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Article

Gulf War Illness Induced Sex-Specific Transcriptional Differences Under Stressful Conditions

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Abstract: Gulf War Illness (GWI) is a multi-symptom disorder affecting 1990–1991 Persian Gulf War veterans and is characterized by post-exertional malaise, neurological symptoms, immune deregulation, and exhaustion. Causation is not understood, and effective diagnostics and therapies have not yet been developed. In this work, we analyzed stress-related, sex-specific transcriptomic shifts in GWI subjects and healthy controls through RNA sequencing of peripheral blood mononuclear cells (PBMCs). Blood samples at baseline (T0), at maximal exertion (T1), and four hours post-exertion (T2) were analyzed. In female subjects with GWI proinflammatory processes, and in male subjects IL-12 signaling and lymphocytic activation were deregulated at T1 compared to T0. During recovery from stress, HCMV activity and microglia activation increased in female GWI subjects, and apoptotic signaling changed in males with GWI. Documented sex-specific immune deregulation warrants personalized markers and therapies. Targeting sex-specific transcriptomic markers of disease could lead to new therapies for GWI.

Keywords: Gulf War Illness (GWI); transcriptomics; sex differences; Peripheral Blood Mononuclear Cells (PBMCs); RNA Sequencing (RNA-seq); immune dysregulation; oxidative stress; inflammation; cytokine signaling

1. Introduction

Gulf War Illness (GWI) is a multi-symptom, complex, and debilitating disease, which has been reported by the veterans who were deployed in the 1990-1991 Persian Gulf War [1,2]. Veterans with GWI commonly present with fatigue, neurological dysfunction, neuromuscular and joint pain, symptoms related to the gastrointestinal tract, impaired immune functions, sensitivity to odors, chemicals, and light, post-exertional malaise, and other related symptoms that were reported during or immediately after the Gulf War [3–6]. It is suggested that GWI is caused by a combination of factors such as toxic exposure and modifications of epigenetic profiles [7–9]. While some advances have been made to determine the underlying mechanisms of GWI, the exact causes of GWI remain unclear [10] and no effective diagnostic approaches and/or treatment modalities have been made available to date [11,12].

While the underlying mechanisms of GWI are not fully elucidated, research efforts have highlighted some novel approaches to better understand the pathophysiology. As part of our

ongoing research efforts, we carried out transcriptomic profiling of circulating immune cells from male GWI patients and healthy controls (HCs) to gain insights into the gene expression dysregulation tied to GWI and stress [13]. Using an exercise challenge protocol, we collected blood samples from male GWI and HC subjects at baseline (T0) before the stress event (exercise), at the point of maximal exertion (T1), and four hours after recovery from maximal exertion (T2). Peripheral blood mononuclear cells (PBMCs) were used to extract RNA and perform transcriptomics profiling using RNA sequencing (RNA-seq) at each time point. Our data identified dysregulation in immune and inflammatory pathways and novel immune and inflammatory markers in GWI-male compared to HCs [13].

In the current study, we extended our exercise challenge strategy to both male and female GWI patients and HCs to identify sex-dependent transcriptional changes in response to stress in GWI. We performed RNA-seq on PBMCs from female and male GWI patients and HCs under the same modeled stress conditions to evaluate transcriptomic alterations and affected molecular pathways. We identified sex-dependent pathways at maximal exertion (such as regulation of leukocyte chemotaxis, oxidative stress pathways, leukocyte activation response to a toxic substance, IL12, and proinflammatory and profibrotic mediators) and at recovery from maximal exertion (such as human cytomegalovirus (HCMV) or human herpes virus 5 pathways (HHV-5), oxidative stress-induced senescence, the regulation of microglial cell activation, negative regulation of the extrinsic apoptotic signaling pathway and negative regulation of type II interferon production).

Our findings highlight the sex-dependent nature of GWI pathogenesis, which could be used to more effectively diagnose and treat GWI patients. It is our overarching goal to target treatment more effectively, outside symptom amelioration, and identify therapeutic targets for GWI.

2. Results

2.1. Participant Characteristics

25 female and 19 male GWI subjects and 17 female and 25 male age- and body mass index (BMI)-matched HCs participated in this study. We found no significant differences based on age and BMI between male and female GWI subjects or male and female HCs (Table 1). Levels of mental and physical disability were evaluated via the Short Form 36-item Survey (SF-36) questionnaire [14]. Scores were evaluated on a 100-point scale with lower scores indicating greater levels of disability. Female GWI subjects showed significant differences ($p < 0.05$) in physical and mental health when compared to HCs. We have previously published on the effects of GWI on the levels of mental and physical disability of male patients [13].

Table 1. Demographic information comparing Female GWI subjects and HCs, including SF-36 questionnaire data. Data are shown as mean \pm standard error of the mean, $*-p \leq 0.05$, Student t-test.

Category	GWI Females	Healthy Controls	<i>p</i> -Value
Age	53.2 \pm 1.4	49.8 \pm 1.4	0.09
BMI	29.0 \pm 1.0	26.6 \pm 1.2	0.16
Physical Health			
Physical Function	49.0 \pm 4.8	96.3 \pm 1.5	<0.00*
Role-Physical	20.0 \pm 6.6	98.4 \pm 1.5	<0.00*
Body Pain	33.6 \pm 4.0	90.7 \pm 2.9	<0.00*
General Health	37.7 \pm 4.4	79.0 \pm 4.6	<0.00*

Mental-Health				
	Vitality	32.8±4.1	65.7±6.3	<0.00*
	Social Function	24.9±5.0	95.3±2.2	<0.00*
	Role Emotional	36±8.5	95.8±2.8	<0.00*

2.2. Transcriptomic Changes Between Maximum Exertion (T1) and Baseline Before Exercise Challenge (T0) Stratified by Sex

Gene expression in PBMCs of both GWI subjects and HCs was compared at T1 (maximal exertion) versus T0 (baseline). These results were stratified by sex and differentially expressed genes (DEG) were analyzed using Metascape [15] for functional comparative analysis.

Female GWI subjects had 27 significant DEGs at T1 vs. T0, six of which were underexpressed and 21 of which were overexpressed (Supplementary Table S1). When comparing female HCs at T1 vs. T0, significant differential expression in 60 genes was found. Of these 60 genes, 59 were overexpressed and one was underexpressed (Supplementary Table S2). DEGs in male HCs and GWI subjects were compared in our previous publication [13].

To identify sex differences in affected pathways and ontologies in both GWI and HC cohorts in response to the maximal exertion, DEGs were analyzed through comparative functional analysis of gene networks using Metascape [15]. Specifically, we focused on the functional groups related to immune responses, detoxification, and oxidative stress, because these processes have been shown to be disrupted in GWI models [16]. We compared the most affected pathways and ontologies between male and female cohorts (Figures 1A,B and 2A).

In female GWI patients compared to males, pathways of high significance included the proinflammatory and profibrotic mediators, regulation of leukocyte chemotaxis, response to toxic substances, and secretion by cells were the pathways of high significance (Figure 2A). In male GWI patients, unlike females, pathways that were observed to have high significance included the interleukin (IL)-12 pathway, natural killer (NK)-mediated cytotoxicity, immunoregulatory interactions between a lymphoid and non-lymphoid cell, regulation of lymphocyte-mediated immunity, regulation of leukocyte-mediated immunity, lymphocyte activation, regulation of NK cell-mediated immunity, and positive regulation of NK cell-mediated cytotoxicity (Figure 2A).

Of note, both female and male GWI patients demonstrated significant to various degrees changes in cell activation, leukocyte activation, positive regulation of immune effector process, positive regulation of defense response, innate immune response, IL-10 signaling, cell killing and regulation of leukocyte chemotaxis (Figure 2A).

In female HCs, unlike their healthy male counterparts, demonstrated highly significant changes in regulated exocytosis, cellular response to oxidative stress, and secretion by cells (Figure 2A). At the same time, in male HCs, we found highly significant positive regulation of NK cell-mediated cytotoxicity, development and heterogeneity of the innate lymphoid cell (ILC) family, immune response-activating signaling pathway, immune response-activating cell surface receptor signaling pathway, regulation of chemotaxis, and viral protein interaction with cytokine and cytokine receptor (Figure 2A).

Both female and male HCs were found to have significantly to various degrees changed pathways of negative regulation of leukocyte activation, immunoregulatory interactions between a lymphoid and a non-lymphoid cell, and the primary interaction database (PID) IL-12 pathway (Figure 1A,B and 2A).

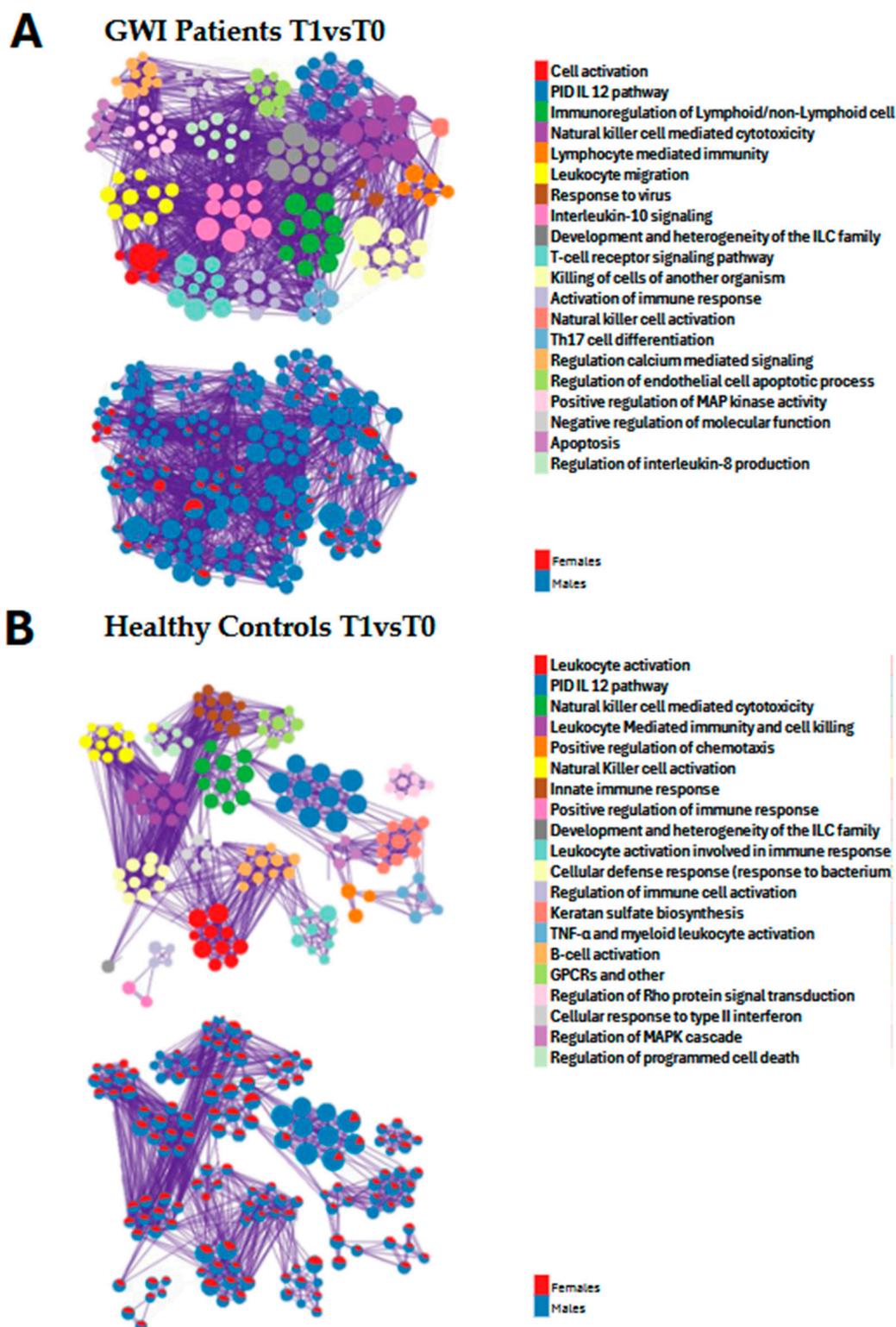


Figure 1. Metascape [15] express analysis of DEGs between male and female GWI patients (A) and HCs (B) between T1 and T0. Each node represents an enriched term (right). The larger the representation of color the more enriched it is in each cohort. Cutoff values included a p-value <0.01, a minimum count of 3 in differential expression, and an enrichment factor of >1.5 GO biological processes, KEGG pathways, Reactome gene sets, CORUM complexes, and canonical pathways from MSigDB were included in the search; (left) nodes are colored by ratios between male (blue) and female (red) participants.

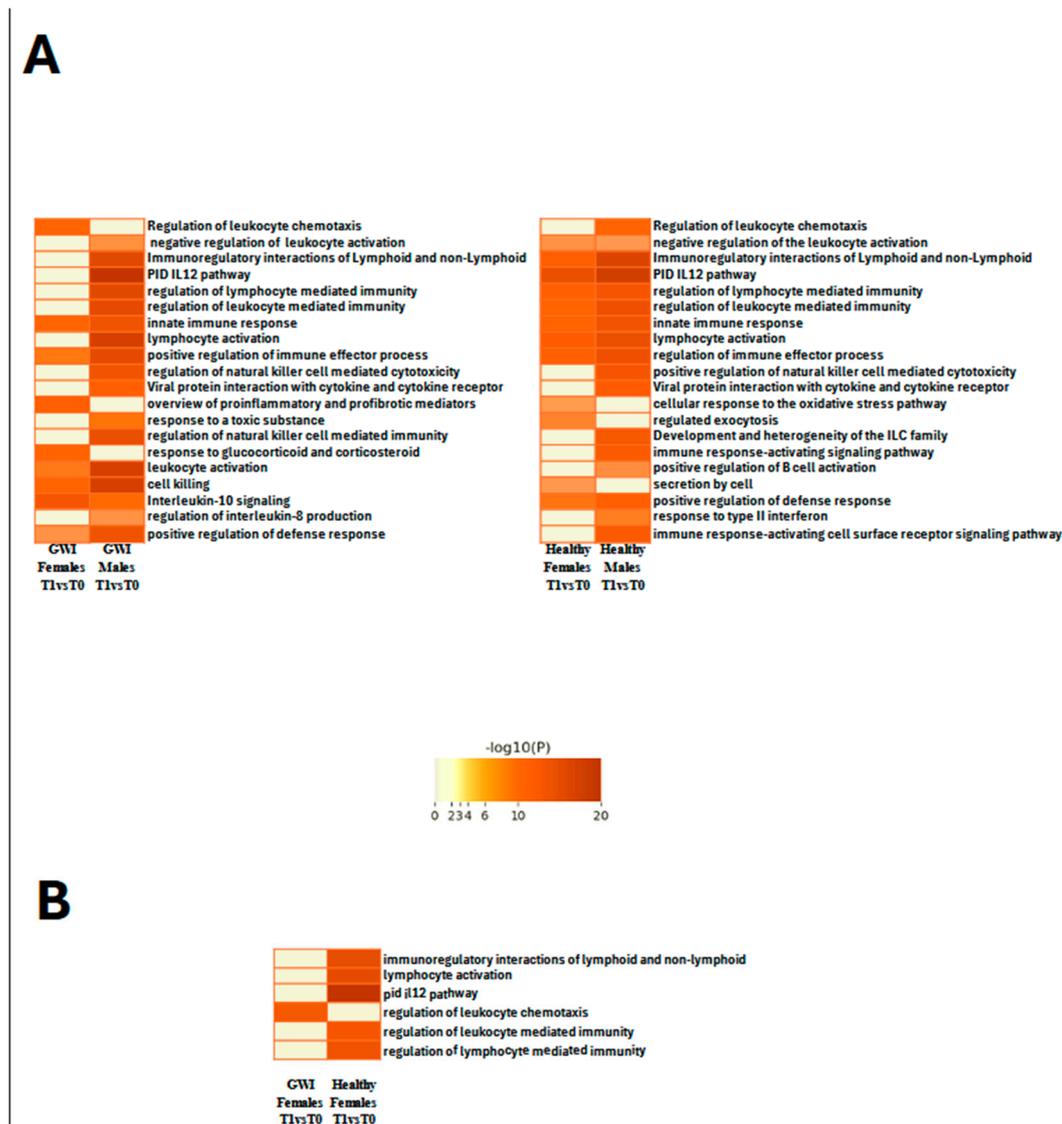


Figure 2. (A) heatmap of the individual pathways that showed the largest difference between T1 and T0 in PBMCs of male and female participants which were found to match those pathways discovered in HCs or exhibited a change in which cohort was affected compared to HCs. (B) comparison of enriched pathways between female GWI veterans and healthy controls.

2.3. Transcriptomic Changes Between T2 (4h After Maximal Exertion) and T1 (Maximal Exertion)

Transcriptomic changes in PBMCs were compared in GWI patients and HCs between recovery from exercise challenge (T2) and maximal exertion (T1) time points. The results were stratified by sex and functional comparative analysis, which was performed using Metascape [15].

Female GWI subjects had 675 DEGs between T2 and T1, 442 of which were underexpressed and 232 were overexpressed (Supplementary Table S3). However, when comparing gene expression in female HCs at T2 vs. T1, 585 DEGs were found. Of these 584 DEGs, 232 were overexpressed and 352 were underexpressed (Supplementary Table S4). Results of differential gene expression in male GWI patients and HC have been published previously [13].

Functional pathway analysis conducted on DEGs between male and female GWI cohorts at T2 and T1 demonstrated significant changes between the sexes (Figures 3A and 4A). In the male GWI cohort, unlike females, pathways of high significance included those associated with the mucosal immune response, tissue-specific immune response, antibacterial humoral response, immunoregulatory interactions between a lymphoid and a non-lymphoid cell, regulation of NK cell-

mediated immunity and cytotoxicity, positive regulation of IL-12 production, and positive regulation of intrinsic apoptotic signaling pathway (Figure 4A).

In the female GWI cohort compared to males, pathways of high significance included those associated with negative regulation of type II interferon production, leukocyte differentiation, regulation of neuroinflammatory response, regulation of microglial cell activation, regulation of inflammatory response, negative regulation of extrinsic apoptotic signaling pathway, and negative regulation of oxidative stress-induced senescence pathway (Figure 4A).

In addition, pathways of high significance to various degrees in both male and female GWI patients included those tied to systemic lupus erythematosus, activated *PKN1*, HCMV events, activation of HOX genes during differentiation, oxidative stress-induced senescence, lymphocyte activation, and immune response-activating signaling pathways (Figure 4A).

Regulation of the neuroinflammatory response was highly significant in female HCs, unlike their male HC counterparts (Figure 4A). We also observed that in both male and female HCs, pathways of high significance included those tied to immunoregulatory interactions between a lymphoid and a non-lymphoid cell, regulation of NK cell-mediated immunity and cytotoxicity, lymphocyte activation and differentiation, immune response-activating signaling pathway, regulation of microglial cell activation, regulation of extrinsic apoptotic signaling pathway via death domain receptors, and regulation of intrinsic apoptotic signaling were pathways (Figures 3A,B and 4A).

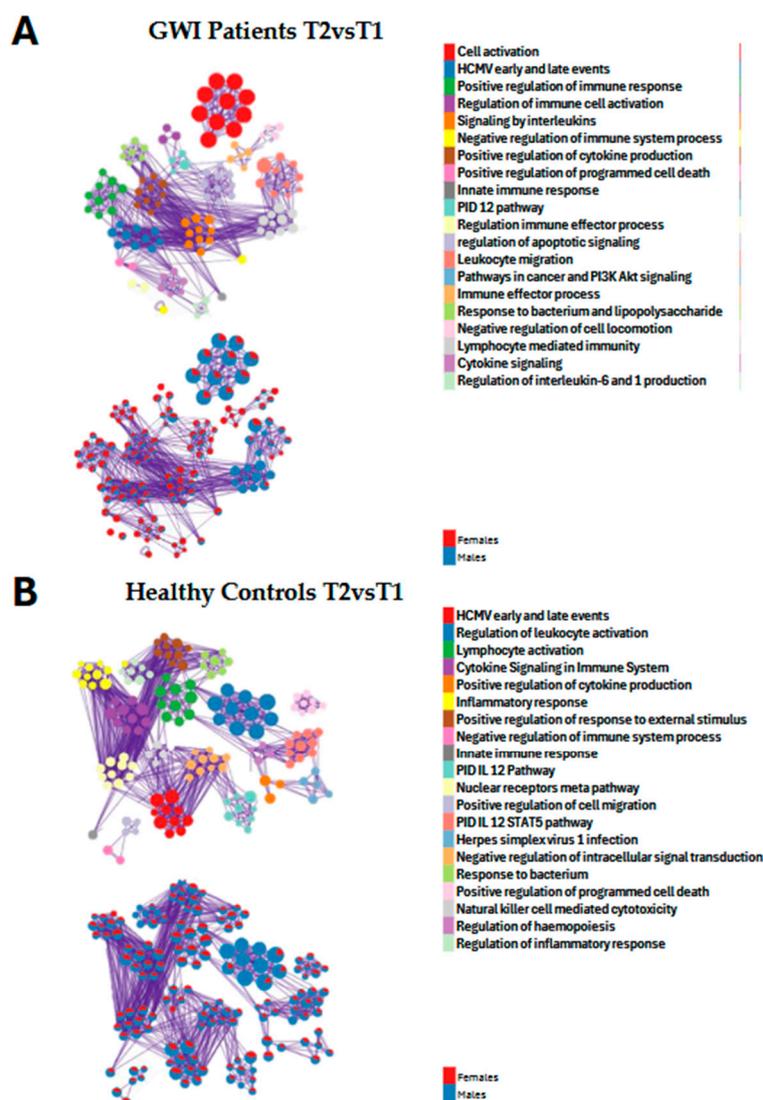


Figure 3. Metascape [15] express analysis of DEGs between male and female GWI patients (A) and HCs (B) between T2 and T1. Each node represents an enriched term (right). The larger the representation of color the more enriched it is in each cohort. Cutoff values included a p-value <0.01, a minimum count of 3 in differential expression, and an enrichment factor of >1.5 GO biological processes, KEGG pathways, Reactome gene sets, CORUM complexes, and canonical pathways from MSigDB were included in the search; (left) nodes are colored by ratios between male (blue) and female (red) participants.

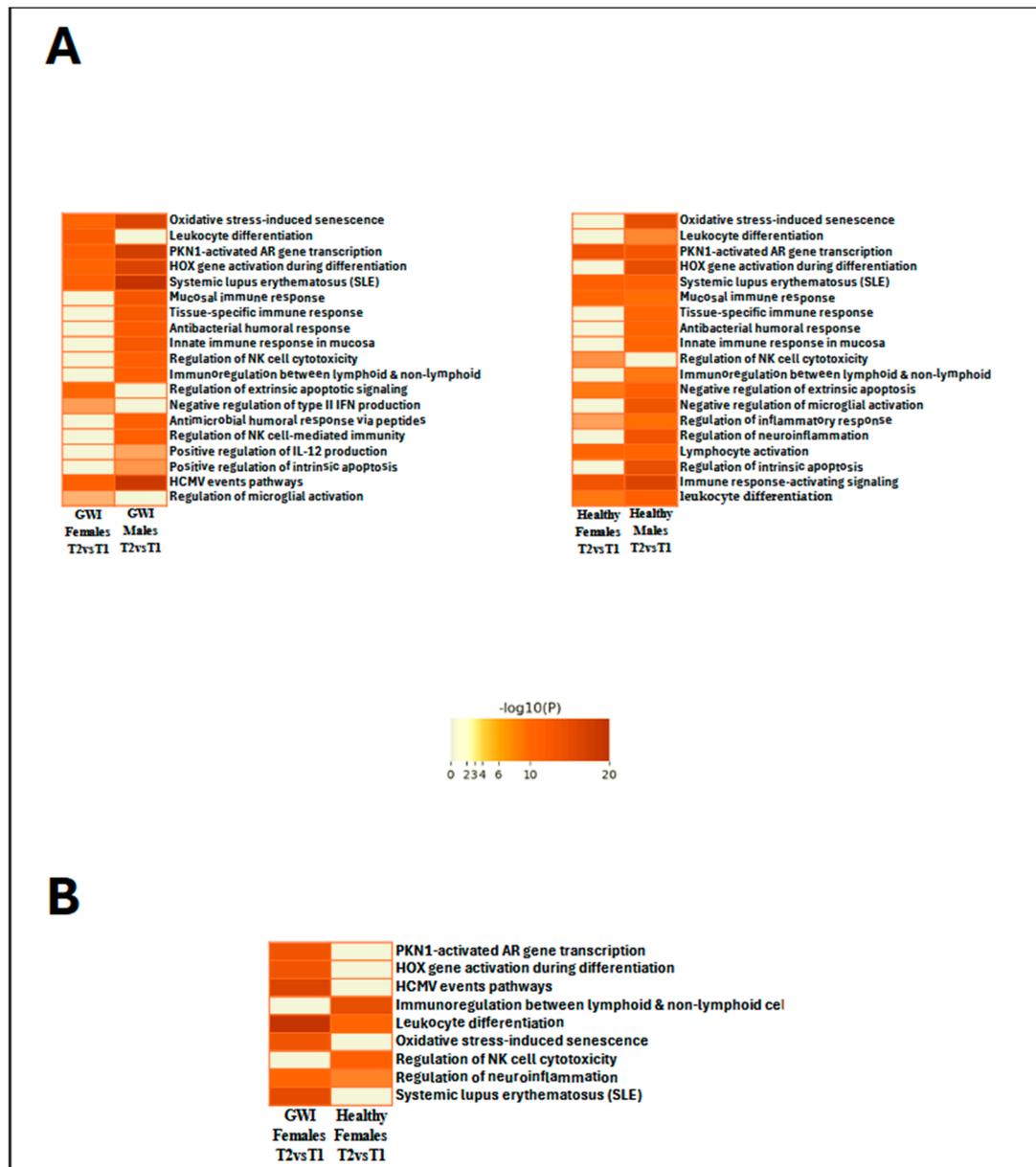


Figure 4. (A) heatmap of the individual pathways that showed the largest difference between T1 and T0 in PBMCs of male and female participants which were found to match those pathways discovered in HCs, or exhibited a change in which cohort was affected compared to HCs. (B) comparison of enriched pathways between female GWI veterans and healthy controls.

2.4. Nanostring Validation

95 percent of the DEGs identified by RNA-Seq were validated by Nanostring technology (Supplementary Table S5a–d). Genes with discrepancies between Nanostring and RNA-seq were not used in the functional analysis.

3. Discussion

This project was the first effort to analyze sex differences in the transcriptomes of circulating immune cells among GWI patients under stress. Specifically, we evaluated transcriptomic changes provoked by exercise challenge, first, between the point of maximal exertion (T1) and baseline (T0), and second, between recovery (T2) and the point of maximal exertion (T1). Our results show that biological sex is likely to influence gene expression of PBMCs in GWI.

3.1. Transcriptomic Changes Between Maximal Exertion (T1) and Baseline Before Exercise (T0) in GWI Patients and HCs

We found significant changes in response to the exercise challenge in the PBMCs of all participants. Functional analysis of the DEGs revealed a broad number of affected pathways.

Functional analysis of DEGs in male GWI patients, as compared to their female GWI counterparts, in response to maximal exertion (T1 vs. T0) showed that positive regulation of NK cell-mediated cytotoxicity, and regulation of leukocyte chemotaxis were highly affected. These pathways were also significantly affected in the HCs.

3.1.1. Regulation of Leukocyte Chemotaxis

We found that gene networks tied to the regulation of leukocyte chemotaxis were of significance among female GWI (Figures 1A and 2A) and male HC subjects (Figures 1B and 2A), unlike their male GWI and female HC counterparts. This could allude to potential underlying sex-dependent regulatory mechanisms in GWI. Pro-inflammatory cytokines were found to be increased in GWI veterans, contributing to the dysregulation of leukocyte chemotaxis from a cytokine imbalance [17]. In addition, in patients with Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS), a disease that shares many symptoms with GWI, pro-chemotactic factors are significantly deregulated [18–20]. Given what has been published, this could indicate the potential for a biomarker of GWI in female patients. The absence of an effect on this pathway in male GWI patients may require more research.

3.1.2. Oxidative Stress

We observed that pathways related to the cellular response to oxidative stress are uniquely activated in female HCs compared to males, however, changes in these pathways are not significant enough in GWI subjects (Figure 2A). The absence of changes in oxidative stress upon exertion in GWI cohorts could indicate that at baseline, GWI patients are already experiencing a level of exertion similar to the T1 time point of HCs.

Oxidative stress pathways were found to be significantly deregulated in the rat model of GWI. In addition, malondialdehyde (MDA), a by-product of peroxidation, is an oxidative stress marker that was also found to be increased in rat models of GWI [16]. Furthermore, stress studies in ME/CFS patients also found increased markers of oxidative stress at rest that further rose after exercise [21].

3.1.3. Response to a Toxic Substance

Response to a toxic substance was shown to be uniquely deregulated in male GWI subjects compared to females (Figure 2A). It has been well established that Gulf War veterans were exposed to various environmental toxins [22]. To reduce cellular damage from chemical exposures, redox balance pathways are utilized [23]. Considering that female GWI subjects have deregulated changes in oxidative stress pathways (see above), this could hint at a sex-mediated mechanism, in which GWI females are unable to maintain redox balance and detoxification processes.

3.1.4. Immune Dysfunction

Functional analysis of DEGs showed that the negative regulation of leukocyte activation, immunoregulatory interactions between a lymphoid and a non-lymphoid cell, regulation of NK cell-mediated cytotoxicity, and the PID IL-12 pathway were significantly enriched in both HC male and female subjects and in male GWI subjects, but not among female GWI subjects (Figure 2A). This could suggest that there is a dysregulation of these functions in female GWI subjects.

3.1.5. Leukocyte Activation and Lymphoid/Non-Lymphoid

Leukocyte activation and lymphoid/non-lymphoid cell pathways are significantly changed between T1 and T0 in all cohorts, except for GWI females (Figure 2A). Although regulation of the leukocyte activation pathway isn't fully understood in GWI patients, research suggests that factors such as immune system dysregulation can contribute to negative leukocyte regulation. Specifically, it was found that the immune dysfunction in GWI subjects results from decreased NK cytotoxicity, immune cell impairment, changes in the levels of pro-inflammatory cytokines, and cytokine signaling [24]. Together, these immune changes can cause defects in the immune cell signaling of regulatory pathways that serve to dampen the immune response, contributing to higher levels of inflammation [24]. Dysregulated communication between lymphoid and non-lymphoid cells was observed when analyzing the innate immune system among GWI subjects [25]. The pathophysiology of ME/CFS is also associated with dysregulation of the innate immune system [26]. Despite the overall increased levels of inflammation seen in ME/CFS, aspects of negative regulation of leukocyte activation are also present [27,28].

3.1.6. IL-12

IL-12 pathways are changed between T1 and T0 in all cohorts, except for GWI females (Figures 1B and 2A). IL-12 is a pro-inflammatory cytokine that participates in various immune functions and responses, particularly in antigen presentation and differentiation of naive T cells. These cytokines also regulate the functions of various effector cells and are often therapeutic targets in inflammatory and autoimmune diseases [29]. Prior studies have shown that the IL-12 response demonstrates significant sex-specific differences. Male GWI subjects showed significant alterations to the IL-12 signaling pathway. IL-12 stimulates the proliferation and activation of T cells and NK cells [30,31] and the release of IFN- γ [31]. IFN- γ has also been characterized as a pro-apoptotic factor [32], which may be related to the dysregulated intrinsic apoptotic signaling pathway in male GWI patients (cell killing in Figure 2D). Patients with ME/CFS were also found to have dysregulation in the IL-12 pathway, most prominently in males [33]. Though further study is warranted, it has been found that in ME/CFS patients who were exposed to toxic substances showed greater levels of immune dysregulation [34].

3.1.7. Proinflammatory and Profibrotic Mediators

Only in the female GWI cohort, pathways related to proinflammatory and profibrotic mediators were enriched (Figure 2A). This enrichment could be associated with immune responses among female GWI patients. Previous studies have found that estrogen may inhibit cytokine storm [35]. This hints at a potential sex hormone-mediated mechanism that results in a deregulated response to proinflammatory and profibrotic mediators in female GWI subjects (Figure 2C). Furthermore, recent literature on GWI has shown that patients with GWI were found to have increased levels of pro-inflammatory cytokines [31,36]. It was also published that the pathophysiology of ME/CFS is related to the activation of multiple immune-inflammatory pathways [37].

3.1.8. Overall

Based on these findings, GWI patients appear to exhibit multiple pathways contributing to their pathological response to stress. Notably, genes related to toxic exposure and oxidative stress, key

mechanisms for combating environmental toxins, are significantly altered in GWI patients during their initial stress response. These findings align with GWI patients' documented exposure to hazardous substances such as organophosphates, pyridostigmine bromide, and combustion products and suggest that an inability of their body to adapt and or break down these hazards may be a key driver of the disease. Additionally, chronic immune system dysregulation likely plays a central role in their impaired stress response, as genes associated with immune cell function, immune mediators, and signaling molecules show significant alterations.

3.2. Transcriptomic Changes Between Four Hour After Maximal Exertion (T2) and Maximal Exertion (T1) in GWI Patients and HCs

3.2.1. HCMV Events

Functional analysis of DEGs in GWI subjects during recovery (T2 vs. T1) showed that HCMV events were found to be significantly enriched among female GWI subjects, as compared to female HCs (Figure 4B). HCMV is a member of the herpes virus family, otherwise known as HHV-5. These findings could suggest that HCMV progression is a potential avenue for additional research into biomarkers of GWI disease activity among female patients.

The reason HCMVs could be more active among females with GWI compared to healthy females has yet to be elucidated. Meanwhile, previous studies on other herpes viruses, such as HHV-6, revealed that patients have higher serum antibody levels of IgG. This implies that viral enzyme activity contributes to the hormone response [38].

HCMV gene changes in female subjects with GWI imply that viral reactivation could contribute to disease, similar to long COVID and ME/CFS [39].

3.2.2. Oxidative Stress-Induced Senescence

Oxidative stress-induced senescence occurs at high concentrations of reactive oxygen species (ROS) in a cell and contributes to cellular senescence through various downstream protein interactions [40,41]. The changes of this pathway during recovery among healthy females but not females with GWI (Figures 3A and 4B), suggests that it could be involved in sex-specific GWI onset and/or progression. The underlying mechanisms tied to the oxidative stress-induced senescence pathway associated with GWI have yet to be fully elucidated. Higher levels of oxidative stress can decrease antioxidant levels, promoting cellular damage and senescence [41,42].

3.2.3. Leukocyte Differentiation

In addition, we found that leukocyte differentiation was significantly enriched in female GWI patients, as compared to male and female HCs in response to recovery (Figure 4A). As such, this pathway could provide a method for differentiating between GWI patients and HCs. Previous publications have observed that the differentiation of various immune cells is broadly inhibited by anti-estrogens, which could indicate that leukocyte differentiation is upregulated by high estrogen levels [43]. This could provide a reasonable explanation as to why leukocyte differentiation was enriched in female GWI subjects and not their male counterparts. While leukocyte differentiation hasn't been explicitly studied in GWI, it may be assumed that pro-inflammatory cytokines excessively promote the differentiation of leukocytes [6].

3.2.4. Regulation of Extrinsic Apoptotic Signaling

The regulation of the extrinsic apoptotic signaling pathway is significantly deregulated in GWI females (Figure 4A). It is unclear how this correlates to the progression of GWI, however, previous studies among ME/CFS patients have shown that reduced levels of adenosine triphosphate (ATP) likely contribute to the impairment of the extrinsic apoptotic pathway [44]. It is known that testosterone can increase levels of ATP [45], and male GWI subjects have been predicted to exhibit

lower levels of testosterone [46]. This could provide a reasonable explanation for why male GWI subjects did not exhibit enrichment of this pathway.

3.2.5. Negative Regulation of Type II Interferon

Negative regulation of type II interferon production was significantly changed in female GWI subjects compared to males with GWI in response to recovery but did not change in either male or female HC subjects. Genes associated with this pathway were found to be significantly underexpressed (Supplementary Table S3, Figure 4A), which could indicate that the production of type II interferons is increased. This discovery could be of interest in further research into the biomarkers of GWI in female patients. Previous publications have shown that type II interferon levels were significantly increased in GWI patients [47]. Meanwhile, it remains unclear why females with GWI, but not males, experience increased production of type II interferons, highlighting the need for further research.

3.2.6. Immune Dysfunction

Immunoregulatory interactions between a lymphoid and non-lymphoid cell, positive regulation of NK cell-mediated cytotoxicity, lymphocyte activation, and the immune response-activating signaling pathway were found to be similarly enriched at T2 vs T1 (Figure 4A) as discussed in T1 vs T0 (3.1).

3.2.7. Overall

These findings emphasize the significant sex-dependent changes in a variety of biological pathways that could underlie a failure of GWI patients' ability to recover following various stressors. Sex-specific inflammation, oxidative stress, and deregulation of the immune system were detected. In both males and females, compromised cellular repair, differentiation, and immune processes could underlie the chronic nature of GWI. These findings point to the immediate need for personalized diagnostic markers and treatment strategies, accounting for sex-based immune, metabolic, and viral response differences to improve targeted interventions for GWI patients.

4. Materials and Methods

4.1. Cohort

This project was conducted in the Miami/Fort Lauderdale area and included 25 female subjects clinically diagnosed with GWI and 17 female HCs. It also included 19 male GWI subjects and 25 male HCs. All individuals with GWI and HCs were recruited as part of a larger study. This study was conducted under the Declaration of Helsinki. Protocols were approved by the Nova Southeastern University (NSU)'s Institutional Review Board (2016-2-NSU). All subjects provided informed consent. GWI subjects that met the criteria for diagnosis (including Centers for Disease Control and Prevention (CDC) [48] and Kansas criteria for GWI [1]) were included in the study. Subjects were excluded if they were presented with a history of active smoking or alcohol, diabetes, immunodeficiency, cardiovascular disease, stroke, autoimmunity, malignancy, or systemic infection present within two weeks of blood collection. Both cohorts were matched for age and BMI. Female subjects were also required to complete a gynecologic questionnaire to ensure that blood collection occurred during the first two weeks of their menstrual cycle.

SF-36 questionnaires [14] were used to evaluate the physical and mental health of GWI and HC subjects. SF-36 scores ranging from zero to 100 were evaluated where higher scores indicated lower levels of disability. SF-36 questionnaires were used to compare individuals with GWI and HCs including "Physical Health," (which is comprised of eight domains of well-being including physical functioning, physical role functioning, bodily pain, and general health perception) as well as "Mental Health," (which includes vitality, social function, emotional role, and mental health). Each of the eight

listed domains were transformed into a zero-to-100-point scale, with each question being weighed equally. Higher scores on this scale indicated lower levels of disability. The SF-36 questionnaire includes eight scaled scores, which are weighted sums of the questions in their respective section. All scales were directly transformed into a zero to 100 scale on the assumption that each question holds equal weight.

Participants were fed a standardized breakfast (yogurt and banana) and rested for 30 minutes in reclining chairs before blood draws (T0). Following this, a standard maximal graded exercise test (GXT) was conducted according to McArdle's protocol [49]. This protocol was used as a part of a larger study to determine the biological mechanisms underlying neuroimmune diseases. GXTs involved participants pedaling at 60 W for two minutes, with an increase of 30 W every two minutes until maximal exertion was reached. A second blood draw was taken at the point of maximal exertion (T1) and a third blood draw was performed four hours after maximal exertion (T2).

4.2. PBMC Isolation and RNA Extraction

All Participants donated up to 8 ml of whole blood, which was then collected in K2EDTA tubes and diluted in a 1:1 ratio (v/v) in phosphate-buffered saline (PBS) within two hours. The resulting solution was layered with Ficoll-Paque Premium (GE Healthcare, Chicago, IL, USA). From there, samples were subjected to centrifugation at 500 x g for 30 minutes at 20 degrees Celsius without brakes. The PBMC layer was then washed with PBS and resuspended in a freezing medium. Aliquots of 10⁷ cells per mL were frozen in liquid nitrogen until preparations for analysis were completed. Total RNA was extracted using RNeasy (Qiagen, Crawfordsville, IN, USA), and quality was assessed using Agilent TapeStation 4200, with all samples returning an RNA integrity number >7.

4.3. RNA-Seq

300 ng of RNA was submitted to the Genomic Core Facility of NSU and to the Center of Genome Technology (CGT) at the University of Miami for RNA-Seq. TruSeq Stranded Total RNA library Prep Kit (Illumina Inc., San Diego, CA, USA) was used to generate libraries with a paired-end sequencing reading length of 150 base pairs. Illumina RNA-Seq pipeline was employed to determine genomic coverage, alignment percentage, and nucleotide quality for quality control testing. After quality control, raw sequencing data was converted to a fastq format.

4.4. RNA-Seq Analysis

GSNAP (version 2021-02-22) [50], HISAT2 (version 2.1.0) [51], and STAR (version 2.7.8a) [52] software were used to map raw reads to the GRCh38/release 103 reference genome. After alignment, analysis of data produced by each aligner was conducted separately through batch effect mitigation and differential expression. Read counts aligned by GSNAP and HISAT2 were evaluated using HTSeq (version 0.13.5) [53], while STAR read counts were determined using the "--quantMode Transcriptome SAM" option. Following read counts by HTSeq and STAR, genes from sex chromosomes were removed, and ComBat-seq [54] was utilized to adjust false positives generated from batch effects. Gene counts produced by each aligner were imported into Bioconductor/R (version 4.2.1) package DESeq2 (version 1.36.0) [55] for differential gene expression analysis. Genes with counts of 50 or higher in all samples in one or more groups (GWI patients at T0, GWI patients at T1, GWI patients at T2, HC at T0, HC at T1, HC at T2) were used for differential analysis by DESeq2, resulting in a total of 14986 genes for GSNAP, 11794 genes for HISAT2, and 11768 genes for STAR. DEGs were selected based on two criteria: (1) fold change (FC) >1.5 and false discovery rate (FDR) < 0.10 in one out of three aligners, and FC > 1.4 and FDR < 0.15 in the remaining two aligners; or (2) FC > 1.5 and FDR < 0.10 in at least two out of three aligners.

Raw data have been submitted to GEO omnibus with the accession number GSE286345.

After selection, DEGs were uploaded to Metascape [15], where analysis was conducted according to default parameters, with a p-value threshold of <0.01, a minimum count of three, and an enrichment factor of >1.5. Cytoscape was then employed to perform visualization of GO networks.

4.5. Validation of RNA-Seq Results

A custom panel was used on the Nanostring *nCounter* platform to validate RNA-Seq findings. Hybridization and processing of 100 ng total RNA in conjunction with counting was carried out according to manufacturer guidelines. Raw counts were analyzed using NanoString nSolver v.4 software. Geometric means of negative controls plus two standard deviations were computed for all samples. Counts below the threshold value were removed from normalization and analysis. All procedures were executed following the manufacturer's instructions.

5. Conclusions

In conclusion, our results illustrate sex dependent transcriptomic differences in the effects of stressors on male and female GWI patients, which warrants further investigation. This was found to hold in both response to maximal exertion and recovery where the symptoms of GWI are known to be exacerbated [56]. It is important to note that our study findings have limitations. Specifically, larger scale cohorts and reproduced data would allow for a comprehensive examination of transcriptomic profiles and confirm potential biomarkers that could be associated with disease progression under stress. Greater lengths of 48 hours of exposure to exercise challenges could also yield valuable results. Furthermore, the inclusion of further multi omics data in the same participants could provide a more encompassing understanding of molecular mechanisms and pathways involved. As far as we know this is the first and only study on sex differences at a transcriptomic level in GWI veterans. Thus, more studies directly on GWI veterans are needed to provide a valid and comprehensive understanding of GWI pathophysiology

Supplementary Materials: The following supporting information can be downloaded at: Preprints.org, Supplementary Table S1: Differentially expressed genes in GWI females between maximal exertion (T1) and baseline before exercise (T0); Supplementary Table S2: Differentially expressed genes in healthy females between maximal exertion (T1) and baseline before exercise (T0); Supplementary Table S3: Differentially expressed genes in GWI females between 4 hours post maximal exertion (T2) and maximal exertion (T1); Supplementary Table S4: Differentially expressed genes in healthy females between 4 hours post maximal exertion (T2) and maximal exertion (T1); Supplementary Table S5: NanoString validation of RNA-seq results in (a) GWI females between maximal exertion (T1) and baseline before exercise (T0), (b) healthy females between maximal exertion (T1) and baseline before exercise (T0), (c) GWI females between 4 hours post maximal exertion (T2) and maximal exertion (T1), (d) healthy females between 4 hours post maximal exertion (T2) and maximal exertion (T1) time points.

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Data Availability Statement: Raw data have been submitted to GEO omnibus with the accession number GSE286345 and is available to reviewers with the token "evmjucskrbobfw".

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Abbreviations

The following abbreviations are used in this manuscript:

ATP	Adenosine Triphosphate
BMI	Body Mass Index
CDC	Centers for Disease Control and Prevention
CGT	Center of Genome Technology
CDMRP	Congressionally Directed Medical Research Programs
DEG	Differentially Expressed Genes
FC	Fold Change
FDR	False Discovery Rate
GEO	Gene Expression Omnibus
GO	Gene Ontology
GRECC	Geriatric Research Education and Clinical Center
GSE	Gene Series Expression
GWI	Gulf War Illness
GXT	Graded Exercise Test
HC	Healthy Control
HCMV	Human Cytomegalovirus
HHV-5	Human Herpes Virus 5
HISAT2	Hierarchical Indexing for Spliced Alignment of Transcripts 2
IFN- γ	Interferon Gamma
IL-10	Interleukin-10
IL-12	Interleukin-12
ILC	Innate Lymphoid Cell
KEGG	Kyoto Encyclopedia of Genes and Genomes
MDA	Malondialdehyde

References

1. White RF, Steele L, O'Callaghan JP, Sullivan K, Binns JH, Golomb BA, et al. Recent research on Gulf War illness and other health problems in veterans of the 1991 Gulf War: Effects of toxicant exposures during deployment. *Cortex*. 2016;74:449-75.
2. Steele L. Prevalence and patterns of Gulf War illness in Kansas veterans: association of symptoms with characteristics of person, place, and time of military service. *Am J Epidemiol*. 2000;152(10):992-1002.

3. Kerr KJ. Gulf War illness: an overview of events, most prevalent health outcomes, exposures, and clues as to pathogenesis. *Rev Environ Health*. 2015;30(4):273-86.
4. Ramirez-Sanchez I, Navarrete-Yanez V, Garate-Carrillo A, Loredó M, Lira-Romero E, Estrada-Mena J, et al. Development of muscle atrophy and loss of function in a Gulf-War illness model: underlying mechanisms. *Sci Rep*. 2020;10(1):14526.
5. Haley RW, Vongpatanasin W, Wolfe GI, Bryan WW, Armitage R, Hoffmann RF, et al. Blunted circadian variation in autonomic regulation of sinus node function in veterans with Gulf War syndrome. *Am J Med*. 2004;117(7):469-78.
6. Whistler T, Fletcher MA, Lonergan W, Zeng XR, Lin JM, Laperriere A, et al. Impaired immune function in Gulf War Illness. *BMC Med Genomics*. 2009;2:12.
7. Trivedi MS, Abreu MM, Sarria L, Rose N, Ahmed N, Beljanski V, et al. Alterations in DNA Methylation Status Associated with Gulf War Illness. *DNA Cell Biol*. 2019;38(6):561-71.
8. Ashbrook DG, Hing B, Michalovicz LT, Kelly KA, Miller JV, de Vega WC, et al. Epigenetic impacts of stress priming of the neuroinflammatory response to sarin surrogate in mice: a model of Gulf War illness. *J Neuroinflammation*. 2018;15(1):86.
9. Mozhui K, O'Callaghan JP, Ashbrook DG, Prins P, Zhao W, Lu L, et al. Epigenetic analysis in a murine genetic model of Gulf War illness. *Front Toxicol*. 2023;5:1162749.
10. Nugent SM, Freeman M, Ayers CK, Winchell KA, Press AM, O'Neil ME, et al. A Systematic Review of Therapeutic Interventions and Management Strategies for Gulf War Illness. *Mil Med*. 2021;186(1-2):e169-e78.
11. Dickey B, Madhu LN, Shetty AK. Gulf War Illness: Mechanisms Underlying Brain Dysfunction and Promising Therapeutic Strategies. *Pharmacol Ther*. 2021;220:107716.
12. Chester JE, Rowneki M, Van Doren W, Helmer DA. Progression of intervention-focused research for Gulf War illness. *Mil Med Res*. 2019;6(1):31.
13. Van Booven D, Zarnowski O, Perez M, Sarria L, Collado F, Hansotia K, et al. The effect of stress on the transcriptomes of circulating immune cells in patients with Gulf War Illness. *Life Sci*. 2021;281:119719.
14. Ware JE, Jr., Sherbourne CD. The MOS 36-item short-form health survey (SF-36). I. Conceptual framework and item selection. *Med Care*. 1992;30(6):473-83.
15. Zhou Y, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, et al. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. *Nat Commun*. 2019;10(1):1523.
16. Shetty GA, Hattiangady B, Upadhyaya D, Bates A, Attaluri S, Shuai B, et al. Chronic Oxidative Stress, Mitochondrial Dysfunction, Nrf2 Activation and Inflammation in the Hippocampus Accompany Heightened Systemic Inflammation and Oxidative Stress in an Animal Model of Gulf War Illness. *Front Mol Neurosci*. 2017;10:182.
17. Cohen J, Mathew A, Dourvetakis KD, Sanchez-Guerrero E, Pangen RP, Gurusamy N, et al. Recent Research Trends in Neuroinflammatory and Neurodegenerative Disorders. *Cells*. 2024;13(6).
18. Maksoud R, Magawa C, Eaton-Fitch N, Thapaliya K, Marshall-Gradisnik S. Biomarkers for myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS): a systematic review. *BMC Med*. 2023;21(1):189.
19. Montoya JG, Holmes TH, Anderson JN, Maecker HT, Rosenberg-Hasson Y, Valencia IJ, et al. Cytokine signature associated with disease severity in chronic fatigue syndrome patients. *Proc Natl Acad Sci U S A*. 2017;114(34):E7150-E8.
20. Yang T, Yang Y, Wang D, Li C, Qu Y, Guo J, et al. The clinical value of cytokines in chronic fatigue syndrome. *J Transl Med*. 2019;17(1):213.
21. Wood E, Hall KH, Tate W. Role of mitochondria, oxidative stress and the response to antioxidants in myalgic encephalomyelitis/chronic fatigue syndrome: A possible approach to SARS-CoV-2 'long-haulers'? *Chronic Dis Transl Med*. 2021;7(1):14-26.
22. Elhaj R, Reynolds JA-O. Chemical exposures and suspected impact on Gulf War Veterans. (2054-9369 (Electronic)).
23. Pearson JN, Patel M. The role of oxidative stress in organophosphate and nerve agent toxicity. (1749-6632 (Electronic)).

24. Coughlin SS. A Neuroimmune Model of Gulf War Illness. LID - 10.15436/2378-6841.17.1665 [doi]. (2378-6841 (Print)).
25. Trageser KJ, Sebastian-Valverde M, Naughton SX, Pasinetti GM. The Innate Immune System and Inflammatory Priming: Potential Mechanistic Factors in Mood Disorders and Gulf War Illness. (1664-0640 (Print)).
26. Lande A, Fluge Ø, Strand EB, Flåm ST, Sosa DD, Mella O, et al. Human Leukocyte Antigen alleles associated with Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/CFS). *Scientific Reports*. 2020;10(1):5267.
27. Maksoud R, Magawa C, Eaton-Fitch N, Thapaliya K, Marshall-Gradisnik S. Biomarkers for myalgic encephalomyelitis/chronic fatigue syndrome (ME/CFS): a systematic review. *BMC Medicine*. 2023;21(1):189.
28. Gabrysová L, Nicolson Ks Fau - Streeter HB, Streeter Hb Fau - Verhagen J, Verhagen J Fau - Sabatos-Peyton CA, Sabatos-Peyton Ca Fau - Morgan DJ, Morgan Dj Fau - Wraith DC, et al. Negative feedback control of the autoimmune response through antigen-induced differentiation of IL-10-secreting Th1 cells. (1540-9538 (Electronic)).
29. Sun L, He C, Nair L, Yeung J, Egwuagu CE. Interleukin 12 (IL-12) family cytokines: Role in immune pathogenesis and treatment of CNS autoimmune disease. (1096-0023 (Electronic)).
30. Liu J Fau - Cao S, Cao S Fau - Kim S, Kim S Fau - Chung EY, Chung Ey Fau - Homma Y, Homma Y Fau - Guan X, Guan X Fau - Jimenez V, et al. Interleukin-12: an update on its immunological activities, signaling and regulation of gene expression. (1573-3955 (Print)).
31. Zwirner NW, Ziblat A. Regulation of NK Cell Activation and Effector Functions by the IL-12 Family of Cytokines: The Case of IL-27. *Frontiers in Immunology*. 2017;8.
32. Yoo JK, Cho Jh Fau - Lee SW, Lee Sw Fau - Sung YC, Sung YC. IL-12 provides proliferation and survival signals to murine CD4+ T cells through phosphatidylinositol 3-kinase/Akt signaling pathway. (0022-1767 (Print)).
33. Gamer J, Van Booven DJ, Zarnowski O, Arango S, Elias M, Kurian A, et al. Sex-Dependent Transcriptional Changes in Response to Stress in Patients with Myalgic Encephalomyelitis/Chronic Fatigue Syndrome: A Pilot Project. *Int J Mol Sci*. 2023;24(12).
34. Racciatti D, Vecchiet J, Ceccomancini A, Ricci F, Pizzigallo E. Chronic fatigue syndrome following a toxic exposure. *Science of The Total Environment*. 2001;270(1):27-31.
35. Zhang H, Tang Y, Tao J. Sex-Related Overactivation of NLRP3 Inflammasome Increases Lethality of the Male COVID-19 Patients. *Frontiers in Molecular Biosciences*. 2021;8.
36. Ohs I, van den Broek M, Nussbaum K, Münz C, Arnold SJ, Quezada SA, et al. Interleukin-12 bypasses common gamma-chain signalling in emergency natural killer cell lymphopoiesis. *Nature Communications*. 2016;7(1):13708.
37. Morris G, Anderson G, Maes M. Hypothalamic-Pituitary-Adrenal Hypofunction in Myalgic Encephalomyelitis (ME)/Chronic Fatigue Syndrome (CFS) as a Consequence of Activated Immune-Inflammatory and Oxidative and Nitrosative Pathways. (1559-1182 (Electronic)).
38. Halpin P, Williams MV, Klimas NG, Fletcher MA, Barnes Z, Ariza MA-O. Myalgic encephalomyelitis/chronic fatigue syndrome and gulf war illness patients exhibit increased humoral responses to the herpesviruses-encoded dUTPase: Implications in disease pathophysiology. (1096-9071 (Electronic)).
39. Annesley SJ, Missailidis D, Heng B, Josev EK, Armstrong CW. Unravelling shared mechanisms: insights from recent ME/CFS research to illuminate long COVID pathologies. *Trends in Molecular Medicine*. 2024;30(5):443-58.
40. He Y, Liu Y, Zhang M. The beneficial effects of curcumin on aging and age-related diseases: from oxidative stress to antioxidant mechanisms, brain health and apoptosis. (1663-4365 (Print)).
41. Nousis L, Kanavaros P, Barbouti AA-OX. Oxidative Stress-Induced Cellular Senescence: Is Labile Iron the Connecting Link? LID - 10.3390/antiox12061250 [doi] LID - 1250. (2076-3921 (Print)).
42. Tan BL, Norhaizan ME, Liew WP, Sulaiman Rahman H. Antioxidant and Oxidative Stress: A Mutual Interplay in Age-Related Diseases. (1663-9812 (Print)).

43. Bhatia A, Sekhon HK, Kaur G. Sex hormones and immune dimorphism. (1537-744X (Electronic)).
44. Morris G, Berk M, Galecki P, Maes M. The Emerging Role of Autoimmunity in Myalgic Encephalomyelitis/Chronic Fatigue Syndrome (ME/cfs). *Molecular Neurobiology*. 2014;49(2):741-56.
45. Zhang T, Wang Y, Kang Y, Wang L, Zhao H, Ji X, et al. Testosterone enhances mitochondrial complex V function in the substantia nigra of aged male rats. (1945-4589 (Electronic)).
46. Craddock TJ, Fritsch P, Rice MA, Jr., del Rosario RM, Miller DB, Fletcher MA, et al. A role for homeostatic drive in the perpetuation of complex chronic illness: Gulf War Illness and chronic fatigue syndrome. *PLoS One*. 2014;9(1):e84839.
47. Broderick G, Kreitz A Fau - Fuite J, Fuite J Fau - Fletcher MA, Fletcher Ma Fau - Vernon SD, Vernon Sd Fau - Klimas N, Klimas N. A pilot study of immune network remodeling under challenge in Gulf War Illness. (1090-2139 (Electronic)).
48. Maule AL, Janulewicz PA, Sullivan KA, Kregel MH, Yee MK, McClean M, et al. Meta-analysis of self-reported health symptoms in 1990-1991 Gulf War and Gulf War-era veterans. *BMJ Open*. 2018;8(2):e016086.
49. McArdle WD, Katch FI, Katch VL. *Exercise Physiology: Nutrition, Energy, and Human Performance*. Philadelphia: Lippincott Williams & Wilkins; 2001.
50. Wu TD, Nacu S. Fast and SNP-tolerant detection of complex variants and splicing in short reads. *Bioinformatics*. 2010;26(7):873-81.
51. Kim D, Langmead B, Salzberg SL. HISAT: a fast spliced aligner with low memory requirements. *Nat Methods*. 2015;12(4):357-60.
52. Dobin A, Gingeras TR. Mapping RNA-seq Reads with STAR. *Curr Protoc Bioinformatics*. 2015;51:11 4 1- 4 9.
53. Anders S, Pyl PT, Huber W. HTSeq--a Python framework to work with high-throughput sequencing data. *Bioinformatics*. 2015;31(2):166-9.
54. Zhang Y, Parmigiani G, Johnson WE. ComBat-seq: batch effect adjustment for RNA-seq count data. *NAR Genom Bioinform*. 2020;2(3):lqaa078.
55. Love MI, Huber W, Anders S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol*. 2014;15(12):550.
56. Washington SD, Rayhan RU, Garner R, Provenzano D, Zajur K, Addiego FM, et al. Exercise alters cerebellar and cortical activity related to working memory in phenotypes of Gulf War Illness. (2632-1297 (Electronic)).

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