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Article

Chemical Synthesis of Silver Nanoparticles: A Comparative Study of Antibacterial Properties

Selection of Chemical Reduction Methods for Synthesizing AgNPs Affects the Bactericidal Properties

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Abstract

Background: Recent studies have suggested significant antimicrobial properties of silver nanoparticles (AgNPs), offering a ray of hope during the height of antibiotic resistance. However, their efficacy largely depends on the particle size and colloidal stability. Although higher stability of AgNPs is associated with improved antimicrobial activity, excessive stability weakens their reactivity with the bacterial membrane. The objective of the research is to evaluate different reducing agent combinations to achieve optimal particle size and colloidal stability for maximum bactericidal efficacy. **Materials and Methods:** The synthesis of AgNPs was carried out using silver nitrate using five different chemical reduction methods to compare their antimicrobial activity. After purification, the freeze-dried AgNPs were characterized by UV-visible spectroscopy, thermogravimetric analysis (TGA), scanning electron microscopy (SEM), and dynamic light scattering (DLS). The minimum bactericidal concentration (MBC) of the AgNP formulations were investigated by plate counting method against the methicillin-resistant *E. coli*. Finally, the synthesized AgNPs were tested against the resistant strains of *E. coli*, *Salmonella*, *Klebsiella*, *Bacillus*, and *Staphylococcus* by the well diffusion method to evaluate and compare their inhibition zones. **Results and Discussions:** The average particle size for different formulations of nanoparticles ranges from 30.91 nm to 93.85 nm, while the zeta potential ranges from -8.0 mV to -41.1 mV. The minimum bactericidal concentration (MBC) for all AgNP formulations was determined against gram-negative *E. coli*, and AgNPs synthesized with only trisodium citrate were found to be the most effective with 99.75% bactericidal efficacy at 20 ppm concentration due to their optimal particle size and stability. However, AgNPs synthesized with polyethylene glycol and polyvinyl pyrrolidine were found to be the most effective against the resistant bacterial strains in the well diffusion assay. **Conclusion:** The reducing agents affect the particle size and stability of synthesized AgNPs, resulting in significant variations in their antibacterial activity, warranting further study.

Keywords: silver nanoparticle; chemical reduction; reducing agent; particle size; colloidal stability; antibacterial activity

1. Introduction

Antimicrobial resistance continues to reduce the effectiveness of conventional antibiotics, reviving interest in alternative or adjunctive antimicrobials, including silver in nanoscale form [1,2]. Silver nanoparticles (AgNPs) exhibit broad-spectrum antibacterial activity and have been explored in wound care, device coatings, and as potentiators of existing antibiotics [3]. Their nanoscale dimensions confer a high surface-area-to-volume ratio and size-dependent physicochemical properties that can enhance antimicrobial effects relative to bulk silver [4,5]. AgNPs provides antibacterial activity through the sustained release of silver ions, which interact with bacterial membranes, and bind to ribosomal subunits or key enzymes to inhibit protein synthesis [6,7].

The antibacterial performance of AgNPs is not an inherent property of silver but a function of particle size, morphology, surface chemistry, and colloidal stability [8,9]. Parameters such as core size and dispersity (nucleation or growth control), capping environment, and zeta potential affect the nanoparticles approach to adhere and disrupt bacterial envelopes [8–12]. While higher colloidal stability generally preserves dispersion, bioavailability, and shelf life, excessive stabilization can decrease reactivity at the cell-nanoparticle interface [13–16]. Thus, identifying formulations that balance stability and reactivity is the key to optimizing bactericidal efficacy [14,17].

Chemical reduction remains the most accessible and scalable route to AgNPs [18]. However, the choice and combination of reductants and stabilizers—such as trisodium citrate, sodium borohydride (NaBH_4), polyethylene glycol (PEG), polyvinylpyrrolidone (PVP), and oxidants like hydrogen peroxide (H_2O_2)—affect the nucleation rates, growth pathways, particle yield, size distribution, and surface charge [19,20]. Literature reports often contradict one another regarding structure–property–activity relationships because synthesis variables are not systematically controlled across methods [14,21,22]. A comparison across different AgNP formulations, physicochemical characterization, and microbiological testing is required to clarify how specific reduction chemistries affect antibacterial outcomes.

To address this gap, AgNPs was synthesized via five different chemical routes—(i) citrate-only reduction, (ii) NaBH_4 reduction, (iii) NaBH_4 with citrate as a secondary reductant, (iv) citrate/ NaBH_4 in the presence of H_2O_2 , and (v) a PEG/PVP-assisted method—followed by standardized purification and characterization [23–27]. Bactericidal performance was evaluated by minimum bactericidal concentration (MBC) assays against methicillin-resistant *Escherichia coli*. The antibacterial activity was tested against *Escherichia coli*, *Salmonella*, *Klebsiella*, *Bacillus*, and *Staphylococcus*, using agar well diffusion method.

The present study is aimed to synthesize silver nanoparticles (AgNPs) using different reducing agents and assess their antimicrobial activity against methicillin-resistant *E. coli*. The research examines how different sizes and shapes of nanoparticles affect the way they interact with bacterial cells, ultimately aiming to find the most suitable features that would allow nanoparticles to kill bacteria most effectively. The objective of the research is to co-optimize size and stability of chemically synthesized AgNPs using different formulation approaches to achieve reliable antibacterial outcomes.

2. Materials and Methods

2.1. Materials

Silver nitrate (AgNO_3) was collected from Honeywell Fulka, polyvinylpyrrolidone (PVP 40000), Polyethylene glycol, and sodium borohydride from Sigma-Aldrich. In addition, hydrogen peroxide and trisodium citrate was purchased from Roth, whereas ethanol and ammonium hydroxide were purchased from Merck. All chemicals and reagents were used as received without further purification. Water used in all procedures was obtained from a water purification system (Purelab from ELGA) and had a measured resistivity of $18.2 \text{ M}\Omega \text{ cm}^{-1}$.

2.2. Instruments

Experiments were conducted using various instruments like UV-vis double beam spectrophotometer (PG Instrument, UK T60 U), Scanning Electron Microscope (SEM) (Model Joel JSM-7600FE, Japan). In addition, Ultra Sonicator (Power-Sonic, Korea), Hotplate Stirrer (Stuart), Micro Centrifuge Machine (Tomy, MX-305), Orbital shaker Incubator (JSR) and Laboratory incubator (Incucell), Particle Size Analyzer (Malvern), Freeze Dryer, TGA (Hitachi instrument - TG/DTA 7200) are also used as per manufacturer's instructions.

2.3. Experimental Methods

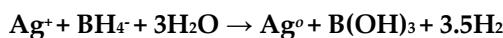
2.3.1. Formula One (F1). AgNP Synthesis Using Trisodium Citrate as a Reducing Agent

150 ml of 7 mM Trisodium Citrate solution was prepared in a beaker at room temperature and placed on a magnetic stirrer. The solution was first brought to boil while being stirred and then 1.5 ml of 0.1 M aqueous silver nitrate was added to start the reaction. The color of the reaction was observed to change slowly from colorless to yellow, then turbid. At last, no more change took place indicating that the reaction was completed [23].



2.3.2. Formula Two (F2). AgNP Synthesis Using Sodium Borohydride as Reducing Agent

About 30 ml of 2 mM Sodium Borohydride solution was prepared and chilled at 4°C for 20 minutes. The solution was poured into a beaker and placed on a magnetic stirrer. A 2.5 ml solution of 0.01M Silver Nitrate was added dropwise to the beaker while the mixture was continuously stirred. The solution turned pale yellow after only a few drops and brighter yellow after all the solution had been poured, following which stirring was stopped [24].



2.3.3. Formula Three (F3). AgNP Synthesis by a Combined Method Using Sodium Borohydride as Primary Reductant and Trisodium Citrate as Secondary Reductant

A mixture of 72 mL of distilled water, 2 mL of 0.01M trisodium citrate, and 2 mL of 0.01 M silver nitrate was stirred for 3 minutes at 10°C. Then, 3 mL of 0.002 M sodium borohydride was added to the mixture, and immediately after the addition, stirring was stopped. Following that, the color of the solution turned golden yellow, indicating the formation of nanoparticles [25].

2.3.4. Formula Four (F4). AgNP Synthesis by a Combined Method Using Trisodium Citrate and Sodium Borohydride as Reducing Agent in the Presence of Hydrogen Peroxide

At first, an aqueous solution combining 0.05 M, 50 µL silver nitrate, 0.075 M, 0.5 mL trisodium citrate, and 30% (v/v) 20 µL H₂O₂, each prepared separately, was stirred vigorously at room temperature to allow homogenous mixing. Then 100 mM, 250 µL sodium borohydride was injected into the solution mixture to initiate the reaction. Initially, the color of the solution was light yellow, which gradually became deep reddish yellow after 3-4 minutes under continuous stirring [26].

2.3.5. Formula Five (F5). AgNP Synthesis Using Polyethylene Glycol as a Reducing Agent

0.5 g of polyethene glycol (PEG) was completely dissolved in 60 mL of distilled water under magnetic stirring by heating at 120°C. Then, 1.5 g of Polyvinyl pyrrolidine (PVP) and 0.05 g AgNO₃ were added to the solution and stirred for 45 minutes, continuing heating. The solution's color slowly changed from light to brighter yellow, and the stirring was stopped [27].

2.4. Purification

The previously synthesized silver nanoparticle has been purified using centrifugation at 14000 rpm for 15 minutes. For all methods where sodium borohydride was used as a reducing agent, the nanoparticle formulation was freeze-dried before centrifugation to reduce two-thirds of the volume. After centrifugation, the supernatant was discarded, and the pellet was resuspended in 80 ml ethanol and centrifuged again at 14000 rpm for another 15 minutes. Finally, the pellet was collected.

In the formula where PEG was used as a reducing agent, the pellet obtained after the first centrifugation was suspended in 150 ml acetone and centrifuged at 14000 rpm for 15 minutes. After that, a small amount of orange floating pellet appeared at the bottom of the falcon tube, carefully collected by discarding the supernatant from the top. Then, 20 ml of ethanol was added to the pellet and centrifuged under the same parameters, and the final pellet was collected.

2.5. Chemical Characterization

The optical absorption spectra for nanoparticles in colloidal preparation were recorded using a double beam PG UV-Visible spectrophotometer, UK-T60 U. Rectangular quartz cells of path length 1 cm were used throughout the investigation. The Malvern DLS measurement Instrument measured the size and size distribution of the nanoparticles synthesized in the colloidal solution. The same equipment was used to determine zeta potential (ζ) for a surface charge of Ag nanoparticles. The cuvette used in this experiment was the universal dip cell. The Huckel equation was used to calculate the electrophoretic mobilities of silver nanoparticles.

Morphological analyses of nanoparticles were carried out using the JEOL analytical scanning electron microscope (SEM) model JSM-7600FE. Dried purified AgNP powder was spread on the carbon-coated aluminum stubs for the measurements at an acceleration voltage of 20 kV. TGA was studied by a Hitachi instrument (TG/DTA 7200) in the 20–800°C temperature range under a Nitrogen gas atmosphere at a per-minute heating rate of 10°C.

2.6. Microbiological Characterization of AgNP Formulations

The antimicrobial studies of this research were carried out in the Laboratory of Microbiology of the Department of Mathematics and Natural Sciences of BRAC University. The antibacterial activities of AgNPs synthesized by five different methods were compared through the well diffusion method. The efficiency of the same concentration of silver nitrate was also measured, as silver nitrate was the basis of silver nanoparticle synthesis in all five methods. Apart from that, the Minimum Bactericidal Concentration (MBC) of five nanoparticles was determined against an *E. coli* isolate.

2.6.1. Collection of Bacterial Strains

Klebsiella spp. was collected from Mohakhali TB Hospital, and a few sample strains of *E. coli* were collected at Uttara Adhunik Medical College Hospital. *Salmonella Typhi*, *Staphylococcus* spp., and *Bacillus* spp. were also collected from BRAC University Microbiology laboratory. These strains were subcultured in Nutrient Agar Media every week without fail, ensuring their viability and preventing contamination. The freshly cultured colonies were then used in the procedure, providing reliable and consistent results.

2.6.2. Well Diffusion Assay

Mueller-Hinton agar was utilized as the culture medium for the agar well diffusion assay. two isolates of *Klebsiella* species, two isolates of *E. coli*, one isolate of *Bacillus* species, one isolate of *Salmonella* species, and two isolates of *Staphylococcus* were inoculated onto a separate petri dish. In every dish, six wells were created to assess the antimicrobial activity of five different nanoparticle formulations and a silver nitrate solution, all at the same concentration.

Here, same quantity of silver nitrate was incorporated into each silver nanoparticle synthesis, ensuring that the presumed concentration of silver nanoparticles remained consistent across all

samples, assuming complete conversion of silver nitrate to nanoparticles. The nanoparticle concentration for each method was standardized to approximately 80 ppm, and a silver nitrate solution of matching concentration was also prepared. For each well, 40 μ l of the respective nanoparticle solution was added, with the final well receiving an equal volume of silver nitrate solution. Following overnight incubation, the zones of inhibition produced by each nanoparticle type and the silver nitrate were measured to evaluate antimicrobial efficacy (Figure 1).

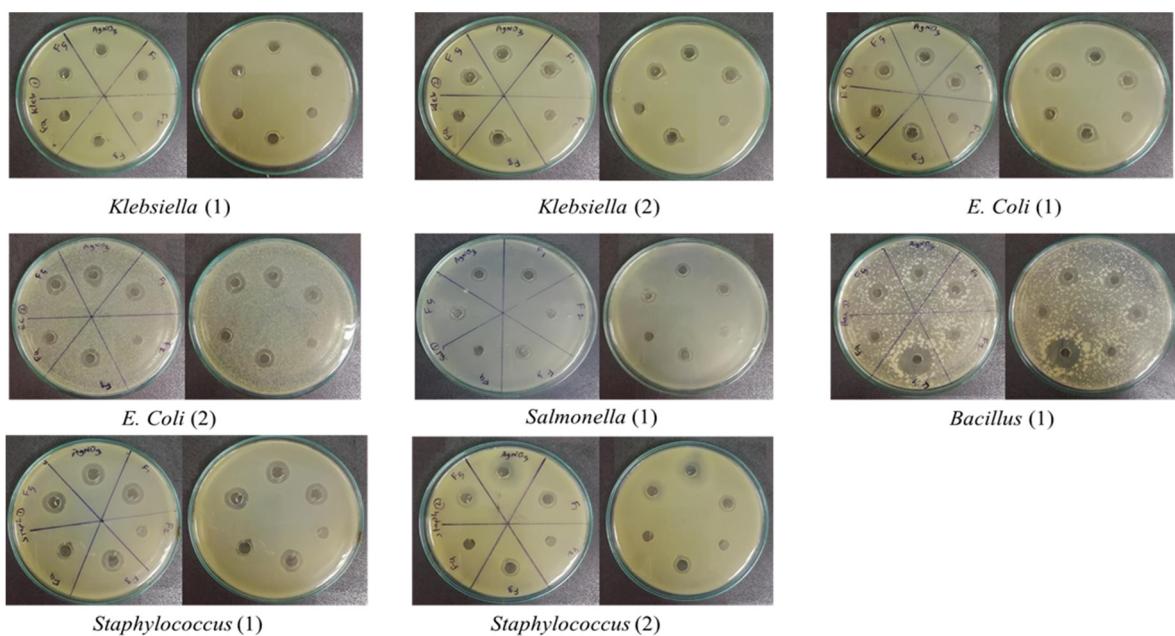


Figure 1. Zone of inhibition of AgNPs formulations against different bacterial strains.

2.6.3. Determination of Minimum Bactericidal Concentration (MBC)

The methicillin-resistant *E. coli* culture was grown on Luria Bertani broth till the desired range of Optical Density was reached. 1 ml of culture was diluted ten times before using the organism in a series of test tubes containing different concentrations of AgNPs. For each type of AgNPs formulations, a series of test tubes containing Luria Bertani broth was prepared using the serial dilution method. Five test tubes contained 0 ppm, 5 ppm, 10 ppm, 20 ppm, 40 ppm of AgNPs, and an additional test tube contained only LB broth. 50 μ l of the cell from the diluted culture was applied to each test tube except the last one, which contains LB broth only. After 2 hours of incubation, 5 μ l from each test tube were spread into separate petri dishes containing nutrient agar. This step was followed by overnight incubation before counting the colonies per petri dish (Figure 2). The LB broth was cultured to ensure a contamination-free procedure in determining MBC for each type of AgNPs formulation.

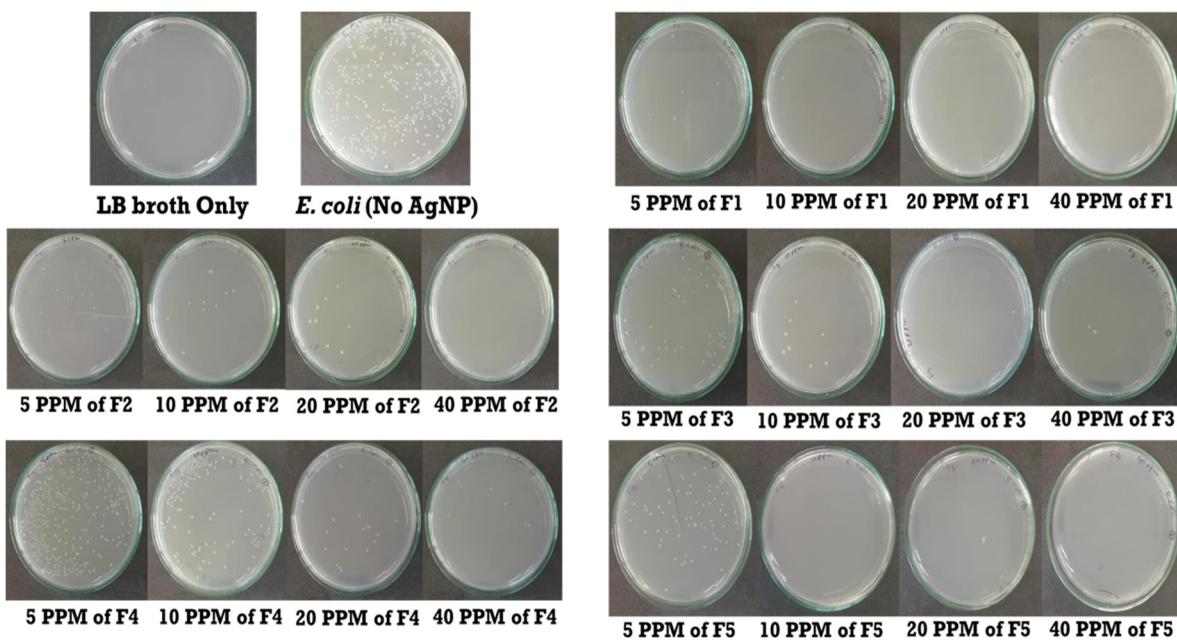


Figure 2. Counting the number of surviving *E. coli* colonies assayed against different concentrations of AgNP formulations.

3. Results and Discussions

3.1. Quantitative Analysis

Due to surface plasmon resonance, Ag nanoparticle shows a strong absorption peak around 400 nm (Figure 3). Formulation of AgNPs produced from different methods showed absorption peaks exactly in that region which confirmed nanoparticle formation.

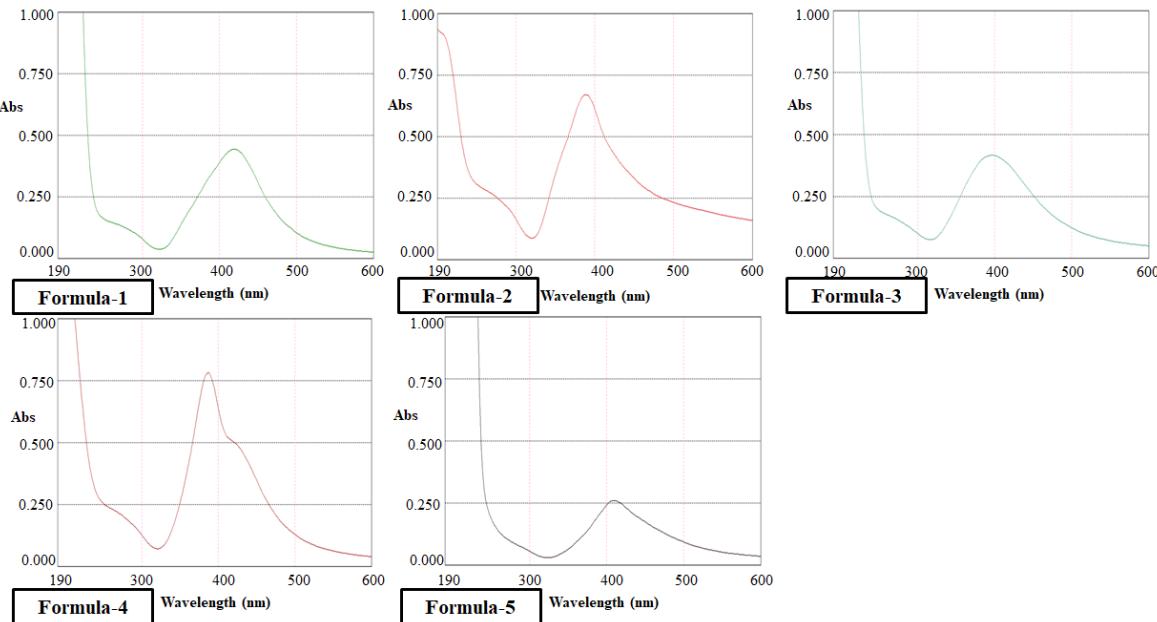


Figure 3. UV-visible absorption spectra of AgNP formulations.

Formulation four due to the presence of H_2O_2 showed the highest absorbance and the second highest absorbance was found for formulation one. This confirmed the highest abundance of nanoparticle formation for these two methods. Analysis for Total Dissolved Solids (TDS) by TDS Meter (Table 1) also supports the change in AgNPs concentration among different formulations i.e., $\text{F2} < \text{F5} < \text{F3} < \text{F1} < \text{F4}$.

Table 1. Electrical Conductivity, Total Dissolved Solid, Zeta potential and Particle size of synthesized AgNP Formulations.

Formula (Reducing Agent)	Electrical Conductivity	Total Dissolved Solid (Solution Volume = 30 ml)	Zeta Potential	Particle Size (Diameter)
Formula-1 (TSC)	818 μ S	314 ppm	-39.4 mV	87.15 nm
Formula-2 (NaBH_4)	848 μ S	176 ppm	-29.1 mV	91.71 nm
Formula-3 (NaBH_4 & TSC)	779 μ S	297 ppm	-41.1 mV	65.07 nm
Formula-4 (NaBH_4 & TSC, H_2O_2)	758 μ S	609 ppm	-36.7 mV	30.91 nm
Formula-5 (PEG & PVP)	871 μ S	191 ppm	-8.0 mV	93.85 nm

3.2. Thermogravimetric Analysis

TG thermograms of AgNP (Figure 4) provided information about the percent weight loss of silver nanoparticles upon increasing temperature. To decompose the AgNPs, the samples were heated from 20° C to 800° C; at 270 - 350°C the decomposition of the samples starts and its size decreases gradually up to 470 - 590°C.

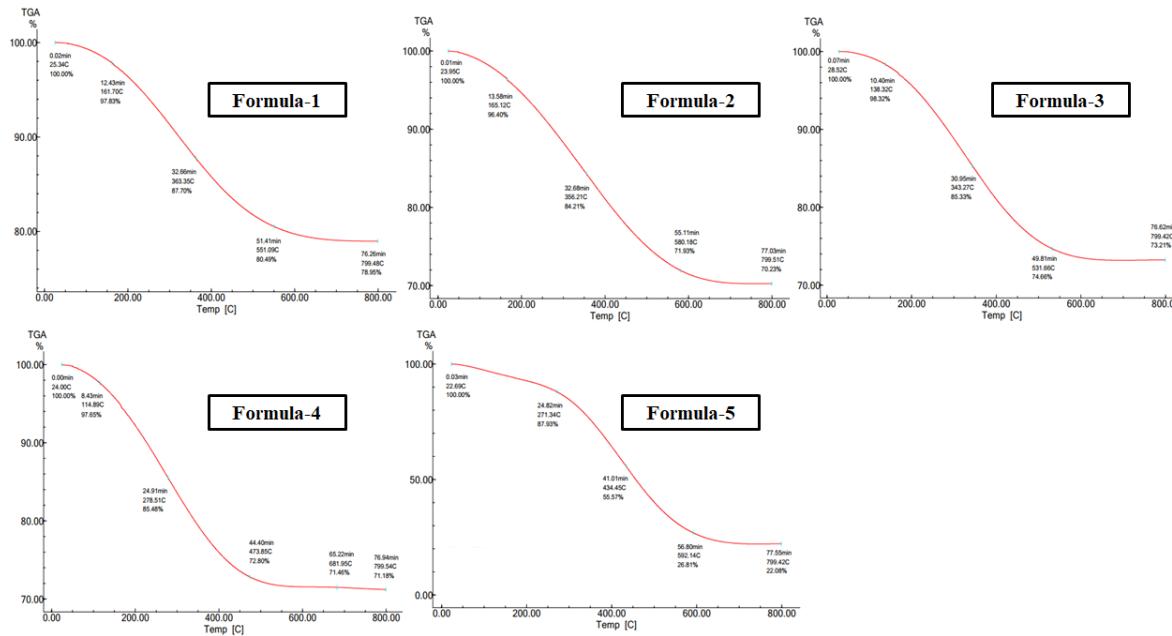


Figure 4. Thermogravimetric Analysis (TGA) of synthesized and freeze-dried AgNPs.

3.3. Morphological Analysis

SEM image of nanoparticle (Figure 5) formed Provides an idea on the size and shape of particle synthesized by five methods, which also supported with the DLS data (Table 1).

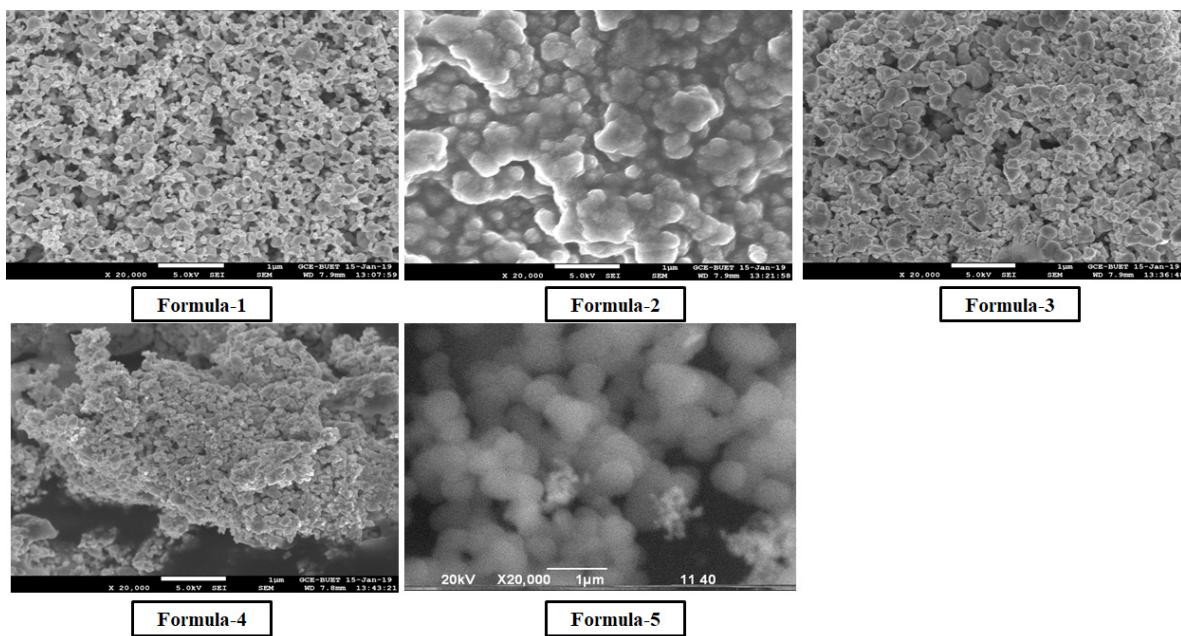


Figure 5. Scanning Electron Microscopic (SEM) images of freeze-dried AgNPs.

Particle size analysis using DLS (Figure 6) confirmed nanoparticle formation and conjugation. The size of nanoparticles for all five methods was below or around and 100 nm and the smallest particle was formed for method four. Electrical Conductivity (EC) Analysis (Table 1) by EC Meter also indicates the change in the mean nanoparticle size and size distribution among different formulations i.e., **F4 < F3 < F1 < F2 < F5**.

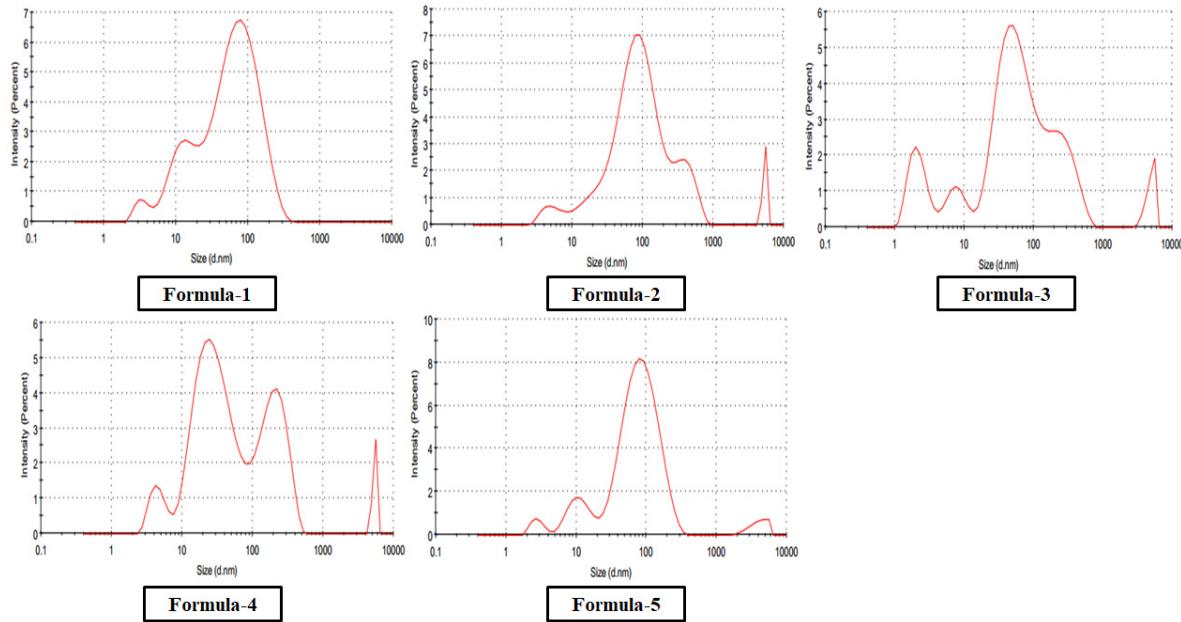


Figure 6. Size distribution by the intensity of AgNP formulations using DLS.

3.4. Stability Analysis

Zeta potentials of the nanoparticle (Figure 7) formed using the first four methods were below or around -30 mV, which confirms the good stability of the colloidal preparations. Besides, for the fifth method, it was -8 mV, which indicates that this formulation was less stable than others (Table 1).

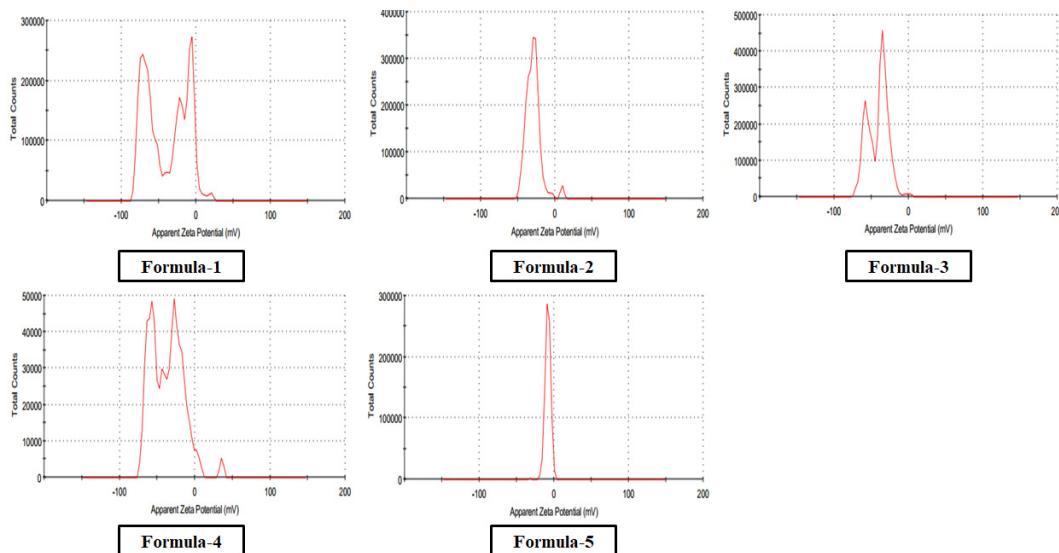


Figure 7. Zeta potential distribution of AgNP formulations.

3.5. Bactericidal Efficacy of AgNPs

Before inoculation, the OD of the *E. coli* culture was set to 0.440. The numbers of colonies of each petri dish were counted (Table 2). The petri dish containing only LB broth showed no visible microbial colony growth. Thus, it was ensured that the procedure was contamination-free. Also, 402 colonies were found for 0 ppm concentration of silver nanoparticles, which contained no nanoparticles (Table 2). The number of colonies for other ppm concentrations was also counted to determine the survival rate of *E. coli*. The number of colonies significantly declined even on a lower concentration of silver nitrate (5 ppm) for formula one. Besides, the number of colonies moderately declined at 5 ppm concentration of formulas two, three, and five. Lastly, the number of colonies very slowly declined for formula four (Table 2).

Table 2. Number of *E. coli* colonies at different concentrations of AgNP formulations.

AgNP Formula Concentration	Number of Colonies				
	0 PPM	5 PPM	10 PPM	20 PPM	40 PPM
Formula- 1	402	7	2	1	0
Formula- 2	402	39	10	8	0
Formula- 3	402	106	8	6	4
Formula- 4	402	395	176	18	4
Formula- 5	402	71	4	2	0

The survival percentage of *E. coli* for AgNPs synthesized by formula one, two, and five were least at the 20 ppm concentration, whereas it declined to zero at 40 ppm concentration (Table 2). Thus, it can be concluded that 40 ppm or a lower ppm concentration of AgNPs was MBC of formula one, two, and five (Figure 8). On the other hand, nanoparticles of formula three and four had the lowest survival rate at 40 ppm concentration. Similarly, MBC for formula three and four nanoparticles would be more than 40 ppm concentration (Figure 8). After analyzing MBC, AgNPs synthesized by formula-one found to be the most effective, followed by formula-five and formula-two respectively. On the contrary, formula-three and formula-four provided least bactericidal efficiency in the case of MBC.

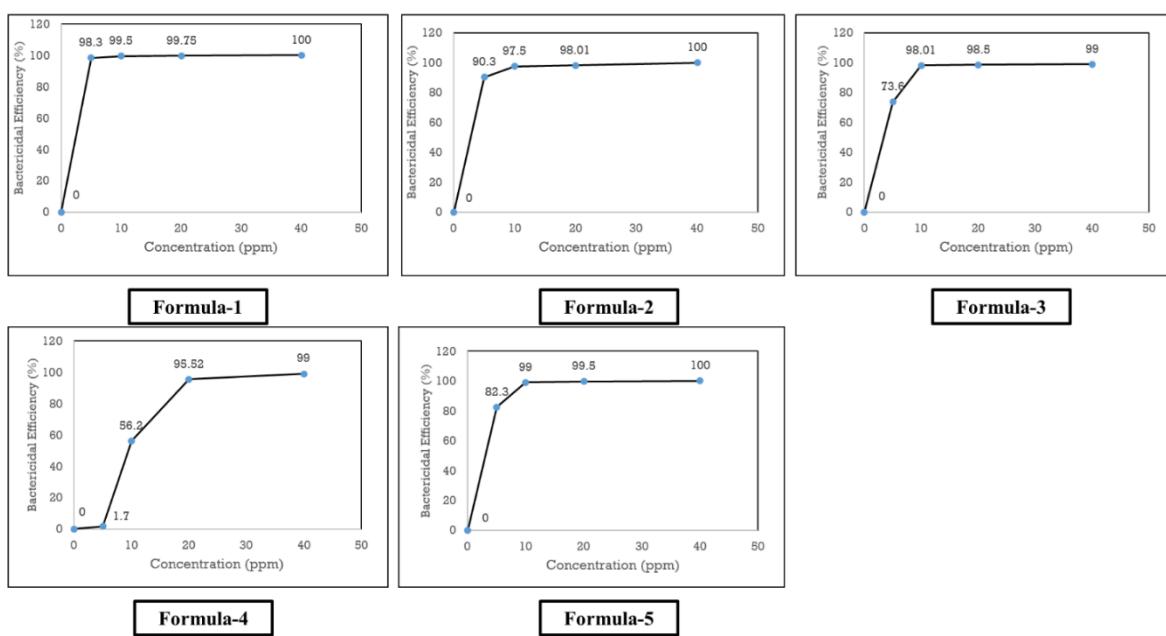


Figure 8. Bactericidal efficacy of different concentrations of five AgNP formulations.

3.6. Antibacterial Activity of AgNPs

Nanoparticle synthesized by formula one was most effective against the second isolate of *E. coli*, *Bacillus*, and the first isolate of *Staphylococcus* species in the well diffusion assay (Figure 9). It was found to be least efficient against the first isolate of *Klebsiella*. It should be mentioned that silver nitrate showed almost the same efficiency as formula one except for *Bacillus* isolate (Figure 9). On the contrary, formula one is least effective against the first isolate of *Klebsiella* (Figure 9). However, nanoparticles synthesized by formula two seemed ineffective against any isolates.

Apart from that, nanoparticles synthesized by formula three were observed to be most effective against the *Bacillus* isolate and least effective against the first isolate of *Klebsiella*. Besides, nanoparticles of formula four were most effective against *Bacillus* and the second isolate of *E. coli* and least effective against the first isolate of *Klebsiella* and the second isolate of *Staphylococcus* (Figure 9). Furthermore, nanoparticles synthesized by formula five were observed to be the most effective against all the clinical strains except *Bacillus*.

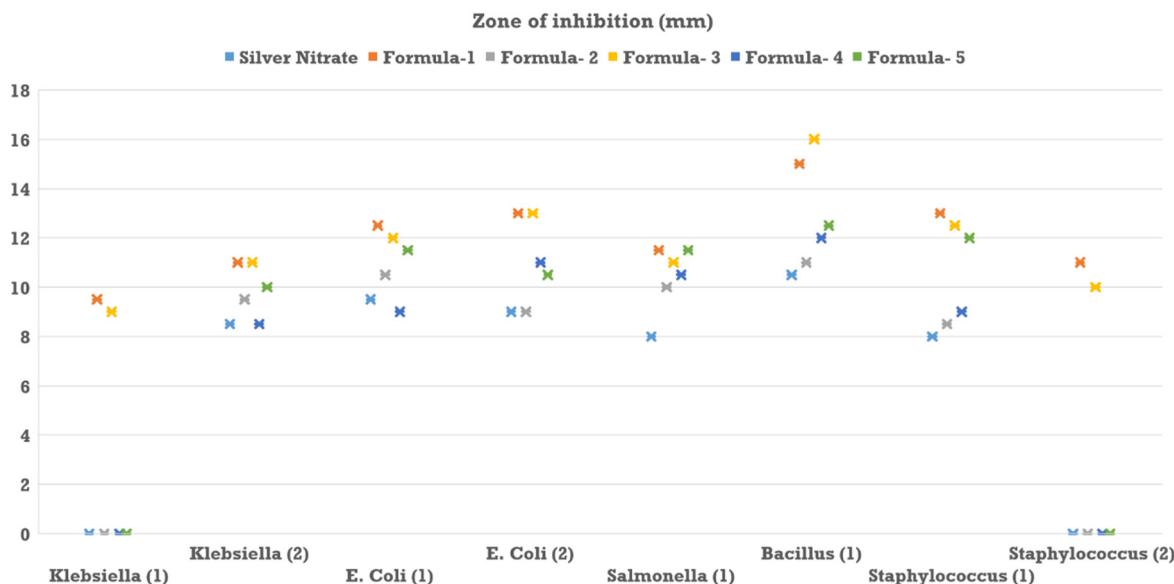


Figure 9. Comparing the anti-bacterial activities of AgNPs synthesized by five formulations.

3.7. Interpretation of Findings

AgNPs synthesized with trisodium citrate were most effective against methicillin-resistant *E. coli* due to their optimum particle size and stability (Figure 10). AgNPs synthesized with PEG and PVP were the least stable; however, their optimal particle size led to significant bactericidal efficacy (Figure 10). Although AgNPs synthesized with only sodium borohydride demonstrated optimal particle size and moderate stability, their inferior yield value may lead to subsidized antimicrobial efficacy (Figure 10). In contrast, AgNPs synthesized with a combination of trisodium citrate and sodium borohydride enhanced the yield but reduced the particle size and, subsequently, the antibacterial activity (Figure 10). Meanwhile, the addition of hydrogen peroxide caused a further decline in particle size and bacteria-killing efficacy while improving the yield (Figure 10).

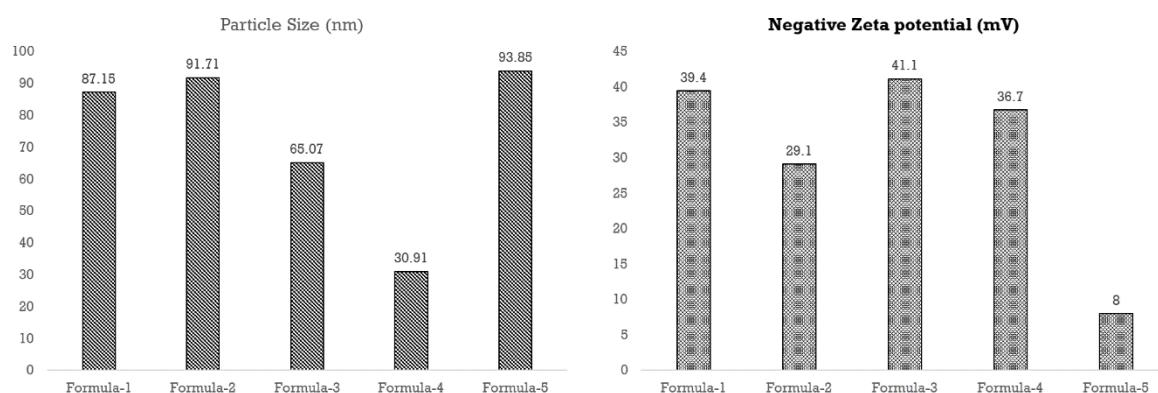


Figure 10. Comparison of Particle size and Zeta potential of different AgNPs formulations.

The efficiency of silver nanoparticles (AgNPs) in demonstrating antimicrobial behavior is mainly due to their size and shape, which influence their interaction with bacterial cells. However, these properties may vary based on interactions with other components in microbial cells. The stability of AgNPs in relation to their antimicrobial activity depends on morphology, surface variations, and conditions of preparation. In general, higher stability of AgNPs is linked to improved antimicrobial efficiency but too much stability might impair their reactivity and effectiveness. Hence, optimizing those parameters to establish the equilibrium between stability and reactivity holds the key to effectively using them in antimicrobial strategies.

4. Conclusion

The study showed that the choice of reduction chemistry leads to significant, formulation-dependent differences in antibacterial activity, with citrate-reduced AgNPs demonstrating the most effective bactericidal performance against methicillin-resistant *E. coli*, due to an optimal combination of particle size and stability. These findings underscore the importance of optimizing size and stability to achieve reliable antibacterial outcomes from chemically synthesized AgNPs. Further investigation of the impact of particle size and stability on the antibacterial activity of AgNPs is necessary to optimize the formulation.

Data Availability Statement: The data presented in this study are available in the supporting materials.

Conflict of Interest: The authors declare no conflict of interest.

Abbreviations

AgNPs = Silver nanoparticles, NaBH₄ = Sodium borohydride, H₂O₂ = hydrogen peroxide, PEG = Polyethylene glycol, PVP = Polyvinyl pyrrolidone

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