

Review

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Keywords: Cancer; phytomolecules; fused oxygen-based heterocycles; anticancer; flavonoids; coumarins



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Review

# Emerging Oxygen Based Heterocyclic Scaffolds as Potential Anticancer Candidates

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**Abstract:** Oxygen based heterocyclic moieties hold an ample range of therapeutic activities. Heterocyclic molecules are nominated as vital components of an extensive array of structural motifs with both biological and pharmaceutical significance. The oxygen-based scaffolds act as anticancer candidates and are also present in numerous phytochemicals viz; irinotecan, camptothecin, topotecan, taxol, taxotere, podophylotoxin, etoposide, daunorubicin and teniposide. The architectural design of numerous structural motifs for amelioration of cancer has become progressively amplified in recent past years. Until now presently there is no strategic treatment which is so capable that can cure cancer from its roots. Henceforth, it is very indispensable to design novel anticancer structural motifs with least side effects. The oxygen containing heterocyclic scaffolds includes flavonoids, pyrans, xanthenes and coumarins are of utmost importance in medical chemistry for the mitigation of cancer. This assemblage offers several recent developments as anticancer oxygen containing heterocyclic molecules all-round the globe and attracted the structural motifs of auspicious molecules, along with their mechanistic insights, IC<sub>50</sub> values, structure–activity relationships, and molecular docking studies. The encouraging properties discovered by these oxygen-based scaffolds unconditionally engaging them at frontline in invention of potential drug candidates. Consequently, these probably will be of amazing attention to scientists working on the design and synthesis of antitumor candidates.

**Keywords:** cancer; phytochemicals; fused oxygen-based heterocycles; anticancer; flavonoids; coumarins

## 1. Introduction

Natural Products are the gold standard excellent prosperity of nature. Natural phytochemicals play a chief role in the alleviation of ample range of ailments and diseases viz; cancer [1,2] antioxidant, wound healing [3–5], microbial infections, antiviral, HIV [6,7], inflammation, liver, and respiratory disorders [8,9]. The therapeutic use of herbal remedies is owing to the presence of phytochemicals such as flavonoids, alkaloids, tannins, glycosides, triterpenoids, and phenolic molecules [10,11].

Various tribal societies have been consuming herbal products since ancient times for the mitigation of cancer. To authenticate the traditional use of natural plants and their products, researchers from the globe were working diligently to establish the role of plants and their phytomolecules in the management of cancer because of their fewer side effects, economical, ease of accessibility are the topmost benefits of the natural products [12,13]. The hunt for phytomolecules in medicine, especially in the field of anticancer agents from the prehistoric periods. But the meticulous era for such type of research is momentarily recent. Traditionally, plants-based products and plant extracts were the roots of numerous medicinal agents from which the budding lead molecules were designed to offer birth of potential therapeutic candidates. Several numbers of natural plants have been employed for the amelioration of cancer, few are in clinical use and certain in clinical development.

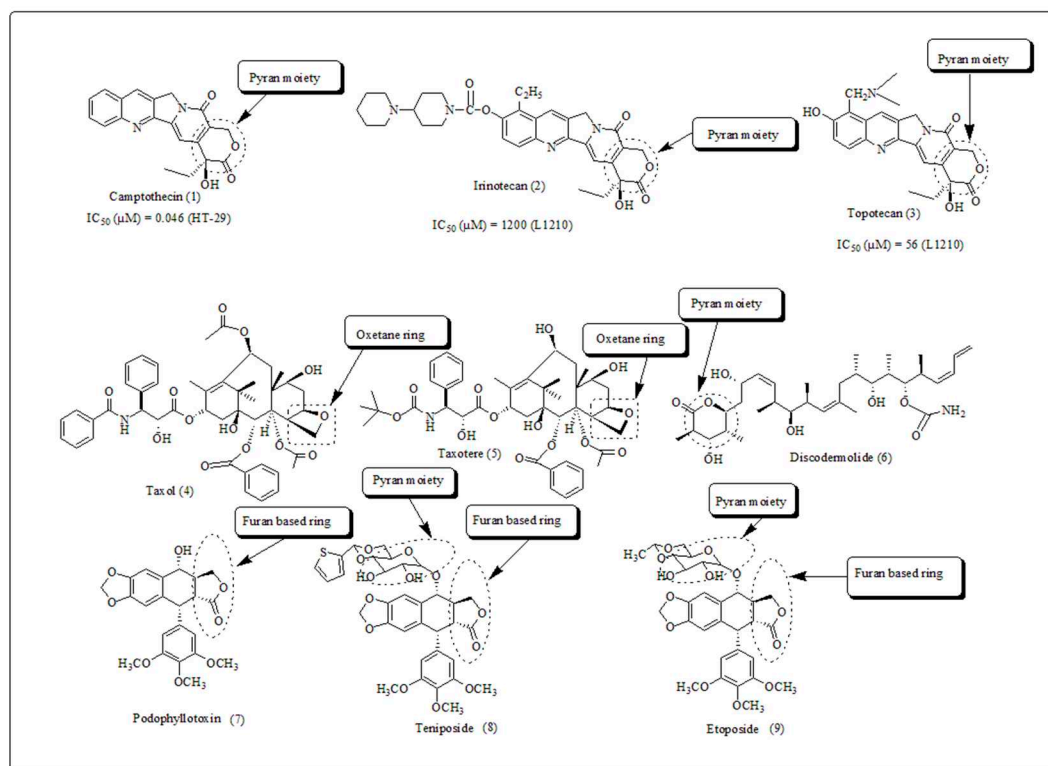
Cancer is an assemblage of diseases concerning abnormal cell growth with the uninterrupted spread to the other parts of the body. This illness is triggered by speckled agents such as, chemical compound and radiant energy [14–17]. There are numerous drugs which are used for management of this disease either by killing cancer cells or modify their progression. Oxygen containing heterocyclic scaffolds are regularly being used for the architecting of new chemical entities. Many of them are clinically approved drugs contain the oxygen based heterocyclic ring and acting as anti-cancer agents such as daunorubicin, irinotecan, camptothecin, topotecan, taxol, taxotere, podophylotoxin, etoposide and teniposide etc [18,19]. Introduction of oxygen heteroatoms enhances polarity, solubility, and hydrogen bonding capabilities leading to ADMET optimization for drug-gable physiognomies [20]. Heterocyclic molecules are nominated as vital components on an extensive array of structural motifs with both biological and therapeutic significance. The oxygen-based heterocycles are of utmost importance as these found as a great cohort of structures thru gigantic reputation in medicinal chemistry.

This assemblage offers several recent developments as anticancer oxygen containing heterocyclic molecules all-round the globe and attracted the structural motifs of auspicious molecules, along with their mechanistic insights,  $IC_{50}$  values, structure–activity relationships, molecular docking studies and interesting key findings of biological activities in the mitigation of several human cancer cell lines. The encouraging properties discovered by these oxygen-based scaffolds unconditionally engaging them at frontline for the invention of potential drug candidates. Consequently, these possibly will be of amazing attention to scientists facilitating them to embrace a most demanding and speedy target focused on drug discovery process as antitumor candidates.

In this era most commonly used antitumor drugs are analogs of phytomolecules which contains oxygen-based heterocycles such as oxitane, oxirane, furan, pyran, chromone and xanthone. Several booming phytomolecules includes camptothecin, irinotecan, topotecan, taxol, taxotere, podophylotoxin, teniposide and etoposide (Figure 1) have been emerged as drugs candidates after structural modification on the leads of natural origin [21–28].

Camptothecin (CPT) is an alkaloid isolated [29] from the stem wood of *Camptotheca acuminata* also documented as the ‘tree of love’ or ‘tree of joy’. It has also been obtained from *Mapia foetida* and *Ophiorrhiza pumila*. The anticancer potential of plants has been discovered using in vitro studies and in mouse leukemia cells. The results revealed in creation as a prospective antitumor agent. Currently, the foremost discovery leads to CPT scaffolds, such as irinotecan and topotecan are used in treatment of colon cancers and ovarian cancers [30,31].

Taxol (paclitaxel) and Taxotere (docetaxel) shown in (Figure 1) binds at taxane binding site. Paclitaxel was initially obtained from the bark of *Taxus brevifolia* which is famously known as the pacific yew tree. Docetaxel, isolated from *Taxus baccata* recognized as a European yew tree, is a semi-synthetic analog of Taxol and is clinically employed in the treatment of prostate, breast, non small cell lung and ovarian cancer. At low doses taxanes cause mitotic arrest and at high doses they act as microtubule stabilizing agents [32,33]. Several researchers have tried to synthesize its analogs and only a few of them were found to be active as compared to taxol. Georg *et al.* synthesized novel taxol analogs and demonstrated cytotoxicity towards B16 melanoma cells like taxol [34,35].



**Figure 1.** Oxygen-based heterocyclic phytomolecules with anticancer potential.

Discodermolide is isolated from the marine Caribbean deep sea sponge *Discodermia dissolute* [36]. It was primarily exhibited antifungal and immunosuppressive properties, but later on found to be a microtubule stabilizing candidate [37,38]. Discodermolide exhibiting anticancer potential against a diverse cell lines and found to be more potent than taxol ( $EC_{50}$  value of  $2\ \mu\text{M}$  [39]). Discodermolide is active against taxol resistant cell lines [39]. It can inhibit cell mitosis thru stimulating tubulin polymerization and thus persuade microtubule bundles in cells using in vitro studies [40,41].

Podophyllotoxin (PDT), a bioactive lignin, documented as May apple or American mandrake, obtained from *Podophyllotoxin peltatum* [25]. Podophyllotoxin has also been isolated from other species *P. emodi* and *P. pleianthum*. The semisynthetic derivatives of Podophyllotoxin, Teniposide and Etoposide are now widely recommended in the mitigation of numerous cancers [26]. Over the years, several derivatives of podophyllotoxin, have been synthesized and evaluated for cytotoxicity against A-549 and P-388 human cancer cell lines [27]. Zhang *et al* synthesized a new series of podophyllotoxin analogs and screened against human malignant cell lines such as RPMI-8226, HL-60 and A-549 [28].

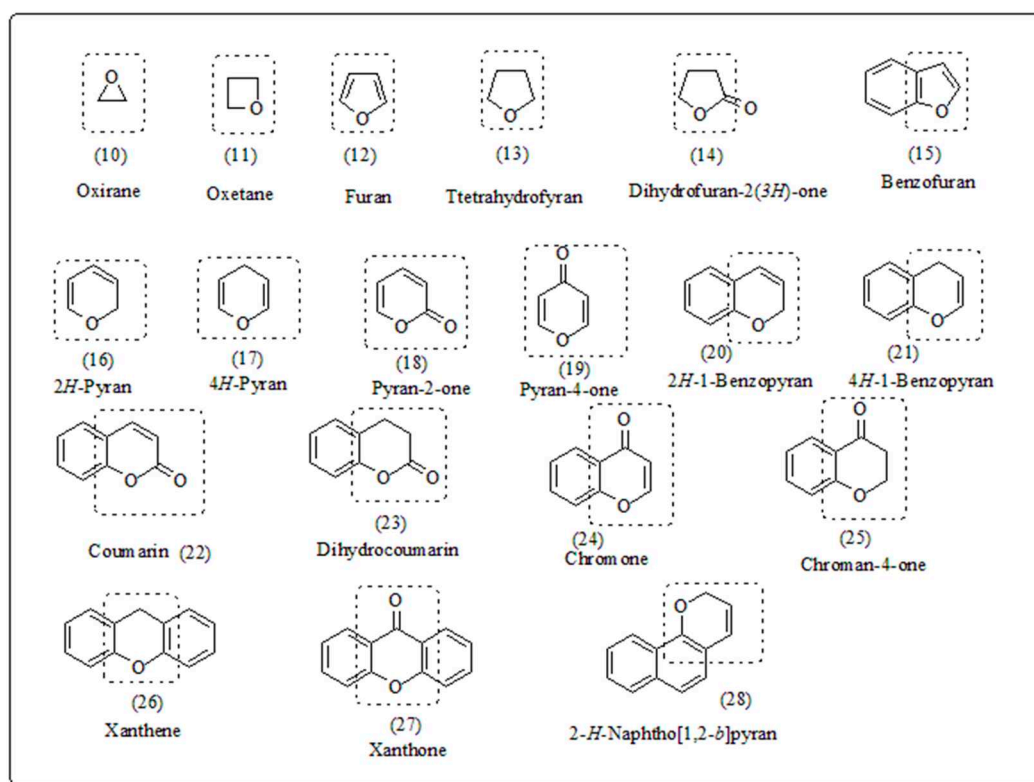
This article also presents some recent advancement in the field of anticancer agents around the globe. The focus is on the structure-activity relationship in addition to the structure of the most promising molecules along with  $IC_{50}$  values against diverse human tumor cell lines and some interesting key findings. For presenting the different types of heterocycles and fused heterocycles, we have basically categorized these based on one of the hub functionalities of their chemical architecture. It also attempts to provide a comprehensive summary on the chemistry and structure of the most promising agent as anticancer agent along with their  $IC_{50}$  values and structure activity relationship. To the paramount of our understanding, this is the first comprehensive review of recent advancements in the domain of anticancer heterocycles which includes literature published during the previous years. We believe that the present review will provide medicinal and bioorganic chemists collective insights into the biological profiles of heterocycles having anti-proliferative activity, thus helping them adopt a more focused and speedier target-oriented drug discovery process. An in-depth summary of the structure-activity relationships and mechanistic insights exposed throughout the pharmacological screening of the potential oxygen containing heterocyclic



compounds. The structures of the synthesized compounds discussed in this assemblage clearly highlight the promising and interesting antitumor profiles, structure of potent molecules, therapeutic actions, important key finding of molecular docking and mechanistic studies. For representing the antitumor potential of the oxygen-based heterocycles, we have categorized the significance of oxygen based heterocyclic structural moiety of other significant heterocycles such as pyran, furan, oxirane, oxitane and fused oxygen-based heterocycles. This organization is established on one of the central functionalities of their chemical structural architecture.

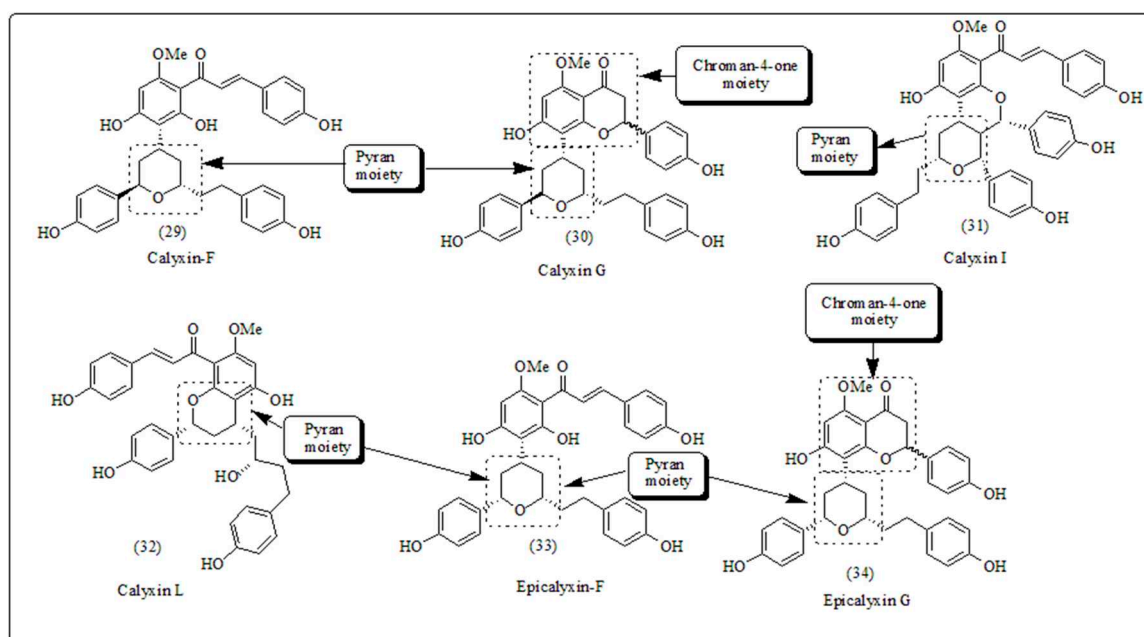
## 2. Pyran based oxygen containing heterocyclic scaffolds

The oxygen heteroatom is the building component of pyran, chromone, benzopyran, flavanoids, xanthenes, coumarin, naphthoquinones, furan, benzofuran, oxirane and oxitane which display assorted therapeutic potential. Several natural compounds comprise pyrans and benzopyrans, exhibiting interesting therapeutic activities, have encouraged the researchers to design and synthesize novel scaffolds. Numerous pyran based heterocycles have been presented in (Figure 2) [42].



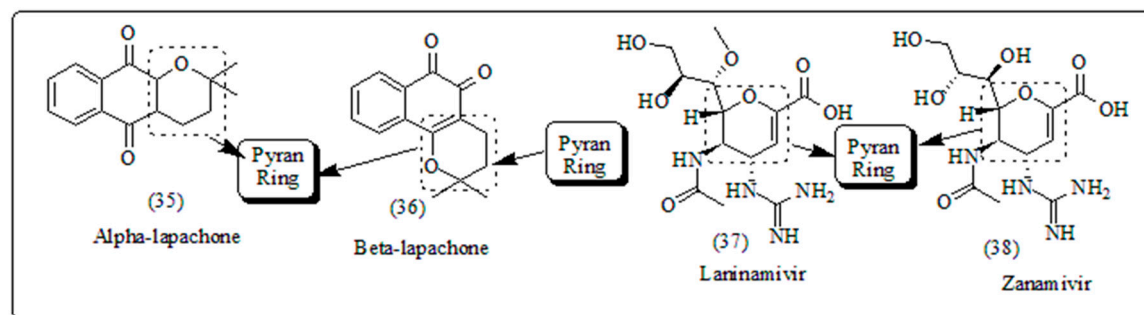
**Figure 2.** Structures of oxygen-based heterocycles.

It is well established that oxygen based heterocyclic compounds are chief building blocks of biologically active phytochemicals [43–45]. Oxygen based heterocyclic skeletons are vital structural components found extensively in natural compounds like sugars, flavonoids [46], anthraquinones [47] and coumarins [48]. Oxygen containing flavonoid-based pyran scaffolds (Figure 3), including calyxin F, calyxin G, calyxin I, calyxin L, epicalyxin G and epicalyxin F, obtained from seeds of *Alpinia blepharocalyx*. Epicalyxin F is a significant compound of this series, as an antitumor agent towards murine 26-L5 carcinoma and HT-1080 fibrosarcoma [49].



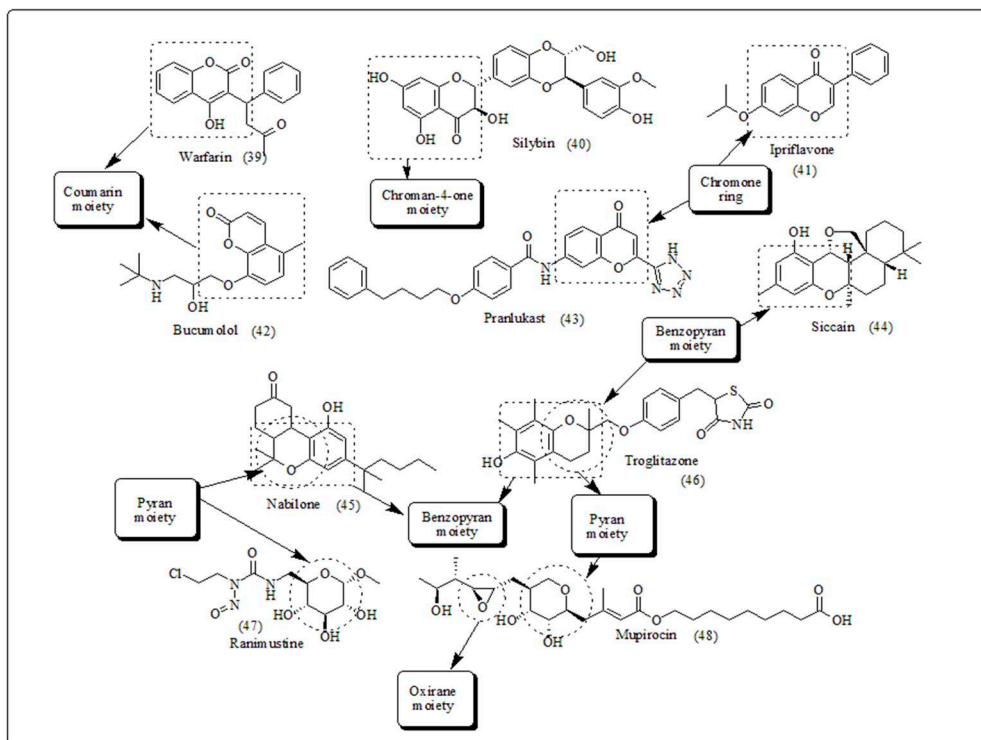
**Figure 3.** Pyran-based compounds isolated from plant species with cell killing potential.

The  $\beta$ -lapachone depicted in (Figure 4), is a pyran derivative, which displayed the ample range of therapeutic potential (including anti-inflammatory, anticancer and antibacterial) and thus having a significant role in drug development. Laninamivir presented in (Figure 4), administered by oral route and is structurally similar to Zanamivir which is prodrug of Laninamivir octanoate screened for its antiviral and anticancer potential. Zanamivir (Figure 4), was permitted for cure of influenza A and B virus. The drug is marketed by GlaxoSmithKline with the trade name of “Relenza” [50,51].



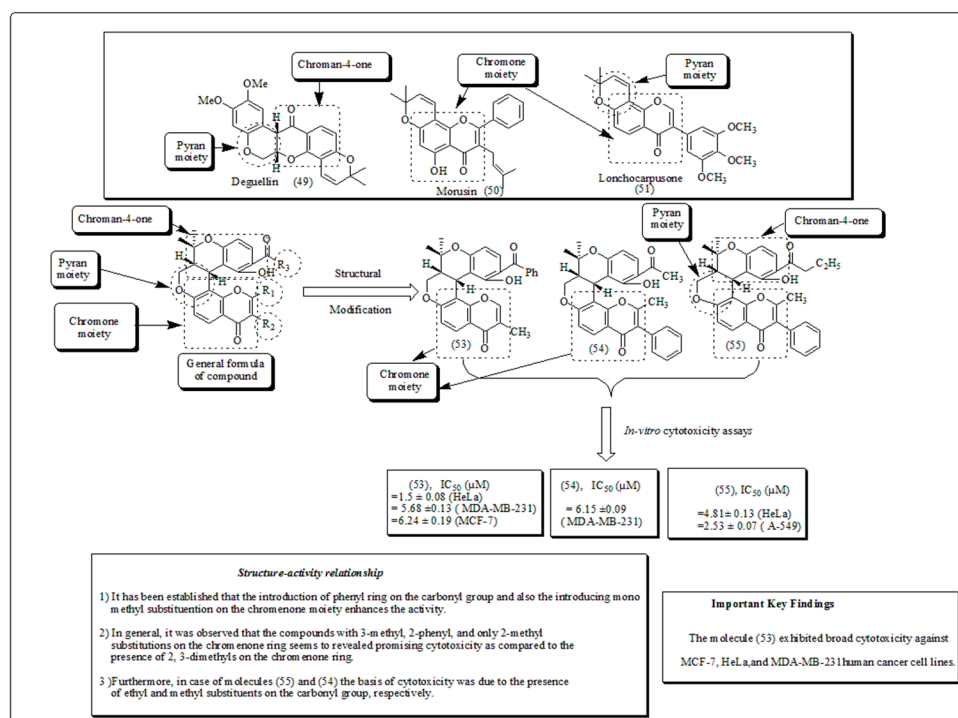
**Figure 4.** Pyran-based marketed drugs in preclinical/clinical phase.

Literature reports have revealed the richness of commercially existing therapeutic candidates containing pyran heterocycle. Oxygen based heterocyclic compounds (Figure 5) which having a chief part of structural motif in numerous synthetic natural and compounds, owning potential therapeutic profiles, due to their extensive range of pharmacological activities such as antituberculosis [52], anticancer [53,54], anti- human immunodeficiency virus (HIV) [55], antifungal [56], antimicrobial [57], calcium channel antagonist activity [58], antidiabetic, [59] and antiviral activities [60].



**Figure 5.** Structure of compounds entrenched with oxygen-based heterocycles.

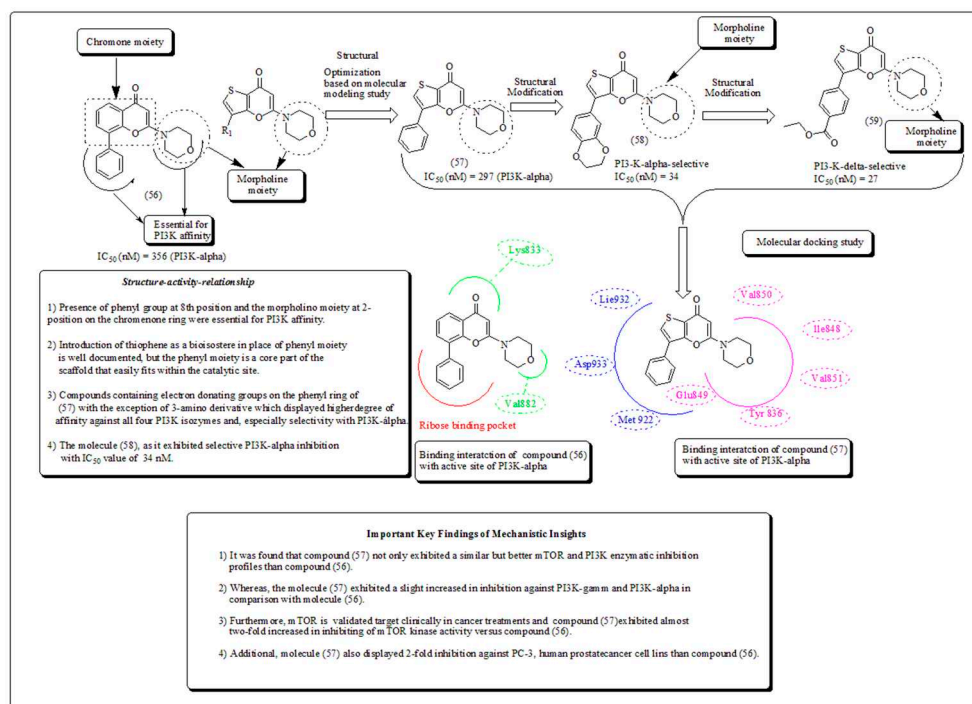
Madda *et al.* established a new class of oxygen-based pyran analogs, and screened using *in vitro* assays towards diverse human malignant cells. The outcomes revealed that compounds displayed significant cytotoxicity against HeLa cells, human cervical malignant cells (Figure 6) [61].



**Figure 6.** Structures of compounds along with their SAR reported by Madda *et al* [61].

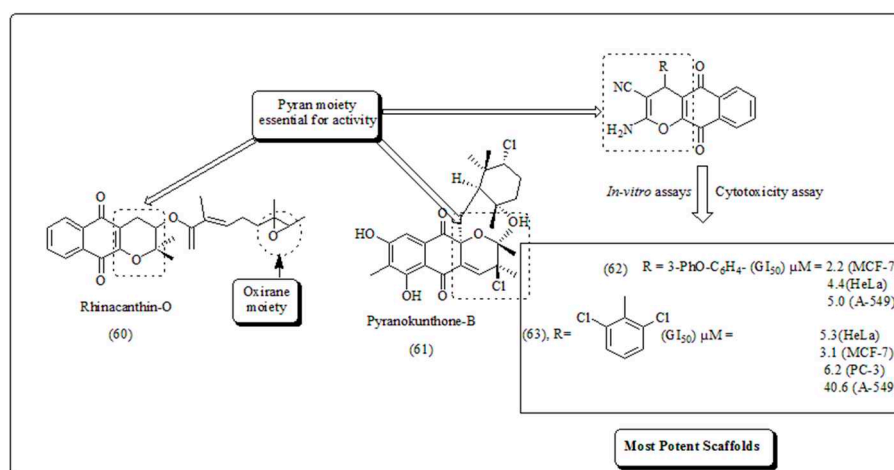
Compound (53) demonstrated marked inhibitory action in both breast cancer cell lines, MCF-7 and MDA-MB-231. Moreover, compound (54) presented cytotoxicity only against MDA-MB-231,

whereas compound (55) confirmed encouraging effects against A549, human lung cancer cell line, with  $IC_{50}$  of 2.53  $\mu$ M [61]. In another study, Morales *et al.* synthesized and screened morpholine based pyran motifs as potential PI3K inhibitors. Compounds presented in (Figure 7) which showed a remarked mTOR and PI3K enzymatic inhibition properties. The compound also exhibited aqueous solubility, and better activity when screened against PC-3 cells, while down-regulating the PI3K pathway as displayed through restraining pAKT-S473 expressions [62].



**Figure 7.** Structure of compound along with mechanistic insights and docking studies established by Morales *et al.* [62].

Naturally occurring pyranonaphthoquinones and their derived analogs, with capable antitumor potentials. Rhinacanthin O (Figure 8) isolated from *Rhinocanthus nasutus*, an Asian medicinal plant and its structure is naphthoquinone based compound consisting of oxirane and pyran heterocyclic compound, pyranokunthone B presented in (Figure 8) obtained from marine actinomycetes [63]. The compounds depicted in (Figure 8) have been examined for the mitigation of tumors connected with heaved NADH quinone oxidoreductase expressions.



**Figure 8.** Structure of oxirane containing compound with anticancer capability.



Frolova *et al.* synthesized and screened a series of compounds, exhibiting anti-proliferative activity and apoptotic potential against a panel of malignant cells [64]. Selected analogues displayed promising results against human cancer. The compound (62) and (63), exhibited anti-proliferative effects superior than  $\alpha$ -lapachone [64].

Natural dihydropyranonaphthoquinones can be obtained from fungi, bacteria, and higher plants. Numerous scaffolds have certainly been found to hold diverse and noticeable biological actions, including antiparasitic, antimicrobial, anticancer and antiviral properties [65]. Eleutherin (64) pentalongin (66) and psychorubrin (65) are of compounds, which exhibited an interesting antimicrobial, phytotoxic antiparasitic, and antineoplastic properties. A new class of compounds (67-70) and their analogs have been screened towards human cancer cell lines by Thi *et al.* The results revealed that compound (67) exhibited  $IC_{50}$  value of 3.6  $\mu$ M and 1.5  $\mu$ M against Hep-G2 and KB malignant cells (Figure 9) [65].

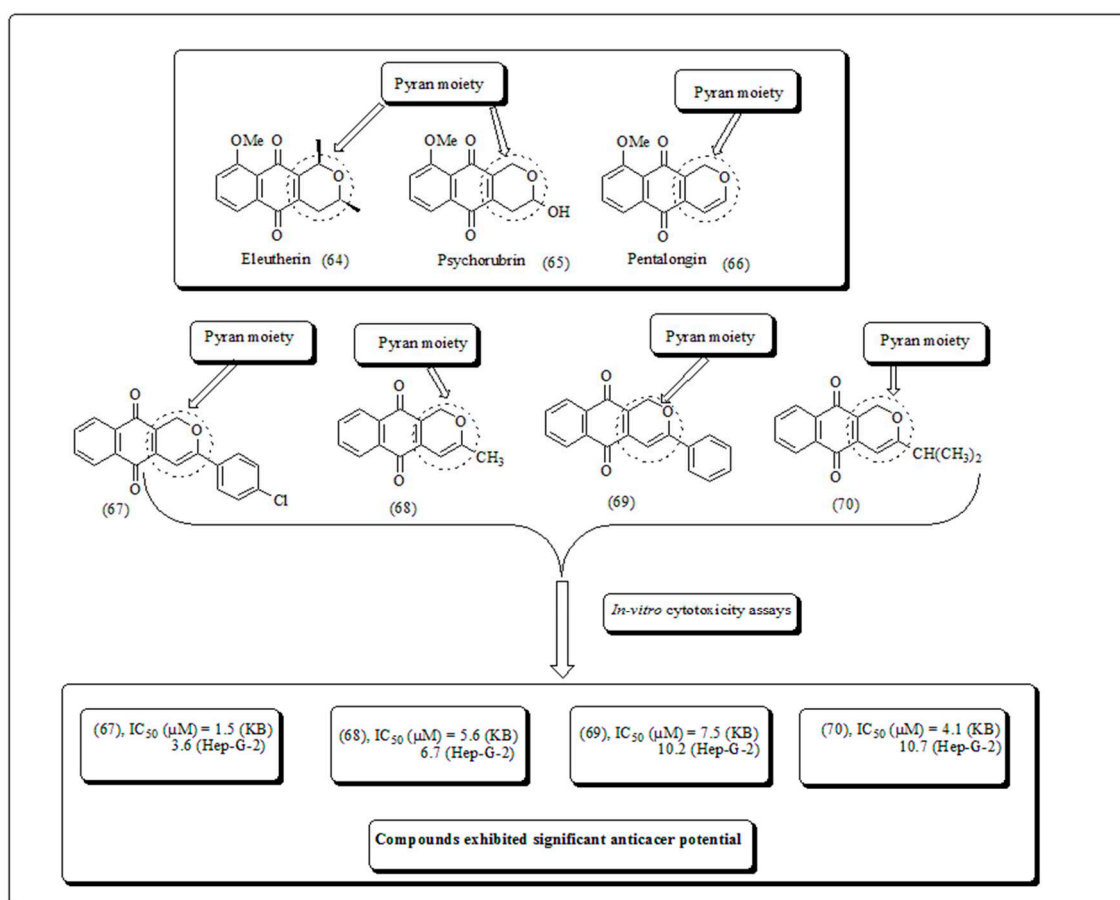


Figure 9. Bioactive natural and synthetic molecules with cell killing potential.

### 1. Flavone-based oxygen containing heterocyclic scaffolds

Flavonoids are plant-based poly phenolic compounds and are termed as secondary metabolites. Flavonoids are precursors of chalcones and present in numerous foods [66]. These are often described as their structures having (C<sub>6</sub>–C<sub>3</sub>–C<sub>6</sub>) phenylbenzopyrone linkage. Flavonoids are categorized into such as flavones, flavanones, isoflavones, flavanols and flavanonols (Figure 10) [67]. Flavonoids exhibit an extensive range of pharmacological actions [68–70] including anti-mutagenic, antioxidant, and anti-proliferative activities. The antioxidants flavonoids are generally involved in angiogenesis, cell signaling and cell cycle regulation [71–74].

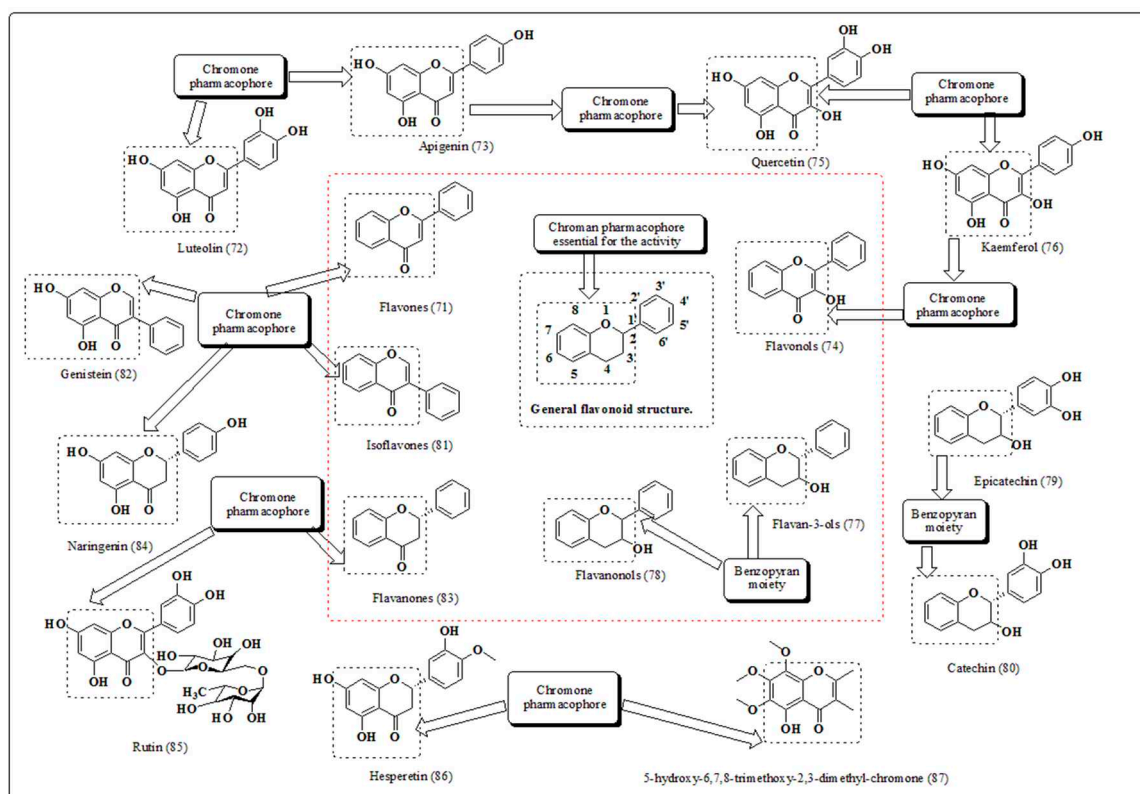


Figure 10. Structures of oxygen-based promising flavonoids.

Hesperitin and Naringenin belongs to flavanones, quercetin and kaempferol are flavonol derivative, luteolin and apigenin belongs to flavone, whereas, genistein sub classified into isoflavone and thought to be talented in the discovery of new lead chemotherapeutic agent for mitigation of cancer. Few years ago, Hsiao *et al.* recognized that flavanone and 2'-hydroxyflavanone decreased cell growth of A549, HA22T, and SK-Hep1 cancer cells, whereas other flavanones (4'-hydroxy flavanone and 6-hydroxy flavanone) displayed little or almost no inhibition [75]. Furthermore, Choi *et al.* reported that 4',7-dimethoxyflavanone, exhibited convincing anti-proliferative actions apoptosis and cell cycle in MCF-7 human breast cancer cell lines [76]. The outcomes of another research evaluated the anticancer actions of flavanone derivatives on human breast malignant cells and exhibited actions through p53-mediated cell and induce apoptosis in G1 phase of cycle arrest [77]. In another approach, Usman *et al.* presented the cytotoxic potential of flavanones obtained from bark of plant *Cryptocarya costata* [78]. Similarly, in another study, flavanones exhibited significant cytotoxic action on colorectal cancer cells via formation of apoptotic bodies and DNA fragmentation [79]. Flavonoids (Figure 11) exhibiting cell killing potential [77,80–83].

Kumar *et al.* reported the synthesis and evaluation of naphthoflavones (92-95) as anti-proliferative agents against a panel of human cancer cell lines. Compound (92) displayed noteworthy cytotoxicity towards MiaPaCa-2 cell lines, with IC<sub>50</sub> values of 1.93  $\mu$ M and 5.63  $\mu$ M against MCF-7 cell lines. The molecule (92) was found to induce apoptosis which was established by DAPI staining, MMP loss, phase contrast microscopy and cell cycle arrest of 55.19 % at 20  $\mu$ M in MiaPaCa-2 pancreatic cancer cells (Figure 12) [84].

Myricetin is one of the flavonoid-based phytomolecule and found in numerous natural sources. Outstandingly, those myricetin based subordinates are thought to exhibit anticancer potential, which have diminished pancreatic cancer via apoptosis [85,86]. In another finding, Xue *et al.* established a series of novel myricetin analogues [87]. It was established that analog (98) displayed significant action in MDA-MB-231 human breast cancer cells. The outcomes from the telomerase inhibition experiment also confirmed that compound (98) exhibited extraordinary action towards human MDA-MB-231 cells, with IC<sub>50</sub> value in 0.91  $\mu$ M. The molecular docking of scaffold 98, towards target site,

revealed that the heterocyclic nucleus was intensely embedded into dynamic site, establishing hydrophobic associations with build-ups of Phe568, Pro627, with four methoxy groups having hydrophobic interactions with amino acid residues Phe568, Lys902, Pro627, Pro929 and Val904 presented in (Figure 13) [87].

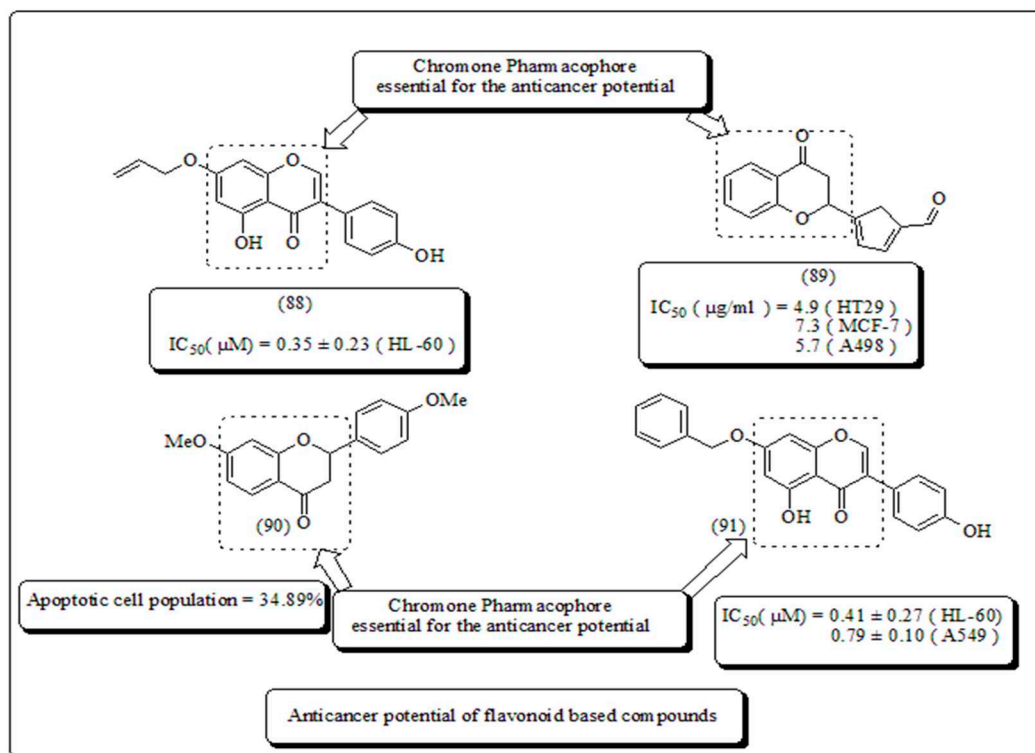


Figure 11. Anticancer potential of some flavonoid-based compounds [88–91].

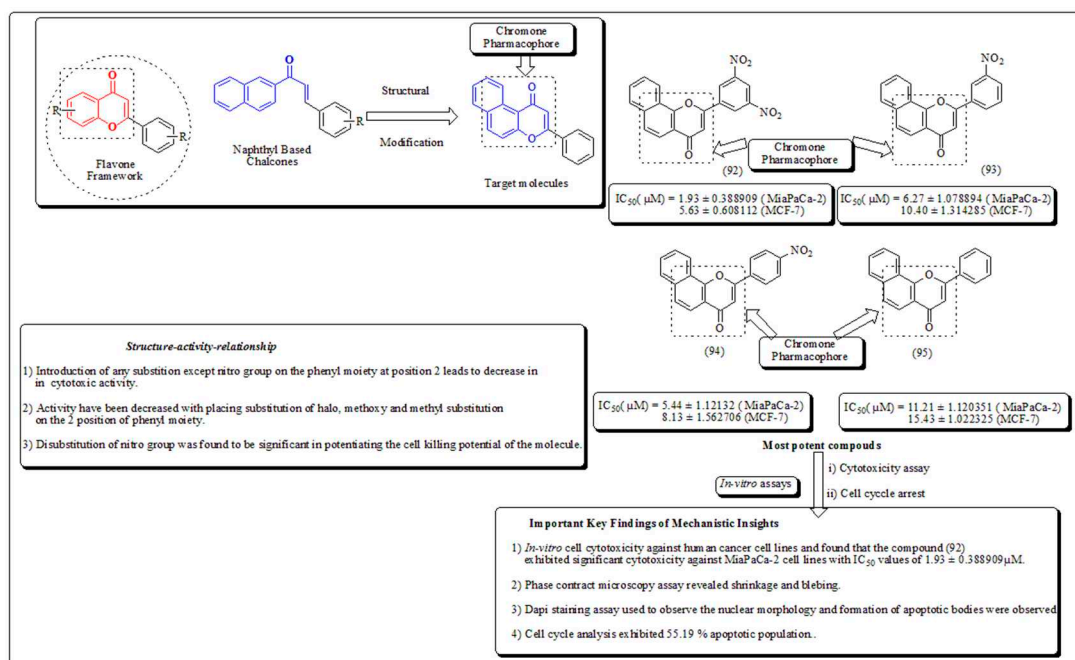
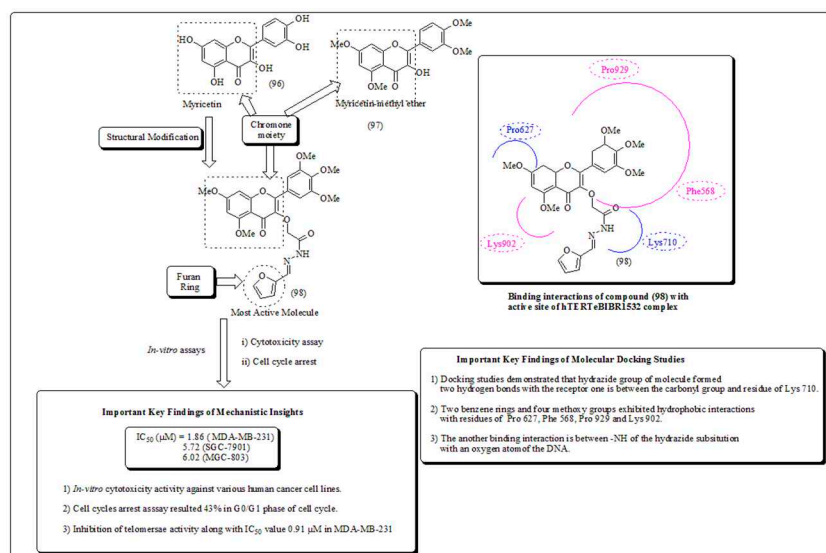
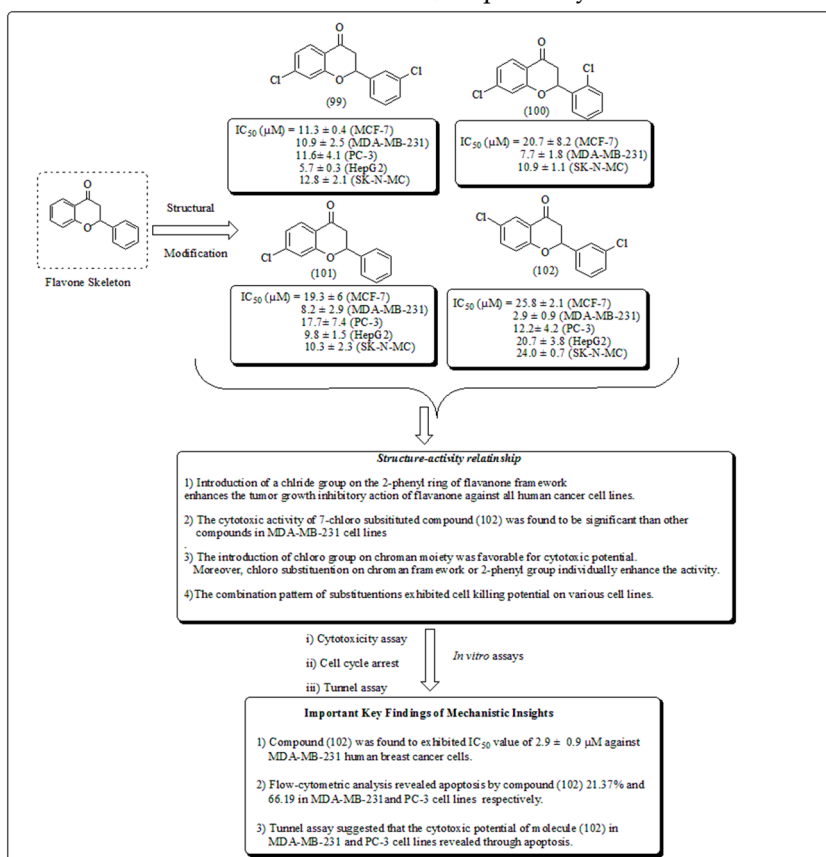


Figure 12. Structure of compounds 92-95, along with mechanistic insights as anticancer agents.



**Figure 13.** Structure of myricetin anticancer scaffold along with docking studies.

Safavi *et al.* conceded the synthesis and evaluated halogenated flavanones towards human cancer cells [88]. From all the synthesized analogs, 3',7- dichloroflavanone (99) displayed potential activity in MCF-7, PC-3, LNCaP, Hep-G2, SK-NMC, and KB cells. However, compound (102) presented in (Figure 14) exhibited  $IC_{50}$  of 2.9  $\mu$ M and was the most significant analog against MDA-MB-231 breast cells, and approximately 12 times more powerful, than etoposide. Thus, introduction of chloro group on chromanone moiety and on C-2 attached phenyl nucleus was used as structural modification to form a lead pharmacophore of flavanones. The analog (102) induces apoptosis from 21.37% and 66.19% in MDA-MB-231 and PC-3 cells, respectively.



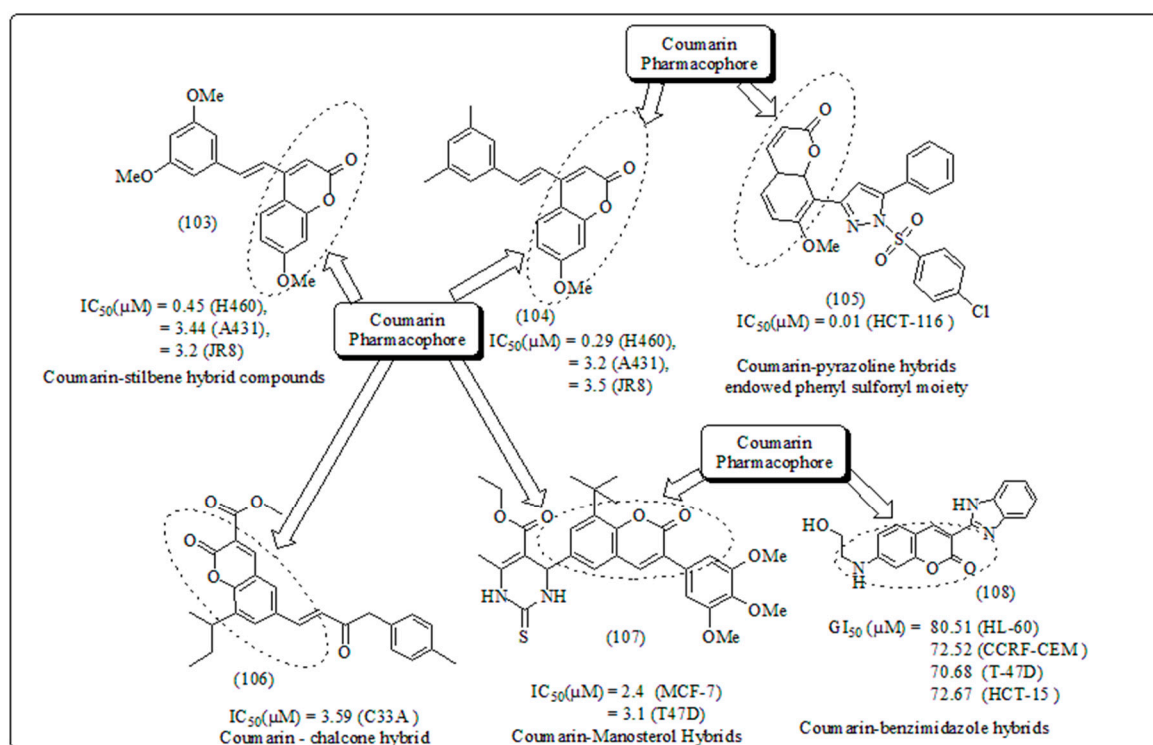
**Figure 14.** Structure flavanones as apoptosis-inducing compounds along with their important key findings.



The results of TUNEL assays recommended that the cytotoxic potential of this analog in MDA-MB-231 and PC-3 cell lines exerted through apoptosis [88].

## 2. Coumarin-based oxygen containing heterocyclic scaffolds

Oxygen plays a chief role in the architecting of coumarins moiety which is an exceptional class of oxygen based heterocyclic scaffold, presenting a significant role in medicinal chemistry, owing to their ample range of pharmacological activities and structural diversity [89]. Coumarins perform a distinctive role in nature [90,91]. The incidence of coumarin heterocycle in phytomolecules exhibiting extensive activities such as anticancer [92,93] antitubercular [94], anti-HIV, [95] anti-influenza [96], antiviral [97], anti-Alzheimer [98,99], antimicrobial [100] and anti-inflammatory [101] actions makes it a fortunate structural motif [102]. Coumarin scaffolds explored through established biological actions of coumarin based hybrids as promising molecules (Figure 15) [103–107].



**Figure 15.** Structure of coumarin hybrids (103-108).

Coumarin hybrids are pyranocoumarin derivatives, having several structural arrangements between pyran and coumarin rings. The few important pyranocoumarins such as khellactone (109) obtained from *Ligusticum elatum* and xanthyletin (110) predominantly extracted from *Zanthoxylum americanum*, along with pyripyropenes (111) and arisugacins (112) presented in (Figure 16) [108].

A few years ago, Kumar and co-workers synthesized and evaluated coumarin hybrids [109]. The design strategy involved the merging of chalcone and coumarin hiring a pyran as a connector. All the analogs were evaluated against a panel of cancer cell lines. Compound 113 (Figure 17) exhibited probable effects in MiaPaCa-2 and HCT 116 cells with  $IC_{50}$  values of 4.3 and 1.4  $\mu M$ , respectively. Compound 113 initiated apoptosis which was confirmed by MMP loss, Hoechst 33258 staining and cell cycle arrest with apoptotic population of 57.19% in a dose of 20  $\mu M$  [109].

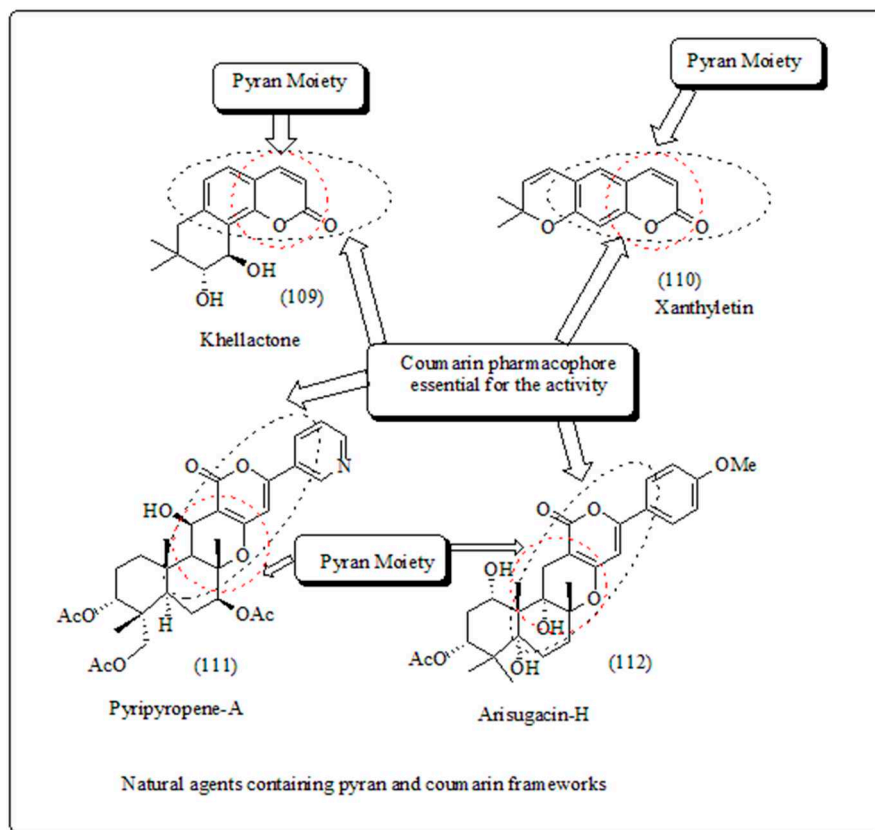


Figure 16. Naturally-occurring compounds (109-112) consisting pyran and coumarin scaffolds.

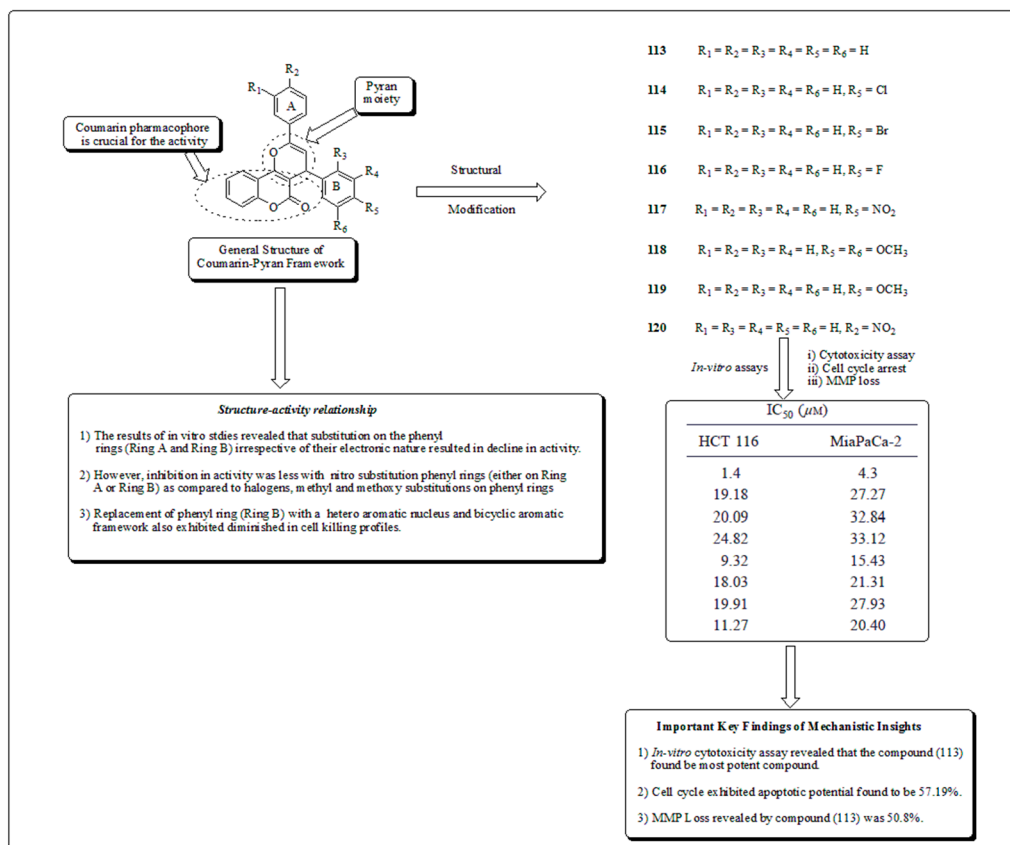
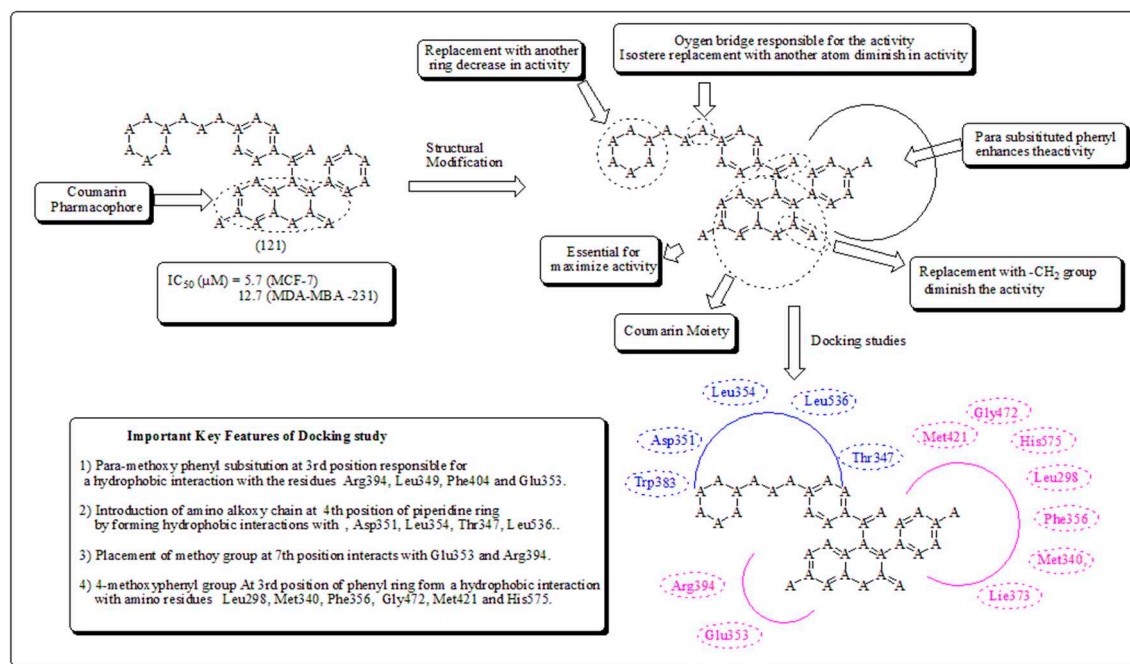


Figure 17. Most significant coumarins scaffolds reported by Kumar et al. [109].

A decade ago, Hussain *et al.* further carried out a synthesis of coumarin molecules as potent anticancer agents towards breast cancer [110]. Compound (121) was established as ER- $\alpha$  selective and most potential from all synthesized analogs. The docking study displayed that analog (121) satisfactorily fits well into the receptor pocket of ER- $\alpha$ . The coumarin moiety and the p-methoxyphenyl substitution on the third position formed a hydrophobic binding interaction with amino acids such as Glu353, Phe404, Leu349 and Arg394, the introduction of amino alkoxy substitution, fixed the piperidine nucleus by forming hydrophobic interaction with residue like Trp383, Thr347, Asp351, Leu536 and Leu354. The introduction of methoxy on coumarin at the 7th position interacted with Arg394 and Glu353 presented in (Figure 18).

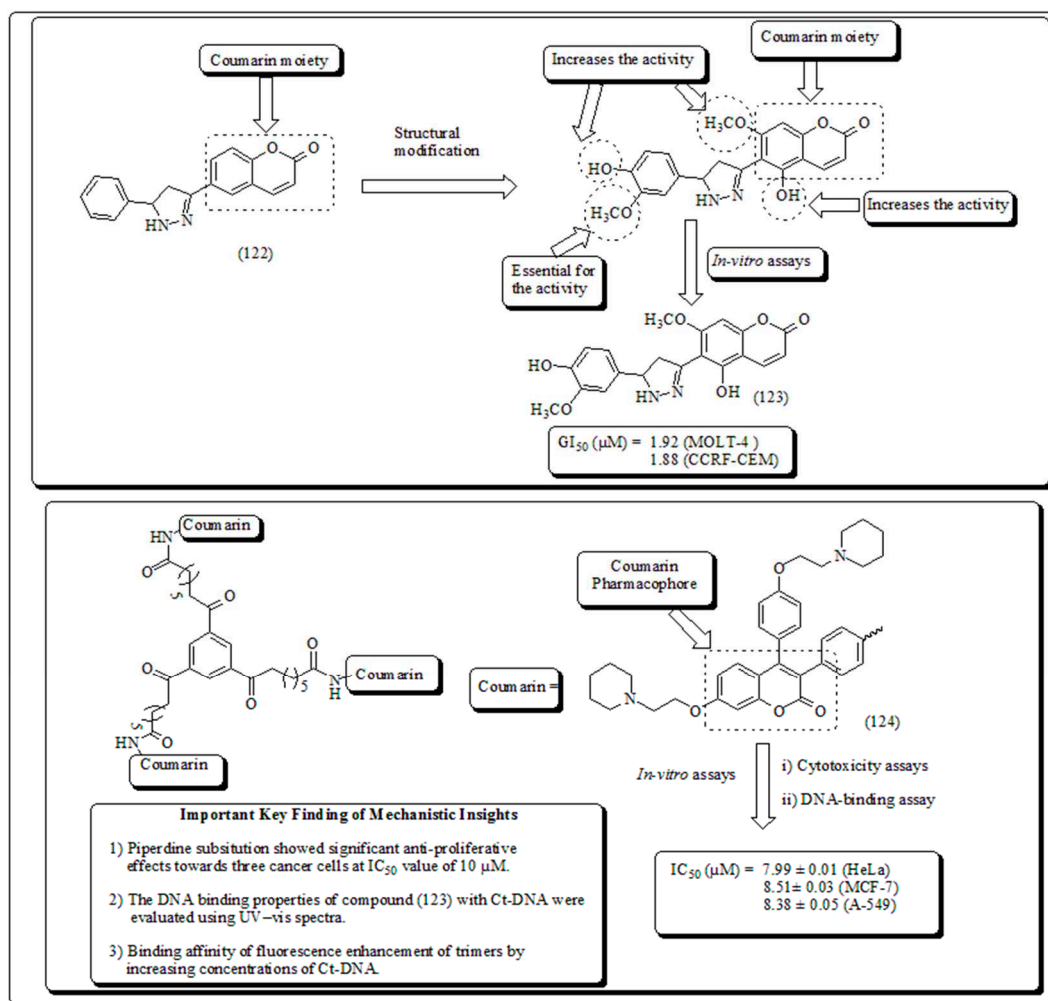


**Figure 18.** Structure of selective coumarin analog along with molecular modeling study.

The analog (121) had a comparable binding pattern for ER- $\beta$  site, coumarin pharmacophore developing hydrophobic binding interactions with residues Glu305, Leu343, Arg346, Leu339 and Leu301. The 4-methoxyphenyl group formed a hydrophobic interaction with Met421, Met340, Gly472, Phe356, His575 and Leu298, which are indispensable for ER- $\beta$  binding [110].

Coumarin is an amendment of benzopyran-2-one through introducing a direct heterocyclic substituent. For instance, a heteroaryl substitution is introduced at 3 or 4 position of coumarin moiety. Consequently, 3 and 4-heteroarylcoumarins are described to display significant therapeutic activities including anticancer [111], antibacterial, antimicrobial [112], DNA cleavage [113], antioxidant, anticholinesterase [114] and monoamine oxidase inhibitory action [115]. Encouraged from this, Yana and coworkers presented the synthesis and anti-proliferative screening against a panel of human cancer cell lines. The results of the study revealed that compound (123) exhibited significant anti-mitotic actions presented in (Figure 19) [116].

Another study showed the synthesis of coumarin derivatives with enhanced anticancer properties [117,118]. The resultant dimeric product was revealed to have more effective than of monomeric compound (with  $IC_{50} \sim 70 \mu\text{mol/L}$ ) [119,120]. Prompted from this, the conception of molecular oligomerization lead to invention of two novel classes of dimeric analogs of triphenylethylene-coumarin hybrids [121,122]. The dimeric analogs displayed antitumor activities [123,124]. Similarly, Zhang and his coworkers further exposed new trimers of triphenylethylene-coumarin hybrids. The trimeric analog (124) revealed significant anti-proliferative activity with  $IC_{50}$  value of 7-9  $\mu\text{M}$  range presented in (Figure 19) [125].

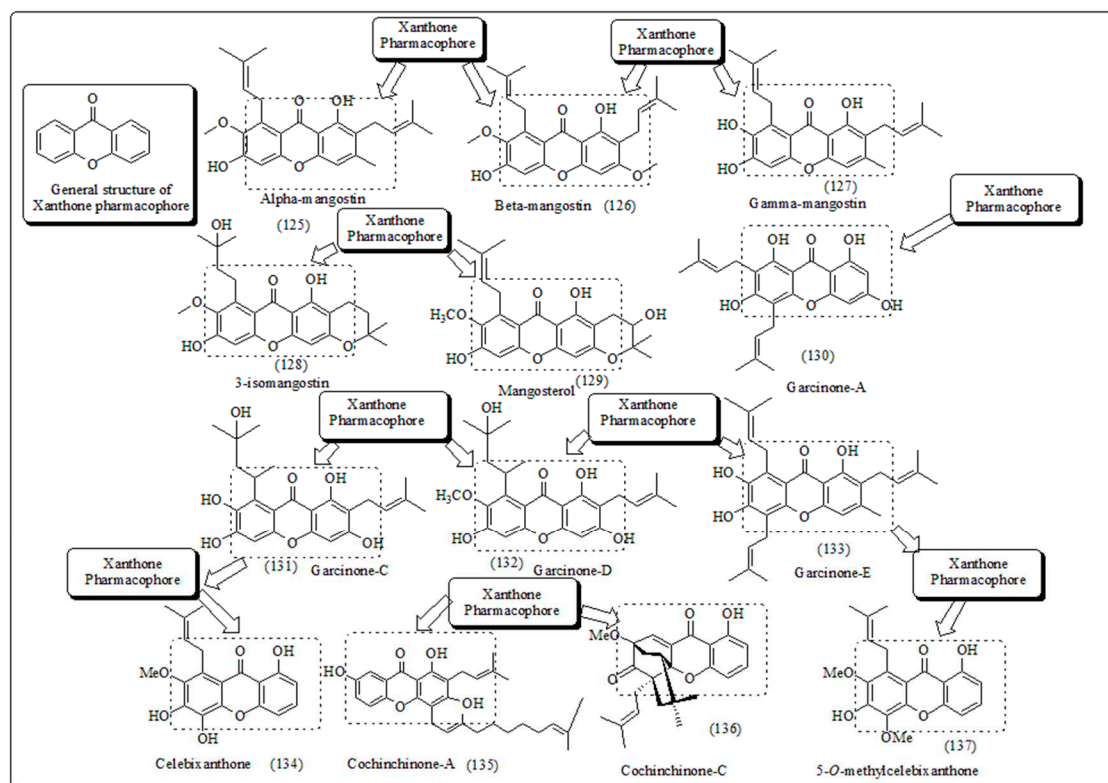


**Figure 19.** Structure of coumarin analogs and triphenylethylene-coumarin hybrids with cell killing potential reported by Garazd *et al.* [116] and Zhang *et al.* [125].

### 3. Xanthone-based oxygen-containing heterocyclic scaffolds

Xanthenes and xanthenes are an exceptional class of oxygen containing tricyclic compounds in which pyran ring is fused with two benzene rings on both the sides and exhibited diverse attractive pharmacological effects, depending on nature and types of substituents [126–128]. Recently, xanthenes and xanthenes have been appreciated as effective pharmacophore in the arena of medicinal world [129]. Earlier, xanthenes were publicized as larvicides, bug sprays, and ovicides [130]. Presently, several studies recognized that xanthone scaffolds are proficient to halt the progression of tumor cells and also holding anti-inflammatory and antioxidant actions [131]. Xanthenes are primarily available in plants belong to Clusiaceae, Bonnetiaceae, Gentianaceae and Podostemaceae families [132]. Structures of natural xanthenes presented in marketed formulations and potent cytotoxic compounds as reported by Lee *et al.* [133] and Laphookhieo *et al.* [134] were depicted in (Figure 20).





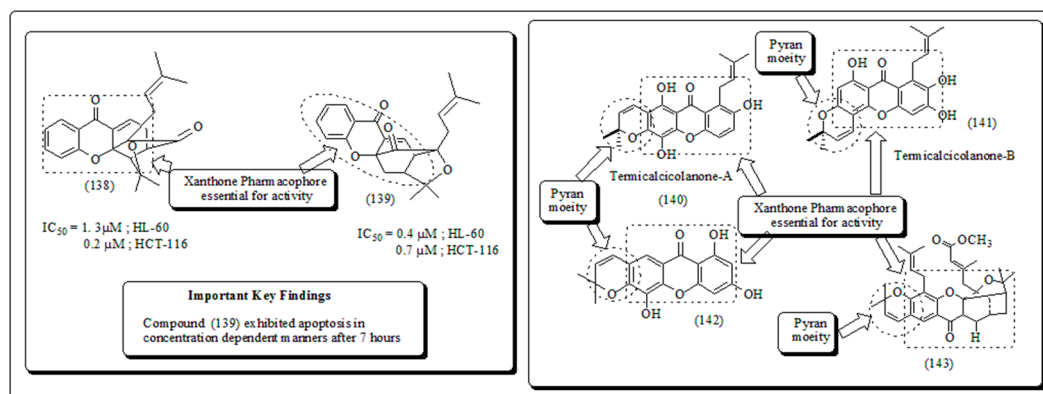
**Figure 20.** Structures of xanthenes based compounds in marketed formulations.

Numerous man-made and naturally occurring xanthene scaffolds have been described to reveal anti-malarial [135], antitumor [136], anti-trypanosomal [137], and antileishmanial properties. In recent past years, the synthesis and evaluation of this class of molecules have been increased, both in the arena of material science and medicinal world. Predominantly, xanthenes are well explored heterocyclic scaffolds with dibenzo- $\gamma$ -pyrone structural motif [138–141]. Several xanthenes isolated containing plant-based extracts are widely used in folklore medicines [142–146]. Furthermore, some marketed preparations containing xanthone as their structural unit presented in (Figure 20).

Inspired from previous findings Laphookhieo and his co-workers extracted  $\alpha$ -mangostin, 5-*O*-methylcelebixanthone,  $\beta$ -mangostin, celebixanthone, cochinchinone A and cochinchinone C from roots of plant *Cratoxylum cochinchinense* [134]. These molecules were evaluated cytotoxic effects against human lung cancer, NCI-H187 cell line. Amongst these,  $\alpha$ -mangostin, celebixanthone, cochinchinone A displayed cytotoxicity along with  $IC_{50}$  in a range of 0.65 to 5.2  $\mu$ g/mL. Similarly, Chantarasriwong *et al.* recognized the set of Garcinia xanthenes and tested for their anti-proliferative actions and inducing apoptosis towards human leukemic and colon HL-60 and HCT-116 cells respectively. Molecule (138) demonstrated to have maximum action against human colon cancer along with  $IC_{50}$  value 0.2  $\mu$ M towards HCT-116, while another molecule (139) was the most effective Leukemia, HL-60 cells with  $IC_{50}$  value of 0.4  $\mu$ M (Figure 21) [147].

In a parallel study, Matsumoto along with his coworkers established the xanthenes isolated from *Garcinia mangostana* revealed a remarkable anticancer activity [148]. Nevertheless,  $\alpha$ ,  $\beta$  and  $\gamma$ -mangostins (126–127) were found to be active in a dose of 10  $\mu$ M. The most potent molecule at such concentration was  $\alpha$ -mangostin. The  $\alpha$ -mangostin exhibited anti-proliferative actions leukemia cells like K562, U937 and NB4. Chiang *et al.* stated that concentrate of mangostin-organic pericarp displayed an intense anti-leukemic action, with  $IC_{50}$  of 159 and 61  $\mu$ g/mL towards Raji and K562 cells respectively [149]. Prompted from these results, Balunas *et al.* have also evaluated all the three mangostins using a non-cell, chemical based microsomal aromatase hindrance experiment assay in  $IC_{50}$  value 4.97  $\mu$ M against breast cancer, SK-BR-3 cells [150]. Recently, Jung *et al.* presented antitumor effect of these molecules in pre-neoplastic injuries persuaded using 7,12-dimethylbenz[a]anthracene

(DMBA) in mouse mammary organ enlargement [151]. In another approach, Suksamrarn *et al.* extracted distinctive xanthenes from mangosteen pericarp of fruit and screened it for antineoplastic activity against human cancer cells, including breast, small cell lung and mouth carcinoma; BC-1, NCI-H187 and KB cells with  $IC_{50}$  values of 3.53, 3.72 and 2.8  $\mu\text{g/mL}$  respectively. However,  $\alpha$ -mangostin (125) displayed the most noticeable result on BC-1 cell line, with  $IC_{50}$  value 0.92  $\mu\text{g/mL}$  [152].



**Figure 21.** Structure of compounds, along with anti-proliferative potential.

Chen *et al.* proved that  $\gamma$  and  $\alpha$ -mangostins produced noticeably cytotoxic effects towards RAW 264 cells and  $IC_{50}$  values of  $\gamma$  and  $\alpha$ -mangostins were found to be 10.1 and 12.4  $\mu\text{M}$  respectively [153]. Similarly, Watanapokasin *et al.* studied the anticancer action of mangostin xanthenes, on colon malignant cells [154]. In addition, minor doses of the extract of mangostin were diminished the tumor volume in xenograft model. The mangosteen pericarp concentrate comprises of 25%  $\alpha$ -mangostin reducing 50%–70% of tumor size in balb/c mice bearing colon tumor NL-17 xenografts model [154–157].

Cao *et al.* extracted two novel cytotoxic xanthenes named termicalcicolanone B (141) and termicalcicolanone from the ethanolic extract of the plant *Terminalia calcicola* presented in (Figure 21) [158]. These analogs were screened for their cytotoxic activity in ovarian cancer, A2780 cells having  $IC_{50}$  values of 8.1 and 40.6  $\mu\text{M}$  respectively [158]. In another discovery, Han *et al.* isolated new prenylated xanthenes, in addition to a few compounds from bark of *Garcinia lancilimba* [159]. These analogs were established their apoptotic potential through caspase-3 activation in HeLa-C3 cells. Similarly, Tao *et al.* extracted new xanthenes, a pair of novel natural compounds from resin of *Garcinia hanburyi* [160]. These compounds were assessed for their cytotoxic effect against HeLa cervical carcinoma, along with adriamycin as reference standard and compound (143) exhibited cytotoxicity with  $IC_{50}$  value of 111  $\mu\text{M}$  (Figure 21) [160].

*Garcinia hanburyi*, is a resin initially employed as folk medicine and pigment. In modern era, a group of xanthenes (called *Garcinia* xanthenes) were recognized as bioactive phytomolecules with potent therapeutic properties such as anti-HIV-1, antitumor, anti-inflammatory, antibacterial activities. The compounds were available in fruit, resin, and in other parts of the plant. Additionally, manifold mechanisms of cytotoxic activity were documented including apoptosis induction, cell cycle arrest, anti-angiogenesis and telomerase inhibition [161]. Caged xanthenes obtained from *G. hanburyi* were examined for cytotoxic activities towards HCT-116, A 549, K 562/R, SMMC-7221, HepG2, Huh7 and SH-SY5Y cells of colon, lungs, doxorubicin-resistant K 562, hepatoma, hepatocellular, liver and neuroblastoma human cell carcinoma respectively. The modified xanthenes were induced apoptosis in HepG2 cells in a concentration dependent pattern [162,163]. Jang *et al.* described modified xanthenes presented in (Figure 22) were selective against TrkA receptor, displaying a strong neurotrophic activity by selective binding to TrkA, through its provoking Akt/PI3-kinase/ and MAPK activation, tyrosine phosphorylation and thus inhibiting neuronal cell death [164].

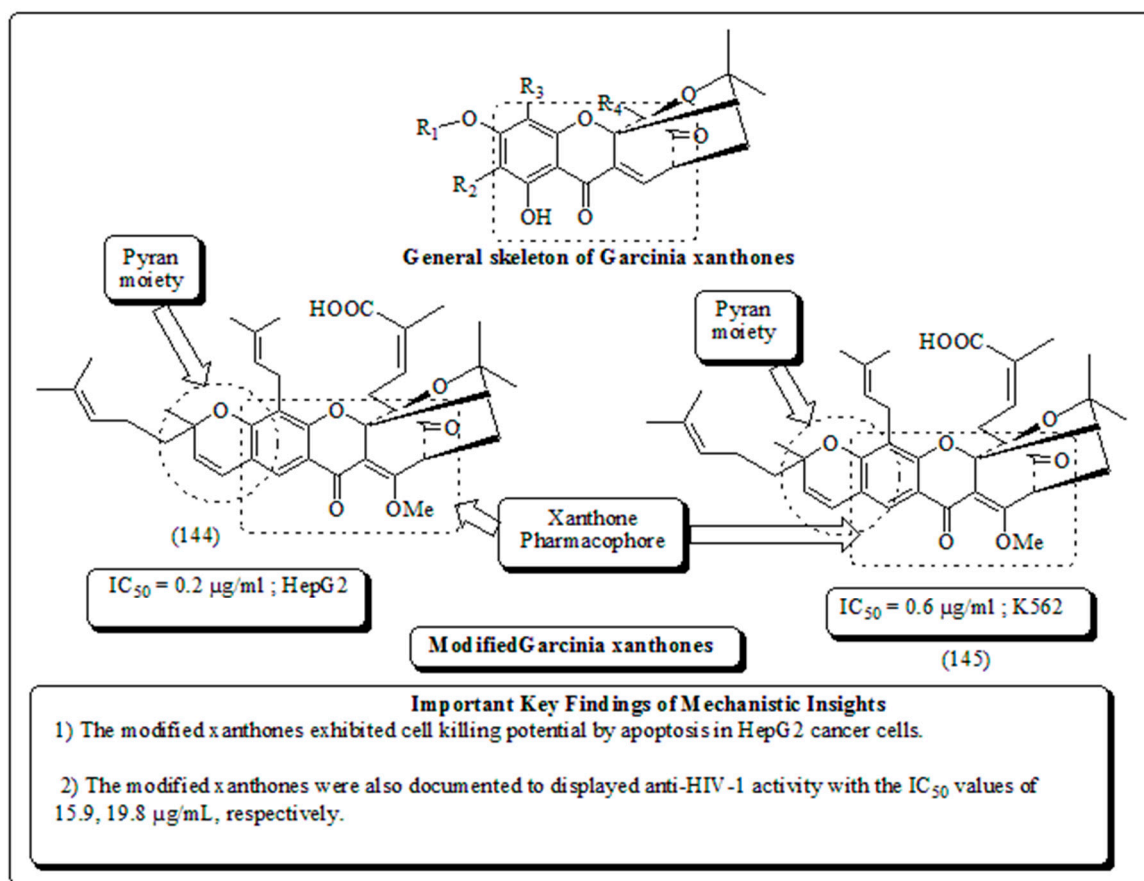
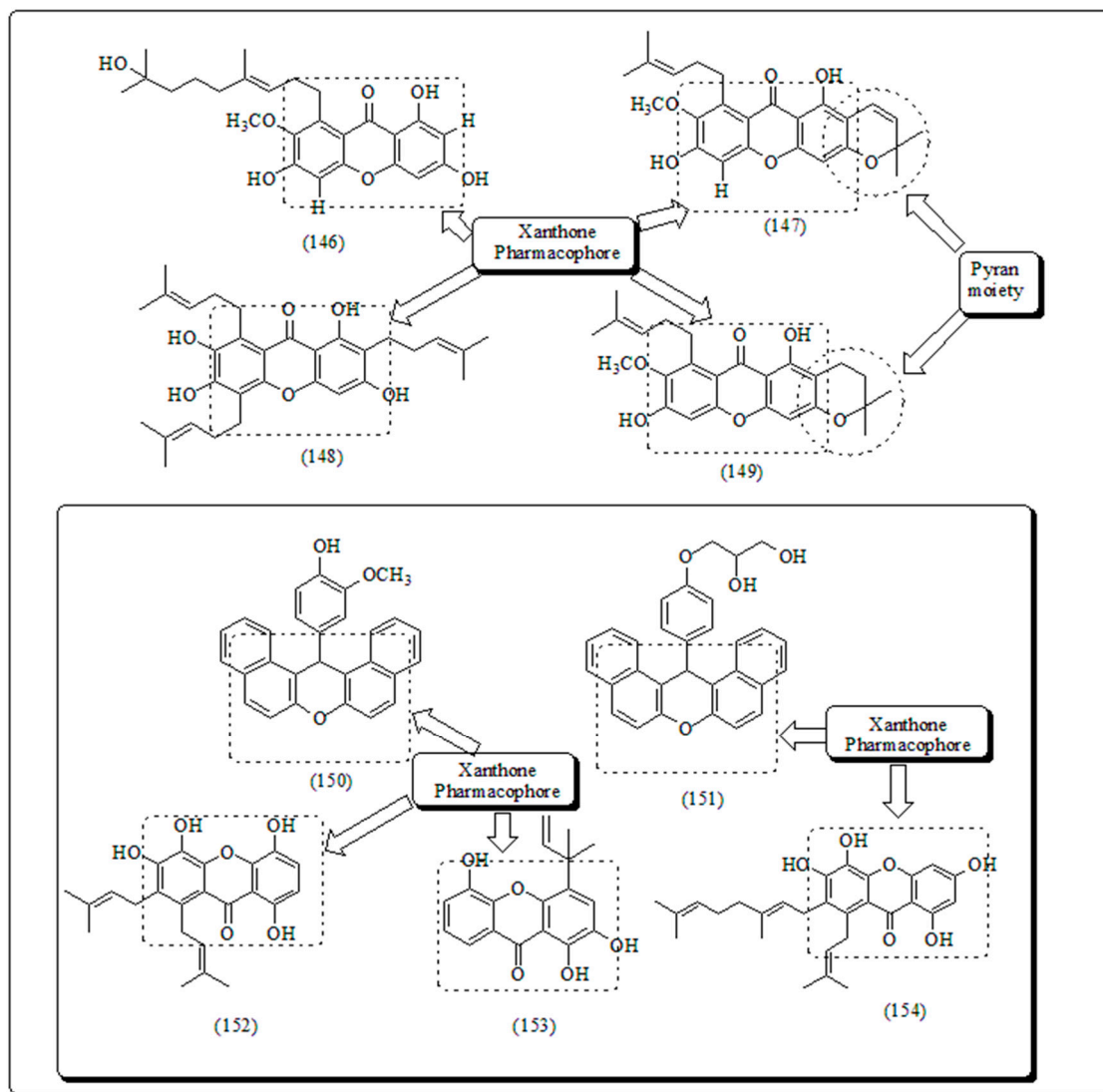


Figure 22. Structure of compounds as screened by Jang *et al.* [164].

Inspired by previous findings Zelefacek *et al.* found butyraxanthenes A-D, four known xanthenes (146-149) from the bark of *Pentadesma butyracea* [165]. These compounds were assessed for in vitro anti-plasmodial actions against *Plasmodium falciparum* chloroquine-resistant strain along with cytotoxic effect in MCF-7 human breast tumor cell lines. It was found that among all screened analogs, only butyraxanthone D (149) was inactive with  $IC_{50} > 10 \mu\text{g/mL}$ , while compound 147 displayed significant cytotoxic effectiveness (Figure 23) [165,166].

Bhattacharya and his coworkers synthesized xanthenes from aryl aldehydes and  $\beta$ -naphthol catalyzed by  $\text{TaCl}_5$  in solvent-free one-pot condensation conventional heating method [167]. All these xanthenes (Figure 23) were screened towards group human tumor cells including SW-620, Colo-205 (Colon), 502713, SKNSH (CNS), PC-3 (Prostate) and A-549 (Lung) sulforhodamine B assay. The compound 150 displayed  $IC_{50}$  values from 41.3 and 37.9  $\mu\text{M}$  against 502713 and Colo-205 respectively, however compound (151) presented  $IC_{50}$  of 41.9  $\mu\text{M}$  towards Colo-205 cells. Niu *et al.* isolated bracteaxanthenes and 1,4,5,6-tetrahydroxanthenes together along with 26 known molecules from ethanolic extract of stem bark of plant *Garcinia bracteata*. All compounds were assessed cell killing potential against HL-60 human leukaemic cells. The prenylated xanthenes depicted in (Figure 23) displayed potent effects. Compounds 152-154 were found to be the most effective with  $GI_{50}$  values of 2.8, 3.4 and 3.1  $\mu\text{M}$  respectively against HL-60 malignant cells [168].

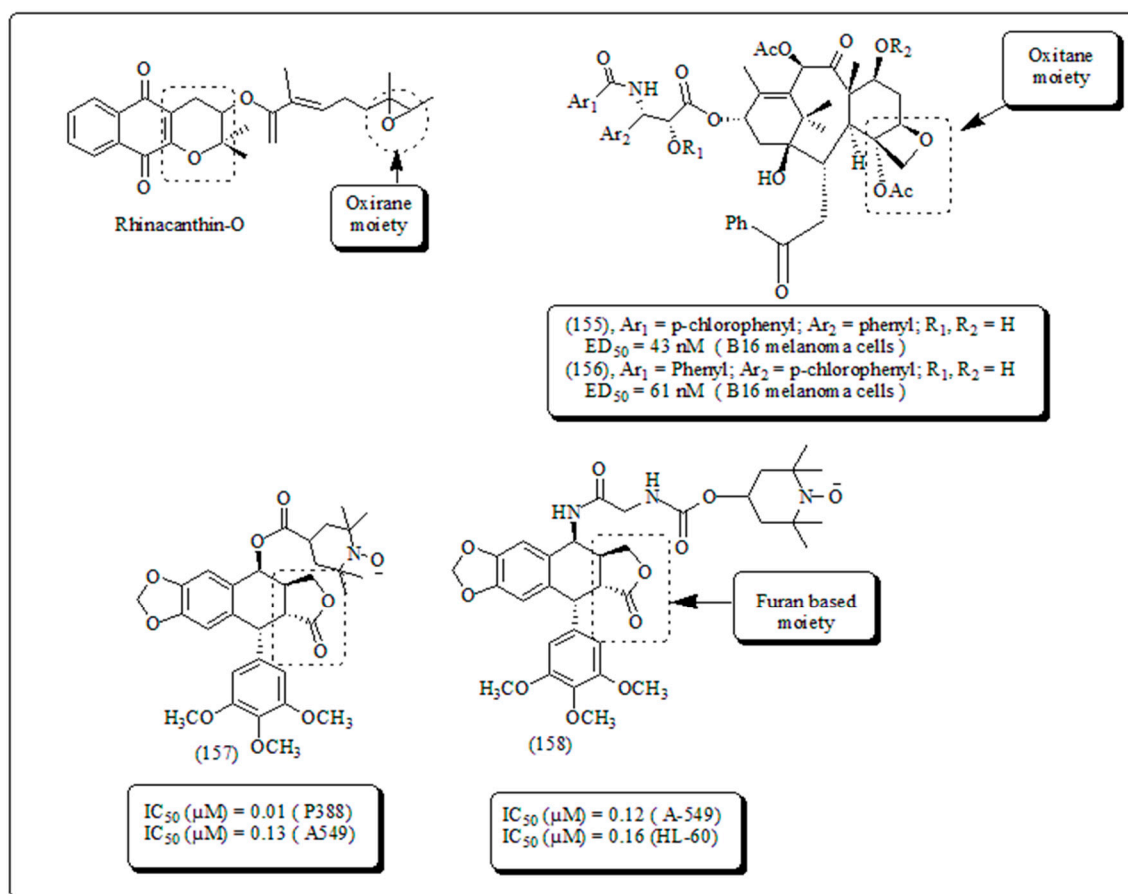


**Figure 23.** Most effective cytotoxic analogs (146-154) documented by Zelefsky *et al.* [165], Mosoophon *et al.* [166], Bhattacharya *et al.* [167] and Niu *et al.* [168].

#### 4. Miscellaneous oxygen-containing heterocyclic scaffolds

Numerous researchers around the globe were involved in the hunting of novel entities for the mitigation of diverse types of cancers. Several phytomolecules and their derivatives were clinically used against various cancers. Rhinacanthin O is pyran and oxirane based compound which (Figures 8 and 24) is isolated from *Rhinocanthus nasutus* and is therapeutically active towards breast cancer [63]. Similarly, taxols and phodophyllotoxins its derivatives consist of oxitane and furan rings, respectively were also used clinically nowadays against the treatment of various cancers. Structure of compounds containing oxirane, oxitane and furan moieties presented in (Figure 24). Additionally, oxygen and nitrogen containing rings such as oxazole and iso-oxazole along with their anticancer potential were also discussed as follows.

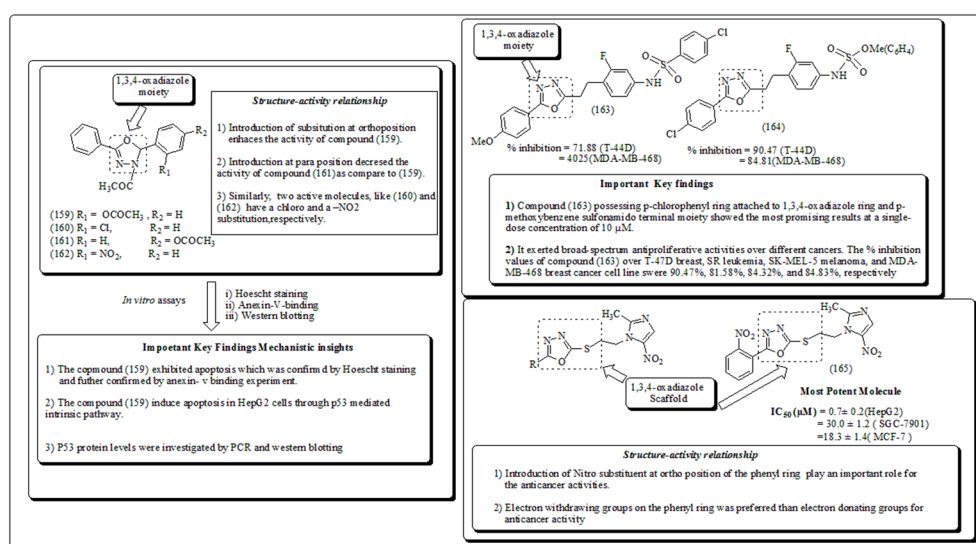




**Figure 24.** Structure of compounds containing oxirane, oxitane and furan moieties.

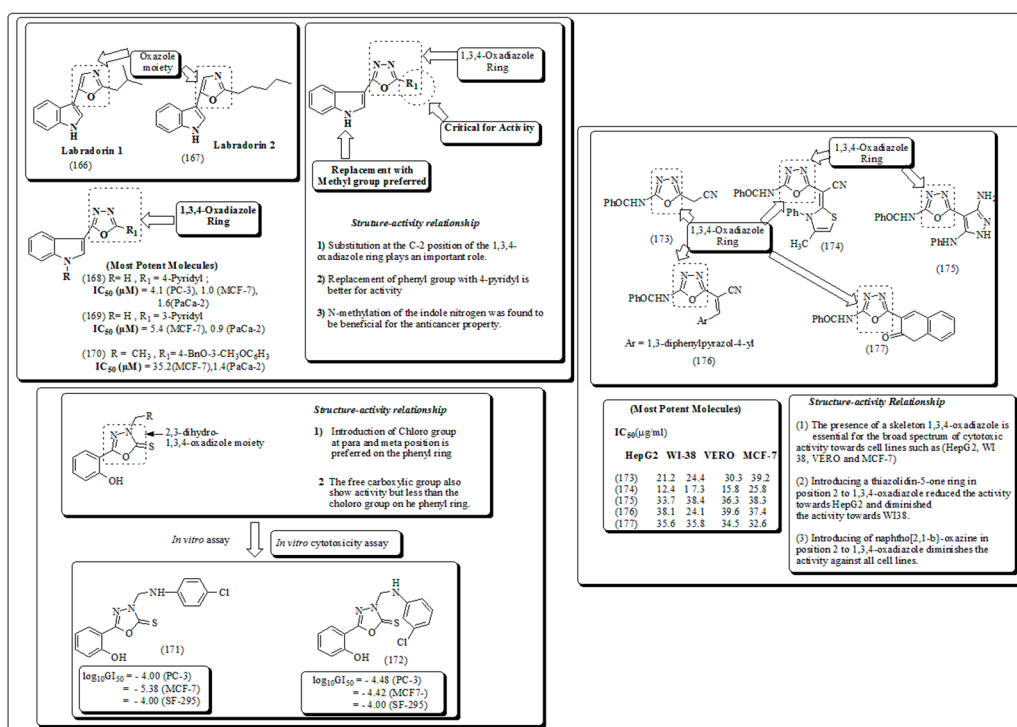
1,3,4-Oxadiazoles are noteworthy class of heterocyclic molecules with an extensive array of therapeutic activities including anticancer [169–172], antineoplastic [173], inhibition of tyrosinase [174] antiviral [175], fungicidal [176]. These play an important intermediate role in various organic synthesis reactions [177] and are usually used engaged as hole-blocking and electron transporting agents [178]. These structural motifs are extensively employed in medicinal chemistry and documented as privileged groups of structures. Furthermore, 1,3,4-oxadiazole, substituted 1,3,4-oxadiazoles and 1,2,4-oxadiazoles have been attracted because of their uses in photoluminescence, organic light-emitting diodes, material science and polymers. Moreover, 1,3,4-oxadiazole are exceptional bio-isosteres of esters and amides, which can impact noticeably in intensifying biological activities by involving in hydrogen bonding interactions with amino acid residue receptors [179].

Sankhe *et al.* recognized a series of substituted-1,3,4-oxadiazole analogs and evaluated for their antineoplastic action towards numerous human cancer cell lines using *in vitro* assays. Structure of several oxygen based heterocyclic compounds along with their mechanistic insights IC<sub>50</sub> value and structural-activity relationship have been presented in Figures 25–29 [180].



**Figure 25.** Structures of miscellaneous oxygen containing heterocyclic compounds along with mechanistic insights.

Mahmoud *et al.* designed a class of 1,3,4-oxadiazole analogs possessing sulfonamide moiety and screened against a panel of NCI-58 human cancer cell lines [181]. A library of new 1,3,4-oxadiazole thioether analogs based on 1-(2-hydroxyethyl)-2-methyl-5-nitroimidazole scaffold have been synthesized and evaluated for anticancer activity against three human cancer cell lines by Qian and coworkers. [182]. Naturally occurring oxazoles are recognized to exhibit diverse pharmacological activities. Labradorin 1 (166) and Labradorin 2 (167), shown in Figure 26, were established to be cytotoxic against lung-NSC, malignant cells. Dalip *et al.* have presented compounds (168-170) as cytotoxic agents [183].



**Figure 26.** Structures of compounds (166-177) with anticancer potential and structure-activity relationships.

Inspired from previous studies Ahmed *et al.* have described a class of substituted-1,3,4-oxadiazole analogs having anticancer activity. [184]. Samir and coworkers [185]. Presented 1,3,4-oxadiazole-based Heterocyclic analogs along with their mechanistic insights, IC<sub>50</sub>, SAR and docking studies exhibited binding interactions with amino acid residues. 1,2,4-Oxadiazoles are formerly designated for selective inhibition of receptors including and benzodiazepines [186,187], 5-hydroxytry- ptamine (5-HT1B/D) [188,189] histamine-H3 [190], 5-HT4 [191] and muscarinic [192]. These also revealed anticancer [193,194] anti-inflammatory and anticancer properties [195,196]. Kumar *et al.* have established a set of substituted-1,2,4-oxadiazoles, 1,3,4- thiadiazole and their bioisosters and evaluated for their cytotoxic potential using *in vitro* studies and presented in Figure 27 [197,198].

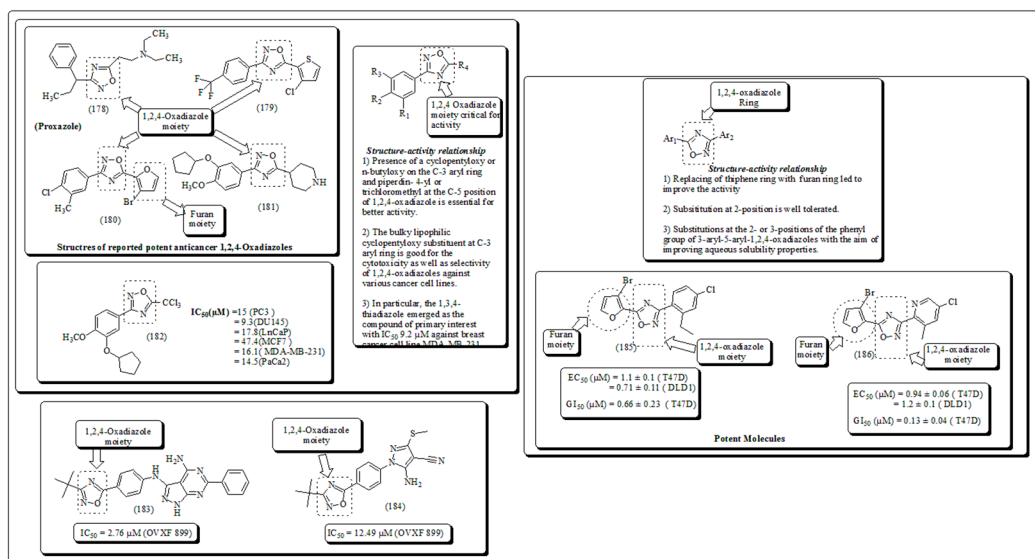


Figure 27. Structure of 1,2,4-oxadiazoles analogs and SAR studies.

Kemnitz and coworkers [199] reported the synthesis and evaluation of substituted 1,2,4-oxadiazoles with the intention to enhance the water solubility. Moreover, the structural activity relationship revealed that introduction of furan ring accelerating the anticancer activity and presented in Figure 28 [200,201].

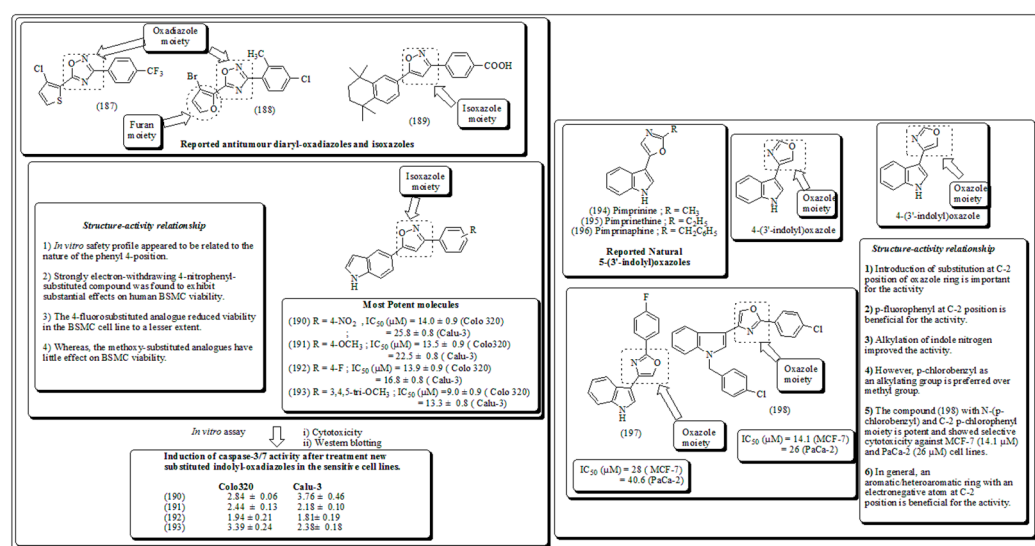


Figure 28. Structures of apoptotic antitumor agents.

Puthiyapurayil, *et al.* have architected and synthesized a novel combinatorial class of S-substituted-1,3,4-oxadiazole bearing N-methyl-4-(trifluoromethyl) phenyl pyrazole moiety and screened for in-vitro cytotoxic effects by MTT assay [202]. Recently, a series of quinoline derivatives have been screened as potential telomerase inhibitors by Juan *et al.* presented in Figure 29 [203,204]

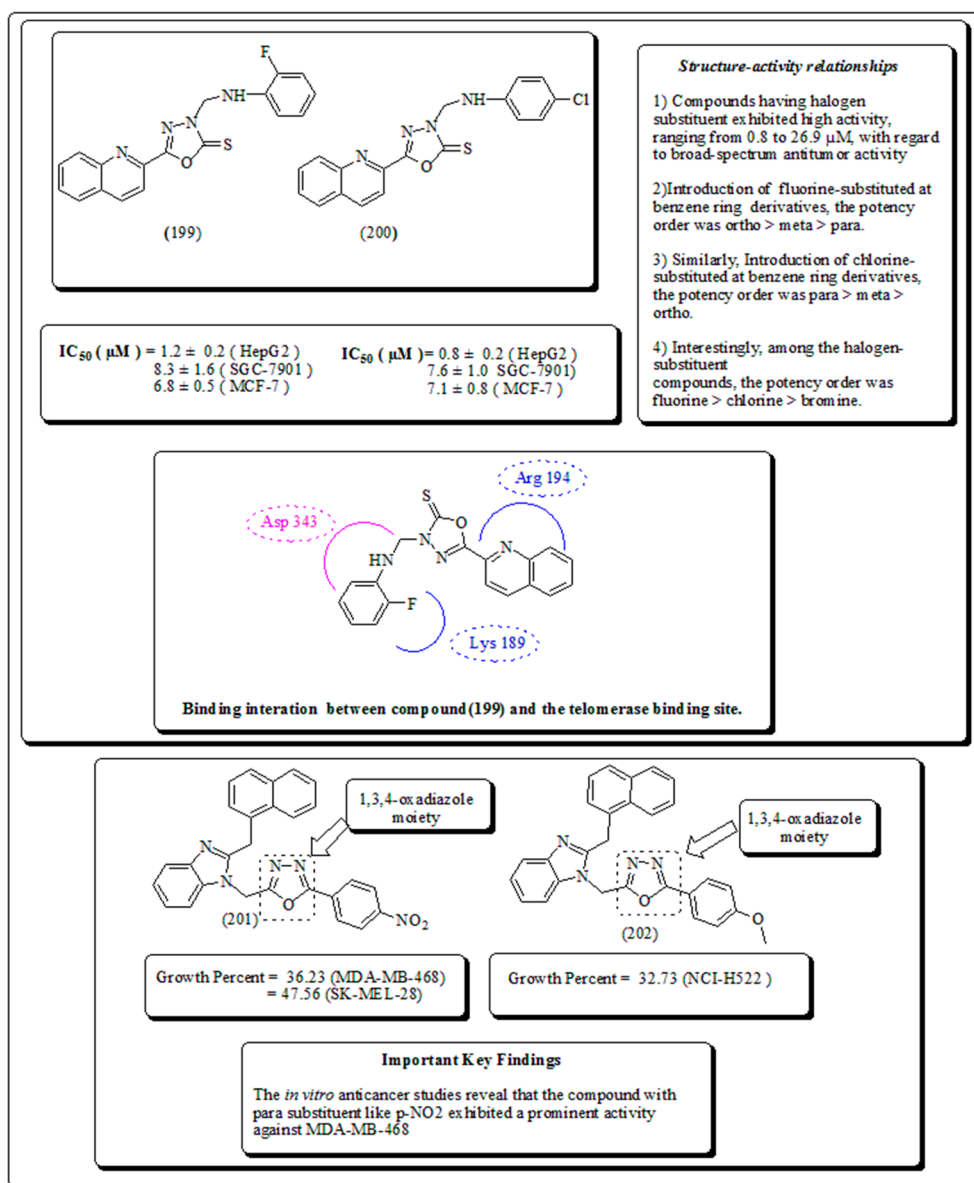


Figure 29. Structures of analogs reported by Juan *et al.* [203] and Mohammad *et al.* [204].

## 5. Conclusions

The abovementioned report endeavored to ascertain that the oxygen containing heterocyclic scaffolds were considered as privileged structures and has accredited considerable attention in last decade from pharmaceutical, industrial and academic scientists. As revealed from several cited reports, the oxygen-based scaffold is essential in architecting of innumerable flavonoids, phenolic compounds, coumarins and xanthenes present in numerous phytomolecules. Numerous oxygen based structural motifs exhibited astounding inhibitory actions along with their IC<sub>50</sub> values in nano and micromolar range. The universal assumption is that oxygen-based compounds are the privileged heterocycles recognized to have comprehensive potential therapeutic activities, predominantly against diverse human cancers. There are ample evidence that, the deployment of oxygen based substituted scaffolds including furan, pyran, chromone, coumarin and xanthone analogs have offered the platform for innovative chemical entities which might be act as potential therapeutic candidates

with diverse array of pharmacological properties. Several *in vitro*, *in silico* and *in vivo* studies have publicized that the oxygen based heterocyclic scaffolds with hypothetically useable structural motifs for development of anticancer and cytotoxic properties. Additionally, the structures of designed and synthesized compounds discussed in this assemblage noticeably highlighted the remarkable and promising cytotoxic profiles along with IC50 values, mechanistic insights, and their structure-activity relationships studies. The article also presented binding interactions with key amino acid residues in designated binding pockets of receptors, as validated by docking studies. In nutshell, the recorded activities, and recognized mechanisms of action, and can utilize hodgepodge of these oxygen based heterocyclics in the design of numerous novel structural frameworks.

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## References

- Chin, Y.W.; Balunas, M.J.; Chai, H.B.; Kinghorn, A.D. Drug discovery from natural sources. *AAPS J.* **2006**, *8*, E239-E253.
- Sharma, P.; Kumar, D.; Shri, R.; Kumar, S. Mechanistic insights and docking studies of phytomolecules as potential candidates in the management of cancer. *Curr. Pharm. Des.* **2022**, *28*, 2704-2724.
- Dua, R.; Shrivastava, S.; Sonwane, S.K.; Srivastava, S.K. Pharmacological significance of synthetic heterocycles scaffold: a review. *Adv. Biol. Res.* **2011**, *5*, 120-144.
- Sharma, P.; Shri, R.; Ntie-Kang, F.; Kumar, S. Phytochemical and Ethnopharmacological Perspectives of *Ehretia laevis*. *Molecules*, **2021**, *26*, 3489.
- Sharma, P.; Shri, R.; Kumar, S. Phytochemical and In Vitro Cytotoxic Screening of Chloroform Extract of *Ehretia Microphylla* Lamk. *Stresses*, **2022**, *2*, 384-394.
- Kaur, R.; Sharma, P.; Gupta, G.K.; Ntie-Kang, F.; Kumar, D. Structure Activity Relationship and Mechanistic Insights for Anti- HIV Natural Products. *Molecules*, **2020**, *25*, 2070.
- Kumar, D.; Sharma, P.; Kaur, R.; Lobe, M.M.; Gupta, G.K.; Ntie-Kang, F. In search of therapeutic candidates for HIV/AIDS: Rational approaches, design strategies, structure-activity relationship and mechanistic insights. *RSC Adv.* **2021**, *11*, 17936-17964.
- Singla, R.K.; Sharma, P.; Dubey, A.K.; Gundamaraju, R.; Kumar, D.; Kumar, S.; Madaan, R.; Shri, R.; Tsagkaris, C.; Parisi, S.; et al. Natural Product-Based Studies for the Management of Castration-Resistant Prostate Cancer: Computational to Clinical Studies. *Front. Pharmacol.* **2021**, *12*, 732266.
- Singla R.K.; Sharma, P.; Kumar, D.; Gautam, R.K.; Goyal, R.; Tsagkaris, C.; Dubey, A.K.; Bansal, H.; Sharma, R.; Shen, B. The role of nanomaterials in enhancing natural product translational potential and modulating endoplasmic reticulum stress in the treatment of ovarian cancer. *Front. Pharmacol.* **2022**, 13:987088. doi: 10.3389/fphar.2022.987088.
- Kumar, D.; Sharma, P.; Singh, H.; Nepali, K.; Gupta, G.K.; Jain, S.K.; Ntie-Kang, F. The value of pyrans as anticancer scaffolds in medicinal chemistry. *RSC Adv.* **2017**, *7*, 36977.
- Kumar, D.; Jain, S.K. A Comprehensive Review of N-Heterocycles as Cytotoxic Agents. *Curr. Med. Chem.* **2016**, *23*, 4338-4394.
- Gomtsyan, A. Heterocycles in drugs and drug discovery. *Chem Heterocycle Compd* **2012**, *48*, 7–10.



13. Von Angerer, E.; Biberger, C.; Leichlt, S. Studies on Heterocycle-Based Pure Estrogen Antagonist. *Ann. N. Y. Acad. Sci.* **1995**, *761*, 176–191.
14. Kumar, D.; Nepali, K.; Bedi, P.M.S.; Kumar, S.; Malik, F.; Jain, S. 4,6-diaryl Pyrimidones as Constrained Chalcone Analogues: Design, Synthesis and Evaluation as Antiproliferative Agents, *Anti-Cancer Agents Med. Chem.* **2015**, *15*, 793-803.
15. Kumar, D.; Singla, R.K.; Sharma, P.; Kumar, L.; Kaur, N.; Dhawan, R.K.; Sharma, S.; Dua, K.; Sharma, R. Phytochemistry and Polypharmacological Potential of Colebrookea Oppositifolia Smith, *Curr. Top. Med. Chem.* **2022**.
16. Kumar, D.; Singh, G.; Sharma, P.; Qayum, A.; Mahajan, G.; Mintoo, M.J.; Singh, S.K.; Mondhe, D.M.; Bedi, P.M.S.; Jain, S.K.; Gupta, G.K. 4-aryl/heteroaryl-4H-fused pyrans as Anti-proliferative Agents: Design, Synthesis and Biological Evaluation. *Anti-Cancer Agents Med.* **2018**, *18*, 57-73.
17. Kumar, D.; Sharma, P.; Nepali, K.; Mahajan, G.; Mintoo, M.J.; Singh, A.; Singh, G.; Mondhe, D.M.; Singh, G.; Jain, S.K. Gupta, G.K.; Ntie-Kang, F. Antitumour, acute toxicity and molecular modeling studies of 4-(pyridine-4-yl)-6-(thiophen-2-yl)pyrimidin-2(1H)-one against Ehrlich ascites Carcinoma and sarcoma-180. *Heliyon.* **2018**, *4*, e0061.
18. Sharma, P.; Sharma, R.; Rao, H.S.; Kumar, D. Phytochemistry and Medicinal Attributes of A. Scholaris: A Review. *Int. J. Pharm. Sci. Res.* **2015**, *6*, 1000-10.
19. Kaur, T.; Sharma, P.; Gupta, G.; Ntie-Kang, F.; Kumar, D. Treatment of tuberculosis by natural drugs: A review. *Plant Archives*, **2019**, *19*, 2168-2176.
20. Kumar, D.; Sharma, P.; Mahajan, A.; Dhawan, R.; Dua, K. Pharmaceutical interest of in-silico approaches, *Phys. Sci. Rev.* **2022**.
21. Damayanthi, Y.; Lown, J.W. Podophyllotoxins: Current status and recent developments. *Curr. Med. Chem.* **1998**, *5*, 205-252.
22. Wani, M.C.; Taylor, H.L.; Wall, M.E. Plant antitumor agents. VI. The isolation and structure of Taxol, a novel anti-leukemic and antitumor agent from *Taxus brevifolia*. *J. Am. Chem. Soc.* **1971**, *93*: 2325-2327.
23. Jordan, A.; Hadfield, J.A.; Lawrence, N.J.; McGown, A.T. Tubulin as a target for anticancer drugs: Agents which interact with the mitotic spindle. *Med. Res. Rev.* **1998**, *18*, 259-296.
24. Luduena, R.F.; Roach, M.C. Tubulin sulfhydryl groups as probes and targets for antimitotic and antimicrotubule agents. *Pharmacol. Ther.* **1991**, *49*, 133-152.
25. Podowysstzki, V. Pharmacological studies of Podophyllum peltatum. *Arch. Exp. Pathol. Pharmacol.* **1880**, *13*, 29-52.
26. Dwyer, P.J.; Leyland-Jones, B.; Alonso, M.T.; Marsoni, S.; Wittes, R.E. Etoposide (VP-16-213): Current status of an active anticancer drug. *N. Engl. J. Med.* **1985**, *312*, 692-700.
27. Jin, Y.; Chena, S.W.; Tiana, X. Synthesis and biological evaluation of new spin-labeled derivatives of podophyllotoxin. *Bioorg. Med. Chem.* **2006**, *14*, 3062-3068.
28. Zhang, J.Q.; Zhang, Z.W.; Hui, L.; Chen, S.W.; Tian, X. Novel semi synthetic spin-labeled derivatives of podophyllotoxin with cytotoxic and anti-oxidative activity. *Bioorg. Med. Chem.* **2010**, *20*, 983-986.
29. Wall, M.E. Camptothecin and taxol: discovery to clinic. *Med. Res. Rev.* **1998**, *18*(5), 299-314.
30. Saltz, L.B.; Cox, J.V.; Blanke, C.; Rosen, L.S.; Fehrenbacher, L.; Moore, M.J.; Maroun, J.A.; Ackland, S.P.; Locker, P.K.; Pirota, N.; Elfring, G.L.; Miller, L.L.N. Irinotecan plus fluorouracil and leucovorin for metastatic colorectal cancer. Irinotecan Study Group. *Engl. J. Med.* **2000**, *343*(13), 905-914.
31. Gore, M.; Bokkel Huinink, W.; Carmichael, J.; Gordon, A.; Davidson, N.; Coleman, R.; Spaczynski, M.; Heron, J.F.; Bolis, G.; Malmstrom, H.; Malfetano, J.; Acarabelli, C.; Vennin, P.; Ross, G.; Fields, S.Z.J. Clinical evidence for topotecan-paclitaxel non-cross-resistance in ovarian cancer. *Clin. Oncol.* **2001**, *19*(7), 1893-1900.
32. Downing, K.H.; Nogales, E. Tubulin structure: Insights into microtubule properties and functions. *Curr. Opin. Struct. Biol.* **1998**, *8*, 785- 91.
33. Schmidt, M.; Basthians, H. Mitotic drug targets and the development of novel anti-mitotic anticancer drugs. *Drug. Resist. Updat.* **2007**, *10*, 162-168.
34. Georg, G.I.; Harriman, G.C.B.; Himes, R.H.; Mejillano, M.R. 7(p- Azidobenzoyl)-taxol synthesis and biological evaluation. *Bioorg. Med. Chem. Lett.* **1992**, *2*, 735-738.
35. Georg, G.I.; Cheruvallath, Z.S.; Himes, R.H.; Mejillano, M.R. Semi synthesis and biological activity of taxol analogs: Baccatin III 13-(Nbenzoyl-( 2'R,3'S)-3'-(p-tolyl)isoserinate), Baccatin III 13-(N-ptoluoyl)-( 2'R,3'S)-3'-phenylisoserinate), Baccatin III 13-(N-benzoyl-( 2'R,3'S)-3'-(p- trifluoromethylphenyl)isoserinate), and Baccatin III 13-(N-(p trifluoromethylbenzoyl)-(2'R,3'S)-3' phenylisoserinate). *Bioorg. Med. Chem. Lett.* **1992**, *2*, 1751-1754.
36. Longley, R.E.; Caddigan, D.; Harmody, D.; Gunasekera, M.; Gunasekera, S.P. Discodermolide: A new, marine-derived immunosuppressive compound. I. *In vitro* studies. *Transplantation*, **1991**, *52*, 650-656.
37. Ter-Haar, E.; Kowalski, R.J.; Hamel, E.; Lin, C.M.; Longley, R.E.; Gunasekera, S.P. Discodermolide toxic marine agent that stabilizes microtubules more potently than taxol. *Biochemistry*, **1996**, *35*, 243-250.

38. Song, W.; Lei, M.; Zhao, K.; Hu, L.; Meng, Y.; Guo, D. Ceric ammonium nitrate-promoted oxidative coupling reaction for the synthesis and evaluation of a series of anti-tumor amide anhydro vinblastine analogs. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 387-390.
39. Kowalski, R.J.; Giannakakou, P.; Gunasekera, S.P.; Longley, R.E.; Day, B.W.; Hamel, E. The microtubule-stabilizing agent discodermolide competitively inhibits the binding of paclitaxel (Taxol) to tubulin polymers, enhances tubulin nucleation reactions more potently than paclitaxel, and inhibits the growth of paclitaxel-resistant cells. *Mol. Pharmacol.* **1997**, *52*, 613-22.
40. Nicolaou, K.C.; Roshangar, F.; Vourlouinis, D. Chemical biology of the epothilones. *Angew. Chem. Int. Ed.* **1998**, *37*, 2014-2045.
41. Sasse, F. Microtubule binding. *Curr. Biol.* **2000**, *10*, R469-R469.
42. Nakhi, A.; Adepu, R.; Rambabu, D.; Kishore, R.; Vanaja, G.R.; Kalle, A.M.; Pal, M. Thieno [3, 2-c] pyran-4-one based novel small molecules: Their synthesis, crystal structure analysis and *in vitro* evaluation as potential anticancer agents. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 4418-4427.
43. Tietze, L.F. Domino reactions in organic synthesis. *Chem. Rev.* **1996**, *96*, 115-136.
44. Enders, D.; Grondal, C.; Huttel, M.R.M. Asymmetric organocatalytic domino reactions. *Angew. Chem. Int. Ed.* **2007**, *46*, 1570-1581.
45. Kazemi, B.; Javanshir, S.; Maleki, A.; Safari, M.; Khavasi, H.R. An efficient synthesis of 4H-chromene, 4H-pyran, and oxepine derivatives via one-pot three-component tandem reactions. *Tetrahedron Lett.* **2012**, *53*, 6977-6981.
46. Moumou, Y.; Vasseur, J.; Trotin, F.; Dubois, J.; Catechin production by callus cultures of *Fagopyrum esculentum*. *Phytochemistry* **1992**, *31*, 1239-1241.
47. Soria-Mercado, I.E.; Prieto-Davo, A.; Jensen, P.R.; Fenical, W.J. Antibiotic terpenoid chlorodihydroquinones from a new marine actinomycete. *J. Nat. Prod.* **2005**, *68*, 904-910.
48. Nicolaou, K.C.; Pfeifferkorn, J.A.; Cao, G.Q. Selenium-based solid-phase synthesis of benzopyrans I: applications to combinatorial synthesis of natural products. *Angew. Chem. Int. Ed.* **2000**, *39*, 734-739.
49. Gewali, M.B.; Tezuka, Y.; Banskota, A.H.; Ali, M.S.; Saiki, I.; Dong, H.; Kadota, S. Epicalyxin F and calyxin I: two novel antiproliferative diarylheptanoids from the seeds of *Alpinia blepharocalyx*. *Org. Lett.* **1999**, *1*, 1733-1736.
50. Koyama, K.; Takahashi, M.; Oitate, M.; Nakai, N.; Takakusa, H.; Miura, S.; Okazaki, O. CS-8958, a prodrug of the novel neuraminidase inhibitor R-125489, demonstrates a favourable long-retention profile in the mouse respiratory tract. *Antimicrob. Agents Chemother.* **2009**, *53*, 4845-4851.
51. Kiso, M.; Kubo, S.; Ozawa, M.; Le, Q.M.; Nidom, C.A.; Yamashita, M.; Kawaoka, Y. Efficacy of the new neuraminidase inhibitor CS-8958 against H5N1 influenza viruses. *PLoS Pathog.* **2010**, *6*, 1000786.
52. Ferreira, S.B.; da Silva, F.C.; Bezerra, F.A.; Lourenco, M.; Kaiser, C.R.; Pinto, A.C.; Ferreira, V.F. Synthesis of  $\alpha$ - and  $\beta$ -pyran naphthoquinones as a new class of antitubercular agents. *Arch. Pharm.* **2010**, *343*, 81-90.
53. da Rocha, D.R.; de Souza, A.C.; Resende, J.A.; Santos, W.C.; dos Santos, E.A.; C. Pessoa, M.O. de Moraes, L.V. Costa-Lotufo, R.C. Montenegro, V.F. Ferreira, Synthesis of new 9-hydroxy- $\alpha$ - and 7-hydroxy- $\beta$ -pyran naphthoquinones and cytotoxicity against cancer cell lines, *Org. Biomol. Chem.* **2011**, *9*, 4315-4322.
54. Dong, Y.; Shi, Q.; Nakagawa-Goto, K.; Wu, P.C.; Morris-Natschke, S.L.; Brossi, A.; Bastow, K.F.; Lang, J.Y.; Hung, M.C.; Lee, K.H. Antitumor agents 270. Novel substituted 6-Phenyl-4H-furo[3,2-c]pyran-4-one derivatives as potent and highly selective anti-breast cancer agents. *Bioorg. Med. Chem.* **2010**, *18*, 803-808.
55. He, M.Z.; Yang, N.; Sun, C.L.; Yao, X.J.; Yang, M. Modification and biological evaluation of novel 4-hydroxy-pyrone derivatives as non-peptidic HIV-1 protease inhibitors. *Med. Chem. Res.* **2010**, *20*, 200-209.
56. Schiller, R.; Tichotova, L.; Pavlik, J.; Buchta, V.; Melichar, B.; Votruba, I.; Kunes, J.; Spulak, M.; Pour, M. 3, 5-Disubstituted pyranone analogues of highly antifungally active furanones: Conversion of biological effect from antifungal to cytostatic. *Bioorg. Med. Chem. Lett.* **2010**, *20*, 7358-7360.
57. Hussain, H.; Aziz, S.; Schulz, B.; Krohn, K. Synthesis of a 4H-anthra [1, 2-b] pyran derivative and its antimicrobial activity. *Nat. Prod. Commun.* **2011**, *6*, 841-843.
58. Shahrisa, A.; Zirak, M.; Mehdipour, A.R.; Miri, R. Synthesis and calcium channel antagonist activity of new symmetrical and asymmetrical 4-[2-chloro-2-(4-chloro-6-methyl-2-oxo-2H-pyran-3-yl) vinyl]-substituted 1, 4-dihydropyridines. *Chem. Heterocycl. Compd.* **2011**, *46*, 1354-1363.
59. Bisht, S.S.; Jaiswal, N.; Sharma, A.; Fatima, S.; Sharma, R.; Rahuja, N.; Srivastava, A.K.; Bajpai, V.; Kumar, B.; Tripathi, R.P. A convenient synthesis of novel pyranosyl homo-C-nucleosides and their antidiabetic activities. *Carbohydrates Res.* **2011**, *346*, 1191-1201.
60. Wang, S.M.; Milne, G.W.A.; Yan, X.J.; Posey, I.J.; Nicklaus, M.C.; Graham, L.; Rice, W.G. Discovery of novel, non-peptide HIV-1 protease inhibitors by pharmacophore searching. *J. Med. Chem.* **1996**, *39*, 2047-2054.
61. Madda, J.; Venkatesham, A.; Kumar, B.N.; Nagaiah, K.; Sujitha, P.; Kumar, C.G.; Rao, T.P.; Babu, N.J. Synthesis of novel chromeno-annulated cis-fused pyrano[3,4-c]benzopyran and naphtho pyran derivatives via domino aldol-type/hetero Diels-Alder reaction and their cytotoxicity Evaluation. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 4428-4434.

62. Morales, G.A.; Garlich, J.R.; Su, J.; Peng, X.; Newblom, J.; Weber, K.; Durden, D.L. Synthesis and cancer stem cell-based activity of substituted 5-morpholino-7H-thieno[3,2-b]pyran-7-ones designed as next generation PI3K inhibitors. *J. Med. Chem.* **2013**, *56*, 1922–1939.
63. Siripong, P.; Kanokmedakul, K.; Piyaviriyagul, S.; Yahuaifai, J.; Chanpai, R.; Ruchirawat, S.; Oku, N. Antiproliferative naphthoquinone esters from *Rhinacanthus nasutus* Kurz. roots on various cancer cells. *J. Trad. Med.* **2006**, *23*, 166–172.
64. Frolova, L.V.; Magedov, I.V.; Romero, A.E.; Karki, M.; Otero, I.; Hayden, K.; Evdokimov, N.M.; Banuls, L.M.Y.; Rastogi, S.K.; Smith, W.R.; Lu, S.L. Structural simplification of bioactive natural products with multicomponent synthesis. 4. 4H-Pyrano-[2,3-b] naphthoquinones with anticancer activity. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 5195–5198.
65. Thi, T.A.D.; Thi, T.H.V.; Phuong, H.T.; Nguyen, T.H.; Duc, C.V.; Depetter, Y.; Van Nguyen, T.; D'hooghe, M. Synthesis and anticancer properties of new (dihydro) pyranonaphthoquinones and their epoxy analogs. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 3355–3358.
66. Kowalski, K.; Chyła, A.K.; Szczupak, L.; Hikisz, P.; Bernasińska, J.; Rajnisz, A.; Solecka, J.; Therrien, B. Ferrocenylvinyl-flavones: Synthesis, structure, anticancer and antibacterial activity studies. *J. Organomet. Chem.* **2013**, *741*, 153–161.
67. Huang, W.Y.; Cai, Y.Z. Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention. *Nutr. Cancer* **2010**, *62*, 1–20.
68. De Groot, H.; Rauen, U. Tissue injury by reactive oxygen species and the protective effects of flavonoids. *Fundam. Clin. Pharmacol.* **1998**, *12*, 249–255.
69. Middleton, J.E. Effect of plant flavonoids on immune and inflammatory cell function. *Adv. Exp. Med. Biol.* **1998**, *439*, 175–182.
70. Yoon, J.S.; Lee, M.K.; Sung, S.H.; Kim, Y.C. Neuroprotective 2- (2 phenylethyl) chromones of *Imperata cylindrical*. *J. Nat. Prod.* **2006**, *69*, 290–291.
71. Neuhouwer, M.L. Dietary flavonoids and cancer risk: evidence from human population studies. *Nutr. Cancer* **2004**, *50*, 1–7.
72. Min, L.W. New therapeutic aspects of flavones: the anticancer properties of *Scutellaria* and its main active constituents Wogonin, Baicalein and Baicalin. *Cancer Treat. Rev.* **2009**, *35*, 57–68.
73. Akao, Y.; Itoh, T.; Ohguchi, K.; Iinuma, M.; Nozawa, Y. Interactive effects of polymethoxy flavones from Citrus on cell growth inhibition in human neuroblastoma SH-SY5Y cells. *Bioorg. Med. Chem.* **2008**, *16*, 2803–2810.
74. Mays, J.R.; Hill, S.A.; Moyers, J.T.; Blagg, B.S. The synthesis and evaluation of flavone and isoflavone chimeras of novobiocin and derrubone. *Bioorg. Med. Chem.* **2010**, *18*, 249–266.
75. Hsiao, Y.C.; Hsieh, Y.S.; Kuo, W.H.; Chiou, H.L.; Yang, S.F.; Chiang, W.L.; Chu, S.C.; The tumor-growth inhibitory activity of flavanone and 2'-OH flavanone *in vitro* and *in vivo* through induction of cell cycle arrest and suppression of cyclins and CDKs. *J. Biomed. Sci.* **2007**, *14*, 107–119.
76. Choi, E.J.; Lee, J.I.; Kim, G.H. Anti-carcinogenic effect of a new analogue 4'-chloroflavanone from flavanone in human breast cancer cells. *Int. J. Mol. Med.* **2010**, *25*, 293–298.
77. Choi, E.J.; Lee, J.I.; Kim, G.H. Effects of 4',7-dimethoxyflavanone on cell cycle arrest and apoptosis in human breast cancer MCF-7 cells. *Arch. Pharm. Res.* **2011**, *34*, 2125–2130.
78. Usman, H.; Hakim, E.H.; Harlim, T.; Jalaluddin, M.N.; Syah, Y.M.; Achmad, S.A.; Takayama, H. Cytotoxic chalcones and flavanones from the tree bark of *Cryptocarya costata*. *Naturforsch., C. J. Biosci.* **2006**, *61*, 184–188.
79. Shen, S.C.; Ko, C.H.; Tseng, S.W.; Tsai, S.H.; Chen, Y.C. Structurally related antitumor effects of flavanones in vitro and in vivo: involvement of caspase 3 activation, p21 gene expression, and reactive oxygen species production. *Toxicol. Appl. Pharmacol.* **2004**, *197*, 84–95.
80. Liao, S.Y.; Chen, J.C.; Qian, L.; Shen, Y.; Zheng, K.C. QSAR, action mechanism and molecular design of flavone and isoflavone derivatives with cytotoxicity against HeLa. *Eur. J. Med. Chem.* **2008**, *43*, 2159–70.
81. Hirunuma, M.; Shoyama, Y.; Sasaki, K.; Sakamoto, S.; Taura, F.; Shoyama, Y.; Tanaka, H.; Morimoto, S. Flavone-catalyzed apoptosis in *Scutellaria baicalensis*. *Phytochemistry* **2011**, *72*, 752–60.
82. Switalska, M.; Gryniewicz, G.; Strzadala, L.; Wietrzyk, J. Novel genistein derivatives induce cell death and cell cycle arrest through different mechanisms. *Nutr. Cancer* **2013**, *65*, 874–884.
83. Murti, Y.; Mishra, P. Synthesis and evaluation of flavanones as anticancer agents. *Indian J. Pharm. Sci.* **2014**, *76*, 163–166.
84. Kumar, D.; Singh, O.; Nepali, K.; Bedi, P.M.S.; Qayum, A.; Singh, S.; Jain, S.K. Naphthoflavones as antiproliferative agents: design, synthesis and biological evaluation. *Anti-Cancer Agent Med. Chem.* **2016**, *16*, 881–890.
85. Aghdassi, A.; Phillips, P.; Dudeja, V.; Dhaulakhandi, D.; Sharif, R.; Dawra, R.; Lerch, M.M.; Saluja, S. Heat shock protein 70 increases tumorigenicity and inhibits apoptosis in pancreatic adenocarcinoma. *Cancer Res.* **2007**, *67*, 616–625.

86. Mouria, M.; Gukovskaya, A.S.; Jung, Y.; Buechler, P.; Hines, O.J.; Reber, H.A.; Pandol, S.J. Food-derived polyphenols inhibit pancreatic cancer growth through mitochondrial cytochrome C release and apoptosis. *Int. J. Canc.* **2002**, *98*, 761-769.
87. Xue, W.; Song, B.A.; Zhao, H.J.; Qi, X.B.; Huang, Y.J.; Liu, X.H., Novel myricetin derivatives: Design, synthesis and anticancer activity *Eur. J. Med. Chem.* **2015**, *97*, 155-163.
88. Safavi, M.; Esmati, N.; Ardestani, S.K.; Emami, S.; Ajdari, S.; Davoodi, J.; Shafiee, A.; Foroumadi, A. Halogenated flavanones as potential apoptosis-inducing agents: synthesis and biological activity evaluation. *Eur. J. Med. Chem.* **2012**, *58*, 573-80.
89. Al-Kawkabani, A.; Boutemour-Kheddis, B.; Makhoulfi-Chebli, M.; Hamdi, M.; Talhi, O.; Silva, A.M.; Synthesis of novel 2H,8H-pyrano[2,3-f]chromene-2,8-diones from 8-formyl-7-hydroxy-4-methylcoumarin. *Tetrahedron Lett.* **2013**, *54*, 5111-5114.
90. Shi, Y., Zhou, C. Synthesis and evaluation of a class of new coumarin triazole derivatives as potential antimicrobial agents. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 956-960
91. Hur, S.Y.; Kim, T.E.; Park, Y.G.; Kim, J.R.; Kim, J.W. Natural compounds, fraxin and chemicals structurally related to fraxin protect cells from oxidative stress. *Exp. Mol. Med.* **2005**, *37*, 436-446.
92. Devji, T.; Reddy, C.; Woo, C.; Awale, S.; Kadota, S.; Carrico-Moniz, D. Pancreatic anticancer activity of a novel geranylgeranylated coumarin derivative. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 5770-5773.
93. Reddy, N.S.; Mallireddigari, M.R.; Cosenza, S.; Gumireddy, K.; Bell, S.C. Reddy, E.P.; Reddy, M.V. Synthesis of new coumarin 3-(N-aryl) sulfonamides and their anticancer activity. *Bioorg. Med. Chem. Lett.* **2004**, *14*, 4093-4097.
94. Manvar, A.; Bavishi, A.; Radadiya, A.; Patel, J.; Vora, V.; Dodia, N.; Rawal, K.; Shah, A. Diversity oriented design of various hydrazides and their *in vitro* evaluation against *Mycobacterium tuberculosis* H37 Rv strains. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 4728-4731.
95. Xue, H.; Lu, X.; Zheng, P.; Liu, L.; Han, C.; Hu, J.; Liu, Z.; Ma, T.; Li, Y.; Wang, L.; Chen, Z.; Liu, G. Highly suppressing wild-type HIV-1 and Y181C mutant HIV-1 strains by 10-chloromethyl-11-demethyl-12-oxo-calanolide A with druggable profile. *J. Med. Chem.* **2010**, *53*, 1397-1401.
96. Yeh, J.Y.; Coumar, M.S.; Horng, J.T.; Shiao, H.Y.; Kuo, F.M.; Lee, H.L.; Chen, I.C.; Chang, C.W.; Tang, W.F.; Tseng, S.N.; Chen, C.J. Anti-Influenza drug discovery: structure-activity relationship and mechanistic insight into novel angelicin derivatives. *J. Med. Chem.* **2010**, *53*, 1519-1533.
97. Gonzales, B.S.P.; Rodriguez B.J.C. Synthesis of collinin, an antiviral coumarin. *Aust. J. Chem.* **2003**, *56*, 59-60.
98. Anand P.; Singh, B.; Singh, N. review on coumarins as acetylcholinesterase inhibitors for Alzheimer's disease. *Bioorg. Med. Chem.* **2012**, *20*, 1175-1180.
99. Piazzzi, L.; Cavalli, A.; Colizzi, F.; Belluti, F.; Bartolini, M.; Mancini, F.; Recanatini, M.; Andrisano, V.; Rampa, A. Multi-target-directed coumarin derivatives: hAChE and BACE1 inhibitors as potential anti-Alzheimer compounds. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 423-426.
100. Shi, Y., Zhou, C. Synthesis and evaluation of a class of new coumarin triazole derivatives as potential antimicrobial agents. *Bioorg. Med. Chem. Lett.* **2011**, *21*, 956-960
101. Lin, C.M.; Huang, S.T.; Lee, F.W.; Kuo, H.S.; Lin, M.H. 6-Acyl-4-aryl/alkyl-5, 7-dihydroxycoumarins as anti-inflammatory agents. *Bioorg. Med. Chem.* **2006**, *14*, 4402-4409
102. Nepali, K.; Sharma, S.; Kumar, D.; Budhiraja, A.; Dhar, K.L. Anticancer Hybrids- A Patent Survey. *Recent Pat. Anticancer Drug Discov.* **2014**, *9*, 303-339.
103. Amin, K.M.; Eissa, A.A.; Abou-Seri, S.M.; Awadallah, F.M.; Hassan, G.S. Synthesis and biological evaluation of novel coumarin-pyrazoline hybrids endowed with phenylsulfonyl moiety as antitumor agents. *Eur. J. Med. Chem.* **2013**, *60*, 187-198.
104. Belluti, F.; Fontana, G.; Dal Bo, L.; Carenni, N.; Giommarelli, C.; Zunino, F. Design, synthesis and anticancer activities of stilbene-coumarin hybrid compounds: Identification of novel proapoptotic agents. *Bioorg. Med. Chem.* **2010**, *18*, 3543-3550.
105. Paul, K.; Bindal, S.; Luxami, V. Synthesis of new conjugated coumarin-benzimidazole hybrids and their anticancer activity. *Bioorg. Med. Chem.* **2013**, *23*, 3667-3672.
106. Sashidhara, K.V.; Kumar, A.; Kumar, M.; Sarkar, J.; Sinha, S. Synthesis and in vitro evaluation of novel coumarin-chalcone hybrids as potential anticancer agents. *Bioorg. Med. Chem.* **2010**, *20*, 7205-7211.
107. Sashidhara, K.V.; Avula, S.R.; Sharma, K. Palnati, G.R.; Bathula, S.R. Discovery of coumarin-monastrol hybrid as potential antitumor tumor-specific agent. *Eur. J. Med. Chem.* **2013**, *60*, 120-127.
108. Bagdi, A.K.; Majee, A.; Hajra, A. Regioselective synthesis of pyrano[3,2-c]coumarins via Cu(II)-catalyzed tandem reaction. *Tetrahedron Lett.* **2013**, *54*, 3892-3895.
109. Kumar, D.; Malik, F.; Bedi, P.M.S.; Jain, S. 2,4-Diarylpyrano[3,2-c]chromen-5(4H)-ones as antiproliferative agents: design, synthesis and biological evaluation. *Chem. Pharm. Bull.* **2016**, *64*, 399-409.
110. Hussain, M.K.; Ansari, M.I.; Yadav, N.; Gupta, P.K.; Gupta, A.K.; Saxena, R.; Fatima, I.; Manohar, M.; Kushwaha, P.; Khedgikar, V. Design and synthesis of ERa/ERb selective coumarin and chromene derivatives as potential antitumor cancer and anti-osteoporotic agents. *RSC Adv.* **2014**, *4*, 8828-8845.



111. Ganina, O.G.; Daras, E.; Bourgarel-Rey, V.; Peyrot, V.; Andresyuk, A.N.; Finet, J.P.; Fedorov, A.Y.; Beletskaya, I.P.; Combes, S. Synthesis and biological evaluation of polymethoxylated 4-heteroaryl coumarins as tubulin assembly inhibitor. *Bioorg. Med. Chem.* **2008**, *16*, 8806–8812.
112. Arshad, A.; Osman, H.; Bagley, M.C.; Lam, C.K.; Mohamad, S.; Zahariluddin, A.S.M.; Synthesis and antimicrobial properties of some new thiazolyl coumarin derivatives. *Eur. J. Med. Chem.* **2011**, *46*, 3788–3794.
113. Gali, R.; Banothu, J.; Gondru, R.; Bavantula, R.; Velivela, Y.; Crooks, P.A. One-pot multicomponent synthesis of indole incorporated thiazolyl coumarins and their antibacterial, anticancer and DNA cleavage studies. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 106–112.
114. Kurt, B.Z.; Gazioglu, I.; Sonmez, F.; Kucukislamoglu, M. Synthesis, antioxidant and anticholinesterase activities of novel coumaryl thiazole derivatives. *Bioorg. Chem.* **2015**, *59*, 80–90.
115. Delogu, G.; Picciau, C.; Ferino, G.; Quezada, E.; Podda, G.; Uriarte, E.; Vina, D. Synthesis, human monoamine oxidase inhibitory activity and molecular docking studies of 3-heteroaryl coumarin derivatives. *Eur. J. Med. Chem.* **2011**, *46*, 1147–1152.
116. Garazd, Y.; Garazd, M.; Lesyk, R. Synthesis and evaluation of anticancer activity of 6-pyrazolinyl coumarin derivatives. *Saudi Pharm J* **2017**, *25*(2), 214–223.
117. Zhao, H.; Donnelly, A.C.; Kusuma, B.R.; Brandt, G.E.; Brown, D.; Rajewski, R.A.; Blagg, B.S. Engineering an antibiotic to fight cancer: optimization of the novobiocin scaffold to produce anti-proliferative agents. *J. Med. Chem.* **2011**, *54*, 3839–3853.
118. Siddiqui, Z.N.; TN, M.M.; Ahmad, A.; Khan, A.U. Synthesis of 4-Hydroxycoumarin Heteroarylhybrids as Potential Antimicrobial Agents. *Arch. Pharm.* **2011**, *344*, 394–401.
119. Kusuma, B.R.; Peterson, L.B.; Zhao, H.; Vielhauer, G. Holzbeierlein, J.; Blagg, B.S. Targeting the heat shock protein 90 dimer with dimeric inhibitors. *J. Med. Chem.* **2011**, *54*, 6234–6253.
120. Burlison, J.A.; Blagg, B.S.J. Synthesis and evaluation of coumermycin A1 analogues that inhibit the Hsp90 protein folding machinery. *Org. Lett.* **2006**, *8*, 4855–4858.
121. Tan, G.; Yao, Y.; Gu, Y.; Li, S.; Lv, M.; Wang, K.; Li, X. Cytotoxicity and DNA binding property of the dimers of triphenylethylene–coumarin hybrid with one amino side chain. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 2825–2830.
122. Zhu, M.; Zhou, L.K.; Yao, Y.C.; Li, S.; Lv, M.; Wang, K.; Li, X.; Chen, H. Anticancer activity and DNA binding property of the dimers of triphenylethylene–coumarin hybrid with two amino side chains. *Med. Chem. Res.* **2015**, *24*, 2314–2324.
123. Zhao, L.; Yao, Y.C.; Lv, S.M.; Chen, H.; Li, X. Cytotoxicity and DNA binding property of triphenylethylene–coumarin hybrids with two amino side chains. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 900–904.
124. Chen, H.; Li, S.; Yao, Y.; Zhou, L.; Zhao, J.; Gu, Y.; Li, X. Design, synthesis, and anti-tumor activities of novel triphenylethylene–coumarin hybrids, and their interactions with Ct-DNA. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 4785–4789.
125. Zhang, L.; Yao, Y.C.; Gao, M.Y.; Rong, R.X.; Wang, K.R.; Li, X.L.; Chen, H. Anticancer activity and DNA binding property of the trimers of triphenylethylene–coumarin hybrids. *Chin. Chem. Lett.* **2016**, *27*(11), 1708–1716.
126. Kupchan, S.M.; Streelman, D.R.; Sneden, A.T. Psorospermin, a new antileukemic xanthone from *Psorospermum febrifugum*. *J. Nat. Pro.* **1980**, *43*, 296–301.
127. Winter, D.K.; Sloman, D.L.; Porco Jr, J.A. Polycyclic xanthone natural products: structure, biological activity and chemical synthesis. *J. Nat. Pro. Rep.* **2013**, *30*, 382–391.
128. Lin, C.N.; Liou, S.J.; Lee, T.H.; Chuang, Y.C.; Won, S.J. Xanthone derivatives as potential anti-cancer drugs. *J. Pharm. Pharmacol.* **1996**, *48*, 539–544.
129. Chen, C.A.; Yeh, R.H.; Lawrence, D.S. Design and synthesis of a fluorescent reporter of protein kinase activity. *J. Am. Chem. Soc.* **2002**, *124*, 3840–3841.
130. Steiner, L.F.; Summerland, S.A.; Xanthone as an ovicide and larvicide for the codling moth. *J. Econ. Entomol.* **1943**, *36*, 435–439.
131. Cheng, J.H.; Huang, A.M.; Hour, T.C.; Yang, S.C.; Pu, Y.S.; Lin, C.N. Antioxidant xanthone derivatives induce cell cycle arrest and apoptosis and enhance cell death induced by cisplatin in NTUB1 cells associated with ROS. *Eur. J. Med. Chem.* **2011**, *46*, 1222–1231.
132. Chase, M. Angiosperm Phylogeny Group. *Bot. J. Lin. Soc.* **2003**, *141*, 399–436.
133. Lee, K.H.; Chai, H.B.; Tamez, P.A.; Pezzuto, J.M.; Cordell, G.A.; Win, K.K.; Tin Wa, M. Biologically active alkylated coumarins from *Kayea assamica*. *Phytochem.* **2003**, *64*, 535–539.
134. Laphookhieo, S.; Syers, J.K.; Kiattansakul, R.; Chantapromma, K. Cytotoxic and antimalarial prenylated xanthenes from *Cratoxylum cochinchinense*. *Chem. Pharm. Bull.* **2006**, *54*, 745–747.
135. Wu, C.P.; Van Schalkwyk, D.A.; Taylor, D.; Smith, P.J.; Chibale, K.; Reversal of chloroquine resistance in *Plasmodium falciparum* by 9H-xanthene derivatives. *Int. J. Antimicrob. Agent* **2005**, *26*, 170–175.
136. Campos-Esparza, M.R.; Sanchez-Gomez, M.V.; Matute, C.; Molecular mechanisms of neuroprotection by two natural antioxidant polyphenols. *Cell Cal.* **2009**, *45*, 358–368.



137. Chibale, K.; Visser, M.; Schalkwyk, D.; Smith, P.J.; Saravanamuthu, A.; Fairlamb, A.H. Exploring the potential of xanthene derivatives as trypanothione reductase inhibitors and chloroquine potentiating agents. *Tetrahedron*, **2003**, *59*, 2289-2296.
138. Martinez, A.; Galano, A.; Vargas, R. Free radical scavenger properties of  $\alpha$ -mangostin: thermodynamics and kinetics of HAT and RAF mechanisms. *J. Phys. Chem.* **2011**, *115*, 12591-12598.
139. Pedraza-Chaverri, J.; Reyes-Fermin, L.M.; Nolasco-Amaya, E.G.; Orozco-Ibarra, M.; Medina-Campos, O.N.; Gonzalez-Cuahutencos, O.; Rivero-Cruz, I.; Mata, R. ROS scavenging capacity and neuroprotective effect of  $\alpha$ -mangostin against 3-nitropropionic acid in cerebellar granule neurons. *Exp. Toxicol. Pathol.* **2009**, *61*, 491-501.
140. Buelna-Chontal, M.; Correa, F.; Hernandez-Resendiz, S.; Zazueta, C.; Pedraza-Chaverri, J. Protective effect of  $\alpha$ -mangostin on cardiac reperfusion damage by attenuation of oxidative stress. *J. Med. Food* **2011**, *14*, 1370-1374.
141. Reyes-Fermin, L.M.; González-Reyes, S.; Tarco-Álvarez, N.G.; Hernández-Nava, M.; Orozco-Ibarra, M.; Pedraza-Chaverri, J. Neuroprotective effect of  $\alpha$ -mangostin and curcumin against iodoacetate-induced cell death. *Nutr. Neurosci.* **2012**, *15*, 34-41.
142. Schwaebe, M.K.; Moran, T.J.; Whitten, J.P. Total synthesis of psorospermin. *Tetrahedron Lett.* **2005**, *46*, 827-829.
143. Matsumoto, K.; Akao, Y.; Yi, H.; Ohguchi, K.; Ito, T.; Tanaka, T.; Kobayashi, E.; Iinuma, M.; Nozawa, Y. Preferential target is mitochondria in  $\alpha$ -mangostin-induced apoptosis in human leukemia HL60 cells. *Bioorg. Med. Chem.* **2004**, *12*, 5799-5806.
144. Pedro, M.; Cerqueira, F.; Sousa, M.E.; Nascimento, M.S.J.; Pinto, M. Xanthonenes as inhibitors of growth of human cancer cell lines and their effects on the proliferation of human lymphocytes *in vitro*. *Bioorg. Med. Chem.* **2002**, *10*, 3725-3730.
145. Gnerre, C.; Thull, U.; Gaillard, P.; Carrupt, P.A.; Testa, B.; Fernandes, E.; Silva, F.; Pinto, M.; Wolfender, J.L.; Hostettmann, K.; Cruciani, G. Natural and synthetic xanthonenes as monoamine oxidase inhibitors: Biological assay and 3D-QSAR. *Helv. Chim. Acta* **2001**, *84*, 552-570.
146. Poondru, S.; Zhou, S.; Rake, J.; Shackleton, G.; Corbett, T.H.; Parchment, R.E.; Jasti, B.R.; High-performance liquid chromatographic method for the estimation of the novel investigational anti-cancer agent SR271425 and its metabolites in mouse plasma. *J. Chromatogr. B. Biomed. Sci. Appl.* **2001**, *759*, 175-178.
147. Chantarasriwong, O.; Cho, W.C.; Batova, A.; Chavasiri, W.; Moore, C.; Rheingold, A.L.; Theodorakis, E.A. Evaluation of the pharmacophoric motif of the caged *Garcinia* xanthonenes. *Org. Biomol. Chem.* **2009**, *7*, 4886-4894.
148. Matsumoto, K.; Akao, Y.; Kobayashi, E.; Ohguchi, K.; Ito, T.; Tanaka, T.; Iinuma, M.; Nozawa, Y. Induction of apoptosis by xanthonenes from mangosteen in human leukemia cell lines. *J. Nat. Prod.* **2003**, *66*, 1124-1127.
149. Chiang, L.C.; Cheng, H.Y.; Liu, M.C.; Chiang, W.; Lin, C.C. In vitro evaluation of antileukemic activity of 17 commonly used fruits and vegetables in Taiwan. *Lebensm Wiss Technol.* **2004**, *37*, 539-544.
150. Balunas, M.J.; Su, B.; Brueggemeier, R.W.; Kinghorn, A.D. Xanthonenes from the botanical dietary supplement mangosteen (*Garcinia mangostana*) with aromatase inhibitory activity. *J. Nat. Prod.* **2008**, *71*, 1161-1166.
151. Jung, H.A.; Su, B.N.; Keller, W.J.; Mehta, R.G.; Kinghorn, A.D. Antioxidant xanthonenes from the pericarp of *Garcinia mangostana* (Mangosteen). *J. Agric. Food Chem.* **2006**, *54*, 2077-2082.
152. Suksamrarn, S.; Komutiban, O.; Ratananukul, P.; Chimnoi, N.; Lartpornmatulee, N.; Suksamrarn, A. Cytotoxic prenylated xanthonenes from the young fruit of *Garcinia mangostana*. *Chem. Pharm. Bull.* **2006**, *54*, 301-305.
153. Chen, L.G.; Yang, L.L.; Wang, C.C. Anti-inflammatory activity of mangostins from *Garcinia mangostana*. *Food Chem. Toxicol.* **2008**, *46*, 688-693.
154. Watanapokasin, R.; Jarinthan, F.; Jerusalmi, A.; Suksamrarn, S.; Nakamura, Y.; Sukserree, S.; Uthaisang-Tanethpongamb, W.; Ratananukul, P.; Sano, T. Potential of xanthonenes from tropical fruit mangosteen as anti-cancer agents: caspase-dependent apoptosis induction *in vitro* and in mice. *Appl. Biochem. Biotechnol.* **2010**, *162*, 1080-1094.
155. Aisha, A.F.; Abu-Salah, K.M.; Ismail, Z.; Majid, A.M.S.A. *In vitro* and *in vivo* anti-colon cancer effects of *Garcinia mangostana* xanthonenes extract. *BMC Complementary Altern. Med.* **2012**, *12*, 1-10.
156. Kosem, N.; Ichikawa, K.; Utsumi, H.; Moongkarndi, P. *In vivo* toxicity and antitumor activity of mangosteen extract. *J. Nat. Med.* **2013**, *67*, 255-263.
157. Kim, S.J.; Hong, E.H.; Lee, B.R.; Park, M.H.; Kim, J.W.; Pyun, A.R.; Kim, Y.J.; Chang, S.Y.; Chin, Y.W.; Ko, H.J.  $\alpha$ -Mangostin reduced ER stress-mediated tumor growth through autophagy activation. *Immune Netw.* **2012**, *12*, 253-260.
158. Cao, S.; Brodie, P.J.; Miller, J.S.; Randrianaivo, R.; Ratovoson, F.; Birkinshaw, C.; Andriantsiferana, R.; Rasamison, V.E.; Kingston, D.G. Antiproliferative xanthonenes of *Terminalia calcicola* from the Madagascar Rain Forest. *J. Nat. Prod.* **2007**, *70*, 679-681.

159. Han, Q.B.; Tian, H.L.; Yang, N.Y.; Qiao, C.F.; Song, J.Z.; Chang, D.C.; Luo, K.Q.; Xu, H.X. Polyprenylated xanthenes from *Garcinia lancilimba* showing apoptotic effects against HeLa-C3 Cells. *Chem. Biodiver.* **2008**, *5*, 2710-2717.
160. Tao, S.J.; Guan, S.H.; Wang, W.; Lu, Z.Q.; Chen, G.T.; Sha, N.; Yue, Q.X.; Liu, X.; Guo, D.A. Cytotoxic polyprenylated xanthenes from the resin of *Garcinia hanburyi*. *J. Nat. Prod.* **2009**, *72*, 117-124.
161. Han, Q.B.; Xu, H.X. Caged *Garcinia* xanthenes: development since 1937. *Curr. Med. Chem.* **2009**, *16*, 3775-3796.
162. Mu, R.; Lu, N.; Wang, J.; Yin, Y.; Ding, Y.; Zhang, X.; Gui, H.; Sun, Q.; Duan, H.; Zhang, L.; Zhang, Y. An oxidative analogue of gambogic acid-induced apoptosis of human hepatocellular carcinoma cell line HepG2 is involved in its anticancer activity *in vitro*. *Eur. J. Canc. Prev.* **2010**, *19*, 61-67.
163. Zhou, W.; Cai, B.; Shan, J.; Wang, S.; Di, L. Discovery and current status of evaluation system of bioavailability and related pharmaceutical technologies for Traditional Chinese Medicines—Flos Lonicerae Japonicae Fructus Forsythiae herb couples as an example. *AAPS Pharm. Sci. Tech.* **2015**, *16*, 28812-28840.
164. Jang, S.W.; Okada, M.; Sayeed, I.; Xiao, G.; Stein, D.; Jin, P.; Ye, K. Gambogic amide, a selective agonist for TrkA receptor that possesses robust neurotrophic activity, prevents neuronal cell death. *Proc. Natl. Acad. Sci.* **2007**, *104*, 16329-16334.
165. Zelefsack, F.; Guilet, D.; Fabre, N.; Bayet, C.; Chevalley, S.; Ngouela, S.; Lenta, B.N.; Valentin, A.; Tsamo, E.; Dijoux-Franca, M.G. Cytotoxic and antiplasmodial xanthenes from *Pentadesma butyracea*. *J. Nat. Prod.* **2009**, *72*, 954-957.
166. Moosophon, P.; Kanokmedhakul, S.; Kanokmedhakul, K.; Soyong, K. Prenylxanthenes and a bicyclo [3.3.1] nona-2, 6-diene derivative from the fungus *Emericella rugulosa*. *J. Nat. Prod.* **2009**, *72*, 1442-1446.
167. Bhattacharya, A.K.; Rana, K.C.; Mujahid, M.; Sehar, I.; Saxena, A.K.; Synthesis and *in vitro* study of 14-aryl-14H-dibenzo[a,j]xanthenes as cytotoxic agents. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 5590-5593.
168. Niu, S.L.; Li, Z.L.; Ji, F.; Liu, G.Y.; Zhao, N.; Liu, X.Q.; Jing, Y.K.; Hua, Y.M. Xanthenes from the stem bark of *Garcinia bracteata* with growth inhibitory effects against HL-60 cells. *Phytochemistry*, **2012**, *77*, 280-286.
169. Loetchutin, C.; Chau, F.; Mankhetkorn, S. Synthesis and evaluation of 5-Aryl-3-(4-hydroxyphenyl)-1,3,4-oxadiazole-2-(3H)-thiones as P-glycoprotein inhibitors. *Chem. Pharm. Bull.* **2003**, *51*(6), 728-730.
170. Abadi, A.H.; Eissa, A.A.; Hassan, G.S. Synthesis of novel 1,3,4-trisubstituted pyrazole derivatives and their evaluation as antitumor and antiangiogenic agents. *Chem. Pharm. Bull.* **2003**, *51*(7), 838-844.
171. Szczepankiewicz, B.G.; Liu, G.; Jae, H.S.; Tasker, A.S.; Gunawardana, I.W.; Geldern, T.W.V.; Gwaltney, S.L.; Wu-Wong, J.R.; Gehrke, L.; Chiou, W.J.; Credo, R.B.; Alder, J.D.; Nukkala, M.A.; Zielinski, N.A.; Jarvis, K.; Mollison, K.W.; Frost, D.J.; Bauch, J.L.; Hui, Y.H.; Claiborne, A.K.; Li, Q.; Rosenberg, S.H. New antimitotic agents with activity in multi-drug-resistant cell lines and *in vivo* efficacy in murine tumor models. *J. Med. Chem.* **2001**, *44*(25), 4416-4430.
172. Kumar, D.; Sundaree S.; Johnson, E.O.; K. Shah. An efficient synthesis and biological study of novel indolyl-1,3,4-oxadiazoles as potent anticancer agents. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4492-4494.
173. Aboraia, A.S.; Abdel-Rahman, H.M.; Mahfouz, N.M.; El-Gendy, M.A. Novel 5-(2-hydroxyphenyl)-3-substituted-2,3-dihydro-1,3,4-oxadiazole-2-thione derivatives: promising anticancer agents. *Bioorg. Med. Chem.* **2006**, *14*(4), 1236-1246.
174. Khan, M.T.; Choudhary M.I.; Khan, K.M.; Rani, M.; Rahman, A.U.; Structure-activity relationships of tyrosinase inhibitory combinatorial library of 2,5-disubstituted-1,3,4-oxadiazole analogues. *Bioorg. Med. Chem.* **2005**, *13*, 3385-3395.
175. Tan, T.M.C.; Chen, Y.; Kong, K.H.; Bai, J.; Li, Y.; Lim, S.G.; Ang, T.H.; Lam, Y. Synthesis and the biological evaluation of 2-benzenesulfonylalkyl-5-substituted-sulfanyl-[1,3,4]-oxadiazoles as potential anti-hepatitis B virus agents. *Antiviral Res.* **2006**, *71*, 7-14.
176. Li, Y.; Liu, J.; Zhang, H.; Yang, X.; Liu, Z. Stereo selective synthesis and fungicidal activities of (E)-alpha-(methoxyimino)-benzene acetate derivatives containing 1,3,4-oxadiazole ring. *Bioorg. Med. Chem. Lett.* **2006**, *16*(8), 2278-2282.
177. Warener, R.N. New Adventures in the Synthesis of Hetero-Bridged *syn*-Facially Fused Norbornadienes ("[n]Polynorbornadienes") and Their Topological Diversity. *Eur. J. Org. Chem.* **2000**, *65*, 3363-3380.
178. Guan, M.; Bian, Z.Q.; Zhou, Y.F.; Li, F.Y.; Li, Z.L.; Huang, C.H. High-performance blue electroluminescent devices based on 2-(4-biphenyl)-5-(4-carbazole-9-yl)phenyl-1,3,4-oxadiazole. *Chem. Commun.* **2003**, *9*, 2708-2709.
179. Guimaraes, C.R.W.; Boger, D.L.; Jorgensen, W.L. Elucidation of fatty acid amide hydrolase inhibition by potent  $\alpha$ -ketoheterocycle derivatives from Monte Carlo simulations. *J. Am. Chem. Soc.* **2005**, *127*(49), 17377-17384.
180. Sankhe, N.M.; Durgashivaprasad, E.; Kutty, N.G. Rao, J.V.; Narayanan, K.; Kumar, N.; Raj, P.V. Novel 2, 5-disubstituted-1, 3, 4-oxadiazole derivatives induce apoptosis in HepG2 cells through p53 mediated intrinsic pathway. *Arab. J. Chem.* **2019**, *12*, 2548-2555.

181. Mahmoud, M.; Gamal, E.D.; Mohammed, I.E.G.; Mohammed, S.; Abdel, M.; Kyung, H. Y.; Chang-Hyun, O. Synthesis and in vitro anti-proliferative activity of new 1,3,4-oxadiazole derivatives possessing sulfonamide moiety. *Eur. J. Med. Chem.* **2015**, *90*, 45-52.
182. Qian-Ru, D.; Dong-Dong, L.; Ya-Zhou, P.; Jing-Ran, L.; Jian, S.; Fei, F.; Wei-Qing, Z.; Gong, H.B.; Zhu, H.L. Novel 1,3,4-oxadiazole thioether derivatives targeting thymidylate synthase as dual anticancer/antimicrobial agents. *Bioorg. Med. Chem.* **2013**, *21*, 2286-2297.
183. Dalip, K.; Swapna, S.; Johnson, E.O.; Kavita, S. An efficient synthesis and biological study of novel indolyl-1,3,4-oxadiazoles as potent anticancer agents. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4492-4494.
184. Ahmed, S.; Aboraia, H.M.; Rahman, A.; Nadia, M.; Mahmoud, A.; Gendy, E.L. Novel 5-(2-hydroxyphenyl)-3-substituted-2,3-dihydro-1,3,4-oxadiazole-2-thione derivatives: Promising anticancer agents. *Bioorg. Med. Chem.* **2006**, *14*, 1236-1246.
185. Samir, B.; Shymaa, A.; Hassan, A.; Etman, F.; Badria, A. Synthesis and antitumor evaluation of some new 1,3,4-oxadiazole-based heterocycles. *Eur. J. Med. Chem.* **2012**, *48*, 192-199.
186. Clitherow, J.W.; Beswick, P.; Irving, W.J.; Scopes, D.I.C.; Barnes, J.C.; Clapham, J.; Brown, J.D.; Evans, D.J.; Hayes, A.G. Novel 1, 2, 4-oxadiazoles as potent and selective histamine H3 receptor antagonists. *Bioorg. Med. Chem. Lett.* **1996**, *6*, 833-838.
187. Acri, J.B.; Wong, G.; Witkin, J.M. Stereospecific transduction of behavioral effects via diazepam-insensitive GABAA receptors. *Eur. J. Pharmacol.* **1995**, *278*, 213-223.
188. Gaster, L.M.; Blaney, F.E.; Davies, S.; Duckworth, D.M.; Ham, P.; Jenkins, S.; Jennings, A.J.; Joiner, G.F.; King, F.D.; Mulholland, K.R. The selective 5-HT1B receptor inverse agonist 10-Methyl-5-[[20-methyl-40-(5-methyl-1,2,4-oxadiazol-3-yl)biphenyl-4-yl]carbonyl]-2,3,6,7-tetrahydro-spiro[furo [2,3-f]indole-3, 40-piperidine] potentially blocks terminal 5-HT auto receptor function both in vitro and in vivo. *J. Med. Chem.* **1998**, *41*, 1218-1235.
189. Selkirk, J.V.; Scott, C.; Ho, M.; Burton, M.J.; Watson, J.; Gaster, L.M.; Collin, L.; Jones, B.J.; Middlemiss, D.N.; Price, G.W.; SB-224289 novel selective (human) 5-HT1B receptor antagonist with negative intrinsic activity. *J. Pharmacol.* **1998**, *125*, 202-208.
190. Macor, J.E.; Ordway, T.; Smith, R.L.; Verhoest, P.R.; Mack, R.A. Synthesis and use of 5-vinyl-1,2,4-oxadiazoles as michael acceptors. A rapid synthesis of the potent muscarinic agonist L-670, 548, *J. Org. Chem.* **1996**, *61*, 3228-3229.
191. Suzuki, T.; Iwaoka, K.; Imanishi, N.; Nagakura, Y.; Miyata, K.; Nakahara, H.; Ohta, M.; Mase, T. Synthesis of the Selective 5-Hydroxytryptamine 4 (5-HT4) Receptor Agonist (p)-(S)-2-Chloro-5-methoxy-4-[5-(2-piperidylmethyl)-1, 2, 4-oxadiazol-3-yl] aniline. *Chem. Pharm. Bull.* **1999**, *47*, 120-122.
192. Manfredini, S.; Lampronti, I.; Vertuani, S.; Solaroli, N.; Recanatini, M.; Bryan, D.; McKinney, M. Design, synthesis and binding at cloned muscarinic receptors of N-[5-(10-substituted-acetoxymethyl)-3-oxadiazolyl] and N-[4-(10-substitutedacetoxymethyl)-2-dioxolanyl] dialkyl amines. *Bioorg. Med. Chem.* **2000**, *8*, 1559-1566.
193. Nicolaides, D.N.; Fylaktakidou, K.C.; Litinas, K.E.; Hadjipavlou-Litina, D. Synthesis and biological evaluation of several coumarin-4-carboxamidoxime and 3-(coumarin-4-yl)-1,2,4-oxadiazole derivatives. *Eur. J. Med. Chem.* **1998**, *33*, 715-724.
194. Matsumoto, J.; Takahashi, T.; Agata, M.; Toyofuku, H.; Sasada, N. A study of the biological pharmacology of IFO, a new selective and reversible monoamine oxidase-B inhibitor. *J. Pharmacol.* **1994**, *65*, 51-57.
195. Zhang, H.Z.; Kasibhatla, S.; Kuemmerle, J.; Kemnitzer, W.; Ollis-Mason, K.; Qiu, L.; Crogan-Grundy, C.; Tseng, B.; Drewe, J.; Cai, S.X. Discovery and structure-activity relationship of 3-aryl-5-aryl-1,2,4-oxadiazoles as a new series of apoptosis inducers and potential anticancer agents. *J. Med. Chem.* **2005**, *48*, 5215-5223.
196. Anjos, J.V.; Neves, Filho, R.A.W.; Nascimento, S.C.; Srivastava, R.M.; Melo, S.J.; Sinou, D. Synthesis and cytotoxic profile of glycosyl-triazole linked to 1,2,4-oxadiazole moiety at C-5 through a straight-chain carbon and oxygen atoms. *Eur. J. Med. Chem.* **2009**, *44*, 3571-3576.
197. Kumar, D.; Patel, G.; Chavers, A.K.; Chang, K.H.; Shah, K. Synthesis of novel 1,2,4-oxadiazoles and analogues as potential anticancer agents. *Eur. J. Med. Chem.* **2011**, *46*, 3085-3092.
198. Maftai, C.V.; Fodor, E.; Jones, P.G.; Daniliuc, C.G.; Franz, M.H. Kelter, G.; Neda, I. Novel 1, 2, 4-oxadiazoles and trifluoromethylpyridines related to natural products: Synthesis, structural analysis and investigation of their antitumor activity. *Tetrahedron* **2016**, *72*(9), 1185-1199.
199. Kemnitzer, W.; Kuemmerle, J.; Zhang, H.Z.; Kasibhatla, S.; Tseng, B.; Drewe, J.; Cai, S.X. Discovery of 3-aryl-5-aryl-1,2,4-oxadiazoles as a new series of apoptosis inducers. 2. Identification of more aqueous soluble analogs as potential anticancer agents. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4410-4415.
200. Tohid, S.F.M.; Ziedan, N.I.; Stefanelli, F.; Fogli, S.; Westwell, A.D. Synthesis and evaluation of indole-containing 3,5-diarylisoxazoles as potential pro-apoptotic antitumour agents. *Eur. J. Med. Chem.* **2012**, *56*, 263-270.
201. Kumar, D.; Maruthi N.; Sundaree, S.; Johnson, E.O.; Shah, K. An expeditious synthesis and anticancer activity of novel 4-(3'-indolyl)oxazoles. *Eur. J. Med. Chem.* **2010**, *45*, 1244-1249.

202. Puthiyapurayil, P.; Poojary, B.; Chikkanna, C.; Kumar, B.S. Design, synthesis and biological evaluation of a novel series of 1,3,4-oxadiazole bearing N-methyl-4-(trifluoromethyl)phenyl pyrazole moiety as cytotoxic agents. *Eur. J. Med. Chem.* **2012**, *53*, 203-210.
203. Juan, S.; Hui, Z.; Zhong-Ming, Y.; Hai-Liang, Z. Synthesis, molecular modeling and biological evaluation of 2-aminomethyl-5-(quinolin-2-yl)-1,3,4-oxadiazole-2(3H)-thione quinolone derivatives as novel anticancer agent. *Eur. J. Med. Chem.* **2013**, *60*, 23-28.
204. Mohammad, S.; Avijit, M.; Mohamed, J.A. Synthesis, characterization and anticancer evaluation of 2-(naphthalen-1-ylmethyl/naphthalen-2-yloxymethyl)-1-[5-(substitutedphenyl)-[1,3,4]oxadiazol-2-ylmethyl]-1H-benzimidazole. *Arab. J. Chem.* **2014** *7*, 418–424.

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