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Article

Zebrafish (*Danio rerio*) Prefer Undisturbed Shoals over Shoals Exposed to the Synthetic Alarm Substance Hypoxanthine-3N-Oxide (C₅H₄N₄O₂).

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Simple Summary: This study investigates how zebrafish, a small freshwater fish known for social behavior, respond to visual indicators of potential threats in their environment. Animals often form groups to reduce the risk of predation, benefiting from safety in numbers. Zebrafish are highly social and can communicate threats through chemical signals; however, these signals are limited to short distances and can break down quickly in water. This experiment explored whether zebrafish can use visual cues alone to distinguish between groups of fish which are alarmed (indicating a potential threat) and fish which are not. Using a 3-tank experimental setup, researchers found that zebrafish preferred to stay near non-alarmed groups, spending more time and exhibiting calmer behavior in these zones. Males showed more freezing behavior than females, indicating differences in how each sex responds to threats. These findings highlight zebrafish's ability to visually assess danger and choose safer social environments. This research has broader implications for understanding group behavior and decision-making under threat, offering insights into how animals, including humans, respond to social and environmental cues. By studying zebrafish, scientists can uncover mechanisms that may help address social- and anxiety-related disorders in humans.

Abstract: As an anti-predation behavior, shoaling enhances survival among prey species by reducing individual predation risk through mechanisms like the dilution effect and collective vigilance. Zebrafish – a highly social and genetically tractable species – are valuable for studying these behaviors. The present study examined zebrafish social preferences in a 3-chamber open-tank free-swim task, assessing whether visual cues alone could distinguish between an intact and an alarmed shoal exposed to the synthetic alarm substance H3NO. Subjects were allowed to freely associate with either shoal while their behavior was recorded and analyzed. Results revealed a significant preference for proximity to the intact shoal, indicating zebrafish's ability to visually discern threat levels. Subjects spent nearly twice as much time in the zone near the intact shoal, with reduced freezing and faster movement velocities compared to the alarmed shoal zone. Males exhibited more freezing behavior than females, consistent with sex-specific strategies in threat response. These findings underscore zebrafish's reliance on visual cues for social responding under predatory threat and highlight sex-based differences in threat perception. This research expands the understanding of zebrafish social dynamics and provides a robust framework for future exploration of the neural mechanisms underlying social behavior and threat assessment in zebrafish.

Keywords: alarm substance; predation threat; 3-chamber open-tank free-swim task; shoaling; social preference

1. Introduction

Anti-predation adaptations evolved through natural selection to reduce risk among members of prey species and increase survival/fitness. Many animals exhibit behavioral strategies – such as playing dead or evading capture – to reduce predation risk [1–3]. Another common behavioral adaptation is social collection; prey often form complex social groups which provide multiple advantages in reducing predation risk [4]. First, while predators can locate groups of prey more readily than solitary prey and can proficiently identify which prey in a group are easier targets [5–8], group living decreases the likelihood of any one individual being successfully captured due to the dilution effect [9,10], the “many eyes” effect [11,12], and the confusion effect [13,14]. Additionally, a group of vigilant individuals is more likely to detect a potential predatory threat than a solitary individual [15,16]. Once detected, threats can be communicated to other group members in a number of different ways, including alarm movements e.g. [17,18] and chemical messaging e.g. [19,20]. While chemosignaling of predatory threat can be highly specific [21,22], this signaling process requires the receiver’s proximity to the signaling animal. Chemical signals can break down quickly in aquatic environments [23–25], and the dynamics of concentration changes as chemical signals disperse can be quite complex [26]. It would also be adaptive for an aquatic species such as zebrafish to communicate a potential predatory threat at greater distances through visual display rather than relying solely on short-range chemosignaling. The goal of the present study is to determine whether zebrafish can use visual-only cues to detect the communication of simulated predatory threat, and if so, what social preference zebrafish will form in response to the visual detection of the communicated threat.

Zebrafish (*Danio rerio*) are highly social freshwater fish widely recognized as a valuable model for biomedical and behavioral neuroscience research [27,28]. This species is particularly advantageous as a model organism due to its genetic similarity to humans [29], standardization of husbandry e.g. [30], rapid reproduction [31], and distinctly observable social behaviors [32]. Zebrafish exhibit a broad range of complex social interactions, many of which parallel human social behaviors, making zebrafish a powerful tool for studying neuropsychiatric disorders such as anxiety e.g. [33], depression e.g. [34], and social impairments e.g. [35]. Their use in research enables scientists to investigate behavior and responses to social cues in a controlled setting without the ethical and practical challenges involving human subjects e.g., [36]. A key component of zebrafish social behavior is shoaling – the formation of loosely coordinated groups composed of a few to several hundred individuals [15,37]. Shoaling provides important survival benefits such as increased mating opportunities, improved foraging efficiency, and added protection from predators through the safety-in-numbers effect [38–40]. These social behaviors, combined with their genetic tractability, make zebrafish an ideal model for understanding the neural and behavioral mechanisms underlying social dynamics, offering valuable insights into human health and disease.

Schreckstoff, which translates to “scary substance” in German, is a chemical message released by an injured fish that serves as a warning to nearby conspecifics of a potential predatory threat [41,42]. When epidermal injury results from a predatory attack, chemical compounds are released from specialized epidermal club cells [19,43,44], then detected by the olfactory receptors of surrounding conspecifics [42]. Olfactory sensory neurons transmit excitatory signals to the olfactory bulb, triggering specific anxiety-like behaviors such as freezing, darting, and jumping [22,45–47]. Previous literature indicates that the synthetic chemical hypoxanthine-3N-oxide (H3NO) elicits alarm responses in zebrafish in a manner similar to schreckstoff [48]. Additionally, more recent research suggests that specific combinations of epidermal secretions (e.g. a fear signal plus a conspecific signal) are responsible for eliciting fear responses within a shoal [21,22]. When isolated from their home shoal and seeking new shoaling opportunities, solitary fish may avoid shoals that have experienced recent predatory attack, instead preferring to join shoals that are undisturbed [48,49].

Previous research indicates that multi-chamber tasks and multidimensional analyses enhance our understanding of zebrafish behavior [50–53]. A recent study used an adjacent-tank apparatus [54] to examine zebrafish responses to naturally-derived alarm signals [52] in order to study social

contagion – a phenomena in which undisturbed subjects respond to conspecifics exposed to an alarm signal. In this study, both undisturbed male and female subjects demonstrated freezing responses when exposed to the sight of alarmed conspecifics, with males demonstrating more freezing than females. Subjects of both sexes also demonstrated other responses to the sight of alarmed conspecifics. While the adjacent tank configuration is useful for testing detection of visual-only cues between subjects and stimulus shoals, it is limited in the extent to which social preference can be studied as only one social stimulus is presented to the solitary subject. More recently, the 3-chamber open-tank free-swim task (OTFST) emerged as a validated method for testing social preference in zebrafish as it increases the amount of potential data which can be obtained and allows dichotomous preference analyses [55]. Subsequent research examined zebrafish social preference between two different shoals of fish using the 3-chamber OTFST, demonstrating that zebrafish spend more time near real shoals than artificial shoals [56] and prefer established shoals over newly-formed shoals [57]. These findings indicate that zebrafish can effectively detect subtle differences in neighboring shoals through visual cues during the 3-chamber OTFST. To further explore and develop the utility of this technique, the present study expands upon earlier two-tank approaches in assessing the shoaling preference of solitary subjects. Unlike chemosignaling of predatory threats, there is limited research on zebrafish's ability to detect visual-movement signals associated with predatory threats, particularly when subjects rely solely upon visual signals in the context of the 3-chamber OTFST. The present experiment tests the hypothesis that zebrafish can detect differences between alarmed and unalarmed shoals and will spend more time near the intact (i.e. unalarmed) versus the alarmed shoal using the 3-chamber OTFST.

2. Materials and Methods

2.1. Animals and Housing

The experimental subjects (N = 20, 10 males & 10 females) as well as all stimulus fish were adult (> 6 mos age) short-fin wild-type (SFWT) zebrafish obtained from a regional supplier (Quinn's Fins, Palm Bay, FL, USA). Several previous studies have established the efficacy of using SFWT zebrafish [58–60] as SFWT fish are expected, “to be more representative of the species and to possess fewer idiosyncratic features that may have developed during the inbreeding process of laboratory strains” [48] (p. 338); as such, this particular population is an excellent choice for behavioral studies. Upon arrival at the facility, fish were separated into same-sex group-housing tanks (38 L capacity, 50.8 cm X 25.4 cm X 30.5 cm) equipped with active bio-filters (“Penguin 100”, Model PF00100B, MarineLand - Spectrum Brands Pet, LLC, Blacksburg, VA, USA) and conditioned water (balanced pH of 7.0 - 8.0, 400 - 700 μ S, temperature 25 - 27 °C, < 40 ppm nitrates, < 0.2 ppm nitrites, & 0.01 - 0.1 ammonia) with an average density of 1.5 fish per gallon (3.79 L). Fish were held in same-sex group-housing tanks until selection as either individual subjects or members of stimulus shoals. All fish were kept under a 14L:10D photoperiod and fed once daily (“TetraPro Tropical Crisps” - Product #77070, Tetra - Spectrum Brands Pet, LLC, Blacksburg, VA, USA). Animals were kept under these conditions for 1 to 6 weeks before being transferred to smaller staging tanks prior to experimentation.

Subjects and stimulus fish were selected on a weekly basis for the experiment and transferred to staging tanks on a stand-alone recirculating-flow rack system (#ZS660, Aquaneering, Inc., San Diego, CA, USA) maintained within the same parameters as the group-housing tanks. A high-density polyethylene tank (90 L capacity, NorthStar #2691, Northern Tool & Equipment, Burnsville, MN, USA) was custom fitted with a flow-through manifold on the rear of the rack system to supply water for filling testing tanks. Subjects were housed individually in 1.4L tanks (#ZT080, Aquaneering, Inc., San Diego, CA, USA) on one row of the rack system with opaque dividers placed between the tanks. Other fish were selected to form small stimulus shoals of four fish (2 males & 2 females); shoals were housed in 2.8L tanks (#ZT280, Aquaneering, Inc., San Diego, CA, USA) on different rows of the rack system. Subjects and stimulus fish were kept under these conditions for three to seven days before testing. Following transfer of subjects and stimulus fish to the testing apparatus, all 1.4L and 2.8L

tanks were washed and sterilized in a standard laboratory washer before being returned to the rack for subsequent re-use. s. All housing, caretaking, and other procedures involving the animals were performed under appropriate animal welfare guidelines [61].

2.2. H₃NO Solution Preparation

100 mg of H₃NO (152.11 g/mol) was synthesized to 99% purity by a scientific laboratory (Synchem UG, Altenburg, Germany). A 50 μ M stock solution was prepared in a 1L volumetric flask by adding 7.6055 mg H₃NO to reagent-grade RO water (> 18 M Ω) and stored at 1°C under dark conditions. A 5 μ M working solution was prepared by adding 100 mL of stock solution to 900 mL reagent-grade RO water and also stored at 1°C under dark conditions. Weekly, 5 mL aliquots were taken from the working solution and stored at -20°C under dark conditions until needed for experimental sessions.

2.3. Apparatus and Software

Previous studies have successfully demonstrated the efficacy of the 3-chamber OTFST and include detailed descriptions of the apparatus [56,57,62]. The 3-chamber OTSFT apparatus in the present study consisted of a central testing tank ("Rimless 5.5 Gallon" - #100541424, Aqueon, Inc., Franklin, WI, USA - dimensions 41.275 cm L x 21.273 cm W x 26.67 cm H; max capacity 23.4 L) flanked by two stimulus tanks identical to the central tank. Linear polarizing filters (dimensions, Rosco #7300, obtained from B&H Foto & Electronics, Inc., New York, NY USA) were placed in a 90° orientation to each other on the outer surfaces of the central tank to ensure the fish of one stimulus shoal tank could not see the members of the stimulus shoal in the opposite flanking tank while still allowing the test subject in the central tank a view of both stimulus shoals. The central tank provided a free-swim environment for observing the behavioral responses of test subjects to the shoaling stimuli contained in either flanking tank. Each tank was fitted with a custom clear polycarbonate lid (21 cm x 41 cm), and the lid for the rightmost tank had a 5 mm diameter hole drilled in the center to allow for insertion of a syringe-like Positive-Displacement Dispenser Tip (PDDT; 10 ml "Combitip" #0030089464, Eppendorf Corp., Hamburg, Germany).

Uniform backlighting for the 3-chamber OTFST apparatus was provided by an LED dimmable flat-panel light (model - CPANL 1X4 40LM SWW7 120 TD DCMK, Lithonia Lighting - Acuity Brands, Inc., Conyers, GA, USA) held in a custom PVC frame and controlled with a dimmer switch (model CTCL-153P-WH, Lutron Electronics, Inc., Coopersburg, PA, USA) to maintain 350 Lux at surface of the light. The rear walls of the testing chamber and flanking tanks were covered with a self-adhesive white plastic film (#16F-C9A952-06, Con-Tact Brand, La Mirada, CA, USA) which further diffused the backlighting, reducing the brightness to 15 lux at the front of each tank. One digital video camera (model - acA 1300-60gc, Basler AG, Ahrensburg, Germany) was clamped to a custom stationary frame positioned 66 cm in front of the center tank of the 3-chamber OTFST apparatus, and the camera was positioned such that the image of the entire front of the tank filled the frame. Two additional digital video cameras were clamped to the same stationary frame and positioned such that the image of each flanking tank filled the frame for its respective camera. The cameras were connected via CAT-5 ethernet cable to a PC-compatible computer (Dell Precision 3630, Dell Computer, Inc., Round Rock, TX, USA) running Microsoft Windows 10. During experimental sessions, video from the cameras was obtained using MediaRecorder 6.0 (Noldus, Inc., Wageningen, the Netherlands). For quantification of movement variables, digital videos were analyzed using Ethovision XT 16.0 (Noldus, Inc., Wageningen, the Netherlands). The entire area for the testing apparatus and camera frame was surrounded by three custom-built floor-standing screens covered in black cloth to prevent animals on the testing bench from seeing the experimenters, and the computer was contained on a rack outside of the screened area.

2.4. Experimental Procedure

At the beginning of the testing day, opaque barriers attached to strings on a pulley system were placed between the testing tanks. To maintain identical water-quality parameters from the staging tanks to the testing tanks and reduce transfer-stress on animals [63], water was obtained from the reservoir on the flow-through rack system and used to fill the testing tanks at the beginning of each experimental day. Because net-capture and air-transfer can act as stressors and negatively affect the subsequent behavior of zebrafish [64], each subject and its associated stimulus shoals were volume-transferred by carefully pouring the entire contents of each staging tank into its respective tank on the 3-chamber OTFST apparatus at 09:00 on the testing day. After the transfer of staging tank contents, each tank on the testing apparatus had a final volume of 26.7 L of water. Because no main effects or interactions of side position have been revealed in previous studies by our laboratory [56,57], and counterbalancing stimulus shoal positions across the flanking tanks would result in unnecessary duplication [61] by doubling the number of animals needed for the experiment, the positions of the intact and alarmed shoals were not counterbalanced. The intact shoal was always placed in the left flanking tank while the shoal to be alarmed was always placed in the right flanking tank. Following addition of the test subject and stimulus shoals to their respective tanks, an aerator was placed in each tank to maintain oxygen levels during the day, and a clear lid was placed on each tank. One 5 mL aliquot of the H₃NO working solution was removed from the freezer and thawed to room temperature (22 - 24 °C). Subjects and stimulus shoals were then acclimated for 4 hours before testing at 13:00.

Before starting the experimental session, a welfare check was performed to ensure that all animals in the apparatus were responding normally, and aerators were removed from each testing tank and lids replaced. Because H₃NO degrades quickly in acidic conditions [48,65], the pH was obtained for the water in the right flanking tank and tested with a bench meter to ensure it was 7.0 or higher prior to starting the experimental session. A PDDT was filled with the contents of the aliquot. The experimenter opened a session in MediaRecorder and began recording as the dividers between the testing tanks were removed. Within 30 seconds, the experimenter delivered the H₃NO by reaching from behind the screen to fully insert the PDDT through the hole in the lid, quickly dispensing the contents into the tank to provide a 1.5 nM final concentration of H₃NO. This 1.5 nM concentration has been demonstrated to activate detectable alarm responses in zebrafish, including erratic movements [48]. Recording continued for 10 minutes to obtain digital video of each subject's movement throughout the session. At the conclusion of the session, the test subject and stimulus shoal exposed to H₃NO were transferred to an outbound holding tank. The intact stimulus shoal was transferred to a 2.8 L staging tank and placed on another row of the flow-through rack system to be used as an alarmed shoal in a future session at least one week later. This step allowed for further reduction of the total number of animals necessary to run the experiment [66–68]. After all animals were removed from the testing tanks, the tanks were drained, rinsed thoroughly, and sprayed with a 5% citric acid solution to sanitize the tanks and degrade any residual H₃NO. Tanks were allowed to air dry overnight before being rinsed again at the beginning of the subsequent experimental day and returned to the testing bench.

At the conclusion of all testing sessions, the digital video from subjects' sessions was analyzed in Ethovision XT 16.0. The arena settings for Ethovision XT 16.0 were set to divide the screen into the leftmost third of the arena (closest to the intact shoal), the middle third of the arena, and the rightmost third of the arena (closest to the alarmed shoal). These zones were used for subsequent data extraction and movement analyses.

2.5. Design and Analysis of Subjects' Responding During the 3-Chamber OTFST

The experiment was a 2 (Sex: Male or Female - between subjects) × 3 (Vertical Zone: Intact Shoal Third, Middle Third, and Alarmed Shoal Third - within subjects) mixed-factorial design. The following measures were obtained using Ethovision XT 16.0 to analyze the digital videos from subjects' sessions:

- Percent duration within each zone: The relative amount of total time each subject spent in each zone during the entire session.
- Percent of session time spent moving: The relative amount of time each subject was moving at any velocity in each zone
- Percent of session time spent freezing: The relative amount of time each subject had ceased any detectable movement for a minimum of 3 seconds in each zone.
- Average movement velocity: The average movement speed in mm/s for each subject's movement within each zone.

Data were organized into a 2 (Sex) \times 3 (Vertical Zone) design and analyzed with SPSS 29.01 (IBM Corp., Armonk, NY, USA) using a Linear Mixed Model (LMM) with Type III Sums of Squares at $\alpha = 0.05$; for advantages of the LMM over mixed-factorial ANOVA, see [69]. The model was set with diagonal covariance structure for heterogeneous variance. Degrees of freedom for the denominator of mixed-model F-ratios were adjusted according to the Maximum Likelihood Estimator for LMM. When appropriate, unplanned comparisons were made using the Bonferroni correction for familywise error. Experimental effect sizes are reported as partial- η^2 values as well as Cohen's f values.

3. Results

3.1. Manipulation Check

To verify that H3NO exposure had a demonstrable effect on the activity of shoal members, digital video footage of shoal responses was analyzed with Ethovision 16.0 to obtain the mean inter-individual distances (IID) for each stimulus shoal during each session. These data were organized into a between-subjects design (alarmed shoal vs intact shoal) and analyzed using an independent-samples t -test with SPSS 29.01 at $\alpha = 0.05$. The mean IID during each session was calculated for both types of shoals. As Levene's test for equality of variance was not significant, $F(1,38) = 3.37$, $p = 0.07$, equality of variances was assumed for the subsequent independent-samples t -test. The mean IID for members of intact shoals ($M = 10.42$ cm, $SEM = 0.56$) was significantly greater than the mean IID for members of alarmed shoals ($M = 8.67$ cm, $SEM = 0.83$), $t(38) = 1.75$, $p = 0.044$, $d = 0.55$, $\eta_p^2 = 0.081$. The effect size on IID due to the administration of the synthetic alarm substance is considered "medium" under Cohen's criteria [71] and accounts for 8.1% of the variance in IID across sessions (see Figure 1). Alarmed shoals had a lower mean IID than intact shoals, which is consistent with previous reports e.g. [51,70].

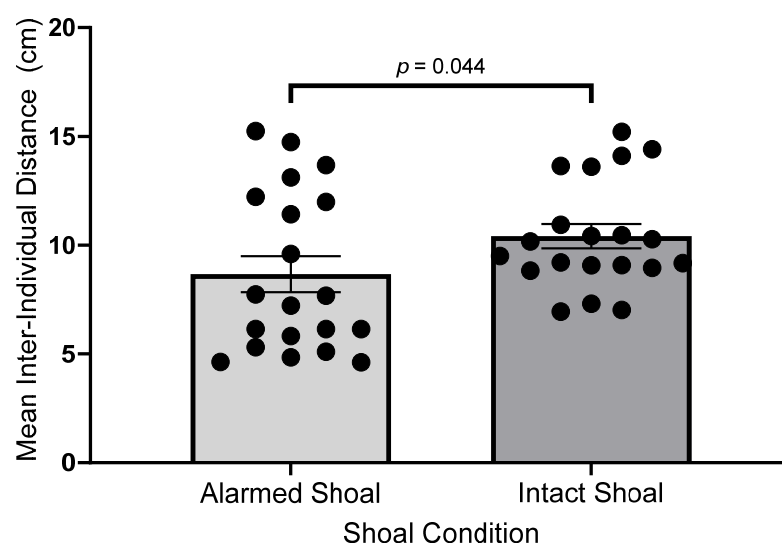


Figure 1. Dot-plot for the comparison of mean inter-individual distance (IID) between alarmed shoals and intact shoals. Error bars represent ± 1 SEM.

3.2. Percent of Session Time in Zones

The analysis of subjects' percent duration in the different zones revealed a significant main effect of Zone, $F(2, 32.25) = 43.5$, $p \leq .001$, $\eta_p^2 = 0.73$, $f = 1.55$; the effect size is considered "very large" under Cohen's criteria [71] and accounts for 73% of the variance in percent duration in different zones. However, there was neither a significant main effect of Sex nor an interaction of Sex by Zone (both $F_s \leq 0.2$, p 's ≥ 0.682). On average, subjects spent almost twice as much time in the zone closer to the intact shoal ($M = 64.7\%$, $SEM = 7.24\%$) than they spent in the zone closer to the alarmed shoal ($M = 34.67\%$, $SEM = 7.46\%$). Subjects spent the least amount of time in the middle zone of the arena ($M = 3.56\%$, $SEM = 0.75\%$; see Figure 2).

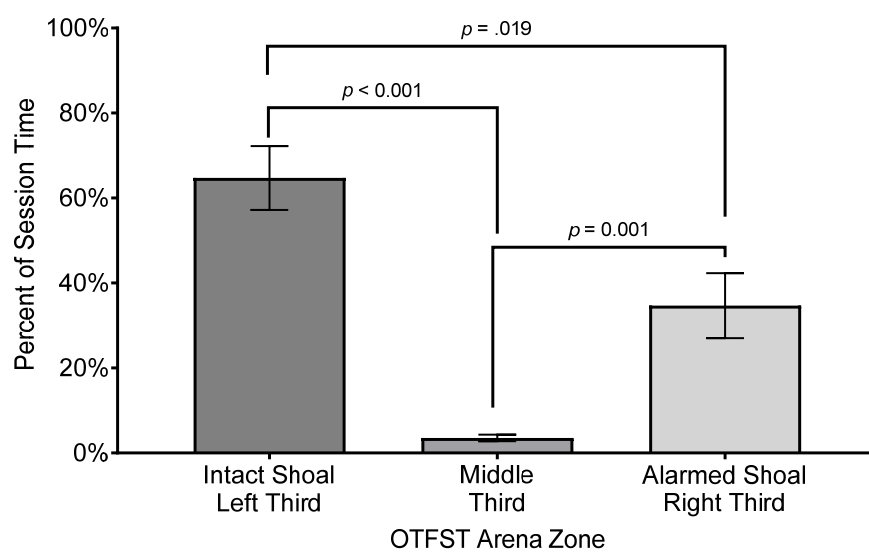


Figure 2. Comparison of mean percent session time subjects spent in each zone of the testing arena. Difference bars between zones are reported using the Bonferroni adjustment for family-wise error. Error bars represent ± 1 SEM.

3.3. Percent of Time in Motion

The analysis of subjects' percent of session time in motion in the different zones revealed a significant effect of Zone, $F(2, 32.35) = 31.84$, $p \leq .001$, $\eta_p^2 = 0.66$, $f = 1.32$; the effect size is considered "large" under Cohen's criteria [71] and accounts for 66% of the variance in percent time in motion across the different zones. However, there was neither a significant main effect of Sex nor an interaction of Sex by Zone (both $F_s \leq 0.14$, p 's ≥ 0.136). Subjects spent a greater percentage of time moving in zones closer to either the intact shoal ($M = 46.97\%$, $SEM = 6.04\%$) or the alarmed shoal ($M = 26.06\%$, $SEM = 6.23\%$) than they did in the middle zone of the arena ($M = 3.41\%$, $SEM = 0.710$; see Figure 3).

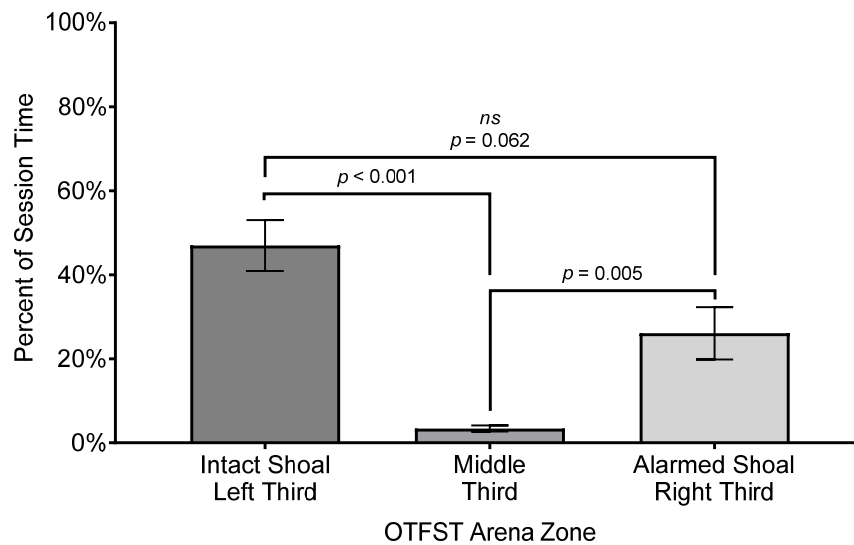


Figure 3. Comparison of mean percent session time subjects spent in motion in each zone of the testing arena. Difference bars between zones are reported using the Bonferroni adjustment for family-wise error. Error bars represent ± 1 SEM.

3.4. Percent of Time Without Motion (Freezing)

The analysis of subjects' percent of session time without motion in the different zones revealed a significant effect of Zone, $F(2, 32.42) = 49.04$, $p \leq .001$, $\eta_p^2 = 0.75$, $f = 1.65$; the effect size is considered "very large" under Cohen's criteria [71] and accounts for 75% of the variance in percent duration of freezing across the different zones. Regarding the main effect of Zone, subjects spent more time motionless in the zone closer to the intact shoal ($M = 17.68\%$, $SEM = 1.96\%$) than they did when closer to the alarmed shoal ($M = 8.52\%$, $SEM = 1.97\%$). Subjects spent the least amount of time motionless in the middle zone of the arena ($M = 0.14\%$, $SEM = 0.05\%$; see Figure 4).

The ANOVA also revealed a significant effect of Sex, $F(1, 39.01) = 4.14$, $p = 0.049$, $\eta_p^2 = 0.10$, $f = 0.28$; the effect size is considered "medium" under Cohen's criteria [71] and accounts for 10% of the variance in percent duration of freezing across the different zones. Regarding the main effect of subject Sex, males ($M = 10.6\%$, $SEM = 1.33\%$) spent more time motionless than females ($M = 6.9\%$, $SEM = 1.29\%$). The interaction of Sex by Zone on percent time freezing was not significant, $F(2, 32.42) = 2.46$, $p = 0.101$, $\eta_p^2 = 0.103$, $f = 0.29$.

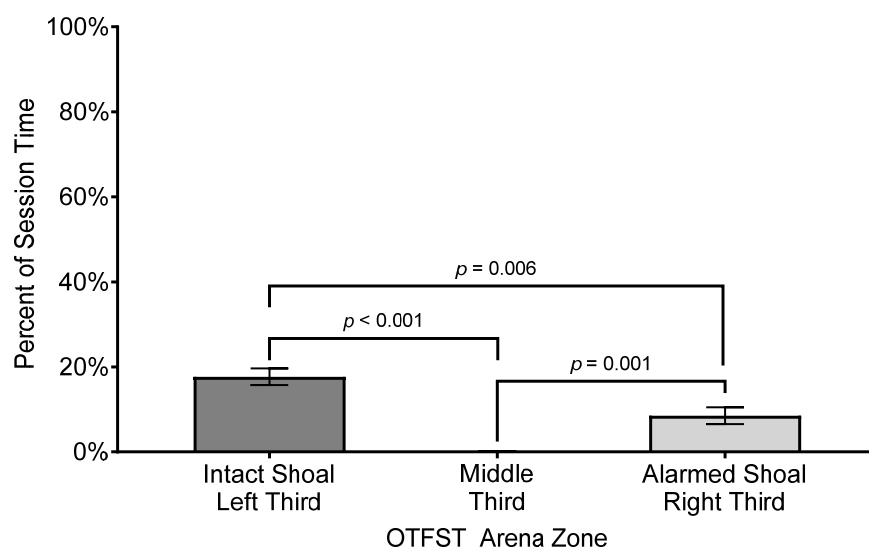


Figure 4. Comparison of mean percent session time subjects spent freezing in each zone of the testing arena. Difference bars between zones are reported using the Bonferroni adjustment for family-wise error. Error bars represent ± 1 SEM.

3.5. Velocity During Movement

The analysis of subjects' movement velocity revealed a significant effect of Zone, $F(2, 23.63) = 14.28$, $p \leq .001$, $\eta_p^2 = 0.17$, $f = 0.33$; the effect size is considered "medium" under Cohen's criteria [71] and accounts for 17% of the variance in velocity during movement across the different zones. However, there was neither a significant main effect of Sex nor an interaction of Sex by Zone (both $F_s \leq 0.83$, $p's \geq 0.38$). Regarding the main effect of Zone, subjects swam the slowest average speed in the zone closer to the intact shoal ($M = 4.57$ mm/s, $SEM = 0.26$ mm/s). There was no significant difference in average swim speed between the middle zone ($M = 10.30$ mm/s, $SEM = 1.10$ mm/s) and the zone proximal to the alarmed shoal ($M = 8.54$ mm/s, $SEM = 2.16$ mm/s; see Figure 5)

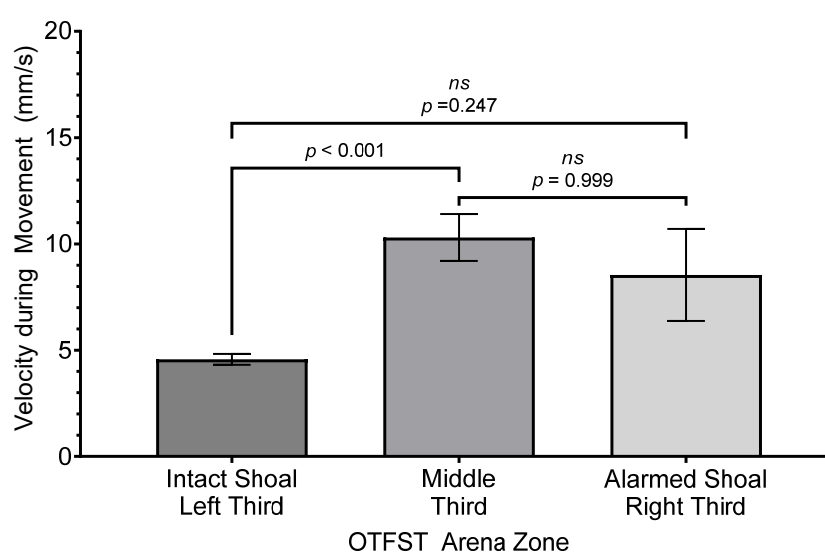


Figure 5. Comparison of mean velocity during subjects' movements in each zone of the testing arena. Difference bars between zones are reported using the Bonferroni adjustment for family-wise error. Error bars represent ± 1 SEM.

4. Discussion

In a previous experiment on social contagion and social buffering in zebrafish [52], males were responsive to visual cues indicating threat perception from alarmed conspecifics, while females demonstrated greater sensitivity to chemical alarm substances. However, the choice architecture in that study offered only an asymmetrical comparison, i.e. a "Hobson's Choice" [72], where conspecifics were presented visually on one side of the experimental chamber while the other side lacked any stimulus. This design provided limited insight into preference or avoidance responses, as subjects were basically choosing between the presence and the absence of conspecifics rather than evaluating among two concurrently-available social stimuli. The present study addresses this limitation by using a symmetrical choice architecture, presenting zebrafish with two shoaling options: an intact shoal and an alarmed shoal. The present approach provides a more comprehensive assessment of social preference under conditions specifically designed to test for social cueing of fear responses and predatory threat.

Our findings are consistent with prior research [52], demonstrating that both male and female zebrafish in the present study demonstrate social contagion of fear when visually exposed to alarmed conspecifics. Specifically, subjects of both sexes exhibited a clear preference for proximity to the intact

shoal over the alarmed shoal, spending significantly more time in zones closer to the intact shoal. These results suggest that zebrafish can effectively differentiate between alarmed and unalarmed groups solely by relying on visual cues to assess threat levels. By preferentially associating with less-alarmed conspecifics, zebrafish can mitigate predation risk while maintaining the benefits of social grouping [9,13,15]. These behaviors highlight the utility of zebrafish as a model organism for studying complex social processes in response to environmental threats.

Interestingly, our study also revealed a significant sex difference in freezing behavior, with males freezing more than females. This finding aligns with previous research [52] which demonstrated that males are more responsive to visual information related to predatory threat. This sex-specific behavior may reflect differences in life-history strategies, where males adopt higher-risk approaches that prioritize quick reactions to visual cues, even at the expense of greater energy expenditure [74,75]. In contrast, females may rely more heavily on chemical alarm cues, which are slower to disseminate but may provide more reliable information about the presence of a predator. These sex differences underscore the importance of considering individual differences in threat perception and response when interpreting zebrafish behavior.

The present findings not only contribute to our understanding of zebrafish social behavior but also offer valuable insights into the dynamics of social contagion in the context of threat perception. Future research should investigate symmetrical choice dynamics under conditions that combine social buffering and social contagion to explore potential interactions between these two phenomena. Additionally, exploring dose-dependent effects of alarm substance exposure could provide deeper insights into how varying levels of threat cues modulate zebrafish behavior. Such studies could establish the thresholds at which zebrafish shift from avoidance to engagement behaviors and enhance our understanding of the neural mechanisms underlying these responses. This area of research holds promise for advancing our knowledge of adaptive group behaviors and their implications for social responding in both zebrafish and other social species.

5. Conclusions

Previous research on social contagion and buffering in zebrafish [52] revealed that males respond strongly to visual threat cues from alarmed conspecifics when choosing to either affiliate or avoid a shoal, whereas females appear to rely more on chemical alarm substances when choosing to either affiliate or avoid a shoal. By adding a second shoaling option, the present study employed a symmetrical choice architecture which offered zebrafish subjects the option to associate with either an intact shoal or an alarmed shoal, thereby enabling a clearer evaluation of social preferences under predatory threat. Consistent with prior findings, both sexes preferred proximity to the intact shoal, highlighting zebrafish's ability to differentiate threat levels via visual cues and mitigate predation risk by associating with less-alarmed conspecifics. Notably, males exhibited more freezing behavior than females, reflecting sex-specific differences in threat perception: males may prioritize rapid visual responses and females may rely on slower, but perhaps more reliable, chemical cues. These results underscore zebrafish's utility as a model for studying social and threat-related behaviors and suggest directions for future research, including the interplay between social buffering and contagion, dose-dependent alarm cue effects, and the neural mechanisms underlying adaptive group behavior.

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References

1. Vidal-García, M.; O'Hanlon, J. C.; Svenson, G. J.; Umbers, K. D. The evolution of startle displays: A case study in praying mantises. *Proc. R. Soc. B.*, **2020**, *287*, 20201016. <https://doi.org/10.1098/rspb.2020.1016>
2. Humphreys, R. K.; Ruxton, G. D. A review of thanatosis (death feigning) as an anti-predator behaviour. *Behav. Ecol. Sociobiol.* **2018**, *72*, 1-16. <https://doi.org/10.1007/s00265-017-2436-8>.
3. Ortega J.C.; Figueiredo B.R.; da Graça W.J.; Agostinho A.A.; Bini L.M. Negative effect of turbidity on prey capture for both visual and non-visual aquatic predators. *J. Anim. Ecol.* **2020**, *89*, 2427-2439. <https://doi.org/10.1111/1365-2656.13329>.
4. Majolo, B.; de Bortoli Vizioli, A.; Schino, G. Costs and benefits of group living in primates: group size effects on behaviour and demography. *Animal Behaviour*, **2008**, *76*, 1235-1247. <https://doi.org/10.1016/j.anbehav.2008.06.008>.
5. Ioannou, C.C.; Guttal, V.; Couzin, I.D. Predatory fish select for coordinated collective motion in virtual prey. *Science* **2012**, *337*, 1212-1215. <https://doi.org/10.1126/science.121891>.
6. Ioannou, C.C.; Krause, J. Searching for prey: the effects of group size and number. *Anim. Behav.* **2008**, *75*, 1383-1388. <https://doi.org/10.1016/j.anbehav.2007.09.012>.
7. Kruuk, H. Predators and anti-predator behaviour of the black-headed gull (*Larus ridibundus* L.). *Behaviour: Supplement 11*, **1964**, 1-129. <https://www.jstor.org/stable/30039149>.
8. Kruuk, H. The biological function of gulls' attraction towards predators. *Anim. Behav.* **1976**, *24*, 146-153. [https://doi.org/10.1016/S0003-3472\(76\)80108-X](https://doi.org/10.1016/S0003-3472(76)80108-X).
9. Rubenstein, D. *Animal Behavior*, 12th ed.; Oxford University Press: Sinauer Associates: Oxford, England, 2022
10. Lehtonen, J.; Jaatinen, K. Safety in numbers: the dilution effect and other drivers of group life in the face of danger. *Behav. Ecol. Sociobiol.* **2016**, *70*, 449-458. <https://doi.org/10.1007/s00265-016-2075-5>.
11. Hammer, T.L.; Bize, P.; Gineste, B.; Robin, J.P.; Groscolas, R.; Viblanc, V.A. Disentangling the “many-eyes”, “dilution effect”, “selfish herd”, and “distracted prey” hypotheses in shaping alert and flight initiation distance in a colonial seabird. *Behav. Proc.* **2023**, *210*, 104919. <https://doi.org/10.1016/j.beproc.2023.104919>.
12. Ward, A.; Webster, M. Chapter 4: Social Foraging and Predator-Prey Interactions. In *Sociality: The Behaviour of Group-Living Animals*. Ward, A., Webster, M., Eds; Springer International, Cham, Switzerland; 2016, pp. 55-87. https://doi.org/10.1007/978-3-319-28585-6_4

13. Tan, M.; Zhang, S.; Stevens, M.; Li, D.; Tan, E.J. Antipredator defences in motion: animals reduce predation risks by concealing or misleading motion signals. *Biol. Rev.* **2024**, *99*, 778-96. <https://doi.org/10.1111/brv.13044>.
14. Cattelan, S.; Griggio, M. Within-shoal phenotypic homogeneity overrides familiarity in a social fish. *Behav. Ecol. Sociobiol.* **2020**, *74*, 48. <https://doi.org/10.1007/s00265-020-2826-1>
15. Pitcher T.J. Chapter 12, Functions of shoaling behaviour in teleosts. In *The Behavior of Teleost Fishes*. Springer: Boston, MA, USA; 1986; pp. 294–337. https://doi.org/10.1007/978-1-4684-8261-4_12.
16. Lima, S.L.; Dill, L.M. Behavioral decisions made under the risk of predation: A review and prospectus. *Can. J. Zool.* **1990**, *68*, 619-640. <https://doi.org/10.1139/z90-092>.
17. Burbano, D.; Senthilkumar, S.; Manzini, M.C. Exploring emotional contagion in zebrafish: A virtual-demonstrator study of positive and negative emotions. *Behav. Processes* **2023**, *213*, 104961. <https://doi.org/10.1016/j.beproc.2023.104961>.
18. Thapa, H.; Salahinejad, A.; Crane, A.L.; Ghobeishavi, A.; Ferrari, M.C. Background predation risk induces anxiety-like behaviour and predator neophobia in zebrafish. *Anim. Cogn.* **2024**, *27*, 69. <https://doi.org/10.1007/s10071-024-01908-z>.
19. Chivers, D.P.; Smith, R.J. Chemical alarm signalling in aquatic predator-prey systems: A review and prospectus. *Ecoscience* **1998**, *1*, 338-352. <https://doi.org/10.1080/11956860.1998.11682471>.
20. Crane, A.L.; Bairos-Novak, K.R.; Goldman, J.A.; Brown, G.E. Chemical disturbance cues in aquatic systems: A review and prospectus. *Ecol. Monogr.* **2022**, *92*, e01487. <https://doi.org/10.1002/ecm.1487>.
21. Li, Y.; Yan, Z.; Lin, A.; Yang, X.; Li, X.; Yin, X.; Li, W.; Li, K. Epidermal oxysterols function as alarm substances in zebrafish. *iScience*, **2024**, *27*, 109660 <https://doi.org/10.1016/j.isci.2024.109660>.
22. Masuda, M.; Ihara, S.; Mori, N.; Koide, T.; Miyasaka, N.; Wakisaka, N.; Yoshikawa, K.; Watanabe, H.; Touhara, K.; Yoshihara, Y. Identification of olfactory alarm substances in zebrafish. *Current Biology*, **2024**, *34*, 1377-1389. <https://doi.org/10.1016/j.cub.2024.02.003>.
23. Wisenden, B.D.; Rugg, M.L.; Korpi, N.L.; Fuselier, L.C. Lab and field estimates of active time of chemical alarm cues of a cyprinid fish and an amphipod crustacean. *Behav.*, **2009**, *1*, 1423-42. <https://doi.org/10.1163/156853909X440998>.
24. Crane, A.L.; Achtymichuk, G.H.; Rivera-Hernández, I.A.E.; Preagola, A.A.; Thapa, H.; Ferrari, M.C.O. Uncertainty about old information results in differential predator memory in tadpoles. *Proc. R. Soc. B*, **2023**, *290*: 20230746. <https://doi.org/10.1098/rspb.2023.0746>.
25. Chivers, D.P.; Dixon, D.L.; White, J.R.; McCormick, M.I.; Ferrari, M.C.. Degradation of chemical alarm cues and assessment of risk throughout the day. *Ecol. Evol.*, **2013**, *3*, 3925-3934. <https://doi.org/10.1002/ece3.760>.
26. Michaelis, B.T.; Leathers, K.W.; Bobkov, Y.V.; Ache, B.W.; Principe, J.C.; Baharloo, R.; Park I.M.; Reidenbach, M.A. Odor tracking in aquatic organisms: The importance of temporal and spatial intermittency of the turbulent plume. *Sci. Rep.* **2020**, *10*:7961. <https://doi.org/10.1038/s41598-020-64766-y>.
27. Fontana, B.D.; Mezzomo, N.J.; Kalueff, A.V.; Rosenberg, D.B. The developing utility of zebrafish models of neurological and neuropsychiatric disorders: A critical review. *Exp. Neuro.* **2018**, *299*, 157-171. <https://doi.org/10.1016/j.expneurol.2017.10.004>.
28. Gerlai, R. Zebrafish (*Danio rerio*): A newcomer with great promise in behavioral neuroscience. *Neurosci. Biobehav. Rev.* **2023**, *144*, 104978. <https://doi.org/10.1016/j.neubiorev.2022.104978>.
29. Choi, T.Y.; Choi, T.I.; Lee, Y.R.; Choe, S.K.; Kim, C.H. Zebrafish as an animal model for biomedical research. *Exp. Mol. Med.* **2021**, *53*, 310-317. <https://doi.org/10.1038/s12276-021-00571-5>.
30. Aleström, P.; D'Angelo, L.; Midtlyng, P.J.; Schorderet, D.F.; Schulte-Merker, S.; Sohm, F.; Warner, S. Zebrafish: Housing and husbandry recommendations. *Lab. Anim.* **2020**, *54*, 213-224. <https://doi.org/10.1177/0023677219869037>.
31. Castranova, D.; Wang, C. Chapter 31: Zebrafish breeding and colony management. In *The Zebrafish in Biomedical Research*; Cartner, S.C., Eisen, J.S., Farmer, S.C., Guillemin, K.J., Kent, M.L., Sanders, G.E; Eds.; Elsevier Academic Press: Cambridge, MA, USA, **2020**, pp 357-364. <https://doi.org/10.1016/B978-0-12-812431-4.00031-2>.

32. Stewart, A. M.; Braubach, O.; Spitsbergen, J.; Gerlai, R.; Kalueff, A. V. Zebrafish models for translational neuroscience research: from tank to bedside. *Trends Neurosci.* **2014**, *37*, 264-278. <https://doi.org/10.1016/j.tins.2014.02.011>.
33. Chahardehi, A.M.; Hosseini, Y.; Mahdavi, S.M.; Naseh, I. Zebrafish, a biological model for pharmaceutical research for the management of anxiety. *Mol. Biol. Rep.* **2023**, *50*, 3863-3872. <https://doi.org/10.1007/s11033-023-08263-1>.
34. Lachowicz, J.; Niedziałek, K.; Rostkowska, E.; Szopa, A.; Świąder, K.; Szponar, J.; Serefko, A. Zebrafish as an animal model for testing agents with antidepressant potential. *Life* **2021**, *11*, 792. <https://doi.org/10.3390/life11080792>.
35. Rea, V.; Van Raay, T.J. Using zebrafish to model autism spectrum disorder: A comparison of ASD risk genes between zebrafish and their mammalian counterparts. *Front. Mol. Neurosci.*, **2020**, *13*, 575575. <https://doi.org/10.3389/fnmol.2020.575575>.
36. Mathur, P.; Guo, S. Use of zebrafish as a model to understand mechanisms of addiction and complex neurobehavioral phenotypes. *Neurobiol. Dis.* **2010**, *40*, 66–72. <https://doi.org/10.1016/j.nbd.2010.05.016>.
37. Ruhl, N.; McRobert, S.P. The effect of sex and shoal size on shoaling behaviour in *Danio rerio*. *J. Fish Biol.* **2005**, *67*, 1318-1326. <https://doi.org/10.1111/j.0022-1112.2005.00826.x>
38. Pyke, G. H. Optimal foraging theory: A critical review. *Annu. Rev. Ecol. Evol. Syst.* **1984**, *15*, 523-575. <https://doi.org/10.1146/annurev.es.15.110184.002515>.
39. Miller N.Y.; Gerlai R. Shoaling in zebrafish: What we don't know. *Rev. Neurosci.*, **2011**, *22*, 17-25. <https://doi.org/10.1515/rns.2011.004>.
40. Ettinger, A.; Lebron, J.; Palestis, B.G.; Sex-assortative shoaling in zebrafish (*Danio rerio*). *Bios.* **2009**, *80*, 153-158. <https://doi.org/10.1893/011.080.0402>.
41. Von Frisch, K. Zur psychologie des fisch-schwarmes [Psychology of preference in fish]. *Naturwissenschaften*, **1938**, *26*, 601–606. <https://doi.org/10.1007/BF01590598>.
42. Mathuru, A.S.; Kibat, C.; Cheong, W.F.; Shui, G.; Wenk, M.R.; Friedrich, R.W.; Jesuthasan, S. Chondroitin fragments are odorants that trigger fear behavior in fish. *Curr. Biol.* **2012**, *22*, 538-544. <https://doi.org/10.1016/j.cub.2012.01.061>.
43. Goodall, J.; Rincón-Camacho, L.; Pozzi, A.G.; Epidermal club cells in the cardinal tetra (*Paracheirodon axelrodi*): Presence, distribution, and relationship to antipredator behavior. *J. Zool.* **2024**, *164*, 126170. <https://doi.org/10.1016/j.zool.2024.126170>.
44. Pfeiffer W. Über die schreckreaktion bei fischen und die herkunft des schreckstoffes [On the fear reaction in fish and the origin of alarm substance]. *Z. Vgl. Physiol.* **1960**, *43*, 578-614. <https://doi.org/10.1007/BF00298105>.
45. Blaser, R. E.; Chadwick, L.; McGinnis, G. C. Behavioral measures of anxiety in zebrafish (*Danio Rerio*). *Behav. Brain Res.* **2010**, *208*, 56-62. <https://doi.org/10.1016/j.bbr.2009.11.009>
46. Suboski, M. D.; Bain, S.; Carty, A. E.; McQuoid, L. M.; Seelen, M. I.; Seifert, M. Alarm reaction in acquisition and social transmission of simulated-predator recognition by zebra danio fish (*Brachydanio rerio*). *J. Comp. Psychol.* **1990**, *104*, 101–112. <https://doi.org/10.1037/0735-7036.104.1.101>.
47. Speedie, N.; Gerlai, R. Alarm substance induced behavioral responses in zebrafish (*Danio rerio*). *Behav. Brain Res.* **2008**, *188*, 168–177. <https://doi.org/10.1016/j.bbr.2007.10.031>.
48. Parra, K. V.; Adrian, J. C.; Gerlai, R. The synthetic substance hypoxanthine 3-N-oxide elicits alarm reactions in Zebrafish (*Danio rerio*). *Behav. Brain Res.* **2009**, *205*, 336–341. <https://doi.org/10.1016/j.bbr.2009.06.037>.
49. Wisenden, B.D.; Andebrhan, A.A.; Anderson, C.M.; Angus, J.M.; Coffman, I.C.; Cloutier, M.E.; et al. Olfactory cues of risk and visual cues of safety interact with sympatry and phylogeny in shaping behavioral responses by littoral fishes. *Behav. Ecol. Sociobiol.*, **2023**, *77*, 91. <https://doi.org/10.1007/s00265-023-03367-x>.
50. Ogi, A.; Licitra, R.; Naef, V.; Marchese, M.; Fronte, B.; Gazzano, A.; Santorelli, F.M.. Social preference tests in zebrafish: A systematic review. *Front. Vet. Sci.*, **2021**, *7*, 590057. <https://doi.org/10.3389/fvets.2020.590057>.
51. Rosa, L. V.; Costa, F. V.; Canzian, J.; Borba, J. V.; Quadros, V. A.; Rosemberg, D. B. Three- and bi-dimensional analyses of the shoaling behavior in Zebrafish: Influence of modulators of anxiety-like responses. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2020**, *102*. <https://doi.org/10.1016/j.pnpbp.2020.109957>.

52. Akinrinade, I. D.; Varela, S. A. M.; Oliveira, R. F. Sex differences in social buffering and social contagion of alarm responses in zebrafish. *Animal Cognition* **2023**, *26*, 1307-1318. <https://doi.org/10.1007/s10071-023-01779-w>.
53. Kuroda, T.; Ritchey, C.M.; Podlesnik, C.A. Selective effects of conspecific movement on social preference in zebrafish (*Danio rerio*) using real-time 3D tracking and 3D animation. *Scientific Reports* **2023**, *13*, 10502. <https://doi.org/10.1038/s41598-023-37579-y>
54. Faustino A.I.; Tacão-Monteiro A.; Oliveira R.F. Mechanisms of social buffering of fear in zebrafish. *Sci. Rep.* **2017**, *7*, 44329. <https://doi.org/10.1038/srep44329>.
55. Ariyasiri K.; Choi T.-I.; Kim O.-H.; Hong T.-I.; Gerlai R.; Kim C.-H. Pharmacological (ethanol) and mutation (sam2 KO) induced impairment of novelty preference in zebrafish quantified using a new three-chamber social choice task. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2019**, *88*, 53-65. <https://doi.org/10.1016/j.pnpbp.2018.06.009>.
56. Velkey, A. J.; Boles, J.; Betts, T. K.; Kay, H.; Henenlotter, R.; Wiens, K. M. High fidelity: Assessing Zebrafish (*Danio rerio*) responses to social stimuli across several levels of realism. *Behav. Processes*, **2019**, *164*, 100-108. <https://doi.org/10.1016/j.beproc.2019.04.012>.
57. Velkey, A. J.; Koon, C. H.; Danstrom, I. A.; Wiens, K. M. Female zebrafish (*Danio rerio*) demonstrate stronger preference for established shoals over newly-formed shoals in the three-tank open-swim preference test. *Plos one*, **2022**, *17*, e0265703. <https://doi.org/10.1371/journal.pone.0265703>
58. Canzian, J.; Fontana, B. D.; Quadros, V. A.; Rosemberg, D. B. Conspecific alarm substance differently alters group behavior of zebrafish populations: Putative involvement of vholinergic and purinergic signaling in anxiety- and fear-like responses. *Behav. Brain Res.* **2017**, *320*, 255-263. <https://doi.org/10.1016/j.bbr.2016.12.018>.
59. Sison, M.; Gerlai, R. Behavioral performance altering effects of MK-801 in zebrafish (*Danio rerio*). *Behav. Brain Res.*, **2011**, *220*, 331-337. <https://doi.org/10.1016/j.bbr.2011.02.019>.
60. do Nascimento, B.G.; Maximino, C. Social investigation and social novelty in zebrafish: Roles of salience and novelty. *Behav. Processes*, **2023**, *210*, 104903. <https://doi.org/10.1016/j.beproc.2023.104903>.
61. National Research Council. *Guide for the Care and Use of Laboratory Animals*: 8th ed.; The National Academies Press: Washington, DC, USA, 2011 <https://doi.org/10.17226/12910>.
62. Ariyasiri K.; Choi T.-I.; Kim O.-H.; Hong T.-I.; Gerlai R.; Kim C.-H. Pharmacological (ethanol) and mutation (sam2 KO) induced impairment of novelty preference in zebrafish quantified using a new three-chamber social choice task. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2019**, *88*, 53-65. <https://doi.org/10.1016/j.pnpbp.2018.06.009>.
63. Tsang, B.; Gerlai, R. Nature versus laboratory: How to optimize housing conditions for zebrafish neuroscience research. *Trends Neurosci.* **2024**, *47*, 985-993, <https://doi.org/10.1016/j.tins.2024.08.013>.
64. Shishis, S.; Tsang, B.; Ren, G.J.; Gerlai, R. Effects of different handling methods on the behavior of adult zebrafish. *Physiol. Behav.*, **2023**, *262*, 114106. <https://doi.org/10.1016/j.physbeh.2023.114106>.
65. Brown, G.E.; Adrian, Jr., J.C.; Lewis, M.G.; Tower, J.M. The effects of reduced pH on clark73
66. chemical alarm signalling in *Ostariophysan* fishes. *Can. J. Fish. Aquat. Sci.* **2002**, *59*, 1331-1338. <https://doi.org/10.1139/f02-104>.
67. MacArthur Clark, J. The 3Rs in research: A contemporary approach to replacement, reduction and refinement. *Br. J. Nutr.*, **2017**, *120(s1)*, S1-S7. <https://doi.org/10.1017/s0007114517002227>.
68. Tadich, T.; Tarazona, A.M.. Replacement, Reduction and Refinement: Ethical Considerations in the Current Applications of the 3Rs. In *Handbook of Ethical Decisions*; Springer eBooks. 2023, Volume 1, pp. 667-683. https://doi.org/10.1007/978-3-031-29451-8_35.
69. Zhang, T.; Zhang, N.; Feng, R.; Wang, H.; Bi, F.; Wang, S. Promoting the welfare of animals utilized in neuroscience research. *Brain-X*, **2024**, *2*, E70002. <https://doi.org/10.1002/brx2.70002>.
70. Bolker, B.M.; Brooks, M.E.; Clark, C.J.; Geange, S.W.; Poulsen, J.R.; Stevens, M.H.H.; White, J.S.S. Generalized linear mixed models: A practical guide for ecology and evolution. *Trends Ecol. Evol.* **2009**, *24*, 127-135. <https://doi.org/10.1016/j.tree.2008.10.008>.
71. Miller N.; Gerlai R. Quantification of shoaling behaviour in zebrafish (*Danio rerio*). *Behav. Brain Res.* **2007**, *184*, 157-166. <https://doi.org/10.1016/j.bbr.2007.07.007>.

72. Cohen, J. *Statistical Power Analysis for the Behavioral Sciences* 2nd Ed.; Routledge: New York, NY, USA, 1988; pp.284-288. <https://doi.org/10.4324/9780203771587>.
73. Clark, C.W. Antipredator behavior and the asset-protection principle. *Beh. Ecol.* **1994**, *5*, 159-170. <https://doi.org/10.1093/beheco/5.2.159>.
74. Roy, T.; Shukla, R.; Bhat, A. Risk-taking during feeding: Between- and within-population variation and repeatability across contexts among wild zebrafish. *Zebrafish*, **2017**, *14*, 393–403. <https://doi.org/10.1089/zeb.2017.1442>.
75. Roy, T.; Bhat, A. Repeatability in boldness and aggression among wild zebrafish (*Danio rerio*) from two differing predation and flow regimes. *J. Comp. Psychol.* **2018**, *132*, 349-360. <https://doi.org/10.1037/com0000150>.

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