

Article

Not peer-reviewed version

Integrins Can Act As Suppressors of Ras-Mediated Oncogenesis in the Drosophila Wing Disc Epithelium

[Maria D. Martín-Bermudo](#)^{*} and Ana Martínez-Abarca Millán

Posted Date: 9 October 2023

doi: 10.20944/preprints202310.0373.v1

Keywords: drosophila integrins; cancer; cell growth; invasion



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Article

Integrins Can Act as Suppressors of Ras-Mediated Oncogenesis in the *Drosophila* Wing Disc Epithelium

Ana Martínez-Abarca Millán ¹ and María D. Martín-Bermudo ^{1,*}

¹ Centro Andaluz de Biología del Desarrollo CSIC-University Pablo de Olavide, Sevilla 41013, Spain.

* Correspondence: mdmarber@upo.es

Abstract: Cancer is the second leading cause of death worldwide. Key to cancer initiation and progression is the crosstalk between cancer cells and their microenvironment. The extracellular matrix (ECM) is a major component of the tumour microenvironment and integrins, main cell-ECM adhesion receptors, are involved in every step of cancer progression. However, accumulating evidence has shown that integrins can act as tumour promoter but also as tumour suppressor factors, revealing that the biological roles of integrins in cancer are complex. This incites a better understating of integrin function in cancer progression. To achieve this goal, simple model organisms, such as *Drosophila*, offer a great potential to unravel underlying conceptual principles. Here, we find that in the *Drosophila* wing disc epithelium the bPS integrins act as suppressors of tumours induced by a gain of function of the oncogenic form of *Ras*, *Ras*^{V12}. We show that bPS integrin depletion enhances the growth, delamination and invasive behaviour of *Ras*^{V12} tumour cells, as well as their ability to affect the tumour microenvironment. These results strongly suggest that integrin function as tumour suppressors might be evolutionary conserved. *Drosophila* can be used to understand the complex tumour modulating activities conferred by integrins, thus facilitating drug development.

Keywords: *Drosophila* integrins; cancer; cell growth; invasion

Introduction

Cancer is the second leading cause of death worldwide. In 2020, there were 19.3 million new cancer cases, and by 2040, this number is expected to rise to 29.5 million [1]. Thus, cancer research studies are crucial towards diminishing the tremendous human and economic tolls of cancer.

The crosstalk between cancer cells and their microenvironment plays a key role in cancer initiation and progression. The extracellular matrix (ECM) is a major component of the tumour microenvironment and integrins, main cell adhesion receptors for components of the ECM, are involved in almost every step of cancer progression, including cancer initiation, proliferation, metastasis and survival of circulating tumour cells (reviewed in [2]). Integrins are heterodimeric molecules containing an α and a β subunit. The mammalian family consists of 24 heterodimers generated from a combination of 18 α integrin and 8 β integrin subunits (reviewed in [3]). Much attention has been devoted to the $\beta 1$ family of integrins. These studies have demonstrated that the biological roles of $\beta 1$ integrins in cancer, as signalling molecules, mechanotransducers and essential components of the cell migration machinery, are quite complex and highly dependent on the type and developmental stage of the tumour. In different types of cancers, the expression of $\beta 1$ integrin facilitates the growth and survival of cancer cells. However, regarding metastasis, the effects of $\beta 1$ integrin is controversial and $\beta 1$ integrins can act as tumour promoters or tumour suppressors. For example, while most $\beta 1$ integrins, and more specifically $\alpha 3\beta 1$, are essential for mammary tumorigenesis [4, 5], $\alpha 2\beta 1$ integrin acts as a metastasis suppressor in breast cancer [6]. Furthermore, when cultured in three dimensional ECM scaffolds, block or knockdown of $\beta 1$ integrin function elicits a pro-metastatic switch in human and mouse E-cadherin-positive triple-negative breast cancer cells [7]. Thus, given the complexity of $\beta 1$ integrins and their antagonistic roles in some types of cancer

cells, therapeutic targeting of these receptors is a challenge and only a small number of clinical trials have been successful. This incites a better understating of the role of integrins in cancer progression, which could lead to the development of new targeting approaches. To achieve this goal, simple model organisms, such as *Drosophila*, with fewer genes compared to the human genome and lower genetic redundancy, have great potential to unravel underlying conceptual principles.

In contrast to vertebrates, the *Drosophila* genome encodes only two β subunits, β PS and β v, and five α subunits, α PS1 to α PS5. The β PS subunit, encoded by the gene *myospheroid* (*mys*), the orthologue of vertebrate b1, is widely expressed and forms heterodimers with all α -subunits (reviewed in [8, 9]). The β PS integrins have been shown to regulate many cellular processes both during embryonic and larval development and in the adult. In particular, the primordium of the *Drosophila* wing, the larval wing imaginal disc has been successfully used to decipher the function of integrins during epithelial morphogenesis and homeostasis (reviewed in [10-14]). The formation of the wing starts during early embryonic development when about 30 cells are allocated to form the primordium of the wing imaginal disc (henceforth the wing disc). During larval life, disc cells divide and form an epithelial sac composed of a squamous layer of epithelial cells called the peripodial epithelium (PE) and a columnar epithelial layer called the disc proper, with myoblasts, tracheal cells and a few neurons attach to the disc proper. Underlying the epithelial cells of the disc proper there is a basement membrane (BM). The wing disc epithelium is often divided into four broad domains, the pouch, the hinge, the notum and the PE. The pouch and the hinge give rise to the wing blade and wing hinge, respectively, while the notum gives rise to most of the back of the fly in the thorax, and the PE, which gives rise to some of the pleura (reviewed in [15]). Integrins have been shown to be required for adhesion, shape and survival of the wing disc epithelial cells [14, 16-18]. The wing imaginal disc also constitutes an excellent and productive choice for investigating the biology of carcinoma, the most common cancer type that originates from epithelial cells (reviewed in [19]). More specifically, the wing imaginal disc has been extensively employed to study the progression of tumours due to mutations in the proto-oncogene *Ras* [20], mutated in 30% of human cancers [21]. The induction of groups of cells over-expressing an oncogenic activated form of *Ras* (*Ras^{V12}*) in the wing disc gives rise to hyperplastic growth. This has been exploited in genetic screens to identify additional genes that can either suppress or enhance the growth of cells overexpressing *Ras^{V12}* [20]. In this context, we have recently found that elimination of integrins from transformed *Ras^{V12}* wing disc epithelial cells stimulate their basal extrusion [14]. However, little more is known about the role of the β PS integrins in cancer initiation and progression.

In this work, we have used the wing disc to analyse the role of integrins on the progression of *Ras^{V12}* epithelial tumour cells. We find that downregulation of the β PS integrins enhances *Ras^{V12}*-mediated tissue hyperplasia. Elimination of integrins promotes the growth of *Ras^{V12}* tumour cells. In addition, using live imaging and confocal microscopy, we find that reduction of β PS integrin levels induces the delamination and invasion of tumour cells to neighbouring wild type regions. These results demonstrate that, as it is the case in some mouse models and human cancer cell lines, in *Drosophila*, integrins inhibit the metastatic capacity of tumour cells. These results also strongly suggest that the role of the β 1 integrin family as suppressor of tumour metastasis might be evolutionary conserved.

Materials and Methods

Fly strains

The following stocks were used: *UAS-Ras^{V12}* [22]; *Ap>myrT/ Cyo*, *Ap>GFP/Cyo*, *Ap-Gal4-UAS-flyfucci/Cyo* (gifts from Dr. Marco Milán's laboratory); *UAS-mys^{RNAi}* and *UAS-hid^{RNAi}* (Vienna *Drosophila* Center). Flies were raised at 25°C.

Immunocytochemistry and Imaging

Wing imaginal disc from instar 3 larvae were dissected and stained following standard procedures and mounted in Vectashield (Vector Laboratories, Burlingame, CA, United States). The

following primary antibodies were used: goat anti-GFP (Abcam, 1:500), rabbit anti-caspase Dcp1 (Cell Signaling; 1:100), rabbit anti-pJNK (Promega, 1:200), mouse anti- β PS (Developmental Studies Hybridoma Bank, DSHB, University of Iowa, United States, 1:50), rabbit anti-PH3 (EMD Millipore, 1:250), rabbit anti-Perlecan (Dr. A. González-Reyes, 1:850), rabbit anti-aPKC (Promega, 1:200), Rat-anti-RFP (Chromotek, 1:500). The secondary antibodies used were Alexa fluor 488, (Molecular ProbesTM) and Cy3 and Cy5 (Jackson ImmunoResearch Laboratories, Inc.) at 1: 200. DNA was labelled using Hoechst (Molecular Probes, 1:1000). F-actin was visualized using Rhodamine Phalloidin (Molecular Probes, 1:50).

Confocal images were obtained using a Leica SPE microscope, equipped with a 20X oil objective (NA 0.7) and a Leica Stellaris microscope equipped with 20x, 40x and 63x oil objective with NA 0.75, 1.4 and 1.4 respectively.

For live imaging, wing disc samples were prepared as described in [23] and filmed for 2-3 hours using a Leica Stellaris microscope and a 40x oil objective.

Quantification of fluorescence intensity

To quantify the fluorescent intensity of the different markers, fluorescent signaling was measured on several confocal images per genotype using the square tool of FIJI-Image J. The microscope settings, such as laser parameters and z-confocal interval, were maintained between imaging sessions in each experiment. Measurements of whole fluorescence intensity were done by using the "mean gray value" tool of FIJI. This tool divides the mean of all included pixels intensity by the outlined cell area. The different areas of the wing disc, apterous (dorsal) and control (ventral) domains, were selected using the wand tool.

To quantify cell proliferation, dots of fluorescent intensity of wing discs stained with an anti-PH3 antibody were quantified using the the FIJI-Image J tool analyze particles and the Trainable Weka 2D Segmentation plug-in, which transforms 8-bit images into a binary system, was used to quantify using the FIJI-Image J tool analyze particles. the number of mitotic cells we segmented the image in the Cy3 channel by adjusting the threshold and used "analyze particles" tool to count the PH3⁺ cells.

Cell areas were calculated by manually delineating the apical and basal surfaces of the cells and measuring the resulting surfaces using FIJI-ImageJ software. A similar procedure was used to quantify cell height. A vertical line was drawn from the apical to the basal surface in a region of interest of a wing disc, stained for F-actin to visualize cell limits, and the length of this line was measured using FIJI.

Flyfucci analysis were performed using the "merge channels" tool of FIJI to generate an image containing the green and red channels together. The "TrackMate" tool was used to measure the fluorescence intensity of both channels in the same 3D coordinates. Spots showing similar levels of fluorescence intensity of both channels correspond to G2 and those with higher levels, at least 3 times greater, in the red channel with respect to the green one and vice versa correspond to S and G1, respectively.

All graphics and mathematical analysis were generated using R software and ggplot2 package. For statistical comparisons, the p-value was calculated using the Welch test and assuming that all distributions were normal with unequal variances. Multiple comparisons p-values were adjusted following BH procedure (controlled False Discovery Rate).

Results

Integrin knockdown enhances Ras^{V12} hyperplastic phenotype

We and other have previously shown that the ectopic expression of Ras^{V12} in the dorsal compartment of wing discs, by means of an *apterous*-Gal4 line (*Ap>GFP; Ras^{V12}*), induced overgrowth of the tissue and the formation of ectopic folds [24, 25] (Figure 1B, B'). To analyse the role of integrins in Ras^{V12}-mediated oncogenesis, we decided to reduce integrin levels in tumour cells by expressing a *mys* specific RNAi, a method that was shown to cause a strong reduction in bPS protein levels in wing

discs [14]; Sup. Figure 1A-B'''). In agreement with previous results, we found that the expression of a *mys* RNAi in the dorsal compartment of wing discs (*Ap>GFP; mys^{RNAi}*) induced the formation of ectopic folds (Figure 1C, C'; [14]. In addition, we found that the downregulation of β PS integrin function in *Ras^{V12}* cells (*Ap>GFP; mys^{RNAi}; Ras^{V12}*) enhanced the formation of folds due to *Ras^{V12}* overexpression alone (Figure 1D, D'). We have previously shown that reducing integrin levels in the central region of the wing disc induced caspase-dependent cell death [14]. In addition, we found that integrins promoted cell survival by inhibiting the expression of the pro-apoptotic gene *hid*, as reducing *hid* levels rescued apoptosis due to integrin knockdown [14]. Furthermore, we demonstrated that the increased apoptosis due to reduction of integrin levels was rescued by ectopic expression of *Ras^{V12}* [14]. In this context, the increase in fold formation observed in *Ras^{V12}* wing discs when reducing integrin levels could be just due to the ability of *Ras^{V12}* to rescue the apoptosis of *mys^{RNAi}* cells. To test this possibility, we analysed the effects of directly inhibiting apoptosis in wing discs carrying integrin mutant cells on fold formation. To do this, we expressed a *hid* specific RNAi in *ap>GFP; mys^{RNAi}* wing discs (*Ap>GFP; mys^{RNAi}; hid^{RNAi}*). We found that although the over expression of a *hid^{RNAi}* in *Ap>GFP; mys^{RNAi}* wing discs led to an increase in fold formation (Figure 1E, E'), the increase was smaller than that found when over expressing *Ras^{V12}* (Figure 1D, D'). Altogether these results suggest that the ability of *Ras^{V12}* overexpression to induce the overgrowth of integrin mutant tissues goes beyond its capacity to rescue the survival of integrin mutant cells.

The expression of β 1 integrins in cancer cells is altered, being up-or downregulated depending on tumour type, stage and microenvironment, an issue that remains controversial [2]. Here, we found that the levels of β PS integrin in *Ras^{V12}* wing disc epithelial cells were significantly higher than those found in wild type neighbouring cells (Sup. Figure 1C-C''', E). As expected, the expression of a *mys^{RNAi}* in these tumour cells led to a reduction in β PS levels (Sup. Figure 1D-D''', E).

Even though the ectopic expression of *Ras^{V12}* in wing disc cells alone does not affect cell polarity [26]; Sup. Figure 2B-C'), the removal of polarity genes enhances the hyperplastic phenotype of *Ras^{V12}* [27]. In addition, there is increasing evidence that integrin-mediated polarity is implicated in cancer (reviewed in [28]. In fact, integrin-mediated cell polarity suppresses oncogenesis in the fly midgut [29]. Thus, we tested whether cell polarity was affected in *Ras^{V12}; mys^{RNAi}* disc cells. First, we analysed whether cell polarity was perturbed in the absence of integrins in wing epithelial disc cells and found it was not (Sup. Figure 2D, D'). In addition, we found that downregulation of integrins did not alter the polarity of *Ras^{V12}* cells (Sup. Figure 2E, E').

Similarly, while expression of *Ras^{V12}* on its own does not grossly affect the BM, loss of cell polarity in these tumour cells induces BM degradation and the formation of aggressive, malignant tumours (reviewed in [30]. In addition, recent studies have proven that integrins present in tumour cells, or tumour-associated cells, are involved in the remodelling of the ECM (reviewed in [31]. Here, using an antibody against the BM component Perlecan, we found that elimination of integrins did not affect the deposition of the BM in either normal or tumour cells (Sup. Figure 2F-I'').

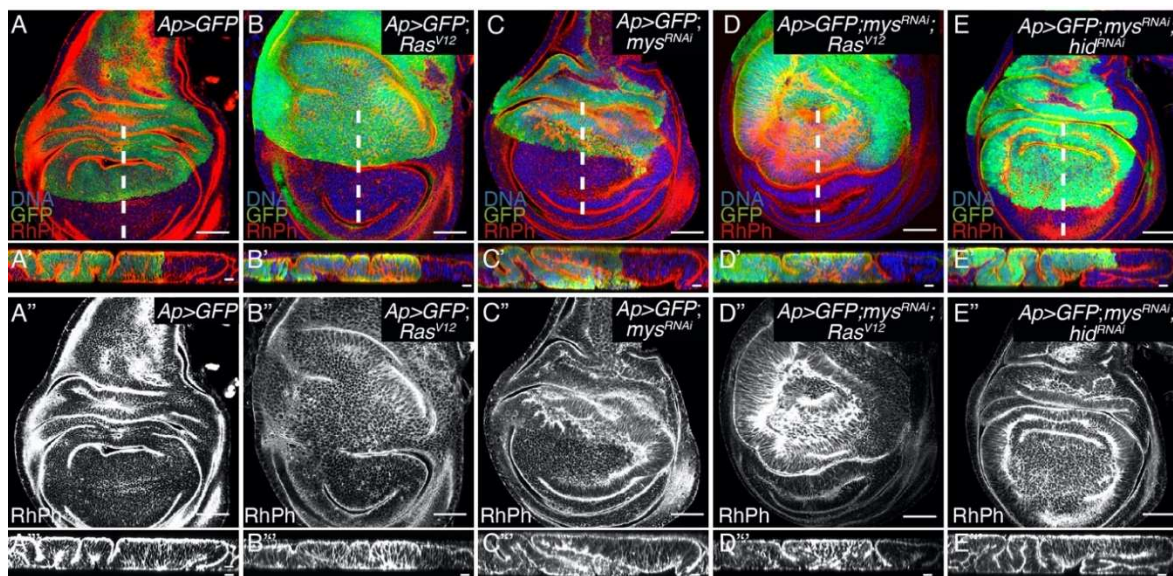


Figure 1. Integrin knockdown enhances Ras^{V12} hyperplasia in the *Drosophila* wing disc. (A-E') Confocal views of third-instar larvae wing discs of the indicated genotypes, stained with anti-GFP (green), Rhodamine Phalloidin (RhPh) to detect F-actin (red in A-E, white in A'-E') and Hoechst for DNA detection (blue). (A'-E' and A'''-E''') Confocal yz sections along the white straight line shown in A-E. Scale bars 50 μm (A-F, A''-F'') and 10 μm (A'-F', A'''-F''').

Integrin knockdown enhances Ras^{V12} -associated cell shape changes and growth

The formation of additional folds could be due to increased cell proliferation, changes in cell shape, cell growth or to a combination of these factors. Staining with an antibody against phosphorylated Histone H3 (PH3) showed that, in agreement with previous results [24, 25, 32, 33], overexpression of Ras^{V12} resulted in a reduction in the number of PH3⁺ cells (Sup.Figure 3A-B', E). This was not affected by downregulation of integrin expression (Sup.Figure 3C-D, E). To analyse possible cell shape changes, cell plasma membranes were visualized using the F-actin marker Rhodamine-Phalloidin (RhPh) (Figure 2). We found that, in agreement with previous results, Ras^{V12} and integrin mutant expressing cells underwent a columnar to cuboidal cell shape change, showing reduced height and increased apical and basal surfaces compared to control cells (Figure 2A-C''', F-H, [18, 25]). Furthermore, we found that elimination of integrins exacerbated the cell shape change of Ras^{V12} cells (Figure 2D-D''', F-H). Again, this was not just due to the ability of Ras^{V12} to rescue apoptosis of integrin mutant cells, as inhibiting apoptosis in mys^{RNAi} cells does not cause the same shape changes as overexpressing Ras^{V12} (Figure 2E-E''', F-H).

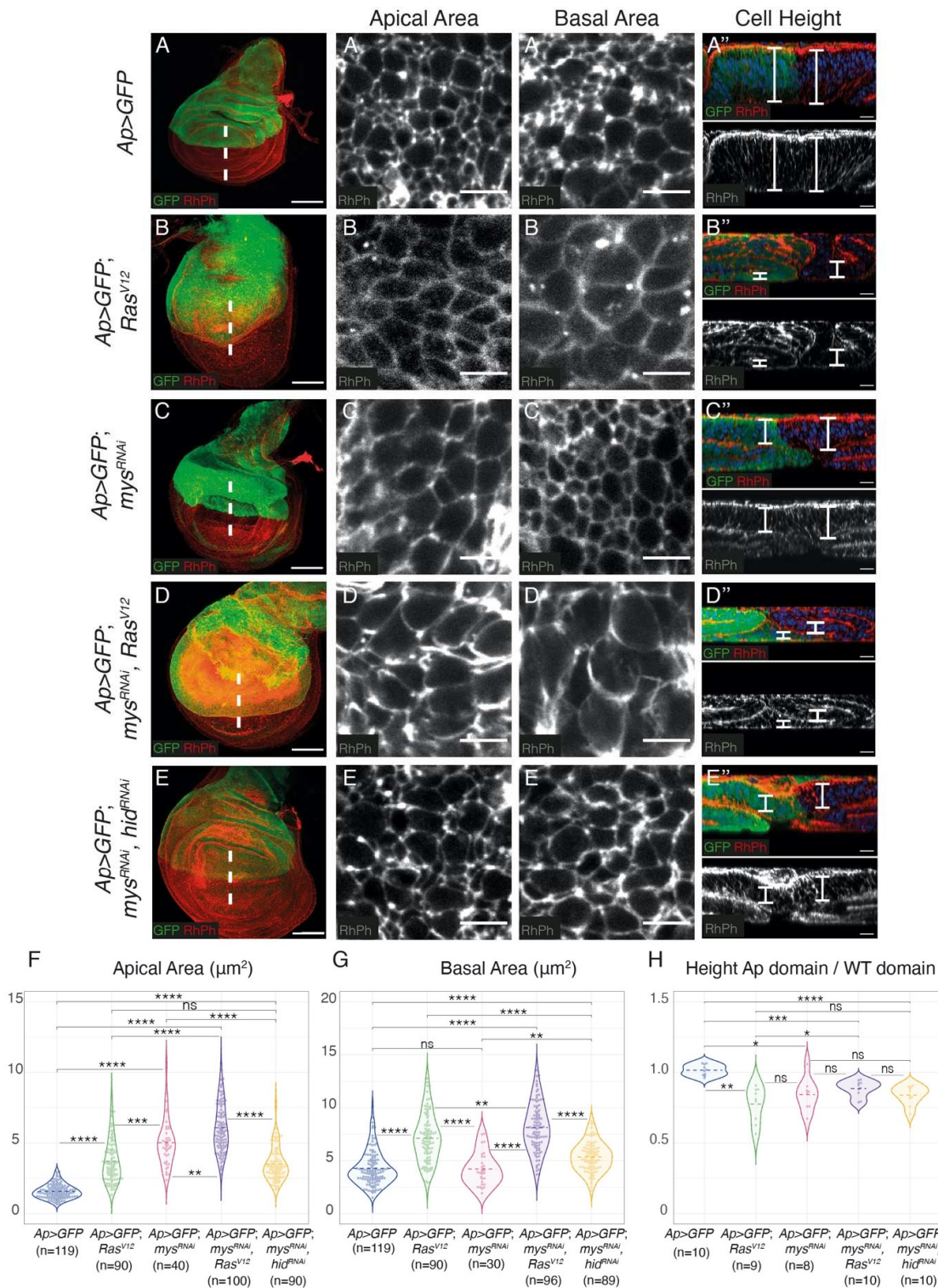


Figure 2. Integrin expression downregulation increases *Ras*^{V12}-dependent cell shape changes and growth in *Drosophila* wing discs. (A-E) Maximal projection of confocal images of third-instar larvae wing discs of the designated genotypes, stained with anti-GFP (green), RhPh (red in A-E, white in A'-E'') and Hoechst for DNA detection (blue, A'''-E'''). (A'-E') Apical and (A''-E'') basal surface views of GFP⁺ cells of wing discs of the indicated genotypes. (A'''-E''') Confocal *xz* sections along the white dotted lines of wing discs shown in A-F. The apical side of wing discs is at the top. White brackets indicate cell height. (F-H) Violin plots of the apical cell area (F), basal cell area (G) and cell height (H) of the indicated genotypes. Differences were measured with welch-test, ****, ***, **, and * P values <0.0001, <0.001, <0.01 and <0.05 respectively. Scale bars 50 μm (A-E), 5 μm (A'-E', A''-E'') and 10 μm (A'''-E''').

Prolonged expression of *Ras*^{V12} promotes G1-S phase transition, as a consequence of E2F activation and cyclin E induction [24]. The use of Fly-Fucci, which relies on fluorochrome-tagged degrons from the cyclin B -degraded during mitosis- and E2F1 -degraded at the onset of S phase-, confirmed the role of *Ras*^{V12} in driving transition from G1 to S phase and also showed that *Ras*^{V12} expressing tissues exhibited a reduction of the number of cells in S phase and an accumulation in G2, compared to control tissues [33]. These results are consistent with the previously reported role of *Ras*^{V12} in promoting cellular growth and senescence in *Drosophila* imaginal epithelia [24, 34]. Using Fly-Fucci, we found that, similar to *Ras*^{V12} overexpression [33], Figure 3B-C'), downregulation of integrin levels resulted in an increase in the number of cells in G2 (Figure 3D, D'), an increase that did not depend on cell death (Figure 3F, F'). In addition, reducing integrin levels increased the number of *Ras*^{V12} tumour cells in G2 (Figure 3E, E').

All together, these results suggest that integrins can limit *Ras*^{V12}-mediated tissue hyperplasia by regulating cell shape changes and cellular growth. Given the constraints imposed by the peripodial membrane, the observed cell shape changes and the increase in cellular growth size observed in all experimental conditions could explain the formation of extra folds.

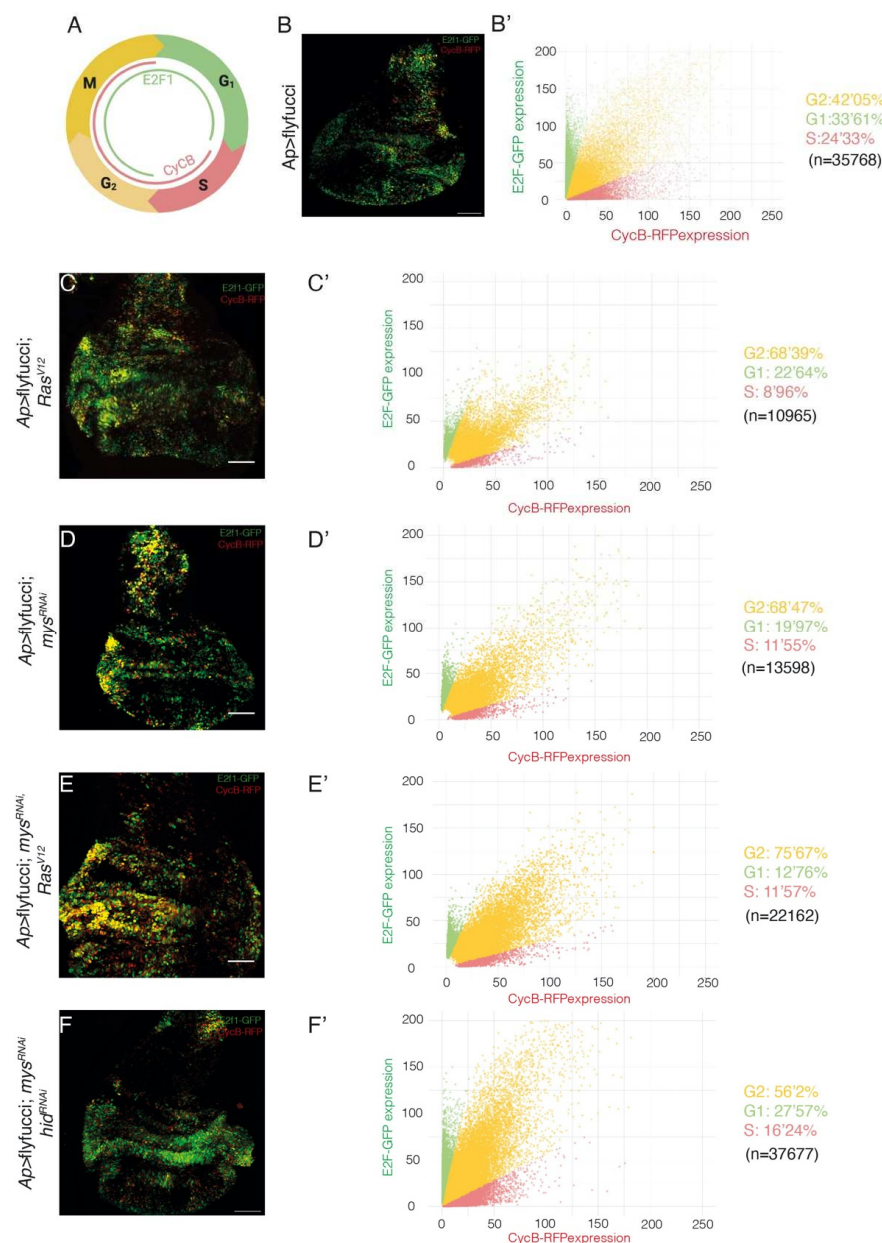


Figure 3. Integrin knockdown enhances the changes in cell cycle progression in *Drosophila Ras*^{V12} wing disc cells. (A) Scheme depicting CycB-RFP and E2F1-GFP nuclear expression during the cell

cycle. (B-F) Maximal projection of confocal images of third-instar larvae wing discs of the indicated genotypes, expressing E2f1-GFP (green) and CycB-RFP (red). (B'-E') Scatter plots representing the fluorescence intensity of E2f1-GFP (green) and CycB-RFP (red). Scale bars 50 μ m (B-E).

Integrins restrict the ability of Ras^{V12} tumour cells to stimulate apoptosis in neighbouring wild type cells

Even though the overexpression of Ras^{V12} can promote cell survival, several studies have shown that it can also stimulate the death of nearby wild-type cells [35, 36]. Accordingly, in a recent study, we detected a clear enrichment in the levels of the apoptotic marker cleaved Dcp1 in wild-type (GFP negative) ventral cells located at the D/V boundary in *Ap>GFP; Ras^{V12}* discs [25]; Sup. Figure 4B, B', F). In addition, as mentioned above, we have previously shown that reducing integrin levels in the whole wing pouch induces caspase-dependent cell death, which is suppressed by oncogenic Ras [14]. Here, we found that although apoptosis was reduced in the dorsal region of *Ap>GFP; Ras^{V12}; mys^{RNAi}* (Sup. Figure 4D, D', E), compared to *Ap>GFP; mys^{RNAi}* discs (Sup. Figure 4C, C', E), in agreement with our previous results [14], there was an increase in Dcp1 levels at the D/V boundary, similar to what happens in *Ap>GFP; Ras^{V12}* discs (Sup. Figure 4B-C', F). Furthermore, elimination of integrins increased the ability of Ras^{V12} cells to induce apoptosis in neighbouring wild type cells, as measured by Dcp1 levels (Sup. Figure 4B, B', D, D', F).

The capacity of Ras^{V12} cells to induce apoptosis of neighboring wild type cells has been shown to be mediated by activation of the JNK pathway [35, 36]. Accordingly, in a recent study, using an antibody against phosphorylated JNK (pJNK), we detected a clear enrichment of JNK activity in wild-type (GFP negative) ventral cells located at the D/V boundary in *Ap>GFP; Ras^{V12}* discs [25], Figure 4A-B', F). We have also shown that JNK mediates apoptosis induced by loss of integrin function [14]. Here, we found that although JNK activity was reduced in the dorsal region of *Ap>GFP; Ras^{V12}; mys^{RNAi}* (Figure 4D, D', F), compared to *Ap>GFP; mys^{RNAi}* discs (Figure 4C, C', F), pJNK levels at the D/V boundary were higher than those found in *Ap>GFP; Ras^{V12}* discs (Figure 4B, B', F).

Induction of apoptosis and activation of the JNK pathway in wild-type neighbors of Ras^{V12} cells have been partially explained by an increase in tissue compaction due to the enhanced growth of tumour cells [37]. Thus, the rise in apoptosis (Sup. Figure 4) and JNK activity (Figure 4) we observed in wild type cells located at the D/V boundary in *Ap>GFP; Ras^{V12}; mys^{RNAi}*, compared to *Ap>GFP; Ras^{V12}* discs, could be due to the increase in the formation of folds in the tumorigenic tissue upon integrin removal.

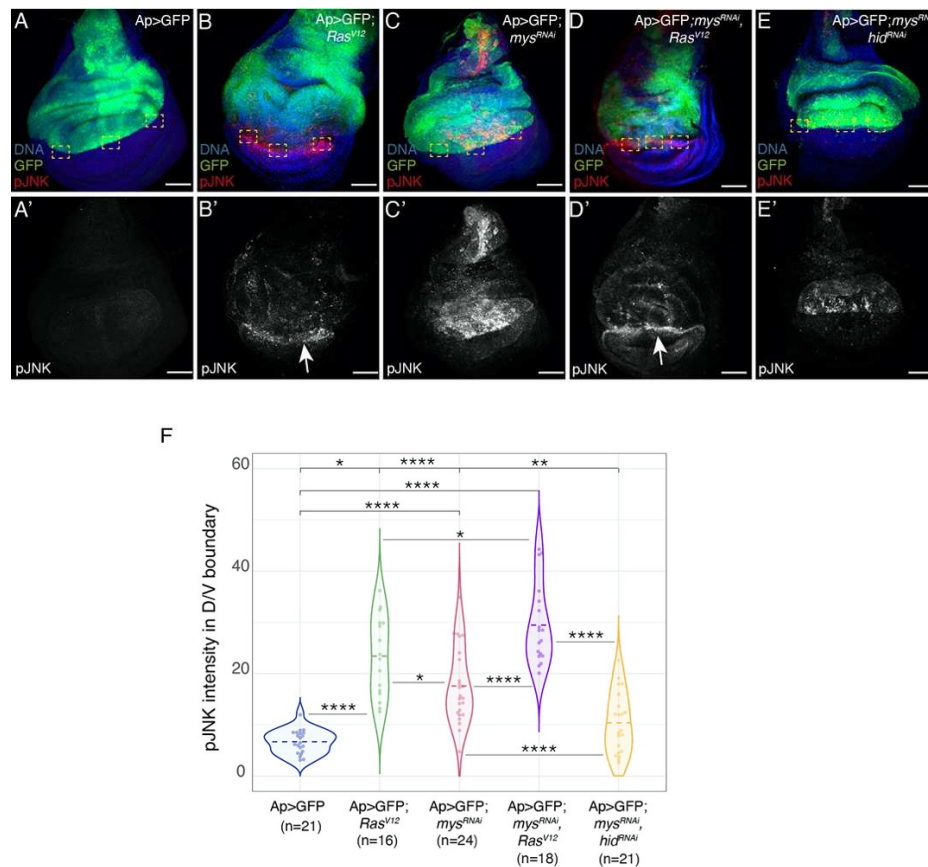


Figure 4. Integrin knock down increases the ability of *Ras^{V12}* to induce JNK activity in wild type adjacent cells. (A-E') Maximal projection of confocal views of third-instar wing discs of the specified genotypes, stained with anti-GFP (green), anti-pJNK (red in A-E, white in A'-E') and Hoechst (DNA, blue). (F) Violin plots of mean fluorescent pJNK intensity at the dorsal-ventral boundary of wing discs of the designated genotypes. The statistical significance of differences was assessed with a welch-test, ****, ** and * P values <0.0001, <0.01 and <0.05, respectively. Scale bars, 50 μ m (A-E').

*Integrins limit the ability of epithelial *Ras^{V12}* tumour cells to leave the epithelium and move*

We have recently found that a proportion of wing discs expressing GFP and *Ras^{V12}* in the dorsal domain showed GFP⁺ *Ras^{V12}* cells in the ventral domain, some of which were positive for the apoptotic marker cleaved Dcp1 [25]; Figure 5A, B, F). This result led us to suggest that the expression of *Ras^{V12}* in wing disc cells could induce an invasive behaviour. Here, using the Dcp1 marker to distinguish between dead and alive cells, we found that 39,3% of *Ap>GFP; Ras^{V12}* wing discs presented alive GFP⁺ cells in the ventral domain (Figure 5B-B''', F), in contrast to the 8.2% found in control wing discs (Figure 5A-A''', F). In addition, alive ventral GFP⁺ cells were only seen in 14,3% of wing discs with reduced integrin levels (Figure 5C-C''', F). However, here, we found that reducing integrin function increased up to 63,3% the percentage of *Ras^{V12}* wing discs showing invading GFP⁺ cells (Figure 5D-D''', F). Once more, this was not just due to the ability of *Ras^{V12}* to rescue the death of integrin mutant cells, as preventing apoptosis in *mys^{RNAi}* cells only increased the penetrance of the invasive phenotype up to 20,6% (Figure 5E-E''', F).

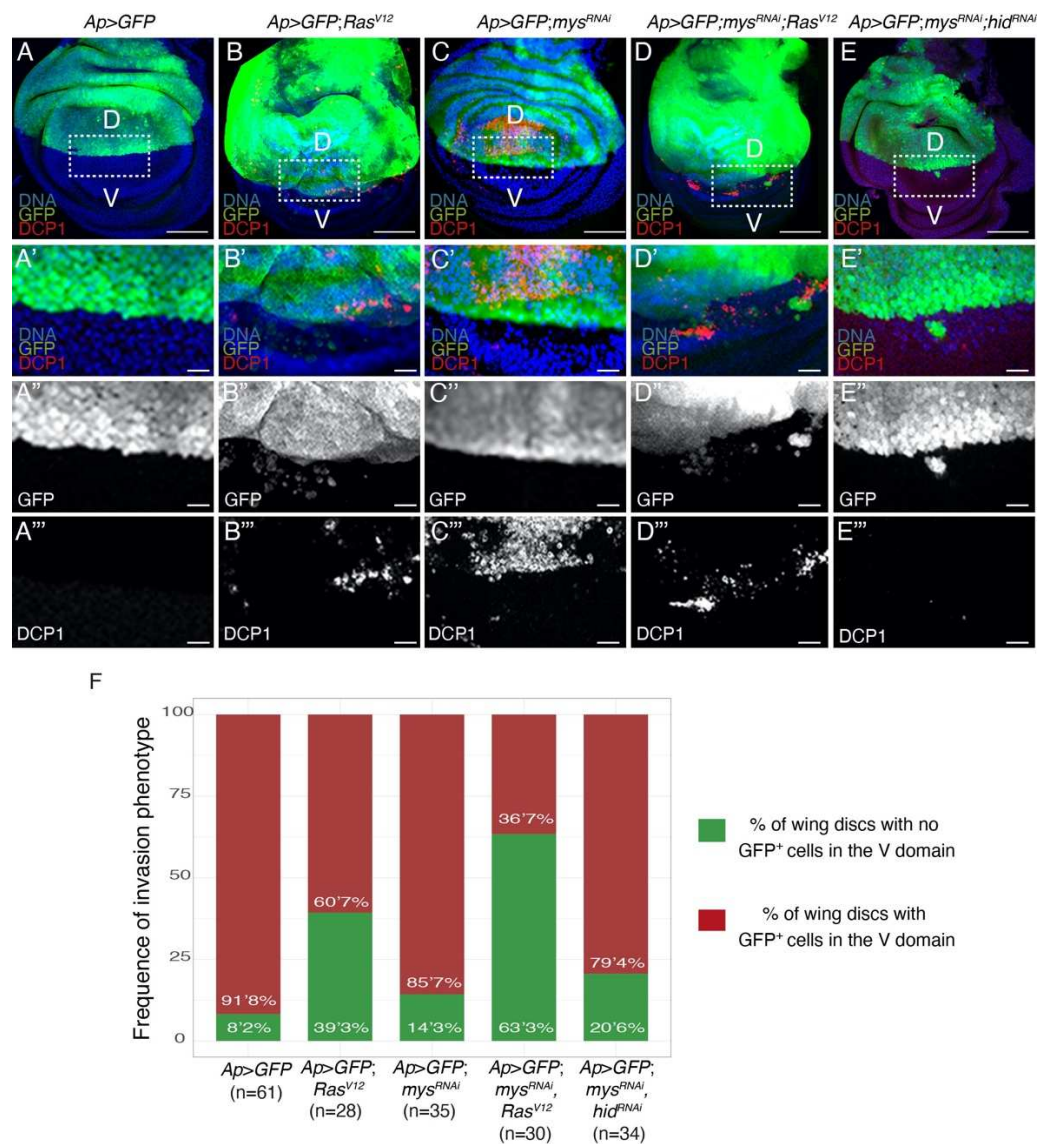


Figure 5. Integrins downregulation enhances the invasive behavior of *Ras^{V12}* cells. (A-E) Confocal views of third-instar larvae wing discs of the designated genotypes, stained with anti-GFP (green in A-E' and white in A''-E'''), DCP1 (red in A-E' and white in A'''-E''') and Hoechst (DNA, blue) (A-E'). (A'-E''') Magnifications of the white boxes in A-E, respectively. (F) Bar plot of the frequency of appearance of non-dead GFP+ cells in the ventral domain. Scale bars 50 μ m (A-E) and 10 μ m (A'-E''').

Our previous analysis had shown that eliminating integrin function in *Ras^{V12}* cells promoted their basal extrusion [14, 25]. To study in more detail the behaviour of these tumour cells, we dissected, cultured and monitored lively control and experimental third instar wing discs expressing GFP in the dorsal domain (see Materials and Methods). Live imaging analysis of confocal sections of cultured wing discs allowed us to watch the behaviour of dorsal GFP+ cells up to 180 minutes (Figure 6).

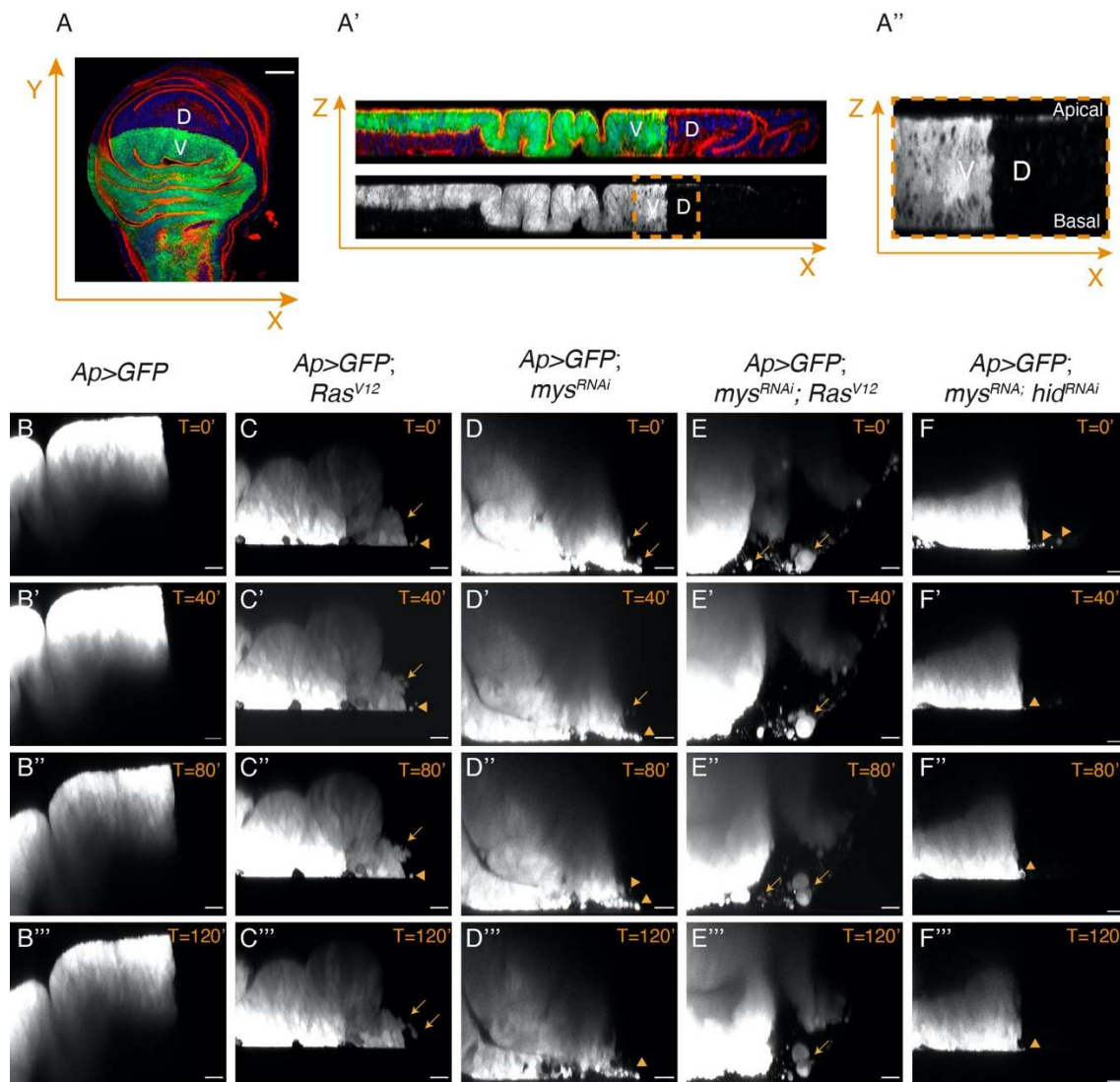


Figure 6. Integrins regulate the invasive capacity of Ras^{V12} cells. (A-A'') Scheme illustrating the region of the wing disc where movies had been taken. (A) Confocal view of a third-instar larvae wing disc, stained with Hoechst (blue), RhPh (red) and anti-GFP (green), and (A') an orthogonal view along the dotted white line in A. (A'') Zoom-in of the region in the yellow dotted box in A', corresponding to the region where movies had been taken. (B-F'') Confocal images taken with a 40 min difference of live wing discs of the designated genotypes.

Analysis of control *Ap>GFP* wing discs showed that dorsal cells stayed confined within their compartment throughout the incubation period (Figure 6B-B'', Movie S1). Conversely, a few GFP^+ cells from *Ap>GFP; Ras^{V12}* wing discs were observed leaving the dorsal compartment and moving into the GFP^- ventral compartment (Figure 6C-C'', Movie S2). Some of these cells looked smaller and rounder than control cells (arrowheads in C-C'') and we assumed these were dying cells. This was supported by live imaging analysis of *Ap>GFP; mys^{RNAi}* wing discs, showing that cells moving out of the ventral domain were also small and round (Figure 6D-D'', Movie S3). In contrast, we found that most *mys^{RNAi}; Ras^{V12}* wing disc cells looked normal and were able to move into the ventral domain further than *Ras^{V12}* cells (Figure 6E-D'', Movie S4). This was not just a consequence of the ability of *Ras^{V12}* to rescue the death of integrin mutant cells, as only a few small GFP^+ cells were found moving into the ventral compartment in *mys^{RNAi}; hid^{RNAi}* wing discs (Figure 6F-F'', Movie S5).

Together, this data demonstrates that integrins can restrict the ability of *Ras^{V12}* tumour cells to invade adjacent tissues.

Discussion

Integrins have been implicated in nearly every step of cancer progression, including initiation, proliferation, survival and metastasis. However, accumulating evidence has shown that integrins can act not only as tumour promoters but also as tumour suppressors (reviewed in [2]. This is especially the case of integrins that mediate cell adhesion to laminins, major components of the basement membranes surrounding most organs and tissues [38]. The ability of integrins to promote or suppress oncogenesis depends, among other factors, on the cell type, the oncogene context and the stage of the tumour, (reviewed in [39]. Here, we find that in the *Drosophila* wing disc epithelium the bPS containing integrins act as suppressors of tumours induced by a gain of function of the oncogenic form of *Ras*, *Ras^{V12}*. We show that while depletion of bPS integrins does not affect the polarity and proliferation of *Ras^{V12}* tumour cells, it increases their growth and invasive behaviour. In addition, we show that elimination of integrins enhances the ability of *Ras^{V12}* tumour cells to affect the tumour microenvironment.

To better understand the genetic bases and the development of cancer, researchers have used small animal models, such as mice, zebrafish and *Drosophila* [40]. In particular, mice are the most widely used to analyse the role of integrins in tumour development. The use of conditional genetic mouse models has pointed to opposing roles for the β_1 family of integrins as tumour enhancer or suppressors. Deletion of β_1 integrins blocks and delays cancer initiation in the Polyoma middle T (PyMT) and ErbB2 oncogene models, respectively [5, 41]. Likewise, in the RIPTag model for pancreatic cancer and in squamous cell carcinomas, deletion of β_1 integrins leads to impaired tumour growth and metastasis [42, 43]. In contrast, in other cancer models, β_1 integrins appear to play a tumour suppressor-like role. Thus, deletion of β_1 integrins in the TRAMP prostate adenocarcinoma model led to an increase in the expansion of the tumour cell population, an enhancement of the rate of tumour progression and a reduction in overall animal survival [44]. In contrast to the wide use of mice to study of the role of integrins in cancer development, few studies have been reported using the zebrafish or the *Drosophila* models to analyse this issue. The zebrafish is a vertebrate promising model rising in recent years, as it is small, transparent, low cost and reproduces fast. At present, zebrafish xenografts have been used to analyse the role of integrins in metastasis. These studies have shown that disrupting the expression of some integrins, such as β_1 , α_3 or α_6 in cancer cells reduces their ability to extravasate or metastasize when transplanted in zebrafish, suggesting a role for integrins as tumour promoters in this context [45-47]. Intriguingly, with respect to *Drosophila*, despite the fact that it has been extensively used to study cancer, there is little evidence for its use as a platform to analyse the role of integrins in tumour development. Using *Drosophila* Intestinal Stem Cell tumours in the adult midgut, it was shown that integrins were required for the initiation of hyperplasia upon loss of adenomatous polyposis coli and for the growth of tumours induced by gain of an oncogenic form of the transcription co-factor Yki [48, 49]. These results proposed a role for integrins as tumour promoting factors. In contrast, here, we show that integrins can act as tumour suppressors in the *Drosophila* wing imaginal disc epithelium, suggesting that, the dual role of integrins as tumour modulators is conserved. In the future, *Drosophila* could be used to understand the opposing tumour modulating activities conferred by integrins. This will facilitate further development of drugs targeting integrin signalling pathways.

The role of integrins in tumour growth is also complex. Thus, while loss of the $\alpha_2\beta_1$ integrin does not affect tumour growth *in vivo* or *in vitro*, the α_2 -null/Neu tumour cells demonstrate enhanced anchorage-independent growth [6]. In *Drosophila*, loss of integrin function inhibits the proliferation of Yki-driven intestinal stem cell tumours in the adult [49]. As mentioned above, ectopic expression of *Ras^{V12}* in the *Drosophila* wing disc produces hyperplasia, as a consequence of increased cell growth, accelerated G1-S transition and cell shape changes [24]. In agreement with this, we have recently shown that ectopic expression of *Ras^{V12}* in wing disc cells induces a cell shape change from columnar to cuboidal [25]. Here, we show that downregulation of integrin expression increases the formation

of extra folds due to *Ras^{V12}* overexpression, by enhancing cell shape changes and cellular growth. We have previously shown that removal of integrins on its own induces a change in cell shape similar to that caused by *Ras^{V12}* overexpression [18]. In addition, we have recently found that integrins regulate cell shape in the wing disc by modulating Myosin II activity and dynamics (<https://ssrn.com/abstract=4542879>). Similarly, increased EGFR/Ras signalling has been shown to affect cell shape through the regulation of Myosin II dynamics [50, 51]. Thus, the increase in the folding phenotype we observed in *Ras^{V12}* wing discs when downregulating integrin expression could just be an additive effect of disrupting simultaneously two pathways regulating cell shape independently. Alternatively, integrins could modulate oncogenic *Ras* signalling. In fact, many crosstalk between integrins and oncogenic *Ras* have been proposed, including the regulation of RTK activity by integrins [52]. However, in most cases these have been shown to increase *Ras* signalling (reviewed in [53], rather than limit it, which is what we find here. In the future, it will be interesting to investigate the mechanisms by which integrins restrict oncogenic *Ras* function.

Integrins not only regulate the properties of the cancer cells but also their ability to alter the tumour microenvironment, which includes the extracellular matrix and surrounding cells, stimulating non-autonomous tumour progression (reviewed in [54]. Here, we find that while elimination of integrins in *Ras^{V12}* wing disc cells does not seem to clearly affect the adjacent basement membrane, it increases the ability of the tumour cells to cause activation of the JNK pathway and non-autonomous death in neighbouring wild type tissue. This effect has been proposed to increase proliferation of tumour cells through the secretion of growth factors and Eiger (tumour necrosis factor homolog) [35, 36]. Here, we would like to propose that this non-autonomous effect could also contribute to tumour invasion by secreting factors that could potentiate the migratory behaviour of tumour cells. The contribution of the non-autonomous effects of cancer cells to tumour invasion is an exciting area for future investigation.

Most studies have shown that upregulation or overexpression of integrins associates with cancer metastasis, conferring integrins a pro-metastatic activity (reviewed in [39]. Furthermore, integrin inhibitors, such as monoclonal antibodies or RGD peptide analogs, are in clinical trials as metastatic suppressors [55]. However, other studies have reported that integrins can also inhibit metastasis, demonstrating that the role of integrins in cancer metastasis is also complex. For instance, the $\alpha_3\beta_1$ and $\alpha_2\beta_1$ integrins have been shown to suppress metastasis of prostate and breast cancer [6, 56-58]. Moreover, the same integrin can play opposite roles in the progression of different types of cancers. Depletion of the integrin α_9 reduces tumour metastasis in triple-negative breast cancer [59], while its overexpression suppresses hepatocarcinoma invasion [60]. Here, we find that while the expression of *Ras^{V12}* alone is not sufficient to create invasive tumours in wing disc epithelial cells, elimination of integrins in these tumour cells triggers an invasive behaviour. As integrins are the main receptor implicated in cell migration, how can their elimination induce invasion? Even though collective cell movement of primary melanoma explants requires β_1 -integrin mediated cell-matrix adhesion, impairment of cluster cohesion resulted in detachment and dissemination of amoeboid single tumour cells [61]. Similarly, when cultured in three-dimensional ECM scaffolds, antibodies that blocked β_1 integrin function or knockdown β_1 switched the migratory behaviour of human and mouse E-cadherin-positive triple-negative breast cancer (TNBC) cells from integrin-dependent collective movement to integrin-independent “amoeboid” crawling and dissemination [7]. We have previously shown that eliminating integrins from *Ras^{V12}* wing disc epithelial cells induces their detachment [14]. Here, we show that these tumour cells are able to move and invade nearby regions. Based on these results, we would like to suggest a conserved role for the β_1 family of integrins as promoters of epithelial phenotype and suppressors of dissemination. Understanding the apparent metastatic suppression activity conferred by integrins in certain cancer contexts is crucial to develop proper treatment strategies targeting integrins.

A variety of *Drosophila* cancer models has been established in several tissues besides the wing imaginal disc, such as the gut, the brain and the eye imaginal disc, by means of transgenesis, genome editing, transplantation and drug-induced toxic damage. These tumour models have provided new concepts and findings in cancer biology that have significant parallels and relevance to human cancer.

Our findings establish the *Drosophila* wing imaginal disc as a platform to address the complex role of integrins in cancer. We can now extend these studies to the other *Drosophila* cancer models to further explore the multiple biological functions mediated by integrins in cancers. Because targeting integrins has shown great potential in diagnosis and treatment of cancer, more mechanistic insights are warranted to fully understand the mechanisms of integrin-mediated biological behaviour of cancer cells.

Conclusions

Using the *Drosophila* wing disc epithelium, we show that the bPS integrin, the orthologue of vertebrate b1, can act as a suppressor of tumours induced by a gain of function of the oncogenic form of *Ras*, *Ras^{V12}*. We show that the bPS integrin restrains the growth, delamination and invasiveness of *Ras^{V12}* tumour cells, as well as their ability to affect the tumour microenvironment. These results strongly suggest that the role of integrin of the b1 family as tumour suppressors might be evolutionary conserved. In the future, it will be interesting to study the contribution of the bPS integrin to other *Drosophila* cancer models to further explore the multiple biological functions mediated by integrins in cancers. *Drosophila* can be used to understand the mechanisms underlying the complex tumour modulating activities conferred by integrins, thus facilitating the design of cancer drugs targeting integrins.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

Author Contributions: Conceptualization: M. D. Martín-Bermudo; Methodology: A. Martínez-Abarca Millán and M. D. Martín-Bermudo; Formal analysis: A. Martínez-Abarca Millán and M. D. Martín-Bermudo; Supervision: M. D. Martín-Bermudo ; Writing original draft & editing: A. Martínez-Abarca Millán and M. D. Martín-Bermudo; Funding acquisition: M. D. Martín-Bermudo.

Funding: This work was funded by the Spanish Minister of Science and Innovation (MCIN), grant numbers PID2019-109013GB-I00 and BFU2016-80797R, to MDMB and by the regional Agency Fundación Pública Progreso y Salud (Junta de Andalucía), grant number P20_00888, to MDMB. AMAM received a salary from MCIN (pre2020-092568). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Conflicts of interest: The authors declare no conflict of interest.

Acknowledgments: We thank Bloomington and Kyoto Stock Centers and the *Drosophila* community for fly stocks and reagents. We are also particularly grateful to Tomas Navarro Alvarez for his invaluable help in analyzing the statistical significance of our analysis.

References

1. Ferlay J, Colombet M, Soerjomataram I, Parkin DM, Pineros M, Znaor A, et al. Cancer statistics for the year 2020: An overview. *Int J Cancer*. 2021. Epub 2021/04/06. doi: 10.1002/ijc.33588. PubMed PMID: 33818764.
2. Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. *Nat Rev Cancer*. 2018;18(9):533-48. Epub 2018/07/14. doi: 10.1038/s41568-018-0038-z. PubMed PMID: 30002479; PubMed Central PMCID: PMC629548.
3. Humphries JD, Byron A, Humphries MJ. Integrin ligands at a glance. *J Cell Sci*. 2006;119(Pt 19):3901-3. Epub 2006/09/22. doi: 10.1242/jcs.03098. PubMed PMID: 16988024; PubMed Central PMCID: PMC629548.
4. Cagnet S, Faraldo MM, Kreft M, Sonnenberg A, Raymond K, Glukhova MA. Signaling events mediated by $\alpha 3 \beta 1$ integrin are essential for mammary tumorigenesis. *Oncogene*. 2014;33(34):4286-95. Epub 2013/10/01. doi: 10.1038/onc.2013.391. PubMed PMID: 24077284.
5. White DE, Kurpios NA, Zuo D, Hassell JA, Blaess S, Mueller U, et al. Targeted disruption of $\beta 1$ -integrin in a transgenic mouse model of human breast cancer reveals an essential role in mammary tumor induction. *Cancer Cell*. 2004;6(2):159-70. Epub 2004/08/25. doi: 10.1016/j.ccr.2004.06.025. PubMed PMID: 15324699.
6. Ramirez NE, Zhang Z, Madamanchi A, Boyd KL, O'Rear LD, Nashabi A, et al. The $\alpha 2 \beta 1$ integrin is a metastasis suppressor in mouse models and human cancer. *J Clin Invest*. 2011;121(1):226-37. Epub 2010/12/08. doi: 10.1172/JCI42328. PubMed PMID: 21135504; PubMed Central PMCID: PMC3007139.

7. Truong HH, Xiong J, Ghotra VP, Nirmala E, Haazen L, Le Devedec SE, et al. beta1 integrin inhibition elicits a prometastatic switch through the TGFbeta-miR-200-ZEB network in E-cadherin-positive triple-negative breast cancer. *Sci Signal*. 2014;7(312):ra15. Epub 2014/02/13. doi: 10.1126/scisignal.2004751. PubMed PMID: 24518294.
8. Brown N, Gregory SL, Martin-Bermudo MD. Integrins as mediators of morphogenesis in *Drosophila*. *Developmental Biology*. 2000;223:1-16.
9. Brower DL. Platelets with wings: the maturation of *Drosophila* integrin biology. *Current Opinion in Cell Biol*. 2003;15:607-13.
10. Bokel C, Brown NH. Integrins in Development: moving on, responding to, and sticking to the extracellular matrix. *Developmental Cell*. 2002;3:311-21.
11. Fernández-Miñán A, Martín-Bermudo MD, González-reyes A. Integrin signaling regulates spindle orientation in *Drosophila* to preserve the follicular-epithelium monolayer. *Current Biology*. 2007;17:683-8.
12. Lovegrove HE, Bergstralh DT, St Johnston D. The role of integrins in *Drosophila* egg chamber morphogenesis. *Development*. 2019;146(23). Epub 2019/12/01. doi: 10.1242/dev.182774. PubMed PMID: 31784458; PubMed Central PMCID: PMC6918751.
13. Santa-Cruz Mateos C, Valencia-Exposito A, Palacios IM, Martin-Bermudo MD. Integrins regulate epithelial cell shape by controlling the architecture and mechanical properties of basal actomyosin networks. *PLoS Genet*. 2020;16(6):e1008717. Epub 2020/06/02. doi: 10.1371/journal.pgen.1008717. PubMed PMID: 32479493; PubMed Central PMCID: PMC697263567.
14. Valencia-Exposito A, Gomez-Lamarca MJ, Widmann TJ, Martin-Bermudo MD. Integrins Cooperate With the EGFR/Ras Pathway to Preserve Epithelia Survival and Architecture in Development and Oncogenesis. *Front Cell Dev Biol*. 2022;10:892691. Epub 2022/07/01. doi: 10.3389/fcell.2022.892691. PubMed PMID: 35769262; PubMed Central PMCID: PMC9234701.
15. Tripathi BK, Irvine KD. The wing imaginal disc. *Genetics*. 2022;220(4). Epub 2022/03/05. doi: 10.1093/genetics/iyac020. PubMed PMID: 35243513; PubMed Central PMCID: PMC8982031.
16. Brower DL, Jaffe SM. Requirements for integrins during *Drosophila* wing development. *Nature*. 1989;342:285-7.
17. Wilcox M, DiAntonio A, Leptin M. The function of PS integrins in *Drosophila* wing morphogenesis. *Development*. 1989;107:891-7.
18. Dominguez-Gimenez P, Brown NH, Martin-Bermudo MD. Integrin-ECM interactions regulate the changes in cell shape driving the morphogenesis of the *Drosophila* wing epithelium. *J Cell Sci*. 2007;120(Pt 6):1061-71. doi: 10.1242/jcs.03404. PubMed PMID: 17327274.
19. Gonzalez C. *Drosophila melanogaster*: a model and a tool to investigate malignancy and identify new therapeutics. *Nat Rev Cancer*. 2013;13(3):172-83. Epub 2013/02/08. doi: 10.1038/nrc3461. PubMed PMID: 23388617.
20. Mirzoyan Z, Sollazzo M, Allocca M, Valenza AM, Grifoni D, Bellosta P. *Drosophila melanogaster*: A Model Organism to Study Cancer. *Front Genet*. 2019;10:51. doi: 10.3389/fgene.2019.00051. PubMed PMID: 30881374; PubMed Central PMCID: PMC6405444.
21. Forbes SA, Beare D, Gunasekaran P, Leung K, Bindal N, Boutselakis H, et al. COSMIC: exploring the world's knowledge of somatic mutations in human cancer. *Nucleic Acids Res*. 2015;43(Database issue):D805-11. doi: 10.1093/nar/gku1075. PubMed PMID: 25355519; PubMed Central PMCID: PMC4383913.
22. Lee T, Feig L, Montell DJ. Two distinct roles for Ras in a developmentally regulated cell migration. *Development*. 1996;122:409-18.
23. Villa-Fombuena G, Lobo-Pecellin M, Marin-Menguiano M, Rojas-Rios P, Gonzalez-Reyes A. Live imaging of the *Drosophila* ovarian niche shows spectrosome and centrosome dynamics during asymmetric germline stem cell division. *Development*. 2021;148(18). Epub 2021/08/10. doi: 10.1242/dev.199716. PubMed PMID: 34370012; PubMed Central PMCID: PMC8489027.
24. Prober DA, Edgar BA. Ras1 promotes cellular growth in the *Drosophila* wing. *Cell*. 2000;100(4):435-46. doi: 10.1016/s0092-8674(00)80679-0. PubMed PMID: 10693760.
25. Soler Beatty J, Molnar C, Luque CM, de Celis JF, Martin-Bermudo MD. EGFRAP encodes a new negative regulator of the EGFR acting in both normal and oncogenic EGFR/Ras-driven tissue morphogenesis. *PLoS Genet*. 2021;17(8):e1009738. Epub 2021/08/20. doi: 10.1371/journal.pgen.1009738. PubMed PMID: 34411095; PubMed Central PMCID: PMC8407591.

26. Genevet A, Polesello C, Blight K, Robertson F, Collinson LM, Pichaud F, et al. The Hippo pathway regulates apical-domain size independently of its growth-control function. *J Cell Sci.* 2009;122(Pt 14):2360-70. Epub 2009/06/18. doi: 10.1242/jcs.041806. PubMed PMID: 19531586; PubMed Central PMCID: PMCPMC2704876.
27. Brumby AM, Richardson HE. scribble mutants cooperate with oncogenic Ras or Notch to cause neoplastic overgrowth in *Drosophila*. *EMBO J.* 2003;22(21):5769-79. Epub 2003/11/01. doi: 10.1093/emboj/cdg548. PubMed PMID: 14592975.
28. Lee JL, Streuli CH. Integrins and epithelial cell polarity. *J Cell Sci.* 2014;127(Pt 15):3217-25. Epub 2014/07/06. doi: 10.1242/jcs.146142. PubMed PMID: 24994933; PubMed Central PMCID: PMCPMC4117227.
29. Goulas S, Conder R, Knoblich JA. The Par complex and integrins direct asymmetric cell division in adult intestinal stem cells. *Cell Stem Cell.* 2012;11(4):529-40. Epub 2012/10/09. doi: 10.1016/j.stem.2012.06.017. PubMed PMID: 23040479; PubMed Central PMCID: PMCPMC3465556.
30. Dillard C, Reis JGT, Rusten TE. Ras(V12); scrib(-/-) Tumors: A Cooperative Oncogenesis Model Fueled by Tumor/Host Interactions. *Int J Mol Sci.* 2021;22(16). Epub 2021/08/28. doi: 10.3390/ijms22168873. PubMed PMID: 34445578; PubMed Central PMCID: PMCPMC8396170.
31. Winkler J, Abisoye-Ogunniyan A, Metcalf KJ, Werb Z. Concepts of extracellular matrix remodelling in tumour progression and metastasis. *Nature communications.* 2020;11(1):5120. Epub 2020/10/11. doi: 10.1038/s41467-020-18794-x. PubMed PMID: 33037194; PubMed Central PMCID: PMCPMC7547708.
32. Karim FD, Rubin GM. Ectopic expression of activated Ras1 induces hyperplastic growth and increased cell death in *Drosophila* imaginal tissues. *Development.* 1998;125(1):1-9. PubMed PMID: 9389658.
33. Murcia L, Clemente-Ruiz M, Pierre-Elies P, Royou A, Milan M. Selective Killing of RAS-Malignant Tissues by Exploiting Oncogene-Induced DNA Damage. *Cell Rep.* 2019;28(1):119-31 e4. doi: 10.1016/j.celrep.2019.06.004. PubMed PMID: 31269434.
34. Nakamura M, Ohsawa S, Igaki T. Mitochondrial defects trigger proliferation of neighbouring cells via a senescence-associated secretory phenotype in *Drosophila*. *Nature communications.* 2014;5:5264. Epub 2014/10/28. doi: 10.1038/ncomms6264. PubMed PMID: 25345385.
35. Chabu C, Xu T. Oncogenic Ras stimulates Eiger/TNF exocytosis to promote growth. *Development.* 2014;141(24):4729-39. Epub 2014/11/21. doi: 10.1242/dev.108092. PubMed PMID: 25411211; PubMed Central PMCID: PMCPMC4299269.
36. Perez-Garijo A, Fuchs Y, Steller H. Apoptotic cells can induce non-autonomous apoptosis through the TNF pathway. *Elife.* 2013;2:e01004. Epub 2013/09/26. doi: 10.7554/eLife.01004. PubMed PMID: 24066226; PubMed Central PMCID: PMCPMC3779319.
37. Moreno E, Valon L, Levillayer F, Levayer R. Competition for Space Induces Cell Elimination through Compaction-Driven ERK Downregulation. *Curr Biol.* 2019;29(1):23-34 e8. Epub 2018/12/18. doi: 10.1016/j.cub.2018.11.007. PubMed PMID: 30554899; PubMed Central PMCID: PMCPMC6331351.
38. Ramovs V, Te Molder L, Sonnenberg A. The opposing roles of laminin-binding integrins in cancer. *Matrix Biol.* 2017;57-58:213-43. Epub 2016/08/27. doi: 10.1016/j.matbio.2016.08.007. PubMed PMID: 27562932.
39. Su CY, Li JQ, Zhang LL, Wang H, Wang FH, Tao YW, et al. The Biological Functions and Clinical Applications of Integrins in Cancers. *Front Pharmacol.* 2020;11:579068. Epub 2020/10/13. doi: 10.3389/fphar.2020.579068. PubMed PMID: 33041823; PubMed Central PMCID: PMCPMC7522798.
40. Li Z, Zheng W, Wang H, Cheng Y, Fang Y, Wu F, et al. Application of Animal Models in Cancer Research: Recent Progress and Future Prospects. *Cancer Manag Res.* 2021;13:2455-75. Epub 2021/03/25. doi: 10.2147/CMAR.S302565. PubMed PMID: 33758544; PubMed Central PMCID: PMCPMC7979343.
41. Huck L, Pontier SM, Zuo DM, Muller WJ. beta1-integrin is dispensable for the induction of ErbB2 mammary tumors but plays a critical role in the metastatic phase of tumor progression. *Proc Natl Acad Sci U S A.* 2010;107(35):15559-64. Epub 2010/08/18. doi: 10.1073/pnas.1003034107. PubMed PMID: 20713705; PubMed Central PMCID: PMCPMC2932605.
42. Tran T, Barlow B, O'Rear L, Jarvis B, Li Z, Dickeson K, et al. Loss of the alpha2beta1 integrin alters human papilloma virus-induced squamous carcinoma progression in vivo and in vitro. *PLoS One.* 2011;6(10):e26858. Epub 2011/11/03. doi: 10.1371/journal.pone.0026858. PubMed PMID: 22046385; PubMed Central PMCID: PMCPMC3203166.
43. Kren A, Baeriswyl V, Lehembre F, Wunderlin C, Strittmatter K, Antoniades H, et al. Increased tumor cell dissemination and cellular senescence in the absence of beta1-integrin function. *EMBO J.* 2007;26(12):2832-42. Epub 2007/06/02. doi: 10.1038/sj.emboj.7601738. PubMed PMID: 17541405; PubMed Central PMCID: PMCPMC1894776.

44. Moran-Jones K, Ledger A, Naylor MJ. beta1 integrin deletion enhances progression of prostate cancer in the TRAMP mouse model. *Sci Rep.* 2012;2:526. Epub 2012/07/26. doi: 10.1038/srep00526. PubMed PMID: 22829980; PubMed Central PMCID: PMC3402831.
45. Du Q, Wang W, Liu T, Shang C, Huang J, Liao Y, et al. High Expression of Integrin alpha3 Predicts Poor Prognosis and Promotes Tumor Metastasis and Angiogenesis by Activating the c-Src/Extracellular Signal-Regulated Protein Kinase/Focal Adhesion Kinase Signaling Pathway in Cervical Cancer. *Front Oncol.* 2020;10:36. Epub 2020/03/03. doi: 10.3389/fonc.2020.00036. PubMed PMID: 32117712; PubMed Central PMCID: PMC7033469.
46. Stoletov K, Kato H, Zardouzian E, Kelber J, Yang J, Shattil S, et al. Visualizing extravasation dynamics of metastatic tumor cells. *J Cell Sci.* 2010;123(Pt 13):2332-41. Epub 2010/06/10. doi: 10.1242/jcs.069443. PubMed PMID: 20530574; PubMed Central PMCID: PMC2886748.
47. Tanaka S, Senda N, Iida A, Sehara-Fujisawa A, Ishii T, Sato F, et al. In silico analysis-based identification of the target residue of integrin alpha6 for metastasis inhibition of basal-like breast cancer. *Genes Cells.* 2019;24(9):596-607. Epub 2019/07/12. doi: 10.1111/gtc.12714. PubMed PMID: 31295752.
48. Lin G, Zhang X, Ren J, Pang Z, Wang C, Xu N, et al. Integrin signaling is required for maintenance and proliferation of intestinal stem cells in *Drosophila*. *Dev Biol.* 2013;377(1):177-87. doi: 10.1016/j.ydbio.2013.01.032. PubMed PMID: 23410794.
49. Bajpai A, Quazi TA, Tang HW, Manzar N, Singh V, Thakur A, et al. A *Drosophila* model of oral peptide therapeutics for adult intestinal stem cell tumors. *Dis Model Mech.* 2020;13(7). Epub 2020/06/17. doi: 10.1242/dmm.044420. PubMed PMID: 32540914; PubMed Central PMCID: PMC7390633.
50. Saxena A, Denholm B, Bunt S, Bischoff M, VijayRaghavan K, Skaer H. Epidermal growth factor signalling controls myosin II planar polarity to orchestrate convergent extension movements during *Drosophila* tubulogenesis. *PLoS Biol.* 2014;12(12):e1002013. Epub 2014/12/03. doi: 10.1371/journal.pbio.1002013. PubMed PMID: 25460353; PubMed Central PMCID: PMC4251826.
51. Tamada M, Zallen JA. Square Cell Packing in the *Drosophila* Embryo through Spatiotemporally Regulated EGF Receptor Signaling. *Dev Cell.* 2015;35(2):151-61. Epub 2015/10/28. doi: 10.1016/j.devcel.2015.09.015. PubMed PMID: 26506305; PubMed Central PMCID: PMC4939091.
52. Desgrosellier JS, Cheresh DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer.* 2010;10(1):9-22. Epub 2009/12/24. doi: 10.1038/nrc2748. PubMed PMID: 20029421; PubMed Central PMCID: PMC4383089.
53. Bianconi D, Unseld M, Prager GW. Integrins in the Spotlight of Cancer. *Int J Mol Sci.* 2016;17(12). Epub 2016/12/09. doi: 10.3390/ijms17122037. PubMed PMID: 27929432; PubMed Central PMCID: PMC5187837.
54. Enomoto M, Vaughen J, Igaki T. Non-autonomous overgrowth by oncogenic niche cells: Cellular cooperation and competition in tumorigenesis. *Cancer Sci.* 2015;106(12):1651-8. Epub 2015/09/13. doi: 10.1111/cas.12816. PubMed PMID: 26362609; PubMed Central PMCID: PMC4714670.
55. Avraamides CJ, Garmy-Susini B, Varner JA. Integrins in angiogenesis and lymphangiogenesis. *Nat Rev Cancer.* 2008;8(8):604-17. Epub 2008/05/24. doi: 10.1038/nrc2353. PubMed PMID: 18497750; PubMed Central PMCID: PMC2577722.
56. Zutter MM, Santoro SA, Staatz WD, Tsung YL. Re-expression of the alpha 2 beta 1 integrin abrogates the malignant phenotype of breast carcinoma cells. *Proc Natl Acad Sci U S A.* 1995;92(16):7411-5. Epub 1995/08/01. doi: 10.1073/pnas.92.16.7411. PubMed PMID: 7638207; PubMed Central PMCID: PMC41349.
57. Varzavand A, Hacker W, Ma D, Gibson-Corley K, Hawayek M, Tayh OJ, et al. alpha3beta1 Integrin Suppresses Prostate Cancer Metastasis via Regulation of the Hippo Pathway. *Cancer Res.* 2016;76(22):6577-87. Epub 2016/11/05. doi: 10.1158/0008-5472.CAN-16-1483. PubMed PMID: 27680681; PubMed Central PMCID: PMC5290210.
58. Ramos-Lewis W, Page-McCaw A. Basement membrane mechanics shape development: Lessons from the fly. *Matrix Biol.* 2019;75-76:72-81. Epub 2018/04/16. doi: 10.1016/j.matbio.2018.04.004. PubMed PMID: 29656148; PubMed Central PMCID: PMC6185827.
59. Wang Z, Li Y, Xiao Y, Lin HP, Yang P, Humphries B, et al. Integrin alpha9 depletion promotes beta-catenin degradation to suppress triple-negative breast cancer tumor growth and metastasis. *Int J Cancer.* 2019;145(10):2767-80. Epub 2019/04/23. doi: 10.1002/ijc.32359. PubMed PMID: 31008533; PubMed Central PMCID: PMC6750961.

60. Zhang YL, Xing X, Cai LB, Zhu L, Yang XM, Wang YH, et al. Integrin alpha9 Suppresses Hepatocellular Carcinoma Metastasis by Rho GTPase Signaling. *J Immunol Res.* 2018;2018:4602570. Epub 2018/06/29. doi: 10.1155/2018/4602570. PubMed PMID: 29951557; PubMed Central PMCID: PMC5989280.
61. Hegerfeldt Y, Tusch M, Brocker EB, Friedl P. Collective cell movement in primary melanoma explants: plasticity of cell-cell interaction, beta1-integrin function, and migration strategies. *Cancer Res.* 2002;62(7):2125-30. Epub 2002/04/04. PubMed PMID: 11929834.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.