

Review

Not peer-reviewed version

# Unraveling the Cave: A Seventy-Year Journey into Caveolar Network, Cellular Signaling, and Human Disease.

<u>Alessio D'Alessio</u>\*

Posted Date: 30 October 2023

doi: 10.20944/preprints202310.1924.v1

Keywords: calcium signaling, caveolae; caveolin-1; cell senescence; ECs, eNOS, endocytosis; lipid rafts; signal transduction.



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Remiero

# Unraveling the Cave: A Seventy-Year Journey into Caveolar Network, Cellular Signaling, and Human Disease

#### Alessio D'Alessio 1,2

- Dipartimento di Scienze della Vita e Sanità Pubblica, Sezione di Istologia ed Embriologia, Università Cattolica del Sacro Cuore, 00168 Roma, Italy
- <sup>2</sup> Fondazione Policlinico Universitario "Agostino Gemelli", IRCCS, 00168 Rome, Italy
- \* Correspondence: alessio.alessio@unicatt.it or alessio.dalessio@unicatt.it

Abstract: In the mid-1950s, a groundbreaking discovery revealed the fascinating presence of caveolae, referred to as flask-shaped invaginations of the plasma membrane, sparking renewed excitement in the field of cell biology. Caveolae are small, flask-shaped invaginations in the cell membrane that play crucial roles in diverse cellular processes, including signal transduction, endocytosis, lipid homeostasis, and signal transduction. The structural stability and functionality of these specialized membrane microdomains are attributed to the coordinated activity of scaffolding proteins, including caveolins and cavins. While caveolae and caveolins have been long appreciated for their integral roles in cellular physiology, the accumulating scientific evidence throughout the years reaffirms their association with a broad spectrum of human disorders. This review article aims to offer a thorough account of the historical advancements in caveolae research, spanning from their initial discovery to the recognition of caveolin family proteins and their intricate contributions to cellular functions. Furthermore, it will examine the consequences of a dysfunctional caveolar network in the development of human diseases.

**Keywords:** calcium signaling; caveolae; caveolin-1; cell senescence; ECs; eNOS; endocytosis; lipid rafts; signal transduction

#### 1. Introduction

Caveolae are small, flask-shaped invaginations in the cell membrane that belong to a specialized subgroup of non-planar (invaginated) lipid rafts in the cell plasma membrane of eukaryotic cells, involved in various cellular processes. Lipid rafts and caveolae are both specialized cell membrane microdomains, but they have distinct characteristics and functions. Both structures are enriched in specific lipid components, particularly cholesterol and sphingolipids, that make these membrane regions more ordered compared to the surrounding lipid bilayer. Lipid rafts are flat, smaller, less organized, and more fluid in nature compared to caveolae [1]. They serve as central hubs for bringing together signaling molecules, which are essential for facilitating favorable interactions required for signal transduction. Conversely, caveolae have a well-defined, stable, and rigid structure, owing to the presence of distinct scaffolding proteins that grant them their characteristic omega-shaped appearance [2]. According to electron microscopy studies, caveolae have a diameter ranging from 60 to 80 nm, with a narrow neck of 10 to 50 nm. These microscopic structures serve crucial roles in numerous physiological functions, including the regulation of lipid metabolism, intricate intracellular signaling adjustments, and participation in vital processes such as membrane repair, mechanosensation, and cellular trafficking [3-6]. Furthermore, their implication in a variety of pathologies, such as cancer, cardiovascular disorders, and neurodegenerative conditions, offers intriguing prospects for future research [7-9]. This article seeks to provide a concise overview of pivotal discoveries in the caveolae field, highlighting their profound significance in cellular biology and their potential as promising therapeutic targets for a spectrum of human diseases.

#### 2. Historical milestones in caveolae research

The study of caveolae has seen several significant milestones since their discovery, enhancing our comprehension of these unique membrane features (Figure 1). In the mid-1950s early electron microscopy investigations into cell membranes in continuous endothelium unveiled characteristic striated flask-shaped structures by George Emil Palade, who referred to them as "plasmalemmal vesicles" [10]. In 1953, Eichi Yamada found similar features in the gall bladder epithelium and named them "caveolae intracellulares" due to their resemblance to "little caves" [11]. From the time they were first identified, increasing number of studies has consistently detected caveolae-like structures in many cell types, including adipocytes, endothelial cells (ECs), fibroblasts, muscle cells, and epithelial cells [12-15]. ECs show a high prevalence of caveolae, and in adipocytes they can make up 30 to 50% of the cell surface [16]. However, some eukaryotic cells, including red blood cells (erythrocytes), lymphocytes [17], and some epithelial cells [18] do not possess these structures. In the nervous system, the presence of caveolin proteins had remained elusive for a long time, contributing to the prevailing notion that caveolin proteins were absent in brain tissue and neurons. The perspective shifted in the late 1990s with the discovery of caveolins in different types of brain cells, including brain ECs, glial cells, and neurons [19-21]. It is noteworthy that neurons seemingly express all three caveolins, although certain authors concur that they might be devoid of caveolae, thus implying a potential function for these proteins beyond the confines of the plasma membrane caveolae [22,23]. These hypotheses find support in studies indicating that caveolins can be found in both buoyant and heavy fractions resulting from density gradient centrifugation, implying a more extensive distribution than solely within caveolae. Consideration of these findings is imperative when conducting investigations focused on endogenous caveolin protein expression, such as through RNA interference (RNAi). In such scenarios, it becomes challenging to distinguish between the effects of caveolins localized within caveolae and those found outside, in cells that exhibit caveolar structures. In the ever-evolving landscape of cellular biology, the continuous investigation of the caveolar network promises to reveal new insights into the complex mechanisms that govern cell function and has significant implications for our understanding of human diseases and their treatment.

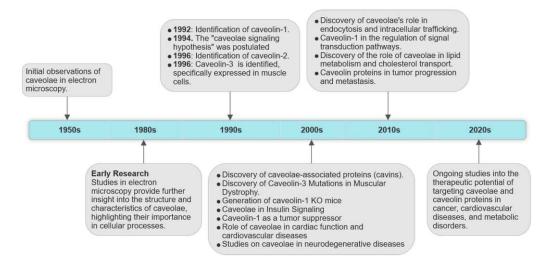


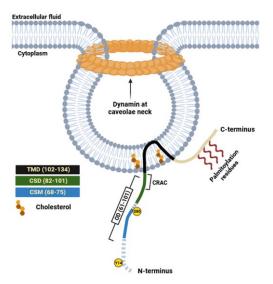
Figure 1. Timeline of major discoveries in the field of caveolae research.

# 3. Caveolins and cavins: synthesis and post-translational modifications

#### 3.1. Caveolin-1

The first recognized protein marker of caveolae, called caveolin-1, was not discovered until the early 1990s (Figure 1). This finding offered valuable insights into the structural and functional characteristics of caveolae [13]. The same year Glenney cloned and sequenced a human cDNA encoding the caveolin-1 (CAV1) gene [24], translating a full-length protein of 178 amino acids that was expressed on the surface and in the cytoplasm of ECs of pulmonary arteries [25]. This was a

pivotal discovery, as this monotopic membrane protein was found to be a major structural coat protein playing a crucial role in caveolae biogenesis [26]. A structural examination has unveiled the organization of caveolin-1 monomers into functional oligomers within the membrane [27]. Each caveolin monomer consists of distinct regions, which include: i) a cytosolic N-terminal domain that faces the cytoplasm; ii) a C-terminal domain, encompassing residues 135 to 178; iii) a transmembrane domain (TMD) spanning residues 102 to 134; iv) an oligomerization domain (OD), spanning residues 61 to 101, that incorporates both the highly conserved "FEDVIAEP" caveolin signature motif (CSM, residues 68 to 75) and the caveolin scaffolding domain (CSD, residues 82 to 101) [28-31] (Figure 2). Notably a cholesterol recognition/interaction amino acid consensus (CRAC) "VTKYWFYR" spans residues 94-101 within the CSD, allows caveolin-1 to interact with cholesterol molecules. Two isoforms of caveolin-1 have been elucidated, referred to as caveolin-1 $\alpha$  (including residues 1–178) and caveolin- $1\beta$  (spanning residues 32–178). While the precise role of these two protein variants in caveolae formation is still largely obscure, it appears that the  $\alpha$  isoform demonstrates greater efficacy in driving the genesis of caveolae compared to the  $\beta$  isoform [32]. Caveolin-1 undergoes posttranslational modification (PTM) through palmitoylation, which involves the covalent attachment of palmitic acid to three distinct cysteine residues within the TMD domain (Figure 2). Palmitoylation of proteins by palmitoyl transferases is generally a reversible process, playing a crucial role in regulating various cellular functions. This includes the control of protein localization, membrane organization, and signal transduction processes, ultimately contributing to the overall stability and functionality of the cell. Studies involving a palmitate-deficient mutant of caveolin-1 have revealed that palmitoylation of caveolin-1 is not essential for its localization to caveolae [33], and is irreversible in ECs [34]. Caveolin-1 can also undergo phosphorylation [35], occurring both at tyrosine 14 and at serine 80 (Figure 2). Although the functional implications of caveolin-1 phosphorylation have been relatively understudied, the two phosphorylation sites seem to serve distinct purposes. Tyrosine phosphorylation of caveolin-1 can be facilitated by various kinases, including those in the Src family, and it can influence the response to oxidative stress, promote cell senescence in chondrocytes, and modulate autophagy [36-38]. In contrast, phosphorylation of the serine residue in caveolin-1 influences its cellular localization within the endoplasmic reticulum, directing it towards the secretory pathway [39]. Ubiquitination can also regulate the expression of caveolin-1. When caveolae assembly is dysfunctional or the caveolin-1 levels increase, it can be ubiquitinated and subsequently degraded within lysosomes [40]. More recently, it has been shown that degradation of caveolin-1 by the E3 ubiquitin ligase ZNRF1 is vital for the regulation of the TLR4-mediated inflammatory pathway [41].



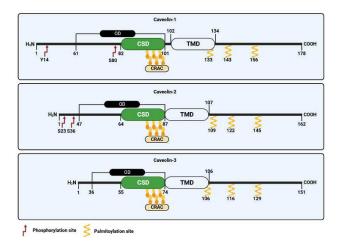
**Figure 2.** Schematic illustration of caveolin-1. CRAC sequence "VTKYWFYR" spanning residues 94-101; CSD, Caveolin Scaffolding Domain; OD, Oligomerization Domain; TDM, transmembrane domain. Two phosphorylation sites at tyrosine 14 (Y14) and serine 80 (S80) are indicated.

#### 3.2. Caveolin-2.

In 1996, caveolin-2, the second member of the caveolin family, was identified and cloned by Scherer and collaborators [29]. Caveolin-2 forms hetero-oligomers with caveolin-1 [2], and both proteins are found typically co-expressed in the same cells [29,42]. Unlike caveolin-1, the lack of caveolin-2 does not affect the formation of caveolae. However, in specific cell types, the concurrent presence of both proteins appears to enhance caveolae formation more effectively than caveolin-1 alone [32]. Like caveolin-1, caveolin-2 show multiple isoforms characterized by distinct but partially overlapping subcellular distributions [43]. These include the full-length caveolin- $2\alpha$  and two truncated variants known as caveolin- $2\beta$  and caveolin- $2\gamma$ . Nonetheless, the functional significance of these caveolin-2 isoforms remains largely unknown [44]. Like caveolin-1, caveolin-2 undergoes posttranslational modifications, including myristoylation at a single glycine residue and palmitoylation at cysteine 109, 122, and 145, which mainly control protein localization in the plasma membrane (Figure 3). Recently, it has been revealed that acylation of caveolin-2 directs the protein to be phosphorylated by the insulin receptor tyrosine kinase, emphasizing its potential role in regulating insulin signaling [45]. Caveolin-2 can be also phosphorylated at serine 23 and 36, and this modification is crucial to sustain caveolin-1 activity during caveolae biogenesis [46]. Two additional sites of phosphorylation, namely tyrosine 19 and 27, have been identified within caveolin-2 that play crucial function in regulating cell signaling by promoting protein interaction with SH2 bearing domains [47,48], and is involved in adipocyte hypertrophy [49].

# 3.3. Caveolin-3.

The third member of the caveolin gene family, termed caveolin-3 was discovered in 1996 by Tang and collaborators [30]. Caveolin-3 is predominantly expressed in mature smooth, cardiac, and skeletal muscle cells [50]. Like caveolin-1, caveolin-3 shows distinct structural domains including i) an N-terminal domain, spanning residues 1-54, which includes the FEDVIAEP CSM (residues 41-48); ii) a CSD spanning residues 55–74; iii) a TMD (residues 75–106); and iv) a C-terminal domain (residues 107-151, facing the cytoplasm) [51] (Figure 3). Caveolin-3 is also subjected to post translation palmitoylation at three different cysteine residues [52]. While caveolin-3 is shorter in length compared to caveolin-1, it exhibits significant structural and functional similarities to caveolin-1 and possesses the ability to independently form caveolae [30]. The caveolin-3 (CAV3) gene encodes an open reading frame of 151 amino acids that is 65% identical and 85% like caveolin-1. In addition to palmitoylation, caveolin-3 undergoes post-translational modification through SUMOylation, which is a process involving the attachment of Small Ubiquitin-like Modifier (SUMO) to its Nterminus. This modification of caveolin-3 plays a crucial role in regulating the expression of βadrenergic receptors [53]. Mutations in CAV3 gene lead to a series of a family genetic disorders known as caveolinopathies [54], leading to some rare forms of muscle dysfunction, such as muscular dystrophies, myopathies, and arrhythmias. Four different caveolinopathies have been identified, including Limb-Girdle Muscular Dystrophy (LGMD) 1C, Rippling Muscle Disease (RMD), isolated HyperCKemia, and Distal Myopathy [54,55]. Within skeletal myofibers caveolin-3 takes part into the dystrophin-glycoprotein complex (DGC), serving as a link between the extracellular matrix and intracellular cytoskeletal elements. The connection between caveolin-3 and DGC within muscle cells plays a pivotal role in upholding the structural integrity of the sarcolemma, enabling the normal functioning of muscle cells [50]. Although post-translational modifications of caveolin-3 have received limited attention, there is evidence to suggest that mutations in the CAV3 gene, which result in muscular dystrophy, are broken down through ubiquitination and proteasomal pathways. Blocking proteasomal degradation could potentially restore normal caveolin-3, which holds promise for the treatment of LGMD-1C from a clinical perspective [55].



**Figure 3.** Schematic representation of the caveolin family of proteins. CRAC, cholesterol recognition/interaction amino acid consensus; OD, oligomerization domain; TDM, transmembrane domain; CSD, caveolin scaffolding domain.

#### 3.3. The cavin family of proteins.

In the 2000s, the identification of novel structural constituents of caveolae, known as caveolae-associated proteins (cavins), has augmented our comprehension of the intricate architecture of caveolae [56–58] (Figure 1). Cavin proteins, including cavins 1-4, encoded by the polymerase I and transcript release factor (PTRF), serum-deprivation-response protein (SDPR), SDPR-related gene product that binds to C-kinase (SRBC), and muscle-related coiled-coil protein (MURC) genes, respectively, assume a pivotal role in the biogenesis of caveolae. Among cavin proteins, only cavin-1 appears crucial for the correct biogenesis of caveolae [59], while the other three members of the family have been suggested to contribute to caveolae stabilization [60–62], and appear to display tissue-specific distributions. Their intricate interplay with caveolin proteins and their involvement in membrane dynamics impart this family of proteins with essential significance in many cellular processes and signal transduction mechanisms associated with the presence and functionality of caveolae [63,64]. The structure and function of the cavin family of proteins are reviewed by Kovtun et al. [64].

# 4. Functional roles of caveolae and caveolins.

# 4.1. Caveolae and caveolin in endocytosis and transcytosis

Endocytosis is a fundamental cellular process through which cells internalize molecules, particles, or fluids from their external surroundings for various purposes, including nutrient uptake, receptor internalization, and removal of waste. This cellular uptake mechanism encompasses three primary forms: phagocytosis (engulfment of large particles), pinocytosis (the taking up of small, dissolved molecules and fluids), and receptor-mediated endocytosis [65]. Transcytosis refers to the transport of materials from one side of the cell to the other (e.g., from the apical to the basolateral membrane or vice versa), thereby making them accessible to neighboring cells and different tissue compartments. It is particularly involved in polarized epithelial cells that line body surfaces like endothelia and intestinal epithelium. Functionally, transcytosis combines endocytosis (uptake at one membrane) and exocytosis (release at the opposite membrane) [66,67]. The strategic positioning of caveolae in the plasma membrane, their unique shape and composition, and the ability of their scaffolding proteins to interact with multiple molecules all make them ideal for aiding in the uptake of substances into the cell. While early studies have depicted caveolae as stationary structures within the cell [68,69], an increasing body of evidence now proves their capacity to drive internalization of molecules that interact with the caveolar network. The observation revealed a notably slow rate of exchange between the plasma membrane and the intracellular compartment for a fusion protein consisting of caveolin-1 tagged with green fluorescent protein (GFP). This suggests that the caveolae

located within the plasma membrane are relatively immobile structures. Moreover, the study indicates that an increased expression of caveolin-1 acts as a rate-limiting factor for the internalization of caveolae, implying that the protein itself may have a role in stabilizing caveolae on the plasma membrane. A widely accepted hypothesis suggests that caveolae generally do not participate in endocytosis. Nonetheless, when specific ligands interact with caveolae or caveolins, they can induce the rapid internalization of these structures. Therefore, in addition to the well-known endocytic pathway mediated by clathrin-coated vesicles (CCV) [70], caveolae-dependent endocytosis stands for an alternative high specific component of cellular uptake. This mechanism appears to rely on the phosphorylation of caveolin-1 and caveolin-2, as indicated by studies conducted in the presence of phosphatase inhibitors that significantly enhance the internalization of caveolae [71,72]. Notably, fibroblasts from caveolin1-deficient mice cannot internalize albumin compared to their wild-type counterparts, supplying further confirmation of the integral role played by this protein in transcytosis [73]. Molecules that undergo internalization via caveolae include albumin, folic acid, alkaline phosphatase, lipids, insulin, low-density lipoproteins (LDL), chemokines [74-78], and certain pathogens such as toxins, viruses and bacteria [79-90]. It is important to mention that the GTPase protein dynamin, recognized for its role in easing the pinching off clathrin-coated vesicles [91] is also recruited to the caveolar neck [92,93]. This implies that dynamin oversees both clathrin-dependent endocytosis and the internalization of caveolae. On the other hand, in recent reports, it has been suggested that dynamin plays a role in stabilizing caveolae by keeping them restricted to the plasma membrane and preventing their release into the cell [94]. In conclusion, the pivotal roles of the caveolar network in cellular uptake suggest that targeting these components could hold great promise for advancing therapies in drug delivery and disease treatment.

# 4.2. Caveolins in cell signaling.

The "caveolae signaling hypothesis" proposed in the mid-1990s (Figure 1), is a concept related to the discovery of the caveolin family of proteins and their role in cellular signaling [95]. This hypothesis implies that caveolae and their associated proteins may act as organizers or modulators of signaling events within the cell. This organization facilitates the spatial regulation of various signaling pathways, and their dysfunction or dysregulation could have implications for various physiological processes or disease states. This hypothesis stems from the observation of specific structural and molecular traits of the caveolar platform. As caveolin-1 expression controls the abundance of caveolae at the plasma membrane [96], it could potentially change the functioning of signaling molecules by regulating the number of caveolae present on the cell surface. Moreover, the aptitude of caveolin-1 to interact with signaling molecules containing a caveolin-binding motif, thereby leading to their confinement within the caveolae network, has been shown to regulate downstream intercellular processes. Caveolin-1 has been shown to interact with RTKs like the epidermal growth factor receptor (EGFR) [97] and insulin receptor (IR) [98,99], thereby modulating their downstream signaling. These interactions have significant implications for cellular proliferation and survival. Caveolins also influence GPCR-mediated signaling [100]. The role of caveolin-1 in GPCR endocytosis and desensitization is well-established, impacting processes ranging from neurotransmission to immune responses. Additional signaling molecules can be regulated through their confinement within the caveolae network or interaction with caveolin-1 [101–108]. In the context of endothelial function, caveolins play a crucial role in regulating nitric oxide (NO) signaling. Caveolin-1 physically interacts with eNOS, restricting its enzymatic activity and preventing the untimely production of nitric oxide (NO). In situations of physiological stimulation, such as increased shear stress or specific agonists, caveolin-1 can dissociate from eNOS, allowing the enzyme to become activated and generate NO. This dynamic interaction between caveolin-1 and eNOS serves as a crucial mechanism for maintaining vascular homeostasis, as NO serves as a potent vasodilator and regulator of blood pressure, impacting various physiological and pathological processes, including cardiovascular health [109-111]. In addition, post-translation phosphorylation of caveolin-1 can affect downstream signaling. For instance, phosphorylation of caveolin-1 plays a crucial role in regulating autophagy under conditions of oxidative stress and cerebral ischemic injury, resulting in protecting

against cellular damage caused by oxidative stress, and potentially reducing cerebral infarct damage during ischemic events [37]. Moreover, tyrosine phosphorylation of caveolin-1 in ECs exposed to oxidative stress or alterations in flow conditions can govern both cell permeability and mechanotransduction responses [112,113], and its inhibition hinders Rac1/Cdc42-mediated axonal growth in human neuronal progenitor cells [114].

Like caveolin-1, caveolin-2 phosphorylation plays a critical role in the regulation of signal transduction, specifically in the context of cellular communication and intracellular signaling. This post-translational modification of caveolin-2 has been shown to impact its interactions with various signaling molecules [115]. In conclusion, modulation of signaling cascades through caveolins phosphorylation is a key mechanism for fine-tuning cellular responses and proper cellular functions that deserves further investigation.

#### 4.3. Caveolins in host cell response and inflammation

Inflammation is the body's response to infections and tissue damage in vascularized tissues. The function of this process is to transfer cells and molecules responsible for host defense from the circulation to the injured sites, ultimately leading to the elimination of the offending agents. It is a complex and tightly regulated physiological response triggered by the host immune system to combat various harmful stimuli. The typical inflammatory reaction involves a series of coordinated steps, including the recognition of harmful stimulus (e.g., injury, infection, or tissue damage), recruitment of white blood cells, particularly neutrophils and macrophages to the site of inflammation, leukocyte activation, and resolution of inflammation. Interestingly, the caveolar network can take part to these events. First, pathogens like viruses and bacteria can use all the molecules found in caveolae, which are essential for vesicle formation and fusion, to enter the host cell [72,85,86,92,116-124]. Of note, since caveolae don't follow the typical path to lysosomes, pathogens that enter through these structures can remain viable and avoid getting broken down in lysosomes [125]. A crucial aspect of inflammation involves leukocyte adhesion to ECs [126]. During this process, leukocytes first interact with adhesion molecules on EC surface, such as E- and Pselectin, and later with intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion protein 1 (VCAM-1). Interestingly, there are reports indicating that ICAM-1 plays a role in helping white blood cells exit the bloodstream by moving to areas rich in caveolin-1 within the lateral portion of ECs. This relocation of ICAM-1 is believed to promote the formation of pseudopodia by leukocytes, facilitating their exit into the surrounding tissue [127,128]. A study by Michell et al. found that exposure to high pressure led to increased leukocyte adhesion and inflammation in rat carotid arteries is triggered by the mechanosensory capability of caveolin-1. Consequently, mice and ECs lacking caveolin-1 were protected from pressure-induced inflammation, highlighting the significance of caveolin-1 in pressure-induced vascular inflammation [129]. Supporting the role of the caveolar network in recruiting white blood cells, it has been shown that caveolin-1 reduces excessive NOrelated permeability in blood vessels and facilitates white blood cell adhesion through a specific mechanism involving ICAM-1 and Src activation [130]. Besides caveolin-1, caveolin-2 has been also found to facilitate leukocyte infiltration in brain ECs, contributing to the development of common neurodegenerative diseases [131]. In addition, this protein has been suggested to play a protective role in reducing postischemic tissue injury by dampening plasminogen activator inhibitor-1 (PAI-1) expression, leading to reduced leukocyte recruitment by ECs. Authors suggest that targeting PAI-1 or enhancing caveolin-2 expression could be potential therapeutic strategies to reduce tissue damage caused by ischemia/reperfusion injury [132]. These findings highlight the intricate involvement of caveolae and caveolins in the inflammatory response and underscore their vital role in regulating cellular signaling and immune processes.

# 4.4. Caveolin proteins and mechanotransduction

Mechanotransduction, the process by which cells sense and respond to mechanical forces, plays a pivotal role in numerous physiological and pathological processes [133–135]. Caveolins not only modulate intracellular signaling cascades [136] but influence the mechanical properties of the plasma

membrane itself [7]. These features make caveolins crucial players in the ability of cells to sense and respond to mechanical cues, which is fundamental to various physiological processes, including tissue development, vascular homeostasis, and cancer metastasis [137]. Caveolin-1 is the most extensively studied isoform in this context, thanks to its capability to regulate and organize signaling molecules in caveolae, facilitating their rapid response to mechanical stimuli. Its involvement in mechanosensing has been evidenced in several studies, such as one by Drab et al., that highlighted the role of caveolin-1 in the activation of eNOS in ECs in response to shear stress [138]. Furthermore, caveolins can regulate the activation of integrins and focal adhesion kinase, influencing cell adhesion and migration in response to mechanical cues [139]. In ECs and muscle cells, caveolae can serve as a repository for membranes, which can undergo flattening when the cell experiences mechanical stress [139]. Therefore, unstimulated cells have excess membrane confined within invaginated caveolae, granting them the ability to alleviate plasma membrane tension, thereby effectively counteracting mechanical stress. In such a scenario, the reconfiguration of the plasma membrane after the breakdown of caveolae would thereby facilitate the release/activation of mechanoreceptors that had been previously trapped within the caveolae. More recently Moreno-Vicente and collaborators have demonstrated that caveolin-1 can regulate the activity of the YAP (Yes-associated protein) and TAZ (transcriptional coactivator with PDZ-binding motif), two central components of the Hippo signaling pathway, which governs cell proliferation, organ, and mechanosensing [140,141]. The recent identification of caveolin-1 invaginations, distinct from conventional caveolae, and devoid of cavins, referred to as "dolines", has introduced a new layer of complexity to our knowledge of mechanotransduction [142]. The authors propose that these structures, alongside caveolae, function distinctly and complementarily to mediate the cell responses to mechanical stimuli originating from the extracellular environment. In summary, elucidating the precise mechanisms by which caveolins contribute to mechanotransduction is a growing area of research, with implications for understanding cell biology, disease pathogenesis, and potential therapeutic interventions.

# 4.5. Caveolae in the regulation of calcium signal

Calcium ion (Ca<sup>2+</sup>) is perhaps the most prevalent and adaptable intracellular messenger across various cell types [143]. Intracellular (Ca<sup>2+</sup>) signaling plays a pivotal role in a multitude of cellular processes, ranging from muscle contraction and neurotransmission to cell proliferation and gene expression. The precise control of Ca<sup>2+</sup> concentration within various cellular compartments is essential for these diverse functions, and cells have developed an intricate network of regulatory mechanisms, among which caveolae have emerged as a critical component [2]. The first indications of the involvement of caveolae in mobilizing intracellular Ca<sup>2+</sup> come from studies conducted in the mid-1970s by Popescu et al [144]. The authors showed that following the treatment of smooth muscle cells with calcium oxalate, the compound was found to be distributed in various cellular compartments, including invaginated caveolae. Further studies have shown the localization of essential signaling molecules regulating Ca<sup>2+</sup> mobilization, such as the IP3 receptor (IP3R) and Ca<sup>2+</sup>-ATPase, within caveolae both in ECs and smooth muscle cells [145]. In 1998 Isshiki demonstrated that caveolin-rich cell edges, are involved in the initiation of ATP-induced Ca<sup>2+</sup> waves in bovine aortic ECs [146], and two years later the same group utilized a modified calcium sensor to illustrate the central function of caveolae in facilitating thapsigargin-induced Ca<sup>2+</sup> entry [147]. They reported that interfering with these domains using a cholesterol-extracting agent impeded this process. In ECs, caveolin-1 expression is necessary for the relaxation associated with endothelium-derived hyperpolarizing factor (EDHF), as it influences the membrane positioning and functioning of transient receptor potential vanilloid (TRPV) channels and connexins, two crucial players in the EDHF-signaling pathway [148]. More recently, Medvedev et al., have shown that the role of caveolae in regulating Ca<sup>2+</sup> signaling in atrial cardiomyocytes, and that their disruption leads to an increase in cAMP levels, which in turn enhances the phosphorylation of Prtein Kinase A (PKA)mediated Ca<sup>2+</sup> signaling [149]. Specific isoforms of Protein Kinase C (PKC) are activated by calcium, thereby regulating diverse cellular functions, including cell growth, differentiation, and apoptosis. Among the different isoforms of PKC, PKC $\alpha$  has been shown mainly localized within caveolae

[150,151]. In addition, the scaffolding domain of both caveolin-1 and caveolin-3 (but not caveolin-2) binds to PKC, regulating its kinase activity [152,153]. A broad family of proteins, which plays a crucial role in determining how calcium ions are redistributed within cells, is represented by G-proteincoupled receptors (GPCRs) [154]. When ligands bind to GPCRs, they initiate a series of intracellular events that ultimately lead to the production of inositol-1,4,5-trisphosphate (IP3). IP3 then binds to its specific receptor within the endoplasmic (or sarcoplasmic) reticulum, causing a rapid release of Ca<sup>2+</sup> from these compartments. In this context, there is substantial evidence supporting the involvement of caveolae as well as the caveolin-1 CSD in sequestering various GPCR subtypes. Nicotinic acid adenine dinucleotide phosphate (NAADP) is a pivotal signaling molecule, serving as a second messenger [155]. It initiates the release of calcium from lysosomal reservoirs by binding to the calcium channels within the two-pore channel (TPC) family, thereby inducing the efflux of Ca<sup>2+</sup> into the cytoplasm. Our research has illustrated the participation of caveolae and lipid rafts in mediating NAADP-induced calcium release via the endothelin receptor subtype B (ETB) in smooth muscle cells [104]. Therefore, the role of caveolae in calcium signaling is a fascinating area of research that has provided valuable insights into the intricate mechanisms governing cellular communication. Ongoing research into the precise functions of caveolae, their dynamic interactions with calcium signaling pathways, and their potential as drug targets is likely to yield exciting discoveries.

# 5. Caveolin and cardiovascular diseases

#### 4.5. Caveolins in Cardiac Function

The cardiovascular system is a complex network responsible for pumping blood throughout the body, ensuring the delivery of essential nutrients and oxygen to various tissues and organs. In recent years, caveolins, primarily caveolin-3, that are abundant in cardiac myocytes, have emerged as a subject of significant interest in the field of cardiology. These proteins have been implicated in various aspects of cardiac function, including the regulation of contractile signaling pathways, ion channel function, and mechanosensory processes within the heart [156]. Overall, the presence of caveolin-3 in cardiac tissue appears essential to provide adequate cardiac protection [157]. It has been reported that increasing caveolin expression in the heart, potentially through PI3K-related pathways, may offer a protective mechanism against injury caused by ischemia and subsequent reperfusion. Mice lacking caveolin-3, exhibiting a total absence of caveolae, display a cardiac dysfunction pattern that closely resembles what is observed in mice lacking caveolin-1 [158,159]. Bryant and collaborators have showed that the lack of caveolin-3 induces cardiac dysfunction in vivo, as well as myocytes hypertrophy, defective t-tubule structure, and decreased t-tubular L-type Ca<sup>2+</sup> current density in vitro [160]. These findings are consistent with an earlier study by Woodman and collaborators that links the expression of caveolin-3 to the inhibition of the hypertrophic p42/44 MAPK pathway in the heart [158]. Interestingly, although caveolin-3 is the predominant caveolin in cardiomyocytes, caveolin-1 knock-out mice progressively develop a severe cardiomyopathy, show abnormal ventricular wall thickness, hypertrophy, decreased contractility, increased activity of eNOS, and severely shortened lifespan [161-163]. Researchers also generated double knockouts mice for caveolin-1 and caveolin-3 genes [164]. These mice are both viable and fertile and exhibit a broader reduction of caveolae in various cell types, including ECs, adipocytes, smooth muscle cells, skeletal muscle fibers. Interestingly, despite this more extensive depletion, the observed phenotype is like that of mice with single gene knockouts, although more severe. The discovery that the ablation of caveolin protein in the heart is linked to heart dysfunction supports the broader idea of caveolar network playing a crucial role in cardiac protection. Future research on the role of caveolin proteins in heart functions holds significant promise. With coronary artery disease (CAD) and ischemic heart disease continuing to be leading causes of mortality in industrialized countries, understanding the molecular mechanisms that underlie ischemia-reperfusion injury (I/R injury) is of paramount importance. Exploring how caveolin proteins and other molecular factors contribute to cardiac protection in different contexts, like hypertension, hyperlipidemia, diabetes, aging, and heart failure, can provide

valuable insights for the development of targeted therapeutic interventions to mitigate heart-related diseases and complications.

#### 4.5. Caveolin-1 and Hypertension

Hypertension, or high blood pressure, is a medical condition characterized by prolonged elevated blood pressure in arteries, posing a significant risk for cardiovascular events. Nitric oxide (NO), a lipophilic, colorless, and odorless gas with a short half-life in biological fluids, plays a crucial role in various physiological processes within the human body, particularly in the cardiovascular system. One of its key functions is the regulation of blood vessel tone and diameter. Caveolin-1 has been implicated in the pathophysiology of hypertension, mainly through its influence on ECs and vascular smooth muscle cells. In particular, caveolin-1 is known to modulate NO signaling, a key vasodilator, by sequestering eNOS and preventing its activation [165,166]. Elevated caveolin-1 expression in ECs can bind to eNOS and inhibit its activity, resulting in reduced NO production, blood vessel constriction, and increased blood pressure. Conversely, when caveolin-1 expression is low, eNOS operates more efficiently, generating more NO, promoting vasodilation, reducing vascular resistance, and aiding in blood pressure reduction. Consequently, any disruption in this system can contribute to conditions such as hypertension and impaired blood flow, which are known cardiovascular disease risk factors. Wang and colleagues established a hypertension model to elucidate the specific involvement of caveolin-1 in the control of hypertensive vascular remodeling through its influence on the Notch pathway. Their findings suggest that targeting the caveolin-1/Notch1 signaling pathways could offer promising therapeutic avenues for hypertension management [167]. Moreover, a recent study has elucidated the pivotal mechanosensory function of caveolin-1 in the regulation of inflammation induced by pressure in vascular and renal systems [129]. It is worth highlighting how mutations in the proximal portion of the C-terminal domain of caveolin-1 (crucial for the formation of oligomers from monomers), have been observed in patients diagnosed with heritable and idiopathic pulmonary arterial hypertension [25]. In conclusion, caveolin-1 plays a significant role in the pathophysiology of hypertension, shedding light on potential therapeutic targets for managing this complex cardiovascular condition.

# 6. Caveolae and cellular senescence

Researchers first noticed the limited lifespan of primary human cells in the 1960s, which led to the development of the concept of senescence [168]. Cellular senescence is a unique biological occurrence observed in somatic cells, marked by a range of distinct features, including irreversible growth arrest, morphological changes, increased senescence-associated β-Galactosidase (SA-β-gal) activity, DNA damage, telomere shortening, and resistance to apoptosis [169–172]. It also serves as a critical mechanism to ensure proper embryogenesis [173], and has been proposed as a possible therapeutic target in cardiovascular diseases [174]. Recent research has revealed that caveolin-1 regulates multiple pathways involved in senescence, affecting cell signaling, oxidative stress, and the maintenance of cellular structures [175]. Caveolin-1 expression can either promote or inhibit senescence, depending on the cellular context and the signaling pathways involved. Studies have shown that caveolin-1 overexpression has the remarkable ability to inhibit EGF-mediated signaling in senescent fibroblasts. Conversely, downregulation of caveolin-1 expression through siRNA technology effectively restored EGF-signaling and reactivated cell cycle in senescent fibroblasts [176,177]. Apart from fibroblasts, the influence of caveolin-1 expression on replicative senescence has also been noted in other types of cells, such as human mesenchymal stem cells, macrophages, and chondrocyte [38,178-180]. Caveolin-1 has also been associated with stress-induced premature senescence (SIPS). Volonte and colleagues have presented empirical evidence illustrating the direct involvement of caveolin-1 in oxidative stress-induced cellular senescence within human NIH 3T3 cells. This investigation proved that the extent of caveolin-1 expression in NIH 3T3 cells plays a pivotal role in determining whether oxidative stress-mediated senescence or apoptosis is induced [181]. Likewise, it has been reported that smoker patients that were diagnosed coronary atherosclerosis, showed that elevated level of caveolin-1 expression correlated with the expression of

the oxidative stress marker 4-hydroxynonenal, resulting in oxidative stress-induced EC senescence, compared with older nonsmokers [182]. From a molecular point of view, among the major molecular pathways by which caveolin-1 mediates SIPS is the p53/p21 signaling pathway [183]. This is due to the ability of caveolin-1 to regulate, directly or indirectly p53 inhibitory molecules, including Mdm2, Ataxia telangiectasia-mutated (ATM), sirtuin 1, nuclear erythroid 2 p45-related factor-2 (Nrf2), and others [175,184-186]. Of note, cavin-1, a well-known caveolae associated protein, has been also found increased in senescent fibroblasts, suggesting its role as an additional regulator of cellular senescence acting through the p53/p21 and pathways [187]. Nevertheless, it should be noted that the role of caveolin-1 in cellular senescence is a complex and debated topic. While earlier reported studies suggest that the expression of caveolin-1 can induce cellular senescence, others propose that the downregulation of caveolin-1 is responsible for promoting this process [188]. The morphological and functional alterations seen in ECs when caveolin-1 is silenced by siRNA align with the role of caveolin-1 in diminishing the senescence-like phenotype [189]. This conflicting evidence underscores the intricate interplay between caveolin-1 and cellular senescence, highlighting the need for further research to elucidate the precise mechanisms and conditions under which caveolin-1 exerts its opposite effects on this process.

# 7. Caveolin in tumor progression and stromal cell biology

The multifaceted involvement of caveolins in cancer encompasses both pro-tumorigenic and anti-tumorigenic functions, depending on the specific context and type of cancer. On one hand, caveolins can function as tumor suppressors by inhibiting cell proliferation, promoting apoptosis, and regulating the activity of certain signaling pathways. Reasonably, the tumor-suppressive function of caveolin-1 is associated with its ability to engage with numerous proteins situated in the caveolae of the plasma membrane. For example by sequestering growth-inhibitory proteins, such as p53 and transforming growth factor-β (TGF-β), within caveolae [190]. Nevertheless, a recent study has uncovered that the expression of caveolin-1 within the endoplasmic reticulum inhibits the unfolded protein response (UPR) in both in vitro and solid tumors. This repression leads to a reduction in PERK and IRE1 $\alpha$  signaling while enhancing cellular vulnerability to endoplasmic reticulum stress and hypoxia. Notably, this tumor-suppressive role relies on the phosphorylation of serine-80 in caveolin-1 [191]. In a study conducted on different trancer cell lines, it has been established that caveolin-1-dependent tumor suppression, especially in the absence of E-cadherin, is linked to reduced HIF1α transcriptional activity via diminished NOS-mediated HIF1α Snitrosylation [192]. On the other hand, caveolin is known to promote tumor progression by facilitating the activation of pro-oncogenic signaling pathways, such as those mediated by receptor tyrosine kinases [193]. The first evidence highlighting the involvement of caveolin-1 in cell proliferation and tumor cell survival was first observed in Rous Sarcoma Virus (RSV)-transformed cells [194]. In these cells, the tyrosine phosphorylation of caveolin-1 by protein-tyrosine kinase v-SRC resulted in a reduction in both caveolin-1 expression and caveolae [36,195], leading to the transformation process by disrupting normal cell signaling and cellular homeostasis [196]. A similar effect of reduced caveolin-1 has been documented in some tumor types [197-200]. Conversely, increased caveolin-1 expression has been seen in melanoma cell lines, where caveolin-1 Y14 phosphorylation plays a role in promoting tumor progression [201], or in Ewing's sarcoma where its increased expression triggers MMPs-induced metastatic invasion [202,203]. A similar effect of caveolin-1 overexpression on cell proliferation and invasion has been reported in hepatocarcinoma and lung adenocarcinoma [204-208]. This duality of caveolin's function in cancer highlights its complex nature, making it a subject of ongoing research aimed at better understanding its precise contributions to tumor progression and metastasis.

It is worth mentioning that tromal cells have a crucial role in influencing the development and control of cancer. Found within the tumor microenvironment, these non-cancerous cells, including fibroblasts, mesenchymal stem cells, macrophages, lymphocytes, ECs, and pericytes, are not passive observers but actively engage in the cancer process. Stromal cells can produce growth factors, cytokines, and extracellular matrix components that fuel tumor growth and promote the invasion of

cancer cells into surrounding tissues. Within the tumor microenvironment, cancer-associated fibroblasts (CAFs) play a pivotal role in advancing cancer by inhibiting cell apoptosis and promoting the proliferation of cancer cells, induce epithelial-mesenchymal transition, and foster neo angiogenesis [209-214]. Likewise, they can also influence immune responses, either suppressing or enhancing the body's ability to fight cancer [215]. Unlike what is seen in cancer cells, where caveolin-1 exhibits varying effects across different transformed tissues, its expression in the stroma cells in tumor microenvironment seems to follow a more consistent pattern in relation to tumor progression. For instance, the reduced expression of caveolin-1 in CAFs is commonly associated with adverse clinical outcomes, including tumor recurrence, metastasis formation, and resistance to chemotherapy [209,210,216–220]. Sotgia and collaborators have reported that stromal fibroblasts collected from mice lacking caveolin-1 display many resemblances to human CAFs [221]. Overall, multiple research studies concur with the hypothesis that the reduction of caveolin-1 in the stromal microenvironment of breast cancer signifies an adverse clinical prognosis [220,222,223]. A diminished level of caveolin-1 expression has also been shown in stromal cells across several types of cancers, such as prostate, gastric, and melanoma [216,218,224,225]. The expression of caveolin proteins in tumor stromal cells exerts profound effects on cancer development and therapeutic responses, making them attractive targets for future research and potentially novel anti-cancer therapies.

# 7. Caveolin and neurodegenerative disorders

Neurodegenerative disorders (NDDs) represent a class of debilitating diseases characterized by the progressive degeneration of neurons in the nervous system. These disorders, which include Alzheimer's disease (AD), Parkinson's disease (PD), and amyotrophic lateral sclerosis (ALS), have far-reaching implications for the affected individuals and their families, making it imperative to decipher the intricate mechanisms underlying their pathogenesis. Emerging research has suggested that caveolins play a pivotal role in the pathogenesis of NDDs by influencing various cellular processes, including protein aggregation, oxidative stress, and synaptic dysfunction. Studies conducted on rats have shown that caveolin-1 is extensively distributed across various brain compartments [226], and all three caveolin proteins are expressed in brain ECs and astrocytes. This observation marks the first instance of caveolin-3 expression occurring beyond the confines of the muscular system [21]. Wu et al. has shown that caveolae-like domains are prominently present in neuronal plasma membranes and have a high concentration of signaling molecules, including tyrosine kinases [227]. Consistent with these findings, a growing body of evidence has been gathered to support the regulatory role of caveolins in intracellular signaling within neurons [228]. Biochemical analysis has uncovered the presence of both caveolin-1 and caveolin-2 in dorsal root ganglion and PC12 cells, with their levels increasing following treatment with nerve growth factor (NGF) [229]. In more recent studies, Wang and colleagues found that an increased expression of caveolin-1 in human neuronal progenitor cells promotes the growth of axons. Conversely, when caveolin-1 phosphorylation is inhibited, it negatively affects both the differentiation of neuronal progenitor cells and the growth of axons. An effect due to the blunting of caveolin-1 mediated Rac1/Cdc42 signaling, a crucial signal regulating filopodia and neurite outgrowth [114]. In a different study, Koh et al., have shown that caveolin-1 is found within lipid rafts situated at nerve terminals in primary cultured hippocampal neurons, and plays a vital role in presynaptic processes. The authors proved that caveolin-1 silencing using siRNA significantly impairs synaptic transmission as well as significantly decrease the rate of both synaptic vesicle exocytosis and endocytosis [230]. Caveolin-1 plays a crucial neuroprotective role in the brain, as shown by studies conducted on caveolin-1 knockout mice. A study by Jasmin and collaborators [231] revealed that the levels of caveolin-1 and 2 increased during cerebral ischemia. Conversely, when caveolin-1 was genetically removed, it was associated with larger areas of brain tissue damage, impaired angiogenesis, and increased cell death in comparison to wild-type mice. These findings support the idea that caveolin-1 serves a neuroprotective function. Other studies have further highlighted the importance of this protein in maintaining and regulating the permeability of the blood-brain barrier [232].

Cholesterol metabolism in the brain is crucial for supporting normal brain function, and imbalances in this process can have significant implications for neurological health, including the development of neurodegenerative diseases like Alzheimer's. Due to their role in maintaining cellular cholesterol balance and overall lipid homeostasis, caveolins and caveolae are expected to play a significant role in cholesterol metabolism in the nervous system. Scientific and epidemiological evidence suggest that the interaction between cholesterol levels and the processing of the amyloid precursor protein (APP) plays a significant role in AD. It is widely recognized that only about 25% of the cholesterol in the bloodstream can penetrate the blood-brain barrier [233]. As a result, the brain is self-sustaining and capable of producing the cholesterol it needs, thanks to a finely controlled cholesterol metabolism and supporting glial cells, primarily astrocytes. One of the most relevant mechanisms that is believed to contribute to the development of AD is the accumulation of the betaamyloid protein in the brain, resulting in the formation of plaques that disrupt communication between nerve cells and trigger inflammation. Notably, beta-amyloid protein has been found localized in a detergent-insoluble glycolipid-enriched membrane domain where it is supposed to be processed from the cleavage of the larger amyloid precursor protein (APP) [234,235]. Moreover, caveolin-1 is recognized for its physical interaction with APP through its C-terminal tail, and when caveolin-1 is overexpressed, it reduces the proteolysis of APP that is mediated by  $\gamma$ -secretase [234,236]. Beyond the role of caveolin-1, some studies have shown that caveolin-2 may play roles in neurodegenerative diseases, particularly AD. It's worth noting that the expression of caveolin-2 decreases when activated by the Rac1b signaling pathway, resulting in the formation of neurofibrillary tangles (NFTs) in neurons [237]. NFTs are considered a neuropathological hallmark of AD characterized by a dysfunctional cytoskeleton, along with the loss of microtubules and tubulinassociated proteins [238]. Like caveolin-1, caveolin-3 has been discovered to be physically linked with APP, and it showed a notable increase in reactive astrocytes around senile plaques in tissue derived from patients with AD [239]. These findings supply support for the involvement of caveolin-1 and 3 in the regulation of amyloid production and open new avenues to decipher the amyloid biogenesis in neurodegenerative disorders.

There is a growing body of evidence that shows a strong link between increased levels of caveolin-1 and the key molecular aspects of Parkinson's disease (PD). Caveolin-1 plays a pleiotropic role in PD, showing both positive and negative impacts on its development. Analysis of human neuroblastoma cells has shown that overexpressing  $\alpha$ -Synuclein, a presynaptic neuronal protein, disrupts the ERK signaling through an elevation in caveolin-1 levels resulting in reduced neurite outgrowth and cell adhesion ultimately contributing to the development of PD [240]. Moreover, the increase of  $\alpha$ -Synuclein inhibits cell survival by upregulating caveolin-1 level [241]. However, neither of these studies was able to pinpoint the exact molecular mechanisms through which  $\alpha$ -Synuclein regulates the expression of caveolin-1. A similar mechanism leading to caveolin-1 upregulation is seen in reduced expression of parkin, belonging to a group of genes whose mutations have been clearly linked to PD [242]. This leads to the inhibition of caveolin-1 degradation by proteasomedependent pathway, resulting in imbalanced cholesterol level, alteration of membrane fluidity and dysfunctional lipid raft-dependent endocytosis that finally contribute to PD [243]. Conversely, research by Cataldi et al. discovered that 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) (a common lipophilic substance used to induce PD) reduces caveolin-1 expression, therefore contributing to PD development [244]. In addition, deficiency of DJ-1 (also known as Protein deglycase or PARK 7), a gene associated with familial PD, induces downregulation of caveolin-1 expression through its ability to regulate flotillin-1, leading to dysfunctional cholesterol metabolism, lipid rafts-mediated endocytosis, and impairment of glutamate uptake into astrocytes [245]. Of note, a comparable situation has also been seen by the same authors in DJ-1 knockout mouse embryonic fibroblasts, showing that the mechanism through which DJ-1 regulates the expression of caveolin-1 and flotillin-1 is not exclusively limited to astrocytes. Furthermore, a diminished presence of caveolin proteins has been documented as diminishing the protective benefits of natural tocotrienols, a member of the vitamin E family and a natural compound, which has shown its potential in averting neurodegeneration and lowering the risk of Parkinson's disease [246,247]. The growing body of

evidence linking caveolin proteins to fundamental aspects of nervous system function makes them promising candidates for future research in the pathophysiology of neurodegenerative disorders. This research has the potential to substantially illuminate the underlying mechanisms and, ultimately, contribute to the development of innovative therapeutic approaches.

# 6. Conclusions

Targeting caveolins and caveolae holds great promise in the field of cell signaling and disease management. Their pivotal roles in various diseases, from cardiovascular conditions to cancer and neurodegenerative disorders, have opened new avenues for innovative therapeutic approaches. Additionally, the development of drugs targeting caveolae, caveolin proteins, and the myriad of signaling molecules they regulate, including src kinase, cavins, lipid metabolism enzymes, eNOS, and ion channels, just to mention a few, may offer potential solutions for numerous human diseases. On the other hand, the complex and multifaceted nature of the caveolar network poses challenges, from their intricate biogenesis to context-dependent effects on disease progression. Therefore, future research should focus on uncovering the precise mechanisms behind these structures and harnessing their potential as therapeutic targets, thereby advancing medical science.

**Author Contributions:** writing, review and editing, A.D.

**Funding:** This work was supported by the Catholic University of the Sacred Heart within the Framework of its Programs for the Promotion and Dissemination of Scientific Research, Linea D.1 2019 (R4124500822) to A.D.

Conflicts of Interest: Author declares no conflicts of interest.

#### References

- 1. Simons, K.; Ikonen, E. Functional rafts in cell membranes. *Nature* **1997**, *387*, 569-572.
- 2. Parton, R.G.; Simons, K. The multiple faces of caveolae. Nat Rev Mol Cell Biol 2007, 8, 185-194.
- 3. Sezgin, E.; Levental, I.; Mayor, S.; Eggeling, C. The mystery of membrane organization: composition, regulation and roles of lipid rafts. *Nat Rev Mol Cell Biol.* **2017**, *18*, 361-376.
- 4. Martinez-Outschoorn, U.E.; Sotgia, F.; Lisanti, M.P. Caveolae and signalling in cancer. *Nat Rev Cancer* **2015**, 15, 225-237, doi:10.1038/nrc3915.
- 5. Lingwood, D.; Simons, K. Lipid rafts as a membrane-organizing principle. *Science* **2010**, 327, 46-50, doi:10.1126/science.1174621.
- Simons, K.; Toomre, D. Lipid rafts and signal transduction. Nat Rev Mol Cell Biol 2000, 1, 31-39, doi:10.1038/35036052.
- 7. Parton, R.G.; del Pozo, M.A. Caveolae as plasma membrane sensors, protectors and organizers. *Nat Rev Mol Cell Biol* **2013**, *14*, 98-112, doi:10.1038/nrm3512.
- 8. Ariotti, N.; Parton, R.G. SnapShot: caveolae, caveolins, and cavins. *Cell* **2013**, *154*, 704-704 e701, doi:10.1016/j.cell.2013.07.009.
- 9. Hansen, C.G.; Nichols, B.J. Exploring the caves: cavins, caveolins and caveolae. *Trends Cell Biol* **2010**, 20, 177-186, doi:10.1016/j.tcb.2010.01.005.
- 10. Palade, G.E. Fine structure of blood capillaries. J Appl Phys. 1953, 24.
- 11. Yamada, E. The fine structure of the gall bladder epithelium of the mouse. *J Biophys Biochem Cytol* **1955**, *1*, 445-458.
- 12. Dupree, P.; Parton, R.G.; Raposo, G.; Kurzchalia, T.V.; Simons, K. Caveolae and sorting in the trans-Golgi network of epithelial cells. *Embo J* **1993**, *12*, 1597-1605.
- 13. Rothberg, K.G.; Heuser, J.E.; Donzell, W.C.; Ying, Y.S.; Glenney, J.R.; Anderson, R.G. Caveolin, a protein component of caveolae membrane coats. *Cell* **1992**, *68*, *673*-682.
- 14. Kurzchalia, T.V.; Dupree, P.; Parton, R.G.; Kellner, R.; Virta, H.; Lehnert, M.; Simons, K. VIP21, a 21-kD membrane protein is an integral component of trans-Golgi-network-derived transport vesicles. *J Cell Biol* **1992**, *118*, 1003-1014.
- 15. Glenney, J.R., Jr. The sequence of human caveolin reveals identity with VIP21, a component of transport vesicles. *FEBS Lett* **1992**, *314*, 45-48.
- 16. Thorn, H.; Stenkula, K.G.; Karlsson, M.; Ortegren, U.; Nystrom, F.H.; Gustavsson, J.; Stralfors, P. Cell surface orifices of caveolae and localization of caveolin to the necks of caveolae in adipocytes. *Mol Biol Cell* **2003**, *14*, 3967-3976, doi:10.1091/mbc.e03-01-0050.
- 17. Fra, A.M.; Williamson, E.; Simons, K.; Parton, R.G. De novo formation of caveolae in lymphocytes by expression of VIP21-caveolin. *Proc Natl Acad Sci U S A* **1995**, 92, 8655-8659, doi:10.1073/pnas.92.19.8655.

- 18. Mirre, C.; Monlauzeur, L.; Garcia, M.; Delgrossi, M.H.; Le Bivic, A. Detergent-resistant membrane microdomains from Caco-2 cells do not contain caveolin. *Am J Physiol* **1996**, 271, C887-894, doi:10.1152/ajpcell.1996.271.3.C887.
- 19. Niesman, I.R.; Zemke, N.; Fridolfsson, H.N.; Haushalter, K.J.; Levy, K.; Grove, A.; Schnoor, R.; Finley, J.C.; Patel, P.M.; Roth, D.M., et al. Caveolin isoform switching as a molecular, structural, and metabolic regulator of microglia. *Mol Cell Neurosci* **2013**, *56*, 283-297, doi:10.1016/j.mcn.2013.07.002.
- 20. Cameron, P.L.; Liu, C.; Smart, D.K.; Hantus, S.T.; Fick, J.R.; Cameron, R.S. Caveolin-1 expression is maintained in rat and human astroglioma cell lines. *Glia* **2002**, *37*, 275-290, doi:10.1002/glia.10036.
- 21. Ikezu, T.; Ueda, H.; Trapp, B.D.; Nishiyama, K.; Sha, J.F.; Volonte, D.; Galbiati, F.; Byrd, A.L.; Bassell, G.; Serizawa, H., et al. Affinity-purification and characterization of caveolins from the brain: differential expression of caveolin-1, -2, and -3 in brain endothelial and astroglial cell types. *Brain Res* 1998, 804, 177-192, doi:10.1016/s0006-8993(98)00498-3.
- 22. Lang, D.M.; Lommel, S.; Jung, M.; Ankerhold, R.; Petrausch, B.; Laessing, U.; Wiechers, M.F.; Plattner, H.; Stuermer, C.A. Identification of reggie-1 and reggie-2 as plasmamembrane-associated proteins which cocluster with activated GPI-anchored cell adhesion molecules in non-caveolar micropatches in neurons. *J Neurobiol* 1998, 37, 502-523, doi:10.1002/(sici)1097-4695(199812)37:4<502::aid-neu2>3.0.co;2-s.
- 23. Shyng, S.L.; Heuser, J.E.; Harris, D.A. A glycolipid-anchored prion protein is endocytosed via clathrin-coated pits. *J Cell Biol* **1994**, *125*, 1239-1250, doi:10.1083/jcb.125.6.1239.
- Glenney, J.R., Jr.; Soppet, D. Sequence and expression of caveolin, a protein component of caveolae plasma membrane domains phosphorylated on tyrosine in Rous sarcoma virus-transformed fibroblasts. *Proc Natl Acad Sci U S A* 1992, 89, 10517-10521.
- 25. Austin, E.D.; Ma, L.; LeDuc, C.; Berman Rosenzweig, E.; Borczuk, A.; Phillips, J.A., 3rd; Palomero, T.; Sumazin, P.; Kim, H.R.; Talati, M.H., et al. Whole exome sequencing to identify a novel gene (caveolin-1) associated with human pulmonary arterial hypertension. *Circ Cardiovasc Genet* **2012**, *5*, 336-343, doi:10.1161/CIRCGENETICS.111.961888.
- 26. Allen, K.N.; Entova, S.; Ray, L.C.; Imperiali, B. Monotopic Membrane Proteins Join the Fold. *Trends Biochem Sci* **2019**, 44, 7-20, doi:10.1016/j.tibs.2018.09.013.
- 27. Porta, J.C.; Han, B.; Gulsevin, A.; Chung, J.M.; Peskova, Y.; Connolly, S.; McHaourab, H.S.; Meiler, J.; Karakas, E.; Kenworthy, A.K., et al. Molecular architecture of the human caveolin-1 complex. *Sci Adv* **2022**, *8*, eabn7232, doi:10.1126/sciadv.abn7232.
- 28. Li, S.; Okamoto, T.; Chun, M.; Sargiacomo, M.; Casanova, J.E.; Hansen, S.H.; Nishimoto, I.; Lisanti, M.P. Evidence for a regulated interaction between heterotrimeric G proteins and caveolin. *J Biol Chem* **1995**, 270, 15693-15701, doi:10.1074/jbc.270.26.15693.
- 29. Scherer, P.E.; Okamoto, T.; Chun, M.; Nishimoto, I.; Lodish, H.F.; Lisanti, M.P. Identification, sequence, and expression of caveolin-2 defines a caveolin gene family. *Proc Natl Acad Sci U S A* **1996**, 93, 131-135.
- 30. Tang, Z.; Scherer, P.E.; Okamoto, T.; Song, K.; Chu, C.; Kohtz, D.S.; Nishimoto, I.; Lodish, H.F.; Lisanti, M.P. Molecular cloning of caveolin-3, a novel member of the caveolin gene family expressed predominantly in muscle. *J Biol Chem* **1996**, 271, 2255-2261.
- 31. Machleidt, T.; Li, W.P.; Liu, P.; Anderson, R.G. Multiple domains in caveolin-1 control its intracellular traffic. *J Cell Biol* **2000**, 148, 17-28, doi:10.1083/jcb.148.1.17.
- 32. Fujimoto, T.; Kogo, H.; Nomura, R.; Une, T. Isoforms of caveolin-1 and caveolar structure. *J Cell Sci* **2000**, 113 *Pt* 19, 3509-3517.
- 33. Dietzen, D.J.; Hastings, W.R.; Lublin, D.M. Caveolin is palmitoylated on multiple cysteine residues. Palmitoylation is not necessary for localization of caveolin to caveolae. *J Biol Chem* **1995**, 270, 6838-6842.
- 34. Parat, M.O.; Fox, P.L. Palmitoylation of caveolin-1 in endothelial cells is post-translational but irreversible. *J Biol Chem* **2001**, 276, 15776-15782.
- 35. Li, S.; Seitz, R.; Lisanti, M.P. Phosphorylation of caveolin by src tyrosine kinases. The alpha-isoform of caveolin is selectively phosphorylated by v-Src in vivo. *J Biol Chem* **1996**, *271*, 3863-3868.
- 36. Aoki, T.; Nomura, R.; Fujimoto, T. Tyrosine phosphorylation of caveolin-1 in the endothelium. *Exp Cell Res* **1999**, 253, 629-636, doi:10.1006/excr.1999.4652.
- 37. Nah, J.; Yoo, S.M.; Jung, S.; Jeong, E.I.; Park, M.; Kaang, B.K.; Jung, Y.K. Phosphorylated CAV1 activates autophagy through an interaction with BECN1 under oxidative stress. *Cell Death Dis* **2017**, *8*, e2822, doi:10.1038/cddis.2017.71.
- 38. Goutas, A.; Outskouni, Z.; Papathanasiou, I.; Satra, M.; Koliakos, G.; Trachana, V. Dysregulation of Caveolin-1 Phosphorylation and Nuclear Translocation Is Associated with Senescence Onset. *Cells* **2021**, *10*, doi:10.3390/cells10112939.
- 39. Schlegel, A.; Arvan, P.; Lisanti, M.P. Caveolin-1 binding to endoplasmic reticulum membranes and entry into the regulated secretory pathway are regulated by serine phosphorylation. Protein sorting at the level of the endoplasmic reticulum. *J Biol Chem* **2001**, *276*, 4398-4408, doi:10.1074/jbc.M005448200.

- 40. Hayer, A.; Stoeber, M.; Ritz, D.; Engel, S.; Meyer, H.H.; Helenius, A. Caveolin-1 is ubiquitinated and targeted to intralumenal vesicles in endolysosomes for degradation. *J Cell Biol* **2010**, *191*, 615-629, doi:10.1083/jcb.201003086.
- 41. Lee, C.Y.; Lai, T.Y.; Tsai, M.K.; Chang, Y.C.; Ho, Y.H.; Yu, I.S.; Yeh, T.W.; Chou, C.C.; Lin, Y.S.; Lawrence, T., et al. The ubiquitin ligase ZNRF1 promotes caveolin-1 ubiquitination and degradation to modulate inflammation. *Nat Commun* **2017**, *8*, 15502, doi:10.1038/ncomms15502.
- 42. Scherer, P.E.; Lewis, R.Y.; Volonte, D.; Engelman, J.A.; Galbiati, F.; Couet, J.; Kohtz, D.S.; van Donselaar, E.; Peters, P.; Lisanti, M.P. Cell-type and tissue-specific expression of caveolin-2. Caveolins 1 and 2 co-localize and form a stable hetero-oligomeric complex in vivo. *J Biol Chem* **1997**, 272, 29337-29346.
- 43. Scherer, P.E.; Tang, Z.; Chun, M.; Sargiacomo, M.; Lodish, H.F.; Lisanti, M.P. Caveolin isoforms differ in their N-terminal protein sequence and subcellular distribution. Identification and epitope mapping of an isoform-specific monoclonal antibody probe. *J Biol Chem* **1995**, 270, 16395-16401, doi:10.1074/jbc.270.27.16395.
- 44. Kogo, H.; Ishiguro, K.; Kuwaki, S.; Fujimoto, T. Identification of a splice variant of mouse caveolin-2 mRNA encoding an isoform lacking the C-terminal domain. *Arch Biochem Biophys* **2002**, *401*, 108-114, doi:10.1016/S0003-9861(02)00009-7.
- 45. Kwon, H.; Lee, J.; Jeong, K.; Jang, D.; Pak, Y. Fatty acylated caveolin-2 is a substrate of insulin receptor tyrosine kinase for insulin receptor substrate-1-directed signaling activation. *Biochim Biophys Acta* **2015**, 1853, 1022-1034, doi:10.1016/j.bbamcr.2015.02.002.
- 46. Sowa, G.; Pypaert, M.; Fulton, D.; Sessa, W.C. The phosphorylation of caveolin-2 on serines 23 and 36 modulates caveolin-1-dependent caveolae formation. *Proc Natl Acad Sci U S A* **2003**, *100*, 6511-6516.
- 47. Wang, X.B.; Lee, H.; Capozza, F.; Marmon, S.; Sotgia, F.; Brooks, J.W.; Campos-Gonzalez, R.; Lisanti, M.P. Tyrosine phosphorylation of caveolin-2 at residue 27: differences in the spatial and temporal behavior of phospho-Cav-2 (pY19 and pY27). *Biochemistry* **2004**, *43*, 13694-13706, doi:10.1021/bi049295+.
- 48. Lee, H.; Park, D.S.; Wang, X.B.; Scherer, P.E.; Schwartz, P.E.; Lisanti, M.P. Src-induced phosphorylation of caveolin-2 on tyrosine 19. Phospho-caveolin-2 (Tyr(P)19) is localized near focal adhesions, remains associated with lipid rafts/caveolae, but no longer forms a high molecular mass hetero-oligomer with caveolin-1. *J Biol Chem* **2002**, *277*, 34556-34567, doi:10.1074/jbc.M204367200.
- 49. Choi, M.; Kwon, H.; Pak, Y. Caveolin-2 in association with nuclear lamina controls adipocyte hypertrophy. *FASEB J* **2023**, 37, e22745, doi:10.1096/fj.202201028RR.
- 50. Song, K.S.; Scherer, P.E.; Tang, Z.; Okamoto, T.; Li, S.; Chafel, M.; Chu, C.; Kohtz, D.S.; Lisanti, M.P. Expression of caveolin-3 in skeletal, cardiac, and smooth muscle cells. Caveolin-3 is a component of the sarcolemma and co-fractionates with dystrophin and dystrophin-associated glycoproteins. *J Biol Chem* **1996**, 271, 15160-15165.
- 51. Root, K.T.; Plucinsky, S.M.; Glover, K.J. Recent progress in the topology, structure, and oligomerization of caveolin: a building block of caveolae. *Curr Top Membr* **2015**, *75*, 305-336, doi:10.1016/bs.ctm.2015.03.007.
- 52. Kim, J.H.; Peng, D.; Schlebach, J.P.; Hadziselimovic, A.; Sanders, C.R. Modest effects of lipid modifications on the structure of caveolin-3. *Biochemistry* **2014**, *53*, 4320-4322, doi:10.1021/bi5005238.
- 53. Fuhs, S.R.; Insel, P.A. Caveolin-3 undergoes SUMOylation by the SUMO E3 ligase PIASy: sumoylation affects G-protein-coupled receptor desensitization. *J Biol Chem* **2011**, 286, 14830-14841, doi:10.1074/jbc.M110.214270.
- 54. Gazzerro, E.; Sotgia, F.; Bruno, C.; Lisanti, M.P.; Minetti, C. Caveolinopathies: from the biology of caveolin-3 to human diseases. *Eur J Hum Genet* **2010**, *18*, 137-145.
- 55. Galbiati, F.; Volonte, D.; Minetti, C.; Bregman, D.B.; Lisanti, M.P. Limb-girdle muscular dystrophy (LGMD-1C) mutants of caveolin-3 undergo ubiquitination and proteasomal degradation. Treatment with proteasomal inhibitors blocks the dominant negative effect of LGMD-1C mutanta and rescues wild-type caveolin-3. *J Biol Chem* **2000**, *275*, *37702-37711*, doi:10.1074/jbc.M006657200.
- 56. Parton, R.G.; McMahon, K.A.; Wu, Y. Caveolae: Formation, dynamics, and function. *Curr Opin Cell Biol* **2020**, *65*, 8-16, doi:10.1016/j.ceb.2020.02.001.
- 57. Busija, A.R.; Patel, H.H.; Insel, P.A. Caveolins and cavins in the trafficking, maturation, and degradation of caveolae: implications for cell physiology. *Am J Physiol Cell Physiol* **2017**, 312, C459-C477.
- 58. Nassar, Z.D.; Parat, M.O. Cavin Family: New Players in the Biology of Caveolae. *Int Rev Cell Mol Biol* **2015**, 320, 235-305, doi: https://doi.org/10.1016/j.biochi.2010.03.022.
- 59. Hill, M.M.; Bastiani, M.; Luetterforst, R.; Kirkham, M.; Kirkham, A.; Nixon, S.J.; Walser, P.; Abankwa, D.; Oorschot, V.M.; Martin, S., et al. PTRF-Cavin, a conserved cytoplasmic protein required for caveola formation and function. *Cell* **2008**, *132*, 113-124.
- 60. Bastiani, M.; Liu, L.; Hill, M.M.; Jedrychowski, M.P.; Nixon, S.J.; Lo, H.P.; Abankwa, D.; Luetterforst, R.; Fernandez-Rojo, M.; Breen, M.R., et al. MURC/Cavin-4 and cavin family members form tissue-specific caveolar complexes. *J Cell Biol* **2009**, *185*, 1259-1273, doi:10.1083/jcb.200903053.
- 61. Hansen, C.G.; Bright, N.A.; Howard, G.; Nichols, B.J. SDPR induces membrane curvature and functions in the formation of caveolae. *Nat Cell Biol* **2009**, *11*, 807-814, doi:10.1038/ncb1887.

- 62. McMahon, K.A.; Zajicek, H.; Li, W.P.; Peyton, M.J.; Minna, J.D.; Hernandez, V.J.; Luby-Phelps, K.; Anderson, R.G. SRBC/cavin-3 is a caveolin adapter protein that regulates caveolae function. *EMBO J* **2009**, 28, 1001-1015, doi:10.1038/emboj.2009.46.
- 63. Briand, N.; Dugail, I.; Le Lay, S. Cavin proteins: New players in the caveolae field. *Biochimie* **2011**, 93, 71-77, doi:10.1016/j.biochi.2010.03.022.
- 64. Kovtun, O.; Tillu, V.A.; Ariotti, N.; Parton, R.G.; Collins, B.M. Cavin family proteins and the assembly of caveolae. *J Cell Sci* **2015**, *128*, 1269-1278.
- 65. Doherty, G.J.; McMahon, H.T. Mechanisms of endocytosis. *Annu Rev Biochem* **2009**, *78*, 857-902, doi:10.1146/annurev.biochem.78.081307.110540.
- 66. Serra, N.D.; Sundaram, M.V. Transcytosis in the development and morphogenesis of epithelial tissues. *EMBO J* **2021**, 40, e106163, doi:10.15252/embj.2020106163.
- 67. Garcia-Castillo, M.D.; Chinnapen, D.J.; Lencer, W.I. Membrane Transport across Polarized Epithelia. *Cold Spring Harb Perspect Biol* **2017**, 9, doi:10.1101/cshperspect.a027912.
- 68. Thomsen, P.; Roepstorff, K.; Stahlhut, M.; van Deurs, B. Caveolae are highly immobile plasma membrane microdomains, which are not involved in constitutive endocytic trafficking. *Mol Biol Cell* **2002**, *13*, 238-250, doi:10.1091/mbc.01-06-0317.
- 69. van Deurs, B.; Roepstorff, K.; Hommelgaard, A.M.; Sandvig, K. Caveolae: anchored, multifunctional platforms in the lipid ocean. *Trends Cell Biol* **2003**, *13*, 92-100, doi:10.1016/s0962-8924(02)00039-9.
- 70. Kaksonen, M.; Roux, A. Mechanisms of clathrin-mediated endocytosis. *Nat Rev Mol Cell Biol* **2018**, *19*, 313-326, doi:10.1038/nrm.2017.132.
- 71. Kiss, A.L.; Botos, E.; Turi, A.; Mullner, N. Ocadaic acid treatment causes tyrosine phosphorylation of caveolin-2 and induces internalization of caveolae in rat peritoneal macrophages. *Micron* **2004**, *35*, 707-715, doi:10.1016/j.micron.2004.04.003.
- 72. Parton, R.G.; Joggerst, B.; Simons, K. Regulated internalization of caveolae. J Cell Biol 1994, 127, 1199-1215.
- 73. Razani, B.; Engelman, J.A.; Wang, X.B.; Schubert, W.; Zhang, X.L.; Marks, C.B.; Macaluso, F.; Russell, R.G.; Li, M.; Pestell, R.G., et al. Caveolin-1 null mice are viable but show evidence of hyperproliferative and vascular abnormalities. *J Biol Chem* **2001**, *276*, 38121-38138.
- 74. Zhang, X.; Sessa, W.C.; Fernandez-Hernando, C. Endothelial Transcytosis of Lipoproteins in Atherosclerosis. *Front Cardiovasc Med* **2018**, *5*, 130, doi:10.3389/fcvm.2018.00130.
- 75. Moriyama, T.; Sasaki, K.; Karasawa, K.; Uchida, K.; Nitta, K. Intracellular transcytosis of albumin in glomerular endothelial cells after endocytosis through caveolae. *Journal of cellular physiology* **2017**, 232, 3565-3573, doi:10.1002/jcp.25817.
- 76. Pavlides, S.; Gutierrez-Pajares, J.L.; Iturrieta, J.; Lisanti, M.P.; Frank, P.G. Endothelial caveolin-1 plays a major role in the development of atherosclerosis. *Cell Tissue Res* **2014**, *356*, 147-157, doi:10.1007/s00441-013-1767-7.
- 77. Ge, S.; Song, L.; Serwanski, D.R.; Kuziel, W.A.; Pachter, J.S. Transcellular transport of CCL2 across brain microvascular endothelial cells. *J Neurochem* **2008**, *104*, 1219-1232, doi:10.1111/j.1471-4159.2007.05056.x.
- 78. Ghitescu, L.; Fixman, A.; Simionescu, M.; Simionescu, N. Specific binding sites for albumin restricted to plasmalemmal vesicles of continuous capillary endothelium: receptor-mediated transcytosis. *J Cell Biol* **1986**, *102*, 1304-1311, doi:10.1083/jcb.102.4.1304.
- 79. Pietiainen, V.; Marjomaki, V.; Upla, P.; Pelkmans, L.; Helenius, A.; Hyypia, T. Echovirus 1 endocytosis into caveosomes requires lipid rafts, dynamin II, and signaling events. *Mol Biol Cell* **2004**, *15*, 4911-4925, doi:10.1091/mbc.e04-01-0070.
- 80. Pelkmans, L.; Puntener, D.; Helenius, A. Local actin polymerization and dynamin recruitment in SV40-induced internalization of caveolae. *Science* **2002**, *296*, 535-539.
- 81. Richterova, Z.; Liebl, D.; Horak, M.; Palkova, Z.; Stokrova, J.; Hozak, P.; Korb, J.; Forstova, J. Caveolae are involved in the trafficking of mouse polyomavirus virions and artificial VP1 pseudocapsids toward cell nuclei. *J Virol* **2001**, *75*, 10880-10891, doi:10.1128/JVI.75.22.10880-10891.2001.
- 82. Puri, V.; Watanabe, R.; Singh, R.D.; Dominguez, M.; Brown, J.C.; Wheatley, C.L.; Marks, D.L.; Pagano, R.E. Clathrin-dependent and -independent internalization of plasma membrane sphingolipids initiates two Golgi targeting pathways. *J Cell Biol* **2001**, *154*, 535-547, doi:10.1083/jcb.200102084.
- 83. Pelkmans, L.; Kartenbeck, J.; Helenius, A. Caveolar endocytosis of simian virus 40 reveals a new two-step vesicular-transport pathway to the ER. *Nat Cell Biol* **2001**, *3*, 473-483.
- 84. Campbell, S.M.; Crowe, S.M.; Mak, J. Lipid rafts and HIV-1: from viral entry to assembly of progeny virions. *J Clin Virol* **2001**, 22, 217-227, doi:10.1016/s1386-6532(01)00193-7.
- 85. Shin, J.S.; Gao, Z.; Abraham, S.N. Involvement of cellular caveolae in bacterial entry into mast cells. *Science* **2000**, *289*, 785-788, doi:10.1126/science.289.5480.785.
- 86. Werling, D.; Hope, J.C.; Chaplin, P.; Collins, R.A.; Taylor, G.; Howard, C.J. Involvement of caveolae in the uptake of respiratory syncytial virus antigen by dendritic cells. *J Leukoc Biol* **1999**, *66*, 50-58, doi:10.1002/jlb.66.1.50.

- 87. Benlimame, N.; Le, P.U.; Nabi, I.R. Localization of autocrine motility factor receptor to caveolae and clathrin-independent internalization of its ligand to smooth endoplasmic reticulum. *Mol Biol Cell* **1998**, *9*, 1773-1786, doi:10.1091/mbc.9.7.1773.
- 88. Schnitzer, J.E.; Oh, P.; Pinney, E.; Allard, J. Filipin-sensitive caveolae-mediated transport in endothelium: reduced transcytosis, scavenger endocytosis, and capillary permeability of select macromolecules. *J Cell Biol* 1994, 127, 1217-1232.
- 89. Anderson, R.G.; Kamen, B.A.; Rothberg, K.G.; Lacey, S.W. Potocytosis: sequestration and transport of small molecules by caveolae. *Science* **1992**, *255*, 410-411.
- 90. Rothberg, K.G.; Ying, Y.S.; Kolhouse, J.F.; Kamen, B.A.; Anderson, R.G. The glycophospholipid-linked folate receptor internalizes folate without entering the clathrin-coated pit endocytic pathway. *J Cell Biol* **1990**, *110*, 637-649, doi:10.1083/jcb.110.3.637.
- 91. Hinshaw, J.E.; Schmid, S.L. Dynamin self-assembles into rings suggesting a mechanism for coated vesicle budding. *Nature* **1995**, *374*, 190-192, doi:10.1038/374190a0.
- 92. Henley, J.R.; Krueger, E.W.; Oswald, B.J.; McNiven, M.A. Dynamin-mediated internalization of caveolae. *J Cell Biol* 1998, 141, 85-99, doi:10.1083/jcb.141.1.85.
- 93. Oh, P.; McIntosh, D.P.; Schnitzer, J.E. Dynamin at the neck of caveolae mediates their budding to form transport vesicles by GTP-driven fission from the plasma membrane of endothelium. *J Cell Biol* **1998**, *141*, 101-114, doi:10.1083/jcb.141.1.101.
- 94. Larsson, E.; Moren, B.; McMahon, K.A.; Parton, R.G.; Lundmark, R. Dynamin2 functions as an accessory protein to reduce the rate of caveola internalization. *J Cell Biol* **2023**, 222, doi:10.1083/jcb.202205122.
- 95. Lisanti, M.P.; Scherer, P.E.; Tang, Z.; Sargiacomo, M. Caveolae, caveolin and caveolin-rich membrane domains: a signalling hypothesis. *Trends Cell Biol* **1994**, *4*, 231-235.
- 96. Shatz, M.; Lustig, G.; Reich, R.; Liscovitch, M. Caveolin-1 mutants P132L and Y14F are dominant negative regulators of invasion, migration and aggregation in H1299 lung cancer cells. *Exp Cell Res* **2010**, *316*, 1748-1762, doi:10.1016/j.yexcr.2010.02.006.
- 97. Couet, J.; Sargiacomo, M.; Lisanti, M.P. Interaction of a receptor tyrosine kinase, EGF-R, with caveolins. Caveolin binding negatively regulates tyrosine and serine/threonine kinase activities. *J Biol Chem* **1997**, 272, 30429-30438.
- 98. Pilch, P.F.; Liu, L. Fat caves: caveolae, lipid trafficking and lipid metabolism in adipocytes. *Trends Endocrinol Metab* **2011**, 22, 318-324, doi:10.1016/j.tem.2011.04.001.
- 99. Saltiel, A.R.; Pessin, J.E. Insulin signaling in microdomains of the plasma membrane. *Traffic* **2003**, *4*, 711-716, doi:10.1034/j.1600-0854.2003.00119.x.
- 100. Patel, H.H.; Murray, F.; Insel, P.A. G-protein-coupled receptor-signaling components in membrane raft and caveolae microdomains. *Handb Exp Pharmacol* **2008**, 167-184.
- 101. Boscher, C.; Nabi, I.R. Caveolin-1: role in cell signaling. *Adv Exp Med Biol* **2012**, 729, 29-50, doi:10.1007/978-1-4614-1222-9\_3.
- 102. D'Alessio, A.; Esposito, B.; Giampietri, C.; Ziparo, E.; Pober, J.S.; Filippini, A. Plasma membrane micro domains regulate TACE-dependent TNFR1 shedding in human endothelial cells. *J Cell Mol Med* **2011**.
- 103. D'Alessio, A.; Kluger, M.S.; Li, J.H.; Al-Lamki, R.; Bradley, J.R.; Pober, J.S. Targeting of tumor necrosis factor receptor 1 to low density plasma membrane domains in human endothelial cells. *J Biol Chem* **2010**, 285, 23868-23879.
- 104. Gambara, G.; Billington, R.A.; Debidda, M.; D'Alessio, A.; Palombi, F.; Ziparo, E.; Genazzani, A.A.; Filippini, A. NAADP-induced Ca(2+) signaling in response to endothelin is via the receptor subtype B and requires the integrity of lipid rafts/caveolae. *Journal of cellular physiology* **2008**, *216*, 396-404.
- 105. D'Alessio, A.; Al-Lamki, R.S.; Bradley, J.R.; Pober, J.S. Caveolae Participate in Tumor Necrosis Factor Receptor 1 Signaling and Internalization in a Human Endothelial Cell Line. *Am J Pathol* **2005**, *166*, 1273-1282.
- 106. Labrecque, L.; Royal, I.; Surprenant, D.S.; Patterson, C.; Gingras, D.; Beliveau, R. Regulation of vascular endothelial growth factor receptor-2 activity by caveolin-1 and plasma membrane cholesterol. *Mol Biol Cell* **2003**, *14*, 334-347.
- 107. Feng, X.; Gaeta, M.L.; Madge, L.A.; Yang, J.H.; Bradley, J.R.; Pober, J.S. Caveolin-1 associates with TRAF2 to form a complex that is recruited to tumor necrosis factor receptors. *J Biol Chem* **2001**, *276*, 8341-8349.
- 108. Garcia-Cardena, G.; Fan, R.; Stern, D.F.; Liu, J.; Sessa, W.C. Endothelial nitric oxide synthase is regulated by tyrosine phosphorylation and interacts with caveolin-1. *J Biol Chem* **1996**, 271, 27237-27240.
- 109. Bucci, M.; Gratton, J.P.; Rudic, R.D.; Acevedo, L.; Roviezzo, F.; Cirino, G.; Sessa, W.C. In vivo delivery of the caveolin-1 scaffolding domain inhibits nitric oxide synthesis and reduces inflammation. *Nat Med* **2000**, *6*, 1362-1367.
- 110. Ju, H.; Zou, R.; Venema, V.J.; Venema, R.C. Direct interaction of endothelial nitric-oxide synthase and caveolin-1 inhibits synthase activity. *J Biol Chem* **1997**, 272, 18522-18525, doi:10.1074/jbc.272.30.18522.

- 111. Garcia-Cardena, G.; Martasek, P.; Masters, B.S.; Skidd, P.M.; Couet, J.; Li, S.; Lisanti, M.P.; Sessa, W.C. Dissecting the interaction between nitric oxide synthase (NOS) and caveolin. Functional significance of the nos caveolin binding domain in vivo. *J Biol Chem* **1997**, 272, 25437-25440.
- 112. Sun, Y.; Hu, G.; Zhang, X.; Minshall, R.D. Phosphorylation of caveolin-1 regulates oxidant-induced pulmonary vascular permeability via paracellular and transcellular pathways. *Circ Res* **2009**, *105*, 676-685, 615 p following 685, doi:10.1161/CIRCRESAHA.109.201673.
- 113. Rizzo, V.; Morton, C.; DePaola, N.; Schnitzer, J.E.; Davies, P.F. Recruitment of endothelial caveolae into mechanotransduction pathways by flow conditioning in vitro. *American journal of physiology. Heart and circulatory physiology* **2003**, 285, H1720-1729, doi:10.1152/ajpheart.00344.2002.
- 114. Wang, S.; Zhang, Z.; Almenar-Queralt, A.; Leem, J.; DerMardirossian, C.; Roth, D.M.; Patel, P.M.; Patel, H.H.; Head, B.P. Caveolin-1 Phosphorylation Is Essential for Axonal Growth of Human Neurons Derived From iPSCs. *Front Cell Neurosci* **2019**, *13*, 324, doi:10.3389/fncel.2019.00324.
- 115. Abel, B.; Willoughby, C.; Jang, S.; Cooper, L.; Xie, L.; Vo-Ransdell, C.; Sowa, G. N-terminal tyrosine phosphorylation of caveolin-2 negates anti-proliferative effect of transforming growth factor beta in endothelial cells. *FEBS Lett* **2012**, *586*, 3317-3323, doi:10.1016/j.febslet.2012.07.008.
- 116. Gatfield, J.; Pieters, J. Essential role for cholesterol in entry of mycobacteria into macrophages. *Science* **2000**, *288*, 1647-1650, doi:10.1126/science.288.5471.1647.
- 117. Lauer, S.; VanWye, J.; Harrison, T.; McManus, H.; Samuel, B.U.; Hiller, N.L.; Mohandas, N.; Haldar, K. Vacuolar uptake of host components, and a role for cholesterol and sphingomyelin in malarial infection. *EMBO J* **2000**, *19*, 3556-3564, doi:10.1093/emboj/19.14.3556.
- 118. Mordue, D.G.; Desai, N.; Dustin, M.; Sibley, L.D. Invasion by Toxoplasma gondii establishes a moving junction that selectively excludes host cell plasma membrane proteins on the basis of their membrane anchoring. *J Exp Med* **1999**, 190, 1783-1792, doi:10.1084/jem.190.12.1783.
- 119. Stang, E.; Kartenbeck, J.; Parton, R.G. Major histocompatibility complex class I molecules mediate association of SV40 with caveolae. *Mol Biol Cell* 1997, 8, 47-57, doi:10.1091/mbc.8.1.47.
- 120. Baorto, D.M.; Gao, Z.; Malaviya, R.; Dustin, M.L.; van der Merwe, A.; Lublin, D.M.; Abraham, S.N. Survival of FimH-expressing enterobacteria in macrophages relies on glycolipid traffic. *Nature* **1997**, *389*, 636-639, doi:10.1038/39376.
- 121. Anderson, H.A.; Chen, Y.; Norkin, L.C. Bound simian virus 40 translocates to caveolin-enriched membrane domains, and its entry is inhibited by drugs that selectively disrupt caveolae. *Mol Biol Cell* **1996**, *7*, 1825-1834, doi:10.1091/mbc.7.11.1825.
- 122. Schnitzer, J.E.; Oh, P.; McIntosh, D.P. Role of GTP hydrolysis in fission of caveolae directly from plasma membranes. *Science* **1996**, *274*, 239-242.
- 123. Montesano, R.; Roth, J.; Robert, A.; Orci, L. Non-coated membrane invaginations are involved in binding and internalization of cholera and tetanus toxins. *Nature* **1982**, 296, 651-653.
- 124. Schnitzer, J.E.; Liu, J.; Oh, P. Endothelial caveolae have the molecular transport machinery for vesicle budding, docking, and fusion including VAMP, NSF, SNAP, annexins, and GTPases. *J Biol Chem* **1995**, 270, 14399-14404, doi:10.1074/jbc.270.24.14399.
- 125. Anderson, R.G. The caveolae membrane system. Annu Rev Biochem 1998, 67, 199-225.
- 126. Butcher, E.C. Leukocyte-endothelial cell recognition: three (or more) steps to specificity and diversity. *Cell* **1991**, *67*, 1033-1036, doi:10.1016/0092-8674(91)90279-8.
- 127. Nourshargh, S.; Hordijk, P.L.; Sixt, M. Breaching multiple barriers: leukocyte motility through venular walls and the interstitium. *Nat Rev Mol Cell Biol* **2010**, *11*, 366-378, doi:10.1038/nrm2889.
- 128. Millan, J.; Hewlett, L.; Glyn, M.; Toomre, D.; Clark, P.; Ridley, A.J. Lymphocyte transcellular migration occurs through recruitment of endothelial ICAM-1 to caveola- and F-actin-rich domains. *Nat Cell Biol* **2006**, *8*, 113-123, doi:10.1038/ncb1356.
- 129. Michell, D.L.; Shihata, W.A.; Andrews, K.L.; Abidin, N.A.Z.; Jefferis, A.M.; Sampson, A.K.; Lumsden, N.G.; Huet, O.; Parat, M.O.; Jennings, G.L., et al. High intraluminal pressure promotes vascular inflammation via caveolin-1. *Sci Rep* **2021**, *11*, 5894, doi:10.1038/s41598-021-85476-z.
- 130. Xu, S.; Zhou, X.; Yuan, D.; Xu, Y.; He, P. Caveolin-1 scaffolding domain promotes leukocyte adhesion by reduced basal endothelial nitric oxide-mediated ICAM-1 phosphorylation in rat mesenteric venules. *American journal of physiology. Heart and circulatory physiology* **2013**, 305, H1484-1493, doi:10.1152/ajpheart.00382.2013.
- 131. Park, H.; Shin, J.A.; Lim, J.; Lee, S.; Ahn, J.H.; Kang, J.L.; Choi, Y.H. Increased Caveolin-2 Expression in Brain Endothelial Cells Promotes Age-Related Neuroinflammation. *Mol Cells* **2022**, *45*, 950-962, doi:10.14348/molcells.2022.0045.
- 132. Liu, Y.; Wang, M.; Wang, D.; Fay, W.P.; Korthuis, R.J.; Sowa, G. Elevated postischemic tissue injury and leukocyte-endothelial adhesive interactions in mice with global deficiency in caveolin-2: role of PAI-1. *American journal of physiology. Heart and circulatory physiology* **2021**, 320, H1185-H1198, doi:10.1152/ajpheart.00682.2020.

- 133. Ingber, D.E. Mechanobiology and diseases of mechanotransduction. *Ann Med* **2003**, *35*, 564-577, doi:10.1080/07853890310016333.
- 134. Discher, D.E.; Janmey, P.; Wang, Y.L. Tissue cells feel and respond to the stiffness of their substrate. *Science* **2005**, *310*, 1139-1143, doi:10.1126/science.1116995.
- 135. Wozniak, M.A.; Chen, C.S. Mechanotransduction in development: a growing role for contractility. *Nat Rev Mol Cell Biol* **2009**, *10*, 34-43, doi:10.1038/nrm2592.
- 136. Smart, E.J.; Graf, G.A.; McNiven, M.A.; Sessa, W.C.; Engelman, J.A.; Scherer, P.E.; Okamoto, T.; Lisanti, M.P. Caveolins, liquid-ordered domains, and signal transduction. *Mol Cell Biol* 1999, 19, 7289-7304, doi:10.1128/MCB.19.11.7289.
- 137. Cheng, J.P.; Mendoza-Topaz, C.; Howard, G.; Chadwick, J.; Shvets, E.; Cowburn, A.S.; Dunmore, B.J.; Crosby, A.; Morrell, N.W.; Nichols, B.J. Caveolae protect endothelial cells from membrane rupture during increased cardiac output. *J Cell Biol* **2015**, *211*, 53-61, doi:10.1083/jcb.201504042.
- 138. Drab, M.; Verkade, P.; Elger, M.; Kasper, M.; Lohn, M.; Lauterbach, B.; Menne, J.; Lindschau, C.; Mende, F.; Luft, F.C., et al. Loss of caveolae, vascular dysfunction, and pulmonary defects in caveolin-1 gene-disrupted mice. *Science* **2001**, 293, 2449-2452.
- 139. Sinha, B.; Koster, D.; Ruez, R.; Gonnord, P.; Bastiani, M.; Abankwa, D.; Stan, R.V.; Butler-Browne, G.; Vedie, B.; Johannes, L., et al. Cells respond to mechanical stress by rapid disassembly of caveolae. *Cell* **2010**, 144, 402-413.
- 140. Moreno-Vicente, R.; Pavon, D.M.; Martin-Padura, I.; Catala-Montoro, M.; Diez-Sanchez, A.; Quilez-Alvarez, A.; Lopez, J.A.; Sanchez-Alvarez, M.; Vazquez, J.; Strippoli, R., et al. Caveolin-1 Modulates Mechanotransduction Responses to Substrate Stiffness through Actin-Dependent Control of YAP. *Cell Rep* **2018**, 25, 1622-1635 e1626, doi:10.1016/j.celrep.2018.10.024.
- 141. Dupont, S.; Morsut, L.; Aragona, M.; Enzo, E.; Giulitti, S.; Cordenonsi, M.; Zanconato, F.; Le Digabel, J.; Forcato, M.; Bicciato, S., et al. Role of YAP/TAZ in mechanotransduction. *Nature* **2011**, 474, 179-183.
- 142. Lolo, F.N.; Walani, N.; Seemann, E.; Zalvidea, D.; Pavon, D.M.; Cojoc, G.; Zamai, M.; Viaris de Lesegno, C.; Martinez de Benito, F.; Sanchez-Alvarez, M., et al. Caveolin-1 dolines form a distinct and rapid caveolae-independent mechanoadaptation system. *Nat Cell Biol* **2023**, 25, 120-133, doi:10.1038/s41556-022-01034-3.
- 143. Berridge, M.J.; Lipp, P.; Bootman, M.D. The versatility and universality of calcium signalling. *Nat Rev Mol Cell Biol* **2000**, *1*, 11-21, doi:10.1038/35036035.
- 144. Popescu, L.M.; Diculescu, I.; Zelck, U.; Ionescu, N. Ultrastructural distribution of calcium in smooth muscle cells of guinea-pig taenia coli. A correlated electron microscopic and quantitative study. *Cell Tissue Res* **1974**, 154, 357-378.
- 145. Fujimoto, T.; Nakade, S.; Miyawaki, A.; Mikoshiba, K.; Ogawa, K. Localization of inositol 1,4,5-trisphosphate receptor-like protein in plasmalemmal caveolae. *J Cell Biol* 1992, 119, 1507-1513, doi:10.1083/jcb.119.6.1507.
- 146. Isshiki, M.; Ando, J.; Korenaga, R.; Kogo, H.; Fujimoto, T.; Fujita, T.; Kamiya, A. Endothelial Ca2+ waves preferentially originate at specific loci in caveolin-rich cell edges. *Proc Natl Acad Sci U S A* **1998**, *95*, 5009-5014, doi:10.1073/pnas.95.9.5009.
- 147. Isshiki, M.; Ying, Y.S.; Fujita, T.; Anderson, R.G. A molecular sensor detects signal transduction from caveolae in living cells. *J Biol Chem* **2002**, 277, 43389-43398, doi:10.1074/jbc.M205411200.
- 148. Saliez, J.; Bouzin, C.; Rath, G.; Ghisdal, P.; Desjardins, F.; Rezzani, R.; Rodella, L.F.; Vriens, J.; Nilius, B.; Feron, O., et al. Role of caveolar compartmentation in endothelium-derived hyperpolarizing factor-mediated relaxation: Ca2+ signals and gap junction function are regulated by caveolin in endothelial cells. *Circulation* **2008**, *117*, 1065-1074, doi:10.1161/CIRCULATIONAHA.107.731679.
- 149. Medvedev, R.Y.; Turner, D.G.P.; DeGuire, F.C.; Leonov, V.; Lang, D.; Gorelik, J.; Alvarado, F.J.; Bondarenko, V.E.; Glukhov, A.V. Caveolae-associated cAMP/Ca(2+)-mediated mechano-chemical signal transduction in mouse atrial myocytes. *J Mol Cell Cardiol* 2023, 184, 75-87, doi:10.1016/j.yjmcc.2023.10.004.
- 150. Smart, E.J.; Ying, Y.S.; Anderson, R.G. Hormonal regulation of caveolae internalization. *J Cell Biol* **1995**, *131*, 929-938, doi:10.1083/jcb.131.4.929.
- 151. Smart, E.J.; Foster, D.C.; Ying, Y.S.; Kamen, B.A.; Anderson, R.G. Protein kinase C activators inhibit receptor-mediated potocytosis by preventing internalization of caveolae. *J Cell Biol* **1994**, 124, 307-313, doi:10.1083/jcb.124.3.307.
- 152. Mineo, C.; Ying, Y.S.; Chapline, C.; Jaken, S.; Anderson, R.G. Targeting of protein kinase Calpha to caveolae. *J Cell Biol* **1998**, *141*, 601-610, doi:10.1083/jcb.141.3.601.
- 153. Oka, N.; Yamamoto, M.; Schwencke, C.; Kawabe, J.; Ebina, T.; Ohno, S.; Couet, J.; Lisanti, M.P.; Ishikawa, Y. Caveolin interaction with protein kinase C. Isoenzyme-dependent regulation of kinase activity by the caveolin scaffolding domain peptide. *J Biol Chem* **1997**, 272, 33416-33421, doi:10.1074/jbc.272.52.33416.
- 154. Dhyani, V.; Gare, S.; Gupta, R.K.; Swain, S.; Venkatesh, K.V.; Giri, L. GPCR mediated control of calcium dynamics: A systems perspective. *Cell Signal* **2020**, *74*, 109717, doi:10.1016/j.cellsig.2020.109717.
- 155. Patel, S.; Churchill, G.C.; Galione, A. Coordination of Ca2+ signalling by NAADP. *Trends Biochem Sci* **2001**, 26, 482-489, doi:10.1016/s0968-0004(01)01896-5.

- 156. Balijepalli, R.C.; Kamp, T.J. Caveolae, ion channels and cardiac arrhythmias. *Prog Biophys Mol Biol* **2008**, *98*, 149-160, doi:10.1016/j.pbiomolbio.2009.01.012.
- 157. Tsutsumi, Y.M.; Horikawa, Y.T.; Jennings, M.M.; Kidd, M.W.; Niesman, I.R.; Yokoyama, U.; Head, B.P.; Hagiwara, Y.; Ishikawa, Y.; Miyanohara, A., et al. Cardiac-specific overexpression of caveolin-3 induces endogenous cardiac protection by mimicking ischemic preconditioning. *Circulation* **2008**, *118*, 1979-1988, doi:10.1161/CIRCULATIONAHA.108.788331.
- 158. Woodman, S.E.; Park, D.S.; Cohen, A.W.; Cheung, M.W.; Chandra, M.; Shirani, J.; Tang, B.; Jelicks, L.A.; Kitsis, R.N.; Christ, G.J., et al. Caveolin-3 knock-out mice develop a progressive cardiomyopathy and show hyperactivation of the p42/44 MAPK cascade. *J Biol Chem* **2002**, 277, 38988-38997, doi:10.1074/jbc.M205511200.
- 159. Galbiati, F.; Engelman, J.A.; Volonte, D.; Zhang, X.L.; Minetti, C.; Li, M.; Hou, H., Jr.; Kneitz, B.; Edelmann, W.; Lisanti, M.P. Caveolin-3 null mice show a loss of caveolae, changes in the microdomain distribution of the dystrophin-glycoprotein complex, and t-tubule abnormalities. *J Biol Chem* **2001**, *276*, 21425-21433, doi:10.1074/jbc.M100828200.
- 160. Bryant, S.M.; Kong, C.H.T.; Watson, J.J.; Gadeberg, H.C.; Roth, D.M.; Patel, H.H.; Cannell, M.B.; James, A.F.; Orchard, C.H. Caveolin-3 KO disrupts t-tubule structure and decreases t-tubular I(Ca) density in mouse ventricular myocytes. *American journal of physiology. Heart and circulatory physiology* **2018**, 315, H1101-H1111, doi:10.1152/ajpheart.00209.2018.
- 161. Park, D.S.; Cohen, A.W.; Frank, P.G.; Razani, B.; Lee, H.; Williams, T.M.; Chandra, M.; Shirani, J.; De Souza, A.P.; Tang, B., et al. Caveolin-1 null (-/-) mice show dramatic reductions in life span. *Biochemistry* **2003**, 42, 15124-15131, doi:10.1021/bi0356348.
- 162. Cohen, A.W.; Park, D.S.; Woodman, S.E.; Williams, T.M.; Chandra, M.; Shirani, J.; Pereira de Souza, A.; Kitsis, R.N.; Russell, R.G.; Weiss, L.M., et al. Caveolin-1 null mice develop cardiac hypertrophy with hyperactivation of p42/44 MAP kinase in cardiac fibroblasts. *Am J Physiol Cell Physiol* 2003, 284, C457-474, doi:10.1152/ajpcell.00380.2002.
- 163. Zhao, Y.Y.; Liu, Y.; Stan, R.V.; Fan, L.; Gu, Y.; Dalton, N.; Chu, P.H.; Peterson, K.; Ross, J., Jr.; Chien, K.R. Defects in caveolin-1 cause dilated cardiomyopathy and pulmonary hypertension in knockout mice. *Proc Natl Acad Sci U S A* **2002**, *99*, 11375-11380, doi:10.1073/pnas.172360799.
- 164. Park, D.S.; Woodman, S.E.; Schubert, W.; Cohen, A.W.; Frank, P.G.; Chandra, M.; Shirani, J.; Razani, B.; Tang, B.; Jelicks, L.A., et al. Caveolin-1/3 double-knockout mice are viable, but lack both muscle and non-muscle caveolae, and develop a severe cardiomyopathic phenotype. *Am J Pathol* **2002**, *160*, 2207-2217.
- 165. Shaul, P.W.; Smart, E.J.; Robinson, L.J.; German, Z.; Yuhanna, I.S.; Ying, Y.; Anderson, R.G.; Michel, T. Acylation targets emdothelial nitric-oxide synthase to plasmalemmal caveolae. *J Biol Chem* **1996**, 271, 6518-6522
- 166. Garcia-Cardena, G.; Oh, P.; Liu, J.; Schnitzer, J.E.; Sessa, W.C. Targeting of nitric oxide synthase to endothelial cell caveolae via palmitoylation: implications for nitric oxide signaling. *Proc Natl Acad Sci U S A* **1996**, *93*, 6448-6453.
- 167. Wang, Q.; Lao, M.; Xu, Z.; Ding, M.; Guo, S.; Li, L. Caveolin-1 modulates hypertensive vascular remodeling via regulation of the Notch pathway. *Mol Med Rep* **2020**, 22, 4320-4328, doi:10.3892/mmr.2020.11508.
- 168. Hayflick, L. The Limited in Vitro Lifetime of Human Diploid Cell Strains. *Exp Cell Res* **1965**, *37*, 614-636, doi:10.1016/0014-4827(65)90211-9.
- 169. Zhang, L.; Pitcher, L.E.; Yousefzadeh, M.J.; Niedernhofer, L.J.; Robbins, P.D.; Zhu, Y. Cellular senescence: a key therapeutic target in aging and diseases. *J Clin Invest* **2022**, *132*, doi:10.1172/JCI158450.
- 170. Di Micco, R.; Krizhanovsky, V.; Baker, D.; d'Adda di Fagagna, F. Cellular senescence in ageing: from mechanisms to therapeutic opportunities. *Nat Rev Mol Cell Biol* **2021**, 22, 75-95, doi:10.1038/s41580-020-00314-w.
- 171. Kumari, R.; Jat, P. Mechanisms of Cellular Senescence: Cell Cycle Arrest and Senescence Associated Secretory Phenotype. *Front Cell Dev Biol* **2021**, *9*, 645593, doi:10.3389/fcell.2021.645593.
- 172. Gorgoulis, V.; Adams, P.D.; Alimonti, A.; Bennett, D.C.; Bischof, O.; Bischof, C.; Campisi, J.; Collado, M.; Evangelou, K.; Ferbeyre, G., et al. Cellular Senescence: Defining a Path Forward. *Cell* **2019**, 179, 813-827, doi:10.1016/j.cell.2019.10.005.
- 173. Munoz-Espin, D.; Canamero, M.; Maraver, A.; Gomez-Lopez, G.; Contreras, J.; Murillo-Cuesta, S.; Rodriguez-Baeza, A.; Varela-Nieto, I.; Ruberte, J.; Collado, M., et al. Programmed cell senescence during mammalian embryonic development. *Cell* **2013**, *155*, 1104-1118, doi:10.1016/j.cell.2013.10.019.
- 174. Suda, M.; Paul, K.H.; Minamino, T.; Miller, J.D.; Lerman, A.; Ellison-Hughes, G.M.; Tchkonia, T.; Kirkland, J.L. Senescent Cells: A Therapeutic Target in Cardiovascular Diseases. *Cells* **2023**, 12, doi:10.3390/cells12091296.
- 175. Volonte, D.; Galbiati, F. Caveolin-1, a master regulator of cellular senescence. *Cancer Metastasis Rev* **2020**, 39, 397-414, doi:10.1007/s10555-020-09875-w.
- 176. Cho, K.A.; Ryu, S.J.; Park, J.S.; Jang, I.S.; Ahn, J.S.; Kim, K.T.; Park, S.C. Senescent phenotype can be reversed by reduction of caveolin status. *J Biol Chem* **2003**, 278, 27789-27795, doi:10.1074/jbc.M208105200.

- 178. Lim, J.S.; Nguyen, K.C.; Nguyen, C.T.; Jang, I.S.; Han, J.M.; Fabian, C.; Lee, S.E.; Rhee, J.H.; Cho, K.A. Flagellin-dependent TLR5/caveolin-1 as a promising immune activator in immunosenescence. *Aging Cell* **2015**, *14*, 907-915, doi:10.1111/acel.12383.
- 179. Sun, C.; Wang, N.; Huang, J.; Xin, J.; Peng, F.; Ren, Y.; Zhang, S.; Miao, J. Inhibition of phosphatidylcholine-specific phospholipase C prevents bone marrow stromal cell senescence in vitro. *J Cell Biochem* **2009**, *108*, 519-528, doi:10.1002/jcb.22282.
- 180. Park, J.S.; Kim, H.Y.; Kim, H.W.; Chae, G.N.; Oh, H.T.; Park, J.Y.; Shim, H.; Seo, M.; Shin, E.Y.; Kim, E.G., et al. Increased caveolin-1, a cause for the declined adipogenic potential of senescent human mesenchymal stem cells. *Mech Ageing Dev* **2005**, *126*, 551-559, doi:10.1016/j.mad.2004.11.014.
- 181. Dasari, A.; Bartholomew, J.N.; Volonte, D.; Galbiati, F. Oxidative stress induces premature senescence by stimulating caveolin-1 gene transcription through p38 mitogen-activated protein kinase/Sp1-mediated activation of two GC-rich promoter elements. *Cancer Res* **2006**, *66*, 10805-10814, doi:10.1158/0008-5472.CAN-06-1236.
- 182. Farhat, N.; Thorin-Trescases, N.; Voghel, G.; Villeneuve, L.; Mamarbachi, M.; Perrault, L.P.; Carrier, M.; Thorin, E. Stress-induced senescence predominates in endothelial cells isolated from atherosclerotic chronic smokers. *Can J Physiol Pharmacol* **2008**, *86*, 761-769, doi:10.1139/Y08-082.
- 183. Galbiati, F.; Volonte, D.; Liu, J.; Capozza, F.; Frank, P.G.; Zhu, L.; Pestell, R.G.; Lisanti, M.P. Caveolin-1 expression negatively regulates cell cycle progression by inducing G(0)/G(1) arrest via a p53/p21(WAF1/Cip1)-dependent mechanism. *Mol Biol Cell* **2001**, 12, 2229-2244.
- 184. Ding, L.; Zeng, Q.; Wu, J.; Li, D.; Wang, H.; Lu, W.; Jiang, Z.; Xu, G. Caveolin-1 regulates oxidative stress-induced senescence in nucleus pulposus cells primarily via the p53/p21 signaling pathway in vitro. *Mol Med Rep* 2017, 16, 9521-9527, doi:10.3892/mmr.2017.7789.
- 185. Volonte, D.; Kahkonen, B.; Shapiro, S.; Di, Y.; Galbiati, F. Caveolin-1 expression is required for the development of pulmonary emphysema through activation of the ATM-p53-p21 pathway. *J Biol Chem* **2009**, 284, 5462-5466, doi:10.1074/jbc.C800225200.
- 186. Bartholomew, J.N.; Volonte, D.; Galbiati, F. Caveolin-1 regulates the antagonistic pleiotropic properties of cellular senescence through a novel Mdm2/p53-mediated pathway. *Cancer Res* **2009**, *69*, 2878-2886, doi:10.1158/0008-5472.CAN-08-2857.
- 187. Bai, L.; Deng, X.; Li, J.; Wang, M.; Li, Q.; An, W.; A, D.; Cong, Y.S. Regulation of cellular senescence by the essential caveolar component PTRF/Cavin-1. *Cell Res* **2011**, *21*, 1088-1101, doi:10.1038/cr.2011.56.
- 188. Yu, D.M.; Jung, S.H.; An, H.T.; Lee, S.; Hong, J.; Park, J.S.; Lee, H.; Lee, H.; Bahn, M.S.; Lee, H.C., et al. Caveolin-1 deficiency induces premature senescence with mitochondrial dysfunction. *Aging Cell* **2017**.
- 189. Madaro, L.; Antonangeli, F.; Favia, A.; Esposito, B.; Biamonte, F.; Bouche, M.; Ziparo, E.; Sica, G.; Filippini, A.; D'Alessio, A. Knock down of caveolin-1 affects morphological and functional hallmarks of human endothelial cells. *J Cell Biochem* **2013**, *114*, 1843-1851.
- 190. Razani, B.; Woodman, S.E.; Lisanti, M.P. Caveolae: from cell biology to animal physiology. *Pharmacol Rev* **2002**, *54*, 431-467.
- 191. Diaz, M.I.; Diaz, P.; Bennett, J.C.; Urra, H.; Ortiz, R.; Orellana, P.C.; Hetz, C.; Quest, A.F.G. Caveolin-1 suppresses tumor formation through the inhibition of the unfolded protein response. *Cell Death Dis* **2020**, 11, 648, doi:10.1038/s41419-020-02792-4.
- 192. Sanhueza, C.; Bennett, J.C.; Valenzuela-Valderrama, M.; Contreras, P.; Lobos-Gonzalez, L.; Campos, A.; Wehinger, S.; Lladser, A.; Kiessling, R.; Leyton, L., et al. Caveolin-1-Mediated Tumor Suppression Is Linked to Reduced HIF1alpha S-Nitrosylation and Transcriptional Activity in Hypoxia. *Cancers* (*Basel*) **2020**, 12, doi:10.3390/cancers12092349.
- 193. Williams, T.M.; Lisanti, M.P. The caveolin proteins. Genome Biol 2004, 5, 214.
- 194. Galbiati, F.; Volonte, D.; Engelman, J.A.; Watanabe, G.; Burk, R.; Pestell, R.G.; Lisanti, M.P. Targeted downregulation of caveolin-1 is sufficient to drive cell transformation and hyperactivate the p42/44 MAP kinase cascade. *Embo J* 1998, 17, 6633-6648.
- 195. Ko, Y.G.; Liu, P.; Pathak, R.K.; Craig, L.C.; Anderson, R.G. Early effects of pp60(v-src) kinase activation on caveolae. *J Cell Biochem* **1998**, *71*, 524-535.
- 196. Glenney, J.R., Jr. Tyrosine phosphorylation of a 22-kDa protein is correlated with transformation by Rous sarcoma virus. *J Biol Chem* **1989**, *264*, 20163-20166.
- 197. Wiechen, K.; Diatchenko, L.; Agoulnik, A.; Schafff, K.M.; Schober, H.; Arlt, K.; Zhumabayeva, B.; Siebert, P.D.; Dietel, M.; Schafer, R., et al. Caveolin-1 is down-regulated in human ovarian carcinoma and acts as a candidate tumor suppressor gene. *Am J Pathol* **2001**, *159*, 1635-1643, doi:10.1016/S0002-9440(10)63010-6.
- 198. Lee, S.W.; Reimer, C.L.; Oh, P.; Campbell, D.B.; Schnitzer, J.E. Tumor cell growth inhibition by caveolin reexpression in human breast cancer cells. *Oncogene* **1998**, *16*, 1391-1397, doi:10.1038/sj.onc.1201661.

- 199. Engelman, J.A.; Wykoff, C.C.; Yasuhara, S.; Song, K.S.; Okamoto, T.; Lisanti, M.P. Recombinant expression of caveolin-1 in oncogenically transformed cells abrogates anchorage-independent growth. *J Biol Chem* **1997**, 272, 16374-16381, doi:10.1074/jbc.272.26.16374.
- 200. Koleske, A.J.; Baltimore, D.; Lisanti, M.P. Reduction of caveolin and caveolae in oncogenically transformed cells. *Proc Natl Acad Sci U S A* **1995**, 92, 1381-1385.
- 201. Felicetti, F.; Parolini, I.; Bottero, L.; Fecchi, K.; Errico, M.C.; Raggi, C.; Biffoni, M.; Spadaro, F.; Lisanti, M.P.; Sargiacomo, M., et al. Caveolin-1 tumor-promoting role in human melanoma. *Int J Cancer* 2009, 125, 1514-1522, doi:10.1002/ijc.24451.
- 202. Lagares-Tena, L.; Garcia-Monclus, S.; Lopez-Alemany, R.; Almacellas-Rabaiget, O.; Huertas-Martinez, J.; Sainz-Jaspeado, M.; Mateo-Lozano, S.; Rodriguez-Galindo, C.; Rello-Varona, S.; Herrero-Martin, D., et al. Caveolin-1 promotes Ewing sarcoma metastasis regulating MMP-9 expression through MAPK/ERK pathway. *Oncotarget* 2016, 7, 56889-56903, doi:10.18632/oncotarget.10872.
- 203. Sainz-Jaspeado, M.; Lagares-Tena, L.; Lasheras, J.; Navid, F.; Rodriguez-Galindo, C.; Mateo-Lozano, S.; Notario, V.; Sanjuan, X.; Garcia Del Muro, X.; Fabra, A., et al. Caveolin-1 modulates the ability of Ewing's sarcoma to metastasize. *Mol Cancer Res* **2010**, *8*, 1489-1500, doi:10.1158/1541-7786.MCR-10-0060.
- 204. Zhang, C.; Huang, H.; Zhang, J.; Wu, Q.; Chen, X.; Huang, T.; Li, W.; Liu, Y.; Zhang, J. Caveolin-1 promotes invasion and metastasis by upregulating Pofut1 expression in mouse hepatocellular carcinoma. *Cell Death Dis* 2019, *10*, 477, doi:10.1038/s41419-019-1703-1.
- 205. Moreno-Caceres, J.; Caballero-Diaz, D.; Nwosu, Z.C.; Meyer, C.; Lopez-Luque, J.; Malfettone, A.; Lastra, R.; Serrano, T.; Ramos, E.; Dooley, S., et al. The level of caveolin-1 expression determines response to TGF-beta as a tumour suppressor in hepatocellular carcinoma cells. *Cell Death Dis* **2017**, *8*, e3098, doi:10.1038/cddis.2017.469.
- 206. Tse, E.Y.; Ko, F.C.; Tung, E.K.; Chan, L.K.; Lee, T.K.; Ngan, E.S.; Man, K.; Wong, A.S.; Ng, I.O.; Yam, J.W. Caveolin-1 overexpression is associated with hepatocellular carcinoma tumourigenesis and metastasis. *J Pathol* 2012, 226, 645-653, doi:10.1002/path.3957.
- 207. Cokakli, M.; Erdal, E.; Nart, D.; Yilmaz, F.; Sagol, O.; Kilic, M.; Karademir, S.; Atabey, N. Differential expression of Caveolin-1 in hepatocellular carcinoma: correlation with differentiation state, motility and invasion. *BMC Cancer* 2009, 9, 65, doi:10.1186/1471-2407-9-65.
- 208. Ho, C.C.; Huang, P.H.; Huang, H.Y.; Chen, Y.H.; Yang, P.C.; Hsu, S.M. Up-regulated caveolin-1 accentuates the metastasis capability of lung adenocarcinoma by inducing filopodia formation. *Am J Pathol* **2002**, *161*, 1647-1656, doi:10.1016/S0002-9440(10)64442-2.
- 209. Yang, D.; Liu, J.; Qian, H.; Zhuang, Q. Cancer-associated fibroblasts: from basic science to anticancer therapy. *Exp Mol Med* **2023**, *55*, 1322-1332, doi:10.1038/s12276-023-01013-0.
- 210. Glabman, R.A.; Choyke, P.L.; Sato, N. Cancer-Associated Fibroblasts: Tumorigenicity and Targeting for Cancer Therapy. *Cancers* (*Basel*) **2022**, *14*, doi:10.3390/cancers14163906.
- 211. Rizzolio, S.; Giordano, S.; Corso, S. The importance of being CAFs (in cancer resistance to targeted therapies). *J Exp Clin Cancer Res* **2022**, *41*, 319, doi:10.1186/s13046-022-02524-w.
- 212. Wang, F.T.; Sun, W.; Zhang, J.T.; Fan, Y.Z. Cancer-associated fibroblast regulation of tumor neo-angiogenesis as a therapeutic target in cancer. *Oncol Lett* **2019**, *17*, 3055-3065, doi:10.3892/ol.2019.9973.
- 213. Angelucci, C.; D'Alessio, A.; Iacopino, F.; Proietti, G.; Di Leone, A.; Masetti, R.; Sica, G. Pivotal role of human stearoyl-CoA desaturases (SCD1 and 5) in breast cancer progression: oleic acid-based effect of SCD1 on cell migration and a novel pro-cell survival role for SCD5. *Oncotarget* 2018, 9, 24364-24380, doi:10.18632/oncotarget.25273.
- 214. Angelucci, C.; Maulucci, G.; Colabianchi, A.; Iacopino, F.; D'Alessio, A.; Maiorana, A.; Palmieri, V.; Papi, M.; De Spirito, M.; Di Leone, A., et al. Stearoyl-CoA desaturase 1 and paracrine diffusible signals have a major role in the promotion of breast cancer cell migration induced by cancer-associated fibroblasts. *Br J Cancer* 2015, *112*, 1675-1686, doi:10.1038/bjc.2015.135.
- 215. Guo, S.; Deng, C.X. Effect of Stromal Cells in Tumor Microenvironment on Metastasis Initiation. *Int J Biol Sci* **2018**, *14*, 2083-2093, doi:10.7150/ijbs.25720.
- 216. Zhao, X.; He, Y.; Gao, J.; Fan, L.; Li, Z.; Yang, G.; Chen, H. Caveolin-1 expression level in cancer associated fibroblasts predicts outcome in gastric cancer. *PLoS One* **2013**, *8*, e59102, doi:10.1371/journal.pone.0059102.
- 217. Simpkins, S.A.; Hanby, A.M.; Holliday, D.L.; Speirs, V. Clinical and functional significance of loss of caveolin-1 expression in breast cancer-associated fibroblasts. *J Pathol* **2012**, 227, 490-498, doi:10.1002/path.4034.
- 218. Wu, K.N.; Queenan, M.; Brody, J.R.; Potoczek, M.; Sotgia, F.; Lisanti, M.P.; Witkiewicz, A.K. Loss of stromal caveolin-1 expression in malignant melanoma metastases predicts poor survival. *Cell Cycle* **2011**, *10*, 4250-4255, doi:10.4161/cc.10.24.18551.
- 219. Witkiewicz, A.K.; Dasgupta, A.; Sotgia, F.; Mercier, I.; Pestell, R.G.; Sabel, M.; Kleer, C.G.; Brody, J.R.; Lisanti, M.P. An absence of stromal caveolin-1 expression predicts early tumor recurrence and poor clinical outcome in human breast cancers. *Am J Pathol* **2009**, *174*, 2023-2034, doi:10.2353/ajpath.2009.080873.

- 220. Sloan, E.K.; Ciocca, D.R.; Pouliot, N.; Natoli, A.; Restall, C.; Henderson, M.A.; Fanelli, M.A.; Cuello-Carrion, F.D.; Gago, F.E.; Anderson, R.L. Stromal cell expression of caveolin-1 predicts outcome in breast cancer. *Am J Pathol* 2009, 174, 2035-2043, doi:10.2353/ajpath.2009.080924.
- 221. Sotgia, F.; Del Galdo, F.; Casimiro, M.C.; Bonuccelli, G.; Mercier, I.; Whitaker-Menezes, D.; Daumer, K.M.; Zhou, J.; Wang, C.; Katiyar, S., et al. Caveolin-1-/- null mammary stromal fibroblasts share characteristics with human breast cancer-associated fibroblasts. *Am J Pathol* **2009**, *174*, 746-761, doi:10.2353/ajpath.2009.080658.
- 222. Witkiewicz, A.K.; Dasgupta, A.; Sammons, S.; Er, O.; Potoczek, M.B.; Guiles, F.; Sotgia, F.; Brody, J.R.; Mitchell, E.P.; Lisanti, M.P. Loss of stromal caveolin-1 expression predicts poor clinical outcome in triple negative and basal-like breast cancers. *Cancer Biol Ther* **2010**, *10*, 135-143, doi:10.4161/cbt.10.2.11983.
- 223. El-Gendi, S.M.; Mostafa, M.F.; El-Gendi, A.M. Stromal caveolin-1 expression in breast carcinoma. Correlation with early tumor recurrence and clinical outcome. *Pathol Oncol Res* **2012**, *18*, 459-469, doi:10.1007/s12253-011-9469-5.
- 224. He, Y.; Zhao, X.; Gao, J.; Fan, L.; Yang, G.; Cho, W.C.; Chen, H. Quantum dots-based immunofluorescent imaging of stromal fibroblasts Caveolin-1 and light chain 3B expression and identification of their clinical significance in human gastric cancer. *Int J Mol Sci* **2012**, *13*, 13764-13780, doi:10.3390/ijms131113764.
- 225. Di Vizio, D.; Morello, M.; Sotgia, F.; Pestell, R.G.; Freeman, M.R.; Lisanti, M.P. An absence of stromal caveolin-1 is associated with advanced prostate cancer, metastatic disease and epithelial Akt activation. *Cell Cycle* **2009**, *8*, 2420-2424, doi:10.4161/cc.8.15.9116.
- 226. Cameron, P.L.; Ruffin, J.W.; Bollag, R.; Rasmussen, H.; Cameron, R.S. Identification of caveolin and caveolin-related proteins in the brain. *J Neurosci* **1997**, *17*, 9520-9535, doi:10.1523/JNEUROSCI.17-24-09520.1997.
- 227. Wu, C.; Butz, S.; Ying, Y.; Anderson, R.G. Tyrosine kinase receptors concentrated in caveolae-like domains from neuronal plasma membrane. *J Biol Chem* **1997**, *272*, 3554-3559, doi:10.1074/jbc.272.6.3554.
- 228. Stern, C.M.; Mermelstein, P.G. Caveolin regulation of neuronal intracellular signaling. *Cell Mol Life Sci* **2010**, 67, 3785-3795, doi:10.1007/s00018-010-0447-y.
- 229. Galbiati, F.; Volonte, D.; Gil, O.; Zanazzi, G.; Salzer, J.L.; Sargiacomo, M.; Scherer, P.E.; Engelman, J.A.; Schlegel, A.; Parenti, M., et al. Expression of caveolin-1 and -2 in differentiating PC12 cells and dorsal root ganglion neurons: caveolin-2 is up-regulated in response to cell injury. *Proc Natl Acad Sci U S A* 1998, 95, 10257-10262, doi:10.1073/pnas.95.17.10257.
- 230. Koh, S.; Lee, W.; Park, S.M.; Kim, S.H. Caveolin-1 deficiency impairs synaptic transmission in hippocampal neurons. *Mol Brain* **2021**, *14*, 53, doi:10.1186/s13041-021-00764-z.
- 231. Jasmin, J.F.; Malhotra, S.; Singh Dhallu, M.; Mercier, I.; Rosenbaum, D.M.; Lisanti, M.P. Caveolin-1 deficiency increases cerebral ischemic injury. *Circ Res* **2007**, *100*, 721-729, doi:10.1161/01.RES.0000260180.42709.29.
- 232. Gu, Y.; Zheng, G.; Xu, M.; Li, Y.; Chen, X.; Zhu, W.; Tong, Y.; Chung, S.K.; Liu, K.J.; Shen, J. Caveolin-1 regulates nitric oxide-mediated matrix metalloproteinases activity and blood-brain barrier permeability in focal cerebral ischemia and reperfusion injury. *J Neurochem* **2012**, *120*, 147-156, doi:10.1111/j.1471-4159.2011.07542.x.
- 233. Bjorkhem, I.; Meaney, S. Brain cholesterol: long secret life behind a barrier. *Arterioscler Thromb Vasc Biol* **2004**, 24, 806-815, doi:10.1161/01.ATV.0000120374.59826.1b.
- 234. Ikezu, T.; Trapp, B.D.; Song, K.S.; Schlegel, A.; Lisanti, M.P.; Okamoto, T. Caveolae, plasma membrane microdomains for alpha-secretase-mediated processing of the amyloid precursor protein. *J Biol Chem* **1998**, 273, 10485-10495, doi:10.1074/jbc.273.17.10485.
- 235. Lee, S.J.; Liyanage, U.; Bickel, P.E.; Xia, W.; Lansbury, P.T., Jr.; Kosik, K.S. A detergent-insoluble membrane compartment contains A beta in vivo. *Nat Med* **1998**, *4*, 730-734, doi:10.1038/nm0698-730.
- 236. Kapoor, A.; Hsu, W.M.; Wang, B.J.; Wu, G.H.; Lin, T.Y.; Lee, S.J.; Yen, C.T.; Liang, S.M.; Liao, Y.F. Caveolin-1 regulates gamma-secretase-mediated AbetaPP processing by modulating spatial distribution of gamma-secretase in membrane. *J Alzheimers Dis* **2010**, 22, 423-442, doi:10.3233/JAD-2010-100531.
- 237. Perez, S.E.; Getova, D.P.; He, B.; Counts, S.E.; Geula, C.; Desire, L.; Coutadeur, S.; Peillon, H.; Ginsberg, S.D.; Mufson, E.J. Rac1b increases with progressive tau pathology within cholinergic nucleus basalis neurons in Alzheimer's disease. *Am J Pathol* **2012**, *180*, 526-540, doi:10.1016/j.ajpath.2011.10.027.
- 238. Metaxas, A.; Kempf, S.J. Neurofibrillary tangles in Alzheimer's disease: elucidation of the molecular mechanism by immunohistochemistry and tau protein phospho-proteomics. *Neural Regen Res* **2016**, *11*, 1579-1581, doi:10.4103/1673-5374.193234.
- 239. Nishiyama, K.; Trapp, B.D.; Ikezu, T.; Ransohoff, R.M.; Tomita, T.; Iwatsubo, T.; Kanazawa, I.; Hsiao, K.K.; Lisanti, M.P.; Okamoto, T. Caveolin-3 upregulation activates beta-secretase-mediated cleavage of the amyloid precursor protein in Alzheimer's disease. *J Neurosci* **1999**, *19*, 6538-6548, doi:10.1523/JNEUROSCI.19-15-06538.1999.
- 240. Hashimoto, M.; Takenouchi, T.; Rockenstein, E.; Masliah, E. Alpha-synuclein up-regulates expression of caveolin-1 and down-regulates extracellular signal-regulated kinase activity in B103 neuroblastoma cells:

- role in the pathogenesis of Parkinson's disease. *J Neurochem* **2003**, *85*, 1468-1479, doi:10.1046/j.1471-4159.2003.01791.x.
- 241. Madeira, A.; Yang, J.; Zhang, X.; Vikeved, E.; Nilsson, A.; Andren, P.E.; Svenningsson, P. Caveolin-1 interacts with alpha-synuclein and mediates toxic actions of cellular alpha-synuclein overexpression. *Neurochem Int* **2011**, *59*, 280-289, doi:10.1016/j.neuint.2011.05.017.
- 242. Dawson, T.M.; Dawson, V.L. The role of parkin in familial and sporadic Parkinson's disease. *Mov Disord* **2010**, 25 *Suppl 1*, S32-39, doi:10.1002/mds.22798.
- 243. Cha, S.H.; Choi, Y.R.; Heo, C.H.; Kang, S.J.; Joe, E.H.; Jou, I.; Kim, H.M.; Park, S.M. Loss of parkin promotes lipid rafts-dependent endocytosis through accumulating caveolin-1: implications for Parkinson's disease. *Mol Neurodegener* 2015, *10*, 63, doi:10.1186/s13024-015-0060-5.
- 244. Cataldi, S.; Codini, M.; Hunot, S.; Legeron, F.P.; Ferri, I.; Siccu, P.; Sidoni, A.; Ambesi-Impiombato, F.S.; Beccari, T.; Curcio, F., et al. e-Cadherin in 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine-Induced Parkinson Disease. *Mediators Inflamm* **2016**, 2016, 3937057, doi:10.1155/2016/3937057.
- 245. Kim, J.M.; Cha, S.H.; Choi, Y.R.; Jou, I.; Joe, E.H.; Park, S.M. DJ-1 deficiency impairs glutamate uptake into astrocytes via the regulation of flotillin-1 and caveolin-1 expression. *Sci Rep* **2016**, *6*, 28823, doi:10.1038/srep28823.
- 246. Kumari, M.; Ramdas, P.; Radhakrishnan, A.K.; Kutty, M.K.; Haleagrahara, N. Tocotrienols Ameliorate Neurodegeneration and Motor Deficits in the 6-OHDA-Induced Rat Model of Parkinsonism: Behavioural and Immunohistochemistry Analysis. *Nutrients* **2021**, *13*, doi:10.3390/nu13051583.
- 247. Nakaso, K.; Tajima, N.; Horikoshi, Y.; Nakasone, M.; Hanaki, T.; Kamizaki, K.; Matsura, T. The estrogen receptor beta-PI3K/Akt pathway mediates the cytoprotective effects of tocotrienol in a cellular Parkinson's disease model. *Biochim Biophys Acta* **2014**, *18*42, 1303-1312, doi:10.1016/j.bbadis.2014.04.008.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.