

Review

Not peer-reviewed version

---

# Nanomedicine Strategies to Overcome Multi-Drug Resistance in Cancer: Innovations in Targeted Delivery, Tumor Microenvironment Modulation and Synergistic Therapies

---

Shiva Kumar Yadav , Nandakumar UP , [Vinay BC](#) , Manoj S Dikkatar , [Kala Bahadur Rawal](#) \*

Posted Date: 1 July 2025

doi: [10.20944/preprints202507.0081.v1](https://doi.org/10.20944/preprints202507.0081.v1)

Keywords: nanomedicine; multidrug resistance; co-delivery systems; tumor microenvironment (TME); drug efflux inhibition; stimuli-responsive nanoparticles; cancer immunotherapy; siRNA delivery; ferroptosis; targeted drug delivery



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

# Nanomedicine Strategies to Overcome Multi-Drug Resistance in Cancer: Innovations in Targeted Delivery, Tumor Microenvironment Modulation and Synergistic Therapies

Shiva Kumar Yadav <sup>1</sup>, Nandakumar UP <sup>2</sup>, Vinay BC <sup>3</sup>, Manoj S Dikkatar <sup>4</sup>  
and Kala Bahadur Rawal <sup>5,\*</sup>

<sup>1</sup> Assistant Professor, Department of Pharmacy Practice, R.C. Patel Institute of Pharmaceutical Education and Research Shirpur, Maharashtra, India

<sup>2</sup> Assistant Professor, Department of Pharmacy Practice, NGSM Institute of Pharmaceutical Sciences, Nitte (Deemed to be University), Mangaluru, Karnataka 575018, India

<sup>3</sup> Assistant Professor, Department of Pharmacy Practice, Bapuji Pharmacy College, Rajiv Gandhi University of Health Sciences, Davangere- 577004, Karnataka, India

<sup>4</sup> Associate Professor, Department of Pharmacy practice, Parul Institute of Pharmacy, Faculty of Pharmacy, Parul University, Vadodara- 391760, Gujarat

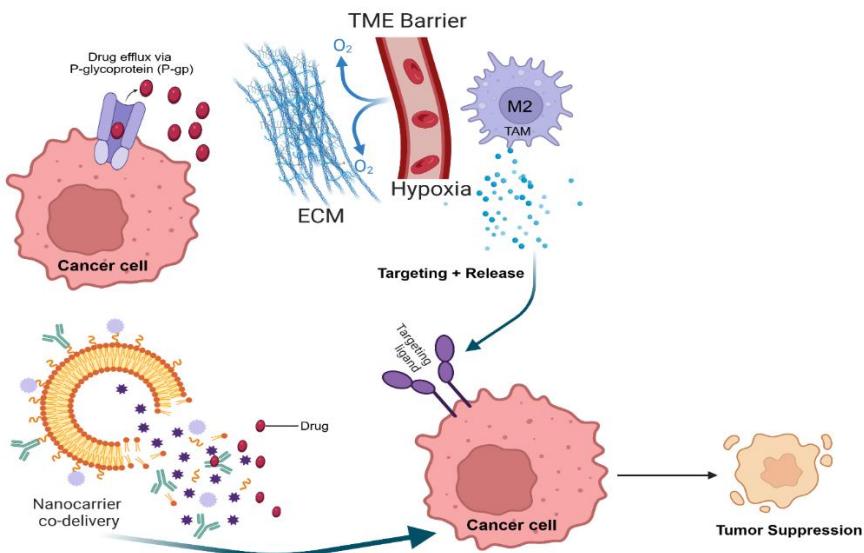
<sup>5</sup> Assistant Professor, Department of Pharmacy Practice, MM College of Pharmacy, Maharishi Markandeshwar (Deemed to be University), Mullana, Ambala-133207, Haryana, India

\* Correspondence: kbrawal52@gmail.com; Tel.: +91-7975141896

## Abstract

Multi-drug resistance (MDR) remains a major health challenge in the cancer treatment, leading to treatment failure and disease recurrence. Recent advancements in nanomedicine have introduced innovative approaches to treat MDR by improving drug delivery, reducing systemic toxicity, and re-sensitizing resistant cancer cells. This review provides a comprehensive summary of various nanocarrier systems that have been developed to bypass drug efflux mechanisms, promote intracellular drug accumulation, and permit controlled release. These nanocarrier systems include liposomes, polymeric nanoparticles, metal-based nanoparticles, and supramolecular constructs. Additionally, we discuss approaches targeting the tumor microenvironment, such as reprogramming tumor-associated macrophages (TAMs), reversing immunosuppression, and manipulating cancer stem cell differentiation. Special attention is paid to co-delivery systems that combine chemotherapeutics with gene therapies, redox-active compounds, autophagy inhibitors, and nitric oxide donors to produce synergistic anticancer effects. Novel strategies such as ferroptosis-inducing nanodrugs, stimuli-responsive platforms, and ultrasonic or photothermal based improved therapies are emphasized for their ability to evade typical resistance pathways. We also go over the important examples where nanotechnology has been utilized to counter MDR specifically in colorectal, ovarian, glioblastoma, and non-small cell lung cancer, targeting mechanisms such as P-glycoprotein overexpression, MRP2 transport, MGMT-mediated repair, and EGFR-TKI resistance. While promising preclinical results highlight the translational potential of nanomedicine for overcoming MDR, clinical integration remains a challenge. Key obstacles include scalable manufacturing, regulatory alignment, and thorough safety validation. This review aims to inform the rational design and clinical translation of nanotechnology-enabled therapeutics for drug-resistant cancers by integrating mechanistic insights with nanoplateform innovation.

**Keywords:** nanomedicine; multidrug resistance; co-delivery systems; tumor microenvironment (TME); drug efflux inhibition; stimuli-responsive nanoparticles; cancer immunotherapy; siRNA delivery; ferroptosis; targeted drug delivery



### Graphical Abstract

## 1. Introduction

Cancer is a leading cause of global mortality, with ~20 million new cases and 9.7 million deaths in 2022 [1–3]. A major obstacle in effective cancer therapy is multidrug resistance (MDR), where tumors become refractory to chemotherapy, targeted agents, or radiation. In fact, up to ~90% of chemotherapy failures are attributed to acquired resistance mechanisms [4,5]. MDR arises from complex biochemical and cellular adaptations that allow cancer cells to evade drug-induced cytotoxicity. These include upregulated drug efflux pumps (e.g. P-glycoprotein), enhanced DNA repair, apoptosis suppression, and influences from the tumor microenvironment (TME) [6–8]. Overcoming MDR is critical to improve patient outcomes. In recent years, nanomedicine has emerged as a transformative approach to counter MDR by improving drug delivery and enabling novel combination therapies. The clinical shift toward precision, low-toxicity interventions—such as HPV-HR DNA testing for posttreatment cancer monitoring [9,10] parallels the rise of nanomedicine strategies for overcoming treatment resistance. Engineered nanoparticles (NPs) can preferentially accumulate in tumors via the enhanced permeability and retention (EPR) effect, be surface modified with targeting ligands, and co-encapsulate multiple agents (e.g. a chemotherapeutic plus an MDR modulator or gene therapy) for synergistic action [11,12]. Nanocarriers can also be designed to release cargo in response to tumor-specific stimuli (pH, enzymes, light, ultrasound), and to modulate the TME (e.g. reprogramming macrophages or degrading stroma) [11,13,14]. These properties help to evade efflux pumps, bring back drug sensitivity, and improve antitumor immunity. These capabilities uniquely position nanomedicine as a multifaceted tool to counteract resistance at the molecular, cellular, and microenvironmental levels.

This review provides a comprehensive, up-to-date analysis of the molecular mechanisms underlying MDR in cancer alongside emerging nano-strategies to address them. We begin by discussing the cellular and molecular and biochemical foundation of MDR, including drug efflux transporters, apoptotic evasion, and tumor heterogeneity. We then explore a range of advanced nanocarrier platforms, targeted delivery tactics, co-delivery systems, and stimuli-responsive therapies that jointly overcome resistance. Throughout, we emphasize molecular mechanisms and translational potential, highlighting biochemical targets (e.g. ABC transporters, apoptosis regulators) and preclinical/clinical innovations in nanomedicine for MDR [6,15,16]. Emerging modalities such as ferroptosis-inducing nanoparticles, nitric oxide (NO) releasing platforms, and photothermal/ultrasound-triggered nanotherapies are also examined. Finally, we outline the key

translational challenges and propose future directions to facilitate the clinical adoption of nanomedicine in the therapy of MDR cancers.

## 2. Mechanisms of MDR in Cancer

Tumor cells evade therapy through multiple, often overlapping mechanisms, as detailed in recent comprehensive reviews [6]. The principal categories are:

### 2.1. Drug Efflux Transporters

Upregulation of ATP-binding cassette (ABC) pumps is a hallmark of MDR. P-glycoprotein (P-gp/ABCB1) is the most studied efflux pump [6]. Cryo-EM studies reveal conformational changes during substrate transport that underline its broad specificity [6,17]. Other ABC proteins such as MRP1/ABCC1 and BCRP/ABCG2 contribute to resistance against chemotherapies and targeted drugs [6,16]. For example, MRP1 can export kinase inhibitor metabolites, and BCRP confers resistance to several targeted agents [17,18]. Notably, cells can dynamically regulate transporter levels: recent data show that resistant cells reprogram ABC expression profiles under drug stress [6]. Stromal signals also impact efflux: cancer-associated fibroblasts (CAFs) can induce tumor cell ABC expression via paracrine pathways, creating a protective niche [6,19]. High efflux activity prevents intracellular drug accumulation, so inhibiting or bypassing these pumps is key to overcoming MDR. However, most platforms remain in the early translational phase, with few having progressed to clinical trials.

### 2.2. Apoptosis Evasion

Cancer cells often disable programmed cell death to survive therapy [6,20,21]. Overexpression of anti-apoptotic BCL-2 family proteins (BCL-2, BCL-XL, MCL-1) is commonly observed in resistant tumors [22]. Advanced proteomics show these proteins are post-translationally stabilized in MDR cells, strengthening survival [23]. Similarly, the IAP family (e.g. XIAP, cIAP1) inhibits caspases; elevated XIAP correlates with poor response to platinum chemo in ovarian cancer [24]. Tumors also inactivate p53: beyond gene mutations, recent studies identified novel posttranslational modifications of p53 that impair its apoptotic function [25]. Tumor cells can further rewire mitochondrial dynamics (increasing fusion, reducing cytochrome c release) to block intrinsic apoptosis [26,27]. They may also downregulate death receptors (FAS, TRAIL receptors) on the cell surface, evading extrinsic apoptosis [28,29]. Importantly, the TME enforces survival: CAFs and other stromal cells secrete cytokines (e.g. IL-6, IGFs) that upregulate anti-apoptotic proteins in cancer cells [30,31]. Together, these adaptations mean drugs that rely on apoptosis (most chemotherapies) become ineffective.

### 2.3. Enhanced DNA Repair

Many cytotoxic agents work by inducing DNA damage; resistant tumors often boost repair pathways. For instance, repeated genotoxic stress selects for cancer cells with upregulated homologous recombination and non-homologous end joining (NHEJ) repair proteins [32]. Single-cell analyses reveal plasticity: subclones with heightened DNA damage response (DDR) survive therapy, leading to radio- and chemo-resistant populations [33,34]. Cancer stem-like cells, often drug-resistant, inherently exhibit superior DNA repair and antioxidant defenses [33]. Enhanced nucleotide excision repair or base-excision repair can remove drug-induced lesions before apoptosis is triggered. Thus, inhibiting key repair enzymes (e.g. PARP, ATR) is explored to sensitize MDR tumors.

### 2.4. Tumor Microenvironment (TME)-Induced Resistance

The TME comprises stromal fibroblasts, immune cells (macrophages, myeloid cells), extracellular matrix (ECM), and factors like hypoxia and acidity, all of which influence drug response [35,36]. Hypoxic regions in tumors stabilize HIF-1 $\alpha$ , which promotes cell survival pathways and selection of aggressive, stem-like cells [37]. Hypoxia also impairs drug penetration and immune

function [38]. CAFs secrete ECM components (collagen, fibronectin) that increase tissue stiffness and form a physical barrier, limiting drug diffusion. They also release growth factors (TGF- $\beta$ , IL-6) that activate pro-survival signaling in cancer cells [39]. Similarly, tumor-associated macrophages (TAMs) often polarize to an M2-like phenotype under TME cues, releasing IL-10, VEGF and proteases that support tumor growth, angiogenesis, and matrix remodeling [40]. MDSCs and regulatory T cells in the TME suppress anti-tumor immunity and can secrete metabolites (e.g. arginase, IDO) that reduce drug efficacy. Metabolic competition (e.g. for glucose) in the TME also stresses effector immune cells while tumor cells adapt [41]. These TME factors effectively create a fortress around tumor cells, promoting quiescence and resistance.

### 2.5. Epigenetic Reprogramming

Reversible changes in gene expression contribute to MDR. For example, alterations in DNA methylation or histone modification can silence tumor suppressors or activate survival genes. Recent findings show that epigenetic changes can swiftly reprogram cells to use bypass pathways under therapy [42]. Cancer cells undergoing drug tolerance states display unique chromatin landscapes that prime them for resistance. For instance, histone methylation changes may activate drug efflux or DNA repair genes. MicroRNAs and long noncoding RNAs also modulate MDR by targeting ABC transporters or apoptotic genes. Such epigenetic plasticity enables tumors to adapt transiently to drugs and later re-sensitize after drug withdrawal, complicating therapy [43,44].

In summary, MDR arises from the interplay of intrinsic tumor cell adaptations and extrinsic TME factors, enabling tumors to escape multiple therapies. Effective strategies must therefore multitask; suppress efflux, restore apoptosis, block repair, and re-engineer the TME. Nanomedicine is uniquely suited to this challenge.

## 3. Strategies to Overcome Multidrug Resistance

### 3.1. Nanocarriers Inhibiting Drug Efflux Pumps (P-gp, MRP2, etc.)

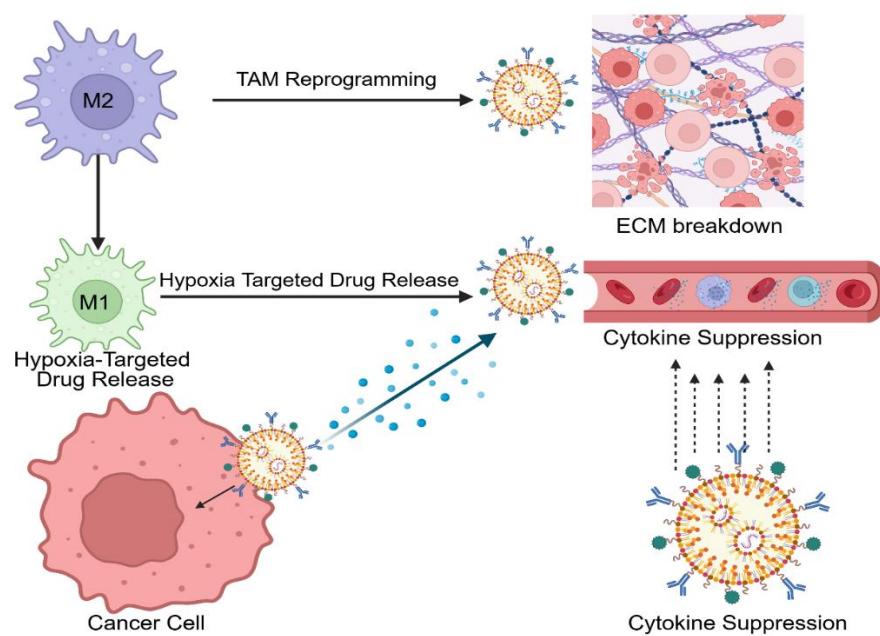
One fundamental approach to overcoming multidrug resistance (MDR) is to inhibit or evade the ATP-binding cassette (ABC) efflux transporters such as P-glycoprotein (P-gp/ABCB1), MRP1/MRP2, and BCRP. Recent nanocarrier systems have been designed to co-deliver chemotherapeutics with efflux pump inhibitors or siRNAs to suppress these transporters' function or expression [45–47]. Encapsulating drugs in nanoparticles can bypass recognition by efflux pumps and even target the cell nucleus, thereby increasing intracellular drug retention. For example, polymeric micelles carrying doxorubicin (DOX) together with a P-gp siRNA showed effective P-gp gene silencing and restored drug sensitivity in resistant cancer cells [48,49]. Another strategy is using nanocarriers that disrupt cancer cell energy supply: a mitochondria-targeted hybrid nanoparticle was shown to generate ROS and consume ATP under near-infrared (NIR) light, transiently impairing P-gp function and creating a therapeutic window for chemotherapy [50,51]. Such multifaceted nano-formulations significantly increase drug accumulation in MDR tumor cells by inhibiting efflux pump activity at the protein or gene level.

### 3.2. Modulating the Tumor Microenvironment (TME)

The tumor microenvironment plays a critical role in MDR, contributing factors like hypoxia, dense extracellular matrix, and immunosuppressive cells (e.g. M2 tumor-associated macrophages, TAMs) [52]. Nanomedicine strategies increasingly target these components to reverse the supportive niche that fosters drug resistance. One approach is re-educating TAMs: delivering Toll-like receptor agonists or small interfering RNAs via nanoparticles can polarize macrophages from an M2 (protumor, repair) phenotype to an M1 (pro-inflammatory, anti-tumor) phenotype [53]. This immunomodulation can heighten the tumor's response to therapy and reduce MDR, as M2-like TAMs are known to promote tumor growth and drug resistance [52]. Another strategy involves

extracellular matrix (ECM) remodeling to improve drug penetration. Nanocarriers functionalized with enzymes like collagenase or hyaluronidase have been used to locally degrade collagen and hyaluronic acid in tumors. For instance, collagenase-decorated nanoparticles carrying DOX showed enhanced tissue penetration and reduced tumor fibrosis, thereby overcoming the ECM barrier to drug delivery [54]. As TME-driven immune evasions such as T and NK cell suppression in non-small cell lung cancer, contributes significantly to therapeutic failure [55], nanomedicine strategies that reprogram immunosuppressive niches offer a promising route to overcome such resistance. By modulating TME factors, normalizing abnormal vasculature, reducing interstitial pressure, re-polarizing macrophages, and enzymatically softening the stroma, nanotherapies can significantly improve the efficacy of chemotherapy in otherwise resistant tumors [52,56].

As summarized in Figure 1, multiple nanomedicine strategies have been developed to target key components of the tumor microenvironment, including TAM reprogramming, ECM remodeling, hypoxia-triggered drug release, and cytokine suppression.



**Figure 1.** TME modulation via Nanoparticles.

### 3.3. Dual and Multi-Drug Co-Delivery Nanosystems

Co-delivery of multiple therapeutic agents in a single nanocarrier has emerged as a powerful method to tackle MDR. Advanced nanoparticles (liposomes, polymeric nanoparticles, dendrimers, etc.) can be engineered to carry two or more drugs simultaneously, allowing synergistic action and synchronized release. By incorporating a chemotherapeutic together with a chemosensitizer or a second drug, these nanosystems can attack cancer cells on multiple fronts. Key examples include:

#### 3.3.1. Chemotherapy–chemosensitizer combination

Nanoparticles co-loaded with a conventional anticancer drug and an efflux pump inhibitor (e.g., DOX + tariquidar) or a reversal agent like verapamil have shown increased intracellular drug retention and cytotoxicity in resistant cell lines [57]. Similarly, the rational design of small-molecule chemotherapeutics, including benzofuran–piperazine derivatives with demonstrated cytotoxic activity, can complement nanocarrier-based delivery strategies by providing mechanistically potent payloads [58].

### 3.3.2. Dual chemotherapies

Co-encapsulation of two therapeutics that act via different mechanisms can produce synergistic killing. For instance, a cRGD-targeted lipid nanoparticle was developed to deliver gemcitabine and paclitaxel together, achieving enhanced breast cancer cell kill rates compared to either drug alone [59]. Such co-delivery ensures both drugs are present at the tumor in the optimal ratio and timing.

### 3.3.3. Drug–gene combinations

Nano-carriers can concurrently deliver a drug and a genetic therapy. Quantum-dot nanoconjugates have been reported that adsorb DOX together with siRNA targeting MDR genes; this approach successfully downregulated P-gp expression and resensitized cervical cancer cells (HeLa) to chemotherapy [60,61]. Similarly, mesoporous silica nanoparticles carrying DOX plus an siRNA against MDR1 gene achieved higher tumor inhibition in an MDR breast cancer model by blocking drug efflux at the gene level. Similarly, nanocarriers co-loaded with therapeutics and immunostimulatory agents, such as IL-2, represent a promising strategy to simultaneously debulk tumors and enhance immune surveillance. Previous viral vector-based studies have demonstrated the feasibility and antitumor efficacy of IL-2-mediated immunotherapy in solid tumors [62].

**Table 1.** Examples of nano co-delivery systems overcoming MDR.

Nanoformulation	Drugs/Agents	Cancer Model	Key Outcomes (synergy)
Polymeric NP (mPEG-PLGA)	Paclitaxel dimer prodrug + Tetrandrine	MDR HeLa cells (cervical)	Enhanced uptake and ROS; ≈50% higher apoptosis vs. single drug
Polymeric NP (PEG-coated)	SN-38 (prodrug) + Ko143 (BCRP inhibitor)	BCRP-overexpressing CRC xenograft	Reversed irinotecan resistance; ~10-fold ↓ IC <sub>50</sub>
Transferrin-PLGA NP	Gefitinib (EGFR-TKI) + Thymoquinone	Gefitinib-resistant NSCLC (A549/GR)	Re-sensitized to gefitinib; suppressed EMT (increased E-cadherin)
cRGD–Heparin NP	Cisplatin + Olaparib (PARP inhibitor)	Cisplatin-resistant ovarian	Inhibited P-gp/MRP2, ↑ DNA damage; overcame cisplatin resistance
Doxorubicin liposome + HCQ*	Doxorubicin + Hydroxychloroquine	DOX-resistant breast cancer	Restored apoptosis; polarized TAMs to M1 (↑TNF $\alpha$ , IL-12)

By tailoring nanocarrier release profiles and surface chemistry, dual-drug nanoparticles can ensure spatial and temporal co-localization of therapeutic agents in tumor cells, effectively bypassing mechanisms of resistance and yielding greater cytotoxic effect than single-drug treatments.

### 3.4. Tumor-Specific Targeting and Active Delivery Systems

To maximize drug delivery to cancer cells while sparing normal tissue, researchers are functionalizing nanocarriers with tumor-targeting ligands [63]. Such active targeting helps overcome MDR by strengthening the effective drug concentration at the tumor site. Common targeting moieties include transferrin (targets transferrin receptors often overexpressed in cancers), folic acid (targets folate receptors), antibodies or fragments (against tumor antigens), and peptides like cyclic RGD (cRGD) which bind integrin  $\alpha_v\beta_3$  on tumor endothelium and cells [64,65].

Transferrin-conjugated nanoparticles are a prominent example: in one study, Tf-decorated PLGA nanoparticles loaded with an experimental organoselenium drug were tested against drug-resistant tumor cells. The Tf-NPs showed significantly higher cytotoxicity in P-gp overexpressing cancer cell lines compared to non-targeted NPs, indicating enhanced uptake via Tf receptor-mediated

endocytosis [66,67]. In 3D tumor spheroid models of an MDR ovarian cancer line (NCI/ADR-RES), the Tf-NPs likewise penetrated better and reduced spheroid growth more effectively [68,69]. Similarly, cRGD functionalization of liposomes or polymeric NPs improves their accumulation in tumors by targeting neovasculature and invasive tumor cells. cRGD-modified nanoparticles were observed to undergo receptor-mediated endocytosis into  $\alpha_v\beta_3$ -expressing cancer cells, achieving higher intracellular drug delivery than untargeted particles [70,71]. In vivo imaging confirmed that cRGD-NPs concentrate preferentially in tumors, with reduced off-target distribution to liver and lungs [70].

Other ligand-targeted nanocarriers include folate-NPs (effective in folate receptor-positive ovarian and breast cancers), EGFR-targeted immunoliposomes, HER2-targeted nanoparticles for resistant breast cancer, and aptamer-guided NPs. By actively homing to tumor cells or the tumor microenvironment, these systems increase drug efficacy against MDR tumors and mitigate systemic toxicity [70,72]. The result is a higher therapeutic index and the ability to kill resistant cancer subpopulations that might evade passive delivery. Additionally, receptor-mediated delivery strategies are increasingly leveraged in nanoparticle design to enhance tumor targeting while minimizing off-target toxicity. For instance, receptor-based frameworks have been applied to bypass efflux-mediated resistance and improve intracellular drug retention [73].

### 3.5. Nanotechnology in Specific Cancers: Colorectal, Breast, Ovarian, and Kidney

Multi-drug resistance manifests differently across cancer types, and nanomedicine strategies have been tailored accordingly in colorectal, breast, ovarian, and kidney cancers:

#### 3.5.1. Colorectal Cancer (CRC)

MDR in CRC (e.g., resistance to 5-fluorouracil or oxaliplatin) is often linked to efflux pumps and cancer stem cells. Nanocarriers have been explored to deliver combination therapies and siRNAs to overcome these mechanisms [74]. For instance, lipid nanoparticles co-loading 5-FU with curcumin (a natural chemosensitizer) have shown the ability to reverse 5-FU resistance in CRC cells [74,75]. Polymeric nanosystems targeting colon cancer stem cell markers are also under investigation to prevent recurrence and MDR. Early results demonstrate that nano-delivery can enhance drug uptake in CRC and downregulate survival pathways, improving chemosensitivity [74,75]. Additionally, preclinical studies such as those using CF10 polymer formulations have shown remarkable efficacy against colorectal cancer liver metastasis model, highlighting the translational potential of optimized nanopolymers[76–78].

#### 3.5.2. Breast Cancer

Breast tumors (especially triple-negative or recurrent tumors) commonly develop MDR through P-gp overexpression. Many studies use the doxorubicin-resistant MCF-7/ADR cell model to evaluate nano-therapies. Successful approaches include PEGylated liposomes carrying DOX plus P-gp inhibitors, and pH-responsive micelles delivering dual drugs [79,80]. A notable example combining DOX with siRNA against MDR1 in a pH-sensitive chitosan-based micelle, achieving dramatically higher tumor suppression in an MCF-7/ADR mouse model (87% tumor inhibition) compared to free DOX (~50%) [81,82]. Another approach used a hyaluronic-acid modified MoS<sub>2</sub> nanosheet to deliver DOX and perform photothermal therapy; under NIR laser, the nano-system generated heat and downregulated P-gp expression, leading to near-complete tumor ablation in an MDR breast cancer model [83–85]. These examples highlight that in breast cancer, nanomedicines can restore drug potency by both chemical and physical mechanisms (gene silencing, heat, etc.).

#### 3.5.3. Ovarian Cancer

Ovarian tumors often exhibit MDR to platinum drugs and taxanes. Nanocarriers are being designed for intraperitoneal delivery in ovarian cancer to achieve high local drug levels. As

mentioned, transferrin-targeted nanoparticles have shown promise in sensitizing ovarian cancer cells (like NCI/ADR-RES) to drugs [86]. Another strategy is delivering small molecule inhibitors of anti-apoptotic pathways (e.g., BCL-2 or PI3K/Akt) alongside chemo in a single nanoplatform [87–89]. In preclinical studies, dual-loaded NPs (e.g., paclitaxel + Akt siRNA) demonstrated the ability to overcome paclitaxel resistance in ovarian cancer xenografts by inducing apoptosis in otherwise refractory tumors [90–93]. Ongoing translational research in ovarian cancer focuses on nano-formulations of platinum drugs, PARP inhibitors, and gene therapies to bypass resistance and reduce systemic toxicity [94–96].

### 3.5.4. Kidney (Renal Cell) Cancer

Metastatic renal cell carcinoma (RCC) can develop resistance to targeted therapies like tyrosine kinase inhibitors (TKIs) [97]. Nanomedicine efforts here aim to deliver novel therapeutics or re-sensitize tumors to TKIs. One innovative example is the use of cuprous oxide ( $\text{Cu}_2\text{O}$ ) nanoparticles to overcome resistance to sunitinib (a common RCC TKI) [98]. It is reported that  $\text{Cu}_2\text{O}$  NPs induce endoplasmic reticulum stress and ROS-mediated apoptosis in renal cancer cells, thereby restoring their sensitivity to sunitinib [99]. This nanoparticle effectively modulated copper trafficking inside tumor cells, pointing to a unique ferroptosis-like mechanism to kill drug-resistant RCC cells [100]. Additionally, dual-ligand liposomes targeting both RCC cells and angiogenic endothelium have shown promise in drug-resistant kidney tumors, by concentrating drugs in the tumor microvasculature and tumor tissue simultaneously [101]. While nanotherapy in kidney cancer is still nascent, these approaches suggest that overcoming resistance to targeted agents (like TKIs) is feasible with nanoparticle delivery that triggers alternative cell-death pathways or improved drug localization.

## 3.6. Emerging Strategies: Ferroptosis, Autophagy Modulation, Nitric Oxide, and Gene Therapy

Beyond conventional chemo, several cutting-edge approaches leverage nanotechnology to induce non-traditional cell death pathways or to modulate cellular survival mechanisms in MDR cancers:

### 3.6.1. Ferroptosis Induction

Ferroptosis is an iron-dependent form of programmed cell death characterized by lipid peroxidation. Recent studies highlight that inducing ferroptosis can help kill drug-resistant cancer cells that evade apoptosis [102]. Nanomedicines are being engineered as “ferroptosis nanoinducers” for example, ultrasmall iron oxide or magnetite nanoparticles that release  $\text{Fe}^{2+}$  to catalyze lipid ROS generation, or nanocarriers delivering ferroptosis-triggering drugs (like erastin or RSL3) [103]. The versatile ferroptosis-targeted nanotherapies have shown broad potential in reversing therapy resistance by directly triggering this pathway and by feedback regulation of cellular redox state [104]. In practice, combining ferroptosis inducers with chemo or immunotherapy (via co-loaded NPs) has yielded synergistic anti-tumor effects in refractory tumors, and is a hot area of preclinical research [105,106]. Additionally, targeting DNA repair checkpoints, such as CHK1 or PARP, offers another avenue to overcome resistance. CHK1 inhibition has been shown to potentiate DNA damage and sensitize chemoresistant ovarian tumors by modulating PARPylation and metabolic stress [107,108].

### 3.6.2. Autophagy Inhibition

Autophagy, a cellular recycling process, is a double-edged sword in cancer; in established tumors, moderate autophagy often promotes survival under therapeutic stress, contributing to MDR [109]. Thus, blocking cytoprotective autophagy can re-sensitize cancer cells to treatment. However, systemic autophagy inhibitors (like chloroquine derivatives) have off-target effects. Nanocarriers offer a way to target autophagy inhibitors to tumors. Researchers have loaded agents such as hydroxychloroquine (HCQ) or novel autophagy blockers into tumor-targeted nanoparticles to

concentrate their action in tumor cells [110–112]. For example, Wuliji et al. formulated amphiphilic polymer micelles co-encapsulating HCQ and DOX; these NPs preferentially accumulated in tumors and effectively overcame DOX resistance by inhibiting autophagy flux in cancer cells [112]. Such nano-formulations ensure that autophagy is sufficiently inhibited in the tumor (preventing cancer cells from “recycling” damaged organelles and drugs) while minimizing toxicity elsewhere. Notably, some teams are also exploring autophagy activation in certain contexts (since excessive autophagy can induce cell death); both inhibition and promotion of autophagy via nanomedicines are being tested as strategies to collapse the defenses of MDR tumors [112–114].

### 3.6.3. Nitric Oxide (NO) Delivery

Nitric oxide is a gaseous signaling molecule that, at high concentrations, can kill cancer cells and modulate MDR pathways. NO has been found to downregulate P-glycoprotein expression and sensitize tumors to drugs. To exploit this, researchers have developed NO-releasing nanocarriers [115,116]. One innovative system is a supramolecular peptide hydrogel loaded with a NO prodrug and DOX: when injected into a tumor, the hydrogel slowly releases NO (especially in the presence of high glutathione levels in the TME) alongside DOX [117,118]. The burst of NO not only directly damages tumor cells via oxidative stress, but also reverses P-gp-mediated MDR, making cancer cells more susceptible to DOX [119]. In vivo studies in MDR breast cancer models showed that this NO-releasing co-delivery platform significantly enhanced tumor shrinkage compared to DOX alone [120]. Another study encapsulated an NO donor and camptothecin in a PLGA nanoparticle; under the acidic conditions of the tumor, NO was generated, which destabilized the NP and triggered drug release in situ. The released NO reduced P-gp levels by ~45%, thereby permitting a much higher intracellular concentration of camptothecin [121]. These results underscore NO’s potential as a chemosensitizing agent when delivered via a controlled nanocarrier.

### 3.6.4. Gene Therapy and RNA Interference

Nanotechnology enables gene-based therapies to overcome MDR, such as siRNA, shRNA, mRNA or CRISPR/Cas9 systems directed against resistance-related genes [122]. Because nucleic acids alone have poor stability and delivery, nano-formulations (lipid nanoparticles, polymeric polyplexes, etc.) are essential for their clinical use [123,124]. Numerous studies have shown that siRNA against MDR1 (ABCB1) delivered by nanoparticles can knock down P-gp expression and restore drug sensitivity in vitro and in vivo [125–127]. For example, a hierarchical mesoporous silica nanocarrier was used to co-deliver DOX plus an anti-Pgp siRNA: the MSN protected the siRNA from degradation and released both payloads inside multidrug-resistant breast cancer cells, leading to P-gp silencing and significantly higher chemotherapy efficacy [128]. Beyond P-gp, researchers are targeting other resistance genes (BCL-2, Akt, MYC, etc.) using siRNA or even CRISPR delivered by viral-mimicking nanoparticles [129]. Similarly, modulation of oncogenic miRNAs like miR-221-5p has been implicated in chemoresistance, further justifying nanoparticle-mediated miRNA therapies in resistant ovarian cancers [130]. The upregulation of lymphoblastic leukemia-derived sequence-1 (LYL1) has been implicated in the progression and metastatic potential of ovarian cancer, highlighting it as a potential target for gene-silencing approaches. Sah et al. demonstrated the oncogenic role of LYL1 in ovarian cancer models, supporting the rationale for RNAi-loaded nanocarriers designed to reverse oncogene-driven drug resistance [131]. Early results are promising, showing that silencing or editing genes can directly reverse MDR phenotypes. However, gene therapy approaches must surmount delivery challenges and potential off-target effects and thus are often combined with nanoparticle strategies for tumor-specific, controlled delivery [132,133].

In summary, these emerging nano-strategies aim to attack MDR cancer cells via novel mechanisms inducing ferroptotic cell death, shutting down autophagy survival pathways, using bioactive gases like NO, or reprogramming gene expression – thereby supplementing conventional cytotoxic drugs and overcoming resistance that arises from classic mechanisms.

### 3.7. Stimuli-Responsive Nanotherapies: Ultrasound, Photothermal, and Sonodynamic Approaches

Stimuli-responsive nanomedicine adds another layer of innovation to combat MDR by using external triggers (such as light or sound) to activate drug delivery or kill tumor cells by non-chemical means [134,135]. These approaches can bypass drug efflux and damage resistant cells through physical mechanisms:

#### 3.7.1. Photothermal Therapy (PTT)

This involves nanoparticles that absorb NIR laser light and convert it to heat, selectively killing cancer cells or disrupting their membranes. Photothermal agents (gold nanoshells, nanorods, carbon nanomaterials, graphene oxide, etc.) can be loaded with chemotherapeutics for a combined effect. Under NIR irradiation, the localized heating can enhance drug release from the nanocarrier and also sensitize tumor cells (e.g., by causing protein denaturation or ATP depletion) [136–138]. PTT has been shown to downregulate efflux pumps: for example, mild hyperthermia ( $\approx 52^{\circ}\text{C}$ ) generated by MoS<sub>2</sub> nanosheets caused a significant reduction in P-gp levels in MDR breast cancer cells [139,140]. In one study, PLGA nanoparticles co-loaded with DOX and indocyanine green (ICG, an FDA-approved NIR dye) were used to treat MCF-7/ADR tumors. NIR laser exposure of these NP in tumors led to heat generation (from ICG) and triggered DOX release; the combination of chemotherapy + PTT effectively destroyed drug-resistant breast cancer cells in vitro and in vivo [141]. Photothermal nano-therapy thus provides a way to physically eradicate cancer cells that might survive chemical therapy, and when integrated with drug delivery, it produces a synergistic anti-MDR effect.

#### 3.7.2. Ultrasound and Sonodynamic Therapy (SDT)

Ultrasound can penetrate deep into tissues and be focused on tumor sites, making it a valuable trigger for drug release or activation of sonosensitizers [142,143]. In SDT, a sonosensitizer compound (often a porphyrin, IR780, or other ROS-generating molecule) is delivered by a nanoparticle and produces cytotoxic reactive oxygen species when exposed to low-intensity ultrasound. This strategy is particularly useful against hypoxic or deep-seated MDR tumors [144,145]. A recent breakthrough involved cRGD-targeted gold nanoparticles loaded with the EGFR inhibitor gefitinib and the sonosensitizer IR780 [146]. The gold shell provided a photothermal effect (for on-demand drug release) and IR780 generated ROS under ultrasound; together, this low-temperature PTT + SDT combination was able to overcome acquired TKI resistance in an EGFR-mutant NSCLC model [146]. The ultrasound-triggered ROS helped kill cancer cells independent of drug action, while also impairing cellular defense mechanisms, and the mild heating facilitated drug uptake [147,148]. Beyond this, ultrasound is also used to trigger drug release from acoustically responsive nanocarriers (like liposomes or microbubbles) at the tumor site, ensuring a high local drug concentration that can overwhelm efflux pumps [149,150]. Sonodynamic and ultrasound-triggered therapies are still in translational stages, but they exhibit tremendous potential to non-invasively eliminate MDR cancer cells and can be repeated or spatially controlled with imaging guidance (many ultrasound-sensitive NPs also serve as contrast agents) [151].

#### 3.7.3. Other External Triggers

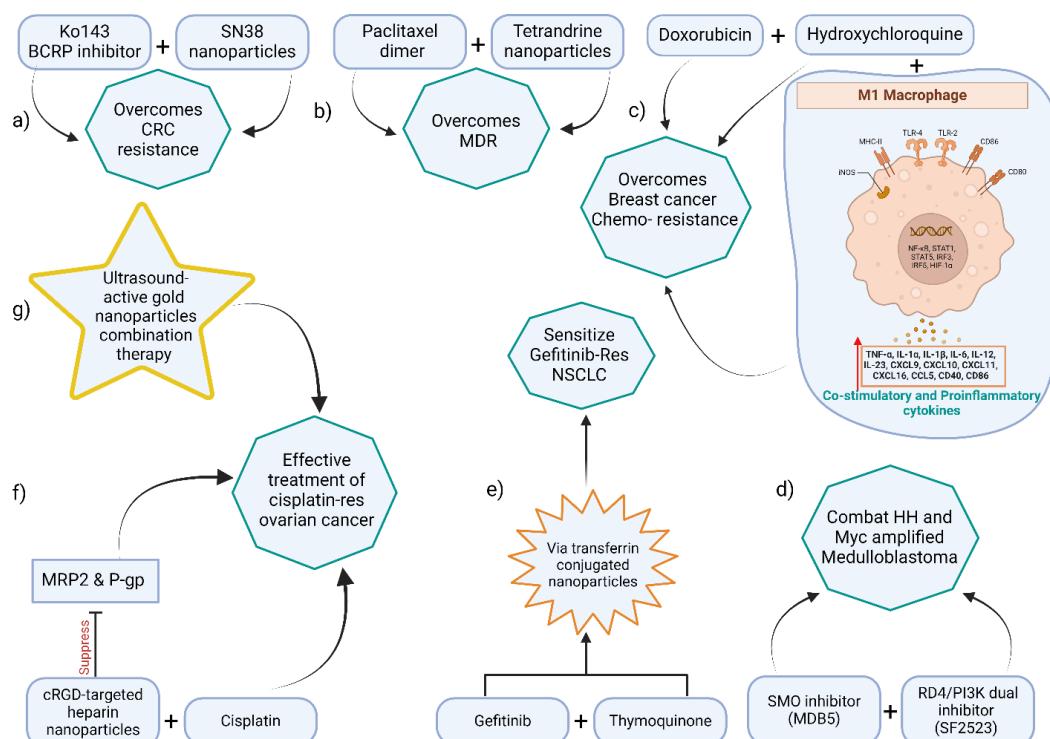
Magnetic field-responsive nanoparticles (e.g., iron oxide NP for magnetic hyperthermia or drug targeting), photodynamic therapy (light-activated generation of singlet oxygen using NP-delivered photosensitizers), and radiation-triggered nanoparticle release are additional modalities under exploration [152,153]. For instance, magnetically heated nanoparticles or X-ray-activated liposomes could help in circumventing resistance by causing direct tumor cell damage or by releasing drugs in a burst when the tumor vasculature is most permeable. These methods, often termed “externally responsive nanotherapies,” can be combined with chemotherapy to potentiate treatment effects [154]. By carefully tuning the stimulus (laser wavelength, ultrasound frequency, magnetic field strength),

clinicians can activate the nanoparticle only at the tumor site, thereby achieving high precision [154,155].

In summary, stimuli-responsive nanotherapies offer a way to attack MDR tumors using physical forces (heat, sound, light) that cancer cells are less equipped to resist. When coupled with smart nanocarriers, these approaches release drugs at the optimal time or bypass biochemical resistance entirely, and they have demonstrated impressive results in preclinical MDR cancer models [156]. To illustrate the diversity and mechanisms of nanocarrier-based co-delivery systems that successfully circumvent MDR in different cancers, a schematic representation of selected strategies is shown in Figure 2, corresponding to the formulations detailed in Table 2.

**Table 2.** Stimuli-responsive nanotherapy modalities for MDR cancer.

Stimulus	Mechanism (effects)	Representative Cancer Model/Type
<i>Ultrasound (US)/SDT</i>	Cavitation increases drug uptake; triggers ROS from sonosensitizers (deep penetration)	Pancreatic cancer, breast cancer, brain (glioblastoma)
<i>Photothermal (PTT)</i>	NIR light → nanoparticle heats tumor (45–60°C), causing protein denaturation and cell death (efflux-independent)	Skin, head/neck, breast tumors (where NIR penetrates)
<i>Chemophotodynamic (PDT)</i>	Light activates photosensitizer → ROS (singlet O <sub>2</sub> ); synergizes with chemo	Superficial tumors, MDR melanoma models
<i>Sonodynamic (US + sensitizer)</i>	Ultrasound activates sensitizer → ROS; deep-tissue effect	Deep tumors (brain, pancreas)
<i>Hyperthermia</i>	Elevated temperature triggers drug release (thermosensitive liposomes) and tumor cell death	Liver (HCC), prostate



**Figure 2.** Mechanistic Pathways of Nanomedicine Co-Delivery Platforms Targeting Multidrug Resistance Across Diverse Cancer Types.

### 3.8. Innovative and Underexplored Nanomedicine Strategies for MDR Cancers

While significant attention has been given to co-delivery systems, tumor microenvironment modulation, and stimuli-responsive release strategies, recent studies have spotlighted a new wave of highly promising nanomedicine innovations. These approaches, while underrepresented in earlier clinical trials, are gaining traction for their ability to circumvent traditional resistance mechanisms through novel physicochemical properties and delivery modalities.

#### 3.8.1. Magnetic Nanoparticles for Theranostics and Targeted Therapy

Magnetic nanoparticles (MNPs), particularly superparamagnetic iron oxide nanoparticles (SPIONs), offer dual functionality as both therapeutic and diagnostic agents [157]. When guided by an external magnetic field, MNPs can accumulate at the tumor site with high specificity, allowing for concentrated drug delivery [158]. Moreover, magnetic hyperthermia where MNPs generate localized heat in response to an alternating magnetic field has demonstrated synergy with chemotherapeutics and the capacity to downregulate drug efflux pumps like P-gp [159]. This form of image-guided, magnetically targeted therapy is especially promising for difficult-to-treat or deep-seated tumors.

#### 3.8.2. Intratumoral Administration of Nano Drug Delivery Systems

Intratumoral injection of nano-formulations bypasses systemic pharmacokinetic barriers and delivers high drug concentrations directly into tumor tissue [160]. This strategy enhances local therapeutic efficacy while reducing off-target toxicity. Various delivery systems such as hydrogels, polymeric micelles, and dendritic nanogels have been engineered for sustained and localized drug release following intratumoral injections. This modality is particularly effective in poorly vascularized or desmoplastic tumors, where systemic delivery fails to achieve adequate penetration [161,162].

#### 3.8.3. Polydopamine (PDA) Nanoparticles in Drug Delivery and Immune Modulation

Polydopamine-based nanoparticles exhibit excellent biocompatibility, surface adhesion, and responsiveness to oxidative environments [163]. PDA nanocarriers have been to provide immune modulation, photothermal effect and chemotherapeutic effect simultaneously. [164]. These nanoparticles generate heat when exposed to near-infrared light which can damage cancer cell membranes or sensitize resistant cells to chemotherapy [165]. Additionally, PDA systems have shown promise in modulating immune responses by carrying Toll-like receptor agonists or checkpoint inhibitors, thus bridging drug delivery with tumor immunotherapy [166].

#### 3.8.4. Inorganic Nanoparticles in Overcoming Resistance

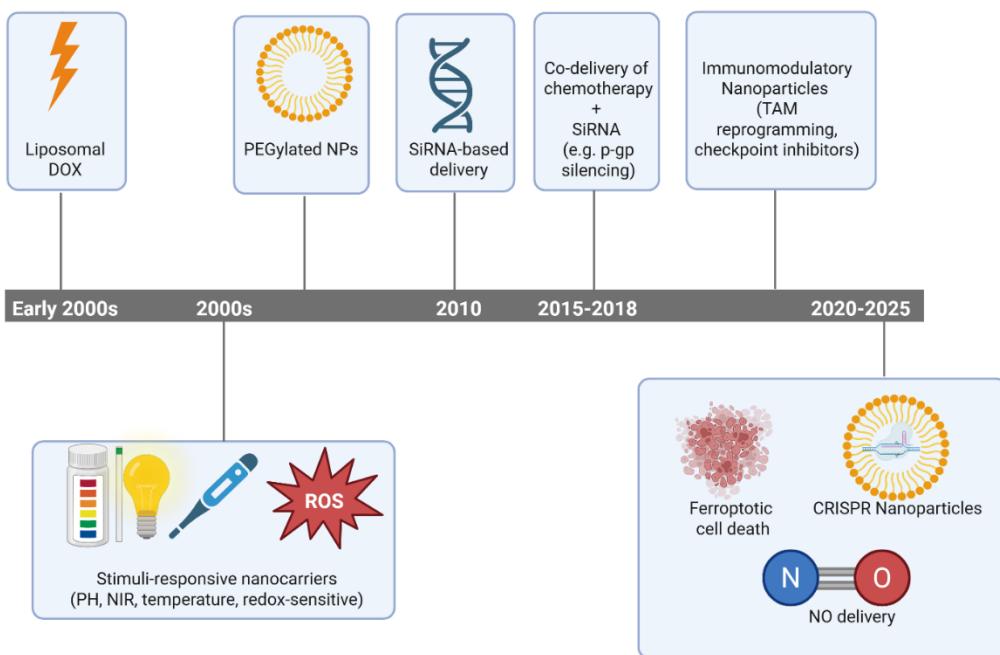
Inorganic nanomaterials, including gold nanoparticles, mesoporous silica, and quantum dots offer unique optical, electrical, and structural features [167]. These carriers are not only effective in delivering therapeutics but also enhance imaging and diagnosis. Several studies have demonstrated that inorganic NPs can induce ROS generation, promote apoptosis, and even directly interfere with intracellular resistance pathways [168–170]. Quantum-dot conjugates, for instance, allow real-time tracking while co-delivering siRNA and chemotherapeutic agents to downregulate MDR-associated genes [171,172].

#### 3.8.5. Oral Nanoformulations for Colorectal Cancer

Oral administration of nanomedicine is particularly valuable in colorectal cancer therapy [173]. Engineered nanoparticles with pH-sensitive or mucoadhesive properties can bypass gastric degradation and release drugs specifically in the colon [174,175]. Co-formulations combining 5-FU with natural chemosensitizers like curcumin have shown enhanced drug retention and MDR reversal [176]. These systems offer improved patient compliance and the potential to deliver both cytotoxic

and immunomodulatory agents directly to the tumor site, which is crucial in colorectal tumors with localized resistance profiles [177].

Together, these underexplored strategies highlight the evolving landscape of nanomedicine for drug-resistant cancers. Their distinct mechanisms ranging from magnetic targeting and localized hyperthermia to oxidative disruption and site-specific oral delivery complement established approaches and provide new opportunities to overcome therapeutic resistance in challenging cancer subtypes.



**Figure 3.** The Historical development of nanomedicine platforms aimed at reversing MDR, highlighting the evolution from conventional liposomal drugs to gene-editing and ferroptosis-inducing nanocarriers.

#### 4. Clinical Translation: Limitations and Challenges

Despite the plethora of promising nanomedicine strategies in preclinical studies, translating these innovations into clinical use faces significant challenges. The first consideration is the complexity of regulatory approval for nanodrugs. Nanomedicines often don't fit neatly into existing regulatory categories, they may be classified as drugs, biologics, devices, or combination products, which complicates the approval pathway [178]. Regulatory agencies like the FDA and EMA have been cautious, requiring extensive characterization of nanoparticle physicochemical properties, manufacturing consistency, and toxicity profiles. Key hurdles include:

##### 4.1. Standardization and Characterization:

There is a lack of standardized guidelines specific to nanomedicines [178]. Small variations in formulation (particle size, surface charge, etc.) can dramatically alter a nanoparticle's behavior *in vivo* [179]. Thus, companies must implement advanced analytical techniques and strict quality controls to ensure batch-to-batch consistency and well-characterized products [178,180]. Demonstrating reproducibility and stability of complex nanoparticle formulations (which may comprise multiple components like polymers, lipids, targeting ligands, and drugs) is much more involved than for small-molecule drugs. This also affects the development of "nanosimilars" (generic versions of nanodrugs), where establishing equivalence is non-trivial [180,181].

#### 4.2. Safety and Toxicity Concerns

While nanocarriers can reduce the systemic toxicity of potent drugs, the long-term fate of nanoparticles in the body is not fully understood. Issues such as accumulation in the reticuloendothelial system (liver, spleen), unforeseen immunogenic reactions, or off-target effects of the nanoparticle components must be addressed [182]. Extensive preclinical studies are needed to evaluate biocompatibility, for example, whether the NP triggers complement activation or how it is excreted or metabolized [183]. Some high-profile nanodrugs have failed in trials due to toxicity or immune-related adverse events not predicted by animal models [184,185]. Rare but severe immune-mediated reactions to pharmacological agents, such as sulfasalazine-induced DRESS syndrome, underscore the importance of vigilant safety profiling and post-marketing surveillance in both traditional and nanomedicine therapies [186]. In addition to manufacturing and scalability, successful integration of nanomedicine into real-world clinical settings requires alignment with hospital practices, prescriber behaviors, and resource availability. System-level challenges, such as noncompliance with prescription-writing standards, delayed IV-to-oral transitions, and inconsistent infection management protocols highlight the barriers to adopting complex therapeutic innovations like nanomedicine in routine care [187–189]. Moreover, including pharmacists in clinical audit workflows has shown to improve adherence to treatment protocols and could support future implementation of nanomedicine [190]. Overcoming MDR in a tumor is controversial if nanotherapy cannot be used safely in patients, so ensuring safety profiles and conducting rigorous toxicity studies (including looking at chronic exposure) is essential [191,192].

#### 4.3. Manufacturing and Scalability

Producing nanomedicines at a commercial scale with uniform quality is challenging [193]. Techniques used in the lab (like microfluidic mixing or small-batch emulsification) may not easily translate to industrial scale. Additionally, sterile filtering or validating complex nanoconjugates can be difficult [194]. Scaling up often requires significant engineering and process development, and any process changes might alter the NP properties, sending developers back to perform comparability studies [195]. For example, standardized methods for the growth, purification, and titration of virus-based therapies, such as oncolytic herpes simplex viruses, have been developed to improve process reproducibility and safety, offering insights applicable to viral and gene-based nanocarriers [196]. Scaling up contributes to the high cost of bringing nanodrugs to the market [197]. Manufacturing complexities also tie into regulatory hurdles, as changes in a nanomedicine's production may trigger the need for new safety evaluations [198,199].

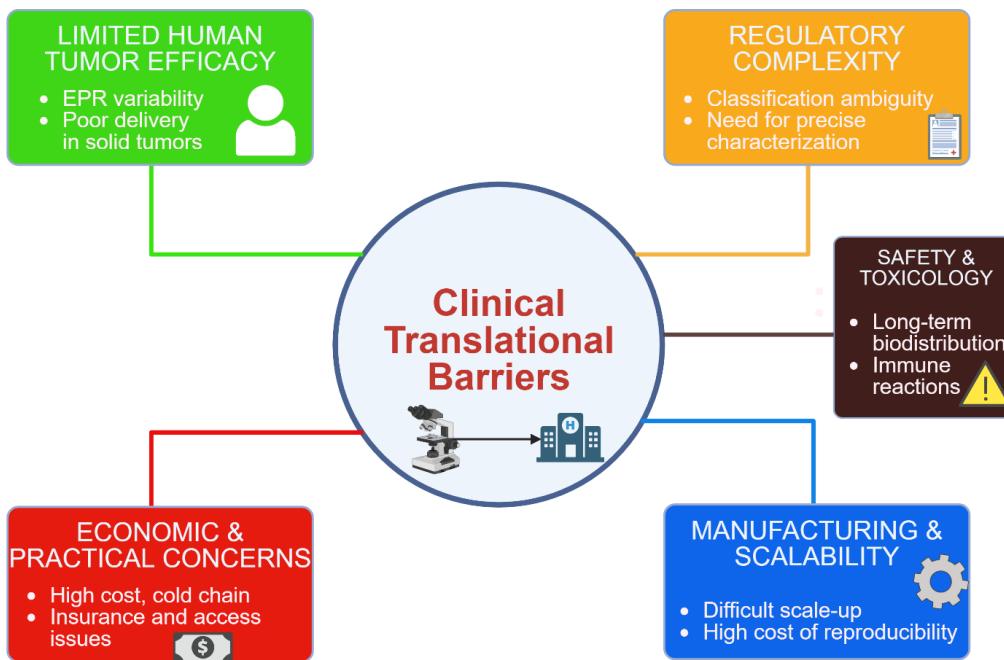
#### 4.4. Efficacy in Human Tumors

A well-known challenge in cancer nanomedicine is the gap between preclinical efficacy (often in mouse xenograft models) and clinical efficacy in human patients [200]. The enhanced permeability and retention (EPR) effect, which allows nanoparticles to accumulate in tumors due to leaky vasculature, is variable in humans – many human tumors have irregular blood supply or higher interstitial pressure that limits nanoparticle penetration [201,202]. Consequently, some nanoparticles that work in mice don't achieve sufficient tumor delivery in patients [203]. Heterogeneity among patients and tumor types means that nanomedicines might benefit some patients more than others [204]. For clinical translation, better methods to image and verify NP delivery in human tumors are needed (e.g., PET or MRI-labeled nanoparticles) to select patients most likely to benefit [204].

#### 4.5. Economic and Practical Considerations

The cost of developing and producing nanomedicines can be substantially higher than for traditional drugs [205]. This includes the cost of specialized facilities and analytical assays. These costs can make nanodrugs expensive, potentially limiting patient access or insurance acceptance [206]. Additionally, regulatory approval times can be longer due to the need for expert consultations

on nanotech aspects [207]. From a practical standpoint, the shelf-life and storage conditions of nanomedicines (some may require cold storage or have shorter stability) can be a logistical challenge for distribution [208,209]. Real-world implementation of nanomedicine must also account for disparities in patient comprehension, as limited health literacy has been shown to hinder cancer care engagement and decision-making in vulnerable populations [210].



**Figure 4.** Key challenges in the clinical translation of nanomedicine for multidrug-resistant (MDR) cancers. The schematic summarizes five major hurdles: (1) regulatory complexity due to overlapping classification and lack of standardization; (2) safety and toxicity concerns including unpredictable biodistribution and immune responses; (3) manufacturing and scalability issues affecting reproducibility and cost; (4) limited clinical efficacy stemming from tumor heterogeneity and variable nanoparticle delivery; and (5) economic and logistical barriers related to high production costs, cold chain requirements, and limited accessibility. Addressing these interconnected limitations is essential for successful bench-to-bedside translation.

Despite these challenges, progress is being made. A growing number of nanomedicines have achieved clinical approval (including many liposomal drugs, polymer-drug conjugates, and nanoparticle-albumin-bound drugs), proving that regulatory and manufacturing hurdles are surmountable [211]. As of mid-2025, over 100 nanotechnology-based therapeutics or diagnostics have been approved for clinical use [212], though most are reformulations of existing drugs rather than novel MDR-targeting systems [213]. Importantly, lessons from these successes are informing next-generation designs: for instance, using biodegradable materials to improve safety, simplifying nanoparticle compositions to ease manufacturing, and developing clear regulatory standards [214]. Collaborations between academia, industry, and regulatory bodies are ongoing to refine guidelines specific to nanomedicine [215]. In parallel, clinical trial designs are adapting, sometimes incorporating patient stratification (e.g., selecting patients with leaky tumors or certain biomarkers for nanoparticle uptake) to demonstrate benefit [216,217].

## 5. Conclusions

In conclusion, while nanomedicine offers innovative solutions to overcome cancer MDR – through targeted delivery, co-therapy, microenvironment modulation, and externally-triggered release – the journey from bench to bedside requires careful navigation of regulatory, safety, and

production challenges. Ongoing innovations and a better understanding of how nanoparticles behave in patients will help ensure that the most promising laboratory breakthroughs can be translated into real-world cancer therapies [218]. The continued success of nanomedicine in the clinic will pave the way for these multi-modal MDR-reversing strategies to become part of standard cancer care in the future.

**Author Contributions:** For research articles with several authors, a short paragraph specifying their individual contributions must be provided. The following statements should be used “Conceptualization, S.K.Y., V.B.C., K.B.R.; methodology, S.K.Y., V.B.C., K.B.R.; software, N.U.P., M.S.D.; validation, N.U.P., M.S.D., K.B.R.; formal analysis, M.S.D., K.B.R.; investigation, N.U.P., M.S.D., K.B.R.; resources, N.U.P., M.S.D., K.B.R.; data curation, S.K.Y., V.B.C.; writing—original draft preparation, S.K.Y., V.B.C.; writing—review and editing, N.U.P., M.S.D., K.B.R.; visualization, N.U.P., M.S.D.; supervision, N.U.P., M.S.D., K.B.R.; project administration, N.U.P., M.S.D., K.B.R.; funding acquisition, S.K.Y., K.B.R.

**Funding:** NIL.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Acknowledgments:** Not applicable.

**Conflicts of Interest:** NIL.

## References

1. Bray, F.; Laversanne, M.; Sung, H.; Ferlay, J.; Siegel, R.L.; Soerjomataram, I.; Jemal, A. Global Cancer Statistics 2022: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* **2024**, *74*, 229–263, doi:10.3322/caac.21834.
2. Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J Clin* **2021**, *71*, 209–249, doi:10.3322/caac.21660.
3. Ferlay, J.; Colombet, M.; Soerjomataram, I.; Parkin, D.M.; Piñeros, M.; Znaor, A.; Bray, F. Cancer Statistics for the Year 2020: An Overview. *Int J Cancer* **2021**, doi:10.1002/ijc.33588.
4. Emran, T.B.; Shahriar, A.; Mahmud, A.R.; Rahman, T.; Abir, M.H.; Siddiquee, M.F.-R.; Ahmed, H.; Rahman, N.; Nainu, F.; Wahyudin, E.; et al. Multidrug Resistance in Cancer: Understanding Molecular Mechanisms, Immunoprevention and Therapeutic Approaches. *Front. Oncol.* **2022**, *12*, doi:10.3389/fonc.2022.891652.
5. Ward, R.A.; Fawell, S.; Floc'h, N.; Flemington, V.; McKerrecher, D.; Smith, P.D. Challenges and Opportunities in Cancer Drug Resistance. *Chem Rev* **2021**, *121*, 3297–3351, doi:10.1021/acs.chemrev.0c00383.
6. Roszkowska, M. Multilevel Mechanisms of Cancer Drug Resistance. *International Journal of Molecular Sciences* **2024**, *25*, 12402, doi:10.3390/ijms252212402.
7. Wang, X.; Zhang, H.; Chen, X. Drug Resistance and Combating Drug Resistance in Cancer. *Cancer Drug Resist* **2019**, *2*, 141–160, doi:10.20517/cdr.2019.10.
8. Vasan, N.; Baselga, J.; Hyman, D.M. A View on Drug Resistance in Cancer. *Nature* **2019**, *575*, 299–309, doi:10.1038/s41586-019-1730-1.
9. Huddleston, C.; Mani, C.; Sah, N.; Courtney, E.; Reese, K.; Stroever, S.; Palle, K.; Reedy, M.B. Evaluating Efficacy of Cervical HPV-HR DNA Testing as Alternative to PET/CT Imaging for Posttreatment Cancer Surveillance: Retrospective Proof-of-Concept Study. *Cancer Epidemiology, Biomarkers & Prevention* **2025**, OF1–OF5, doi:10.1158/1055-9965.EPI-24-1828.
10. Huddleston, C.; Fakhreddine, A.B.; Stroever, S.; Young, R.; Sah, N.; Palle, K.; Reedy, M. Abstract 1305: Comparison of HPV-DNA Testing to PET-CT Imaging as Prognostic Test Following Definitive Treatment for Cervical Cancer: A Retrospective Proof-of-Concept Study. *Cancer Research* **2024**, *84*, 1305, doi:10.1158/1538-7445.AM2024-1305.

11. Sabir, S.; Thani, A.S.B.; Abbas, Q. Nanotechnology in Cancer Treatment: Revolutionizing Strategies against Drug Resistance. *Front Bioeng Biotechnol* **2025**, *13*, 1548588, doi:10.3389/fbioe.2025.1548588.
12. Souri, M.; Soltani, M.; Moradi Kashkooli, F.; Kiani Shahvandi, M. Engineered Strategies to Enhance Tumor Penetration of Drug-Loaded Nanoparticles. *J Control Release* **2022**, *341*, 227–246, doi:10.1016/j.jconrel.2021.11.024.
13. Chen, D.; Liu, X.; Lu, X.; Tian, J. Nanoparticle Drug Delivery Systems for Synergistic Delivery of Tumor Therapy. *Front. Pharmacol.* **2023**, *14*, doi:10.3389/fphar.2023.1111991.
14. Rehman, M.U.; Khan, A.; Imtiyaz, Z.; Ali, S.; Makeen, H.A.; Rashid, S.; Arafah, A. Current Nano-Therapeutic Approaches Ameliorating Inflammation in Cancer Progression. *Semin Cancer Biol* **2022**, *86*, 886–908, doi:10.1016/j.semcan.2022.02.006.
15. Rehman, M.U.; Khan, A.; Imtiyaz, Z.; Ali, S.; Makeen, H.A.; Rashid, S.; Arafah, A. Current Nano-Therapeutic Approaches Ameliorating Inflammation in Cancer Progression. *Semin Cancer Biol* **2022**, *86*, 886–908, doi:10.1016/j.semcan.2022.02.006.
16. Wilkens, S. Structure and Mechanism of ABC Transporters. *F1000Prime Rep* **2015**, *7*, 14, doi:10.12703/P7-14.
17. Alfarouk, K.O.; Stock, C.-M.; Taylor, S.; Walsh, M.; Muddathir, A.K.; Verduzco, D.; Bashir, A.H.H.; Mohammed, O.Y.; Elhassan, G.O.; Harguindeguy, S.; et al. Resistance to Cancer Chemotherapy: Failure in Drug Response from ADME to P-Gp. *Cancer Cell Int* **2015**, *15*, 71, doi:10.1186/s12935-015-0221-1.
18. Wang, J.; Seebacher, N.; Shi, H.; Kan, Q.; Duan, Z. Novel Strategies to Prevent the Development of Multidrug Resistance (MDR) in Cancer. *Oncotarget* **2017**, *8*, 84559–84571, doi:10.18632/oncotarget.19187.
19. Targeting Interleukin-6 as a Strategy to Overcome Stroma-Induced Resistance to Chemotherapy in Gastric Cancer - PubMed Available online: <https://pubmed.ncbi.nlm.nih.gov/30927911/> (accessed on 28 May 2025).
20. Chen, S.-Q.; Li, J.-Q.; Wang, X.-Q.; Lei, W.-J.; Li, H.; Wan, J.; Hu, Z.; Zou, Y.-W.; Wu, X.-Y.; Niu, H.-X. EZH2-Inhibitor DZNep Enhances Apoptosis of Renal Tubular Epithelial Cells in Presence and Absence of Cisplatin. *Cell Div* **2020**, *15*, 8, doi:10.1186/s13008-020-00064-3.
21. Deng, Y.W.; Hao, W.J.; Li, Y.W.; Li, Y.X.; Zhao, B.C.; Lu, D. Hsa-miRNA-143-3p Reverses Multidrug Resistance of Triple-Negative Breast Cancer by Inhibiting the Expression of Its Target Protein Cytokine-Induced Apoptosis Inhibitor 1 In Vivo. *J Breast Cancer* **2018**, *21*, 251–258, doi:10.4048/jbc.2018.21.e40.
22. Venkatadri, R.; Muni, T.; Iyer, A.K.V.; Yakisich, J.S.; Azad, N. Role of Apoptosis-Related miRNAs in Resveratrol-Induced Breast Cancer Cell Death. *Cell Death Dis* **2016**, *7*, e2104, doi:10.1038/cddis.2016.6.
23. Archer, T.C.; Ehrenberger, T.; Mundt, F.; Gold, M.P.; Krug, K.; Mah, C.K.; Mahoney, E.L.; Daniel, C.J.; LeNail, A.; Ramamoorthy, D.; et al. Proteomics, Post-Translational Modifications, and Integrative Analyses Reveal Molecular Heterogeneity within Medulloblastoma Subgroups. *Cancer Cell* **2018**, *34*, 396–410.e8, doi:10.1016/j.ccr.2018.08.004.
24. Devi, G.R.; Finetti, P.; Morse, M.A.; Lee, S.; de Nonneville, A.; Van Laere, S.; Troy, J.; Geraerts, J.; McCall, S.; Bertucci, F. Expression of X-Linked Inhibitor of Apoptosis Protein (XIAP) in Breast Cancer Is Associated with Shorter Survival and Resistance to Chemotherapy. *Cancers* **2021**, *13*, 2807, doi:10.3390/cancers13112807.
25. Targeting P53 Pathways: Mechanisms, Structures and Advances in Therapy | Signal Transduction and Targeted Therapy Available online: <https://www.nature.com/articles/s41392-023-01347-1> (accessed on 29 May 2025).
26. Peiris-Pagès, M.; Bonuccelli, G.; Sotgia, F.; Lisanti, M.P. Mitochondrial Fission as a Driver of Stemness in Tumor Cells: mDIV1 Inhibits Mitochondrial Function, Cell Migration and Cancer Stem Cell (CSC) Signalling. *Oncotarget* **2018**, *9*, 13254–13275, doi:10.18632/oncotarget.24285.
27. Kashatus, D.F. The Regulation of Tumor Cell Physiology by Mitochondrial Dynamics. *Biochem Biophys Res Commun* **2018**, *500*, 9–16, doi:10.1016/j.bbrc.2017.06.192.
28. Fas Receptor - an Overview | ScienceDirect Topics Available online: <https://www.sciencedirect.com/topics/biochemistry-genetics-and-molecular-biology/fas-receptor> (accessed on 29 May 2025).
29. Leonard, B.C.; Johnson, D.E. Signaling by Cell Surface Death Receptors: Alterations in Head and Neck Cancer. *Adv Biol Regul* **2018**, *67*, 170–178, doi:10.1016/j.jbior.2017.10.006.
30. Henke, E.; Nandigama, R.; Ergün, S. Extracellular Matrix in the Tumor Microenvironment and Its Impact on Cancer Therapy. *Front Mol Biosci* **2020**, *6*, 160, doi:10.3389/fmolb.2019.00160.

31. Tao, L.; Huang, G.; Song, H.; Chen, Y.; Chen, L. Cancer Associated Fibroblasts: An Essential Role in the Tumor Microenvironment. *Oncol Lett* **2017**, *14*, 2611–2620, doi:10.3892/ol.2017.6497.
32. Sobanski, T.; Rose, M.; Suraweera, A.; O'Byrne, K.; Richard, D.J.; Bolderson, E. Cell Metabolism and DNA Repair Pathways: Implications for Cancer Therapy. *Front Cell Dev Biol* **2021**, *9*, 633305, doi:10.3389/fcell.2021.633305.
33. Bhat, G.R.; Sethi, I.; Sadida, H.Q.; Rah, B.; Mir, R.; Algehainy, N.; Albalawi, I.A.; Masoodi, T.; Subbaraj, G.K.; Jamal, F.; et al. Cancer Cell Plasticity: From Cellular, Molecular, and Genetic Mechanisms to Tumor Heterogeneity and Drug Resistance. *Cancer Metastasis Rev* **2024**, *43*, 197–228, doi:10.1007/s10555-024-10172-z.
34. Torborg, S.R.; Li, Z.; Chan, J.E.; Tammela, T. Cellular and Molecular Mechanisms of Plasticity in Cancer. *Trends Cancer* **2022**, *8*, 735–746, doi:10.1016/j.trecan.2022.04.007.
35. Wang, Q.; Shao, X.; Zhang, Y.; Zhu, M.; Wang, F.X.C.; Mu, J.; Li, J.; Yao, H.; Chen, K. Role of Tumor Microenvironment in Cancer Progression and Therapeutic Strategy. *Cancer Med* **2023**, *12*, 11149–11165, doi:10.1002/cam4.5698.
36. Anderson, N.M.; Simon, M.C. Tumor Microenvironment. *Curr Biol* **2020**, *30*, R921–R925, doi:10.1016/j.cub.2020.06.081.
37. Hassan Venkatesh, G.; Abou Khouzam, R.; Shaaban Moustafa Elsayed, W.; Ahmed Zeinelabdin, N.; Terry, S.; Chouaib, S. Tumor Hypoxia: An Important Regulator of Tumor Progression or a Potential Modulator of Tumor Immunogenicity? *Oncoimmunology* **10**, 1974233, doi:10.1080/2162402X.2021.1974233.
38. Zheng, Z.; Bian, C.; Wang, H.; Su, J.; Meng, L.; Xin, Y.; Jiang, X. Prediction of Immunotherapy Efficacy and Immunomodulatory Role of Hypoxia in Colorectal Cancer. *Ther Adv Med Oncol* **2022**, *14*, 17588359221138383, doi:10.1177/17588359221138383.
39. Kazakova, A.N.; Lukina, M.M.; Anufrieva, K.S.; Bekbaeva, I.V.; Ivanova, O.M.; Shnaider, P.V.; Slonov, A.; Arapidi, G.P.; Shender, V.O. Exploring the Diversity of Cancer-Associated Fibroblasts: Insights into Mechanisms of Drug Resistance. *Front Cell Dev Biol* **2024**, *12*, 1403122, doi:10.3389/fcell.2024.1403122.
40. Pan, Y.; Yu, Y.; Wang, X.; Zhang, T. Tumor-Associated Macrophages in Tumor Immunity. *Front Immunol* **2020**, *11*, 583084, doi:10.3389/fimmu.2020.583084.
41. Sui, H.; Dongye, S.; Liu, X.; Xu, X.; Wang, L.; Jin, C.Q.; Yao, M.; Gong, Z.; Jiang, D.; Zhang, K.; et al. Immunotherapy of Targeting MDSCs in Tumor Microenvironment. *Front Immunol* **2022**, *13*, 990463, doi:10.3389/fimmu.2022.990463.
42. Gregg, C. Starvation and Climate Change—How to Constrain Cancer Cell Epigenetic Diversity and Adaptability to Enhance Treatment Efficacy. *Front. Ecol. Evol.* **2021**, *9*, doi:10.3389/fevo.2021.693781.
43. Hajji, N.; García-Domínguez, Daniel J; Hontecillas-Prieto, Lourdes; O'Neill, Kevin; de Álava, Enrique; and Syed, N. The Bitter Side of Epigenetics: Variability and Resistance to Chemotherapy. *Epigenomics* **2021**, *13*, 397–403, doi:10.2217/epi-2017-0112.
44. Lai, T.-Y.; Ko, Y.-C.; Chen, Y.-L.; Lin, S.-F. The Way to Malignant Transformation: Can Epigenetic Alterations Be Used to Diagnose Early-Stage Head and Neck Cancer? *Biomedicines* **2023**, *11*, 1717, doi:10.3390/biomedicines11061717.
45. Zhang, Q.; Li, F. Combating P-Glycoprotein-Mediated Multidrug Resistance Using Therapeutic Nanoparticles. *Current Pharmaceutical Design* **2013**, *19*, 6655–6666.
46. Wu, Z.; Zhang, J.; Hao, J.; Liu, P.; Liu, X. Understanding Efflux-Mediated Multidrug Resistance in *Botryotis Cinerea* for Improved Management of Fungicide Resistance. *Microbial Biotechnology* **2025**, *18*, e70074, doi:10.1111/1751-7915.70074.
47. Qiao, D.; Tang, S.; Aslam, S.; Ahmad, M.; To, K.K.W.; Wang, F.; Huang, Z.; Cai, J.; Fu, L. UMMS-4 Enhanced Sensitivity of Chemotherapeutic Agents to ABCB1-Overexpressing Cells via Inhibiting Function of ABCB1 Transporter.
48. Nie, Y.; Fu, G.; Leng, Y. Nuclear Delivery of Nanoparticle-Based Drug Delivery Systems by Nuclear Localization Signals. *Cells* **2023**, *12*, 1637, doi:10.3390/cells12121637.
49. Kievit, F.M.; Wang, F.Y.; Fang, C.; Mok, H.; Wang, K.; Silber, J.R.; Ellenbogen, R.G.; Zhang, M. Doxorubicin Loaded Iron Oxide Nanoparticles Overcome Multidrug Resistance in Cancer *in Vitro*. *Journal of Controlled Release* **2011**, *152*, 76–83, doi:10.1016/j.jconrel.2011.01.024.

50. Jin, L.; Xu, Y.; Chen, F.; Yu, D.; Liang, H.; Liang, Z.; Liu, Z.; Li, H.; Liu, J.; Tan, H.; et al. Mitochondria-Targeted and pH-Triggered Charge-Convertible Polymeric Micelles for Anticancer Therapy. *Materials & Design* **2022**, *224*, 111290, doi:10.1016/j.matdes.2022.111290.

51. He, W.; Dong, S.; Zeng, Q. Functional Nucleic Acid Nanostructures for Mitochondrial Targeting: The Basis of Customized Treatment Strategies. *Molecules* **2025**, *30*, 1025, doi:10.3390/molecules30051025.

52. Wang, S.; Wang, J.; Chen, Z.; Luo, J.; Guo, W.; Sun, L.; Lin, L. Targeting M2-like Tumor-Associated Macrophages Is a Potential Therapeutic Approach to Overcome Antitumor Drug Resistance. *npj Precis. Onc.* **2024**, *8*, 1–19, doi:10.1038/s41698-024-00522-z.

53. Mendes, B.B.; Sousa, D.P.; Connio, J.; Conde, J. Nanomedicine-Based Strategies to Target and Modulate the Tumor Microenvironment. *Trends in Cancer* **2021**, *7*, 847–862, doi:10.1016/j.trecan.2021.05.001.

54. Extracellular Matrix Remodeling in Tumor Progression and Immune Escape: From Mechanisms to Treatments | Molecular Cancer | Full Text Available online: <https://molecular-cancer.biomedcentral.com/articles/10.1186/s12943-023-01744-8> (accessed on 29 May 2025).

55. Mestiri, S.; Sami, A.; Sah, N.; El-Ella, D.M.A.; Khatoon, S.; Shafique, K.; Raza, A.; Mathkor, D.M.; Haque, S. Cellular Plasticity and Non-Small Cell Lung Cancer: Role of T and NK Cell Immune Evasion and Acquisition of Resistance to Immunotherapies. *Cancer Metastasis Rev* **2025**, *44*, 27, doi:10.1007/s10555-025-10244-8.

56. Wang, Y.; Zhou, Q.; Luo, W.; Yang, X.; Zhang, J.; Lou, Y.; Mao, J.; Chen, J.; Wu, F.; Hou, J.; et al. A Collagenase-Decorated Cu-Based Nanotheranostics: Remodeling Extracellular Matrix for Optimizing Cuproptosis and MRI in Pancreatic Ductal Adenocarcinoma. *J Nanobiotechnology* **2024**, *22*, 689, doi:10.1186/s12951-024-02968-6.

57. Iqubal, M.K.; Kaur, H.; Md, S.; Alhakamy, N.A.; Iqubal, A.; Ali, J.; Baboota, S. A Technical Note on Emerging Combination Approach Involved in the Onconanotherapeutics. *Drug Deliv* **29**, 3197–3212, doi:10.1080/10717544.2022.2132018.

58. Schumacher, T.J.; Sah, N.; Palle, K.; Rumbley, J.; Mereddy, V.R. Synthesis and Biological Evaluation of Benzofuran Piperazine Derivatives as Potential Anticancer Agents. *Bioorganic & Medicinal Chemistry Letters* **2023**, *93*, 129425, doi:10.1016/j.bmcl.2023.129425.

59. Gabizon, A.; Ohana, P.; Amitay, Y.; Gorin, J.; Tzemach, D.; Mak, L.; Shmeeda, H. Liposome Co-Encapsulation of Anti-Cancer Agents for Pharmacological Optimization of Nanomedicine-Based Combination Chemotherapy. *Cancer Drug Resist* **2021**, *4*, 463–484, doi:10.20517/cdr.2020.87.

60. Fang, G.; Zhang, A.; Zhu, L.; Wang, Q.; Sun, F.; Tang, B. Nanocarriers Containing Platinum Compounds for Combination Chemotherapy. *Front Pharmacol* **2022**, *13*, 1050928, doi:10.3389/fphar.2022.1050928.

61. Xu, M.; Han, X.; Xiong, H.; Gao, Y.; Xu, B.; Zhu, G.; Li, J. Cancer Nanomedicine: Emerging Strategies and Therapeutic Potentials. *Molecules* **2023**, *28*, 5145, doi:10.3390/molecules28135145.

62. Wang, H.; Borlongan, M.; Hemminki, A.; Basnet, S.; Sah, N.; Kaufman, H.L.; Rabkin, S.D.; Saha, D. Viral Vectors Expressing Interleukin 2 for Cancer Immunotherapy. *Human Gene Therapy* **2023**, *34*, 878–895, doi:10.1089/hum.2023.099.

63. Chehelgerdi, M.; Chehelgerdi, M.; Allela, O.Q.B.; Pecho, R.D.C.; Jayasankar, N.; Rao, D.P.; Thamaraikani, T.; Vasanthan, M.; Viktor, P.; Lakshmaiya, N.; et al. Progressing Nanotechnology to Improve Targeted Cancer Treatment: Overcoming Hurdles in Its Clinical Implementation. *Mol Cancer* **2023**, *22*, 169, doi:10.1186/s12943-023-01865-0.

64. YU, B.; TAI, H.C.; XUE, W.; LEE, L.J.; LEE, R.J. Receptor-Targeted Nanocarriers for Therapeutic Delivery to Cancer. *Mol Membr Biol* **2010**, *27*, 286–298, doi:10.3109/09687688.2010.521200.

65. Nethi, S.K.; Bhatnagar, S.; Prabha, S. Synthetic Receptor-Based Targeting Strategies to Improve Tumor Drug Delivery. *AAPS PharmSciTech* **2021**, *22*, 93, doi:10.1208/s12249-021-01919-w.

66. Altuwaijri, N.; Atef, E. Transferrin-Conjugated Nanostructured Lipid Carriers for Targeting Artemisone to Melanoma Cells. *Int J Mol Sci* **2024**, *25*, 9119, doi:10.3390/ijms25169119.

67. Scheeren, L.E.; Nogueira-Librelootto, D.R.; Macedo, L.B.; de Vargas, J.M.; Mitjans, M.; Vinardell, M.P.; Rolim, C.M.B. Transferrin-Conjugated Doxorubicin-Loaded PLGA Nanoparticles with pH-Responsive Behavior: A Synergistic Approach for Cancer Therapy. *J Nanopart Res* **2020**, *22*, 72, doi:10.1007/s11051-020-04798-7.

68. Tosca, E.M.; Ronchi, D.; Facciolo, D.; Magni, P. Replacement, Reduction, and Refinement of Animal Experiments in Anticancer Drug Development: The Contribution of 3D In Vitro Cancer Models in the Drug Efficacy Assessment. *Biomedicines* **2023**, *11*, 1058, doi:10.3390/biomedicines11041058.

69. Nunes, A.S.; Barros, A.S.; Costa, E.C.; Moreira, A.F.; Correia, I.J. 3D Tumor Spheroids as in Vitro Models to Mimic in Vivo Human Solid Tumors Resistance to Therapeutic Drugs. *Biotechnol Bioeng* **2019**, *116*, 206–226, doi:10.1002/bit.26845.

70. Zhang, J.; Zhang, P.; Zou, Q.; Li, X.; Fu, J.; Luo, Y.; Liang, X.; Jin, Y. Co-Delivery of Gemcitabine and Paclitaxel in cRGD-Modified Long Circulating Nanoparticles with Asymmetric Lipid Layers for Breast Cancer Treatment. *Molecules* **2018**, *23*, 2906, doi:10.3390/molecules23112906.

71. Xing, H.; Hwang, K.; Lu, Y. Recent Developments of Liposomes as Nanocarriers for Theranostic Applications. *Theranostics* **2016**, *6*, 1336–1352, doi:10.7150/thno.15464.

72. Macedo, L.B.; Nogueira-Librelootto, D.R.; Mathes, D.; Pieta, T.B.; Mainardi Pillat, M.; Rosa, R.M. da; Rodrigues, O.E.D.; Vinardell, M.P.; Rolim, C.M.B. Transferrin-Decorated PLGA Nanoparticles Loaded with an Organoselenium Compound as an Innovative Approach to Sensitize MDR Tumor Cells: An In Vitro Study Using 2D and 3D Cell Models. *Nanomaterials* **2023**, *13*, 2306, doi:10.3390/nano13162306.

73. Sah, N.; Shaik, A.A.; Acharya, G.; Dunna, M.; Silwal, A.; Sharma, S.; Khan, S.; Bagchi, S. Receptor-Based Strategies for Overcoming Resistance in Cancer Therapy. *Receptors* **2024**, *3*, 425–443, doi:10.3390/receptors3040021.

74. Ashique, S.; Bhowmick, M.; Pal, R.; Khatoon, H.; Kumar, P.; Sharma, H.; Garg, A.; Kumar, S.; Das, U. Multi Drug Resistance in Colorectal Cancer- Approaches to Overcome, Advancements and Future Success. *Advances in Cancer Biology - Metastasis* **2024**, *10*, 100114, doi:10.1016/j.adcanc.2024.100114.

75. Yang, R.; Liu, Y.; Yang, N.; Zhang, T.; Hou, J.; He, Z.; Wang, Y.; Sun, X.; Shen, J.; Jiang, H.; et al. Combination of miR159 Mimics and Irinotecan Utilizing Lipid Nanoparticles for Enhanced Treatment of Colorectal Cancer. *Pharmaceutics* **2024**, *16*, 570, doi:10.3390/pharmaceutics16040570.

76. Sah, N.; Luna, P.; Mani, C.; Gmeiner, W.; Palle, K. Abstract 6178: A Novel Second-Generation Nano-Fluoropyrimidine to Treat Metastatic Colorectal Cancer and Overcome 5-Fluorouracil Resistance. *Cancer Research* **2023**, *83*, 6178, doi:10.1158/1538-7445.AM2023-6178.

77. Sah, N.; Luna, P.; Mani, C.; Gmeiner, W.; Palle, K. A Novel Fluoropyrimidine Drug to Treat Recalcitrant Colorectal Cancer. *The Journal of Pharmacology and Experimental Therapeutics* **2023**, *385*, 441, doi:10.1124/jpet.122.224620.

78. Okechukwu, C.C.; Ma, X.; Sah, N.; Mani, C.; Palle, K.; Gmeiner, W.H. Enhanced Therapeutic Efficacy of the Nanoscale Fluoropyrimidine Polymer CF10 in a Rat Colorectal Cancer Liver Metastasis Model. *Cancers* **2024**, *16*, 1360, doi:10.3390/cancers16071360.

79. Lainetti, P. de F.; Leis-Filho, A.F.; Laufer-Amorim, R.; Battazza, A.; Fonseca-Alves, C.E. Mechanisms of Resistance to Chemotherapy in Breast Cancer and Possible Targets in Drug Delivery Systems. *Pharmaceutics* **2020**, *12*, 1193, doi:10.3390/pharmaceutics12121193.

80. Murren, J.R.; Durivage, H.J.; Buzaid, A.C.; Reiss, M.; Flynn, S.D.; Carter, D.; Hait, W.N. Trifluoperazine as a Modulator of Multidrug Resistance in Refractory Breast Cancer. *Cancer Chemother Pharmacol* **1996**, *38*, 65–70, doi:10.1007/s002800050449.

81. Zlotnikov, I.D.; Streltsov, D.A.; Ezhov, A.A.; Kudryashova, E.V. Smart pH- and Temperature-Sensitive Micelles Based on Chitosan Grafted with Fatty Acids to Increase the Efficiency and Selectivity of Doxorubicin and Its Adjuvant Regarding the Tumor Cells. *Pharmaceutics* **2023**, *15*, 1135, doi:10.3390/pharmaceutics15041135.

82. Butt, A.M.; Amin, M.C.I.M.; Katas, H.; Abdul Murad, N.A.; Jamal, R.; Kesharwani, P. Doxorubicin and siRNA Codelivery via Chitosan-Coated pH-Responsive Mixed Micellar Polyplexes for Enhanced Cancer Therapy in Multidrug-Resistant Tumors. *Mol Pharm* **2016**, *13*, 4179–4190, doi:10.1021/acs.molpharmaceut.6b00776.

83. Hu, L.; Song, Z.; Wu, B.; Yang, X.; Chen, F.; Wang, X. Hyaluronic Acid-Modified and Doxorubicin-Loaded Au Nanorings for Dual-Responsive and Dual-Imaging Guided Targeted Synergistic Photothermal Chemotherapy Against Pancreatic Carcinoma. *Int J Nanomedicine* **2024**, *19*, 13429–13442, doi:10.2147/IJN.S476936.

84. Intelligent MoS<sub>2</sub> Nanotheranostic for Targeted and Enzyme-/pH-/NIR-Responsive Drug Delivery To Overcome Cancer Chemotherapy Resistance Guided by PET Imaging | ACS Applied Materials & Interfaces Available online: <https://pubs.acs.org/doi/10.1021/acsami.7b17506>.

85. Cao, Y.; Ren, Q.; Hao, R.; Sun, Z. Innovative Strategies to Boost Photothermal Therapy at Mild Temperature Mediated by Functional Nanomaterials. *Materials & Design* **2022**, *214*, 110391, doi:10.1016/j.matdes.2022.110391.

86. Macedo, L.B.; Nogueira-Librelootto, D.R.; Mathes, D.; Pieta, T.B.; Mainardi Pillat, M.; Rosa, R.M. da; Rodrigues, O.E.D.; Vinardell, M.P.; Rolim, C.M.B. Transferrin-Decorated PLGA Nanoparticles Loaded with an Organoselenium Compound as an Innovative Approach to Sensitize MDR Tumor Cells: An In Vitro Study Using 2D and 3D Cell Models. *Nanomaterials* **2023**, *13*, 2306, doi:10.3390/nano13162306.

87. Gong, J.; Shi, T.; Liu, J.; Pei, Z.; Liu, J.; Ren, X.; Li, F.; Qiu, F. Dual-Drug Codelivery Nanosystems: An Emerging Approach for Overcoming Cancer Multidrug Resistance. *Biomedicine & Pharmacotherapy* **2023**, *161*, 114505, doi:10.1016/j.biopha.2023.114505.

88. Yoo, H.; Kim, Y.; Kim, J.; Cho, H.; Kim, K. Overcoming Cancer Drug Resistance with Nanoparticle Strategies for Key Protein Inhibition. *Molecules* **2024**, *29*, 3994, doi:10.3390/molecules29173994.

89. Luobin, L.; Wanxin, H.; Yingxin, G.; Qinzhou, Z.; Zefeng, L.; Danyang, W.; Huaqin, L. Nanomedicine-Induced Programmed Cell Death in Cancer Therapy: Mechanisms and Perspectives. *Cell Death Discov.* **2024**, *10*, 1–13, doi:10.1038/s41420-024-02121-0.

90. Yang, H.; Mao, W.; Rodriguez-Aguayo, C.; Mangala, L.S.; Bartholomeusz, G.; Iles, L.R.; Jennings, N.B.; Ahmed, A.A.; Sood, A.K.; Lopez-Berestein, G.; et al. Paclitaxel Sensitivity of Ovarian Cancer Can Be Enhanced by Knocking down Pairs of Kinases That Regulate MAP4 Phosphorylation and Microtubule Stability. *Clin Cancer Res* **2018**, *24*, 5072–5084, doi:10.1158/1078-0432.CCR-18-0504.

91. Demuytere, J.; Carlier, C.; Van de Sande, L.; Hoorens, A.; De Clercq, K.; Giordano, S.; Morosi, L.; Matteo, C.; Zucchetti, M.; Davoli, E.; et al. Preclinical Activity of Two Paclitaxel Nanoparticle Formulations After Intraperitoneal Administration in Ovarian Cancer Murine Xenografts. *Int J Nanomedicine* **2024**, *19*, 429–440, doi:10.2147/IJN.S424045.

92. Nagumo, Y.; Villareal, M.O.; Isoda, H.; Usui, T. RSK4 Confers Paclitaxel Resistance to Ovarian Cancer Cells, Which Is Resensitized by Its Inhibitor BI-D1870. *Biochemical and Biophysical Research Communications* **2023**, *679*, 23–30, doi:10.1016/j.bbrc.2023.08.060.

93. Nunes, M.; Silva, P.M.A.; Coelho, R.; Pinto, C.; Resende, A.; Bousbaa, H.; Almeida, G.M.; Ricardo, S. Generation of Two Paclitaxel-Resistant High-Grade Serous Carcinoma Cell Lines With Increased Expression of P-Glycoprotein. *Front Oncol* **2021**, *11*, 752127, doi:10.3389/fonc.2021.752127.

94. Summey, R.; Uyar, D. Ovarian Cancer Resistance to PARPi and Platinum-Containing Chemotherapy. *Cancer Drug Resist* **2022**, *5*, 637–646, doi:10.20517/cdr.2021.146.

95. The Future of Ovarian Cancer: Innovation, Treatment and Hope Available online: <https://www.icr.ac.uk/about-us/icr-news/detail/the-future-of-ovarian-cancer--innovation--treatment-and-hope> (accessed on 29 May 2025).

96. Kaur, P.; Singh, S.K.; Mishra, M.K.; Singh, S.; Singh, R. Nanotechnology for Boosting Ovarian Cancer Immunotherapy. *Journal of Ovarian Research* **2024**, *17*, 202, doi:10.1186/s13048-024-01507-z.

97. Kamli, H.; Li, L.; Gobe, G.C. Limitations to the Therapeutic Potential of Tyrosine Kinase Inhibitors and Alternative Therapies for Kidney Cancer. *Ochsner J* **2019**, *19*, 138–151, doi:10.31486/toj.18.0015.

98. Yang, Q.; Wang, Y.; Yang, Q.; Gao, Y.; Duan, X.; Fu, Q.; Chu, C.; Pan, X.; Cui, X.; Sun, Y. Cuprous Oxide Nanoparticles Trigger ER Stress-Induced Apoptosis by Regulating Copper Trafficking and Overcoming Resistance to Sunitinib Therapy in Renal Cancer. *Biomaterials* **2017**, *146*, 72–85, doi:10.1016/j.biomaterials.2017.09.008.

99. Lai, Y.; Zeng, T.; Liang, X.; Wu, W.; Zhong, F.; Wu, W. Cell Death-Related Molecules and Biomarkers for Renal Cell Carcinoma Targeted Therapy. *Cancer Cell International* **2019**, *19*, 221, doi:10.1186/s12935-019-0939-2.

100. Lei, G.; Tang, L.; Yu, Y.; Bian, W.; Yu, L.; Zhou, J.; Li, Y.; Wang, Y.; Du, J. The Potential of Targeting Cuproptosis in the Treatment of Kidney Renal Clear Cell Carcinoma. *Biomedicine & Pharmacotherapy* **2023**, *167*, 115522, doi:10.1016/j.biopha.2023.115522.

101. Li, J.; Wu, K.; Zhang, J.; Gao, H.; Xu, X. Progress in the Treatment of Drug-Loaded Nanomaterials in Renal Cell Carcinoma. *Biomedicine & Pharmacotherapy* **2023**, *167*, 115444, doi:10.1016/j.biopha.2023.115444.
102. Zhou, Q.; Meng, Y.; Li, D.; Yao, L.; Le, J.; Liu, Y.; Sun, Y.; Zeng, F.; Chen, X.; Deng, G. Ferroptosis in Cancer: From Molecular Mechanisms to Therapeutic Strategies. *Sig Transduct Target Ther* **2024**, *9*, 1–30, doi:10.1038/s41392-024-01769-5.
103. Jing, L.; Xiao, W.; Hu, Z.; Liu, X.; Yuan, M. A Systematic Review of Nanoparticle-Mediated Ferroptosis in Glioma Therapy. *Int J Nanomedicine* **2025**, *20*, 5779–5797, doi:10.2147/IJN.S523008.
104. Carlos, A.; Mendes, M.; Cruz, M.T.; Pais, A.; Vitorino, C. Ferroptosis Driven by Nanoparticles for Tackling Glioblastoma. *Cancer Letters* **2025**, *611*, 217392, doi:10.1016/j.canlet.2024.217392.
105. Wu, B.; Zhang, B.; Li, B.; Wu, H.; Jiang, M. Cold and Hot Tumors: From Molecular Mechanisms to Targeted Therapy. *Sig Transduct Target Ther* **2024**, *9*, 1–65, doi:10.1038/s41392-024-01979-x.
106. Mohapatra, A.; Mohanty, A.; Park, I.-K. Inorganic Nanomedicine—Mediated Ferroptosis: A Synergistic Approach to Combined Cancer Therapies and Immunotherapy. *Cancers* **2024**, *16*, 3210, doi:10.3390/cancers16183210.
107. Acharya, G.; Mani, C.; Sah, N.; Saamarthy, K.; Young, R.; Reedy, M.B.; Sobol, R.W.; Palle, K. CHK1 Inhibitor Induced PARylation by Targeting PARG Causes Excessive Replication and Metabolic Stress and Overcomes Chemoresistance in Ovarian Cancer. *Cell Death Discov.* **2024**, *10*, 1–15, doi:10.1038/s41420-024-02040-0.
108. Targeting Metabolic and DNA Damage Checkpoint Indu...: Full Text Finder Results Available online: <https://resolver-ebscohost-com.ezproxy.ttuhs.edu/openurl?sid=google&auinit=G&aulast=Acharya&atitle=Targeting+Metabolic+an+d+DNA+Damage+Checkpoint+Induces+Excessive+DNA+Damage+and+Causes+Synergistic+Lethality+in+GLShigh+Chemoresistant+Ovarian+Cancer+Cells&title=Environmental+and+Molecular+Mutagenesis&volume=65&date=2024&spage=18&issn=0893-6692> (accessed on 10 June 2025).
109. Long, X.; Yan, J.; Zhang, Z.; Chang, J.; He, B.; Sun, Y.; Liang, Y. Autophagy-Targeted Nanoparticles for Effective Cancer Treatment: Advances and Outlook. *NPG Asia Mater* **2022**, *14*, 1–16, doi:10.1038/s41427-022-00422-3.
110. Wei, W.; Rosenkrans, Z.T.; Luo, Q.-Y.; Lan, X.; Cai, W. Exploiting Nanomaterial-Mediated Autophagy for Cancer Therapy. *Small Methods* **2019**, *3*, 1800365, doi:10.1002/smtd.201800365.
111. Yuan, Z.; He, J.; Li, Z.; Fan, B.; Zhang, L.; Man, X. Targeting Autophagy in Urological System Cancers: From Underlying Mechanisms to Therapeutic Implications. *Biochimica et Biophysica Acta (BBA) - Reviews on Cancer* **2024**, *1879*, 189196, doi:10.1016/j.bbcan.2024.189196.
112. Long, X.; Yan, J.; Zhang, Z.; Chang, J.; He, B.; Sun, Y.; Liang, Y. Autophagy-Targeted Nanoparticles for Effective Cancer Treatment: Advances and Outlook. *NPG Asia Mater* **2022**, *14*, 1–16, doi:10.1038/s41427-022-00422-3.
113. He, K.; Chen, M.; Liu, J.; Du, S.; Ren, C.; Zhang, J. Nanomedicine for Cancer Targeted Therapy with Autophagy Regulation. *Front Immunol* **2024**, *14*, 1238827, doi:10.3389/fimmu.2023.1238827.
114. Dowaidar, M. Guidelines for the Role of Autophagy in Drug Delivery Vectors Uptake Pathways. *Helix* **2024**, *10*, e30238, doi:10.1016/j.helix.2024.e30238.
115. Wang, L.; Chang, Y.; Feng, Y.; Li, X.; Cheng, Y.; Jian, H.; Ma, X.; Zheng, R.; Wu, X.; Xu, K.; et al. Nitric Oxide Stimulated Programmable Drug Release of Nanosystem for Multidrug Resistance Cancer Therapy. *Nano Lett* **2019**, *19*, 6800–6811, doi:10.1021/acs.nanolett.9b01869.
116. de la Cruz-Ojeda, P.; Flores-Campos, R.; Dios-Barbeito, S.; Navarro-Villarán, E.; Muntané, J. Role of Nitric Oxide in Gene Expression Regulation during Cancer: Epigenetic Modifications and Non-Coding RNAs. *Int J Mol Sci* **2021**, *22*, 6264, doi:10.3390/ijms22126264.
117. Cauwenbergh, T.; Ballet, S.; Martin, C. Peptide Hydrogel-Drug Conjugates for Tailored Disease Treatment. *Materials Today Bio* **2025**, *31*, 101423, doi:10.1016/j.mtbio.2024.101423.
118. Falcone, N.; Ermis, M.; Tamay, D.G.; Mecwan, M.; Monirizad, M.; Mathes, T.G.; Jucaud, V.; Choroomi, A.; Barros, N.; Zhu, Y.; et al. Peptide Hydrogels as Immunomaterials and Their Use in Cancer Immunotherapy Delivery. *Adv Healthc Mater* **2023**, *12*, e2301096, doi:10.1002/adhm.202301096.

119. Yadav, P.; Ambudkar, S.V.; Rajendra Prasad, N. Emerging Nanotechnology-Based Therapeutics to Combat Multidrug-Resistant Cancer. *J Nanobiotechnology* **2022**, *20*, 423, doi:10.1186/s12951-022-01626-z.

120. Ashique, S.; Garg, A.; Hussain, A.; Farid, A.; Kumar, P.; Taghizadeh-Hesary, F. Nanodelivery Systems: An Efficient and Target-specific Approach for Drug-resistant Cancers. *Cancer Med* **2023**, *12*, 18797–18825, doi:10.1002/cam4.6502.

121. Wang, L.; Chang, Y.; Feng, Y.; Li, X.; Cheng, Y.; Jian, H.; Ma, X.; Zheng, R.; Wu, X.; Xu, K.; et al. Nitric Oxide Stimulated Programmable Drug Release of Nanosystem for Multidrug Resistance Cancer Therapy. *Nano Lett* **2019**, *19*, 6800–6811, doi:10.1021/acs.nanolett.9b01869.

122. Esmaeilpour, D.; Ghomi, M.; Zare, E.N.; Sillanpää, M. Recent Advances in DNA Nanotechnology for Cancer Detection and Therapy: A Review. *International Journal of Biological Macromolecules* **2025**, *307*, 142136, doi:10.1016/j.ijbiomac.2025.142136.

123. Sousa, C.; Videira, M. Dual Approaches in Oncology: The Promise of siRNA and Chemotherapy Combinations in Cancer Therapies. *Oncotarget* **2025**, *5*, 2, doi:10.3390/oncotarget.5010002.

124. Tong, L.; Liu, D.; Cao, Z.; Zheng, N.; Mao, C.; Liu, S.; He, L.; Liu, S. Research Status and Prospect of Non-Viral Vectors Based on siRNA: A Review. *Int J Mol Sci* **2023**, *24*, 3375, doi:10.3390/ijms24043375.

125. Yhee, J.Y.; Song, S.; Lee, S.J.; Park, S.-G.; Kim, K.-S.; Kim, M.G.; Son, S.; Koo, H.; Kwon, I.C.; Jeong, J.H.; et al. Cancer-Targeted MDR-1 siRNA Delivery Using Self-Cross-Linked Glycol Chitosan Nanoparticles to Overcome Drug Resistance. *Journal of Controlled Release* **2015**, *198*, 1–9, doi:10.1016/j.jconrel.2014.11.019.

126. Mir, S.A.; Hamid, L.; Bader, G.N.; Shoaib, A.; Rahamathulla, M.; Alshahrani, M.Y.; Alam, P.; Shakeel, F. Role of Nanotechnology in Overcoming the Multidrug Resistance in Cancer Therapy: A Review. *Molecules* **2022**, *27*, 6608, doi:10.3390/molecules27196608.

127. Maimaitijiang, A.; He, D.; Li, D.; Li, W.; Su, Z.; Fan, Z.; Li, J. Progress in Research of Nanotherapy for Overcoming Multidrug Resistance in Cancer. *International Journal of Molecular Sciences* **2024**, *25*, 9973, doi:10.3390/ijms25189973.

128. Meng, H.; Mai, W.X.; Zhang, H.; Xue, M.; Xia, T.; Lin, S.; Wang, X.; Zhao, Y.; Ji, Z.; Zink, J.I.; et al. Co-Delivery of an Optimal Drug/siRNA Combination Using Mesoporous Silica Nanoparticle to Overcome Drug Resistance in Breast Cancer In Vitro and In Vivo. *ACS Nano* **2013**, *7*, 994–1005, doi:10.1021/nn3044066.

129. Kim, H.-K.; Cheong, H.; Kim, M.-Y.; Jin, H.-E. Therapeutic Targeting in Ovarian Cancer: Nano-Enhanced CRISPR/Cas9 Gene Editing and Drug Combination Therapy. *Int J Nanomedicine* **2025**, *20*, 3907–3931, doi:10.2147/IJN.S507688.

130. Omy, T.R.; Sah, N.; Reedy, M.; Acharya, G.; Palle, K. Abstract 3090: miRNA-221-5p-Mediated Epigenetic Regulation Promotes Chemoresistance and Offers Therapeutic Potential in Ovarian Cancer. *Cancer Research* **2025**, *85*, 3090, doi:10.1158/1538-7445.AM2025-3090.

131. Sah, N.; Peddibhotla, S.; Richardson, B.; Luna, P.; Bansal, N.A.; Mani, C.; Reedy, M.; Palle, K. Abstract A084: Oncogenic Role for Upregulated Lymphoblastic Leukemia Derived Sequence-1 in the Progression of Ovarian Cancer and Its Metastasis. *Cancer Epidemiology, Biomarkers & Prevention* **2023**, *32*, A084, doi:10.1158/1538-7755.DISP22-A084.

132. Tong, L.; Liu, D.; Cao, Z.; Zheng, N.; Mao, C.; Liu, S.; He, L.; Liu, S. Research Status and Prospect of Non-Viral Vectors Based on siRNA: A Review. *Int J Mol Sci* **2023**, *24*, 3375, doi:10.3390/ijms24043375.

133. Maimaitijiang, A.; He, D.; Li, D.; Li, W.; Su, Z.; Fan, Z.; Li, J. Progress in Research of Nanotherapy for Overcoming Multidrug Resistance in Cancer. *International Journal of Molecular Sciences* **2024**, *25*, 9973, doi:10.3390/ijms25189973.

134. Agiba, A.M.; Arreola-Ramírez, J.L.; Carbajal, V.; Segura-Medina, P. Light-Responsive and Dual-Targeting Liposomes: From Mechanisms to Targeting Strategies. *Molecules* **2024**, *29*, 636, doi:10.3390/molecules29030636.

135. Rout, B.; Maharana, S.K.; Jain, A. Stimuli-Responsive Nanoparticles: A Novel Approach for Melanoma Treatment. *J Nanopart Res* **2025**, *27*, 34, doi:10.1007/s11051-025-06231-3.

136. Szwed, M.; Jost, T.; Majka, E.; Gharibkandi, N.A.; Majkowska-Pilip, A.; Frey, B.; Bilewicz, A.; Fietkau, R.; Gaipl, U.; Marczak, A.; et al. Pt-Au Nanoparticles in Combination with Near-Infrared-Based Hyperthermia Increase the Temperature and Impact on the Viability and Immune Phenotype of Human Hepatocellular Carcinoma Cells. *Int J Mol Sci* **2025**, *26*, 1574, doi:10.3390/ijms26041574.

137. He, X.; Zhang, S.; Tian, Y.; Cheng, W.; Jing, H. Research Progress of Nanomedicine-Based Mild Photothermal Therapy in Tumor. *Int J Nanomedicine* **2023**, *18*, 1433–1468, doi:10.2147/IJN.S405020.

138. Gold Nanoparticles in Photothermal Therapy - CD Bioparticles Available online: <https://www.cd-bioparticles.com/support/gold-nanoparticles-in-photothermal-therapy.html> (accessed on 29 May 2025).

139. Ayana, G.; Ryu, J.; Choe, S. Ultrasound-Responsive Nanocarriers for Breast Cancer Chemotherapy. *Micromachines (Basel)* **2022**, *13*, 1508, doi:10.3390/mi13091508.

140. Pourmadadi, M.; Tajiki, A.; Hosseini, S.M.; Samadi, A.; Abdouss, M.; Daneshnia, S.; Yazdian, F. A Comprehensive Review of Synthesis, Structure, Properties, and Functionalization of MoS<sub>2</sub>; Emphasis on Drug Delivery, Photothermal Therapy, and Tissue Engineering Applications. *Journal of Drug Delivery Science and Technology* **2022**, *76*, 103767, doi:10.1016/j.jddst.2022.103767.

141. Chen, H.-H.; Lu, I.-L.; Liu, T.-I.; Tsai, Y.-C.; Chiang, W.-H.; Lin, S.-C.; Chiu, H.-C. Indocyanine Green/Doxorubicin-Encapsulated Functionalized Nanoparticles for Effective Combination Therapy against Human MDR Breast Cancer. *Colloids Surf B Biointerfaces* **2019**, *177*, 294–305, doi:10.1016/j.colsurfb.2019.02.001.

142. Awad, N.S.; Paul, V.; AlSawaftah, N.M.; ter Haar, G.; Allen, T.M.; Pitt, W.G.; Husseini, G.A. Ultrasound-Responsive Nanocarriers in Cancer Treatment: A Review. *ACS Pharmacol Transl Sci* **2021**, *4*, 589–612, doi:10.1021/acsptsci.0c00212.

143. Al Refaai, K.A.; AlSawaftah, N.A.; Abuwatfa, W.; Husseini, G.A. Drug Release via Ultrasound-Activated Nanocarriers for Cancer Treatment: A Review. *Pharmaceutics* **2024**, *16*, 1383, doi:10.3390/pharmaceutics16111383.

144. Pan, M.; Hu, D.; Yuan, L.; Yu, Y.; Li, Y.; Qian, Z. Newly Developed Gas-Assisted Sonodynamic Therapy in Cancer Treatment. *Acta Pharm Sin B* **2023**, *13*, 2926–2954, doi:10.1016/j.apsb.2022.12.021.

145. Yu, J.; Hu, J.-R.; Tian, Y.; Lei, Y.-M.; Hu, H.-M.; Lei, B.-S.; Zhang, G.; Sun, Y.; Ye, H.-R. Nanosensitizer-Assisted Sonodynamic Therapy for Breast Cancer. *J Nanobiotechnology* **2025**, *23*, 281, doi:10.1186/s12951-025-03311-3.

146. Lv, W.; Wu, H.; Zhang, Y.; Li, H.; Shu, H.; Su, C.; Zhu, Y.; Wang, T.; Nie, F. cRGD-Targeted Gold-Based Nanoparticles Overcome EGFR-TKI Resistance of NSCLC via Low-Temperature Photothermal Therapy Combined with Sonodynamic Therapy. *Biomater. Sci.* **2023**, *11*, 1677–1691, doi:10.1039/D2BM01825J.

147. Collins, V.G.; Hutton, D.; Hossain-Ibrahim, K.; Joseph, J.; Banerjee, S. The Abscopal Effects of Sonodynamic Therapy in Cancer. *Br J Cancer* **2025**, *132*, 409–420, doi:10.1038/s41416-024-02898-y.

148. Zhang, Y.; Zhao, Y.; Zhang, Y.; Liu, Q.; Zhang, M.; Tu, K. The Crosstalk between Sonodynamic Therapy and Autophagy in Cancer. *Front. Pharmacol.* **2022**, *13*, doi:10.3389/fphar.2022.961725.

149. Jeong, Y.-G.; Park, J.-H.; Khang, D. Sonodynamic and Acoustically Responsive Nanodrug Delivery System: Cancer Application. *Int J Nanomedicine* **2024**, *19*, 11767–11788, doi:10.2147/IJN.S496028.

150. Foglietta, F.; Giacone, M.; Durando, G.; Canaparo, R.; Serpe, L. Sonodynamic Treatment Triggers Cancer Cell Killing by Doxorubicin in P-Glycoprotein-Mediated Multidrug Resistant Cancer Models. *Advanced Therapeutics* **2024**, *7*, 2400070, doi:10.1002/adtp.202400070.

151. Tharkar, P.; Varanasi, R.; Wong, W.S.F.; Jin, C.T.; Chrzanowski, W. Nano-Enhanced Drug Delivery and Therapeutic Ultrasound for Cancer Treatment and Beyond. *Front Bioeng Biotechnol* **2019**, *7*, 324, doi:10.3389/fbioe.2019.00324.

152. Bañobre-López, M.; Teijeiro, A.; Rivas, J. Magnetic Nanoparticle-Based Hyperthermia for Cancer Treatment. *Rep Pract Oncol Radiother* **2013**, *18*, 397–400, doi:10.1016/j.rpor.2013.09.011.

153. Li, G.; Wang, C.; Jin, B.; Sun, T.; Sun, K.; Wang, S.; Fan, Z. Advances in Smart Nanotechnology-Supported Photodynamic Therapy for Cancer. *Cell Death Discov.* **2024**, *10*, 1–18, doi:10.1038/s41420-024-02236-4.

154. Deng, W.; Chen, W.; Clement, S.; Guller, A.; Zhao, Z.; Engel, A.; Goldys, E.M. Controlled Gene and Drug Release from a Liposomal Delivery Platform Triggered by X-Ray Radiation. *Nat Commun* **2018**, *9*, 2713, doi:10.1038/s41467-018-05118-3.

155. Gibney, M. X-Ray-Triggered Liposomes Deliver Precisely Controlled Cancer Treatment | Fierce Pharma Available online: <https://www.fiercepharma.com/r-d/x-ray-triggered-liposomes-deliver-precisely-controlled-cancer-treatment> (accessed on 4 June 2025).

156. Lv, W.; Wu, H.; Zhang, Y.; Li, H.; Shu, H.; Su, C.; Zhu, Y.; Wang, T.; Nie, F. cRGD-Targeted Gold-Based Nanoparticles Overcome EGFR-TKI Resistance of NSCLC via Low-Temperature Photothermal Therapy Combined with Sonodynamic Therapy. *Biomater. Sci.* **2023**, *11*, 1677–1691, doi:10.1039/D2BM01825J.

157. Spoială, A.; Ilie, C.-I.; Motelica, L.; Ficai, D.; Semenescu, A.; Oprea, O.-C.; Ficai, A. Smart Magnetic Drug Delivery Systems for the Treatment of Cancer. *Nanomaterials* **2023**, *13*, 876, doi:10.3390/nano13050876.

158. BSN, A.B.B.L. Magnetic Nanoparticles for Drug Delivery Available online: <https://www.news-medical.net/health/Magnetic-Nanoparticles-for-Drug-Delivery.aspx> (accessed on 5 June 2025).

159. Ren, Y.; Zhang, H.; Chen, B.; Cheng, J.; Cai, X.; Liu, R.; Xia, G.; Wu, W.; Wang, S.; Ding, J.; et al. Multifunctional Magnetic Fe<sub>3</sub>O<sub>4</sub> Nanoparticles Combined with Chemotherapy and Hyperthermia to Overcome Multidrug Resistance. *Int J Nanomedicine* **2012**, *7*, 2261–2269, doi:10.2147/IJN.S29357.

160. Yun, W.S.; Kim, J.; Lim, D.-K.; Kim, D.-H.; Jeon, S.I.; Kim, K. Recent Studies and Progress in the Intratumoral Administration of Nano-Sized Drug Delivery Systems. *Nanomaterials* **2023**, *13*, 2225, doi:10.3390/nano13152225.

161. Katopodi, T.; Petanidis, S.; Floros, G.; Porpodis, K.; Kosmidis, C. Hybrid Nanogel Drug Delivery Systems: Transforming the Tumor Microenvironment through Tumor Tissue Editing. *Cells* **2024**, *13*, 908, doi:10.3390/cells13110908.

162. Koirala, M.; DiPaola, M. Overcoming Cancer Resistance: Strategies and Modalities for Effective Treatment. *Biomedicines* **2024**, *12*, 1801, doi:10.3390/biomedicines12081801.

163. Acter, S.; Moreau, M.; Ivkovic, R.; Viswanathan, A.; Ngwa, W. Polydopamine Nanomaterials for Overcoming Current Challenges in Cancer Treatment. *Nanomaterials* **2023**, *13*, 1656, doi:10.3390/nano13101656.

164. Dong, Z.; Gong, H.; Gao, M.; Zhu, W.; Sun, X.; Feng, L.; Fu, T.; Li, Y.; Liu, Z. Polydopamine Nanoparticles as a Versatile Molecular Loading Platform to Enable Imaging-Guided Cancer Combination Therapy. *Theranostics* **2016**, *6*, 1031–1042, doi:10.7150/thno.14431.

165. Li, Z.; Wang, B.; Zhang, Z.; Wang, B.; Xu, Q.; Mao, W.; Tian, J.; Yang, K.; Wang, F. Radionuclide Imaging-Guided Chemo-Radioisotope Synergistic Therapy Using a <sup>131</sup>I-Labeled Polydopamine Multifunctional Nanocarrier. *Mol Ther* **2018**, *26*, 1385–1393, doi:10.1016/j.mtthe.2018.02.019.

166. Zhou, X.; Wang, L.; Wu, Y.; Lin, X.; Liu, F.; Sun, X.; Song, K.; Jiang, C.; Zhao, F.; Li, X. Bioactive Polydopamine Nanomedicines-Assisted Cancer Immunotherapy. *Materials Today Bio* **2025**, *32*, 101864, doi:10.1016/j.mtbiol.2025.101864.

167. Długosz, O.; Matyjasik, W.; Hodacka, G.; Szostak, K.; Matysik, J.; Krawczyk, P.; Piasek, A.; Pilit-Prociak, J.; Banach, M. Inorganic Nanomaterials Used in Anti-Cancer Therapies: Further Developments. *Nanomaterials* **2023**, *13*, 1130, doi:10.3390/nano13061130.

168. Newton Amaldoss, M.J.; Sorrell, C.C. ROS Modulating Inorganic Nanoparticles: A Novel Cancer Therapeutic Tool. *Recent Adv Drug Deliv Formul* **2022**, *16*, 84–89, doi:10.2174/2667387816666220506203123.

169. Mohapatra, A.; Mohanty, A.; Park, I.-K. Inorganic Nanomedicine—Mediated Ferroptosis: A Synergistic Approach to Combined Cancer Therapies and Immunotherapy. *Cancers* **2024**, *16*, 3210, doi:10.3390/cancers16183210.

170. Bharde, A. Targeting Redox Homeostasis in Tumor Cells Using Nanoparticles. In *Handbook of Oxidative Stress in Cancer: Therapeutic Aspects*; Springer, Singapore, 2022; pp. 1–17 ISBN 9789811612473.

171. Pareek, A.; Kumar, D.; Pareek, A.; Gupta, M.M. Advancing Cancer Therapy with Quantum Dots and Other Nanostructures: A Review of Drug Delivery Innovations, Applications, and Challenges. *Cancers (Basel)* **2025**, *17*, 878, doi:10.3390/cancers17050878.

172. He, Z.-Y.; Jin, Z.-H.; Zhan, M.; Qin, Z.; Li, Z.; Xu, T. Advances in Quantum Dot-Mediated siRNA Delivery. *Chinese Chemical Letters* **2017**, *28*, 1851–1856, doi:10.1016/j.cclet.2017.07.012.

173. Wang, N.; Chen, L.; Huang, W.; Gao, Z.; Jin, M. Current Advances of Nanomaterial-Based Oral Drug Delivery for Colorectal Cancer Treatment. *Nanomaterials* **2024**, *14*, 557, doi:10.3390/nano14070557.

174. Kanojia, N.; Thapa, K.; Verma, N.; Rani, L.; Sood, P.; Kaur, G.; Dua, K.; Kumar, J. Update on Mucoadhesive Approaches to Target Drug Delivery in Colorectal Cancer. *Journal of Drug Delivery Science and Technology* **2023**, *87*, 104831, doi:10.1016/j.jddst.2023.104831.

175. Gvozdeva, Y.; Staynova, R. pH-Dependent Drug Delivery Systems for Ulcerative Colitis Treatment. *Pharmaceutics* **2025**, *17*, 226, doi:10.3390/pharmaceutics17020226.

176. Jakobušić Brala, C.; Karković Marković, A.; Kugić, A.; Torić, J.; Barbarić, M. Combination Chemotherapy with Selected Polyphenols in Preclinical and Clinical Studies—An Update Overview. *Molecules* **2023**, *28*, 3746, doi:10.3390/molecules28093746.

177. C. de S. L. Oliveira, A.L.; Schomann, T.; de Geus-Oei, L.-F.; Kapiteijn, E.; Cruz, L.J.; de Araújo Junior, R.F. Nanocarriers as a Tool for the Treatment of Colorectal Cancer. *Pharmaceutics* **2021**, *13*, 1321, doi:10.3390/pharmaceutics13081321.

178. 4dm1n\_diverse technologies Overcoming Regulatory Hurdles in Clinical Translation of Nanomedicine. *DIVERSA* 2025.

179. Foulkes, R.; Man, E.; Thind, J.; Yeung, S.; Joy, A.; Hoskins, C. The Regulation of Nanomaterials and Nanomedicines for Clinical Application: Current and Future Perspectives. *Biomater Sci* **2020**, *8*, 4653–4664, doi:10.1039/d0bm00558d.

180. Agrahari, V.; and Hiremath, P. Challenges Associated and Approaches for Successful Translation of Nanomedicines Into Commercial Products. *Nanomedicine* **2017**, *12*, 819–823, doi:10.2217/nnm-2017-0039.

181. Desai, N. Challenges in Development of Nanoparticle-Based Therapeutics. *AAPS J* **2012**, *14*, 282–295, doi:10.1208/s12248-012-9339-4.

182. Safety Concerns with Nanotechnology in Medicine - Society for Brain Mapping and Therapeutics 2024.

183. Dobrovolskaia, M.A.; Aggarwal, P.; Hall, J.B.; McNeil, S.E. Preclinical Studies To Understand Nanoparticle Interaction with the Immune System and Its Potential Effects on Nanoparticle Biodistribution. *Mol Pharm* **2008**, *5*, 487–495, doi:10.1021/mp800032f.

184. Desai, N. Challenges in Development of Nanoparticle-Based Therapeutics. *AAPS J* **2012**, *14*, 282–295, doi:10.1208/s12248-012-9339-4.

185. Bedőcs, P.; Szebeni, J. The Critical Choice of Animal Models in Nanomedicine Safety Assessment: A Lesson Learned From Hemoglobin-Based Oxygen Carriers. *Front Immunol* **2020**, *11*, 584966, doi:10.3389/fimmu.2020.584966.

186. Sah, N.; Ramaiah, B.; Koneri, R. Sulfasalazine-Induced Drug Rash with Eosinophilia and Systemic Symptoms Syndrome in a Seronegative Spondyloarthritis Patient: A Case Report. *Indian Journal of Pharmacology* **2021**, *53*, 391, doi:10.4103/ijp.IJP\_129\_18.

187. Varghese, J.; Sah, N.; Ramaiah, B. PDG50 Obstacles in the IV to ORAL Antibiotic Shift for Eligible Patients at a Tertiary Care Hospital. *Value in Health* **2020**, *23*, S527, doi:10.1016/j.jval.2020.08.733.

188. Sah, N.; Ramaiah, B.; Abdulla; Gupta, A.K.; Thomas, S.M. Noncompliance with Prescription-Writing Guidelines in an OutpatientDepartment of a Tertiary Care Hospital: A Prospective, Observational Study. *rjps* **2020**, *10*, doi:10.26463/rjps.10\_1\_5.

189. Varghese, J.; Sah, N.; Thomas, S.M.; Jose, J.C.; Ramaiah, B.; Koneri, R. PDG6 Clinical Evaluation of Skin and Soft Tissue Infections in Inpatient Versus Outpatient Setting at a Tertiary Care Hospital - a Prospective Study. *Value in Health* **2020**, *23*, S521–S522, doi:10.1016/j.jval.2020.08.689.

190. Sah, N.; Ramaiah, B.; Koneri, R. The Pharmacist Role in Clinical Audit at an Indian Accredited Hospital: An Interventional Study. *IJOPP* **2019**, *12*, 117–125, doi:10.5530/ijopp.12.2.25.

191. 4dm1n\_diverse technologies Overcoming Regulatory Hurdles in Clinical Translation of Nanomedicine. *DIVERSA* 2025.

192. Agrahari, V.; and Hiremath, P. Challenges Associated and Approaches for Successful Translation of Nanomedicines Into Commercial Products. *Nanomedicine* **2017**, *12*, 819–823, doi:10.2217/nnm-2017-0039.

193. Agrahari, V.; and Hiremath, P. Challenges Associated and Approaches for Successful Translation of Nanomedicines Into Commercial Products. *Nanomedicine* **2017**, *12*, 819–823, doi:10.2217/nnm-2017-0039.

194. Clark, B. Lab to Industry: Scaling Up Nanotechnology for Practical Applications. *Journal of Nanomaterials & Molecular Nanotechnology* **2024**, 2024.

195. Policy Statements & Letters | Advocacy | ASGCT - American Society of Gene & Cell Therapy | ASGCT - American Society of Gene & Cell Therapy Available online: <https://www.asgct.org/advocacy/policy-statement-landing/2023/manufacturing-changes-and-comparability-for-human> (accessed on 5 June 2025).

196. Nguyen, H.-M.; Sah, N.; Humphrey, M.R.M.; Rabkin, S.D.; Saha, D. Growth, Purification, and Titration of Oncolytic Herpes Simplex Virus. *J Vis Exp* **2021**, 10.3791/62677, doi:10.3791/62677.

197. Nanomedicine Market Share, Size & Growth Report | 2032 Available online: <https://www.skyquestt.com/report/nanomedicine-market> (accessed on 5 June 2025).

198. Đorđević, S.; Gonzalez, M.M.; Conejos-Sánchez, I.; Carreira, B.; Pozzi, S.; Acúrcio, R.C.; Satchi-Fainaro, R.; Florindo, H.F.; Vicent, M.J. Current Hurdles to the Translation of Nanomedicines from Bench to the Clinic. *Drug Deliv Transl Res* **2022**, *12*, 500–525, doi:10.1007/s13346-021-01024-2.

199. Navigating Nanomedicine Regulatory Requirements in 2024 Available online: <https://www.izon.com/news/navigating-nanomedicine-regulatory-requirements-in-2023> (accessed on 5 June 2025).

200. Richmond, A.; Su, Y. Mouse Xenograft Models vs GEM Models for Human Cancer Therapeutics. *Dis Model Mech* **2008**, *1*, 78–82, doi:10.1242/dmm.000976.

201. Shen, X.; Pan, D.; Gong, Q.; Gu, Z.; Luo, K. Enhancing Drug Penetration in Solid Tumors via Nanomedicine: Evaluation Models, Strategies and Perspectives. *Bioactive Materials* **2024**, *32*, 445–472, doi:10.1016/j.bioactmat.2023.10.017.

202. Yuan, Z.; Li, Y.; Zhang, S.; Wang, X.; Dou, H.; Yu, X.; Zhang, Z.; Yang, S.; Xiao, M. Extracellular Matrix Remodeling in Tumor Progression and Immune Escape: From Mechanisms to Treatments. *Molecular Cancer* **2023**, *22*, 48, doi:10.1186/s12943-023-01744-8.

203. Kim, J.; Cho, H.; Lim, D.-K.; Joo, M.K.; Kim, K. Perspectives for Improving the Tumor Targeting of Nanomedicine via the EPR Effect in Clinical Tumors. *Int J Mol Sci* **2023**, *24*, 10082, doi:10.3390/ijms241210082.

204. Swetha, K.L.; Roy, A. Tumor Heterogeneity and Nanoparticle-Mediated Tumor Targeting: The Importance of Delivery System Personalization. *Drug Deliv Transl Res* **2018**, *8*, 1508–1526, doi:10.1007/s13346-018-0578-5.

205. Ph.D, D.P.B. Nanomedicine: Advantages and Disadvantages Available online: <https://www.azonano.com/article.aspx?ArticleID=6707> (accessed on 5 June 2025).

206. Bosetti, R.; and Jones, S.L. Cost-Effectiveness of Nanomedicine: Estimating the Real Size of Nano-Costs. *Nanomedicine* **2019**, *14*, 1367–1370, doi:10.2217/nmm-2019-0130.

207. Hejmady, S.; Singhvi, Gautam; Saha, Ranendra Narayan; and Dubey, S.K. Regulatory Aspects in Process Development and Scale-up of Nanopharmaceuticals. *Therapeutic Delivery* **2020**, *11*, 341–343, doi:10.4155/tde-2020-0034.

208. Das, A.; Adhikari, S.; Deka, D.; Bailhya, N.; Sahare, P.; Banerjee, A.; Paul, S.; Bisgin, A.; Pathak, S. An Updated Review on the Role of Nanoformulated Phytochemicals in Colorectal Cancer. *Medicina (Kaunas)* **2023**, *59*, 685, doi:10.3390/medicina59040685.

209. Degobert, G.; Aydin, D. Lyophilization of Nanocapsules: Instability Sources, Formulation and Process Parameters. *Pharmaceutics* **2021**, *13*, 1112, doi:10.3390/pharmaceutics13081112.

210. Richardson, B.; Anderson, A.; Lakey, K.; Hickham, L.; Sah, N.; Mani, C.; Palle, K.; Reedy, M. Assessing the Need and Implementation of Health Literacy Testing for New Gynecologic Oncology Patients in West Texas: A Prospective Pilot Study. *Texas Public Health Journal* **2025**, *77*, 18–24.

211. Wei, A.; Mehtala, J.G.; Patri, A.K. Challenges and Opportunities in the Advancement of Nanomedicines. *J Control Release* **2012**, *164*, 236–246, doi:10.1016/j.jconrel.2012.10.007.

212. Deb, T. Nanomedicine Statistics and Facts (2025). *Market.us Media* 2025.

213. Rodríguez-Gómez, F.D.; Monferrer, D.; Penon, O.; Rivera-Gil, P. Regulatory Pathways and Guidelines for Nanotechnology-Enabled Health Products: A Comparative Review of EU and US Frameworks. *Front Med (Lausanne)* **2025**, *12*, 1544393, doi:10.3389/fmed.2025.1544393.

214. Tracey, S.R.; Smyth, P.; Barelle, C.J.; Scott, C.J. Development of next Generation Nanomedicine-Based Approaches for the Treatment of Cancer: We've Barely Scratched the Surface. *Biochem Soc Trans* **2021**, *49*, 2253–2269, doi:10.1042/BST20210343.

215. 4dm1n\_diversatechnologies Overcoming Regulatory Hurdles in Clinical Translation of Nanomedicine. *DIVERSA* 2025.

216. Clinical Trial Designs Incorporating Predictive Biomarkers. Available online: <https://www.broadinstitute.org/publications/broad7862> (accessed on 5 June 2025).

217. Wang, Y.; Ma, Z.; Wong, K.-C.; Li, X. Nature-Inspired Multiobjective Patient Stratification from Cancer Gene Expression Data. *Information Sciences* **2020**, *526*, 245–262, doi:10.1016/j.ins.2020.03.095.
218. Mao, Y.; Xie, J.; Yang, F.; Luo, Y.; Du, J.; Xiang, H. Advances and Prospects of Precision Nanomedicine in Personalized Tumor Theranostics. *Front Cell Dev Biol* **2024**, *12*, 1514399, doi:10.3389/fcell.2024.1514399.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.