

## Review

# Bacterial plant biostimulants: A sustainable way towards improving growth, productivity, and health of crops

Basharat Hamid<sup>1</sup>, Muzafar Zaman<sup>1</sup>, Shabeena Farooq<sup>1</sup>, Sabah Fatima<sup>1\*</sup>, R. Z. Sayyed<sup>2</sup>, Zahoor Ahmad Baba<sup>3</sup>, Tahir Ahmad Sheikh<sup>4</sup>, M. S. Reddy<sup>5\*</sup>, Hesham El Enshasy<sup>6,7</sup>, Abdul Gafur<sup>8</sup> and Ni Luh Suriani<sup>9</sup>

- 1 Department of Environmental Science, University of Kashmir, Hazratbal, Srinagar-190006, Jammu and Kashmir, India; [basharat384@gmail.com](mailto:basharat384@gmail.com) (B.H.); [muzafarzaman@gmail.com](mailto:muzafarzaman@gmail.com) (M.Z.); [shabeenafarooq188@gmail.com](mailto:shabeenafarooq188@gmail.com) (S.F.); [sabah.fatima333@gmail.com](mailto:sabah.fatima333@gmail.com) (S.F.).
- 2 Department of Microbiology, PSGVP Mandal's Arts, Science, and Commerce College, Shahada -425409, Maharashtra, India; [sayyedrz@gmail.com](mailto:sayyedrz@gmail.com)
- 3 Division of Basic Science and Humanities, FOA, Wadura, Sher-e-Kashmir University of Agricultural Sciences and Technology-193201, Wadura, Jammu and Kashmir, India; [baba.zahor@gmail.com](mailto:baba.zahor@gmail.com) (Z.A.B.)
- 4 Division of Agronomy, FOA, Wadura, Sher-e-Kashmir University of Agricultural Sciences and Technology-193 201, India; [tahirkmr@gmail.com](mailto:tahirkmr@gmail.com) (T.A.S.)
- 5 Asian PGPR Society for Sustainable Agriculture & Auburn Ventures, Department of Plant Pathology and Entomology, Auburn University, Alabama, Auburn, USA; [prof.m.s.reddy@gmail.com](mailto:prof.m.s.reddy@gmail.com)
- 6 Institute of Bioproduct Development (IBD), Universiti Teknologi Malaysia (UTM), Skudai, Johor Bahru, 81310, Malaysia; [henshasy@ibd.utm.my](mailto:henshasy@ibd.utm.my)
- 7 City of Scientific Research and Technology Applications (SRTA), New Borg Al-Arab, Alexandria, 21934, Egypt
- 8 Sinarmas Forestry Corporate Research and Development, Perawang 28772, Indonesia; [gafur@uwalumni.com](mailto:gafur@uwalumni.com)
- 9 Biology study program. Mathematics and natural sciences faculty. Udayana University, Bali. Indonesia; [niluhsuriani@unud.ac.id](mailto:niluhsuriani@unud.ac.id)

Corresponding author: Prof. M. S. Reddy; [prof.m.s.reddy@gmail.com](mailto:prof.m.s.reddy@gmail.com); Sabah Fatima; [sabah.fatima333@gmail.com](mailto:sabah.fatima333@gmail.com)

**Citation:** Lastname, F.; Last-name, F.; Last-name, F. Title. *Sustainability* **2021**, *13*, x. <https://doi.org/10.3390/xxxxx>

**Abstract:** This review presents a comprehensive and systematic study of the field of bacterial plant biostimulants and considers the fundamental and innovative principles underlying this technology. Plant biostimulants are an important tool for modern agriculture as part of an integrated crop management (ICM) system; helping make agriculture more sustainable and resilient. Plant biostimulants contain substance(s) and/or microorganisms whose function when applied to plants or the rhizosphere, is to stimulate natural processes to enhance plant nutrient uptake, nutrient use efficiency, tolerance to abiotic stress, biocontrol, and crop quality. The use of plant biostimulants has gained substantial and significant heed worldwide as an environment-friendly alternative for sustainable agricultural production. Presently, there is an increasing curiosity of industry and researchers in microbial biostimulants especially, bacterial plant biostimulants (BPs) to improve crop growth and productivity. The BPs that are based on PGPR (plant growth-promoting rhizobacteria) play plausible roles to promote/stimulate the crop plant growth through several mechanisms that include, i) nutrient acquisition by nitrogen (N<sub>2</sub>) fixation and solubilization of insoluble minerals (P, K, Zn), organic acids and siderophores, ii) antimicrobial metabolites and various lytic enzymes, iii) action of growth regulators and stress-responsive/induced phytohormones, iv) ameliorating abiotic stress like drought, high soil salinity, extreme temperatures, oxidative stress, and heavy metals by using different modes

---

of action, and v) plant defense induction modes. Presenting here is a brief review emphasizing the applicability of BPBs as an innovative exertion to fulfill the current food crisis.

**Keywords:** Abiotic Stress; Ethylene; Jasmonic acid; Mineral solubilization; Phytostimulants

---

## 1. Introduction

The global environment is changing continuously and the incidence of global warming caused by extreme climatic events is also on the rise consequently disturbing the world ecosystems including agroecosystems [1]. Such extreme changes in climate can affect the quality and quantity of crops severely by inducing various environmental stresses to crops threatening food security worldwide [2]. An increase in global temperature, atmospheric CO<sub>2</sub> level, tropospheric O<sub>3</sub>, and acid rains can cause multifarious chronic stresses to the plants, reducing their capability to respond in case of pathogen attacks [3]. Among these stresses drought, water scarcity and soil salinization are the most problematic and complicated factors of agricultural losses resulting from man-induced climate changes [4]. Fluctuations in temperature and rainfall variations are key indicators of environmental stresses [5]. Elevated temperatures lead to amplification in the rates of respiration and evapotranspiration in crops, a higher infestation of pests, shifts in weed flora patterns, and reduction in crop duration [6]. Water scarcity is also considered as one of the prime global issues that have direct effects on agricultural systems and according to climate projections, its severity will increase in the future [7]. Water scarcity piercingly influences a crop's gaseous exchange capacity causing the closure of stomata [8]. This leads to the impairment of the evapotranspiration and photosynthetic activities of plants, affecting the overall biomass production [9]. Impaired evapotranspiration reductions also affect the nutrient uptake ability of plants [8]. In semi-arid and arid climatic zones where rainfalls are already less intense and sporadic, the damages caused by drought stress can exacerbate due to excessive accumulation of salts in soil [10].

Furthermore, the liberal use of inorganic fertilizers and pesticides to increase crop productivity and meet the food requirement of the ever-growing human population which is projected to reach 9.7 billion by 2050 has severely affected the health of agroecosystem and human beings. Confrontational challenges of improving agriculture production with limited arable land rely on sustainable technologies. Several technical advances have been suggested in the past three decades to increase the productivity of agricultural production processes by reducing toxic agrochemical substances such as pesticides and fertilizers. An emerging technology tackling these critical problems includes the creation of novel plant biostimulants and successful methods for their application [11-15]. Plant biostimulants differ from other agricultural inputs such as fertilizers and plant protection products because they utilize different mechanisms and work regardless of the presence of nutrients in the products. They also do not have any direct action against pests or diseases and therefore complement the use of fertilizers and

plant protection products. According to the latest European Regulation (EU 2019/1009) biostimulant is an EU fertilizer that seeks to promote processes for plant feeding, regardless of the product's nutrient quality, solely to boost the following plant or plant rhizosphere characteristics; i) increasing nutrient utilization efficiency, ii) abiotic stress alleviation/tolerance, iii) quality traits and iv) soil or rhizosphere supply of stored nutrients [16-17]. Over the past decade, microbiome research has changed our understanding of the complexity and composition of microbial communities. The intense interest of industry and academics in bio-stimulants based on live microbes has increased due to the reason that the growth and development of a plant can be improved under field conditions more effortlessly than other biostimulants [18-19]. Biostimulants are not nutrients, but encourage the utilization of nutrients or help foster plant growth or plant's resistance/tolerance to various types of stresses [9,20]. Plant beneficial fungi and bacteria can be considered as the most promising microbial biostimulants [21]. The recent trend has underscored the fact that plants are not autonomous agents in their environments but are associated with bacterial and fungal microorganisms, and that many external and internal microbial interactions respond to biotic and abiotic stresses [22-23]. Therefore, biostimulants are gradually being incorporated into production systems to alter physiological processes in plants to maximize productivity [24].

Bacterial plant biostimulants (BPBs) comprise a major category of plant biostimulants. Plant growth-promoting rhizobacteria (PGPR) that colonize the plant rhizosphere are the most prominent group of this category [24]. These PGPR improve plant growth, control plant pathogens, improve nutrient and mineral uptake in plants and increase plant's resistance to various types of biotic stresses and tolerance towards abiotic stresses (Figure 1). The representative beneficial groups of PGPRs based BPBs include nitrogen-fixing *Rhizobium*, *Azotobacter* spp., *Azospirillum* spp., *Pseudomonas* spp., and *Bacillus* spp. [25-26]. The present review describes the recent knowledge concerning the beneficial BPBs and their role in improving crop health through various mechanisms. The article concludes by highlighting the main findings of an in-depth analysis of research articles published between 2015 and 2020, sorted using different databases like Google Scholar, Science direct, Pub Med, Web of Science, etc.

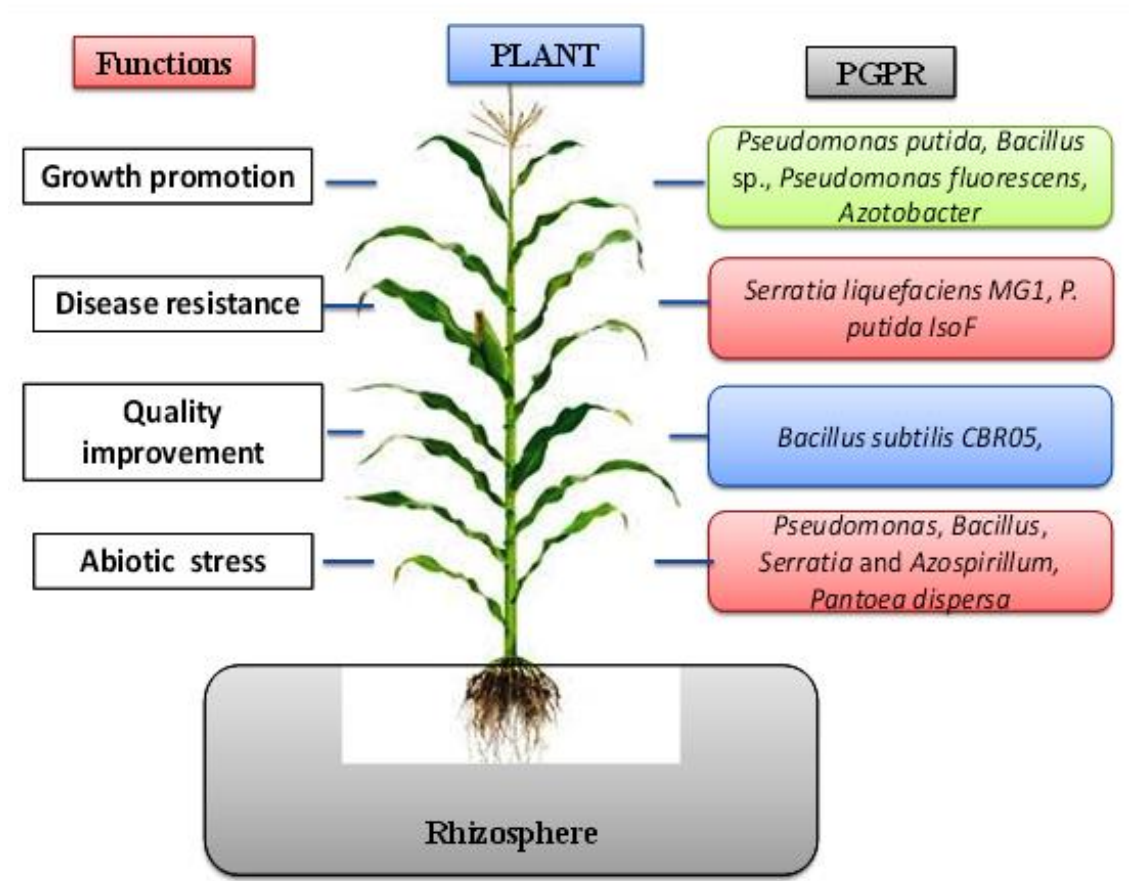


Figure 1. The beneficial influence of PGPR’s on crop plants

2. Global market for PGPR based biostimulants

Biostimulants are emerging as an essential component in sustainable agricultural practices. Instances of environmental hazards and soil contamination from injudicious and excessive application of chemical-based products on crops have been a key issue for the industry in recent times. The global biostimulants market size was estimated at USD 1.74 billion in 2016, projected to expand at a CAGR of 10.2% from 2017 to 2025. A rising focus on enhancing productivity, coupled with rapid soil degradation, is likely to drive the market over the forecast period. The **global biostimulants market** size was estimated at USD 2.30 billion in 2019 and is expected to reach USD 2.53 billion in 2020. The **global biostimulants market** is expected to grow at a compound annual growth rate of 10.2% from 2017 to 2025 to reach USD 4.14 billion by 2025 [27]. Although all biostimulants are not biological in nature [28], the bacteria are ancestral companions of a plant in all conditions. Moreover, according to the presently available literature, less than 25% of the commercial products of biostimulants are microbial-based [9]. As shown in Table 1 provides a list of some popular PGPR-based commercial biostimulants [29-31]. Although, some formulations contain fungal associations also, however, the preparations are mainly based on PGPR.

Table 1. Examples of commercial PGPR-based plant biostimulants [29-31].

Commercial Products (Manufacturer)	PGPR strains	Target crops for use	Target of function
FZB24® Rhizovital 42® (ABiTEP GmbH, Germany)	<i>Bacillus amyloliquefaciens</i> , <i>B. amyloliquefaciens</i> sp. <i>plantarum</i>	Ornamentals, vegetable Field crops	Phosphate availability and protection against pathogens
Inomix® Biostimulant; Inomix® phosphore; and Inomix® Biofertilisant (IAB (Iabiotec), Spain)	<i>B. subtilis</i> (IAB/BS/F1), <i>B. polymyxa</i> (IAB/BP/01); <i>Saccharomyces cerevisiae</i> <i>B. megaterium</i> , <i>P. fluorescens</i> ; and <i>Rhizobium leguminosarum</i> , <i>Azotobacter vinelandii</i> , <i>B. megaterium</i> , <i>Saccharomyces cerevisiae</i>	Cereals	Plant growth promotion increases root and shoot weight, strong root system
BactoFil B10® (AGRO. bio Hungary Kft., Hungary)	<i>Azotobacter vinelandii</i> , <i>Azospirillum lipoferum</i> , <i>P. fluorescens</i> , <i>B. circulans</i> , <i>B. megaterium</i> , <i>B. subtilis</i>	Dicotyledons (potato, sunflower, rapeseed)	Soil amelioration, produce plant growth-promoting hormones auxin, gibberellins and kinetin, N <sub>2</sub> fixation, a biocontrol agent
Bio-Gold (BioPower, Sri Lanka)	<i>Pseudomonas fluorescens</i> and <i>Azotobacter chroococcum</i>	All agricultural and horticultural crops	Growth promotion via nitrogen fixation, drought tolerance, control of root rot and wilt diseases, phosphorus solubilization
Cedomon® (Lantmannen BioAgri AB, Sweden)	<i>P. chlororaphis</i>	Barley and Oats	Highly effective against various types of seed-borne diseases
Rhizosum N Liquid PSA (Mapleton Agri Biotec Pty Limited, Australia)	<i>Azotoformans</i> (N <sub>2</sub> -fixing bacteria), <i>Pseudomonas</i> sp	Wheat	Phosphate availability, N <sub>2</sub> fixation, plant growth promotion
BactoFil A10® (AGRO. bio Hungary Kft., Hungary)	<i>Azotobacter vinelandii</i> , <i>Azospirillum brasilense</i> , <i>P. fluorescens</i> , <i>B. polymyxa</i> , <i>B. megaterium</i> <i>Agrobacterium radiobacter</i>	Monocotyledons (cereals)	Increased soil nutrient content that results in plant growth promotion
Micosat F® Uno; Micosat F® Cereali (CCS Aosta Srl, Italy)	AR 39, <i>Streptomyces</i> sp. SB 14, <i>B. subtilis</i> BA 41; <i>Paenibacillus durus</i> PD 76, <i>B. subtilis</i> BR 62, <i>Streptomyces</i> spp. ST 60	Fruits, vegetables, flowers Cereals, soybeans, beet, tomatoes, sunflowers	Increased nutrient and water absorption, increases stress tolerance and enhances ISR

Bioscrop BT16 (Motivos Campestres, Portugol)	<i>Bacillus thuringiensis</i> var. kurstaki	Deciduous fruit trees, horticultural brassicas, cotton, citrus, cauliflower, olives, pepper, banana, and tomato	Protection against pests ( beetles)
Amase® (Lantmannen Bioagri, Sweden)	<i>Rhizobium</i> , <i>Azotobacter</i> , <i>Pseudomonas</i> , <i>Bacillus</i> , <i>Chaetomium</i>	Cucumber, lettuce, tomato, pepper, eggplant, cabbage, and broccoli	Growth promotion, quick production of the large and strong root system, and increases stress tolerance
PGA® (Organica technologies, USA)	<i>Bacillus</i> sp.	Fruits, vegetables	Improved biomass accumulation, stress tolerance
Nitroguard®	<i>Azorhizobium caulinodens</i> NAB38, <i>Azospirillum</i> <i>brasiliense</i> NAB317, <i>Azoarcus indigens</i> NAB04, <i>Bacillus</i> sp.	Cereals, seed rape, sugar	Growth promotion via nitrogen fixation
TwinN® (Mapleton Agri Biotec Pty Ltd. Australia)	<i>Azospirillum brasiliense</i> NAB317, <i>Azoarcus</i> <i>indigens</i> NAB04, <i>A.</i> <i>caulinodens</i> NAB38,	Beet, sugarcane, vegetables	Helps in nitrogen fixation, phosphorus solubilization and produces growth- promoting hormones
Symbion®-N; Symbion®-P; and Symbion®-K (T. Stanes & Company Ltd, India)	<i>Rhizobium</i> , <i>Azotobacter</i> , <i>Azospirillum</i> , <i>Acetobacter</i> ; <i>B. megaterium</i> var. <i>phosphaticum</i> ; and <i>Frateuria aurantia</i>		Promotion of plant growth improved root and shoot weight, the more strong root system
Ceres® (Biovitis, France)	<i>Pseudomonas fluorescens</i>	Field and horticultural crops	Biocontrol agent against pathogens
Gmax® PGPR (Greenmax AgroTech, India)	<i>P. fluorescens</i> , <i>Azotobacter</i> , <i>Phosphobacteria</i>	Field crops	Nitrogen and phosphatic nutrition, disease prevention and helps in plant growth promotion.

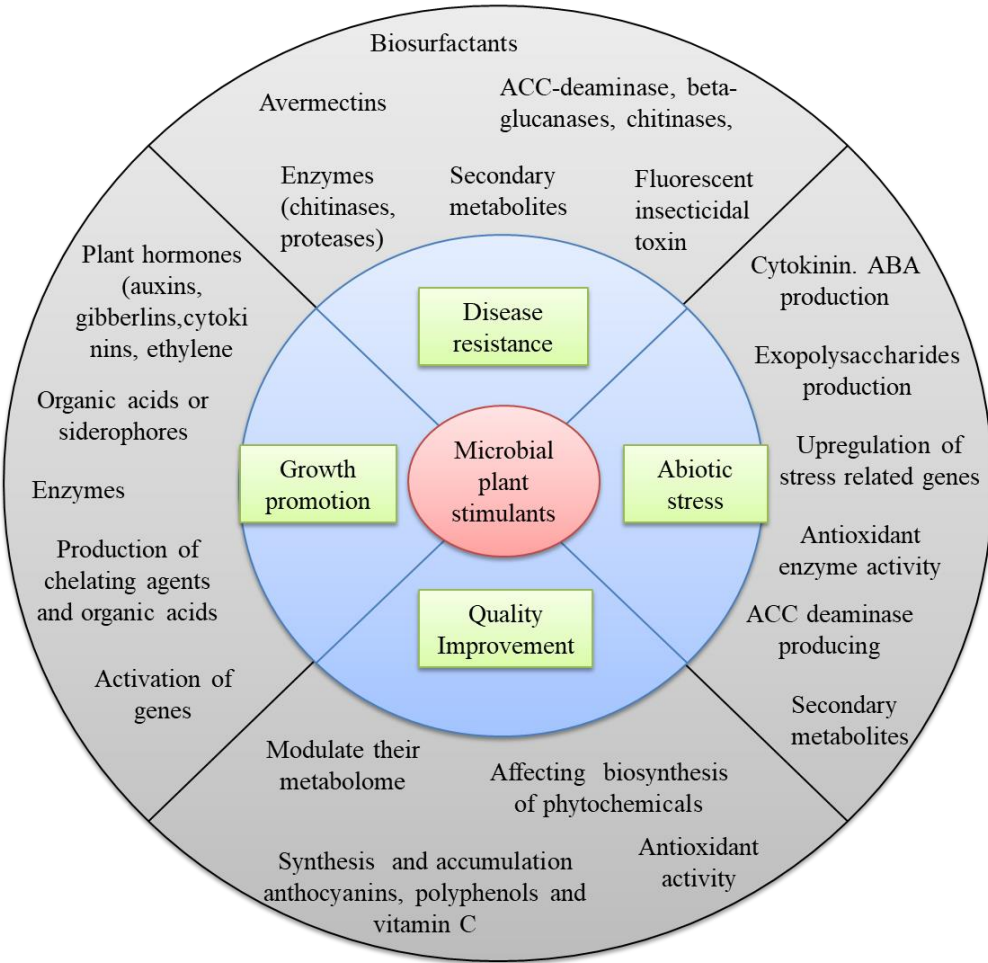
### 3. Bacterial plant biostimulants, beneficial effects, and mode of action

Bacteria are known to interact with the plants in all the possible ways [32] including i) continuum of symbiosis, ii) bacteria niches extending from substrate to interior of cells,



which are called intermediate locations for rhizosphere and rhizoplane, iii) associations that are transient or lifelong and iv) functions that affect lots include engagement in biogeochemical cycles, the supply of nutrients, increased nutrient consumer efficiency, induction of resistance, increased stress tolerance, plant growth regulators and morphogenesis control. In this regard, a large amount of work presented in recent literature has its sharp emphasis on potential applications of the bacterial association of plants largely as agents for promoting plant growth and maintaining soil and crop health [33-36]. The plant growth-promoting bacteria are generally associated with numerous (if not all) crop plant species and are habitually present in varied environments. The most extensively investigated category of PGPB is the plant growth-promoting rhizobacteria (PGPR) primarily colonizing the surfaces of roots and closely adhering to the soil interface namely, the rhizosphere. As overviewed by recent reviews [37-39], several PGPR can enter the root interior thereby establishing endophytic associations. Some of them can even surpass the endodermis barrier, transcending from root cortex to vascular system, and afterward thrive as the endophytes (inside stem, tubers, leaves, and other organs). The extent of the endophytic associations of host plant tissues (and/or organs) reflects the capability of these bacteria to selectively acclimatize to various specific ecological-niches [40-41]. As a result, such intimate bacterial associations with host plants are formed with no damage to the plant [42-43]. In regards to taxonomic, functional, and ecological diversity in developing agriculture biostimulants, the PGPR seize the most prominent place.

While numerous soil bacteria were documented to help plant growth promotion and production, it is hardly understood the mode(s) of action by which the bacteria exhibit beneficial activities. The molecular basis for association processes between bacteria and crop plants that induce/stimulate physiological modifications is starting to be understood, primarily because of the emerging approaches to "omics." A varied number of pathways have been employed to aid the acquisition of plant nutrients, including improved plant root surface, phosphorus solubilization, nitrogen fixation, production of HCN, and development of siderophores which are further discussed under subsections [44]. PGPR differ and has consequences for all facets of the plant life cycle: promoting growth and nutraceutical values of plants, morphological and physiological development, stress responses (biotic and abiotic), interactions of agro-ecosystem with other species forms, and enhanced production. Numerous direct and indirect mechanisms are involved in the development of these responses that are shown in Figure 2.



**Figure 2.** Mode of action of PGPR on the growth of crop plants

**3.1. Plant growth promotion and nutrient acquisition**

The modulation of bacterial behavior has tremendous potential in the procurement of nutrition for plants. PGPR formulations are a significant biostimulant class, as they allow root growth, mineral availability, and efficiencies in the utilization of nutrients in crop rhizosphere to increase crop growth [45]. Many PGPR is known to stimulate phytohormone production through a combination of various mechanisms [46-53] represented in Table 2.

*3.1.1. Phytohormone stimulation*

Auxins such as IAA are involved in the processes such as germination of seeds, control processes for the vegetative increase, establishment of lateral or adventurous roots and can mediate light and heavy reaction, photosynthesis biosynthesis of metabolites, and stress tolerance [54]. It has been observed that PGPR produces hormones that provide protection and wall-related transcription changes [55]; induce long roots, increases



the biomass of roots, reduce the density and dimensions of stomata [56]; besides activating auxin reaction genes which enhance plant development [21]. As IAA producers, separate PGPR genera have been recognized, such as *Rhizobium* [57], *Aeromonas* and *Azotobacter* [32], *Bacillus* [21], and *Pseudomonas* [50]. A great number of PGPRs produce cytokinins and gibberellins [59], though the roles of bacteria in the regulation of plant hormones and the bacterial mechanism involved in their synthesis are largely not understood yet. Some strains of PGPR can support relatively large quantities of gibberellins, which contribute to increased growth in plants [60]. The PGPR also regulates proper amounts of ethylene to maintain plant growth as confirmed by previous studies [61].

### 3.1.2. Nitrogen

Nitrogen (N) is a very essential macronutrient needed for plant growth and development, but it is not available to most plants due to its inertness. Atmospheric nitrogen ( $N_2$ ) gets converted into ammonia by PGPR by nitrogen fixation and this source of nitrogen (ammonia) can be utilized by crop plants for productivity purposes [62]. The application of  $N_2$ -fixing bacteria as growth enhancers has become known as one of the effective and environmentally feasible methods and concurrently replacing the use of inorganic nitrogen fertilizers [63]. Biological Nitrogen Fixation (BNF) is accomplished by free-living microorganisms like *Azotobacter*, *Azospirillum*, *Bacillus*, *Enterobacter*, *Pseudomonas*, *Burkholderia*, and *Serratia*, etc. and symbiotic or associated microorganisms like *Rhizobium*, *Bradyrhizobium*, and certain species of *Azospirillum* sp. which contribute fixed nitrogen for the associated crop plants [64-65]. Moreover, a small group of woody, non-legume, known as actinorhizal plants, can also be colonized by Diazotrophs belonging to *Frankia* sp. which can induce the development of nitrogen-fixing root nodules. Leguminous inoculants are the first example of industrial bacterial products in agriculture and now the most commonly used inoculants in agriculture [66]. Beginning in the early 21st century, the interest began rising around the mass production of the commercial inoculants from wild, live N-fixing bacteria, including *Azoarcus* sp., *Burkholderia* sp., *Gluconacetobacter* sp., *Diazotrophicus* sp. These free-living diazotrophs are more efficient in providing N to a wider variety of crops than rhizobia. *Azospirillum* sp. based commercial inoculants from small and medium-sized businesses worldwide have improved the production yields of different cereal crops effectively [67]. Other bacteria which do not primarily fix  $N_2$  have also shown increased N in many plants and possibly due to root growth enhancement, allowing plants to gain more soil [68] thus, increase the efficiency of nitrogen usage.

### 3.1.3. Phosphorus

Phosphorus is another essential macronutrient in metabolic and physiological processes in plants such as photosynthesis, biological oxidation, and cell division [69],

and is also an important nutrient for crop growth and productivity. Chemical phosphorus fertilizers are subjected to chemical fixation (in soil) with some other metal cations and are lost by leaching and their unavailability to plants limits their ability to perform these crucial functions [70]. The application of stimulants that contain PGPR that are capable of solubilizing insoluble phosphate by discharging organic acids increases the accessibility of this element to crop plants thereby improving soil fertility and productivity [71-72]. Numerous strains among bacterial genera including *Pseudomonas*, *Rhizobium*, *Bacillus*, and *Enterobacter* are the most potent P-solubilizers. The phosphorus solubilizing bacteria (PSB) may facilitate plants' access to the non-labile phosphorus reserve by liberation of its recalcitrant form and making it more accessible to the crops by secreting organic acids and/or HC ions. Likewise, PSB-manufactured phytase can release reactive phosphates from organic compounds [73].

#### 3.1.4. Potassium

Potassium is another essentially important macronutrient required for crop growth and improvement, owing to its rhizospheric deficiency and consequently, has always been a major constraint in crop production [74-75]. The shortage of solubilized form of rhizospheric potassium is also because it tends to form insoluble complexes when applied as inorganic fertilizer. However, PGPR can solubilize the insoluble potassium through the secretions of inorganic acids and making it available to the crop plants thus improving agricultural productivity and health of crops [76-77]. Hence, they offer an attractive option as biostimulators in place of conventional fertilizers. PGPR, like *Bacillus edaphicus*, *Acidithiobacillus* sp., *Ferrooxidans* sp., *Pseudomonas* sp., *Bacillus mucilaginosus*, *Burkholderia* sp. and *Paenibacillus* sp. have been known to release the potassium in its available form from the potassium-bearing minerals in soils [78].

#### 3.1.5. Micronutrients

Many strains of bacteria improve Fe (iron) availability by generating siderophores or organic acids. The commercial preparation of the genus *Acidithiobacillus ferrooxidans* developed and produced by AgriLife (India) [79] solubilizes the Fe through the release of organic acids [80]. Zinc is another crucial micronutrient that is needed in smaller quantities for healthy growth and improved production of the crops. About 96-99% of zinc applied to the crop plants is converted into an insoluble form that depends on soil type and other physiological reactions [81]. Several bacteria strains increase Zn mobilization, thereby increasing Zn uptake by plants and thus boost the yield in many crops [82]. Although the mechanisms involving Zn-mobilizers still remain uncertain, they are more likely similar to PSBs and Fe-mobilizers and involve mainly the production of organic acids and chelating agents.

**Table 2.** Beneficial effects of reported PGPR biostimulants on different crops and their modes of action.

PGPR biostimulant	Crop	Beneficial effects	Mode of action	References
<i>Bacillus</i> sp.	Lettuce	Growth, biomass, and yield of plants	Increased production of phytohormones and availability of nutrients	[46]
<i>Azospirillum brasilense</i> , <i>Gluconacetobacter diazotrophicus</i> , <i>Herbaspirillum seropedicae</i> , and <i>Burkholderia ambifaria</i>	Onion	Plant growth, crop yield, and increased number of bulbs	Production of plant hormone and solubilization of nutrients that causes uptake of nutrients	[47]
<i>Bacillus pumilus</i> , <i>B. mojavensis</i> , <i>B. amyloliquefaciens</i> , <i>P. putida</i> .	Tomato	Growth and production and nutrient uptake	Synthesis of indole-3-acetic acid $N_2$ -fixation and P solubilization	[48]
PGPR ( <i>Bacillus subtilis</i> )	Tomato	Improved fruit quality	Enhanced production of phenols, flavonoids, carotenoids, and antioxidants	[49]
<i>Pseudomonas aeruginosa</i>	Wheat	Nutrient uptake	$N_2$ fixation involving many reactions and synthesis of organic acids	[50]
<i>Azospirillum brasilense</i> (Sp7b and Sp245b)	Cucumber, lettuce, and tomato	Enhanced germination, root length, and weight, vigor index of germinating seeds	Production of a substantial amount of phytohormones such as IAA	[51]
<i>Bacillus pumilus</i> , <i>Pseudomonas pseudoalcaligenes</i>	Rice	Stimulated growth and production	Phosphate solubilization, production of IAA,	[52]

<i>Azospirillum brasiliense</i>	Maize, sorghum, wheat, barley, legumes	Bio-stimulated growth and production	gibberellins, siderophores, and ACC utilization		[53]
			Synthesis of indoleacetic acid (IAA), nitric oxide, carotenoids, and numerous cell surface components		

3.2. Quality improvement of crop and yield by bacterial plant biostimulants

Plant biostimulants, which increase plant evolution, flowering, fruit forming, crop production, can provide a desirable and environmentally friendly agricultural modernization [83]. A variety of living and non-living bacterial isolates like *Bacillus licheniformis*, *Bacillus megaterium*, *Bacillus pumilus*, *Bacillus safensis*, *Microbacterium* sp., *Nocardia globerula*, *Pseudomonas fluorescens*, *Pseudomonas fulva*, *Pseudoxanthomonas dajeonensis*, *Rhodococcus coprophilus*, *Lactobacillus plantarum*, *Sphingopyxis macrogoltabida*, *Streptomyces* sp., *Bifidobacterium bifidus*, *Lactobacillus acidophilus*, *Lactobacillus* sp., *Lactobacillus buchneri*, *Lactobacillus paraplantarum*, *Lactobacillus delbrueckii*, *Lactobacillus pentosus*, have been reported to increase concentrations of total carbohydrates, nutrients (magnesium, nitrogen, and phosphorus, etc.), pigments (like chlorophyll, carotenoids) and antioxidant substances and therefore, improve plant quality, productivity, and yield [21,83-84]. As, an example, the impact on bean common plants cultivated under water stress shows substantial enhancement in the phenolic contents of the inoculated plants of four biostimulant products having *Bacillus subtilis* in their formulations [84]. Also, by inoculation of PGPR strain, *Bacillus subtilis* CBR05, the quality of tomatoes is known to get better for carotenoid profile (carotene and lycopene) [49]. The influence of the bio-preparation containing some bacterial species like *Streptomyces* sp., *Bacillus subtilis*, and *Pseudomonas fluorescens* onto the growth enhancement of fruits through organic farming reported that the growth of sour cherry and apple trees was improved by these bio-preparations [85]. The regulation of horticultural primary and secondary metabolisms in microbial biostimulants, culminating in synthesis and build-up of lipophilic as well as hydrophilic antioxidant molecules also referred to as phytochemicals [86-87]. The microbial biostimulant applications containing beneficial bacterial cultures often improve fruit quality by suppressing diseases that may cause economic loss [88].

3.3. Abiotic stress tolerance induced by bacterial plant biostimulants

The global climate changes dictated that abiotic stresses, particularly, nutrient deficiency, salinity, drought, hypoxia, and heat stress are responsible for 60–70% of yield deficit, [14].

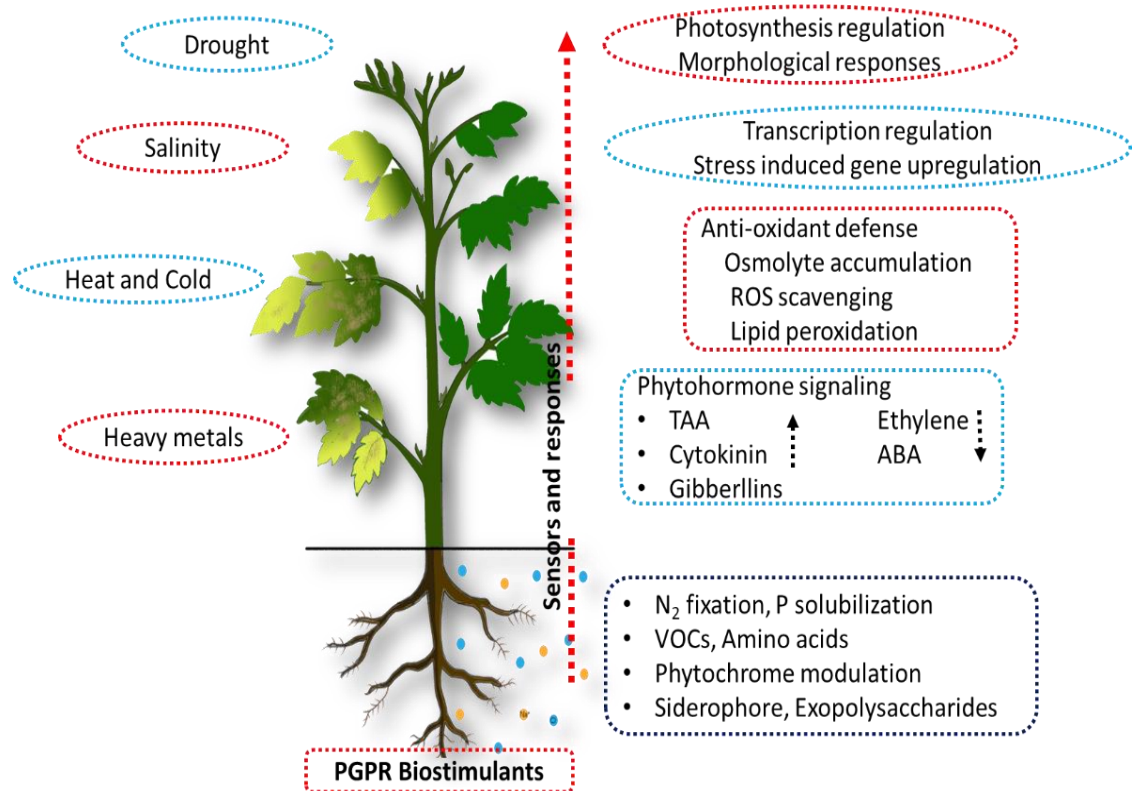
Under these situations plant, biostimulant application is suggested as an effective agronomic method to improve tolerance to adverse soil, harsh environmental conditions and to address the adverse effects of the suboptimal conditions on agricultural and horticulture crops [9]. Plant growth rhizobacteria (PGPRs) can enhance plant reactions to abiotic pressures (Figure 3), promote physical, chemical, and biological activities [89] through various mechanisms [90-100] as presented in Table 3. A voluminous work has been done on bacterial isolates that can be employed to promote the mitigation of abiotic stress in various crops.

#### *3.3.1. Drought stress*

Recent attention has turned to the application of beneficial microorganisms that mediate drought tolerance and improve plant water-use efficiency. These efforts have been augmented due to technological advances in next-generation sequencing and microbiomics [101-102]. The application of plant growth-promoting rhizobacteria (PGPR) is considered a sustainable synergistic biological approach to cope with water deficiency in crop production [103]. PGPR can impart tolerance to the drought stress by releasing phytohormones, volatile compounds, ACC deaminase, exopolysaccharides, and antioxidants, by the regulation of osmolytes and stress-responsive genes, and aggravating modifications in the roots [102-104].

#### *3.3.2. Salinity Stress*

Soil salinization accounts for above 6% of global land rendering 22% and 33% of total cultivated and irrigated agrarian land, respectively under stress that adversely affects crop productivity [105]. By the year 2050, approximately 50% of the arable area will be under threat due to soil salinity as it increases rapidly at the rate of 10% annually, because of numerous reasons including implausible irrigation practices, irrational fertilization, poor drainage, and climate change [106-107].



**Figure 3.** Illustration of abiotic stress tolerance induced by bacterial plant biostimulants

PGPR can alleviate salinity stress in plants by many synergistic mechanisms including osmotic regulation by prompting accumulation of osmolytes and signaling of phytohormones, increasing nutrient uptake and attaining homeostasis of ions, and reducing oxidative stress through enhanced antioxidant activity, volatile organic compounds (VOCs), and photosynthesis amelioration [108-109].

### 3.3.3. Heat stress

The prime alarming effect of climate change is the rise in global temperature and is directly linked to crop productivity. High temperatures increase respiration and transpiration rates, alter the allocation of photosynthates and affect photosynthesis (particularly C<sub>3</sub> plants) thereby, influencing plant physiology [110]. Intense heat can cause plant cell protein denaturation or affect cell wall and membrane permeability [111]. PGPR help in mitigating the heat stress in plants through their properties like the production of osmolytes and reduction of carbon flux [112]. They can secrete several polysaccharides involved in biofilm formation covering root nodules that enhance the water retaining capability of plant roots. PGPR especially, the heat-tolerant/evolved strains possess the ability of enhanced the production of lipopolysaccharides (LPS) and exopolysaccharides (EPS) and specific proteins known as heat shock proteins (HSPs) [113]. The application of ethylene reducing bacteria especially, with ACC deaminase activity can avoid the detrimental effects of heat stress in plants [3].



#### 3.3.4. Cold stress

Cold stress is detrimental to plants as it directly affects the rate of nutrient and water uptake, which may lead to cell starvation, desiccation, and consequent death. Reduced metabolism, which occurs in cold tension, results in a photo-inhibition, inhibition of the activity of the photosystem II and destabilization of the phosphorus lipid bilayers thereby, affecting the normal architecture of cell membranes [44,114]. In harsh environments, psychrophilic (cold-adapted) microorganisms can thrive and have possible resistance enhancement pathways that benefit plants [115]. Cold-adapted PGPR belongs to various genera including *Pseudomonas*, *Bacillus*, *Arthrobacter*, *Exiguobacterium*, *Paenibacillus*, *Providencia*, and *Serratia*. There are several attributes of psychrotolerant PGPR that make their application as biostimulants beneficial in alleviating cold stress. These attributes include solubilization of nutrients, Fe-chelating compounds, ACC deaminase production, IAA, and bioactive compounds. In plants, cold tolerance can be imparted by PGPR through the enhanced accumulation of carbohydrates, regulation of stress-responsive genes for modulation of osmolytes, and increasing specific protein including cold shock proteins (CSP) [113]. Also, the application of such biostimulants having the ability to outcompete the ice-nucleating activity of microorganisms is becoming an effective method to overcome the losses caused by cold/frost damages [3].

#### 3.3.5. Heavy metal stress

Heavy metal stress due to hyper-accumulation of toxic metals including Hg, As, Cd, Pb, and Al greatly decrease crop productivity. Their accumulation in the soil directly affects its texture and pH which consequently reduces the crop growth by exerting negative effects on several biological processes [116]. In plants, heavy metal stress shows both direct effects including, cytoplasmic enzyme inhibitions and cell structure damages as well as indirect consequences including, oxidative stress by several indirect mechanisms (e.g. glutathione depletion or binding to protein's -SH groups) or inhibiting anti-oxidative enzymes, inducing ROS producing enzymes (e.g. NADPH oxidases) [117]. Heavy metal tolerant PGPRs such as *Pseudomonas*, *Bacillus*, *Methylobacterium*, *Streptomyces* can reduce the deleterious effects of heavy metals and improve the growth and yield of crops. PGPR biostimulants are very effective in alleviating the toxicity of heavy metals in plants. They reduce the translocation of heavy metals to different parts of the plant by altering their mobilization through chelation, precipitation, complexation, redox reactions, and adsorption [118-119]. Rhizospheric bacteria also release extracellular polymeric substances (EPS) [93] such as polysaccharides, glycoprotein, lipopolysaccharide, and soluble peptide which possess a substantial quantity of anion binding sites to help in the removal or recovery of heavy metals from the rhizosphere via biosorption. However, in highly contaminated sites, the mobilization and consequent bioavailability of heavy metals in excess, by siderophores, organic acids, or through bioleaching remains debatable.

**Table 3.** Influence of PGPR biostimulants on abiotic stress tolerance in various crop plants.

PGPR biostimulants	Crop plants	Type of abiotic stress	Mode of Action	References
<i>Glutamicibacter</i> sp. YD01	Rice	Salt tolerance	Ethylene mediation, ROS accumulation, maintaining photosynthetic efficiency and ion homeostasis, increasing expression of stress-related genes, the activity of ACC oxidase, and acquisition of K <sup>+</sup>	[90]
<i>Bacillus</i> sp., <i>Azospirillum lipoferum</i> , <i>Azospirillum brasilense</i> and <i>Pseudomonas stutzeri</i>	Wheat	Salt stress	Production of phytohormones, osmoregulators, and enzymes (ROS scavenging) activation	[91]
<i>Gluconacetobacter diazotrophicus</i> Pal5	Red rice	Drought stress alleviation	Increased production of ABA, osmoprotectants (proline and glycine betaine) and AHLs	[92]
<i>Gluconacetobacter diazotrophicus</i> Pal5	Red rice	Water stress alleviation	Increased ABA production, enhanced chlorophyll synthesis, increased trehalose, and $\alpha$ -tocopherol content in roots.	[93]
<i>Azospirillum</i> spp. (Az19)	Maize	Water/drought stress alleviation	Increased production of proline, trehalose (Glutamate) and Glycine-betaine	[94]

<i>Bacillus</i> spp XT13, XT38, and XT110	Maize	Drought stress	Increased proline content accompanied by reduced APX and GR activities; increased nutrient uptake	[95]
<i>Pseudomonas entomophila</i> (PE3)	Sunflower	Salinity stress alleviation	Exopolysaccharides, IAA, gibberellic acid, and siderophores	[96]
<i>P. fragi</i> , <i>P. proteolytica</i> , <i>P. fluorescens</i> , <i>P. chloropaphis</i> , <i>Brevibacterium frigoritolerans</i>	Bean	Cold stress	Reduced chill injury, lipid peroxidation, and ice-nucleating activity corresponding to ROS level, stimulation of apoplastic antioxidant enzyme activities	[97]
<i>Pseudochrobactrum kiredjianiae</i>	Wheat	Cold stress	Growth promotion, biocontrol	[98]
<i>Pseudomonas fluorescens</i>	Maize	Heavy metal stress	Production of IAA	[99]
<i>Azotobacter chroococcum</i>	Maize	Heavy metal stress	Production of siderophores, ammonia, and ACCD	[100]

### 3.5. Disease suppression/defense against plant pathogens through antagonism

Nowadays, the biological control of pathogens is managed by the activities of several microbiomes. Additionally, PGPR are known to develop resistance to various diseases through various direct or indirect mechanisms [120-128] shown in Table 4. The application of bacterial biostimulants encourages the healthy growth of crops by suppression of different plant pathogens and pests. The PGPR inhibition of microbial/pathogen growth occurs synergistically by several chief mechanisms including antibiosis, volatile organic compound (VOC) production, extracellular enzymatic lysis, bacteriocin, and siderophore mediated inhibition [129].

#### 3.5.1. Antibiosis

PGPR produce antibiotics that are the most significant antagonistic agents effective against phytopathogens. Antibiotics produced by PGPR are known to have antimicrobial, antiviral, cytotoxic, insecticidal, antihelminthic, and phytotoxic (against weeds) effects [130-131]. Antibiotic production usually allows better competition between microbes and thus enhances the efficiency of beneficial PGPR associations [132]. Numerous species of *Pseudomonas* produce a broad range of antifungal antibiotics including butyrolactones, cepaciamide A, ecomycins, 2,4-diacetylphloroglucinol (2,4-DAPG), phenazines, pyrrolnitrin, pyocyanin, pyoluteorin, oomycin A, rhamnolipids, N-butylbenzene sulfonamide, and viscosinamide [133]. *Bacillus* species also secrete a large variety of antibiotics including bacilysin, bacillaene, difficidin, mycobacillin, rhizoctins, sublancin, subtilint A, subtilosin A, etc. They also produce numerous lipopeptide biosurfactants, such as bacillomycin, iturins, surfactin, etc. having antibiotic activity [134].

### 3.5.2. VOC antagonism

In plants, VOCs help in the biocontrol of bacteria, fungi nematodes and also act as elicitors of the induced systemic resistance against phytopathogens [135]. Several VOC metabolites having antagonistic activities are secreted by PGPR. These include benzene, cyclohexane, 2-(benzyloxy)-1-ethanamine, methyl, dodecane, decane, 1-(N-phenyl carbamyl)-2-morpholinocyclohexene, benzene (1-methylnonadecyl), dotriacontane, 1-chlorooctadecane, tetradecane, and 11-decyldocosane although, their type and quantities released varies among different species [136]. Among VOCs, HCN produced by rhizospheric bacteria is known to play an important function in the biocontrol of phytopathogens and pests [137]. *Pseudomonas* sp. synthesizing HCN can inhibit some pathogenic fungi [138]. HCN released by *P. chlororaphis* O6 is known to show nematicidal activity [139]. Also, VOCs (acetoin and 2,3-Butanediol) secreted by *Bacillus* spp. are very effective fungal inhibitors [140]. In addition to biological control, VOCs are associated with beneficial tradeoffs in attracting pollinators via the mediation of communication signals [141].

### 3.5.3. Lysis by extracellular enzymes

Lytic enzymes produced by PGPR provide another effective mechanism for combating pathogen attack. Rhizobacteria release extracellular enzymes like chitinase and  $\beta$ -1,3-glucanase which are involved in cell wall lysis, killing pathogens [142]. Since the fungal cell wall is mainly composed of chitin and  $\beta$ -1,4-N-acetyl-glucosamine, rhizobacteria secreting chitinase and  $\beta$ -1,3-glucanase are potent antifungals. For example, *P. fluorescens* LPK2 and *S. fredii* KCC5 release  $\beta$ -glucanases and chitinases and suppress wilts caused by *Fusarium udum* and *F. oxysporum* [133]. Bacteria with protease, lipase, and chitinolytic activities have been reported to show insecticidal activity [143]. PGPR having ACC deaminase activity also plays a very important role in all types of stresses including biocontrol.

#### 3.5.4. Bacteriocins

Bacteriocins or bacterial toxins are narrow-spectrum antimicrobial peptides produced by bacteria including PGPR. Their production is another mechanism for eliminating competitor strains which are narrow-spectrum, proteinaceous antibiotics that target and kill related bacterial species [144]. Bacteriocins are produced by both Gram –negative (colicins, S-piocins, microcins, etc) as well as Gram –positive (nisin, helvecin, mersadecin, etc) bacteria [145]. Direct application of bacteriocins has shown promising results under laboratory conditions against bacterial spot disease of tomato [146]. Typically, bacteriocins are highly selective towards their targets without affecting off-targets and provide a safer substitute to field applications of chemicals [147].

#### 3.5.5. Siderophores

Siderophores are the largest class of known compounds that can bind and transport, or shuttle, iron (Fe). These low-molecular-weight coordination molecules are excreted by a wide variety of fungi and bacteria to aid Fe assimilation [148]. Siderophore production by PGPR is an indirect mechanism involving the reduction or prevention of destructive effects caused by phytopathogens [149]. Siderophores possess an antagonistic effect and prevent the escalation of other pathogenic bacteria and fungi in the plant's rhizosphere [150]. Their low molecular weight and ability to sequester Fe<sup>3+</sup> ions in the rhizospheric zone makes iron inaccessible to the plant pathogens thus, preventing their growth.

**Table 4.** Influence of PGPR biostimulants on biotic stress resistance in different crop plants.

PGPR Biostim- ulants	Crop	Biotic stress	Mode of action	References
<i>Bacillus cereus</i> (PX35), <i>Serratia</i> sp. XY21 and <i>Bacillus subtilis</i> SM21	Tomato	Root-knot nematodes	Synergistic biocontrol	[120]
<i>Pseudomonas aeruginosa</i> LV	Tomato	Bacterial stem rot	Extracellular-bioactive compounds (phytoalexins, flavonoids, defensins, proteins, and phenolics)	[121]
<i>B. subtilis</i> 26DCryChS	Potato	Late blight agent and damaged by Colorado	Production of Cry1Ia $\delta$ -endotoxin, stimulating transcription of jasmonate reliant genes	[122]

		potato beetle larvae	promoting transcription of salicylate reliant gene (PR1)	
<i>Lactobacillus plantarum</i> PM411 and <i>Lactobacillus plantarum</i> TC92	Strawberry	Disease prevention in strawberry and kiwi fruit	Antimicrobial metabolites (lactic acid) production that disrupts pathogen's cell membranes	[123]
<i>B. subtilis</i> BS2	Tomato	Tomato wilt	Production of defense enzymes such as peroxidase, polyphenol oxidase, chitinase, and phenylalanine IAA production	[124]
<i>Bacillus safensis</i> , <i>Bacillus altitudinis</i> <i>B. velezensis</i> , <i>B. mojavensis</i> and <i>B. safensis</i>	Cabbage	Black rot		[125]
<i>Bacillus cereu</i> , <i>B. subtilis</i> BSV <i>B. subtilis</i> BSP	Soybean	Phytophthora root rot	IAA production	[126]
	Ginger	Blister blight	1-aminocyclopropane,1, carboxylic acid production	[127]
<i>B. cepacia</i> GRB35	Ginger	Soft rot in ginger	Fungicide production	[128]

3.6. Induction of systemic resistance (ISR)

The first line of the defense system of plants is comprised of a precise surveillance system that by perceiving several elicitors, allows them to switch on plant defense mode and reject potentially dangerous pathogens or microbes. The elicitors are small structures referred to as pathogen/microbe-associated molecular patterns (PAMPs or MAMPs) which are recognized by the pattern recognition receptors (PRRs) of the plant's innate immune systems [151]. Similar to this innate mechanism, PGPR are also capable of stimulating the defense system of their associated plants against pathogen attack through induction of systemic resistance (ISR) by SAR (system acquired resistance) and ISR (induced systemic resistance) pathways [152]. Furthermore, PGPR can be exploited for stimulation of induced systemic tolerance (IST) against various abiotic stresses including water scarcity, drought, salinity, osmolyte stress, temperature extremes,



heavy-metal stress, and mechanical injuries, etc are some examples of stresses [153]. Therefore, the application of multi-stress-resistant PGPRs biostimulants has become important to enhance agricultural production, to resolve global climate change concerns, to low annual crop yields, and to combat increasing food demands [13].

### 3.6.1. Systemic Acquired Resistance (SAR)

Systemic acquired resistance (SAR) is a mechanism of induced defense that confers long-lasting protection against a broad spectrum of microorganisms. It is an induced immune mechanism found in plants having a broad spectrum that is not specific to the initial infection [154] and can be systematically expressed in all organs [155]. SAR requires the salicylic acid (SA) signaling that accumulates within the infected plant tissues after pathogen attack, which stimulates immune responses such as pathogenesis-related (PR) gene expression and antimicrobial substance encoding [156]. The SA signal transduction requires activation of PR (pathogenesis-related) genes of which NPR1 regulatory (activator) protein is an essential gene that operates within the terminal of the SAR signal pathway [157].

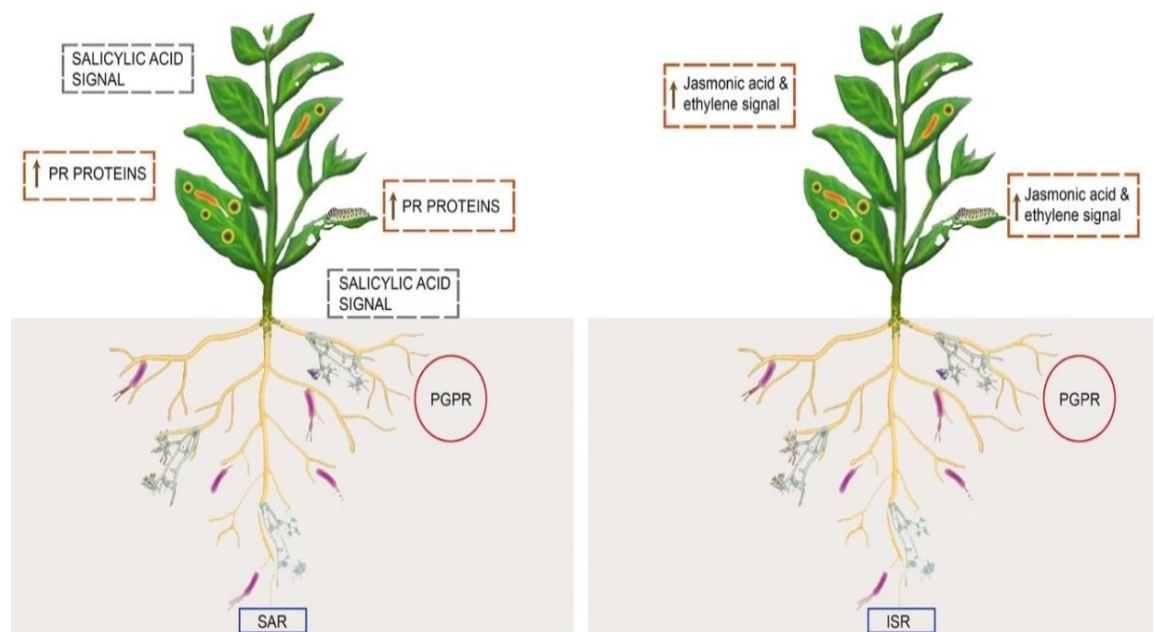
SAR is generally activated by pathogens or chemical stimuli however, some PGPR are also known to trigger the SA (salicylic acid)-dependent pathway through the production of SA at the root surface [158]. Treatment of tomato plants with *Bacillus amyloliquefaciens* (strain MBI600) which is an active component of a fungicide, Serifel® was shown to produce antiviral action against *Potato virus Y* (PVY) and *Tomato spotted wilt virus* (TSWS) in tomato plants through SA-dependent signaling pathway [159]. In another example, leaf infiltration with *Bacillus cereus* (AR156), a PGPR has been reported to enhance disease resistance against *Pst* (*P. syringae* pv. tomato) in Arabidopsis through the activation of a SAR pathway [160]. However, the salicylic acid released by rhizobacteria does not need to necessarily mediate the SAR mechanism as SA produced by rhizobacteria may require siderophores for their assimilation [161].

### 3.6.2. Induced Systemic Resistance (ISR)

Induced systemic resistance (ISR) emerged as an important mechanism by which selected plant growth-promoting bacteria and fungi in the rhizosphere for enhanced defense against a broad range of pathogens and insect herbivores [162]. ISR induction requires components of the jasmonic acid (JA) signaling pathway followed by the ethylene signaling pathway [163]. For many biological control agents, ISR has been recognized as the mechanism that at least partly explains disease suppression. It is of significant importance from an agronomic perspective for its effectiveness against a wide range of microbial pathogens, nematodes, and insects that damage crops [164-165]. It was reported that the PGPR *Bacillus amyloliquefaciens* induces systemic resistance in bean plants against the aphids through the production of higher contents of Jasmonic acid [166]. The attack

of insect herbivores on the plant roots and leaves imposes different selection pressures on plants, which in turn produces contrasting responses in terms of gene expression and production of secondary metabolites and wound hormones [167]. PGPR-triggered ISR does not involve severe defense-related gene changes and assists the plant in the induction of resistance against various pathogens by the production of several extracellular metabolites that act as elicitors [153]. Several PGPR metabolites such as N-Acyl-Homoserine Lactones [168], siderophores [169], VOCs [170], rhamnolipids [171] and cyclic lipopeptides [172], etc. However, most of these elicitors have been identified from strains of *Bacillus* and *Pseudomonas* sp. and elicitors from many other species remain mostly undiscovered.

These elicitors require higher  $\mu\text{M}$  concentrations to activate the immune responses as compared to the MAMPs indicating that they may not be sensed through high-affinity receptors [173]. Quorum sensing molecules such as acyl-homoserine lactones produced by PGPR represent novel elicitors of biotic stress resistance in plants. In a recent study, a halotolerant plant growth-promoting bacterium, *Staphylococcus equorum* EN21 triggered ISR against *Pseudomonas syringae* (pv. Tomato) through quorum quenching of acyl-homoserine in Arabidopsis and tomato plants [174]. ISR activity of the elicitor, oxo-C14-HSL was observed in tomato and wheat against *Phytophthora infestans* and *Puccinia graminis* f., respectively [175]. In monocots (such as rice) cyclic lipopeptides released by *Pseudomonas* are crucial in eliciting the ISR. For example, cyclic lipopeptides such as lokisin, endolysin, and WLIP were described recently to successfully induce resistance against *Magnaporthe oryzae* [176] while orfamide (at 25  $\mu\text{M}$  concentration) was known as an elicitor of ISR against *Cochliobolus miyabeanus* [177]. Accumulation of ROS following the inoculation of bacteria *Gluconacetobacter diazotrophicus* has also been observed at the early stages of rice root colonization. The study indicates that with the help of bacterial ROS-scavenging enzymes, glutathione reductase and superoxide dismutase, triggering a typical ISR plant defense response against pathogens [178].



**Figure 4.** Types of induced resistance in plants by bacterial biostimulants.

### 3.6.3. Induced Systemic Tolerance (IST)

Similar to ISR against biotic stresses, the defense responses induced by different PGPR to withstand abiotic stresses generally involve highly-regulated mechanisms including regulation of phytohormones, ROS accumulation, EPS (exopolysaccharides) production, ACC-deaminase activity, secretion of secondary metabolites, VOCs, antioxidant machinery, and activation of defense-related genes that lead to induced systemic tolerance (IST) and has been well documented by [153]. Such responses also involve a web of highly coordinated plant hormones like abscisic acid (ABA), gibberellins (GA), ethylene (ET), auxins (indole acetic acid, IAA), cytokinins (CK), jasmonic acid (JA), salicylic acid (SA) and brassinosteroids (BRs). These plant hormones habitually act as the key signaling molecules triggering intricate signaling cascades that subsequently lead to stimulation of physiological and morphological changes, eventually leading to tolerance or resistance of abiotic stresses [179]. Several molecular studies have described that the PGPR induce stress tolerance (biotic as well abiotic) through crosstalk between various phytohormones and proper signaling network [180].

Different mechanisms of IST by several elicitors stimulated by inoculation of PGPR have been also demonstrated for the mitigation of abiotic stresses [92-93]. Under the conditions of salt stress the inoculation of tomato by PGPR *Sphingobacterium* BHU-AV3 showing whole plant protection through IST was due to reduced ROS levels, increased antioxidant enzyme activities, and the multiple-isoform expression of superoxide dismutase (SOD), polyphenol oxidase (PPO), and peroxidase (POD) in the plant roots [181]. In wheat, IST was elicited by a halotolerant *Aeromonas* sp. (strains, SAL17 and

SAL21) via the production of many acyl-homoserine lactones (AHLs) to mitigate salt stress [182]. During heavy metal stress, *Pseudomonas* SFP1 which is a metal tolerant species produces IAA [183]. It also secretes many enzymes for degradation of the cell wall that include chitinases, cellulose, protease, glucanase, lipopolypeptides, and HCN which provide inhibition to plant pathogenic fungi, bacteria, viruses, and also restrain nematodes [180]. Different PGPR treatments known to induce systemic tolerance in wheat against abiotic stresses including salinity, drought, heat, and cold have been well studied [184].

#### 4. Conclusions

Feeding the world's rising population is one of the big challenges especially when the agriculture system is facing a multitude of complex problems rising from changing environments due to global climate change. This global phenomenon triggers and worsens already existing abiotic stresses due to the shifting of normal climatic patterns like water budgets resulting in frequent droughts, floods, salinization, and temperature extremes. These problems become factors for shifting patterns of weeds and phytopathogens and also reduce the beneficial microbial population associated with plants that affect plant health while leaving plants susceptible to biotic stress. Further, to guarantee and ensure a sufficient yield and biocontrol of pests, agriculture is increasingly relying on chemical fertilizers and pesticides, which unfortunately has a very negative environmental effect. Therefore, in recent years, to establish environmentally sustainable alternatives to such agrochemicals the use of PGPR plant biostimulants (PBs) is attracting worldwide interest. The PB market is rising rapidly, with an expected exponential growth rate in the next future. PGPR-based BPBs have shown effectiveness in nutrition use, mitigation of abiotic/biotic stress, and/or crop quality characteristics when applied to agricultural and horticultural crop plants (fruits, vegetables, ornamental plants, and medicinal plants). The PGPR makes soil elements, such as iron, phosphorus, and potassium, zinc more available to plants through the phytohormone regulation, production, and release of siderophores, organic acids, and enzymes.

Furthermore, PGPR fight various abiotic and biotic stresses through a multitude of mechanisms or a combination of an array of mechanisms such as phytohormone regulation, signaling pathways, gene regulation, and expression, secondary metabolites, VOCs, bioactive compound enhancement, ROS enzyme activities, etc. However, detailed work also needs to be carried out for a further explanation of mechanisms related to plant-microbe interactions, their bilateral "molecular dialogue" and the "omics" approaches particularly under the synergistic pressures of abiotic and biotic stress under field conditions. Such cognizance will expound on the development of new biostimulants formulations and their implementation as an innovative solution to the current food crisis.

**Acknowledgment:** The authors extend their appreciation to Universiti Teknologi Malaysia (UTM), Malaysia, and All Cosmos Industries Sdn. Bhd. for providing support to this study.

**Funding:** The authors extend their appreciation to Universiti Teknologi Malaysia (UTM) for project No. 526 QJ130000.3609.02M43, QJ130000.3609.02M39, and All Cosmos Industries Sdn. Bhd. through project No. R.J130000.7344.4B200.

## References

- [1]. Ebert, A.W.; Engels, J.M. Plant Biodiversity and Genetic Resources Matter!. *Plants*. **2020**, *9*, 1706.
- [2]. Shahzad, A.; Ullah, S.; Dar, A.A.; Sardar, M.F.; Mehmood, T.; Tufail, M.A.; Shakoor, A.; Haris, M. Nexus on climate change: agriculture and possible solution to cope future climate change stresses *Environ. Sci. Pollut. Res.* **2021**, 1-22.
- [3]. Sangiorgio, D.; Cellini, A.; Donati, I.; Pastore, C.; Onofrietti, C.; Spinelli, F. Facing climate change: application of microbial biostimulants to mitigate stress in horticultural crops. *Agronomy*. **2020**, *10*, 6, 794.
- [4]. Del Buono, D. Can biostimulants be used to mitigate the effect of anthropogenic climate change on agriculture? It is time to respond. *Sci. Total. Environ.* **2020**, 141763.
- [5]. Raza, A.; Razzaq, A.; Mehmood, S.S.; Zou, X.; Zhang, X.; Lv, Y.; Xu, J. Impact of climate change on crops adaptation and strategies to tackle its outcome: A review. *Plants*. **2019**, *8*, 34.
- [6]. Malhi, G.S.; Kaur, M.; Kaushik, P. Impact of Climate Change on Agriculture and Its Mitigation Strategies: A Review. *Sustainability*. **2021**, *13*, 1318.
- [7]. Mancosu, N.; Snyder, R.L.; Kyriakakis, G.; Spano, D. Water scarcity and future challenges for food production. *Water*. **2015**, *7*, 975–992.
- [8]. Kałużewicz, A.; Bosiacki, M.; Spizewski, T. Influence of biostimulants on the content of macro-and micronutrients in broccoli plants exposed to drought stress. *J. Elem.* **2018**, *23*, 287–296.
- [9]. Bulgari, R.; Franzoni, G.; Ferrante, A. Biostimulants application in horticultural crops under abiotic stress conditions. *Agronomy*. **2019**, *9*, 306.
- [10]. Daneshmand, H.; Alaghmand, S.; Camporese, M.; Talei, A.; Yeh, P.J.F.; Daly, E. Long-term impacts of partial afforestation on water and salt dynamics of an intermittent catchment under climate change. *Water*. **2020**, *12*, 1067.
- [11]. Bulgari, R.; Cocetta, G.; Trivellini, A.; Vernieri, P.; Ferrante, A. Biostimulants and crop responses: a review. *Biol. Agric. & Hort.* **2015**, *31*, 1-17.
- [12]. Colla G.; Rouphael Y. Biostimulants in horticulture. *Sci. Hort.* **2015**, *196*, 1–2.
- [13]. Yakhin, O.I.; Lubyantsev, A.A.; Yakhin, I.A.; Brown, P.H. Biostimulants in plant science: a global perspective. *Front Plant Sci.* **2017**, *7*, 2049.
- [14]. Rouphael, Y.; Colla, G. Synergistic biostimulatory action: Designing the next generation of plant biostimulants for sustainable agriculture. *Front Plant Sci.* **2018**, *9*, 1655.
- [15]. Chiaiese, P.; Corrado, G.; Colla, G.; Kyriacou, M.C.; Rouphael, Y. Renewable sources of plant biostimulation: microalgae as a sustainable means to improve crop performance. *Front Plant Sci.* **2018**, *9*, 1782.
- [16]. Rouphael, Y.; Colla, G. Toward sustainable agriculture through plant biostimulants: From experimental data to practical applications. **2020**, *10*, 1461.
- [17]. E.U. Regulation of the European Parliament and the Council Laying Down Rules on the Making Available on the Market of EU Fertilising Products and Amending Regulations (EC) No 1069/2009 and (EC) No. 1107/2009 and Repealing Regulation (EC) No 2003/2003. **2019**. Available online: <https://eur-lex.europa.eu/legal-content/EN/TXT/?uri=OJ:L:2019:170:TOC>.
- [18]. Szczałba, M.; Kopta, T.; Gąstoł, M.; Sękara, A. Comprehensive insight into arbuscular mycorrhizal fungi, *Trichoderma* spp. and plant multilevel interactions with emphasis on biostimulation of horticultural crops. *J. Appl. Microbiol.* **2019**, *127*, 630-647.
- [19]. Wozniak, E.; Blaszcak, A.; Wiatrak, P.; Canady, M. Biostimulant Mode of Action: Impact of Biostimulant on Whole-Plant Level. *The Chem Biol of Plant Biostimu.* **2020**, 205-227.
- [20]. Brown, P.; Saa, S. Biostimulants in agriculture. *Front Plant Sci.* **2015**, *6*, 671.
- [21]. Ruzzi, M.; Aroca, R. Plant growth-promoting rhizobacteria act as biostimulants in horticulture. *Scient Hort.* **2015**, *196*, 124-134.



- [22]. Vandenkoornhuyse, P.; Quaiser, A.; Duhamel, M., Le Van,; Dufresne, A. The importance of the microbiome of the plant holobiont. *New Phytol.* **2015**, *206*, 1196-1206.
- [23]. Ratiu, I.A.; Al-Suod, H.; Ligor, M.; Monedeiro, F.; Buszewski, B. Effects of growth conditions and cultivability on the content of cyclitols in *Medicago sativa*. *International Journal of Environmental Science and Technology*. **2020**, *18*, 33–48.
- [24]. Du Jardin, P. Plant biostimulants: definition, concept, main categories and regulation. *Scient Hort.* **2015**, *196*, 3-14.
- [25]. Lugtenberg, B. Principles of plant-microbe interactions: microbes for sustainable agriculture. *Princ Plant-Microbe Interact Microbes Sustain Agric*. **2015**.
- [26]. Rouphael, Y.; Colla, G. Biostimulants in Agriculture. *Front Plant Sci*. **2020**, *11*, 40.
- [27]. Dunhamtrimmer.com, **2018**. Available from: <http://dunhamtrimmer.com/products-services>.
- [28]. Woo, S.L.; Pepe, O. Microbial consortia: promising probiotics as plant biostimulants for sustainable agriculture. *Front Plant Sci*. **2018**, *9*, 1801.
- [29]. Le Mire, G.; Nguyen, M.; Fassotte, B.; du Jardin, P.; Verheggen, F.; Delaplace, P.; Jijakli, H. Implementing biostimulants and biocontrol strategies in the agroecological management of cultivated ecosystems. *Biotechnologie, Agronomie, Société et Environnement*. **2016**, *20*, 1-15.
- [30]. Aamir, M., Rai, K.K., Zehra, A., Dubey, M.K., Kumar, S., Shukla, V. and Upadhyay, R.S. Microbial bioformulation-based plant biostimulants: a plausible approach toward next generation of sustainable agriculture. In *Microbial Endophytes*; Woodhead Publishing, **2020**, pp. 195-225.
- [31]. Mishra, J.; Arora, N.K. Bioformulations for plant growth promotion and combating phytopathogens: a sustainable approach. In *Bioformulations: For sustainable agriculture*; Springer, New Delhi, **2016**, pp. 3-33.
- [32]. Dellagi, A.; Quillere, I.; Hirel, B. Beneficial soil-borne bacteria, and fungi: a promising way to improve plant nitrogen acquisition. *J Exp Bot.* **2020**, *7*, 4469-4479.
- [33]. Sayyed, R. Z.; Patel, P. R.; Shaikh, S. S. Plant Growth Promotion and Root Colonization by EPS Producing *Enterobacter* sp. RZS5 under Heavy Metal Contaminated Soil, *Indian J Exp Biol.* **2015**, *53*, 116-123.
- [34]. Souza, R. D.; Ambrosini, A.; Passaglia, L. M. Plant growth-promoting bacteria as inoculants in agricultural soils. *Genetics and Mol Biol*. **2015**, *38*, 401-419.
- [35]. Santoyo, G.; Moreno-Hagelsieb, G.; Del Carmen Orozco-Mosqueda, M.; Glick, B. R. Plant growth-promoting bacterial endophytes. *Microbiol Res*. **2016**, *183*, 92-99.
- [36]. Patel, P. R.; Shaikh, S. S.; Sayyed, R. Z. Dynamism of PGPR in bioremediation and plant growth promotion in heavy metal contaminated soil. *Indian J Exp Biol*. **2016**, *54*, 286-290.
- [37]. Woźniak, M.; Gałazka, A.; Tyśkiewicz, R.; Jaroszek-Ścisł, J. Endophytic Bacteria Potentially Promote Plant Growth by Synthesizing Different Metabolites and their Phenotypic/Physiological Profiles in the Biolog GEN III MicroPlate™ Test. *Intl J Mol Sci*, **2019**, *20*, 5283.
- [38]. Kabiraj, A.; Majhi, K.; Halder, U.; Let, M.; Bandopadhyay, R. Role of Plant Growth-Promoting Rhizobacteria (PGPR) for Crop Stress Management. In *Sustainable Agriculture in the Era of Climate Change, 2020*; Springer, Cham, **2020**, 367-389.
- [39]. Papik, J.; Folkmanova, M.; Polivkova, M.; Suman, J.; Uhlik, O. The invisible life inside plants: Deciphering the riddles of endophytic bacterial diversity. *Biotechnol Adv.* **2020**, 107614.
- [40]. Miliute, I.; Buzaitė, O.; Baniulis, D.; Stanys, V. Bacterial endophytes in agricultural crops and their role in stress tolerance: a review. *Zemdirbyste-Agriculture* **2015**, *102*, 465-478.
- [41]. Wemheuer, F.; Kaiser, K.; Karlovsky, P.; Daniel, R.; Vidal, S.; Wemheuer, B. Bacterial endophyte communities of three agricultural important grass species differ in their response towards management regimes. *Scienti Rep*. **2017**, *7*, 1-13.
- [42]. Liu, H., Carvalhais, L. C., Crawford, M., Singh, E., Dennis, P. G., Pieterse, C. M., & Schenk, P. M. Inner plant values: diversity, colonization, and benefits from endophytic bacteria. *Front In Microbiol*. **2017**, *8*, 2552.
- [43]. Pii, Y.; Penn, A.; Gattullo, C.E.; Allegratta, I.; Terzano, R.; Crecchio, C.; Mimmo, T.; Cesco, S. The interactions between plant, microorganism, and soil affect Fe acquisition in cucumber plants. *Center for Soil Ecol*. **2015**.
- [44]. Sayyed, R.Z.; Arora, N.K.; Reddy, M.S. *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management: Volume 1: Rhizobacteria in Abiotic Stress Management*, Springer Nature, Singapore, **2019**; pp. 1-362.
- [45]. Nguyen, M.L.; Spaepen, S.; du Jardin, P.; Delaplace, P. Biostimulant effects of rhizobacteria on wheat growth and nutrient uptake depend on nitrogen application and plant development. *Arch of Agro and Soil Sci*, **2019**, *65*, 58-73.
- [46]. Vetrano, F.; Miceli, C.; Angileri, V.; Frangipane, B.; Moncada, A.; Miceli, A. Effect of Bacterial Inoculum and Fertigation Management on Nursery and Field Production of Lettuce Plants. *Agronomy* **2020**, *10*, 1477.
- [47]. Pellegrini, M.; Spera, D.; Ercole, C.; del Gallo, M. Allium cepa L. seed inoculation with a consortium of plant growth-promoting bacteria: effects on plant growth and development and soil fertility status and microbial community. **2020**.
- [48]. He, Y.; Pantigoso, H.A.; Wu, Z.; Vivanco, J.M. Co-inoculation of *Bacillus* sp. and *Pseudomonas putida* at different development stages acts as a biostimulant to promote growth, yield, and nutrient uptake of tomato. *J Applied Microbiol.* **2019**, *127*, 196-207.
- [49]. Chandrasekaran, M.; Chun, S.C.; Oh, J.W.; Paramasivan, M.; Saini, R.K.; Sahayarayan, J.J. *Bacillus subtilis* CBR05 for Tomato (*Solanum lycopersicum*) Fruits in South Korea as a Novel Plant Probiotic Bacterium (PPB): Implications from Total Phenolics, Flavonoids, and Carotenoids Content for Fruit Quality. *Agronomy* **2019**, *9*, 838.



- [50]. Islam, S.; Akanda, A.M.; Prova, A.; Islam, M.T.; Hossain, M.M. Isolation and identification of plant growth-promoting rhizobacteria from cucumber rhizosphere and their effect on plant growth promotion and disease suppression. *Front In Microbiol.* **2016**, *6*, 1360.
- [51]. Mangmang, J.S.; Deaker, R.; Rogers, G. Optimal plant growth-promoting concentration of *Azospirillum brasilense* inoculated to cucumber, lettuce, and tomato seeds varies between bacterial strains. *Israel J Plant Sci.* **2015**, *62*, 145-152.
- [52]. Jha, Y.; Subramanian, R.B. PGPR regulate the caspase-like activity, programmed cell death, and antioxidant enzyme activity in paddy under salinity. *Physiology and Molecular Biology of Plants* **2014**, *20*, 201-207.
- [53]. Fibach-Paldi, S.; Burdman, S.; Okon, Y. Key physiological properties contributing to rhizosphere adaptation and plant growth promotion abilities of *Azospirillum brasilense*. *FEMS Microbiol Lett.* **2012**, *326*, 99-108.
- [54]. Fahad, S.; Hussain, S.; Matloob, A.; Khan, F.A.; Khaliq, A.; Saud, S.; Hassan, S.; Shan, D.; Khan, F.; Ullah, N.; Faiq, M. Phytohormones and plant responses to salinity stress: a review. *Plant Growth Reg.* **2015**, *75*, 391-404.
- [55]. Spaepen, S.; Bossuyt, S.; Engelen, K.; Marchal, K.; Vanderleyden, J. Phenotypical and molecular responses of *Arabidopsis thaliana* roots as a result of inoculation with the auxin-producing bacterium *Azospirillum brasilense*. *New Phytol.* **2014**, *201*, 850-861.
- [56]. Llorente, B.E.; Alasia, M.A.; Larraburu, E.E. Biofertilization with *Azospirillum brasilense* improves in vitro culture of *Handroanthus ochraceus*, a forestry, ornamental and medicinal plant. *New Biotechnol.* **2016**, *33*, 32-40.
- [57]. Faruque, O. M., Miwa, H., Yasuda, M., Fujii, Y., Kaneko, T., Sato, S., & Okazaki, S. Identification of *Bradyrhizobium elkanii* genes involved in incompatibility with soybean plants carrying the Rj4 allele. *Appl and Env Microbiol.* **2015**, *81*, 6710-6717.
- [58]. Hernández-León, R.; Rojas-Solís, D.; Contreras-Pérez, M.; del Carmen Orozco-Mosqueda, M.; Macías-Rodríguez, L.I.; Reyes-de la Cruz, H.; Valencia-Cantero, E.; Santoyo, G. Characterization of the antifungal and plant growth-promoting effects of diffusible and volatile organic compounds produced by *Pseudomonas fluorescens* strains. *Biol. Cont.* **2015**, *81*, 83-92.
- [59]. Gupta, G.; Parihar, S.S.; Ahirwar, N.K.; Sneh, S.K.; Singh, V. Plant growth-promoting rhizobacteria (PGPR): current and prospects for the development of sustainable agriculture. *J Microbiol Biochem Technol.* **2015**, *7*, 096-102.
- [60]. Jha, C.K.; Saraf, M. Plant growth-promoting rhizobacteria (PGPR): a review. *J of Agric Res and Dev.* **2015**, *5*, 108-119.
- [61]. Heydarian, Z.; Yu, M.; Gruber, M.; Glick, B.R.; Zhou, R.; Hegedus, D.D. Inoculation of soil with plant growth-promoting bacteria producing 1-aminocyclopropane-1-carboxylate deaminase or expression of the corresponding *acdS* gene in transgenic plants increases salinity tolerance in *Camelina sativa*. *Front in Microbiol.* **2016**, *7*, 196674.
- [62]. Bucuo, D. S.; Yapit, R. H.; Gabriel, M. L. S. Biochemical Characterization of Microbials and Their Effects on the Growth and Yield of Multiplier Onion (*Allium ascalonicum* L.) in Northwestern Philippines. In *Plant Growth Promoting Rhizobacteria (PGPR): Prospects for Sustainable Agriculture*; Springer, Singapore, **2019**, pp. 75-91.
- [63]. Kumar, R.; Saurabh, K.; Kumawat, N.; Sundaram, P.K.; Mishra, J.S.; Singh, D.K.; Hans, H.; Krishna, B.; Bhatt, B.P. Sustaining Productivity Through Integrated Use of Microbes in Agriculture. In *Role of Microbial Communities for Sustainability*; Springer, Singapore, **2021**, pp. 109-145.
- [64]. Singh, M.; Dotaniya, M.L.; Mishra, A.; Dotaniya, C.K.; Regar, K.L.; Lata, M. Role of biofertilizers in conservation agriculture. *In Conser Agric.* **2016**, 113-134.
- [65]. Thilakarathna, M.S.; Raizada, M.N. A meta-analysis of the effectiveness of diverse rhizobia inoculants on soybean traits under field conditions. *Soil Bioland Biochem* **2017**, *105*, 177-196.
- [66]. Ramakrishna, W., Yadav, R., & Li, K). Plant growth-promoting bacteria in agriculture: two sides of a coin. *Appl Soil Ecol.* **2019**, *138*, 10-18.
- [67]. Bashan, Y.; de-Bashan, L.E. Inoculant Preparation, and Formulations for *Azospirillum* spp. In *Handbook for Azospirillum* Springer, Cham. **2015**, 469-485.
- [68]. Beattie, G.A. Microbiomes: curating communities from plants. *Nature* **2015**, *528*, 340-341.
- [69]. Gupta, M.; Kiran, S.; Gulati, A.; Singh, B.; Tewari, R. Isolation and identification of phosphate solubilizing bacteria able to enhance the growth and aloin-A biosynthesis of *Aloe barbadensis* Miller. *Microbiol Res.* **2012**, *167*, 358-363.
- [70]. Sharma, S.B.; Sayyed, R.Z.; Trivedi, M.H.; Gobi, T.A. Phosphate solubilizing microbes: sustainable approach for managing phosphorus deficiency in agricultural soils. *Springer Plus.* **2013**, *2*, 587.
- [71]. Souza, R.D.; Ambrosini, A.; Passaglia, L.M. Plant growth-promoting bacteria as inoculants in agricultural soils. *Genet and Mol Biol.* **2015**, *38*, 401-419.
- [72]. Suleman, M.; Yasmin, S.; Rasul, M.; Yahya, M.; Atta, B.M.; Mirza, M.S. Phosphate solubilizing bacteria with glucose dehydrogenase gene for phosphorus uptake and beneficial effects on wheat. *PloS One* **2018**, *13*, 9, 0204408.
- [73]. Bechtaoui, N.; Raklami, A.; Benidire, L.; Tahiri, A.I.; Göttfert, M.; Oufdou, K. Effects of PGPR co-inoculation on growth, phosphorus nutrition and phosphatase/phytase activities of faba bean under different phosphorus availability conditions. *Polish Journal of Environmental Studies*, **2020**, *29*, pp. 1557-1565.
- [74]. Nath, D.; Maurya, B.R.; Meena, V.S. Documentation of five potassium-and phosphorus-solubilizing bacteria for their K and P-solubilization ability from various minerals. *Biocatal and Agri Biotechnol.* **2017**, *10*, 174-181.
- [75]. Meena, V.S.; Meena, S.K.; Verma, J.P.; Kumar, A.; Aeron, A.; Mishra, P.K.; Bisht, J.K.; Pattanayak, A.; Naveed, M.; Dotaniya, M.L. Plant beneficial rhizospheric microorganism (PBRM) strategies to improve nutrients use efficiency: A review. *Ecol Engg.* **2017**, *107*, 8-32.

- [76]. Sindhu, S.S.; Parmar, P.; Phour, M.; Sehrawat, A. Potassium-solubilizing microorganisms (KSMs) and its effect on plant growth improvement. In Potassium solubilizing microorganisms for sustainable agriculture. Springer. **2016**, 171-185.
- [77]. Bahadur, I.; Maurya, B.R.; Meena, V.S.; Saha, M.; Kumar, A.; Aeron, A. Mineral release dynamics of tricalcium phosphate and waste muscovite by mineral-solubilizing rhizobacteria isolated from Indo-Gangetic plain of India. *Geomicrobiol J.* **2017**, *34*, 454-466.
- [78]. Liu, J.; Aronsson, H.; Ulen, B.; Bergström, L. Potential phosphorus leaching from sandy topsoils with different fertilizer histories before and after application of pig slurry. *Soil Use and Mgmt.* **2012**, *28*(4), 457-467.
- [79]. Mehnaz, S. An overview of globally available bioformulations. In Bioformulations: For Sustainable Agriculture. **2016**, 267-281.
- [80]. Wei, Y.; Zhao, Y.; Shi, M.; Cao, Z.; Lu, Q.; Yang, T.; Fan, Y.; Wei, Z. Effect of organic acids production and bacterial community on the possible mechanism of phosphorus solubilization during composting with enriched phosphate-solubilizing bacteria inoculation. *Biores Technol.* **2018**, *247*, 190-199.
- [81]. Schultz, N.; Pereira, W.; de Albuquerque Silva, P.; Baldani, J. I.; Boddey, R. M.; Alves, B. J. R.; Urquiaga, S.; Reis, V. M. Yield of sugarcane varieties and their sugar quality grown in different soil types and inoculated with a diazotrophic bacteria consortium. *Plant Prod Sci.* **2018**, *20*, 366-374.
- [82]. Shakeel, M.; Rais, A.; Hassan, M.N.; Hafeez, F.Y. Root associated *Bacillus* sp. improves growth, yield, and zinc translocation for basmati rice (*Oryza sativa*) varieties. *Front in Microbiol.* **2015**, *6*, 1286.
- [83]. Bhupenchandra, I.; Devi, S.H.; Basumatary, A.; Dutta, S.; Singh, L.K.; Kalita, P. Bora, S.S.; Devi, S.R.; Saikia, A. Sharma, P.; Bhagwati, S. Biostimulants: Potential and Prospects in Agriculture. *Intl Res J of Pure and Appl Chem.* **2020**, 20-35.
- [84]. Colla, G.; Nardi, S.; Cardarelli, M.; Ertani, A.; Lucini, L.; Canaguier, R.; Roupheal, Y. Protein hydrolysates as biostimulants in horticulture. *Scient Hort.* **2015**, *196*, 28-38.
- [85]. Grzyb, Z.S.; Paszt, L.S.; Piotrowski, W.; Malusa, E. The influence of mycorrhizal fungi on the growth of apple and sour cherry maidens fertilized with different bioproducts in the organic nursery. *J Life Sci.* **2015**, *9*, 221-228.
- [86]. Drobek, M.; Frac, M.; Cybulska, J. Plant biostimulants: the importance of the quality and yield of horticultural crops and the improvement of plant tolerance to abiotic stress—a review. *Agronomy.* **2019**, *9*, 335.
- [87]. Fiorentino, N.; Ventorino, V.; Woo, S.L.; Pepe, O.; De Rosa, A.; Gioia, L.; Romano, I.; Lombardi, N.; Napolitano, M.; Colla, G.; Roupheal, Y. Trichoderma-based biostimulants modulate rhizosphere microbial populations and improve N uptake efficiency, yield, and nutritional quality of leafy vegetables. *Front in Plant Sci.-* **2018**, *9*, 743.
- [88]. Caradonia, F.; Ronga, D.; Flore, A.; Barbieri, R.; Moulin, L.; Terzi, V.; Francia, E., Biostimulants and cherry rootstock increased tomato fruit yield and quality in sustainable farming systems. *Italian J Agron.* **2020**, *15*, 121-131.
- [89]. Turan, M.; Yildirim, E.; Kitir, N.; Unek, C.; Nikerel, E.; Ozdemir, B.S.; Güneş, A.; Mokhtari, N.E.P. Beneficial role of plant growth-promoting bacteria in vegetable production under abiotic stress. *Microbial Strat for Veg Prod.* **2017**, 151-166.
- [90]. Ji, J.; Yuan, D.; Jin, C.; Wang, G.; Li, X.; Guan, C. Enhancement of growth and salt tolerance of rice seedlings (*Oryza sativa* L.) by regulating ethylene production with a novel halotolerant PGPR strain *Glutamicibacter* sp. YD01 containing ACC deaminase activity. *Acta Physiol Plant.* **2020**, *42*, 1-17.
- [91]. Ilyas, N.; Mazhar, R.; Yasmin, H.; Khan, W.; Iqbal, S.; Enshasy, H.E.; Dailin, D.J. Rhizobacteria isolated from saline soil induce systemic tolerance in wheat (*Triticum aestivum* L.) against salinity stress. *Agronomy.* **2020**, *10*, 989.
- [92]. Filgueiras, L.; Silva, R.; Almeida, I.; Vidal, M.; Baldani, JI; Meneses, CHSG. *Gluconacetobacter diazotrophicus* mitigates drought stress in *Oryza sativa* L. *Plant Soil.* **2019**, *451*, 57–73.
- [93]. Silva, R.; Filgueiras, L.; Santos, B.; Coelho, M.; Silva, M.; Estrada-Bonilla, G.; Meneses, C. *Gluconacetobacter diazotrophicus* changes the molecular mechanisms of root development in *Oryza sativa* L. growing under water stress. *Int. J. Mol. Sci.* **2020**, *21*, 333.
- [94]. Garcia, J.E.; Maroniche, G.; Creus, C.; Suarez-Rodriguez, R.; Ramirez-Trujillo, J.A.; Groppa, M.D. In vitro PGPR properties and osmotic tolerance of different *Azospirillum* native strains and their effects on growth of maize under drought stress. *Microbiol Res.* **2017**, *202*, 21–29.
- [95]. Moreno-Galván, A.; Romero-Perdomo, F. A.; Estrada-Bonilla, G.; Meneses, C.H.S.G.; Bonilla, R.R. Dry-caribbean *Bacillus* spp. strains ameliorate drought stress in maize by a strain-specific antioxidant response modulation. *Microorganisms*, **2020**, *8*, 823.
- [96]. Fatima, T.; Arora, N.K. *Pseudomonas entomophila* PE3 and its exopolysaccharides as biostimulants for enhancing growth, yield, and tolerance responses of sunflower under saline conditions. *Microbiological Research.* **2021**, *244*, 126671.
- [97]. Tiriyaki, D.; Aydın, İ.; Atıcı, Ö. Psychrotolerant bacteria isolated from the leaf apoplast of cold-adapted wild plants improve the cold resistance of bean (*Phaseolus vulgaris* L.) under low temperature. *Cryobiology.* **2019**, *86*, 111-119.
- [98]. Qin, Y.; Fu, Y.; Kang, W.; Li, H.; Gao, H.; Vitalievitch K. S.; Liu, H. Isolation and identification of a cold-adapted bacterium and its characterization for biocontrol and plant growth-promoting activity. *Ecol Eng.* **2017**, *105*, 362–369.
- [99]. Zerrouk, I. Z.; Benchabane, M.; Khelifi, L.; Yokawa, K.; Ludwig-Muller, J.; Baluska, F. *Pseudomonas* strain isolated from date-palm rhizospheres improves root growth and promotes root formation in maize exposed to salt and aluminum stress. *J Plant Physiol.* **2016**, *191*, 111–119.

- [100]. Rizvi, A.; Khan, M. S. Heavy metal-induced oxidative damage and root morphology alterations of maize (*Zea mays* L.) plants and stress mitigation by metal tolerant nitrogen-fixing *Azotobacter chroococcum*. *Ecotoxicol Environ Saf.* **2018**, *157*, 9–20
- [101]. Ngumbi, E.; Kloepper, J. Bacterial-mediated drought tolerance: current and future prospects. *Appl Soil Ecol.* **2016**, *10*, 109–125.
- [102]. Vurukonda, S.S.K.P.; Vardharajula, S.; Shrivastava, M.; Ali, S.Z. Enhancement of drought stress tolerance in crops by plant growth-promoting rhizobacteria. *Microbiol Res.* **2016**, *184*, 13–24.
- [103]. Jochum, M. D.; McWilliams, K. L.; Borrego, E. J.; Kolomiets, M. V.; Niu, G.; Pierson, E. A.; Jo, Y. K. Bioprospecting plant growth-promoting rhizobacteria that mitigate drought stress in grasses. *Front in Microbiol.* **2019**, *10*, 2106.
- [104]. Khan, N.; Bano, A.; Ali, S.; Babar, M. A. Crosstalk amongst phytohormones from planta and PGPR under biotic and abiotic stresses. *Plant Growth Reg.* **2020**, *90*, 189–203.
- [105]. Abbas, R.; Rasul, S.; Aslam, K.; Baber, M.; Shahid, M.; Mubeen, F.; Naqqash, T. Halotolerant PGPR: A hope for the cultivation of saline soils. *J King Saud Univ Sci.* **2019**, *31*, 1195–1201.
- [106]. Ilangumaran, G.; Smith, D. L. Plant growth-promoting rhizobacteria in amelioration of salinity stress: a systems biology perspective. *Front in Plant Sci.* **2017**, *8*, 1768.
- [107]. Wen, W.; Timmermans, J.; Chen, Q.; van Bodegom, P. M. A Review of Remote Sensing Challenges for Food Security with Respect to Salinity and Drought Threats. *Remote Sensing* **2021**, *13*, 6.
- [108]. Fazeli-Nasab, B.; Sayyed, R. Z. Plant Growth-Promoting Rhizobacteria and Salinity Stress: A Journey into the Soil. In *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management*, Springer, Singapore, 2019, pp. 21–34.
- [109]. Pan, J.; Peng, F.; Xue, X.; You, Q.; Zhang, W.; Wang, T.; Huang, C. The growth promotion of two salt-tolerant plant groups with PGPR inoculation: a meta-analysis. *Sustainability.* **2019**, *11*, 378.
- [110]. Malhotra, S.K. Horticultural crops and climate change—A review. *Indian J. Agric. Sci.* **2017**, *87*, 12–22.
- [111]. Pereira, F. Rhizobacteria as Bioprotectants against stress conditions. In *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management*; Springer, Singapore, 2019, pp. 157–177.
- [112]. Canarini, A.; Dijkstra, F.A. Dry-rewetting cycles regulate wheat carbon rhizodeposition, stabilization, and nitrogen cycling. *Soil Biol Biochem.* **2015**, *81*, 195–203.
- [113]. Singh, J.; Singh, P.; Ray, S.; Rajput, R.S.; Singh, H.B. Plant Growth-Promoting Rhizobacteria: Benign and Useful Substitute for Mitigation of Biotic and Abiotic Stresses. In *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management*; Springer, Singapore, 2019, pp. 81–101.
- [114]. Huang, S.; Zuo, T.; Ni, W. Important roles of glycine betaine in stabilizing the structure and function of the photosystem II complex under abiotic stresses. *Planta.* **2020**, *251*, 1–14.
- [115]. Bharti, N.; Pandey, S.S.; Barnawal, D.; Patel, V.K.; Kalra, A. Plant growth-promoting rhizobacteria *Dietzia natronolimnaea* modulates the expression of stress-responsive genes providing protection of wheat from salinity stress. *Sci Rep.* **2016**, *6*, 1–16.
- [116]. Kang, S.M.; Asaf, S.; Khan, A.L.; Khan, A.; Mun, B.G.; Khan, M.A.; Gul, H.; Lee, I.J. Complete genome sequence of *Pseudomonas psychrotolerans* CS51, a plant growth-promoting bacterium, under heavy metal stress conditions. *Microorganisms*, **2020**, *8*, 382.
- [117]. Mishra, J.; Singh, R.; Arora, N.K. Alleviation of heavy metal stress in plants and remediation of soil by rhizosphere microorganisms. *Front in Microbiol.* **2017**, *8*, 1706.
- [118]. Hassan, T.U.; Bano, A.; Naz, I. Alleviation of heavy metals toxicity by the application of plant growth-promoting rhizobacteria and effects on wheat grown in saline-sodic field. *Intl J of Phytoremed.* **2017**, *19*, 522–529.
- [119]. Khanna, K.; Jamwal, V. L.; Gandhi, S. G.; Ohri, P.; Bhardwaj, R. Metal resistant PGPR lowered Cd uptake and expression of metal transporter genes with improved growth and photosynthetic pigments in *Lycopersicon esculentum* under metal toxicity. *Scienti Rep.* **2019**, *9*, 1–14.
- [120]. Niu, D.D.; Zheng, Y.; Zheng, L.; Jiang, C.H.; Zhou, D.M.; Guo, J.H. Application of PSX biocontrol preparation confers root-knot nematode management and increased fruit quality in tomato under field conditions. *Biocont Sci and Technol.* **2016**, *26*, 174–180.
- [121]. Munhoz, L.D.; Fontequ, J.P.; Santos, I.M.O.; Navarro, M.O.P.; Simionato, A.S.; Goya, E.T.; Rezende, M.I.; Balbi-Peña, M.L.; de Oliveira, A.G.; Andrade, G. Control of bacterial stem rot on tomato by extracellular bioactive compounds produced by *Pseudomonas aeruginosa* LV strain. *Cog. Food & Agri.* **2017**, *31*, 1282592.
- [122]. Sorokan, A.; Benkovskaya, G.; Burkhanova, G.; Blagova, D.; Maksimov, I. Endophytic Strain *Bacillus subtilis* 26DCryChS Producing CryIIa Toxin from *Bacillus thuringiensis* Promotes Multifaceted Potato Defense against *Phytophthora infestans* (Mont.) de Bary and Pest *Leptinotarsa decemlineata* Say. *Plants.* **2020**, *9*, 1115.
- [123]. Daranas, N.; Badosa, E.; Francés, J.; Montesinos, E.; Bonaterra, A. Enhancing water stress tolerance improves fitness in biological control strains of *Lactobacillus plantarum* in-plant environments. *PLoS One* **2018**, *13*, 0190931.
- [124]. Meena, M.; Zehra, A. Tomato: a model plant to study plant-pathogen interactions. *Food Sci Nutr Technol.* **2019**, *4*, 000171.
- [125]. Liu, K.; Garrett, C.; Fadamiro, H.; Kloepper, J.W. Induction of systemic resistance in Chinese cabbage against black rot by plant growth-promoting rhizobacteria. *Biol. Control-* **2016**, *99*, 8–13.



- [126]. Zehra, A.; Meena, M.; Dubey, M.K.; Aamir, M., Upadhyay, R.S. Synergistic effects of plant defense elicitors and *Trichoderma harzianum* on enhanced induction of antioxidant defense system in tomato against Fusarium wilt disease. *Bot Stud.* **2017**, *58*, 44.
- [127]. Goutam, J.; Singh, R.; Vijayaraman, R.S.; Meena, M. Endophytic fungi: carrier of potential antioxidants. In *Fungi and their role in sustainable development: current perspectives.*; Gehlot, P., Singh, J.; Springer, Singapore, **2018**, pp. 539-51.
- [128]. Dinesh, R.; Anandaraj, M.; Kumar, A.; Bini, Y.K.; Subila, K.P.; Aravind, R. Isolation, characterization, and evaluation of multi-trait plant growth-promoting rhizobacteria for their growth-promoting and disease suppressing effects on ginger. *Microbiol Res.* **2015**, *173*, 34-43.
- [129]. Sayyed, R.Z. *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management: Volume 2: Rhizobacteria in Biotic Stress Management*, Springer Nature, Singapore, **2019**; pp. 1-419.
- [130]. Ulloa-Ogaz AL, Muñoz-Castellanos LN, Nevárez-Moorillón GV. Biocontrol of phytopathogens: antibiotic production as a mechanism of control. In *The battle against microbial pathogens: basic science, technological advances, and educational programs.*; Méndez-Vilas, A.; Formatex Research Center, Badajoz, Spain, **2015**, *1*, pp. 305-309.
- [131]. Fernando, W.; Nakkeeran, S.; Zhang, Y.; Savchuk, S. Biological control of *Sclerotinia Sclerotiorum* (lib.) de Bary by *Pseudomonas* and *Bacillus* species on canola petals. *Crop Prot* **2018**, *26*, 100-107.
- [132]. Sharma, S.; Kumar, V.; Tripathi, R. B. Isolation of phosphate solubilizing microorganism (PSMs) from the soil. *J Microbiol Biotechnol Res.* **2017**, *1*, 90-95.
- [133]. Ramadan, E.M.; AbdelHafez, A.A.; Hassan, E.A.; Saber, F.M. Plant growth-promoting rhizobacteria and their potential for biocontrol of phytopathogens. *Afr J Microbiol Res* **2016**, *10*, 486-504.
- [134]. Wang, X.; Mavrodi, D.V.; Ke, L.; Mavrodi, O.V.; Yang, M.; Thomashow, L.S.; Zhang, N.; Weller, D.M.; Zhang, J. Biocontrol and plant growth-promoting activity of rhizobacteria from Chinese fields with contaminated soils. *Microb Biotechnol.* **2015**, *8*, 404-418.
- [135]. Raza, W.; Yousaf, S.; Rajer, F.U. Plant growth-promoting activity of volatile organic compounds produced by biocontrol strains. *Sci Lett.* **2016**, *4*, 40-43.
- [136]. Kanchiswamy, C.N.; Malnoy, M.; Maffei, M.E. Chemical diversity of microbial volatiles and their potential for plant growth and productivity. *Front Plant Sci.* **2015**, *6*, 151
- [137]. Sindhu, S.S.; Sharma, R. Amelioration of biotic stress by application of rhizobacteria for agriculture sustainability. In *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management.*; Springer, Singapore, **2019**, pp. 111-168.
- [138]. Suryadi, Y.; Susilowati, D. N.; Fauziah, F. Management of Plant Diseases by PGPR-Mediated Induced Resistance with Special Reference to Tea and Rice Crops. *Plant Growth Promoting Rhizobacteria for Sustainable Stress Management.* **2019**, 65-110.
- [139]. Kang, B.R.; Anderson, A.J.; Kim, Y.C. Hydrogen cyanide produced by *Pseudomonas chlororaphis* o6 exhibits nematocidal activity against *Meloidogyne hapla*. *Plant Pathol J.* **2018**, *34*, 35-43.
- [140]. Santoro, M.V.; Bogino, P.C.; Nocelli, N.; Cappellari, L.R.; Giordano, W.F.; Banchio, E. Analysis of plant growth-promoting effects of Fluorescent *Pseudomonas* strains isolated from *Mentha piperita* rhizosphere and effects of their volatile organic compounds on essential oil composition. *Front Microbiol.* **2016**, *7*, 1085.
- [141]. Liu, H.; Brettell, L.E. Plant defense by VOC-induced microbial priming. *Trends in plant science.* **2019**, *24*, 187-189.
- [142]. Goswami, D.; Thakker, J.N.; Dhandhukia, P.C. Portraying mechanics of plant growth-promoting rhizobacteria (PGPR): a review. *Cogent Food Agric*, **2016**, *2*, 1-19.
- [143]. Rakshiya, Y.S.; Verma, M.K.; Sindhu, S.S. Efficacy of antagonistic soil bacteria in the management of subterranean termites (Isoptera). *Res Environ Life Sci.* **2016**, *9*, 949-955.
- [144]. Rooney, W.M.; Grinter, R.W.; Correia, A.; Parkhill, J.; Walker, D.C.; Milner, J.J. Engineering bacteriocin-mediated resistance against the plant pathogen *Pseudomonas syringae*. *Plant Biotechnol. J.* **2020**, *18*, 1296-1306.
- [145]. Zimina, M.; Babich, O.; Prosekov, A.; Sukhikh, S.; Ivanova, S.; Shevchenko, M.; Noskova, S. Overview of global trends in classification, methods of preparation and application of bacteriocins. *Antibiotics* **2020**, *9*, 553.
- [146]. Principe, A.; Fernandez, M.; Torasso, M.; Godino, A.; Fischer, S. Effectiveness of tailocins produced by prin in controlling the bacterial-spot disease in tomatoes caused by *Xanthomonas vesicatoria*. *Microbiol. Res.* **2018**, *213*, 94-102.
- [147]. Rooney, W.M.; Chai, R.; Milner, J.J.; Walker, D. Bacteriocins targeting Gram-negative phytopathogenic bacteria: Plantibiotics of the future. *Front in Microbiol.* **2020**, *11*, 2283.
- [148]. Riaz, U.; Murtaza, G.; Anum, W.; Samreen, T.; Sarfraz, M.; Nazir, M.Z. Plant Growth-Promoting Rhizobacteria (PGPR) as Biofertilizers and Biopesticides. In *Microbiota and Biofertilizers*; Springer, Cham, **2021**, pp. 181-196.
- [149]. Reed, S.C.; Yang, X.; Thornton, P.E. Incorporating phosphorus cycling into global modeling efforts: a worthwhile, tractable endeavor. *New Phytol.* **2015**, *208*, 324-329.
- [150]. Sayyed, R.Z.; Ilyas, N.; Tabassum, B.; Hashem, A.; Abd\_Allah, E.F.; Jadhav, H. P. Plausible role of plant growth-promoting Rhizobacteria in future climatic scenario. In *Env Biotechnol: for sustainable future*; Sobti, R., Arora, N., Kothari, R.; Springer, Singapore, **2019**, pp. 175-197.
- [151]. Abdul Malik, N.A.; Kumar, I.S.; Nadarajah, K. Elicitor and receptor molecules: Orchestrators of plant defense and immunity. *Int. J. Mol. Sci.* **2020**, *21*, 963.

- [152]. Maithani, D.; Singh, H.; Sharma, A. Stress Alleviation in Plants Using SAR and ISR: Current Views on Stress Signaling Network. In *Microbes and Signaling Biomolecules Against Plant Stress*. Springer, Singapore, 2021, p. 7-36.
- [153]. Meena, M.; Swapnil, P.; Divyanshu, K.; Kumar, S.; Tripathi, Y.N.; Zehra, A.; Marwal, A.; Upadhyay, R.S. PGPR-mediated induction of systemic resistance and physiochemical alterations in plants against the pathogens: Current perspectives. *J. Basic. Microbiol.* **2020**, *60*, 828-861.
- [154]. David, L.; Kang, J.; Dufresne, D.; Zhu, D.; Chen, S. Multi-Omics Revealed Molecular Mechanisms Underlying Guard Cell Systemic Acquired Resistance. *International Journal of Molecular Sciences*, **2021**, *22*, 191.
- [155]. Nishad, R.; Ahmed, T.; Rahman, V.J.; Kareem, A. Modulation of Plant Defense System in Response to Microbial Interactions. *Frontiers in Microbiology*. **2020**, *11*, 1298.
- [156]. Shaikh, S.S.; Sayyed, R.Z.; Reddy, M.S. Plant growth-promoting rhizobacteria: an eco-friendly approach for sustainable agroecosystem. In *Plant, soil, and microbes*. Springer, Cham, **2016**, pp. 181-201.
- [157]. Backer, R.; Rokem, J.S.; Ilangumaran, G.; Lamont, J.; Praslickova, D.; Ricci, E et al. Plant growth-promoting rhizobacteria: context, mechanisms of action, and roadmap to the commercialization of biostimulants for sustainable agriculture. *Front. Plant Sci.* **2018**, *9*, 1473.
- [158]. Choudhary, D.K.; Prakash, A.; Johri, B.N. Induced systemic resistance (ISR) in plants: mechanism of action. *Ind. J. Microbiol.* **2007**, *47*, 289-297.
- [159]. Beris, D.; Theologidis, I.; Skandalis, N.; Vassilakos, N. *Bacillus amyloliquefaciens* strain MBI600 induces salicylic acid-dependent resistance in tomato plants against Tomato spotted wilt virus and Potato virus Y. *Scientific reports*. **2018**, *8*, 1-11.
- [160]. Niu, D.; Wang, X.; Wang, Y.; Song, X.; Wang, J.; Guo, J.; Zhao, H. *Bacillus cereus* AR156 activates PAMP-triggered immunity and induces a systemic acquired resistance through a NPR1- and SA-dependent signaling pathway. *Biochem. Biophys. Res.* **2016**, *469*, 120-125.
- [161]. Bukhat, S.; Imran, A.; Javaid, S.; Shahid, M.; Majeed, A.; Naqqash, T. Communication of plants with microbial world: Exploring the regulatory networks for PGPR mediated defense signaling. *Microb. Res.* **2020**, 126486.
- [162]. Zebelo, S.; Song, Y.; Kloepper, J. W.; Fadamiro, H. Rhizobacteria activates (+)- $\delta$ -cadinene synthase genes and induces systemic resistance in cotton against beet armyworm (*Spodoptera exigua*). *Plant Cell Environ.*, **2016**, *39*, 935-943.
- [163]. Khoshru, B.; Mitra, D.; Khoshmanzar, E.; Myo, E.M.; Uniyal, N.; Mahakur, B.; Mohapatra, P.K.D.; Panneerselvam, P.; Boutaj, H.; Alizadeh, M.; Cely, M.V.T. Current scenario and future prospects of plant growth-promoting rhizobacteria: an economic valuable resource for the agriculture revival under stressful conditions. *Journal of Plant Nutrition*. **2020**, *43*, 3062-3092.
- [164]. Rashid, M.H.O.; Chung, Y.R.; .. Induction of systemic resistance against insect herbivores in plants by beneficial soil microbes. *Front. Plant Sci.* **2017**, *8*, 1816.
- [165]. Mhatre, P. H.; Karthik, C.; Kadirvelu, K.; Divya, K. L.; Venkatasalam, E. P.; Srinivasan, S et al. Plant growth-promoting rhizobacteria (PGPR): a potential alternative tool for nematodes bio-control. *Biocatal. Agricult. Biotechnol.* **2018**, *17*, 119-128.
- [166]. Serteyn, L.; Quaghebeur, C.; Ongena, M et al. Induced Systemic Resistance by a Plant Growth-Promoting Rhizobacterium Impacts Development and Feeding Behavior of Aphids. *Insects*. **2020**, *11*, 234.
- [167]. Johnson, S.N.; Erb, M.; Hartley, S.E. Roots under attack: contrasting plant responses to below- and aboveground insect herbivory. *New Phytologist*. **2016**, *210*, 413-418.
- [168]. Schikora, A.; Schenk, S.T.; Hartmann, A. Beneficial effects of bacteria plant communication based on quorum sensing molecules of the N-acyl homoserine lactone group. *Plant Mol. Biol.* **2016**, *90*, 605-612.
- [169]. Kramer, J.; Özkaya, Ö.; Kümmerli, R. Bacterial siderophores in community and host interactions. *Nature Rev Microbiol.* **2020**, *18*, 152-163.
- [170]. Kang, S. M.; Shahzad, R.; Bilal, S.; Khan, A. L.; Park, Y. G.; Lee, K. E et al. Indole-3-acetic-acid and ACC deaminase producing *Leclercia adecarboxylata* MO1 improves *Solanum lycopersicum* L. growth and salinity stress tolerance by endogenous secondary metabolites regulation. *BMC. Microbiol.* **2019**, *19*, 80.
- [171]. Monnier, N.; Furlan, A.; Botcazon, C.; Dahi, A.; Mongelard, G.; Cordelier, S. Rhamnolipids from *Pseudomonas aeruginosa* are elicitors triggering Brassica napus protection against Botrytis cinerea without physiological disorders. *Front. Plant Sci.* **2018**, *9*, 1170.
- [172]. Mejri, S.; Siah, A.; Coutte, F.; Magnin-Robert, M.; Randoux, B.; Tisserant, B.; Krier, F.; Jacques, P.; Reignault, P.; Halama, P. Biocontrol of the wheat pathogen *Zymoseptoria tritici* using cyclic lipopeptides from *Bacillus subtilis*. *Environmental science and pollution research*, **2018** *25*, 29822-29833.
- [173]. Pršić, J.; Ongena, M. Elicitors of plant immunity triggered by beneficial bacteria. *Front. Plant. Sci.* **2020**, *11*.
- [174]. Vega, C.; Rodríguez, M.; Llamas, I.; Béjar, V.; Sampedro, I. Silencing of Phytopathogen Communication by the Halotolerant PGPR *Staphylococcus Equorum* Strain EN21. *Microorganisms*. **2020**, *8*, 42.
- [175]. Hernández-Reyes, C.; Schenk S. T.; Neumann C.; Kogel K. H.; Schikora, A. N-acyl-homoserine lactones-producing bacteria protect plants against plant and human pathogens. *Microb. Biotechnol.* **2014**, *7*, 580-588.
- [176]. Omoboye, O.O.; Oni, F.E.; Batool, H.; Yimer, H.Z.; De Mot, R.; Höfte, M. *Pseudomonas* cyclic lipopeptides suppress the rice blast fungus *Magnaporthe oryzae* by induced resistance and direct antagonism. *Front. Plant.Sci.* **2019**, *10*, 901.
- [177]. Ma, Z.; Ongena, M.; Höfte, M. The cyclic lipopeptide orfamide induces systemic resistance in rice to *Cochliobolus miyabeanus* but not to *Magnaporthe oryzae*. *Plant Cell Rep.* **2017**, *36*, 1731-1746.

- 
- [178]. Alquéres, S.; Meneses, C.; Rouws, L.; Rothballer, M.; Baldani, I.; Schmid, M.; Hartmann, A. The bacterial superoxide dismutase and glutathione reductase are crucial for endophytic colonization of rice roots by *Gluconacetobacter diazotrophicus* PAL5. *Molecular plant-microbe interactions*, **2013**, *26*, 937-945.
- [179]. Nephali, L.; Piater, L.A.; Dubery, I.A.; Patterson, V.; Huyser, J.; Burgess, K.; Tugizimana, F. Biostimulants for plant growth and mitigation of abiotic stresses: A metabolomics perspective. *Metabolites*. **2020**, *10*, 505.
- [180]. Khan, N.; Bano, A.; Ali, S.; Babar, M.A. Crosstalk amongst phytohormones from plant and PGPR under biotic and abiotic stresses. *Plant Growth Regul.* **2020**, *90*, 189-203.
- [181]. Vaishnav, A.; Singh, J.; Singh, P.; Rajput, R.S.; Singh, H.B.; Sarma, B.K. *Sphingo bacterium* sp. BHU-AV3 Induces Salt Tolerance in tomato by enhancing antioxidant activities and energy metabolism. *Front in Microbiol.* **2020**, *11*, 443.
- [182]. Nawaz, M.S.; Arshad, A.; Rajput, L.; Fatima, K.; Ullah, S.; Ahmad, M.; Imran, A. Growth-Stimulatory Effect of Quorum Sensing Signal Molecule N-Acyl-Homoserine Lactone-Producing Multi-Trait *Aeromonas* spp. on Wheat Genotypes Under Salt Stress. *Frontiers in microbiology*. **2020**, *11*, 553621.
- [183]. Saif, S.; Khan, M.S. Assessment of toxic impact of metals on proline, antioxidant enzymes, and biological characteristics of *Pseudomonas aeruginosa* inoculated *Cicer arietinum* grown in chromium and nickel-stressed sandy clay loam soils. *Environ. Monit. Assess.* **2018**, *190*, 1-18.
- [184]. Abd El-Daim, I. A.; Bejai, S.; Meijer, J. *Bacillus velezensis* 5113 Induced Metabolic and Molecular Reprogramming during Abiotic Stress Tolerance in Wheat. *Sci Rep.* **2019**, *9*, 16282.