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Article

A G311E Mutant Gene of MATE Family Protein DTX6 Confers Bipyrindyl Herbicide Resistance and Increases Seed Amino Acid Level in Rice

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Abstract: Weeds present a pervasive challenge in agricultural fields. The integration of herbicide-resistant crops with chemical weed management offers an effective solution for sustainable weed control while reducing labor inputs, particularly in large-scale intensive farming systems. In this study, we found that a G311E mutant variant of the *Arabidopsis* MATE (multidrug and toxic compound extrusion) family transporter DTX6, designated DTX6m, confers robust resistance to bipyrindyl herbicides in rice. *DTX6m*-overexpression lines exhibited marked resistance to paraquat and diquat, tolerating diquat concentrations up to 5 g/L, which is five-fold higher than the recommended field application dosage. Agronomic assessments demonstrated that grain yields of *DTX6m*-overexpression plants were statistically equivalent to those of wild-type plants. Moreover, the plants displayed beneficial phenotypic changes, such as accelerated flowering and a slight reduction in plant height. Seed morphometric analysis indicated that, in comparison with the wild-type control, *DTX6m*-transgenic lines exhibited altered grain dimensions while maintaining consistent 1000-grain weight. Nutritional assays further demonstrated that DTX6m increased the free amino acid levels in seeds, while normal protein and starch contents were retained. Collectively, these results establish that *DTX6m* effectively boosts rice resistance to bipyrindyl herbicides, validating DTX6m as a candidate gene for engineering plant herbicide resistance and also implying a potential role for *DTX6m* in amino acid homeostasis in plants.

Keywords: Weed control; DTX6m; bipyrindyl herbicide; yield; nutrition

1. Introduction

Weeds pose a major threat to agricultural productivity in cultivated areas. They compete with crops for essential resources such as water, nutrients, and sunlight, leading to decreased yields and overall agricultural output [1]. Traditional manual weeding methods, while effective, are highly labor-intensive and becoming increasingly unsustainable due to rural labor shortage and declining birth rate [1,2]. As a result, chemical herbicides have become the primary weed control method in modern agriculture, offering both high efficacy and cost efficiency [3]. This shift is particularly crucial for rice, a staple crop for over half of the global population, which faces escalating yield losses from weed infestation, especially as cultivation transitions from transplanting to direct seeding [4,5]. Herbicide application, combined with herbicide-resistant rice varieties, offers an effective weed management solution with minimal labor demands, making it indispensable for large-scale rice production [6].

The identification of herbicide-resistance genes is crucial for developing herbicide-resistant crops. By isolating these genes, which often are derived from resistant weeds, bacteria, or other plant

species, researchers can employ genetic engineering to introduce herbicide tolerance into crops, ultimately producing elite cultivars [7,8]. Through the introduction and upregulation of genes responsible for herbicide detoxification, scientists can enhance the herbicide resistance of crops without negatively affecting their agronomic performance or yield [8,9].

To date, numerous synthetic herbicides with diverse selectivity and modes of action have been commercialized and widely adopted [10,11]. Developing crop cultivars resistant to various herbicides and implementing rotations of crops with different herbicide resistances are essential strategies to mitigate the risk of evolved weed resistance, which often results from the overreliance on single herbicide-resistant crops [2,12]. Diquat, a bipyridyl herbicide with structural similarity to paraquat, exerts its phytotoxic effects by accepting electrons from chloroplast photosystem I (PSI) under light conditions. The subsequent transfer of these electrons to molecular oxygen generates reactive oxygen species, inducing severe oxidative stress and leading to rapid plant tissue necrosis [13,14]. Compared to paraquat, diquat maintains comparable herbicidal efficacy but exhibits lower mammalian toxicity. In recent years, diquat has been widely used to replace paraquat in many countries [15–17]. Additionally, diquat can be absorbed by soil particles and undergoes degradation, resulting in reduced environmental persistence and enhanced ecological safety [13].

In our previous study, we identified a bipyridyl herbicide-resistance gene named *DTX6* in *Arabidopsis* [18]. This gene encodes a MATE (multidrug and toxic compound extrusion) family transporter. A glycine-to-glutamic acid substitution at position 311 (G311E) confers enhanced paraquat resistance in *Arabidopsis* [18,19]. *DTX6* comprises 12 transmembrane domains with dual localization to both vacuolar and plasma membranes. Though its *in vivo* substrate remains undetermined, it mediates bipyridyl herbicide detoxification through vacuolar sequestration and exocytotic extrusion [18]. While these findings established the molecular basis of resistance in dicots, whether this mutation confers cross-species resistance or induces pleiotropic effects remains unknown. In this study, we demonstrate that the G311E-mutated *DTX6* (designated *DTX6m*) confers robust resistance to bipyridyl herbicides in rice. Furthermore, *DTX6m* induces mild dwarfism and accelerates flowering time without compromising grain yield, while significantly increasing seed amino acid content. These results highlight its agricultural potential for developing herbicide-resistant crops through genetic breeding without sacrificing agronomic performance, and also provide mechanistic insights for investigating its physiological substrates.

2. Materials and Methods

2.1. Rice Variety and Growth Conditions

All rice materials used in this study are of *Oryza sativa* L. cv. Zhonghua 11 (ZH11) background. For green house assay, they were grown under a 12-h-light (30°C)/12-h-dark (22°C) photoperiod with 50% humidity unless otherwise specified. For field assay, rice seeds were germinated and grown in green house for three weeks, and then transplanted to the paddy field within the rice transgene experimental base located in Fudan university campus on June and harvested on October.

2.2. Generation of Transgenic Lines

The open reading frames of *DTX6-G311E* were cloned into the pCAMBIA1301-N-eYFP vector by Kpn I / BamH I sites. The construct was transformed into rice using the *Agrobacterium tumefaciens*-mediated transformation method [20], with the assistance provided by EDGENE Biotechnology Co., Ltd (Wuhan, China). The primers used in clone construction for overexpression are listed in Supplemental Table S1.

2.3. RNA Extraction and RT-qPCR

Total RNA was extracted using the TRIzol reagent (TaKaRa, Japan) and reverse-transcribed with PrimeScript RT reagent (TaKaRa, Japan). qPCR was performed with the SYBR Premix Ex Taq kit

(TaKaRa, Japan) using the CFX96 real-time PCR detection system (Bio-Rad, USA). Rice *UBQ5* gene was used as an internal control to normalize different samples. RT-qPCR primer sequences are given in Supplemental Table S1.

2.4. Herbicide Spraying Assay of Soil-Grown Seedlings

For paraquat resistance assay in greenhouse, the homozygous overexpression lines and ZH11 wild type were grown in soil for one month, and then sprayed with 0.5 g/L mM paraquat. 10 days after spraying, the survival rates and wilted leaves of different lines were documented and plants were photographed. For diquat resistance assay, plants were grown for 25 days and then sprayed with 0, 0.5, 1, 2, and 5 g/L diquat. 3 days post-application, the phenotypes were recorded and the survival rate was calculated. For the field test, the overexpression lines and ZH11 wild type were grown side-by-side for two months till the reproductive stage, then 2 g/L diquat was sprayed and pictures were taken after 10 days.

2.5. Agronomic Trait Analysis of Transgenic Rice Plants

Rice plants were grown in the paddy field within the rice transgene experimental base located in Fudan university campus from June to October. At least 12 plants of each line were randomly chosen in the field for further agronomic trait analysis, including plant height, flowering time, effective panicle number per plant, grain number per panicle, panicle length, one thousand-grain weight and grain size. One thousand-grain weight was measured using the harvested seeds that were air-dried for two weeks. Grain size including the length and width were analyzed by the automatic seed-examining high-speed document camera (Shenzhen Liangtian, China) and the thickness were measured manually by a vernier caliper.

2.6. Nutritional Component Determination of Rice Seeds

About 2 g dehulled rice seeds were ground into fine powder using a grinding mill and then used for the following nutritional component determination.

2.6.1. Free Fatty Acids Determination

Free fatty acids (FFAs) were extracted and quantified according to the established protocol with modifications [21,22]. Briefly, 0.1 g of rice powder was homogenized in 1 mL of chloroform-heptane-methanol (200:150:7, v/v/v) containing silicic acid (0.05 g/mL) to eliminate phospholipids. The mixture was agitated for 3 h at room temperature, followed by centrifugation at $8,000 \times g$, 4°C for 10 min. The supernatant was collected and reacted with freshly prepared Cu-TEA solution [0.05 M $\text{Cu}(\text{NO}_3)_2$, 0.1 M triethanolamine, NaCl-saturated (~ 33 g/100 mL), pH 8.1] to form FFA-Cu complexes. The FFA content was then determined using a commercial Free Fatty Acid Determination Kit (Suzhou Keming Co. Ltd., China).

2.6.2. Total Sugar Determination

Total sugar content was determined using the 3,5-dinitrosalicylic acid (DNS) method [23]. Briefly, 0.1 g of rice powder was homogenized in 1.5 mL of ddH₂O, followed by the addition of 1 mL of 6 N HCl. The mixture was incubated in a 95°C water bath for 30 min to hydrolyze polysaccharides. After cooling to room temperature, the sample was neutralized with ~ 1 mL of 6 N NaOH, diluted to a final volume of 10 mL with ddH₂O, and centrifuged at $8000 \times g$ for 10 min at 25°C . The supernatant was then analyzed for sugar content using a Total Sugar Determination Kit (Suzhou Keming Co., Ltd., China) according to the manufacturer's instructions.

2.6.3. Soluble Sugar Determination

Soluble sugar content was determined using the anthrone colorimetric method [24] with a Plant Soluble Sugar Determination Kit (Suzhou Keming Co. Ltd., China). The soluble sugar was extracted

from 0.1 g of rice powder with 1 mL of distilled water at 95°C for 10 min. After cooling, samples were centrifuged at 8000 × g for 10 min at 25°C. The supernatant was collected, diluted to 10 mL with ddH₂O, and 40 μL aliquots were used for soluble sugar quantification according to the manufacturer's protocol.

2.6.4. Starch Content Determination

Starch were extracted and determined as described previously [25] with some modifications: 0.1g rice powder was extracted with 1 mL of 80% ethanol at 80°C for 30 min. After centrifugation at 3,000 × g for 5 min, the supernatant was discarded and the pellet was air-dried. The air-dried residue was resuspended in 0.5 mL ddH₂O, heated in a boiling water bath for 15 min, and cooled. For starch extraction, 0.35 mL of 9.2 M perchloric acid was added and the mixture was incubated for 15 min at room temperature. The solution was then diluted with 0.85 mL ddH₂O and centrifuged at 3,000 × g for 10 min. The resulting supernatant was analyzed for starch content using Plant Starch Content Determination Kit (Suzhou Keming Co. Ltd., China) according to the manufacturer's instructions.

2.6.5. Protein Level Determination

0.02 g of rice powder was subjected to sequential extractions to obtain the four major rice storage proteins (albumin, globulin, prolamin, and glutelin) according to the method described previously [26]. The contents of different protein ingredients were determined using the Modified Bradford Protein Assay Kit (Sangon Biotech, China).

2.6.6. Free Amino Acid Level Determination

0.1 g rice powder was extracted with 1 mL of distilled water in a boiling water bath for 15 min. After cooled to room temperature, samples were centrifuged at 10,000×g for 10 min. The supernatant was used for total amino acid determination by ninhydrin method [27].

2.7. Phylogenetic Analysis

The amino acid sequences of Arabidopsis and rice MATE family proteins were extracted from the TAIR (www.arabidopsis.org) and RAP-DB (<https://rapdb.dna.affrc.go.jp/>) websites. 56 *Arabidopsis* MATE family proteins and 46 rice MATE proteins (for genes with multiple transcripts, only the first transcript was chosen) were analyzed for their phylogenetic relationship using *selaginella_moellendorffii* as the alien species. Multiple sequence alignment was performed by Clustal X using default parameters. The resulting alignment was output in Phylip format and uploaded to the online PhyML (<http://www.atgc-montpellier.fr/>) website to construct the phylogenetic tree using the maximal likelihood method by standard bootstrap analysis with 1000 replicates [28]. The resulting phylogenetic tree was visualized using the iTOL tool (<https://itol.embl.de>).

3. Results

3.1. Generation of *DTX6m*-Expressing Rice Lines

The G311E-mutated *DTX6* gene allele, *DTX6m*, was cloned into the pCAMBIA-N-eYFP vector under the promoter of CaMV 35S and transformed into ZH11 (Figure 1A). Screening of T₀ transgenic plants identified 20 independent lines harboring the *DTX6m* transgene, which were designated OE1 to OE20. Quantitative RT-PCR analysis revealed substantial variation in *DTX6m* expression level across these lines (Figure 1B), with the majority exhibiting significant transgene expression. To assess the correlation between expression levels and herbicide resistance, three representative lines: OE-3 (highest level), OE-5 (intermediate level), and OE-13 (relatively low level), were chosen for phenotypic characterization.

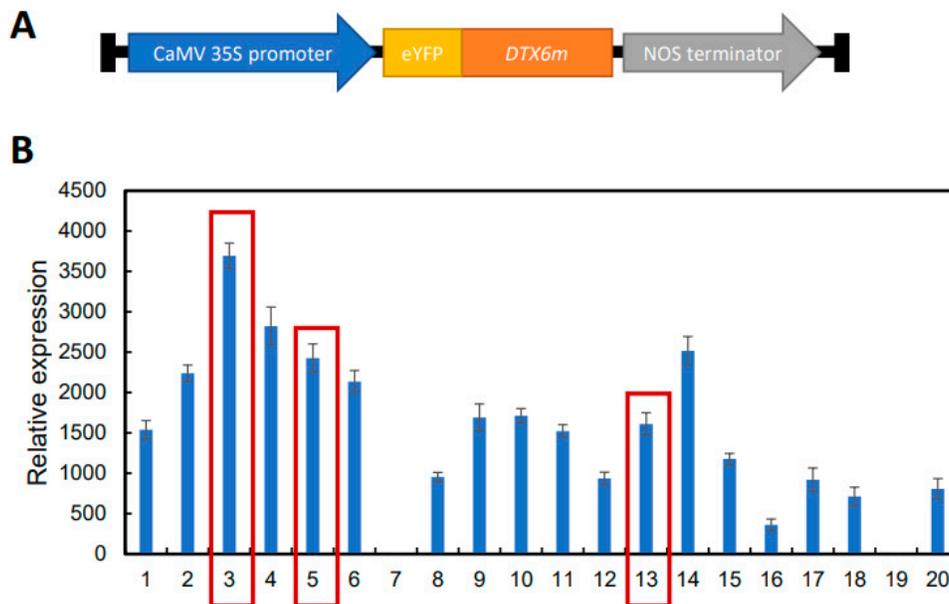


Figure 1. Generation of DTX6m-overexpression lines. (A) Schematic diagram of the 35S::eYFP-DTX6m vector; (B) Identification of the DTX6m-overexpression lines by RT-qPCR. RNA were extracted from leaves of two-week old transgenic seedlings and then used for the examination. The expression level of DTX6m relative to UBQ5 was shown. Data represent means \pm SD of three technical replicates. Scale bar=10 cm.

3.2. DTX6m-Overexpression Lines Exhibit Strong Bipyrindyl Herbicide Resistance

To evaluate resistance to bipyrindyl herbicides, we utilized the T3 generation of the representative lines whose homozygosity were confirmed by the absence of segregation in hygromycin resistance. To preliminarily observe the herbicide resistance on culture media, OE3, 5, 13 and the ZH11 control were grown on 1/2 MS media supplemented with 0.3 μ M paraquat for 8 days. As shown in Figure S1, all overexpression lines had strikingly higher seedling heights and longer primary roots compared to the wild type control, which were severely stunted by the herbicide in the media. To compare their resistance to foliar-applied herbicide, seedlings grown in soil for 25 days were foliar-sprayed with 0.5 g/L paraquat. Ten days after application, leaves of the ZH11 wild type exhibited severe necrosis, whereas all three overexpression lines maintained green foliage with minimal phytotoxicity symptoms (Figure 2A). Quantitative analysis confirmed significantly higher survival rates and more retention of functional green leaves in transgenic lines relative to the wild type (Figure 2B). These findings conclusively demonstrate that constitutive expression of DTX6m confers substantial paraquat tolerance in rice.

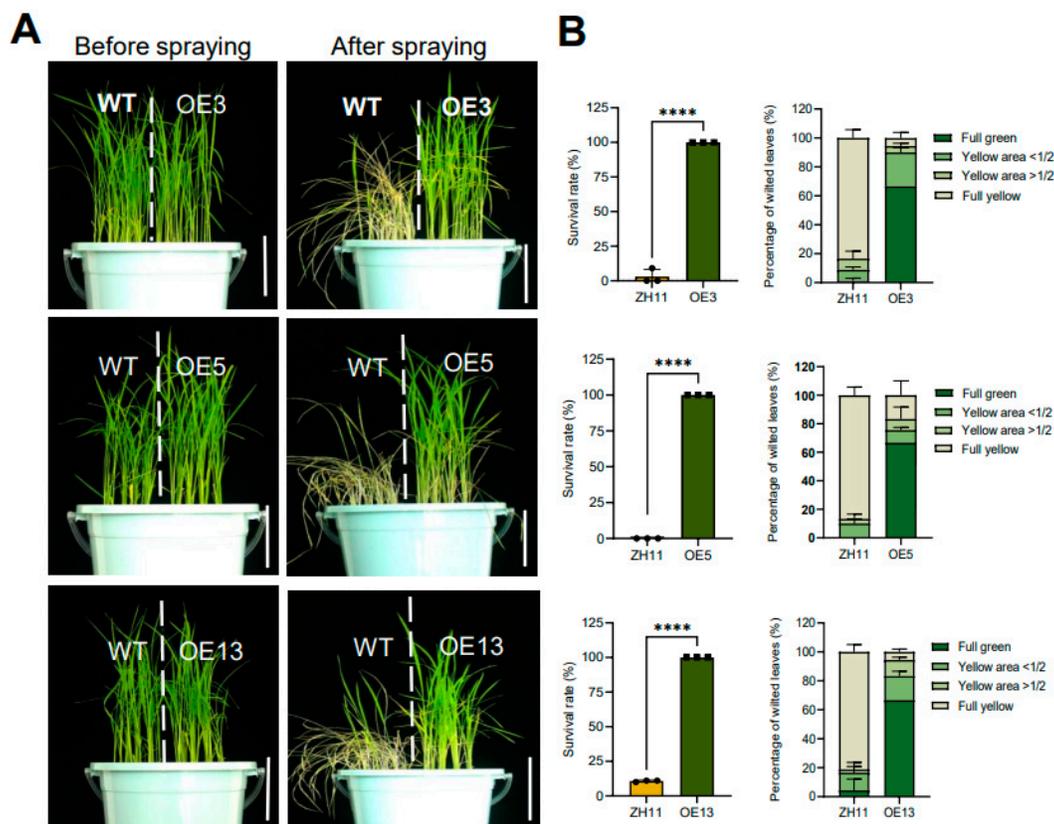


Figure 2. Paraquat resistance phenotypes of *DTX6m*-overexpression lines. (A) Growth conditions of plants before and after spraying with paraquat. ZH11 and three *DTX6m*-overexpression lines were grown in soil for one month, and then sprayed with 0.5 g/L paraquat. Plants were photographed 10 days after spraying. (B) The survival rate and wilted leaves were documented. Data represent means \pm SD of three biological replicates. Each replicate includes 10 rice plants. One-way ANOVA was performed to analyze the significance of difference between ZH11 and *DTXm*-overexpression lines. **** $P \leq 0.0001$.

Given the prevalent agricultural usage of diquat as an alternative bipyridyl herbicide to paraquat in many countries, we evaluated diquat resistance in rice plants through multi-stage physiological assessments. Initial screening at the seedling stage employed 25-day-old soil-grown plants subjected to foliar application of diquat at escalating concentrations (0, 0.5, 1, 2, and 5 g/L). Post-treatment evaluation after three days revealed a concentration-dependent response: ZH11 plants exhibited severe chlorosis at all tested concentrations, whereas transgenic lines maintained significant chlorophyll integrity even at 5 g/L (5 \times the commercial field application rate) (Figure 3A). Quantitative analysis demonstrated differential tolerance among transgenic lines, with OE3 and OE5 showing significantly stronger resistance compared to OE13 (Figure 3B), correlating with the higher expression level of *DTX6m* in OE3 and OE5 (Figure 1). Field validation experiments involved diquat application (2 g/L) at the reproductive stage (2 months post-transplant in the paddy field). 10 days after treatment, ZH11 plants displayed complete systemic necrosis and mortality, while all transgenic lines survived without chlorosis (Figure 4). This translational evidence confirms that *DTX6m* overexpression confers robust cross-environmental tolerance to diquat.

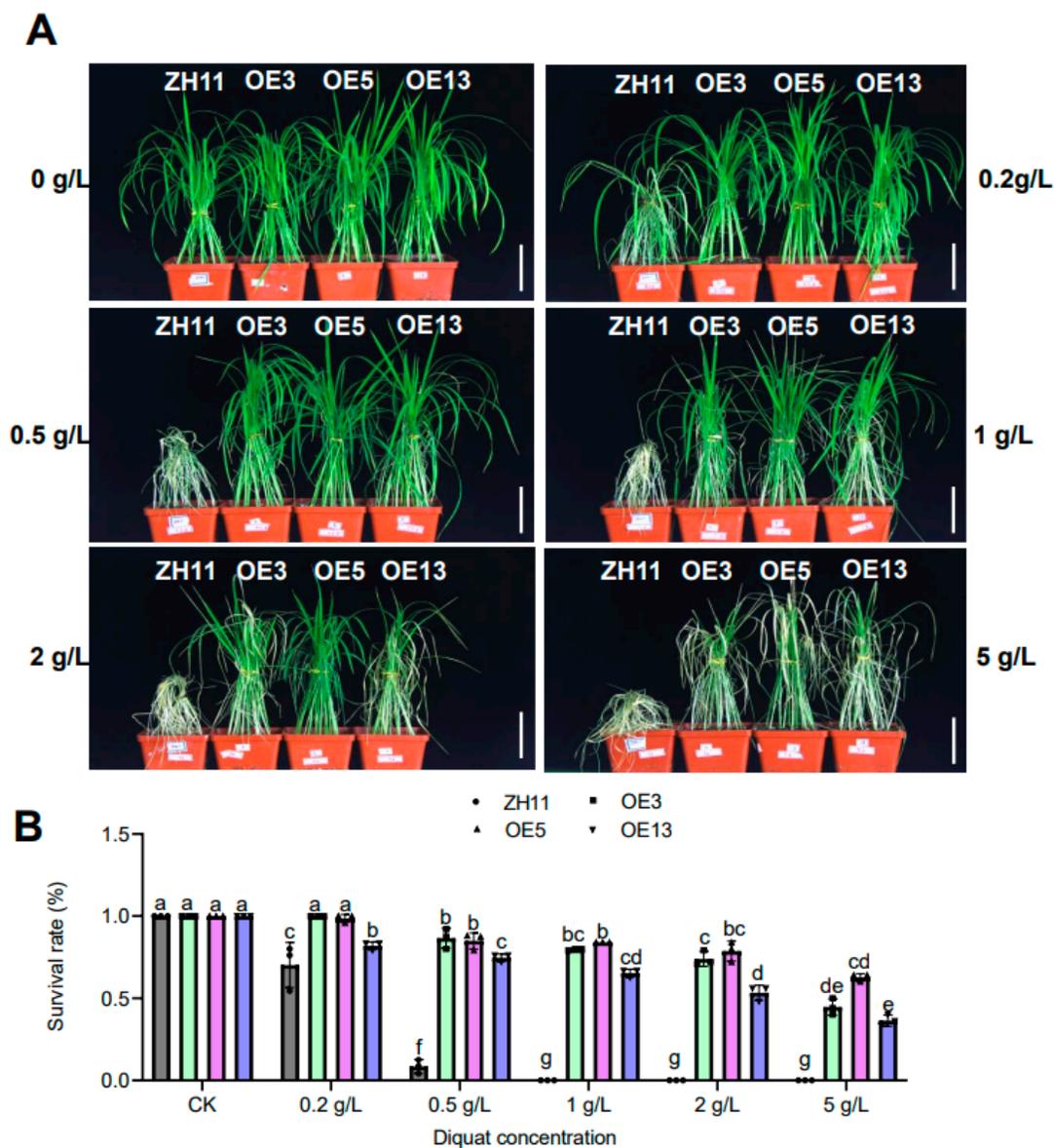


Figure 3. Evaluation of the resistance of the *DTX6m*-transgenic lines to different concentrations of diquat. (A) Diquat resistance phenotypes of ZH11 and *DTX6m*-transgenic lines. ZH11 and three *DTX6m*-overexpression lines were grown in soil for 25 days, and then sprayed with diquat at different concentrations (0, 0.5, 1, 2, and 5 g/L). Plants were photographed 3 days after herbicide application. (B) The survival rates of ZH11 and *DTX6m*-transgenic lines. Data represent means \pm SD of three biological replicates. Each replicate includes 20 rice plants. Two-way ANOVA was performed to analyze the significance of difference, with Fisher's LSD test. Scale bar=5 cm.



Figure 4. Diquat resistance phenotypes of three *DTX6m*-overexpression lines in the field. Rice plants were grown in the field for two months and then sprayed with 2 g/L diquat. Pictures were taken 10 days after spraying. Scale bar=10 cm.

3.3. *DTX6m*-Overexpression Lines Had Early Flowering and Semi-Dwarf Growth Phenotypes in the Field

To systematically characterize the possible pleiotropic effects of *DTX6m*-overexpression in rice, we conducted side-by-side field trials of transgenic lines and ZH11 wild-type controls. While vegetative growth showed no significant differences between the groups, pronounced phenotypic variations emerged during maturation. The *DTX6m*-overexpressing lines displayed reduced plant height and an approximately 3-4 days earlier flowering time, accompanied by earlier leaf senescence (Figure 5A–C). Analysis of primary tillers revealed that transgenic lines produced significantly shorter tillers with precocious maturation relative to ZH11 (Figure 5D). These findings demonstrate that *DTX6m* also functions as a developmental regulator in rice, in addition to the herbicide-detoxifying capacity.

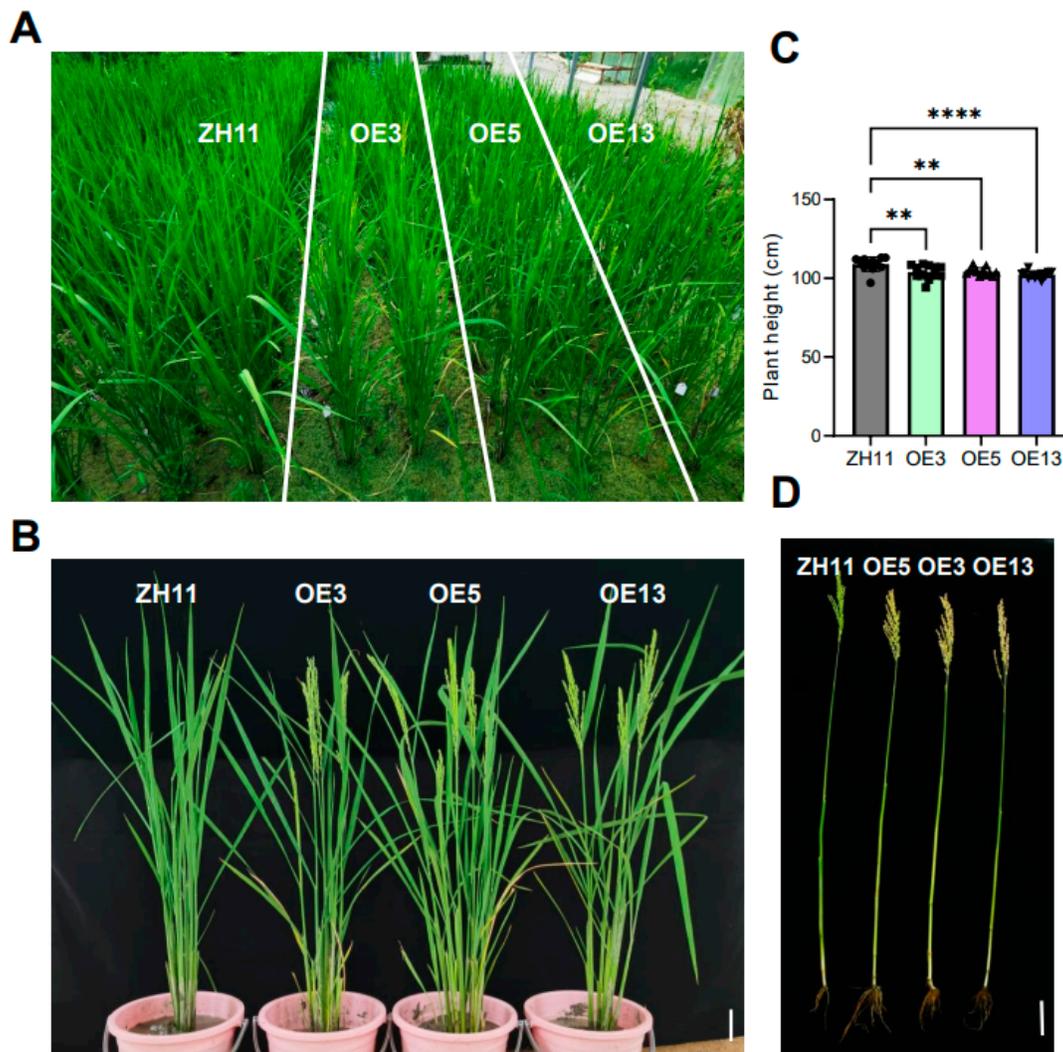


Figure 5. Phenotypes of the transgenic lines in the field. (A) Flowering stage rice plants in the field. (B) Phenotype comparison of ZH11 and three *DTX6m*-overexpression lines. (C) Comparison of the plant height in the field. Data represent means \pm SD (n=12 plants). One-way ANOVA was performed to analyze the significance of difference between ZH11 and different *DTXm*-overexpression lines. **** $P \leq 0.0001$. ** $P \leq 0.01$. (D) Phenotype of the main tiller at day 110 after breeding. Scale bar=10 cm.

3.4. Analysis of the Field Agronomic Traits of *DTX6m*-Transgenic Lines

Yield traits are essential considerations in the application of genetically modified crops. We therefore evaluated the impact of *DTX6m* overexpression on agronomic traits associated with grain yield, including grain size, 1000-grain weight, number of effective panicles per plant, panicle length, and grain number per panicle. As shown in Figure 6A–F, OE3, OE5, and OE13 lines displayed three distinct morphological characteristics: (1) increased grain length, (2) comparable grain width, and (3) a slight reduction in grain thickness compared to ZH11. Statistical analysis revealed no significant differences in 1000-grain weight between *DTX6m*-OE lines and ZH11 (Figure 6G). Evaluation of panicle architecture showed consistent patterns in both main panicle length and grain number per panicle across all lines (Figure 6H,I). However, phenotypic divergence emerged in effective panicle numbers, with OE3 and OE13 showing significant reductions relative to ZH11, while OE5 maintained wild-type levels (Figure 6J). This differential performance suggests that field-effective panicle

formation exhibits environmental sensitivity, with the OE5 line demonstrating better field adaptability, likely attributable to the preserved tillering capacity compared to other transgenic lines.

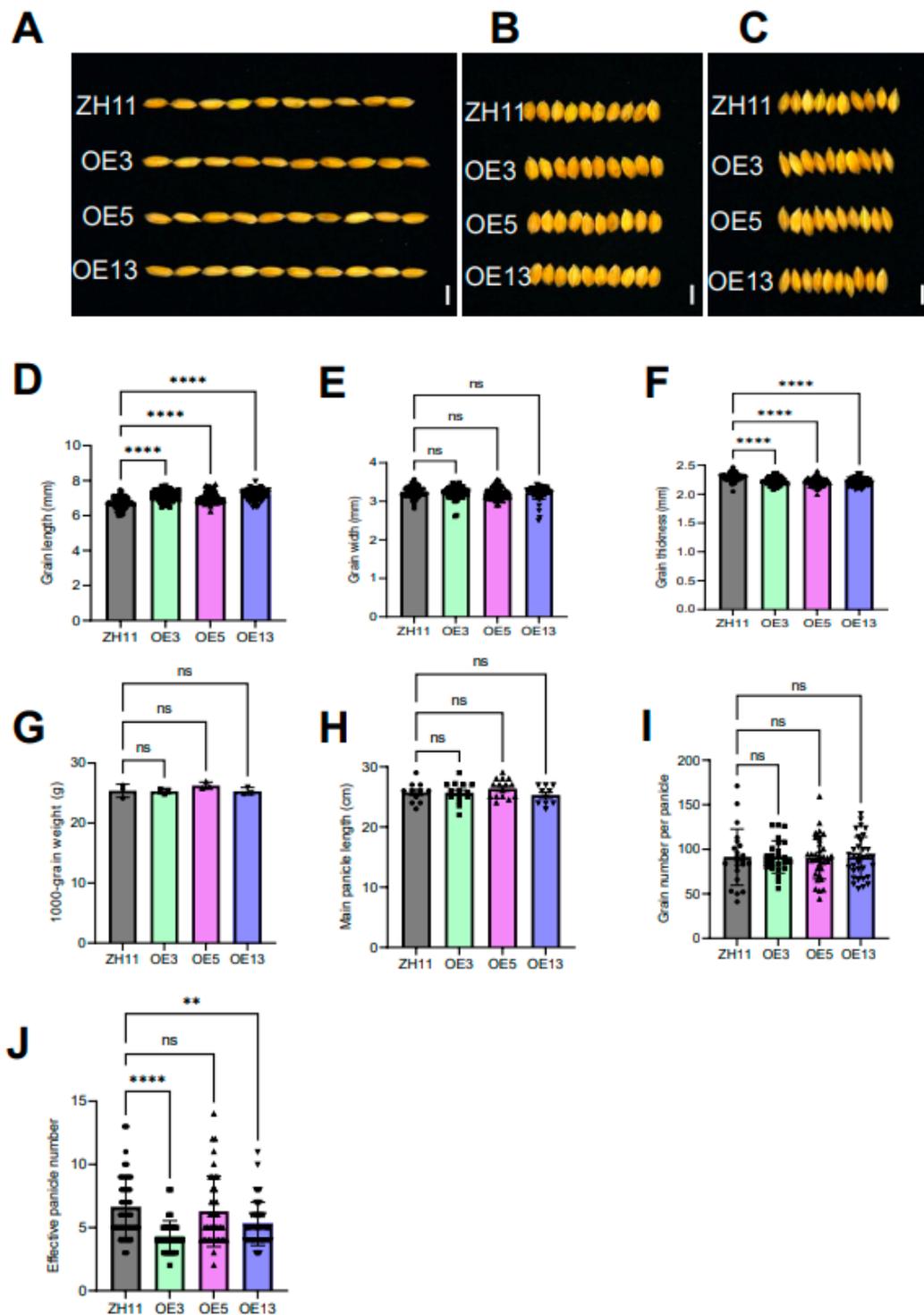


Figure 6. Comparative analyses of the grain size and yield traits of ZH11 and *DTX6m*-overexpression lines. (A) Grain length. (B) Grain width. (C) Grain thickness. (D) Statistical analysis of the grain length. (E) Statistical analysis of the grain width. (F) Statistical analysis of the grain thickness. In (D-F) Data represent means \pm SD ($n=65$ seeds from different plants). (G) Statistical analysis of 1000-grain weight ($n=3$). (H) Statistical analysis of the main panicle length ($n\geq 10$). (I) Statistical analysis of the

grain number per panicle ($n \geq 22$). One-way ANOVA was used to analyze the significance of difference between ZH11 and different *DTX6m*-overexpression lines. **** $P \leq 0.0001$. ** $P \leq 0.01$. In A-C, Scale bar=5 mm.

3.5. The Seed Storage Reserve Accumulation Is Not Consistently Affected in the Transgenic Lines Except the Free Amino Acid Level

Rice seeds, as a staple food, serve as a source of carbohydrates, proteins, and lipids for human consumption. To determine whether the overexpression of *DTX6m* altered the nutritional quality of rice seeds, we analyzed the levels of starch, total sugar, soluble sugars, free lipidic acids, free amino acids and proteins in transgenic rice lines. The results revealed that the transformation of *DTX6m* did not change the starch content (Figure 7A), but did elevate the free amino acid level in all three transgenic lines (Figure 7B). As regards the total sugar, soluble sugar and free lipidic acid levels, there were no consistent changes in three transgenic lines (Figure 7C–E), suggesting that their content might be affected by factors other than *DTX6m*. The protein content such as albumin, globulin, prolamin and glutelin levels remained unchanged in the transgenic lines (Figure 7F). Among the three *DTX6m*-overexpression lines, OE5 performed exceptionally well (Figure 7A–E), exhibiting significantly higher levels of free fatty acids, free amino acids and soluble sugar compared to ZH11.

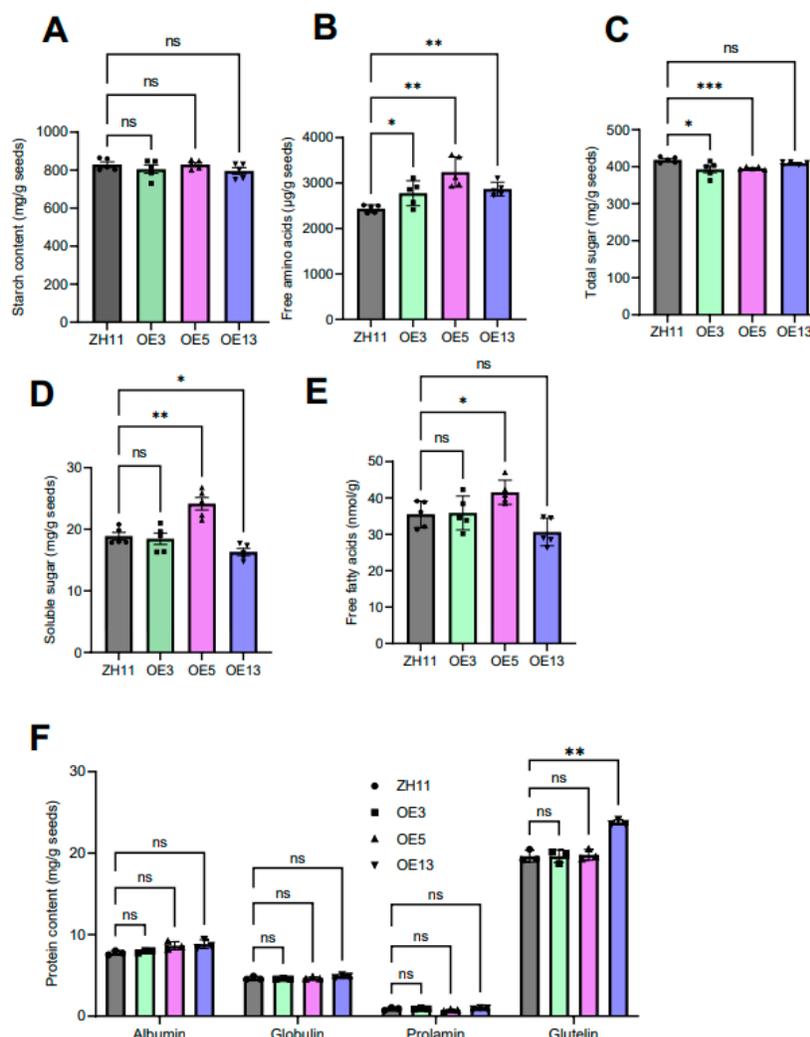


Figure 7. A comparative analysis of the nutritional components of seeds from ZH11 and *DTX6m*-overexpression lines. (A) Free fatty acid level comparison. (B) Free amino acid level comparison. (C) Total sugar comparison.

(D) Soluble sugar comparison. (E) Starch content comparison. (F) Albumin, Globulin, Prolamine and Glutelin level comparison. Each determination was performed with 5 replicates. One-way ANOVA was used to analyze the significance of difference between ZH11 and *DTX6m*-overexpression lines. *** $P \leq 0.001$. ** $P \leq 0.01$. * $P \leq 0.1$.

4. Discussion

4.1. Evolutionary Analysis of *Arabidopsis* and rice MATE Family Genes

MATE family proteins, belonging to a large superfamily of evolutionarily conserved membrane transporters, are widely distributed across diverse organisms ranging from bacteria to plants and animals [29]. In plants, these proteins exhibit distinct subcellular localizations, including the plasma membrane [30,31], tonoplast [32–35], Golgi apparatus [36], and late endosome/prevacuolar compartment [37]. They mediate a wide spectrum of physiological processes such as heavy metal and herbicide detoxification [18,19,30]; aluminum tolerance [38,39]; iron homeostasis regulation [31,36,40]; vacuolar sequestration of secondary metabolites [32,33,35,41]; hormone delivery [42–45]; leaf senescence and turgor modulation [34,37]; and pathogen resistance [46–48]. Notably, the substrate specificity of MATE family members shows remarkable diversity, necessitating individual characterization for each transporter.

The MATE transporter family comprises 56 members in *Arabidopsis thaliana* [49] and 46 members in rice (*Oryza sativa*) [50]. Phylogenetic analysis of MATE proteins from both species revealed that the rice homologs LOC_Os01g31980, LOC_Os01g49120 and LOC_Os05g48040 show closest evolutionary proximity to *Arabidopsis* DTX6 (Figure S2). Although clustered within the same phylogenetic subgroup, these proteins occupy distinct branches with relatively low sequence identities to DTX6 (55.8%, 53.9% and 50.7%, respectively), highlighting substantial divergence of MATE transporters between monocots and dicots. To functionally explore their relationship, we employed single-base editing technique [51] to introduce a G311E substitution (equivalent to DTX6's critical glycine residue) in LOC_Os01g31980 (Figure S3). Unexpectedly, the edited rice plants exhibited no altered resistance phenotypes to paraquat or diquat herbicides, suggesting that LOC_Os01g31980 may not serve as the functional ortholog of DTX6 in rice.

4.2. *DTX6m* Largely Increased the Bipyrindyl Herbicides Resistance of Rice

Weed infestation poses a significant threat to global rice production, with yield losses exceeding 40% in severe cases [8]. In large-scale rice cultivation, the integration of herbicide-resistant rice cultivars with compatible herbicides has emerged as an effective weed control strategy. Recent advances in understanding plant-herbicide interactions have driven the development of herbicide-tolerant rice through three principal strategies including traditional mutagenesis and phenotypic screening, precise editing of endogenous resistance genes and transgenic introduction of exogenous resistance determinants [2,37]. In previous studies, we found that DTX6 mediates paraquat and diquat detoxification through exocytosis and vacuolar sequestration mechanisms. Notably, when glycine at position 311 in *Arabidopsis* DTX6 is substituted with acidic amino acid (G311E or G311D), the protein exhibits enhanced detoxification capacity [18]. In this study, we investigated the potential application of the *DTX6m* gene and its side effects in monocot crop rice. Three independent *DTX6m*-overexpression lines (OE3, OE5 and OE13) demonstrated remarkable bipyrindyl herbicide resistance compared to the ZH11 wild-type. Greenhouse trials with rice seedlings showed that while ZH11 plants completely wilted in 3 days after 1 g/L diquat treatment, all three transgenic lines maintained >80% survival rates. Even at 5 g/L diquat (5× concentration), a significant number of the transgenic plants survived, with OE5 showing the strongest resistance (over 60%). Field trials conducted on two-month-old plants revealed complete mortality of ZH11 within 10 days after 2 g/L diquat application, whereas all transgenic lines survived, demonstrating stable, long-term diquat resistance that appears more effective in mature plants. These results suggest that *DTX6m* functions similarly in rice and *Arabidopsis*, likely through the same mechanisms.

4.3. The Transgenic Rice Has Slightly Early Flowering and Dwarf phenotype Without Compromised Yield and Nutritional Value

The potential pleiotropic effects of *DTX6m* transgenesis on plant growth, development, yield, and nutrient assimilation were systematically investigated. In field trials, all three transgenic lines exhibited two phenotypes with significant agricultural value: slightly early flowering and semi-dwarfism. Early flowering shortens the growth cycle, optimizing land use efficiency. Meanwhile, the semi-dwarf trait enhances lodging resistance, which is particularly advantageous in typhoon-prone regions.

Before implementing *DTX6m* in genetic manipulation, it is crucial to determine its impact on crop yield and grain quality. Therefore, we evaluated key rice yield components, including the number of panicles per plant, the number of grains per panicle, and 1000-grain weight. The results showed that the OE3 and OE13 transgenic lines produced fewer productive panicles than the wild-type ZH11, while the OE5 line did not exhibit such a reduction. In all three transgenic lines, the number of grains per panicle and 1000-grain weight remained unchanged. However, compared to ZH11, all *DTX6m*-overexpression lines had altered seed morphology, with increased grain length, unchanged grain width, and reduced grain thickness. Additionally, no significant differences were detected in starch or total protein content between the transgenic and wild-type plants. Overall, among the transgenic lines, OE5 exhibited the most superior performance, demonstrating robust resistance to bipyridyl herbicides while maintaining rice yield and nutritional quality. These findings highlight the critical need for screening large transgenic populations to identify optimally performing lines for agricultural implementation. The significantly elevated accumulation of free amino acids in all three independent *DTX6m*-OE lines (Figure 7B) implies a potential role for DTX6 in amino acid homeostasis. However, the precise function of DTX6 in *Arabidopsis* still remains elusive at present. Future research will focus on identifying the endogenous substrates of DTX6 to clarify its physiological role.

5. Conclusions

Our results reveal that the *DTX6m* gene effectively confers resistance to bipyridyl herbicides in rice without compromising yield potential. Moreover, its overexpression leads to early flowering, semi-dwarfism, and increased free amino acid levels in rice grains. The phenotypes induced by *DTX6m* in rice suggest generalizability to other monocot cereal crops. Collectively, our findings in rice and previous results in *Arabidopsis* indicate that *DTX6m* holds broad utility for improving herbicide resistance in both monocot and dicot crops.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

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