

Review

Not peer-reviewed version

Understanding Cancer as a Systemic Disease by Understanding Neural Stemness as the Core Property of Cancer Cell

[Ying_Cao](#)*

Posted Date: 14 August 2025

doi: 10.20944/preprints202508.1001.v1

Keywords: neural stemness; general stemness; pluripotency; tumorigenicity; tumorigenesis; unicellular state; neural induction; neural default model



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Understanding Cancer as a Systemic Disease by Understanding Neural Stemness as the Core Property of Cancer Cell

Ying Cao

The MOE Key Laboratory of Model Animals for Disease Study, Model Animal Research Center, Medical School of Nanjing University, Nanjing, China; caoying@nju.edu.cn; Tel.: +86-25-58641537

Abstract

Cancer is a systemic disease with multilayered complexity. Some theories/hypotheses have been proposed to explain cancer. They are successful in explaining certain aspects of cancer, but meet serious challenges in other aspects. Inappropriate understanding of cancer cell and cancer also hinders the development of more effective strategies of cancer therapy. My previous studies demonstrated that the core property of cancer (tumorigenic) cells is neural stemness. The finding led to the subsequent discovery about the central role of neural stemness during tumorigenesis and a novel paradigm that can hopefully explain systemic complexity of cancer as a whole. In the review, I summarize the evidence from evolutionary, developmental biology and cancer biology research supporting that neural stemness, referring to the collective property of embryonic neural/neural stem cells, represents the general stemness, the cellular property that determines both pluripotency and tumorigenicity. I make detailed discussion about the key role of neural stemness in understanding the core property and phenotypic traits of cancer cell and in understanding cancer complexity. These pieces of evidence and discussion reveal that cancer is the manifestation of the power of general rules dictating both embryogenesis and tumorigenesis. Briefly, acquiring neural stemness in ectodermal cells during embryogenesis, i.e., neural induction, leads to body axis formation, and ectopic neural induction causes a conjoined twin in an embryo; whereas acquirement of neural stemness in cells of a postnatal animal/human results in a degenerated conjoined twin-like structure, i.e., a tumor. Due to the causal role of neural stemness in tumorigenesis, novel strategies of cancer therapy can be developed by targeting neural stemness using the principle of pluripotent cell differentiation. In addition, some essential issues worth considering in cancer research are also discussed.

Keywords: neural stemness; general stemness; pluripotency; tumorigenicity; tumorigenesis; unicellular state; neural induction; neural default model

Introduction

Cancer is recognized as a systemic disease because of its multi-dimensional complexity (Swanton et al., 2024). Cancer research has revealed that almost all aspects of biological research are accountable for cancer, such as from the finest molecular details to evolutionary ecosystem (Bhattacharya et al., 2025; Merlo et al., 2006), from embryonic development to aging (Rubin, 1985; Stanger and Wahl, 2024; Swanton et al., 2024), from genetic heterogeneity (Turajlic et al., 2019) to phenotypic heterogeneity (Meacham and Morrison, 2013), and to the heterogeneity of a particular biological process involved in cancer, for example, metabolic heterogeneity (Tong et al., 2020). Molecular studies on cancer cells and different elements in tumor microenvironment (TME), including different types of cells and even microbiome, revealed that nearly every gene can be associated with cancer (de Magalhães, 2022). Such a scenario of systemic complexity means that dealing with cancer is almost dealing with the whole animal world. In the past time, cancer has been

understood primarily based on reductionist cell biology, and molecular mechanism-driven cancer research has produced extraordinary discoveries that help understanding cancer cells and their local microenvironment. But frustratingly, very few innovative new therapeutic strategies that are broadly beneficial across different cancers and significantly prolong overall survival have been developed, and therapy resistance is always the insurmountable obstacle (Swanton et al., 2024). Dilemma of this reality prompts to rethink of cancer more inclusively to reflect the multidimensional complexity of disease mechanisms, instead of focusing only on the reductionist view of shared hallmarks of cancer. Novel and more efficient therapeutic strategies should be developed based on the systemic complexity of cancer (Bhattacharya et al., 2025; Swanton et al., 2024). Cancer is a disease with multidimensional complexity. If looking at the complex elements, e.g., cell types, interactions, etc., in the scenario of tumor ecosystem, it raises the question what is the causality in the ecosystem and what are the causal and supporting elements. The answer to the question will improve the understanding cancer as a systemic disease and help to identify the key factor to consider for developing novel therapeutic strategies. Cancer is a complex derivative from normal cells and contains almost all elements in animals. Therefore, tumorigenesis should also follow the rules operating in the animal kingdom. My studies identified neural stemness, which represents the general stemness, as the core property of cancer (tumorigenic) cells, which led to the subsequent finding of general rules that governs both tumorigenesis and embryogenesis. These rules explain complexity of cancer as the derivative from neural stemness. In the review, I will discuss the role of neural stemness as the cornerstone in understanding tumorigenesis. Moreover, I will also discuss misconceptions and ambiguities that may complicate the research and understanding of cancer.

1. Neural Stemness Represents the General Stemness

By intuition, neural stemness is recognized as a type of tissue stemness. But further analysis and integration of information from evolutionary and developmental biology research suggest otherwise.

1.1. Pluripotency of Neural Stem Cells (NSCs)

Embryonic pluripotent cells, such as amphibian blastula ectodermal cells and mammalian embryonic stem cells (ESCs), are considered as the basis for differentiation because of their ability of differentiation into all types of cells during normal embryogenesis and in experimental conditions. Neural stem/progenitor cells and neural crest cells (NCCs) appear later than embryonic pluripotent cells during embryonic development and their obvious contribution to embryonic development is formation of the nervous system. Therefore, neural stemness, referring to the collective property of primitive NSCs, NCCs, adult NSCs and neural progenitor cells, was naturally considered as a type of tissue stemness. This view has considered simply the obvious properties of embryonic pluripotent cells and neural stemness. The hidden relationship between the stemness of embryonic pluripotent cells and neural stemness has been overlooked. During gastrulation of amphibian embryogenesis, neuroectoderm is formed via a process called “neural induction”, in which the ectoderm loses its epidermal fate and acquires neuroectodermal fate in response to inhibition of TGF β signaling (Cao, 2023). Neuroectoderm gives rise to neural plate, which contributes to formation of not only the central nervous system, but non-neural cells as well during later developmental stage. In the most posterior region of elongating embryos, neuromesodermal progenitors, which are presumably originated from anterior neural plate, generate both spinal cord and paraxial mesoderm (Henrique et al., 2015; Sambasivan and Steventon, 2021). Locating between neural plate and epidermal ectoderm, neural crest is induced by interactions between neural plate and adjacent tissues (Selleck and Bronner-Fraser, 1995; Knecht and Bronner-Fraser, 2002; Pla and Monsoro-Burq, 2018). It is well characterized that NCCs are pluripotent because of they are not just the precursors of the peripheral nervous system. They also contribute to a variety of non-neural tissues, such as melanocytes, skeletal and connective tissues, and medulla cells of the adrenal gland (Le Douarin and Dupin, 2016). Meanwhile, pluripotent property of NCCs is supported by a pluripotency-like molecular program in these cells (Pajanoja et al., 2023; Zalc et al., 2021). Developmental relationship between neural plate and neural

crest means that pluripotency of NCCs is manifestation of the property of neural plate cells, i.e., the primitive NSCs. Pluripotency of NSCs is not obvious during embryogenesis, but was experimentally verified (Cao, 2022; Clarke et al., 2000; Tropepe et al., 2001; Xu et al., 2021; Zhang et al., 2022). NSCs also have pluripotency-like molecular program. The four original reprogramming factors, Sox2, c-Myc, Oct4 and Klf4, subsequent alternative reprogramming factors, and reprogramming co-regulators, are all enriched in neural precursor/progenitor cells during vertebrate embryonic development (Cao, 2022).

1.2. The 'Neural Default State' of Embryonic Pluripotent Cells

Pluripotency of NSCs can be traced back to the 'neural default model' of embryonic pluripotent cells during embryogenesis (Muñoz-Sanjuán and Brivanlou, 2002). Ectoderm is the germ layer that gives rise to both epidermis and nervous system, and other tissues/organs are mostly derived from either endoderm or mesoderm. How the neural tissue in an early embryo is induced to form had been a major topic of research in developmental biology. Spemann and Mangold in 1924 demonstrated that dorsal blastopore lip, or the Spemann-Mangold organizer, of an early newt gastrula embryo was able to induce neural plate in ectoderm when transplanted into the ventral side of a host embryo, while the dorsal lip itself developed into mesodermal notochord. After a tortuous process in exploration of the mechanisms underlying neural inducing activity by the organizer, it was elucidated that absence, but not presence, of an extracellular signal is prerequisite for neural fate decision, suggesting that neural fate might be the default fate of ectoderm (Godsave and Slack, 1989; Grunz and Tacke, 1989; Sato and Sargent, 1989). In agreement, the organizer is the rich source of secreted factors antagonizing BMP4, a TGF β ligand that transduces epidermis-inducing and anti-neural signaling in ectoderm (De Robertis, 2006; De Robertis and Kuroda, 2004; Harland, 2000). It was concluded that neural fate is achieved by default and epidermal fate is induced during ectodermal cell fate decision, i.e., the 'neural default model' of ectoderm (Muñoz-Sanjuán and Brivanlou, 2002). Amphibian blastula ectodermal cells were further validated to be the equivalent of mammalian ESCs, which also follow the 'neural default model'. They adopt neural fate and turn into primitive NSCs in absence of extracellular inducers (Smukler et al., 2006; Tropepe et al., 2001).

1.3. Unicellular Origin of Neural Stemness

The neural default state of pluripotency is rooted in the evolutionary advantage of neural genes and neural stemness. During evolution, origin of ectoderm is the earliest, followed sequentially by endoderm and mesoderm. Comparison of evolutionary origin of neural and non-neural genes in ectoderm showed that one peak of emergence of neural genes is already present in the time point representing the last common ancestors of eukaryotes and the other is at the time of emergence of eumetazoa (Domazet-Loso et al., 2007), indicating that neural genes have an earlier evolutionary origin and hence, play a critical role in multicellularity. Detailed analysis on more than 5,000 neural genes in vertebrates demonstrated that most of these genes can be traced back to *Monosiga brevicollis*, *Amphimedon queenslandica* and *Trichoplax adhaerens*, which are the closest species representing transition from unicellularity to multicellularity, and share a last common unicellular ancestor in more than 600 million years ago. *M. brevicollis* represents the closest unicellular relatives of metazoans, *A. queenslandica* is the oldest surviving metazoan and an evolutionary intermediary between unicellular choanoflagellate protists and eumetazoans, and *T. adhaerens* is the basal species of eumetazoan. Therefore, most ancestral neural genes had emerged during the transition from unicellularity to multicellularity. Importantly, more than 60% of genes in *M. brevicollis* that are homologous to vertebrate genes are ancestral neural genes, suggesting that the last common unicellular ancestor was biased towards a neural state (Xu et al., 2021). The notion is further supported by identification of neurosecretory apparatus in *M. brevicollis* and unicellular origin of neurosecretory cell-cell signalling (Burkhardt et al., 2011; Göhde et al., 2021).

This means that neural biased state is the ground state or the starting cellular state for the transition from unicellularity to multicellularity during evolution. In addition, genes coding for the

components of machineries required for basic cellular physiological functions and developmental programs are mostly enriched in embryonic neural cells, such as cell cycle, ribosome, spliceosome, proteasome, epigenetic modification, reprogramming, DNA damage and repair. They work concerted together to define neural stem/embryonic neural cells as a highly proliferative and pluripotent state. These basic machineries are common to eukaryotes, hence, have a unicellular origin. Embryonic neural cells represent the direct descendant cells of the unicellular ancestors (Cao, 2022; Chen et al., 2021; Xu et al., 2021).

Emergence of TGF β signaling during evolution also suggests that neural state represents the ground state of differentiation. TGF β signaling is required for inhibition of neural differentiation and promotion of non-neural differentiation of ESCs or during germ layer differentiation (Itoh et al., 2014; Meyers and Kessler, 2017; Ozair et al., 2013). Meanwhile, BMP4, a TGF β family member, is also required for maintenance of ESC pluripotency, because ESCs adopting a neural fate in the absence of BMP4 signaling has been considered as a differentiation effect (Malaguti et al., 2013; Ying et al., 2003). As a prime signaling promoting non-neural differentiation during embryogenesis, emergence of TGF β pathway coincided with the onset of multicellularity during evolution (Nicols et al., 2006), suggesting its role in cell type diversification. The pathway is not present in *M. brevicollis*, but present in *A. queenslandica* (Adamska et al., 2007; King et al., 2008; Srivastava et al., 2010). Moreover, study on pluripotent cells in *A. queenslandica* also confirmed that pluripotency has a unicellular origin (Sogabe et al., 2019). Therefore, evolutionary studies indicate that neural-biased state of the last common unicellular ancestors represents the ground state of pluripotency (Cao, 2022). Adoption of neural fate of ESCs in the absence of TGF β signaling should not be interpreted as a differentiation effect, but rather a reversal effect to the most initial state of pluripotency. Evolutionary advantage of neural genes and neural stemness explains why the default fate of embryonic pluripotent cells is neural. TGF β signaling has been considered to maintain pluripotency. But paradoxically, its inhibition improves reprogramming to generate pluripotent state (Woltjen and Stanford, 2009). The paradox can be resolved if the evolutionary origin of pluripotency and the relationship between ESCs and primitive NSCs are considered. The induced pluripotent stem cells (iPSCs) are equivalent to primitive NSCs rather than ESCs.

In addition to evolutionary advantage, neural genes are characteristic of over-representation of long genes with more exons and introns, as compared with non-neural genes. Neural genes are twice as long as non-neural genes, and have four more exons/introns than non-neural genes in average (Cao, 2022; Gabel et al., 2015; Xu et al., 2021; Zylka et al., 2015). Longer genes with more exon/introns facilitate binding of different regulators or forming secondary structures, and hence, can serve as more flexible scaffolds for diverse regulatory signals during differentiation. More exon/intron compositions mean these genes can generate more splice variants via splicing. Accordingly, components of the machinery of alternative splicing, a mechanism contributing to phenotypic novelty during evolution, and to cell differentiation, lineage determination and organogenesis during development (Baralle and Giudice, 2017; Bush et al., 2017), are enriched in embryonic neural cells (Cao, 2022). By contrast, shorter genes should have no such advantages. Enrichment of longer genes makes neural stemness a more flexible scaffold and an appropriate initial state for cell differentiation. By integrating the evidence from studies on evolution, cellular properties and regulatory networks, and intrinsic association between ESCs and NSCs, **it can be concluded that neural stemness represents general stemness**, i.e., the ground state of pluripotency. Neural induction is in fact the process of returning to the neural ground state of embryonic pluripotent cells. Unicellular origin of neural stemness and pluripotent state, and neural default state of embryonic pluripotent cells is summarized in Figure 1 (Figure 1).

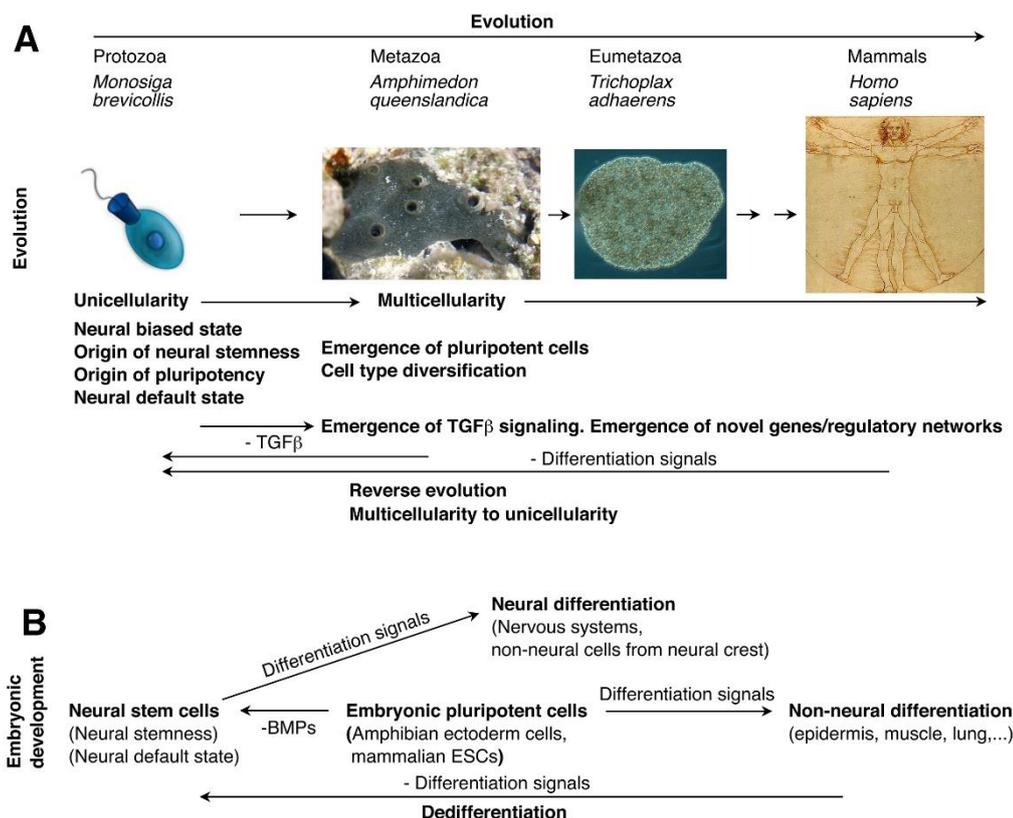


Figure 1. Evidence from evolutionary and developmental biology studies supporting neural stemness as the general stemness. (A) Neural-biased state of the last common unicellular ancestors indicating unicellular origin of neural stemness and pluripotency, which serves as the initial state from unicellularity to multicellularity during animal evolution. Emergence of TGFβ signaling in the basal species of metazoan and other new genes and regulatory networks during evolution promotes cell type diversification in animals. (B) The ‘neural default model’ of embryonic pluripotent cells. Embryonic pluripotent cells are capable of neural and non-neural differentiation. For neural differentiation, BMP signaling must be inhibited, e.g., by secreted signals from the Spemann-Mangold organizer, thereby embryonic pluripotent cells turning into primitive neural stem cells, a process determined by evolution as depicted in (A). Loss of BMP signaling in embryonic pluripotent cells and loss of differentiation signals in differentiated cells, i.e., dedifferentiation, reflects an effect of reverse evolution.

2. Neural Stemness as the Core Property of Cancer Cell

Cancer cell was ever the primary focus of cancer research before 1980s because mutations in oncogenes and tumor suppressor genes are seemingly sufficient to determine cancer initiation and progression. But this cancer cell-intrinsic view has met difficulties in explaining the mechanisms that govern cancer metastasis. Studies on crosstalks in TME and interactions between tumor and host tissues might provide reasonable explanations, leading to a shift of focus on cancer cell to tumor environment (Garner and de Visser, 2020; Maman and Witz, 2018; Vogelstein and Kinzler, 1993). Cell and molecular biology research has revealed a number of phenotypic traits of cancer cell that are usually supposed to be not manifested by normal cells, including fast proliferation, invasion/migration, stemness, evasion of cell death and immune destruction, therapy resistance, dysregulated epigenetics and metabolism. Development of novel therapeutic strategies based on molecular mechanisms that regulate these phenotypic traits has achieved very limited effects. The frustrating situation also incites researchers to consider more comprehensively the complexity of cancer instead of cancer cells themselves (Swanton et al., 2024). It is believed that phenotypic traits of cancer cell, e.g., metastasis, are the consequence of acquirement of ‘mesenchymal state’ via process called ‘epithelial-mesenchymal transition (EMT)’ during tumorigenesis. However, after more than 50 years of extensive EMT study and publication of more than 50K EMT papers, nothing is clear about

the basics of EMT, including epithelial and mesenchymal states, EMT-specific markers, clear evidence of EMT, and even basic rationale of EMT, rendering EMT a scientifically groundless and meaningless concept (Cao, 2024; Yang et al., 2020). Understanding cancer cell with a poorly defined 'concept' is an obvious violation of scientific reasoning. This raises a question how much cancer cell has been really understood. Major cancer promoting factors are best investigated for their roles in regulating of phenotypic traits of cancer cell. It is common to see that one factor is able to regulate different phenotypic traits of cancer cells. For example, EZH2 plays oncogenic role in different types of cancers, promotes cancer cell stemness (Balint et al., 2022), proliferation (Bryant et al., 2007), metastasis (Zingg et al., 2015), chemoresistance (Ougolkov et al., 2008; Crea et al., 2012), metabolic dysregulation (Ahmad et al., 2017), immune evasion (Kim et al., 2020; Zhou et al., 2020), etc. c-Myc regulates almost all phenotypic traits, including immune evasion, of cancer cells (Dhanasekaran et al., 2022; Fatma et al., 2022; Llombart and Mansour, 2022). These mean that different phenotypic traits of cancer cells are not independent from each other, but rather, intrinsically connected. Understanding how these traits are intrinsically connected also needs better understanding of the property of cancer cell.

2.1. Cancer (Tumorigenic) Cells Are Characteristic of Neural Stem/Embryonic Neural Cells

Cancer cells are immature cells, and differentiation is expected to suppress malignancy of cancer cells. In an initial study, we tried to find out whether it was possible to drive terminal differentiation of different types of cancer cells using a same approach. Instead of using a technical strategy for screening some common molecules that might be involved in differentiation/dedifferentiation of cancer cells, we set up a few limiting rules, which were inferred from characteristics of cancer cells, to narrow down the number of candidate factors (Zhang et al., 2017). These restrictions led us to focusing on HDAC1, HDAC3, EZH2, LSD1 and DNMT1, the best-known epigenetic modification factors that are pan-cancer promoting proteins (Zhang et al., 2017). Interestingly, combined inhibition of these oncoproteins led to post-mitotic neuronal-like differentiation in cells of different types of cancer, including hepatocellular carcinoma, prostate cancer, breast cancer, colon cancer, melanoma, osteosarcoma, glioblastoma, and lung cancer. As expected, differentiated cancer cells showed decrease in expression in cancer promoting proteins, malignant features and tumorigenicity. This was the first piece of evidence that, in contrast to extensive heterogeneity, different cancer cells might share the property of neural stem/progenitor cells, i.e., neural stemness. In agreement, expression of the genes for these epigenetic factors are all enriched in embryonic neural cells during early neural development, and they play essential role in maintaining stemness of neural stem cells (Cao, 2017; Zhang et al., 2017). Further analysis revealed that the 'core EMT factors/markers', SNAIL1/2, ZEB2, TWIST1, N-Cadherin and Vimentin, which promote cancer or generally upregulated in cancer cells, are also specifically expressed or enriched in embryonic neural cells. In fact, these are markers for neural stem/crest cells. It seemed that embryonic neural expression of cancer promoting genes should not be just accidental. To clarify whether it is a general rule that cancer promoting genes are embryonic neural/neural stemness genes, more than 3,000 cancer genes were categorized into genes promoting cancer/upregulated in cancer cells, genes suppressing cancer/downregulated in cancer cells, and genes playing dual roles in cancer, and their embryonic tissue expression was analyzed. The analysis led to the generalization that most (if not all) cancer promoting genes are embryonic neural/neural stemness genes, and by contrast, a majority of cancer suppressor genes are non-neural genes (Zhang et al., 2017). Therefore, cancer cells share regulatory networks with embryonic neural cells, which confer cancer cells the property of neural stemness (Cao, 2017; Zhang et al., 2017). The link between neural stemness and cancer cells is also manifested by many lineage-tracing studies. For example, CD133 (or PROM1), Msi1, Sox2, and Dcl1 were used as tracing markers to identify cancer-initiating cells or cancer stem cells (CSCs) in different types of cancer, including colon cancer, pancreatic cancer, squamous-cell carcinoma (Boumahdi et al., 2014; Fox et al., 2016; Nakanishi et al., 2013; Ricci-Vitiani et al., 2007). CD133, Sox2, Msi1 or Dcl1, which are frequently used as CSC markers, are either typical markers for NSCs and/or their genes are specifically expressed in neural tissues in vertebrate embryos. Considering that neural stemness represents the ground state for cell

differentiation, neural stemness and expression of genes involved in promoting/maintaining neural stemness genes are diluted in differentiated cells. Differentiated cells in postnatal animal/human may experience intracellular/extracellular insults, including mutations, chromosomal instability, aneuploidy, microenvironmental changes, gene misregulation, etc., which may accidentally cause downregulation/silencing of tissue-specific genes or differentiation genes, or upregulation/activation of neural genes or both. Then differentiated cells will return progressively back to their original ground state, i.e., neural stemness. These results shed the light on the unified principle underlying tumorigenesis beyond enormous inter- and intra-tumoral heterogeneities: **tumorigenesis represents a process of progressive loss of original cell identity and acquirement of neural stemness in postnatal animal/human cells along the default route determined by evolution** (Cao, 2017; Cao, 2022) (Figure 2). Such a paradigm has been validated by increasing studies. For instances, loss of a transcriptional repressor causes transition of intestinal stem cells into NSC-like state and drives neuroendocrine tumor formation (Li et al., 2020); dedifferentiation of neuron into a neural stem-like state initiates tumorigenesis (Southall et al., 2014); Tuft cells transdifferentiate to neural-like progenitor cells during progression of pancreatic cancer (Salas-Escabillas et al., 2025); loss of muscle differentiation factor Myod1 leads to gain of neural stemness and tumorigenicity in myoblasts (Xu et al., 2021); neural-like dedifferentiation in cancer cells was observed during melanoma tumorigenesis (McGrail et al., 2025). Single cell RNA sequencing data also revealed dedifferentiation from the melanocytic toward the neural crest-like state during uveal melanoma progression (Xu et al., 2025), malignant cells with a neural crest-like state during gliomagenesis (Hamed et al., 2025), and neural cell state in different cancer cells (Pascual et al., 2021; Xing et al., 2025).

Cancer cells share not only regulatory networks with neural stem/embryonic neural cells, but also various phenotypic traits, such as single-cell migration (Cao, 2017; Cao, 2022; Zhang et al., 2017). Both are tumorigenic, defined by or dependent on activation of ancestral regulatory networks, and prone to genomic instability. Both exhibit neural stemness and pluripotent differentiation potential. Their metabolism is characterized by aerobic glycolysis. Both cancer genes and genes defining neural stemness are characteristic of over-representation of long genes with more exon/intron compositions that facilitate generation of more splice variants. Cancer cells are characteristic of neural stemness, which has a unicellular origin (Xu et al., 2021; Cao, 2022; Zhang et al., 2017). This agrees with that cancer cells are formed via a process of reverse evolution, i.e., multicellular to unicellular state, and cancer cells are characteristic of unicellular-like state (Alfarouk et al., 2011; Anatskaya et al., 2020; Chen et al., 2015; Vinogradov and Anatskaya, 2025) (Figure 2). Both cancer cells and neural stem cells are immune privileged cells. Embryonic pluripotent cells also share most of these features because the default fate of pluripotent cells is neural stemness and pluripotency has a unicellular origin (Cao, 2022).

2.2. Neural Stemness as the Source of Cell Tumorigenicity

Among different types of stem cells, ESCs and iPSCs are tumorigenic, but tissue stem cells, such as hematopoietic stem cells, mesenchymal stem cells, are not. NSCs were considered as a type of tissue stem cells and hence non-tumorigenic, although there were sporadic reports about tumorigenic potential of primitive NSCs derived from ESCs or iPSCs. It was routinely explained by incomplete change of ESCs or iPSCs into NSCs or by the expression of MYC oncoprotein in iPSCs (Deng et al., 2018; Germain et al., 2012). The standard in vivo tumorigenicity assay is xenograft tumor formation in immunodeficient mice. My studies demonstrated that either primitive NSCs derived from ESCs, NSCs from E9 mouse embryos, and neural progenitor cells isolated from cortices of E13.5 mouse embryos were able to form xenograft tumors in immunodeficient mice. By contrast, loss of neural stemness via differentiation of NSCs into neuronal cells leads to reduced tumorigenicity (Chen et al., 2021; Xu et al., 2021). Myoblast cells are not tumorigenic. Loss of Myod1 causes acquirement of neural stemness and tumorigenicity, and loss of neural stemness via differentiation into neuronal cells also causes reduced tumorigenicity (Xu et al., 2021). These are the direct evidence that neural stemness determines tumorigenicity of cells. ESCs in absence of TGF β signaling, which suppresses

tumorigenesis, adopt their default fate, i.e., the primitive NSCs. Correspondingly, ESCs are less tumorigenic than primitive NSCs (Xu et al., 2021). In combination with the evolutionary advantage of neural genes and neural stemness and the emergence of TGF β signaling at the start point of multicellularity during evolution, it can be concluded that neural stemness, but not other cellular properties/states, is the cellular property conferring tumorigenicity in cells (Cao, 2017; Cao, 2021; Xu et al., 2021).

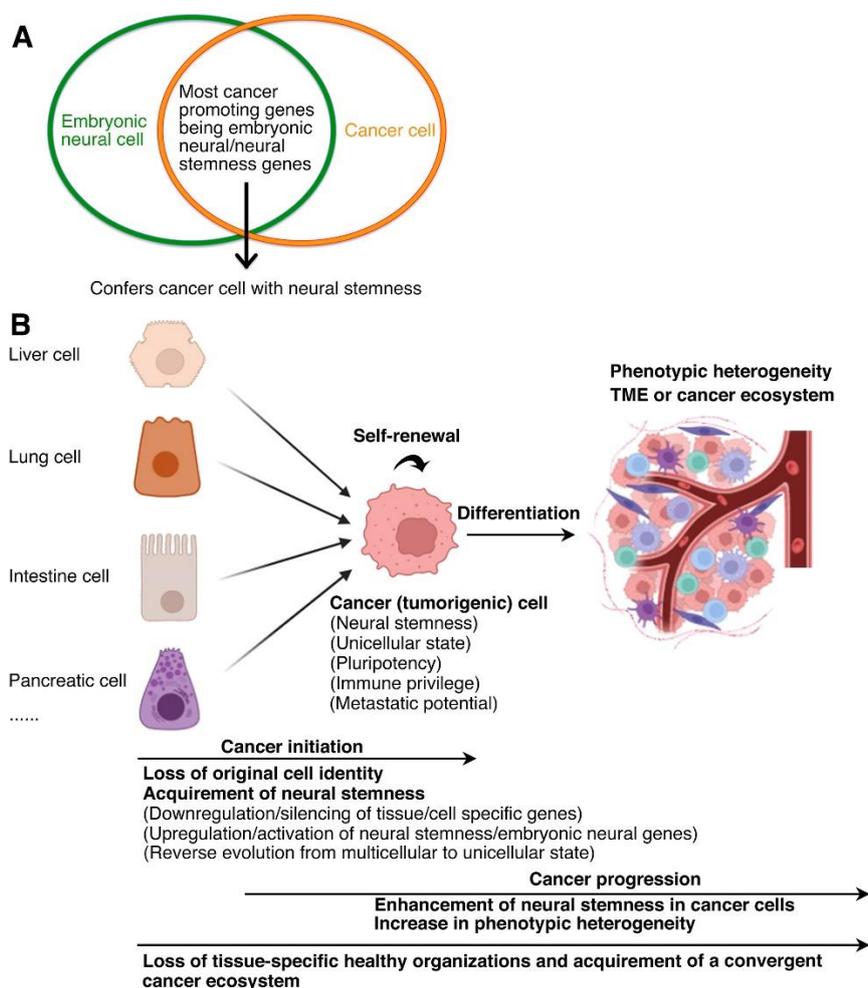


Figure 2. Neural stemness as the core property of cancer cell. (A) Most (if not all) cancer promoting genes are specifically expressed or enriched in embryonic neural cells, which confers cancer cell with neural stemness. (B) Tumorigenesis represents a process of progressive loss of original cell identity and acquirement of neural stemness. This is a process of reverse evolution from differentiated state to the unicellular state. Acquirement of neural stemness means acquiring pluripotent differentiation potential in cancer cell, which is capable of multilineage differentiation, leading to different types of cells that contribute at least partially to formation of tumor microenvironment (TME) or cancer ecosystem. Acquirement of neural stemness means acquiring immune privilege in cancer cell, which helps cancer cells to evade from immune killing. Acquirement of neural stemness also means acquiring metastatic potential. During cancer progression, there is a tendency of enhancement in neural stemness of cancer cells, hence the increase in differentiation potential or cell plasticity, and consequently, leading to increase in phenotypic heterogeneity. Regardless of the type of tissue of cancer origin, healthy organization of the tissue is disrupted. Meanwhile, neural stemness as the core property of cancer cells gives rise to convergence of cancer ecosystem formation. ((B) was created partially with Biorender.com).

As mentioned above, genes coding for the components of machineries required for basic cellular physiological functions and developmental programs, such as cell cycle, ribosome, spliceosome, proteasome, epigenetic modification, reprogramming, DNA damage and repair, are mostly enriched

in embryonic neural cells. These machineries are also enriched in cancer cells and play roles in promoting cancer. Cancer cells are characteristic of fast cell cycle/proliferation. They need more protein syntheses to sustain cell growth, more protein turnover program to maintain protein homeostasis; they need more DNA synthesis, and hence more DNA damage and repair mechanism; they undergo differentiation, hence they need more proteins involved in developmental programs, and so on. These machineries work concerted together to define neural stemness in neural stem/embryonic neural cells, and define tumorigenic property in cancer cells (Cao, 2021; Chen et al., 2021).

2.3. Pluripotency and Tumorigenicity: Two Sides of a Same Coin

Pluripotency and tumorigenicity are usually considered as distinct cellular properties because they are the most fundamental cellular properties dictating embryogenesis and tumorigenesis, separately. However, numerous studies on pluripotency have implied the intrinsic link between the two cellular properties. The earliest evidence was the pluripotent differentiation potential of embryonal carcinoma cells derived from teratocarcinoma, a type of cancer that can originate from many types of tissues/organs. After identification of embryonal carcinoma cells in more than a decade later, mouse ESCs were isolated and the property of ESCs were observed to be very comparable with those of embryonal carcinoma cells. Both cell types display pluripotent differentiation potential because they form teratomas in immunodeficient mice and contribute to formation of chimeric embryos (Solter, 2006). Teratoma formation in immunodeficient mice is a standard assay of pluripotency. Xenograft tumor formation performed in the same way is a standard assay of tumorigenicity. Histologically identifiable tissues/organs from different germ layers that are visible in teratomas formed by pluripotent cells, such as nerves, gut and glandular tissues, and cartilaginous tissues, are usually not visible in xenograft tumors formed by cancer cells. However, cell types that are derived from all three germ layers are present in the tumors (Xu et al., 2021; Zhang et al., 2022). This means that xenograft tumors are degenerated forms of teratomas. Besides teratocarcinoma cells, a variety of other cancer cells, including leukemia, neuroblastoma and melanoma cells, can contribute to chimeric formation or be induced to differentiate into different types of cells when transplanted into an embryo. The differentiated offspring cells are similar to host cells and not tumorigenic anymore (Brinster, 1974; Cooper and Pinkus, 1977; Gerschenson et al., 1986; Gootwine et al., 1982; Hendrix et al., 2007; Illmensee and Mintz, 1976; Kulesa et al., 2006; Papaioannou et al., 1975; Podesta et al., 1984; Webb et al., 1984; Wells and Miotto, 1986). Moreover, transplantation of the nuclei of different cancer cells into enucleated oocytes led to development of normal embryos (DiBerardino et al., 1983; Hochedlinger et al., 2004; Li et al., 2003; King and DiBerardino, 1965; McKinnell et al., 1969), suggesting the pluripotent nature of cancer cells. Characterization of cancer cells and NSCs demonstrates that variants of pluripotent state can be numerous and are present throughout the life of an animal/human, from a pre-implantation blastocyst to adult stage. **It can be seen that, historically, it was a type of cancer cells, the teratocarcinoma cell, that inspired the study of pluripotent property of ESCs. Now pluripotency is a key word in developmental/stem cell biology, but the pluripotent property of cancer cell has faded into oblivion.** Ironically, phenotypic alteration of cancer cells during tumorigenesis has been primarily understood with cell plasticity driven by 'EMT' (Bakir et al., 2020; Chaffer et al., 2016; Kalluri and Weinberg, 2009; Lu and Kang, 2019; Nieto et al., 2016), a poorly defined concept in which no basic scientific rationale can be found (Cao, 2024; Yang et al., 2020).

Pluripotency and tumorigenicity are both determined by neural stemness, implying that they are coupled cellular properties. Experimental evidence showed that it is the case. Blocking an endogenous factor, which promotes neural stemness and cancer, in neural stem cells and cancer cells led to a neuronal differentiation effect and loss of neural stemness. The resulting cells showed a simultaneous decrease in both tumorigenicity and pluripotency. Vice versa, enhancing neural stemness in cancer cells caused a simultaneous increase (Zhang et al., 2022). Pluripotency manifested by chimeric formation means that, in the presence of embryonic inducing factors, pluripotent cells,

including cancer cells, NSCs and embryonic pluripotent cells, can be induced to differentiate into normal cells in an embryonic milieu and integrated into the development of an embryo. In a postnatal animal/human, they differentiate and form tumor structures that cannot be integrated into normal differentiated tissues/organs because of no correct embryonic differentiation signals. The different behavior of pluripotent cells in embryonic milieu and in a postnatal animal/human suggests that tumorigenicity is actually the manifestation of pluripotent state in a postnatal animal/human. In summary, **pluripotency and tumorigenicity are both but different manifestations of the same cellular property, neural stemness, in embryonic and postnatal stage of animal/human life, respectively.**

2.4. Neural Stemness or General Stemness Represents Cancer Stemness

CSC is an important concept in cancer biology because it is believed that CSCs are capable of differentiation, thereby contributing to TME. However, properties of CSCs and their regulatory network have been poorly characterized. Cancer stemness is characteristic of some common features or hallmarks, including self-renewal and differentiation, multipotency, tumor initiation, immune evasion, etc. (Agudo and Miao, 2024; Loh and Ma, 2024). Critical questions still remain. Answers to these questions are essential for understanding the property of cancer cells, TME formation, and for development of novel strategies for cancer therapy. It cannot be determined or inferred from these criteria whether CSCs from different types of cancer share a common type of stemness, or CSCs of different cancer types exhibit the tissue stemness of their respective tissues of cancer origin, or CSCs are not comparable with any known stem/progenitor cell types (Cao, 2022). CSCs are isolated by using a few specific surface markers (Agudo and Miao, 2024). This would lead to pinpointing only a subset of CSC populations because of high heterogeneity of CSCs, and thus leading to misinterpretation of the properties of cancer stemness. Identification of CSCs also relies on approaches commonly used for identifying adult tissue stem cells. Moreover, CSCs are thought to be comparable with tissue stem cells because CSCs and some, but not all, tissue stem cells are immune privileged (Agudo and Miao, 2024). This implies that CSCs are characteristic of adult tissue stemness, a premise that has not been confirmed. If this is true, it can be deduced that CSCs of different types of cancer should have the differentiation potential similar to their respective tissue stem cells. However, there is no such evidence to show lineage-specific differentiation hierarchy of CSCs of a particular cancer (Cao, 2022; Cao, 2023). By contrast, CSCs show multi-lineage differentiation capacity, such as CSCs in colon cancer (Cao, 2022; Vermeulen et al., 2008). Vice versa, there is also no evidence to show that any type of adult tissue stemness reveals the features of tumor initiation, metastasis, or therapy resistance, which are the hallmarks of CSCs (Loh and Ma, 2024), and no evidence to show that CSCs share the regulatory networks with certain adult tissue stem cells. EMT explains that cancer stemness is a consequence of acquirement of mesenchymal state in cancer cells (Celià-Terrassa and Jolly, 2020; Dongre and Weinberg, 2019; Tanabe, 2022). It is hard to understand how an unknown and indefinable 'mesenchymal state' can help to understand cancer stemness (Cao, 2017; Cao, 2023; Cao, 2024; Yang et al., 2020). The analyses above revealed that cancer (tumorigenic) cells share regulatory networks, cellular properties and even evolutionary advantage with neural stem/embryonic neural cells, clarifying that neural stemness or general stemness, but not other types of stemness, represents cancer stemness. As cancer progresses, neural stemness of cancer cells will progressively increase, meaning the enhancement in differentiation potential or plasticity (Moorman et al., 2025; Zhang et al., 2022).

2.5. Neural Stemness Unifies Phenotypic Traits of Cancer Cells

Cancer cells display some phenotypic traits, such as stemness, high proliferation, invasion/migration, evasion of death and anti-cancer immunity, dysregulated metabolism and epigenetics, therapy resistance. Predominant research of cancer cell biology seems to be the regulation of a particular trait by a specific gene/factor, creating an impression that different traits of cancer cells are independent from each other. It should not be the case. Major cancer promoting

factors are best investigated for their roles in regulating cancer cell phenotypic traits. It is common to see that one factor is able to regulate different traits of cancer cells. For an instance, the epigenetic factor EZH2 plays oncogenic role in different types of cancers, promotes cancer cell stemness (Balinth et al., 2022), proliferation (Bryant et al., 2007), metastasis (Zingg et al., 2015), chemoresistance (Ougolkov et al., 2008; Crea et al., 2012), metabolism (Ahmad et al., 2017), etc. Of course, its upregulation in cancer cells leads to global change in epigenetic modification in genome and proteins. Similarly, studies on the oncoprotein C-MYC revealed that it regulates almost all traits of cancer cells (Dhanasekaran et al., 2022; Fatma et al., 2022; Llombart and Mansour, 2022). Cancer immunity and metabolism are prevailing research fields in cancer biology. Typical oncoproteins that play multiple roles in cancer are also major regulators of cancer cell metabolism and immunogenicity, e.g., EZH2 (Kim et al., 2020; Nylund et al., 2021; Zhou et al., 2020), C-MYC (Miller et al., 2012; Zimmerli et al., 2022), KRAS (Kerk et al., 2021; Watterson and Coelho, 2023; Lasse-Opsahl et al., 2025). A same factor being able to regulate different phenotypic traits of cancer cells suggests that these traits are intrinsically interconnected. SNAI1/2, ZEB2 and TWIST1 could serve as an additional example. These 'core EMT factors' have not been and cannot be clarified to be specific markers for 'mesenchymal state', because the cellular state is unknown and indefinable (Cao, 2024; Yang et al., 2020). Instead, they are markers for neural stem or neural crest cells, and their roles in regulating neural stemness are well documented. They were initially employed to explain single-cell migration of cancer cells in the context of the 'EMT concept'. In addition to their role in promoting cancer cell migration, they also regulate or are regulated by many other cancer promoting factors that are involved in regulation of almost all phenotypic traits of cancer cells, including stemness, proliferation, therapy resistance, metabolism, epigenetics, immune evasion (Cao, 2024). All these cancer promoting factors are components of embryonic neural regulatory network, which endows cancer cells with neural stemness (Cao, 2017; Cao, 2022; Xu et al., 2021; Zhang et al., 2017; Zhang et al., 2022). This means that different phenotypic traits of cancer cells are ultimately determined and coupled together by neural stemness and its corresponding regulatory networks, similar to the unification of tumorigenicity and pluripotency of cancer cells by neural stemness (Zhang et al., 2022). Disrupting one phenotypic trait will inevitably affect one or more, if not all, other traits of cancer cells.

2.6. Neural Stemness and Immune Privilege of Cancer Cells

Cancer cells are tumorigenic and immune privileged cells that are capable of immune evasion. How tumorigenicity and immune evasion are correlated was major topic to investigate. A relevant research focus on cancer immunotherapy is to find out ways to boost the sensitivity of cancer cells to anti-cancer immunity. Extensive studies on major oncoproteins have revealed their critical roles in promoting immune evasion and immunotherapy resistance of cancer cells via pairwise molecular regulatory mechanisms. Here are some examples. C-MYC suppresses STING-IFN signaling, thereby weakening immune cell infiltration in triple-negative breast cancer (Zimmerli et al., 2022); Inhibition of KRAS(G12D) induces FAS expression in cancer cells and facilitates CD8+ T cell-mediated death (Mahadevan et al., 2023); Inhibition of EZH2 upregulates MHC class I expression, leading to increase of antigen-specific CD8+ T-cell proliferation, IFN γ production, and tumor cell cytotoxicity (Zhou et al., 2020); Inhibition of CDK4/6 enhances T cell activation as a result of de-repression of NFAT family proteins (Deng et al, 2018); beta-catenin represses *CCL4* transcription, thus inhibiting anti-cancer immunity (Spranger et al., 2015); HDAC1 brakes anti-cancer immunity via suppression of type I dendritic cell maturation (De Sá Fernandes et al., 2024). These pairwise regulatory mechanisms might be countless because both cancer and immunity are complex. Still, "oncogenic signaling is the least understood aspect of functional immunogenicity" (Karasarides et al., 2022). The molecular mechanisms above appear nothing in common. When considering the fact that most cancer promoting proteins are embryonic neural proteins, including C-MYC, EZH2, KRAS, beta-catenin, CDK4/6, and HDAC1, it can be deduced that it is a cellular property, i.e., neural stemness, that is the ultimate factor to determine immune evasion. Cancer stemness is considered as a key factor driving

immune evasion and immunotherapy resistance (Galassi et al., 2021; Agudo and Miao, 2024). The link now becomes clear because neural stemness represents cancer stemness.

My latest research revealed that the link between neural stemness and cancer cell immune evasion should be understood according to principles of developmental biology (Liu et al., 2025). Neural stemness is determined by genes specifically expressed in or enriched in neural stem cells/embryonic neural cells, which are generally repressed or silenced in non-neural cells. Vice versa, genes specifying non-neural cells or maintaining their identities/functions, including immune related genes, are not highly expressed in embryonic neural cells. Interestingly, genes enhancing immunogenicity, including IFN- γ response genes and genes involved in antigen processing and presentation, are not or only weakly expressed in neural stem or embryonic stem cells, but they are highly expressed in immune cells and other non-neural cells, e.g., muscle and fat cells. The difference in expression of immune related genes is in agreement with that neural stem cells and embryonic stem cells, whose default fate is neural stem cells, are immune privileged, but other types of cells are not (Fändrich et al., 2002; Hori et al., 2003; Drukker et al., 2006; Magliocca et al., 2006; Itakura et al., 2017; Ozaki et al., 2017). This means that neural stemness endows cancer cells with tumorigenicity, pluripotency and other malignant features, and immune privilege as well. In general, most (if not all) oncoproteins are embryonic neural proteins and play roles in specifying neural stem/precursor cells and/or maintaining neural stemness. On one hand, they promote immune evasion by promoting the cancer regulatory network, i.e., the embryonic neural network, and on the other they are involved in repressing non-neural genes, including immune related genes in neural stem cells and cancer cells. Induced differentiation of cancer cells, either by forced expression of a lineage-specific differentiation factors or by inhibition of endogenous cancer promoting factors, led to reprogramming the regulatory networks of cancer cells into those of differentiated cells (Ascić et al., 2024; Linde et al., 2023; Liu et al., 2025; Zimmermannova et al., 2023), and reprogramming the cellular properties of cancer cells into the properties of differentiated cells with reduced neural stemness and tumorigenicity. Differentiation also generates the general tendency of decreased expression of cancer promoting genes, which promotes immune evasion, and increased expression of immune related genes including those enhancing immunogenicity, and enhancing immunogenicity of cancer cells (Ascić et al., 2024; Linde et al., 2023; Liu et al., 2025; Zimmermannova et al., 2023). In summary, neural stemness confers cancer cells with the capability of immune evasion. Tumorigenicity and immunogenicity are inversely correlated properties of cancer cells.

Analysis above revealed the shared characteristics between neural stem/progenitor cells and cancer cell, indicating that neural stemness determines phenotypic traits of cancer cells (Table 1).

Table 1. Comparison of the properties between neural stem/progenitor cells, cancer cells and pluripotent stem cells.

Neural stem/progenitor cells (References)	Cancer cells (References)	Pluripotent stem cell (PSCs) (References)
Tumorigenic (Xu et al., 2021)	Tumorigenic	Tumorigenic (Ben-David and Benvenisty, 2011)
Migratory	Migratory	Migratory
Immune privileged (Hori et al., 2003; Itakura et al., 2017; Ozaki et al., 2017)	Immune privileged	Immune privileged (Drukker et al., 2006; Fändrich et al., 2002; Magliocca et al., 2006;)
Defined by ancestral regulatory networks (Domazet-Lošo et al., 2007; Xu et al., 2021)	Dependent on activation of ancestral regulatory networks (Bussey et al., 2017; Domazet-Lošo and Tautz, 2010; Trigos et al., 2017; Trigos et al., 2018)	Unknown
Neural stemness	Neural stemness (Cao, 2017; Cao, 2022; Chen et al., 2021; Lei et al., 2019; Xu et al., 2021; Zhang et al., 2017; Zhang et al., 2022)	Neural stemness as the default state of PSC (Malaguti et al., 2013; Muñoz-Sanjuán and Brivanlou, 2002; Smukler et al.,

		2006; Tropepe et al., 2001; Ying et al., 2003)
Pluripotent differentiation potential (Clarke et al., 2000; Tropepe et al., 2001; Xu et al., 2021)	Pluripotent differentiation potential (Mintz and Illmensee, 1975; Papaioannou et al., 1975; Xu et al., 2021; Zhang et al., 2022)	Pluripotent differentiation potential
Characteristic of aerobic glycolysis. Differentiation into neurons decreases glycolysis (Kim et al., 2014; Zheng et al., 2016)	Characteristic of aerobic glycolysis	Characteristic of aerobic glycolysis. Turning into NSCs does not change or increases glycolysis; differentiation into mesoderm and endoderm decreases glycolysis (Intlekofer and Finley, 2019; Zheng et al., 2016)
Unicellular origin (Cao, 2022; Xu et al., 2021)	Resulting from loss of original cell identity and acquirement of neural stemness, and reverse evolution from multicellular to unicellular state (Alfarouk et al., 2011; Anatskaya et al., 2020; Bussey et al., 2017; Cao, 2022; Chen et al., 2015; Vinogradov and Anatskaya, 2025; Xu et al., 2021)	Unicellular origin of pluripotency (Sogabe et al., 2019)
Prone to genomic instability (Varela et al., 2012)	Genomic instability	Prone to genomic instability (Peterson and Loring, 2014)
Enriched in long genes with more splice variants (Gabel et al., 2015; Xu et al., 2021; Zylka et al., 2015)	Enriched in long genes with more splice variants (Sahakyan and Balasubramanian, 2016)	Unknown

3. Neural Stemness Unifies Embryogenesis and Tumorigenesis

Cancer was proposed as a disorder of developmental dynamics (Rubin, 1985). But how the complex process of embryogenesis is intrinsically linked with the complex process of tumorigenesis has remained a challenging question, particularly when considering that normal embryogenesis needs a process of fusion of gametes. The central role of neural stemness contributing to both pluripotency and tumorigenicity reminds of the intrinsic association between embryonic development and tumorigenesis. Before understanding the association, it needs to look back again on a paramount research work in developmental biology in history.

3.1. Embryonic Neural Induction, Body Axis Formation and Embryogenesis

A major question in developmental biology was to understand how the nervous system is induced to form and body axis is established during embryogenesis. The most inspiring work was done by Spemann and Mangold in 1924. They found that transplantation of the dorsal blastopore lip, which was named Spemann-Mangold organizer later, of a newt gastrula embryo to the ventral side of a host gastrula embryo was able to induce a complete secondary body axis or a conjoined twin. The secondary body axis contained neural tube, somites, pronephros and gut that were derived from the host, and the transplanted dorsal blastopore lip differentiated mostly into notochord (Spemann and Mangold, 1924; Spemann and Mangold, 2001). By contrast, an embryo without organizer activity forms only a 'belly piece' that contains no neural and dorsal structures (Spemann, 1938; Gerhart, 2001; De Robertis, 2009; Sosa et al., 2019). These experiments demonstrated the critical role of the organizer activity in inducing neural tissue and body axis during embryogenesis. The mechanisms underlying the induction of neural tissue and body axis by organizer began to be understood progressively until six decades after the dorsal blastopore transplantation experiment (De Robertis, 2009). It was concluded that neural fate is actually the default fate of blastula ectodermal cells, but epidermal fate is induced. The organizer promotes neural fate in ectoderm and dorsalization of primary germ layers by secreting a number of factors, such as Noggin, Chordin, and Cerberus, which inactivate the signaling pathways promoting epidermalization of ectoderm and ventralization of body axis, i.e.,

TGF β and Wnt signaling (Anderson and Stern, 2016; Bouwmeester et al., 1996; De Robertis, 2006; De Robertis and Kuroda, 2004; Sasai et al., 1994; Smith and Harland, 1992; Harland, 2000). Disruption of TGF β and Wnt signaling led to secondary axis formation, neuralization of ectoderm in absence of inducing factors, and rescue of ventralized embryos (Hemmati-Brivanlou and Melton, 1994; Glinka et al., 1997). Due to the epidermal inhibitory activity of the organizer, **neural induction during embryogenesis is a process of loss of epidermal fate and acquirement of the fate neuroectodermal cells, i.e., the primitive NSCs, in ectoderm along a default route determined by evolution.** The pluripotent neuroectodermal cells further contribute to not only formation of the nervous system, but also differentiation of non-neural cells during establishment of body axis.

Functional homologue known as the node has been identified in embryos of all classes of vertebrates, such as fish, birds and mammals. Similar to the organizer, the node displays the inducing activity for neural development and body axis formation through conserved molecular mechanisms (Gerhart, 2001; Martinez Arias and Steventon, 2018). Neural induction means activation or upregulation of a spectrum of neural genes, forming regulatory networks that define embryonic neural tissues. Thus, the effect of neural induction and formation of secondary axis or conjoined twin can also be mimicked by ectopic activation of genes that are enriched in embryonic neural cells. For example, ectopic activation of *eed*, *yy1*, *ski*, *egfr*, *erbb2*, *erbb4*, or *gsn* in *Xenopus* or zebrafish embryos causes formation of a partial secondary body axis that contains neural and non-neural tissues (Amaravadi et al., 1997; Kanungo et al., 2003; Nie and Chang, 2006; Satijn et al., 2001). On the contrary, disruption of embryonic neural genes causes defects in neural and axial differentiation in mouse embryos, ultimately leading to developmental arrest at early stages (Berk et al., 1997; Britsch et al., 1998; Donohoe et al., 1999; Faust et al., 1995; Gassmann et al., 1995; Threadgill et al., 1995). As a general rule, disruption of embryonic neural genes almost always causes severe disruption of embryonic development.

Neural induction, a process leading to the effect of acquirement of neural stemness in ectoderm, is the paradigm for understanding how neural tissue and body axis are initiated to form during early embryogenesis. Neural induction or a similar effect might aberrantly occur and be associated with some most complex pathological phenomena. Similar to the result of blastopore lip transplantation experiment, if a secondary organizer-like activity occurs erroneously in a gastrulating embryo, such as a human embryo, a conjoined twin will form (Levin, 1999). **Moreover, a neural induction-like process could also occur in cells of a postnatal animal or human, which would essentially lead to formation of a conjoined twin-like structure, i.e., a tumor.**

3.2. Neural Induction-Like Process During Tumorigenesis, and the Hallmarks of Tumorigenesis as a Conjoined Twin-Like Formation in a Postnatal animal/human

The proposal that tumorigenesis represents the process of progressive loss of original cell identity and gain of neural stemness in postnatal cells reminds of neural induction during embryogenesis, i.e., the loss of epidermal fate and gain of neural stemness in ectoderm, which can cause formation of a conjoined twin if it occurs ectopically. Acquirement of neural stemness in cells means acquirement of pluripotency and tumorigenicity. In the microenvironment in a postnatal animal/human, cancer cells undergo proliferation and differentiation. However, differentiated cells cannot integrate into normal tissues/organs because of lack of embryonic differentiation signals. Consequently, a conjoined twin-like structure, i.e., a tumor that contains various types of cells, is formed. A tumor being analogous to a conjoined twin-like structure can be best exemplified by teratocarcinomas, which can occur in many different types of tissues/organs. Teratocarcinomas are composed of tissues derived from all three germ layers, including undifferentiated neural epithelial tissue, differentiated nerves, gut and glandular tissues, cartilaginous and muscle tissues, similar to teratomas formed by pluripotent cells in immunodeficient mice. Unlike teratocarcinomas, other tumors that are diagnosed in most tissues/organs, e.g., lung, breast, colon cancer, usually do not contain well differentiated histologically identifiable tissues. But still, different types of cells that are derived from all three germ layers can be detected across tumors, for example, SOX1 or SOX2-

expressing cells representing cells with neural stemness and derived from ectoderm, ACTA2-expressing cells derived from mesoderm, and AFP-expressing cells derived from endoderm (Xu et al., 2021; Cao, 2022; Zhang et al., 2022). These cells can be detected across a wide range of cancer types. A similar trend can be found in public databases, which show that a majority of transcripts and their protein products have low cancer specificity (www.proteinatlas.org) (Uhlen et al., 2017), for example, BMI1, CDH2, DCLK1, FGFR4, MSI2, and SMARCA4 that indicate neural stemness; MAP2, NEUROG2, and TUBB3 that indicate neuronal differentiation; AFP, FOXA3, GATA6, and KRT8 that indicate endodermal tissue differentiation; and ACTA1, ACTA2, COL1A1, FXR1, and MEF2D that indicate mesodermal tissue differentiation (Cao, 2023). Olfactory neuroblastoma and small-cell lung cancer are originated from distinct organs. Nevertheless, they are similar in molecular heterogeneity and lineage trajectories (Finlay et al., 2024). Recent research on pan-cancer single-cell transcriptomic atlas demonstrated that, as tumor progresses, tissues gradually lose tissue-specific healthy organizations and acquire a convergent cancerous ecosystem (Shi et al., 2025), strengthening the commonality of components and component interactions across different tumors (Figure 2). The common theme of cell differentiation in different tumors reflects that they follow rules similar to embryonic development, and tumors (including teratocarcinoma) are degenerated forms of conjoined twin-like structures. In fact, tumorigenesis resembles an ectopic neural induction process, leading to gain of neural stemness and hence pluripotency in cells, which then differentiate long different lineages via interactions between tumor cells and interaction between tumor and host cells.

Tumors being comparable with a conjoined twin-like structure can be further supported by partial secondary axis (conjoined twin) formation resulting from ectopic activation of oncogenes in embryos, such as *eed*, *yy1*, *ski*, *egfr*, *erbb2*, *erbb4*, or *gsn* (Amaravadi et al., 1997; Kanungo et al., 2003; Nie and Chang, 2006; Satijn et al., 2001). Secondary axis formation was also observed when Hif-1 α was activated in early embryos (MacColl Garfinkel et al., 2023). Ectopic activation of β -catenin in embryos generates twinned embryo is a classical finding in developmental biology (Funayama et al., 1995; Kelly et al., 1995). Mutations leading to stabilization and nuclear accumulation of β -catenin in cells are usually the cause of some cancers, e.g., colorectal cancer. Correspondingly, these mutations are able to induce formation of a twinned mouse embryo (Brickman and Burdon, 2002). These results confirmed that genes or mutations leading to tumorigenesis can also induce formation of a conjoined twin in embryos, reinforcing the intrinsic link between neural induction, conjoined twin formation and tumorigenesis. To add more support, transplantation of cancer cells into the appropriate position of a blastula stage of zebrafish embryo can induce secondary body axis formation (Hendrix et al., 2007). Secondary body axis or conjoined twin formation involves most genes and developmental programs for cell differentiation and tissue/organ formation, and involves complex interactions between tissues/organs within the conjoined twin, and interactions with the primary body axis. A conjoined twin at postnatal stages should also contain microbes and experience physiological/pathological effects, such as aging, senescence, and inflammation. Long history of cancer research has revealed that most genes are associated with cancer (de Magalhães, 2022), and nearly all elements found in an animal/human, including aging, senescence, inflammation, and even microbes, can be found in cancer. Meanwhile, complex interactions or crosstalks within the tumor, and interactions between tumor and the host, have been extensively documented. This magnitude of complexity of cancer is only comparable to the complexity at the level of an organism.

The analysis above revealed the intrinsic association between embryonic neural induction and tumorigenesis. The former means the loss of epidermal fate and acquirement of neural stemness, hence the pluripotency in ectodermal cells, leading to neural development and body axis formation during embryogenesis. The latter means the loss of original cell identity and acquirement of neural stemness, hence the tumorigenicity in postnatal cells, leading to the formation of a tumor. Pluripotency and tumorigenicity are exchangeable cellular properties. Alignment of tumorigenesis with embryonic neural induction, hence conjoined twin formation, have been experimentally validated (Figure 3). **This paradigm, in which neural stemness functions as the cornerstone and the causal factor, can hopefully interpret the complexity of tumorigenesis and cancer ecosystem as a**

whole. It is interesting to note that the paramount work on embryonic induction by Spemann and Mangold was almost never mentioned in cancer research. Instead, the epigenetic landscape by Conrad Waddington, who was a great admirer of Spemann and Mangold's discovery of the principle of embryonic induction, has been frequently cited to interpret the link between tumorigenesis and embryonic development (Aranda-Anzaldo and Dent, 2018; Nicoglou, 2018).

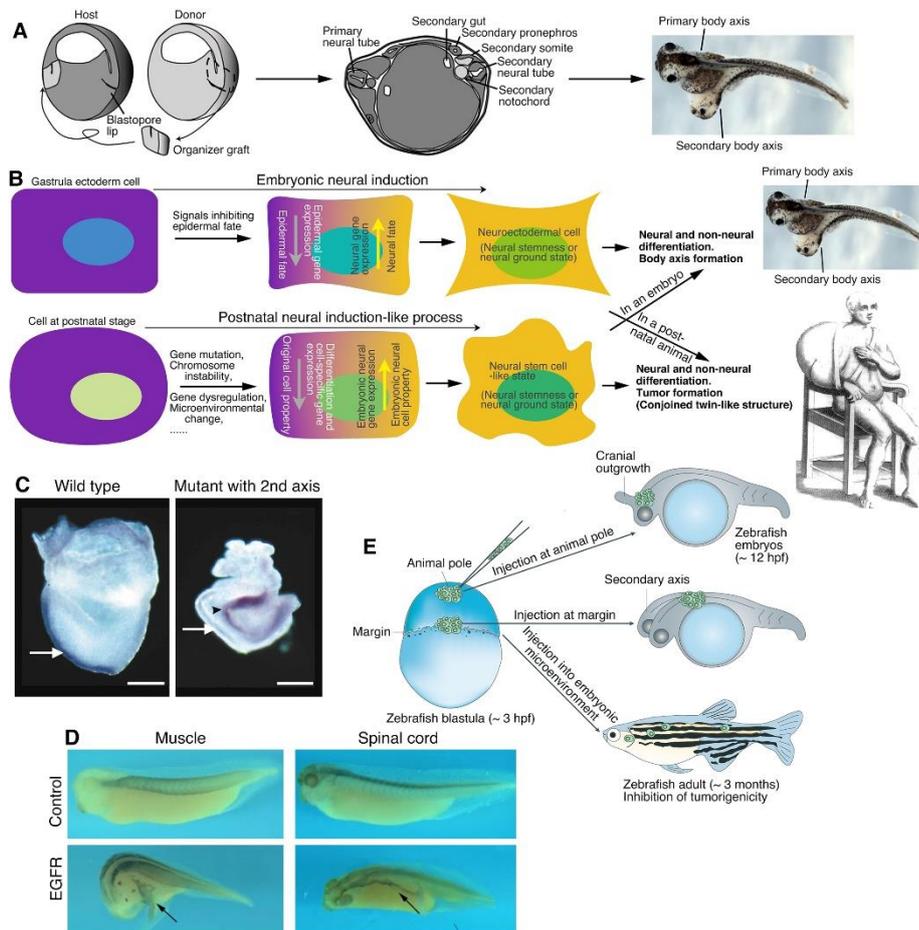


Figure 3. Neural stemness unifies embryogenesis and tumorigenesis. (A) (Left) A schematic illustration depicting the organizer grafting experiment done by Spemann and Mangold, in which the organizer (the dorsal blastopore lip) of an early gastrula of a light-gray newt (*Triturus cristatus*) was grafted to the site opposite to the dorsal lip of an early gastrula of a dark-gray newt (*Triturus taeniatus*). (Middle) An illustration showing secondary axis (or conjoined twin) formation by organizer grafting. A section through the trunk of a twinned embryo demonstrated that the graft contributed to the notochord, medial somites and floor plate of the secondary body axis, but the secondary neural tube, somites, pronephros, and archenteron cavity were induced from the host. (Right) Conjoined twin formation by Spemann-Mangold organizer grafting performed at early gastrula stage of frog (*Xenopus laevis*) embryo. (B) Acquisition of neural stemness is prerequisite for both embryogenesis and tumorigenesis. (Upper) Acquisition of neural stemness via embryonic neural induction and body axis formation. During embryonic development, the organizer or node secretes proteins inhibiting epidermal fate of gastrula ectoderm, leading to acquisition of neural fate in ectoderm and formation of neuroectoderm (neural stemness). The neural induction process is required for differentiation of both the nervous system and many non-neural tissues that are necessary for body axis formation. Ectopic neural induction or neural induction-like event in an embryo causes formation of secondary embryonic structures or a conjoined twin. (Lower) Acquisition of neural stemness via a neural induction-like process in postnatal animal/human and tumorigenesis. Cells of a postnatal animal/human may suffer various extracellular (e.g., microenvironmental change) and/or intracellular (e.g., gene mutations) insults, which could, occasionally or due to 'bad luck', cause upregulation/activation of embryonic neural genes or/and downregulation/silencing of

tissue-specific or differentiation genes. This results in progressive loss of original cell identity and acquirement of neural stemness along the default route pre-determined by evolution. The resulting cells are capable of self-renewal and differentiation into tissue/cell types of all three germ layers, and consequently, leading to formation of a conjoined twin-like structure, i.e., a tumor in a postnatal animal/human. The rightmost is the first clinical illustration of tumor, a large scapulohumeral tumor, in a book by a surgeon Marco Aurelio Severino published in 1632 (Hajdu, 2011). Embryonic pluripotent cells (ESCs, NSCs) and cancer cell are exchangeable. Both can contribute to chimera formation when put into an embryo but form xenograft tumors in a postnatal animal because of lack of embryonic inducing signals. (C-E) Either an oncogenic mutant, an oncogene or cancer cell is capable of inducing formation of a (partial) secondary body axis in embryos. (C) Mutation causing ectopic expression of β -catenin produces a twinned embryonic axis. The left is a normal E7.5 mouse embryo, the right is E8.5 embryo homozygous with respect to a mutation of axin ($axin^{T81}$ allele). White arrows indicate the primary axis and black arrowhead indicates the secondary axis. (D) Forced expression of EGFR in early frog (*Xenopus laevis*) embryos generates partial secondary axis, as indicated by black arrows, that contains ectopic neural and non-neural tissues. Embryos were analyzed with immunohistochemistry using antibodies against a muscle epitope and a neural epitope. (E) Injection of green fluorescence protein-labeled aggressive human melanoma cells into zebrafish embryos causes formation of cranial outgrowth when injected at the animal pole, or a secondary body axis when injected at the marginal zone, or contributes to chimera formation when injected into the embryonic microenvironment. hpf: hours post-fertilization. (A) is adapted from Harland (2008) (Harland, 2008), (B) is from Cao (2023), Hajdu (2011) and Harland (2008) (Cao, 2023; Harland 2008; Hajdu, 2011), (C) from Brickman and Burdon (2002) (Brickman and Burdon, 2002), (D) is from Nie and Chang (2006) (Nie and Chang, 2006), and (E) is from Hendrix et al. (2007) (Hendrix et al., 2007).

4. Important Issues to Consider or Re-Consider in Cancer Research

It's no doubt that great progresses have been achieved in cancer research, which deepens essentially the understanding of cancer and leads to development of innovative cancer therapies, such as immune checkpoint inhibition. But 'frustratingly, few of these innovative new therapeutic strategies are broadly beneficial across the spectrum of human cancers and, with many avenues for tumors to evolve resistance, even fewer significantly prolong overall survival' (Swanton et al., 2024). Development of efficient therapeutic strategies depends on the understanding of cancer cell. Nevertheless, scrutiny of some important 'concepts' that are used as a basic tool to measure phenotypic traits of cancer cell, such as 'EMT', revealed that they are just misconceptions (Cao, 2017; Cao, 2023; Cao, 2024; Yang et al., 2020). In the context of 'EMT', loss of epithelial state, e.g., by loss of the epithelial protein E-cadherin, will inevitably cause transition to mesenchymal state, thereby promoting metastasis of cancer cells. A latest study (and also other previous research) showed no 'EMT' but upregulation of embryonic neural factors GRPR and YAP1 after E-cadherin loss (Raymond et al., 2025). As a matter of fact, 'EMT factors/markers' that are upregulated/activated during tumorigenesis, including SNAI1/2, ZEB2, TWIST1, N-cadherin, Vimentin, are also embryonic neural factors, which are mistakenly labeled as mesenchymal proteins/markers (Cao, 2024). The consequence is the misattribution of the key role of neural stemness in determining phenotypic traits of cancer cells to the indefinable 'mesenchymal state'. 'EMT' and its related cellular state transitions as groundless and scientifically meaningless concepts was ever discussed in detail (Cao, 2017; Cao, 2023; Cao, 2024). Cancer cell is resulted from the change of cellular property and the change in expression of genes/proteins, and tumorigenesis is a highly flexible and dynamic process. Making correct association between gene/protein symbols and cellular state/property or making correct interpretation of expression of genes/proteins in cancer is critical for correct understanding cancer cell and tumor microenvironments. Wrong association or interpretation generates only misunderstanding and confusion.

As mentioned above, a same oncoprotein regulates different phenotypic traits of cancer cells. For example, it is frequently reported that inhibition of an oncoprotein in cancer cells leads to reduced tumorigenicity as shown by suppression of xenograft tumor formation in immunodeficient mice that lack T, NK and macrophage cells, and suppression of tumorigenesis. Detailed molecular mechanisms

are usually found to be responsible for such an effect. In separate studies, inhibition of the same oncoprotein in cancer cells causes enhanced immunogenicity or sensitivity to antitumor immunity as shown by suppression of tumor formation in syngeneic mice with functional immune cells, and thus suppression of tumorigenesis. The effect is also interpreted as the result of elegant but different molecular signaling cascades. Typically, blocking an oncoprotein de-represses a particular immune related gene that is involved in regulation of immune cell activity or infiltration in a tumor. If putting these studies together side by side, it is interesting to find that blocking an oncoprotein suppresses tumorigenesis no matter whether functional immune cells are present, raising a question what is the prime factor to consider, and whether it is appropriate to focus only on immunity when interpreting tumor suppression effect in response to blocking an oncoprotein. My recent results demonstrated that tumorigenicity and immunogenicity of cancer cells are inversely correlated with neural stemness. Induced differentiation of cancer cells causes loss of neural stemness and tumorigenicity, but enhancement of immunogenicity (Liu et al., 2025). The studies on differentiation of cancer cells into antigen-presenting cells analyzed their ability to promote anti-cancer immunity and underlying mechanisms in the context of immunity, but leaving tumorigenicity untested (Asic et al., 2024; Linde et al., 2023). However, it can be predicted that cancer cells after differentiation into immune cells will inevitably lose their tumorigenicity even in an environment of severely impaired immunity, as shown by another study (Zimmermannova et al., 2023). This effect should not be interpreted purely as a consequence of anti-tumor immunity.

Quite many theories/hypothesis have been proposed to explain cancer, including the somatic mutation theory, the ecosystem theory, the 'tissue organization field theory (TOFT)' (Bhattacharya et al., 2025; Huang et al., 2025). Somatic mutations are among the many factors that drive cancer initiation. There are also many cancers contain no consistent driver mutations, and tissues free of cancer are found to harbor oncogenic mutations (Huang et al., 2025). Meanwhile, the mechanisms governing cancer metastasis cannot be explained smoothly by mutations only. The importance of somatic mutation can be interpreted in the following way. Cancer is a disease of cellular property change: loss of original cell identity and acquirement of neural stemness. Therefore, whether a mutation (or generally, any intra-/extracellular insults a cell suffers) drives cancer depends on its ability to confer or promote neural stemness in cells. This is highly cellular context sensitive. A mutation driving cancer in one type of cells does not necessarily mean that it drives cancer in another because of the difference in intra-/extracellular regulatory signals. The ecosystem theory tries to interpret multiple cell populations, their complex interactions, and populations' evolvability by analogy with an ecosystem using ecological terminology (Aguadé-Gorgorió et al., 2024; Bhattacharya et al., 2025). The complex TME is indeed similar to an ecosystem to some extent. However, it seems that this analogy makes cancer TME, including concrete cell types, their interactions and evolving cell phenotypes, more abstract to understand. In a broader sense, any complex system can be considered as an ecosystem. Growth and differentiation of an embryo into an animal/human is also an good example of evolving ecosystem. As discussed earlier, the principle of neural induction during embryogenesis, which leads to generation of various types of cells/tissues, is also applicable to tumorigenesis. This might be helpful for understanding cancer ecosystem. TOFT was proposed to substitute for the somatic mutation theory. It emphasizes that 1) cancer is the disease resulting from tissue organization comparable to organogenesis, but not the disease of the cell, and 2) proliferation is the default state of all cells. Whether a cell becomes cancerous depends on the healthiness of the stroma of tissues/organs (Huang et al., 2025; Soto and Sonnenschein, 2005). But what tissue organization means remains to be clearly defined. Organogenesis of different organs is different processes and follows distinct hierarchy of differentiation and morphogenesis. This raises the question whether tumorigenesis resembles the organogenesis of the tissue/organ from which cancer is initiated. So far, there is no evidence to support the idea. By contrast, cancers with distinct origin are similar in molecular heterogeneity and lineage trajectories (Finlay et al., 2024). As tumor progresses, tissues gradually lose tissue-specific healthy organizations and acquire a convergent cancerous ecosystem (Shi et al., 2025). Additionally, there is also no evidence to support proliferation

as the default state of all cells. Although TOFT was proposed to explain cancer as “development gone awry” (Soto and Sonnenschein, 2005), it does not consider any key principles in developmental biology. Besides these concepts, some pathological/physiological processes, such as inflammation, aging, have been proposed explain cancer. The effectiveness of a theory/hypothesis does not depend on the similarity in some particular details, which are convenient to find because of the systemic complexity of cancer, but depend on the following considerations: 1) Whether it can clearly define cancer cell and cancer; 2) Whether it can be experimentally verified; 3) Whether it can explain cancer complexity as a whole, or integrate different aspects rather than specific details of cancer initiation and progression; 4) Whether it can be translated into cancer therapeutic strategies.

5. Neural Stemness Being the Core Property of Cancer Cell Paves the Road to Differentiation Therapy of Cancer

Differentiation therapy was suggested 50 years ago (Pierce and Wallace, 1971). Neural growth factors, all trans retinoic acid, arsenic trioxide, butyric acid or cAMP, showed some degree of differentiation-inducing capability in cancer cells. But differentiation therapy has been not applied as widely as other therapies. The best-known case of differentiation therapy might be the treatment of acute promyelocytic leukemia with all-trans retinoic acid (de Thé, 2018). The major obstacle should be the inappropriate understanding of the key property of cancer cells despite enormous studies. The core property of cancer cells being neural stemness, which endows cells with pluripotency, provides a general framework for differentiation therapy of different types of cancers: cancer cells can be induced to differentiate into different cell types by differentiation factors, particularly those driving embryonic tissue differentiation. A series of studies mainly in the 1970-80s demonstrated differentiation of cancer cells into benign cells within embryonic environment (Brinster, 1974; Cooper and Pinkus, 1977; Gerschenson et al., 1986; Gootwine et al., 1982; Hendrix et al., 2007; Illmensee and Mintz, 1976; Kulesa et al., 2006; Mintz and Illmensee, 1975; Papaioannou et al., 1975; Podesta et al., 1984; Webb et al., 1984; Wells and Miotto, 1986). Unfortunately, these results didn't get more attention later. Our studies have demonstrated that cancer cells can be induced to differentiate into neuronal-like cells by blocking endogenous oncoproteins, leading to loss of neural stemness and tumorigenicity in cancer cells (Chen et al., 2021; Lei et al., 2019; Zhang et al., 2017; Zhang et al., 2022). Embryonic non-neural lineage differentiation factors, GATA3, HNF4A, HHEX and FOXA3, were shown to suppress tumorigenicity via inhibition of neural stemness of cancer cells and suppress tumorigenesis in a mouse model of colorectal cancer (Yang et al., 2021). Immune cells are the key player in anti-cancer immunity. Differentiation of cancer cells into cells resembling immune cells was predicted based on cancer cell pluripotency (Cao, 2023). Indeed, in vitro and in vivo experimental data verified that cancer cells can be reprogrammed into antigen-presenting cells by lineage-specific specification factors. The resulting cells efficiently present tumor antigens and elicit systemic and safe antitumor immunity (Ascic et al., 2024; Linde et al., 2023; Zimmermann et al., 2023). Colorectal cancer cells can be induced to differentiate into normal-like enterocytes, thereby suppressing malignancy (Gong et al., 2025). Moreover, tumorigenicity and immune privilege of cancer cells are coupled together by neural stemness. Induced differentiation, either by forced expression of lineage differentiation factors or by blocking endogenous oncoproteins, leads to loss of neural stemness, reprograms cancer cells from a tumorigenic and immune privileged state into a non-tumorigenic and immunogenic state, and reprograms the regulatory networks of cancer cells into those of differentiated cells (Liu et al., 2025). Most cancer promoting proteins are neural stemness factors, targeted therapies via inhibition of cancer promoting factors are in essence differentiation therapies. The efficiency relies on the uniformity of presence of a target in cancer cells and the degree of differentiation of cancer cells in response to target inhibition. In addition, cancer with inhibition of a target gene/protein might be more prone to signal feedback loops in cancer regulatory networks, which will ultimately lead to therapy resistance. By contrast, targeting neural stemness via induced differentiation driven by embryonic non-neural differentiation factors does not depend on a specific target molecule, can achieve more efficient differentiation effect, and reprogram overall cellular properties and regulatory

networks of cancer cells. Such a strategy might avoid or alleviate therapy resistance effect elicited from complex signal feedback loops.

Conclusion

Cancer is a systemic disease. Understanding a systemic disease needs the understanding of systemic rules. In my opinion, cancer is the manifestation of the power of the rules operating in the animal kingdom: 1) The core property of cancer (tumorigenic) cell is neural stemness, which represents general stemness. Neural stemness unifies phenotypic traits of cancer cell, such as proliferation, metastasis, stemness, immune privilege; 2) Evolutionarily predetermined advantage of neural genes and unicellular origin of neural stemness determines and unifies pluripotent differentiation potential and tumorigenicity; 3) Pluripotency and tumorigenicity are both but different manifestations of the same cellular property, the neural stemness, during embryonic and postnatal stages of animal life, respectively; 4) Tumorigenicity is by nature the manifestation of aberrant occurrence of pluripotent state or neural stemness in a postnatal animal/human; 5) Tumorigenesis represents a process of progressive loss of original cell identity and acquirement of neural stemness; 6) Neural induction drives body axis formation during embryogenesis (and ectopic neural induction causes a conjoined twin), whereas a neural induction-like process drives tumorigenesis in postnatal animals/human. Therefore, tumors are degenerated forms of conjoined twin structures; 7) Induced differentiation of cancer cells by embryonic differentiation factors reprograms cellular property and regulatory network of cancer cells, leading to loss of neural stemness, tumorigenicity, and differentiation potential, and enhancement of sensitivity to anti-tumor immunity. I suggest understanding cancer as a systemic disease by understanding neural stemness as the core property of cancer cells and the principle of embryonic differentiation, and suggest developing cancer therapy by targeting neural stemness via efficient differentiation.

Acknowledgement: I would like to thank all who gave me support and encouragement during the research of such a topic.

Conflict of interest: None.

Author contributions: Y.C. conceived and wrote the review.

References

- Adamska M, Degnan SM, Green KM, Adamski M, Craigie A, Larroux C, Degnan BM. Wnt and TGF-beta expression in the sponge *Amphimedon queenslandica* and the origin of metazoan embryonic patterning. *PLoS One*. 2007 Oct 10;2(10):e1031.
- Aguadé-Gorgorió G, Anderson ARA, Solé R. Modeling tumors as complex ecosystems. *iScience*. 2024 Aug 10;27(9):110699.
- Agudo J, Miao Y. Stemness in solid malignancies: coping with immune attack. *Nat Rev Cancer*. 2025 Jan;25(1):27-40.
- Ahmad F, Patrick S, Sheikh T, Sharma V, Pathak P, Malgulwar PB, Kumar A, Joshi SD, Sarkar C, Sen E. Telomerase reverse transcriptase (TERT) - enhancer of zeste homolog 2 (EZH2) network regulates lipid metabolism and DNA damage responses in glioblastoma. *J Neurochem*. 2017 Dec;143(6):671-683.
- Alfarouk KO, Shayoub ME, Muddathir AK, Elhassan GO, Bashir AH. Evolution of Tumor Metabolism might Reflect Carcinogenesis as a Reverse Evolution process (Dismantling of Multicellularity). *Cancers (Basel)*. 2011 Jul 22;3(3):3002-17.
- Amaravadi LS, Neff AW, Sleeman JP, Smith RC. Autonomous neural axis formation by ectopic expression of the protooncogene *c-ski*. *Dev Biol*. 1997 Dec 15;192(2):392-404.
- Anatskaya OV, Vinogradov AE, Vainshelbaum NM, Giuliani A, Erenpreisa J. Phylostratic Shift of Whole-Genome Duplications in Normal Mammalian Tissues towards Unicellularity Is Driven by Developmental Bivalent Genes and Reveals a Link to Cancer. *Int J Mol Sci*. 2020 Nov 19;21(22):8759.
- Anderson C, Stern CD. Organizers in Development. *Curr Top Dev Biol*. 2016;117:435-54.

- Aranda-Anzaldo A, Dent MAR. Landscaping the epigenetic landscape of cancer. *Prog Biophys Mol Biol*. 2018 Dec;140:155-174.
- Ascić E, Åkerström F, Sreekumar Nair M, Rosa A, Kurochkin I, Zimmermannova O, Catena X, Rotankova N, Vesper C, Rudnik M, Ballocci T, Schärer T, Huang X, de Rosa Torres M, Renaud E, Velasco Santiago M, Met Ö, Askmyr D, Lindstedt M, Greiff L, Ligeon LA, Agarkova I, Svane IM, Pires CF, Rosa FF, Pereira CF. In vivo dendritic cell reprogramming for cancer immunotherapy. *Science*. 2024 Oct 18;386(6719):eadn9083.
- Bakir B, Chiarella AM, Pitarresi JR, Rustgi AK. EMT, MET, Plasticity, and Tumor Metastasis. *Trends Cell Biol*. 2020 Oct;30(10):764-776.
- Balint S, Fisher ML, Hwangbo Y, Wu C, Ballon C, Sun X, Mills AA. EZH2 regulates a SETDB1/ Δ Np63 α axis via RUNX3 to drive a cancer stem cell phenotype in squamous cell carcinoma. *Oncogene*. 2022 Aug;41(35):4130-4144.
- Baralle FE, Giudice J. Alternative splicing as a regulator of development and tissue identity. *Nat Rev Mol Cell Biol*. 2017 Jul;18(7):437-451.
- Ben-David U, Benvenisty N. The tumorigenicity of human embryonic and induced pluripotent stem cells. *Nat Rev Cancer*. 2011 Apr;11(4):268-77.
- Berk M, Desai SY, Heyman HC, Colmenares C. Mice lacking the ski proto-oncogene have defects in neurulation, craniofacial, patterning, and skeletal muscle development. *Genes Dev*. 1997 Aug 15;11(16):2029-39.
- Bhattacharya R, Avdieiev SS, Bukkuri A, Whelan CJ, Gatenby RA, Tsai KY, Brown JS. The Hallmarks of Cancer as Eco-Evolutionary Processes. *Cancer Discov*. 2025 Apr 2;15(4):685-701.
- Boumahdi S, Driessens G, Lapouge G, Rorive S, Nassar D, Le Mercier M, Delatte B, Caauwe A, Lenglez S, Nkusi E, Brohée S, Salmon I, Dubois C, del Marmol V, Fuks F, Beck B, Blanpain C. SOX2 controls tumour initiation and cancer stem-cell functions in squamous-cell carcinoma. *Nature*. 2014 Jul 10;511(7508):246-50.
- Bouwmeester T, Kim S, Sasai Y, Lu B, De Robertis EM. Cerberus is a head-inducing secreted factor expressed in the anterior endoderm of Spemann's organizer. *Nature*. 1996 Aug 15;382(6592):595-601.
- Brickman JM, Burdon TG. Pluripotency and tumorigenicity. *Nat Genet*. 2002 Dec;32(4):557-8.
- Brinster RL. The effect of cells transferred into the mouse blastocyst on subsequent development. *J Exp Med*. 1974 Oct 1;140(4):1049-56.
- Britsch S, Li L, Kirchoff S, Theuring F, Brinkmann V, Birchmeier C, Riethmacher D. The ErbB2 and ErbB3 receptors and their ligand, neuregulin-1, are essential for development of the sympathetic nervous system. *Genes Dev*. 1998 Jun 15;12(12):1825-36.
- Bryant RJ, Cross NA, Eaton CL, Hamdy FC, Cunliffe VT. EZH2 promotes proliferation and invasiveness of prostate cancer cells. *Prostate*. 2007 Apr 1;67(5):547-56.
- Burkhardt P, Stegmann CM, Cooper B, Kloepper TH, Imig C, Varoqueaux F, Wahl MC, Fasshauer D. Primordial neurosecretory apparatus identified in the choanoflagellate *Monosiga brevicollis*. *Proc Natl Acad Sci U S A*. 2011 Sep 13;108(37):15264-9.
- Bush SJ, Chen L, Tovar-Corona JM, Urrutia AO. Alternative splicing and the evolution of phenotypic novelty. *Philos Trans R Soc Lond B Biol Sci*. 2017 Feb 5;372(1713):20150474.
- Bussey KJ, Cisneros LH, Lineweaver CH, Davies PCW. Ancestral gene regulatory networks drive cancer. *Proc Natl Acad Sci U S A*. 2017 Jun 13;114(24):6160-6162.
- Cao Y. Tumorigenesis as a process of gradual loss of original cell identity and gain of properties of neural precursor/progenitor cells. *Cell Biosci*. 2017 Nov 7;7:61.
- Cao Y. Neural is Fundamental: Neural Stemness as the Ground State of Cell Tumorigenicity and Differentiation Potential. *Stem Cell Rev Rep*. 2022 Jan;18(1):37-55.
- Cao Y. Neural induction drives body axis formation during embryogenesis, but a neural induction-like process drives tumorigenesis in postnatal animals. *Front Cell Dev Biol*. 2023 May 9;11:1092667.
- Cao Y. Lack of basic rationale in epithelial-mesenchymal transition and its related concepts. *Cell Biosci*. 2024 Aug 20;14(1):104.
- Celià-Terrassa T, Jolly MK. Cancer Stem Cells and Epithelial-to-Mesenchymal Transition in Cancer Metastasis. *Cold Spring Harb Perspect Med*. 2020 Jul 1;10(7):a036905.
- Chaffer CL, San Juan BP, Lim E, Weinberg RA. EMT, cell plasticity and metastasis. *Cancer Metastasis Rev*. 2016 Dec;35(4):645-654.

- Chen H, Lin F, Xing K, He X. The reverse evolution from multicellularity to unicellularity during carcinogenesis. *Nat Commun.* 2015 Mar 9;6:6367.
- Chen L, Zhang M, Fang L, Yang X, Cao N, Xu L, Shi L, Cao Y. Coordinated regulation of the ribosome and proteasome by PRMT1 in the maintenance of neural stemness in cancer cells and neural stem cells. *J Biol Chem.* 2021 Nov;297(5):101275.
- Clarke DL, Johansson CB, Wilbertz J, Veress B, Nilsson E, Karlström H, Lendahl U, Frisén J. Generalized potential of adult neural stem cells. *Science.* 2000 Jun 2;288(5471):1660-3.
- Cooper M, Pinkus H. Intrauterine transplantation of rat basal cell carcinoma as a model for reconversion of malignant to benign growth. *Cancer Res.* 1977 Aug;37(8 Pt 1):2544-52.
- Crea F, Paolicchi E, Marquez VE, Danesi R. Polycomb genes and cancer: time for clinical application? *Crit Rev Oncol Hematol.* 2012 Aug;83(2):184-93.
- de Magalhães JP. Every gene can (and possibly will) be associated with cancer. *Trends Genet.* 2022 Mar;38(3):216-217.
- Deng J, Wang ES, Jenkins RW, Li S, Dries R, Yates K, Chhabra S, Huang W, Liu H, Aref AR, Ivanova E, Paweletz CP, Bowden M, Zhou CW, Herter-Sprie GS, Sorrentino JA, Bisi JE, Lizotte PH, Merlino AA, Quinn MM, Bufe LE, Yang A, Zhang Y, Zhang H, Gao P, Chen T, Cavanaugh ME, Rode AJ, Haines E, Roberts PJ, Strum JC, Richards WG, Lorch JH, Parangi S, Gunda V, Boland GM, Bueno R, Palakurthi S, Freeman GJ, Ritz J, Haining WN, Sharpless NE, Arthanari H, Shapiro GI, Barbie DA, Gray NS, Wong KK. CDK4/6 Inhibition Augments Antitumor Immunity by Enhancing T-cell Activation. *Cancer Discov.* 2018 Feb;8(2):216-233.
- Deng J, Zhang Y, Xie Y, Zhang L, Tang P. Cell Transplantation for Spinal Cord Injury: Tumorigenicity of Induced Pluripotent Stem Cell-Derived Neural Stem/Progenitor Cells. *Stem Cells Int.* 2018 Feb 4;2018:5653787.
- De Sá Fernandes C, Novoszel P, Gastaldi T, Krauß D, Lang M, Rica R, Kutschat AP, Holcman M, Ellmeier W, Seruggia D, Strobl H, Sibilia M. The histone deacetylase HDAC1 controls dendritic cell development and anti-tumor immunity. *Cell Rep.* 2024 Jun 25;43(6):114308.
- de Thé H. Differentiation therapy revisited. *Nat Rev Cancer.* 2018 Feb;18(2):117-127.
- De Robertis EM. Spemann's organizer and self-regulation in amphibian embryos. *Nat Rev Mol Cell Biol.* 2006 Apr;7(4):296-302.
- De Robertis EM. Spemann's organizer and the self-regulation of embryonic fields. *Mech Dev.* 2009 Dec;126(11-12):925-41.
- De Robertis EM, Kuroda H. Dorsal-ventral patterning and neural induction in *Xenopus* embryos. *Annu Rev Cell Dev Biol.* 2004;20:285-308.
- Dhanasekaran R, Deutzmann A, Mahauad-Fernandez WD, Hansen AS, Gouw AM, Felsher DW. The MYC oncogene - the grand orchestrator of cancer growth and immune evasion. *Nat Rev Clin Oncol.* 2022 Jan;19(1):23-36.
- DiBerardino MA, Mizell M, Hoffner NJ, Friesendorf DG. Frog larvae cloned from nuclei of pronephric adenocarcinoma. *Differentiation.* 1983;23(3):213-7.
- Domazet-Loso T, Brajković J, Tautz D. A phylostratigraphy approach to uncover the genomic history of major adaptations in metazoan lineages. *Trends Genet.* 2007 Nov;23(11):533-9.
- Domazet-Loso T, Tautz D. Phylostratigraphic tracking of cancer genes suggests a link to the emergence of multicellularity in metazoa. *BMC Biol.* 2010 May 21;8:66. doi: 10.1186/1741-7007-8-66.
- Dongre A, Weinberg RA. New insights into the mechanisms of epithelial-mesenchymal transition and implications for cancer. *Nat Rev Mol Cell Biol.* 2019 Feb;20(2):69-84.
- Donohoe ME, Zhang X, McGinnis L, Biggers J, Li E, Shi Y. Targeted disruption of mouse Yin Yang 1 transcription factor results in peri-implantation lethality. *Mol Cell Biol.* 1999 Oct;19(10):7237-44.
- Drukker M, Katchman H, Katz G, Even-Tov Friedman S, Shezen E, Hornstein E, Mandelboim O, Reisner Y, Benvenisty N. Human embryonic stem cells and their differentiated derivatives are less susceptible to immune rejection than adult cells. *Stem Cells.* 2006 Feb;24(2):221-9.
- Fändrich F, Dresske B, Bader M, Schulze M. Embryonic stem cells share immune-privileged features relevant for tolerance induction. *J Mol Med (Berl).* 2002 Jun;80(6):343-50.
- Fatma H, Maurya SK, Siddique HR. Epigenetic modifications of c-MYC: Role in cancer cell reprogramming, progression and chemoresistance. *Semin Cancer Biol.* 2022 Aug;83:166-176.

- Faust C, Schumacher A, Holdener B, Magnuson T. The eed mutation disrupts anterior mesoderm production in mice. *Development*. 1995 Feb;121(2):273-85.
- Finlay JB, Ireland AS, Hawgood SB, Reyes T, Ko T, Olsen RR, Abi Hachem R, Jang DW, Bell D, Chan JM, Goldstein BJ, Oliver TG. Olfactory neuroblastoma mimics molecular heterogeneity and lineage trajectories of small-cell lung cancer. *Cancer Cell*. 2024 Jun 10;42(6):1086-1105.e13.
- Fox RG, Lytle NK, Jaquish DV, Park FD, Ito T, Bajaj J, Koechlein CS, Zimdahl B, Yano M, Kopp J, Kritzik M, Sicklick J, Sander M, Grandgenett PM, Hollingsworth MA, Shibata S, Pizzo D, Valasek M, Sasik R, Scadeng M, Okano H, Kim Y, MacLeod AR, Lowy AM, Reya T. Image-based detection and targeting of therapy resistance in pancreatic adenocarcinoma. *Nature*. 2016 Jun 16;534(7607):407-411.
- Funayama N, Fagotto F, McCrean P, Gumbiner BM. Embryonic axis induction by the armadillo repeat domain of beta-catenin: evidence for intracellular signaling. *J Cell Biol*. 1995 Mar;128(5):959-68.
- Gabel HW, Kinde B, Stroud H, Gilbert CS, Harmin DA, Kastan NR, Hemberg M, Ebert DH, Greenberg ME. Disruption of DNA-methylation-dependent long gene repression in Rett syndrome. *Nature*. 2015 Jun 4;522(7554):89-93.
- Galassi C, Musella M, Manduca N, Maccafeio E, Sistigu A. The Immune Privilege of Cancer Stem Cells: A Key to Understanding Tumor Immune Escape and Therapy Failure. *Cells*. 2021 Sep 8;10(9):2361.
- Garner H, de Visser KE. Immune crosstalk in cancer progression and metastatic spread: a complex conversation. *Nat Rev Immunol*. 2020 Aug;20(8):483-497.
- Gassmann M, Casagrande F, Orioli D, Simon H, Lai C, Klein R, Lemke G. Aberrant neural and cardiac development in mice lacking the ErbB4 neuregulin receptor. *Nature*. 1995 Nov 23;378(6555):390-4.
- Gerhart J. Evolution of the organizer and the chordate body plan. *Int J Dev Biol*. 2001;45(1):133-53.
- Germain ND, Hartman NW, Cai C, Becker S, Naegele JR, Grabel LB. Teratocarcinoma formation in embryonic stem cell-derived neural progenitor hippocampal transplants. *Cell Transplant*. 2012;21(8):1603-11.
- Gerschenson M, Graves K, Carson SD, Wells RS, Pierce GB. Regulation of melanoma by the embryonic skin. *Proc Natl Acad Sci U S A*. 1986 Oct;83(19):7307-10.
- Glinka A, Wu W, Onichtchouk D, Blumenstock C, Niehrs C. Head induction by simultaneous repression of Bmp and Wnt signalling in *Xenopus*. *Nature*. 1997 Oct 2;389(6650):517-9.
- Godsave SF, Slack JM. Clonal analysis of mesoderm induction in *Xenopus laevis*. *Dev Biol*. 1989 Aug;134(2):486-90.
- Göhde R, Naumann B, Laundon D, Imig C, McDonald K, Cooper BH, Varoqueaux F, Fasshauer D, Burkhardt P. Choanoflagellates and the ancestry of neurosecretory vesicles. *Philos Trans R Soc Lond B Biol Sci*. 2021 Mar 29;376(1821):20190759.
- Gong JR, Lee CK, Kim HM, Kim J, Jeon J, Park S, Cho KH. Control of Cellular Differentiation Trajectories for Cancer Reversion. *Adv Sci (Weinh)*. 2025 Jan;12(3):e2402132.
- Gootwine E, Webb CG, Sachs L. Participation of myeloid leukaemic cells injected into embryos in haematopoietic differentiation in adult mice. *Nature*. 1982 Sep 2;299(5878):63-5.
- Grunz H, Tacke L. Neural differentiation of *Xenopus laevis* ectoderm takes place after disaggregation and delayed reaggregation without inducer. *Cell Differ Dev*. 1989 Dec;28(3):211-7.
- Hajdu SI. A note from history: landmarks in history of cancer, part 2. *Cancer*. 2011 Jun 15;117(12):2811-20.
- Hamed AA, Hua K, Trinh QM, Simons BD, Marioni JC, Stein LD, Dirks PB. Gliomagenesis mimics an injury response orchestrated by neural crest-like cells. *Nature*. 2025 Feb;638(8050):499-509.
- Harland R. Induction into the Hall of Fame: tracing the lineage of Spemann's organizer. *Development*. 2008 Oct;135(20):3321-3.
- Harland R. Neural induction. *Curr Opin Genet Dev*. 2000 Aug;10(4):357-62.
- Hemmati-Brivanlou A, Melton DA. Inhibition of activin receptor signaling promotes neuralization in *Xenopus*. *Cell*. 1994 Apr 22;77(2):273-81.
- Henrique D, Abranches E, Verrier L, Storey KG. Neuromesodermal progenitors and the making of the spinal cord. *Development*. 2015 Sep 1;142(17):2864-75.
- Hendrix MJ, Seftor EA, Seftor RE, Kasemeier-Kulesa J, Kulesa PM, Postovit LM. Reprogramming metastatic tumour cells with embryonic microenvironments. *Nat Rev Cancer*. 2007 Apr;7(4):246-55.

- Hori J, Ng TF, Shatos M, Klassen H, Streilein JW, Young MJ. Neural progenitor cells lack immunogenicity and resist destruction as allografts. *Stem Cells*. 2003;21(4):405-16.
- Hochedlinger K, Blelloch R, Brennan C, Yamada Y, Kim M, Chin L, Jaenisch R. Reprogramming of a melanoma genome by nuclear transplantation. *Genes Dev*. 2004 Aug 1;18(15):1875-85.
- Huang S, Soto AM, Sonnenschein C. The end of the genetic paradigm of cancer. *PLoS Biol*. 2025 Mar 18;23(3):e3003052.
- Illmensee K, Mintz B. Totipotency and normal differentiation of single teratocarcinoma cells cloned by injection into blastocysts. *Proc Natl Acad Sci U S A*. 1976 Feb;73(2):549-53.
- Intlekofer AM, Finley LWS. Metabolic signatures of cancer cells and stem cells. *Nat Metab*. 2019 Feb;1(2):177-188.
- Itakura G, Ozaki M, Nagoshi N, Kawabata S, Nishiyama Y, Sugai K, Iida T, Kashiwagi R, Ookubo T, Yastake K, Matsubayashi K, Kohyama J, Iwanami A, Matsumoto M, Nakamura M, Okano H. Low immunogenicity of mouse induced pluripotent stem cell-derived neural stem/progenitor cells. *Sci Rep*. 2017 Oct 11;7(1):12996.
- Itoh F, Watabe T, Miyazono K. Roles of TGF- β family signals in the fate determination of pluripotent stem cells. *Semin Cell Dev Biol*. 2014 Aug;32:98-106.
- Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest*. 2009 Jun;119(6):1420-8.
- Kanungo J, Kozmik Z, Swamynathan SK, Piatigorsky J. Gelsolin is a dorsalizing factor in zebrafish. *Proc Natl Acad Sci U S A*. 2003 Mar 18;100(6):3287-92.
- Karasarides M, Cogdill AP, Robbins PB, Bowden M, Burton EM, Butterfield LH, Cesano A, Hammer C, Haymaker CL, Horak CE, McGee HM, Monette A, Rudqvist NP, Spencer CN, Sweis RF, Vincent BG, Wennerberg E, Yuan J, Zappasodi R, Lucey VMH, Wells DK, LaVallee T. Hallmarks of Resistance to Immune-Checkpoint Inhibitors. *Cancer Immunol Res*. 2022 Apr 1;10(4):372-383.
- Kelly GM, Erezyilmaz DF, Moon RT. Induction of a secondary embryonic axis in zebrafish occurs following the overexpression of beta-catenin. *Mech Dev*. 1995 Oct;53(2):261-73.
- Kerk SA, Papagiannakopoulos T, Shah YM, Lyssiotis CA. Metabolic networks in mutant KRAS-driven tumours: tissue specificities and the microenvironment. *Nat Rev Cancer*. 2021 Aug;21(8):510-525.
- Kim DY, Rhee I, Paik J. Metabolic circuits in neural stem cells. *Cell Mol Life Sci*. 2014 Nov;71(21):4221-41.
- Kim HJ, Cantor H, Cosmopoulos K. Overcoming Immune Checkpoint Blockade Resistance via EZH2 Inhibition. *Trends Immunol*. 2020 Oct;41(10):948-963.
- King N, Westbrook MJ, Young SL, Kuo A, Abedin M, Chapman J, Fairclough S, Hellsten U, Isogai Y, Letunic I, Marr M, Pincus D, Putnam N, Rokas A, Wright KJ, Zuzow R, Dirks W, Good M, Goodstein D, Lemons D, Li W, Lyons JB, Morris A, Nichols S, Richter DJ, Salamov A, Sequencing JG, Bork P, Lim WA, Manning G, Miller WT, McGinnis W, Shapiro H, Tjian R, Grigoriev IV, Rokhsar D. The genome of the choanoflagellate *Monosiga brevicollis* and the origin of metazoans. *Nature*. 2008 Feb 14;451(7180):783-8.
- King TJ, DiBerardino MA. Transplantation of nuclei from the frog renal adenocarcinoma. I. Development of tumor nuclear-transplant embryos. *Ann N Y Acad Sci*. 1965 Aug 10;126(1):115-26.
- Knecht AK, Bronner-Fraser M. Induction of the neural crest: a multigene process. *Nat Rev Genet*. 2002 Jun;3(6):453-61.
- Kulesa PM, Kasemeier-Kulesa JC, Teddy JM, Margaryan NV, Seftor EA, Seftor RE, Hendrix MJ. Reprogramming metastatic melanoma cells to assume a neural crest cell-like phenotype in an embryonic microenvironment. *Proc Natl Acad Sci U S A*. 2006 Mar 7;103(10):3752-7.
- Lasse-Opsahl EL, Barravecchia I, McLintock E, Lee JM, Ferris SF, Espinoza CE, Hinshaw R, Cavanaugh S, Robotti M, Rober L, Brown K, Abdelmalak KY, Galban CJ, Frankel TL, Zhang Y, Pasca di Magliano M, Galban S. KRASG12D drives immunosuppression in lung adenocarcinoma through paracrine signaling. *JCI Insight*. 2025 Jan 9;10(1):e182228.
- Le Douarin NM, Dupin E. The Pluripotency of Neural Crest Cells and Their Role in Brain Development. *Curr Top Dev Biol*. 2016;116:659-78.
- Lei A, Chen L, Zhang M, Yang X, Xu L, Cao N, Zhang Z, Cao Y. EZH2 Regulates Protein Stability via Recruiting USP7 to Mediate Neuronal Gene Expression in Cancer Cells. *Front Genet*. 2019 May 3;10:422.
- Levin M. Twinning and embryonic left-right asymmetry. *Laterality*. 1999 Jul;4(3):197-208.

- Li L, Connelly MC, Wetmore C, Curran T, Morgan JI. Mouse embryos cloned from brain tumors. *Cancer Res.* 2003 Jun 1;63(11):2733-6.
- Li Z, Guo X, Huang H, Wang C, Yang F, Zhang Y, Wang J, Han L, Jin Z, Cai T, Xi R. A Switch in Tissue Stem Cell Identity Causes Neuroendocrine Tumors in *Drosophila* Gut. *Cell Rep.* 2020 Feb 11;30(6):1724-1734.e4.
- Linde MH, Fan AC, Köhnke T, Trotman-Grant AC, Gurev SF, Phan P, Zhao F, Haddock NL, Nuno KA, Gars EJ, Stafford M, Marshall PL, Dove CG, Linde IL, Landberg N, Miller LP, Majzner RG, Zhang TY, Majeti R. Reprogramming Cancer into Antigen-Presenting Cells as a Novel Immunotherapy. *Cancer Discov.* 2023 May 4;13(5):1164-1185.
- Liu Y, Wang C, Li J, Cao Y. Differentiation status determines tumorigenicity and immunogenicity of cancer cells. *bioRxiv* 2025.05.30.656250; doi: <https://doi.org/10.1101/2025.05.30.656250>
- Llombart V, Mansour MR. Therapeutic targeting of "undruggable" MYC. *EBioMedicine.* 2022 Jan;75:103756.
- Loh JJ, Ma S. Hallmarks of cancer stemness. *Cell Stem Cell.* 2024 May 2;31(5):617-639.
- Lu W, Kang Y. Epithelial-Mesenchymal Plasticity in Cancer Progression and Metastasis. *Dev Cell.* 2019 May 6;49(3):361-374.
- MacColl Garfinkel A, Mnatsakanyan N, Patel JH, Wills AE, Shteyman A, Smith PJS, Alavian KN, Jonas EA, Khokha MK. Mitochondrial leak metabolism induces the Spemann-Mangold Organizer via Hif-1 α in *Xenopus*. *Dev Cell.* 2023 Nov 20;58(22):2597-2613.e4.
- Magliocca JF, Held IK, Odorico JS. Undifferentiated murine embryonic stem cells cannot induce portal tolerance but may possess immune privilege secondary to reduced major histocompatibility complex antigen expression. *Stem Cells Dev.* 2006 Oct;15(5):707-17.
- Mahadevan KK, McAndrews KM, LeBleu VS, Yang S, Lyu H, Li B, Sockwell AM, Kirtley ML, Morse SJ, Moreno Diaz BA, Kim MP, Feng N, Lopez AM, Guerrero PA, Paradiso F, Sugimoto H, Arian KA, Ying H, Barekatin Y, Sthanam LK, Kelly PJ, Maitra A, Heffernan TP, Kalluri R. KRASG12D inhibition reprograms the microenvironment of early and advanced pancreatic cancer to promote FAS-mediated killing by CD8 $^{+}$ T cells. *Cancer Cell.* 2023 Sep 11;41(9):1606-1620.e8.
- Malaguti M, Nistor PA, Blin G, Pegg A, Zhou X, Lowell S. Bone morphogenic protein signalling suppresses differentiation of pluripotent cells by maintaining expression of E-Cadherin. *Elife.* 2013 Dec 17;2:e01197.
- Maman S, Witz IP. A history of exploring cancer in context. *Nat Rev Cancer.* 2018 Jun;18(6):359-376.
- Martinez Arias A, Steventon B. On the nature and function of organizers. *Development.* 2018 Mar 9;145(5):dev159525.
- McGrail K, González-Sánchez E, Granado-Martínez P, Orsenigo R, Ding Y, Ferrer B, Hernández-Losa J, Ortega I, Martín-Caballero J, Muñoz-Couselo E, García-Patos V, Recio JA. Loss of Lkb1 cooperates with BrafV600E and ultraviolet radiation, increasing melanoma multiplicity and neural-like dedifferentiation. *Mol Oncol.* 2025 Feb;19(2):329-343.
- Meacham CE, Morrison SJ. Tumour heterogeneity and cancer cell plasticity. *Nature.* 2013 Sep 19;501(7467):328-37.
- Merlo LM, Pepper JW, Reid BJ, Maley CC. Cancer as an evolutionary and ecological process. *Nat Rev Cancer.* 2006 Dec;6(12):924-35.
- Meyers EA, Kessler JA. TGF- β Family Signaling in Neural and Neuronal Differentiation, Development, and Function. *Cold Spring Harb Perspect Biol.* 2017 Aug 1;9(8):a022244.
- Miller DM, Thomas SD, Islam A, Muench D, Sedoris K. c-Myc and cancer metabolism. *Clin Cancer Res.* 2012 Oct 15;18(20):5546-53.
- Mintz B, Illmensee K. Normal genetically mosaic mice produced from malignant teratocarcinoma cells. *Proc Natl Acad Sci U S A.* 1975 Sep;72(9):3585-9.
- Moorman A, Benitez EK, Cambulli F, Jiang Q, Mahmoud A, Lumish M, Hartner S, Balkaran S, Bermeo J, Asawa S, Firat C, Saxena A, Wu F, Luthra A, Burdzyak C, Xie Y, Sgambati V, Lockett K, Li Y, Yi Z, Masilionis I, Soares K, Pappou E, Yaeger R, Kingham TP, Jarnagin W, Paty PB, Weiser MR, Mazutis L, D'Angelica M, Shia J, Garcia-Aguilar J, Nawy T, Hollmann TJ, Chaligné R, Sanchez-Vega F, Sharma R, Pe'er D, Ganesh K. Progressive plasticity during colorectal cancer metastasis. *Nature.* 2025 Jan;637(8047):947-954.
- Muñoz-Sanjuán I, Brivanlou AH. Neural induction, the default model and embryonic stem cells. *Nat Rev Neurosci.* 2002 Apr;3(4):271-80.

- Nakanishi Y, Seno H, Fukuoka A, Ueo T, Yamaga Y, Maruno T, Nakanishi N, Kanda K, Komekado H, Kawada M, Isomura A, Kawada K, Sakai Y, Yanagita M, Kageyama R, Kawaguchi Y, Taketo MM, Yonehara S, Chiba T. Dcl1 distinguishes between tumor and normal stem cells in the intestine. *Nat Genet.* 2013 Jan;45(1):98-103.
- Nicoglou A. Waddington's epigenetics or the pictorial meetings of development and genetics. *Hist Philos Life Sci.* 2018 Sep 27;40(4):61.
- Nie S, Chang C. Regulation of early *Xenopus* development by ErbB signaling. *Dev Dyn.* 2006 Feb;235(2):301-14.
- Nieto MA, Huang RY, Jackson RA, Thiery JP. EMT: 2016. *Cell.* 2016 Jun 30;166(1):21-45.
- Nylund P, Atienza Párraga A, Haglöf J, De Bruyne E, Menu E, Garrido-Zabala B, Ma A, Jin J, Öberg F, Vanderkerken K, Kalushkova A, Jernberg-Wiklund H. A distinct metabolic response characterizes sensitivity to EZH2 inhibition in multiple myeloma. *Cell Death Dis.* 2021 Feb 12;12(2):167.
- Ougolkov AV, Bilim VN, Billadeau DD. Regulation of pancreatic tumor cell proliferation and chemoresistance by the histone methyltransferase enhancer of zeste homologue 2. *Clin Cancer Res.* 2008 Nov 1;14(21):6790-6.
- Ozair MZ, Kintner C, Brivanlou AH. Neural induction and early patterning in vertebrates. *Wiley Interdiscip Rev Dev Biol.* 2013 Jul;2(4):479-98.
- Ozaki M, Iwanami A, Nagoshi N, Kohyama J, Itakura G, Iwai H, Nishimura S, Nishiyama Y, Kawabata S, Sugai K, Iida T, Matsubayashi K, Isoda M, Kashiwagi R, Toyama Y, Matsumoto M, Okano H, Nakamura M. Evaluation of the immunogenicity of human iPS cell-derived neural stem/progenitor cells in vitro. *Stem Cell Res.* 2017 Mar;19:128-138.
- Pajanoja C, Hsin J, Olinger B, Schiffmacher A, Yazejian R, Abrams S, Dapkunas A, Zainul Z, Doyle AD, Martin D, Kerosuo L. Maintenance of pluripotency-like signature in the entire ectoderm leads to neural crest stem cell potential. *Nat Commun.* 2023 Sep 23;14(1):5941.
- Papioannou VE, McBurney MW, Gardner RL, Evans MJ. Fate of teratocarcinoma cells injected into early mouse embryos. *Nature.* 1975 Nov 6;258(5530):70-73.
- Pascual G, Domínguez D, Elosúa-Bayes M, Beckedorff F, Laudanna C, Bigas C, Douillet D, Greco C, Symeonidi A, Hernández I, Gil SR, Prats N, Bescós C, Shiekhatar R, Amit M, Heyn H, Shilatifard A, Benitah SA. Dietary palmitic acid promotes a prometastatic memory via Schwann cells. *Nature.* 2021 Nov;599(7885):485-490.
- Peterson SE, Loring JF. Genomic instability in pluripotent stem cells: implications for clinical applications. *J Biol Chem.* 2014 Feb 21;289(8):4578-84.
- Pierce GB, Wallace C. Differentiation of malignant to benign cells. *Cancer Res.* 1971 Feb;31(2):127-34.
- Pla P, Monsoro-Burq AH. The neural border: Induction, specification and maturation of the territory that generates neural crest cells. *Dev Biol.* 2018 Dec 1;444 Suppl 1:S36-S46.
- Podesta AH, Mullins J, Pierce GB, Wells RS. The neurula stage mouse embryo in control of neuroblastoma. *Proc Natl Acad Sci U S A.* 1984 Dec;81(23):7608-11.
- Raymond JH, Aktary Z, Pouteaux M, Petit V, Luciani F, Wehbe M, Gizzi P, Bourban C, Decaudin D, Nemati F, Martianov I, Davidson I, Tomasetto CL, White RM, Mahuteau-Betzer F, Vergier B, Larue L, Delmas V. Targeting GRPR for sex hormone-dependent cancer after loss of E-cadherin. *Nature.* 2025 Jul;643(8072):801-809.
- Rubin H. Cancer as a dynamic developmental disorder. *Cancer Res.* 1985 Jul;45(7):2935-42.
- Ricci-Vitiani L, Lombardi DG, Pilozzi E, Biffoni M, Todaro M, Peschle C, De Maria R. Identification and expansion of human colon-cancer-initiating cells. *Nature.* 2007 Jan 4;445(7123):111-5.
- Sahakyan AB, Balasubramanian S. Long genes and genes with multiple splice variants are enriched in pathways linked to cancer and other multigenic diseases. *BMC Genomics.* 2016 Mar 12;17:225.
- Salas-Escabillas DJ, Hoffman MT, Brender SM, Moore JS, Wen HJ, Benitz S, Davis ET, Long D, Wombwell AM, Chianis ERD, Allen-Petersen BL, Steele NG, Sears RC, Matsumoto I, DelGiorno KE, Crawford HC. Tuft cells transdifferentiate to neural-like progenitor cells in the progression of pancreatic cancer. *Dev Cell.* 2025 Mar 24;60(6):837-852.e3.
- Sambasivan R, Steventon B. Neuromesodermal Progenitors: A Basis for Robust Axial Patterning in Development and Evolution. *Front Cell Dev Biol.* 2021 Jan 15;8:607516.

- Sasai Y, Lu B, Steinbeisser H, Geissert D, Gont LK, De Robertis EM. Xenopus chordin: a novel dorsalizing factor activated by organizer-specific homeobox genes. *Cell*. 1994 Dec 2;79(5):779-90.
- Satijn DP, Hamer KM, den Blaauwen J, Otte AP. The polycomb group protein EED interacts with YY1, and both proteins induce neural tissue in *Xenopus* embryos. *Mol Cell Biol*. 2001 Feb;21(4):1360-9.
- Sato SM, Sargent TD. Development of neural inducing capacity in dissociated *Xenopus* embryos. *Dev Biol*. 1989 Jul;134(1):263-6.
- Selleck MA, Bronner-Fraser M. Origins of the avian neural crest: the role of neural plate-epidermal interactions. *Development*. 1995 Feb;121(2):525-38.
- Shi Q, Chen Y, Li Y, Qin S, Yang Y, Gao Y, Zhu L, Wang D, Zhang Z. Cross-tissue multicellular coordination and its rewiring in cancer. *Nature*. 2025 Jul;643(8071):529-538.
- Smith WC, Harland RM. Expression cloning of noggin, a new dorsalizing factor localized to the Spemann organizer in *Xenopus* embryos. *Cell*. 1992 Sep 4;70(5):829-40.
- Smukler SR, Runciman SB, Xu S, van der Kooy D. Embryonic stem cells assume a primitive neural stem cell fate in the absence of extrinsic influences. *J Cell Biol*. 2006 Jan 2;172(1):79-90.
- Sogabe S, Hatleberg WL, Kocot KM, Say TE, Stoupin D, Roper KE, Fernandez-Valverde SL, Degnan SM, Degnan BM. Pluripotency and the origin of animal multicellularity. *Nature*. 2019 Jun;570(7762):519-522.
- Solter D. From teratocarcinomas to embryonic stem cells and beyond: a history of embryonic stem cell research. *Nat Rev Genet*. 2006 Apr;7(4):319-27.
- Sosa EA, Moriyama Y, Ding Y, Tejada-Muñoz N, Colozza G, De Robertis EM. Transcriptome analysis of regeneration during *Xenopus laevis* experimental twinning. *Int J Dev Biol*. 2019;63(6-7):301-309.
- Soto AM, Sonnenschein C. Emergentism as a default: cancer as a problem of tissue organization. *J Biosci*. 2005 Feb;30(1):103-18.
- Southall TD, Davidson CM, Miller C, Carr A, Brand AH. Dedifferentiation of neurons precedes tumor formation in *Lola* mutants. *Dev Cell*. 2014 Mar 31;28(6):685-96.
- Spemann, H. (1938). *Embryonic development and induction*. New Haven: Yale University Press, 401.
- Spemann, H., and Mangold, H. (2001). Induction of embryonic primordia by implantation of organizers from a different species. *Int. J. Dev. Biol.* 45, 13–38.
- Spemann, H., and Mangold, H. (1924). Über Induktion von Embryonalanlagen durch Implantation artfremder Organisatoren. *Arch. Mikrosk. Anat. EntwMech.* 100, 599–638. doi:10.1007/bf02108133
- Spranger S, Bao R, Gajewski TF. Melanoma-intrinsic β -catenin signalling prevents anti-tumour immunity. *Nature*. 2015 Jul 9;523(7559):231-5.
- Srivastava M, Simakov O, Chapman J, Fahey B, Gauthier ME, Mitros T, Richards GS, Conaco C, Dacre M, Hellsten U, Larroux C, Putnam NH, Stanke M, Adamska M, Darling A, Degnan SM, Oakley TH, Plachetzki DC, Zhai Y, Adamski M, Calcino A, Cummins SF, Goodstein DM, Harris C, Jackson DJ, Leys SP, Shu S, Woodcroft BJ, Vervoort M, Kosik KS, Manning G, Degnan BM, Rokhsar DS. The Amphimedon queenslandica genome and the evolution of animal complexity. *Nature*. 2010 Aug 5;466(7307):720-6.
- Stanger BZ, Wahl GM. Cancer as a Disease of Development Gone Awry. *Annu Rev Pathol*. 2024 Jan 24;19:397-421.
- Swanton C, Bernard E, Abbosh C, André F, Auwerx J, Balmain A, Bar-Sagi D, Bernards R, Bullman S, DeGregori J, Elliott C, Erez A, Evan G, Febbraio MA, Hidalgo A, Jamal-Hanjani M, Joyce JA, Kaiser M, Lamia K, Locasale JW, Loi S, Malanchi I, Merad M, Musgrave K, Patel KJ, Quezada S, Wargo JA, Weeraratna A, White E, Winkler F, Wood JN, Vousden KH, Hanahan D. Embracing cancer complexity: Hallmarks of systemic disease. *Cell*. 2024 Mar 28;187(7):1589-1616.
- Tanabe S. Epithelial-Mesenchymal Transition and Cancer Stem Cells. *Adv Exp Med Biol*. 2022;1393:1-49.
- Threadgill DW, Dlugosz AA, Hansen LA, Tennenbaum T, Lichti U, Yee D, LaMantia C, Mourton T, Herrup K, Harris RC, et al. Targeted disruption of mouse EGF receptor: effect of genetic background on mutant phenotype. *Science*. 1995 Jul 14;269(5221):230-4.
- Tong Y, Gao WQ, Liu Y. Metabolic heterogeneity in cancer: An overview and therapeutic implications. *Biochim Biophys Acta Rev Cancer*. 2020 Dec;1874(2):188421.

- Trigos AS, Pearson RB, Papenfuss AT, Goode DL. Altered interactions between unicellular and multicellular genes drive hallmarks of transformation in a diverse range of solid tumors. *Proc Natl Acad Sci U S A*. 2017 Jun 13;114(24):6406-6411.
- Trigos AS, Pearson RB, Papenfuss AT, Goode DL. How the evolution of multicellularity set the stage for cancer. *Br J Cancer*. 2018 Jan;118(2):145-152.
- Tropepe V, Hitoshi S, Sirard C, Mak TW, Rossant J, van der Kooy D. Direct neural fate specification from embryonic stem cells: a primitive mammalian neural stem cell stage acquired through a default mechanism. *Neuron*. 2001 Apr;30(1):65-78.
- Turajlic S, Sottoriva A, Graham T, Swanton C. Resolving genetic heterogeneity in cancer. *Nat Rev Genet*. 2019 Jul;20(7):404-416.
- Varela C, Denis JA, Polentes J, Feyeux M, Aubert S, Champon B, Piétu G, Peschanski M, Lefort N. Recurrent genomic instability of chromosome 1q in neural derivatives of human embryonic stem cells. *J Clin Invest*. 2012 Feb;122(2):569-74.
- Vermeulen L, Todaro M, de Sousa Mello F, Sprick MR, Kemper K, Perez Alea M, Richel DJ, Stassi G, Medema JP. Single-cell cloning of colon cancer stem cells reveals a multi-lineage differentiation capacity. *Proc Natl Acad Sci U S A*. 2008 Sep 9;105(36):13427-32.
- Vinogradov AE, Anatskaya OV. "Cell dedifferentiation" versus "evolutionary reversal" theories of cancer: The direct contest of transcriptomic features. *Int J Cancer*. 2025 May 1;156(9):1802-1813.
- Vogelstein B, Kinzler KW. The multistep nature of cancer. *Trends Genet*. 1993 Apr;9(4):138-41.
- Watterson A, Coelho MA. Cancer immune evasion through KRAS and PD-L1 and potential therapeutic interventions. *Cell Commun Signal*. 2023 Mar 2;21(1):45.
- Webb CG, Gootwine E, Sachs L. Developmental potential of myeloid leukemia cells injected into midgestation embryos. *Dev Biol*. 1984 Jan;101(1):221-4.
- Wells RS, Miotto KA. Widespread inhibition of neuroblastoma cells in the 13- to 17-day-old mouse embryo. *Cancer Res*. 1986 Apr;46(4 Pt 1):1659-62.
- Woltjen K, Stanford WL. Inhibition of Tgf-beta signaling improves mouse fibroblast reprogramming. *Cell Stem Cell*. 2009 Nov 6;5(5):457-8.
- Xing X, Zhong J, Biermann J, Duan H, Zhang X, Shi Y, Gao Y, He K, Zhai D, Luo F, Lai Y, Xiao F, Wang W, Wang M, Xu J, Liu H, Tang J, Chu L, Chen T, D'Souza EK, Caprio L, Ebel L, Biswas D, Cottarelli A, Mou Y, Izar B, Zhang N, Bai F. Pan-cancer human brain metastases atlas at single-cell resolution. *Cancer Cell*. 2025 Apr 7;S1535-6108(25)00126-6.
- Xu L, Zhang M, Shi L, Yang X, Chen L, Cao N, Lei A, Cao Y. Neural stemness contributes to cell tumorigenicity. *Cell Biosci*. 2021 Jan 19;11(1):21.
- Xu X, Liu X, Dollar JJ, Liu X, Jasani N, Posorske B, Sriramareddy SN, Jarajapu V, Kuznetsoff JN, Sinard J, Bennett RL, Licht JD, Smalley KSM, Harbour JW, Yu X, Karreth FA. A multi-step immune-competent genetic mouse model reveals phenotypic plasticity in uveal melanoma. *bioRxiv [Preprint]*. 2025 Jun 7:2025.06.04.657841. doi: 10.1101/2025.06.04.657841.
- Yang J, Antin P, Berx G, Blanpain C, Brabletz T, Bronner M, Campbell K, Cano A, Casanova J, Christofori G, Dedhar S, Derynck R, Ford HL, Fuxe J, García de Herreros A, Goodall GJ, Hadjantonakis AK, Huang RYJ, Kalchauer C, Kalluri R, Kang Y, Khew-Goodall Y, Levine H, Liu J, Longmore GD, Mani SA, Massagué J, Mayor R, McClay D, Mostov KE, Newgreen DF, Nieto MA, Puisieux A, Runyan R, Savagner P, Stanger B, Stemmler MP, Takahashi Y, Takeichi M, Theveneau E, Thiery JP, Thompson EW, Weinberg RA, Williams ED, Xing J, Zhou BP, Sheng G; EMT International Association (EMTIA). Guidelines and definitions for research on epithelial-mesenchymal transition. *Nat Rev Mol Cell Biol*. 2020 Jun;21(6):341-352.
- Yang X, Cao N, Chen L, Liu L, Zhang M, Cao Y. Suppression of Cell Tumorigenicity by Non-neural Pro-differentiation Factors via Inhibition of Neural Property in Tumorigenic Cells. *Front Cell Dev Biol*. 2021 Sep 14;9:714383.
- Ying QL, Nichols J, Chambers I, Smith A. BMP induction of Id proteins suppresses differentiation and sustains embryonic stem cell self-renewal in collaboration with STAT3. *Cell*. 2003 Oct 31;115(3):281-92.

- Zalc A, Sinha R, Gulati GS, Wesche DJ, Daszczuk P, Swigut T, Weissman IL, Wysocka J. Reactivation of the pluripotency program precedes formation of the cranial neural crest. *Science*. 2021 Feb 5;371(6529):eabb4776.
- Zhang M, Liu Y, Shi L, Fang L, Xu L, Cao Y. Neural stemness unifies cell tumorigenicity and pluripotent differentiation potential. *J Biol Chem*. 2022 Jul;298(7):102106.
- Zhang Z, Lei A, Xu L, Chen L, Chen Y, Zhang X, Gao Y, Yang X, Zhang M, Cao Y. Similarity in gene-regulatory networks suggests that cancer cells share characteristics of embryonic neural cells. *J Biol Chem*. 2017 Aug 4;292(31):12842-12859.
- Zheng X, Boyer L, Jin M, Mertens J, Kim Y, Ma L, Ma L, Hamm M, Gage FH, Hunter T. Metabolic reprogramming during neuronal differentiation from aerobic glycolysis to neuronal oxidative phosphorylation. *Elife*. 2016 Jun 10;5:e13374.
- Zhou L, Mudianto T, Ma X, Riley R, Uppaluri R. Targeting EZH2 Enhances Antigen Presentation, Antitumor Immunity, and Circumvents Anti-PD-1 Resistance in Head and Neck Cancer. *Clin Cancer Res*. 2020 Jan 1;26(1):290-300.
- Zimmerli D, Brambillasca CS, Talens F, Bhin J, Linstra R, Romanens L, Bhattacharya A, Joosten SEP, Da Silva AM, Padrao N, Wellenstein MD, Kersten K, de Boo M, Roorda M, Henneman L, de Bruijn R, Annunziato S, van der Burg E, Drenth AP, Lutz C, Endres T, van de Ven M, Eilers M, Wessels L, de Visser KE, Zwart W, Fehrmann RSN, van Vugt MATM, Jonkers J. MYC promotes immune-suppression in triple-negative breast cancer via inhibition of interferon signaling. *Nat Commun*. 2022 Nov 2;13(1):6579.
- Zimmermannova O, Ferreira AG, Ascic E, Velasco Santiago M, Kurochkin I, Hansen M, Met Ö, Caiado I, Shapiro IE, Michaux J, Humbert M, Soto-Cabrera D, Benonisson H, Silvério-Alves R, Gomez-Jimenez D, Bernardo C, Bauden M, Andersson R, Höglund M, Miharada K, Nakamura Y, Hugues S, Greiff L, Lindstedt M, Rosa FF, Pires CF, Bassani-Sternberg M, Svane IM, Pereira CF. Restoring tumor immunogenicity with dendritic cell reprogramming. *Sci Immunol*. 2023 Jul 14;8(85):eadd4817.
- Zingg D, Debbache J, Schaefer SM, Tuncer E, Frommel SC, Cheng P, Arenas-Ramirez N, Haeusel J, Zhang Y, Bonalli M, McCabe MT, Creasy CL, Levesque MP, Boyman O, Santoro R, Shakhova O, Dummer R, Sommer L. The epigenetic modifier EZH2 controls melanoma growth and metastasis through silencing of distinct tumour suppressors. *Nat Commun*. 2015 Jan 22;6:6051.
- Zylka MJ, Simon JM, Philpot BD. Gene length matters in neurons. *Neuron*. 2015 Apr 22;86(2):353-5.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.