

Case Report

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From Innovation to Complication: A Case Report and Review on Immune-Related Colitis Induced by ICIs

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Case Report

From Innovation to Complication: A Case Report and Review on Immune-Related Colitis Induced by ICIs

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Abstract

Immune checkpoint inhibitors (ICIs) have revolutionized cancer therapy by providing durable responses and a favorable safety profile, ushering in a new era of tumor immunotherapy. However, immune-related adverse events (irAEs) remain a significant clinical challenge. Among these, gastrointestinal irAEs, especially immune-related colitis (ir-colitis), can lead to serious complications if not promptly recognized and managed. Here, we present a case of grade 3 ir-colitis induced by the PD-1 inhibitor sintilimab in a 68-year-old woman with endometrial cancer. The patient developed severe acute diarrhea following ICI administration, which progressed despite initial antidiarrheal and antimicrobial treatments. A multidisciplinary team (MDT) involving gastroenterologists, oncologists, a pathologist, and a clinical pharmacist confirmed the diagnosis and implemented high-dose corticosteroid therapy, yielding significant clinical improvement. Importantly, this report highlights the mechanistic link between PD-1 blockade and ir-colitis pathogenesis, focusing on the dysregulation of the mucosal immune environment and its role in triggering colonic injury. In addition to the case description, we provide a comprehensive review of the literature and clinical guidelines, discussing risk factors, diagnostic approaches, therapeutic strategies, and long-term monitoring. By integrating insights from pharmacology, immunology, and clinical practice, this work emphasizes the importance of early detection, patient education, and MDT collaboration for optimizing therapeutic outcomes and advancing the understanding of ir-colitis in the context of ICI therapy.

Keywords: immune checkpoint inhibitors; diarrhea; colitis; sintilimab; immune-related adverse events

1. Introduction

The advent of immune checkpoint inhibitors (ICIs) has revolutionized cancer therapy, significantly improving survival outcomes and quality of life in patients with advanced malignancies[1]. Despite their clinical benefits, ICIs are associated with a broad spectrum of immune-related adverse events (irAEs), most commonly affecting the skin, thyroid, gastrointestinal tract, liver, and pituitary gland[2]. Among these, immune-related colitis (ir-colitis) presents a particular concern. While the overall incidence of ir-colitis is estimated at approximately 11%, a meta-analysis of PD-1 inhibitor monotherapy reported a much lower incidence of around 0.9%, with grade 3–4 events occurring in only about 0.6% of patients[3,4]. Clinically, ir-colitis typically presents with diarrhea

(with or without mucus or bloody stools), abdominal pain, and weight loss, with endoscopic findings including ulceration, erythema, and loss of vascular architecture. However, its insidious onset and nonspecific gastrointestinal symptoms often overlap with infectious or chemotherapy-induced colitis, contributing to delayed diagnosis and suboptimal management. This delay increases the risk of refractory inflammation, impairs quality of life, and imposes a substantial healthcare burden.

Sintilimab, a novel PD-1 inhibitor, has demonstrated potent antitumor efficacy across multiple solid tumors. Nevertheless, it is also associated with various irAEs, including immune-mediated diabetes mellitus[5], thyroid dysfunction[6], dermatologic toxicities[7], and urologic adverse events[8]. Notably, sintilimab-induced colitis is rarely reported, with an estimated incidence of <1%, and severe (grade \geq 3) cases occurring in fewer than 0.5% of treated patients[9]. This underreporting, combined with the lack of standardized diagnostic and therapeutic protocols, may contribute to delayed clinical recognition, leading to treatment interruptions, disease progression, and diminished patient autonomy.

From a clinical pharmacist's and pharmacotherapeutic management perspective, this case represents a detailed report of sintilimab-induced colitis in an elderly patient with advanced endometrial cancer. By integrating insights from immunology, pharmacology, and clinical practice, and through a multidisciplinary team (MDT) approach, we highlight key aspects of ir-colitis pathogenesis, early detection, and evidence-based treatment strategies. This case highlights the crucial role of clinical pharmacists within the multidisciplinary team (MDT) in facilitating timely pharmacovigilance to optimize the safety and effectiveness of ICI- based immunotherapy.

2. Case Report

2.1. The Patient's History and Clinical Presentation

A 68-year-old woman (156 cm, 51 kg, BMI 20.96 kg/m²) with stage IIIA moderately to poorly differentiated endometrial adenocarcinoma underwent laparoscopic radical surgery in September 2018. Following the surgery, she received 20 cycles of chemotherapy with paclitaxel (270 mg) and carboplatin (600 mg), experiencing nausea and vomiting but no abdominal pain or diarrhea.

In August 2023, tumor recurrence was documented, leading to the initiation of camrelizumab (200 mg) plus chemotherapy. Ten days post-administration, the patient developed grade 1 immune-related diarrhea, characterized by approximately three episodes daily and a fecal volume of about 400 mL, which resolved spontaneously without intervention. We hypothesized that this adverse effect was related to camrelizumab, as this humanized anti-PD-1 monoclonal antibody has a broader tissue distribution in the intestine, potentially triggering excessive activation of the local immune microenvironment[10]. To reduce the risk of further immune-mediated intestinal toxicity, her subsequent immunotherapy cycle (September 14, 2023) switched to sintilimab (200 mg), a fully human PD-1 monoclonal antibody, which is anticipated to be less immunogenic, thereby reducing immune-mediated intestinal inflammation.

However, 40 days after sintilimab administration (October 23, 2023), the patient developed worsening diarrhea of uncertain cause. Initially, she experienced yellow, formed stools (5–10 times per day), which progressed over the next six days to yellow, watery stools (10–15 times per day) of variable volume, accompanied by intermittent fever (up to 37.7°C) and periumbilical colicky pain (VAS score of 3). Treatment with montmorillonite powder and Bacillus licheniformis provided no relief

Ten days later, the patient presented with purulent and bloody stools (without mucus) and persistent abdominal pain. Levofloxacin and symptomatic therapy at an external facility failed to yield significant improvement. Seventeen days later, her diarrhea worsened, characterized by watery, bloody stools with a blood volume of approximately 10 mL, occurring 10–16 times per day, and a total daily volume ranging from 1000 to 1500 mL. The patient experienced intermittent diarrhea for about 20 days, during which anti-diarrheal agents and empirical antibiotic therapy proved ineffective, and her condition continued to worsen. The patient had no history of long-term use of aspirin or

other nonsteroidal anti-inflammatory drugs (NSAIDs). The patient's surgical and prior medication history is summarized in Supplementary Table S1.

2.2. The Patient's Treatment Process

On November 13, 2023, the patient was admitted for further evaluation and management. Upon admission, her vital signs were stable (temperature: 36.4° C, pulse: 68 bpm, respiratory rate: 18 breaths/min, blood pressure: 118/64 mmHg), but the physical examination revealed hyperactive bowel sounds (6 beats/min). Laboratory investigations demonstrated markedly elevated fecal calprotectin ($183.23~\mu g/g$), C-reactive protein (18~mg/L), IL-8 (36.65~pg/mL), IL-9 (1.81~pg/mL), G-CSF (12.38~pg/mL), indicative of active intestinal inflammation, while inflammatory markers, including white blood cell count, procalcitonin, and cytomegalovirus, remained within normal limits. Tests for Clostridium difficile toxins and antigens, antinuclear antibodies (ANA), and T-SPOT were negative, and the patient had no history of lactose intolerance. Whole blood biochemical parameters, electrolyte levels before and during treatment, cytokine and PCT levels are presented in the Supplementary Table S2 and Figures S1–S3.

Colonoscopy revealed extensive mucosal edema from the terminal ileum to the colon, along with spasm, pseudomembrane formation, and friable mucosa prone to hemorrhage (Figure 1 A-H). Histopathological examination confirmed significant colonic inflammation, characterized by mucosal erosion, tissue necrosis, and inflammatory exudates, findings consistent with immune-related colitis (Figure 1 I-J).

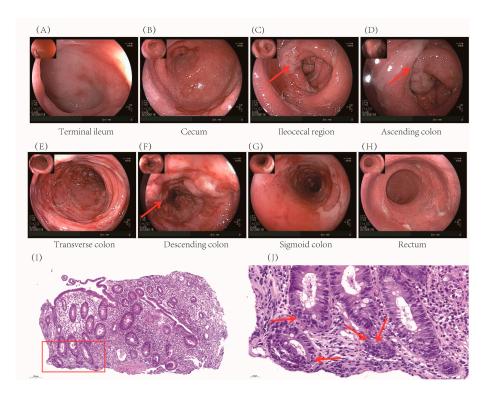


Figure 1. Electronic colonoscopy and pathological images of the patient after Sintilimab. (A)-(H): Colonoscopy revealed diffuse intestinal involvement characterized by hyperemia, edema, ulcers, vesicles, and white pseudomembrane formation from the terminal ileum to the rectum. (I)-(J): Hematoxylin and eosinstained sections of crypt epithelium and a significant reduction of cup cells in colonic tissues, evident in cryptitis, crypt abscesses, crypt atrophy, and structural disturbances in the arrangement of crypt tissues.

The patient was initially treated with intravenous cefoperazone (1 g, 3 times daily) and metronidazole (0.5 g, 2 times daily) for broad-spectrum antibiotic coverage, oral montmorillonite powder (3 g, 3 times daily) for symptomatic control of diarrhea, and bifidobacterium triple viable capsules (840 mg, 2 times daily) to regulate the gut microbiota. Despite treatment, the patient

experienced persistent diarrhea (15–20 episodes/day). On November 17, intravenous methylprednisolone (40 mg/day) was initiated; however, after 3 days, stool frequency remained at 10–20 times/day.

Given the refractory nature of the symptoms, a multidisciplinary team (MDT) consultation, including gastroenterologists, oncologists, a pathologist, and a clinical pharmacist, was conducted to reassess the patient's clinical history, disease progression, and differential diagnosis. From the perspective of pharmacotherapy management, the clinical pharmacist reviewed the patient's medication history, evaluated the clinical trajectory, and strongly suspected grade 3 ir-colitis. Based on the NCCN Guidelines for Management of Immune-Related Adverse Events, the pharmacist recommended escalating methylprednisolone to 1–2 mg/kg/day[11]. Consequently, the dose was increased to 40 mg IV BID (total 80 mg/day) on November 20, with a taper planned over 4–6 weeks. The patient was maintained on a nil per os (NPO) status and received nutritional support with a caloric target of 1,560 kcal/day and protein intake of 1–1.5 g/kg/day. Nutritional supplementation consisted of 27 scoops per day of oral enteral nutrition powder, IV medium- and long-chain triglyceride (MCT/LCT) fat emulsion (250 mL), and a compound amino acid solution (18AA-7).

Given the expected exposure to high-dose corticosteroids (≥20 mg/day prednisone equivalent for ≥4 weeks), Pneumocystis jirovecii pneumonia (PJP) prophylaxis was initiated with oral cotrimoxazole (800 mg twice daily), as advised by the clinical pharmacist. Remarkably, one day after steroid escalation, the patient's stool frequency decreased to <10 times/day, and by day 6, to 0–5 soft, formed stools/day (total volume: 0–500 mL/day). By December 5, 2023, the diarrhea had fully resolved, and the patient's vital signs remained stable. Following a corticosteroid taper, the patient was discharged. The patient's medication history and treatment responses are summarized in Figure 2.

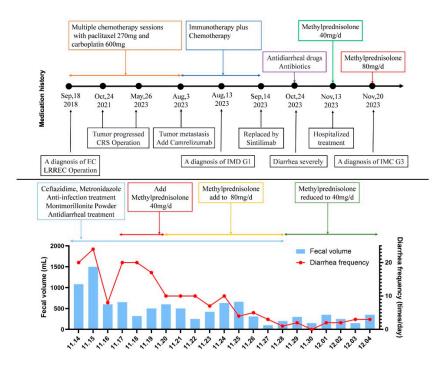


Figure 2. Correlation between diarrhea severity and treatment timeline.

Due to the severity of ir-colitis, ICI therapy was discontinued in accordance with guidelines. Reinitiation was deferred until toxicity resolved to grade ≤1. At the 6-week post-discharge follow-up, the patient reported 3–4 episodes of diarrhea per day, characterized by yellow, pasty stools, with fecal occult blood testing (FOBT) remaining positive. At the 14-week follow-up, the frequency of diarrhea had decreased to 1–2 times per day, with yellow, soft stools; however, the FOBT continued to be positive. At the 1-year follow-up, the patient reported 0–2 normal bowel movements per day, a negative FOBT, and no recurrence of colitis symptoms.

3. Discussion

3.1. Assessment of Adverse Drug Reactions

The patient developed severe, progressive diarrhea on October 23, 2023, which was not observed during previous chemotherapy cycles involving paclitaxel liposomes and carboplatin, agents known to disrupt gut microbiota and sensitize intestinal mucosa. Infectious causes and tumor progression were ruled out, strengthening the likelihood of an immune-related etiology. Notably, symptoms emerged 40 days after the initiation of sintilimab, aligning well with the reported onset times of sintilimab-induced colitis, which range from 6 to 374 days, with a median onset of 112 days[12]. Based on this timeframe, sintilimab was considered the most probable causative agent.

To quantitatively assess the causality of sintilimab-induced colitis, we applied the Naranjo Adverse Drug Reaction Probability Scale, a widely accepted tool for evaluating drug-related adverse events[13,14]. The patient's total score was 6, indicating a "probable" association between sintilimab and the adverse event (details in Supplementary Table S3). The clinical pharmacist diagnosed the patient with grade 3 immune-related colitis (ir-colitis) induced by sintilimab, which was further supported by the prompt resolution of diarrhea following the discontinuation of sintilimab and methylprednisolone therapy (40 mg IV, twice daily). Of note, the patient had previously developed grade 1 self-limiting diarrhea after receiving camrelizumab on August 3, 2023, suggesting a predisposition to ICI-induced gastrointestinal toxicity.

3.2. Mechanisms of Immune-Related Colitis

Sintilimab is a fully humanized IgG4 monoclonal antibody that targets PD-1 with high specificity and affinity for its FG loop region[15]. Under physiological conditions, engagement of PD-1 by its ligand PD-L1 negatively regulates T cell receptor (TCR)-mediated signaling pathways, suppressing the expression of proinflammatory cytokines including IL-2 and IFN- γ [16]. These signaling cascades are essential for maintaining peripheral immune tolerance and preventing autoreactive immune responses. Although the fully human structure of sintilimab is associated with a lower risk of irAEs[15], its potent immunostimulatory activity can override immune homeostasis, increasing the susceptibility to ir-colitis.

PD-1 blockade enhances the proliferation and activation of effector T cells (Teff) while concurrently impairing the suppressive function of regulatory T cells (Tregs)[17]. In patients with ircolitis, histological analyses reveal significant infiltration of CD8+ tissue-resident memory T cells (Trm) in the colonic mucosa, which further differentiate into cytotoxic T lymphocytes (CTLs) capable of secreting high levels of IFN- γ and TNF- α [18]. At the same time, Tregs dysfunction leads to diminished secretion of essential immunosuppressive cytokines, including IL-10 and TGF- β . This imbalance favors the expansion of proinflammatory Th1 and Th17 cells, resulting in increased levels of IL-17 in the gut mucosa and upregulation of chemokines such as TNF- α , IL-6, CXCL9, and CXCL10[19,20]. These inflammatory mediators facilitate the recruitment of additional T cells and neutrophils to inflamed tissues, amplifying the mucosal immune response. In some instances, Tregs undergo phenotypic plasticity, acquiring an IFN- γ -producing Th1-like profile[21], thereby losing their immunosuppressive capacity and further promoting mucosal inflammation.

Histopathologically, one of the defining features of ir-colitis is the increased rate of apoptosis in colonic crypt epithelial cells[22]. This apoptosis is primarily mediated by activated CD8⁺ CTLs through both extrinsic and intrinsic pathways, including Fas/FasL signaling leading to caspase-8–dependent extrinsic apoptosis, and the perforin-granzyme B axis, which induces epithelial cell lysis and caspase-3–dependent intrinsic apoptosis[23]. These processes contribute to crypt architectural distortion and disruption of the epithelial barrier. Additionally, PD-1 inhibitors may induce the production of autoantibodies targeting the integrin $\alpha V\beta 6$, an epithelial adhesion molecule, thereby further impairing epithelial integrity and exacerbating tissue damage[24,25].

As the intestinal barrier becomes increasingly compromised, microbial translocation of luminal bacteria and their metabolites into the lamina propria ensues, activating innate immune cells and

promoting diffuse mucosal inflammation[26]. Clinical studies have demonstrated that patients with ICI-induced colitis exhibit significantly reduced gut microbial diversity[27]. In particular, a marked decrease in the relative abundance of *Faecalibacterium prausnitzii* and *Bacteroides fragilis* has been consistently observed[28]. *F. prausnitzii* is known for its critical role in maintaining colonic mucosal integrity, primarily through the production of butyrate and other short-chain fatty acids that support epithelial barrier function and modulate local immune responses[29,30]. Meanwhile, *B. fragilis* exerts potent anti-inflammatory effects by inducing IL-10 production via its capsular polysaccharide A[31], thereby contributing to the attenuation of mucosal inflammation and the preservation of immune homeostasis.

The combination of immune dysregulation, epithelial damage, and microbial translocation results in impaired absorptive function, increased vascular permeability, and microvascular injury[32]. Clinically, these alterations manifest as acute watery diarrhea, often accompanied by hematochezia, mucus-containing stools, and crampy abdominal pain—hallmarks of sintilimabinduced colitis. (Figure 3. Mechanisms of ir-colitis provides a schematic overview of the pathophysiology, including checkpoint blockade, T-cell dysregulation, epithelial damage, and microbiota-associated factors.)

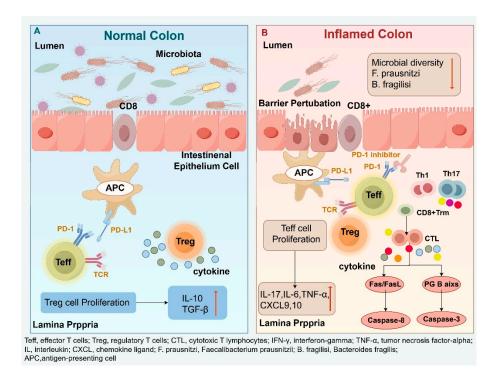


Figure 3. Mechanisms of ir-colitis. (a) Under physiological conditions, the migration of T lymphocytes to the intestinal mucosa is tightly regulated, with preserved epithelial integrity and a diverse, balanced gut microbiota.; (b) PD-1 inhibition disrupts this equilibrium by enhancing the activation and proliferation of Teff while impairing Treg function. In the colonic mucosa, massive infiltration and differentiation of CD8+ T cells into CTLs results in the excessive release of proinflammatory cytokines, such as IFN- γ and TNF- α . Concurrently, activation of Th1 and Th17 cells amplifies local inflammation through the production of IL-17, TNF- α , IL-6, CXCL9, and CXCL10. Activated CTLs induce epithelial cell apoptosis via Fas/FasL signaling and the perforin–granzyme B aixs, leading to mucosal barrier disruption. The consequent increase in epithelial permeability facilitates microbial translocation into the lamina propria, where innate immune activation further exacerbates mucosal inflammation. These synergistic processes culminate in the development of ir-colitis.

3.3. Risk Factors for ir-Colitis Development

The risk of ir-colitis associated with PD-1 blockade is influenced by the specific agent used, treatment regimen, underlying malignancy, and patient-level comorbidities. Combination therapy

with PD-1 and CTLA-4 inhibitors significantly increases the risk, with an overall incidence of up to 30%[33]. In a real-world cohort of 362 lung cancer patients, the incidence of ir-colitis varied across PD-1 inhibitors: sintilimab (3.1%), camrelizumab (3.2%), pembrolizumab (5.0%), and tislelizumab (3.6%). Grade 3–4 colitis occurred in 1.0% and 0.8% of patients treated with camrelizumab and pembrolizumab, respectively, while no high-grade events were reported in the sintilimab group[34]. The risk appears to be dose-dependent, with ir-colitis most commonly emerging after the third treatment cycle[35].

Additional risk factors include tumor biology and patient characteristics. Patients with melanoma have higher ir-colitis rates compared to those with non-small cell lung cancer (NSCLC) or renal cell carcinoma, likely due to distinct tumor–immune microenvironment interactions between the gut microbiome and the tumor-immune microenvironment[36]. Individuals with pre-existing autoimmune diseases, particularly inflammatory bowel disease (IBD), face heightened susceptibility, with colitis incidence rates reaching 40% [37,38]. Gender differences have also been observed: data from the FDA Adverse Event Reporting System indicate that 53.5% of irritable colitis cases occurred in males versus 33.2% in females, although this disparity warrants further study[39].

3.4. Therapeutic Management of ir-Colitis

Gastrointestinal irAEs are among the most common toxicities associated with PD-1 inhibitors, second only to dermatologic events, and more frequent than endocrine toxicities[40]. However, their nonspecific presentation—diarrhea, abdominal pain—can lead to diagnostic delays. Given these challenges, an early MDT intervention is essential, involving oncologists, gastroenterologists, radiologists, pathologists, pharmacists, nutritionists, and psychologists to deliver comprehensive care. Timely identification and intervention not only improve clinical outcomes but also enhance patients' quality of life and treatment adherence (see Figure 4).

Management strategies are guided by evidence-based recommendations from major oncology societies, including the Society for Immunotherapy of Cancer (SITC), American Society of Clinical Oncology (ASCO), National Comprehensive Cancer Network (NCCN), European Society for Medical Oncology (ESMO), and Chinese Society of Clinical Oncology (CSCO)[11,41–44]. A comparative overview is presented in Supplementary Table S4.

For grade 1 ir-colitis, symptomatic management with loperamide, atropine, dietary modifications, and possibly mesalazine is sufficient[11,41,42]. Immunotherapy can typically be continued. In grade 2 ir-colitis, infections should be ruled out, and topical budesonide (9 mg/day) is the preferred initial treatment by NCCN for its enteric-coated formulation allows targeted release in the terminal ileum and right colon, making it theoretically advantageous for localized mucosal inflammation while minimizing systemic corticosteroid exposure, particularly in cases with mild to moderate symptoms[11]. A retrospective study by Machado et al. demonstrated that budesonide achieves remission rates comparable to systemic corticosteroids[45]. However, budesonide has significant limitations. Its efficacy in moderate to severe or extensive colitis remains uncertain, and it is generally not recommended as monotherapy for grade 2 ir-colitis in both international guidelines and expert consensus in China[41–44], initial management still involves the oral corticosteroids, typically prednisone or methylprednisolone at a dose of 1 mg/kg/day. If symptoms fail to improve within 3–5 days, escalation to intravenous corticosteroids or biologics may be necessary.

For grade ≥ 3 ir-colitis, immediate discontinuation of ICIs is mandatory. Patients require hospitalization, continuous monitoring, and urgent IV corticosteroid therapy (methylprednisolone 1–2 mg/kg/day). For steroid-refractory cases, occurring in up to 60% of patients, escalation to biologics is necessary. Infliximab (5 mg/kg)[46]or vedolizumab ($\alpha 4\beta 7$ integrin antagonist) are first-line agents[47]. Additionally, ustekinumab (an IL-12/IL-23 inhibitor) is also effective in select refractory cases[48].

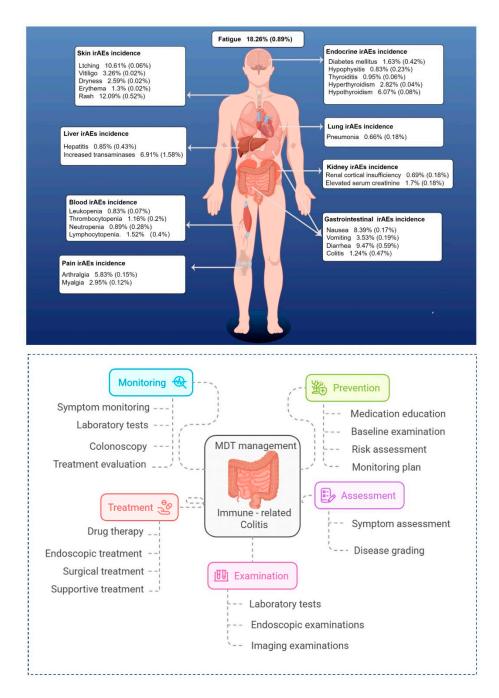


Figure 4. Incidence of irAEs to PD-1 inhibitors and multidisciplinary management of ir-colitis.

Emerging treatment strategies include Janus kinase (JAK) inhibitors and microbiota-targeted interventions. Tofacitinib (10 mg twice daily) has achieved a 75% clinical remission rate in steroid-refractory IBD[49,50], while upadacitinib (45 mg/day) has demonstrated rapid efficacy in severe cases[51]. Fecal microbiota transplantation (FMT) has demonstrated a 58% clinical remission rate within four weeks and is being investigated as a salvage therapy[52].

While novel biologics and small molecules offer hope, their use is limited by potential risks, including infection reactivation (e.g., hepatitis B, tuberculosis), systemic immunosuppression, and incomplete safety profiles. JAK inhibitors, in particular, show promise due to oral administration and rapid onset but require further validation. Future trials should evaluate the safety, efficacy, and durability of these interventions in oncologic populations[53].

4. Conclusions

Sintilimab-induced ir-colitis represents a serious but underrecognized complication of cancer immunotherapy, underscoring the need for timely diagnosis and evidence-based management. Advancing our understanding of its immunopathogenesis, refining risk stratification, and developing targeted therapeutic approaches will be pivotal for optimizing patient outcomes and maximizing the clinical benefits of PD-1 inhibitors. Future prospective studies are warranted to validate emerging treatments and to refine clinical guidelines for managing immune-mediated gastrointestinal toxicities.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org.

Author Contributions: Conceptualization, Funding acquisition, Methodology, Project administration, Resources, Writing – review & editing, H.L.; Data curation, Formal analysis, Visualization, Writing – original draft, Y.P.; Investigation, Supervision, W.L.; Visualization, H.Z.; Data curation Investigation, Supervision, X.T.; Writing – review & editing, R.Z.; Funding acquisition, Writing – review & editing, Y.Z.

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Institutional Review Board Statement: The study was conducted in accordance with the Declaration of Helsinki, and approved by Peking University Third Hospital. The studies were conducted in accordance with the local legislation and institutional requirements.

Informed Consent Statement: The participant provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

Data Availability Statement: The original contributions presented in this study are included in the article/Supplementary Materials. Further inquiries can be directed to the corresponding authors.

Conflicts of Interest: The authors declare no conflicts of interest.

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