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Production of Bioethanol from *Artocarpus integer* (Chempedak) Peel: An Approach Towards Clean Technology

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Abstract

This research focuses on a sustainable way to reduce waste and generate renewable energy by converting Chempedak bio-waste into bioethanol. It undergoes several phases: feedstock collection and pretreatment (mechanical, acid and alkaline pretreatment), hydrolysis, fermentation, and distillation. The main goal of this project is to yield ethanol from 300g Chempedak peel waste by using a functioning and cost-effective way. When processing the project, the distillation process was then changed to ethanol evaporation which used a rotary evaporator to extract the ethanol as it is more effective. The final product for the ethanol yielded 73% for 41ml. This work provided solutions to the society to reduce waste, promote more sustainable energy solutions, and increase economic growth in industries by finding more sustainable options.

Keywords: bioethanol; biofuel; biowaste; clean technology; Chempedak

1. Introduction

Biowaste is the biodegradable organic waste produced by humans, this can be from the animals and plants we consume, or also the products we use that may produce waste products. Solid Waste Management and Public Cleansing Corporation have done an analysis showing that there are about 39,078 tons of waste is produced daily in Malaysia which amounts to 1.17 kg per person [1]. According to some reports, more than 30% of waste is disposed of as food waste, while other reports state that organic solid waste makes up 57% of municipal solid waste [2]. Currently, most of the waste in Malaysia ends up in landfills and the others end up incinerated, only 17.5% of it is recycled [3]. Despite being biodegradable, these wastes may affect the environment and degrade soil quality if they are not properly treated. They can also produce methane, which may harm the environment if it is not stored. The waste in landfills will release toxins from the biowaste and these toxins will lead to the soil or water which will pollute them [4]. Hence, the biowaste could be transformed into valuable products like bioethanol instead of being discarded.

Bioethanol can be produced from these biowaste materials since they include monomeric sugars that are either in monomeric form or are still in polymer form. These polymers can be processed to form glucose which is the main component in making bioethanol. The cost of producing bioethanol is decreased by this method's relative simplicity. It can move away from traditional fossil fuels that pollute the atmosphere by using bioethanol as fuel.

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The gasoline used in cars is made with blended fuels which contain 90% of gasoline and 10% of ethanol [5]. Depending on the kind of vehicle, this composition can differ. For instance, E85 which contains up to 85% of ethanol, can be used in flexible fuel vehicles [6]. The flexibility provides consumers with options to use higher renewable fuel content, contributing to reduced fossil fuel dependency. Bioethanol that is produced by biowaste can be used in this way which in turn up cycles of waste. The conversion of biowaste into bioethanol helps to address the global waste problem by diverting organic materials from landfills, and reducing methane emissions from anaerobic decomposition, while simultaneously creating a renewable energy source. Moreover, bioethanol serves as an excellent alternative to traditional fossil fuels, as it burns cleaner than other hydrocarbons, significantly reducing harmful air pollutants. When combusted, ethanol emits fewer nitrogen oxides, hydrocarbons and carbon monoxide compared to gasoline, leading to improved air quality and reduced smog formation. Cars that use ethanol emit no net carbon emissions due to ethanol's lower combustion temperature [7]. Additionally, some studies indicate that the second-generation bioethanol derived from biomass can reduce carbon emissions by 90% [7]. To determine how an automobile can run entirely on ethanol, programs such as ETHANOL 100 have also been launched [8]. The need for bioethanol is growing as ethanol-fueled technologies are developed worldwide, which opens up the possibility of turning biowaste into bioethanol.

There are several challenges in biowaste conversion which include: variability of biowaste, lack of efficiency throughout the process, high initial cost and lack of policy and incentives support. First, the variability of biowaste is one of the biggest challenges as biowaste consists of food, agriculture residue and kitchen waste or other more. Each of these components contains distinct chemicals, some of which may not be suitable for use in bioethanol production. Furthermore, the pretreatment processes needed for these various chemicals varies, which could affect the components of the others. As a result, the process's efficiency is decreased.

Next, the overall lack of efficiency throughout the process mostly stems from the pretreatment, which can be highly energy intensive. This is particularly true for mechanical pretreatment as it might require a lot of energy to break down biowaste into tiny pieces. Other pretreatments, including chemical pretreatment, are less expensive but they also have neutralisation issues and can be harmful if improperly handled.

Moreover, the high initial cost of the specialized equipment is needed as there are many different components needed in bioethanol plants. This may result high costs, especially for initial capital, which makes it difficult for a plant to be built. Additionally, the incentives for doing so are low as landfills are often cheaper than bioethanol production. There are also no policies which can encourage businesses to adopt biowaste to bioethanol conversion. Hence, the lack of incentives and policy support makes converting biowaste into bioethanol less attractive compared to cheaper methods like landfill disposal.

One tropical fruit that is commonly found in Southeast Asia in Malaysia, Thailand, Myanmar, Indonesia and Vietnam is the Chempedak (*Artocarpus integer*). The significant waste of the Chempedak is its peels which have a thin greenish-yellow colour skin dotted with pentagons [1]. The Chempedak peel usually gets thrown away because it is useless and cannot be eaten. Chempedak peel has carbohydrates within it and can be fermented by yeast to produce ethanol and carbon dioxide [9]. This study provides a solution to both waste management and the production of alternative energy while attempting to solve the problems and challenges that may happen when converting biowaste to biofuel. Additionally, this project provides a secondary goal of optimizing the process and finding optimal methods to increase efficiency in the future.

2. Materials and Methods

2.1. Materials

The materials and chemicals used in this experiment can be seen in **Table 1**.

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Table 1. Chemicals used in this Project.

Materials & Chemicals
Chempedak Skin
Sulfuric Acid
Sodium Hydroxide
Sodium Citrate Dihydrate
Citric Acid
Cellulase
Brewing Yeast (*Saccharomyces cerevisiae)*

2.2. Pretreatment

To produce ethanol, Chempedak or *Artocarpus integer* skin is used. This is due to its abundance in Malaysia as well as its skin being considered waste after eating. Although, this skin is considered waste a lot of sugars, cellulose and starch is present in the chempedak approximately 19.75 ± 0.16%, 27.75 ± 0.06% and 4.12 ± 0.02% [9]. These sugars however are not accessible for ethanol fermentation yet as they are not in its monomeric glucose form.

The sugars found in cellulose are bonded by beta-1-4 glycosidic bonds and is used for the fruits cell walls [10]. Thus, it gives the fruits strong and rigid shape along with lignin and hemicellulose which are also found in cell walls. To be able to access these sugars, pretreatment will be done as it breaks down the cell walls from which allows the sugars found within it to be accessible as well as the cellulose itself turning into smaller sugar polymer chains.

2.2.1. Mechanical Pretreatment

Mechanical pretreatment was done by drying the Chempedak skin to remove its moisture content and was grinded to smaller pieces. Firstly, the Chempedak was cut to smaller pieces to be able to be placed in the oven. Initially, the skin was put in an oven for 60 °C for 6 hours as stated in a study [11]. However, the skin was still moist, thus, the skin was put in the oven for 120 °C for 2 hours which allows moisture to evaporate while the sugars are not burnt as they decompose at 180-185 °C [12]. The skin was then placed in a mechanical grinder and was sieved to obtain a size of 1mm [13]. 300g of this powder was collected to undergo three different pretreatment processes.

2.2.2. Acid Pretreatment

Acid Pretreatment is then done to break down the previously mentioned hemicellulose bonds allowing sugar to be processed further and the hemicellulose bonds being sugars as well [14] [15]. The powder is split into two with 100g each and is mixed into a sulfuric acid (H2SO4) solution of (2% w/w) and is placed for 2 and 3 hours at 160 °C each [16] [17]. After being treated, sodium hydroxide (1M) was added to neutralize the solution to the pH of 4.8 as the next process, hydrolysis, requires a pH of 4.8. Since the hemicellulose is broken, some sugars will be diluted in the water, thus, neutralisation was done instead of washing to not lose the sugar found within it [14]. The pH was then tested with a litmus paper to ensure that is within range.

2.2.3. Alkaline Pretreatment

Unlike acid pretreatment, alkaline pretreatment primarily targets the removal of lignin as it dissolves within the alkaline solution [18]. By doing so, the sugars will be more exposed when put through hydrolysis. Firstly, the sodium hydroxide solution is prepared by mixing the 4g of sodium hydroxide into a 100 ml solution to obtain a 1M solution with its sodium hydroxide content being 4% (w/w). This is then heated for 1 hour at 120 °C [19] [20].

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Since the sugars and hemicellulose are not broken down, they remain within the solid phase of the solution, thus washing is done until the liquid within the solution was clear. The pH was also measured until it reaches the pH of 7 [9].

2.3. Hydrolysis

Hydrolysis refers to a process where the sugars present will be broken down to simpler forms, mainly monomeric glucose, which can be fermented. By doing pretreatment, the hydrolysis process happens faster as these sugars are more accessible for the enzymes and have a higher yield of sugar as the other sugars that hydrolysis cannot break down are broken down by the pretreatment process. This is done with the help of an enzyme known as cellulase which breaks down the cellulose in the Chempedak skin. This enzyme is a combination of many enzymes which all aid in breaking down the different sugars found inside the Chempedak skin, the main enzymes are exoglucanase, glucosidase and endoglucanase [21].

The initial bonds that are broken down by the exoglucanase and the endoglucanase are those within and between the fibres of the chempedak skin which are formed by interactions between OH molecules. The exoglucanase breaks down the reducing bonds mainly aldehyde or ketone bonds as well as the non-reducing bonds which form a crystalline structure mainly consisting of acetals. This process, however, is mostly accomplished when pretreated [21].

The second process, on the other hand, involves the glucosidase which converts the shorter cellulose polymers into monomeric glucose [21]. The second process, on the other hand, involves the glucosidase which converts the shorter cellulose polymers into monomeric glucose [21].

2.4. Buffer and Enzyme Preparation

A buffer solution was prepared for the alkaline pretreatment as its pH is 7 which is not optimal for cellulase which has an optimum pH of 4.8. The buffer solution prepared was a citric buffer solution consisting of and citric acid and sodium citrate dihydrate with a molarity of 0.05M. Due to storage constrains, the solution was made 1 litre unlike the proposed solution of 1.5 litre, where the buffer will be placed in 800 ml and the alkaline pretreated solid phase is diluted to 200 ml. Due to storage constrains, the solution was made 1 litre unlike the proposed solution of 1.5 litre, where the buffer will be placed in 800 ml and the alkaline pretreated solid phase is diluted to 200 ml. The prior citrate buffers where prepared following the calculations in [22].

A cellulase solution is prepared by adding 100 ml to the amount of cellulase needed. Initially, impurities such as starch were expected to be present, thus it was refrigerated to separate the starch. However, there was no starch present in the cellulase we used allowing the cellulase solution to be added to the pretreated solution. Since the units of enzyme is unknown, it was assumed that there is 10000U/gr in the cellulase used.

During hydrolysis, an excess of enzymes will only quicken the process and will have no drawbacks. Since the units of enzyme is unknown and there is no disadvantage in adding excess cellulase, a total of 10g of enzyme was placed in the solution.

The 3 solutions are then put in an incubator at 50 °C for 72 hours. This was chosen as it is the optimum temperature for the enzyme and is the optimum time to fully hydrolyse the solution [20]. Two sample of each solution was taken every day to find the ideal time for the solution to hydrolyse. The 3 solutions are then put in an incubator at 50 °C for 72 hours. This was chosen as it is the optimum temperature for the enzyme and is the optimum time to fully hydrolyse the solution [20]. Two sample of each solution was taken every day to find the ideal time for the solution to hydrolyse.

2.5. Fermentation

Once glucose is obtained, the solution undergoes fermentation to produce ethanol. This process uses *Saccharomyces cerevisiae*, a fungus used when brewing alcohol. It grows best in acidic conditions

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with a pH between 4 and 6 and at temperatures ranging from 30 °C to 35 °C [23]. **Equation 1** shows the reaction is as below,

$$C_6H_{12}O_6(aq) \xrightarrow[yeast]{} 2C_2H_5OH(aq) + 2CO_2 \quad (1)$$

For each solution, 1 gram of yeast is added, and the container is sealed with a seal connected with a pipe to a beaker filled with water to see the carbon dioxide being produced. The seal is done to ensure anaerobic conditions which is crucial for ethanol production. If oxygen is present, microbes will break down carbohydrates more efficiently for energy, reducing ethanol production as they prioritize aerobic respiration [24].

The solution was placed in an incubator at a pH of 4.8 with a temperature of of 30 °C for 7 days [13] [25]. Similar to hydrolysis, excess yeast will help speed up the reaction and will have no negative effect on the process. Thus, 10 grams of yeast is added to the solutions.

2.6. Extraction

After fermentation, the majority of the glucose monomers have been converted into bioethanol. However, the solution remains impure, containing spent yeast, salt, water, and other organic compounds from the peel. Thus, distillation is required to extract the ethanol found. Traditionally, a distiller is used to obtain ethanol, it works by utilizing the difference in boiling points between ethanol and water. Ethanol, with a lower boiling point of 78.37 °C, evaporates first when the solution is heated to around 80 °C, below water's boiling point of 100 °C. However, when using a distiller, the temperature was not able to be controlled causing it not to evaporate. Thus, a rotary evaporator is used instead of which works under a similar premise.

A rotary evaporator also uses the difference in boiling points to extract ethanol, but it is done under a vacuum lowering the boiling point of the solution. In this case, the temperature of the water bath was at 60 °C with a vacuum of 175 mbar allowing the ethanol vapour to be at 40 °C with the condenser being set at 20 °C, allowing the ethanol vapour to travel to the receiving flask [26].

2.7. Data Collection and Analysis

The final ethanol concentration was tested with a refractometer, and hydrometer. The refractometer uses how much the sample refracts to measure its refractive index to find its concentration [27]. Next, a hydrometer is used where the solution can be placed in a graduated cylinder. The hydrometer used measures the alcohol percentage directly and works by the different specific gravities that are found with different alcohol percentages [28]. Additionally, testing was done on the glucose levels after hydrolysis with Benedict's Solution, in order to find which pretreatment method yielded the most glucose.

2.8. Glucose Testing

After hydrolysis, the resulting solution should contain sugars in monomeric form or glucose. This can be tested using Benedict's reagent and will be performed on three solutions to determine which yields the most glucose. Benedict's reagent is prepared by mixing 50 ml of an 8.7g Copper (II) sulphate pentahydrate (CuSO4.5H2O) solution with 400 ml of a solution containing 86.5g sodium citrate and 50g sodium carbonate which is then diluted to 500 ml [29].

To determine glucose concentration, 3 ml of Benedict's solution is mixed with 3 ml of the sample solution in a test tube. The mixture is heated in boiling water for 5 minutes and allowed to cool until the precipitate settles. The glucose concentration is indicated from the solution's colour, with red indicating the highest glucose content and blue showing no glucose present [30].

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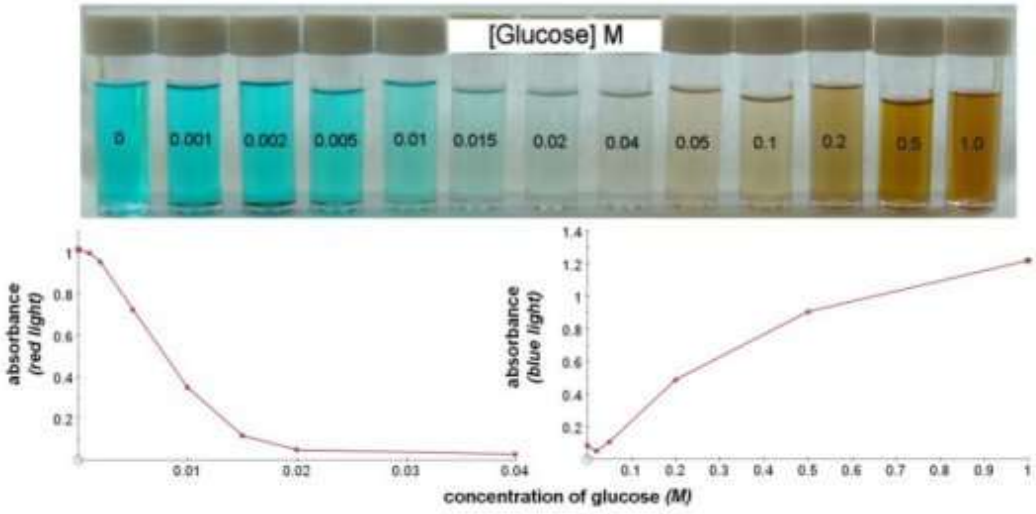

Figure 1. Benedict's Test Chart.

2.9. Refractometer Test

The refractive index of the distillate from the extraction was measured using DR-A1, ATAGO Abbe Refractometer to determine its ethanol concentration. Abbe refractometer uses the principle of prism refraction to measure the refractive index of solutions [31]. By observing and aligning refraction boundary line with the crosshairs, the refractometer displays the measurement value of the sample's refractive index and Brix percentage along with its temperature on the digital display screen [32]. To enhance accuracy, a calibration graph was created using standard ethanol solutions before measuring our samples. This approach ensures precise and tailored measurements while minimizing the reliance on generic data.

Before testing the distillate, a calibration graph of refractive index versus ethanol concentration was measured and plotted using refractive index with known ethanol concentrations of 20%, 40%, 60%, 80%, and 95%. This calibration graph serves as the guideline for determining the ethanol concentration in each of the distillates.

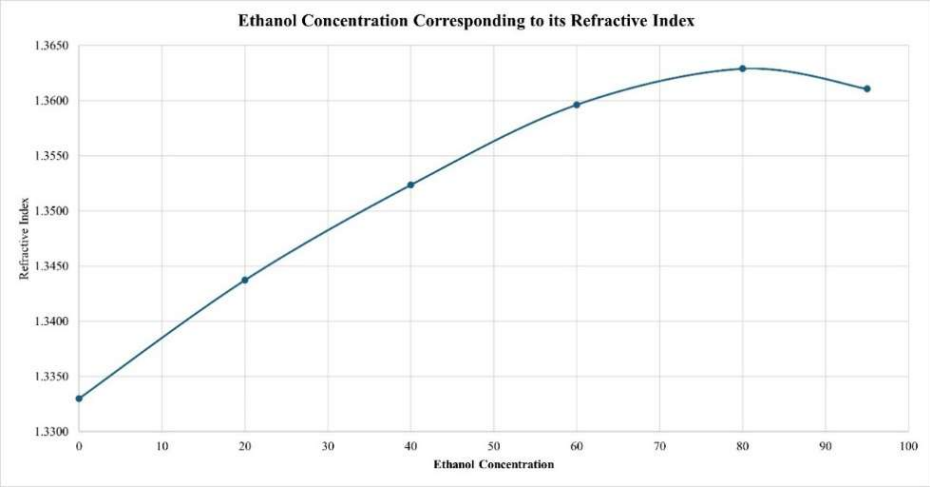

Figure 2. Graph of ethanol concentration corresponding to its refractive index.

2.10. Alcohol Hydrometer Test

The ethanol concentration of the distillate from the extraction was measured using an alcohol hydrometer. The alcohol hydrometer uses Archimede's principle of buoyancy to measure the specific

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gravity of alcoholic solutions which is directly proportional to its concentration [33] . By immersing the alcohol hydrometer into the sample, the concentration of ethanol can be directly determined by reading the value indicated on the hydrometer at the point where it aligns with the surface of the sample solution. This is because alcohol hydrometer is already pre-calibrated to directly display the ethanol concentration based on the solution's specific gravity. Therefore, unlike the previous two test, calibration is not required here. The scientific theory of the test method will be elaborated in the following section.

The alcohol hydrometer used in the lab consist of a set of 3 hydrometer, where the 1st alcohol hydrometer has an alcohol concentration scale from 0% to 40%, 2nd hydrometer has scale from 40% to 70% and the 3rd hydrometer has a scale from 70% to 100%. Depending on the concentration, all three were used in this experiment to find which scale represents the alcohol concentration.

3. Results and Discussion

3.1. Theoretical Yield

The theoretical final yield of ethanol depends on how effective each step is done from pretreatment to extraction. The theoretical yield will assume complete breakdown of lignin and cellulose from pretreatment, complete hydrolysis of the sugars into glucose, complete fermentation for every molecule of sugar and no losses during distillation. This also assumes the composition of Chempedak being as such [9] :

Table 2: Chempedak sugar Composition

Chempedak Composition	Weight (%)
Total sugars	19.750 ± 0.160
Cellulose	27.750 ± 0.060
Starch	4.120 ± 0.020

When converting the composition to glucose and then to ethanol, its molecular weight will be used to find the final calculated weight and volume of ethanol. The calculated result can be seen in the table below,

Table 3. Maximum Ethanol Yield.

Composition	Calculated Values
Glucose (in 300g of Chempedak)	154.860 g
Weight of Ethanol	79.197 g
Volume of Ethanol	100.377 ml

3.2. Glucose

A glucose test was conducted using Benedict's Solution where its colour would indicate the concentration of glucose within the solution. Visually, all the acid pretreatment with different time intervals resulted in a brick red colour which indicates the concentration of 1M of glucose. As for the alkaline pretreatment the time intervals also did not affect the colour, resulting in a concentration of around 0.2M of glucose.

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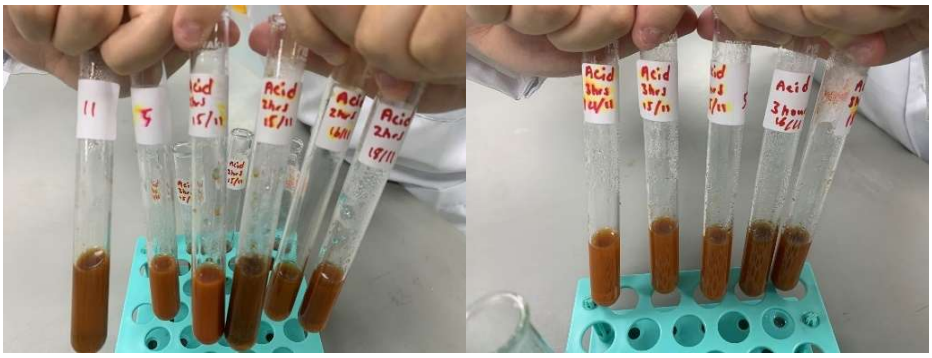

Figure 3. Benedict's Test on Acid Pretreated Solution.

Figure 4. Benedict's Test on Alkaline Pretreated Solution.

Since the acid pretreatment between 2 and 3 hours and the different time intervals produce similar results which is around 1M in glucose concentration, all the calculations for the samples will be made the same.

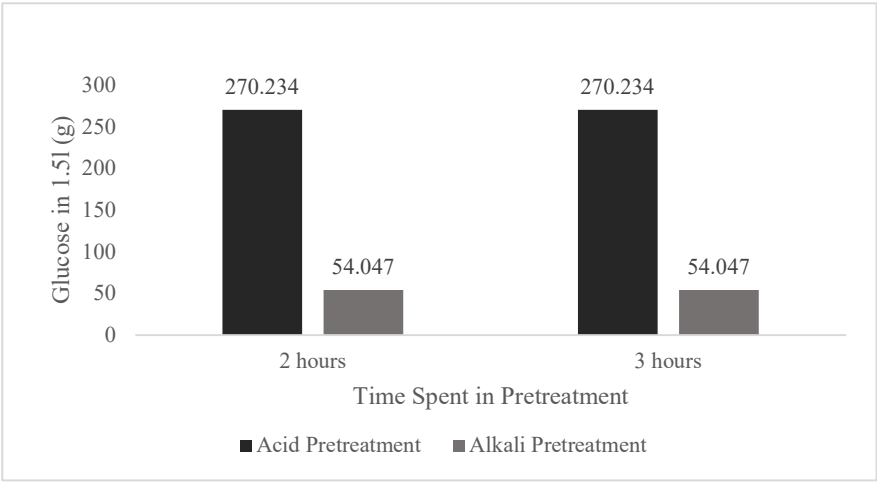

Figure 5. Effect of Time on Alkali and Acid Pretreatments.

As seen in the results, the test being done is not representative as $270.234g$ of glucose is not possible within a 100g sample of biomass. In addition to that, the results may be inaccurate as the solution has a naturally brown, red colour which may affect the colour, thus, making the brick red

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colour originating from the biomass rather than the reaction from the test. Therefore, the glucose test is unsuccessful, however, it may be concluded that the Acid Pretreatment broke down the most amount of sugar.

3.3. Extraction Sets

Multiple extractions were conducted in order to obtain a higher yield and higher amount of ethanol. The sets of extractions are stated below.

Extraction Set 1

- 1st Distillate: Distillate from 2 hours of extraction of fermented solution.
- 2nd Distillate: Distillate from 2 hours of extraction of distillate 1.
- 3rd Distillate: Distillate from 1 hour of extraction of distillate 2.
- 4th Distillate: Distillate from 0.5 hour of extraction of distillate 3.

Extraction Set 2

- 1st Distillate: Distillate from 3 hours of extraction of 1st set extraction distillate 1 waste.
- 2nd Distillate: Distillate from 2 hours of extraction of distillate 1.

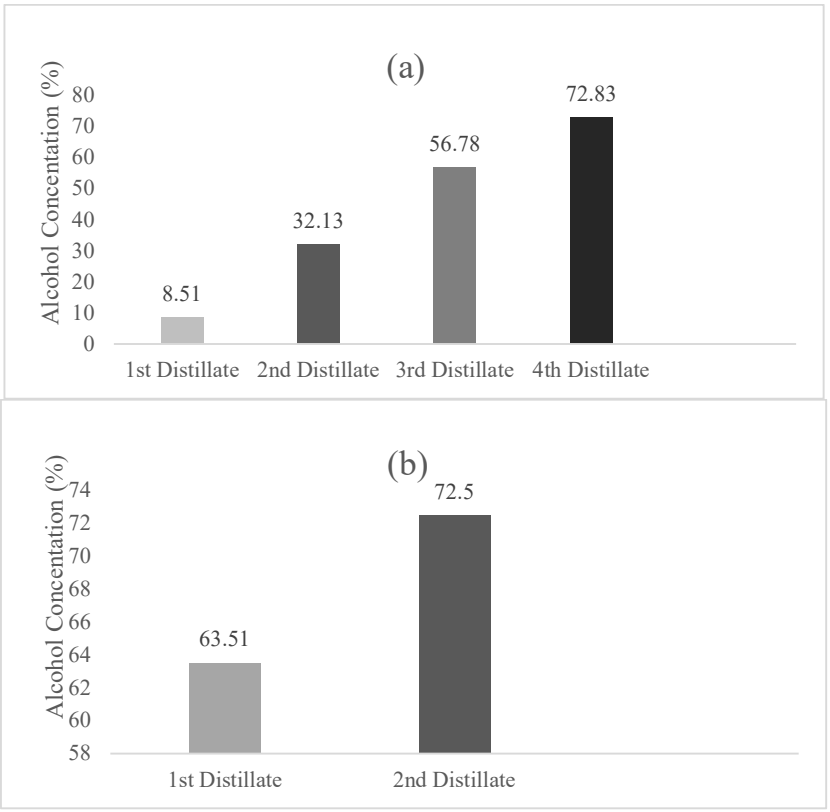
Extraction Set 3

- Combination of 4th distillate from extraction set 1 and 2nd distillate from extraction set 2
- 1st Distillate: Distillate from 1 hour extraction of set 3
- 2nd Distillate: Distillate from 0.5 hour of extraction of distillate 1.

3.4. Refractive Index Test

Based on ***Figure 2***, the refractive index values of the distillates obtained were calculated using **Equation 2** below to determine ethanol concentration. The results are tabulated below.

$$y = (-3 \times 10^{-8})x^3 + (9 \times 10^{-7})x^2 + 0.0005x + 1.3331 \qquad (2)$$



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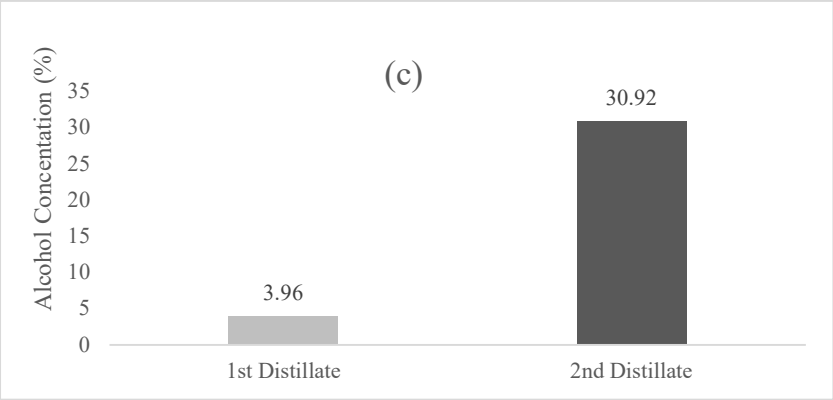

Figure 6. (a) 1st Distillate (b) 2nd Distillate (c) 3rd Distillate.

When calculating **Figure 6**, some results are doubled such as the 4th distillate of extraction set 1, and the 2nd distillate of extraction set 2, this is due to the quadratic equation which dictates its concentration, resulting in multiple results for certain data sets. Due to the azeotropic nature of water and alcohol mixture, it can be assumed that the lower percentage or the one presented was acquired.

3.5. Alcohol Hydrometer Test

The ethanol concentration of distillate was directly read from the scale on the alcohol hydrometer where it aligns with the sample's surface. Additionally, both extractions set 1 and set 2 distilled waste was measured to ensure that there was no ethanol content in it.

Unfortunately, the alcohol hydrometer was obtained only after completing extraction set 1 and set 2. Therefore, we were only able to measure the ethanol concentration of the final distillates from extraction set 1 and set 2. This limitation was also due to the small quantity of distillate obtained in each round from set 1 and set 2. They were directly used in subsequent extractions to avoid wastage, leaving no individual distillates for separate measurements. However, for extraction set 3, both distillates were successfully measured with the alcohol hydrometer, providing the final few results for this extraction process. The results are measured in **Figure 7** below.

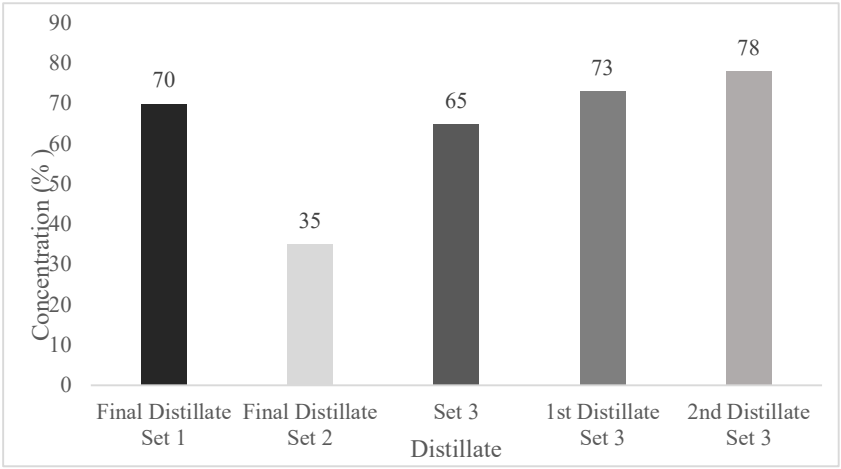

Figure 7. Alcohol Concentration recorded with a Hydrometer.

3.6. Comparison Between Refractive Index Test and Alcohol Hydrometer Test

Although the refractive index test provides a more comprehensive dataset, it has significant limitations that impact its result's accuracy. This test involves manual alignment of the shadow boundary with the crosshairs using the adjustment knob. Additionally, the differences in refractive

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index values for ethanol concentrations can vary by as little as 0.0001, making the test highly sensitive to small errors. Despite these limitations, the refractive index test produced a final concentration result of 72.95%, which closely aligns with the alcohol hydrometer test result of 73.0%, validating the reliability of the hydrometer readings.

In contrast, the alcohol hydrometer test eliminates the need for manual adjustments and complex polynomial calculations. Although it is calibrated for ethanol, its mechanism does not allow for extremely precise results as it assumes of a mixture of mostly pure ethanol and water. However, there are other compounds within the mixtures such as volatiles which may affect the overall specific gravity of the liquid, resulting in slightly off values.

In conclusion, the alcohol concentration selected was 41ml of 73% alcohol as this result was obtained by both the hydrometer and refractometer. This indicates that 29.93 ml of 100% ethanol is present in the final solution, which validates the success of the whole chempedak to bioethanol production process.

3.7. Analysis

After finding its final yield, to test the effectiveness of the process, its theoretical maximum yield will be compared to its actual yield. This is shown as such,

Table 4. Yield Efficiency.

Composition	Volume (ml)
Calculated Ethanol Volume	100.377
Actual Ethanol Volume	29.930
Yield Efficiency	29.818%

The yield efficiency was found to be 29.818%, indicating that the methods chosen were able to produce 29.818% of the theoretical maximum. This relatively low value may be due to various reasons such as inaccuracies in calculated ethanol volume as the exact amount of glucose within the Chempedak skin was not tested but was instead estimated. Another reason could be due to the mixing of both of the pretreated solutions together as the alkali pretreated mixture had a lower glucose concentration compared to the acid pretreated mixture. Due to the nature of the project, this was required to be done, however, to better observe the effectiveness of the pretreatment process, both should be isolated and done separately. Nevertheless, this experiment has achieved its goal of the conversion of bioethanol from Chempedak waste.

4. Failure Analysis and Limitations

Systematic error is a consistent difference between the obtained value and the actual value of something. It shifts data from its actual value resulting in false conclusions. During the feedstock collection and pretreatment stage, contaminants such as dirt and unclean water may be present. To mitigate this issue, proper cleaning techniques, including controlling the number of washes and using a consistent and appropriate amount of water.

In the mechanical pretreatment step, feedstock loss may occur during the transfer from the grinder to the beaker. This loss can reduce overall yield and cause inaccuracies in calculations. To minimize this issue, the transfer pathway from the grinder to the beaker should be enclosed to prevent spillage.

For the chemical pretreatment process, uneven mixing can lead to suboptimal chemical reactions and incomplete treatment. This can be improved by using an overhead mechanical stirrer to ensure uniform mixing throughout the reaction. Additionally, residual alkaline may not be fully washed or neutralized out after pretreatment. This can be mitigated by washing the feedstock and confirming neutrality through pH measurements.

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During the hydrolysis process as well as the buffer and enzyme preparation, enzymes may be used under non-optimal temperature or pH conditions which reduces effectiveness. To prevent this, the mixture should be constantly monitored to ensure optimal pH and temperatures,

Similarly, on the fermentation process, non-optimal temperature, pH, or oxygen levels will reduce the amount of fermentation occurring as the yeast will use the oxygen present instead. The use of an incubator can help maintain optimal temperature and ensuring proper sealing will ensure anaerobic conditions.

Additionally, experiment procedures can be improved to optimize in finding the true amount of ethanol produced for each pretreatment, as this experiment fails to do so due to the project scope requirements of maximizing yield and not a comparison. Along with that, testing can be improved for glucose as the result obtain was not possible, this can be done by dilution to ensure other precipitates do not take over the brick-red colour. A different test that is able to quantify the glucose levels such as glucose test strip or specific refractometers.

Lastly, random errors may also occur on testing and procedural equipment, such as an inaccurate temperature and pH reading. This can be mitigated by repeating the experiment multiple times to ensure consistent results.

5. Future Perspective

The success of our small-scale bioethanol production serves as a foundation for future advancement in turning biowaste into renewable energy. From our experiment we have found various alternatives to convert chempedak peel to bioethanol, with some producing better yield than others. Refining and optimization will be essential in improving yields and lower production cost. If there is more capital, more experiment can be conducted to find the best steps to each process such as enhanced pretreatment methods, increased enzymic efficiency and more.

Due to capital and time constraints, chempedak peel is chosen for their high cellulose content. Diversifying in feedstocks would be the next step. By exploring other biowaste feedstocks such as fruit peels, and agricultural residue can broaden the systems applicability and reduce dependency on a single material. Developing an automated system will further enhance the system. Sensors can be equipped to detect and sort different biowaste types, adjusting enzymes and chemicals inputs accordingly for optimum performance. By integrating machine learning, it allows the system to learn from previous processes and improve its efficiency over time.

Once everything is optimized, the next step will be to scale up the system to commercial applications. This includes larger reactors and establishing infrastructure for biowaste collection and ethanol distribution while meeting environmental and safety standards. In addition, bioethanol's versatility allows it to be used across many industries, including in cosmetics, healthcare, food and beverage, automotive, and pharmaceutical sectors, expanding its market potential. In the future, this system could be integrated with other renewable energy sources to create a diversified and sustainable energy source. With continued research and development, production of bioethanol from biowaste can contribute significantly towards building a more sustainable future and circular economy.

6. Conclusion

In conclusion, this project addresses the challenges of biowaste management by focusing on converting Chempedak (Artocarpus Integer) peel into bioethanol which is a renewable energy source. By employing a structured design-thinking approach, we refined each step of the ethanol production process, including feedstock collection and pretreatment (mechanical, acid and alkaline pretreatment), hydrolysis, fermentation and ethanol extraction.

The key findings of this project include the successful optimization of mechanical pretreatment by turning chempedak peel into smaller pieces and then into powder form of size $1mm^3$ and dehydrated in oven at 120 °C for 2 hours, which effectively removed moisture while preserving

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sugars. Acid pretreatment proved more effective than alkaline pretreatment in enhancing sugar availability for hydrolysis. Fermentation using yeast in sealed containers efficiently produced ethanol, as indicated by active carbon dioxide generation. Additionally, ethanol recovery was significantly improved through the use of a rotary evaporator operating at 60 °C under vacuum, resulting in a final ethanol concentration of 7% and a yield of 41ml. Our method is unique because it turns an environmental burden into a useful resource by using Chempedak peel. When compared to traditional approaches, our process integrates cost-effective mechanical advances and scalable extraction processes, making it for greater commercial applicability.

This project aligns with multiple United Nations Sustainable Development Goals (SDGs). It contributes to SDG 7 (Affordable and Clean Energy) by generating bioethanol to reduce fossil fuel dependency, supports SDG 12 (Responsible Consumption and Production) by transforming waste into valuable energy, and advances SDG 13 (Climate Action) by mitigating methane emissions and providing cleaner combustion fuels.

Through the management of biowaste and the creation of renewable energy, this project promotes environmental responsibility and lessens reliance on landfills while providing sustainable energy options that residents and businesses may use. Along with lowering carbon footprints and fostering economic opportunities, it improves societal well-being. This project demonstrates innovation in sustainable technology by integrating biowaste management with renewable energy production. It reinforces the potential for bioethanol derived from chempedak peel waste to support the circular economy, inspiring future research and commercialization in clean energy solutions.

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References

1. R. A. S. Sivakumar, "DETERMINANTS OF SUSTAINABLE WASTE MANAGEMENT PRACTICES," University Utara Malaysia, Kedah, 2024.
2. C.C. Bong et al., "Towards low carbon society in Iskandar Malaysia: Implementation and feasibility of community organic waste composting," Journal of Environmental Management, vol. 203, no. 2, pp. 679-687, 2017.
3. Salwa Khamis et al., "Characterization of Municipal Solid Waste in Malaysia for Energy Recovery," IOP Conference Series: Earth and Environmental Science, vol. 021003, p. 264, 2019.
4. A. Siddiqua et al., "An Overview of the Environmental Pollution and Health Effects Associated with Waste Landfilling and Open Dumping.," Environmental Science and Pollution Research, vol. 39, p. 29, 2022.
5. Klein E. Ileleji and Nathan S. Mosier, "How Fuel Ethanol Is Made from Corn," Bioenergy, vol. 1, pp. 379-384, 2015.
6. H. Zhai et al., "Comparison of Flexible Fuel Vehicle and Life-Cycle Fuel Consumption and Emissions of Selected Pollutants and Greenhouse Gases for Ethanol 85 Versus Gasoline," Journal of the Air & Waste Management Association, vol. 59, no. 8, pp. 912-924, 2009.
7. Hasbi Priadi et al., "Techno-enviro-economic analysis of second-generation bioethanol at plant-scale by different pre-treatments of biomass from palm oil waste.," Energy Conversion and Management, vol. X, p. 100522–100522, 2024.
8. P. Bielaczyc et al., "Ethanol as an automotive fuel," Combustion Engines, vol. 3, pp. 39-45, 2016.
9. A. A. Sundarraj & T.V. Ranganathan, "Physicochemical characterization of Jackfruit (Artocarpus integer (Thumb.).) Peel.," Research Journal of Pharmaceutical Biological and Chemical Sciences,, vol. 8, no. 3, p. 2285–2295, 7 6 2024.
10. L. P. Johannes & T. D. Xuan., "Comparative Analysis of Acidic and Alkaline Pretreatment Techniques for Bioethanol Production from Perennial Grasses," Energies, vol. 5, p. 17, 2024.

 doi:10.20944/preprints202608.1114.v1

11. D. M. D. J. Miss-Zacarías et al., "Spray Drying of Jackfruit (Artocarpus heterophyllus Lam.) Seeds Protein Concentrate: Physicochemical, Structural, and Thermal Characterization.," Processes, vol. 13, no. 7, 2025.
12. J. W. Lee et al., " Investigation of Thermal Decomposition as the Kinetic Process That Causes the Loss of Crystalline Structure in Sucrose Using a Chemical Analysis Approach (Part II).," Journal of Agricultural and Food Chemistry, vol. 59, no. 2, p. 702, 2011.
13. Irhamni et al., "Bioethanol production from durian shell wastes by saccharification and liquefaction processes," In International Journal of Multidisciplinary Research and Development., vol. 5, no. 6, pp. 48-49, 2018.
14. S. Kumar &. S. Jain, "Advances and challenges in pretreatment technologies for bioethanol production," A comprehensive review. Sustainable Chemistry for Climate Action, vol. 5, p. 100053, 2024.
15. Javanmard et al., "Breaking Barriers for a Green Future: A Comprehensive Study on Pre-treatment Techniques for Empty Fruit Bunches in the Bio-Based Economy.," Process Safety and Environmental Protection, vol. 182, pp. 535-558, 2024.
16. F. R. Amin et al., "Pretreatment methods of lignocellulosic biomass for anaerobic digestion.," AMB Express, vol. 7, no. 1, 2017.
17. V. Ashokkumar et al., "Recent advances in lignocellulosic biomass for biofuels and value-added bioproducts," A critical review. Bioresource Technology, vol. 344, p. 126195, 2022.
18. G. Brodeur et al., "Chemical and Physicochemical Pretreatment of Lignocellulosic Biomass: A Review.," Enzyme Research, pp. 1-17, 2011.
19. D. Madyire &. K. O. Olatunji, "Alkali Pretreatment of Lignocellulose Feedstock Improves Morphological Structure and Biomethane Yield.," Sustainability, vol. 17, no. 2, p. 534, 2025.
20. A. K. Obeng et al., "Improved glucose recovery from durian peel by alkaline-catalyzed steam pretreatment.," PeerJ, vol. 9, p. 12026, 2021.
21. S. Jayasekara & R. Ratnayake, "Microbial Cellulases: An Overview and Applications.," Cellulose, 2019.
22. AAT Bioquest, "Citrate Buffer (pH 3.0 to 6.2) Preparation and Recipe," AAT Bioquest, [Online]. Available: https://www.aatbio.com/resources/buffer-preparations-and-recipes/citrate-buffer-ph-3-to-6-2.
23. N. V. Narendranath & R. Power, "Relationship between pH and Medium Dissolved Solids in Terms of Growth and Metabolism of Lactobacilli and Saccharomyces cerevisiae during Ethanol Production.," Applied and Environmental Microbiology, vol. 71, no. 5, p. 2239–2243, 2005.
24. D. J. Kelly et al., Microaerobic Physiology: Aerobic Respiration, Anaerobic Respiration, and Carbon Dioxide Metabolism, vol. 10, ASM Press, 2001.
25. S. I. Dhas and M. Yuvarani, "Synthesis of Bioethanol from Artocarpus Heterophyllus Peel by Fermentation using Saccharomyces Cerevisiae at Low Cost," Global Research and Development Journal for Engineering, vol. 2, no. 12, 2017.
26. University of Wollongong, "Buchi 20/40/60 rule for Rotary Evaporators," University of Wollongong, 2005.
27. Y. Plugatar, "Prediction of Ethanol Content and Total Extract Using Densimetry and Refractometry," Beverages, vol. 9, no. 2, p. 31, 2023.
28. A. L. J. Rico, "Determining the Quality and Quantity of Bioethanol Production using Golden Shower (Cassica fistula) Fruit," Baghdad Science Journal, vol. 18, no. 1, 2021.
29. C. L. Borgford and L. R. Summerlin, "Chemical of Foods," in Chemical Activites, Washington, D.C., American Chemical Society, 1988, p. 253.
30. J.-S.. Kim et al., "A study on detection of glucose concentration using changes in color coordinates.," Bioengineered, vol. 8, no. 1, pp. 99-104, 2016.
31. F. K. Coradin et al., "Etched fiber bragg gratings sensors for water-ethanol mixtures: a comparative study.," Journal of Microwaves, Optoelectronics and Electromagnetic Applications, vol. 9, no. 2, p. 131–143, 2010.
32. JJS, "Atago Abbe Refractometer - DR-A1.," JJS, 2026. [Online]. Available: https://www.jjstech.com/dr-a1.html?srsltid=AfmBOoouhy9Ai0-tS5e51laT6o9jOJzrJt1dS9AFnvo5jsNMvXEEb4Zw.
33. S. Lorefice & A. Malengeo, "Calibration of hydrometers.," Measurement Science and Technology, vol. 17, no. 10, pp. 2560-2566, 2006.

 doi:10.20944/preprints202608.1114.v1