

Review

Not peer-reviewed version

Recognizing the Role of Insulin Resistance in Polycystic Ovary Syndrome: A Paradigm Shift from a Glucose-Centric Approach to an InsulinCentric Model

<u>Jim Parker</u>*, Lara Briden , <u>Felice Gersh</u>

Posted Date: 19 May 2025

doi: 10.20944/preprints202505.1501.v1

Keywords: Polycystic ovary syndrome; insulin; insulin resistance; hyperinsulinemia; metabolic; hyperandrogenism, chronic inflammation; glucose-centric; insulin-centric



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Article

Recognizing the Role of Insulin Resistance in Polycystic Ovary Syndrome: A Paradigm Shift from a Glucose-Centric Approach to an Insulin-Centric Model

Jim Parker 1,*, Lara Briden 2 and Felice L. Gersh 3

- ¹ School of Medicine, University of Wollongong, Wollongong 2522, Australia
- ² Centre for Menstrual Cycle and Ovulation Research, University of British Columbia, Vancouver, British Columbia, Canada; Lara@larabriden.com
- ³ College of Medicine, University of Arizona, Tucson, AZ 85004, USA; felicelgersh@yahoo.com
- * Correspondence: jimparker@ozemail.com.au

Abstract: Polycystic ovary syndrome (PCOS) is a common metabolic-endocrine disorder affecting women of reproductive age, and insulin resistance (IR) is a key pathophysiological feature. Current medical education and clinical practice emphasize glucose-centric approaches in PCOS management, and IR testing is often overlooked due to inadequate knowledge or perceived lack of reliable assessment methods. Additionally, the glucose-focused paradigm has been the standard of care for decades. But this approach has led to delayed diagnosis of progressive metabolic and reproductive consequences, leaving many patients underdiagnosed and undertreated. Therefore, we propose a paradigm shift towards an insulin-centric model for PCOS management. This new approach aims to diagnose IR at an earlier stage enabling timely implementation of effective lifestyle and treatment strategies. By focusing on IR, clinicians can potentially limit the progression of PCOS-related reproductive and metabolic diseases. The insulin-centric model involves comprehensive IR screening, dynamic insulin testing, personalized lifestyle and insulin-sensitizing interventions, and regular monitoring of insulin and glycemic parameters. Adopting this paradigm in clinical practice could improve patient outcomes, offering a more proactive approach to managing PCOS and related metabolic disorders. Furthermore, this model has broader implications, potentially transforming treatment approaches for various chronic diseases beyond PCOS.

Keywords: polycystic ovary syndrome; insulin; insulin resistance; hyperinsulinemia; metabolic; hyperandrogenism; chronic inflammation; glucose-centric; insulin-centric

1. Introduction

Polycystic ovary syndrome (PCOS) affects 8-13% of women and usually presents in adolescence with a complex mixture of symptoms that result from underlying metabolic and endocrine disturbance of homeostatic networks [1–3]. Developmental programming of inherited gene variants predisposes women with PCOS to reduced insulin sensitivity which provided an adaptive survival advantage in ancestral environments [4–8]. However, in the modern environment, reduced insulin sensitivity predisposes to maladaptive metabolic, hormonal, and symptom responses [1,2,4–11]. In its early stages, PCOS can therefore be considered a reversible disturbance of physiology in response to environmental stressors, rather than a true disease entity. This characterization of PCOS is supported by the International Guidelines, which provide evidence that many of the symptoms and features of PCOS are reversible following diet, exercise, and other lifestyle interventions [12].

PCOS is associated with a number of chronic diseases and complications [1,12,13]. These include reproductive problems (subfertility, implantation failure, miscarriage), pregnancy complications (pre-eclampsia, pre-term labour, fetal growth restriction, gestational diabetes, and stillbirth), and

metabolic diseases (obesity, type 2 diabetes, metabolic syndrome, metabolic-associated liver disease, fatty pancreas disease, dyslipidemia, hypertension, renal and cardiovascular disease, and cancer) [14–17]. In many cases, the pathophysiological effects of insulin resistance (IR) and hyperinsulinemia on the ovary, brain, vascular endothelium, endometrium, placenta, and endocrine and metabolic systems, are already established [17]. As a result, IR and hyperinsulinemia are now recognized to be significant contributors to these long-term health complications [15], in addition to being major drivers of the core features of PCOS, which include hypothalamic and ovulatory dysfunction, and hyperandrogenism [18–22].

A major limitation of the prevailing glucose-centric approach is that measurable changes in serum glucose do not occur until decades after the onset of IR and hyperinsulinemia (Figure 1) [23]. As a result, IR frequently goes undetected during its early, silent phase, when intervention would be most effective [24]. In this paper, we present evidence to support a paradigm shift towards an insulincentric model for the assessment and management of PCOS. We propose that early identification of IR in adolescents and women with PCOS would enable timely intervention and reduce the risk of subsequent metabolic and reproductive complications. This model is evidence-based, feasible, and suitable for integration into routine clinical practice. Further research is needed to improve the predictive accuracy of surrogate markers of IR, refine early intervention strategies, and enhance knowledge translation to both health professionals and women.

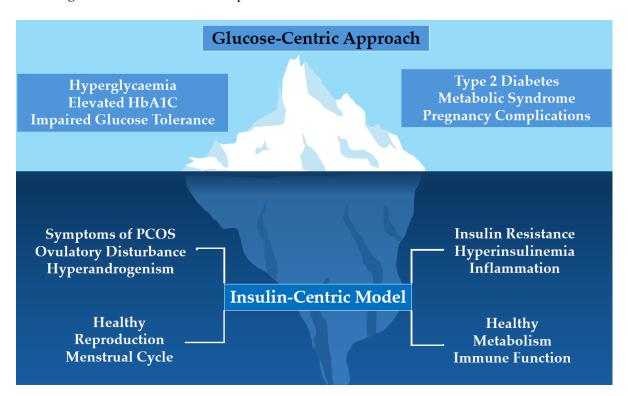


Figure 1. The observed features of the glucose-centric approach represent the late stages of the effects of insulin resistance or "tip of the iceberg". HbA1C = Hemoglobin A1C.

2. Scope and Methodology

The aim of this paper is to present a review of the current state of research on the advantages and disadvantages of adopting an insulin-centric model of PCOS, compared with the longstanding glucose-centric approach. Sections 3 to 7 provide the background and rationale for adopting a new approach to early diagnosis and management of metabolic disturbance in women with PCOS. This model emphasizes the central role of hyperinsulinemia and IR as driving forces that shape disease progression.

Section 3 provides an overview of the glucose-centric paradigm currently used for the assessment and management of women with PCOS. This section describes the history, rationale,

testing, predictive value for detecting complications, treatment strategies, and strengths and limitations of this paradigm in a contemporary environment. Section 4 is an up-to-date summary of the physiological actions of insulin. Section 5 discusses the adaptive significance of reduced insulin sensitivity and IR in women with PCOS. Section 6 discusses the pathophysiology of IR and hyperinsulinemia in PCOS. Section 7 describes the features of the proposed insulin-centric model. This section provides evidence supporting an insulin-centric model in women with PCOS. It includes the rationale for the need to change, recommendations for testing, a plan for phase-based therapeutic interventions, and identification of future research and treatment candidates. Section 8 is the framework for an insulin-centric model and was prepared with the assistance of the Generative Artificial Intelligence (AI) tool "Microsoft Copilot". Copilot was asked the question "Design an insulin-centric model for the assessment and management of PCOS".

The list of bibliographic references is based on PubMed, MEDLINE, Scopus, and Google Scholar databases. Databases were searched from inception to April 2025 repeatedly over many years. This narrative review summarizes the relevant literature and provides a new perspective on the need for greater emphasis on hyperinsulinemia and IR in women with PCOS. We propose a paradigm shift towards an insulin-centric model for the assessment and management of women with PCOS.

3. The Glucose-Centric Model of Insulin Resistance in PCOS

3.1. Origins of the Glucose-Centric Model

PCOS can be a progressive metabolic disease and women with PCOS have a significantly increased risk of developing impaired glucose tolerance (IGT), gestational diabetes mellitis (GDM), and type 2 diabetes mellitis (T2DM) [1,15,25–27]. As a result, the glucose-centric model of diabetes has been used for the assessment and management of PCOS (Figure 2).

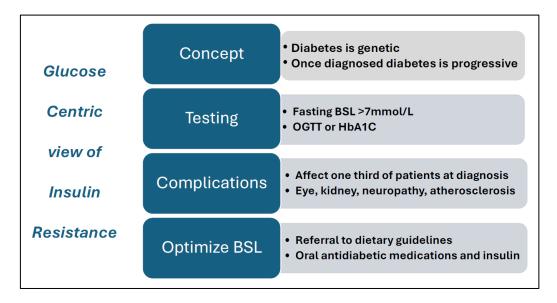


Figure 2. Features of a glucose-centric model of insulin resistance. Abbreviations: BSL = Blood Sugar Levels; mmol/L = millimole per Litre; OGTT = Oral Glucose Tolerance Test; HbA1C = Hemoglobin A1C.

Diabetes has been known since antiquity and the term mellitis was added in the seventeenth century to describe the associated sweet taste of urine that was subsequently determined to be sugar [28]. Diabetes was also known to be associated with dietary foods such as rice, cereals and sweets [28]. Insulin was isolated from the pancreas in 1921 and subsequently became a life-saving treatment for type 1 diabetes [29]. In 1935 Himsworth described the difference between type 1 diabetes (insulin sensitive) and T2DM (insulin insensitive)[30]. Although vaguely defined, the term IR was subsequently applied to people with insulin insensitive T2DM [31,32].

The original description of PCOS by Stein and Levinthal emphasized the clinical features of menstrual irregularity and infertility [33]. PCOS was subsequently characterized as a reproductive disorder until it was recognized that some women with typical PCOS had acanthosis nigricans and IR [34]. Further research showed that obese women with PCOS had elevated blood glucose levels following oral glucose tolerance testing (OGTT). Large cross-sectional studies during the 1990's revealed an increased prevalence of IGT and T2DM in women with PCOS [35,36]. The current glucose-centric model of PCOS therefore originated from the long-standing medical framework used to diagnose and manage type 1 and T2DM, for which blood glucose levels are a well-established and clinically accessible biomarker. From the point at which PCOS was recognized to be linked with metabolic dysfunction and increased risk of diabetes in the 1990's [34], the default approach has been to use glucose-based assessments such as the OGTT. Accumulating evidence now shows that IR and hyperinsulinemia precedes hyperglycemia by decades and plays a key role in the pathophysiology of PCOS and related complications [2,23,37,38].

3.2. Assessment Based on the Glucose-Centric Model of Insulin Resistance in PCOS

Many of the diabetes testing methods introduced during the twentieth century, adopted pragmatically and without robust evidenced to support their accuracy or predictive value. The first clinical tests for the diagnosis of diabetes were qualitative tests to detect glucose in the urine [39]. The OGTT was introduced in 1922, and urine test strips in the 1950's (dextrostix) [40]. Glucose monitors were used in the 1970's, despite poor precision and accuracy [41]. The hemoglobin A1c (HbA1c) test was introduced in the 1970's and provided an estimate of the average blood sugar over months rather than a single point in time [42]. In the 1980's self-monitoring of blood glucose became the standard of care. The first continuous glucose monitor (CGM) was approved in 1999 and has been refined and evaluated over the past 25 years [43].

Tests used in the glucose-centric model in women with PCOS, include fasting plasma glucose, the OGTT, and HbA1c. These tests aim to identify IGT or T2DM, rather than IR and hyperinsulinaemia. Their continued use is due in part to their standardization, accessibility, and clear diagnostic cutoffs. Nevertheless, there are many limitations to the accuracy, reproducibility, and predictive value of all these testing methods [44–46].

The International Guidelines for the assessment and management of PCOS advise screening for glycaemic statis in all adults and adolescents with PCOS [12]. The 75gm OGTT is recommended as a first-line test, regardless of body mass index (BMI), as it is believed to be the most accurate. According to the Guidelines, "If an OGTT cannot be performed, fasting plasma glucose and/or glycated hemoglobin (HbA1c) could be considered, noting significantly reduced accuracy" [12].

The rationale for using these tests is based on their role in diabetes diagnosis, their ease of administration, and the extensive body of research linking elevated glucose to long-term complications such as retinopathy, renal disease, neuropathy, and vascular disease [20,47,48]. All of the testing methods were introduced into clinical practice due to the belief that they would provide improvements in patient care, despite very limited supportive evidence. The current adherence to glucose-centric testing using the OGTT, fasting BSL, and HbA1c testing, has impeded progress in the early diagnosis and prevention of metabolic dysfunction and complications of IR, hyperinsulinaemia, and hyperglycaemia in women with PCOS [19,20].

3.3. Treatments Based on the Glucose-Centric Model

Treatment approaches within the glucose-centric model of PCOS are largely derived from strategies used in T2DM prevention and management. They typically aim to delay the onset of overt hyperglycemia in IGT or reduce elevated blood glucose levels in T2DM. First-line recommendations include lifestyle advice focused on weight loss, increased physical activity, and diet [12]. Diet composition is suggested to be consistent with population guidelines for sustainable healthy eating, tailored to individual preferences and goals.



Pharmacological treatment for impaired blood glucose levels and T2DM in women with PCOS includes metformin, an insulin-sensitizing agent with glucose-lowering effects [49]. Metformin has multiple mechanisms of action that improve IR, symptoms of PCOS, and fertility [50]. Inositol supplementation is now considered a treatment option as it may improve metabolic and fertility outcomes [12].

3.4. Strengths of the Current Approach

The glucose-centric approach can be effective in reducing long-term complications in women with PCOS who have already developed IGT or T2DM [25]. Evidence supports the role of weight loss, dietary modification, and metformin in improving metabolic outcomes and reducing the risk of T2DM in this subset of patients [25,50,51].

Moreover, the glucose-centric model aligns with existing public health frameworks, particularly those aimed at identifying and managing prediabetes and T2DM [52]. For example, glucose-based screening integrates easily into primary care settings [46], facilitating early detection of overt glycemic abnormalities in high-risk PCOS populations [53]. Clinicians are generally familiar with interpreting these tests and treatment pathways are clearly defined in national and international guidelines.

3.5. Limitations and Clinical Consequences of the Translation Gap

By focusing primarily on glycemic thresholds, the glucose-centric model overlooks the early and often silent stage of IR and hyperinsulinemia that drives many of the reproductive, metabolic, and inflammatory manifestations of PCOS [49]. The disconnect between pathophysiological understanding and clinical assessment reflects a broader translation gap. While the research literature consistently identifies IR as a central mechanism in PCOS [2], clinical guidelines and routine practice continue to emphasize glucose assessment and management [12].

In addition, the outdated assumption that T2DM in PCOS is an inevitable and inherited progressive disease, rather than a largely preventable consequence of untreated IR, has delayed recognition of the underlying metabolic problem and hindered opportunities for early targeted intervention to prevent future chronic diseases. This belief contributed to an emphasis on managing blood glucose rather than addressing the upstream metabolic drivers of IR and hyperinsulinemia. Today however, there is growing evidence that lifestyle modification and early targeted interventions can significantly reduce the risk of disease progression [51]. An insulin-centric strategy allows for earlier detection and more proactive management potentially preventing both GDM and T2DM [54].

4. Overview of Insulin's Diverse Biological Actions

Insulin plays a central role in the regulation of human metabolism, energy storage, appetite regulation, immune function and inflammation, hormone regulation and reproduction, and vascular dynamics and blood pressure regulation (Table 1) [55–64]. Insulin facilitates glucose removal from the blood in insulin-dependent tissue such as skeletal muscle, cardiac muscle, adipose tissue, and endothelium and also acts as an anti-inflammatory hormone.

4.1. Cellular Actions of Insulin

Insulin is a peptide hormone only produced and secreted by the beta cells in the Islets of Langerhans of the pancreas [56,65]. Insulin binds with the extracellular domain of the alpha subunit of the insulin receptor, which induces autophosphorylation of tyrosine kinase on the intracellular side of the membrane. This initiates a cascade of signal transduction events via two key pathways that lead to different distal signalling responses in target tissues. These include the phosphatidylinositol-3 kinase (PI-3K) metabolic pathway which activates transcription factors such as forkhead box 01, tuberous sclerosis complex 1/2, sterol regulatory binding protein 1c, and the

mitogen-activated protein kinase (MAPK) pathway that activates cell growth and proliferation [56,66].

Table 1. Physiological actions of insulin.

Functions of Insulin	Mechanism	Reference
Pleiotropic cellular action	Tissue-specific action after binding to insulin receptor	[55]
Energy storage	Adipose: glucose uptake, triglyceride storage, inhibit lipolysis	[56]
	Muscle: glucose uptake, glycogen synthesis, inhibit proteolysis	[56]
	Liver: glycogen synthesis, inhibit gluconeogenesis	[56]
Glucagon antagonist	Pancreas: paracrine suppression of glucagon release	[57]
Anti-inflammatory	BSL: helps keep BSL normal by decreasing ROS and AGE	[58]
	Inhibits NF-κB and MCP-1 activated cytokine production	[59]
	Reduced NLRP3 inflammasome formation and TLR signalling	[58,60]
	Reduced leukocyte adhesion to endothelium	[61]
Kidney	Sodium reabsorption: water retention and volume expansion	[62]
	Reduced excretion of urate	[64]
Vasodilation	Arteriole: increased blood flow via endothelial nitric oxide	[63]
Tissue perfusion	Volume expansion and vasodilation	[62]
Blood pressure regulation	Volume expansion, vasodilation, altered peripheral resistance	[62]
Ovary	Stimulates androgen synthesis via insulin and IGF-1 receptors	[67]
Central Nervous System	Hypothalamus: suppresses appetite, modulates energy expenditure, regulates GnRH pulsatility	[68]
	Liver: CNS mediated regulation of hepatic glucose production	[69]
	Muscle: CNS mediated promotion of glucose uptake	[70]
	Adipose: CNS mediated suppression of lipolysis	[70] [70]

Abbreviations: BSL = Blood Sugar Level; ROS = Reactive Oxygen Species; AGE = Advanced Glycation End Products; NF-kB = Nuclear-Factor Kappa B; MCP-I = Monocyte Chemoattractant Protein-1; NLRP3 = Nucleotide-Binding Leukocyte-Rich Pyrin Domain Containing 3; TLR = Toll-Like Receptor; IGF-1 = Insulin-Like Growth Factor-1; CNS = Central Nervous System.

Activation of the PI-3K pathway induces GLUT4 translocation to the cellular membrane in insulin-dependent tissues such as muscle and adipose tissue [71]. The PI-3K signaling cascade can upregulate the transcription of key steroidogenic enzymes including CYP17A1 (17α -hydroxylase/17,20-lyase), which play a central role in converting androgenic precursors (pregnenolone and progesterone) into androgens (testosterone and androstenedione) [72].

4.2. Tissue-Specific Actions of Insulin

Insulin is an anabolic hormone that regulates a myriad of tissue-specific cellular processes, such as protein, fat, glucose, and glycogen synthesis, RNA and DNA synthesis, as well as cellular proliferation and differentiation. Importantly, insulin inhibits catabolic processes by inhibiting glucagon release from the pancreas, gluconeogenesis in the liver, proteolysis in muscle, and lipolysis in adipose tissue [56]. In this way, insulin acts as a metabolic switch between anabolic and catabolic pathways to control energy production, storage, and utilization, during feeding and fasting [73].

4.3. Anti-Inflammatory Actions of Insulin

Beyond metabolism, insulin links energy regulation and immune modulation, contributing to adaptive survival responses to environmental stressors [74]. Insulin inhibits transcription factor NF-kB and reduces the production of inflammatory cytokines [59], inhibits NRL inflammasome formation [60], decreases leukocyte adhesion to the endothelium [61], and prevents hyperglycaemia-induced production of reactive oxygen species and advanced glycation end products [58]. Under conditions of insulin sensitivity the anti-inflammatory effects of insulin may have an evolutionary



protective function to prevent overactivation of the immune system to small amounts of ingested antigens that cross the gastrointestinal barrier during food intake [75]. Insulin may therefore be part of an extensive network of communication mechanisms that contribute to the systemic regulation of the inflammatory response [2,76,77]. In contrast, IR is proinflammatory and part of an adaptive survival response to a variety of environmental challenges (see section 5) [78–80].

5. The Adaptive Significance of Reduced Insulin Sensitivity and Insulin Resistance in PCOS

5.1. Insulin Sensitivity as a Continuous Variable

Insulin sensitivity reflects the ability of insulin to remove glucose from the blood and restore normoglycaemia [32]. Decreased insulin sensitivity is a continuous variable that is classified as IR once it reaches an arbitrarily defined cut-off value, during an experimental hyperinsulinemic-euglycemic clamp test (eg 4.45 mg/kg/min) [81], or as an elevated surrogate marker test in clinical practice [82]. The functional consequences of reduced insulin sensitivity manifest as altered tissue responsiveness to insulin. This is particularly relevant in PCOS, where a systematic review of clamp studies reported that women with PCOS exhibit a 27% reduced sensitivity to insulin [7].

Despite the widespread use of the term IR, as yet, there is no universally agreed upon normal range for use in clinical practice. Variability in diagnostic cut-off values contributes to inconsistencies in reported prevalence rates of IR in PCOS [83]. Functionally, IR refers to a diminished biological response to insulin stimulation in target tissues that may reflect a physiological adaptation or a pathological condition, depending on the context [84].

5.2. Physiological Insulin Resistance as an Adaptive Survival Mechanism

Decreased tissue sensitivity to the physiological actions of insulin has become synonymous with the pathological effects of IR, despite the fact that reduced sensitivity to insulin is an evolutionary-conserved homeostatic survival mechanism [1,4,84,85]. Reduced insulin sensitivity appears to be a key inherited component in PCOS that improves survival in response to a range of internal and external environmental situations [1,86]. Physiological IR and hyperinsulinemia occur during systemic infection, trauma, starvation, adolescence, and pregnancy, and limit glucose uptake in insulin dependent tissues, such as muscle and adipose tissue [1,17,79,84]. IR therefore functions as an adaptive mechanism to redistribute glucose to tissues in need, such as immune cells, brain, bone, and the fetus [17,78].

From an evolutionary perspective, women with PCOS can be considered "metabolically elite", as they can store energy efficiently, and redirect glucose to tissues with increased demand for energy.

5.3. The Shift to Pathological Insulin Resistance

Insulin resistance becomes pathological when the adaptive reduction in insulin sensitivity persists or is exaggerated in response to modern environmental stressors. In this state, metabolic dysfunction arises from disrupted insulin signaling pathways, often triggered by recurrent dietinduced hyperglycemia and hyperinsulinemia, metabolic intermediates, chronic stress, inflammatory cytokines, hormonal imbalances, and exposure to endocrine-disrupting chemicals [56,87].

Pathological IR is always associated with hyperinsulinemia, which act together and contribute to the symptoms, biochemical, metabolic, immune, and reproductive features of PCOS (see section 6) [1]. Hyperinsulinemia is an early indicator of metabolic dysfunction and is associated with many genetic, nutritional and environmental factors [88]. Dysregulation of insulin biology is also a key pathophysiological component of the initiation and progression of complications and chronic disease in women with PCOS [13,15,89].

6. Insulin Resistance and Hyperinsulinemia as Central Drivers of PCOS and Related Complications

6.1. Bidirectional Relationship Between Insulin Resistance and Chronic Inflammation

The pathogenesis of PCOS in contemporary populations is thought to be due to the epigenetic effects of nutritional, environmental, and lifestyle exposures, on inherited adaptive gene variants [1,2,4–11]. Chronic low-grade systemic inflammation and IR are central drivers of the pathophysiology of PCOS [2,90,91]. The bidirectional relationship between IR and chronic inflammation creates a vicious cycle that exacerbates both the metabolic and reproductive disturbances seen in women with PCOS.

Inflammation is an evolutionary-conserved adaptive survival response of the immune system primarily directed at combating infection, toxins, allergens, and tissue injury [92]. Inflammation can be physiological or pathological and optimal health is achieved by balancing anti- and proinflammatory effects aimed at removing infected, damaged, or aging cells [93]. Inflammatory cytokines, chemokines, and extracellular vesicles help support a successful protective response by connecting neuroendocrine and immunometabolic systems [94–96].

As part of this response, inflammation induces a rapid adaptive metabolic response by downregulating insulin signalling and GLUT4 translocation, resulting in reduced insulin sensitivity and glucose uptake, IR, and increased serum glucose levels [97]. This is a physiological response to redirect energy to immune cells and vital organs [98]. As a result, immune cells can increase their energy demand from 10% of basal energy use, to 30% when required [99]. In addition, inflammation and metabolic adaptation are linked to reproduction to optimize fertility [100]. Consequently, both IR and inflammation can increase ovarian androgen production, which augments IR and downregulates ovulation [101,102].

Women with PCOS are believed to have an evolutionary beneficial "proinflammatory design" that results in a heightened physiological response [8,103]. In a modern environment, poor-quality diet, and a range of environmental and lifestyle factors result in chronic activation of the immune system and metaflammation [104]. The combined effects of IR and chronic low-grade inflammation contribute to the symptoms, biochemical and endocrine features, and progressive metabolic diseases associated with PCOS [1,2].

IR can cause inflammation via direct or indirect mechanisms. IR is accompanied by reduced uptake and oxidation of glucose which results in hyperglycaemia, that in turn triggers oxidative stress [105]. Oxidative stress activates innate intracellular defense systems, such as the endoplasmic reticular stress response and inflammasome formation, that initiate proinflammatory cytokine production and inflammation [96]. Hyperglycaemia-associated advanced glycation end-products (AGE) cause oxidative stress and inflammatory cytokine production, that are protective if short-lived and limited, and pathological if chronic and excessive [106,107]. The bidirectional relationship between IR and chronic inflammation ensures a coordinated and co-operative physiological response to stressors, that become self-reinforcing and pathological following excessive activation from nutritional, environmental, and lifestyle factors in the contemporary environment.

6.2. Insulin Resistance Disrupts Ovarian Function

Ovarian tissue remains sensitive to insulin even in the presence of hyperinsulinemia associated with IR [91]. This heightened ovarian sensitivity may reflect an adaptive mechanism to suppress ovulation under conditions of metabolic stress or scarcity [2]. A maladaptive consequence is that the chronically elevated insulin levels associated with pathological IR can significantly disrupt ovarian function.

Insulin directly stimulates androgen production by theca cells via the PI-3K and MAPK pathways and through the inositolglycan signal transduction system [67,72,108]. Insulin also increases the amplitude of gonadotropin-releasing hormone (GnRH)-stimulated LH pulses [109,110] and acts synergistically with LH to enhance androgen synthesis, primarily via direct activation of

ovarian insulin and IGF-1 receptors [67]. Insulin also promotes serine phosphorylation of insulin receptor substrates and possibly LH receptors, amplifying the androgenic response of theca cells even in the absence of high LH levels [111]. In parallel, insulin suppresses hepatic production of sexhormone binding globulin, increasing the bioavailability of circulating androgens and further enhancing their activity in target tissues [112].

Hyperinsulinemia and IR also disrupt ovarian function indirectly through oxidative stress and inflammatory pathways. Hyperglycemia-associated AGE are elevated in women with PCOS and IR [113] and accumulate in the follicular environment, impairing steroidogenesis and disrupting granulosa cell function [2,114,115]. Together with hyperinsulinemia, AGE contribute to mitochondrial oxidative stress, diminished oocyte quality and maturation, and ovulatory disturbance [91,116].

Additionally, IR alters ovarian adipokine signaling by reduced adiponectin and increased leptin. These changes impair granulosa and theca cell function and promote a pro-inflammatory, insulinresistant local environment [51]. Insulin may increase anti-Müllerian hormone (AMH) production by granulosa cells, potentially contributing to follicular arrest, the accumulation of small antral follicles, and ovulatory disturbance [117]. Another important mechanism involves insulin-mediated impairment of follicle-stimulating hormone (FSH) signaling in granulosa cells, leading to downregulation of aromatase expression [34]. Reduced ovarian aromatase activity decreases ovarian estradiol production and contributes to androgen excess and ovulatory dysfunction, which are all hallmarks of PCOS [118].

6.3. Insulin Resistance Induces Neuroendocrine Disturbance and Dysregulates the Hypothalamic-Pituitary-Ovarian-Axis

The hypothalamic-pituitary-gonadotropin system plays a central role in the regulation of reproduction by integrating different neuroendocrine, metabolic and environmental signals [119]. GnRH secretion is regulated by feedback loops involving gonadal hormones, primarily estrogen and progesterone, which exert both stimulatory and inhibitory effects on the hypothalamic-pituitary-ovarian (HPO) axis and maintain cyclical reproductive function [120]. Insulin receptors are expressed in the hypothalamus, particularly on kisspeptin and pro-opiomelanocortin neurons, where they contribute to the regulation of GnRH pulsatility and energy homeostasis [110,121]. Under conditions of normal insulin sensitivity, these pathways support coordinated reproductive and metabolic function. In states of IR, however, central insulin signalling is disrupted, leading to altered GnRH neuronal activity, increased amplitude of GnRH-stimulated LH pulses and impaired or absent ovulation [91,122]. The absence of ovulation further dysregulates GnRH/LH pulsatility by removing the negative feedback normally provided by luteal-phase progesterone [123].

In a normal ovulatory cycle, progesterone and its neurosteroid metabolites, such as allopregnanolone, interact with GABA-A receptors on kisspeptin neurons to inhibit GnRH secretion and stabilize the HPO axis [123,124]. This inhibitory action is diminished by androgens, which can bind to hypothalamic progesterone receptors [125], but enhanced by ovulatory levels of estradiol, which upregulate progesterone receptor expression in kisspeptin neurons and stimulate astrocytes to synthesize neuroprogesterone [126,127]. Hypothalamus-derived estradiol (neuroestradiol) also contributes to GnRH regulation [128]. In the absence of these regulatory brakes, (due to anovulation, decreased estrogen, and impaired hypothalamic progesterone inhibition), a feedforward loop develops where elevated LH promotes further androgen excess, which exacerbates IR and perpetuates anovulation [129]. By impairing ovulation and interrupting the normal maturation to ovulatory cycling, IR may therefore exaggerate or prolong hyperandrogenism and anovulatory cycles. This may also be a factor that drives the onset of neuroendocrine dysfunction in adolescence and contributes to the onset of symptoms of PCOS [130,131].

In summary, the pathophysiology of PCOS involves a vicious cycle between hypothalamic-pituitary dysfunction and disrupted ovarian steroidogenesis. Increased GnRH pulse frequency leads to elevated LH and relatively low FSH, that results in excess ovarian androgen production. Decreased

ovarian aromatase function, and possibly accelerated degradation of estradiol in ovarian follicles, leads to lower estradiol levels and hyperandrogenism [132,133]. The lower estradiol and elevated androgens, coupled with elevated AMH, contributes to follicular arrest, anovulation, and polycystic ovarian morphology [117]. Anovulation perpetuates the neuroendocrine disruption by preventing the luteal-phase rise in estradiol and progesterone, and the negative feedback needed to regulate GnRH output.

As discussed above and in section 6.2, IR and hyperinsulinemia disrupt neuroendocrine regulation in many ways and are primary drivers of HPO dysfunction. As a result, lifestyle-based interventions are the first line of treatment recommended in the 2023 International Guidelines and in phase 1 of the insulin-centric model (see section 7.4) [12]. Second-line pharmacotherapeutic support may also be required to counteract the effects of IR and hormonal imbalances if lifestyle measures alone are insufficient (discussed in section 7.5). While the combined oral contraceptive pill (COCP) has long been used to manage PCOS symptoms, it suppresses ovulation and disrupts natural cyclical hormone production. In addition, the OCP, particularly those containing androgenic progestins, has a risk of side-effects such thromboembolism [134], and may worsen IR [135]. Emerging alternatives include cyclical human-identical estradiol (17B-estradiol patch or gel) and oral luteal-phase micronized progesterone [136–138]. Although some small studies report promising results, conflicting results have been reported [139] and large-scale randomized trials are required.

6.4. Bidirectional Relationship Between Insulin Resistance and Hyperandrogenism

Androgens have a physiological role in developmental programming [140], adipose tissue differentiation [141], body fat distribution [142], skeletal muscle growth [143], hypothalamic control of food intake and energy balance [144], energy metabolism [145], and insulin signalling in insulintargeted tissues [146]. Increased androgen levels in women with PCOS can result in pathological effects in any of these tissues or physiological systems. Excess androgens promote beta-cell dysfunction and impaired insulin secretion, disruption of insulin signaling in muscle and adipose tissue, and increase visceral adipose tissue [147].

There is clear evidence that elevated androgens can exacerbate IR, and both hyperinsulinaemia and IR can lead to elevated androgens [111,148]. Nevertheless, mildly elevated androgen levels may have a range of adaptive survival functions such as increased physical strength and decreased reproductive function in times of environmental stress [1,4,86]. In addition, the augmentation of IR by elevation of androgens would also provide other survival advantages, such as the redirection of energy to tissues of need [149]. The bidirectional relationship between insulin signalling pathways and androgens may represent a self-reinforcing feedback loop that links metabolic health to optimal reproductive performance.

Although reducing androgen levels would be expected to improve insulin sensitivity, conflicting results have been reported. Several small clinical trials, including studies using GnRH analogues and androgen receptor antagonists such as flutamide, have reported inconsistent effects on insulin sensitivity, with many showing no improvement despite reduced androgen levels [150–157]. Surgical interventions such as ovarian drilling and oophorectomy similarly yielded mixed results [158,159]. These findings are sometimes interpreted to mean that androgens are not the primary cause of IR. However, the interpretation of these conflicting reports is limited by inclusion of small numbers of patients, use of indirect methods of assessing insulin sensitivity, and short duration of treatment in some studies. In addition, the interpretation of the GnRH analogue studies should consider potential compensatory metabolic changes resulting from the estradiol deficiency and neuroendocrine disruption induced by these medications.

Further evidence that hyperinsulinemia and IR are primary causes of hyperandrogenism comes from studies investigating inherited severe insulin resistance syndromes (Rabson-Mendenhall syndrome, type A, B and C insulin resistance syndromes, and lipodystrophies) [160]. Ovarian hyperandrogenism is present in many of these individuals, and is secondary to IR. A prospective study of 31 women with PCOS randomized to 3 treatment groups (flutamide, metformin, or

flutamide/metformin) for 9 months, found that combined treatment with flutamide/metformin resulted in greater improvements in lipid profiles, androgen levels and IR, than with monotherapy alone [161]. Finally, a meta-analysis of 24 randomized trials reported a beneficial impact of using the insulin-sensitizing medication metformin, to reduce both metabolic parameters and serum androgen levels [162].

In summary, although elevated androgen levels clearly exacerbate IR, intervention trials where androgen levels are reduced do not consistently improve IR. This is likely due to the fact that IR has a number of underlying causes such as poor quality diet, gastrointestinal dysbiosis, chronic low-grade inflammation, stress, and circadian disruption, in addition to elevated androgens [2]. On the other hand, improving insulin sensitivity can significantly improve androgen levels and PCOS-related symptoms [162]. Evidence suggests that both IR and chronic inflammation are primary drivers of hyperandrogenism via specific molecular effects in the hypothalamus, ovaries, liver, and metabolic pathways [101,102]. IR and hyperandrogenism reinforce each other, perpetuating both metabolic and reproductive dysfunction. Understanding this bidirectional relationship is key to devising a combined therapeutic strategy to reduce both IR and elevated androgens, in order to provide the most beneficial treatment approach.

6.5. Adverse Effects of Insulin Resistance on the Endometrium, Placenta and Associated Pregnancy Complications

The key pathophysiological features of PCOS (chronic inflammation, IR, and hyperandrogenism), have all been individually associated with adverse endometrial and decidual changes that contribute to altered placental development and function [13,163–169]. The resulting dysfunctional cellular network at the maternal-fetal interface has an adverse impact on bidirectional communication between maternal decidual cells and fetal trophoblast cells involved in formation of the placenta [17,170–172]. As a consequence, women with PCOS are at significantly increased risk of miscarriage and implantation failure and have reduced success rates following assisted fertility treatments [173,174]. In addition, women with PCOS have an increased risk of all of the "great obstetrical syndromes" (spontaneous preterm labour, fetal growth restriction, stillbirth, and preeclampsia), which share common pathophysiological processes and placental abnormalities [175–177]. In addition, women with PCOS have a significantly increased incidence of GDM, related to their underlying IR [178].

A range of possible mechanisms have been proposed to explain the effect of preexisting hyperinsulinaemia and IR on placental development as a result of in-vitro, animal, and human studies. Insulin can inhibit aromatase activity in human trophoblasts [179], which may connect hyperinsulinaemia to excess placental androgens. The majority of reported studies show an adverse impact of maternal hyperandrogenism on pregnancy complications [17,169,180]. Elevated insulin was shown to cause DNA damage, apoptosis and reduced cell survival in trophoblasts in-vitro [165], which could contribute to impaired trophoblast migration and spiral artery remodeling. Pretreatment of trophoblasts with metformin prevented insulin's deleterious effects in a mouse model [181]. IR has been associated with altered transcriptome signatures in pathways that may affect placental development [166]. Interestingly, altered endometrial transcriptome signatures isolated from a variety of endometrial cells were reversed following 16 weeks treatment with metformin and lifestyle, in obese women with PCOS, hyperandrogenism and IR [182]. Placental trophoblasts from obese women are significantly less sensitive to insulin than non-obese women, and obese women have greater placental lipid accumulation (fatty placenta), similar to IR-related maternal lipotoxicity [183,184]. This may contribute to placental inflammation and metabolic and nutrient transport abnormalities.

Adherence to a healthy lifestyle (higher diet quality, regular exercise, maintaining normal weight, non-smoking, and avoidance of alcohol) has been found to reduce the risk of pregnancy complications in women with PCOS [185,186], and is cost-effective [187,188]. Accumulating clinical and molecular evidence therefore suggests that IR and hyperinsulinaemia contribute to abnormal

placental development and pregnancy complications that can be prevented or minimized with preconception and antenatal lifestyle and medical interventions.

7. Introduction of an Evidenced-Based Insulin-Centric Model of Insulin Resistance in PCOS

7.1. Rationale for Changing from a Glucose-Centric View of Glycaemic Disturbance to an Insulin-Centric Model

The limitations of the glucose-centric approach for early diagnosis and prevention of complications of T2DM have been recognized for decades [189–191]. Historically, current testing and management protocols facilitated detection of hyperglycemia and interventions designed to reduce premature morbidity and mortality. However, these interventions tend to occur late in the disease process after 30-50% of individuals diagnosed with T2DM have already developed complications [20]. As a result, there is now a growing consensus that a new approach is necessary for early detection of metabolic precursors to overt T2DM and related complications, such as beta cell failure, IGT and IR [192]. These approaches encompass novel diabetes classification systems [193], diagnosis of diabetes using machine learning (ML)-based predictive models [194], optimized diabetes detection ML models utilizing feature engineering and ensemble learning [195], and interactive network models [196]. They also include a dysglycemia-based framework for managing multi-morbidity [197] a comorbidity-centric algorithm [198], and strategies that target the "ominous octet" to prevent damage across multiple organs [199,200]. In addition, there are diverse models centered on different aspects of the disease, that include glucagonocentric [201], cardiorenal-metabolic [202], TOR-centric [203], beta cell-centric [204], adipocentric [205], or insulin-centric approaches [206].

Many women show no symptoms of hyperglycemia and are diagnosed with T2DM after presenting with other medical problems or during a routine check-up [207]. Adolescents and women with PCOS, however, represent an ideal group for early detection and intervention. They usually present at a young age with apparently non-metabolic symptoms, such as menstrual disturbance, acne, hirsutism, or infertility, and have a high likelihood of underlying metabolic disturbance and IR [2,7]. As a result, we advocate for incorporating an insulin-centric approach in the assessment and management of PCOS (Figure 3). This method builds on the well-established role of hyperinsulinemia and IR in metabolic dysfunction and hyperandrogenism, thereby enabling earlier interventions to prevent progressive morbidity and premature mortality [208]. One advantage of introducing an insulin-centric approach is that available methods for assessing hyperinsulinemia and surrogate markers of IR could easily be incorporated into current management protocols.

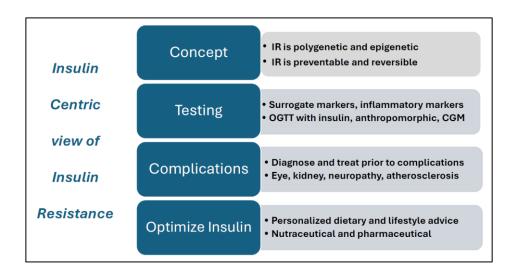


Figure 3. Features of an insulin-centric model of insulin resistance. Abbreviations: IR = Insulin Resistance; OGTT = Oral Glucose Tolerance Test; CGM = Continuous Glucose Monitoring.

7.2. Reasons for Delayed Introduction of an Insulin-Centric Paradigm

Despite widespread acknowledgment of the shortcomings of the glucose-centric paradigm and the possible clinical benefits of an insulin-centric approach, significant barriers hinder changing current practices. These challenges include the inherent physiological complexity and incomplete understanding of insulin biology and IR, difficulties with measurement, standardization, and test validation, gaps in the evidence that contribute to outcome uncertainty, and a persistent clinical inertia that resists altering long-established paradigms [209].

We assert that the current evidence robustly supports shifting to an insulin-centric model. Although our understanding of insulin dynamics and supporting data may have limitations, the overwhelming clinical need for change outweighs these gaps. It is important to note that current glycemic tests were not developed through exhaustive, long-term intervention trials, yet their timely adoption has led to significant improvements in patient care. Nevertheless, it is now widely appreciated that the glucose-centric approach has significant limitations (189–191,193–205). Clinicians accustomed to relying on "clinical judgement" need to embrace "evidence judgement" when there is a significant unmet need, despite an imperfect evidence base.

7.3. Testing for Insulin Resistance – Measurement Challenges and Standardization

7.3.1. Hyperglycaemic-Euglycaemic Clamp Test

The hyperglycaemic-euglycaemic clamp (clamp) test is the gold standard procedure for measuring IR [210]. The clamp test involves simultaneous infusions of high-dose intravenous insulin and glucose in order to suppress hepatic gluconeogenesis and create a steady state blood glucose level (5.5 mmol/L) [211]. Insulin sensitivity is measured over a normal biological range and IR is diagnosed when the whole-body glucose disposal rate reaches an arbitrary cut-off of 4.9 mg/min/kg [81] (or 46.0 +/- 16.9 micromol/min/kg lean body mass) [212].

The clamp test is only suitable for experimental studies in research centres, due to the complexity, associated risks, expense, and time required to conduct the procedure [211]. The clamp test has considerable intra-laboratory variability due to different methods of measuring glucose and insulin and is operator dependent. In addition, the clamp test underestimates glucose disposal in very IR participants due to incomplete suppression of hepatic gluconeogenesis [211]. When taken together, it is clear that the clamp test has many limitations, even if it were suitable for use in clinical settings. Despite these limitations, the clamp test is used as a gold standard to compare the accuracy of surrogate marker tests.

7.3.2. Surrogate Biomedical Markers of Insulin Resistance and Hyperinsulinemia

Fasting insulin levels are useful to provide preliminary information about insulin dynamics but may differ widely within individuals, can have variable performance characteristics at low concentrations (<12 pmol/L), may be impaired by the presence of insulin antibodies, and assay standardization has not been determined internationally [213,214]. Nevertheless, consistent high levels of fasting insulin reflect underlying IR.

Insulin assays are more accurate predictors of IR when combined with other markers [214]. A wide range of surrogate marker indices have been compared to the clamp test to determine accuracy, precision, cutoff values, and reproducibility. It is not the purpose of this review to describe these in detail and we refer the reader to current comprehensive reviews [24,214,215]. Examples include Homeostatic Model Assessment-IR (HOMA-IR), HOMA-Triglyceride index (HOMA-TG), Quantitative Insulin Sensitivity Check index (QUICKI), TG/High Density Lipoprotein (TG/HDL) index, TG/Glucose (TG/G) index, alanine aminotransferase/aspartate aminotransferase (ALT/AST) ratio, and Adipose-IR index. Many of these indices have been shown to be valuable predictors of IR. The most commonly used and tested index is the HOMA-IR, which could be coupled with circulating inflammatory mediators known to be associated with IR, adipose-tissue derived biomarkers, or other emerging novel molecules, to improve diagnostic accuracy and predictive ability [24,216].

Insulin resistance is a global health problem that is associated with significant clinical symptoms and is an early predictive indicator of future chronic disease in women with PCOS [24]. There is an urgent worldwide need for the development of accurate measurement, risk prediction, and evaluation of how surrogate indices change in intervention trials. In order to facilitate a rapid transition from a glucose-centric approach to an insulin-centric paradigm, an international collaborative effort is required to determine a consensus approach to surrogate marker testing, based on the best available evidence. The current evidence base far exceeds that used during the historical introduction of glucose-based testing (discussed in sections 3.1 and 3.2). Clinicians should be informed about the limitations of using surrogate marker indices and learn to interpret the results in the context of other tests, such as dynamic glucose-insulin testing, continuous glucose monitoring, anthropomorphic data, inflammatory cytokines, and other biomarkers. These assessment tools should be used in collaboration with individual clinical history and examination, as is usually done during comprehensive personalized medical assessment. Future research should focus on investigating the correlation between surrogate indices, biomarkers and clinical outcomes.

7.3.3. Dynamic Glucose-Insulin Testing

Dynamic glucose-insulin testing is a modified OGTT where both glucose and insulin are measured at specified intervals (eg. 0, 30, 60, 90, and 120 minutes) [217]. Measurement of insulin at the same time as glucose facilitates a dynamic assessment of the magnitude of insulin response following a standardized glucose load (eg. 75-100 grams) [218]. This timing allows for assessment of the early hyperinsulinemic response pattern, highlighting abnormalities in insulin secretion that static tests might miss [219,220].

In women with early IR, the pancreas may overcompensate by secreting increased insulin at the 30-minute point of the OGTT. This hyperinsulinemic response helps compensate for reduced insulin sensitivity in muscle, fat, and liver tissues, keeping blood glucose levels normal even when the metabolic system is under stress. Dynamic glucose-insulin testing therefore allows a concurrent assessment of insulin secretion with insulin sensitivity and can provide a window into the early pathological changes that precede overt dysglycemia [218,221].

7.3.4. Anthropomorphic Data

Anthropomorphic data aim to assess body composition and fat distribution as a surrogate indicator of metabolic and endocrine disturbance and future risk prediction in PCOS [222]. Commonly used metrics include BMI, waist circumference, waist-hip ratio, waist-height ratio, visceral adiposity index (VAI), and more detailed fat mass assessments with Dual Xray Absorptiometry (DEXA) scans and Magnetic Resonance Imaging (MRI). When coupled with biomedical surrogate markers, anthropomorphic data can help construct a comprehensive profile of metabolic status and risk. Waist-to-height ratio is emerging as a better predictor of hyperandrogenism than BMI, in women with PCOS [223]. VAI is a gender-specific index, based on a anthropomorphic and metabolic parameters, and has been found to correlate with visceral adipose dysfunction and IR in women with PCOS [224].

Multidimensional analysis using anthropomorphic data, surrogate biomedical markers, and comprehensive metabolic assessments could further refine risk stratification and treatment monitoring, especially in response to lifestyle interventions. Prospective interventional studies are required to assess the value of combined indices in clinical management and future chronic disease risk prediction.

7.3.5. Concurrent Testing for Inflammatory Markers

Large systematic reviews confirm the important role of chronic low-grade systemic inflammation as a key pathophysiological process that acts together with IR in the pathogenesis of PCOS [225–227]. The dysbiosis of gut microbiota theory of the pathogenesis of PCOS proposed that poor-quality diet results in increased release of lipopolysaccharide from gram negative bacteria, which traverses the gastrointestinal barrier and activates toll-like receptors on submucosal macrophages. This in turn activates the NF-kB signaling pathway and increases inflammatory cytokine production and secretion [227,228]. It is now recognized that systemic inflammation can be initiated at any mucosal surface [229], in response to microparticulate air pollution [230], microplastics [231], micro-organisms, other environmental antigens, and endogenous factors such as stress [232].

The bidirectional relationship and co-existence of IR and chronic inflammation (discussed in section 6.1) supports the measurement of inflammatory mediators as an aid to the assessment and management of IR in women with PCOS. The most commonly measured markers are C-reactive protein (CRP) and white blood count (WBC) [233]. Other markers of inflammation, such as interleukins, tumour necrosis factor, and homocysteine, are consistently elevated in PCOS, but are not routinely measured in clinical practice [234]. Large databases of the inflammatory proteome now exist (Olink multiplex inflammation panels and Immunology multiplex assay HCYTA 60K-PX48), and have been investigated in women with PCOS [235,236]. Measurement of inflammatory markers could provide a useful adjunct to the assessment of IR and warrant further investigation in clinical trials.

7.4. Targeted, Phase-Based Therapeutic Interventions; Phase 1-Lifestyle

It is important to educate patients about how IR and hyperinsulinemia impact both reproductive and metabolic health. Individual use of tools and training to self-monitor and adjust daily practices according to the feedback from their personalized assessments, will result in empowerment and better autonomy [237]. Personalized dietary advice focused on low-glycemic foods that reduce post-prandial glucose levels will help facilitate better symptom control, weight management, and reduce future risk [238].

Personalized exercise prescriptions tailored to the abilities and interests of the individual will reduce visceral adiposity and increase compliance and quality of life [239]. Behavioural and stress management strategies will reduce cortisol levels and improve hyperinsulinemia and IR [240]. Exercise and stress reduction, coupled with attention to sleep hygiene and circadian re-alignment, improve energy and motivation [241]. Increased emphasis on strengthening personal and community support systems may help reduce anxiety, depression and loneliness [242].

7.5. Targeted, Phase-Based Therapeutic Interventions; Phase 2-Pharmacotherapeutic

A wide range of nutraceutical treatments have been studied for their ability to reduce IR and improve metabolic health [243]. These include resveratrol [244], N-acetyl cysteine [245], berberine [246], curcumin [247], magnesium [248], and inositol [12]. So far, only inositol has been recognized as a treatment option in the International Guidelines, but there is clearly a need for greater inclusion of many of these therapies in future clinical trials.

There is also a growing list of possible pharmaceutical treatment options for reducing IR that require more intensive investigation in women with PCOS. These include metformin, thiazolidinediones, GLP-1 agonists, GIP and GLP-1 dual agonists, and SGLT-2 inhibitors [249]. It is likely that evidence from research in non-PCOS women will initially be translated to women with PCOS, but PCOS-specific trials are needed.

7.6. Targeted, Phase-Based Therapeutic Interventions; Phase 3-Monitoring and Support

Digital health tools, wearable technology, and continuous glucose monitoring can provide real-time insight and be integrated into patient management dashboards [250]. These technologies are readily available and widely used and need to be critically evaluated in clinical trials. Dynamic retesting and data driven lifestyle and pharmacotherapeutic adjustments should contribute to better patient education, compliance and empowerment. Integrated metabolic reprofiling of glucose levels, lipid levels, inflammatory markers, and IR, coupled with diet and lifestyle reassessment, will contribute to the prevention of the health consequences of persistent IR [251].

7.7. Research and Continuous Improvement

The rapid introduction of wearable, digital, personalized monitoring technology is well ahead of rigorous clinical trial evaluation for safety, efficacy, and future disease prediction. These devices are promising tools for gathering large amounts of health data and can use ML to gain valuable insights that assist with individualized healthcare solutions [252]. Large-scale data collection using ML analytic capability should facilitate timely refinement of the insulin-centric model. Interdisciplinary collaboration and longitudinal studies will be needed to ensure the model evolves in line with new scientific insights.

7.8. Active Surveillance of Future Pipeline Assessment Tools and Therapeutic Candidates

The establishment of cooperative interdisciplinary networks will be necessary to keep pace with the rapid nature of scientific progress in technically challenging medical and supportive disciplines. These include microbiome assessment [253], identification of microbiome-related metabolic signatures [254], genetic screening for high-risk single nucleotide polymorphisms (INSR, IRS-1/2, AR, CAPN2) [255], epigenetic monitoring of methylation changes during treatment [256], and integrated multiomics assessments [257]. In addition, adaptive management platforms, that continuously update and consolidate information from multiple sources, will facilitate tailored adjustment to treatment strategies (mobile apps, online dashboards, or integrated clinical software) [258], and ML and AI related tools [259]. There is an increasing need to assess the optimal balance between the use of digital technologies and human involvement in medical care [260].

8. Artificial Intelligence-Generated Insulin-Centric Model for the Assessment and Management of PCOS

Below is a comprehensive proposal by Generative Artificial Intelligence tool "Microsoft Copilot" for an insulin-centric model that rethinks the way we assess and manage PCOS. This approach builds on the well-established role of IR and hyperinsulinemia in driving androgen excess and metabolic and hormonal dysfunction in PCOS. The model—called the Insulin-Centric PCOS Analysis and Management Model (IC-PAMM)—frames insulin as both a diagnostic hallmark and a therapeutic target (Figure 4).

1. Enhanced Diagnostic and Stratification Process

A. Comprehensive Metabolic Profiling

- Baseline Hormonal and Metabolic Laboratory Workup: Every patient undergoes standard
 endocrine evaluations (e.g., LH, FSH, testosterone and SHBG) along with robust metabolic
 panels. However, this model significantly emphasizes fasting insulin levels, fasting glucose,
 and HbA1c. Key parameters might include:
- Dynamic Glucose-Insulin Testing: Implement a modified oral glucose tolerance test (OGTT) where both glucose and insulin are measured at intervals —0, 30, 60, 90, and 120 minutes. This timing allows for understanding the early hyperinsulinemic response pattern, highlighting abnormalities in insulin secretion that static tests might miss.

B. Integration of Novel Biomarkers and Anthropometrics

- Metabolic Inflammatory Markers & Adipokines: Include assessments of adiponectin, resistin, and perhaps inflammatory markers such as C-reactive protein (CRP) to gauge the link between adiposity and insulin resistance.
- Anthropometric Data: Waist circumference, BMI, and body fat distribution provide additional layers for risk stratification.
- Genetic/Epigenetic Screening: Although still emerging, screening for genetic variants
 associated with insulin signaling (e.g., polymorphisms in insulin receptor substrates) could
 help tailor interventions further.

2. Targeted, Phase-Based Therapeutic Interventions

A. Lifestyle Modification as the Cornerstone

- Dietary Interventions: Focus on low glycemic index diets that minimize postprandial insulin spikes. Structured meal planning might incorporate intermittent fasting strategies or timerestricted feeding to enhance insulin sensitivity.
- Exercise Prescriptions: Encourage a mix of high-intensity interval training (HIIT) and
 resistance training, both known to improve insulin sensitivity and reduce visceral adiposity.
- Behavioral and Stress Management: Since chronic stress worsens hyperinsulinemia, integrating mindfulness practices, yoga, or cognitive behavioral therapy can help reduce cortisol levels, which in turn supports better insulin regulation.

B. Pharmacotherapeutic Strategies

Insulin Sensitizers: Medications such as metformin should be front-line therapy—especially
in those with clear hyperinsulinemia. In select cases, thiazolidinediones (e.g., pioglitazone) or
GLP-1 receptor agonists might be employed.

Personalized Medicine Approach: For patients identified — via genetic or biochemical
markers — as having a particularly aggressive insulin-resistance profile, consider early and
combined pharmacological intervention. Novel drugs targeting intracellular insulin signaling
pathways or agents that modulate adipokine profiles could also be explored in clinical trials.

C. Digital Health Tools and Continuous Monitoring

- Wearable Technology & Mobile Apps: Integrate continuous (or frequent periodic) glucose
 monitoring systems and activity trackers that feed data into a management dashboard. This
 real-time insight can help both clinicians and patients adjust strategies promptly.
- Feedback Loops & Adaptive Management: Set up a system of regular re-assessments (every 3–6 months) with repeat dynamic tests and metabolic panels. These data-driven checkpoints allow for timely adjustments in lifestyle guidance or medication plans.

3. Holistic and Iterative Care

A. Integration Across Disciplines

- Coordinated Care Teams: PCOS management under this model isn't limited to
 endocrinologists. It involves nutritionists, exercise physiologists, mental health professionals,
 and even genetic counselors to create an all-around management plan.
- Patient Education and Empowerment: Educate patients about how insulin resistance and
 hyperinsulinemia impact both reproductive and metabolic health. Empower them with tools
 and training to self-monitor and adjust daily practices according to the feedback from their
 personalized management dashboard.

B. Research and Continuous Improvement

- Data Collection and Model Refinement: As more patients are managed using the IC-PAMM, the model can be perpetually refined. Large-scale data collection—integrated with machine learning analytics—could identify novel sub-phenotypes and predictive factors, allowing personalized risk scores and treatment algorithms to emerge.
- Interdisciplinary Collaboration: Encourage regular feedback and collaborative studies between research institutions and clinical centers to validate and improve the model, ensuring it evolves with emergent scientific insights.

Figure 4. AI generated future insulin-centric model for the assessment and management of polycystic ovary syndrome (PCOS): Insulin-Centric PCOS Analysis and Management Model (IC-PAMM).

9. Discussion

We have combined a comprehensive review of the literature on historical aspects of the glucose-centric approach to the diagnosis of PCOS with an up-to-date discussion of insulin physiology, the adaptive significance of IR, and the role of IR and hyperinsulinemia in PCOS, with a state-of-the-art AI generated insulin-centric model for the assessment and management of PCOS (IC-PAMM). We provide evidence supporting growing international interest in using currently available surrogate biomarkers, combined with other clinical assessment tools that focus on identification of IR, to facilitate early intervention and prevention of progressive metabolic disease.

Insulin is a central regulatory hormone in human physiology and there are believed to be insulin receptors on all human cells. Nevertheless, insulin exerts different actions in specific tissues and cells, which are reflected in the variety of physiological and pathological effects of IR and

hyperinsulinemia. Hyperinsulinemia and IR are dynamically intertwined and always co-exist [251]. Hyperinsulinemia can cause IR and IR can cause hyperinsulinemia. Reduced insulin sensitivity is a physiological adaptive survival mechanism that can become maladaptive in a modern environment [8]. An insulin-centric model recognizes altered insulin biology as not just a marker, but a primary driver of PCOS and its associated reproductive dysfunction and metabolic disease.

Most current therapies and guidelines target glycemic control, weight reduction, or broader metabolic improvements rather than focusing on reducing circulating insulin directly. An insulincentric approach shifts the focus to reducing one of the primary upstream drivers of metabolic and endocrine dysfunction, rather than on downstream consequences. Changing a long-standing paradigm, even when it is recognized to have significant limitations, is always difficult and associated with significant barriers to change. While the evidence supporting an insulin-focused approach is complex and still evolving, it is nonetheless more substantial than the evidence that underpinned the adoption of the glucose-centric model. More importantly, there is a significant clinical need for a shift in focus from glucose to insulin to improve clinical management and quality of life and prevent premature morbidity and mortality.

The introduction of an insulin-centric model requires a coordinated international effort, as we have seen with the development of the International Guidelines for PCOS, to develop protocols based on existing evidence. Multidisciplinary collaborative research efforts will be required for data collection and model refinement to improve diagnostic strategies and develop shared therapeutic interventions that are integrated with digital health platforms and delivered in a holistic and personalized way.

10. Strengths and Limitations of the Current Review

10.1. Strengths

The current review provides a detailed discussion of the historical introduction of the longstanding glucose-based approach to the assessment and management of glycemic dysfunction in women with PCOS. The advantages and limitations of this model are explored and discussed in detail. The authors provide evidence and rationale that further supports the existing international momentum for a paradigm shift to a focus on insulin pathophysiology in PCOS. The report covers a broad range of complex topics that need to be examined when considering a change from a well-established traditional approach to an emerging new paradigm.

10.2. Limitations

There are many factors that limit the widespread acceptance of an insulin-centric approach to PCOS. PCOS is a heterogeneous syndrome that involves not only IR, but also hormonal imbalances, such as hyperandrogenism and decreased estrogen and progesterone, inflammatory processes, disturbance of the microbiome, hypothalamic and ovarian alterations, and reproductive and psychological dysfunction. The pathophysiology of PCOS involves disturbance of a complex network of inter-related adaptive physiological systems, and there may be multiple initiating factors besides IR, that drive symptoms and disease progression. This model is predicated on the hypothesis that IR and hyperinsulinemia are fundamental primary drivers of metabolic and reproductive dysfunction in women with PCOS.

11. Conclusions

PCOS is just the visible tip of a much larger global health crisis of metabolic-associated chronic disease. Early diagnosis in adolescents and young women allows for an in-depth assessment and management of the metabolic, hormonal, and psychological challenges that not only trigger symptoms, but drive future health risks. By integrating comprehensive metabolic evaluations, dynamic insulin testing, and targeted lifestyle and medical interventions, this approach provides a versatile yet robust framework for tailoring PCOS treatment to the individual. IC-PAMM is a model



that goes beyond simply correcting the biochemical imbalances – such as IR, chronic inflammation, and hyperandrogenism-to also enhance overall quality of life through holistic care. This multifaceted framework reflects an evolving understanding of PCOS that emphasizes early intervention and personalized treatment strategies, ensuring that care is both proactive and integrative.

Author Contributions: Conceptualization, J.P., L.B., and F.L.G; methodology, J.P., L.B., and F.L.G; writing original draft preparation, J.P. and L.B.; writing - review and editing, J.P. and L.B.; review and editing F.L.G. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Acknowledgments: During the preparation of this manuscript the authors used Generative Artificial Intelligence tool Microsoft Copilot for the purposes of designing a framework for an insulin-centric model for the assessment and management of PCOS (section 8). The authors have amended and edited the output and take full responsibility for the content of this publication.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviations

AGE Advanced Glycation End Products

ALT Alanine Aminotransferase

AMH Antimullerian Hormone

AR Androgen Receptor

AST Aspartate Aminotransferase

BMI **Body Mass Index** BSL **Blood Sugar Level**

CAPN2 Caplain-2 Catalytic Subunit **CGM** Continuous Glucose Monitor Central Nervous System **CNS**

CRP C-reactive Protein

CYP17A1 Cytochrome P450, family 17, subfamily A1

DEXA Dual Xray Absorptiometry

DNA Deoxyribose Nucleic Acid FSH Follicle Stimulating Hormone **GDM** Gestational Diabetes Mellitis

GIP Glucose-Dependent Insulinotropic Peptide

GLP-1 Glucagon-Like Peptide-1 GLUT4Glucose Transporter Type 4

GnRH Gonadotropin Releasing Hormone

HbA1C Hemoglobin A1C

HDL High Density Lipoprotein HIIT High-Intensity Interval Training HOMA-IR Homeostatic Model assessment-IR

IC-PAMM Insulin-Centric PCOS Analysis and Management Model

IGF-1 Insulin-like Growth Factor-1 Impaired Glucose Tolerance **IGT IRS** Insulin Receptor Substrate

INSR Insulin Receptor
IR Insulin Resistance
LH Luteinizing Hormone
ML Machine Learning

MAPK Mitogen-Activated Protein Kinase MCP-1 Monocyte Chemoattractant Protein-1 MRI Magnetic Resonance Imaging

NF-кВ Nuclear Factor Kappa B

NLR Nucleotide-Binding Domain, Leucine-Rich Repeat Containing

NLRP3NLR Family Pyrin Domain Containing 3

OGTT Oral Glucose Tolerance Test PCOS Polycystic Ovary Syndrome

PI-3K Phosphotidylinositol-3 Kinase

pmol/LPicomole per Litre

QUICKI Quantitative Insulin Sensitivity Check Index

ROS Reactive Oxygen Species RNA Ribose Nucleic Acid

SGLT-2 Sodium glucose cotransporter-2

SHBG Sex Hormone Binding Globulin SNP Single nuclear peptide T2DM Type 2 Diabetes Mellitis

TG Triglyceride

VAI Visceral Adiposity Index WBC White Blood Cell Count

References

- 1. Parker J, O'Brien C, Hawrelak J, Gersh FL. Polycystic Ovary Syndrome: An Evolutionary Adaptation to Lifestyle and the Environment. Int J Environ Res Public Health. 2022;19[3]:1336.
- 2. Parker J. Pathophysiological Effects of Contemporary Lifestyle on Evolutionary-Conserved Survival Mechanisms in Polycystic Ovary Syndrome. Life. 2023;13[4]:1056.
- 3. Su P, Chen C, Sun Y. Physiopathology of polycystic ovary syndrome in endocrinology, metabolism and inflammation. J Ovarian Res. 2025;18[34]:1–10.
- 4. Dumesic DA, Padmanabhan V, Abbott DH. Polycystic ovary syndrome: an evolutionary metabolic adaptation. Reproduction. 2025;169(e250021).
- 5. Abbott DH, Dumesic DA, Franks S. Developmental origin of polycystic ovary syndrome A hypothesis. J Endocrinol. 2002;174[1]:1–5.
- 6. Parker J, O'Brien C, Gersh FL. Developmental origins and transgenerational inheritance of polycystic ovary syndrome. Aust New Zeal J Obstet Gynaecol. 2021;61[6]:1–5.
- 7. Cassar S, Misso ML, Hopkins WG, Shaw CS, Teede HJ, Stepto NK. Insulin resistance in polycystic ovary syndrome: A systematic review and meta-analysis of euglycaemic-hyperinsulinaemic clamp studies. Hum Reprod. 2016;31[11]:2619–31.
- 8. Tsatsoulis A, Mantzaris MD, Bellou S, Andrikoula M. Insulin resistance: An adaptive mechanism becomes maladaptive in the current environment An evolutionary perspective. Metabolism [Internet]. 2013;62[5]:622–33. Available from: http://dx.doi.org/10.1016/j.metabol.2012.11.004
- 9. Shaw LMA, Elton S. Polycystic ovary syndrome: A transgenerational evolutionary adaptation. BJOG An Int J Obstet Gynaecol. 2008;115[2]:144–8.
- 10. Azziz R, Dumesic DA, Goodarzi MO. Polycystic ovary syndrome: An ancient disorder? Fertil Steril [Internet]. 2011;95[5]:1544–8. Available from: http://dx.doi.org/10.1016/j.fertnstert.2010.09.032
- 11. Charifson MA, Trumble BC. Evolutionary origins of polycystic ovary syndrome: An environmental mismatch disorder. Evol Med Public Heal. 2019;[1]:50–63.



- 12. Teede HJ, Misso ML, Costello MF, Dokras A, Laven J, Moran L, et al. Recommendations from the international evidence-based guideline for the assessment and management of polycystic ovary syndrome. Eur J Endocrinol [Internet]. 2023;189:G43–64. Available from: https://doi.org/10.1093/ejendo/lvad096
- 13. Parker J, Hofstee P, Brennecke S. Prevention of Pregnancy Complications Using a Multimodal Lifestyle, Screening, and Medical Model. J Clin Med. 2024;13[15]:4344.
- 14. Hoeger KM, Dokras A, Piltonen T. Update on PCOS: Consequences, Challenges, and Guiding Treatment. J Clin Endocrinol Metab. 2021;106[3]:E1071–83.
- 15. Rodgers RJ, Avery JC, Moore VM, Davies MJ, Azziz R, Stener-Victorin E, et al. Complex diseases and comorbidities: Polycystic ovary syndrome and type 2 diabetes mellitus. Endocr Connect. 2019;8[3]:R71–5.
- 16. Du Y, Li F, Li S, Ding L, Liu M. Causal relationship between polycystic ovary syndrome and chronic kidney disease: A Mendelian randomization study. Front Endocrinol (Lausanne). 2023;14:1120119.
- 17. Parker J, O'Brien C, Yeoh C, Gersh FL, Brennecke S. Reducing the Risk of Pre-Eclampsia in Women with Polycystic Ovary Syndrome Using a Combination of Pregnancy Screening, Lifestyle, and Medical Management Strategies. J Clin Med. 2024;13[1774]:1–33.
- 18. Whicher CA, O'Neill S, Holt RIG. Diabetes in the UK: 2019. Diabet Med. 2020;37[2]:242-7.
- 19. Tabák AG, Jokela M, Akbaraly TN, Brunner EJ, Kivimäki M, Witte DR. Trajectories of glycaemia, insulin sensitivity, and insulin secretion before diagnosis of type 2 diabetes: an analysis from the Whitehall II study. Lancet. 2009;373[9682]:2215–21.
- 20. Bonora E, Trombetta M, Dauriz M, Travia D, Cacciatori V, Brangani C, et al. Chronic complications in patients with newly diagnosed type 2 diabetes: Prevalence and related metabolic and clinical features: The Verona Newly Diagnosed Type 2 Diabetes Study (VNDS) 9. BMJ Open Diabetes Res Care. 2020;8[1]:1–7.
- 21. International Diabetes Federation. More than two in three people with diabetes already have complications at diagnosis [Internet]. 2023. Available from: https://idf.org/news/more-than-two-in-three-people-with-diabetes-already-have-complications-at-diagnosis/
- 22. Amisi CA. Markers of insulin resistance in Polycystic ovary syndrome women: An update. World J Diabetes. 2022;13[3]:129–49.
- 23. Tabák, AG; Jokela, M; Akbaraly, TN; Brunner E, Kivimäki, M; Witte D. Trajectories of Glycemia, Insulin Sensitivity and Insulin Secretion Preceding the Diagnosis of Type 2 Diabetes: The Whitehall II Study. Lancet. 2009;373[9682]:2215–21.
- 24. Kosmas CE, Sourlas A, Oikonomakis K, Zoumi EA, Papadimitriou A, Kostara CE. Biomarkers of insulin sensitivity/resistance. J Int Med Res. 2024;52[10]:1–40.
- 25. Celik C, Tasdemir N, Abali R, Bastu E, Yilmaz M. Progression to impaired glucose tolerance or type 2 diabetes mellitus in polycystic ovary syndrome: A controlled follow-up study. Fertil Steril [Internet]. 2014;101[4]:1123-1128.e1. Available from: http://dx.doi.org/10.1016/j.fertnstert.2013.12.050
- 26. Bahri Khomami M, Joham AE, Boyle JA, Piltonen T, Silagy M, Arora C, et al. Increased maternal pregnancy complications in polycystic ovary syndrome appear to be independent of obesity—A systematic review, meta-analysis, and meta-regression. Obes Rev. 2019;20[5]:659–74.
- 27. Reyes-Muñoz E, Castellanos-Barroso G, Ramírez-Eugenio BY, Ortega-González C, Parra A, Castillo-Mora A, et al. The risk of gestational diabetes mellitus among Mexican women with a history of infertility and polycystic ovary syndrome. Fertil Steril. 2012;97[6]:1467–71.
- 28. Karamanou, Marianna. Protogerou, A. Tsoucalas, G. Androutsos G, Poulakou-Rebelakou E. Milestones in the history of diabetes mellitus: The main contributors. World J Diabetes. 2016;7[1]:1–7.
- 29. Tan SY, Merchant J. Frederick Banting [1891–1941]: Discoverer of insulin. Singapore Med J. 2017;58[1]:2–3.
- 30. Himsworth HP. Diabetes mellitus: Its differentiation into insulin-sensitive and insulin-insensitive types. Diabet Med. 2011;28[12]:1440–4.
- 31. Campbell MR, Shokrani M. Introduction, Background and Various Types. Am Soc Clin Lab Sci. 2016;29[2]:106–10.
- 32. Weijers R. The Evolution of Type 2 Diabetes Mellitus and Insulin Resistance. Endocrinol Diabetes Metab J. 2023;7[3]:1–9.
- 33. Stein ILM. Amenorrhoea associated with bilateral polycystic ovaries. Am J Obs Gynecol. 1935;29:181-91.
- 34. Diamanti-Kandarakis E, Dunaif A. Insulin resistance and the polycystic ovary syndrome revisited: An update on mechanisms and implications. Endocr Rev. 2012;33[6]:981–1030.



- 35. Dunaif A, Legro RS. Prevalence and Predictors of Risk for Type 2 Diabetes Mellitus and Impaired Glucose Tolerance in Polycystic Ovary Syndrome-Authors' Response. J Clin Endocrinol Metab. 1999;84[8]:297–2976.
- 36. Ehrmann DA, Liljenquist DR, Kasza K, Azziz R, Legro RS, Ghazzi MN, et al. Prevalence and predictors of the metabolic syndrome in women with polycystic ovary syndrome. J Clin Endocrinol Metab. 2006;91[1]:48–53.
- 37. Stener-Victorin E, Padmanabhan V, Walters KA, Campbell RE, Benrick A, Giacobini P, et al. Animal Models to Understand the Etiology and Pathophysiology of Polycystic Ovary Syndrome. Endocr Rev. 2020;41[4]:538–76.
- 38. He FF, Li YM. Role of gut microbiota in the development of insulin resistance and the mechanism underlying polycystic ovary syndrome: A review. J Ovarian Res. 2020;13[1]:1–13.
- 39. Moodley N, Ngxamngxa U, Turzyniecka MJ, Pillay TS. Historical perspectives in clinical pathology: A history of glucose measurement. J Clin Pathol. 2015;68[4]:258–64.
- 40. Klimek M, Knap J, Reda M, Masternak M. History of glucose monitoring: past, present, future. J Educ Heal Sport. 2019;9[9]:222–7.
- 41. Hirsch I. Introduction: History of Glucose Monitoring. ADA Clin Compend. 2018;1–1.
- 42. Raza SI, Raza SA, Kazmi M, Saad M, Hussain I. 100 Years of Glucose Monitoring in Diabetes Management. J Diabetes Mellit. 2021;11[05]:221–33.
- 43. Didyuk O, Econom N, Guardia A, Livingston K, Klueh U. Continuous Glucose Monitoring Devices: Past, Present, and Future Focus on the History and Evolution of Technological Innovation. J Diabetes Sci Technol. 2020;15[3]:676–83.
- 44. Nelson RL. Oral Glucose Tolerance Test: Indications and Limitations. Mayo Clin Proc [Internet]. 1988;63[3]:263–9. Available from: http://dx.doi.org/10.1016/S0025-6196[12]65100-3
- 45. Jagannathan R, Neves JS, Dorcely B, Chung ST, Tamura K, Rhee M, et al. The oral glucose tolerance test: 100 years later. Diabetes, Metab Syndr Obes. 2020;13:3787–805.
- 46. Duan D, Kengne AP, Echouffo-Tcheugui JB. Screening for Diabetes and Prediabetes. Endocrinol Metab Clin North Am. 2021;50[3]:369–85.
- 47. Zaharia OP, Strassburger K, Strom A, Bönhof GJ, Karusheva Y, Antoniou S, et al. Risk of diabetes-associated diseases in subgroups of patients with recent-onset diabetes: a 5-year follow-up study. Lancet Diabetes Endocrinol. 2019;7[9]:684–94.
- 48. Tomic D, Shaw JE, Magliano DJ. The burden and risks of emerging complications of diabetes mellitus. Nat Rev Endocrinol. 2022;18[9]:525–39.
- 49. Brand KMG, Gottwald-Hostalek U, Andag-Silva A. Update on the therapeutic role of metformin in the management of polycystic ovary syndrome: Effects on pathophysiologic process and fertility outcomes. Women's Heal. 2025;21:1–18.
- 50. Notaro ALG, Neto FTL. The use of metformin in women with polycystic ovary syndrome: an updated review. J Assist Reprod Genet [Internet]. 2022;39[3]:573–9. Available from: https://doi.org/10.1007/s10815-022-02429-9
- 51. Livadas S, Anagnostis P, Bosdou JK, Bantouna D, Paparodis R. Polycystic ovary syndrome and type 2 diabetes mellitus: A state-of-the-art review. World J Diabetes. 2022;13[1]:5–26.
- 52. Gruss, SM. Nhim, K. Gregg, E. Bell, M. Luman, E. Albright A. Public Health Approaches to Type 2 Diabetes Prevention: The US National Diabetes Prevention Program and Beyond. Curr Diab Rep. 2019;19[11]:78.
- 53. Vollmer J, Lacy ME, Christian WJ. Diabetes screening among women with Polycystic Ovary Syndrome: A descriptive study of commercial claims, 2011-2019. Res Sq. 2024;24:194.
- 54. Yang J, Qian F, Chavarro JE, Ley SH, Tobias DK, Yeung E, et al. Modifiable risk factors and long term risk of type 2 diabetes among individuals with a history of gestational diabetes mellitus: prospective cohort study. BMJ. 2022;Sept 21(378:e070312):1–11.
- 55. Haeusler RA, McGraw TE, Accili D. Metabolic Signalling: Biochemical and cellular properties of insulin receptor signalling. Nat Rev Mol Cell Biol [Internet]. 2018;19[1]:31–44. Available from: http://dx.doi.org/10.1038/nrm.2017.89
- 56. Petersen MC, Shulman GI. Mechanisms of insulin action and insulin resistance. Physiol Rev. 2018;98[4]:2133–223.
- 57. Unger RH, Orci L. Paracrinology of islets and the paracrinopathy of diabetes. Proc Natl Acad Sci U S A. 2010;107[37]:16009–12.
- 58. Sun Q, Li J, Gao F. New insights into insulin: The anti-inflammatory effect and its clinical relevance. World J Diabetes. 2014;5[2]:89.



- 59. Aljada A, Ghanim H, Saadeh R, Dandona P. Insulin Inhibits NFκB and MCP-1 Expression in Human Aortic Endothelial Cells. J Clin Endocrinol Metab. 2001;86[1]:450–3.
- 60. Chang YW, Hung LC, Chen YC, Wang WH, Lin CY, Tzeng HH, et al. Insulin Reduces Inflammation by Regulating the Activation of the NLRP3 Inflammasome. Front Immunol. 2021;11(February):1–11.
- 61. Li J, Wu F, Zhang H, Fu F, Ji L, Dong L, et al. Insulin inhibits leukocyte-endothelium adherence via an Akt-NO-dependent mechanism in myocardial ischemia/reperfusion. J Mol Cell Cardiol [Internet]. 2009;47[4]:512–9. Available from: http://dx.doi.org/10.1016/j.yjmcc.2009.07.010
- 62. Tiwari S, Riazi S, Ecelbarger CA. Insulin's impact on renal sodium transport and blood pressure in health, obesity, and diabetes. Am J Physiol Ren Physiol. 2007;293[4]:974–84.
- 63. Steinberg HO, Brechtel G, Johnson A, Fineberg N, Baron AD. Insulin-mediated skeletal muscle vasodilation is nitric oxide dependent: A novel action of insulin to increase nitric oxide release. J Clin Invest. 1994;94[3]:1172–9.
- 64. Mandal AK, Leask MP, Estiverne C, Choi HK, Merriman TR, Mount DB. Genetic and Physiological Effects of Insulin on Human Urate Homeostasis. Front Physiol. 2021;12(August):1–17.
- 65. Rahman MS, Hossain KS, Das S, Kundu S, Adegoke EO, Rahman MA, et al. Role of insulin in health and disease: An update. Int J Mol Sci. 2021;22[12]:1–19.
- 66. Li M, Chi X, Wang Y, Setrerrahmane S, Xie W, Xu H. Trends in insulin resistance: insights into mechanisms and therapeutic strategy. Signal Transduct Target Ther. 2022;7[1]:1–25.
- 67. Nestler JE, Jakubowicz DJ, De Vargas AF, Brik C, Quintero N, Medina F. Insulin stimulates testosterone biosynthesis by human thecal cells from women with polycystic ovary syndrome by activating its own receptor and using inositolglycan mediators as the signal transduction system. J Clin Endocrinol Metab. 1998;83[6]:2001–5.
- 68. Stephen C. Woods, Elizabeth C. Lotter LDM, Jr & DP. Chronic intracerebroventricular infusion of insulin reduces food intake and body weight of baboons. Nature. 1979;282(November):503–5.
- 69. Obici S, Zhang BB, Karkanias G, Rossetti L. Hypothalamic insulin signaling is required for inhibition of glucose production. Nat Med. 2002;8[12]:1376–82.
- 70. Koch L, Wunderlich FT, Seibler J, Könner AC, Hampel B, Irlenbusch S, et al. Central insulin action regulates peripheral glucose and fat metabolism in mice. J Clin Invest. 2008;118[6]:2132–47.
- 71. Dupont J, Scaramuzzi RJ. Insulin signalling and glucose transport in the ovary and ovarian function during the ovarian cycle. Biochem J. 2016;473[11]:1483–501.
- 72. Li T, Mo H, Chen W, Li L, Xiao Y, Zhang J, et al. Role of the PI3K-Akt Signaling Pathway in the Pathogenesis of Polycystic Ovary Syndrome. Reprod Sci. 2017;24[5]:646–55.
- 73. Bedinger DH, Adams SH. Metabolic, anabolic, and mitogenic insulin responses: A tissue-specific perspective for insulin receptor activators. Mol Cell Endocrinol [Internet]. 2015;415:143–56. Available from: http://dx.doi.org/10.1016/j.mce.2015.08.013
- 74. Makhijani, P. Basso, PJ. Chan, YT. Chen, N. Baechle, J. Khan, S. Furman, D. Tsai, S. Winer D. Regulation of the immune system by the insulin receptor in health and disease. Front Endocrinol (Lausanne). 2023;(March):1–18.
- 75. Jacobse J, Li J, Rings EHHM, Samsom JN, Goettel JA. Intestinal Regulatory T Cells as Specialized Tissue-Restricted Immune Cells in Intestinal Immune Homeostasis and Disease. Front Immunol. 2021;12(August):1–23.
- 76. Martelli D. The inflammatory reflex reloaded. Brain Behav Immun [Internet]. 2022;104:137–8. Available from: https://doi.org/10.1016/j.bbi.2022.06.001
- 77. Pavlov VA, Wang H, Czura CJ, Friedman SG, Tracey KJ. The Cholinergic Anti-inflammatory Pathway: A Missing Link in Neuroimmunomodulation. Mol Med. 2003;9[5–8]:125–34.
- 78. Zhou MS, Wang A, Yu H. Link between insulin resistance and hypertension: What is the evidence from evolutionary biology? Diabetol Metab Syndr. 2014;6[1]:1–8.
- 79. Wensveen FM, Šestan M, Turk Wensveen T, Polić B. 'Beauty and the beast' in infection: How immune–endocrine interactions regulate systemic metabolism in the context of infection. Eur J Immunol. 2019;49[7]:982–95.
- 80. Wang P, Mariman ECM. Insulin resistance in an energy-centered perspective. Physiol Behav. 2008;94[2]:198–205.
- 81. Tam CS, Xie W, Johnson WD, Cefalu WT, Redman LM, Ravussin E. Defining insulin resistance from hyperinsulinemic-euglycemic clamps. Diabetes Care. 2012;35[7]:1605–10.
- 82. Park SY, Gautier JF, Chon S. Assessment of insulin secretion and insulin resistance in human. Diabetes Metab J. 2021;45[5]:641–54.

- 83. Tosi F, Bonora E, Moghetti P. Insulin resistance in a large cohort of women with polycystic ovary syndrome: A comparison between euglycaemic-hyperinsulinaemic clamp and surrogate indexes. Hum Reprod. 2017;32[12]:2515–21
- 84. Tsatsoulis, A. Mantzaris, MD. Sofia, B. Andrikoula M. Insulin resistance: An adaptive mechanism becomes maladaptive in the current environment An evolutionary perspective. Metabolism. 2012;62[5]:622–33.
- 85. Parker J, O'Brien C. Evolutionary and genetic antecedents to the pathogenesis of polycystic ovary syndrome (PCOS). J ACNEM. 2021;40[1]:12–20.
- 86. Dumesic DA, Padmanabhan V, Chazenbalk GD, Abbott DH. Polycystic ovary syndrome as a plausible evolutionary outcome of metabolic adaptation. Reprod Biol Endocrinol [Internet]. 2022;20[1]:1–12. Available from: https://doi.org/10.1186/s12958-021-00878-y
- 87. Gorjão R, Takahashi HK, Pan JA, Massao Hirabara S. Molecular mechanisms involved in inflammation and insulin resistance in chronic diseases and possible interventions. J Biomed Biotechnol. 2012;2012:2012–4.
- 88. Thomas DD, Corkey BE, Istfan NW, Apovian CM. Hyperinsulinemia: An early indicator of metabolic dysfunction. J Endocr Soc. 2019;3[9]:1727–47.
- 89. Zore T, Joshi N V., Lizneva D, Azziz R. Polycystic Ovarian Syndrome: Long-Term Health Consequences. Semin Reprod Med. 2017;35[3]:271–81.
- 90. Zhai Y, Pang Y. Systemic and ovarian inflammation in women with polycystic ovary syndrome. J Reprod Immunol [Internet]. 2022;151(February):103628. Available from: https://doi.org/10.1016/j.jri.2022.103628
- 91. Zhao H, Zhang J, Cheng X, Nie X, He B. Insulin resistance in polycystic ovary syndrome across various tissues: an updated review of pathogenesis, evaluation, and treatment. J Ovarian Res [Internet]. 2023;16[1]:1–17. Available from: https://doi.org/10.1186/s13048-022-01091-0
- 92. Okin D, Medzhitov R. Evolution of inflammatory diseases. Curr Biol [Internet]. 2012;22[17]:R733–40. Available from: http://dx.doi.org/10.1016/j.cub.2012.07.029
- 93. Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, et al. Chronic inflammation in the etiology of disease across the life span. Nat Med [Internet]. 2019;25[12]:1822–32. Available from: http://dx.doi.org/10.1038/s41591-019-0675-0
- 94. Kany S, Vollrath JT, Relja B. Cytokines in inflammatory disease. Int J Mol Sci. 2019;20[23]:1–31.
- 95. Buzas EI. The roles of extracellular vesicles in the immune system. Nat Rev. 2023;23(April):236-50.
- 96. Rehman, K. Akash M. Mechanisms Linking Inflammation to Insulin Resistance: How are they interlinked? J Biomed Sci. 2016;23[87]:1–18.
- 97. Leguisamo NM, Lehnen AM, Machado UF, Okamoto MM, Markoski MM, Pinto GH, et al. GLUT4 content decreases along with insulin resistance and high levels of inflammatory markers in rats with metabolic syndrome. Cardiovasc Diabetol [Internet]. 2012;11[1]:1. Available from: Cardiovascular Diabetology
- 98. Carlos R, Anderson E, Durstine JL, Carson JA. In fl ammation, physical activity, and chronic disease: An evolutionary perspective. Sport Med Heal Sci [Internet]. 2020;2[1]:1–6. Available from: https://doi.org/10.1016/j.smhs.2020.03.004
- 99. Gajewski M, Rzodkiewicz P, Maśliński S. The human body as an energetic hybrid? New perspectives for chronic disease treatment? Rheumatologia. 2017;55[2]:94–9.
- 100. Velez LM, Seldin M, Motta AB. Inflammation and reproductive function in women with polycystic ovary syndrome. Biol Reprod. 2021;104[6]:1205–17.
- 101. Unluhizarci K, Karaca Z, Kelestimur F. Role of insulin and insulin resistance in androgen excess disorders. World J Diabetes. 2024;12[5]:616–29.
- 102. Fox CW, Zhang L, Sohni A, Doblado M, Wilkinson MF, Chang RJ, et al. Inflammatory Stimuli Trigger Increased Androgen Production and Shifts in Gene Expression in Theca-Interstitial Cells. Endocrinology. 2019;160[12]:2946–58.
- 103. Straub RH. Insulin resistance, selfish brain, and selfish immune system: an evolutionarily positively selected program used in chronic inflammatory diseases. Arthritis Res Ther [Internet]. 2014;16(Suppl 2):S4. Available from: https://doi-org.ezproxy.uow.edu.au/10.1186/ar4688
- 104. Christ A, Lauterbach M, Latz E. Western Diet and the Immune System: An Inflammatory Connection. Immunity [Internet]. 2019;51[5]:794–811. Available from: https://doi.org/10.1016/j.immuni.2019.09.020

- 105. Giri B, Dey S, Das T, Sarkar M, Banerjee J. Chronic hyperglycemia mediated physiological alteration and metabolic distortion leads to organ dysfunction, infection, cancer progression and other pathophysiological consequences: An update on glucose toxicity. Biomed Pharmacother [Internet]. 2018;107(April):306–28. Available from: https://doi.org/10.1016/j.biopha.2018.07.157
- 106. Kathryn C. B. Tan; Sammy W. M. Shiu; Ying Wong; Xystus Tam. Serum advanced glycation end products (AGEs) are associated with insulin resistance. Diabetes Metab Res Rev. 2011;27:1488–92.
- 107. Palimeri S, Palioura E, Diamanti-Kandarakis E. Current perspectives on the health risks associated with the consumption of advanced glycation end products: Recommendations for dietary management. Diabetes, Metab Syndr Obes Targets Ther. 2015;8:415–26.
- 108. Baillargeon JP, Nestler JE. Commentary: Polycystic ovary syndrome: A syndrome of ovarian hypersensitivity to insulin? J Clin Endocrinol Metab. 2006;91[1]:22–4.
- 109. Soldani R, Cagnacci A, Yen SSC. Insulin insulin-like growth factor I (IGF-I) and IGF-II enhance basal and gonadotrophin-releasing hormone-stimulated luteinizing hormone release from rat anterior pituitary cells in vitro. Eur J Endocrinol. 1994;131[6]:641–5.
- 110. Sliwowska JH, Fergani C, Gawałek M, Skowronska B, Fichna P, Lehman MN. Insulin: Its role in the central control of reproduction. Physiol Behav. 2014;133:197–206.
- 111. Bremer AA, Miller WL. The serine phosphorylation hypothesis of polycystic ovary syndrome: a unifying mechanism for hyperandrogenemia and insulin resistance. Fertil Steril. 2008;89[5]:1039–48.
- 112. Nestler JE, Powers LP, Matt DW, Steingold KA, Plymate SR, Rittmaster RS, et al. A direct effect of hyperinsulinemia on serum sex hormone-binding globulin levels in obese women with the polycystic ovary syndrome. J Clin Endocrinol Metab. 1991;72[1]:83–9.
- 113. Diamanti-Kandarakis E, Piperi C, Kalofoutis A, Creatsas G. Increased levels of serum advanced glycation end-products in women with polycystic ovary syndrome. Clin Endocrinol (Oxf). 2005;62[1]:37–43.
- 114. Tatone C, Di Emidio G, Placidi M, Rossi G, Ruggieri S, Taccaliti C, et al. AGEs-related dysfunctions in PCOS: Evidence from animal and clinical research. J Endocrinol. 2021;251[2]:R1–9.
- 115. Garg D, Merhi Z. Relationship between Advanced Glycation End Products and Steroidogenesis in PCOS. Reprod Biol Endocrinol [Internet]. 2016;14[1]:1–13. Available from: http://dx.doi.org/10.1186/s12958-016-0205-6
- 116. Zuo T, Zhu M, Xu W. Roles of oxidative stress in polycystic ovary syndrome and cancers. Oxid Med Cell Longev. 2016;2016.
- 117. Aydogan Mathyk B, Cetin E YB. Use of anti-Müllerian hormone for understanding ovulatory dysfunction in polycystic ovarian syndrome. Curr Opin Endocrinol Diabetes Obes. 2022;29[6]:528–34.
- 118. Armanini D, Boscaro M, Bordin L, Sabbadin C. Controversies in the Pathogenesis, Diagnosis and Treatment of PCOS: Focus on Insulin Resistance, Inflammation, and Hyperandrogenism. Int J Mol Sci. 2022;23[8]:4110.
- 119. Pedro Marques, Adriana De Sousa Lages, Karolina Skorupskaite, Kavitha S. Rozario, Richard A. Anderson and JTG. Physiology of GnRH and Gonadotrophin Secretion. In: Feingold KR, Ahmed SF, Anawalt B, et al. editors, editor. Endotext ([Internet]. On Line. Endotext [Internet]. South Dartmouth (MA): MDText.com, Inc.; 2000–. PMID: 25905297.; 2024. p. 1–121. Available from: https://www-ncbi-nlm-nih-gov.ezproxy.uow.edu.au/books/NBK279070/
- 120. Herbison AE. A simple model of estrous cycle negative and positive feedback regulation of GnRH secretion. Front Neuroendocrinol [Internet]. 2020;57(March):100837. Available from: https://doi.org/10.1016/j.yfrne.2020.100837
- 121. Kelly MJ, Wagner EJ. Canonical transient receptor potential channels and hypothalamic control of homeostatic functions. J Neuroendocrinol. 2024;(March):1–9.
- 122. Adashi EY, Hsueh AJ, Yen SS. Insulin Enhancement of Luteinizing Hormone and Endocrinology. 1981;108[4]:1441–9.
- 123. Silva MSB, Campbell RE. Polycystic Ovary Syndrome and the Neuroendocrine Consequences of Androgen Excess. Compr Physiol. 2022;12[2]:3347–69.
- 124. Briden L, Shirin S, Prior JC. The central role of ovulatory disturbances in the etiology of androgenic polycystic ovary syndrome (PCOS)—Evidence for treatment with cyclic progesterone. Drug Discov Today Dis Model [Internet]. 2020;32:71–82. Available from: https://doi.org/10.1016/j.ddmod.2020.11.008
- 125. Pielecka J, Quaynor SD, Moenter SM. Androgens increase gonadotropin-releasing hormone neuron firing activity in females and interfere with progesterone negative feedback. Endocrinology. 2006;147[3]:1474–9.

- 126. Micevych PE, Chaban V, Ogi J, Dewing P, Lu JKH, Sinchak K. Estradiol stimulates progesterone synthesis in hypothalamic astrocyte cultures. Endocrinology. 2007;148[2]:782–9.
- 127. Uenoyama Y, Inoue N, Nakamura S, Tsukamura H. Kisspeptin neurons and estrogen–estrogen receptor α signaling: Unraveling the mystery of steroid feedback system regulating mammalian reproduction. Int J Mol Sci. 2021;22[17]:1–16.
- 128. Terasawa E. Neuroestradiol in Regulation of GnRH Release. Horm Behav. 2018;104:138-45.
- 129. Blank SK, McCartney CR, Chhabra S, Helm KD, Eagleson CA, Chang RJ, et al. Modulation of gonadotropin-releasing hormone pulse generator sensitivity to progesterone inhibition in hyperandrogenic adolescent girls Implications for regulation of pubertal maturation. J Clin Endocrinol Metab. 2009;94[7]:2360–6.
- 130. Hannon TS, Janosky J, Arslanian SA. Longitudinal study of physiologic insulin resistance and metabolic changes of puberty. Pediatr Res. 2006;60[6]:759–63.
- 131. Gurule S, Sustaita-Monroe J, Padmanabhan V, Cardoso R. Developmental programming of the neuroendocrine axis by steroid hormones: Insights from the sheep model of PCOS. Front Endocrinol (Lausanne). 2023;14(January):1–11.
- 132. Chauvin S, Cohen-Tannoudji J, Guigon CJ. Estradiol Signaling at the Heart of Folliculogenesis: Its Potential Deregulation in Human Ovarian Pathologies. Int J Mol Sci. 2022;23[1]:1–20.
- 133. Dumesic DA, Oberfield SE, Stener-Victorin E, Marshall JC, Laven JS, Legro RS. Scientific statement on the diagnostic criteria, epidemiology, pathophysiology, and molecular genetics of polycystic ovary syndrome. Endocr Rev. 2015;36[5]:487–525.
- 134. Oguz SH, Yildiz BO. An update on contraception in polycystic ovary syndrome. Endocrinol Metab. 2021;36[2]:296–311.
- 135. Cree JME, Brennan NM, Poppitt SD, Miles-Chan JL. The Effect of the Oral Contraceptive Pill on Acute Glycaemic Response to an Oral Glucose Bolus in Healthy Young Women: A Randomised Crossover Study. Nutrients. 2024;16[20]:1–13.
- 136. Prior J. The Case for A New PCOS Therapy. Clue [Internet]. 2018; Accessed On Line: 13 May 2025. Available from: https://helloclue.com/articles/cycle-a-z/the-case-for-a-new-pcos-therapy
- 137. Shirin S, Murray F, Goshtasebi A, Kalidasan D, Prior JC. Cyclic progesterone therapy in androgenic polycystic ovary syndrome (Pcos)—a 6-month pilot study of a single woman's experience changes. Med. 2021;57[10]:1–5.
- 138. Livadas S, Boutzios G, Economou F, Alexandraki K, Xyrafis X, Christou M, et al. The effect of oral micronized progesterone on hormonal and metabolic parameters in anovulatory patients with polycystic ovary syndrome. Fertil Steril [Internet]. 2010;94[1]:242–6. Available from: http://dx.doi.org/10.1016/j.fertnstert.2009.02.073
- 139. Kim SH, Lundgren JA, Patrie JT, Burt Solorzano CM, McCartney CR. Acute progesterone feedback on gonadotropin secretion is not demonstrably altered in estradiol-pretreated women with polycystic ovary syndrome. Physiol Rep. 2022;10[7]:1–16.
- 140. Parker J, O'Brien C, Gersh FL. Developmental origins and transgenerational inheritance of polycystic ovary syndrome. Aust New Zeal J Obstet Gynaecol. 2021;61[6]:922–6.
- 141. O'Reilly MW, House PJ, Tomlinson. JW. Understanding androgen action in adipose tissue. J Steroid Biochem Mol Biol [Internet]. 2014;143:277–84. Available from: http://dx.doi.org/10.1016/j.jsbmb.2014.04.008
- 142. Montes-Nieto R, Insenser M, Martínez-García MÁ, Escobar-Morreale HF. A nontargeted proteomic study of the influence of androgen excess on human visceral and subcutaneous adipose tissue proteomes. J Clin Endocrinol Metab. 2013;98[3]:576–85.
- 143. Rizk J, Sahu R, Duteil D. An overview on androgen-mediated actions in skeletal muscle and adipose tissue. Steroids [Internet]. 2023;199(June):109306. Available from: https://doi.org/10.1016/j.steroids.2023.109306
- 144. Diamanti-Kandarakis E, Pappalou O, Kandaraki EA. The Role of Androgen Excess on Insulin Sensitivity in Women. Front Horm Res. 2019;53:50–64.
- 145. Navarro G, Allard C, Xu W, Mauvais-Jarvis F. The role of androgens in metabolism, obesity, and diabetes in males and females. Obesity. 2015;23[4]:713–9.
- 146. Corbould A. Effects of androgens on insulin action in women: is androgen excess a component of female metabolic syndrome? Diabetes Metab Res Rev. 2008;24:520–32.
- 147. Kempegowda P, Melson E, Manolopoulos KN, Arlt W, O'Reilly MW. Implicating androgen excess in propagating metabolic disease in polycystic ovary syndrome. Ther Adv Endocrinol Metab. 2020;11:1–24.

- 148. Zuo T, Zhu M, Xu W. Roles of oxidative stress in polycystic ovary syndrome and cancers. Oxid Med Cell Longev. 2016;1–14.
- 149. Fazakerley DJ, Krycer JR, Kearney AL, Hocking SL, James DE. Muscle and adipose tissue insulin resistance: Malady without mechanism? J Lipid Res. 2019;60[10]:1720–32.
- 150. Diamanti-Kandarakis E, Mitrakou A, Hennes MMI, Platanissiotis D, Kaklas N, Spina J, et al. Insulin sensitivity and antiandrogenic therapy in women with polycystic ovary syndrome. Metabolism. 1995;44[4]:525–31.
- 151. Diamanti-Kandarakis E, Mitrakou A, Raptis S, Tolis G, Duleba AJ. The effect of a pure antiandrogen receptor blocker, flutamide, on the lipid profile in the polycystic ovary syndrome. J Clin Endocrinol Metab. 1998;83[8]:2699–705.
- 152. Ibanez, L. Potau, N. Marcos, MV. Zegher FF De. Treatment of hirsutism, Hyperandrogenism, Oligomenorrhea, Dyslipidemia, and Hyperinsulinism in Nonobese, Adolescent Girls: Effect of Flutamide. J Clin Endocrinol Metab. 2000;85[9]:3251–5.
- 153. Lasco A, Cucinotta D, Gigante A, Denuzzo G, Pedulla M, Trifiletti A, et al. No changes of peripheral insulin resistance in polycystic ovary syndrome after long-term reduction of endogenous androgens with leuprolide. Eur J Endocrinol. 1995;133[6]:718–22.
- 154. Dale PO, Tanbo T, Djoseland O, Jervell J, Abyholm T. Persistence of hyperinsulinemia in polycystic ovary syndrome after ovarian suppression by gonadotropin-releasing hormone agonist. Acta Endocrinol (Copenh). 1992;126[2]:132–6.
- 155. Moghetti P, Castello R, Magnani CM, Furlani L. Antiandrogen Treatment: Evidence That Androgens Impair Insulin Action in Women *. J Clin Endocrinol Metab. 1996;61[3]:952–60.
- 156. Elkind-Hirsch KE, Valdes CT, Malinak LR. Insulin resistance improves in hyperandrogenic women treated with Lupron. Fertil Steril [Internet]. 1993;60[4]:634–41. Available from: http://dx.doi.org/10.1016/S0015-0282[16]56213-X
- 157. Shoupe D, Lobo RA. The influence of androgens on insulin resistance. Fertil Steril [Internet]. 1984;41[3]:385–8. Available from: http://dx.doi.org/10.1016/S0015-0282[16]47716-2
- 158. Seow KM, Chang YW, Chen KH, Juan CC, Huang CY, Lin L Te, et al. Molecular mechanisms of laparoscopic ovarian drilling and its therapeutic effects in polycystic ovary syndrome. Int J Mol Sci. 2020;21[21]:1–22.
- 159. Nagamani M, Dinh T Van, Kelver ME. Hyperinsulinemia in hyperthecosis of the ovaries. Am J Obstet Gynecol. 1986;154[2]:384–9.
- 160. Angelidi AM, Filippaios A, Mantzoros CS. Severe insulin resistance syndromes. J Clin Invest. 2021;131[4]:e142245.
- 161. Ibáñez L, Valls C, Ferrer A, Ong K, Dunger DB, De Zegher F. Additive effects of insulin-sensitizing and antiandrogen treatment in young, nonobese women with hyperinsulinism, hyperandrogenism, dyslipidemia, and anovulation. J Clin Endocrinol Metab. 2002;87[6]:2870–4.
- 162. Abdalla MA, Shah N, Deshmukh H, Sahebkar A, Östlundh L, Al-Rifai RH, et al. Impact of metformin on the clinical and metabolic parameters of women with polycystic ovary syndrome: a systematic review and meta-analysis of randomised controlled trials. Ther Adv Endocrinol Metab. 2022;13:1–19.
- 163. Cotechini T, Komisarenko M, Sperou A, Macdonald-Goodfellow S, Adams MA, Graham CH. Inflammation in rat pregnancy inhibits spiral artery remodeling leading to fetal growth restriction and features of preeclampsia. J Exp Med. 2014;211[1]:165–79.
- 164. Matteo M, Serviddio G, Massenzio F, Scillitani G, Castellana L, Picca G, et al. Reduced percentage of natural killer cells associated with impaired cytokine network in the secretory endometrium of infertile women with polycystic ovary syndrome. Fertil Steril [Internet]. 2010;94[6]:2222-2227.e3. Available from: http://dx.doi.org/10.1016/j.fertnstert.2010.01.049
- 165. Vega M, Mauro M, Williams Z. Direct toxicity of insulin on the human placenta and protection by metformin. Fertil Steril [Internet]. 2019;111[3]:489-496.e5. Available from: https://doi.org/10.1016/j.fertnstert.2018.11.032
- 166. Lassance L, Haghiac M, Leahy P, Basu S, Minium J, Zhou J, et al. Identification of early transcriptome signatures in placenta exposed to insulin and obesity. Am J Obstet Gynecol [Internet]. 2015;212[5]:647.e1-647.e11. Available from: http://dx.doi.org/10.1016/j.ajog.2015.02.026
- 167. Tarkun I, Arslan BC, Cantürk Z, Türemen E, Şahin T, Duman C. Endothelial dysfunction in young women with polycystic ovary syndrome: Relationship with insulin resistance and low-grade chronic inflammation. J Clin Endocrinol Metab. 2004;89[11]:5592–6.

- 168. Koster MPH, DeWilde MA, Veltman-Verhulst SM, Houben ML, Nikkels PGJ, Van Rijn BB, et al. Placental characteristics in women with polycystic ovary syndrome. Hum Reprod. 2015;30[12]:2829–37.
- 169. Naver K V., Grinsted J, Larsen SO, Hedley PL, Jørgensen FS, Christiansen M, et al. Increased risk of preterm delivery and pre-eclampsia in women with polycystic ovary syndrome and hyperandrogenaemia. BJOG An Int J Obstet Gynaecol. 2014;121[5]:575–81.
- 170. Kingdom JCP, Drewlo S. Is heparin a placental anticoagulant in high-risk pregnancies? Blood [Internet]. 2011;118[18]:4780–8. Available from: http://dx.doi.org/10.1182/blood-2011-07-319749
- 171. Burton GJ, Jauniaux E. The human placenta: new perspectives on its formation and function during early pregnancy. Proc R Soc B Biol Sci. 2023;290:20230191.
- 172. Dimitriadis E, Rolnik DL, Zhou W, Estrada-Gutierrez G, Koga K, Francisco RPV, et al. Pre-eclampsia. Nat Rev Dis Prim. 2023;9[1]:1–22.
- 173. McDonnell R, Hart RJ. Pregnancy-related outcomes for women with polycystic ovary syndrome. Women's Heal. 2017;13[3]:89–97.
- 174. Bui LM, Aghajanova L, Lathi RB, Sokalska A. Polycystic ovary syndrome and miscarriage: a narrative review. F S Rev [Internet]. 2024;5[4]:100078. Available from: https://doi.org/10.1016/j.xfnr.2024.100078
- 175. Hoffman MK. The great obstetrical syndromes and the placenta. BJOG An Int J Obstet Gynaecol. 2023;130(S3):8–15.
- 176. Brosens I, Puttemans P, Benagiano G. Placental bed research: I. The placental bed: from spiral arteries remodeling to the great obstetrical syndromes. Am J Obstet Gynecol. 2019;221[5]:437–56.
- 177. Brosens I, Pijnenborg R, Vercruysse L, Romero R. The "great Obstetrical Syndromes" are associated with disorders of deep placentation. Am J Obstet Gynecol [Internet]. 2011;204[3]:193–201. Available from: http://dx.doi.org/10.1016/j.ajog.2010.08.009
- 178. Yan Q, Qiu D, Liu X, Xing Q, Liu R, Hu Y. The incidence of gestational diabetes mellitus among women with polycystic ovary syndrome: a meta-analysis of longitudinal studies. BMC Pregnancy Childbirth [Internet]. 2022;22[1]:1–12. Available from: https://doi.org/10.1186/s12884-022-04690-3
- 179. Nestler JE. Regulation of the aromatase activity of human placental cytotrophoblasts by insulin, insulin-like growth factor-I, and -II. J Steroid Biochem Mol Biol. 1993;44[4–6]:449–57.
- 180. de Wilde MA, Lamain-de Ruiter M, Veltman-Verhulst SM, Kwee A, Laven JS, Lambalk CB, et al. Increased rates of complications in singleton pregnancies of women previously diagnosed with polycystic ovary syndrome predominantly in the hyperandrogenic phenotype. Fertil Steril [Internet]. 2017;108[2]:333–40. Available from: http://dx.doi.org/10.1016/j.fertnstert.2017.06.015
- 181. Wu Y, Fang W, Fu M, Cheng W, Quon MJ, Yang P. Cellular stress, excessive apoptosis, and the effect of metformin in a mouse model of type 2 diabetic embryopathy. Diabetes. 2015;64[7]:2526–36.
- 182. Eriksson G, Li C, Sparovec TG, Dekanski A, Torstensson S, Risal S, et al. Single-cell profiling of the human endometrium in polycystic ovary syndrome. Nat Med. 2025; March 20(Epub ahead of print).
- 183. Calabuig-Navarro V, Puchowicz M, Glazebrook P, Haghiac M, Minium J, Catalano P, et al. Effect of ω -3 supplementation on placental lipid metabolism in overweight and obese women. Am J Clin Nutr [Internet]. 2016;103[4]:1064–72. Available from: http://dx.doi.org/10.3945/ajcn.115.124651
- 184. Calabuig-Navarro V, Haghiac M, Minium J, Glazebrook P, Ranasinghe GC, Hoppel C, et al. Effect of maternal obesity on placental lipid metabolism. Endocrinology. 2017;158[8]:2543–55.
- 185. Kinshella MLW, Pickerill K, Bone JN, Prasad S, Campbell O, Vidler M, et al. An evidence review and nutritional conceptual framework for pre-eclampsia prevention. Br J Nutr. 2023;130[6]:1065–76.
- 186. Bahri Khomami M, Moran LJ, Kenny L, Grieger JA, Myers J, Poston L, et al. Lifestyle and pregnancy complications in polycystic ovary syndrome: The SCOPE cohort study. Clin Endocrinol (Oxf). 2019;90[6]:814–21.
- 187. Bailey C, Skouteris H, Harrison CL, Hill B, Thangaratinam S, Teede H, et al. A Comparison of the Cost-Effectiveness of Lifestyle Interventions in Pregnancy. Value Heal. 2022;25[2]:194–202.
- 188. Lloyd M, Morton J, Teede H, Marquina C, Abushanab D, Magliano DJ, et al. Long-term cost-effectiveness of implementing a lifestyle intervention during pregnancy to reduce the incidence of gestational diabetes and type 2 diabetes. Diabetologia. 2023;66[7]:1223–34.

- 189. Therapeutics Initiative. Is the current "glucocentric" approach to management of type 2 diabetes misguided? Therapeutics Letter [Internet]. 2016;1–2. Available from: https://www-ncbi-nlm-nih-gov.ezproxy.uow.edu.au/books/NBK598428/
- 190. Rodríguez-Gutiérrez R, Millan-Alanis JM, Barrera FJ, McCoy RG. Value of Patient-Centered Glycemic Control in Patients with Type 2 Diabetes. Curr Diab Rep. 2021;21[12]:63.
- 191. Rodriguez-Gutierrez R, Gonzalez-Gonzalez JG, Zuñiga-Hernandez JA, McCoy RG. Benefits and harms of intensive glycemic control in patients with type 2 diabetes. BMJ. 2019;367:1–20.
- 192. Han SK, Seo MJ, Lee T, Kim MY. Effectiveness of the ALT/AST ratio for predicting insulin resistance in a Korean population: A large-scale, cross-sectional cohort study. PLoS One [Internet]. 2024;19(5 May):1–13. Available from: http://dx.doi.org/10.1371/journal.pone.0303333
- 193. Schwartz SS, Epstein S, Corkey BE, Grant SFA, Gavin JR, Aguilar RB, et al. A Unified Pathophysiological Construct of Diabetes and its Complications. Trends Endocrinol Metab. 2017;28[9]:645–55.
- 194. Kaviyaadharshani D, Nivedhidha M, Jeyarohini R, Lece Elizabeth Rani J, Ramkumar MP, Emil Selvan GSR. Diagnosing Diabetes using Machine Learning-based Predictive Models. Procedia Comput Sci [Internet]. 2024;233[2023]:288–94. Available from: https://doi.org/10.1016/j.procs.2024.03.218
- 195. Althobaiti T, Althobaiti S, Selim MM. An optimized diabetes mellitus detection model for improved prediction of accuracy and clinical decision-making. Alexandria Eng J [Internet]. 2024;94(February):311–24. Available from: https://doi.org/10.1016/j.aej.2024.03.044
- 196. Bukhari MM, Alkhamees BF, Hussain S, Gumaei A, Assiri A, Ullah SS. An Improved Artificial Neural Network Model for Effective Diabetes Prediction. Complexity. 2021;2021:1–10.
- 197. Mechanick JI, Garber AJ, Grunberger G, Handelsman Y, Timothy Garvey W. Dysglycemia-based chronic disease: An American association of clinical endocrinologists position statement. Endocr Pract. 2018;24[11]:995–1011.
- 198. McEwan P, Foos V, Roberts G, Jenkins RH, Evans M, Wheeler DC, et al. Beyond glycated haemoglobin: Modelling contemporary management of type 2 diabetes with the updated Cardiff model. Diabetes, Obes Metab. 2025;(December 2024):1752–61.
- 199. Defronzo RA. From the triumvirate to the ominous octet: A new paradigm for the treatment of type 2 diabetes mellitus. Diabetes. 2009;58[4]:773–95.
- 200. Grover-Páez F, Maya Gómez A, Hernández Suárez A MEAF a GA to P of M-OD in T 2 D [Internet]. From a Glycocentric Approach to Prevention of Multi-Organ Damage in Type 2 Diabetes. In: Chlup R, editor. Type 2 Diabetes in 2024 From Early Suspicion to Effective Management [Internet]. Intech Open; 2024. p. 99. Available from: http://dx.doi.org/10.5772/intechopen.1002363
- 201. Unger RH, Cherrington AD. Glucagonocentric restructuring of diabetes: A pathophysiologic and therapeutic makeover. J Clin Invest. 2012;122[1]:4–12.
- 202. Chatzis DG, Kolokathis K, Magounaki K, Chatzidakis S, Avramidis K, Leopoulou M, et al. Changing the Concept: From the Traditional Glucose-centric to the New Cardiorenal-metabolic Approach for the Treatment of Type 2 Diabetes. touchREV Endocrinol. 2021;17[2]:92–101.
- 203. Blagosklonny M V. TOR-centric view on insulin resistance and diabetic complications: perspective for endocrinologists and gerontologists. Cell Death Dis. 2013;4[12]:1–8.
- 204. Saisho Y. An emerging new concept for the management of type 2 diabetes with a paradigm shift from the glucose-centric to beta cell-centric concept of diabetes an Asian perspective. Expert Opin Pharmacother [Internet]. 2020;21[13]:1565–78. Available from: https://doi.org/10.1080/14656566.2020.1776262
- 205. Gorgojo Martínez JJ. Glucocentricity or adipocentricity: A critical view of consensus and clinical guidelines for the treatment of type 2 diabetes mellitus. Endocrinol y Nutr (English Ed [Internet]. 2011;58[10]:541–9. Available from: http://dx.doi.org/10.1016/j.endoen.2011.09.002
- 206. Shiffman D, Louie JZ, Meigs JB, Devlin JJ, McPhaul MJ, Melander O. An insulin resistance score improved diabetes risk assessment in the malmö prevention project—a longitudinal population-based study of older europeans. Diabetes Care. 2021;44[10]:e186–7.
- 207. Ogurtsova K, Guariguata L, Barengo NC, Ruiz PLD, Sacre JW, Karuranga S, et al. IDF diabetes Atlas: Global estimates of undiagnosed diabetes in adults for 2021. Diabetes Res Clin Pract. 2022;183[109118]:1–9.

- 208. Handelsman Y, Butler J, Bakris GL, DeFronzo RA, Fonarow GC, Green JB, et al. Early intervention and intensive management of patients with diabetes, cardiorenal, and metabolic diseases. J Diabetes Complications [Internet]. 2023;37[2]:108389. Available from: https://doi.org/10.1016/j.jdiacomp.2022.108389
- 209. Janssen JAMJL. Hyperinsulinemia and its pivotal role in aging, obesity, type 2 diabetes, cardiovascular disease and cancer. Int J Mol Sci. 2021;22[15]:7797.
- 210. DeFronzo RA, Tobin JD, Andres R. Glucose clamp technique: A method for quantifying insulin secretion and resistance. Am J Physiol Endocrinol Metab Gastrointest Physiol. 1979;6[3]:E214–23.
- 211. Gastaldelli A. Measuring and estimating insulin resistance in clinical and research settings. Obesity. 2022;30[8]:1549–63
- 212. Stern SE, Williams K, Ferrannini E, Defronzo RA, Bogardus C, Stern MP. Identification of Individuals With Insulin Resistance Using Routine Clinical Measurements. Diabetes. 2005;54[3]:333–9.
- 213. Staten MA, Stern MP, Miller WG, Steffes MW, Campbell SE. Insulin assay standardization: Leading to measures of insulin sensitivity and secretion for practical clinical care. Diabetes Care. 2010;33[1]:205–6.
- 214. Sharma VR, Matta ST, Haymond MW, Chung ST. Measuring Insulin Resistance in Humans. Horm Res Paediatr. 2021;93[11–12]:577–88.
- 215. Khan SH, Khan AN, Chaudhry N, Anwar R, Fazal N, Tariq M. Comparison of various steady state surrogate insulin resistance indices in diagnosing metabolic syndrome. Diabetol Metab Syndr [Internet]. 2019;11[1]:1–9. Available from: https://doi.org/10.1186/s13098-019-0439-5
- 216. Prashant A, Nataraj SM, Swetha N, Gowda J, Najmunnissa F, Guruswamy KM. Unveiling the Significance of Surrogate Markers of Insulin Resistance in Metabolic Health Assessment. Indian J Med Biochem. 2024;28[2]:45–53.
- 217. Michael Stumvoll MA. Use of the Oral Glucose Tolerance Test to Assess Insulin Release and Insulin S e n s i t i v ity. Ep i d e m i o l o g y / H e a l t h S e r v i c e s / P s y c h o s o c i a l R e s e a r c h. 2000;23 [3]:295–301.
- 218. Kraft JR. Detection of diabetes mellitus in situ (occult diabetes). Lab Med. 1975;6[2]:10-22.
- 219. Hayashi T, Boyko EJ, Sato KK, McNeely MJ, Leonetti DL, Kahn SE, et al. Patterns of insulin concentration during the OGTT predict the risk of type 2 diabetes in Japanese Americans. Diabetes Care. 2013;36[5]:1229–35.
- 220. Chiara Dalla Man, Marco Campioni, Kenneth S. Polonsky, Rita Basu, Robert A. Rizza. Two-Hour Seven-Sample Oral Glucose Tolerance Test and Meal Protocol. Diabetes. 2005;54[11]:3265–73.
- 221. Kattamis C, Ladis V, Skafida M, Iacovidou N, Theodoridis C. The different patterns of insulin response during oral glucose tolerance test (Ogtt) in transfused young patients with β-thalassemia. Acta Biomed. 2021;92[4]:1–7.
- 222. Uysal E, Tammo O, Soylemez E, Incebiyik M, Filiz D, Alci M. Significance of measuring anthropometric and atherogenic indices in patients with polycystic ovary syndrome. BMC Endocr Disord. 2024;24[1]:1–8.
- 223. Mansour A, Noori M, Hakemi MS, Haghgooyan Z, Mohajeri-Tehrani MR, Mirahmad M, et al. Hyperandrogenism and anthropometric parameters in women with polycystic ovary syndrome. BMC Endocr Disord [Internet]. 2024;24[1]:201. Available from: https://doi.org/10.1186/s12902-024-01733-y
- 224. Agrawal H, Aggarwal K, Jain A. Visceral adiposity index: Simple Tool for assessing cardiometabolic risk in women with polycystic ovary syndrome. Indian J Endocrinol Metab. 2019;23[2]:232–7.
- 225. Aboeldalyl S, James C, Seyam E, Ibrahim EM, Shawki HED, Amer S. The role of chronic inflammation in polycystic ovarian syndrome—a systematic review and meta-analysis. Int J Mol Sci. 2021;22[5]:1–31.
- 226. Szukiewicz D, Trojanowski S, Kociszewska A, Szewczyk G. Modulation of the Inflammatory Response in Polycystic Ovary Syndrome (PCOS)—Searching for Epigenetic Factors. Int J Mol Sci. 2022;23[14663]:1–27.
- 227. Tremellen K, Pearce K. Dysbiosis of Gut Microbiota (DOGMA) A novel theory for the development of Polycystic Ovarian Syndrome. Med Hypotheses [Internet]. 2012;79[1]:104–12. Available from: http://dx.doi.org/10.1016/j.mehy.2012.04.016
- 228. Parker, J. O'Brien C, Hawrelak J. A narrative review of the role of gastrointestinal dysbiosis in the pathogenesis of polycystic ovary syndrome. Obstet Gynecol Sci. 2022;65[1]:14–28.
- 229. Zhou X, Wu Y, Zhu Z, Lu C, Zhang C, Zeng L, et al. Mucosal immune response in biology, disease prevention and treatment. Signal Transduct Target Ther [Internet]. 2025;10[1]:1–32. Available from: http://dx.doi.org/10.1038/s41392-024-02043-4
- 230. Stegehuis N, Kotsirilos V, Parker J. The Impact of Microparticulate Air Pollution in Polycystic Ovary Syndrome: A Narrative Review. Clin Exp Obstet Gynecol. 2024;51[10]:233.

- 231. Mahmud F, Sarker DB, Jocelyn JA, Sang QXA. Molecular and Cellular Effects of Microplastics and Nanoplastics: Focus on Inflammation and Senescence. Cells. 2024;13[21]:1–22.
- 232. Liu YZ, Wang YX, Jiang CL. Inflammation: The common pathway of stress-related diseases. Front Hum Neurosci. 2017;11(June):1–11.
- 233. Rudnicka E, Kunicki M, Suchta K, Machura P, Grymowicz M, Smolarczyk R. Inflammatory Markers in Women with Polycystic Ovary Syndrome. Biomed Res Int. 2020;2020:1–10.
- 234. Deng H, Chen Y, Xing J, Zhang N, Xu L. Systematic low-grade chronic inflammation and intrinsic mechanisms in polycystic ovary syndrome. Front Immunol. 2024;15(December):1–20.
- 235. Vasyukova E, Zaikova E, Kalinina O, Gorelova I, Pyanova I, Bogatyreva E, et al. Inflammatory and Anti-Inflammatory Parameters in PCOS Patients Depending on Body Mass Index: A Case-Control Study. Biomedicines. 2023;11[10]:1–13.
- 236. Hatziagelaki E, Pergialiotis V, Kannenberg JM, Trakakis E, Tsiavou A, Markgraf DF, et al. Association between Biomarkers of Low-grade Inflammation and Sex Hormones in Women with Polycystic Ovary Syndrome. Exp Clin Endocrinol Diabetes. 2020;128[11]:723–30.
- 237. Chen J, Mullins CD, Novak P, Thomas SB. Personalized Strategies to Activate and Empower Patients in Health Care and Reduce Health Disparities. Heal Educ Behav. 2016;43[1]:25–34.
- 238. Saadati N, Haidari F, Barati M, Nikbakht R, Mirmomeni G, Rahim F. The effect of low glycemic index diet on the reproductive and clinical profile in women with polycystic ovarian syndrome: A systematic review and meta-analysis. Heliyon [Internet]. 2021;7[11]:e08338. Available from: https://doi.org/10.1016/j.heliyon.2021.e08338
- 239. Sabag A, Patten RK, Moreno-Asso A, Colombo GE, Dafauce Bouzo X, Moran LJ, et al. Exercise in the management of polycystic ovary syndrome: A position statement from Exercise and Sports Science Australia. J Sci Med Sport [Internet]. 2024;27[10]:668–77. Available from: https://doi.org/10.1016/j.jsams.2024.05.015
- 240. Sharma K, Akre S, Chakole S, Wanjari MB. Stress-Induced Diabetes: A Review. Cureus. 2022;14[9]:1-6.
- 241. Olorunfemi Oyewole Babalola, Paul Olamide Ottu, Ebenezer Akinnusi Iwaloye, Precious Olayinka Aturamu O. Lifestyle Interventions to Manage Insulin Resistance [Internet]. On Line. Raghav, Alok. Shaginian R, editor. Glucose and Insulin Homeostasis. IntechOpen; 2024. 122 p. Available from: https://www.intechopen.com/books/1002641
- 242. Buechner H, Toparlak SM, Ostinelli EG, Shokraneh F, Nicholls-Mindlin J, Cipriani A, et al. Community interventions for anxiety and depression in adults and young people: A systematic review. Aust N Z J Psychiatry. 2023;57[9]:1223–42.
- 243. Yuan J, Li Z, Yu Y, Wang X, Zhao Y. Natural compounds in the management of polycystic ovary syndrome: a comprehensive review of hormonal regulation and therapeutic potential. Front Nutr. 2025;12(February):1–21.
- 244. Fadlalmola HA, Elhusein AM, Al-Sayaghi KM, Albadrani MS, Swamy DV, Mamanao DM, et al. Efficacy of resveratrol in women with polycystic ovary syndrome: a systematic review and meta-analysis of randomized clinical trials. Pan Afr Med J. 2023;44[134]:1–17.
- 245. Viña I, Viña JR, Carranza M, Mariscal G. Efficacy of N-Acetylcysteine in Polycystic Ovary Syndrome: Systematic Review and Meta-Analysis. Nutr . 2025;17[2]:1–18.
- 246. Rondanelli M, Infantino V, Riva A, Petrangolini G, Faliva MA, Peroni G, et al. Polycystic ovary syndrome management: a review of the possible amazing role of berberine. Arch Gynecol Obstet [Internet]. 2020;301[1]:53–60. Available from: https://doi.org/10.1007/s00404-020-05450-4
- 247. Mallya P, Lewis SA. Curcumin and its formulations for the treatment of polycystic ovary syndrome: current insights and future prospects. J Ovarian Res [Internet]. 2025;18[78]:1–17. Available from: https://doi.org/10.1186/s13048-025-01660-z
- 248. Shahmoradi S, Chiti H, Tavakolizadeh M, Hatami R, Motamed N, Ghaemi M. The Effect of Magnesium Supplementation on Insulin Resistance and Metabolic Profiles in Women with Polycystic Ovary Syndrome: a Randomized Clinical Trial. Biol Trace Elem Res [Internet]. 2024;202[3]:941–6. Available from: https://doi.org/10.1007/s12011-023-03744-7
- 249. Rashid R, Mir SA, Kareem O, Ali T, Ara R, Malik A, et al. Polycystic ovarian syndrome-current pharmacotherapy and clinical implications. Taiwan J Obstet Gynecol [Internet]. 2022;61[1]:40–50. Available from: https://doi.org/10.1016/j.tjog.2021.11.009
- 250. Helminski D, Sussman JB, Pfeiffer PN, Kokaly AN, Ranusch A, Renji AD, et al. Development, Implementation, and Evaluation Methods for Dashboards in Health Care: Scoping Review. JMIR Med Informatics. 2024;12:1–19.

- 251. Phillips MCL. Metabolic Strategies in Healthcare: A New Era. Aging Dis. 2022;13[3]:655-72.
- 252. Olyanasab A, Annabestani M. Leveraging Machine Learning for Personalized Wearable Biomedical Devices: A Review. J Pers Med. 2024;14[203]:1–21.
- 253. Van Hul M, Cani PD, Petifils C, De Vos WM, Tilg H, El Omar EM. What defines a healthy gut microbiome? Gut. 2024;73:1893–908.
- 254. Yin G, Chen F, Chen G, Yang X, Huang Q, Chen L, et al. Alterations of bacteriome, mycobiome and metabolome characteristics in PCOS patients with normal/overweight individuals. J Ovarian Res [Internet]. 2022;15[1]:1–15. Available from: https://doi.org/10.1186/s13048-022-01051-8
- 255. Luo X, Dong Y, Zheng H, Zhou X, Rong L, Liu X, et al. CAPN2 correlates with insulin resistance states in PCOS as evidenced by multi-dataset analysis. J Ovarian Res. 2024;17[1]:1–13.
- 256. Liu YN, Qin Y, Wu B, Peng H, Li M, Luo H, et al. DNA methylation in polycystic ovary syndrome: Emerging evidence and challenges. Reprod Toxicol [Internet]. 2022;111(April):11–9. Available from: https://doi.org/10.1016/j.reprotox.2022.04.010
- 257. Zhao X, Meng Q, Liu S, Cheng L, Li B, Cheng D. Integrated multi-omics analysis reveals complement component 3 as a central driver of immune dysregulation in polycystic ovary syndrome. Front Endocrinol (Lausanne). 2025;16(March):1–14.
- 258. Percy C, Turner A, Orr C. Developing a Novel Web-Based Self-Management Support Intervention for Polycystic Ovary Syndrome: Mixed Methods Study With Patients and Health Care Professionals. JMIR Form Res. 2024;8:1–22.
- 259. Chen W, Miao J, Chen J, Chen J. Development of machine learning models for diagnostic biomarker identification and immune cell infiltration analysis in PCOS. J Ovarian Res . 2025;18[1]:1–16.
- 260. Awad A, Trenfield SJ, Pollard TD, Ong JJ, Elbadawi M, McCoubrey LE, et al. Connected healthcare: Improving patient care using digital health technologies. Adv Drug Deliv Rev [Internet]. 2021;178:113958. Available from: https://doi.org/10.1016/j.addr.2021.113958

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.