

Review

Not peer-reviewed version

Beyond the Classics: The Synergy of AI and Genomics Reveals A New Army of Pigmentation Genes

[Ehsan Pashay Ahi](#)* and Nidal Karagic

Posted Date: 31 July 2025

doi: 10.20944/preprints202507.2679.v1

Keywords: Pigmentation genetics; Artificial intelligence; Genomic mapping; Non-classical pigmentation genes; Deep learning phenotyping



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Beyond the Classics: The Synergy of AI and Genomics Reveals A New Army of Pigmentation Genes

Ehsan Pashay Ahi ^{1,*} and Nidal Karagic ^{2,*}

¹ Organismal and Evolutionary Biology Research Programme, Faculty of Biological and Environmental Sciences, University of Helsinki, Viikinkaari 9, 00014, Helsinki, Finland

² Helsinki Institute of Life Science (HiLIFE), Institute of Biotechnology, University of Helsinki, Helsinki, Finland

* Correspondence: ehsan.pashayahi@helsinki.fi (E.P.A.); nidal.karagic@helsinki.fi (N.K.)

Abstract

Pigmentation has long served as a powerful system for exploring gene–trait relationships, yet much of the field has focused on a relatively narrow group of well-established genes involved in melanin production and pigment cell differentiation. Recent advances, however, have allowed pigmentation to be studied through a more comprehensive framework. By combining artificial intelligence (AI)–driven phenotyping with genomic mapping approaches such as genome-wide association studies, QTL mapping, and structural variant analysis, a broader range of pigmentation regulators has been identified across diverse animal taxa. This review highlights studies where AI methods, including deep learning, self-supervised modeling, and pattern recognition, have been used to quantify complex pigmentation traits in animals. These approaches have enabled the discovery of non-classical pigmentation genes involved in membrane trafficking, intracellular signaling, structural organization, and non-coding regulation. Rather than displacing the classical pigmentation paradigm, these findings extend it, revealing a wider set of genetic contributors to coloration and pattern diversity. We introduce the term *AI-pigmentomics* to describe the integration of AI-driven phenotyping with genomic mapping, as part of the broader emergence of AI-omics. Together, AI and genomic mapping are reshaping our understanding of pigmentation by uncovering unexpected biological mechanisms and providing a framework for investigating pigmentation in both model and non-model species.

Keywords: Pigmentation genetics; Artificial intelligence; Genomic mapping; Non-classical pigmentation genes; Deep learning phenotyping

1. Introduction

The study of pigmentation has provided key insights into developmental genetics, cell differentiation, and evolutionary biology (Hoekstra, 2006; Saunders et al., 2019; Parichy, 2021; Kratochwil and Mallarino, 2023). Historically, most research in this field has been shaped by discoveries in model organisms, particularly those that display variation in melanin-based traits (Mills and Patterson, 2009; Caro and Mallarino, 2020; McNamara et al., 2021). Genes such as MC1R, TYR, ASIP, and OCA2 have been repeatedly identified as major regulators of melanin synthesis and pigment type-switching, forming the foundation of what has often been referred to as the "classical" pigmentation gene set (Gerstenblith et al., 2010; Wakamatsu and Ito, 2021). These genes have explained substantial phenotypic variation in mammals and birds and have been central to understanding the genetic control of coloration over the past several decades (Galván and Solano, 2016; Caro and Mallarino, 2020; Eizirik and Trindade, 2021; Elkin et al., 2023). However, pigmentation systems are far more complex than initially understood, even in well-studied organisms such as

humans (Martin et al., 2017; Quillen et al., 2019). In many taxa, coloration is determined not only by melanin but also by other pigments such as pteridines, carotenoids, and purines, as well as by structural components like iridophores and reflective crystals (Shawkey and D'Alba, 2017; Toews et al., 2017; Andrade and Carneiro, 2021). In teleost fish, for example, at least five distinct pigment cell types have been described, each with specific pigment content and developmental pathways (Parichy, 2021). Regulators of xanthophore and iridophore differentiation, e.g., *csf1r*, *gch2*, *ltk*, *pnp4a*, and *tfec*, have been studied primarily in zebrafish but remain largely uncharacterized in other species (Irion and Nüsslein-Volhard, 2019; Patterson and Parichy, 2019). Although many of these genes are well described within particular lineages (Baxter et al., 2019), their inclusion in broader pigmentation models has often been limited.

As research has expanded into non-model organisms and more diverse pigmentation systems, the limitations of the classical gene framework have become increasingly apparent (Figon et al., 2021a; Kondo et al., 2021; Vöcking et al., 2022). The use of historically well-known regulatory genes fails to explain a large portion of pigmentation diversity outside of model organisms. A growing number of studies have indicated that genes involved in vesicle trafficking (Ullate-Agote et al., 2020; Liu et al., 2022), membrane transport (Ahi et al., 2020b; Podobnik et al., 2020), organelle-associated signaling (Stuckert et al., 2019; Figon et al., 2021b), cytoskeletal organization (Feiner et al., 2022; Lloyd et al., 2024), epigenomic and epitranscriptomic mechanisms (Strowbridge et al., 2025) and non-coding regulatory functions (Feng et al., 2018; Luo et al., 2019) may play substantial roles in determining pigment cell behavior and spatial patterning. These genes frequently fall outside the melanin biosynthetic pathway yet are consistently implicated in pigmentation phenotypes when higher-resolution phenotyping and broader genomic scans are employed (Baxter et al., 2019). This shift in perspective has been facilitated by the growing use of artificial intelligence and machine learning tools for phenotypic characterization, alongside genome-wide mapping techniques that no longer rely solely on prior knowledge of candidate genes (Karagic and Kratochwil, 2025). As a result, pigmentation genetics is being redefined not by discarding the classical framework but by integrating a wider array of biological mechanisms that govern color and pattern formation across animal taxa. This article focuses on how this integration is being achieved through the combined application of AI-based phenotyping and genomic mapping, and how it is expanding the known landscape of pigmentation genes.

2. From Pattern to Phenotype: How AI is Transforming Pigmentation Trait Analysis

Advances in artificial intelligence have introduced new possibilities for quantifying complex traits in animals, such as pigmentation related traits (Fernandes et al., 2020; Lürig et al., 2021). Traditional approaches to phenotype characterization have relied heavily on manual scoring, categorical classification, or measurements of selected regions of interest (Fernandes et al., 2020). While effective in certain contexts, these methods have often lacked the resolution necessary to capture subtle differences in hue, spatial distribution, pattern complexity, or structural coloration (Hemingson et al., 2024). As a result, large portions of pigmentation variation have remained either unquantified or only coarsely approximated (Butler et al., 2011; Siegenthaler et al., 2017). Recent applications of AI-based tools, particularly deep learning methods, have provided more robust and scalable solutions to these limitations (Wu et al., 2019; Niu et al., 2025).

In several systems, convolutional neural networks (CNNs) have been used to extract detailed features from full-body images, such as in the study of ornamental coloration in male guppies (van der Bijl et al., 2025). In that context, network architectures were trained to quantify only coloration on a global spatial scale with high resolution, which would not be feasible to do manually. Once extracted, the quantitative trait values were used for subsequent genomic association analyses to obtain a spatial map for the genetic control or coloration pattern. Similar techniques have been used in studies of feather pigmentation in ducks and chicks, where region-specific melanin content was quantified using pixel-level segmentation algorithms (Heo et al., 2023; Twumasi et al., 2024; Wang et

al., 2024). In these cases, traits were defined based on measurable output from image data rather than observer-defined categories, thus improving precision and reproducibility.

Beyond standard CNN applications, other AI frameworks such as self-supervised and metric learning have also been employed (Hoyal Cuthill et al., 2019; Xie et al., 2024). In particular, contrastive learning methods have been used to generate latent phenotypic embeddings from human retinal fundus images, providing continuous, data-driven pigmentation scores that are agnostic to predefined labels (Xie et al., 2024). Triplet networks have similarly been used to compare complex wing pattern geometries in butterflies, enabling fine-scale assessment of morphological similarity between individuals or populations (Hoyal Cuthill et al., 2019). These approaches have not only expanded the phenotypic scope of pigmentation research but have also allowed researchers to revisit older datasets with new analytical depth. Archived images or digital specimens can now be reanalyzed with updated models, facilitating retrospective discoveries and longitudinal analyses (Newell et al., 2021; Rosenberg et al., 2021; Hantak et al., 2022). Moreover, as models are trained on increasingly diverse image sets, their generalizability across taxa and environments is expected to improve (Wu et al., 2019). Despite their promise, AI-based phenotyping methods are not without limitations. Models can be biased by training data, may fail to generalize across lighting conditions or image formats, and require significant computational infrastructure (Fernandes et al., 2020). Nonetheless, their ability to detect complex, non-linear, and spatially distributed traits represents a meaningful advancement in the study of pigmentation. When these tools are applied thoughtfully and validated rigorously, they provide a powerful means to expand the trait space available for genomic investigation.

3. Genomic Mapping Methods and Their Integration with AI-Derived Traits

The identification of pigmentation-associated loci has historically depended on either candidate gene studies or classical genetic crosses, which often required prior assumptions about trait architecture. With the advent of genome-wide association studies (GWAS), it became possible to interrogate genotype–phenotype relationships without limiting the analysis to preselected loci (Gerstenblith et al., 2010; Adhikari et al., 2019; Lona-Durazo et al., 2019). However, the resolution and interpretability of GWAS results have always been constrained by the quality of phenotypic data (Bush et al., 2016; Landi et al., 2020; Kim et al., 2024). As AI-derived traits have grown in both resolution and dimensionality, their integration with genomic mapping has resulted in a substantial improvement in the identification of pigmentation genes, particularly those outside classical biosynthetic pathways (see examples in table 1). In recent studies, AI-extracted phenotypes have been used directly as quantitative traits in GWAS pipelines. For example, high resolution color pattern maps extracted from guppy images using CNNs have been employed to identify both autosomal and Y-linked loci influencing complex ornamental patterns (van der Bijl et al., 2025). In the case of human retinal pigmentation, principal components derived from AI-generated pixel embeddings were used to define quantitative pigmentation traits that revealed multiple novel loci through image-GWAS, including genes involved in membrane function and intracellular transport (Rajesh et al., 2025).

Mapping approaches have also been extended beyond GWAS. In scallops and butterflies, where limited reference genomes or recombination maps can present barriers to fine-scale association, quantitative trait locus (QTL) mapping and bulk-segregant analysis have been paired with AI-based pattern scoring to detect non-classic contributors to shell and wing coloration (Woronik et al., 2019; Mao et al., 2020). Similar approaches have been applied in plants, such as peanut, where image-based phenotyping accelerated QTL mapping and QTL \times environment interaction analysis of testa color (Zhang et al., 2021). These include genes involved in structural patterning, chromatin regulation, and non-coding RNAs (Woronik et al., 2019; Mao et al., 2020). Integrative approaches such as eQTL mapping and transcriptome-wide association studies (TWAS) are now increasingly used to connect AI-defined traits to regulatory variation (Ferguson et al., 2021; Jackson et al., 2025). This is particularly relevant for genes with subtle or tissue-specific effects on pigmentation, which may not appear as strong GWAS hits but can influence gene expression in pigment cells (Ioannidis et al., 2018; Zhang et

al., 2018; Lona-Durazo et al., 2021; Song et al., 2024). Functional studies, including CRISPR-mediated validation, can further confirm these novel associations (Crawford et al., 2017; Choi et al., 2020; Bajpai et al., 2023; Xu et al., 2024). Together, these methods are shifting the discovery paradigm away from reliance on known pathways. AI-derived traits, when combined with flexible and comprehensive genomic mapping strategies, are allowing for the identification of previously unrecognized genes involved in coloration. The result is not just a more complete list of pigmentation loci, but a more nuanced understanding of the regulatory, structural, and developmental processes underlying color and pattern formation.

4. AI-Genomics Approaches Reveal Diverse Pigmentation Mechanisms Across Taxa

The integration of AI-driven phenotyping with genomic mapping has led to the discovery of pigmentation-associated genes that had not been previously implicated in classical pigment pathways (see below). These findings span a wide range of animal taxa and reflect a growing recognition that pigmentation traits are regulated by a broader network of cellular and developmental mechanisms than previously appreciated (Table 1). In this section, representative studies are presented to illustrate how this combined approach has enabled the identification of novel genes in fish, birds, mammals and invertebrates.

In humans, AI-derived retinal pigment scores have enabled image-based GWAS that revealed loci associated with pigmentation variation across the fundus (Rajesh et al., 2025). Among the identified genes were involved in ciliary transport and with less known role in cellular signaling. These genes are not part of traditional pigmentation pathways and suggest that pigmentation phenotypes can be influenced by broader cellular physiology.

In guppies, the use of convolutional neural networks to quantify male ornamental color patterns facilitated a high-resolution GWAS that identified both autosomal and Y-linked loci (van der Bijl et al., 2025). Among these were regions containing genes involved in vesicle trafficking and cell polarity, features not typically associated with pigmentation in vertebrates. These genes may affect pigment deposition indirectly through their influence on pigment cell behavior or the spatial organization of pattern elements.

In domestic ducks, melanin content was quantified through automated segmentation of feather images, and GWAS using this trait led to the discovery of *DENND4A* and *PRKG1*. These genes are involved in intracellular transport and signal transduction, respectively, and neither had been linked previously to pigmentation phenotypes (Twumasi et al., 2024). A similar study in Rhode Island Red chicks, using pattern-based classifiers, identified *TMTC3* as a contributor to stripe development (Shen et al., 2022). This gene encodes a transmembrane protein that may influence pattern symmetry through effects on cell adhesion or migration during feather development.

In mollusks, the analysis of shell color in bay scallops using AI-based color classification, followed by GWAS, identified *PKS1*, *GRL101*, and *PLC1*, genes involved in polyketide synthesis and calcium signaling (Zhu et al., 2021). These pathways are not homologous to those involved in melanin or pteridine production and highlight the diversity of pigment chemistries across animal lineages.

In butterflies, AI-driven pattern recognition and QTL mapping have revealed the role of novel genetic regulators and long-range enhancers in the regulation of wing pigmentation (Kronforst and Sheikh, 2023; Fandino et al., 2024; Livraghi et al., 2024). For example, in *Heliconius* species, variation in color pattern elements has been linked to regulatory elements influencing gene expression at loci not previously associated with pigmentation (Kronforst and Sheikh, 2023).

These case studies demonstrate how AI-augmented phenotyping enables the detection of subtle but biologically meaningful trait differences, and how genomic mapping techniques, when freed from candidate gene constraints, are capable of revealing previously unrecognized contributors to pigment variation.

5. Emerging Functions and Biological Roles of Non-Classical Pigmentation Genes

The expansion of pigmentation genetics into non-classical gene space has revealed a more diverse array of biological functions than was previously considered relevant to color formation (Ahi et al., 2020a; Ganguly et al., 2022; Chong et al., 2024; Marin-Recinos and Pucker, 2024; Qi et al., 2025). Many of the genes identified through AI-assisted genomic mapping are not involved in pigment synthesis per se, but instead influence how pigments are transported, deposited, or spatially organized (Heo et al., 2023; Kim et al., 2024; Lay et al., 2024; Twumasi et al., 2024; Xie et al., 2024; Rajesh et al., 2025; van der Bijl et al., 2025). Others act through indirect mechanisms, such as regulating cell migration, vesicle formation, cytoskeletal dynamics, or intracellular signaling in pigment cells or their precursors (Simcoe et al., 2021; Kirchler et al., 2022; Xie et al., 2024; van der Bijl et al., 2025).

One prominent category includes genes involved in membrane trafficking and vesicle transport. For instance, *DENND4A*, identified in ducks, belongs to a family of guanine nucleotide exchange factors known to regulate endosomal trafficking (Twumasi et al., 2024). Alterations in these pathways may affect the delivery of pigment granules to the cell periphery or influence the stability of pigment-containing organelles. Another example is *COMMD3*, identified through AI-assisted image-based GWAS, which plays a role in endosomal trafficking and melanosome regulation (Xie et al., 2024). In another recent human-based deep learning study of polygenic adaptation, genes such as *MREG*, *USP13*, and *BLOC1S3* were identified as pigmentation-linked candidates, likely influencing phenotypic variation through melanosome transport, intracellular trafficking, and organelle organization (Tripathi et al., 2024). Their functions support a growing view that pigmentation can be shaped by cellular infrastructure beyond pigment biosynthesis itself. In fish, additional genes such as *hps4*, a component of the BLOC-3 complex involved in endosomal cargo delivery, and *dnajc6*, a regulator of clathrin-mediated endocytosis, further represent mechanisms of vesicle formation and intracellular transport contributing to pigment pattern diversification (van der Bijl et al., 2025).

Genes involved in intracellular signaling and second messenger pathways have also been implicated. *PRKG1*, associated with feather pigmentation in ducks, encodes a cGMP-dependent protein kinase and may modulate cellular responses to external patterning signals during development (Twumasi et al., 2024). *PDE3A*, identified in human retinal pigmentation studies, encodes a phosphodiesterase that regulates cyclic nucleotide levels, possibly affecting cell-specific pigment expression patterns or signaling thresholds required for pigment cell differentiation (Rajesh et al., 2025). In guppy fish, several genes with roles in cyclic nucleotide and intracellular signaling pathways have been implicated in pigmentation, including *gsk3aa*, a kinase downstream of cAMP/PKG signaling; *crh*, which activates cAMP through GPCR signaling; *adgra1*, a G protein-coupled receptor influencing second messenger levels; and *prex1*, a Rac-GEF linked to signaling cascades regulating cytoskeletal dynamics and pigment cell behavior (van der Bijl et al., 2025).

In some cases, structural proteins or ciliary transport components such as *IFT122* have been implicated (Rajesh et al., 2025). Although primarily studied in the context of developmental signaling, these genes may influence pigmentation by affecting the spatial organization of pigment cells or by modulating morphogen gradients (e.g., *LRMDA*, *MEF2C*, and *PROX1*) during pattern formation (Tripathi et al., 2024). In fish, the gene *ush2a*, better known for its role in sensory cilia and photoreceptor maintenance, has also been associated with pigmentation phenotypes, likely through similar effects on cell positioning or organelle trafficking (van der Bijl et al., 2025). The identification of these genes has begun to shift the understanding of pigmentation from a pigment-centric to a systems-level trait. Rather than being governed solely by pathways that synthesize or degrade pigments, pigmentation emerges as a composite phenotype shaped by diverse biological processes. For instance, genes like *BCL2*, *MYLK*, and *DLG1* may contribute to pigmentation in more indirect ways, including by promoting melanocyte survival, regulating cytoskeletal tension, or maintaining epithelial polarity, thus influencing the persistence and geometry of pigment patterns (Tripathi et al.,

2024). This expanded view invites new hypotheses about how pigment traits evolve, how they are developmentally regulated, and how they might respond to environmental or physiological cues.

Non-coding RNAs have emerged as an additional class of regulators in pigmentation biology. In cashmere goats, a multi-omics and AI-assisted study identified several miRNAs that regulate hair follicle pigmentation-related pathways, including miR-214, miR-29a/b1, and miR-199a-5p (Chunhua et al., 2025). These small RNAs modulate gene expression post-transcriptionally, influencing processes such as melanocyte activity and pigment deposition. By adding a layer of regulatory complexity beyond protein-coding variation, non-coding RNAs may help explain species-specific or sex-specific differences in pigmentation patterns that are not captured by traditional genomic analyses alone.

Although AI tools for RNA modification analysis have not yet been applied directly to pigmentation patterning, their emergence in the field of epitranscriptomics holds significant promise. Recent advances in deep learning models, such as those predicting m⁶A methylation from RNA sequence or direct RNA-seq data (Hendra et al., 2022), demonstrate the potential to uncover post-transcriptional regulatory mechanisms that may influence pigment cell fate, differentiation, or gene expression dynamics (Strowbridge et al., 2025). As these methods mature, they are likely to become powerful tools for exploring how RNA modifications contribute to species-specific or environmentally responsive pigmentation phenotypes.

6. Future Perspectives and Challenges in AI-Enabled Pigmentation Genomics

While the integration of AI-based phenotyping and genomic mapping has led to the discovery of numerous non-classical pigmentation genes, several challenges remain that must be addressed to fully realize the potential of this approach. These challenges are both technical and biological in nature and span the processes of data acquisition, model development, trait validation, and cross-species generalization (Nabwire et al., 2021; Dingemans et al., 2023; Athanasopoulou et al., 2025; Wu and Xie, 2025). A major limitation lies in the standardization of phenotypic data (Ying, 2023; Upadhyay et al., 2024). AI models often rely on large datasets of labeled images or segmentation masks, which may be lacking for many non-model organisms (Koblitz et al., 2025). Variation in lighting, resolution, specimen orientation, and background conditions can introduce noise into training data, potentially reducing model performance and interpretability (Zhang et al., 2017; Yang et al., 2018; Billah et al., 2025). Although data augmentation and domain adaptation strategies can partially address these issues (Orouji et al., 2024; Cao et al., 2025), the creation of high-quality, taxonomically diverse training sets remains a significant bottleneck.

Another concern involves the generalizability of AI models across species or even within populations under different ecological conditions (Norman et al., 2023; Okuley et al., 2025). Models trained on one species or developmental stage may perform poorly when applied to others unless explicitly retrained or fine-tuned (Tabak et al., 2019; Mulero-Pázmány et al., 2025). This challenge is particularly relevant for studies aiming to compare pigmentation traits across evolutionary lineages, where color production mechanisms and cell types may differ considerably. On the genomic side, the quality of genome assemblies and annotation strongly affects the resolution of mapping. Many non-model species lack high-contiguity reference genomes or functional annotation for non-coding regions, limiting the ability to link AI-derived phenotypes to specific regulatory elements or structural variants (Schell et al., 2025). Progress in long-read sequencing, chromatin conformation capture, and single-cell transcriptomics may help bridge this gap by enabling more accurate identification of candidate loci and their functional context (Freedman and Sackton, 2024). Validation of newly discovered genes also presents a persistent challenge. Functional assays such as gene editing or transgenics are often unavailable in non-model systems, and pigment traits are frequently polygenic, making it difficult to isolate the effect of a single gene (Gudmunds et al., 2022; Wattad et al., 2024). In this context, the use of comparative genomics, expression profiling, and co-expression networks may serve as useful tools to build evidence for gene function in the absence of direct manipulation (Martin and Fraser, 2018; Ovens et al., 2021; Zogopoulos et al., 2022). Moreover, the

integration of species-specific interactomes into AI-genomic mapping frameworks could significantly enhance the prioritization of candidate genes (Ahi, 2025). By combining interaction networks with AI-based quantification and GWAS hits, it may become possible to identify pigmentation-relevant gene modules, infer functional pathways, and filter genes based on their network proximity to known pigment regulators (Figure 1). This systems-level context offers a valuable layer of biological interpretation, particularly when applied to taxa where experimental resources are limited.

Looking forward, several opportunities are emerging. The use of AI for real-time, field-based phenotyping may become feasible with the deployment of lightweight mobile imaging systems (Neethirajan and Kemp, 2021; Freitas et al., 2025; Hu et al., 2025). Transfer learning approaches could allow models trained on one species to be adapted to another with minimal retraining (Kutugata et al., 2021), making reproducibility and standardized analysis more reliable. In addition, the integration of multi-omic data, including spatial transcriptomics, chromatin accessibility, and metabolomics, could further enhance the functional interpretation of pigmentation-associated loci. As AI tools become more accessible and genomic resources continue to expand, pigmentation research stands to benefit from a more holistic and data-rich framework. This approach promises not only to identify new genes, but to contextualize them within broader networks of development, physiology, and evolution.

Box 1 | Key concepts in AI-enabled pigmentation genomics

Pigmentation genes (classical): Genes traditionally associated with pigment synthesis or pigment cell development, such as MC1R, TYR, ASIP, and MITF, often focused on melanin pathways.

Pigment cell types: Specialized chromatophores such as melanophores, xanthophores, iridophores, and erythrophores that produce or reflect color through pigments or nanostructures.

Non-classical pigmentation genes: Genes not historically linked to pigmentation but now implicated in color traits, often involved in signaling, membrane trafficking, or regulation.

AI-based phenotyping: The use of machine learning, especially deep learning, to quantify phenotypic traits such as color, pattern, and reflectance from image data.

Convolutional neural networks (CNNs): A deep learning architecture particularly suited to image recognition and feature extraction, widely used in trait analysis.

Self-supervised learning: A machine learning approach that learns structure from unlabeled data, enabling the identification of traits without predefined categories.

Image-based GWAS (iGWAS): A genomic association study that uses AI-extracted phenotypic features from images as input traits instead of manually scored measurements.

Triplet network: A machine learning model that compares trait similarity between images using three-image inputs, useful for pattern-based trait learning.

Retinal pigment score (RPS): A continuous measure of human retinal pigmentation derived from fundus images using AI-based color analysis.

Feather pigmentation quantification: The use of image segmentation or pixel classification to quantify melanin distribution across specific feather regions.

Phenotypic embedding: A numerical representation of trait variation derived from high-dimensional image data, used as input for downstream analysis.

Pattern recognition: An AI task that identifies and classifies visual patterns, used to analyze complex color and shape variation in natural traits.

Transfer learning: A method where a model trained on one dataset or species is adapted to another with minimal retraining, improving generalizability.

Trait dimensionality: The number and complexity of measurable aspects within a phenotype, which is greatly increased by AI-based feature extraction.

Regulatory variant: A genetic variant that influences gene expression rather than protein structure, often found in pigmentation loci identified by AI-assisted methods.

Systems-level trait: A phenotype governed by multiple layers of biological regulation, including gene networks, cell behavior, and environmental inputs.

Table 1. Summary of representative studies combining AI-based pigmentation phenotyping with genomic mapping.

Taxon/Species	Pigmentation Trait	AI/ML Method	Genomic Mapping Tool	Key Gene(s) Identified	Functional Role	Reference
Guppy fish	Male body coloration pattern	CNN pattern extraction	GWAS + CNV	Y-linked regions, novel autosomal loci	Chromatophore patterning, cell polarity	(van der Bijl et al., 2025)
Tianfu duck	Feather melanin distribution	DL segmentation	GWAS	DENND4A, PRKG1	Vesicle trafficking, signal transduction	(Twumasi et al., 2024; Wang et al., 2024)
RIR chick	Back stripe phenotype	Pattern classifier	GWAS	TMTC3	Transmembrane protein, development	(Shen et al., 2022)
Human (UK Biobank)	Retinal pigmentation	PCA + CNN embeddings	iGWAS	IFT122, PDE3A, SIK1	Ciliary transport, signaling	(Rajesh et al., 2025)
Bay scallop	Shell color variation	Color quantification	GWAS	PKS1, GRL101, PLC1	Polyketide synthesis, calcium signaling	(Zhu et al., 2021)

Cattle (Sumatran)	Coat color morphs	AI + SNP modeling	GWAS	CYFIP2 , SGSM1	Membrane dynamics, signaling	(Hartati et al., 2024)
Heliconius butterfly	Wing pattern geometry	Triplet network	QTL	Non-coding RNAs, regulatory elements	Pattern regulation	(Kronforst and Sheikh, 2023)
Papilio butterfly	Eye spot and scale color	Clustering analysis	Genomic scans	white , scarlet , lightoid	Pigment transporters	(Liu et al., 2021)

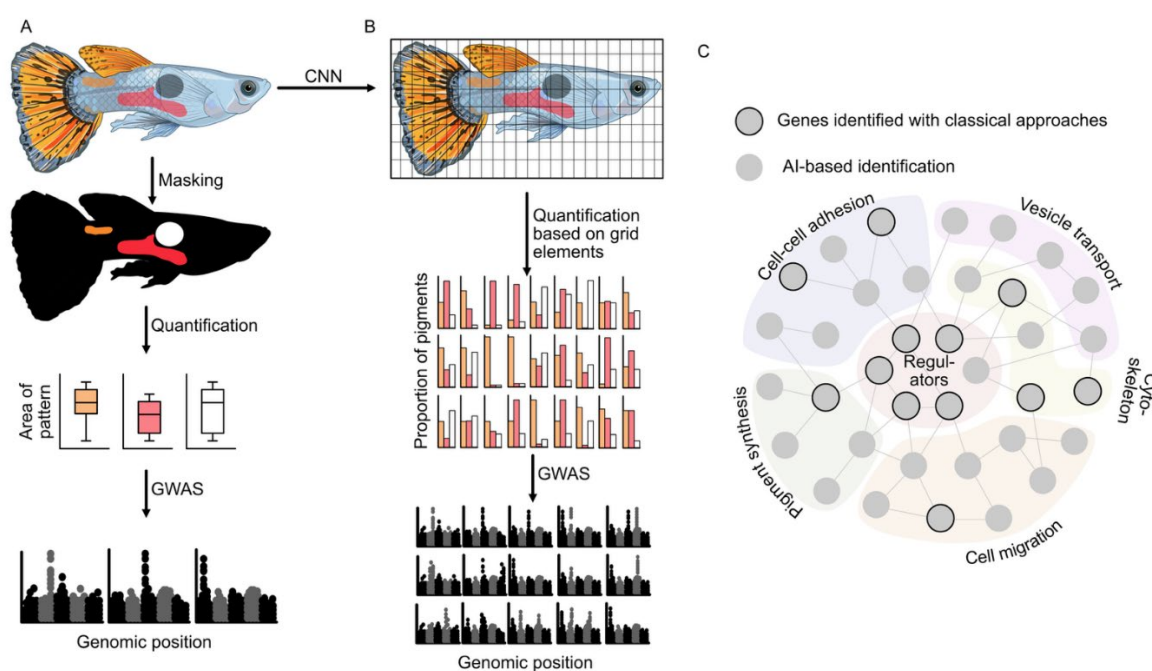


Figure 1. Exemplary comparison of classical approaches and AI-based methods: As exemplified here on a guppy pattern, AI-based methods show far greater throughput and potential to identify diverse gene sets involved in, for example, spatial patterning of color patterns. (A) Classical approach of pattern quantification, where elements of the pattern are masked and individually quantified to be used for GWA analysis. (B) Convolutional neural networks allow for a high-resolution map of a color pattern to identify distinct genetic elements that govern pattern formation and color across the body. (C) AI-based approaches with their high-throughput potential allow for the detection of previously unidentified genes and gene functions/pathways involved in pattern or color formation.

References

- Adhikari K, Mendoza-Revilla J, Sohail A, Fuentes-Guajardo M, Lampert J, Chacón-Duque JC, Hurtado M, Villegas V, Granja V, Acuña-Alonzo V, Jaramillo C, Arias W, Lozano RB, Everardo P, Gómez-Valdés J, Villamil-Ramírez H, Silva de Cerqueira CC, Hunemeier T, Ramallo V, Schuler-Faccini L, Salzano FM, Gonzalez-José R, Bortolini MC, Canizales-Quinteros S, Gallo C, Poletti G, Bedoya G, Rothhammer F, Tobin DJ, Fumagalli M, Balding D, Ruiz-Linares A. 2019. A GWAS in Latin Americans highlights the convergent evolution of lighter skin pigmentation in Eurasia. *Nat Commun* 2019 10:1–16. [Internet] 10:1–16.

- Available from: <https://www.nature.com/articles/s41467-018-08147-0>
2. Ahi EP. 2025. Fish Evo-Devo: Moving Toward Species-Specific and Knowledge-Based Interactome. *J Exp Zool Part B Mol Dev Evol* [Internet]. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1002/jez.b.23287>
 3. Ahi EP, Lecaudey LA, Ziegelbecker A, Steiner O, Glabonjat R, Goessler W, Hois V, Wagner C, Lass A, Sefc KM. 2020a. Comparative transcriptomics reveals candidate carotenoid color genes in an East African cichlid fish. *BMC Genomics* [Internet] 21:54. Available from: <https://bmcgenomics.biomedcentral.com/articles/10.1186/s12864-020-6473-8>
 4. Ahi EP, Lecaudey LA, Ziegelbecker A, Steiner O, Goessler W, Sefc KM. 2020b. Expression levels of the tetratricopeptide repeat protein gene *ttc39b* covary with carotenoid-based skin colour in cichlid fish. *Biol Lett* 16:20200629.
 5. Andrade P, Carneiro M. 2021. Pterin-based pigmentation in animals. *Biol Lett* [Internet] 17. Available from: <https://royalsocietypublishing.org/doi/10.1098/rsbl.2021.0221>
 6. Athanasopoulou K, Michalopoulou VI, Scorilas A, Adamopoulos PG. 2025. Integrating Artificial Intelligence in Next-Generation Sequencing: Advances, Challenges, and Future Directions. *Curr Issues Mol Biol* 2025, Vol 47, Page 470 [Internet] 47:470. Available from: <https://www.mdpi.com/1467-3045/47/6/470/htm>
 7. Bajpai VK, Swigut T, Mohammed J, Naqvi S, Arreola M, Tycko J, Kim TC, Pritchard JK, Bassik MC, Wysocka J. 2023. A genome-wide genetic screen uncovers determinants of human pigmentation. *Science* (80-) [Internet] 381. Available from: <https://www.science.org/doi/10.1126/science.ade6289>
 8. Baxter LL, Watkins-Chow DE, Pavan WJ, Loftus SK. 2019. A curated gene list for expanding the horizons of pigmentation biology. *Pigment Cell Melanoma Res* [Internet] 32:348–358. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/pcmr.12743>
 9. van der Bijl W, Shu JJ, Goberdhan VS, Sherin LM, Jia C, Cortazar-Chinarro M, Corral-Lopez A, Mank JE. 2025. Deep learning reveals the complex genetic architecture of male guppy colouration. *Nat Ecol Evol* 2025 [Internet]:1–12. Available from: <https://www.nature.com/articles/s41559-025-02781-w>
 10. Billah M, Bermann M, Hollifield MK, Tsuruta S, Chen CY, Psota E, Holl J, Misztal I, Lourenco D. 2025. Review: Genomic selection in the era of phenotyping based on digital images. *animal*:101486.
 11. Bush WS, Oetjens MT, Crawford DC. 2016. Unravelling the human genome–phenome relationship using phenome-wide association studies. *Nat Rev Genet* 2016 173 [Internet] 17:129–145. Available from: <https://www.nature.com/articles/nrg.2015.36>
 12. Butler MW, Toomey MB, McGraw KJ. 2011. How many color metrics do we need? Evaluating how different color-scoring procedures explain carotenoid pigment content in avian bare-part and plumage ornaments. *Behav Ecol Sociobiol* [Internet] 65:401–413. Available from: <https://link.springer.com/article/10.1007/s00265-010-1074-1>
 13. Cao J, Wenzel J, Zhang S, Lampe J, Wang H, Yao J, Zhang Z, Zhao S, Zhou Y, Chen C, Schwaninger M, Yang J, Chen DZ, Chen J. 2025. Rethinking deep learning in bioimaging through a data centric lens. *npj Imaging* 2025 31 [Internet] 3:1–7. Available from: <https://www.nature.com/articles/s44303-025-00092-0>
 14. Caro T, Mallarino R. 2020. Coloration in Mammals. *Trends Ecol Evol* [Internet] 35:357–366. Available from: <https://www.cell.com/action/showFullText?pii=S0169534719303532>
 15. Choi J, Zhang T, Vu A, Ablain J, Makowski MM, Colli LM, Xu M, Hennessey RC, Yin J, Rothschild H, Gräwe C, Kovacs MA, Funderburk KM, Brossard M, Taylor J, Pasaniuc B, Chari R, Chanock SJ, Hoggart CJ, Demenais F, Barrett JH, Law MH, Iles MM, Yu K, Vermeulen M, Zon LI, Brown KM. 2020. Massively parallel reporter assays of melanoma risk variants identify MX2 as a gene promoting melanoma. *Nat*

- Commun 2020 111 [Internet] 11:1–16. Available from: <https://www.nature.com/articles/s41467-020-16590-1>
16. Chong Y, Xiong H, Gao Z, Lu Y, Hong J, Wu J, He X, Xi D, Tu X, Deng W. 2024. Genomic and transcriptomic landscape to decipher the genetic basis of hyperpigmentation in Lanping black-boned sheep (*Ovis aries*). *BMC Genomics* [Internet] 25:1–13. Available from: <https://link.springer.com/articles/10.1186/s12864-024-10772-7>
 17. Chunhua Z, Le F, Shengli L, Sachula W, Bao H, Lan M, Antonini M, Haizhou S. 2025. Multi-omics and AI-driven advances in miRNA-mediated hair follicle regulation in cashmere goats. *Front Vet Sci* [Internet] 12:1635202. Available from: <https://www.frontiersin.org/articles/10.3389/fvets.2025.1635202/full>
 18. Crawford NG, Kelly DE, Hansen MEB, Beltrame MH, Fan S, Bowman SL, Jewett E, Ranciaro A, Thompson S, Lo Y, Pfeifer SP, Jensen JD, Campbell MC, Beggs W, Hormozdiari F, Mpoloka SW, Mokone GG, Nyambo T, Meskel DW, Belay G, Haut J, Rothschild H, Zon L, Zhou Y, Kovacs MA, Xu M, Zhang T, Bishop K, Sinclair J, Rivas C, Elliot E, Choi J, Li SA, Hicks B, Burgess S, Abnet C, Watkins-Chow DE, Oceana E, Song YS, Eskin E, Brown KM, Marks MS, Loftus SK, Pavan WJ, Yeager M, Chanock S, Tishkoff SA. 2017. Loci associated with skin pigmentation identified in African populations. *Science* (80-) [Internet] 358. Available from: <https://www.science.org/doi/10.1126/science.aan8433>
 19. Dingemans AJM, Hinne M, Truijien KMG, Goltstein L, van Reeuwijk J, de Leeuw N, Schuurs-Hoeijmakers J, Pfundt R, Diets IJ, den Hoed J, de Boer E, Coenen-van der Spek J, Jansen S, van Bon BW, Jonis N, Ockeloen CW, Vulto-van Silfhout AT, Kleefstra T, Koolen DA, Campeau PM, Palmer EE, Van Esch H, Lyon GJ, Alkuraya FS, Rauch A, Marom R, Baralle D, van der Sluijs PJ, Santen GWE, Kooy RF, van Gerven MAJ, Vissers LELM, de Vries BBA. 2023. PhenoScore: AI-based phenomics to quantify rare disease and genetic variation. *Nat Genet* [Internet] 55:1598. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC11414844/>
 20. Eizirik E, Trindade FJ. 2021. Genetics and Evolution of Mammalian Coat Pigmentation. *Annu Rev Anim Biosci* [Internet] 9:125–148. Available from: <https://www.annualreviews.org/content/journals/10.1146/annurev-animal-022114-110847>
 21. Elkin J, Martin A, Courtier-Orgogozo V, Santos ME. 2023. Analysis of the genetic loci of pigment pattern evolution in vertebrates. *Biol Rev Camb Philos Soc* [Internet] 98:1250–1277. Available from: <https://pubmed.ncbi.nlm.nih.gov/37017088/>
 22. Fandino RA, Brady NK, Chatterjee M, McDonald JMC, Livraghi L, van der Burg KRL, Mazo-Vargas A, Markenscoff-Papadimitriou E, Reed RD. 2024. The ivory lncRNA regulates seasonal color patterns in buckeye butterflies. *Proc Natl Acad Sci U S A* [Internet] 121:e2403426121. Available from: <https://www.pnas.org/doi/abs/10.1073/pnas.2403426121>
 23. Feiner N, Brun-Usan M, Andrade P, Pranter R, Park S, Menke DB, Geneva AJ, Uller T. 2022. A single locus regulates a female-limited color pattern polymorphism in a reptile. *Sci Adv* [Internet] 8. Available from: <https://www.science.org/doi/10.1126/sciadv.abm2387>
 24. Feng D, Li Q, Yu H, Kong L, Du S. 2018. Transcriptional profiling of long non-coding RNAs in mantle of *Crassostrea gigas* and their association with shell pigmentation. *Sci Reports* 2018 81 [Internet] 8:1–10. Available from: <https://www.nature.com/articles/s41598-018-19950-6>
 25. Ferguson JN, Fernandes SB, Monier B, Miller ND, Allen D, Dmitrieva A, Schmuker P, Lozano R, Valluru R, Buckler ES, Gore MA, Brown PJ, Spalding EP, Leakey ADB. 2021. Machine learning-enabled phenotyping for GWAS and TWAS of WUE traits in 869 field-grown sorghum accessions. *Plant Physiol* [Internet] 187:1481–1500. Available from: <https://doi.org/10.1093/plphys/kiab346>
 26. Fernandes AFA, Dórea JRR, Rosa GJ de M. 2020. Image Analysis and Computer Vision Applications in

- Animal Sciences: An Overview. *Front Vet Sci* [Internet] 7:551269. Available from: www.frontiersin.org
27. Figon F, Deravi LF, Casas J. 2021a. Barriers and Promises of the Developing Pigment Organelle Field. *Integr Comp Biol* [Internet] 61:1481–1489. Available from: <https://doi.org/10.1093/icb/icab164>
 28. Figon F, Hurbain I, Heiligenstein X, Trépout S, Lanoue A, Medjoubi K, Somogyi A, Delevoye C, Raposo G, Casas J. 2021b. Catabolism of lysosome-related organelles in color-changing spiders supports intracellular turnover of pigments. *Proc Natl Acad Sci U S A* [Internet] 118:e2103020118. Available from: <https://www.pnas.org/doi/abs/10.1073/pnas.2103020118>
 29. Freedman AH, Sackton TB. 2024. Rethinking eco-evo studies of gene expression for non-model organisms in the genomic era. *Mol Ecol* [Internet]:e17378. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/mec.17378>
 30. Freitas LA, Ferreira REP, Alves AAC, Dórea JRR, Paz CCP, Rosa GJM. 2025. Detection of anemic sheep using ocular conjunctiva images and deep learning algorithms. *Livest Sci* 294:105669.
 31. Galván I, Solano F. 2016. Bird Integumentary Melanins: Biosynthesis, Forms, Function and Evolution. *Int J Mol Sci* 2016, Vol 17, Page 520 [Internet] 17:520. Available from: <https://www.mdpi.com/1422-0067/17/4/520/htm>
 32. Ganguly K, Sengupta D, Sarkar N, Mukherjee N, Dutta T, Saha A, Saha T, Ghosh B, Chatterjee S, Brahmachari P, Kundu A, Sengupta M. 2022. Comprehensive in Silico Analyses of Single Nucleotide Variants of the Human Orthologues of 171 Murine Loci to Seek Novel Insights into the Genetics of Human Pigmentation. *Proc Zool Soc* [Internet] 75:361–380. Available from: <https://link.springer.com/article/10.1007/s12595-022-00449-y>
 33. Gerstenblith MR, Shi J, Landi MT. 2010. Genome-wide association studies of pigmentation and skin cancer: a review and meta-analysis. *Pigment Cell Melanoma Res* [Internet] 23:587–606. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/j.1755-148X.2010.00730.x>
 34. Gudmunds E, Wheat CW, Khila A, Husby A. 2022. Functional genomic tools for emerging model species. *Trends Ecol Evol* [Internet] 37:1104–1115. Available from: <https://www.cell.com/action/showFullText?pii=S0169534722001689>
 35. Hantak MM, Guralnick RP, Zare A, Stucky BJ. 2022. Computer vision for assessing species color pattern variation from web-based community science images. *iScience* [Internet] 25. Available from: <https://www.cell.com/action/showFullText?pii=S2589004222010562>
 36. Hartati H, Putra WPB, Handiwirawan E, Ramon E, Firison J, Zubir Z, Suretno ND, Mariyono M, Yusriani Y, Robba DK, Destomo A, Anggraeni T, Anwar P, Irmawanti S, Aprisal A, Elieser S, Kurniawati D. 2024. Genome-wide association study of genetic markers of coat color patterns in Sumatran native cattle. *Vet World* [Internet] 17:2537. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC11736373/>
 37. Hemingson CR, Cowman PF, Bellwood DR. 2024. Analysing biological colour patterns from digital images: An introduction to the current toolbox. *Ecol Evol* [Internet] 14:e11045. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1002/ece3.11045>
 38. Hendra C, Pratanwanich PN, Wan YK, Goh WSS, Thiery A, Göke J. 2022. Detection of m6A from direct RNA sequencing using a multiple instance learning framework. *Nat Methods* [Internet] 19:1590–1598. Available from: <https://pubmed.ncbi.nlm.nih.gov/36357692/>
 39. Heo S, Cho S, Dinh PTN, Park J, Jin DH, Cha J, Kim YK, Koh YJ, Lee SH, Lee JH. 2023. A genome-wide association study for eumelanin pigmentation in chicken plumage using a computer vision approach. *Anim Genet* [Internet] 54:355–362. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/age.13303>
 40. Hoekstra HE. 2006. Genetics, development and evolution of adaptive pigmentation in vertebrates.

- Heredity (Edinb) [Internet] 97:222–234. Available from: <https://pubmed.ncbi.nlm.nih.gov/16823403/>
41. Hoyal Cuthill JF, Guttenberg N, Ledger S, Crowther R, Huertas B. 2019. Deep learning on butterfly phenotypes tests evolution's oldest mathematical model. *Sci Adv* [Internet] 5:4967–4981. Available from: <https://www.science.org/doi/10.1126/sciadv.aaw4967>
 42. Hu T, Shen P, Zhang Y, Zhang J, Li X, Xia C, Liu P, Lu H, Wu T, Han Z. 2025. OpenPheno: an open-access, user-friendly, and smartphone-based software platform for instant plant phenotyping. *Plant Methods* [Internet] 21:1–12. Available from: <https://link.springer.com/articles/10.1186/s13007-025-01395-4>
 43. Ioannidis NM, Wang W, Furlotte NA, Hinds DA, Agee M, Alipanahi B, Auton A, Bell RK, Bryc K, Elson SL, Fontanillas P, Huber KE, Kleinman A, Litterman NK, McCreight JC, McIntyre MH, Mountain JL, Noblin ES, Northover CAM, Pitts SJ, Sathirapongsasuti JF, Sazonova O V., Shelton JF, Shringarpure S, Tian C, Tung JY, Vacic V, Wilson CH, Bustamante CD, Jorgenson E, Asgari MM, Whittemore AS. 2018. Gene expression imputation identifies candidate genes and susceptibility loci associated with cutaneous squamous cell carcinoma. *Nat Commun* 2018 91 [Internet] 9:1–9. Available from: <https://www.nature.com/articles/s41467-018-06149-6>
 44. Irion U, Nüsslein-Volhard C. 2019. The identification of genes involved in the evolution of color patterns in fish. *Curr Opin Genet Dev* 57:31–38.
 45. Jackson VE, Wu Y, Bonelli R, Owen JP, Scott LW, Farashi S, Kihara Y, Gantner ML, Egan C, Williams KM, Ansell BRE, Tufail A, Lee AY, Bahlo M. 2025. Multi-omic spatial effects on high-resolution AI-derived retinal thickness. *Nat Commun* 2025 161 [Internet] 16:1–19. Available from: <https://www.nature.com/articles/s41467-024-55635-7>
 46. Karagic N, Kratochwil CF. 2025. The next generation of colour pattern genomics. *Nat Ecol Evol* 2025 [Internet]:1–2. Available from: <https://www.nature.com/articles/s41559-025-02816-2>
 47. Kim B, Kim DS, Shin JG, Leem S, Cho M, Kim H, Gu KN, Seo JY, You SW, Martin AR, Park SG, Kim Y, Jeong C, Kang NG, Won HH. 2024. Mapping and annotating genomic loci to prioritize genes and implicate distinct polygenic adaptations for skin color. *Nat Commun* 2024 151 [Internet] 15:1–16. Available from: <https://www.nature.com/articles/s41467-024-49031-4>
 48. Kirchler M, Konigorski S, Norden M, Meltendorf C, Kloft M, Schurmann C, Lippert C. 2022. transferGWAS: GWAS of images using deep transfer learning. *Bioinformatics* [Internet] 38:3621–3628. Available from: <https://doi.org/10.1093/bioinformatics/btac369>
 49. Koblitz J, Reimer LC, Pukall R, Overmann J. 2025. Predicting bacterial phenotypic traits through improved machine learning using high-quality, curated datasets. *Commun Biol* 2025 81 [Internet] 8:1–13. Available from: <https://www.nature.com/articles/s42003-025-08313-3>
 50. Kondo S, Watanabe M, Miyazawa S. 2021. Studies of Turing pattern formation in zebrafish skin. *Philos Trans R Soc A* [Internet] 379. Available from: <https://royalsocietypublishing.org/doi/10.1098/rsta.2020.0274>
 51. Kratochwil CF, Mallarino R. 2023. Mechanisms Underlying the Formation and Evolution of Vertebrate Color Patterns. *Annu Rev Genet* [Internet] 57:135–156. Available from: <https://www.annualreviews.org/content/journals/10.1146/annurev-genet-031423-120918>
 52. Kronforst MR, Sheikh SI. 2023. New molecular insights into butterfly pigmentation. *Cell Rep* [Internet] 42. Available from: <https://www.cell.com/action/showFullText?pii=S2211124723009920>
 53. Kutugata M, Baumgardt J, Goolsby JA, Racelis AE, Kutugata M, Racelis AE, Baumgardt J, Goolsby A. 2021. Automatic Camera-Trap Classification Using Wildlife-Specific Deep Learning in Nilgai Management. *J Fish Wildl Manag* [Internet] 12:412–421. Available from: <https://doi.org/10.3996/JFWM-20-076>

54. Landi MT, Bishop DT, MacGregor S, Machiela MJ, Stratigos AJ, Ghiorzo P, Brossard M, Calista D, Choi J, Fargnoli MC, Zhang T, Rodolfo M, Trower AJ, Menin C, Martinez J, Hadjisavvas A, Song L, Stefanaki I, Scolyer R, Yang R, Goldstein AM, Potrony M, Kypreou KP, Pastorino L, Queirolo P, Pellegrini C, Cattaneo L, Zawistowski M, Gimenez-Xavier P, Rodriguez A, Elefanti L, Manoukian S, Rivoltini L, Smith BH, Loizidou MA, Del Regno L, Massi D, Mandala M, Khosrotehrani K, Akslen LA, Amos CI, Andresen PA, Avril MF, Azizi E, Soyer HP, Bataille V, Dalmasso B, Bowdler LM, Burdon KP, Chen W V., Codd V, Craig JE, Dębniak T, Falchi M, Fang S, Friedman E, Simi S, Galan P, Garcia-Casado Z, Gillanders EM, Gordon S, Green A, Gruis NA, Hansson J, Harland M, Harris J, Helsing P, Henders A, Hočevár M, Höiom V, Hunter D, Ingvar C, Kumar R, Lang J, Lathrop GM, Lee JE, Li X, Lubiński J, Mackie RM, Malt M, Malvey J, McAloney K, Mohamdi H, Molven A, Moses EK, Neale RE, Novaković S, Nyholt DR, Olsson H, Orr N, Fritsche LG, Puig-Butille JA, Qureshi AA, Radford-Smith GL, Randerson-Moor J, Requena C, Rowe C, Samani NJ, et al. 2020. Genome-wide association meta-analyses combining multiple risk phenotypes provide insights into the genetic architecture of cutaneous melanoma susceptibility. *Nat Genet* 2020 525 [Internet] 52:494–504. Available from: <https://www.nature.com/articles/s41588-020-0611-8>
55. Lay L, Khan W, Jo H, Kim SH, Kim Y. 2024. Genome-Wide Association Study on Cowpea seed coat color using RGB images. *Mol Breed* [Internet] 44:1–23. Available from: <https://link.springer.com/article/10.1007/s11032-024-01516-2>
56. Liu G, Liu W, Zhao R, He J, Dong Z, Chen L, Wan W, Chang Z, Wang W, Li X. 2021. Genome-wide identification and gene-editing of pigment transporter genes in the swallowtail butterfly *Papilio xuthus*. *BMC Genomics* [Internet] 22:1–18. Available from: <https://link.springer.com/articles/10.1186/s12864-021-07400-z>
57. Liu L, Wang X, Zhang R, Li H, Zhu H. 2022. Cell Junction and Vesicle Trafficking-Mediated Melanosome/Melanin Transfer Are Involved in the Dynamic Transformation of Goldfish *Carassius auratus* Skin Color. *Int J Mol Sci* [Internet] 23:12214. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC9603685/>
58. Livraghi L, Hanly JJ, Evans E, Wright CJ, Loh LS, Mazo-Vargas A, Kamrava K, Carter A, van der Heijden ESM, Reed RD, Papa R, Jiggins CD, Martin A. 2024. A long noncoding RNA at the cortex locus controls adaptive coloration in butterflies. *Proc Natl Acad Sci U S A* [Internet] 121:e2403326121. Available from: <https://www.pnas.org/doi/abs/10.1073/pnas.2403326121>
59. Lloyd VJ, Burg SL, Harizanova J, Garcia E, Hill O, Enciso-Romero J, Cooper RL, Flenner S, Longo E, Greving I, Nadeau NJ, Parnell AJ. 2024. The actin cytoskeleton plays multiple roles in structural colour formation in butterfly wing scales. *Nat Commun* 2024 151 [Internet] 15:1–15. Available from: <https://www.nature.com/articles/s41467-024-48060-3>
60. Lona-Durazo F, Hernandez-Pacheco N, Fan S, Zhang T, Choi J, Kovacs MA, Loftus SK, Le P, Edwards M, Fortes-Lima CA, Eng C, Huntsman S, Hu D, Gómez-Cabezas EJ, Marín-Padrón LC, Grauholm J, Mors O, Burchard EG, Norton HL, Pavan WJ, Brown KM, Tishkoff S, Pino-Yanes M, Beleza S, Marcheco-Teruel B, Parra EJ. 2019. Meta-analysis of GWA studies provides new insights on the genetic architecture of skin pigmentation in recently admixed populations. *BMC Genet* [Internet] 20:1–16. Available from: <https://link.springer.com/articles/10.1186/s12863-019-0765-5>
61. Lona-Durazo F, Mendes M, Thakur R, Funderburk K, Zhang T, Kovacs MA, Choi J, Brown KM, Parra EJ. 2021. A large Canadian cohort provides insights into the genetic architecture of human hair colour. *Commun Biol* [Internet] 4:1253. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC8568909/>
62. Luo M, Wang L, Yin H, Zhu W, Fu J, Dong Z. 2019. Integrated analysis of long non-coding RNA and mRNA expression in different colored skin of koi carp. *BMC Genomics* [Internet] 20:1–14. Available from:

- <https://link.springer.com/articles/10.1186/s12864-019-5894-8>
63. Lürig MD, Donoughe S, Svensson EI, Porto A, Tsuboi M. 2021. Computer Vision, Machine Learning, and the Promise of Phenomics in Ecology and Evolutionary Biology. *Front Ecol Evol* [Internet] 9:642774. Available from: www.frontiersin.org
 64. Mao J, Zeng Q, Yang Z, Pan H, Yao L, Bao Z, Wang C, Wang S. 2020. High-resolution linkage and quantitative trait locus mapping using an interspecific cross between *Argopecten irradians irradians* (♀) and *A. purpuratus* (♂). *Mar Life Sci Technol* [Internet] 2:123–134. Available from: <https://link.springer.com/article/10.1007/s42995-020-00029-z>
 65. Marin-Recinos MF, Pucker B. 2024. Genetic factors explaining anthocyanin pigmentation differences. *BMC Plant Biol* 2024 241 [Internet] 24:1–20. Available from: <https://link.springer.com/articles/10.1186/s12870-024-05316-w>
 66. Martin AR, Lin M, Granka JM, Myrick JW, Liu X, Sockell A, Atkinson EG, Werely CJ, Möller M, Sandhu MS, Kingsley DM, Hoal EG, Liu X, Daly MJ, Feldman MW, Gignoux CR, Bustamante CD, Henn BM. 2017. An Unexpectedly Complex Architecture for Skin Pigmentation in Africans. *Cell* [Internet] 171:1340–1353.e14. Available from: <https://www.cell.com/action/showFullText?pii=S009286741713247>
 67. Martin T, Fraser HB. 2018. Comparative expression profiling reveals widespread coordinated evolution of gene expression across eukaryotes. *Nat Commun* 2018 91 [Internet] 9:1–9. Available from: <https://www.nature.com/articles/s41467-018-07436-y>
 68. McNamara ME, Rossi V, Slater TS, Rogers CS, Ducrest AL, Dubey S, Roulin A. 2021. Decoding the Evolution of Melanin in Vertebrates. *Trends Ecol Evol* [Internet] 36:430–443. Available from: <https://www.cell.com/action/showFullText?pii=S016953472030375X>
 69. Mills MG, Patterson LB. 2009. Not just black and white: Pigment pattern development and evolution in vertebrates. *Semin Cell Dev Biol* 20:72–81.
 70. Mulero-Pázmány M, Hurtado S, Barba-González C, Antequera-Gómez ML, Díaz-Ruiz F, Real R, Navas-Delgado I, Aldana-Montes JF. 2025. Addressing significant challenges for animal detection in camera trap images: a novel deep learning-based approach. *Sci Reports* 2025 151 [Internet] 15:1–18. Available from: <https://www.nature.com/articles/s41598-025-90249-z>
 71. Nabwire S, Suh HK, Kim MS, Baek I, Cho BK. 2021. Review: Application of Artificial Intelligence in Phenomics. *Sensors* 2021, Vol 21, Page 4363 [Internet] 21:4363. Available from: <https://www.mdpi.com/1424-8220/21/13/4363/htm>
 72. Neethirajan S, Kemp B. 2021. Digital Phenotyping in Livestock Farming. *Anim* 2021, Vol 11, Page 2009 [Internet] 11:2009. Available from: <https://www.mdpi.com/2076-2615/11/7/2009/htm>
 73. Newell C, Walker H, Caro T. 2021. Pig pigmentation: testing Gloger's rule. *J Mammal* [Internet] 102:1525–1535. Available from: <https://doi.org/10.1093/jmammal/gyab090>
 74. Niu J, Li T, Qi K, Liu Y, Deng H, Hu Y, Xu D, Wu L, Amevor FK, Wang Y, Shu G, Zhao X. 2025. Research Note: Application of Convolutional Neural Networks for Feather Classification in Chickens. *Poult Sci*:105254.
 75. Norman DL, Bischoff PH, Wearn OR, Ewers RM, Rowcliffe JM, Evans B, Sethi S, Chapman PM, Freeman R. 2023. Can CNN-based species classification generalise across variation in habitat within a camera trap survey? *Methods Ecol Evol* [Internet] 14:242–251. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/2041-210X.14031>
 76. Okuley OS, Aiello CM, Glad W, Perkins K, Ianniello R, Darby N, Epps CW. 2025. Improving AI performance in wildlife monitoring through species and environment-specific training: A case study on desert Bighorn sheep. *Ecol Inform* 89:103179.

77. Orouji S, Liu MC, Korem T, Peters MAK. 2024. Domain adaptation in small-scale and heterogeneous biological datasets. *Sci Adv* [Internet] 10. Available from: <https://pubmed.ncbi.nlm.nih.gov/39705361/>
78. Ovens K, Eames BF, McQuillan I. 2021. Comparative Analyses of Gene Co-expression Networks: Implementations and Applications in the Study of Evolution. *Front Genet* [Internet] 12:695399. Available from: www.frontiersin.org
79. Parichy DM. 2021. Evolution of pigment cells and patterns: recent insights from teleost fishes. *Curr Opin Genet Dev* 69:88–96.
80. Patterson LB, Parichy DM. 2019. Zebrafish Pigment Pattern Formation: Insights into the Development and Evolution of Adult Form. *Annu Rev Genet* [Internet] 53:505–530. Available from: <https://www.annualreviews.org/content/journals/10.1146/annurev-genet-112618-043741>
81. Podobnik M, Frohnhöfer HG, Dooley CM, Eskova A, Nüsslein-Volhard C, Irion U. 2020. Evolution of the potassium channel gene *Kcnj13* underlies colour pattern diversification in Danio fish. *Nat Commun* 2020 111 [Internet] 11:1–13. Available from: <https://www.nature.com/articles/s41467-020-20021-6>
82. Qi Z, Liu J, Shi J, Yin M, Liu J, Fan J, Bao Z, Ye Z, Hu J. 2025. Integrated Transcriptomic and Epigenomic Analysis Reveals Mechanisms Underlying Melanotic Spot Formation in Red Tilapia (*Oreochromis* spp.). *Int J Mol Sci* [Internet] 26:4370. Available from: <https://www.mdpi.com/1422-0067/26/9/4370/htm>
83. Quillen EE, Norton HL, Parra EJ, Lona-Durazo F, Ang KC, Illiescu FM, Pearson LN, Shriver MD, Lasisi T, Gokcumen O, Starr I, Lin YL, Martin AR, Jablonski NG. 2019. Shades of complexity: New perspectives on the evolution and genetic architecture of human skin. *Am J Phys Anthropol* [Internet] 168:4–26. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1002/ajpa.23737>
84. Rajesh AE, Olvera-Barrios A, Warwick AN, Wu Y, Stuart K V., Biradar MI, Ung CY, Khawaja AP, Luben R, Foster PJ, Cleland CR, Makupa WU, Denniston AK, Burton MJ, Bastawrous A, Keane PA, Chia MA, Turner AW, Lee CS, Tufail A, Lee AY, Egan C, Zheng Y, Yates M, Woodside J, Williams C, Williams K, Weedon M, Vitart V, Viswanathan A, Trucco E, Thomas D, Tapp R, Sun Z, Sudlow C, Strouthidis N, Stratton I, Steel D, Sivaprasad S, Sergouniotis P, Self J, Sattar N, Rudnicka A, Rahi J, Pontikos N, Petzold A, Peto T, Paterson E, Patel P, Owen C, Oram R, O’Sullivan E, Morgan J, Moore T, McKibbin M, McKay G, McGuinness B, Madhusudhan S, Mackie S, MacGillivray T, Luthert P, Lotery A, Littlejohns T, Lascaratos G, Hysi P, Hogg R, Harding S, Hardcastle A, Hammond C, Guggenheim J, Gibson J, Garway-Heath DT, Gallacher J, Ennis S, Doney A, Dick A, Dhillon B, Desai P, Day A, Chua S, Chan M, Chakravarthy U, Carare R, Braithwaite T, Black G, Bishop P, Barrett J, Barman S, Balaskas K, Atan D, Aslam T, Allen N. 2025. Machine learning derived retinal pigment score from ophthalmic imaging shows ethnicity is not biology. *Nat Commun* 2024 161 [Internet] 16:1–14. Available from: <https://www.nature.com/articles/s41467-024-55198-7>
85. Rosenberg AM, Rausser S, Ren J, Mosharov E V., Sturm G, Ogden RT, Patel P, Soni RK, Lacefield C, Tobin DJ, Paus R, Picard M. 2021. Quantitative mapping of human hair greying and reversal in relation to life stress. *Elife* 10.
86. Saunders LM, Mishra AK, Aman AJ, Lewis VM, Toomey MB, Packer JS, Qiu X, McFaline-Figueroa JL, Corbo JC, Trapnell C, Parichy DM. 2019. Thyroid hormone regulates distinct paths to maturation in pigment cell lineages. *Elife* 8.
87. Schell T, Greve C, Podsiadlowski L. 2025. Establishing genome sequencing and assembly for non-model and emerging model organisms: a brief guide. *Front Zool* 2025 221 [Internet] 22:1–26. Available from: <https://link.springer.com/articles/10.1186/s12983-025-00561-7>
88. Shawkey MD, D’Alba L. 2017. Interactions between colour-producing mechanisms and their effects on the integumentary colour palette. *Philos Trans R Soc B Biol Sci* [Internet] 372. Available from:

- <https://royalsocietypublishing.org/doi/10.1098/rstb.2016.0536>
89. Shen Q, Zhou J, Li J, Zhao X, Zheng L, Bao H, Wu C. 2022. Genome-Wide Association Study Identifies Candidate Genes for Stripe Pattern Feather Color of Rhode Island Red Chicks. *Genes (Basel)* [Internet] 13:1511. Available from: <https://www.mdpi.com/2073-4425/13/9/1511/htm>
 90. Siegenthaler A, Mondal D, Benvenuto C. 2017. Quantifying pigment cover to assess variation in animal colouration. *Biol Methods Protoc* [Internet] 2:bp003. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC6994029/>
 91. Simcoe M, Valdes A, Liu F, Furlotte NA, Evans DM, Hemani G, Ring SM, Smith GD, Duffy DL, Zhu G, Gordon SD, Medland SE, Vuckovic D, Girotto G, Sala C, Catamo E, Concas MP, Brumat M, Gasparini P, Toniolo D, Cocca M, Robino A, Yazar S, Hewitt A, Wu W, Kraft P, Hammond CJ, Shi Y, Chen Y, Zeng C, Klaver CCW, Uitterlinden AG, Ikram MA, Hamer MA, van Duijn CM, Nijsten T, Han J, Mackey DA, Martin NG, Cheng CY, Hinds DA, Spector TD, Kayser M, Hysi PG. 2021. Genome-wide association study in almost 195,000 individuals identifies 50 previously unidentified genetic loci for eye color. *Sci Adv* [Internet] 7:24. Available from: <https://www.science.org/doi/10.1126/sciadv.abd1239>
 92. Song S, Wang L, Hou L, Liu JS. 2024. Partitioning and aggregating cross-tissue and tissue-specific genetic effects to identify gene-trait associations. *Nat Commun* 2024 151 [Internet] 15:1–12. Available from: <https://www.nature.com/articles/s41467-024-49924-4>
 93. Strowbridge N, Vieites DR, Ritchie MG, Elmer KR. 2025. Contributions of epigenomic and epitranscriptomic methylation to animal colouration. *Trends Genet* [Internet] 0. Available from: <https://www.cell.com/action/showFullText?pii=S0168952525000988>
 94. Stuckert AMM, Moore E, Coyle KP, Davison I, MacManes MD, Roberts R, Summers K. 2019. Variation in pigmentation gene expression is associated with distinct aposematic color morphs in the poison frog *Dendrobates auratus*. *BMC Evol Biol* [Internet] 19. Available from: <https://pubmed.ncbi.nlm.nih.gov/30995908/>
 95. Tabak MA, Norouzzadeh MS, Wolfson DW, Sweeney SJ, Vercauteren KC, Snow NP, Halseth JM, Di Salvo PA, Lewis JS, White MD, Teton B, Beasley JC, Schlichting PE, Boughton RK, Wight B, Newkirk ES, Ivan JS, Odell EA, Brook RK, Lukacs PM, Moeller AK, Mandeville EG, Clune J, Miller RS. 2019. Machine learning to classify animal species in camera trap images: Applications in ecology. *Methods Ecol Evol* [Internet] 10:585–590. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/2041-210X.13120>
 96. Toews DPL, Hofmeister NR, Taylor SA. 2017. The Evolution and Genetics of Carotenoid Processing in Animals. *Trends Genet* [Internet] 33:171–182. Available from: <https://www.cell.com/action/showFullText?pii=S0168952517300021>
 97. Tripathi D, Bhattacharyya C, Basu A. 2024. Deep learning insights into distinct patterns of polygenic adaptation across human populations. *Nucleic Acids Res* [Internet] 52:e102–e102. Available from: <https://doi.org/10.1093/nar/gkae1027>
 98. Twumasi G, Wang H, Xi Y, Qi J, Li L, Bai L, Liu H. 2024. Genome-Wide Association Studies Reveal Candidate Genes Associated with Pigmentation Patterns of Single Feathers of Tianfu Nonghua Ducks. *Animals* [Internet] 14:85. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC10778472/>
 99. Ullate-Agote A, Burgelin I, Debry A, Langrez C, Montange F, Peraldi R, Daraspe J, Kaessmann H, Milinkovitch MC, Tzika AC. 2020. Genome mapping of a LYST mutation in corn snakes indicates that vertebrate chromatophore vesicles are lysosome-related organelles. *Proc Natl Acad Sci U S A* [Internet] 117:26307–26317. Available from: <https://www.pnas.org/doi/abs/10.1073/pnas.2003724117>
 100. Upadhyay VR, Ramesh V, Kumar H, Somagond YM, Priyadarsini S, Kuniyal A, Prakash V, Sahoo A. 2024. Phenomics in Livestock Research: Bottlenecks and Promises of Digital Phenotyping and Other

- Quantification Techniques on a Global Scale. *OMICS* [Internet] 28:380–393. Available from: <https://europepmc.org/article/med/39012961>
101. Vöcking O, Macias-Muñoz A, Jaeger SJ, Oakley TH. 2022. Deep Diversity: Extensive Variation in the Components of Complex Visual Systems across Animals. *Cells* [Internet] 11:3966. Available from: <https://www.mdpi.com/2073-4409/11/24/3966/htm>
 102. Wakamatsu K, Ito S. 2021. Melanins in Vertebrates. *Pigment Pigment Cells Pigment Patterns* [Internet]:45–89. Available from: https://link.springer.com/chapter/10.1007/978-981-16-1490-3_2
 103. Wang H, Twumasi G, Xu Q, Xi Y, Qi J, Yang Z, Shen Z, Bai L, Li L, Liu H. 2024. Identification of candidate genes associated with primary feathers of tianfu nonghua ducks based on Genome-wide association studies. *Poult Sci* 103:103985.
 104. Wattad H, Molcho J, Manor R, Weil S, Aflalo ED, Chalifa-Caspi V, Sagi A. 2024. Roadmap and Considerations for Genome Editing in a Non-Model Organism: Genetic Variations and Off-Target Profiling. *Int J Mol Sci* [Internet] 25:12530. Available from: <https://www.mdpi.com/1422-0067/25/23/12530/htm>
 105. Woronik A, Tunström K, Perry MW, Neethiraj R, Stefanescu C, Celorio-Mancera M de la P, Brattström O, Hill J, Lehmann P, Käkelä R, Wheat CW. 2019. A transposable element insertion is associated with an alternative life history strategy. *Nat Commun* 2019 101 [Internet] 10:1–11. Available from: <https://www.nature.com/articles/s41467-019-13596-2>
 106. Wu S, Chang CM, Mai GS, Rubenstein DR, Yang CM, Huang YT, Lin HH, Shih LC, Chen SW, Shen SF. 2019. Artificial intelligence reveals environmental constraints on colour diversity in insects. *Nat Commun* 2019 101 [Internet] 10:1–9. Available from: <https://www.nature.com/articles/s41467-019-12500-2>
 107. Wu Y, Xie L. 2025. AI-driven multi-omics integration for multi-scale predictive modeling of genotype-environment-phenotype relationships. *Comput Struct Biotechnol J* 27:265–277.
 108. Xie Z, Zhang T, Kim S, Lu J, Zhang W, Lin CH, Wu MR, Davis A, Channa R, Giancardo L, Chen H, Wang S, Chen R, Zhi D. 2024. iGWAS: Image-based genome-wide association of self-supervised deep phenotyping of retina fundus images. *PLOS Genet* [Internet] 20:e1011273. Available from: <https://journals.plos.org/plosgenetics/article?id=10.1371/journal.pgen.1011273>
 109. Xu XD, Zhou Y, Wang CQ, Huang X, Zhang K, Xu XW, He LW, Zhang XY, Fu XZ, Ma M, Qin QB, Liu SJ. 2024. Identification and effective regulation of *scarb1* gene involved in pigmentation change in autotetraploid *Carassius auratus*. *Zool Res* [Internet] 45:381. Available from: <https://pmc.ncbi.nlm.nih.gov/articles/PMC11017083/>
 110. Yang SJ, Berndl M, Ando DM, Barch M, Narayanaswamy A, Christiansen E, Hoyer S, Roat C, Hung J, Rueden CT, Shankar A, Finkbeiner S, Nelson P. 2018. Assessing microscope image focus quality with deep learning. *BMC Bioinformatics* [Internet] 19:1–9. Available from: <https://link.springer.com/articles/10.1186/s12859-018-2087-4>
 111. Ying W. 2023. Phenomic Studies on Diseases: Potential and Challenges. *Phenomics* 2023 33 [Internet] 3:285–299. Available from: <https://link.springer.com/article/10.1007/s43657-022-00089-4>
 112. Zhang J, Naik HS, Assefa T, Sarkar S, Reddy RVC, Singh A, Ganapathysubramanian B, Singh AK. 2017. Computer vision and machine learning for robust phenotyping in genome-wide studies. *Sci Reports* 2017 71 [Internet] 7:1–11. Available from: <https://www.nature.com/articles/srep44048>
 113. Zhang S, Hu X, Miao H, Chu Y, Cui F, Yang W, Xu S, Guo J, Fu C, Song X, Hou M, Qiu J, Chen J. 2021. Imaged-based phenotyping accelerated QTL mapping and qtl × environment interaction analysis of testa colour in peanut (*Arachis hypogaea*). *Plant Breed* [Internet] 140:884–895. Available from: <https://onlinelibrary.wiley.com/doi/full/10.1111/pbr.12905>

114. Zhang T, Choi J, Kovacs MA, Shi J, Xu M, Consortium MMA, Goldstein AM, Trower AJ, Bishop DT, Iles MM, Duffy DL, MacGregor S, Amundadottir LT, Law MH, Loftus SK, Pavan WJ, Brown KM, Barnabas BB, Bouffard GG, Brooks SY, Coleman H, Dekhtyar L, Guan X, Han J, Ho SL, Legaspi R, Maduro QL, Masiello CA, McDowell JC, Montemayor C, Mullikin JC, Park M, Riebow NL, Schandler K, Schmidt B, Sison C, Smith R, Stantripop S, Thomas JW, Thomas PJ, Vemulapalli M, Young AC. 2018. Cell-type-specific eQTL of primary melanocytes facilitates identification of melanoma susceptibility genes. *Genome Res* 28:1621–1635.
115. Zhu X, Zhang J, Hou X, Liu P, Lv J, Xing Q, Huang X, Hu J, Bao Z. 2021. A Genome-Wide Association Study Identifies Candidate Genes Associated With Shell Color in Bay Scallop *Argopecten irradians irradians*. *Front Mar Sci* [Internet] 8:742330. Available from: www.frontiersin.org
116. Zogopoulos VL, Saxami G, Malatras A, Papadopoulos K, Tsotra I, Iconomidou VA, Michalopoulos I. 2022. Approaches in Gene Coexpression Analysis in Eukaryotes. *Biology (Basel)* [Internet] 11:1019. Available from: <https://www.mdpi.com/2079-7737/11/7/1019/htm>

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.