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# Spectroscopic Study of a Novel Binaphthyl Amine Fluorescent Probe for Chiral Recognition of DL-Lysine

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Keywords: Lysine Chiral Recognition; BINAM-based Fluorescent Probes; Enantioselectivity; Fluorescence Analysis



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Article

# Spectroscopic Study of a Novel Binaphthyl Amine Fluorescent

Running Title: Probe for Chiral Recognition of D/L-Lysine

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**Abstract:** Lysine plays a crucial role in promoting development, enhancing immune function, and improving the function of central nervous system tissues. The two configurational isomers of amino acids have significantly different effects. Currently, methods for chiral recognition of lysine all have drawbacks such as expensive equipment and complicated detection processes, whereas fluorescence analysis boasts high sensitivity, strong selectivity, and simple operation. In this study, we synthesized four novel Binaphthyl-Amine (BINAM)-based fluorescent probes capable of specifically identifying the L-configuration of lysine among the twenty amino acids constituting human proteins. The enantiomeric fluorescence enhancement ratio ( $ef$  or  $\Delta I_L/\Delta I_D$ ) reached up to 15.29, demonstrating high enantioselectivity. We also evaluated the probe's recognition ability under different pH, reaction times, and metal ion conditions, as well as the Limit of Detection of the probe. Our findings indicate that this probe is a highly stable tool for detecting chiral lysine.

**Keywords:** lysine chiral recognition; BINAM-based fluorescent probes; enantioselectivity; fluorescence analysis

## 1. Introduction

The presence of chiral amino acids in living organisms is rooted in the origins of life and the natural selection of molecular synthesis. The early Earth environment likely harbored certain physical or chemical conditions that led to chiral selection, laying the foundation for the formation of life molecules. Within living organisms, the molecular synthesis process exhibits selectivity for specific chiral amino acids, with enzymes and catalysts favoring the efficient synthesis or utilization of L-configured amino acids. Throughout the evolutionary process, organisms gradually developed a preference for specific chiral amino acids, enhancing their chances of survival and reproduction.[1] This selectivity is associated with affinity for particular configurations, catalytic efficiency, and molecular stability, ultimately influencing normal biological functions and adaptation to the environment. In summary, the existence of chiral amino acids is a result of the interplay between the early Earth environment and biological evolution, providing crucial groundwork for the formation and development of life systems.[2,3]

Amino acids are the basic units that constitute the macromolecule proteins in living organisms and are also crucial components in life processes.[4] In the natural amino acids, apart from glycine, all other amino acids exist in two stereoisomeric forms, L and D. During protein synthesis, living organisms selectively utilize L-configured amino acids, highlighting the biological preference for this stereoisomer. The specific spatial arrangement of L-amino acids is essential for proper protein folding, maintaining the structural integrity critical for biological activity.[5] Enzymes, hormones, and other biomolecules often exhibit high selectivity for L-configured amino acids, influencing metabolic pathways and biological functions. Additionally, the role of amino acid chirality extends to drug design, where the preferential use of L-configured amino acids enhances the biological compatibility and efficacy of pharmaceuticals.[6] The role of D-amino acids in living organisms is

relatively limited, primarily demonstrated in specific areas such as antimicrobial activity, drug design, research tools, and applications in veterinary medicine and animal feed, showcasing certain biological functions and potential utility.[7] Overall, the chiral nature of amino acids profoundly impacts the structure, function, and regulation of biological systems, reflecting the evolutionary adaptability and survival advantages associated with specific stereoisomers.

Lysine is a basic amino acid, and most higher animals cannot synthesize lysine.[8] Therefore, lysine is an essential amino acid that must be ingested in sufficient quantities from the diet to maintain protein synthesis.[9] L-lysine is considered an essential amino acid of great importance to human health, playing a significant role in enhancing immunity, promoting calcium absorption, and improving central nervous system function.[10] Abnormal metabolism of L-lysine may lead to certain cancers.[11] L-lysine has multiple functions, including enhancing immunity, promoting skeletal muscle growth, facilitating fat metabolism, and alleviating anxiety.[12–15] Additionally, it can also produce synergistic effects with other nutrients and promote their absorption, improving the utilization efficiency of various nutrients, and thereby better expressing the physiological functions of different nutrients.[16,17] A deficiency in D-lysine may lead to adverse health reactions, including uremia. There are differences in the efficiency with which D-lysine and L-lysine are absorbed and utilized by the body, with D-lysine having a lower absorption and utilization rate, while the primary biological activity is provided by L-lysine.[18] Lysine plays a crucial role in the function of proteins. Due to its widespread and reliable role as a drug target, lysine has received extensive attention.[19] Therefore, it also plays an important role in distinguishing different enantiomers.

Although there are already some methods for detecting lysines, such as the luminol chemiluminescence method,[20] gas chromatography-mass spectrometry,[21] synchrotron X-ray powder diffraction, and thermogravimetric analysis,[22] these methods come with the disadvantages of expensive equipment and cumbersome detection processes. Therefore, there is a need for a simple and efficient method for detecting lysine. Fluorescence analysis, with its advantages of high sensitivity, strong selectivity, low sample requirement, and simplicity, has attracted great interest from researchers. However, the current synthesis of many fluorescent probes is challenging, and they only exhibit limited enantioselectivity.[23] Therefore, preparing efficient fluorescent probes for the chiral recognition of lysine via a simple synthetic route remains a significant challenge.

In recent years, axially chiral 1,1'-Binaphthyl-2,2'-diphenyl phosphine(BINAP), 1,1'-binaphthyl-2,2'-diol(BINOL) (Figure 1), and their derivatives have become one of the most successful chiral ligands/catalysts in asymmetric catalysis and are widely used in various enantioselective catalyzes.[24] However, 2,2'-Diamino-1,1'-binaphthalene (BINAM), which also possesses axial chirality, has seen very limited application in asymmetric synthesis, with relatively few related publications. BINAM has long been considered by researchers as a potential fluorescent detector for the efficient chiral recognition of  $\alpha$ -phenylethylamine and tryptophan enantiomers.[25] Previous studies have shown that probes based on BINAM can highly selectively recognize lysine in aqueous solutions, but the results indicated a significant fluorescent response for both configurations, thus unable to distinguish the configuration of lysine.[26] This article has discovered and synthesized a fluorescent probe that can selectively recognize L-lysine with high selectivity in physiological conditions. The results indicate that other amino acids do not affect the recognition effect, and the probe has the advantages of being acid and alkali resistant, sensitive, and capable of recognition over a longer period. We also studied the recognition effect of the probe in the presence of certain metal ions and the substitution sites of the probe. The results are presented below.

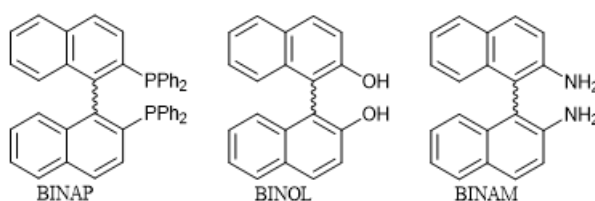


Figure 1. BINAP, BINOL and BINAM.

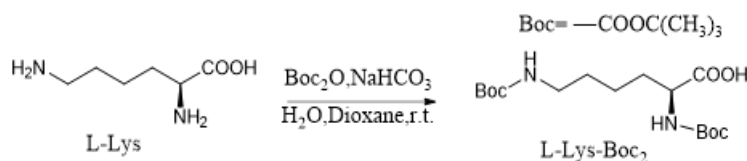
## 2. Experimental

### 2.1. Reagents and Apparatus

$^1\text{H}$ ,  $^{13}\text{C}$ , and  $^{19}\text{F}$  NMR spectra were measured using a Bruker AM600 NMR spectrometer. The chemical shifts of the NMR spectra are given in ppm relative to the internal reference TMS (1H, 0.00ppm) for protons. HRMS spectral data were recorded on a Bruker Micro-TOF-QII mass spectrometer. Fluorescence spectra were obtained at 298 K using a (F97 Pro) spectrofluorophotometer with an excitation wavelength of 365 nm. All chemicals involved in the experiments were purchased from Adamas Chemistry Co., Ltd (Shanghai, China). The chemicals were obtained from commercial suppliers and used without further purification. All solvents used in the optical spectroscopy studies were of HPLC or spectroscopic grade.

### 2.2. Synthesis of L-Lys-Boc<sub>2</sub>

L-Lysine (731 mg, 5 mmol) was added to 100 mL of H<sub>2</sub>O containing NaHCO<sub>3</sub> (840 mg, 10 mmol), and, under stirring, Boc<sub>2</sub>O (1500 mg, 6.87 mmol) dissolved in 50 mL of dioxane was slowly added, followed by stirring at room temperature for 12 hours. Boc<sub>2</sub>O (1500 mg, 6.87 mmol) dissolved in 50 mL of dioxane was then added slowly again under stirring, and the reaction progress was monitored by thin-layer chromatography (TLC). After the reaction proceeded for an additional 12 hours, it was extracted three times with 50 mL of DCM and washed three times with 20 mL of H<sub>2</sub>O. The organic solvent was removed under reduced pressure, and the crude product was purified by silica gel column chromatography, eluting with DCM/MeOH (30:1) to obtain a yellow gel-like L-Lys-Boc<sub>2</sub> with a yield of 84% (1455 mg, 4.2 mmol), as shown in Scheme 1. Similarly, a colorless gel-like D-Lys-Boc<sub>2</sub> was obtained using the same method, with a yield of 80%.

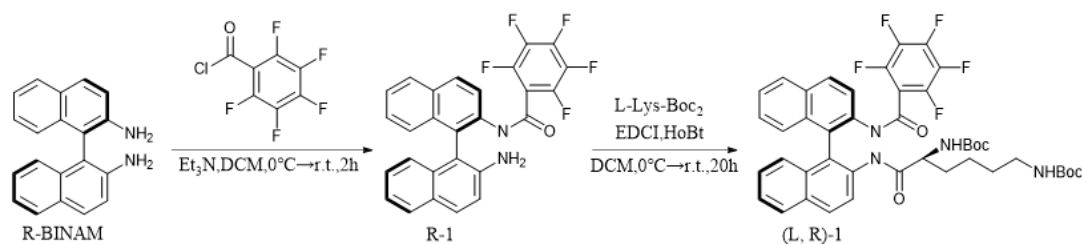


**Scheme 1.** Synthesis of L-Lys-Boc<sub>2</sub>.

### 2.3. Synthesis of (L, R)-1

To a solution of R-BINAM (284 mg, 1 mmol) in DCM (30 mL), triethylamine (200  $\mu\text{L}$ , 1.5 mmol) was added. After cooling the mixture to 0°C, 2,3,4,5,6-pentafluorobenzoyl chloride (230 mg, 1 mmol) was slowly added dropwise, and the mixture was stirred at room temperature for 8 hours. The progress of the reaction was monitored by thin-layer chromatography (TLC). After the reaction was complete, the mixture was washed three times with 20 mL of H<sub>2</sub>O. The organic solvent was removed under reduced pressure, and the crude product was purified by silica gel column chromatography, eluting with DCM/PE (1:1) to obtain a pale-yellow powder R-1, with a yield of 65% (310 mg, 0.65 mmol). The same method yielded a pale-yellow powder S-1, with a yield of 59%. Next, R-1 (239 mg, 0.5 mmol) was added to a DCM solution (30 mL) and stirred at 0°C. After cooling the system to 0°C, HOBt (81 mg, 0.6 mmol) and EDCI (93 mg, 0.6 mmol) were added, and the mixture was stirred for 2 minutes before adding L-Lys-Boc<sub>2</sub> (346 mg, 1 mmol). The system was then transferred to room temperature and stirred for 20 hours, with the progress of the reaction monitored by TLC. After the reaction was complete, the mixture was washed three times with 20 mL of H<sub>2</sub>O. In the back of removing the organic solvent under reduced pressure, the crude product was purified by silica gel column chromatography, eluting with DCM/MeOH (50:1), and following removing the organic solvent under reduced pressure, a white powder defined as (L, R)-1 was obtained, with a yield of 34% (137 mg, 0.17 mmol), as shown in Scheme 2. L/D is the configuration of Lys-Boc<sub>2</sub> and R/S is the configuration of BINAM. The same method yielded (L, S)-1 with a yield of 32%, (D, R)-1 with a yield of 38%, and (D, S)-1 with a yield of 28%.





**Scheme 2.** Synthesis of (L, R)-1.

The four fluorescent probes were characterized by  $^1\text{H}$  NMR,  $^{13}\text{C}$  NMR,  $^{19}\text{F}$  NMR, and MS, with the spectral data provided in the supplemental Information.

#### 2.4. Preparation of Samples for Fluorescence Analysis

Solutions of four sensors were separately prepared at a concentration of 2.0 mM in ethanol, and amino acids were prepared at a concentration of 200 mM in Deionized water as stock solutions, to be used immediately upon preparation. In the study of selectivity towards amino acids, 30  $\mu\text{L}$  of the sensor/EtOH stock solution was mixed with 30  $\mu\text{L}$  of the amino acid/  $\text{H}_2\text{O}$  stock solution, left to stand for 10 minutes, and then diluted with PBS buffer (pH=7.4) to 3000  $\mu\text{L}$  to form the detection solution. In the study of Amino acids selectivity, 30  $\mu\text{L}$  of the L-Lys stock solution was first mixed with 30  $\mu\text{L}$  of the stock solution of another amino acid, left to stand for 10 minutes, followed by the addition of 30  $\mu\text{L}$  of the sensor/EtOH stock solution, and then diluted with PBS buffer to 3000  $\mu\text{L}$  to form the detection solution. For the study recognition in different pH, 30  $\mu\text{L}$  of the sensor/EtOH stock solution was mixed with 30  $\mu\text{L}$  of the Lys/  $\text{H}_2\text{O}$  stock solution, left to stand for 10 minutes, and then diluted with PBS buffer at the corresponding pH to 3000  $\mu\text{L}$  to form the detection solution, adjusting the pH with dilute solutions of NaOH and HCl. In the study of the detection limit of the probes, Lys was re-prepared at a concentration of 20 mM in Deionized water as the stock solution. 30  $\mu\text{L}$  of the sensor/EtOH stock solution was mixed with 3x  $\mu\text{L}$  of the Lys/  $\text{H}_2\text{O}$  stock solution, left to stand for 10 minutes, and then diluted with buffer to 3000  $\mu\text{L}$  to form the detection solution, where x corresponds to the concentration ratio of Lys. In the study of the effect of metal ions on probe recognition, 30  $\mu\text{L}$  of the sensor/EtOH stock solution was first mixed with 30  $\mu\text{L}$  of a 4.0 mM solution of metal chloride, left to stand for 10 minutes, followed by the addition of 30  $\mu\text{L}$  of the L-Lys stock solution, and then diluted with PBS buffer at pH 7.4 to 3000  $\mu\text{L}$  to form the detection solution.

In the experiments described, the volume concentration of EtOH in the detection solution is 1%, and the concentration of the sensor is 0.02 mM. Except for the experiments determining the detection limit, the concentration of amino acids is maintained at 2 mM. All fluorescence data, except for those related to stability studies, were obtained within 3 hours. Unless specifically stated, the data were obtained at room temperature with PBS buffer (pH=7.4, EtOH/PBS=1/99, v/v) solution as the solvent. Reaction time : 30 min. PMT: 650V. Error bars from three independent experiments.  $\lambda_{\text{exc}} = 365$  nm. Slit: 20/10 nm.

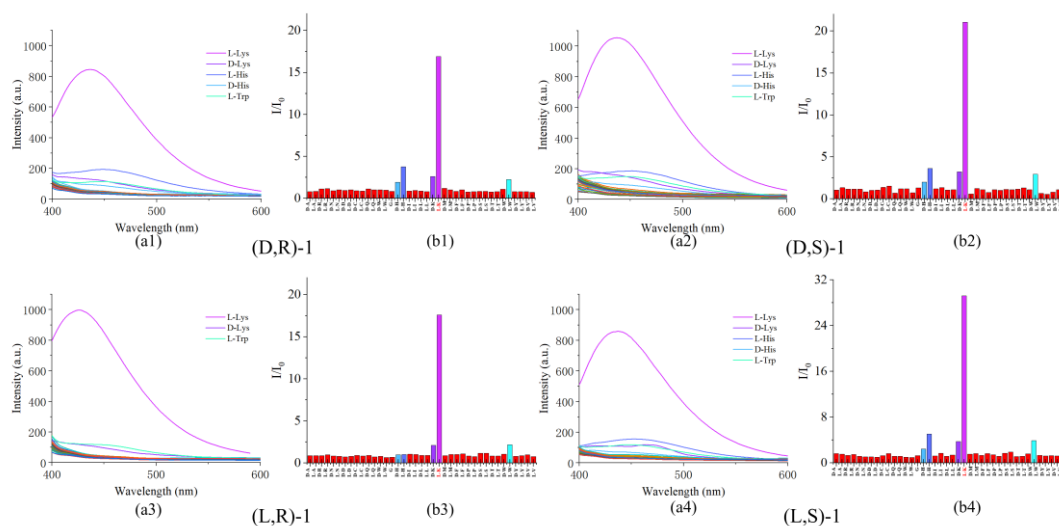
### 3. Result and Discussion

#### 3.1. Chiral Recognition of D/L-Lysine

Based on Binaphthyl Amine, we designed and synthesized four novel probes, and found that all four probes could dissolve in organic solvents, particularly ethanol solution. Subsequently, we diluted them into PBS buffer (pH=7.4) as the reaction buffer (Experimental section).

Firstly, we investigated the fluorescent response of these four probes towards 20 amino acids and their chiral forms. We discovered that all four probes exhibited significant fluorescence enhancement for L-lysine at 437nm (Figure 2. a). Then we use "I" to represent the fluorescence intensity of the probe in response to the component recognition, and "I<sub>0</sub>" denotes the initial fluorescence intensity of the probe. The values of I/I<sub>0</sub> for L-Lys were 16.89, 21.08, 17.60 and 29.23, respectively. In contrast, His and Trp exhibited values between 3.0-5.0, while the remaining amino

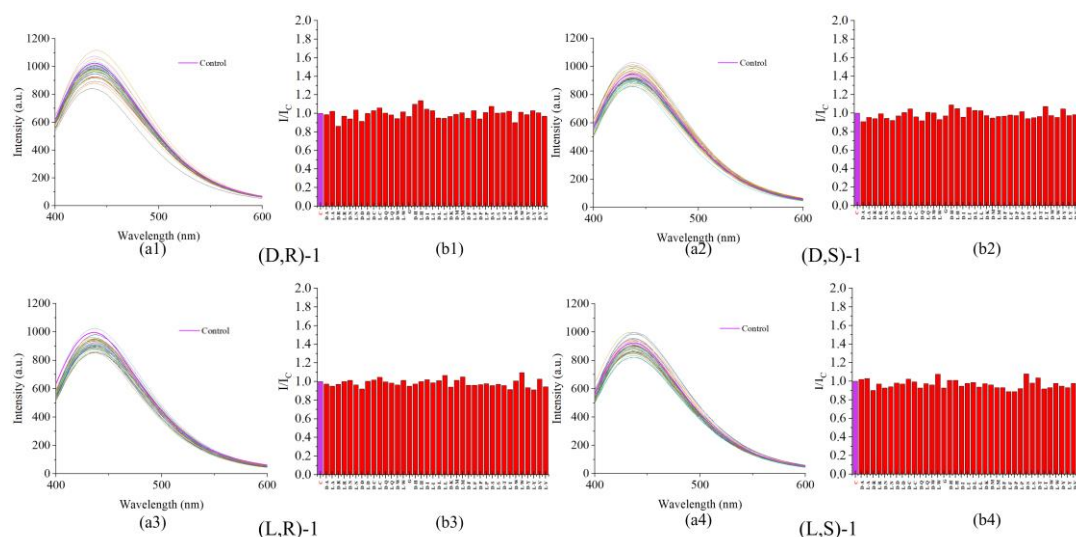
acids ranged between 0.8 and 1.5 (Figure 2. b). The chiral recognition efficacy,  $ef$  ( $ef = \text{enantiomeric fluorescence enhancement ratio} = [I_L - I_0]/[I_D - I_0] = \Delta I_L/\Delta I_D$ ,  $I_0$ , the fluorescence intensity of the probe,  $I_L$  or  $I_D$ , fluorescence intensity with L or D substrate) was calculated as follows:  $ef [(D, R)-1] = 9.99$ ,  $ef [(D, S)-1] = 9.12$ ,  $ef [(L, R)-1] = 15.29$  and  $ef [(L, S)-1] = 10.43$ . This demonstrates that all four probes are capable of efficiently and specifically recognizing L-Lys within chiral amino acids.



**Figure 2.** (a) Fluorescence spectra of probe (0.02 mM) to various amino acids (2 mM). (b) Plot of the fluorescence enhancement  $I/I_0$  at 437 nm. (The sequence of amino acids tested includes: D/L-Ala(A), D/L-Arg(R), D/L-Asp(N), D/L-Asp(P), D/L-Cys(C), D/L-Glu(Q), D/L-Glu(E), Gly(G), D/L-His(H), D/L-Iso(I), D/L-Leu(L), D/L-Lys(K), D/L-Met(M), D/L-Phe(F), D/L-Pro(P), D/L-Ser(S), D/L-Thr(T), D/L-Trp(W), D/L-Tyr(Y), and D/L-Val(V).).

### 3.2. Interference by Other Amino Acids in Probe Recognition

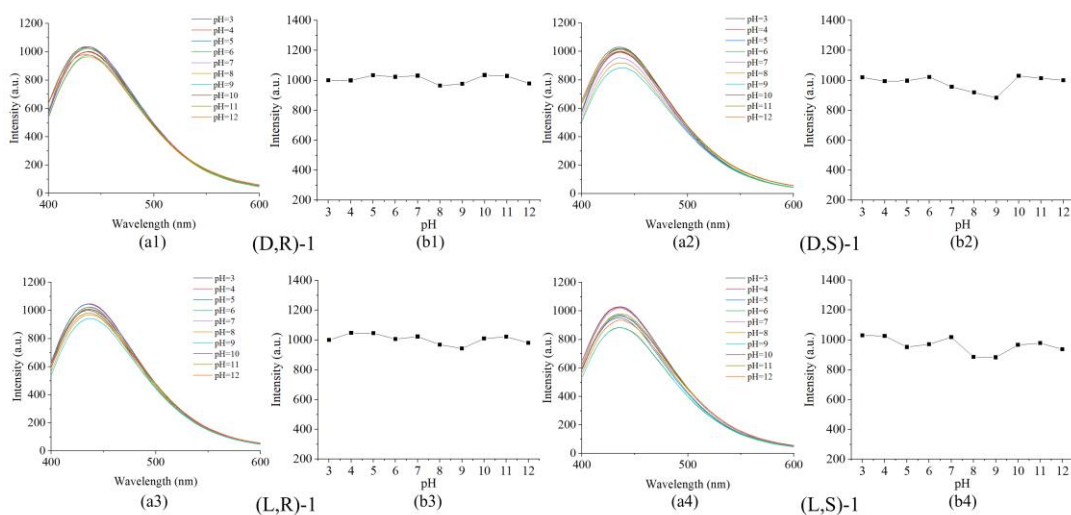
In living organisms, multiple amino acids are simultaneously present. It is crucial to study the recognition of chiral lysine in the presence of various amino acids for newly synthesized probes. Under physiological conditions (PBS buffer, pH=7.4), the fluorescence absorption intensity at 437nm was measured for the other 19 amino acids and their chiral forms, each at the same concentration as L-lysine. Strong intensity peaks were observed for all of them (Figure 3. a). To compare this interference, we set " $I_c$ " as the fluorescence intensity when the probe recognizes L-lysine and " $I$ " as the fluorescence intensity when the probe recognizes L-lysine in the presence of other amino acids. Our research results indicate that all values of  $I/I_c$  range between 0.9 and 1.1. Therefore, in the presence of other amino acids, the probe can effectively recognize L-lysine (Figure 3. b).



**Figure 3.** (a) Fluorescence spectra of probe (0.02mM) with L-Lys (2mM) in the presence of other amino acids (2mM). (b) Plot of the fluorescence enhancement  $I/I_c$  at 437 nm. (The sequence of amino acids tested includes: Control Group(C), D/L-Ala(A), D/L-Arg(R), D/L-Asp(N), D/L-Asp(P), D/L-Cys(C), D/L-Glu(Q), D/L-Glu(E), Gly(G), D/L-His(H), D/L-Iso(I), D/L-Leu(L), D-Lys(K), D/L-Met(M), D/L-Phe(F), D/L-Pro(P), D/L-Ser(S), D/L-Thr(T), D/L-Trp(W), D/L-Tyr(Y), and D/L-Val(V).).

### 3.3. Recognition in Different pH

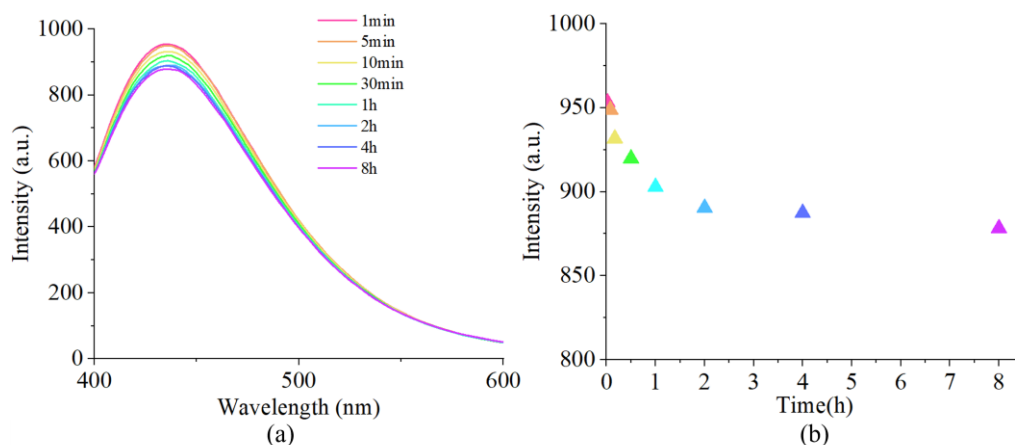
We anticipate that these novel probes can be utilized in multiple fields, as the stability of the probe's recognition of L-lysine across different pH levels is crucial for its widespread applicability. The results indicate that within the pH range of 3 to 12, the recognition efficiency of the probe for L-lysine is similar to its fluorescence intensity in PBS (pH 7.4). Therefore, pH does not affect the probe's ability to recognize L-lysine (Figure 4).



**Figure 4.** (a) Fluorescence spectra of probe (0.02mM) with L-Lys (2mM) with a pH range of 3 to 12. (b) Plot of the fluorescence intensity at 437 nm.

### 3.4. The Reaction Time of the Probes

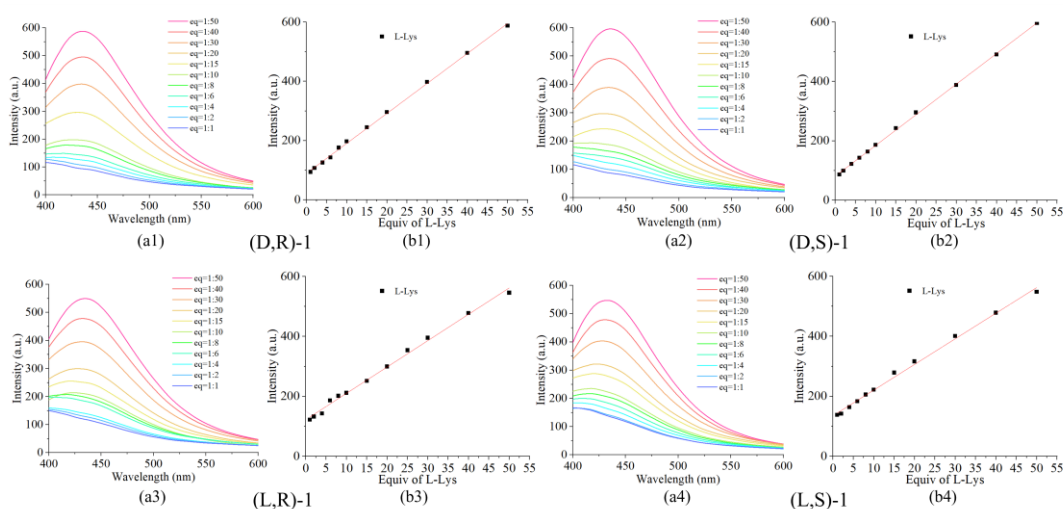
We separately tested the recognition capabilities of four probes towards L-lysine at various time intervals to determine the reaction efficiency and stability of the probes. Taking (L, R)-1 as an example, the study results show that the reaction between the probe and L-Lys is very rapid, allowing for recognition and detection within 1 minute. The recognition effect of the probe at 8 hours still retains 92.2% of its initial efficiency (Figure 5).



**Figure 5.** (a) Fluorescence spectra of probe (0.02mM) with L-Lys (2mM) at 1 min, 5 min, 10 min, 30 min, 1 h, 2 h, 4 h, and 8 h. (b) Plot of the fluorescence intensity at 437 nm.

### 3.5. Limit of Detection

The study examined the fluorescence spectra of probes for L-lysine at different concentrations, as shown in Figure 6. The concentration of the probe was fixed at 0.02 mM, while the concentration of L-lysine ranged from 0.02 mM (eq=1:1) to 1 mM (eq=1:50). The fluorescence intensity significantly increased with the concentration of L-Lys, and the fluorescence intensity at  $\lambda=437$  nm was fitted using Origin. The emission showed a linear positive correlation, with adjusted R-squared values for (D, R)-1, (D, S)-1, (L, R)-1, (L, S)-1 being: 0.99883, 0.9994, 0.99455, 0.99536, respectively. Based on the limit of detection formula  $LOD=3\sigma/K$  (where  $\sigma$  is the standard deviation of the probe samples, and K is the slope of the fitted line), LOD for L-Lys using (D, R)-1, (D, S)-1, (L, R)-1, (L, S)-1 were calculated to be  $2.12\times 10^{-6}M$ ,  $9.38\times 10^{-6}M$ ,  $5.98\times 10^{-6}M$  and  $5.18\times 10^{-6}M$ .

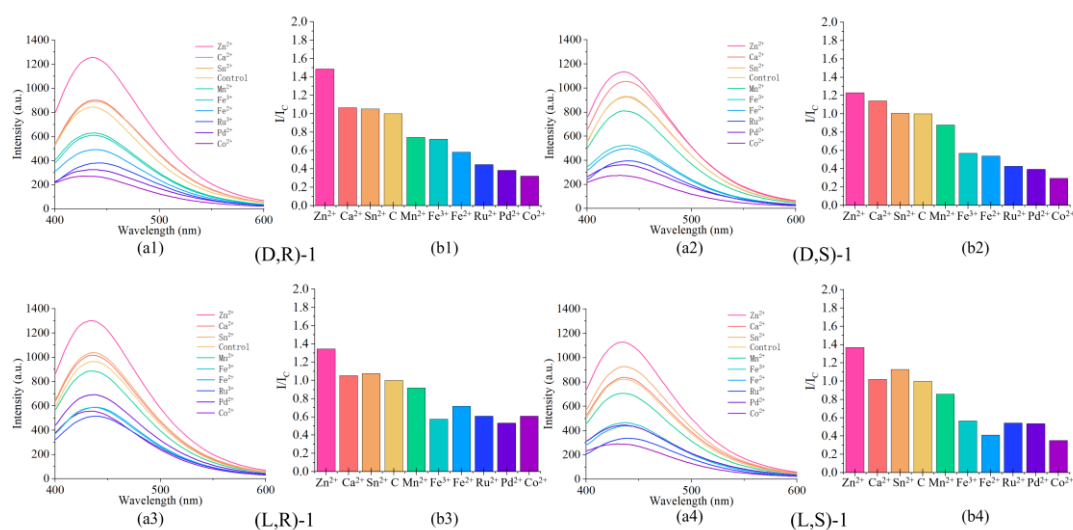


**Figure 6.** (a) Fluorescence spectra for the gradient recognition of 1 to 50 equivalents of L-Lys by the probe (0.02 mM) concentration. (b) Plot of the fluorescence intensity at 437 nm.



### 3.6. The Effect of Metal Ions

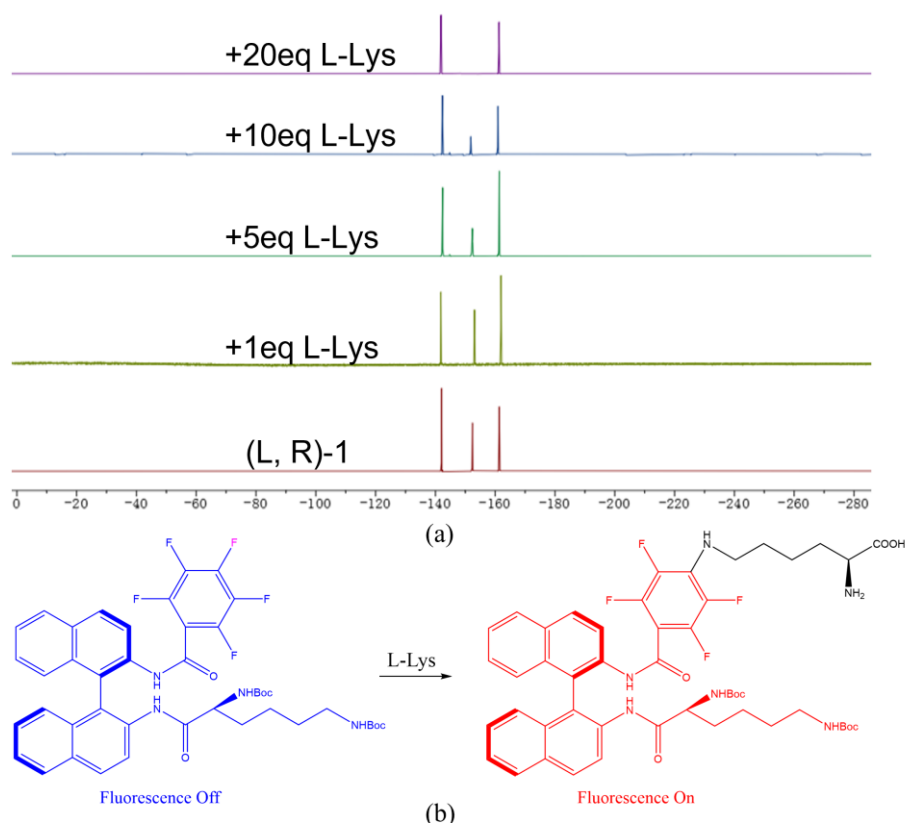
Metal ions can indeed form complexes that may impact the recognition effect of probes. In this study, the recognition effect of the probe on L-lysine was investigated in the presence of various metal ions:  $\text{Co}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Pd}^{2+}$ ,  $\text{Ru}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Sn}^{2+}$  and  $\text{Ca}^{2+}$ . To ensure that anions did not interfere, all metal salts were chosen in their chloride forms. Among these metal ions,  $\text{Zn}^{2+}$  notably enhanced the probe's recognition effect on L-Lys, showing an increase of at least 1.2 to 1.5 times compared to the control group.  $\text{Sn}^{2+}$  and  $\text{Ca}^{2+}$  also slightly enhanced the recognition effect (Figure 7). However, the presence of the remaining metal ions had a negative impact on the probe's recognition effect on L-Lys, indicating interference or inhibition. The presence of other metal ions will reduce the recognition effect of the probe on L-lysine, especially in the presence of  $\text{Co}^{2+}$ ,  $\text{Pd}^{2+}$ ,  $\text{Ru}^{3+}$ ,  $\text{Fe}^{2+}$  and  $\text{Fe}^{3+}$  ions, where the recognition efficiency of the probe for L-lysine is only about half of its original level.



**Figure 7.** (a) Fluorescence spectrum upon addition of 2 equivalents of:  $\text{Co}^{2+}$ ,  $\text{Fe}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Mn}^{2+}$ ,  $\text{Pd}^{2+}$ ,  $\text{Ru}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Sn}^{2+}$ , and  $\text{Ca}^{2+}$  to the probe. (b) Plot of the fluorescence intensity at 437 nm.

### 3.7. Confirm the Reaction Site by $^{19}\text{F}$ NMR

The reaction between (L, R)-1 and L-Lys was investigated using  $^{19}\text{F}$  NMR. Figure 8(a) presents the  $^{19}\text{F}$  NMR spectra of (L, R)-1 treated with 0–20 equivalents of L-Lys (DMSO:  $\text{D}_2\text{O}$  = 3:1). Three different fluorines on the phenyl group of (L, R)-1 exhibited peaks at  $\delta$ -142.2, -152.6, and -161.4 in a 2:1:2 ratio, with the signal at  $\delta$ -152.6 attributed to the para-fluorine. With the addition of L-Lys, the signal at  $\delta$ -152.6 weakened, and upon increasing Lys to 20 equivalents, the signal at  $\delta$ -152.6 completely disappeared. This indicates that the para-fluorine atom of (L, R)-1 is nucleophilically substituted by L-Lys, showing high regioselectivity. Figure 8(b) presents a possible binding mechanism between (L, R)-1 and L-lysine. The  $\epsilon$ -amino group of L-lysine interacts with the para-positioned F atom of the probe. Steric hindrance and hydrogen bonding may be the reasons why D-lysine does not react well with the probe molecule.



**Figure 8.** (a)  $^{19}\text{F}$  NMR spectra of (L, R)-1 treated with 0- 20 equivalents of L-Lys. (b) (L, R)-1 showed significant fluorescence enhancement for L-Lys through nucleophilic substitution.

#### 4. Conclusions

In summary, we discovered a set of BINAM-based fluorescent probes capable of chiral recognition of L-lysine among natural amino acids. The recognition effect is not influenced by other amino acids, and the probe is stable under certain acidic and alkaline conditions, allowing for rapid identification and long-term stability. The probe is sensitive and can identify L-Lys at lower concentrations. The recognition effect of the probe is enhanced in the presence of zinc ions. We also verified through  $^{19}\text{F}$  NMR that the para-fluorine atom of the probe undergoes nucleophilic substitution with L-Lys. This work provides a novel, efficient, and convenient method for the chiral recognition of L-Lys.

**Supplementary Materials:** The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Table S1: ef values for three important amino acids and explanation; Figure S1-S3: experimental data for D-lysine; Figure S4:  $^1\text{H}$  NMR spectra of R/S-1; Figure S5:  $^1\text{H}$  NMR spectra of (D/L, R/S)-1; Figure S6:  $^{13}\text{C}$  NMR spectra of (D/L, R/S)-1; Figure S7:  $^{19}\text{F}$  NMR spectra of (D/L, R/S)-1; Figure S8: MS spectra of (D/L, R/S)-1.

**Author Contributions:** Conceptualization, L.W. and X.L.; Validation, L.W.; Formal analysis, L.W.; Investigation, L.W.; Resources, G.Z.; Writing—original draft, L.W.; Writing—review and editing, W.C., Y.Z., X.Z. and J.Y.; Visualization, L.W.; Validation, W.C.; Supervision, G.Z.; Project administration, X.L.; Funding acquisition, G.Z. All authors have read and agreed to the published version of the manuscript.

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**Conflicts of Interest:** The authors declare no conflicts of interest.

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