

Review

Not peer-reviewed version

---

# Microbiome-Gut-Mucosal-Immune-Brain Axis and Autism Spectrum Disorder (ASD): A Novel Proposal of the Role of Gut Microbiome in ASD Aetiology

---

[Amapola De Sales-Millán](#) , [José Félix Aguirre-Garrido](#) , [Rina María González-Cervantes](#) , [Jose Antonio Velázquez-Aragón](#) \*

Posted Date: 17 May 2023

doi: 10.20944/preprints202305.1223.v1

Keywords: Microbiome-Gut-Brain axis; ASD; dysbiosis; gastrointestinal functions; Immune System; and Neuroimmunogastroenterology



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Review*

# Microbiome-Gut-Mucosal-Immune-Brain Axis and Autism Spectrum Disorder (ASD): A Novel Proposal of the Role of Gut Microbiome in ASD Aetiology

Amapola De Sales-Millán <sup>1</sup>, José Félix Aguirre-Garrido <sup>2</sup>, Rina María González-Cervantes <sup>2</sup>  
and Jose Antonio Velázquez-Aragón <sup>3,\*</sup>

<sup>1</sup> División de Ciencias Biológicas y de la Salud, Universidad Autónoma Metropolitana-Lerma, Estado de México 52006 Lerma, México

<sup>2</sup> Departamento de Ciencias Ambientales, Universidad Autónoma Metropolitana-Lerma, Estado de México 52006 Lerma, México

<sup>3</sup> Experimental Oncology Laboratory, National Institute of Pediatrics, Mexico

\* Correspondence: jvelazqueza@pediatria.gob.mx

**Abstract:** Autism Spectrum Disorder (ASD) is a complex neurodevelopmental disorder characterised by poor social interaction and communication, as well as restricted and stereotyped interests. Due of the high prevalence of gastrointestinal disorders in individuals with ASD, researchers have investigated the gut microbiota as a potential contributor to its aetiology. The relationship between the microbiome, gut, and brain (microbiome-gut-brain axis) has been acknowledged as a key factor in modulating brain function and social behaviour, but its connection to the aetiology of ASD is not well understood. Recently, there has been increasing attention on the relationship between the immune system, gastrointestinal disorders, and neurological issues in ASD, particularly in relation to the loss of specific species or a decrease in microbial diversity. It focuses on how gut microbiota dysbiosis can affect gut permeability, immune function, and microbiota metabolites in ASD. Although a very complete study suggests that dysbiosis is a consequence of the disease and that it has practically no effect on autistic manifestations. This is a review of the relationship between the immune system, microbial diversity, and the microbiome-gut-brain axis in the development of autistic symptoms severity and, a proposal of a novel role of gut microbiome in ASD, where dysbiosis is a consequence of ASD-related behaviour and where dysbiosis in turn accentuates the autistic manifestations of the patients via microbiome-gut-brain axis in a feedback circuit.

**Keywords:** microbiome-gut-brain axis; ASD; dysbiosis; gastrointestinal functions; immune system; neuroimmunogastroenterology

## 1. Introduction

Autism spectrum disorder (ASD) refers to a complex neurodevelopmental disorder characterised by poor social interaction and communication, as well as restricted and stereotyped interests [1]. ASD has a significant financial and health impact on individuals with autism, their families, and society [2]. The cause of ASD is not yet fully understood, but research suggests that a combination of genetic and environmental factors are involved [3]. Different reports propose that, in addition to the core symptoms of ASD, gastrointestinal symptoms, including constipation, abdominal pain, vomiting, diarrhea, and gas, are frequent in people with ASD, with estimates ranging from 9% to 70% [4]. The gut microbiome is a crucial component of human physiology [5], and the frequent gastrointestinal symptoms in individuals with ASD have led to the investigation of a possible connection between gut microbiota and the symptoms of the disorder.

There is increasing evidence supporting the importance of gut microbiota in brain development and function, showing communication along the gut-brain axis [6]. This association is of vital importance within the first 3 years of life as children's brains are developing rapidly at this stage [7] and there are also changes in their gut microbiota as they grow until they are older establish their feeding patterns [8,9]. Recently, the focus has been expanded to the role of the gut microbiota [10],

generating increased interest with findings suggesting specific microorganisms associated with memory, stress, mood, neurodevelopment, and even ASD [11–13].

The gut-microbiome-brain axis is a multidisciplinary research area that has attracted the attention of different researchers from around the world. Multiple studies have found evidence that the gut-microbiome-brain axis is important for the mental and cognitive health of children with ASD [14–16]. Despite this, there have been relatively few clinical studies in humans that provide clear evidence of the role in the aetiology of neurodevelopmental disorders [17,18]. On the other hand, a paper published in 2021, Yap, et al., [19] establishes that the microbiota does not participate in autistic manifestations and that gut dysbiosis is a consequence of central symptoms in ASD.

Understanding the role of the gut-microbiome-brain axis in the development of ASD is crucial to determining its potential as a therapeutic target. This is particularly important because faecal transplants and pre/probiotic interventions have already been used to treat patients with ASD.

## 2. Gut Microbiota

The human gut microbiota is composed of  $10^{13}$  to  $10^{14}$  microorganisms, mostly bacteria [5]. Their collective genome is defined as a gut microbiome [20]. The number of microorganisms varies throughout the intestinal tract, with 95% of symbiotic microorganisms (the largest microecosystem in the human body) residing in the intestine [21]. They are represented by anaerobic bacteria belonging, in order of abundance, to two phyla: Bacteroidetes and Firmicutes, accounting for approximately 90% of the bacterial species [22]; In addition to bacteria belonging to the Proteobacteria, Actinobacteria and Verrucomicrobia phyla, the gut microbiota also includes viruses, fungi, and archaea (mainly in pathological conditions), maintaining symbiotic relationship with the host [23].

The resident microbial community of the human gut has a unique combination of different types and numbers of bacteria that are constantly evolving and can be influenced by diet, environmental factors, drug treatments, and age, and between many other factors. In the European Metagenomics of the Human Intestinal Tract (MetaHit) project, it was possible to identify and characterize three different groups of microbiota or enterotypes [24]. Intercontinentally, enterotype 1 was dominated by the genus *Bifidobacterium*, enterotype 2 was represented by the genus *Prevotella*, and enterotype 3 was represented by the genus *Bacteroides* [25].

The Firmicutes/Bacteroidetes ratio changes with age, with a ratio of approximately 0.4 in infants and up to 10.9 in adults [26]. Imbalances in their abundance cause dysbiosis [27–31]. Dysbiosis is generally associated with pathological conditions such as obesity, type 2 diabetes, cancer, gastrointestinal diseases, and neuropsychiatric and neurodevelopmental diseases [5,21]. However, the composition and functional characteristics of a healthy microbiome remain to be defined.

### 2.1. Functions of Gut Microbiota

The symbiotic relationship between humans and the microbiota has allowed the development of characteristics that play a fundamental role in biological processes, such as nutrient usage, host metabolism, protection against infections, and maturation of the immune system [32,33]; Brain development and behaviour are also part of this symbiotic relationship, as gut microbiota interacts with neuroendocrine, neuroimmune, and autonomic nervous systems (ANS) [5].

The gut microbiota is considered as one of the key elements that contribute to the regulation of host health. In healthy patients, the gut microbiota tends towards homeostasis by resisting change during ecological stress (resistance) or returning to a state of equilibrium after a stress-related disturbance (resilience) [34]. Numerous molecular mechanisms explain how the intestinal microbiota may be causally related to the protection or appearance of disease [21]. For example, celiac disease and inflammatory bowel disorder in infants are related to an increase in several bacterial species, such as *Dialister invisus*, *Parabacteroides* spp., or *Lachnospiraceae* and certain metabolites such as tryptophan, before the disease appears. Conversely, several anti-inflammatory strains, such as *Faecalibacterium prausnitzii* and *Clostridium clostridioforme*, are decreased [35].

The routes of communication between the microbiota and the brain are constantly being discovered, including the vagus nerve, intestinal hormone signalling, the immune system, and tryptophan metabolites or through bacterial metabolites such as short-chain fatty acids (SCFA), which they include propionate, butyrate, and acetate [36]. Butyrate is an epigenetic modulator that acts through histone deacetylases [37]. The gut microbiota also regulates central neurotransmitters by altering the levels of their precursors (Table 1).

**Table 1.** Neurotransmitter synthesis and release altered by dysbiotic gut microbiome in ASD.

Neurotransmitters*	Taxon name of neurotransmitter involved microorganism	Effect on r level in dysbiosis in ASD.	Role of microorganisms in the human body.	References
GABA/Acetylcholine Noradrenaline (norepinephrine)/Dopamine	(g) <i>Lactobacillus</i> sp. <sup>1</sup>	Decrease ↓	Improve the brain function and elevated mood.	[36,61,144]
GABA	(g) <i>Bifidobacterium</i> <sup>1</sup>	Decrease ↓	Regulates emotions and behavior: Maintain gut homeostasis, produce vitamins and antimicrobial substances, and regulates the host immune system.	[30,36,144]
Noradrenaline (norepinephrine)/Serotonin	(g) <i>Escherichia</i> <sup>1</sup>	Decrease ↓	Produce active molecules that may reach and influence the CNS after the secretion into the periphery or by activating afferent neurons.	[12,36]
Noradrenaline (norepinephrine)	(g) <i>Saccharomyces</i> sp. <sup>2</sup>	Increase ↑	Involved in ASD pathogenesis through immune factors and may play an essential role in the development of ASD.	[36,145]
Serotonin	(g) <i>Candida</i> <sup>2</sup>	Increase ↑	In a dysbiotic environment as frequently observed in the autistic population, the yeast proliferates and produces ammonia and toxins, which	[12,36,123]

			increase autistic behavior.
Tryptophan	(g) <i>Streptococcus</i> sp. <sup>1</sup>	Decrease ↓	Protects tissues from oxidative stress. [12,36,144]
	(g) <i>Enterococcus</i> sp. <sup>1</sup>	Decrease ↓	DNA damage in colorectal cancer. [12,36,144]
	(g) <i>Clostridium</i> <sup>1</sup>	Increase ↑	Increases the production of antioxidant and neuro-protectant molecules inside the gut; acts as a biomarker for ASD; inhibits the growth of other gut microbiota, promotes the growth or virulence of gut pathogens. [115,144]

1 Bacteria. 2 Fungi. (g): genus. ↑Increase. ↓Decrease. \*These neurotransmitters produced by bacteria and fungi can cross the intestinal barrier, and potentially play a role in regulating brain functions (neurogenesis, neurodevelopment, mental homeostasis, background emotions and immune response modulation).

2.2. Factors Influencing Gut Microbiota

The transmission of strains (*Prevotella copri*) from mother to child can occur vertically in the first days of life, but the acquisition of strains from a shared environment over time cannot be ruled out, producing host-dependent co-diversification patterns [38]. During the developmental stages of the new-born, from birth to complementary feeding, the complexity and richness of the microbial community fluctuates [39]. At 12 months of age, infants’ gut microbiota is more like their mother’s, showing an increase in alpha diversity and a decrease in beta diversity as a function of time, indicating a more complex and less heterogeneous community [40].

The maturation of the microbial ecosystem depends on the coexistence with family members [41]. There are different related factors that, with individuality, have a crucial role in shaping the microbial composition of the intestine [42]. These factors include age, host genetics, antibiotic use, colonization site physiology, mode of delivery (vaginal section or caesarean section), a type of feeding (human milk or formula), and environment of birth (rural or urban) [43–46].

The colonisation process during the first days of the new-born’s life is critical for correct immunological, cognitive, and physiological development [47–49]. Caesarean section delivery with antibiotic treatment before prophylaxis is related to a decrease in the diversity of Actinobacteria, Bacteroidetes, *Bifidobacterium*, and *Lactobacillus*, and an increase in Proteobacteria, Firmicutes, and *Enterococcus spp.* On the other hand, vaginal delivery with the administration of antibiotics before delivery is related to a decrease in *Bifidobacterium* and an increase in *Clostridium* [50]. The effect of birth by caesarean section on the intestinal microbiota normalizes at the age of 3 to 5 years, however the intestinal microbiota does not reach the complexity of an adult at 5 years of age, since the infant’s gut microbiota is different to his mother and different to adults [39,40]. This indicates that both, the composition (dynamic) and structure of the gut microbiota evolve as infants grow up, in response to environmental factors and the diet that feed the microbial community [40].



The gut microbiota of babies at 4 and 12 months is more heterogeneous and different to an adult but reaches a greater similarity at the age of 3 and 5 years [40,46]. At 4 months of age, the most abundant bacterial genus is *Bifidobacterium* together with lactic acid bacteria (*Enterococcus*, *Streptococcus* and *Lactobacillus*) and Proteobacteria (*Enterobacteriaceae*, *Citrobacter* and *Serratia*) [40]. Some adults may have lower alpha diversity and community type than children, which suggests that they have an immature microbiota, identifying *Bifidobacterium* and *Ruminococcus gnavus* as markers of immature microbiota for both children and adults [40,51]. *R. gnavus* can use the host's mucin and is linked to inflammatory problems of the gastrointestinal tract [39].

### 2.3. Changes in the Composition of Gut Microbiota

The main inhabitants of the intestine in a vaginally born baby approximately 1 month after birth are Actinobacteria (*Bifidobacterium*), Bacteroidetes (*Bacteroides*), and Gammaproteobacteria (*Enterobacteria*), which are essential for the metabolism of specific carbohydrates in breast milk [49,52]. At around the 6 months of life, with the introduction of a solid diet, the infant's microbiota experiences change in relative abundance, decreasing *Bifidobacterium* and *Clostridial*, and an increase in Firmicutes [49]. On the other hand, caesarean section delivery is related to a decrease in *Bacteroides*, with a 41% similarity to the maternal microbiota compared to vaginal delivery [45,47,53].

Caesarean section delivery and formula feeding could be related to an increase in Bacteroides, Proteobacteria, and Firmicutes, compared to vaginally born, breastfed children, who show higher dominance of *Bifidobacterium* and *Bacteroides* dominance [53,54]. Breast milk provides the infant with bioactive molecules such as human milk oligosaccharides, IgA, and essential fatty acids and contributes to the optimal configuration of the gut microbiota, as well as influencing cognitive development and the maturation of the immune system [49].

Throughout life, various environmental factors can alter the gut microbiota [47], but the first 3 years of life are particularly important for the health of the host. However, the microbiota continues to change over the years, becoming more diverse and showing a gradual increase in the proportion of *Bacteroides* and *Clostridium* under normal conditions [55]. When there is an imbalance in microbial abundance, dysbiosis occurs. This imbalance may result from a loss of microbial diversity, either from a loss of beneficial microorganisms or an increase of pathogenic species, which is often linked to pathological states, such as some neurobehavioral and gastrointestinal problems that are more commonly seen in children with ASD [21,28]. Gastrointestinal problems in children with ASD are mainly due to the increase in *Bacteroidetes*, *Clostridium*, and *Suterella* [18,56,57].

### 2.4. Dysbiosis and Gastrointestinal Function in ASD

The prevalence of gastrointestinal symptoms in children with ASD ranges from 9% to 91%; for this reason, the study of the gut microbiota has been a key point in the exploration of gastrointestinal disorders and their relationship with the development of different ASD phenotypes [58]. The severity of the symptoms in ASD, the form of birth, and maternal age are factors that affect the diversity of the intestinal microbiota. Finding a lower abundance of *Bacteroides* and *Faecalibacterium* and higher abundance of *Clostridial* in stool samples from children with autism, and higher levels of *Erysipelotrichaceae* and *Faecalibacterium* were also identified in children with severe autism [29].

Under normal circumstances, the intestinal microbiota is composed of anaerobic bacteria that are essential for protecting against pathogens and performing various functions that benefit the host, such as nutrient absorption, immunity, and production of short-chain fatty acids, vitamins, and amino acid synthesis among others, that help keep the colon healthy [5]. However, under dysbiosis, the absorption of nutrients in the small intestine is low, and therefore more monosaccharides and disaccharides reach the large intestine, benefiting the bacteria that ferment simple sugars and show greater growth compared to bacteria that degrade complex sugars (polysaccharides). More sugars in the gut can cause gas and bloating [59]. Patients with ASD have increased intestinal permeability, which may be related to changes in microbial diversity and a decrease in the amount of *Lactobacillus*, bacteria related to maintaining the union of the epithelial barrier of the intestine [60,61].

Increased intestinal permeability increases circulating bacterial-derived lipopolysaccharides (LPS) that trigger immunological and inflammatory reactions, characterised by a systemic increase in proinflammatory cytokines. The increase in cytokines has been reported in patients with ASD, particularly in those subtypes of mental regression. Cytokines are necessary for normal neurodevelopment, and their disturbances can impact this process [5].

Although the results on the relationship of the neurotransmitter  $\gamma$ -aminobutyric acid (GABA) and the severity of autistic symptoms (Table 1) are not entirely conclusive, the reduction in GABA could be accentuating the neuroinflammatory processes caused by the increase in LPS in ASD. GABA and glutamate are the main excitatory and inhibitory neurotransmitters in the brain, and their balanced interaction is necessary for neuronal function [62]. A recent study found that GABA alleviates sepsis induced by LPS. Therefore, it is possible to relate the decrease in GABA in patients with autism to the attempt to mediate neuroinflammatory processes in these patients [63].

### 3. Gut Microbiota and the Immune System

At birth, the new-born's immune system is not well developed, which gives it the benefit of not generating a severe reaction against the mother's antigens [64]. Different studies suggest that the gut microbiota plays an important role as a source of antigens, including peptidoglycans, lipoproteins, lipopolysaccharides, and flagellin. All these antigens together constitute, activate, and educate the innate and adaptive immune systems [46,65]. A stable microbial composition (homeostasis), as well as a suitable microenvironment (as the body environment affects microbial diversity), is important for healthy metabolic functioning of microbiota. Therefore, the first colonizing microorganisms in the human intestine are of vital importance [66,67].

*Bifidobacterium* is transmitted vertically from mother to child and are the first to colonise and constitute the most abundant group of microorganisms in the intestine of a healthy new-born [52,68,69]. The colonization and establishment of this bacterial group in the intestine of the new-born have been related to the modulation by oligosaccharides, specific nutrients that human milk contained according to a microbe-host coevolution mechanism [53,70]. Different functions with health benefits, modulation of the immune system, and production of metabolites that confer physiological benefits have been proposed [71,72].

Gut bacteria can act as modulators of the immune system. Extracellular structures produced by gut bacteria have been hypothesized to interact with the host's immune system, potentially leading to pro-inflammatory or anti-inflammatory effects [71]. Extracellular structures such as pili/fimbria, exopolysaccharides, and teichoic acids play a crucial role between bacterial communication and the host's immune system. Especially during the first years of life, which are characterised by an immature immune system [68,69].

Considering the *Bifidobacterium* dominance is common until weaning, it has been suggested that these bacteria, followed by *Lactobacillus* and *Veillonella* [73], have a fundamental role in the formation of the host's immune system [52,71]. Early childhood represents a very critical window of time for the formation of human health with lasting effects, especially in those individuals born by caesarean section or with antibiotic treatment who may suffer from altered gut microbiota [72]. Intervention with *Bifidobacterium*-based therapies can be considered a highly suitable approach to promote and maintain a balanced infant gut microbiota [69,72].

#### 3.1. Inflammation and Gut Microbiota

Epidermal growth factor is present in amniotic fluid and, after birth, in colostrum and mature milk. In the infant intestine, it promotes the proliferation and maturation of epithelial cells, which are involved in repairing the intestinal mucosa [47]. The epithelial barrier has the function of maintaining intestinal homeostasis between luminal microbes and the host's immune system. Addition, it is the first site exposed to different environmental factors that can lead to the development of a pathological state [74]. A deterioration in the epithelial barrier can lead to the translocation of intestinal microbes, thus promoting hyperactivation of the mucosal immune system and increased production of proinflammatory cytokines, which together promote inflammation [75].

Inflammation can induce goblet cell depletion, leading to inappropriate mucin secretion, and imbalanced production of IL-17, which together lead to chronic inflammation [75]. The intestinal microbiota is closely related to IL-17 levels of, and dysbiosis can cause chronic inflammation. In hosts genetically susceptible to inflammation, the immune response is dysregulated towards intestinal commensal microorganisms (pathobionts). Hyperactivation of IL-17 can lead to autoimmune-type inflammation in the gut [75,76].

Because genes affect the proper functioning of the immune system and increase susceptibility to disease, certain alleles have been associated with specific types of microbial composition. The rs651821 variant of the *APOA5* gene is related to the presence of *Lactobacillus*, *Sutirella*, and *Methanobrevibacter* in individuals who are at a higher risk of developing metabolic diseases. Additionally, variants in the *SLIT3* gene have been associated with the development of inflammation induced by microbial products [77]. However, it is worth nothing that various studies have shown that the composition of the gut microbiota is primarily influenced by non-genetic factors [78].

### 3.2. Other Immune Alterations Related to Gut Microbiota

The host microbiota is an important environmental factor that can confer maturation and activation to microglia, both in healthy and pathological conditions. During the prenatal stage, some maternal conditions, such as viral or bacterial infections, can activate the maternal immune system (MIA), which can then be passed on to the progeny and cause lasting changes in behaviour [79]. Although the main function of microglia is rapid protection for the brain, dysfunction of immune activity can lead to negative outcomes for surrounding neurons and glia, affecting neurogenesis, synapse, and neuroinflammatory regulation [79,80]. Given that microglia cells are long-lived and part of the early brain structure, microglia with aberrant function during neurodevelopment in infancy could play a key role in modulating cell-cell interaction during life, early life, and the trajectory of health and disease [81].

Gastrointestinal physiology is controlled by the Enteric Nervous System (ENS), which is composed of neurones and glial cells. The gut microbiota and the postnatal mucosal immune system are responsible for conferring maturation on gut neural networks [82]. Molecular interactions between the microbiota, enteric cells, and immune cells (including microglia) are crucial to maintaining gastrointestinal homeostasis. A disruption in these interactions can lead to neurodevelopmental disorders, including ASD [83–85].

## 4. The Microbiota-Gut-Brain Axis

The gut microbiota plays a fundamental role in the physiological functioning of the host and alterations in this microenvironment can have harmful effects on key points in the development of various organs systems, including the brain and digestive system (brain-gut-microbiota or microbiota-gut-brain axis). The brain-gut axis consists of the brain, the spinal cord, ANS, ENS, and hypothalamic-pituitary-adrenal (HPA) axis. Disturbances in the microbiota-gut-brain axis are the principal cause of the most frequent gastrointestinal motility disorders [86]. Studies in germ-free (GF) animals, those treated with antimicrobials, or those exposed to environmental modifications that alter the gut microbiota from the prenatal or postnatal stage, have been related to problems in brain immunity, blood-brain barrier (BBB) permeability, brain architecture, and neural circuits [87].

### 4.1. Animal Models of Altered Gut Microbiota and Effects in CNS

There are bacterial strains such as *Escherichia coli* or *Lactobacillus sp.* that interact directly with the host's CNS through neurotransmitters dopamine, norepinephrine, histamine, acetylcholine, GABA or serotonin. An alteration in the composition of these strains can lead to an alteration in the metabolic state of the microbiome, resulting in metabolic disorders that may be responsible for the severity or progression of neurological disorders, such as Parkinson's, Alzheimer's, ASD and depression, among others [88,89].



Experimental studies have shown that the production of the bacterial metabolite 4-methylphenol (para-cresol or *p*-cresol) can alter the composition of the intestinal microbiota, leading to the recolonisation of *Clostridium difficile*. Fermenting tyrosine via the *p*-hydroxyphenylacetate (*p*-HPA) pathway is how *C. difficile* produces *p*-cresol. Since *C. difficile* is related to decreased growth of Proteobacteria, experimental studies showed that there were animals infected with *C. difficile* mutants (*hpdC*), since they found bacterial families belonging to Proteobacteria. In addition, the studies found that *Bifidobacterium adolescentis* is more sensitive to the presence of *p*-cresol than other Gram-positive species [90,91].

*p*-cresol negatively affects the homeostasis of epithelial cells, and its excess negatively affects the integrity of colonic epithelial cells [92]. As a result of the disruption in epithelial cells, a proinflammatory phenotype involving LPS may be promoted. Inflammation is closely related to the pathophysiology of mental disorders. Multiple communication pathways between the microbiota and the CNS have been identified, including immune pathways [93].

#### 4.2. Effects of Gut Microbiota Metabolites in Immune Cells of CNS

The gut microbiota has a complex and specific communication with the CNS. The communication between the microbiota, the gut, and the brain involves the secretion of different metabolites, including short-chain fatty acids (SCFA), the structural components of bacteria and signalling molecules [94].

Bacterial metabolites such as LPS can easily cross the intestinal barrier and cause inflammation, which affects the brain by altering cytokine levels [95]. Additionally, cytokines produced locally in the gastrointestinal mucosa travel peripherally and can cross the BBB [96]. During inflammation, the brain releases arginine vasopressin, a metabolite that affects social behaviour and is a considered biomarker in ASD [14]. There is a bidirectional relationship between the gut and the brain that includes nerve fibres [88]. Enteroendocrine cells of the intestinal epithelial barrier can detect the composition of the intestinal lumen, as well as nutrients and bacterial metabolites. These cells synapse with afferent fibres that directly connect the intestinal lumen to the brainstem [97].

The SCFA are the result of the fermentation of dietary fibre by anaerobic commensal bacteria in the colon. The host recognizes SCFA (acetate, propionate, and butyrate). Recently, butyrate has been identified as protective of the mucosa via goblet cells, as they regulate the response to the upregulation of *MUC* gene expression [16,98–100].

Altered concentrations of metabolites may have functional consequences in ASD. Different studies have identified various altered metabolites in the urinary profile of children with autism, some even correlate with the severity of autistic behaviour such as *p*-cresol [30,101]. Additionally, the metabolic pathways for tryptophan, vitamin B6, purine, and phenylalanine are altered in ASD [102,103].

Combination therapy with vancomycin and *Bifidobacterium* improved in autistic symptoms. Additionally, the above therapy helped normalize the levels of the metabolites 3-(3-hydroxyphenyl)-3-hydroxypropionic acid, 3-hydroxyphenylacetic acid, and 3-hydroxyhippuric acid in the urine of children with ASD. Indicating an alteration in the production of phenylalanine in ASD [104].

### 5. Gut Microbiome, Immune System, and Neurodevelopment Disorders

In mammals, enteric neurogenesis and gliogenesis occur primarily during the embryonic and foetal stages, but a considerable fraction of enteric neurons and glia are born in the colonizing postnatal gut [82]. Functional maturation of gut neural networks is completed within the microenvironment of the postnatal gut, under the influence of gut microbiota and the mucosal immune system [83].

The gut microbiome lies at the intersection between the environment and the host, with the ability to modify host responses to disease-relevant exposures and stimuli. This is evident in the way that enteric microbes interact with the immune system, for example, by supporting immune maturation in the first years of life, affecting the efficacy of drugs through modulation of immune responses, or influencing the development of immune cell populations and their mediators [105].

### 5.1. Altered Gut Microbiota and Neurodevelopment Disorders

Gut microbiota alterations have been linked to various pathogenic pathways, and an increasing number of studies are linking changes in the gut microbiota to a range of neuropsychiatric diseases. [106,107]. Similarly, the decrease in microbial diversity throughout life could be related to neurodegeneration [108,109].

The neuroinflammation produced by the different metabolites of a dysbiotic microbiota can be a pathogenic factor in severe neurodegenerative disorders [110]. In a severe inflammatory state, the activation of microglia releases proinflammatory cytokines such as TNF- $\alpha$ , IL-6, or MCP-1 or the inflammasome, as well as reactive oxygen species from microglial cells and resident macrophages, which could cause chronic neuroinflammation [17,111,112]. Therefore, the decrease or loss of the integrity of intestinal epithelial cells and chronic inflammation are highlighted as the main consequences of dysbiosis. Neuroinflammation and neurodegenerative and neuropsychiatric disorders can be the result of dysbiosis [36,106,113].

CNS inflammation is related to ASD. TNF- $\alpha$  is found at high levels in children with ASD, which was correlated with the severity of gastrointestinal symptoms [114]. Increases in TNF- $\alpha$  (increase *Lachnoclostridium bolteae*), IL-2, IL-4, IL-6 (increase *Clostridium lituseburense*), IL-8, IL-10, and IL-17 (increase *Clostridium tertium*) indicate a higher level of inflammation in the CNS of children with autism [18,95,115]. Additionally, children with ASD also have dysregulated T-cell production, leading to biases in the Th1 to Th2 ratio and immune cell activation associated with altered behaviour due to further neurodevelopmental impairment [116].

### 5.2. Altered Gut Microbiota and ASD

Microbial colonization of the gastrointestinal tract begins prenatally, as microorganisms are detected in the placenta and meconium [117,118]. A recent study found that mothers of children with ASD harbour altered gut microbiomes [119], supporting the idea that maternal gut microbiota variation and infections during pregnancy may increase the risk of ASD in offspring [120]. In this same study, a clear relationship was found between gut microbiome profiles between children and their mothers.

Children with ASD have unique bacterial biomarkers [119]. Previous studies have suggested that the gut microbiome of children with ASD contains harmful genera or species that contribute to the severity of autism symptoms, such as *Bacteroides* [121,122] or *Desulfovibrio*, which is related to the modulation of *p*-cresol production [30]. Changes in the gut microenvironment caused by the gut microbiome affect the production of signalling substances, leading to inadequate functioning of the brain and thus the prenatal and postnatal CNS [11].

The most common gut microbiota findings in children with ASD were a decreasing trend from Bacteroidetes to Firmicutes [123] and increased abundance of *Clostridium* [27,29,124]. Despite these findings, there are inconsistencies about the phenotypic signature of the gut microbiome of children with ASD, and a reason for these inconsistent results is that the composition of the gut microbiota is influenced by several factors, such as diet, lifestyle, medical history, among others [121]. In this regard, the use of antibiotics showed a correlation with the improvement of symptoms in ASD [126]. Although recently, it has been seen this is not always the case since the prolonged use of oral antibiotics can increase the proliferation of anaerobic bacteria in the intestine. For example, *Clostridia*, *Bacteroidetes*, and *Desulfovibrio* are common bacteria that, in addition to modulating the intestinal immune system, can promote gastrointestinal symptoms and autistic behaviour in ASD [127].

The Fungi *Candida* also appears to play a role in children with ASD [123]. In a dysbiotic environment, as often seen in the population with autism, *Candida* proliferates and produces ammonia and toxins, which increases autistic behaviour. *Candida* also causes malabsorption of minerals and carbohydrates that play an important role in the pathophysiology of ASD [128]. A subset of people with ASD shows gastrointestinal disturbances [129], and results from different studies indicate that eliminating some foods from the diet may help improve gastrointestinal symptoms in ASD. Associating diet is an important factor in the composition of the gut microbiota [130].

On the other hand, in a study with a large group of ASD patients, their gut metagenome showed a relationship with diet, reduced taxonomic diversity, and stool consistency, but no relationship was found between the diagnosis of ASD and the gut microbiome. It was suggested that softer faecal consistency is more closely related to decreased taxonomic diversity and that there is a downstream relationship to reduced dietary diversity, that is a common feature of patients with ASD. In this work, authors proposed that this mechanism could explain the relationship between the increase in gastrointestinal problems and increase in repetitive behaviours in ASD. Maintain that sensory sensitivity could be the basis of restricted diets in ASD but found no relationship between the sensory profile and ASD severity. The study found that all psychometric characteristics had more significant correlations with dietary diversity than with taxonomic diversity. In conclusion, the results suggest that dysbiosis is a consequence of autistic manifestations and that it has no causal role in the disease. Therefore, microbiome-directed treatment is not a suitable therapeutic target for treating comorbidities in children with ASD [19].

The cause of ASD remains undetermined, complex, and incompletely understood, with increasing evidence pointing to abnormal synaptic development and aberrant immune responses as possible effectors of autistic symptoms [85,131]. Microglial cells have been strongly associated with physiological processes and the development of autistic symptoms, as they are part of the main cells of the CNS that provide an innate immune response to the tissue with inflammatory and tissue repair functions [85].

Some metabolites produced by specific bacterial groups have anti-inflammatory effects on microglia, such as butyrate, an essential SCFA for the modulation of excitatory and inhibitory neuronal pathways in ASD [100,132]. Patients with ASD showed low levels of SCFA [133]. The deficiency of these metabolites could be the cause of a disruption in the intercommunication between the ENS and the mucosal immune system [83], what could be generate changes in the intestinal motility of children with ASD [27]. For example, SCFA activate G protein-coupled receptors (GPR41 and GPR43) on enteroendocrine cells of the intestinal epithelium, resulting in increased production of GLP-1 and 5-HT and changes in intestinal motility [83].

Not studies that suggest that dysbiosis has a leading role in the cause of ASD, since it has always been presented as a multifactorial disease with a very high genetic component, which seems to be the main cause of autistic symptoms [119,134–138]. Although, dysbiosis has been proposed as another factor in the cause of ASD, its relevance has not been well clarified since the results of various studies have not been conclusive. Some studies point to dysbiosis as a possible factor in autistic symptoms [29,58], and other studies rule out the possibility that dysbiosis is a determining factor in the aetiology of ASD [19,139].

Recently, it has increased reports of evidence regarding the possible involvement of intestinal dysbiosis (Table 1) as an aetiological factor in ASD with moderate effects. The relevance of dysbiosis as a factor in the aetiology of ASD relies on the fact that it is modifiable. There are reports of interventions that have had beneficial but modest effects on autistic manifestations. For example, faecal transplant therapies and probiotic supplementation. A pioneer group in faecal transplantation in children with ASD found that after ten weeks of intervention, there was a reduction of almost 80% in gastrointestinal symptoms and improvement in behavioural symptoms, and that these improvements persisted after eight weeks of treatment; in addition, there were beneficial changes in the abundance of *Bifidobacterium*, *Prevotella*, and *Desulfovibrio*; these changes also persisted after the suspension of the intervention and two years later [140,141]. Regarding probiotics, a successful study that supplemented with probiotics children with ASD for 6 months found positive effects on some gastrointestinal symptoms, adaptive functions, and sensory profiles versus the placebo group [142]. Another study found that after supplementation with probiotics and prebiotics, there was an increase in beneficial bacteria (*Bifidobacterium* and *B. longum*) and suppression of pathogenic bacteria (*Clostridium*) with a significant reduction in the severity of autistic and gastrointestinal symptoms [143].

ASD is a neurodevelopmental disorder of multifactorial etiology where the main factors are genetic variations interacting with environmental factors (Dysbiosis). Genetics Factors. The genetic

**ASD**

**1. Genetic variants:** *MUC3A, ANKRD20A2/3, ALDH1A, NOXA1 and many others*

**2. Advanced maternal age and antibiotic use during pregnancy**

**Restrictive feeding behaviour**

**Low Food diversity**

**Decrease Microbiome diversity**

**Dysbiosis**

**3 Dysbiotic microbiome factors**

**Microbiome-Gut-Brain Axis**

**3. Inflammatory CNS Cytokines, SCFA, and altered neurotransmitters and gut permeability are caused by dysbiosis**

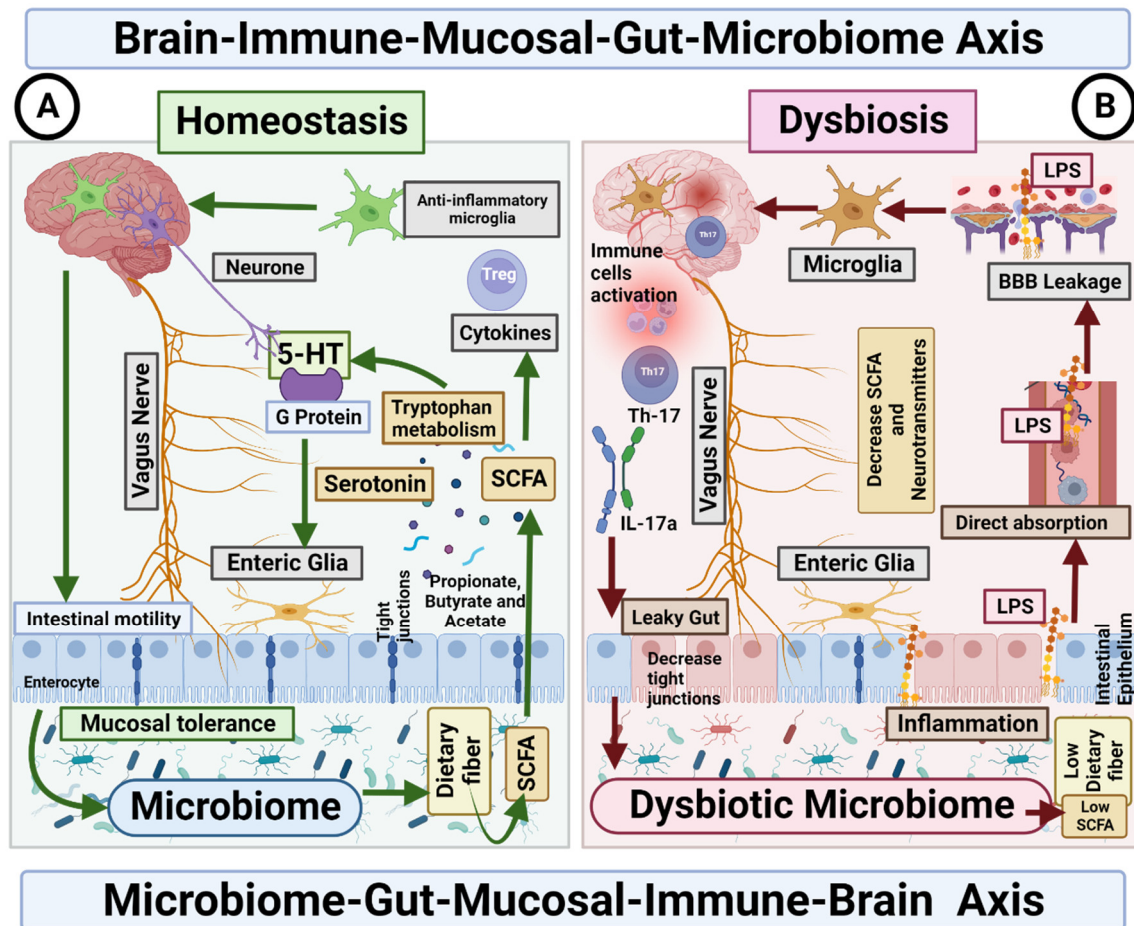
**1 Genetic factors**

**2 Environmental factors**

**Increase Exacerbation of ASD Core Symptoms**

**Figure 1.** Vicious cycle and phenotype relationship in ASD.





**Figure 2. Microbiome-Gut-Mucosal-Immune-Brain bidirectional relationship.** The vagus nerve transfers information on the state of the digestive system to the brain through sensory fibers. It sends neuronal, endocrine, and immune signals. The gut microbiota metabolites are involved in permeability of the intestinal epithelium, neurotransmitter synthesis and cytokines release.

**Homeostasis.** Under homeostasis conditions, gut microbiome and diet control the activity of ENS cells, through SCFA, which in turn activate the enteroendocrine cells of the intestinal epithelium, which increases the production of serotonin and therefore change intestinal motility. In addition, the microbiome is essential for the maintenance of mucosal glial cells that express neurotrophic factor.

**Dysbiosis.** The imbalance in the homeostasis of the gut microbiome is associated with the decreased production of metabolites, such as butyrate, responsible for the modulation of inhibitory and excitatory pathways, and with anti-inflammatory effects on microglia. Deficiency of this microbial metabolite could be the cause of a disruption in the intercommunication between the ENS and the mucosal immune system.

## 6. Conclusions

The mode of delivery at birth (caesarean section or vaginal) will determine the type of microbiota that will colonise the intestine of the new-born. The colonisation process during the first days of life helps configure a correct immunological and cognitive development in the neonate. Subsequently, the type of diet (breast milk or milk formula) will shape the alpha diversity of the infant. During the first months of life, alpha diversity is low, and its diversity increases gradually, when solid foods are introduced to the baby for the first time (approx. 6 months).

During a disease process, the composition, from a dynamic and functional viewpoint, of the microbiome can vary. The changes in this dynamic system can generate the deregulation of the



production of microbial metabolites, generating an imbalance, as reported under the term “dysbiosis”. Gastrointestinal problems are mostly linked to intestinal dysbiosis.

Alterations in the microbiota-gut-brain axis are the main cause of the most frequent gastrointestinal motility disorders in psychiatric and neurodevelopmental diseases. Neuroinflammation produced by the different metabolites of a dysbiotic microbiota may be a pathogenic factor in severe neurodegenerative disorders. Children with ASD have unique bacterial biomarkers, and *Bacteroides* may contribute to the severity of autism symptoms. Despite these findings, there are inconsistencies regarding the phenotypic signature of the gut microbiome of children with ASD, and a reason for these inconsistent results is that the composition of the microbiome is influenced by several factors, such as diet, lifestyle, medical history, and genetic variations. For this reason, there are controversies between the microbiome and the restrictive diet as possible causes of gastrointestinal comorbidities and autistic symptoms in ASD. However, with the model (Figure 2) proposed in this review, it is intended to expand knowledge of the role of gut microbiota in the patients with ASD.

The existence of intercommunication between immune-mediated neurological and gastrointestinal disorders suggests a new microbiome-gut-mucosa-immuno-brain axis (neuroimmunogastroenterology) in patients with ASD that could be part of the vicious cycle in the aetiology of ASD. However, restrictive dietary behaviours are part of the patients with autism, dietary interventions are also critical for modulating the microbiome. Modulating the microbiome with non-invasive interventions, such as diet and probiotics, could be a therapeutic option to attenuate the severity of autistic symptoms.

## References

1. American Psychiatric Association, DSM-5 Task Force. *Diagnostic and statistical manual of mental disorders: DSM-5™*, 5th ed.; American Psychiatric Publishing, Inc.: Washington, DC, USA, 2013. <https://doi.org/10.1176/appi.books.9780890425596>.
2. Rogozin IB, Gertz EM, Baranov PV, Poliakov E, Schaffer AA. Genome-Wide Changes in Protein Translation Efficiency Are Associated with Autism. *Genome Biol. Evol.* **2018**, 10, 1902-1919. <https://doi.org/10.1093/gbe/evy146>.
3. Hallmayer J, Cleveland S, Torres A; et al. Genetic heritability and shared environmental factors among twin pairs with autism. *Arch. Gen. Psychiatry* **2011**, 68, 1095-1102. <https://doi.org/10.1001/archgenpsychiatry.2011.76>.
4. Buie T, Campbell DB, Fuchs GJ 3rd; et al. Evaluation, diagnosis, and treatment of gastrointestinal disorders in individuals with ASDs: A consensus report. *Pediatrics* **2010**, 125 (Suppl 1), S1-S18. <https://doi.org/10.1542/peds.2009-1878C>.
5. Ding HT, Taur Y, Walkup JT. Gut Microbiota and Autism: Key Concepts and Findings. *J. Autism Dev. Disord.* **2017**, 47, 480-489. <https://doi.org/10.1007/s10803-016-2960-9>.
6. Vuong HE, Yano JM, Fung TC, Hsiao EY. The Microbiome and Host Behavior. *Annu. Rev. Neurosci.* **2017**, 40, 21-49. <https://doi.org/10.1146/annurev-neuro-072116-031347>.
7. Rice D, Barone S Jr. Critical periods of vulnerability for the developing nervous system: Evidence from humans and animal models. *Environ. Health Perspect.* **2000**, 108 (Suppl. 3), 511-533. <https://doi.org/10.1289/ehp.00108s3511>.
8. Nagpal R, Tsuji H, Takahashi T; et al. Ontogenesis of the Gut Microbiota Composition in Healthy, Full-Term, Vaginally Born and Breast-Fed Infants over the First 3 Years of Life: A Quantitative Bird's-Eye View. *Front. Microbiol.* **2017**, 8. <https://doi.org/10.3389/fmicb.2017.01388>.
9. Stewart CJ, Ajami NJ, O'Brien JL; et al. Temporal development of the gut microbiome in early childhood from the TEDDY study. *Nature* **2018**, 562, 583-588. <https://doi.org/10.1038/s41586-018-0617-x>.
10. Guo M, Miao M, Wang Y; et al. Developmental differences in the intestinal microbiota of Chinese 1-year-old infants and 4-year-old children. *Sci. Rep.* **2020**, 10. <https://doi.org/10.1038/s41598-020-76591-4>.
11. Sharon G, Sampson TR, Geschwind DH, Mazmanian SK. The Central Nervous System and the Gut Microbiome. *Cell.* **2016**, 167, 915-932. <https://doi.org/10.1016/j.cell.2016.10.027>.
12. Srikantha P, Mohajeri MH. The Possible Role of the Microbiota-Gut-Brain-Axis in Autism Spectrum Disorder. *Int. J. Mol. Sci.* **2019**, 20. <https://doi.org/10.3390/ijms20092115>.
13. Rothenberg SE, Chen Q, Shen J; et al. Neurodevelopment correlates with gut microbiota in a cross-sectional analysis of children at 3 years of age in rural China. *Sci. Rep.* **2021**, 11. <https://doi.org/10.1038/s41598-021-86761-7>.

14. Madore C, Leyrolle Q, Lacabanne C; et al. Neuroinflammation in Autism: Plausible Role of Maternal Inflammation, Dietary Omega 3, and Microbiota. *Neural Plast.* **2016**, 2016, 3597209. <https://doi.org/10.1155/2016/3597209>.
15. Li J, Hu S, Zhang K; et al. A comparative study of the genetic components of three subcategories of autism spectrum disorder. *Mol. Psychiatry* **2019**, 24, 1720–1731. <https://doi.org/10.1038/s41380-018-0081-x>.
16. Tran SM, Mohajeri MH. The Role of Gut Bacterial Metabolites in Brain Development, Aging and Disease. *Nutrients* **2021**, 13. <https://doi.org/10.3390/nu13030732>.
17. Rose DR, Yang H, Serena G; et al. Differential immune responses and microbiota profiles in children with autism spectrum disorders and co-morbid gastrointestinal symptoms. *Brain Behav. Immun.* **2018**, 70, 354–368. <https://doi.org/10.1016/j.bbi.2018.03.025>.
18. Lungba RM, Khan SZA, Ajibawo-Aganbi U; et al. The Role of the Gut Microbiota and the Immune System in the Development of Autism. *Cureus.* **2020**, 12. <https://doi.org/10.7759/cureus.11226>.
19. Yap CX, Henders AK, Alvares GA; et al. Autism-related dietary preferences mediate autism-gut microbiome associations. *Cell* **2021**, 184, 5916–5931.e17. <https://doi.org/10.1016/j.cell.2021.10.015>.
20. Berg G, Rybakova D, Fischer D; et al. Microbiome definition re-visited: Old concepts and new challenges [published correction appears in *Microbiome*. 2020 Aug 20;8, 119]. *Microbiome* **2020**, 8. <https://doi.org/10.1186/s40168-020-00875-0>.
21. de Vos WM, Tilg H, Van Hul M, Cani PD. Gut microbiome and health: Mechanistic insights. *Gut* **2022**, 71, 1020–1032. <https://doi.org/10.1136/gutjnl-2021-326789>.
22. Lepage P, Leclerc MC, Joossens M; et al. A metagenomic insight into our gut's microbiome. *Gut* **2013**, 62, 146–158. <https://doi.org/10.1136/gutjnl-2011-301805>.
23. Lozupone CA, Stombaugh JL, Gordon JI, Jansson JK, Knight R. Diversity, stability and resilience of the human gut microbiota. *Nature* **2012**, 489, 220–230. <https://doi.org/10.1038/nature11550>.
24. Ehrlich, S.D., The MetaHIT Consortium. MetaHIT: The European Union Project on Metagenomics of the Human Intestinal Tract. In *Metagenomics of the Human Body*; Nelson, K., Eds.; Springer: New York, NY, USA, 2011. [https://doi.org/10.1007/978-1-4419-7089-3\\_15](https://doi.org/10.1007/978-1-4419-7089-3_15).
25. Mobeen F, Sharma V, Tulika P. Enterotype Variations of the Healthy Human Gut Microbiome in Different Geographical Regions. *Bioinformation* **2018**, 14. <https://doi.org/10.6026/97320630014560>.
26. Mariat D, Firmesse O, Levenez F; et al. The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. *BMC Microbiol.* **2009**, 9. <https://doi.org/10.1186/1471-2180-9-123>.
27. Ma B, Liang J, Dai M; et al. Altered Gut Microbiota in Chinese Children with Autism Spectrum Disorders. *Front. Cell Infect. Microbiol.* **2019**, 9. <https://doi.org/10.3389/fcimb.2019.00040>.
28. Liu F, Li J, Wu F, Zheng H, Peng Q, Zhou H. Altered composition and function of intestinal microbiota in autism spectrum disorders: A systematic review. *Transl. Psychiatry* **2019**, 9. <https://doi.org/10.1038/s41398-019-0389-6>.
29. Ding X, Xu Y, Zhang X; et al. Gut microbiota changes in patients with autism spectrum disorders. *J. Psychiatr. Res.* **2020**, 129, 149–159. <https://doi.org/10.1016/j.jpsychires.2020.06.032>.
30. Ho LKH, Tong VJW, Syn N; et al. Gut microbiota changes in children with autism spectrum disorder: A systematic review. *Gut Pathog.* **2020**, 12. <https://doi.org/10.1186/s13099-020-0346-1>.
31. Ha S, Oh D, Lee S; et al. Altered Gut Microbiota in Korean Children with Autism Spectrum Disorders. *Nutrients*. **2021**, 13. <https://doi.org/10.3390/nu13103300>.
32. Montiel-Castro AJ, González-Cervantes RM, Bravo-Ruiseco G, Pacheco-López G. The microbiota-gut-brain axis: Neurobehavioral correlates, health and sociality. *Front. Integr. Neurosci.* **2013**, 7. <https://doi.org/10.3389/fnint.2013.00070>.
33. Sharon G, Garg N, Debelius J, Knight R, Dorrestein PC, Mazmanian SK. Specialized metabolites from the microbiome in health and disease. *Cell Metab.* **2014**, 20, 719–730. <https://doi.org/10.1016/j.cmet.2014.10.016>.
34. Bäckhed F, Fraser CM, Ringel Y; et al. Defining a healthy human gut microbiome: Current concepts, future directions, and clinical applications. *Cell Host Microbe* **2012**, 12, 611–622. <https://doi.org/10.1016/j.chom.2012.10.012>.
35. Leonard MM, Valitutti F, Karathia H; et al. Microbiome signatures of progression toward celiac disease onset in at-risk children in a longitudinal prospective cohort study. *Proc. Natl. Acad. Sci. USA* **2021**, 118, e2020322118. <https://doi.org/10.1073/pnas.2020322118>.
36. Dinan TG, Cryan JF. Gut instincts: Microbiota as a key regulator of brain development, ageing and neurodegeneration. *J. Physiol.* **2017**, 595, 489–503. <https://doi.org/10.1113/JP273106>.
37. Paul B, Barnes S, Demark-Wahnefried W; et al. Influences of diet and the gut microbiome on epigenetic modulation in cancer and other diseases. *Clin. Epigenetics* **2015**, 7. <https://doi.org/10.1186/s13148-015-0144-7>.
38. Suzuki TA, Fitzstevens JL, Schmidt VT; et al. Codiversification of gut microbiota with humans. *Science* **2022**, 377, 1328–1332. <https://doi.org/10.1126/science.abm7759>.
39. Bäckhed F, Roswall J, Peng Y; et al. Dynamics and Stabilization of the Human Gut Microbiome during the First Year of Life. *Cell Host Microbe* **2015**, 17, 852. <https://doi.org/10.1016/j.chom.2015.05.012>.

40. Roswall J, Olsson LM, Kovatcheva-Datchary P; et al. Developmental trajectory of the healthy human gut microbiota during the first 5 years of life. *Cell Host Microbe* **2021**, 29, 765776.e3. <https://doi.org/10.1016/j.chom.2021.02.021>.
41. Valles-Colomer M, Bacigalupe R, Vieira-Silva S; et al. Variation and transmission of the human gut microbiota across multiple familial generations. *Nat. Microbiol.* **2022**, 7, 87–96. <https://doi.org/10.1038/s41564-021-01021-8>.
42. Chong CYL, Bloomfield FH, O'Sullivan JM. Factors Affecting Gastrointestinal Microbiome Development in Neonates. *Nutrients*. **2018**, 10. <https://doi.org/10.3390/nu10030274>.
43. Martin R, Makino H, Cetinyurek Yavuz A; et al. Early-Life Events, Including Mode of Delivery and Type of Feeding, Siblings and Gender, Shape the Developing Gut Microbiota. *PLoS ONE* **2016**, 11. <https://doi.org/10.1371/journal.pone.0158498>.
44. Chu DM, Ma J, Prince AL, Antony KM, Seferovic MD, Aagaard KM. Maturation of the infant microbiome community structure and function across multiple body sites and in relation to mode of delivery. *Nat. Med.* **2017**, 23, 314–326. <https://doi.org/10.1038/nm.4272>.
45. Dierikx TH, Berkhout DJC, Visser L; et al. The influence of timing of Maternal administration of Antibiotics during cesarean section on the intestinal Microbial colonization in Infants (MAMI-trial): Study protocol for a randomised controlled trial. *Trials* **2019**, 20. <https://doi.org/10.1186/s13063-019-3552-8>.
46. Kumbhare SV, Patangia DVV, Patil RH, Shouche YS, Patil NP. Factors influencing the gut microbiome in children: From infancy to childhood. *J. Biosci.* **2019**, 44, 49.
47. Moore RE, Townsend SD. Temporal development of the infant gut microbiome. *Open Biol.* **2019**, 9, 190128. <https://doi.org/10.1098/rsob.190128>.
48. Hill JH, Round JL. SnapShot: Microbiota effects on host physiology. *Cell* **2021**, 184, 2796–2796.e1. <https://doi.org/10.1016/j.cell.2021.04.026>.
49. Skillington, O. et al. The Contrasting Human Gut Microbiota in Early and Late Life and Implications for Host Health and Disease. *Nutr. Healthy Aging* **2021**, 157–178. <https://doi.org/10.3233/NHA-210129>.
50. Padilha M, Iaucci JM, Cabral VP, Diniz EMA, Taddei CR, Saad SMI. Maternal antibiotic prophylaxis affects *Bifidobacterium* spp. counts in the human milk, during the first week after delivery. *Benef. Microbes* **2019**, 10, 155–163. <https://doi.org/10.3920/BM2018.0046>.
51. Arzamasov AA, Nakajima A, Sakanaka M; et al. Human Milk Oligosaccharide Utilization in Intestinal *Bifidobacteria* Is Governed by Global Transcriptional Regulator NagR. *mSystems* **2022**, 7, e0034322. <https://doi.org/10.1128/msystems.00343-22>.
52. Turrone F, Milani C, Ventura M, van Sinderen D. The human gut microbiota during the initial stages of life: Insights from *bifidobacteria*. *Curr. Opin. Biotechnol.* **2022**, 73, 81–87. <https://doi.org/10.1016/j.copbio.2021.07.012>.
53. Fernandez-Julia P, Commane DM, van Sinderen D, Munoz-Munoz J. Cross-feeding interactions between human gut commensals belonging to the *Bacteroides* and *Bifidobacterium* genera when grown on dietary glycans. *Microbiome Res. Rep.* **2022**, 1, 12. <https://doi.org/10.20517/mrr.2021.05>.
54. Di Guglielmo MD, Franke KR, Robbins A, Crowgey EL. Impact of Early Feeding: Metagenomics Analysis of the Infant Gut Microbiome. *Front. Cell Infect. Microbiol.* **2022**, 12. <https://doi.org/10.3389/fcimb.2022.816601>.
55. Rinninella E, Raoul P, Cintoni M; et al. What is the Healthy Gut Microbiota Composition? A Changing Ecosystem across Age, Environment, Diet, and Diseases. *Microorganisms* **2019**, 7. <https://doi.org/10.3390/microorganisms7010014>.
56. Finegold SM, Dowd SE, Gontcharova V; et al. Pyrosequencing study of fecal microflora of autistic and control children. *Anaerobe* **2010**, 16, 444–453. <https://doi.org/10.1016/j.anaerobe.2010.06.008>.
57. Liu S, Li E, Sun Z; et al. Altered gut microbiota and short chain fatty acids in Chinese children with autism spectrum disorder. *Sci. Rep.* **2019**, 9. <https://doi.org/10.1038/s41598-018-36430-z>.
58. Liu Z, Mao X, Dan Z; et al. Gene variations in autism spectrum disorder are associated with alteration of gut microbiota, metabolites and cytokines. *Gut Microbes* **2021**, 13, 1–16. <https://doi.org/10.1080/19490976.2020.1854967>.
59. Williams BL, Hornig M, Buie T; et al. Impaired carbohydrate digestion and transport and mucosal dysbiosis in the intestines of children with autism and gastrointestinal disturbances. *PLoS ONE* **2011**, 6, e24585. <https://doi.org/10.1371/journal.pone.0024585>.
60. van De Sande MM, van Buul VJ, Brouns FJ. Autism and nutrition: The role of the gut-brain axis. *Nutr. Res. Rev.* **2014**, 27, 199–214. <https://doi.org/10.1017/S0954422414000110>.
61. Iovene MR, Bombace F, Maresca R; et al. Intestinal Dysbiosis and Yeast Isolation in Stool of Subjects with Autism Spectrum Disorders. *Mycopathologia* **2017**, 182, 349–363. <https://doi.org/10.1007/s11046-016-0068-6>.
62. Kolodny T, Schallmo MP, Gerdt J, Edden RAE, Bernier RA, Murray SO. Concentrations of Cortical GABA and Glutamate in Young Adults with Autism Spectrum Disorder. *Autism Res.* **2020**, 13, 1111–1129. <https://doi.org/10.1002/aur.2300>.

63. Xia Y, He F, Wu X; et al. GABA transporter sustains IL-1 $\beta$  production in macrophages. *Sci. Adv.* **2021**, 7. <https://doi.org/10.1126/sciadv.abe9274>.
64. Gervassi AL, Horton H. Is Infant Immunity Actively Suppressed or Immature? *Virology* **2014**, 2014, 1–9. <https://doi.org/10.4137/VRT.S12248>.
65. Patten DA, Collett A. Exploring the immunomodulatory potential of microbial-associated molecular patterns derived from the enteric bacterial microbiota. *Microbiology* **2013**, 159, 1535–1544. <https://doi.org/10.1099/mic.0.064717-0>.
66. Scholtens PA, Oozeer R, Martin R, Amor KB, Knol J. The early settlers: Intestinal microbiology in early life. *Annu. Rev. Food Sci. Technol.* **2012**, 3, 425–447. <https://doi.org/10.1146/annurev-food-022811-101120>.
67. Stinson LF. Establishment of the early-life microbiome: A DOHaD perspective. *J. Dev. Orig. Health Dis.* **2020**, 11, 201–210. <https://doi.org/10.1017/S2040174419000588>.
68. Nuriel-Ohayon M, Neuman H, Ziv O; et al. Progesterone Increases Bifidobacterium Relative Abundance during Late Pregnancy. *Cell Rep.* **2019**, 27, 730–736.e3. <https://doi.org/10.1016/j.celrep.2019.03.075>.
69. Saturio S, Nogacka AM, Alvarado-Jasso GM; et al. Role of Bifidobacteria on Infant Health. *Microorganisms*. **2021**, 9. <https://doi.org/10.3390/microorganisms9122415>.
70. Laursen MF, Sakanaka M, von Burg N; et al. Bifidobacterium species associated with breastfeeding produce aromatic lactic acids in the infant gut. *Nat. Microbiol.* **2021**, 6, 1367–1382. <https://doi.org/10.1038/s41564-021-00970-4>.
71. Kong Q, Chen Q, Mao X; et al. *Bifidobacterium longum* CCFM1077 Ameliorated Neurotransmitter Disorder and Neuroinflammation Closely Linked to Regulation in the Kynurenine Pathway of Autistic-like Rats. *Nutrients* **2022**, 14. <https://doi.org/10.3390/nu14081615>.
72. Stuivenberg GA, Burton JP, Bron PA, Reid G. Why Are Bifidobacteria Important for Infants? *Microorganisms* **2022**, 10. <https://doi.org/10.3390/microorganisms10020278>.
73. Renz H, Skevaki C. Early life microbial exposures and allergy risks: Opportunities for prevention. *Nat. Rev. Immunol.* **2021**, 21, 177–191. <https://doi.org/10.1038/s41577-020-00420-y>.
74. McCole DF. IBD candidate genes and intestinal barrier regulation. *Inflamm. Bowel Dis.* **2014**, 20, 1829–1849. <https://doi.org/10.1097/MIB.0000000000000090>.
75. Amoroso C, Perillo F, Strati F, Fantini MC, Caprioli F, Facciotti F. The Role of Gut Microbiota Biomodulators on Mucosal Immunity and Intestinal Inflammation. *Cells* **2020**, 9. <https://doi.org/10.3390/cells9051234>.
76. Tilg H, Zmora N, Adolph TE, Elinav E. The intestinal microbiota fuelling metabolic inflammation. *Nat. Rev. Immunol.* **2020**, 20, 40–54. <https://doi.org/10.1038/s41577-019-0198-4>.
77. Al Bander Z, Nitert MD, Mousa A, Naderpoor N. The Gut Microbiota and Inflammation: An Overview. *Int. J. Environ. Res. Public. Health* **2020**, 17. <https://doi.org/10.3390/ijerph17207618>.
78. Rothschild D, Weissbrod O, Barkan E; et al. Environment dominates over host genetics in shaping human gut microbiota. *Nature* **2018**, 555, 210–215. <https://doi.org/10.1038/nature25973>.
79. Tartaglione AM, Villani A, Ajmone-Cat MA; et al. Maternal immune activation induces autism-like changes in behavior, neuroinflammatory profile and gut microbiota in mouse offspring of both sexes. *Transl. Psychiatry* **2022**, 12. <https://doi.org/10.1038/s41398-022-02149-9>.
80. Zhou R, Qian S, Cho WCS; et al. Microbiota-microglia connections in age-related cognition decline. *Aging Cell* **2022**, 21, e13599. <https://doi.org/10.1111/ace1.13599>.
81. Hayes LN, An K, Carloni E; et al. Prenatal immune stress blunts microglia reactivity, impairing neurocircuitry. *Nature* **2022**, 610, 327–334. <https://doi.org/10.1038/s41586-022-05274-z>.
82. Eberl G. A new age for (mucosal) NeuroImmunology. *Mucosal Immunol.* **2022**, 15, 1052–1055. <https://doi.org/10.1038/s41385-022-00573-0>.
83. Obata Y, Pachnis V. The Effect of Microbiota and the Immune System on the Development and Organization of the Enteric Nervous System. *Gastroenterology* **2016**, 151, 836–844. <https://doi.org/10.1053/j.gastro.2016.07.044>.
84. Jacobson A, Yang D, Vella M, Chiu IM. The intestinal neuro-immune axis: Crosstalk between neurons, immune cells, and microbes. *Mucosal Immunol.* **2021**, 14, 555–565. <https://doi.org/10.1038/s41385-020-00368-1>.
85. Davoli-Ferreira M, Thomson CA, McCoy KD. Microbiota and Microglia Interactions in ASD. *Front. Immunol.* **2021**, 12. <https://doi.org/10.3389/fimmu.2021.676255>.
86. Mitrea L, Nemeş SA, Szabo K, Teleky BE, Vodnar DC. Guts Imbalance Imbalances the Brain: A Review of Gut Microbiota Association with Neurological and Psychiatric Disorders. *Front. Med.* **2022**, 9. <https://doi.org/10.3389/fmed.2022.813204>.
87. Agirman G, Hsiao EY. SnapShot: The microbiota-gut-brain axis. *Cell* **2021**, 184, 2524–2524.e1. <https://doi.org/10.1016/j.cell.2021.03.022>.
88. Wang Y, Kasper LH. The role of microbiome in central nervous system disorders. *Brain Behav. Immun.* **2014**, 38:1–12. <https://doi.org/10.1016/j.bbi.2013.12.015>.



89. Marć MA, Jastrzab R, Mytych J. Does the Gut Microbial Metabolome Really Matter? The Connection between GUT Metabolome and Neurological Disorders. *Nutrients* **2022**, 14. <https://doi.org/10.3390/nu14193967>.
90. Passmore IJ, Letertre MPM, Preston MD; et al. Para-cresol production by *Clostridium difficile* affects microbial diversity and membrane integrity of Gram-negative bacteria. *PLoS Pathog.* **2018**, 14. <https://doi.org/10.1371/journal.ppat.1007191>.
91. Harrison MA, Kaur H, Wren BW, Dawson LF. Production of *p*-cresol by Decarboxylation of *p*-HPA by All Five Lineages of *Clostridioides difficile* Provides a Growth Advantage. *Front. Cell Infect. Microbiol.* **2021**, 11. <https://doi.org/10.3389/fcimb.2021.757599>.
92. Rogers AP, Mileto SJ, Lyras D. Impact of enteric bacterial infections at and beyond the epithelial barrier. *Nat. Rev. Microbiol.* **2023**, 21, 260–274. <https://doi.org/10.1038/s41579-022-00794-x>.
93. Cenit MC, Sanz Y, Codoñer-Franch P. Influence of gut microbiota on neuropsychiatric disorders. *World J. Gastroenterol.* **2017**, 23, 5486–5498. <https://doi.org/10.3748/wjg.v23.i30.5486>.
94. Doroszkiewicz J, Groblewska M, Mroczko B. The Role of Gut Microbiota and Gut-Brain Interplay in Selected Diseases of the Central Nervous System. *Int. J. Mol. Sci.* **2021**, 22. <https://doi.org/10.3390/ijms221810028>.
95. Santocchi E, Guiducci L, Prosperi M; et al. Effects of Probiotic Supplementation on Gastrointestinal, Sensory and Core Symptoms in Autism Spectrum Disorders: A Randomized Controlled Trial. *Front. Psychiatry* **2020**, 11. <https://doi.org/10.3389/fpsy.2020.550593>.
96. Banks WA, Kastin AJ, Broadwell RD. Passage of cytokines across the blood-brain barrier. *Neuroimmunomodulation* **1995**, 2, 241–248. <https://doi.org/10.1159/000097202>.
97. Kaelberer MM, Buchanan KL, Klein ME; et al. A gut-brain neural circuit for nutrient sensory transduction. *Science* **2018**, 361, eaat5236. <https://doi.org/10.1126/science.aat5236>.
98. Gaudier E, Jarry A, Blottière HM; et al. Butyrate specifically modulates MUC gene expression in intestinal epithelial goblet cells deprived of glucose. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2004**, 287, G1168–G1174. <https://doi.org/10.1152/ajpgi.00219.2004>.
99. Donohoe DR, Garge N, Zhang X; et al. The microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. *Cell Metab.* **2011**, 13, 517–526. <https://doi.org/10.1016/j.cmet.2011.02.018>.
100. Levy M, Blacher E, Elinav E. Microbiome, metabolites and host immunity. *Curr. Opin. Microbiol.* **2017**, 35, 8–15. <https://doi.org/10.1016/j.mib.2016.10.003>.
101. Vasquez A. Biological plausibility of the gut-brain axis in autism. *Ann. N. Y. Acad. Sci.* **2017**, 1408, 5–6. <https://doi.org/10.1111/nyas.13516>.
102. Clayton TA. Metabolic differences underlying two distinct rat urinary phenotypes, a suggested role for gut microbial metabolism of phenylalanine and a possible connection to autism. *FEBS Lett.* **2012**, 586, 956–961. <https://doi.org/10.1016/j.febslet.2012.01.049>.
103. Gevi F, Zolla L, Gabriele S, Persico AM. Urinary metabolomics of young Italian autistic children supports abnormal tryptophan and purine metabolism. *Mol. Autism* **2016**, 7. <https://doi.org/10.1186/s13229-016-0109-5>.
104. Xiong X, Liu D, Wang Y, Zeng T, Peng Y. Urinary 3-(3-Hydroxyphenyl)-3-hydroxypropionic Acid, 3-Hydroxyphenylacetic Acid, and 3-Hydroxyhippuric Acid Are Elevated in Children with Autism Spectrum Disorders. *Biomed. Res. Int.* **2016**, 2016, 9485412. <https://doi.org/10.1155/2016/9485412>.
105. Hitch TCA, Hall LJ, Walsh SK; et al. Microbiome-based interventions to modulate gut ecology and the immune system. *Mucosal Immunol.* **2022**, 15, 1095–1113. <https://doi.org/10.1038/s41385-022-00564-1>.
106. Santos J, Barbara G. Editorial: Human Intestinal Permeability, Mucosal Inflammation and Diet. *Front. Nutr.* **2022**, 9. <https://doi.org/10.3389/fnut.2022.894869>.
107. Toledo ARL, Monroy GR, Salazar FE; et al. Gut-Brain Axis as a Pathological and Therapeutic Target for Neurodegenerative Disorders. *Int. J. Mol. Sci.* **2022**, 23. <https://doi.org/10.3390/ijms23031184>.
108. Marogianni C, Sokratous M, Dardiotis E, Hadjigeorgiou GM, Bogdanos D, Xiromerisiou G. Neurodegeneration and Inflammation-An Interesting Interplay in Parkinson's Disease. *Int. J. Mol. Sci.* **2020**, 21. <https://doi.org/10.3390/ijms21228421>.
109. Xie J, Van Hoecke L, Vandenbroucke RE. The Impact of Systemic Inflammation on Alzheimer's Disease Pathology. *Front. Immunol.* **2022**, 12. <https://doi.org/10.3389/fimmu.2021.796867>.
110. Giri R, Hoedt EC, Khushi S; et al. Secreted NF-κB suppressive microbial metabolites modulate gut inflammation. *Cell Rep.* **2022**, 39, 110646. <https://doi.org/10.1016/j.celrep.2022.110646>.
111. Saresella M, Piancone F, Marventano I; et al. Multiple inflammasome complexes are activated in autistic spectrum disorders. *Brain Behav. Immun.* **2016**, 57, 125–133. <https://doi.org/10.1016/j.bbi.2016.03.009>.
112. Jyonouchi H, Geng L. Associations between Monocyte and T Cell Cytokine Profiles in Autism Spectrum Disorders: Effects of Dysregulated Innate Immune Responses on Adaptive Responses to Recall Antigens in a Subset of ASD Children. *Int. J. Mol. Sci.* **2019**, 20. <https://doi.org/10.3390/ijms20194731>.



113. Wanchao S, Chen M, Zhiguo S, Futang X, Mengmeng S. Protective effect and mechanism of *Lactobacillus* on cerebral ischemia reperfusion injury in rats. *Braz. J. Med. Biol. Res.* **2018**, *51*, e7172. <https://doi.org/10.1590/1414-431x20187172>.
114. Tomova A, Husarova V, Lakatosova S; et al. Gastrointestinal microbiota in children with autism in Slovakia. *Physiol. Behav.* **2015**, *138*, 179-187. <https://doi.org/10.1016/j.physbeh.2014.10.033>.
115. Luna RA, Oezguen N, Balderas M; et al. Distinct Microbiome-Neuroimmune Signatures Correlate with Functional Abdominal Pain in Children with Autism Spectrum Disorder. *Cell Mol. Gastroenterol. Hepatol.* **2016**, *3*, 218. <https://doi.org/10.1016/j.jcmgh.2016.11.008>.
116. Careaga M, Rogers S, Hansen RL, Amaral DG, Van de Water J, Ashwood P. Immune Endophenotypes in Children with Autism Spectrum Disorder. *Biol. Psychiatry* **2017**, *81*, 434-441. <https://doi.org/10.1016/j.biopsych.2015.08.036>.
117. Groer MW, Gregory KE, Louis-Jacques A, Thibeau S, Walker WA. The very low birth weight infant microbiome and childhood health. *Birth Defects Res. C Embryo Today* **2015**, *105*, 252-264. <https://doi.org/10.1002/bdrc.21115>.
118. Collado MC, Rautava S, Aakko J, Isolauri E, Salminen S. Human gut colonisation may be initiated in utero by distinct microbial communities in the placenta and amniotic fluid. *Sci. Rep.* **2016**, *6*. <https://doi.org/10.1038/srep23129>.
119. Li N, Yang J, Zhang J; et al. Correlation of Gut Microbiome Between ASD Children and Mothers and Potential Biomarkers for Risk Assessment. *Genom. Proteom. Bioinform.* **2019**, *17*, 26-38. <https://doi.org/10.1016/j.gpb.2019.01.002>.
120. Jaini R, Wolf MR, Yu Q, King AT, Frazier TW Jr, Eng C. Maternal genetics influences fetal neurodevelopment and postnatal autism spectrum disorder-like phenotype by modulating in-utero immunosuppression. *Transl. Psychiatry* **2021**, *11*. <https://doi.org/10.1038/s41398-021-01472-x>.
121. Vuong HE, Hsiao EY. Emerging Roles for the Gut Microbiome in Autism Spectrum Disorder. *Biol. Psychiatry* **2017**, *81*, 411-423. <https://doi.org/10.1016/j.biopsych.2016.08.024>.
122. Tamana SK, Tun HM, Konya T; et al. Bacteroides-dominant gut microbiome of late infancy is associated with enhanced neurodevelopment. *Gut Microbes* **2021**, *13*, 1-17. <https://doi.org/10.1080/19490976.2021.1930875>.
123. Strati F, Cavalieri D, Albanese D; et al. New evidences on the altered gut microbiota in autism spectrum disorders. *Microbiome* **2017**, *5*. <https://doi.org/10.1186/s40168-017-0242-1>.
124. Alshammari MK, AlKhulaifi MM, Al Farraj DA, Somily AM, Albarrag AM. Incidence of *Clostridium perfringens* and its toxin genes in the gut of children with autism spectrum disorder. *Anaerobe* **2020**, *61*, 102114. <https://doi.org/10.1016/j.anaerobe.2019.102114>.
125. Vuong HE, Yano JM, Fung TC, Hsiao EY. The Microbiome and Host Behavior. *Annu. Rev. Neurosci.* **2017**, *40*, 21-49. <https://doi.org/10.1146/annurev-neuro-072116-031347>.
126. Taguer M, Maurice CF. The complex interplay of diet, xenobiotics, and microbial metabolism in the gut: Implications for clinical outcomes. *Clin. Pharmacol. Ther.* **2016**, *99*, 588-599. <https://doi.org/10.1002/cpt.366>.
127. Kovtun AS, Averina OV, Alekseeva MG, Danilenko VN. Antibiotic Resistance Genes in the Gut Microbiota of Children with Autistic Spectrum Disorder as Possible Predictors of the Disease. *Microb. Drug Resist.* **2020**, *26*, 1307-1320. <https://doi.org/10.1089/mdr.2019.0325>.
128. Kantarcioglu AS, Kiraz N, Aydin A. Microbiota-Gut-Brain Axis: Yeast Species Isolated from Stool Samples of Children with Suspected or Diagnosed Autism Spectrum Disorders and In Vitro Susceptibility Against Nystatin and Fluconazole. *Mycopathologia* **2016**, *181*, 1-7. <https://doi.org/10.1007/s11046-015-9949-3>.
129. McElhanon BO, McCracken C, Karpen S, Sharp WG. Gastrointestinal symptoms in autism spectrum disorder: A meta-analysis. *Pediatrics* **2014**, *133*, 872-883. <https://doi.org/10.1542/peds.2013-3995>.
130. Ristori MV, Quagliariello A, Reddel S; et al. Autism, Gastrointestinal Symptoms and Modulation of Gut Microbiota by Nutritional Interventions. *Nutrients* **2019**, *11*. <https://doi.org/10.3390/nu11112812>.
131. Goines P, Van de Water J. The immune system's role in the biology of autism. *Curr. Opin. Neurol.* **2010**, *23*, 111-117. <https://doi.org/10.1097/WCO.0b013e3283373514>.
132. Kim CH. Immune regulation by microbiome metabolites. *Immunology* **2018**, *154*, 220-229. <https://doi.org/10.1111/imm.12930>.
133. Adams JB, Johansen LJ, Powell LD, Quig D, Rubin RA. Gastrointestinal flora and gastrointestinal status in children with autism--comparisons to typical children and correlation with autism severity. *BMC Gastroenterol.* **2011**, *11*. <https://doi.org/10.1186/1471-230X-11-22>.
134. Chang J, Gilman SR, Chiang AH, Sanders SJ, Vitkup D. Genotype to phenotype relationships in autism spectrum disorders. *Nat. Neurosci.* **2015**, *18*, 191-198. <https://doi.org/10.1038/nn.3907>.
135. Zhang Y, Li N, Li C; et al. Genetic evidence of gender difference in autism spectrum disorder supports the female-protective effect. *Transl. Psychiatry* **2020**, *10*. <https://doi.org/10.1038/s41398-020-0699-8>.
136. Duda M, Zhang H, Li HD, Wall DP, Burmeister M, Guan Y. Brain-specific functional relationship networks inform autism spectrum disorder gene prediction. *Transl. Psychiatry* **2018**, *8*. <https://doi.org/10.1038/s41398-018-0098-6>.

137. Lefebvre A, Tillmann J, Cliquet F; et al. Tackling hypo and hyper sensory processing heterogeneity in autism: From clinical stratification to genetic pathways. *Autism Res.* **2023**, *16*, 364–378. <https://doi.org/10.1002/aur.2861>.
138. Trost B, Thiruvahindrapuram B, Chan AJS; et al. Genomic architecture of autism from comprehensive whole-genome sequence annotation. *Cell* **2022**, *185*, 4409–4427.e18. <https://doi.org/10.1016/j.cell.2022.10.009>.
139. Nova E, Gómez-Martínez S, González-Soltero R. The Influence of Dietary Factors on the Gut Microbiota. *Microorganisms* **2022**, *10*. <https://doi.org/10.3390/microorganisms10071368>.
140. Kang DW, Adams JB, Gregory AC; et al. Microbiota Transfer Therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: An open-label study. *Microbiome* **2017**, *5*. <https://doi.org/10.1186/s40168-016-0225-7>.
141. Kang DW, Adams JB, Coleman DM; et al. Long-term benefit of Microbiota Transfer Therapy on autism symptoms and gut microbiota. *Sci. Rep.* **2019**, *9*. <https://doi.org/10.1038/s41598-019-42183-0>.
142. Santocchi E, Guiducci L, Prosperi M; et al. Effects of Probiotic Supplementation on Gastrointestinal, Sensory and Core Symptoms in Autism Spectrum Disorders: A Randomized Controlled Trial. *Front. Psychiatry* **2020**, *11*. <https://doi.org/10.3389/fpsy.2020.550593>.
143. Wang Y, Li N, Yang JJ; et al. Probiotics and fructo-oligosaccharide intervention modulate the microbiota-gut brain axis to improve autism spectrum reducing also the hyper-serotonergic state and the dopamine metabolism disorder. *Pharmacol. Res.* **2020**, *157*, 104784. <https://doi.org/10.1016/j.phrs.2020.104784>.
144. Taniya MA, Chung HJ, Al Mamun A, Alam S, Aziz MA, Emon NU, Islam MM, Hong SS, Podder BR, Ara Mimi A, Aktar Suchi S, Xiao J. Role of Gut Microbiome in Autism Spectrum Disorder and Its Therapeutic Regulation. *Front. Cell Infect. Microbiol.* **2022**, *12*, 915701. <https://doi.org/10.3389/fcimb.2022.915701>.
145. Zou R, Wang Y, Duan M, Guo M, Zhang Q, Zheng H. Dysbiosis of Gut Fungal Microbiota in Children with Autism Spectrum Disorders. *J. Autism Dev. Disord.* **2021**, *51*, 267–275. <https://doi.org/10.1007/s10803-020-04543-y>.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.