

Review

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Review

Nanomaterials-Enhanced Electrochemical Biosensors for Epithelial Cancer Diagnosis: Recent Advances

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Abstract

In recent years, the interest in electrochemical biosensors has been constantly growing for epithelial cancer diagnosis and prognosis. The incorporation of the different nanomaterials, specific monoclonal antibodies, and nucleic acids (aptamers) into these methodologies has managed to improve sensitivity and specificity. In this review, we have presented examples of electrochemical biosensors for the determination of different epithelial cancer biomarkers. Based on many published literature examples, we have emphasized the recent application of one and multiplexed platforms for the quantification of epithelial cancer biomarkers. Finally, we discussed the possible way of development, challenges, and future perspectives in the area of electrochemical immuno-, apta-, and geno-sensors.

Keywords: biosensor; electrochemical; nanomaterial; tumor biomarker; cancer diagnosis; real samples

1. Introduction

Cancer is one of the most significant global health challenges today, recognized as a leading cause of morbidity and mortality worldwide. Epithelial cancer is particularly relevant as it encompasses a wide range of common cancers, including breast, prostate, colon, colorectal, lung, ovarian, stomach, pancreatic, and thyroid cancers [1,2]. With the highest mortality rate among all cancer types, epithelial cancer has become a major global threat. This high mortality is primarily due to the lack of specific early-stage symptoms, which often leads to a late diagnosis, typically after the cancer has already metastasized [3,4].

The traditional diagnosis and prognosis of cancer rely on identifying specific tumor markers for each cancer type, which must be correlated with the patient's clinical data [5,6]. Consequently, early

diagnosis and treatment are crucial for improving patient mortality rates and quality of life [7,8]. This has created an urgent clinical need for economical and effective methods for the early diagnosis of these cancers [9]. The rapid analysis and determination of epithelial cancer biomarkers in biological samples such as blood, urine, serum, saliva, or extracellular vesicles (EVs) are essential for early screening, understanding disease progression, and guiding treatment prognosis (Ivanova et al., 2024; Kang et al., 2024).

However, classic diagnostic methods—including immunohistochemistry, histopathology, enzyme-linked immunosorbent assay (ELISA), and radioimmunoassay are often slow, expensive, and require highly trained personnel [10]. For these reasons, developing sensitive, practical, and affordable methods for detecting epithelial biomarkers is critical for advancing early cancer diagnosis [11]. Electrochemical techniques offer a promising alternative due to their inherent sensitivity, selectivity, rapid response times, and potential for miniaturization [12,13].

In recent years, a wide range of sophisticated electrochemical biosensors has been developed for application in oncology. These integrated analytical devices convert the information from the molecular recognition process between a bioreceptor and the target analyte (the cancer biomarker) into a quantifiable signal [14,15]. The type of bioreceptor used, such as antibodies, antigens, oligonucleotides, or aptamers, also defines the biosensors category, leading to different types like immunosensors, genosensors, and aptasensors [16].

These devices employ various detection methods, with the most relevant being cyclic voltammetry (CV), differential pulse voltammetry (DPV), square wave voltammetry (SWV), and electrochemical impedance spectroscopy (EIS). The resulting electronic signals are then correlated with the concentration of a specific biomarker, providing a diagnostic result [17].

These sensors are characterized by their multiplexing capabilities, high sensitivity, excellent selectivity, fast detection, portability, and the potential for integration with advanced electronics [18,19]. To achieve low limits of detection, researchers often utilize various strategies, such as incorporating nanomaterials as immobilization platforms for biomolecules (DNA, RNA, antigens, antibodies, or aptamers) (Shen et al., 2024). Another effective strategy is to combine polymers and nanoparticles with conjugated structures, which synergistically enhances biosensor performance [20].

This review presents an overview of recent electrochemical biosensors developed for the determination of specific epithelial cancer biomarkers. We present examples that illustrate the use of diverse nanomaterials and bioreceptors, including the detection of newly identified biomarkers, together with detailed insights into design strategies, key characteristics, and the types of samples evaluated for both single and multiplex biomarker detection. This work provides valuable information for scientists working on developing new analytical devices with enhanced selectivity and specificity for cancer diagnosis and prognosis. Finally, we discuss the challenges and prospects in this important field.

2. Electrochemical Biosensors

A biosensor is a compact analytical device that integrates a biological recognition element (such as an antigen, antibody, DNA, RNA, enzyme, or aptamer) with a signal transducer, typically an electrochemical or optical detector. This design allows the biosensor to recognize specific target molecules and translate this interaction into a measurable signal (Figure 1) [1,3].

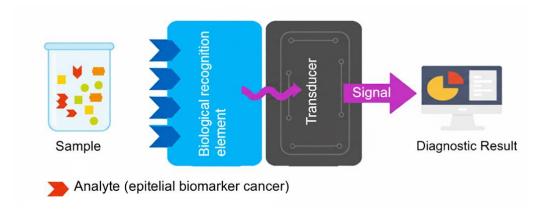


Figure 1. Schematic representation of a biosensor. Figure made by the authors.

Based on their biological recognition elements, electrochemical biosensors can be categorized into several types [2,4]:

- Enzymatic biosensors: These convert substrates into measurable products through enzymecatalyzed reactions.
- Protein receptor-based biosensors: These use cell membrane proteins as receptors.
- Immunosensors: A form of solid-phase immunoassay where antigens or antibodies are selectively attached to a solid support.
- DNA-Aptamer biosensors: These offer a low-cost, highly stable, and specific alternative to traditional antibodies.
- Whole-cell biosensors: These function as chemical reporters to measure the composition of an environment, including toxic or mutagenic substances.

Electrochemical biosensors also differ in their detection techniques, which can be broadly classified as [3,4]:

- Amperometric sensors: Measure the current generated by the oxidation or reduction of species in a biochemical reaction.
- Potentiometric sensors: Measure the electrical potential across electrodes as a result of a biochemical reaction.
- Impedimetric (and conductometric) sensors: Measure changes in impedance or conductivity caused by biochemical reactions.

These sensors are typically constructed with three main components: a working electrode, a reference electrode, and an auxiliary electrode. The working electrode interacts directly with the analyte during detection, while the other two electrodes maintain a stable potential and complete the electrical circuit, respectively [4]. These features make electrochemical biosensors highly suitable for addressing diagnostic needs, particularly for epithelial cancer. In this field, biomarkers are crucial for early detection, guiding treatment selection, and assessing prognosis. Electrochemical biosensors offer significant advantages over traditional methods for detecting these biomarkers, including high sensitivity, selectivity, stability, and low cost [2].

By utilizing diverse recognition elements and advanced materials like nanomaterials, electrochemical biosensors hold immense promise for increasing detection sensitivity, paving the way for groundbreaking diagnostic and therapeutic strategies in epithelial cancer [2–4].

3. Epithelial Cancer Biomarkers

3.1. Cancer Antigen (CA)

Cancer antigen 125 (CA125), also referred to as Mucin 16 (MUC-16), is a well-established biomarker commonly employed in ovarian cancer diagnosis. Elevated serum levels of this protein

(>35 U/mL) are widely recognized as a key indicator of increased ovarian cancer risk. Therefore, the early and reliable detection of CA125 with high sensitivity holds great clinical importance. In recent years, biosensors for biomarker detection have attracted increasing attention owing to their unique advantages, such as high sensitivity, selectivity, stability, and ease of fabrication. However, conventional biosensors relying on immobilized biomolecules as recognition elements often face limitations in stability. In this section, we highlight selected examples of sensing devices (see Table 1) that demonstrate how the incorporation of nanomaterials not only increases versatility but also addresses common limitations of conventional approaches, such as stability and reproducibility. For instance, Yilmaz et al. [21] reported the first fabrication of a rapid, low-cost, disposable, and labelfree CA125 immunosensor achieved by modifying PTB synthesized in a green solvent with AuNPs. The synergistic effect of PTBDES and AuNPs enhanced both the electrode's electrochemical properties and the overall sensing performance. The immunosensor exhibited excellent stability during storage and regeneration, as well as high selectivity in the presence of other tumor markers. CA125 detection in blood serum yielded recoveries with < 5% error, confirming its reliability. Owing to its simplicity, reproducibility, and strong analytical performance, this disposable CA125 immunosensor represents a promising candidate for point-of-care clinical testing of ovarian cancer biomarkers. Similarly, another CA125 immunosensor, based on amino-functionalized carbon nanotube Ti₃C₂Tx/NH₂-CNT-SPCE, exhibited rapid electron transfer, signal amplification, and enhanced stability via chitosan [22]. Compared with the AuNPs/PTB system, this approach offered a wider detection range (1 -500 U/mL) with low LOD (1 mU/mL), good selectivity, and reliable performance in clinical serum, demonstrating strong potential for ovarian cancer diagnostics. Notably, the clinical cut-off value for CA125 is ~35 U/mL in serum, which means that the reported LOD is well below the threshold required for clinical relevance. Guo et al. [23] developed Mn-doped ZnO nanorods functionalized with Au-NPs (ITO-Mn/ZnO-Au). The composite exhibited enhanced conductivity, charge transfer, and electrocatalytic activity, enabling efficient aptamer immobilization. The biosensor achieved a wide linear range (0.002-3 ng/mL), ultralow LOD (0.5 pg/mL), good reproducibility (RSD = 3.5 %), and high recovery (97.2-105 %) in patient serum, highlighting its promise for clinical applications. Besides, traditional materials such as spinel NiFe₂O₄ magnetic nanoparticles, synthesized via the hydrothermal method, were employed as electrode material for CA125 detection [24]. The resulting immunosensor exhibited a low LOD (< 8.5 U/mL), good reproducibility, and strong selectivity against serum proteins and other biomarkers.

Recent advances have explored the use of natural product–derived materials as functional sensing platforms for tumor biomarkers. A notable example is the development of onion oil–based organic–inorganic hybrids (ONOHs) for CA-125 detection in serum [25]. These hybrids, characterized by FT-IR and SEM, integrate onion oil within a porous 3D network, which significantly enhances electrocatalytic activity and reduces charge transfer resistance. Among the reported systems, ONOH-3 exhibited the best electrochemical performance, achieving a low detection limit (0.805 μ U/mL), broad linear ranges (0.5–10 and 10–300 ng/mL), and high selectivity against potential interferents in serum. The improved sensitivity and stability of ONOH-3 highlight the promise of onion oil–based hybrids as cost-effective, biocompatible sensing materials with strong potential for clinical applications in ovarian cancer diagnostics.

A recent strategy for tumor marker detection combined rGO-TEPA/ZIF-67@ZIF-8/Au substrates (ZIF= Zeolitic Imidazol Frameworks; rGO-TEPA= reduced graphene oxide-tetraethylenepentamine) with AuPdRu trimetallic nanoparticles to construct a sandwich-type electrochemical immunosensor for CA72-4 [26]. The system leverages core–shell ZIF nanoparticles, Au-NPs, and rGO-TEPA to achieve high catalytic activity, a wide linear range (0.0001–1000 U/mL), and low detection limits. Another example of an MOF platform employed for CA125 detection is the ratiometric electrochemical immunosensor based on 3DrGO/MWCNTs-Thi and UiO-66-Fc [27]. The 3D rGO/MWCNTs framework enhanced conductivity and Ab1 loading, while UiO-66-NH2 offered high surface area for Fc-COOH/Ab2 immobilization, boosting signal amplification.

Another example of graphene-derived nanomaterial proposed as an immunosensor is the nitrogen-doped graphene oxide decorated platinum cobalt (NGO-PtCo) -modified carbon fiber aptasensor for CA15-3, which showed a low LOD (4.1 × 10-2 U/mL), high sensitivity, and excellent specificity in human serum, with 92–100% recovery. Despite requiring further clinical validation and cost-optimization, it represents a promising platform for breast cancer diagnostics [28]. Regarding the detection of CA15-3, Oliveira and colleagues reported a novel MIP (molecularly imprinted polymers) prepared via SARA ATRP, enabling highly reproducible and sensitive detection, outperforming conventional radical polymerization. The biosensor achieved a wide linear range down to 0.001 U/mL (10× better than conventional methods). It showed excellent selectivity and reproducibility, attributed to the controlled synthesis of PAAm-co-PMBAm (copolymer of acrylamide and N,N'-methylenebisacrylamide) and improved stereochemical recognition at imprinted sites [29]. Moreover, Han et al. (2024) constructed a novel Co-N-C electrocatalyst with atomically dispersed Co sites for CA15-3 detection, achieving signal amplification through Co-N active centers and ascorbic acid-mediated coupling of enzymatic and electrochemical processes [30].

It is important to highlight some dual sensing performance, being the case of Srilikhit et al. [31] approach, where a fluidic dual carbon electrode (Flu-iDCE) platform was developed via stencil printing and laser cutting for the simultaneous detection of Carcinoembryonic antigen (CEA) and CA125. The AuNP-modified DCE exhibited enhanced electrochemical performance, with a higher active surface area and roughness compared to conventional DCE. Applied to human serum samples, the label-free immunosensor showed strong potential for point-of-care cancer screening [31]. Oliveira et al. [32] demonstrate the potential of paper-based printed electrochemical sensors for CA 15-3 determination in biological fluids (see Figure 2). The proposed voltammetric immunosensor, integrating AuNPs and anti-CA 15-3, achieved high sensitivity (0.012 μ A/U mL-1), low LOD (0.56 U/mL), and robust specificity, although with a 1 h response time. The approach highlights the advantages of printed sensors, low cost, real-time analysis, and easy miniaturization, making them promising alternatives for monitoring breast cancer biomarkers [33].

Finally, a pure, dense, and adhesive layer of conductive CuNSs on Whatman® filter paper using ion beam sputtering was developed by Naghib and colleagues [34]. The signal amplification was achieved through enzyme-mimicking catalysis, with Cu-PDA nanoparticles serving as both an antibody immobilization interface and an artificial enzyme for HQ oxidation. The proposed paper-based sensor exhibited a low LOQ (1.3 mU/mL) and wide DLR (5–280 U/mL), covering both healthy and patient ranges. Owing to the low cost, flexibility, and biocompatibility of paper, the device shows strong potential for CA15-3 point-of-care detection. However, being a signal-off system, it is more prone to noise compared to signal-on sensors, which typically offer higher sensitivity and lower LODs.

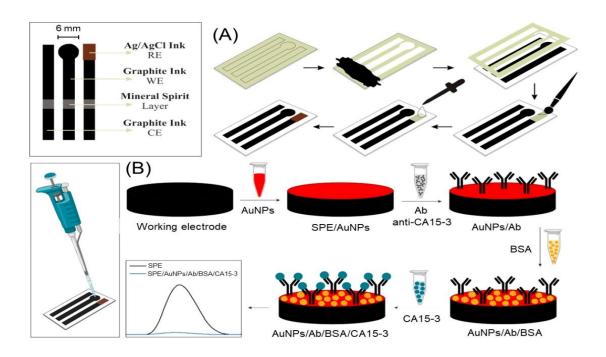


Figure 2. A) Fabrication and (B) modification of SPE with AuNPs, anti-CA 15-3, and BSA to develop an immunosensor for CA 15-3 determination. Reproduce from reference Oliveira et al. [32] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.3390/analytica5010005.

Table 1. Electrochemical biosensors for CA determination.

Nanomaterial	Biosensor type	Linear range	Detection limit	Sample	Ref.
SPCE/PTBDES/AuNPs	Immunosensor	5–100 pg/mL	1.20 pg/mL	Serum	[21]
ONOH-3	Immunosensor	0.5–10 and 10– 300 ng/mL	0.805 μU/mL	Serum	[25]
rGO-TEPA/ZIF-67@ZIF-8/Au	Immunosensor	0.001–1000 U/mL	1.8 × 10-5 U/mL	Serum	[26]
Ti ₃ C ₂ Tx/NH ₂ -CNT	Immunosensor	1–500 U/mL	1 μU/mL	Serum	[22]
NGO-PtCo	Aptasensor	0.05–200 U/mL	4.1 × 10-2 U/mL	Serum	[28]
3DrGO/MWCNTs-Thi)UiO-66- NH ₂ /ferrocenecarboxylic	Immunosensor	0.01–80 U/mL	0.089 U/mL	Serum	[27]
PAAm-co-PMBAm@MIP@3- MPA@Au-SPE	Immunosensor	0.01–100 U/mL	0.001 U/mL	Serum	[29]
ITO-Mn/ZnO-Au	Immunosensor	0.002–3 ng/mL	0.5 pg/mL	Serum	[23]
NiFe ₂ O ₄ NPs	Immunosensor	8.5–70 U/mL	8.5 U/mL	Serum	[24]
FluiDCE	FluiDCE Immunosensor		0.6 U/mL	Serum	[31]
Paper-based electrodes (f-SPE)	Immunosensor	2 - 16 U/mL	0.56 U/mL	Serum Sputum	[32]
CNTs	Immunosensor	0 - 300 U/mL	0.005 ng/mL	Serum	[33]
Ab/HQ/Cu-PDA/CuNSs/FP	Immunosensor	5 - 280 U/mL	1.3 mU/mL	Serum	[34]

Note: PTB/AuNPs: (PTB) (in deep eutectic solvent)/gold nanoparticles (AuNP); ONOH-3: Onion oil-based organic-inorganic hybrids; rGO-TEPA/ZIF-67@ZIF-8/Au: Zeolitic Imidazol Frameworks-67 doped with rGO-TEPA= reduced graphene oxide-tetraethylenepentamine anchored in AuPdRu trimetallic nanoparticles; Ti_3C_2Tx/NH_2 -CNT-SPCE: Ti_3C_2Tx-MX ene/amino-functionalized carbon nanotube (NH2-CNT) modified screen-printed carbon electrode (SPCE); ITO-Mn/ZnO-Au: Mn-doped ZnO nanorod anchored in ITO substrate;

NiFe₂O₄-NPs: NiFe₂O₄ magnetic nanoparticles; 3D-rGO/MWCNTs-Thi)UiO-66-NH₂/ferrocenecarboxylic: UiO-66-NH₂/ferrocenecarboxylic acid (UiO-66-Fc) doped with reduced graphene oxide/multi-walled carbon nanotube carboxylic acid- thionine; NGO-PtCo: nitrogen-doped graphene oxide decorated platinum cobalt nanoparticles; PAAm-co-PMBAm@MIP@3-MPA@Au-SPE: electrode based on gold surface (Au-SPE/3-MPA) doped with a copolymer of acrylamide and N,N'-methylenebisacrylamide (PAAm-co-PMBAm); CNTs: carbon-nanotubes; f-SPE: Screen-printed paper-based electrodes (f-SPE); FluiDCE: fluidic dual carbon electrode; Ab/HQ/Cu-PDA/CuNSs/FP: device based on cellulosic filter paper (FP) coated with Cu nanosponges (CuNSs) and then covered with Cu-doped polydopamine nanospheres (Cu-PDA) and grafted with hydroquinone (HQ).

3.2. CEA

In recent years, significant progress has been made in developing new biosensors for the detection of carcinoembryonic antigen (CEA), with a strong focus on improving sensitivity, stability, and suitability for point-of-care (POCT) applications. The research highlights the use of various advanced materials and innovative sensing mechanisms to achieve these goals.

Some studies have focused on developing highly sensitive electrochemical immunosensors for CEA detection. For instance, Cotchim et al. [35] introduced a novel label-free immunosensor using a nanocomposite of gold nanoparticles, molybdenum trioxide, and chitosan on a porous graphene-modified screen-printed electrode [35]. This dual-electrode sensor was designed for the simultaneous detection of both CEA and carbohydrate antigen 19-9 (CA19-9), a key biomarker for cholangiocarcinoma. The sensor demonstrated excellent performance with a low detection limit and was validated using real human serum samples. In another study, Ranjan et al. [36] developed a high-throughput electrochemical immunosensor utilizing an ionic liquid-functionalized molybdenum trioxide-reduced graphene oxide (MoO3-rGO-IL) nanocomposite. This sensor achieved a remarkably low detection limit for CEA and was also effective in analyzing real serum samples [36]. Similarly, Shamsazar et al. [37] fabricated an immunosensor by modifying a glassy carbon electrode with an iron oxide-multi-walled carbon nanotube composite, leading to an ultra-low detection limit and high stability [37]. In a different approach, Mehta et al. [38] developed a flexible, label-free immunosensor for CEA using nitrogen-rich mesoporous carbon as the substrate, which eliminated the need for coupling agents and simplified the fabrication process [38].

Innovative material synthesis and design have also played a crucial role. For example, Anbalagan et al. [39] created a cost-effective immunosensor using carbon dots synthesized from cow urine, which were bio-functionalized with an HRP-conjugated CEA antibody. This approach not only utilized animal waste but also resulted in a sensor with significantly higher sensitivity [39]. Meanwhile, Janduang et al. [40] synthesized flower-like zinc oxide nanoparticles. They combined them with graphene nanoplatelets to create a label-free immunosensor that could be integrated with a smartphone via NFC for POCT applications [40]. In a similar vein, Lei et al. [41] developed a homogeneous electrochemical sensor using carbon nanotube-bridged MXene electrode arrays integrated with a magnetic-bead-based immunoassay, which provided an ultra-low background signal for the highly sensitive detection of CEA [41].

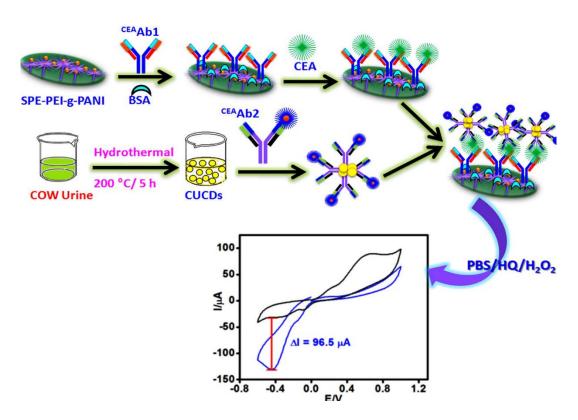


Figure 3. Construction of a sandwich immunosensor for CEA detection and signal amplification. Reproduce from reference Anbalagan et al. [39] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.1186/s11671-024-03980-3.

Several studies have explored aptasensors, which use aptamers instead of antibodies for biomarker recognition, often providing advantages in stability and cost. Erkal-Aytemur et al. [7] presented a quartz crystal microbalance (QCM) based aptasensor for CEA, demonstrating high selectivity and sensitivity suitable for clinical applications [7]. Rana et al. [42] introduced a hybrid of zirconia-gold nanoparticles (ZrO2-AuNPs) to enhance both the sensitivity and stability of their aptasensor for CEA detection, achieving a very low detection threshold and high diagnostic accuracy [42]. Yunussova et al. [43] developed a rapid, label-free aptasensor based on electrochemical impedance spectroscopy (EIS) for CEA, highlighting its potential for quick cancer screening [43]. Furthermore, Shi et al. [19] presented a smartphone-based electrochemical aptasensing platform with a unique dual-signal output strategy to minimize false positives, while Li et al. [44] utilized a hybrid structure of metal-organic frameworks and covalent organic frameworks to create an aptasensor with a remarkably low detection limit.

Finally, some researchers have focused on creating multi-signal or integrated platforms to enhance reliability and user-friendliness. Zhang et al. [45] detailed a dual-mode homogeneous biosensor for CEA that provided both electrochemical and colorimetric signals on a microfluidic paper-based analysis device (μ PAD), with an accompanying smartphone app for signal extraction. This dual-signal approach offers inherent self-verification, leading to more reliable results [45]. Zhou et al. [46] also developed a sensitive electrochemical immunosensor by integrating a biofunctionalized mesoporous silica nanochannel film with a carbon electrode, which effectively blocked the migration of a redox probe in the presence of CEA, leading to a measurable signal change [46].

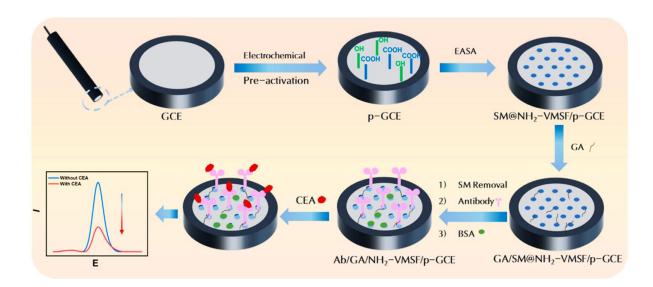


Figure 4. Schematic illustration of the construction of the immunosensor and the electrochemical detection of CEA. Reproduce from reference Zhou et al. [46] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.3390/molecules29040858.

Table 2. Electrochemical biosensors for CEA determination.

Nanomaterial	Biosensor type	Linear range	Detection limit	Sample	Ref.
Au-MoO₃-Chi	Immunosensor	0.001-0.01 ng mL ⁻¹	0.5 pg mL ⁻¹	Serum	[35]
4-mercaptophenyl /AuNPs	Aptasensor	0.5-10 ng mL ⁻¹	0.102 ng mL ⁻¹	Serum	[7]
Fc/PdPt@PCN-224	Aptasensor	1 pg mL ⁻¹ - 100 ng mL ⁻¹	0.27 pg mL ⁻¹	Serum	[19]
AuNPs	Immunosensor	0.1-40 ng mL ⁻¹	0.03 ng mL ⁻¹	Serum	[45]
Au@MoS2@rGO	Aptasensor	0.1 pg mL ⁻¹ - 100 ng mL ⁻¹	0.019 pg mL ⁻¹	Serum	[44]
MNC	Immunosensor	500 fM - 50 nM	500 fM	Serum	[38]
CDs	Immunosensor	0.5-50 ng mL ⁻¹	10 pg mL ⁻¹	Serum	[39]
Fe ₃ O ₄ /MWCNTs	Immunosensor	0.005 - 2.5 ng mL ⁻¹	0.3 pg mL ⁻¹	Serum	[37]
MoO3-rGO-IL	Immunosensor	25 fg mL ⁻¹ - 100 ng mL ⁻¹	1.19 fg mL ⁻¹	Serum	[36]
ZrO2-AuNPs	Aptasensor	0.01-10 ⁴ pg/mL	42.504 fg/mL	Serum	[42]
IDE	Aptasensor	0.002 ng/mL - 2 ng/mL	3.8 pg/mL	Serum	[43]
VMSF	Immunosensor	$0.01~{ m ng~mL^{-1}}$ - $100~{ m ng}$ ${ m mL^{-1}}$	6.3 pg mL ⁻¹	Serum	[46]
ZnONPs/GNPs	Immunosensor	0.5-10.0 ng mL ⁻¹	0.44 ng mL ⁻¹	Serum	[40]
MX@CNT	Immunosensor	0.005 - 1.0 ng mL ⁻¹	1.6 pg mL ⁻¹	Serum	[41]

Note: Au-MoO3-Chi: gold nanoparticles-molybdenum trioxide-chitosan; AuNPs: gold nanoparticles; FcPdPt@PCN-224: ferrocene PdPt nanoparticles@Zr-based porphyrinic metal-organic framework; Au@MoS2@rGO: gold@disulfide@graphene oxide; MNC: nitrogen-rich mesoporous carbon; CDs: carbon dots; Fe3O4: iron oxide nanoparticles; MWCNTs: multi-walled carbon nanotubes; MoO3-rGO-IL: ionic liquid-functionalized molybdenum trioxide-reduced graphene oxide; ZrO2-AuNPs: zirconia-gold nanoparticles; IDE: interdigitated gold electrode; VMSF: ordered mesoporous silica film; ZnONPs: flower-like zinc oxide nanoparticles; GNPs: graphene nanoplatelets; MX@CNT: carbon nanotube-bridged Ti3C2Tx MXene.

3.3. PSA

PSA is a protein marker that is effectively used for prostate cancer diagnosis at the early stage. The concentration of PSA in the serum of healthy individuals is below 4.0 ng mL⁻¹. On the other hand, an elevated PSA level within the range of 4.0-10.0 ng mL⁻¹ could indicate potential abnormality or early stages of prostate cancer. Moreover, PSA levels over 10 ng mL⁻¹ in patients' serum are indicative of prostate cancer development at various stages.

Yaiwong et al. [47] presented a novel dual-mode immunosensor that combines electrochemical (EC) and surface-enhanced Raman spectroscopy (SERS) techniques for the detection of the PSA biomarker. The sensor was constructed using a nanocomposite of gold nanoparticles (AuNPs) deposited on two-dimensional molybdenum disulfide (2D-MoS₂), which modifies a screen-printed carbon electrode (SPE). A primary antibody (Ab1) was immobilized on the modified electrode for the specific detection of the target PSA. Dual-signaling nanotags (TMB/Ab2/AuNPs) were then prepared by conjugating AuNPs with a secondary antibody (Ab2) and a probe molecule (TMB), which exhibit strong responses for both SERS and EC. The formation of a sandwich-type immunocomplex (Ab1-PSA-Ab2) in the presence of PSA enables detection with both techniques. The developed system demonstrated excellent selectivity and sensitivity, with low limits of detection (LOD) of 3.58 pg mL⁻¹ for the EC mode and 4.83 pg mL⁻¹ for the SERS mode, suggesting a high detection capability. In conclusion, the high sensitivity and efficacy of the sensor in analyzing biological samples make it a promising tool for the early diagnosis of cancer.

Also, Uruc et al. (2024) reported a novel label-free immunosensor that has the potential to be utilized for clinical screening and diagnosis of prostate cancer [48]. This sensor was based on combining 3,4-ethylenedioxythiophene (EDOT), 3-methylthiophene (3MT), and gold nanoparticles (AuNPs) applied to the sensitive determination of PSA. The immunosensor platform used [Fe(CN)₆] -3/-4 as a redox probe to obtain the signal, and the DPV technique was used to correlate the signal with the PSA concentration. The linear range was 0.1 to 50000 pg mL⁻¹, with a detection limit of 0.083 pg mL⁻¹. This immunoplatform can be easily converted into point-of-care detection devices due to its low cost, reproducibility, selectivity, and accuracy.

Moreover, Chellachamy Anbalagan et al. (2024) developed a novel electrochemical biosensor for the detection of PSA using a sensing platform based on carbon dots functionalized polyaniline (CDs@PANI) [49]. The sensor was constructed by covalently grafting biomass-derived CDs onto a PANI surface to form a biocompatible and fluorescent nanocomposite, CDs@PANI. This composite served as an immobilization matrix on a screen-printed carbon electrode (SPE). The detection mechanism was a sandwich electrochemical immunoassay, where a capture antibody (PSAAb1) was immobilized on the CDs@PANI platform. Following this, the PSA antigen and a horseradish peroxidase (HRP) conjugated detection antibody (PSAAb2) were sequentially introduced to form an immunocomplex. This sensor showed a broad linear detection range of 0.01–60 ng mL-1 and a low detection limit of 20 pg mL-1. The developed CDs@PANI-based biosensor showed superior analytical performance, including good sensitivity, specificity, stability, and reproducibility for PSA detection in real human serum samples.

In other work, Wang et al. (2024) presented a novel electrochemical sensing strategy for label-free dual-biomarker detection, utilizing a chronopotentiometric nanopore sensor modified with stimulus-responsive molecularly imprinted polymers (MIPs) [50]. The sensor's construction was based on a combination of nanopores and a polymeric membrane chronopotentiometric sensor. The solid-state nanopores, made from anodic aluminum oxide (AAO), were functionalized with two different stimulus-responsive MIPs to recognize two distinct biomarkers selectively. For detecting alpha-fetoprotein (AFP), a pH-sensitive MIP was created using 3-aminobenzeneboronic acid (APBA) as the functional monomer. For PSA detection, a temperature-sensitive MIP was synthesized using an N-isopropylacrylamide (NIPAAm) functionalized aptamer. The sensor demonstrated high sensitivity with low detection limits of 0.17 ng mL-1 for AFP and 0.42 ng mL-1 for PSA. The research demonstrates a significant step toward improving early disease diagnosis, particularly for cancers

like hepatocellular carcinoma and prostate cancer, by allowing for the simultaneous detection of multiple biomarkers from a single sample.

Redondo-Fernández et al. [51] reported an analytical, low-cost, smartphone-based system for the ultrasensitive detection of PSA. In this case, the system offers two detection alternatives: an electrochemical method using a portable potentiostat, and an optical method that uses a smartphone coupled with magnifying lenses. The sensor platform was an indium tin oxide-coated polyethylene terephthalate (ITO-PET) platform, which serves as a solid support for the immunoassay. The detection mechanism is a sandwich-type immunoassay where a specific anti-PSA antibody is immobilized on the ITO-PET surface. A second, anti-PSA antibody, tagged with gold nanoparticles (AuNPs), is then used for the recognition step. A key step for signal amplification is a controlled silver electrodeposition on the surface of the AuNPs, which significantly enhances their size from the nano to the micrometer scale. The proposed strategies exhibited very low limits of detection (LOD) of 102 fg mL-1 for electrochemical detection and 37 fg mL-1 for optical detection. This immunosensor also showed excellent selectivity and performance, capable of quantifying biomarkers at clinically relevant values without any pretreatment of the sample.

Ren et al. [52] showed an electrochemical peptide biosensor, 2FcP-GA-GDY(Fe)@NMIL-B, developed for the highly selective, ultrasensitive, and ultrastable detection of PSA [52]. The sensor's construction was based on a dual chemical bonding strategy to enhance both electron transport and peptide immobilization. The first chemical bond is a C-Fe-O interface created by incorporating high-conductivity Fe-Graphdiyne (Fe-GDY) into a metal-organic framework (MOF) material, NH2-MIL88B(Fe) (NMIL88B). This bond significantly accelerates electron transport. The second chemical bond is a Schiff-base (-N=C-) formed with glutaraldehyde (GA) as a crosslinking agent, which firmly links a ferrocene-labeled peptide (2FcP) to the electrode carrier at high density. When PSA is introduced, it cleaves the specific peptide, releasing ferrocene and leading to a decrease in the electrical signal, which enables sensitive detection. The study reports an ultra-low detection limit of only 0.94 fg mL-1. The sensor also showed an extended linear response range from 10 fg mL-1 to 50 ng mL-1. In conclusion, the 2FcP-GA-GDY(Fe)@NMIL-B sensor demonstrates superior analytical performance, including excellent selectivity and stability in human serum samples, making it a promising tool for clinical use in the early diagnosis of prostate cancer.

Also, Rahman et al. (2024) developed a novel, label-free impedimetric biosensor for the precise detection of tumor-associated biomarkers for prostate cancer [53]. This research used Maackia amurensis (MAA) lectin as a recognition element to identify cancer-associated aberrant glycosylation of PSA. This sensor was constructed by immobilizing MAA lectin onto gold-interdigitated microelectrodes. This was accomplished through a series of surface modifications, including the formation of a self-assembled monolayer using 11-mercaptoundecanoic acid (MUA) and the use of a crosslinking agent to activate carboxylic acid groups for lectin immobilization. The sensing mechanism was based on measuring the impedance response, which changes when the MAA lectin binds to the PSA-containing glycans. The sensor showed a low detection limit of 3.574 pg mL-1 and a concentration range of 0.01-100 ng mL-1. A crucial finding was that the MAA lectin preferentially recognizes α 2,3-linked sialic acid in serum PSA, which is a specific glycan known to be a valuable biomarker for improving the specificity of prostate cancer diagnoses. This biosensor offers a miniaturized and cost-effective platform for detecting PSA biomarker.

Other work reported an aptamer-based electrochemical biosensor for the diagnosis of prostate cancer, utilizing a novel two-dimensional (2D):2D reduced graphene oxide (rGO)/graphitic carbon nitride (g-C₃N₄) composite decorated with gold nanoparticles (Au NPs) [54]. This sensor was constructed by modifying a glassy carbon electrode (GCE) with the rGO/g-C₃N₄/AuNPs composite, upon which aptamer chains were immobilized. This device exhibited remarkable analytical performance, showing high selectivity toward PSA when compared to potential interfering substances like bovine serum albumin (BSA) and glucose. Under optimized conditions, the sensor achieved a rapid detection time of 30 min and a low limit of detection (LOD) of 0.44 fM (femtomolar).

The present method showed a significant advancement in early prostate cancer diagnosis with its validation using real serum samples, demonstrating its potential for clinical use.

On the other hand, Yue et al. [55] published an electrochemical biosensor for the ultrasensitive detection of PSA in human serum, using a platform based on α -Fe₂O₃/Fe₃O₄@Au nanocomposites and magnetically induced self-assembly (MISA) technology [55]. This sensor was constructed in two main steps. First, magnetic α -Fe₂O₃/Fe₃O₄ nanoparticles were synthesized, and then α -Fe₂O₃/Fe₃O₄@Au nanocomposites were prepared by reducing HAuCl₄ with NaBH₄. The self-assembly of the biosensor was then achieved by immobilizing a sulfhydryl-modified PSA aptamer onto the material surface via Au-S bonds. This was followed by the self-assembly of the magnetic nanoparticles onto a magnetic glassy carbon electrode (MGCE) using MISA technology. The developed biosensor demonstrated excellent analytical performance, with a linear response range from 100 fg mL-1 to 100 ng mL-1 and a low detection limit of 0.78 pg mL-1. This biosensor offered a valuable tool for clinical PSA detection, particularly for point-of-care testing (POCT). The present study confirms the sensor's excellent repeatability, stability, selectivity, and reproducibility, and its capability to analyze real samples, laying a strong foundation for future practical applications.

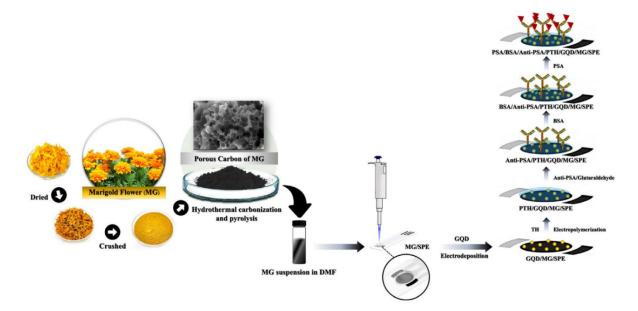


Figure 5. Fabrication process of the proposed label-free electrochemical immunosensor. Reproduce from reference Cotchim et al. [56] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.3390/bios14120589.

Moreover, Cotchim et al. [56] reported an electrochemical immunosensor for the detection of PSA, utilizing activated carbon from marigold flowers (MG) as a key component. The sensor was meticulously constructed using a series of modifications (Figure 5). First, activated carbon from marigold flowers (MG) was synthesized through hydrothermal carbonization and pyrolysis. This material was then modified with graphene quantum dots, which significantly enhanced electron transfer. Poly(thionine) (PTH) was subsequently grafted onto this modified surface, and the amine groups of PTH were used to bond with anti-prostate-specific antigen (Anti-PSA) via glutaraldehyde. This intricate layer-by-layer assembly resulted in a sensor with an improved electron transfer layer and a high affinity for the target biomarker. It exhibited two distinct linear detection ranges: 0.0125 to 1.0 ng mL⁻¹ and 1.0 to 80.0 ng mL⁻¹. The sensor's high sensitivity was evidenced by its low detection limit of 0.005 ng mL⁻¹ and a quantification limit of 0.017 ng mL⁻¹. In conclusion, this research successfully developed a highly sensitive and reliable unlabeled electrochemical immunosensor for PSA detection in serum samples.

Table 3. Electrochemical biosensors for PSA determination.

_						
	Nanomaterial Biosensor type		Linear range	Detection limit	Sample	Ref.
	AuNPs/2D-MoS ₂	Immunosensor	0-10 ng mL ⁻¹	3.58 pg mL ⁻¹	Serum	[47]
	AuNPs	Immunosensor	0.1-50000 pg mL ⁻¹	0.083 pg mL ⁻¹	Serum	[48]
	CDs@PANI	Immunosensor	0.01-60 ng mL ⁻¹	20 pg mL-1	Serum	[49]
	Nanopore/MIPs	Aptasensor	5-100 ng mL ⁻¹	0.42 ng mL ⁻¹	Serum	[50]
	AuNPs	Immunosensor	10-104 pg mL ⁻¹	102 fg mL ⁻¹	Serum	[51]
_	2FcP-GA- GDY(Fe)@NMIL-B	Peptide biosensor	10 fg mL-1-50 ng mL-1	0.94 fg mL ⁻¹	Serum	[52]
	AuNPs	Protein biosensor	0.01-100 ng mL ⁻¹	3.574 pg mL ⁻¹	Serum	[53]
	rGO/g-C3N4/AuNPs	Aptasensor	2.5–12.5 pM	0.44 fM	Serum	[54]
	α-Fe ₂ O ₃ /Fe ₃ O ₄ @Au	Aptasensor	100 fg mL ⁻¹ -100 ng mL ⁻¹	0.78 pg mL ⁻¹	Serum	[55]
	GQDs	Immunosensor	0.0125-1.0 ng mL ⁻¹	0.005 ng mL ⁻¹	Serum	[56]

Note: AuNPs/2D-MoS2: gold nanoparticles/two-dimensional molybdenum disulfide; CDs@PANI: carbon dots functionalized polyaniline; MIPs: molecularly imprinted polymers; 2FcP-GA-GDY(Fe)@NMIL-B: Fe-Graphdiyne into a metal-organic framework material NH2-MIL88B(Fe); rGO/g-C3N4: reduced graphene oxide/graphitic carbon nitride; α -Fe2O3/Fe3O4@Au: magnetic nanoparticles/gold nanoparticles; GQDs: Graphene quantum dots.

3.4. RNA, DNA, EVs y Cells

Electrochemical biosensors are rapidly emerging as a powerful tool in cancer diagnostics, offering sensitive, specific, and often portable platforms for detecting a range of crucial biomarkers, including microRNAs (miRNAs), exosomes, and tumor cells. These innovative approaches aim to overcome the limitations of traditional diagnostic methods, such as their complexity, high cost, and lengthy analysis times. Recent research highlights significant progress in this field, focusing on diverse biomarkers, often integrating novel nanomaterials and amplification strategies to achieve superior analytical performance.

3.4.1. miRNA Detection: A Focus on Early Cancer Biomarkers

MicroRNAs (miRNAs) are small, non-coding RNAs that play critical roles in post-transcriptional gene regulation and have emerged as valuable biomarkers for the early diagnosis and prognosis of various cancers. Their low abundance, short sequence length, and high sequence homology, however, make their detection particularly challenging. Recent advances in electrochemical biosensors, combined with nanomaterials such as gold nanoparticles, carbon-based structures, and metal oxides, have enabled highly sensitive and selective detection of miRNAs in biological samples.

Table 4 summarizes representative electrochemical strategies for miRNA detection, highlighting the diversity of nanomaterials employed, the type of biorecognition element (DNA probe, aptamer, or peptide nucleic acid), analytical performance, and cancer-related applications.

Table 4. Electrochemical biosensors for miRNA determination.

Nanomaterial	Biosensor type	Linear range	Detection limit	Sample	Ref.
AuNRs-AuNPs	Aptasensor	0.1 fM – 100 pM	5.10 aM/9.39 aM	Serum	[57]
PNA-DNA H3WJ	PNA	0.5 fM - 5 nM	0.15 fM	Serum	[58]
AZO nanostars	DNA	1 pM – 10 nM	3.98 pM	Breast cells	[59]
PER-CRISPR	DNA	10- ¹³ – 10- ⁷ M	30.2 fM	Serum	[60]

GO+G	DNA	10 fM – 1 nM	3.18 fM	Serum	[61]
AuNPs	DNA	0.1 – 1000 nM	1 nM	Serum	[62]
Au/MWCNT	DNA	0.001 – 10 nM	0.73 pM	Serum	[63]
AuNPs	DNA	1-100 nM	50 pM	Serum	[64]
Lentil lectin (LCA)-MB	DNA	1 fM - 10 pM	26 aM	Serum	[65]
IONPs	DNA	0.1 pM – 1μM	0.023 pM	Serum	[66]
AuNPs/MBs/COFs	DNA	10 fM - 5 nM	1.2 fM	Serum	[67]
PNA	PNA	0.1 fM – 0.1 nM	12.4 aM	Serum	[68]
AuNPs	DNA	~1 – 100 nM	0.4 nM	Serum	[69]
PNA-MXene (Ti ₃ C ₂ Tx)	PNA	100 aM – 10 nM	40 aM	Serum	[70]
PER-CRISPR	DNA	_	0.43 fM/0.12 fM	Serum	[71]
AuNPs	DNA	10 aM – 100 pM	5.24 aM	Plasma	[72]
GO	DNA	0.1 pM –10 nM	0.029 pM	Serum	[73]
LIG+PNA	PNA	100 aM – 100 nM	0.6 aM	Serum	[74]
PB-COFs nanospheres	DNA	10 fM – 100 nM	2.5 fM	Serum	[75]

Note: AuNPs: gold nanoparticles; AuNRs: gold nanorods; AZO: aluminum-doped zinc oxide; COFs: covalent organic frameworks; G: Graphene; GO: Graphene oxide; H3WJ: hetero-three-way junction; IONPs: Iron oxide nanoparticles; Au/MWCNT: gold/multiwall carbon nanotube; NAL: nucleic acid lock; PB: Prussian Blue; PER: primer exchange reaction; PNA: peptide nucleic acids.

The miRNAs detection is a significant area of research due to their role as critical cancer biomarkers. Several recent studies highlight the development of highly sensitive electrochemical aptasensors and biosensors for miRNA detection. For instance, a novel dual-targeted electrochemical aptasensor utilizes gold nanorods (AuNRs) and aptamer-functionalized gold nanoparticles (AuNPs) on screen-printed carbon electrodes (SPCEs) to simultaneously detect neuroblastoma-associated miRNA-181 and miRNA-184 with remarkable limits of detection (LODs) of 5.10 aM and 9.39 aM, respectively [57]. This amplification-free aptasensor offers a user-friendly and easily engineered platform for early cancer diagnosis and screening.

Another innovative electrochemical biosensor focuses on microRNA-21 (miRNA-21). This biosensor integrates peptide nucleic acid (PNA)-DNA hetero-three-way junction (H3WJ) formation with enzyme-free target-recycling catalytic hairpin assembly (CHA) amplification, achieving an exceptionally low detection limit of 0.15 fM for miRNA-21 [58]. The PNA-DNA H3WJ structure enhances stability and sensitivity, and the biosensor demonstrated excellent reproducibility and long-term stability, proving its utility in human cancer cells. Similarly, aluminum-doped zinc oxide (AZO) nanostars have been employed as a novel electrode modification material to enhance the conductivity and surface area of an electrochemical biosensor for highly sensitive miRNA-21 detection [59]. The unique morphology and high electron transfer efficiency of AZO nanostars contribute to a remarkably low detection limit and a wide linear range for miRNA-21 quantification.

Further advancements in miRNA detection include a CRISPR-empowered electrochemical biosensor, PER-E-CRISPR, for sensitive and target amplification-free detection of miRNA-21 [60]. This biosensor leverages CRISPR/Cas13a and primer exchange reaction (PER) for dual-signal amplification, achieving a low LOD of 30.2 fM without the need for pre-amplification. The practicality of screen-printed electrodes (SPEs) for miRNA detection in liquid biopsy, particularly for lung cancer management, has also been explored, demonstrating their potential for decentralized and user-friendly diagnostic solutions using both commercial and hand-made SPEs [62].

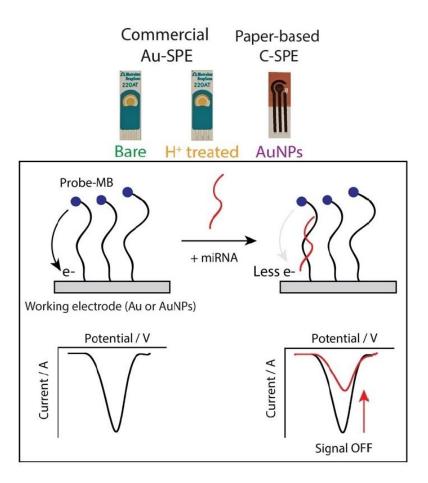


Figure 6. In the upper part of the figure, the three types of SPEs used for this study are reported, namely commercial bare Au-SPE (left), acidic-treated commercial Au-SPE (center), and AuNP-modified paper-based carbon SPE. In the lower part, the mechanism of electrochemical sensing of the presence of miRNA is reported through the development of a signal OFF platform, which corresponds to a decrease in current when the concentration of the target increases. Reproduce from reference Raucci et al. [62] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.1002/open.202300203.

For non-small cell lung cancer (NSCLC), an electrochemical biosensor employing a gold/multiwall carbon nanotube (Au/MWCNT) nanocomposite-based sandwich platform has been developed for the highly sensitive detection of microRNA-223 (miR-223) [63]. This platform offers enhanced sensitivity and selectivity without complex sample preparation. Moreover, a paper-based strategy has been introduced to enhance the sensitivity of electrochemical miRNA detection, focusing on miR-224, a lung cancer biomarker [64]. This approach utilizes an external chromatographic wax-patterned paper-based disk to preconcentrate the sample, significantly lowering the detection limit and offering a more affordable and sustainable method for cancer biomarker detection.

Another notable development for lung adenocarcinoma (LUAD) diagnosis involves an electrochemical biosensor for fucosylated extracellular vesicles (EVs) containing miR-4732-5p [65]. This biosensor employs a dual amplification strategy combining a Mg²⁺-dependent DNAzyme splitting nucleic acid lock (NAL) cycle and hybridization chain reaction (HCR) to achieve high sensitivity.

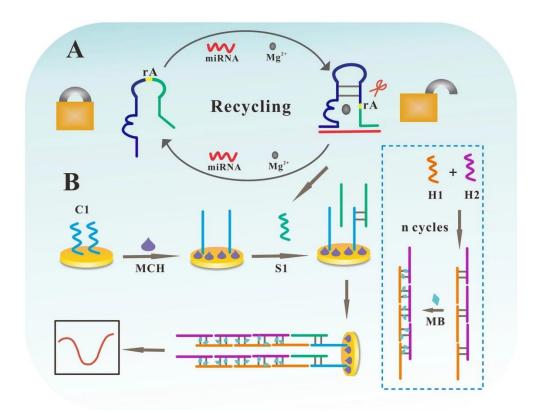


Figure 7. The principle of the electrochemical biosensor for the detection of extracellular vesicle miRNAs. (A) The target miRNA keeps recycling to bind to NAL to release more strands. (B) Strand recognition and signal amplification on the electrode for electrochemical measurements. Reproduce from reference Zhu et al. [65] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.1038/s41598-024-61060-z.

Further advancements in miRNA detection include a paper-based electrochemical device for detecting miRNA-652, a biomarker associated with Triple-Negative Breast Cancer (TNBC) [69]. This device, featuring an AuNP-modified office paper-based screen-printed electrode, demonstrated satisfactory repeatability and selectivity, with an additional 10-fold improvement in detection limit achieved through pre-concentration. Addressing the detection of other crucial miRNAs, Dulgerbaki and Oksuz (2024) presented a label-free electrochemical biosensor for miRNA-21 utilizing a glassy carbon electrode modified with a nanocomposite of iron oxide nanoparticles and PEDOT: PSS, exhibiting high sensitivity and selectivity [66]. Expanding on amplification methods, Yu et al. (2024) developed a photo-induced electron/energy transfer reversible addition-fragmentation chain transfer polymerization (PET-RAFT) signal amplification electrochemical biosensor for miRNA-21 detection, achieving an exceptionally low detection limit of 12.4 aM [68]. Additionally, an electrochemical biosensor based on an atomic layered composite of graphene oxide (GO) and graphene (G) has been developed for selective and sensitive miRNA-21 detection, offering a broad linear detection range and a low LOD of 3.18 fM [61].

In a novel approach to amplification-free detection, Ali et al. [70] engineered a biosensing platform for the ultrasensitive and enzyme-free electrochemical detection of hsa-miR-141 using PNA-functionalized Ti3C2Tx MXene nanosheets, demonstrating an impressive detection limit of 40 aM without the need for complex amplification steps or nanomaterial labels [70]. Furthermore, to enable comprehensive diagnosis, Zhou et al. [71] established a novel electrochemical sensor, PER-CRISPR/Cas9-E, for the sensitive and simultaneous detection of dual miRNAs (miRNA-21 and miRNA-155), integrating primer exchange reaction (PER) for signal amplification with the precise cleavage capabilities of the CRISPR/Cas9 system [71]. Finally, Li et al. (2024) developed a highly sensitive and specific electrochemical biosensor for miRNA-155 detection by integrating magnetic

beads-assisted split DNAzyme cleavage with the assembly of functionalized covalent organic frameworks (COFs), achieving an ultralow detection limit of 1.2 fM through multiple signal amplification [67].

A significant stride in ultrasensitive detection of small extracellular vesicle (sEV)-derived miRNAs has been made with the development of a novel electrochemical biosensor utilizing a turbo-like localized catalytic hairpin assembly (T-CHA) strategy [72]. This biosensor employs a dual localization approach with gold nanoparticles (AuNPs) and sophisticated DNA nanotechnology, enabling robust signal amplification, with an impressive detection limit of 5.24 aM and proven efficacy in real clinical serum samples. Addressing the urgent need for improved early diagnosis of ovarian cancer, a "silent killer" often detected at late stages, another electrochemical nanobiosensor platform has been developed for the simultaneous and sensitive detection of multiple miRNAs [73]. This biosensor leverages carboxylated graphene oxide (GO-COOH) modified screen-printed carbon electrodes (SPCEs) to immobilize specific DNA probes for miR-200c and miR-141. The use of differential pulse voltammetry (DPV) yields linear ranges of 0.1 pM – 10 nM, with corresponding detection limits of 0.029 pM and 0.026 pM.

Beyond enhanced sensitivity and multiplexing, efforts are also focused on creating portable and user-friendly biosensors for POCT settings. A groundbreaking smartphone-interfaced electrochemical biosensor has been introduced for the ultrasensitive, rapid, and robust detection of miRNA cancer biomarkers [74]. This device addresses limitations of high costs and bulky equipment by using laser-induced graphene (LIG) as the sensing platform and a unique π - π stacking immobilization strategy for peptide nucleic acid (PNA) probes. Its single-step fabrication and functionalization, coupled with DPV detection using methylene blue as a redox indicator, achieved an ultra-low detection limit of 48.3 aM and a broad dynamic range.

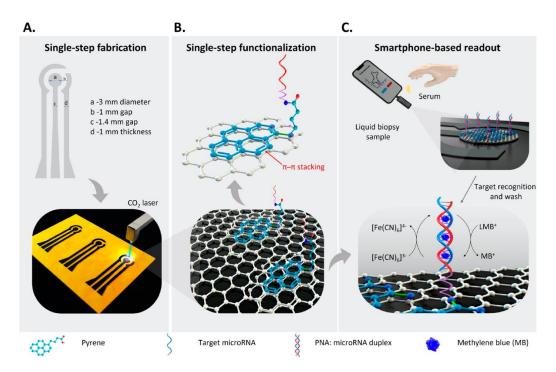


Figure 8. Schematic representation of the proposed PNA-functionalized LIG biosensor featuring (A) a single-step sensor fabrication based on laser burning of a PI film, (B) a single-step functionalization based on π - π stacking between pyrene-modified PNA probes and graphene, and (C) the electrochemical sensing mechanism based on PNA: miRNA duplex-bound MB redox reactions detected using a smartphone-based readout. Reproduce from reference Barman et al. [74] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.1021/acsmaterialslett.3c01225.

Further advancing the frontiers of miRNA detection, an innovative electrochemical biosensor for ultrasensitive detection of microRNA-182-5p (miRNA-182-5p) has been developed [75]. This

biosensor employs a novel strategy based on the target-driven cascade amplified assembly of COFs on a tetrahedral DNA nanostructure (TDN). COFs serve as efficient nanocarriers for electroactive Prussian blue (PB), providing a simple and effective signal reporter. The core of its amplification strategy integrates a chain hybridization amplification (CHA) reaction with a manganese (II)-powered DNAzyme system, ensuring remarkable signal enhancement. The TDN's multi-recognition domains also improve the capture efficiency of the PB-COFs, leading to an impressive detection limit of 2.5 fM for miRNA-182-5p.

3.4.2. Extracellular Vesicles Detection: Non-Invasive Biomarkers

Exosomes are nanoscale extracellular vesicles secreted by cells that carry proteins, lipids, and nucleic acids, reflecting the physiological and pathological state of their cells of origin. Because tumor-derived exosomes are abundant and detectable in body fluids such as serum and urine, they represent promising non-invasive biomarkers for cancer diagnosis and monitoring. Electrochemical biosensors, often coupled with nanomaterials and specific capture probes, have been developed to achieve sensitive and selective exosome detection. These platforms are particularly relevant since they address the limitations of conventional exosome assays (e.g., ultracentrifugation, which is time-consuming and requires specialized equipment). Table 5 presents selected examples that illustrate how nanomaterial-assisted biosensors can reduce assay time while maintaining clinical relevance.

Nanomaterial **Detection limit** Sample Ref. **Biosensor** type Linear range $3.63 \times 10^4 - 7.26 \times 10^8$ GOxS1 (DNAzyme) 3.63×10^4 particles/mL Plasma [76] Aptasensor particles/mL PbS CQDs 10² – 10⁸ particles/mL 19 particles/mL **Immunosensor** Serum [77] Exosomal **DNA** $1 \times 10^2 - 1 \times 10^8$ aM [78] MB-DNA probes 45 aM miRNA GO + AuNPs $500 - 1 \times 10^7 \, \text{exo/} \mu \text{L}$ [79] **Immunosensor** 110 exo/μL Serum **AuNPs** $10^5 - 10^{12} \text{ exo/mL}$ 8.7×10² exo/mL Urine [80] Immunosensor MCH - AuNPs $88 - 8.8 \times 10^7$ particles /µL 22 particles/µL Serum [81] Aptasensor

Table 5. Electrochemical biosensors for Extracellular Vesicles determination.

Note: AuNPs: gold nanoparticles; CQDs: colloidal quantum dots; GO: Graphene oxide; GOx-S1: glucose oxidase-substrate strand S1; MB-DNA: methylene blue - DNA; MCH: 6-mercapto-1-hexanol.

Electrochemical biosensors are proving effective for the detection of exosomes, which are vital non-invasive biomarkers for cancer. A homogeneous and portable electrochemical sensing platform has been developed for the accurate detection of breast cancer exosomes [81]. This platform utilizes a DNAzyme-induced DNA walker and enzyme-catalyzed amplification strategies, eliminating the need for complex washing steps and tedious electrode modifications, making it suitable for point-of-care diagnostics.

The sensitive detection of small extracellular vesicles (EVs), also known as exosomes, has been achieved using a colloidal quantum dots (CQDs)-modified electrochemical sensor [77]. This sensor, modified with a CD63 antibody and utilizing PbS CQDs, demonstrates rapid and highly sensitive electrochemical responses with a remarkably low detection limit of 19 particles per mL for sEVs, indicating its potential for membrane-biomarker-based diagnostics. Furthermore, a dual-amplified electrochemical sensing platform has been developed for the sensitive and accurate detection of exosomal microRNA-21 (miR-21) for breast cancer diagnosis [78]. This platform integrates ternary hybridization-based recognition with a perchlorate-assisted electrocatalytic cycle, offering a highly sensitive and specific method without the need for pre-amplification.

Exosomes, as crucial biomarkers for cancer diagnosis, have been a focal point for the development of electrochemical immunosensors. Sahraei et al. [79] introduced a flexible, paper-based electrochemical immunosensor (Exo-sensing paper) for direct detection of exosomes in serum

samples. This innovative system incorporates a 3D porous nanocomposite of nickel nanofoam with graphene oxide and gold nanoparticles, significantly enhancing the sensor's surface area and conductivity for increased antibody loading. Similarly, Sazaklioglu et al. [80] developed a lab-on-paper-based immunosensor for label-free exosome detection using electrochemical impedance spectroscopy (EIS), offering a portable and disposable solution for early cancer or metastasis detection. This sensor utilizes gold particles on a carbon working electrode conjugated with anti-CD9 antibodies, and the binding of exosomes causes a measurable impedance change [80]. Further pushing the boundaries of sensitivity, Zhang et al. [76] designed an electrochemical aptasensor for exosome detection based on a dual nucleic acid amplification strategy combining primer exchange reaction (PER) and rolling circle amplification (RCA). This innovative approach enables the detection of exosomes at remarkably low concentrations, showcasing the power of amplification techniques in overcoming the challenges of low biomarker abundance.

3.4.3. Tumor Cell and Circulating Tumor DNA Detection: Early Cancer Screening

Circulating tumor cells (CTCs) and circulating tumor DNA (ctDNA) provide direct information about cancer progression and genetic alterations, offering opportunities for minimally invasive "liquid biopsies." Electrochemical biosensors have been engineered to identify these biomarkers with high sensitivity and specificity, exploiting nanostructured electrodes and molecular recognition elements such as aptamers, antibodies, or DNA probes. These systems allow for the detection of extremely low concentrations of tumor-derived material in clinical samples, which is crucial for early diagnosis and disease monitoring. Table 6 summarizes representative electrochemical biosensors for CTCs and ctDNA detection, detailing their analytical performance and potential applications in cancer diagnostics.

Nanomaterial	Biosensor type	Linear range	Detection limit	Sample	Ref.
Fe-MOF	Cytosensor	10–10⁵ (EC)/150-10⁵ (CL)	3 cells/mL (EC)/ 10 cells/mL (CL)	HepG2 cells	[17]
AuNPs	Aptasensor	100-106 cells/mL	21 cells/mL	MUC1+ cells	[82]
Ni@MWNT	Immunosensor	10 ² -10 ⁷ cells/mL	2 cells/mL	Mouse liver tissue	[83]
rGO + IONPs	Immunosensor	10 ² -10 ⁶ cells/mL	5 cells/mL	SKBR3 cells	[84]
AuNPs	DNA	1 pM-50 pM	178 fM/216 fM	Mouse serum	[85]
(Au-Pt-Ag)-PDMS	DNA	2-200 fM	24.1 fM	Carcinoma cells	[86]
Au@Fe ₃ O ₄ NPs	DNA		~3 aM	Plasma	[87]

Table 6. Electrochemical biosensors for DNA/Tumor-Cell/ctDNA determination.

Note: AuNPs: gold nanoparticles; Fe-MOF: Metal-Organic Frameworks; IONPs: Iron oxide nanoparticles; Ni@MWNT: Ni-coated multi-walled carbon nanotube; PDMS: polydimethylsiloxane; rGO: reduced Graphene oxide.

The detection of tumor cells themselves is another critical aspect of early cancer screening and diagnosis. Tang et al. (2024) designed an integrated microfluidic cytosensor for the sensitive and accurate detection of hepatoma cells (HepG2 cells), a biomarker for hepatocellular carcinoma (HCC) [17]. This cytosensor employs a dual-site recognition and dual-mode signal readout strategy, combining electrochemical and colorimetric detection, providing a robust platform for circulating tumor cell (CTC) analysis in liquid biopsies. Additionally, a novel electrochemical method for the highly sensitive detection of MUC1-positive tumor cells utilizes a proximity labeling-assisted multiple signal amplification strategy [82]. This biosensor, which involves the formation of a hemin/G4-DNA complex to catalyze silver deposition, ensures high sensitivity in tumor cell detection, offering a promising tool for early cancer screening.

Yan et al. [83] introduced a highly sensitive cell-based electrochemical biosensor for hepatocellular carcinoma (HCC) detection, leveraging a single-chain variable fragment (scFv)

engineered from a recombinant humanized monoclonal antibody. By immobilizing the His-tagged scFv onto a nickel-based nanomaterial-modified electrode, the biosensor achieved enhanced binding affinity and specificity for HCC cells. The platform demonstrated a wide dynamic range (10²-107 cells mL-¹) and an exceptionally low detection limit of 2 cells mL-¹, with rapid detection within 5 minutes. Its ability to selectively identify HCC cells even in complex biological matrices, such as mouse liver tissue extracts, underscores its potential for early-stage cancer diagnostics and highlights the growing utility of antibody fragment-based biosensors in liquid biopsy applications.

Beyond exosomes and miRNAs, electrochemical biosensors are also being developed for the detection of other critical cancer biomarkers, such as circulating tumor DNA (ctDNA) and specific cancer cell lines. Yang et al. [85] introduced a novel electrochemical paper-based analytical device (ePAD) for the ultrasensitive detection of ctDNA from mice serum, enhanced by the integration of MBene as a novel material and a magnetic clutch probe for efficient pre-enrichment of target ctDNA. This ePAD exhibited an exceptionally low detection limit of 178-216 fM [85]. For the direct and pre-enrichment-free detection of hepatocellular carcinoma (HCC)-specific ctDNA, Arslan et al. (2024) developed a PDMS and MEMS-based microfluidic sensor, demonstrating promising limits of detection in various matrices, including human plasma and clinical samples [86]. Furthermore, for the detection of specific cancer cell lines, Hosseine et al. (2024) developed a label-free electrochemical biosensor for the ultrasensitive detection of the SKBR3 cell line, a key indicator of HER2 breast cancer [84]. This biosensor incorporates a sophisticated nanocomposite of green-synthesized reduced graphene oxide, iron oxide nanoparticles, nafion, and polyaniline, achieving a remarkably low detection limit of 5 cells mL-1.

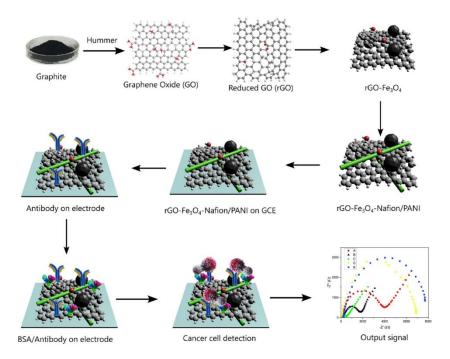


Figure 9. Schematic process diagram for the fabrication and functionalization of sandwich rGO/Fe₃O₄/Nafion/PANI for the detection of SK-BR3 cell line. Reproduce from reference Hosseine et al. [84] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.1038/s41598-024-62231-8.

The detection of circulating tumor DNA (ctDNA) through liquid biopsy is also gaining traction for non-invasive cancer diagnosis and monitoring. A novel method for ultrasensitive and rapid detection of ctDNA utilizes surface-confined gene amplification on dispersible magnetic nanoelectrodes [87]. This system integrates Fe₃O₄-Au core-shell nanoparticles directly into polymerase chain reactions (PCR), allowing these nanoparticles to function as "nano-electrodes" for in situ amplification and accumulation of target ctDNA. This technology achieves detection within one hour

and boasts an exceptional ultra-low detection limit of ~3 aM, making it highly sensitive to even minute quantities of ctDNA. Its successful validation in serum samples from breast cancer patients underscores its promise for earlier and more accurate cancer diagnosis, recurrence monitoring, and assessment of therapeutic efficacy.

3.5. Other Relevant Epithelial Biomarkers

The following paragraphs highlight significant progress in developing electrochemical biosensors for detecting various cancer biomarkers. While each study focuses on a specific cancer type and biomarker, these studies involve novel nanomaterials, innovative sensor architectures, and cost-effective fabrication methods.

EGFR, a type of transmembrane glycoprotein, is frequently found to be overexpressed or to have an aberrant expression in several solid tumors, such as those in the lung, breast, and glioblastoma. Its physiological functions include regulating the development and homeostasis of epithelial tissues. However, it also plays a significant role in several pathological processes, including the proliferation of tumor cells, angiogenesis, invasion, metastasis, and the prevention of apoptosis. Consequently, EGFR's characteristics make it a dependable diagnostic biomarker for cancers where it is involved, mainly lung and breast cancer.

Yue et al. [88] reported an electrochemical biosensing system for the sensitive detection of the EGFR biomarker. This system used magnetic nanocomposites and a magnetic-induced self-assembly (MISA) technique to enhance detection efficiency and enable point-of-care (POC) testing. The sensor was constructed using magnetic Fe₃O₄/ α -Fe₂O₃@Au nanocomposites as both signal amplifiers and the immobilization matrix for specific aptamer probes (ssDNA-APT). This sensor demonstrated a wide linear detection range from 0.1 to 1000 ng mL⁻¹ and a low limit of detection of 0.18 ng mL⁻¹. The sensor's ability to detect EGFR with high precision and its potential for real sample analysis demonstrate its value as a tool for early diagnosis and monitoring of EGFR-related cancers.

In other work, Zhang et al. [89] reported a bimodal biosensor for the detection of the EGFR L858R mutation, a key biomarker for specific non-small cell lung cancer (NSCLC) patients. The sensor's construction was based on a clever dual-amplification mechanism. When the target EGFR L858R mutation is present, it is resistant to digestion by the restriction enzyme Mscl. This resistance activates the first CRISPR-Cas12a protein, which inhibits the binding of nanomagnetic beads with fluorescein-labeled hybridization chain reaction (HCR) products, leading to a "signal-off" fluorescent response. Simultaneously, the activation also inhibits the cleavage activity of a second CRISPR-Cas12a protein, which allows the electrode to sustain a higher electrochemical signal from nanomaterials, creating a "signal-on" electrochemical response. This biosensor demonstrated exceptional analytical performance, with a dynamic detection range spanning from 10 fM to 1 μ M and a detection limit of 372 aM. This system also exhibits excellent specificity, reproducibility, stability, and recovery rates, representing a grand promise for early disease diagnosis and personalized therapeutic strategies for NSCLC patients.

Furthermore, Tang et al. (2024) developed an electrochemical immunosensor for the detection of EGFR [90]. This sensor was fabricated on a gold electrode surface that was first modified with SiO₂ nanospheres through a self-assembly process. Subsequently, the antibody was immobilized on the SiO₂ nanosphere-modified electrode to form an immune sandwich complex with the EGFR antigen and a biotin-labeled nucleic acid aptamer. Finally, an affinity-labeled alkaline phosphatase was immobilized on the electrode, and silver deposition was performed, which served as the signal amplification step. Experimental results showed good linearity for EGFR concentrations ranging from 1 to 1000 ng mL⁻¹ and demonstrated a very low detection limit of 0.06 ng mL⁻¹. In conclusion, the proposed electrochemical immunosensor offers a highly sensitive and stable method for detecting EGFR as an interesting tool for clinical diagnostics.

HER2 is a protooncogene that belongs to the cell surface receptor tyrosine kinase family, which is an important cancer biomarker in the biological behavior and pathogenesis of breast cancer. Liu et al. (2024) presented a novel nanosensor for the detection of breast cancer biomarkers, specifically

HER2, by combining a copper-based metal-organic framework (Cu-MOF) with magnetic beads [91]. The sensor's construction was based on a magnetic enrichment and separation strategy. The porous structure of the Cu-MOF was used to adsorb a large number of aptamers that are designed to capture the target antigens specifically. Subsequently, magnetic beads (MB), which are modified with the corresponding antibodies, are used to enrich and separate the Cu-MOF@antigen complexes. The detection range for the HER2 biomarker was found to be 4.5 fg mL⁻¹ - 20 ng mL⁻¹ with a detection limit of 1.51 fg mL⁻¹.

Furthermore, the sensor showed good performance in detecting biomarkers in real human serum samples, with high recovery rates ranging from 89.00% to 107.57%. These results underscore the sensor's high sensitivity and practical applicability. In conclusion, the developed nanosensor provides a promising solution for the detection of breast cancer biomarkers.

Also, Ai et al. [8] developed a novel photoelectrochemical (PEC) aptasensor for the ultrasensitive detection of the biomarker HER-2. The sensor's construction involved several key components. The photoactive Z-scheme UiO-66/CdIn2S4 heterojunction was synthesized using a hydrothermal method. This material serves as the photoanode and is designed to improve photoelectric conversion efficiency by preventing the recombination of photo-generated electron-hole pairs. The second component is a flower-like PtPdCu nanozyme, fabricated via a wet-chemical method, which acts as a peroxidase-mimicking catalyst. The aptasensor is completed by immobilizing capture DNA and the PtPdCu nanozyme-modified aptamer onto the heterojunction-coated electrode. This leads to a broad linear detection range from 0.1 pg mL $^{-1}$ to 0.1 μ g mL $^{-1}$. The sensor also achieved an impressively low limit of detection of 0.07 pg mL $^{-1}$, confirming its high sensitivity. This research successfully developed a new and highly sensitive PEC aptasensor for the quantitative determination of HER-2 in human serum samples.

Moreover, Zhang et al. [92] developed an innovative electrochemical biosensor for the sensitive detection of HER2. This sensor was constructed using a combination of an aptamer (Apt), peptide nucleic acid (PNA), and magnetic Fe₃O₄/ α -Fe₂O₃ heterogeneous nanorods. The mechanism involves Apt capturing the large HER2 protein, which simultaneously releases single-stranded DNA (ssDNA) chains from a double-stranded DNA (dsDNA) complex. The PNA then captures these released ssDNA chains, which converts the change in the electrochemical signal from a steric hindrance effect to one caused by the number of ssDNA strands, effectively extending the detection range. The use of magnetic Fe₃O₄/ α -Fe₂O₃ nanorods combined with Apt and PNA resulted in an ultra-low detection limit (LOD) of 4.1 fg mL⁻¹. The sensor also showed a wide detection range, from 10 fg mL⁻¹ to 5×10⁶ fg mL⁻¹. The experimental results confirmed that the biosensor has excellent selectivity, reproducibility, and storage stability. Furthermore, the analysis of spiked serum samples demonstrated a high recovery rate of 95.9-115.7%, indicating its great promise for use in real serum samples. In conclusion, this research successfully developed a highly sensitive and reliable electrochemical biosensor for HER2 detection.

CLDN18.2 is a member of the claudin protein family. This biomarker represents a promising target for gastric cancer (GC) immunotherapy. While it is typically expressed in normal gastric mucosal cells, its overexpression is a hallmark of tumor growth and metastasis. A study presented an electrochemical immunosensor applied to GC diagnosis by developing an electrochemical immunosensor for detecting the CLDN18.2 biomarker [93]. The study conducted a comparative analysis to determine the best material for the biosensor's construction, exploring the deposition of polymelamine (PM) on a variety of carbon nanomaterial-based screen-printed electrodes, including carbon (C), graphene oxide (GO), graphene (Gr), and carbon nanotubes (CNT). The results demonstrated that graphene and carbon nanotubes are superior substrates for the polymerization of melamine, showing more significant and stable redox behavior compared to C and GO. The immunosensors were fabricated on the PM-modified Gr and CNT platforms, which showed outstanding analytical performance. This linear response was observed from concentrations of 0.1–100 ng mL-1 in the case of PM-Gr/SPE immunosensor, and from 0.01 ng mL-1 to 100 ng mL-1 in PM-CNT/SPE immunosensor. Limits of detection were achieved at 7.9 pg mL-1 and 0.104 ng mL-1 for the

CNT and Gr-based sensors, respectively, highlighting the platform's high sensitivity. This superior performance is attributed to the enhanced and stable redox activity of PM on the Gr and CNT electrodes, which acts as an in-situ signal transducer. This study not only introduced the electrochemical immunosensor for the CLDN18.2 protein but also established a robust methodology for the selection and modification of carbon nanomaterials.

PG is a relevant biomarker in the digestive process and has become a valuable resource in biochemical diagnosis of GC. PG exists in two immunogenic variants: PG I, secreted by the mucous cells of the gastric fundus, and PG II, which emanates mainly from the mucous cells of the cardiac and pyloric glands of the gastric antrum. Empirical evidence has shown that individuals manifesting early symptoms of GC often present elevated levels of serological biomarkers, particularly PG I, with concentrations around 70.95 µg L⁻¹, along with a mixed PG I/II ratio of approximately 2.99 µg L⁻¹. In this sense, Kanagavalli and Eissa (2024) reported a redox probe-free electrochemical immunosensor for the detection of the PG I biomarker [94]. The use of a polymelamine (PM) electrodeposited on a reduced graphene oxide (rGO) as an in-situ active substrate distinguishes this study from other methods that rely on external redox probes, which simplifies the detection process and makes it more suitable for point-of-care (POC) diagnostics. The electrochemical reduction of graphene oxide (GO) on the screen-printed electrode (SPE) generated a defect-rich surface favorable for the deposition of polymelamine (PM). PG I detection was based on monitoring changes in the PM current signal. The formation of the immune complex between the immobilized anti-PG I antibody and the PG I antigen on the electrode surface caused a significant decrease in the differential pulse voltammetry (DPV) peak current. The biosensor demonstrated a clinically relevant linear concentration range of 0.01 to 200 ng mL⁻¹, with a low limit of detection of 9.1 pg mL⁻¹. This work not only presented the development of a label-free, redox probe-free electrochemical immunosensor for the GC biomarker PG I but also established a robust and promising platform for future clinical applications.

Other work presented an integrated, portable, dual-mode electrochemical immunosensing platform capable of simultaneously detecting two GC biomarkers: PG I and PG II [95]. This platform utilizes screen-printed electrodes (SPEs) with dual working electrodes, each tailored for the detection of a specific biomarker. The electrode surface is coated with nanocomposites of reduced graphene oxide (rGO), tetraethylene pentaamine (TEPA), iron oxide (Fe₃O₄) nanoparticles, and bimetallic AuPt nanoparticles (AuPt NPs). The system provides linear detection ranges from 5 pg mL-1 to 100 ng mL-1 for PG I and from 50 pg mL-1 to 200 ng mL-1 for PG II. The limit of detection was established at 1.67 pg mL-1 for PG I and 16.67 pg mL-1 for PG II. This interesting immunosensor showed minimal cross-interference between the two working electrodes, confirming its ability to perform accurate and simultaneous quantitative measurements for GC diagnosis.

NSE is a multifunctional protein that mainly exists in nerve tissue and neuroendocrine tissue, and it is one of the important biomarkers for the diagnosis of small-cell lung cancer (SCLC). Chen et al. (2024) developed an ultrasensitive photoelectrochemical (PEC) immunosensor for the NSE determination [96]. The core element of this biosensor is a novel heterojunction of cadmium indium sulfide and magnesium indium sulfide (CdIn₂S₄/MgIn₂S₄, CMIS). The mechanism of the immunosensor is a "signal-off" system. A capture antibody (Ab1) was immobilized on the surface of the electrode modified with CMIS and platinum nanoparticles (Pt NPs). The presence of the target NSE led to the formation of a sandwich complex with a detection antibody (Ab2) attached to iron oxide nanoparticles (Fe₃O₄). The Fe₃O₄-Ab2 acts as a signal quencher, competitively consuming electron donors and absorbing light, which results in a decrease in the photocurrent. This strategy enabled the ultrasensitive detection of NSE over a wide range, from 1.0 fg mL⁻¹ to 10 ng mL⁻¹, with an exceptionally low detection limit of 0.34 fg mL⁻¹. This work provides a new perspective for the design of highly sensitive PEC immunosensors for SCLC diagnosis.

The field is rapidly advancing with new materials and designs. For liver cancer, two different approaches for detecting alpha-fetoprotein (AFP) illustrate how material choice directly impacts performance. A nanocomposite of few-layer graphene (FLG) and 3-polythiophene acetic acid (3-PTAA) provided a cost-effective and stable platform with a low detection limit of 0.047 pg/mL [12].

In contrast, hollow PtCoNi bunched nanochains significantly boosted electrocatalytic activity, achieving an even lower detection limit of 0.017 pg/mL and improved long-term stability [97]. This contrast underscores the trade-off between affordability and maximum sensitivity. This comparison illustrates how different nanomaterials can be tailored for enhanced performance. Since the clinical reference cut-off for AFP is ~20 ng/mL, these detection limits in the pg/mL range are several orders of magnitude more sensitive than required, ensuring applicability for early diagnosis.

For ovarian cancer, two distinct biomarkers, HE4 and LPA, have been targeted using different strategies. The disposable HE4 biosensor on a flexible ITO-PET sheet emphasizes affordability and stability for early diagnosis [16]. In contrast, the label-free LPA biosensor, based on a gelsolin–actin complex, simplifies fabrication and aligns better with point-of-care testing [98]. Together, these examples highlight the dual focus of current research: ensuring both cost-effectiveness and clinical practicality. Importantly, the clinical threshold for HE4 is approximately 70 pmol/L (~140 pM), and reported biosensor sensitivities fall well below this cut-off, supporting their clinical applicability.

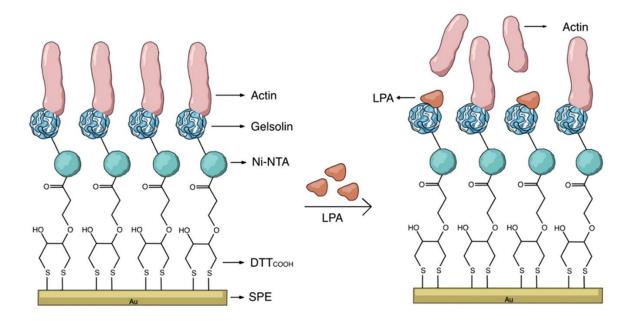


Figure 10. Schematic representation of the label-free electrochemical biosensor for LPA detection. Reproduce from reference Ivanova et al. [98] under the terms and conditions of the Creative Commons Attribution (CC BY) license. https://doi.org/10.3390/electrochem5020015.

For colorectal cancer, a dual immunoplatform on magnetic microsupports was developed to simultaneously detect TIMP-1 and GDF-15, providing a fast and reliable method for analyzing complex clinical samples [11]. Finally, for non-small cell lung cancer, an immunosensor for the CYFRA21-1 antigen utilizes a three-dimensional ordered macroporous carbon framework (3D MCF) with gold-cobalt nanoparticles. This architecture maximizes surface area and electron transport, resulting in a highly sensitive and stable platform suitable for human serum analysis [99]. Considering that the clinical cut-off value for CYFRA21-1 is around 3–3.5 ng/mL, the reported biosensor performance, with detection limits below this threshold, confirms its clinical relevance.

Table 7. Electrochemical biosensors for the determination of other epithelial biomarkers.

Nanomaterial	Biosensor type	Linear range	Detection limit	Sample	Ref.
Fe ₃ O ₄ /α-Fe ₂ O ₃ @Au	Aptasensor	0.1-1000 ng mL ⁻¹	0.18 ng mL ⁻¹	Serum	[88]
Magnetic NPs	Aptasensor	10 fM-1 μM	372 aM	Serum	[89]
SiO ₂ NPs	Immunosensor	1-1000 ng mL ⁻¹	0.06 ng mL ⁻¹	Serum	[90]

Fe ₃ O ₄ /α-Fe ₂ O ₃	Aptasensor	10 fg mL ⁻¹ - 5×10 ⁶ fg mL ⁻¹	4.1 fg mL ⁻¹	Serum	[92]
Gr/CNT	Immunosensor	0.1 – 100 ng mL $^{\text{-}1}$ and 0.01 ng mL $^{\text{-}1}$ - 100 ng mL $^{\text{-}1}$	$7.9~{ m pg~mL^{-1}}$ for CNT and $0.104~{ m ng~mL^{-1}}$ for Gr	Serum	[93]
GO	Immunosensor	0.01 - 200 ng mL ⁻¹	9.1 pg mL ⁻¹	Serum	[94]
rGO/Fe3O4NPs/AuPtNPs	Immunosensor	5 pg mL ⁻¹ - 100 ng mL ⁻¹ for PG I and 50 pg mL ⁻¹ - 200 ng mL ⁻¹ for PG II	$1.67~{ m pg~mL^{-1}}$ for PG I and $16.67~{ m pg~mL^{-1}}$ for PG II	Serum	[95]
PtNPs/Fe ₃ O ₄ NPs	Immunosensor	1.0 fg mL ⁻¹ - 10 ng mL ⁻¹	0.34 fg mL ⁻¹	Serum	[96]
3-PTAA/FLG	Immunosensor	0.0001–250 ng/mL	0.047 pg/mL	Serum	[12]
GH-PtCoNi BNCs	Immunosensor	0.1 to 10⁵ pg mL-¹	0.017 pg mL ⁻¹	Serum	[97]
ITO-PET/3-APTES	Immunosensor	1 pg/mL to 3000 pg/mL	0.094 pg/mL	Serun	n [16]
DTTCOOH	Biosensor	0.01–10 μΜ	0.9 μΜ	Serum	[98]
HOOC-MBs	Immunosensor	43.4–2500 pg mL ⁻¹	13 pg mL ⁻¹	Plasma	[11]
Au/CoNPs-3D MCF	Immunosensor	0.0001–100 ng/mL	0.0224 pg/mL	Serum	[99]

Note: Fe_3O_4/α - Fe_2O_3 : Magnetic nanoparticles; Au: Gold nanoparticles; NPs: nanoparticles. MOF: metal-organic framework; Gr: graphene; CNT: carbon nanotubes; GO: graphene oxide; rGO: reduced graphene oxide; 3-PTAA: 3-polythiophene acetic acid; FLG: few-layer graphene; GH-PtCoNi BNCs: gourd-shaped hollow PtCoNi bunched nanochains; ITO-PET: indium tin oxide-polyethylene terephthalate sheet; 3-APTES: 3-Aminopropyl trimethoxy silane; DTTCOOH: 3-dithiothreitol propanoic acid; HOOC-MBs: magnetic beads; Au/CoNPs-3D MCF: macroporous carbon skeleton material modified with gold-cobalt nanoparticles.

4. Conclusions

Epithelial cancer remains a significant global health concern, with early diagnosis being crucial for improving treatment outcomes and survival rates. Traditional methods like ELISA have notable limitations, but innovative electrochemical biosensors are emerging as a promising solution. These devices offer high sensitivity, specificity, and rapid analysis, making them ideal for cancer applications. We have already seen the successful development of biosensors for numerous biomarkers, including CA125, CEA, and PSA, achieving low detection limits suitable for clinical use. Furthermore, nucleic acid electrochemical sensors, aptasensors, and other sophisticated designs are being developed for the sensitive detection of cells and extracellular vesicles (EVs).

In this review, we have showcased the successful development of advanced electrochemical biosensors based on nanomaterials for determining epithelial cancer biomarkers in patient samples. These sensors are excellent candidates for clinical use, particularly for point-of-care testing. Their ability to enable early detection of primary tumors, metastases, and recurrent diseases contributes to more effective cancer management. The integration of biosensor technology with microfluidic devices has led to the creation of low-cost, adaptable, and efficient diagnostic platforms. These platforms are gaining significant attention due to their selectivity, affordability, and ease of result interpretation. We believe that the biggest challenge lies in developing biosensors capable of simultaneously detecting multiple biomarkers in a single measurement, a goal that the synergy between analytical chemistry and nanotechnology is poised to achieve.

5. Future Perspectives

The future of electrochemical biosensors in cancer diagnostics is auspicious. Future innovations will continue to focus on signal amplification, selective capture, multi-detection, and applications in real-world clinical samples. The integration of novel nanomaterials, sophisticated amplification strategies, and diverse sensing platforms, including paper-based and microfluidic devices, is leading to the development of highly sensitive, selective, and user-friendly tools for early cancer detection and monitoring. These advancements are essential for transitioning toward less invasive, more affordable, and widely accessible diagnostic solutions. Electrochemical biosensors have the immense potential to revolutionize early cancer detection, establish new diagnostic and prognostic platforms, and advance personalized healthcare. Looking ahead, the integration of electrochemical biosensors with emerging technologies holds considerable promise. The combination with microfluidic platforms can enable fully automated lab-on-a-chip devices for rapid, multiplexed analysis. At the same time, portable and even wearable formats may extend cancer diagnostics to point-of-care and continuous monitoring scenarios. Moreover, coupling biosensor readouts with artificial intelligence and machine learning approaches could facilitate real-time data interpretation, improve accuracy, and identify complex biomarker patterns. In the long term, integrating biosensors with omics technologies and digital health systems may pave the way toward personalized cancer diagnostics and precision medicine.

Ultimately, beyond the technological advances summarized in this review, the translational pathway for electrochemical biosensors for epithelial cancer diagnosis still faces critical challenges. Clinical validation in large and diverse patient cohorts is necessary to confirm reliability beyond proof-of-concept studies. Equally important is the reproducibility of results across different laboratories and the establishment of standardized protocols for sample preparation and sensor operation. From a manufacturing perspective, scaling up nanomaterial-based platforms to cost-effective and robust devices remains a challenge. Finally, regulatory approval processes (e.g., FDA, EMA) impose strict requirements for accuracy, stability, and clinical performance before biosensors can be adopted as in vitro diagnostic (IVD) tools. Addressing these challenges is essential to translate current innovations into clinically approved diagnostic devices with a real impact on patient care.

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References

 Malecka-Baturo, K.; Grabowska, I. Efficiency of Electrochemical Immuno- vs. Apta(Geno)Sensors for Multiple Cancer Biomarkers Detection. *Talanta* 2025, 281, doi: 10.1016/j.talanta.2024.126870.

- 2. Kiani, P.; Vatankhahan, H.; Zare-Hoseinabadi, A.; Ferdosi, F.; Ehtiati, S.; Heidari, P.; Dorostgou, Z.; Movahedpour, A.; Baktash, A.; Rajabivahid, M.; et al. Electrochemical Biosensors for Early Detection of Breast Cancer. *Clinica Chimica Acta* 2025, *564*, doi: 10.1016/j.cca.2024.119923.
- 3. Rouhi, S.; Ghasemi, H.; Alizadeh, M.; Movahedpour, A.; Vahedi, F.; Fattahi, M.; Aiiashi, S.; Khatami, S.H. MiRNA-Based Electrochemical Biosensors for Ovarian Cancer. *Clinica Chimica Acta* 2025, *564*, doi: 10.1016/j.cca.2024.119946.
- 4. Wang, Y.; Yao, W.; Sun, W.; Fan, Y.; Xiao, J.; Wang, T.; Huang, K.; Liu, L.; Wang, X.; Jiang, H. Aptamer-Based Electrochemical Analysis Platform for Tumor Cells and Biomarkers Detection. *Journal of Electroanalytical Chemistry* 2024, 960, doi:10.1016/j.jelechem.2024.118194.
- Ahmad, A.; Rabbani, G.; Zamzami, M.A.; Hosawi, S.; Baothman, O.A.; Altayeb, H.; Akhtar, M.S.N.; Ahmad, V.; Khan, M.V.; Khan, M.E.; et al. An Affordable Label-Free Ultrasensitive Immunosensor Based on Gold Nanoparticles Deposited on Glassy Carbon Electrode for the Transferrin Receptor Detection. *Int J Biol Macromol* 2024, 273, doi:10.1016/j.ijbiomac.2024.133083.
- 6. Fortunati, S.; Giannetto, M.; Pedrini, F.; Nikolaou, P.; Donofrio, G.; Bertucci, A.; Careri, M. A Novel Magnetic Ligand-Based Assay for the Electrochemical Determination of BRD4. *Talanta* 2024, 279, doi:10.1016/j.talanta.2024.126577.
- 7. Erkal-Aytemur, A.; Mülazımoğlu, İ.E.; Üstündağ, Z.; Caglayan, M.O. A Novel Aptasensor Platform for the Detection of Carcinoembryonic Antigen Using Quartz Crystal Microbalance. *Talanta* 2024, 277, doi:10.1016/j.talanta.2024.126376.
- 8. Ai, Q.Y.; Xu, B.F.; Xu, F.; Wang, A.J.; Mei, L.P.; Wu, L.; Song, P.; Feng, J.J. Dual Amplification for PEC Ultrasensitive Aptasensing of Biomarker HER-2 Based on Z-Scheme UiO-66/CdIn2S4 Heterojunction and Flower-like PtPdCu Nanozyme. *Talanta* 2024, 274, doi:10.1016/j.talanta.2024.126034.
- 9. Yuan, L.; Li, M.; Li, J.; Zhu, T.F.; Dong, M.; Liu, L. Aggregation-Induced Signal Amplification Strategy Based on Peptide Self-Assembly for Ultrasensitive Electrochemical Detection of Melanoma Biomarker. *Anal Chim Acta* 2024, 1289, doi:10.1016/j.aca.2024.342214.
- 10. Lakshmi Narayanan, M.; Prabhu, K.; Ponpandian, N.; Viswanathan, C. Cu Encrusted RF Sputtered ZnO Thin Film Based Electrochemical Immunosensor for Highly Sensitive Detection of IL-6 in Human Blood Serum. *Microchemical Journal* 2024, 199, doi:10.1016/j.microc.2024.110061.
- 11. Tejerina-Miranda, S.; Gamella, M.; Pedrero, M.; Montero-Calle, A.; Rejas, R.; Pingarrón, J.M.; Barderas, R.; Campuzano, S. Dual Immunoplatform to Assess Senescence Biomarkers TIMP-1 and GDF-15: Advancing in the Understanding of Colorectal Cancer. *Electrochim Acta* 2024, 503, doi:10.1016/j.electacta.2024.144822.
- Gangopadhyay, B.; Roy, A.; Paul, D.; Panda, S.; Das, B.; Karmakar, S.; Dutta, K.; Chattopadhyay, S.; Chattopadhyay, D. 3-Polythiophene Acetic Acid Nanosphere Anchored Few-Layer Graphene Nanocomposites for Label-Free Electrochemical Immunosensing of Liver Cancer Biomarker. ACS Appl Bio Mater 2024, 7, 485–497, doi:10.1021/acsabm.3c01126.
- 13. Verma, D.; Dubey, N.; Yadav, A.K.; Saraya, A.; Sharma, R.; Solanki, P.R. Disposable Paper-Based Screen-Printed Electrochemical Immunoplatform for Dual Detection of Esophageal Cancer Biomarkers in Patients' Serum Samples. *Mater Adv* 2024, *5*, 2153–2168, doi:10.1039/d3ma00438d.
- 14. Jarić, S.; Schobesberger, S.; Velicki, L.; Milovančev, A.; Nikolić, S.; Ertl, P.; Bobrinetskiy, I.; Knežević, N. Direct Electrochemical Reduction of Graphene Oxide Thin Film for Aptamer-Based Selective and Highly Sensitive Detection of Matrix Metalloproteinase 2. *Talanta* 2024, 274, doi:10.1016/j.talanta.2024.126079.
- 15. Mokni, M.; Tlili, A.; Khalij, Y.; Attia, G.; Zerrouki, C.; Hmida, W.; Othmane, A.; Bouslama, A.; Omezzine, A.; Fourati, N. Designing a Simple Electrochemical Genosensor for the Detection of Urinary PCA3, a Prostate Cancer Biomarker. *Micromachines* (*Basel*) 2024, 15, doi:10.3390/mi15050602.
- Vural, B.; Çalışkan, M.; Bilgi Kamaç, M.; Sezgintürk, M.K. A Disposable and Ultrasensitive Immunosensor for the Detection of HE4 in Human Serum Samples. *Chemical Papers* 2024, 78, 3871–3882, doi:10.1007/s11696-024-03359-9.
- 17. Tang, X.; Tang, J.; Zhang, Q.; Suonanmu, D.; Zhang, Y.; Ren, Q.; Tao, F.; Li, C.; Wang, F. An Integrated Microfluidic Cytosensor Coupled with Self-Calibration Electrochemical/Colorimetric Dual-Signal Output for Sensitive and Accurate Detection of Hepatoma Cells. *Sens Actuators B Chem* 2024, 403, doi:10.1016/j.snb.2023.135176.

- 18. Jin, Z.; Xiao, W.; Shen, L.; Shi, X.; Li, J. An Electrochemical Method Based on CRISPR-Cas12a and Enzymatic Reaction for the Highly Sensitive Detection of Tumor Marker MUC1 Mucin. *Analyst* 2024, 149, 3920–3927, doi:10.1039/d4an00595c.
- 19. Shi, S.S.; Li, X.J.; Ma, R.N.; Shang, L.; Zhang, W.; Zhao, H.Q.; Jia, L.P.; Wang, H.S. A Novel Dual-Signal Output Strategy for POCT of CEA Based on a Smartphone Electrochemical Aptasensing Platform. *Microchimica Acta* 2024, 191, doi:10.1007/s00604-024-06493-z.
- Quinchia, J.; Blázquez-García, M.; Torrente-Rodríguez, R.M.; Ruiz-Valdepeñas Montiel, V.; Serafín, V.; Rejas-González, R.; Montero-Calle, A.; Orozco, J.; Pingarrón, J.M.; Barderas, R.; et al. Disposable Electrochemical Immunoplatform to Shed Light on the Role of the Multifunctional Glycoprotein TIM-1 in Cancer Cells Invasion. *Talanta* 2024, 267, doi:10.1016/j.talanta.2023.125155.
- 21. Yılmaz, M.; Bilgi, M. A Disposable Impedimetric Immunosensor for the Analysis of CA125 in Human Serum Samples. *Biomed Microdevices* 2024, 26, doi:10.1007/s10544-023-00691-x.
- 22. Yang, M.; Wang, L.; Xie, C.; Lu, H.; Wang, J.; Li, Y.; Li, H.; Yang, J.; Zhang, T.; Liu, S. A Disposable Ultrasensitive Immunosensor Based on MXene/NH2-CNT Modified Screen-Printed Electrode for the Detection of Ovarian Cancer Antigen CA125. *Talanta* 2025, 281, doi:10.1016/j.talanta.2024.126893.
- 23. Guo, H. Application of a Novel and Effective Aptasensor for Electrochemical Monitoring of Cancer Antigen 125 in Human Serum. *Alexandria Engineering Journal* 2024, *109*, 11–17, doi:10.1016/j.aej.2024.08.084.
- 24. Maghiani, I.; Souza, L. V.; Bach-Toledo, L.; Faria, A.M.; Ortega, P.P.; Amoresi, R.A.C.; Simões, A.Z.; Mazon, T. Application of NiFe2O4 Nanoparticles towards the Detection of Ovarian Cancer Marker. *Mater Res Bull* 2024, 177, doi:10.1016/j.materresbull.2024.112835.
- 25. Er, O.F.; Kivrak, H.; Alpaslan, D.; Dudu, T.E. One-Step Electrochemical Sensing of CA-125 Using Onion Oil-Based Novel Organohydrogels as the Matrices. *ACS Omega* 2024, *9*, 17919–17930, doi:10.1021/acsomega.3c09149.
- 26. Yan, Q.; Wu, R.; Wang, J.; Zeng, T.; Yang, L. An Ultrasensitive Sandwich-Type Electrochemical Immunosensor Based on RGO-TEPA/ZIF67@ZIF8/Au and AuPdRu for the Detection of Tumor Markers CA72-4. *Bioelectrochemistry* 2024, 160, doi:10.1016/j.bioelechem.2024.108755.
- 27. Wu, F.; Gao, H.; Qiu, R.; Zhang, H.; Ren, X.; Qi, X.; Miao, M.; Rui, C.; Chang, D.; Pan, H. A Novel Ratiometric Electrochemical Immunosensor for the Detection of Cancer Antigen 125 Based on Three-Dimensional Carbon Nanomaterial and MOFs. *Microchemical Journal* 2024, 200, doi:10.1016/j.microc.2024.110372.
- 28. Tariq, A.; Bilal, S.; Naz, I.; Nawaz, M.H.; Andreescu, S.; Jubeen, F.; Arif, A.; Hayat, A. A Multifunctional N-GO/PtCo Nanocomposite Bridged Carbon Fiber Interface for the Electrochemical Aptasensing of CA15-3 Oncomarker. *Anal Biochem* 2024, 695, doi:10.1016/j.ab.2024.115640.
- 29. dos Santos Oliveira, D.; Oliveira, A.S.R.; Mendonça, P.V.; Coelho, J.F.J.; Moreira, F.T.C.; Sales, M.G.F. An Innovative Approach for Tailoring Molecularly Imprinted Polymers for Biosensors Application to Cancer Antigen 15-3. *Biosensors (Basel)* 2024, 14, doi:10.3390/bios14050222.
- 30. Han, L.; Cai, S.; Chen, X. Atomically Co-Dispersed Nitrogen-Doped Carbon for Sensitive Electrochemical Immunoassay of Breast Cancer Biomarker CA15-3. *Microchimica Acta* 2024, 191, doi:10.1007/s00604-024-06448-4.
- 31. Srilikhit, A.; Kongkeaw, S.; Cotchim, S.; Janduang, S.; Wannapob, R.; Kanatharana, P.; Thavarungkul, P.; Limbut, W. Carcinoembryonic Antigen and Cancer Antigen 125 Simultaneously Determined Using a Fluidics-Integrated Dual Carbon Electrode. *Microchemical Journal* 2024, 204, doi:10.1016/j.microc.2024.110911.
- 32. Oliveira, A.E.F.; Pereira, A.C.; Resende, M.A.C.; Ferreira, L.F. Disposable Voltammetric Immunosensor for Determination and Quantification of Biomarker CA 15-3 in Biological Specimens. *Analytica* 2024, *5*, 74–89, doi:10.3390/analytica5010005.
- 33. Yan, Y.; Liu, Z.; Pang, W.; Huang, S.; Deng, M.; Yao, J.; Huang, Q.; Jin, M.; Shui, L. Integrated Biosensor Array for Multiplex Biomarkers Cancer Diagnosis via In-Situ Self-Assembly Carbon Nanotubes with an Ordered Inverse-Opal Structure. *Biosens Bioelectron* 2024, 262, doi:10.1016/j.bios.2024.116528.
- 34. Farzin, M.A.; Naghib, S.M. Paper-Based Immunosensor Integrated with Bioinspired Cu-Polydopamine Nanozyme for Voltammetric Detection of CA-15-3 Tumor Marker. *Sensors and Actuators Reports* 2024, 8, doi:10.1016/j.snr.2024.100209.

- 35. Cotchim, S.; Kongkaew, S.; Thavarungkul, P.; Kanatharana, P.; Limbut, W. A Dual-Electrode Label-Free Immunosensor Based on in Situ Prepared Au–MoO3-Chi/Porous Graphene Nanoparticles for Point-of-Care Detection of Cholangiocarcinoma. *Talanta* 2024, 272, doi:10.1016/j.talanta.2024.125755.
- 36. Ranjan, P.; Abubakar Sadique, M.; Yadav, S.; Khan, R.; Kumar Srivastava, A. Electrochemical Nanobiosensor of Ionic Liquid Functionalized MoO3-RGO for Sensitive Detection of Carcinoembryonic Antigen. *Chempluschem* 2024, 89, doi:10.1002/cplu.202300625.
- 37. Shamsazar, A.; Moghaddam, M.S.; Asadi, A.; Mahdavi, M. Development and Performance Assessment of a Multi-Walled Carbon Nanotube/Iron Oxide Nanocomposite-Based Electrochemical Immunosensor for Precise and Ultrasensitive Detection of Carcinoembryonic Antigen. *J Appl Electrochem* 2024, *54*, 1713–1726, doi:10.1007/s10800-023-02063-y.
- 38. Mehta, D.; Kaur, S.; Nagaiah, T.C. Realizing the Label-Free Sensitive Detection of Carcinoembryogenic Antigen (CEA) in Blood Serum via a MNC-Decorated Flexible Immunosensor. *Analytical Methods* 2024, *16*, 1473–1479, doi:10.1039/d3ay02073h.
- Chellachamy Anbalagan, A.; Korram, J.; Doble, M.; Sawant, S.N. Bio-Functionalized Carbon Dots for Signaling Immuno-Reaction of Carcinoembryonic Antigen in an Electrochemical Biosensor for Cancer Biomarker Detection. *Discover Nano* 2024, 19, doi:10.1186/s11671-024-03980-3.
- 40. Janduang, S.; Cotchim, S.; Kongkaew, S.; Srilikhit, A.; Wannapob, R.; Kanatharana, P.; Thavarungkul, P.; Limbut, W. Synthesis of Flower-like ZnO Nanoparticles for Label-Free Point of Care Detection of Carcinoembryonic Antigen. *Talanta* 2024, 277, doi:10.1016/j.talanta.2024.126330.
- 41. Lei, L.; Zhang, J.; Zhang, W.; Hao, J.; Wu, K. Ultrasensitive Carbon Nanotube-Bridged MXene Conductive Network Arrays for One-Step Homogeneous Electrochemical Immunosensing of Tumor Markers. *Biosens Bioelectron* 2024, 263, doi:10.1016/j.bios.2024.116609.
- 42. Rana, M.S.; Meng, F.; Zhang, R.; Huang, L.; Qian, K. Integrating Zirconia-Gold Hybrid into Aptasensor Capable of Ultrasensitive and Robust Carcinoembryonic Antigen Determination. *Electrochim Acta* 2024, 507, doi:10.1016/j.electacta.2024.145063.
- 43. Yunussova, N.; Tilegen, M.; Pham, T.T.; Kanayeva, D. Rapid Detection of Carcinoembryonic Antigen by Means of an Electrochemical Aptasensor. *iScience* 2024, 27, doi:10.1016/j.isci.2024.109637.
- 44. Li, S.; Cao, T.; Zhang, H.; Sun, Y.; Luo, C. An Electrochemical Aptasensor Based on Ce-MOF@COF to Detect Carcinoembryonic Antigen. *New Journal of Chemistry* 2024, 48, 10628–10635, doi:10.1039/d4nj01743a.
- 45. Zhang, Y.; Xu, J.; Shen, J.; Zhang, B.; Xue, T.; Lv, X.; Zhang, X.; Zhu, G. A Dual-Mode Homogeneous Electrochemical-Colorimetric Biosensing Sensor for Carcinoembryonic Antigen Detection Based on a Microfluidic Paper-Based Analysis Device. *Analytical Methods* 2024, 16, 7372–7380, doi:10.1039/d4ay01480d.
- Zhou, Y.; Wang, H.; Xi, F.; Lu, C. Sensitive Electrochemical Detection of Carcinoembryonic Antigen Based on Biofunctionalized Nanochannel Modified Carbonaceous Electrode. *Molecules* 2024, 29, doi:10.3390/molecules29040858.
- 47. Yaiwong, P.; Jakmunee, J.; Pimalai, D.; Ounnunkad, K.; Bamrungsap, S. An Electrochemical/SERS Dual-Mode Immunosensor Using TMB/Au Nanotag and Au@2D-MoS2 Modified Screen-Printed Electrode for Sensitive Detection of Prostate Cancer Biomarker. *Colloids Surf B Biointerfaces* 2024, 243, doi:10.1016/j.colsurfb.2024.114124.
- 48. Uruc, S.; Dokur, E.; Gorduk, O.; Sahin, Y. Disposable and Ultrasensitive Label-Free Gold Nanoparticle Patterned Poly(3,4-Ethylenedioxythiophene-Co-3-Methylthiophene) Electrode for Electrochemical Immunosensing of Prostate-Specific Antigen. *New Journal of Chemistry* 2024, 48, 10415–10426, doi:10.1039/d4nj02034k.
- 49. Chellachamy Anbalagan, A.; Korram, J.; Koyande, P.; Khan, S.; Agrawal, R.; Sawant, S.N. Carbon Dots Functionalized Polyaniline as Efficient Sensing Platform for Cancer Biomarker Detection. *Diam Relat Mater* 2024, 147, doi:10.1016/j.diamond.2024.111276.
- 50. Wang, J.; Zhou, H.; Liang, R.; Qin, W. Chronopotentiometric Nanopore Sensor Based on a Stimulus-Responsive Molecularly Imprinted Polymer for Label-Free Dual-Biomarker Detection. *Anal Chem* 2024, 96, 9370–9378, doi:10.1021/acs.analchem.3c05817.
- 51. Redondo-Fernández, G.; Cid-Barrio, L.; Fernández-Argüelles, M.T.; de la Escosura-Muñiz, A.; Soldado, A.; Costa-Fernández, J.M. Controlled Silver Electrodeposition on Gold Nanoparticle Antibody Tags for

- Ultrasensitive Prostate Specific Antigen Sensing Using Electrochemical and Optical Smartphone Detection. *Talanta* 2024, 275, doi:10.1016/j.talanta.2024.126095.
- 52. Ren, Y.; Li, J.; Wu, W.; Yu, X.; Tao, M.; Han, Y.; Lin, Y.; Shi, Z.; Zhou, L.; Zhao, Z.; et al. Dual Chemical Bonding Construction of Electrochemical Peptide Sensor Based on GDY/MOFs(Fe) Composite for Ultra-Low Determination of Prostate-Specific Antigen. *Talanta* 2024, 278, doi:10.1016/j.talanta.2024.126459.
- 53. Rahman, S.F.A.; Arshad, M.K.M.; Gopinath, S.C.B.; Fathil, M.F.M.; Sarry, F.; Ibau, C.; Elmazria, O.; Hage-Ali, S. Interdigitated Impedimetric-Based Maackia Amurensis Lectin Biosensor for Prostate Cancer Biomarker. *Microchimica Acta* 2024, 191, doi:10.1007/s00604-024-06189-4.
- 54. Saeidi Tabar, F.; pourmadadi, M.; Yazdian, F.; Rashedi, H.; Rahdar, A.; Fathi-karkan, S.; Romanholo Ferreira, L.F. Ultrasensitive Aptamer-Based Electrochemical Nanobiosensor in Diagnosis of Prostate Cancer Using 2D:2D Reduced Graphene Oxide/Graphitic Carbon Nitride Decorated with Au Nanoparticles. *European Journal of Medicinal Chemistry Reports* 2024, 12, doi:10.1016/j.ejmcr.2024.100192.
- 55. Yue, Y.; Zhang, X.; Xu, Z.; Sun, L.; Li, S.; Liu, R. Ultrasensitive Detection of PSA in Human Serum Using Label-Free Electrochemical Biosensor with Magnetically Induced Self-Assembly Based on α-Fe2O3/Fe3O4@Au Nanocomposites. *Microchemical Journal* 2024, 201, doi:10.1016/j.microc.2024.110487.
- 56. Cotchim, S.; Kongkaew, S.; Thavarungkul, P.; Kanatharana, P.; Limbut, W. An Unlabeled Electrochemical Immunosensor Uses Poly(Thionine) and Graphene Quantum Dot-Modified Activated Marigold Flower Carbon for Early Prostate Cancer Detection. *Biosensors (Basel)* 2024, *14*, doi:10.3390/bios14120589.
- 57. Sasa, G.B.K.; He, B.; Chen, C.; Chen, Z.; Li, S.; Tan, C.S. A Dual-Targeted Electrochemical Aptasensor for Neuroblastoma-Related MicroRNAs Detection. *Talanta* 2024, 280, doi:10.1016/j.talanta.2024.126772.
- Yu, X.; Ding, S.; Zhao, Y.; Xu, M.; Wu, Z.; Zhao, C. A Highly Sensitive and Robust Electrochemical Biosensor for MicroRNA Detection Based on PNA-DNA Hetero-Three-Way Junction Formation and Target-Recycling Catalytic Hairpin Assembly Amplification. *Talanta* 2024, 266, doi:10.1016/j.talanta.2023.125020.
- 59. Sun, B.; Tang, H.; Gao, G.; Qian, Y.; Shen, Q.; Liang, L.; Cao, H. Al-Doped ZnO Nanostars for Electrochemical MiRNA-21 Biosensors. *J Electrochem Soc* 2024, *171*, 087509, doi:10.1149/1945-7111/ad6a9b.
- 60. Ma, C.; Zhou, Q.; Shi, J.; Gao, H.; Huang, D.; Xue, H.; Wang, H.; Zhang, Z.; Yang, S.; Zhang, J.; et al. CRISPR-Empowered Electrochemical Biosensor for Target Amplification-Free and Sensitive Detection of MiRNA. *Talanta* 2024, 266, doi:10.1016/j.talanta.2023.125125.
- 61. Huang, M.S.; Govindasamy, M.; Chinnapaiyan, S.; Lin, Y.T.; Lu, S.Y.; Samukawa, S.; Huang, C.H. Electrochemical Biosensor Based on an Atomic Layered Composite of Graphene Oxide/Graphene as an Electrode Material towards Selective and Sensitive Detection of MiRNA-21. *Microchemical Journal* 2024, 199, doi:10.1016/j.microc.2024.110112.
- 62. Raucci, A.; Cimmino, W.; Romanò, S.; Singh, S.; Normanno, N.; Polo, F.; Cinti, S. Electrochemical Detection of MiRNA Using Commercial and Hand-Made Screen-Printed Electrodes: Liquid Biopsy for Cancer Management as Case of Study. *ChemistryOpen* 2024, 13, doi:10.1002/open.202300203.
- 63. Bhattacharjee, S.; George, M.; Neppolian, B.; Das, J. Electrochemical Detection of Non-Small Cell Lung Cancer (NSCLC) Mir-223 Biomarker Employing Gold/MWCNT Nanocomposite–Based Sandwich Platform. *Journal of Solid State Electrochemistry* 2025, 29, 669–680, doi:10.1007/s10008-024-06094-z.
- 64. Cimmino, W.; Raucci, A.; Grosso, S.P.; Normanno, N.; Cinti, S. Enhancing Sensitivity towards Electrochemical MiRNA Detection Using an Affordable Paper-Based Strategy. *Anal Bioanal Chem* 2024, 416, 4227–4236, doi:10.1007/s00216-024-05406-6.
- 65. Zhu, S.; Chen, J.; Yu, L.; Li, J.; You, S.; Zheng, Y.; Zhuang, W.; Qiu, B.; Huang, Y. Detection of Fucosylated Extracellular Vesicles MiR-4732-5p Related to Diagnosis of Early Lung Adenocarcinoma by the Electrochemical Biosensor. *Sci Rep* 2024, 14, doi:10.1038/s41598-024-61060-z.
- Dulgerbaki, C.; Uygun Oksuz, A. Iron Oxide/Poly(3,4-Ethylenedioxythiophene): Poly(Styrene Sulfonate)
 Glassy Carbon Electrode as a Novel Label-Free Electrochemical MicroRNA-21 Sensor. *Instrum Sci Technol* 2024, 52, 67–79, doi:10.1080/10739149.2023.2221339.
- 67. Li, Z.; Liu, J.; Bao, T.; Wu, Z.; Wen, W.; Zhang, X.; Wang, S. Magnetic Beads-Assisted Split DNAzyme Cleavage-Driven Assembly of Functionalized Covalent Organic Frameworks for Highly Sensitive Electrochemical Detection of MicroRNA. Sens Actuators B Chem 2024, 409, doi:10.1016/j.snb.2024.135594.

- 68. Yu, S.; Wei, G.; Zhao, P.; Zhang, J.; Kong, J.; Zhang, X. Organic Small Molecule Catalytic PET-RAFT Electrochemical Signal Amplification Strategy for MiRNA-21 Detection. *Chemical Engineering Journal* 2024, 492, doi:10.1016/j.cej.2024.152380.
- 69. Raucci, A.; Cimmino, W.; Grosso, S.P.; Normanno, N.; Giordano, A.; Cinti, S. Paper-Based Screen-Printed Electrode to Detect MiRNA-652 Associated to Triple-Negative Breast Cancer. *Electrochim Acta* 2024, 487, doi:10.1016/j.electacta.2024.144205.
- 70. Ali, M.; Hasan, E.; Barman, S.C.; Hedhili, M.N.; Alshareef, H.N.; Alsulaiman, D. Peptide Nucleic Acid-Clicked Ti3C2Tx MXene for Ultrasensitive Enzyme-Free Electrochemical Detection of MicroRNA Biomarkers. *Mater Horiz* 2024, *11*, 5045–5057, doi:10.1039/d4mh00714j.
- 71. Zhou, Y.; Che, S.; Wang, Z.; Zhang, X.; Yuan, X. Primer Exchange Reaction Assisted CRISPR/Cas9 Cleavage for Detection of Dual MicroRNAs with Electrochemistry Method. *Microchimica Acta* 2024, 191, doi:10.1007/s00604-024-06548-1.
- 72. Zhu, H.; Chen, S.; Lan, F.; Li, W.; Ji, T.; Zhang, L.; Guo, Y.; Pan, W.; Luo, S.; Xie, R. Sensitive Electrochemical Biosensor for Rapid Detection of SEV-MiRNA Based Turbo-like Localized Catalytic Hairpin Assembly. *Anal Chim Acta* 2024, 1311, doi:10.1016/j.aca.2024.342704.
- 73. Kivrak, E.; Kara, P. Simultaneous Detection of Ovarian Cancer Related MiRNA Biomarkers with Carboxylated Graphene Oxide Modified Electrochemical Biosensor Platform. *Bioelectrochemistry* 2025, 161, doi:10.1016/j.bioelechem.2024.108806.
- 74. Barman, S.C.; Ali, M.; Hasan, E.A.; Wehbe, N.; Alshareef, H.N.; Alsulaiman, D. Smartphone-Interfaced Electrochemical Biosensor for MicroRNA Detection Based on Laser-Induced Graphene with π - π Stacked Peptide Nucleic Acid Probes. *ACS Mater Lett* 2024, *6*, 837–846, doi:10.1021/acsmaterialslett.3c01225.
- 75. Yang, H.; Jin, Y.; Qian, H.; Wang, Y.; Bao, T.; Wu, Z.; Wen, W.; Zhang, X.; Wang, S. Target-Driven Cascade Amplified Assembly of Covalent Organic Frameworks on Tetrahedral DNA Nanostructure with Multiplex Recognition Domains for Ultrasensitive Detection of MicroRNA. *Anal Chim Acta* 2024, 1311, doi:10.1016/j.aca.2024.342743.
- 76. Zhang, J.; Chen, J.; Xie, Q.; Chu, Z.; Zhang, F.; Wang, Q. Sensitive Detection of Exosomes by an Electrochemical Aptasensor Based on Double Nucleic Acid Amplifications. *Microchemical Journal* 2024, 201, doi:10.1016/j.microc.2024.110667.
- 77. Huang, J.; Chen, T.; Zhao, Y.; Li, D.; Huang, Q.; Cao, L.; Chen, J.; Chen, D.; Hu, L.; Liu, H. Colloidal Quantum Dots-Modified Electrochemical Sensor for High-Sensitive Extracellular Vesicle Detection. *Chemical Engineering Journal* 2024, 487, doi:10.1016/j.cej.2024.150616.
- 78. Wang, L.; Wu, X.; Chen, G.; Chen, Y.; Xu, L.; Wang, J.; Chen, J. Dual Amplified Electrochemical Sensing Coupling of Ternary Hybridization-Based Exosomal MicroRNA Recognition and Perchlorate-Assisted Electrocatalytic Cycle. *Biosens Bioelectron* 2024, 243, doi:10.1016/j.bios.2023.115783.
- 79. Sahraei, N.; Mazloum-Ardakani, M.; Moradi, A.; Hoseynidokht, F. Flexible Electrochemical Paper-Based Device for Detection of Breast Cancer-Derived Exosome Using Nickel Nanofoam 3D Nanocomposite. *J Appl Electrochem* 2024, 54, 2817–2829, doi:10.1007/s10800-024-02141-9.
- 80. Sazaklioglu, S.A.; Torul, H.; Tamer, U.; Ensarioglu, H.K.; Vatansever, H.S.; Gumus, B.H.; Çelikkan, H. Sensitive and Reliable Lab-on-Paper Biosensor for Label-Free Detection of Exosomes by Electrochemical Impedance Spectroscopy. *Microchimica Acta* 2024, 191, doi:10.1007/s00604-024-06644-2.
- 81. Zhang, M.; Zhang, T.; Mei, W.; Wang, H.; Wang, Q.; Yang, X.; Wang, K. A Homogeneous Electrochemical Sensing Platform Based on DNAzyme Walker for Accurate Detection of Breast Cancer Exosomes. *Sens Actuators B Chem* 2024, 404, doi:10.1016/j.snb.2023.135252.
- 82. Zhou, G.; Zhou, F.; Yu, X.; Zhou, D.; Wang, J.; Bo, B.; Cao, Y.; Zhao, J. Electrochemical Detection of Tumor Cells Based on Proximity Labelling-Assisted Multiple Signal Amplification. *Sensors and Diagnostics* 2024, *3*, 1542–1550, doi:10.1039/d4sd00217b.
- 83. Yan, Z.; Che, Z.; Shi, Z.; Lv, J.; Yang, L.; Lu, T.; Lu, Y.; Shan, J.; Liu, Q. Hepatocellular Carcinoma Detection by Cell Sensor Based on Anti-GPC3 Single-Chain Variable Fragment. *Advanced Devices and Instrumentation* 2024, 5, doi:10.34133/adi.0043.

- 84. Hosseine, M.; Naghib, S.M.; Khodadadi, A. Label-Free Electrochemical Biosensor Based on Green-Synthesized Reduced Graphene Oxide/Fe3O4/Nafion/Polyaniline for Ultrasensitive Detection of SKBR3 Cell Line of HER2 Breast Cancer Biomarker. *Sci Rep* 2024, *14*, doi:10.1038/s41598-024-62231-8.
- 85. Yang, S.; Zhao, L.; Yang, X.; Yang, L.; Fa, H.; Wang, Y.; Huo, D.; Hou, C.; Zhong, D.; Yang, M. MBene as Novel and Effective Electrochemical Material Used to Enhance Paper-Based Biosensor for Point-of-Care Testing CtDNA from Mice Serum. *Chemical Engineering Journal* 2024, 498, doi:10.1016/j.cej.2024.155345.
- 86. Çağlayan Arslan, Z.; Okan, M.; Külah, H. Pre-Enrichment-Free Detection of Hepatocellular Carcinoma-Specific CtDNA via PDMS and MEMS-Based Microfluidic Sensor. *Microchimica Acta* 2024, 191, doi:10.1007/s00604-024-06315-2.
- 87. Park, B.C.; Soh, J.O.; Choi, H.J.; Park, H.S.; Lee, S.M.; Fu, H.E.; Kim, M.S.; Ko, M.J.; Koo, T.M.; Lee, J.Y.; et al. Ultrasensitive and Rapid Circulating Tumor DNA Liquid Biopsy Using Surface-Confined Gene Amplification on Dispersible Magnetic Nano-Electrodes. *ACS Nano* 2024, *18*, 12781–12794, doi:10.1021/acsnano.3c12266.
- 88. Yue, Y.; Zhang, X.; Zhao, S.; Wang, X.; Wang, J.; Liu, R. Construction of a Label-Free Electrochemical Biosensing System Utilizing Fe3O4/α-Fe2O3@Au with Magnetic-Induced Self-Assembly for the Detection of EGFR Glycoprotein. *Vacuum* 2024, 222, doi:10.1016/j.vacuum.2024.112975.
- 89. Zhang, H.; Gao, H.; Mu, W.; Que, L.; Gu, X.; Rong, S.; Ma, H.; Miao, M.; Qi, X.; Chang, D.; et al. Electrochemical-Fluorescent Bimodal Biosensor Based on Dual CRISPR-Cas12a Multiple Cascade Amplification for CtDNA Detection. *Anal Chem* 2024, doi:10.1021/acs.analchem.4c03012.
- 90. Tang, S.; Xu, Q.; Liu, M.; Zhu, Y.; Zhang, G.; Tang, X. Highly Sensitive Electrochemical Immunosensor Based on SiO2 Nanospheres for Detection of EGFR as Colorectal Cancer Biomarker. *Alexandria Engineering Journal* 2024, 89, 53–59, doi:10.1016/j.aej.2024.01.016.
- 91. Liu, X.; Zhang, Y.; Li, N.; Yang, Z.; Yang, S.; Li, J.; Chen, J.; Huo, D.; Hou, C. Cu-MOF/Aptamer/Magnetic Bead Sensor for Detection of Breast Cancer Biomarkers HER2 and ER. *ACS Appl Nano Mater* 2024, 7, 9768–9776, doi:10.1021/acsanm.4c01671.
- 92. Zhang, Y.; Wang, J.; Liu, M.; Ni, Y.; Yue, Y.; He, D.; Liu, R. Magnetically Induced Self-Assembly Electrochemical Biosensor with Ultra-Low Detection Limit and Extended Measuring Range for Sensitive Detection of HER2 Protein. *Bioelectrochemistry* 2024, 155, doi:10.1016/j.bioelechem.2023.108592.
- 93. Kanagavalli, P.; Eissa, S. Exploring Various Carbon Nanomaterials-Based Electrodes Modified with Polymelamine for the Reagentless Electrochemical Immunosensing of Claudin18.2. *Biosens Bioelectron* 2024, 259, doi:10.1016/j.bios.2024.116388.
- 94. Kanagavalli, P.; Eissa, S. Redox Probe-Free Electrochemical Immunosensor Utilizing Electropolymerized Melamine on Reduced Graphene Oxide for the Point-of-Care Diagnosis of Gastric Cancer. *Talanta* 2024, 270, doi:10.1016/j.talanta.2023.125549.
- 95. Wei, S.; Wang, Z.; Li, S.; Ren, H.; Wang, Y.; Xiao, H.; Zhao, F.; Zhu, J.; Chen, Z. Ultrasensitive and Multiplexed Gastric Cancer Biomarkers Detection with an Integrated Electrochemical Immunosensing Platform. *Talanta* 2025, 282, doi:10.1016/j.talanta.2024.126961.
- 96. Chen, L.; Guo, J.; Zhou, Y.; Yu, W.Q.; Jin, Y.S.; Fu, Y.Z.; Yuan, R. Efficient CdIn2S4/MgIn2S4 Heterojunction for Ultrasensitive Detection of Lung Cancer Marker Neuron-Specific Enolase. *Talanta* 2024, 280, doi:10.1016/j.talanta.2024.126669.
- 97. Xu, F.; Ai, Q.Y.; Wang, A.J.; Mei, L.P.; Song, P.; Liu, W.; Feng, J.J.; Cheang, T.Y. Pronounced Signal Enhancement with Gourd-Shaped Hollow PtCoNi Bunched Nanochains for Electrochemical Immunosensing of Alpha-Fetoprotein. *Sens Actuators B Chem* 2025, 422, doi:10.1016/j.snb.2024.136608.

- 98. Ivanova, N.; Ahmadi, S.; Chan, E.; Fournier, L.; Spagnolo, S.; Thompson, M. Detection of Ovarian Cancer Biomarker Lysophosphatidic Acid Using a Label-Free Electrochemical Biosensor. *Electrochem* 2024, *5*, 243–257, doi:10.3390/electrochem5020015.
- 99. Shen, Q.; Ding, J.; Guo, Z.; Yang, X.; Zhang, Y.; Xu, B.; Yang, H.; Sun, Y.; Hang, L. Dual-Responsive Electrochemical Immunosensor for CYFRA21-1 Detection Based on Au/Co Co-Loaded 3D Ordered Macroporous Carbon Interconnected Framework. *Colloids Surf B Biointerfaces* 2024, 242, doi:10.1016/j.colsurfb.2024.114111.

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