The relationship between carotid intima-media thickness and serum secreted frizzled-related protein-4 and dipeptidyl peptidase-4 in the diabetic patients with cardiovascular diseases

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ABSTRACT

Objective: Diabetes is associated with increased cardiovascular and cerebrovascular diseaserelated mortality. We investigated the association between carotid intima-media thickness (CIMT) with clusterin (CLU), amylin, secreted frizzled-related protein-4 (SFRP-4), total and active glucagon-like peptide-1 (GLP-1) levels, and dipeptidyl peptidase-4 (DPP-4) in type 2 diabetes mellitus (T2DM) individuals with or without coronary artery disease (CAD). Methods: This study consisted of four groups in Medicine Hospital between April- October, 2017: Control group (mean ages: 50.3 ± 10.7 years, 20 females and 15 males), diabetic group (DM) (mean ages: 53.9 ± 11.1 , 14 females and 23 males), CAD group-(mean ages: 62.6 ± 11.8 , 17 females and 17 males) and CAD+ DM group: (mean ages: 62.6±11.8 years, 18 females and 18 males). Results: Both CAD and CAD+DM groups have higher CIMT levels than controls. CAD+DM group have also significantly higher CIMT levels than DM group. Left external carotid artery (ECA) was only found different in DM group from controls. When compared with controls, DM, CAD groups have low DPP-4 and GLP-1_{total} concentration. Control group have significantly lower SFRP-4 levels than DM, CAD and CAD+DM (p<0.001) groups. Serum GLP-1_{total} levels were found to be significantly low in CAD+DM group when compared to control group. Conclusion: DPP-4 and SFRP-4 levels are predictive marker for atherosclerosis in diabetes, correlates well with HOMA-IR particularly in diabetes. CIMT has the potential to be a clinically useful predictor of vascular risk in diabetic patients with CAD. Large cohorts and at-risk populations are needed to confirm the predictive value of these findings.

Keywords: Type 2 Diabetes Mellitus; Carotid intima-media thickness; glucagon-like peptide-1; dipeptidyl peptidase-4; clusterin; amylin; secreted Frizzled-related protein-4

INTRODUCTION

Diabetes mellitus (DM) has reached epidemic proportions worldwide, and its prevalence is rising. The implications of a diagnosis of DM are as severe as a diagnosis of coronary artery disease (CAD). DM, CAD and heart failure are interacting dynamically. There has been considerable improvement in the management of patients with CAD, coronary event rates remain heightened among patients with DM. Enhanced cardiovascular risk stratification based on biomarkers, symptoms and classical risk factors should be performed in patients with pre-existing DM (1,2).

Carotid intima-media thickness (CIMT), arterial stiffness, and epicardial fat thickness are useful non-invasive markers of subclinical atherosclerosis (3). All carotid B-mode real-time ultrasound measurements were performed by the same experienced physician, who was blinded to the patient's urine albumin status. Measurements of the IMT were performed in both the right and left common carotid arteries (CCAs), external carotid arteries (ECA) and internal carotid arteries (ICAs), as previously described (4). It has been reported that the CIMT remained stable in type 2 DM (T2DM) patients who received comprehensive intensive therapy, suggesting that multi-factorial intensive therapies might have potential in reducing macro-vascular events in these patients (5).

Amylin, or islet amyloid polypeptide (IAPP), is a neuroendocrine hormone colocalized, co-secreted and co-packaged with insulin from pancreatic β cells. Amylin functions as part of the neuroendocrine pancreas and contributes to glucose homeostasis with other two pancreatic islet hormones insulin and glucagon (6).

Secreted frizzled related protein 4 (SFRP-4) is a member of the SFRP family. SFRPs act as modulators of the wingless-type mouse mammary tumor virus integration site family (Wnt) signaling pathway. A large number of diabetes-associated factors are studied in the Wnt signaling pathway (6). Individuals having increased levels of SFRP-4 in the blood are five times more likely to develop diabetes in the coming years (7).

The two major incretin hormones, glucagon-like peptide (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP, previously known as gastric inhibitory polypeptide), are secreted from the small intestine in response to meal ingestion and act on specific receptors on the β-cells. Both are metabolized by the enzyme dipeptidyl peptidase-4 (DPP-4). The cleavage can be blocked by specific DPP-4 inhibitors, resulting in increased plasma concentrations of the intact peptides and improved glucose tolerance. DPP-4 inhibitors have been proposed as a possible pharmacological treatment of T2DM (8), and many

compounds are presently in clinical development (8-10). GLP-1 circulates in many different (degraded) forms in the blood, some of which are biologically active and others are not.

Clusterin (CLU) apolipoprotein J (here after CLU) is a 449–amino acid disulfide-linked heterodimeric glycoprotein composed of α and β subunits and generated by a single cleavage in the single-chain precursor protein. However, CLU was not found in the normal aorta but was rather localized in aortas with diffuse, intimal thickening or atherosclerotic lesions; the extend of CLU distribution in the aortic wall increased during the progression of the disease from fatty acid streaks to advanced atherosclerosis (11-13).

CAD is the cause of death in more than half of all diabetic patients, and many are debilitated by symptoms of congestive heart failure or angina. Therefore, this study aimed to explore the association between the concentrations of serum amylin, SFRP-4, GLP-1_{total}, GLP-1_{active}, DPP-4 and CLU and CIMT and to investigate whether these parameters have been atherosclerotic effects in T2DM individuals.

MATERIALS AND METHODS

Subjects

The protocol was approved by the Ethics Committee of Cerrahpasa Medical Faculty and was conducted in accordance with the Declaration of Helsinki. All participants were informed about the survey and voluntarily signed and dated the consent form. This case-control study was conducted in Department of Internal Medicine, Medicine Hospital, and Istanbul between April-October 2017. All subjects were Turkish descent. Pregnant women, patients with renal, hepatic, rheumatic, malign or endocrine diseases, smokers and subjects who were taking drugs which could affect our results were excluded.

Studied group are classified as follows;

General characteristics of studied group were given in Table 1.

Control group: 35 healthy subjects who have no any endocrine, vascular, cardiac or inflammatory disease were accepted as control group (mean ages: 50.3 ± 10.7 years, 20 females and 15 males). An oral questionnaire was applied to the subjects and none of our subjects declared evidence of family history of diabetes. They did not have diabetes, or glucose intolerance confirmed with oral glucose tolerance test (OGTT).

Type 2 Diabetic group (DM): Patients with newly diagnosed T2DM (mean ages: 53.9 ± 11.1 years, 14 females and 23 males) were included in this study. For the diagnosis of DM, guidelines of American Diabetes Association (ADA) criterions were used (14). Diabetic patients involved in our study have no under medical therapy.

CAD group: 34 patients (mean ages: 62.6±11.8 years, 17 females and 17 males) with coronary artery disease were studied. 55% of the patients have hypertension and they were under the therapy with beta blockers (60%), thiazide (35%) and/or ACE inhibitors (13%).

CAD+DM group: 36 diabetic patients (mean ages: 62.6±11.8 years, 18 females and 18 males) with coronary artery diseases were enrolled in our study. All of the diabetic patients had under the therapy for diabetes with insulin (23%) and/or metformin (80%). 86% of diabetic patients in this group have hypertension and they were under the therapy with beta blockers (50%), thiazide (30%) and /or ACE inhibitors (15%). Dyslipidemic diabetic patients (72%) were used antihyperlipidemic drugs such as statins.

Ultrasonographic measurement of carotid intima-media thickness (CIMT)

The extracranial carotid arteries were examined using a standardized protocol by the same radiologist. Ultrasonographic examinations were performed in a quiet, temperature controlled room (22 °C). After 10 min of rest, the examinations were performed with a color Doppler ultrasound unit [General electrics (GE) Logiq S7 Expert, 9 L MHz transducer (prob), USA] equipped with a 5–10-MHz transducer. External carotid artery (ECA) was scanned only for atherosclerotic plaques. Carotid artery for measurement of IMT was not measured. IMT was measured across a 1-cm segment of both the right and left sides of the near and far walls of the distal common carotid artery (CCA), the far wall of the carotid bulb ECA and the internal carotid artery (ICA). The proximal 1.5 cm of the ICA was measured. Atherosclerotic plaque was defined as a distinct area protruding into the vessel lumen. This protrusion had to be at least 50% thicker than the surrounding areas. When plaques were present, measurements were made from their outside borders. The mean of all measurements from eight locations was taken as an overall measure of CIMT. The location, size, number and hemodynamic effects of the atherosclerotic plaques were determined with the help of grayscale, color Doppler and spectral Doppler ultrasound. All of the measurements were made at the time of scanning of the frozen images from the longitudinal scans using the machine's electronic caliper. The radiologist was blinded to the clinical diagnoses. Intra-observer was assessed by a repeated evaluation of 15 randomly selected participants after 2 weeks. Intra-observer agreement was good (κ =0,82).

Sample collection and measurements

Fasting venous blood samples were drawn between 8 and 10 am, after an overnight fasting (10-12 hours). Blood samples were drawn via brachial veins in brachial fossa into plain tubes and into anticoagulant [ethylenediaminetetraacetic acid (EDTA)] containing tubes. Samples were centrifuged for 10 minutes at 4000 rpm at 40C. Biochemical tests were

performed immediately. For the determination of other parameters serum aliquots were frozen and stored at -80°C immediately until further analysis.

Measurement of serum GLP-1 concentrations

Serum GLP-1_{total} and GLP-1_{active} were assayed by antibody sandwich ELISA kit (Cat. EZGLP1T-36K, and Cat. EGLP-35K, EMP Millipore Corporation, USA). Results were expressed as pM. The sensitivity of GLP-1 total ELISA kit was 1.5 pM. Intra and inter-CV for GLP-1_{total} were %6.7 and %11.3, respectively. The lowest level of GLP-1_{active} that can be detected by this assay was 2 pM. Intra and inter-CV for GLP-1 levels were 6.5% and 10.4 %, respectively.

Measurement of serum DPP-4 activity

Levels of serum DPP-4 were also assayed by antibody sandwich ELISA kit (Human DPP-4 kit Cat. No. YHB1023 Hu, ARP American Research Products, Inc. USA). Results of DPP-4 were expressed as pg per mL of serum. The lowest level of DPP-4 that can be detected by this assay was 25 pg/mL. Intra and inter-CV are 8.1% and 10.5%, respectively.

Measurement of serum CLU concentrations

Levels of serum CLU were determined by antibody sandwich ELISA kit assay (Human CLU ELISA Kit, Cat.No: YHB0754 Hu, ARP American Research Products, Inc. USA). Results of CLU levels were expressed as μg per mL of serum ($\mu g/mL$). The lowest level of CLU that can be detected by this assay was 0.24 $\mu g/mL$. Intra and inter variation of the coefficient (CV) were 6.2% and 8.4%, respectively.

Measurement of serum amylin concentrations

Levels of serum amylin were assayed by ELISA kit (Human Amylin Cat No. YHB0161 Hu, ARP American Research Products, Inc. USA). Results were expressed as pg per ml of serum (pg/mL). The sensitivity of this kit was 1.36 pg/mL. Intra and inter-CV are 7.1 % and 9.0 %, respectively.

Measurement of serum SFRP-4 concentrations

Levels of serum SFRP-4 were determined by antibody sandwich ELISA kit (Human SFRP-4 ELISA Kit, Cat No. E2327 Hu, Bioassay Technology Laboratory, USA). Results were expressed as ng per ml of serum (ng/mL). The lowest level of SFRP4 that can be detected by this assay was 1.5 pM. CV for intraassay and interassay were %5.5 and %11.2, respectively.

Glucose, total cholesterol, HDL cholesterol, LDL cholesterol and triglyceride levels were determined by the enzymatic methods (Roche Cobas İntegra 400, Roche Diagnostics Ltd. Germany). Insulin concentrations were measured by electrochemiluminescence

immunoassay (ECLIA) method on Roche-Hitachi E170 (Roche/Hitachi MODULAR Analytics Combination Systems, Roche Diagnostics, USA). HbA1c determination was based on HPLC (Variant Turbo II, Bio-Rad Laboratories, Inc. USA).

HOMA-IR (Homeostatic Model Assessment for Insulin Resistance) was calculated according to the formula: fasting insulin (microU/L) x fasting glucose (nmol/L)/22.5.

Statistical analysis

Statistical analysis was performed using SPSS 20.0 version for Windows Statistical Program (SPSS, Chicago, IL, USA). All data were expressed as means ± standard deviation (SD). Descriptive statistics were obtained and data were tested for normality using the Kolmogorov-Smirnov test for Gaussian distribution. For comparison of parameters with normal distribution parametric tests and comparison of parameters with abnormal distribution non- parametric tests were used. For this purpose, One-Way ANOVA, unpaired student- t, Kruskal-Wallis and Mann-Whitney U tests were used. CLU, amylin, DDP-4, SFRP-4, GLP-1total, GLP-1active and HOMA-IR were shown abnormal distribution. Tukey's (for parametric analysis) and Dunn's tests (for non-parametric analysis) were used as post-hoc tests. For parametric tests, continuous variables are expressed in mean ± standard deviation, while for non-parametric tests, data are expressed in median and interquartile range (25th and 75th percentiles). Relationships between variables were assessed with Pearson's or Spearman's correlation coefficient. Power analysis was used to perform calculations on sample size, effect size, and statistical power. The minimal significance (α) and statistical power (1 – β) were set at 0.05 and 0.80 respectively. A p value equal to or lower than 0.05 was considered statistically significant.

RESULTS

Patients in DM group, CAD and CAD+DM groups have significantly higher fasting plasma glucose concentration than controls (for each p<0.001). The highest plasma glucose levels were obtained from CAD+ DM groups. HbA1c levels in DM and CAD+DM groups were significantly higher control group (P<0.001). There was also significant difference in HbA1c levels between CAD and CAD+DM groups (p<0.001). Plasma total cholesterol levels in controls and CAD groups were significantly higher than DM and CAD+DM groups (respectively, p<0.005 and p<0.005). HDL cholesterol levels is found to be higher in control group than DM, CAD and CAD+DM groups (p<0.05, p<0.001 and p<0.005). Plasma triglycerides levels were only higher in DM group then control group (P<0.01). There was no significant difference in LDL cholesterol levels among groups. HOMA-IR levels were higher in DM and CAD+DM group than controls (p<0.01 and p<0.01). Duration of diabetes in DM

group was not different from CAD+DM group. Uric acid and homocysteine levels in CAD were significantly higher than DM group (for each p<0.01) and control group (for each p<0.005). Uric acid and homocysteine levels in DM group were not different from CAD+DM group. Systolic blood pressure in DM and CAD+DM group were significantly higher than control group (for each p<0.01).

CIMT levels in studied groups were given in table 2. Left ECA was only found different in DM group from controls (p<0.05). Both CAD and CAD+DM groups have higher ICA levels than controls (for each comparison p<0.001). There was no significant difference in ICA levels between CAD and CAD+DM groups. CAD+DM group have significantly higher ICA levels than DM group (for each comparison p<0.001).

We found that serum CLU, amylin and GLP-1_{active} levels were not different among groups (Table 3). When compared with controls, DM, CAD groups have low DPP-4 (p<0.01 and p<0.005) and GLP-1_{total} (p<0.001 and p<0.001) concentration. Control group have significantly lower SFRP-4 levels than DM (p<0.005), CAD (p<0.005) and CAD+DM (p<0.001) groups. Serum GLP-1_{total} levels were found to be significantly lowers in CAD+DM group than in control group. There was also significant difference in GLP-1_{total} between CAD+DM and DM groups (p<0.05).

When we performed correlation analysis in sum of the groups, we were no found any correlation between CIMT and studied parameters, as well as lipids parameters. Significant positive correlation was found between CLU levels (figure 1) and amylin (r: 0.804, p<0.01), DPP-4 (r:0.524, p<0.01) and SFRP-4 levels (p<0.800, p<0.01). Serum amylin levels were correlated with DPP-4 (r: 0.644, p<0.01) and (r: 0.528, p<0.01) (figure 2). A significant weak positive correlation was found between DPP-4 and GLP-1 total (r: 0.205, p<0.05). GLP-1total levels were negative correlated with GLP-1active levels (r: -0.222, p<0.05). There was a significant weak positive correlation between HOMA-IR and SFRP-4 levels (r: 0.244, p<0.05).

DISCUSSION

Type 2 DM is a complex disease with concomitant risk factors for the development of cardiovascular disorders, such as atherosclerosis and hypertension. Atherosclerotic macrovascular disease is the leading cause of death in type 2 diabetes and CIMT is increased in patients with T2DM (4). This study showed that left ECA was only found different in DM group from controls. Both CAD and CAD+DM groups have higher ICA levels than controls. CAD+DM group have higher ICA levels than DM group. This is supported by the

independent association between studied parameters, as well as lipids parameters and CIMT in T2DM patients which is likely due to atherosclerosis characterized by the pathogenesis of vascular complications of diabetes.

Amylin, which is considered the primary culprit for β -cell loss in T2DM patients, is synthesized in β -cells of the pancreas from its precursor proamylin and plays an important role in early intracellular amyloid formation as well (15). Serum amylin levels were also not different among groups in present study as with our previous study (16). However, results of studies in T2DM patients in recent years are conflicting. Zheng et al. (15) found that the serum levels of amylin in the three groups (normal glucose tolerance (NGT) group, patients with impaired glucose regulation (IGR) and T2DM) had no significant differences. The serum proamylin were significantly higher in patients with IGR and T2DM than in control subjects. It appears that proamylin is more important and exert more significant effect than amylin. Skovronsky et al. (17) found that proamylin might have more severe cell toxicity than amylin and then played an important role in the deposition of islet amyloid. Qiu et al. (18) showed that subjects with a long and chronic duration of diabetes were more likely to take insulin treatment and have reduced secretion of amylin. However, further experiments are needed to clarify the role of proamylin and amylin.

The role of CLU in attenuation of inflammation and reverse cholesterol transfer makes this molecule a potential candidate as a marker for cancer, CVD, DM, and metabolic syndrome. An important source of CLU in plasma is associated with HDL particles. In present study, HDL cholesterol levels are found to be higher in control group than DM, CAD and CAD+DM groups. However, we found that serum CLU levels were not different among groups. Significant positive correlation was found between CLU levels and amylin levels in our study. Trougakos et al. (19) increased serum CLU levels in T2DM was found and CLU might be a useful biomarker for detecting the early stage of diabetic retinopathy. They have also demonstrated that plasma CLU levels increase significantly in patients with T2DM which is a well characterized risk factor for atherosclerosis. Study of Cai et al. (20) suggested that plasma CLU concentration increased and was negatively correlated with memory performance in T2DM patients with mild cognitive impairment (MCI). Circulating CLU is associated with insulin resistance in human subjects (21). Future studies will need to clarify the exact role of CLU associated with atherosclerosis in T2DM patients with or without CAD.

SFRP-4 is a regulator of insulin exocytosis in murine and human islet cells. Our data demonstrate that control group has lower SFRP-4 levels than DM, CAD and CAD+DM groups. We found that there was also significant difference in SFRP-4 levels between DM,

CAD and CAD+DM groups. There was also significant weak positive correlation between HOMA-IR and SFRP-4 levels. Mahdi et al. (22) found that serum SFRP-4 in was associated with elevated fasting glucose and reduced disposition index. However, it was also associated with impaired insulin sensitivity, indicating that the protein could have a plethora of metabolic effects and might be released from several tissues involved in glucose homeostasis. They declared increased serum SFRP-4 levels several years before the clinical diagnosis of T2DM was made, proposing the possibility of SFRP-4 as an early risk predictor and it could be a therapeutic target for specific treatment of islet dysfunction. Hoffmann et al. (23) showed that elevated SFRP-4 levels were associated with T2DM, metabolic syndrome, and severity of diabetes. The primary outcome was the composite of cardiovascular death and cardiovascular hospitalization within 48 months' follow-up. Comparison of event-free survival between SFRP-4 tertiles showed that SFRP-4 levels were not predictive for cardiovascular outcome in patients with stable CAD on treatment. Ji et al. (24) found that plasma SFRP-4 levels were increased in CAD patients compared to non-CAD patients. Our results are similar; plasma SFRP-4 levels were positively correlated with BMI, fasting insulin levels and HOMA-IR values. CAD was an independent predictor of the increased plasma SFRP-4 levels. All results and our result suggest that SFRP-4 is a novel biomarker of CAD and might play a role in the development of CAD+DM due to SFRP-4 was up-regulated in patients with T2DM (15, 22-28).

GLP-1 has short half-lives, since they are rapidly degraded by DPP-4, a ubiquitous enzyme found in soluble form in plasma or as a membrane component of many cells (29), including endothelial cells (30). Elevated DPP-4 in patients with diabetes may justify, at least partially, the status of incretin deficiency/resistance related to T2DM. DPP-4 inhibitors may potentially reduce cardiovascular (CV) risk. GLP-1, DPP-4 acts on other substrates, many of which are associated with cardioprotection in experimental models. Inhibition of DPP-4 may also lead to elevations in several substrates with potentially favorable effects on vascular function and anti-coagulation (31, 32). We have shown for the first time in previous our study (15) that diabetic patients with microvascular complications have higher DPP-4 activity and GLP-1_{total} levels than diabetic patients without such complications. DPP-4 activity in CAD and CAD+DM groups has lower than controls in present study. Control group have significantly higher GLP-1_{total} levels than DM, CAD and CAD+DM groups. There was also significant difference in GLP-1_{total} between CAD+DM and DM groups. A significant weak positive correlation was found between DPP-4 activity in patients with T2DM have shown conflict

results such as reduced (33, 34) or increased activity (15, 35-38). However, these disparate results may have occurred due to the use of drugs such as metformin and glitazones, which are both able to promote a decrease in DPP-4 activity (31,39). Thus, whether increased/decreased DPP-4 and GLP-1 levels have beneficial or adverse effects of the pleiotropic effects of GLP-1 and DPP-4 on the CV system remains inconclusive. Different treatments may improve the pleiotropic effects of GLP-1 and DPP-4 on the CV system in patients with CAD+DM.

The power point of our study is that we evaluated the association between the concentrations of serum multiple biomarkers and CIMT and to investigate whether these parameters have been atherosclerotic effects in T2DM individuals. However, our study has some limitations. First, our sample size is relatively small. Second, dietary habits, physical activity and the exercise level of the subjects were not documented. Third, we did not investigate cardiovascular comorbidities and drugs which could affect our results. Due to the cross-sectional design of our study, we cannot make any suggestions about the association between the laboratory and clinical parameters of the subjects.

Conclusion

Patients with T2DM are at increased risk of cardiovascular disease. In addition to hyperglycemia that contributes to increased CV risk, patients with T2DM often have other conditions, such as hypertension and dyslipidemia that contribute to the development of cardiovascular complications (32). DPP-4 and SFRP-4 levels are predictive marker for atherosclerosis in diabetes, correlates well with HOMA-IR particularly in diabetes. CIMT has the potential to be a clinically useful predictor of vascular risk in diabetic patients with CAD. Large cohorts and at-risk populations are needed to confirm the predictive value of these findings.

Abbreviations

CIMT:carotid intima-media thickness; CLU:clusterin; SFRP-4:secreted frizzled-related protein-4; GLP-1:glucagon-like peptide-1; DPP-4:dipeptidyl peptidase-4; T2DM:type 2 diabetes mellitus; CAD:coronary artery disease; IAPP:islet amyloid polypeptide; ADA:American Diabetes Association; EDTA:ethylenediaminetetraacetic acid; ECLIA:electrochemiluminescence immunoassay; HOMA-IR:Homeostatic Model Assessment for Insulin Resistance; NGT:normal glucose tolerance; IGR:impaired glucose regulation; MCI:mild cognitive impairment; CV:cardiovascular; CCA:common carotid artery; ICA:internal carotid artery; ECA:external carotid artery

Authors' contributions

All authors wrote, revised and approved the manuscript. All authors read and approved the final manuscript.

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Not applicable.

Competing interests

The authors declare that they have no competing interests.

Availability of data and materials

All data generated and analyzed during this study are included in this published article.

Consent for publication

Not applicable.

Ethics approval and consent to participate

The protocol was approved by the Ethics Committee of Cerrahpasa Medical Faculty and was conducted in accordance with the Declaration of Helsinki. All participants were informed about the survey and voluntarily signed and dated the consent form.

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Table 1. General characteristics of studied groups

	Controls	DM	CAD	CAD+ DM
	(n:35)	(n:37)	(n:34)	(n:36)
Ages (years)	50.3 ± 10.7	53.9 ± 11.1	60.1 ± 13.5	62.6±11.8
Sex (F/M)	20/15	14/23	17/17	18/18
Systolic Blood Pressure	115.9 ± 10.6	131.7 ± 10.2^{b}	124.6 ± 14.4	132.7 ± 15.4^{b}
(mmHg)				
Diastolic Blood Pressure	72.3 ± 6.8	79.4 ± 7.2	75.4 ± 9.4	77.7 ± 10.2
(mmHg)				
Duration of diabetes	-	4.8 ± 4.6	-	6.2 ± 5.8
(years)				
HOMA-IR*	3.32	4,41	3.22	3.87
	(1.76-4.34)	(3.29-5.98) ^b	(2.21-5.81)	$(2.96-7.64)^{c}$
Total cholesterol (mg/dL)	150.0 ± 48.5	202.9 ± 39.2^{c}	168.6±41.9°	196.3±39.1 °
HDL-cholesterol (mg/dL)	49.9± 16.9	42.2 ± 15.4^{a}	36.8 ± 10.4^{d}	$38.8 \pm 9.1^{d. \text{ f}}$
LDL-cholesterol (mg/dL)	94.5 ± 34.9	104.8 ± 36.9	104.8±36.8	99.8±40.1
Triglycerides (mg/dL)	119.6± 47.7	211.5 ± 54.8^{b}	138.6 ± 84.8	135.5±70.9
Uric Acid (mg/dL)	5.26 ± 1.28	5.42± 1.39	6.40 ±1.64 ^{c,e}	5.54 ± 1.31
Homocysteine (µmol/L)	9.77 ± 3.83	10.41 ± 2.48	$14.3 \pm 8.08^{c,e}$	11.86 ± 4.81

DM; Diabetes mellitus, **CAD**; coronary artery diseases, **CAD+DM**; Diabetic patient with coronary artery diseases, **HOMA-IR**; Homeostatic Model Assessment for Insulin Resistance

Comparison with control group ^ap<0.05, ^bp<0.015, ^cp<0.005, ^dp<0.001

Comparison with diabetes mellitus group ep<0.01, fp<0.005

Comparison with CAD group ^gp<0.001

Values other than HOMO-IR are given as mean \pm standard deviation.

^{*}Result were given as median and interquartile range (25th and 75th percentiles)

Table 2. Carotid intima-media thickness (mm) in groups of controls, type 2 diabetic patients with (CAD+DM) or without coronary artery disease (DM) and patients with coronary artery disease (CAD).

Groups	Right CCA	Right ICA	Right ECA	Left CCA	Left ICA	Left ECA
Controls	0.68 ± 1.06	0.77±1.23	0.51±0.65	0.58 ± 0.80	0.56 ± 0.77	0.47 ± 0.51
DM	0.75 ± 0.26	0.73 ± 0.24	0.66 ± 0.20	$0,78 \pm 0.22$	0.78 ± 0.26	0.66 ± 0.16^{a}
CAD	0.97 ± 0.26^{b}	1.01 ± 0.28^{b}	0.85 ± 0.19^{b}	1.05 ± 2.79^{b}	1.05 ± 0.45^{b}	0.88 ± 0.18^{b}
CAD+DM	$0.90\pm0.32^{b,c}$	$0.96\pm0.29^{b,c}$	0.83±0.23 ^{b,c}	$1.03\pm0.37^{b,c}$	$1.06\pm0.31^{b,c}$	$0.87\pm0.30^{b,c}$

CCA, Common carotid artery; ICA, internal carotid artery; ECA, external carotid artery Comparison with control group ap<0.05, bp<0.001

^b Comparison n with diabetes mellitus group ^cp<0.001

Table 3. Serum levels of clusterin (CLU), amylin, dipeptidly- peptidase-4 (DPP-4), secreted frizzled related protein 4 (SFRP-4), glucagon like peptide -1(GLP-1)_{total} and GLP-1_{active} in groups of controls, diabetic patients with (CAD+DM) and without coronary artery disease (DM) and patients with coronary artery disease (CAD).

	Control	DM	CAD	CAD+ DM
	(n:35)	(n:37)	(n:34)	(n:36)
CLU (µ/mL)	22.2	27.1	20.4	28.1
,	(19.6-37.5)	(19.8-38.2)	(19.1-20.3)	(19.4-59.9)
Amylin (pg/mL)	410 .1	436.9	416.2	428.9
	(358.5-691.0)	(328.6-611.1)	(361.2-567.8)	(350.9-767.4)
DPP-4 (ng/mL)	3496	2861	2617	3191.4
	(3169-6387)	$(2213.1-5004.1)^{a}$	$(2265.7-4445.4)^{b}$	(2017.3-7157.4)
SFRP-4 (ng/mL)	1.41	1.70	1.58	2.02
	(1.24-1.67)	$(1.51-2.45)^{c}$	$(1.31-2.55)^{b}$	$(1.47-3.47)^{c}$
GLP-1 _{total} (pM)	91,9	39.0	43.4	51.4
	(73.3-106.3)	$(33.9-49.9)^{c}$	$(31.2-52.9)^{c}$	$(34.8-71.5)^{c,d}$
GLP-1 _{active} (pM)	3.92	4.71	5.25	4.25
	(3.40-5.16)	(3.50-5.93)	(3.63-6.10)	(3.38-7.64)

DM; Type 2 diabetes mellitus, **CAD**; Coronary artery disease, **CAD**+ **DM**; Type 2 diabetic patients with CAD

Comparison with control group ap<0.01, bp<0.005, cp<0.001

Comparison with DM group, dp<0.05

Result were given as median and interquartile range (25th and 75th percentiles)

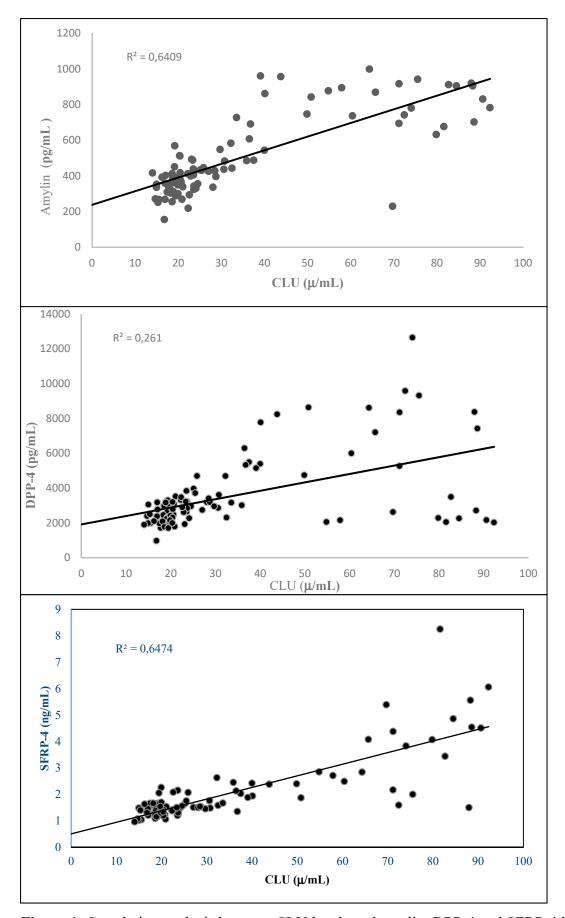


Figure 1. Correlation analysis between CLU levels and amylin, DPP-4 and SFRP-4 levels

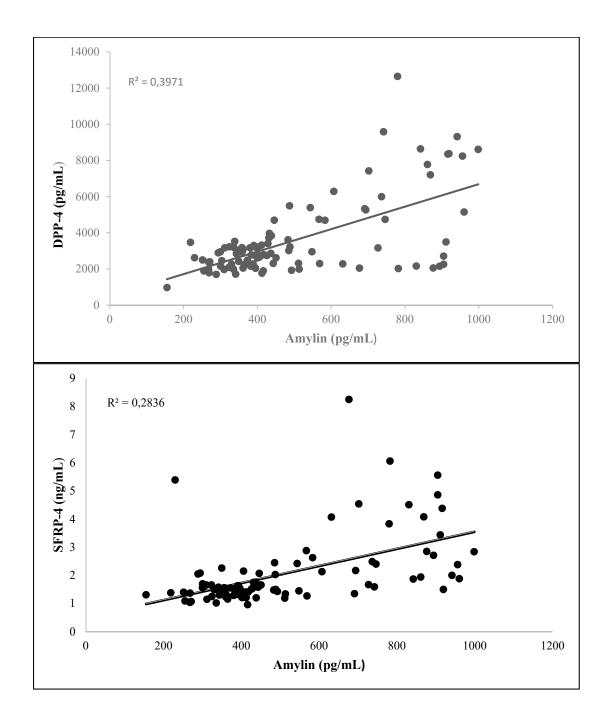


Figure 2. Correlation analysis between amylin levels and DPP-4 and SFRP-4 levels.