

Review

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Posted Date: 24 June 2025

doi: 10.20944/preprints202506.1931.v1

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Review

Extracellular Vesicles as Mediators of Intercellular Communication: Implications for Drug Discovery and Targeted Therapies

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Abstract

Extracellular vesicles (EVs) are emerging as versatile mediators of intercellular communication and promising tools for drug discovery and targeted therapies. These lipid bilayer-bound nanovesicles facilitate the transfer of functional proteins, RNAs, and other biomolecules between cells, thereby influencing various physiological and pathological processes. This review outlines the molecular mechanisms governing EV biogenesis and cargo sorting, including the role of ubiquitin-like 3 (UBL3) and its functional implications in disease microenvironments, such as cancer, neurodegeneration, and immune regulation. We further discuss recent advances in engineering EVs for the delivery of therapeutic RNAs, proteins, and small molecules, and their growing utility in noninvasive diagnostics. Despite their clinical promise, challenges such as EV isolation, heterogeneity, and regulatory standardization persist. Emerging strategies targeting EV pathways and molecular modulators such as UBL3 offer exciting prospects for precision medicine. This review highlights the current progress and future direction of positioning EVs at the forefront of next-generation drug delivery and biomarker research.

Keywords: Extracellular vesicles; ubiquitin-like 3; disease microenvironment; precision medicine; next-generation drug delivery

1. Introduction

Extracellular vesicles (EVs) are lipid bilayer-enclosed nanostructures secreted by almost all cell types and classified into subtypes—mainly exosomes (30–150 nm, endosomal origin), microvesicles (100–1000 nm, plasma membrane origin), and apoptotic bodies—based on their biogenesis and size [1–3]. These vesicles carry molecular cargo such as proteins, lipids, DNA, mRNA, miRNAs, and other non-coding RNAs that influence intercellular communication [4–6]. EVs are present in nearly all biological fluids, including the blood, urine, saliva, and cerebrospinal fluid, and function as mediators of both physiological and pathological processes [2,7,8]. Under homeostatic conditions, EVs contribute to tissue regeneration, immune modulation, and neural development [9,10]. However, in pathological settings, EVs have been implicated in cancer metastasis, neurodegeneration, cardiovascular disease, and immune dysregulation [11,12]. Their dual role as disease biomarkers and delivery vehicles highlights their relevance in diagnostics and therapeutics, particularly in precision medicine [13–15].

One of the most attractive features of EVs is their natural ability to cross biological barriers, such as the blood-brain barrier, making them suitable candidates for delivering therapeutic agents to the central nervous system (CNS) [16,17]. Their surface molecules, including integrins, tetraspanins, and other receptors, confer targeting specificity, allowing selective delivery to recipient cells and tissues [18–20].

In pharmacology and drug discovery, EVs are gaining traction as next-generation therapeutic platforms owing to their biocompatibility, low immunogenicity, and intrinsic targeting capacity [21,22]. These properties have inspired numerous preclinical and clinical investigations exploring EVs in cancer, neurodegenerative diseases, inflammation, and regenerative medicine [23–25].

Recent studies have demonstrated the engineering of EVs to encapsulate various therapeutic agents, including small molecules, siRNAs, miRNAs, proteins, and CRISPR/Cas9 components [26–29]. For example, exosomes engineered to express brain-targeting ligands (e.g., Lamp2b-RVG peptide) have shown enhanced central nervous system delivery via systemic administration [16]. Additionally, surface-functionalized EVs equipped with targeting peptides, antibodies, or aptamers exhibit significantly improved biodistribution and cellular uptake [30–32].

The translational potential of EVs has been demonstrated in various diseases. In oncology, EVs loaded with chemotherapeutics or siRNAs targeting oncogenes, such as KRAS, have shown antitumor efficacy [11]. In stroke and neurodegeneration, BDNF- or catalase-loaded exosomes reduce brain damage and improve functional recovery [33]. Notably, our recent study developed label-free imaging of EVs in breast cancer, highlighting their potential applications in targeted breast cancer therapies [34]. Another study conducted in 2023 reported the neuroprotective effects of EVs in neurodegenerative disease models [14]. Furthermore, neurological disorders like Parkinson's and Alzheimer's diseases have been shown to benefit from EV-based therapies, given their ability to modulate neuroinflammation and synaptic function [35,36]. A recent integrative molecular study uncovered how EVs mediate drug transport and metabolic reprogramming, further supporting their therapeutic utility [37,38].

Despite their promise, technical challenges remain, including scalable and standardized isolation, purity control, in vivo tracking, and storage stability [39,40]. The adoption of international guidelines such as MISEV2018 and MISEV2023 has provided a much-needed framework for EV characterization and nomenclature [41]. The integration of multi-omics technologies, bioinformatics, and nanotechnology is essential for overcoming the current barriers and unlocking the full clinical potential of EVs [41–43].

2. Molecular Mechanisms of EV Biogenesis and Cargo Sorting

The biogenesis and cargo loading of EVs are controlled by coordinated molecular pathways that determine their structures and functions. EVs are broadly categorized as exosomes, formed via inward budding of late endosomes into multivesicular bodies (MVBs), and microvesicles, generated by outward budding from the plasma membrane [4,7,44].

Exosome formation is primarily governed by the Endosomal Sorting Complex Required for Transport (ESCRT) machinery, which includes ESCRT-0 to -III and accessory proteins such as ALIX and TSG101 [45–48]. These components enable membrane budding and cargo selection. ESCRT-independent mechanisms also contribute to EV biogenesis, involving tetraspanins (CD9, CD63, and CD81), ceramides, and lipid rafts, which influence both membrane curvature and selective cargo sorting [49–51].

Rab GTPases, such as Rab27a/b, Rab11, and Rab35, regulate MVB trafficking, docking, and fusion with the plasma membrane, thereby modulating EV secretion [52–54]. For instance, Rab27a positions MVBs at the membrane, whereas Rab11 mediates the recycling pathways [55,56].

Among the emerging regulators, UBL3 (Ubiquitin-like protein 3) plays a non-canonical role in post-translational modifications and cargo sorting. UBL3 localizes to the plasma membrane and directs S-prenylation-dependent protein modifications, facilitating their selective packaging into EVs [34,57,58]. It influences the secretion of immune-regulatory and tumor-related proteins, and UBL3 deficiency disrupts EV composition in pathological contexts [34,57]. Thus, UBL3 is a promising target for the production of engineered EV.

The cargo content of EVs is dynamically modulated by their cellular state. Stress conditions such as hypoxia, inflammation, or oxidative stress alter EV composition, enriching them with proteins like HIF-1 α , VEGF, or pro-inflammatory miRNAs [59–61]. Immune activation, for example, triggers the

release of EVs carrying checkpoint proteins and miR-155, whereas neuronal activity affects EV cargo during synaptic signaling and injury responses [62,63].

Together, these tightly regulated processes ensure that EVs carry highly specific molecular signatures, enabling precise intercellular communication and presenting opportunities for the therapeutic customization of drug delivery systems. The process of EV biogenesis and selective cargo loading is visually summarized in **Figure 1** [64].

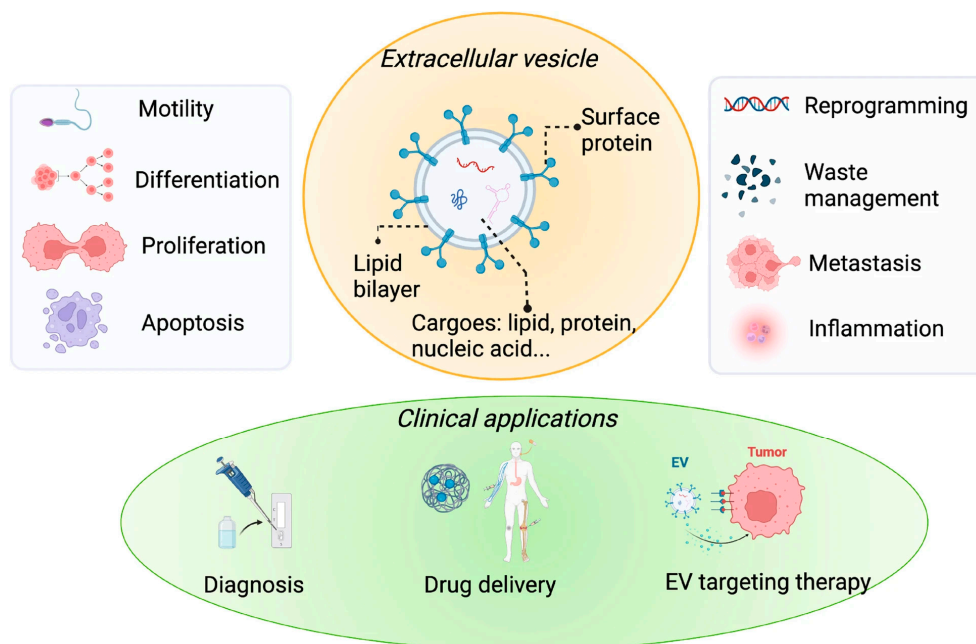


Figure 1. Molecular mechanisms of EV biogenesis and cargo sorting. EVs are generated through multiple pathways, including ESCRT-dependent and ESCRT-independent mechanisms. Rab GTPases and UBL3 also regulate EV trafficking and protein loading. Adapted and modified with permission from Anand et al., *Cell Communication and Signaling* (2023) [64].

3. EV-Mediated Intercellular Communication in Disease Microenvironments

EVs are critical modulators of the disease microenvironment, contributing to cancer progression, neurodegeneration, and immune modulation through the transfer of bioactive molecules, such as proteins, lipids, mRNAs, and non-coding RNAs [65–67]. Their ability to deliver specific cargo into recipient cells enables EVs to shape the behavior of neighboring or distant cells, supporting pathological processes such as tumor metastasis, inflammation, and the propagation of toxic proteins [34,57].

In the tumor microenvironment (TME), EVs mediate bidirectional communication between cancer cells and stromal, endothelial, and immune cells [68]. Tumor-derived EVs (TDEs) carry oncogenic proteins (e.g., EGFRvIII), immunosuppressive molecules (e.g., PD-L1), and pro-angiogenic factors (e.g., VEGF), promoting tumor growth, immune escape, and vascular remodeling [69–71]. TDEs also reprogram fibroblasts and recruit tumor-associated macrophages (TAMs), further amplifying their metastatic potential [72,73]. For example, exosomal integrins ($\alpha 6 \beta 4$ and $\alpha v \beta 5$) have been implicated in organotropic metastasis by preconditioning distant tissues [74].

In neurodegenerative diseases, EVs act as vectors for cell-to-cell transmission of pathogenic proteins [14,75]. Studies have shown that EVs transport misfolded tau, amyloid- β ($A\beta$), α -synuclein, and TDP-43, facilitating their propagation in Alzheimer's disease (AD), Parkinson's disease (PD), and amyotrophic lateral sclerosis (ALS) [76–79]. This prion-like spread contributes to disease progression and neural network dysfunctions. Moreover, neuron- and astrocyte-derived EVs influence microglial activation, contributing to neuroinflammation during the early stages of neurodegeneration [80,81].

Extracellular vesicles (EVs) play a dual role in the immune system. On one hand, they mediate antigen presentation and T-cell activation; on the other hand, cancer-derived EVs often suppress immunity by delivering PD-L1, FasL, or miRNAs that target immune checkpoints [82,83]. Furthermore, EVs secreted by dendritic cells and T cells carry MHC-peptide complexes, costimulatory molecules, and miRNAs that modulate the activity of effector and regulatory cells, thereby influencing inflammation and tolerance [84,85]. An overview of EV functions across different disease microenvironments is shown in **Figure 2** [64].

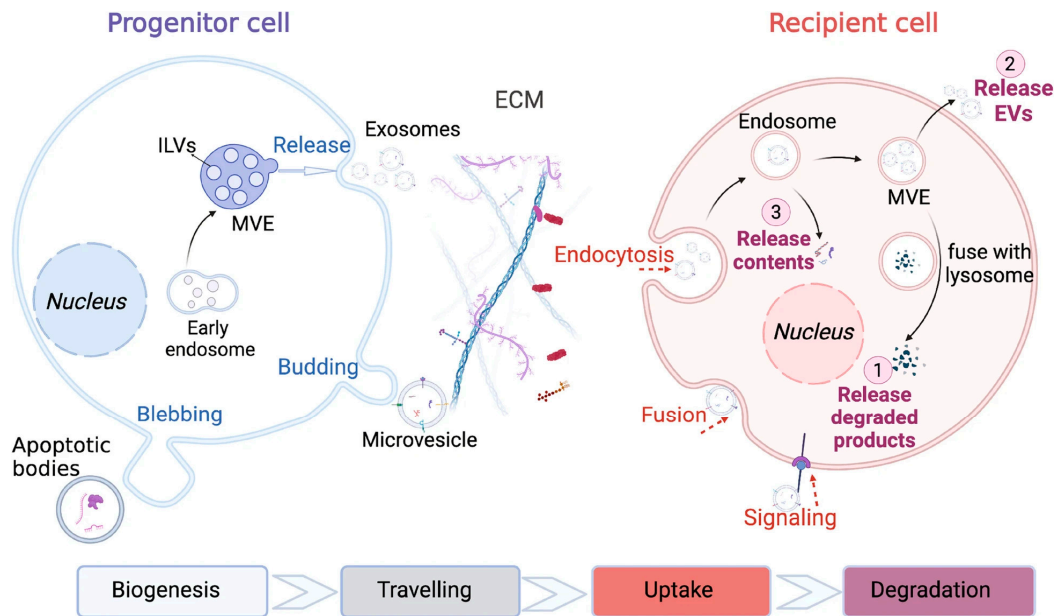


Figure 2. Schematic illustration of extracellular vesicle functions in disease microenvironments, including cancer, neurodegeneration, and immune modulation. EVs mediate intercellular communication by transferring bioactive molecules, influencing disease progression and immune responses. Adapted and modified with permission from Anand et al., *Cell Communication and Signaling* (2023) [64].

Together, EVs act as intercellular shuttles that remodel the disease microenvironment by enhancing malignancy, spreading pathogenic proteins, and modulating the immune response. Understanding these EV-mediated mechanisms opens new avenues for novel therapeutic interventions and biomarker discovery in complex diseases [86].

4. Pharmacological Targeting of EV Pathways

Pharmacological modulation of extracellular vesicle (EV) pathways is a promising strategy for both inhibiting pathological EV activity and enhancing therapeutic EV function. Targeting the key steps in EV biogenesis, release, uptake, and cargo packaging opens new avenues for drug development and disease intervention.

Several small-molecule inhibitors that suppress EV release have been identified. GW4869, a neutral sphingomyelinase inhibitor, is widely used to block ceramide-mediated exosome biogenesis and has been shown to reduce EV-mediated inflammation and tumor progression [87,88]. Similarly, imipramine, an FDA-approved tricyclic antidepressant, inhibits exosome secretion via endolysosomal disruption [89,90]. In contrast, certain compounds such as monensin and forskolin have been reported to enhance EV release, which could be leveraged for therapeutic EV production [91,92].

The uptake of EVs by recipient cells can be pharmacologically regulated. Molecules such as dynasore and chlorpromazine inhibit clathrin- or caveolin-mediated endocytosis, thereby offering tools to prevent unwanted EV-mediated signal propagation [86,93].

Beyond trafficking, emerging methods enable the manipulation of EV content. Engineered donor cells or post-isolation techniques like electroporation, sonication, and chemical transfection allow for the loading of therapeutic cargos such as siRNAs, CRISPR-Cas systems, or chemotherapeutics [90,94].

Among the molecular targets, UBL3 has recently gained attention for its role in S-prenylation-dependent EV cargo loading. UBL3 facilitates the selective inclusion of immune and disease-related proteins into small EVs and represents a druggable pathway for cargo-level modulation [34,95]. Altering UBL3 activity may allow researchers to reprogram EV content for cancer immunotherapy or neuroinflammation modulation [34,57,96].

Together, these strategies emphasize the therapeutic potential of EV pathway modulation, positioning pharmacological EV targeting as the next-generation modality in precision medicine.

5. EVs as Drug Delivery Vehicles and Biomarker Reservoirs

EVs have emerged as powerful platforms for therapeutic delivery and diagnostic applications owing to their inherent biocompatibility, low immunogenicity, and natural targeting ability [97]. Their ability to transport a variety of bioactive molecules—including RNAs, proteins, and small-molecule drugs—makes them particularly promising for precision drug delivery and noninvasive biomarker discovery [98].

Engineered EVs have been extensively explored for the delivery of therapeutic RNAs (e.g., siRNA, miRNA, and mRNA), proteins, and genome-editing tools such as CRISPR/Cas9. Loading strategies include donor cell transfection, electroporation, sonication, and extrusion [99–102]. For example, Alvarez-Erviti et al. (2011) successfully delivered siRNA across the blood-brain barrier using exosomes engineered with the Lamp2b-RVG fusion peptide [16]. Similarly, MSC-derived EVs have been used to deliver anti-inflammatory miRNAs and neuroprotective factors in models of stroke, spinal cord injury, and myocardial infarction [103].

Surface modification techniques, such as ligand display, aptamer conjugation, and peptide anchoring, can further enhance targeted delivery. Ligands like GE11 (for EGFR) or RGD (for integrins) have been conjugated to EV surfaces to improve tissue-specific accumulation [104,105]. These strategies significantly increase therapeutic efficacy while minimizing off-target effects and systemic toxicity.

In parallel, EVs are increasingly being recognized as rich reservoirs of diagnostic biomarkers, particularly in liquid biopsy platforms. Circulating EVs in the blood, urine, or CSF contain disease-specific proteins, lipids, and RNAs reflective of the physiological or pathological state of the originating cells [106–108]. In cancer, EV-derived miRNAs (e.g., miR-21, miR-1246), proteins (e.g., EpCAM, CD63), and DNA fragments have shown strong diagnostic and prognostic potential across various tumor types [109,110]. Similarly, EVs in neurodegenerative diseases carry α -synuclein, tau, or A β species that can distinguish between disease stages and subtypes [111].

Together, these properties position EVs as multifunctional agents in drug delivery and clinical diagnostics. Their use in ongoing clinical trials further highlights their potential as the next-generation precision tools.

6. Challenges and Future Perspectives in EV-Based Drug Discovery

Despite their promise as drug carriers and diagnostic tools, extracellular vesicles (EVs) face several technical and translational challenges that must be overcome for their successful clinical implementation. These include issues with isolation and purification, cargo heterogeneity, dosing standardization, and an uncertain regulatory framework.

One of the primary challenges is the lack of standardized isolation protocols. Current methods, such as ultracentrifugation, size-exclusion chromatography, and precipitation, vary widely in efficiency and purity [112]. This inconsistency affects the reproducibility and downstream functional analyses. Moreover, the heterogeneous nature of EV populations—even within the same biofluid

complicates characterization and therapeutic efficacy [113]. New microfluidic platforms and affinity-based purification systems offer improved selectivity but are not yet scalable or cost-effective for clinical applications [114].

Another hurdle is the regulatory and translational gap. The lack of global consensus on EV classification, potency assays, and quality control has hindered the development of Good Manufacturing Practice (GMP)-compliant EV therapeutics [115,116]. The field is currently guided by position papers such as MISEV2022, but clear regulatory frameworks from agencies like the FDA or EMA remain in development [114].

However, dosing strategies and biodistribution profiling pose significant challenges. Quantifying EVs remains difficult due to overlapping size ranges with other nanoparticles and variability in protein-to-vesicle ratios [117,118]. Additionally, the long-term effects of EV administration and immune clearance mechanisms are not yet fully understood, which raises safety concerns regarding repeated dosing.

Personalized EV-based therapeutics are expected to gain attention in the future. Patient-derived or engineered EVs tailored to individual genetic or proteomic profiles could revolutionize precision medicine [119,120]. In this context, UBL3 has emerged as a promising druggable regulator for EV cargo sorting. By modulating S-prenylation of surface proteins, UBL3 controls the selective inclusion of immune and disease-associated factors in small EVs [58]. Targeting UBL3 and its downstream pathways may allow for the precise reprogramming of EV content, particularly in cancer and neuroinflammatory conditions [34].

In conclusion, although significant hurdles remain, continued advancements in EV engineering, standardization, and molecular targeting (including UBL3) are expected to accelerate the clinical translation of EV-based therapeutics in the near future.

Author Contributions: Conceptualization- M.M.H; writing—original draft preparation- M.A.M; Review and editing- M.A.M, S.M.S, A.S.M.W and M.M.H. All authors have read and agreed to the published version of this manuscript.

Funding: This study received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

Abbreviations

The following abbreviations are used in this manuscript.

AD	Alzheimer’s Disease
ALS	Amyotrophic Lateral Sclerosis
Aβ	Amyloid beta
BBB	Blood-Brain Barrier
CNS	Central Nervous System
CM	Conditioned Medium
CRC	Colorectal Cancer
DC	Dendritic Cell
DNA	Deoxyribonucleic Acid
EV	Extracellular Vesicle
FDA	Food and Drug Administration
GSC	Glioma Stem Cell
GTPase	Guanosine Triphosphatase
HNSCC	Head and Neck Squamous Cell Carcinoma

HSP	Heat Shock Protein
ILV	Intraluminal Vesicle
ISEV	International Society for Extracellular Vesicles
KO	Knockout
LAMP	Lysosomal-Associated Membrane Protein
miRNA	MicroRNA
miR	microRNA (generic notation)
MISEV	Minimal Information for Studies of Extracellular Vesicles
MSC	Mesenchymal Stem Cell
NSCLC	Non-Small Cell Lung Cancer
PD	Parkinson’s Disease
PD-L1	Programmed Death-Ligand 1
RNA	Ribonucleic Acid
siRNA	Small Interfering RNA
sEV	Small Extracellular Vesicle
TME	Tumor Microenvironment
TNBC	Triple-Negative Breast Cancer
UBL3	Ubiquitin-Like Protein 3
WT	Wild-Type

References

1. Moghassemi, S.; Dadashzadeh, A.; Sousa, M.J.; Vlieghe, H.; Yang, J.; León-Félix, C.M.; Amorim, C.A. Extracellular Vesicles in Nanomedicine and Regenerative Medicine: A Review over the Last Decade. *Bioact Mater* **2024**, *36*, 126–156. <https://doi.org/10.1016/j.bioactmat.2024.02.021>.

2. Chronopoulos, A.; Kalluri, R. Emerging Role of Bacterial Extracellular Vesicles in Cancer. *Oncogene* **2020**, *39*, 6951–6960. <https://doi.org/10.1038/s41388-020-01509-3>.

3. El Andaloussi, S.; Mäger, I.; Breakefield, X.O.; Wood, M.J.A. Extracellular Vesicles: Biology and Emerging Therapeutic Opportunities. *Nature Reviews Drug Discovery* **2013**, *12*, 347–357. <https://doi.org/10.1038/nrd3978>.

4. Van Niel, G.; D’Angelo, G.; Raposo, G. Shedding Light on the Cell Biology of Extracellular Vesicles. *Nat Rev Mol Cell Biol* **2018**, *19*, 213–228. <https://doi.org/10.1038/NRM.2017.125>;SUBJMETA=2155,2162,313,631,80,820,86;KWRD=ESCRT,EXTRACELLULAR+SIGNALLING+MOLECULES,MULTIVESICULAR+BODIES.

5. Anand, S.; Samuel, M.; Kumar, S.; Mathivanan, S. Ticket to a Bubble Ride: Cargo Sorting into Exosomes and Extracellular Vesicles. *Biochim Biophys Acta Proteins Proteom* **2019**, 1867. <https://doi.org/10.1016/j.bbapap.2019.02.005>.

6. Yáñez-Mó, M.; Siljander, P.R.M.; Andreu, Z.; Zavec, A.B.; Borràs, F.E.; Buzas, E.I.; Buzas, K.; Casal, E.; Cappello, F.; Carvalho, J.; et al. Biological Properties of Extracellular Vesicles and Their Physiological Functions. *J Extracell Vesicles* **2015**, *4*, 27066. <https://doi.org/10.3402/JEV.V4.27066>.

7. Colombo, M.; Raposo, G.; Théry, C. Biogenesis, Secretion, and Intercellular Interactions of Exosomes and Other Extracellular Vesicles. *Annu Rev Cell Dev Biol* **2014**, *30*, 255–289. <https://doi.org/10.1146/ANNUREV-CELLBIO-101512-122326>.

8. Raposo, G.; Stoorvogel, W. Extracellular Vesicles: Exosomes, Microvesicles, and Friends. *Journal of Cell Biology* **2013**, *200*, 373–383. <https://doi.org/10.1083/JCB.201211138>.

9. Pascucci, L.; Coccè, V.; Bonomi, A.; Ami, D.; Ceccarelli, P.; Ciusani, E.; Viganò, L.; Locatelli, A.; Sisto, F.; Doglia, S.M.; et al. Paclitaxel Is Incorporated by Mesenchymal Stromal Cells and Released in Exosomes That Inhibit in Vitro Tumor Growth: A New Approach for Drug Delivery. *Journal of Controlled Release* **2014**, *192*, 262–270. <https://doi.org/10.1016/j.jconrel.2014.07.042>.

10. Roefs, M.T.; Sluijter, J.P.G.; Vader, P. Extracellular Vesicle-Associated Proteins in Tissue Repair. *Trends Cell Biol* **2020**, *30*, 990–1013. <https://doi.org/10.1016/J.TCB.2020.09.009>;ASSET/73AF1E10-5A4D-4B01-BC9A-10AE3F47D5D8/MAIN.ASSETS/GR1.JPG.

11. Kamerkar, S.; Lebleu, V.S.; Sugimoto, H.; Yang, S.; Ruivo, C.F.; Melo, S.A.; Lee, J.J.; Kalluri, R. Exosomes Facilitate Therapeutic Targeting of Oncogenic KRAS in Pancreatic Cancer. *Nature* **2017**, *546*, 498–503. <https://doi.org/10.1038/NATURE22341>;TECHMETA=13,31;SUBJMETA=1504,1647,631,67;KWRD=BIOLOGICAL+TECHNIQUES,GASTROINTESTINAL+CANCER.
12. Skog, J.; Würdinger, T.; van Rijn, S.; Meijer, D.H.; Gainche, L.; Curry, W.T.; Carter, B.S.; Krichevsky, A.M.; Breakefield, X.O. Glioblastoma Microvesicles Transport RNA and Proteins That Promote Tumour Growth and Provide Diagnostic Biomarkers. *Nat Cell Biol* **2008**, *10*, 1470–1476. <https://doi.org/10.1038/NCB1800>.
13. Doyle, L.M.; Wang, M.Z. Overview of Extracellular Vesicles, Their Origin, Composition, Purpose, and Methods for Exosome Isolation and Analysis. *Cells* **2019**, *8*. <https://doi.org/10.3390/CELLS8070727>.
14. Chen, B.; Hasan, M.M.; Zhang, H.; Zhai, Q.; Waliullah, A.S.M.; Ping, Y.; Zhang, C.; Oyama, S.; Mimi, M.A.; Tomochika, Y.; et al. UBL3 Interacts with Alpha-Synuclein in Cells and the Interaction Is Downregulated by the EGFR Pathway Inhibitor Osimertinib. *Biomedicines* **2023**, *11*. <https://doi.org/10.3390/BIOMEDICINES11061685>.
15. Oyama, S.; Zhang, H.; Ferdous, R.; Tomochika, Y.; Chen, B.; Jiang, S.; Islam, M.S.; Hasan, M.M.; Zhai, Q.; Waliullah, A.S.M.; et al. UBL3 Interacts with PolyQ-Expanded Huntingtin Fragments and Modifies Their Intracellular Sorting. *Neurol Int* **2024**, *16*, 1175–1188. <https://doi.org/10.3390/NEUROLINT16060089/S1>.
16. Alvarez-Erviti, L.; Seow, Y.; Yin, H.; Betts, C.; Lakhali, S.; Wood, M.J.A. Delivery of siRNA to the Mouse Brain by Systemic Injection of Targeted Exosomes. *Nature Biotechnology* **2011**, *29*, 341–345. <https://doi.org/10.1038/nbt.1807>.
17. Haney, M.J.; Klyachko, N.L.; Zhao, Y.; Gupta, R.; Plotnikova, E.G.; He, Z.; Patel, T.; Piroyan, A.; Sokolsky, M.; Kabanov, A. V.; et al. Exosomes as Drug Delivery Vehicles for Parkinson's Disease Therapy. *Journal of Controlled Release* **2015**, *207*, 18–30. <https://doi.org/10.1016/j.jconrel.2015.03.033>.
18. Kwok, Z.H.; Wang, C.; Jin, Y. Extracellular Vesicle Transportation and Uptake by Recipient Cells: A Critical Process to Regulate Human Diseases. *Processes (Basel)* **2021**, *9*, 273. <https://doi.org/10.3390/PR9020273>.
19. French, K.C.; Antonyak, M.A.; Cerione, R.A. Extracellular Vesicle Docking at the Cellular Port: Extracellular Vesicle Binding and Uptake. *Semin Cell Dev Biol* **2017**, *67*, 48. <https://doi.org/10.1016/J.SEMCDB.2017.01.002>.
20. Hoshino, A.; Costa-Silva, B.; Shen, T.L.; Rodrigues, G.; Hashimoto, A.; Tesic Mark, M.; Molina, H.; Kohsaka, S.; Di Giannatale, A.; Ceder, S.; et al. Tumour Exosome Integrins Determine Organotropic Metastasis. *Nature* **2015**, *527*, 329–335. <https://doi.org/10.1038/NATURE15756>.
21. Vader, P.; Mol, E.A.; Pasterkamp, G.; Schiffelers, R.M. Extracellular Vesicles for Drug Delivery. *Adv Drug Deliv Rev* **2016**, *106*, 148–156. <https://doi.org/10.1016/j.addr.2016.02.006>.
22. Du, S.; Guan, Y.; Xie, A.; Yan, Z.; Gao, S.; Li, W.; Rao, L.; Chen, X.; Chen, T. Extracellular Vesicles: A Rising Star for Therapeutics and Drug Delivery. *J Nanobiotechnology* **2023**, *21*, 231. <https://doi.org/10.1186/S12951-023-01973-5>.
23. Wang, L.; Zhang, X.; Yang, Z.; Wang, B.; Gong, H.; Zhang, K.; Lin, Y.; Sun, M. Extracellular Vesicles: Biological Mechanisms and Emerging Therapeutic Opportunities in Neurodegenerative Diseases. *Transl Neurodegener* **2024**, *13*, 60. <https://doi.org/10.1186/S40035-024-00453-6>.
24. Yan, J.; Kahyo, T.; Zhang, H.; Ping, Y.; Zhang, C.; Jiang, S.; Ji, Q.; Ferdous, R.; Islam, M.S.; Oyama, S.; et al. Alpha-Synuclein Interaction with UBL3 Is Upregulated by Microsomal Glutathione S-Transferase 3, Leading to Increased Extracellular Transport of the Alpha-Synuclein under Oxidative Stress. *Int J Mol Sci* **2024**, *25*, 7353. <https://doi.org/10.3390/IJMS25137353/S1>.
25. Klyachko, N.L.; Arzt, C.J.; Li, S.M.; Gololobova, O.A.; Batrakova, E. V. Extracellular Vesicle-Based Therapeutics: Preclinical and Clinical Investigations. *Pharmaceutics* **2020**, *12*, 1171. <https://doi.org/10.3390/PHARMACEUTICS12121171>.
26. Aureliano, M.; Maihemuti, M.; Afsana Mimi, M.; Sohag, S.M.; Hasan, M.M. Single-Cell Transcriptomics in Spinal Cord Studies: Progress and Perspectives. *BioChem* **2025**, *Vol. 5*, Page 16 **2025**, *5*, 16. <https://doi.org/10.3390/BIOCHEM5020016>.
27. Barile, L.; Vassalli, G. Exosomes: Therapy Delivery Tools and Biomarkers of Diseases. *Pharmacol Ther* **2017**, *174*, 63–78. <https://doi.org/10.1016/j.pharmthera.2017.02.020>.

28. O'Brien, K.; Breyne, K.; Ughetto, S.; Laurent, L.C.; Breakefield, X.O. RNA Delivery by Extracellular Vesicles in Mammalian Cells and Its Applications. *Nature Reviews Molecular Cell Biology* 2020 21:10 **2020**, 21, 585–606. <https://doi.org/10.1038/s41580-020-0251-y>.
29. Sluijter, J.P.G.; Davidson, S.M.; Boulanger, C.M.; Buzás, E.I.; De Kleijn, D.P.V.; Engel, F.B.; Giricz, Z.; Hausenloy, D.J.; Kishore, R.; Lecour, S.; et al. Extracellular Vesicles in Diagnostics and Therapy of the Ischaemic Heart: Position Paper from the Working Group on Cellular Biology of the Heart of the European Society of Cardiology. *Cardiovasc Res* **2018**, 114, 19–34. <https://doi.org/10.1093/CVR/ CVX211>.
30. Gong, Z.; Cheng, C.; Sun, C.; Cheng, X. Harnessing Engineered Extracellular Vesicles for Enhanced Therapeutic Efficacy: Advancements in Cancer Immunotherapy. *Journal of Experimental & Clinical Cancer Research* 2025 44:1 **2025**, 44, 1–32. <https://doi.org/10.1186/S13046-025-03403-W>.
31. Murphy, D.E.; de Jong, O.G.; Brouwer, M.; Wood, M.J.; Lavieu, G.; Schiffelers, R.M.; Vader, P. Extracellular Vesicle-Based Therapeutics: Natural versus Engineered Targeting and Trafficking. *Exp Mol Med* **2019**, 51, 1–12. <https://doi.org/10.1038/S12276-019-0223-5>; SUBJMETA=631,80,86; KWRD=CELL+SIGNALLING.
32. Pham, T.C.; Jayasinghe, M.K.; Pham, T.T.; Yang, Y.; Wei, L.; Usman, W.M.; Chen, H.; Pirisinu, M.; Gong, J.; Kim, S.; et al. Covalent Conjugation of Extracellular Vesicles with Peptides and Nanobodies for Targeted Therapeutic Delivery. *J Extracell Vesicles* **2021**, 10, e12057. <https://doi.org/10.1002/JEV2.12057>; PAGE:STRING:ARTICLE/CHAPTER.
33. Doeppner, T.R.; Herz, J.; Görgens, A.; Schlechter, J.; Ludwig, A.-K.; Radtke, S.; de Miroshedji, K.; Horn, P.A.; Giebel, B.; Hermann, D.M. Extracellular Vesicles Improve Post-Stroke Neuroregeneration and Prevent Postischemic Immunosuppression. *Stem Cells Transl Med* **2015**, 4, 1131–1143. <https://doi.org/10.5966/SCTM.2015-0078/-/DC1>.
34. Mimi, M.A.; Hasan, M.M.; Takanashi, Y.; Waliullah, A.S.M.; Mamun, M. Al; Chi, Z.; Kahyo, T.; Aramaki, S.; Takatsuka, D.; Koizumi, K.; et al. UBL3 Overexpression Enhances EV-Mediated Achilles Protein Secretion in Conditioned Media of MDA-MB-231 Cells. *Biochem Biophys Res Commun* **2024**, 738. <https://doi.org/10.1016/J.BBRC.2024.150559>.
35. Han, J.; Zhang, X.; Kang, L.; Guan, J. Extracellular Vesicles as Therapeutic Modulators of Neuroinflammation in Alzheimer's Disease: A Focus on Signaling Mechanisms. *J Neuroinflammation* **2025**, 22, 120. <https://doi.org/10.1186/S12974-025-03443-1>.
36. Cabrera-Pastor, A. Extracellular Vesicles as Mediators of Neuroinflammation in Intercellular and Inter-Organ Crosstalk. *Int J Mol Sci* **2024**, 25, 7041. <https://doi.org/10.3390/IJMS25137041>.
37. Kumar, M.A.; Baba, S.K.; Sadida, H.Q.; Marzooqi, S. Al; Jerobin, J.; Altemani, F.H.; Algehainy, N.; Alanazi, M.A.; Abou-Samra, A.B.; Kumar, R.; et al. Extracellular Vesicles as Tools and Targets in Therapy for Diseases. *Signal Transduction and Targeted Therapy* 2024 9:1 **2024**, 9, 1–41. <https://doi.org/10.1038/s41392-024-01735-1>.
38. Tang, Y.; Liu, X.; Sun, M.; Xiong, S.; Xiao, N.; Li, J.; He, X.; Xie, J. Recent Progress in Extracellular Vesicle-Based Carriers for Targeted Drug Delivery in Cancer Therapy. *Pharmaceutics* **2023**, 15, 1902. <https://doi.org/10.3390/PHARMACEUTICS15071902>.
39. Sódar, B.W.; Kittel, Á.; Pálóczi, K.; Vukman, K. V.; Osteikoetxea, X.; Szabó-Taylor, K.; Németh, A.; Sperlágh, B.; Baranyai, T.; Giricz, Z.; et al. Low-Density Lipoprotein Mimics Blood Plasma-Derived Exosomes and Microvesicles during Isolation and Detection. *Sci Rep* **2016**, 6, 1–12. <https://doi.org/10.1038/SREP24316>; TECHMETA=14,16,28,58,82; SUBJMETA=1407,1492,1647,2230,631; KWRD=FLOW+CYTOMETRY,ISOLATION.
40. Lener, T.; Gimona, M.; Aigner, L.; Börger, V.; Buzas, E.; Camussi, G.; Chaput, N.; Chatterjee, D.; Court, F.A.; del Portillo, H.A.; et al. Applying Extracellular Vesicles Based Therapeutics in Clinical Trials - An ISEV Position Paper. *J Extracell Vesicles* **2015**, 4. <https://doi.org/10.3402/JEV.V4.30087>.
41. Zhang, Y.; Lan, M.; Chen, Y. Minimal Information for Studies of Extracellular Vesicles (MISEV): Ten-Year Evolution (2014–2023). *Pharmaceutics* **2024**, 16, 1394. <https://doi.org/10.3390/PHARMACEUTICS16111394>.
42. Liu, J.; Ren, L.; Li, S.; Li, W.; Zheng, X.; Yang, Y.; Fu, W.; Yi, J.; Wang, J.; Du, G. The Biology, Function, and Applications of Exosomes in Cancer. *Acta Pharm Sin B* **2021**, 11, 2783–2797. <https://doi.org/10.1016/j.apsb.2021.01.001>.

43. Zhang, Y.; Yang, H.; An, X.; Wang, Z.; Yang, X.; Yu, M.; Zhang, R.; Sun, Z.; Wang, Q. Controlled Synthesis of Ag₂Te@Ag₂S Core–Shell Quantum Dots with Enhanced and Tunable Fluorescence in the Second Near-Infrared Window. *Small* **2020**, *16*, 2001003. <https://doi.org/10.1002/SMLL.202001003>.
44. Zhao, Y.; Li, X.; Zhang, W.; Yu, L.; Wang, Y.; Deng, Z.; Liu, M.; Mo, S.; Wang, R.; Zhao, J.; et al. Trends in the Biological Functions and Medical Applications of Extracellular Vesicles and Analogues. *Acta Pharm Sin B* **2021**, *11*, 2114–2135. <https://doi.org/10.1016/J.APSB.2021.03.012>.
45. Lee, Y.J.; Shin, K.J.; Chae, Y.C. Regulation of Cargo Selection in Exosome Biogenesis and Its Biomedical Applications in Cancer. *Experimental & Molecular Medicine* **2024**, *56*, 877–889. <https://doi.org/10.1038/s12276-024-01209-y>.
46. Ju, Y.; Bai, H.; Ren, L.; Zhang, L. The Role of Exosome and the ESCRT Pathway on Enveloped Virus Infection. *Int J Mol Sci* **2021**, *22*, 9060. <https://doi.org/10.3390/IJMS22169060>.
47. Tanaka, N.; Kyuuma, M.; Sugamura, K. Endosomal Sorting Complex Required for Transport Proteins in Cancer Pathogenesis, Vesicular Transport, and Non-endosomal Functions. *Cancer Sci* **2008**, *99*, 1293. <https://doi.org/10.1111/J.1349-7006.2008.00825.X>.
48. Gurung, S.; Perocheau, D.; Touramanidou, L.; Baruteau, J. The Exosome Journey: From Biogenesis to Uptake and Intracellular Signalling. *Cell Communication and Signaling* **2021**, *19*, 1–19. <https://doi.org/10.1186/S12964-021-00730-1>.
49. Van Deun, J.; Mestdag, P.; Sormunen, R.; Cocquyt, V.; Vermaelen, K.; Vandesompele, J.; Bracke, M.; De Wever, O.; Hendrix, A. The Impact of Disparate Isolation Methods for Extracellular Vesicles on Downstream RNA Profiling. *J Extracell Vesicles* **2014**, *3*. <https://doi.org/10.3402/JEV.V3.24858>.
50. Andreu, Z.; Yáñez-Mó, M. Tetraspanins in Extracellular Vesicle Formation and Function. *Front Immunol* **2014**, *5*, 109543. <https://doi.org/10.3389/FIMMU.2014.00442/XML/NLM>.
51. Trajkovic, K.; Hsu, C.; Chiantia, S.; Rajendran, L.; Wenzel, D.; Wieland, F.; Schwille, P.; Brügger, B.; Simons, M. Ceramide Triggers Budding of Exosome Vesicles into Multivesicular Endosomes. *Science (1979)* **2008**, *319*, 1244–1247. <https://doi.org/10.1126/SCIENCE.1153124>.
52. Homma, Y.; Hiragi, S.; Fukuda, M. Rab Family of Small GTPases: An Updated View on Their Regulation and Functions. *FEBS J* **2020**, *288*, 36. <https://doi.org/10.1111/FEBS.15453>.
53. Villarroya-Beltri, C.; Baixauli, F.; Gutiérrez-Vázquez, C.; Sánchez-Madrid, F.; Mittelbrunn, M. SORTING IT OUT: REGULATION OF EXOSOME LOADING. *Semin Cancer Biol* **2014**, *28*, 3. <https://doi.org/10.1016/J.SEMCANCER.2014.04.009>.
54. Ostrowski, M.; Carmo, N.B.; Krumeich, S.; Fanget, I.; Raposo, G.; Savina, A.; Moita, C.F.; Schauer, K.; Hume, A.N.; Freitas, R.P.; et al. Rab27a and Rab27b Control Different Steps of the Exosome Secretion Pathway. *Nat Cell Biol* **2010**, *12*, 19–30. <https://doi.org/10.1038/NCB2000;KWRD=LIFE+SCIENCES>.
55. Han, Q.F.; Li, W.J.; Hu, K.S.; Gao, J.; Zhai, W.L.; Yang, J.H.; Zhang, S.J. Exosome Biogenesis: Machinery, Regulation, and Therapeutic Implications in Cancer. *Mol Cancer* **2022**, *21*, 207. <https://doi.org/10.1186/S12943-022-01671-0>.
56. Alenquer, M.; Amorim, M.J. Exosome Biogenesis, Regulation, and Function in Viral Infection. *Viruses* **2015**, *7*, 5066. <https://doi.org/10.3390/V7092862>.
57. Takanashi, Y.; Kahyo, T.; Kamamoto, S.; Zhang, H.; Chen, B.; Ping, Y.; Mizuno, K.; Kawase, A.; Koizumi, K.; Satou, M.; et al. Ubiquitin-like 3 as a New Protein-Sorting Factor for Small Extracellular Vesicles. *Cell Struct Funct* **2022**, *47*, 1–18. <https://doi.org/10.1247/CSF.21078>.
58. Ageta, H.; Ageta-Ishihara, N.; Hitachi, K.; Karayel, O.; Onouchi, T.; Yamaguchi, H.; Kahyo, T.; Hatanaka, K.; Ikegami, K.; Yoshioka, Y.; et al. UBL3 Modification Influences Protein Sorting to Small Extracellular Vesicles. *Nature Communications* **2018**, *9*, 1–12. <https://doi.org/10.1038/s41467-018-06197-y>.
59. Menck, K.; Sönmezer, C.; Worst, T.S.; Schulz, M.; Dihazi, G.H.; Streit, F.; Erdmann, G.; Kling, S.; Boutros, M.; Binder, C.; et al. Neutral Sphingomyelinases Control Extracellular Vesicles Budding from the Plasma Membrane. *J Extracell Vesicles* **2017**, *6*, 1378056. <https://doi.org/10.1080/20013078.2017.1378056;PAGE=STRING:ARTICLE/CHAPTER>.
60. Keerthikumar, S.; Gangoda, L.; Liem, M.; Fonseka, P.; Atukorala, I.; Ozcitti, C.; Mechler, A.; Adda, C.G.; Ang, C.S.; Mathivanan, S. Proteogenomic Analysis Reveals Exosomes Are More Oncogenic than Ectosomes. *Oncotarget* **2015**, *6*, 15375–15396. <https://doi.org/10.18632/ONCOTARGET.3801>.

61. King, H.W.; Michael, M.Z.; Gleadle, J.M. Hypoxic Enhancement of Exosome Release by Breast Cancer Cells. *BMC Cancer* **2012**, *12*, 1–10. <https://doi.org/10.1186/1471-2407-12-421/FIGURES/5>.
62. Mittelbrunn, M.; Gutiérrez-Vázquez, C.; Villarroya-Beltri, C.; González, S.; Sánchez-Cabo, F.; González, M.Á.; Bernad, A.; Sánchez-Madrid, F. Unidirectional Transfer of MicroRNA-Loaded Exosomes from T Cells to Antigen-Presenting Cells. *Nature Communications* **2011**, *2*, 1–10. <https://doi.org/10.1038/ncomms1285>.
63. Budnik, V.; Ruiz-Cañada, C.; Wendler, F. Extracellular Vesicles Round off Communication in the Nervous System. *Nature Reviews Neuroscience* **2016**, *17*, 160–172. <https://doi.org/10.1038/nrn.2015.29>.
64. Liu, Y.J.; Wang, C. A Review of the Regulatory Mechanisms of Extracellular Vesicles-Mediated Inter cellular Communication. *Cell Communication and Signaling* **2023**, *21*, 1–12. <https://doi.org/10.1186/S12964-023-01103-6>.
65. Prieto-Vila, M.; Yoshioka, Y.; Ochiya, T. Biological Functions Driven by MRNAs Carried by Extracellular Vesicles in Cancer. *Front Cell Dev Biol* **2021**, *9*. <https://doi.org/10.3389/FCELL.2021.620498>.
66. Bao, Q.; Huang, Q.; Chen, Y.; Wang, Q.; Sang, R.; Wang, L.; Xie, Y.; Chen, W. Tumor-Derived Extracellular Vesicles Regulate Cancer Progression in the Tumor Microenvironment. *Front Mol Biosci* **2022**, *8*, 796385. <https://doi.org/10.3389/FMOLB.2021.796385/XML/NLM>.
67. Jurj, A.; Zanoaga, O.; Braicu, C.; Lazar, V.; Tomuleasa, C.; Irimie, A.; Berindan-neagoe, I. A Comprehensive Picture of Extracellular Vesicles and Their Contents. Molecular Transfer to Cancer Cells. *Cancers (Basel)* **2020**, *12*. <https://doi.org/10.3390/CANCERS12020298>.
68. Bao, Q.; Huang, Q.; Chen, Y.; Wang, Q.; Sang, R.; Wang, L.; Xie, Y.; Chen, W. Tumor-Derived Extracellular Vesicles Regulate Cancer Progression in the Tumor Microenvironment. *Front Mol Biosci* **2022**, *8*, 796385. <https://doi.org/10.3389/FMOLB.2021.796385/XML/NLM>.
69. Choi, D.; Montermini, L.; Kim, D.K.; Meehan, B.; Roth, F.P.; Rak, J. The Impact of Oncogenic EGFRvIII on the Proteome of Extracellular Vesicles Released from Glioblastoma Cells. *Mol Cell Proteomics* **2018**, *17*, 1948. <https://doi.org/10.1074/MCP.RA118.000644>.
70. Lawler, S.E.; Nowicki, M.O.; Ricklefs, F.L.; Chiocca, E.A. Immune Escape Mediated by Exosomal PD-L1 in Cancer. *Adv Biosyst* **2020**, *4*. <https://doi.org/10.1002/ADBI.202000017>.
71. Raimondo, S.; Pucci, M.; Alessandro, R.; Fontana, S. Extracellular Vesicles and Tumor-Immune Escape: Biological Functions and Clinical Perspectives. *Int J Mol Sci* **2020**, *21*, 2286. <https://doi.org/10.3390/IJMS21072286>.
72. Belgiovine, C.; Digifico, E.; Anfray, C.; Ummarino, A.; Andón, F.T. Targeting Tumor-Associated Macrophages in Anti-Cancer Therapies: Convincing the Traitors to Do the Right Thing. *J Clin Med* **2020**, *9*, 1–24. <https://doi.org/10.3390/JCM9103226>.
73. Mir, M.A.; Mehraj, U. Double-Crosser of the Immune System: Macrophages in Tumor Progression and Metastasis. *Current Immunology Reviews (Discontinued)* **2019**, *15*, 172–184. <https://doi.org/10.2174/1573395515666190611122818>.
74. Hoshino, A.; Costa-Silva, B.; Shen, T.L.; Rodrigues, G.; Hashimoto, A.; Tesic Mark, M.; Molina, H.; Kohsaka, S.; Di Giannatale, A.; Ceder, S.; et al. Tumour Exosome Integrins Determine Organotropic Metastasis. *Nature* **2015**, *527*, 329–335. <https://doi.org/10.1038/NATURE15756>; SUBJMETA=304,322,327,631,67,80; KWRD=CANCER+MICROENVIRONMENT,MECHANISMS+OF+DISEASE,METASTASIS.
75. Takeuchi, T. Pathogenic and Protective Roles of Extracellular Vesicles in Neurodegenerative Diseases. *J Biochem* **2021**, *169*, 181–186. <https://doi.org/10.1093/JB/MVAA131>.
76. Lee, J.Y.; Kim, H.S. Extracellular Vesicles in Neurodegenerative Diseases: A Double-Edged Sword. *Tissue Engineering and Regenerative Medicine* **2017**, *14*, 667–678. <https://doi.org/10.1007/S13770-017-0090-X>.
77. Yuan, Q.; Li, X. dong; Zhang, S. miao; Wang, H. wei; Wang, Y. liang Extracellular Vesicles in Neurodegenerative Diseases: Insights and New Perspectives. *Genes Dis* **2021**, *8*, 124–132. <https://doi.org/10.1016/j.gendis.2019.12.001>.
78. Hill, A.F. Extracellular Vesicles and Neurodegenerative Diseases. *Journal of Neuroscience* **2019**, *39*, 9269–9273. <https://doi.org/10.1523/JNEUROSCI.0147-18.2019>.

79. Raghav, A.; Singh, M.; Jeong, G.B.; Giri, R.; Agarwal, S.; Kala, S.; Gautam, K.A. Extracellular Vesicles in Neurodegenerative Diseases: A Systematic Review. *Front Mol Neurosci* **2022**, *15*, 1061076. <https://doi.org/10.3389/FNMOL.2022.1061076>.
80. Oyarce, K.; Cepeda, M.Y.; Lagos, R.; Garrido, C.; Vega-Letter, A.M.; Garcia-Robles, M.; Luz-Crawford, P.; Elizondo-Vega, R. Neuroprotective and Neurotoxic Effects of Glial-Derived Exosomes. *Front Cell Neurosci* **2022**, *16*. <https://doi.org/10.3389/FNCEL.2022.920686>.
81. Basso, M.; Bonetto, V. Extracellular Vesicles and a Novel Form of Communication in the Brain. *Front Neurosci* **2016**, *10*. <https://doi.org/10.3389/FNINS.2016.00127>.
82. Poggio, M.; Hu, T.; Pai, C.C.; Chu, B.; Belair, C.D.; Chang, A.; Montabana, E.; Lang, U.E.; Fu, Q.; Fong, L.; et al. Suppression of Exosomal PD-L1 Induces Systemic Anti-Tumor Immunity and Memory. *Cell* **2019**, *177*, 414–427.e13. <https://doi.org/10.1016/j.cell.2019.02.016>.
83. Czernek, L.; D  chler, M. Functions of Cancer-Derived Extracellular Vesicles in Immunosuppression. *Arch Immunol Ther Exp (Warsz)* **2017**, *65*, 311–323. <https://doi.org/10.1007/S00005-016-0453-3>.
84. Robbins, P.D.; Morelli, A.E. Regulation of Immune Responses by Extracellular Vesicles. *Nature Reviews Immunology* **2014**, *14*, 195–208. <https://doi.org/10.1038/nri3622>.
85. Hodge, A.L.; Baxter, A.A.; Poon, I.K.H. Gift Bags from the Sentinel Cells of the Immune System: The Diverse Role of Dendritic Cell-Derived Extracellular Vesicles. *J Leukoc Biol* **2022**, *111*, 903–920. <https://doi.org/10.1002/JLB.3RU1220-801R>.
86. Mulcahy, L.A.; Pink, R.C.; Raul, D.; Carter, F.; David, D.; Carter, R.F. Routes and Mechanisms of Extracellular Vesicle Uptake. *J Extracell Vesicles* **2014**, *3*, 24641. <https://doi.org/10.3402/JEV.V3.24641>.
87. Essandoh, K.; Yang, L.; Wang, X.; Huang, W.; Qin, D.; Hao, J.; Wang, Y.; Zingarelli, B.; Peng, T.; Fan, G.C. Blockade of Exosome Generation with GW4869 Dampens the Sepsis-Induced Inflammation and Cardiac Dysfunction. *Biochim Biophys Acta* **2015**, *1852*, 2362. <https://doi.org/10.1016/J.BBADIS.2015.08.010>.
88. Menck, K.; S  nmezer, C.; Worst, T.S.; Schulz, M.; Dihazi, G.H.; Streit, F.; Erdmann, G.; Kling, S.; Boutros, M.; Binder, C.; et al. Neutral Sphingomyelinases Control Extracellular Vesicles Budding from the Plasma Membrane. *J Extracell Vesicles* **2017**, *6*. <https://doi.org/10.1080/20013078.2017.1378056>.
89. Catalano, M.; O'Driscoll, L. Inhibiting Extracellular Vesicles Formation and Release: A Review of EV Inhibitors. *J Extracell Vesicles* **2019**, *9*, 1703244. <https://doi.org/10.1080/20013078.2019.1703244>.
90. O'Brien, K.; Breyne, K.; Ughetto, S.; Laurent, L.C.; Breakefield, X.O. RNA Delivery by Extracellular Vesicles in Mammalian Cells and Its Applications. *Nature Reviews Molecular Cell Biology* **2020**, *21*, 585–606. <https://doi.org/10.1038/s41580-020-0251-y>.
91. He, Y.; Wang, K.; Lu, Y.; Sun, B.; Sun, J.; Liang, W. Monensin Enhanced Generation of Extracellular Vesicles as Transfersomes for Promoting Tumor Penetration of Pyropheophorbide-a from Fusogenic Liposome. *Nano Lett* **2022**, *22*, 1415–1424. https://doi.org/10.1021/ACS.NANOLETT.1C04962/SUPPL_FILE/NL1C04962_SI_001.PDF.
92. Hao, Y.; Song, H.; Zhou, Z.; Chen, X.; Li, H.; Zhang, Y.; Wang, J.; Ren, X.; Wang, X. Promotion or Inhibition of Extracellular Vesicle Release: Emerging Therapeutic Opportunities. *Journal of Controlled Release* **2021**, *340*, 136–148. <https://doi.org/10.1016/j.jconrel.2021.10.019>.
93. Jackson Cullison, S.R.; Flemming, J.P.; Karagoz, K.; Wermuth, P.J.; Mahoney, M.G. Mechanisms of Extracellular Vesicle Uptake and Implications for the Design of Cancer Therapeutics. *Journal of extracellular biology* **2024**, *3*, e70017. <https://doi.org/10.1002/JEX2.70017>;PAGE:STRING:ARTICLE/CHAPTER.
94. Lener, T.; Gimona, M.; Aigner, L.; B  rger, V.; Buzas, E.; Camussi, G.; Chaput, N.; Chatterjee, D.; Court, F.A.; del Portillo, H.A.; et al. Applying Extracellular Vesicles Based Therapeutics in Clinical Trials - An ISEV Position Paper. *J Extracell Vesicles* **2015**, *4*. <https://doi.org/10.3402/JEV.V4.30087>.
95. Ageta, H.; Ageta-Ishihara, N.; Hitachi, K.; Karayel, O.; Onouchi, T.; Yamaguchi, H.; Kahyo, T.; Hatanaka, K.; Ikegami, K.; Yoshioka, Y.; et al. UBL3 Modification Influences Protein Sorting to Small Extracellular Vesicles. *Nat Commun* **2018**, *9*. <https://doi.org/10.1038/S41467-018-06197-Y>.
96. Zhang, H.; Chen, B.; Waliullah, A.S.M.; Aramaki, S.; Ping, Y.; Takanashi, Y.; Zhang, C.; Zhai, Q.; Yan, J.; Oyama, S.; et al. A New Potential Therapeutic Target for Cancer in Ubiquitin-Like Proteins—UBL3. *International Journal of Molecular Sciences* **2023**, *24*, 1231. <https://doi.org/10.3390/IJMS24021231>.

97. Stremersch, S.; De Smedt, S.C.; Raemdonck, K. Therapeutic and Diagnostic Applications of Extracellular Vesicles. *Journal of Controlled Release* **2016**, *244*, 167–183. <https://doi.org/10.1016/j.jconrel.2016.07.054>.
98. Du, S.; Guan, Y.; Xie, A.; Yan, Z.; Gao, S.; Li, W.; Rao, L.; Chen, X.; Chen, T. Extracellular Vesicles: A Rising Star for Therapeutics and Drug Delivery. *Journal of Nanobiotechnology* **2023**, *21*, 1–51. <https://doi.org/10.1186/S12951-023-01973-5>.
99. Han, Y.; Jones, T.W.; Dutta, S.; Zhu, Y.; Wang, X.; Narayanan, S.P.; Fagan, S.C.; Zhang, D. Overview and Update on Methods for Cargo Loading into Extracellular Vesicles. *Processes* **2021**, *9*, 1–19. <https://doi.org/10.3390/PR9020356>.
100. Lu, Y.; Godbout, K.; Lamothe, G.; Tremblay, J.P. CRISPR-Cas9 Delivery Strategies with Engineered Extracellular Vesicles. *Mol Ther Nucleic Acids* **2023**, *34*. <https://doi.org/10.1016/j.omtn.2023.102040>.
101. Sutaria, D.S.; Badawi, M.; Phelps, M.A.; Schmittgen, T.D. Achieving the Promise of Therapeutic Extracellular Vesicles: The Devil Is in Details of Therapeutic Loading. *Pharm Res* **2017**, *34*, 1053–1066. <https://doi.org/10.1007/S11095-017-2123-5>.
102. Dooley, K.; McConnell, R.E.; Xu, K.; Lewis, N.D.; Haupt, S.; Youniss, M.R.; Martin, S.; Sia, C.L.; McCoy, C.; Moniz, R.J.; et al. A Versatile Platform for Generating Engineered Extracellular Vesicles with Defined Therapeutic Properties. *Molecular Therapy* **2021**, *29*, 1729–1743. <https://doi.org/10.1016/j.ymthe.2021.01.020>.
103. Wang, L.; Wang, D.; Ye, Z.; Xu, J. Engineering Extracellular Vesicles as Delivery Systems in Therapeutic Applications. *Advanced Science* **2023**, *10*, 2300552. <https://doi.org/10.1002/ADVS.202300552>.
104. Li, Z.; Zhao, R.; Wu, X.; Sun, Y.; Yao, M.; Li, J.; Xu, Y.; Gu, J. Identification and Characterization of a Novel Peptide Ligand of Epidermal Growth Factor Receptor for Targeted Delivery of Therapeutics. *The FASEB Journal* **2005**, *19*, 1978–1985. <https://doi.org/10.1096/FJ.05-4058COM>.
105. Walker, S.; Busatto, S.; Pham, A.; Tian, M.; Suh, A.; Carson, K.; Quintero, A.; Lafrence, M.; Malik, H.; Santana, M.X.; et al. Extracellular Vesicle-Based Drug Delivery Systems for Cancer Treatment. *Theranostics* **2019**, *9*, 8001. <https://doi.org/10.7150/THNO.37097>.
106. Liu, J.; Chen, Y.; Pei, F.; Zeng, C.; Yao, Y.; Liao, W.; Zhao, Z. Extracellular Vesicles in Liquid Biopsies: Potential for Disease Diagnosis. *Biomed Res Int* **2021**, *2021*, 6611244. <https://doi.org/10.1155/2021/6611244>.
107. Revenfeld, A.L.S.; Bæk, R.; Nielsen, M.H.; Stensballe, A.; Varming, K.; Jørgensen, M. Diagnostic and Prognostic Potential of Extracellular Vesicles in Peripheral Blood. *Clin Ther* **2014**, *36*, 830–846. <https://doi.org/10.1016/j.clinthera.2014.05.008>.
108. Boukouris, S.; Mathivanan, S. Exosomes in Bodily Fluids Are a Highly Stable Resource of Disease Biomarkers. *Proteomics Clin Appl* **2015**, *9*, 358–367. <https://doi.org/10.1002/PRCA.201400114>.
109. Yoshioka, Y.; Kosaka, N.; Konishi, Y.; Ohta, H.; Okamoto, H.; Sonoda, H.; Nonaka, R.; Yamamoto, H.; Ishii, H.; Mori, M.; et al. Ultra-Sensitive Liquid Biopsy of Circulating Extracellular Vesicles Using ExoScreen. *Nature Communications* **2014**, *5*, 1–8. <https://doi.org/10.1038/ncomms4591>.
110. Holtzman, J.; Lee, H. Emerging Role of Extracellular Vesicles in the Respiratory System. *Exp Mol Med* **2020**, *52*, 887–895. <https://doi.org/10.1038/S12276-020-0450-9>; SUBJMETA=1784,1785,443,631,692,699; KWRD=RESPIRATION,RESPIRATORY+TRACT+DISEASES.
111. Thompson, A.G.; Gray, E.; Heman-Ackah, S.M.; Mäger, I.; Talbot, K.; El Andaloussi, S.; Wood, M.J.; Turner, M.R. Extracellular Vesicles in Neurodegenerative Disease — Pathogenesis to Biomarkers. *Nature Reviews Neurology* **2016**, *12*, 346–357. <https://doi.org/10.1038/nrneurol.2016.68>.
112. Li, P.; Kaslan, M.; Lee, S.H.; Yao, J.; Gao, Z. Progress in Exosome Isolation Techniques. *Theranostics* **2017**, *7*, 789–804. <https://doi.org/10.7150/THNO.18133>.
113. Welsh, J.A.; Goberdhan, D.C.I.; O'Driscoll, L.; Buzas, E.I.; Blenkiron, C.; Bussolati, B.; Cai, H.; Di Vizio, D.; Driedonks, T.A.P.; Erdbrügger, U.; et al. Minimal Information for Studies of Extracellular Vesicles (MISEV2023): From Basic to Advanced Approaches. *J Extracell Vesicles* **2024**, *13*. <https://doi.org/10.1002/JEV2.12404>.
114. Contreras-Naranjo, J.C.; Wu, H.J.; Ugaz, V.M. Microfluidics for Exosome Isolation and Analysis: Enabling Liquid Biopsy for Personalized Medicine. *Lab Chip* **2017**, *17*, 3558. <https://doi.org/10.1039/C7LC00592J>.
115. Riazifar, M.; Mohammadi, M.R.; Pone, E.J.; Yeri, A.; Lasser, C.; Segaliny, A.I.; McIntyre, L.L.; Shelke, G.V.; Hutchins, E.; Hamamoto, A.; et al. Stem Cell-Derived Exosomes as Nanotherapeutics for Autoimmune and Neurodegenerative Disorders. *ACS Nano* **2019**, *13*, 6670–6688. <https://doi.org/10.1021/ACS.NANO.9B01004>.

116. Buschmann, D.; Mussack, V.; Byrd, J.B. Separation, Characterization, and Standardization of Extracellular Vesicles for Drug Delivery Applications. *Adv Drug Deliv Rev* **2021**, *174*, 348–368. <https://doi.org/10.1016/J.ADDR.2021.04.027>.
117. Mateescu, B.; Kowal, E.J.K.; van Balkom, B.W.M.; Bartel, S.; Bhattacharyya, S.N.; Buzás, E.I.; Buck, A.H.; de Candia, P.; Chow, F.W.N.; Das, S.; et al. Obstacles and Opportunities in the Functional Analysis of Extracellular Vesicle RNA - An ISEV Position Paper. *J Extracell Vesicles* **2017**, *6*. <https://doi.org/10.1080/20013078.2017.1286095>.
118. Gupta, D.; Zickler, A.M.; El Andaloussi, S. Dosing Extracellular Vesicles. *Adv Drug Deliv Rev* **2021**, *178*, 113961. <https://doi.org/10.1016/J.ADDR.2021.113961>.
119. Beetler, D.J.; Di Florio, D.N.; Bruno, K.A.; Ikezu, T.; March, K.L.; Cooper, L.T.; Wolfram, J.; Fairweather, D.L. Extracellular Vesicles as Personalized Medicine. *Mol Aspects Med* **2023**, *91*. <https://doi.org/10.1016/j.mam.2022.101155>.
120. Armstrong, J.P.K.; Stevens, M.M. Strategic Design of Extracellular Vesicle Drug Delivery Systems. *Adv Drug Deliv Rev* **2018**, *130*, 12–16. <https://doi.org/10.1016/J.ADDR.2018.06.017>.

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