in unhindered growth. Research by Iura et al. found similar results. Of 93 confirmed cases of myxoid liposarcomas, a sarcoma subtype with classically poorer prognoses, 90% were PRAME positive. In the less aggressive dedifferentiated and well-differentiated liposarcomas, only 43% and 9% expressed PRAME, respectively (Iura et al., 2015). Studies like these display the promising uses of PRAME in the prognostication of cancer. It should be noted that the role of PRAME in the prognosis of hematologic malignancies, by contrast, is still inconclusive. There are three main categories of hematologic malignancies—leukemias of the leukocytes, lymphomas of the lymphocytes, and myelomas of the plasma cells (Xu et al., 2020). Multiple studies have found PRAME to be associated with a poor prognosis in patients with diffuse large B-cell lymphoma, Hodgkin's lymphoma, multiple myeloma, and chronic leukemia through the promotion of drug resistance and disease progression (Xu et al., 2020). Given that increased PRAME expression is associated with poorer prognosis in other cancers, this data is not surprising. The convoluted nature of PRAME in hematologic malignancies comes from research citing the opposite effects. For example, Tajeddine et al. induced overexpression of PRAME in leukemia cell lines and found decreased tumorigenicity by way of induction of caspase-independent cell death in vitro and reduction in the levels of proteins inhibiting apoptosis (Hsp27, p21, and S100A4) in vivo (Tajeddine et al., 2005).

Another study that contradicts the association of increased PRAME expression with poorer prognosis is the work by Zhang et al., which found overexpression of PRAME to predict better overall outcomes in pediatric B-cell acute lymphoblastic leukemia (ALL) (Zhang et al., 2017). The contradiction in the literature emphasizes the importance of continued research on the use of PRAME in the prognostication of hematologic malignancies.

As previously discussed, many studies cite the positive association between PRAME expression and the prognosis of individual cancer types. Study designs like these are highly useful and necessary, but the utilization of meta-analysis techniques to evaluate the prognostic implications of PRAME on a broader scale, irrespective of cancer type, provides a unique understanding of the potential of PRAME. One such analysis by Jiaqiang Li et al. assesses the association of PRAME expression with cancer prognosis in 2,421 patients. In the study, the parameters of disease-free survival (DFS), progression-free survival (PFS), metastasis-free survival (MFS), and overall survival (OS) were evaluated using both fixed-effects and random-effects models. PRAME expression was shown to be positively associated with all these values. The data showed reductions in DFS ($P < 0.001$), PFS ($P = 0.042$), MFS ($P = 0.034$), and OS ($P < 0.001$) which were all statistically significant ($P \leq 0.05$). Additionally, increased PRAME expression was found to be associated with advanced clinical stages (III-IV) as well as positive lymph node metastases (Li et al., 2020). More meta-analyses looking at prognostic effects irrespective of cancer type should be conducted for stronger evidence in utilizing PRAME staining in clinical practice.

4. PRAME as a Therapeutic Target in Immunotherapy

4.1. Overview of Immunotherapy

Despite advances in surgical techniques, chemotherapeutics, and radiation therapies, cancer continues to kill numerous patients each year. Overall survival for cancer patients has improved, but these patients are still subject to metastasis and recurrence. The difficulties in treating cancer lie in the similarities between somatic cells and cancer cells. Cancer cells are almost structurally and genetically identical to their somatic counterparts but with upregulated genetic programming resulting in malignant phenotypes such as immune evasion, replicative immortality, tumor-

promoting inflammation, sustained proliferation, and evasion of cell death (Hanahan, 2022). Chemo and radiation therapies are excellent at destroying tumor cells, but they also damage healthy tissues in the process. The real difficulty is treating the malignant cells while avoiding harming the somatic cells. This need for well-tolerated treatments with robust clinical responses inspired research into a new, remarkably selective cancer therapeutic: immunotherapy. The first immunotherapies to become FDA-approved were anti-tumor cytokines interferon-alpha 2 (IFN- α 2) and interleukin-2 (IL-2), molecules that stimulate T-cell proliferation (Eno, 2017). In 2014, more than 20 years later, the FDA approved a new class of cancer therapeutics called immune checkpoint inhibitors (ICI). ICIs are antibodies against the immune checkpoint molecules Cytotoxic T-Lymphocyte-Associated Antigen 4 (CTLA-4) and Programed Death Ligand 1 (PD-L1), preventing cancer cells from deactivating T-cells (Al-Khadairi and Decock, 2019, Eno, 2017, Nin and Deng, 2023). The more recent therapies that have been developed are the use of genetically engineered T-cells (CAR-T) and ex vivo expansion of circulating antigen-specific T-cells, which work by programming T-lymphocytes to express a surface antigen receptor recognizing a tumor-specific antigen. Crucial to developing T-cell-based immunotherapies is the identification of tumor-selective antigens, those presented by HLA-class 1 molecules of tumor cells and not somatic cells. In recent years, multiple CTAs, including PRAME, have been evaluated as potential targets for immunotherapy as they are selectively presented on the HLA-1 surface proteins of tumor cells. Currently, PRAME is being researched as a target for immunotherapy for cancer vaccines and T-cell-based therapies.

4.2. PRAME as a Target in Cancer Vaccines

Cancer vaccines work by administering a synthesized antigen or nucleic acid sequence with adjuvant molecules to mount an immune response against tumors expressing the antigen. The mechanisms are similar to viral vaccines but cancer vaccines are used therapeutically instead of prophylactically (Paston et al., 2021). The cancer vaccine technologies currently being utilized are peptide vaccines, dendritic cell vaccines, and nucleic acid vaccines. CTA proteins are prime targets for peptide-based cancer vaccines due to their tumor selectivity. Trials exploring the efficacy of MAGE-A and NY-ESO-1 vaccines are ongoing, many eliciting both cellular and humoral responses (Nin and Deng, 2023). For instance, data from phase I and II trials for these CTAs have shown antibody, CD4+, and CD8+ responses with minimal off-target toxicities (Al-Khadairi and Decock, 2019, Gjerstorff et al., 2015). However, when the trials progressed to phase III, different conclusions emerged. Two phase III clinical trials evaluating the efficacy of MAGE-A3 peptide-based vaccines in the treatment of non-small cell lung cancer (NSCLC) (n=2,272 patients) and melanoma (n=1,345 patients) have been performed. Both studies were terminated from a lack of improvement in disease-free survival in treatment groups compared with controls (Gjerstorff et al., 2015). A recurring situation in the field of CTA cancer vaccines has been the problem of little efficacy being demonstrated in clinical trials. Similarly, PRAME vaccine trials have not progressed past phase I trials. In murine samples treated with PRAME antigen vaccines, CD4+ and CD8+ T cell responses were observed which encouraged the escalation to primate model studies. Phase I studies in primates, metastatic melanoma patients, and NSCLC patients found antibody responses but limited CD4+ and no CD8+ responses (Gutzmer et al., 2016, Pujol et al., 2016). The lack of CD8+ responses has discouraged the escalation to phase II or III clinical trials. CD8+ responses, as previously discussed, are crucial to the destruction of tumor cells. Several plausible explanations for the observed lack of efficacy exist and will be discussed in the limitations section. In brief, a melanoma patient's immune cells may have become tolerate to PRAME as an antigen to stimulate killing of tumor cells. A more important strategy would be to hone or re-induce killer cells to PRAME+ melanoma cells. To improve the CD8+ response, investigators are exploring the use of polyvalent vaccines that target multiple antigens simultaneously. Research by Weber et al. found that a polyvalent vaccine targeting PRAME and prostate-specific membrane antigen induced expansion of CD8+ T-cells recognizing PRAME and PSMA in 15 out of 24 prostate cancer patients. In addition to these findings, the researchers found that seven patients showed stable disease for six months or

longer (Weber et al., 2011). Further research must be performed to expand upon this knowledge of polyvalent vaccine technologies targeting CTAs.

4.3. PRAME as a Target in Adoptive T-Cell Therapy

T-cell therapy can be described as the utilization of cytotoxic T lymphocytes (CTLs or CD8+ cells) to target and destroy tumor cells expressing specific antigens. CTLs circulate through the body and screen for aberrant gene products expressed on HLA-1 molecules. Circulating CTLs with unique T-cell receptors (TCRs) recognize specific antigen epitopes. When the CTL identifies a cell with a concerning antigen, it releases granzymes and perforin to destroy the cell harboring the antigen (Dafni et al., 2019, Durgeau et al., 2018). T-cells can recognize foreign antigens with remarkable specificity, posing an exciting solution to the grandest challenge to cancer therapeutics, which is maximizing tumor destruction while minimizing harm to somatic tissues. Chemotherapeutics are efficient at harming cancer cells, but they also have significant side effects that cannot be ignored. The remarkable ability of the cell-mediated immune response to recognize foreign invaders while limiting harm to self has led researchers to develop therapies utilizing CTLs in cancer treatment.

The two classes of adoptive cell transfer, commonly known as T-cell therapy, being researched are ex vivo expansion of tumor-infiltrating lymphocytes (TILs) and chimeric antigen receptor (CAR) T-cell therapy (Table 2) (Al-Khadairi and Decock, 2019, Durgeau et al., 2018). Ex vivo expansion of TILs is a process where lymphocytes that have successfully infiltrated and recognized TAAs on tumor cells are removed from patients, expanded through treatment with T-cell growth factors IL-2, IL-7 or IL-21, and once reaching sufficient population size, readministered to the patient. This process was first developed by Rosenberg et al. in 1988 for the treatment of metastatic melanoma patients. They found potent clinical response and durable regression in 11 out of 20 patients (Rosenberg et al., 1988). In the study, TILs from melanoma tumors were harvested following surgical resection. Once collected, the TILs were selected to recognize specific tumor antigens. The selected TILs were then expanded and infused into patients with adjuvant IL-2 (Rosenberg et al., 1988). Since Rosenberg pioneered ACT with TILs, it has been researched in both solid organ tumors and hematologic cancers. A recent meta-analysis by Dafni et al. investigating TIL therapy for melanoma patients found an objective response rate of 41% in 410 melanoma patients previously treated with chemotherapy or radiation (Dafni et al., 2019). Because of the aggressive nature of cancer, response rates in any form are encouraging. Thus, the 41% response rate for TIL therapy is a promising result. To improve the strength and durability of responses, several studies have experimented with TILs recognizing tumors expressing CTAs. With proteasome cleavage analysis of PRAME, four high-affinity HLA-A*201- restricted epitopes were identified to be recognizable by T-cells (Kessler et al., 2001). Two different labs have searched for circulating TILs recognizing these four PRAME-restricted epitopes in melanoma and AML patients. The researchers found these PRAME-specific TILs in 36% of melanoma patients and 70% of AML patients (Greiner et al., 2006, Griffioen et al., 2006). Trials exploring the efficacy of ACT with TILs targeting PRAME epitopes are ongoing with encouraging pre-clinical results, but substantial clinical data is lacking.

Table 2. PRAME in Immunotherapy Clinical Trials.

Therapy Type	Phase	Target Cancer	Clinical Outcome
PRAME-targeted vaccine	I	Melanoma, NSCLC	CD4+ response but limited/no CD8+ response
PRAME-specific TILs	Preclinical	Melanoma, AML	36% (Melanoma) and 70% (AML) patients showed PRAME-specific TILs
PRAME-CD3 bispecific molecules	III	Cutaneous Melanoma	58% disease control rate, promising PFS (4.2 months)
PRAME-CAR T cells	Preclinical	Various cancers	Promising preclinical results but high risk of off-target toxicities

The other type of ACT is called CAR T-cell therapy. CAR T-cell therapy involves genetically modifying the receptors of T-cells to recognize specific antigen epitopes with high affinity. CAR T-cells function similarly to endogenous CTLs, releasing granzymes and perforin in cells identified as

foreign. What makes CAR T-cell therapy unique is that its chimeric antigen receptor recognizes antigens irrespective of expression on HLA-1 molecules (Sternier and Sternier, 2021). Chimeric antigen receptors can be programmed to recognize any antigen on the surface of cells, resulting in a powerful antitumor response (Sternier and Sternier, 2021). Since its development in 1989, six CAR T-cell therapies have been approved by the FDA. Four of the therapies target cluster of differentiation-19 (CD19) surface proteins and two of the therapies target B-cell maturation antigen (BCMA), both of which are found on the surface of B-cells (Chen et al., 2023, Haslauer et al., 2021). CAR T-cell therapy has had remarkable efficacy in the management of hematological malignancies but little efficacy in the treatment of solid organ tumors. Very little research has been published exploring the targeting of PRAME-expressing cells with CAR T cell therapy. Several trials have been performed exploring other CTAs like NY-ESO-1 and MAGE-A3 with promising preliminary results but, unfortunately, with several cases of severe toxicity as well. Due to the CAR T-cell's ability to recognize surface antigens irrespective of HLA-1, the risk of off-target toxicities is higher. In two separate studies of CAR T-cells targeting MAGE-A3 epitopes, cross-reactivity occurred in cells expressing MAGE-A12 resulting in the destruction of healthy brain tissues in two patients and the cardiac protein titin in two patients resulting in death in all four cases (Al-Khadairi and Decock, 2019, Gjerstorff et al., 2015). The causes of these toxicities must be understood before continued experimentation with PRAME, as the antigen is known to be expressed on the surface of several different somatic tissues (Cassalia et al., 2024). The advances made in the field of immunotherapy through the discovery of ACT techniques are remarkable, having exciting implications for the future of cancer therapeutics. Unfortunately, significant limitations persist in the routine use of these therapeutics in clinical practice.

4.4. PRAME-CD3+ Bispecific Molecules (ImmTACs)

Brenetafusp (IMC-F106C) is an ImmTAC (Immune Mobilizing Monoclonal TCRs Against Cancer) bispecific molecule developed by Immunocore (Figure 3, Table 3). It targets the PRAME (Preferentially Expressed Antigen in Melanoma) antigen while also pulling T-cells to the tumor directly. Brenetafusp is designed to redirect T cells to recognize and kill PRAME-positive cancer cells. This molecule works by utilizing a soluble T cell receptor (TCR) that binds to intracellular cancer antigens, like PRAME, with high specificity and affinity. It then recruits T cells through an anti-CD3 effector function, activating the immune system to attack and destroy the cancerous cells. Brenetafusp has shown promising results in early clinical trials, both as a monotherapy and in combination with anti-PD1 therapies for treating late-stage melanoma patients. In the Phase 1 clinical trials of brenetafusp (IMC-F106C), an ImmTAC bispecific targeting PRAME, for patients with late-line, immune checkpoint pre-treated cutaneous melanoma, the treatment demonstrated promising disease control. The results showed a 58% disease control rate and a median progression-free survival (PFS) of 4.2 months in PRAME-positive patients. The treatment was well-tolerated, with manageable adverse events, primarily mild cytokine release syndrome (CRS) and rash. Additionally, 42% of ctDNA-evaluable, PRAME-positive patients had a molecular response, suggesting potential long-term benefits. Brenetafusp also showed better outcomes in earlier lines of therapy, and it was observed that T cell fitness was associated with increased clinical activity. The combination of brenetafusp with anti-PD1 therapy, such as pembrolizumab, also demonstrated efficacy in patients with heavily pre-treated cutaneous melanoma. The ongoing Phase 3 trial (PRISM-MEL-301) aims to further evaluate the efficacy of brenetafusp combined with nivolumab in first-line advanced melanoma patients.

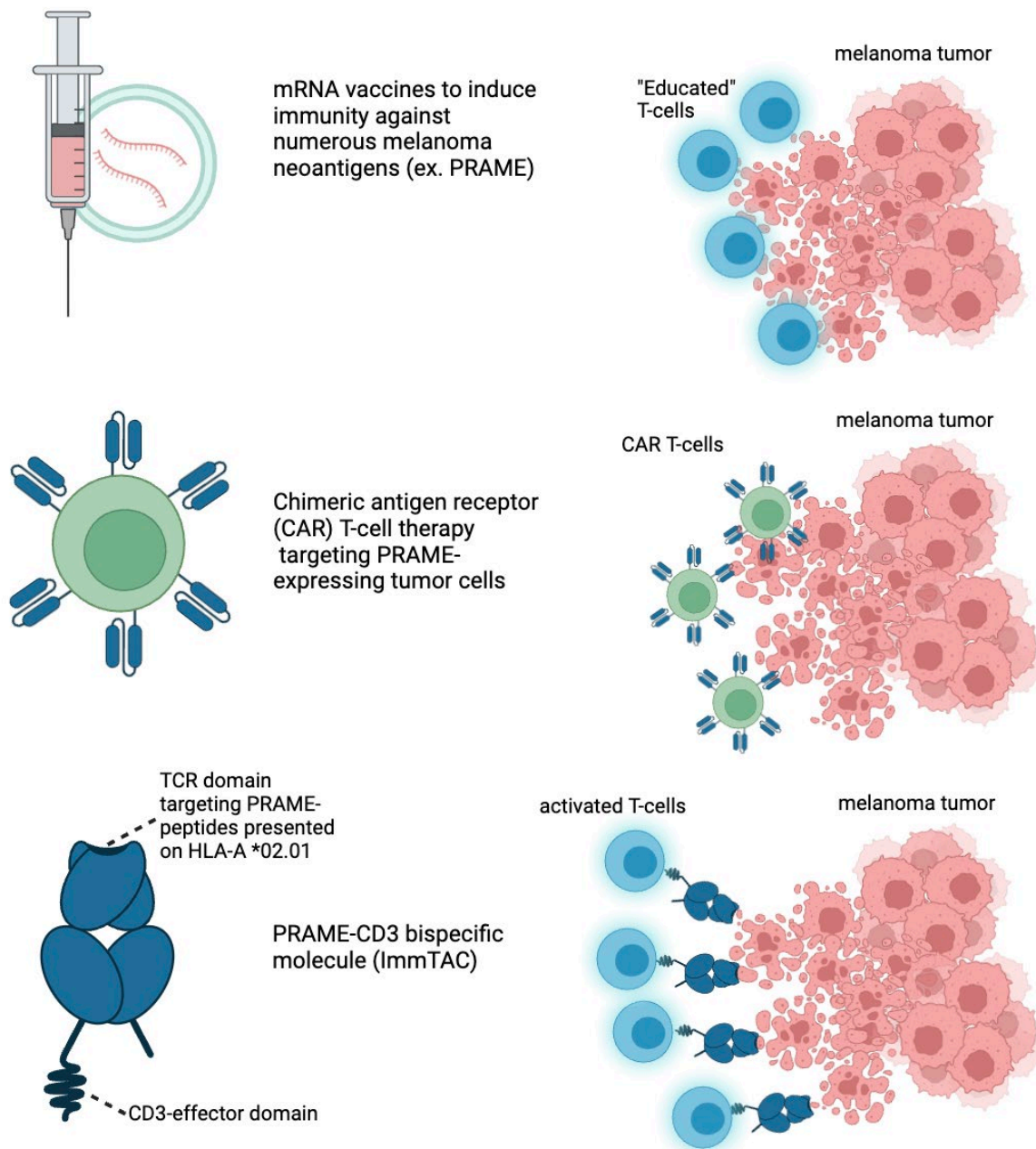


Figure 3. New immunotherapeutic strategies to target melanoma. mRNA vaccines can stimulate expression of neoantigens that are unique to patients' tumor profiles. This stimulates "educated" T-cells to target and kill melanoma tumor cells. CAR T-cells are engineered to attack patients tumor's based on engineered chimeric receptors; CAR-T therapy works by collecting a patient's T-cells, genetically modifying them to express a chimeric antigen receptor (CAR) that can specifically recognize and bind to patients' cancer cells, and then re-infusing these engineered T-cells back into the patient. Once inside the body, the modified T-cells can recognize and attack cancer cells that express the targeted antigen, leading to their destruction. CAR-T therapy works by collecting a patient's T-cells, genetically modifying them to express a chimeric antigen receptor (CAR) that can specifically recognize and bind to cancer cells, and then re-infusing these engineered T-cells back into the patient. Once inside the body, the modified T-cells recognize and attack cancer cells that express the targeted antigen, leading to their destruction. This approach harnesses the immune system's natural ability to fight cancer, with the CAR providing enhanced specificity and efficacy.

Table 3. Challenges in PRAME-Targeted Therapies.

Challenge	Description	Potential Solutions
Tumor Heterogeneity	Not all tumor cells express PRAME, leading to selective destruction of some but not all tumor cells.	Use of demethylating agents like Decitabine to induce PRAME expression across all tumor cells.
Immune Suppressive Tumor Microenvironment (TME)	Tumor cells recruit immune-suppressive cells (e.g., T-regs, MDSCs) that block immune response.	Combination therapies with immune checkpoint inhibitors like anti-PD1 (nivolumab) and anti-CTLA-4 (ipilimumab).
Negative Thymic Selection	T-cells recognizing PRAME as “self” may be destroyed during thymic selection, reducing the immune response.	Further research into overcoming thymic selection with engineered T-cells or improved antigen presentation.
Downregulation of HLA-1 Molecules	Tumor cells reduce the expression of HLA-1 molecules, preventing recognition by CD8+ T cells.	Use of MEK inhibitors or HDAC inhibitors to induce HLA-1 expression on tumor cells.

4.5. mRNA Vaccines

Merck and Moderna’s mRNA-4157/V940 vaccine, developed in combination with pembrolizumab (Keytruda), is showing promising results as a new approach to melanoma treatment. This personalized mRNA-based cancer vaccine works by encoding up to 34 neoantigens, tailored to the unique mutational profile of a patient’s tumor (Figure 3, Table 3). The vaccine is designed to trigger an immune response specifically against these cancer-related mutations. Key recent developments include the results from the Phase 2b KEYNOTE-942 trial, which demonstrated that combining the mRNA-4157/V940 vaccine with pembrolizumab significantly improved recurrence-free survival (RFS) in patients with high-risk resected melanoma compared to pembrolizumab alone (<https://www.aacr.org/about-the-aacr/newsroom/news-releases/adding-a-personalized-mrna-cancer-vaccine-to-immunotherapy-may-prolong-recurrence-free-survival-in-patients-with-high-risk-melanoma/>; <https://www.nature.com/articles/d41591-023-00072-0>; <https://www.merck.com/news/merck-and-moderna-initiate-phase-3-study-evaluating-v940-mrna-4157-in-combination-with-keytruda-pembrolizumab-for-adjuvant-treatment-of-patients-with-resected-high-riskstage-iiib-iv-melanom/>). Importantly, this benefit was observed regardless of the tumor mutational burden, which suggests that this vaccine could be effective across different patient profiles. The safety profile of the combination therapy was consistent with known adverse effects, such as mild cytokine release syndrome and skin rash. Following these encouraging results, the vaccine has received FDA Breakthrough Therapy Designation, and a Phase 3 trial has been initiated to further explore its efficacy in patients with melanoma. This combination therapy represents a potentially groundbreaking advance in melanoma treatment, offering a new personalized therapeutic option for patients at high risk of recurrence. As highly expressed as PRAME is within melanoma cells, it is likely that this approach will likely take advantage of PRAME along with other cancer-associated antigens.

4.6. Current Limitations in Immunotherapeutic Targeting of PRAME

The main limitations to the clinical use of PRAME-targeted vaccines and adoptive cell transfer are tumor heterogeneity, immune suppressive tumor microenvironment (TME), negative thymic selection, and downregulation of HLA-1 molecules. In any one tumor site, there is often heterogeneity in the expression of cancer specific antigens on HLA-1 molecules. In a melanoma tumor, for example, some cells may express PRAME on the surface while other cells do not. When immunotherapies are used targeting PRAME, the tumor cells not expressing PRAME are spared and continue to grow unharmed. Several methods are being developed to sidestep this issue. As mentioned previously, PRAME expression is the result of hypomethylation of the promoter region for the PRAME gene in the nucleus of cells. By utilizing molecules that *hypomethylate* the PRAME gene promoter, PRAME expression in tumor cells can be artificially induced, thereby limiting the heterogeneity of antigen expression. Demethylating agents and histone deacetylase (HDAC) inhibitors like Decitabine,

Sebularine, and DZNep could potentially be utilized as adjuvants for ACT and PRAME-targeted vaccines (Al-Khadairi and Decock, 2019, Gjerstorff et al., 2015, Nin and Deng, 2023). The second key limitation of PRAME-targeted immunotherapy is the immune suppressive TME. Tumor cells are often recognized by CD8⁺ cells, but the cancer cells suppress the response through various mechanisms. Myeloid-derived suppressor cells (MDSCs), tumor-associated macrophages, and regulatory T cells, recruited by tumor cells, secrete molecules promoting immunosuppression (Sterner and Sterner, 2021). Another way tumor cells cultivate an immunosuppressive TME is through the production of checkpoint molecules. To prevent frequent autoimmune reactions, somatic cells produce cytotoxic T lymphocyte-associated protein 4 (CTLA-4), programmed cell death protein 1 (PD-1) and programmed death ligand 1 (PD-L1) which signal to a T-cell to stop the release of cytotoxic enzymes. Tumor cells utilize these molecules to prevent their destruction by T-cells. To limit the immune suppression, combination therapies of cancer vaccines and checkpoint inhibitors like nivolumab and ipilimumab are being researched. Multiple preclinical and phase I trials have found increased efficacy compared to monotherapy. Several other trials exploring combination therapy are ongoing (Paston et al., 2021, Thomas et al., 2018). Another key limitation is the frequently observed lack of CD8⁺ proliferation secondary to negative thymic selection. Thymic selection of T-cells is a process where T-cells recognizing human antigens are destroyed to prevent autoimmunity. This allows T-cells recognizing foreign pathogens to proliferate and enter the circulation while T-cells recognizing human antigens are eliminated. Because members of the CTA family are expressed in normal germline tissues, it is likely that T-cells recognizing these endogenous antigens are eliminated to prevent autoimmunity (Al-Khadairi and Decock, 2019). The final limitation to discuss is the tumor cell's ability to downregulate the expression of HLA-1 molecules, thereby preventing recognition by CD8⁺ cells. One possible solution is to induce the expression of HLA-1 molecules in tumor cells through the administration of MEK inhibitors, demethylating agents, and HDAC inhibitors (Al-Khadairi and Decock, 2019, Spel et al., 2015). Additional ways to circumvent tumor heterogeneity, immune suppressive tumor microenvironment, negative thymic selection, and downregulation of HLA class 1 molecules should continued to be studied to improve the efficacy of immunotherapy in cancer patients.

5. Conclusion

PRAME is a remarkable protein with unique properties that aid in the diagnosis, prognosis, and treatment of melanoma. The selective presentation of PRAME on the HLA-1 molecules of tumor cells, not somatic cells, makes it a tumor-associated antigen of interest. Staining melanocytic lesions with PRAME IHC stains provides aid in accurate diagnoses with excellent sensitivity and specificity for melanoma versus benign melanocytes or dysplastic melanocytes. PRAME can also aid in margin assessment in melanoma excisions and in determining Breslow thickness for melanoma staging, particularly in superficial melanoma arising within a background of benign or dysplastic nevi. The utilization of PRAME as a marker for prognosis will likely become relevant as more biomarkers trials are performed. Multiple studies demonstrated increased PRAME expression to be associated with Class II UM, the more aggressive subtype, and associated with stage III-IV cancers. Using PRAME staining in the cancer staging process could yield a more accurate prognosis, guiding clinicians to make better treatment decisions. Moreover, we highlighted the prospect of PRAME as a therapeutic target in immunotherapy. The literature for PRAME-targeted immunotherapy is not as developed as it is for other TAAs but is still promising. The limitations including negative thymic selection, immune-suppressive TME, tumor heterogeneity, and downregulation of HLA-1 molecules are significant barriers to progress in the field (Figure 3, Table 3). By improving our understanding of PRAME, we can better harness the power of immunotherapy and related modalities to prolong survival in patients with metastatic melanoma, and hopefully, detect melanoma earlier with strategies to prevent development of metastases.

References

1. Al-Khadairi G, Decock J. Cancer Testis Antigens and Immunotherapy: Where Do We Stand in the Targeting of PRAME? *Cancers* 2019;11(7):984.
2. Al-Khadairi G, Naik A, Thomas R, Al-Sulaiti B, Rizly S, Decock J. PRAME promotes epithelial-to-mesenchymal transition in triple negative breast cancer. *Journal of Translational Medicine* 2019;17(1).
3. Alomari AK, Tharp AW, Umphress B, Kowal RP. The utility of <sc>PRAME</sc> immunohistochemistry in the evaluation of challenging melanocytic tumors. *Journal of Cutaneous Pathology* 2021;48(9):1115-23.
4. Brown G. Deregulation of All-Trans Retinoic Acid Signaling and Development in Cancer. *International Journal of Molecular Sciences* 2023;24(15):12089.
5. Cassalia F, Danese A, Tudurachi I, Federico S, Zambello A, Guidotti A, et al. PRAME Updated: Diagnostic, Prognostic, and Therapeutic Role in Skin Cancer. *International Journal of Molecular Sciences* 2024;25(3):1582.
6. Chen Y-J, Abila B, Mostafa Kamel Y. CAR-T: What Is Next? *Cancers* 2023;15(3):663.
7. Dafni U, Michielin O, Lluesma SM, Tsourti Z, Polydoropoulou V, Karlis D, et al. Efficacy of adoptive therapy with tumor-infiltrating lymphocytes and recombinant interleukin-2 in advanced cutaneous melanoma: a systematic review and meta-analysis. *Annals of Oncology* 2019;30(12):1902-13.
8. Duarte AF, Sousa-Pinto B, Azevedo LF, Barros AM, Puig S, Malvey J, et al. Clinical ABCDE rule for early melanoma detection. *European Journal of Dermatology* 2021;31(6):771-8.
9. Durgeau A, Virk Y, Corgnac S, Mami-Chouaib F. Recent Advances in Targeting CD8 T-Cell Immunity for More Effective Cancer Immunotherapy. *Frontiers in Immunology* 2018;9.
10. Eno J. Immunotherapy Through the Years. *J Adv Pract Oncol* 2017;8(7):747-53.
11. Epping MT, Hart AAM, Glas AM, Krijgsman O, Bernards R. PRAME expression and clinical outcome of breast cancer. *British Journal of Cancer* 2008;99(3):398-403.
12. Field MG, Decatur CL, Kurtenbach S, Gezgin G, Van Der Velden PA, Jager MJ, et al. PRAME as an Independent Biomarker for Metastasis in Uveal Melanoma. *Clinical Cancer Research* 2016a;22(5):1234-42.
13. Field MG, Durante MA, Decatur CL, Tarlan B, Oelschlager KM, Stone JF, et al. Epigenetic reprogramming and aberrant expression of PRAME are associated with increased metastatic risk in Class 1 and Class 2 uveal melanomas. *Oncotarget* 2016b;7(37):59209-19.
14. Fratta E, Coral S, Covre A, Parisi G, Colizzi F, Danielli R, et al. The biology of cancer testis antigens: Putative function, regulation and therapeutic potential. *Molecular Oncology* 2011;5(2):164-82.
15. Giaquinto AN, Sung H, Miller KD, Kramer JL, Newman LA, Minihan A, et al. Breast Cancer Statistics, 2022. *CA: A Cancer Journal for Clinicians* 2022;72(6):524-41.
16. Gjerstorff MF, Andersen MH, Ditzel HJ. Oncogenic cancer/testis antigens: prime candidates for immunotherapy. *Oncotarget* 2015;6(18):15772-87.
17. Googe PB, Flanagan KL, Miedema JR. Preferentially Expressed Antigen in Melanoma Immunostaining in a Series of Melanocytic Neoplasms. *Am J Dermatopathol* 2021;43(11):794-800.
18. Greiner J, Schmitt M, Li L, Giannopoulos K, Bosch K, Schmitt A, et al. Expression of tumor-associated antigens in acute myeloid leukemia: implications for specific immunotherapeutic approaches. *Blood* 2006;108(13):4109-17.
19. Griffioen M, Kessler JH, Borghi M, Van Soest RA, Van Der Minne CE, Nouta J, et al. Detection and Functional Analysis of CD8+ T Cells Specific for PRAME: a Target for T-Cell Therapy. *Clinical Cancer Research* 2006;12(10):3130-6.
20. Gutzmer R, Rivoltini L, Levchenko E, Testori A, Utikal J, Ascierto PA, et al. Safety and immunogenicity of the PRAME cancer immunotherapeutic in metastatic melanoma: results of a phase I dose escalation study. *ESMO Open* 2016;1(4):e000068.
21. Hanahan D. Hallmarks of Cancer: New Dimensions. *Cancer Discovery* 2022;12(1):31-46.
22. Haslauer T, Greil R, Zaborsky N, Geisberger R. CAR T-Cell Therapy in Hematological Malignancies. *Int J Mol Sci* 2021;22(16).
23. Ikeda H, Lethé B, Lehmann F, Van Baren N, Baurain J-F, De Smet C, et al. Characterization of an Antigen That Is Recognized on a Melanoma Showing Partial HLA Loss by CTL Expressing an NK Inhibitory Receptor. *Immunity* 1997;6(2):199-208.
24. Iura K, Kohashi K, Hotokebuchi Y, Ishii T, Maekawa A, Yamada Y, et al. Cancer-testis antigens PRAME and NY-ESO-1 correlate with tumour grade and poor prognosis in myxoid liposarcoma. *The Journal of Pathology: Clinical Research* 2015;1(3):144-59.
25. Kessler JH, Beekman NJ, Bres-Vloemans SA, Verdijk P, Van Veelen PA, Kloosterman-Joosten AM, et al. Efficient Identification of Novel Hla-A*0201-Presented Cytotoxic T Lymphocyte Epitopes in the Widely Expressed Tumor Antigen Prame by Proteasome-Mediated Digestion Analysis. *The Journal of Experimental Medicine* 2001;193(1):73-88.
26. Lezcano C, Jungbluth AA, Busam KJ. Comparison of Immunohistochemistry for PRAME With Cytogenetic Test Results in the Evaluation of Challenging Melanocytic Tumors. *Am J Surg Pathol* 2020a;44(7):893-900.

27. Lezcano C, Jungbluth AA, Busam KJ. PRAME Immunohistochemistry as an Ancillary Test for the Assessment of Melanocytic Lesions. *Surg Pathol Clin* 2021;14(2):165-75.
28. Lezcano C, Jungbluth AA, Nehal KS, Hollmann TJ, Busam KJ. PRAME Expression in Melanocytic Tumors. *American Journal of Surgical Pathology* 2018;42(11):1456-65.
29. Lezcano C, Pulitzer M, Moy AP, Hollmann TJ, Jungbluth AA, Busam KJ. Immunohistochemistry for PRAME in the Distinction of Nodal Nevi From Metastatic Melanoma. *Am J Surg Pathol* 2020b;44(4):503-8.
30. Li J, Yin J, Zhong J, Yang Z, Tang A, Li S. Clinicopathological and Prognostic Significance of PRAME Overexpression in Human Cancer: A Meta-Analysis. *BioMed Research International* 2020;2020:1-9.
31. McBride JD, McAfee JL, Piliang M, Bergfeld WF, Fernandez AP, Ronen S, et al. Preferentially expressed antigen in melanoma and p16 expression in acral melanocytic neoplasms. *J Cutan Pathol* 2022;49(3):220-30.
32. Moore LD, Le T, Fan G. DNA Methylation and Its Basic Function. *Neuropsychopharmacology* 2013;38(1):23-38.
33. Moore MW, Gasparini R. FISH as an effective diagnostic tool for the management of challenging melanocytic lesions. *Diagnostic Pathology* 2011;6(1):76.
34. Nagai Y, Ambinder AJ. The Promise of Retinoids in the Treatment of Cancer: Neither Burnt Out Nor Fading Away. *Cancers* 2023;15(14):3535.
35. Naik A, Thomas R, Al-Khadairi G, Bacha R, Hendrickx W, Decock J. Cancer testis antigen PRAME: An anti-cancer target with immunomodulatory potential. *Journal of Cellular and Molecular Medicine* 2021;25(22):10376-88.
36. Nin DS, Deng L-W. Biology of Cancer-Testis Antigens and Their Therapeutic Implications in Cancer. *Cells* 2023;12(6):926.
37. Parra O, Ma W, Li Z, Coffing BN, Linos K, LeBlanc RE, et al. PRAME expression in cutaneous melanoma does not correlate with disease-specific survival. *J Cutan Pathol* 2023;50(10):903-12.
38. Paston SJ, Brentville VA, Symonds P, Durrant LG. Cancer Vaccines, Adjuvants, and Delivery Systems. *Frontiers in Immunology* 2021;12.
39. Pop AM, Monea M, Olah P, Moraru R, Cotoi OS. The Importance of Immunohistochemistry in the Evaluation of Tumor Depth of Primary Cutaneous Melanoma. *Diagnostics* 2023;13(6):1020.
40. Pujol J-L, De Pas T, Rittmeyer A, Vallières E, Kubisa B, Levchenko E, et al. Safety and Immunogenicity of the PRAME Cancer Immunotherapeutic in Patients with Resected Non-Small Cell Lung Cancer: A Phase I Dose Escalation Study. *Journal of Thoracic Oncology* 2016;11(12):2208-17.
41. Ronchi A, Zito Marino F, Moscarella E, Brancaccio G, Argenziano G, Troiani T, et al. PRAME Immunocytochemistry for the Diagnosis of Melanoma Metastases in Cytological Samples. *Diagnostics* 2022;12(3):646.
42. Rosenberg SA, Packard BS, Aebersold PM, Solomon D, Topalian SL, Toy ST, et al. Use of tumor-infiltrating lymphocytes and interleukin-2 in the immunotherapy of patients with metastatic melanoma. A preliminary report. *N Engl J Med* 1988;319(25):1676-80.
43. Roszik J, Wang W-L, Livingston JA, Roland CL, Ravi V, Yee C, et al. Overexpressed PRAME is a potential immunotherapy target in sarcoma subtypes. *Clinical Sarcoma Research* 2017;7(1).
44. Santandrea G, Valli R, Zanetti E, Ragazzi M, Pampena R, Longo C, et al. Comparative Analysis of PRAME Expression in 127 Acral and Nail Melanocytic Lesions. *Am J Surg Pathol* 2022;46(5):579-90.
45. Scheffler AC, Koca E, Bernicker EH, Correa ZM. Relationship between clinical features, GEP class, and PRAME expression in uveal melanoma. *Graefes Archive for Clinical and Experimental Ophthalmology* 2019;257(7):1541-5.
46. Schenk T, Stengel S, Goellner S, Steinbach D, Saluz HP. Hypomethylation of PRAME is responsible for its aberrant overexpression in human malignancies. *Genes Chromosomes Cancer* 2007;46(9):796-804.
47. Scolyer RA, Long GV, Thompson JF. Evolving concepts in melanoma classification and their relevance to multidisciplinary melanoma patient care. *Molecular Oncology* 2011;5(2):124-36.
48. Seedor RS, Orloff M. Treatment of Metastatic Melanoma in the Elderly. *Current Oncology Reports* 2022;24(7):825-33.
49. Shang B, Gao A, Pan Y, Zhang G, Tu J, Zhou Y, et al. CT45A1 acts as a new proto-oncogene to trigger tumorigenesis and cancer metastasis. *Cell Death & Disease* 2014;5(6):e1285-e.
50. Simpson AJG, Caballero OL, Jungbluth A, Yao-Tseng C, Old LJ. Cancer/testis antigens, gametogenesis and cancer. *Nature Reviews Cancer* 2005;5(8):615-25.
51. Spel L, Boelens J-J, Van Der Steen DM, Blokland NJG, Van Noesel MM, Molenaar JJ, et al. Natural killer cells facilitate PRAME-specific T-cell reactivity against neuroblastoma. *Oncotarget* 2015;6(34):35770-81.
52. Sterner RC, Sterner RM. CAR-T cell therapy: current limitations and potential strategies. *Blood Cancer Journal* 2021;11(4).
53. Szymański Ł, Skopek R, Palusińska M, Schenk T, Stengel S, Lewicki S, et al. Retinoic Acid and Its Derivatives in Skin. *Cells* 2020;9(12):2660.

54. Tajeddine N, Gala J-L, Louis M, Van Schoor M, Tombal B, Gailly P. Tumor-Associated Antigen Preferentially Expressed Antigen of Melanoma (PRAME) Induces Caspase-Independent Cell Death *In vitro* and Reduces Tumorigenicity *In vivo*. *Cancer Research* 2005;65(16):7348-55.
55. Tan P, Zou C, Yong B, Han J, Zhang L, Su Q, et al. Expression and prognostic relevance of PRAME in primary osteosarcoma. *Biochem Biophys Res Commun* 2012;419(4):801-8.
56. Thomas R, Al-Khadairi G, Roelands J, Hendrickx W, Dermime S, Bedognetti D, et al. NY-ESO-1 Based Immunotherapy of Cancer: Current Perspectives. *Front Immunol* 2018;9:947.
57. Toyama A, Siegel L, Nelson AC, Najmuddin M, Bu L, LaRue R, et al. Analyses of molecular and histopathologic features and expression of PRAME by immunohistochemistry in mucosal melanomas. *Mod Pathol* 2019;32(12):1727-33.
58. Van Not OJ, Van Den Eertwegh AJM, Haanen JB, Blank CU, Aarts MJB, Van Breeschoten J, et al. Improving survival in advanced melanoma patients: a trend analysis from 2013 to 2021. *eClinicalMedicine* 2024;69:102485.
59. Weber JS, Vogelzang NJ, Ernstoff MS, Goodman OB, Cranmer LD, Marshall JL, et al. A Phase 1 Study of a Vaccine Targeting Preferentially Expressed Antigen in Melanoma and Prostate-specific Membrane Antigen in Patients With Advanced Solid Tumors. *Journal of Immunotherapy* 2011;34(7):556-67.
60. Wen M, Ren H, Zhang S, Li T, Zhang J, Ren P. CT45A1 promotes the metastasis of osteosarcoma cells in vitro and in vivo through β -catenin. *Cell Death & Disease* 2021;12(7).
61. Wolchok JD, Chiarion-Sileni V, Gonzalez R, Grob J-J, Rutkowski P, Lao CD, et al. Long-Term Outcomes With Nivolumab Plus Ipilimumab or Nivolumab Alone Versus Ipilimumab in Patients With Advanced Melanoma. *Journal of Clinical Oncology* 2022;40(2):127-37.
62. Xu Y, Zou R, Wang J, Wang ZW, Zhu X. The role of the cancer testis antigen PRAME in tumorigenesis and immunotherapy in human cancer. *Cell Proliferation* 2020;53(3).
63. Zhang YH, Lu AD, Yang L, Li LD, Chen WM, Long LY, et al. PRAME overexpression predicted good outcome in pediatric B-cell acute lymphoblastic leukemia patients receiving chemotherapy. *Leuk Res* 2017;52:43-9.

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