

Hypothesis

Not peer-reviewed version

Targeted Nanotechnology and AI-Driven Strategies to Penetrate Caseous Lesions and Overcome Immune Evasion in Tuberculosis: A Comprehensive Review

[Amr Ahmed](#)*

Posted Date: 6 March 2026

doi: 10.20944/preprints202603.0540.v1

Keywords: tuberculosis; caseous lesions; nanomedicine; macrophage targeting; lipid nanoparticles; artificial intelligence; immune evasion; extracellular vesicles; bioorthogonal diagnostics; spatial pharmacokinetics



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a [Creative Commons CC BY 4.0 license](#), which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Hypothesis

Targeted Nanotechnology and AI-Driven Strategies to Penetrate Caseous Lesions and Overcome Immune Evasion in Tuberculosis: A Comprehensive Review

Amr Ahmed

Public Health Department, Riyadh First Health Cluster, Ministry of Health, Riyadh, Saudi Arabia;
drmedahmed@gmail.com

Abstract

Despite the availability of antibiotics, pulmonary tuberculosis (TB) remains a leading infectious cause of mortality globally. Treatment failure and the emergence of drug-resistant strains are largely driven by the heterogeneous architecture of caseating granulomas and the complex biophysical mechanisms by which *Mycobacterium tuberculosis* (Mtb) evades host immunity. Highly lipophilic frontline drugs, such as bedaquiline and clofazimine, exhibit severe sequestration within the lipid-rich necrotic caseum, preventing them from reaching the dormant persister bacilli at the lesion's core.⁴ Furthermore, recent biophysical discoveries reveal that Mtb utilizes extracellular vesicles and specialized lipids to mechanically stiffen host macrophage membranes, thereby arresting phagosome-lysosome fusion.⁶ This review proposes an AI-optimized, "Trojan Horse" hybrid nanocarrier strategy—comprising a lipidic core, a mucoadhesive chitosan shell, mannose-targeted ligands, and pH-responsive release mechanisms—to bypass these dual barriers.⁸ By bridging lesion-centric pharmacokinetics (f_{u_caseum} , D_{eff}), novel bioorthogonal diagnostic probes, and machine learning formulation designs, we present a translational roadmap aimed at achieving complete sterilization of caseous cavities.⁴

Keywords: tuberculosis; caseous lesions; nanomedicine; macrophage targeting; lipid nanoparticles; artificial intelligence; immune evasion; extracellular vesicles; bioorthogonal diagnostics; spatial pharmacokinetics

1. Introduction: Historical Context and the Drug Resistance Crisis

Tuberculosis has afflicted humanity since antiquity, with genomic and phylogenetic evidence suggesting that the *Mycobacterium tuberculosis* complex co-evolved with early human migrations from Africa tens of thousands of years ago.¹ Paleopathological analyses of Egyptian mummies and ancient texts from Greece, India, and China document the historical devastation of the disease, historically termed "Phthisis" or the "White Plague".²

A monumental shift in the medical understanding of TB occurred on March 24, 1882, when Robert Koch isolated the tubercle bacillus, definitively proving its infectious etiology and establishing the foundation for modern bacteriology.³ Early treatments were confined to sanatoria and surgical interventions until the dawn of the antibiotic era, heralded by the discovery of streptomycin in 1943 by Selman Waksman and his team.³ This breakthrough was rapidly followed by the introduction of isoniazid, pyrazinamide, ethambutol, and rifampicin, which transformed TB into a curable condition.³

However, this therapeutic triumph has been severely undermined by the emergence of multidrug-resistant (MDR-TB) and extensively drug-resistant (XDR-TB) strains, exacerbated by lengthy treatment regimens and patient non-compliance.¹² While recent approvals of novel agents like bedaquiline, pretomanid, and repurposed clofazimine offer new hope, their in vivo efficacy is

frequently hindered by profound physiological barriers.⁴ Overcoming these obstacles requires a paradigm shift from traditional plasma-centric pharmacology to targeted nanomedicine designed specifically for the granuloma microenvironment.⁸

2. Spatial Localization: The Caseum as a Pharmacological Sanctuary

The hallmark of pulmonary TB is the formation of the granuloma, a highly organized immune structure. In cavitary TB, the center of the granuloma undergoes necrosis, forming a cheese-like substance known as caseum.⁴ This caseous core is a biologically extreme environment: it is completely avascular, highly hypoxic, and densely packed with lipid-rich debris derived from necrotic foamy macrophages.⁵

Subjected to this nutrient-starved and acidic milieu, *Mtb* undergoes a metabolic shift, transitioning into non-replicating, dormant persister cells.¹³ These persisters do not rely on genetic mutations for survival; rather, they exhibit extreme "phenotypic tolerance" to standard bactericidal antibiotics designed to target rapidly dividing cells.¹³ Eradicating these subpopulations requires drugs that can physically penetrate the dense, lipid-rich caseum matrix—a challenge that dictates the success or failure of current therapeutic regimens.⁵

3. Biophysical Mechanisms of Immune Evasion

Beyond the physical barrier of the caseum, *Mtb* employs highly sophisticated, physically driven mechanisms to hijack host macrophages and evade intracellular destruction.

3.1. Extracellular Vesicles and Membrane Stiffening

Groundbreaking biophysical research elucidated a mechanical evasion strategy: *Mtb* actively secretes nanoscale extracellular vesicles (EVs) laden with pathogenic lipids that fuse with host macrophage membranes.⁶ This fusion induces a drastic physical alteration, significantly increasing the surface tension and rigidity of the host's cellular and phagosomal membranes.⁶ Because phagosome-lysosome fusion requires membrane flexibility to form intermediate structures like pores and stalks, the pathogen-induced membrane stiffening physically arrests phagosome maturation.⁶ Consequently, the *Mtb* bacilli are shielded within a protective intracellular bunker, avoiding the lethal acidic and enzymatic environment of the lysosome.⁶

3.2. Lipid-Mediated Disruption of Phagocytosis and Maturation

Mtb's unique cell envelope lipids act as powerful virulence factors:

- **Lipoarabinomannan (LAM):** LAM inserts into the host membrane, altering the molecular packing of sphingomyelin-cholesterol microdomains. This rigidifies the domains in a stable 3D conformation, effectively preventing the vesicle association required for lysosomal fusion.¹⁴
- **Phthiocerol dimycocerosates (DIM):** Exhibiting a conical geometry, DIM promotes the formation of inverted hexagonal non-bilayer phases in the host membrane. This reduces the energy required for membrane curvature, facilitating smooth phagocytosis without triggering robust immune signaling.¹⁵
- **Trehalose dimycolate (TDM):** TDM disrupts membrane fusion molecularly by forcing host SNARE proteins into non-canonical complexes (e.g., complexing with VAMP2 instead of VAMP8), paralyzing the phagosomal maturation pathway.¹⁶

4. Lesion Pharmacokinetics: The Lipophilic Drug Paradox

Traditional drug development relies heavily on plasma pharmacokinetics. However, studies have definitively proven that plasma exposure correlates poorly with drug distribution inside the granuloma.⁵

By adapting the Rapid Equilibrium Dialysis (RED) assay to measure drug binding to caseum homogenates, researchers uncovered a critical paradox regarding modern anti-TB drugs.¹⁸ Highly lipophilic drugs, which form the backbone of modern MDR-TB regimens (e.g., bedaquiline and clofazimine), exhibit massive non-specific binding to the lipid debris in the necrotic core.⁵ The unbound, freely diffusing fraction (f_{u_caseum}) of these drugs is often $< 0.01\%$.⁵ Matrix-assisted laser desorption/ionization mass spectrometry imaging (MALDI-MSI) visually confirms this phenomenon, showing clofazimine intensely accumulated in the cellular rim but entirely absent from the central caseum where persisters reside.¹⁷ Conversely, smaller, less lipophilic drugs like pyrazinamide demonstrate high free fractions ($f_{u_caseum} > 99.9\%$) and diffuse readily throughout the lesion.⁵

5. The "Trojan Horse" Strategy: AI-Optimized Hybrid Nanocarriers

To bypass both the biophysical immune evasion of Mtb and the severe caseum binding of lipophilic drugs, we propose a rational, targeted "Trojan Horse" nanocarrier system designed specifically for the TB microenvironment.⁴

5.1. Nanocarrier Architecture

1. **Lipidic Core:** The core utilizes solid-lipid nanoparticles (SLNs) or nanostructured lipid carriers (NLCs) to encapsulate highly lipophilic drugs (bedaquiline, clofazimine) at high payloads.²⁰ This physically shields the drug from premature binding to caseum debris, artificially circumventing the low f_{u_caseum} limitation.⁴
2. **Chitosan Shell:** A polymeric coating of chitosan provides colloidal stability and mucoadhesive properties, making the formulation highly suitable for localized pulmonary delivery via dry powder inhalation.²¹
3. **Active Macrophage Targeting:** The surface is functionalized with mannose ligands. Because infected alveolar macrophages overexpress mannose receptors (CD206), the nanocarriers are selectively phagocytosed.⁸ The infected macrophage then acts as a Trojan horse, migrating deep into the inflamed granuloma tissue carrying the protected drug payload.⁴
4. **pH-Responsive Release Trigger:** To prevent systemic toxicity, the nanocarrier utilizes stimuli-responsive linkages (e.g., hydrazone bonds). These bonds remain intact in the blood (pH 7.4) but rapidly degrade in the acidic microenvironment of the phagolysosome and caseum (pH 5.5-6.5), releasing a massive, localized dose of bactericidal agents directly upon the intracellular bacilli.²²

5.2. AI-Driven Formulation and Synergy

Designing this multi-component nanoparticle is an intrinsically high-dimensional problem. Utilizing Artificial Intelligence (AI), specifically Bayesian Optimization and Machine Learning algorithms like XGBoost, allows researchers to navigate complex design spaces efficiently.²³ These algorithms can predict critical quality attributes (size, zeta potential, encapsulation efficiency) with over 94% accuracy, significantly accelerating the path to scalable manufacturing.²³ Furthermore, AI chemo-genomics platforms have successfully predicted unprecedented synergistic combinations (e.g., bedaquiline, clofazimine, pretomanid, and thioridazine) that, when co-encapsulated into nanocarriers, could overwhelm Mtb resistance mechanisms.²⁵

6. Diagnostic Innovations: Bioorthogonal Targeting of Rare Glycans

The effectiveness of advanced nanomedicines relies on equally advanced diagnostic capabilities. A 2025 breakthrough by Kiessling and colleagues at MIT provides a novel method for identifying Mtb.²⁶ The virulent Mtb glycan ManLAM is capped with a rare, unique sugar called methylthioxylofuranose (MTX), which contains a highly specific thioether bond.²⁶

By utilizing a small-molecule oxaziridine probe, the team achieved highly selective, bioorthogonal labeling of MTX on live mycobacteria without genetic modification.²⁶ This innovation allows for the real-time spatial imaging of ManLAM dynamics during macrophage infection and paves the way for rapid, ultrasensitive diagnostic tests that detect pathogen-specific glycans in urine or sputum long before traditional cultures yield results.²⁸

7. Experimental Roadmap and Conclusion

Translating these nanotechnologies to clinical practice requires abandoning generic pharmacokinetic testing in favor of lesion-mimetic criteria.⁴ Future investigations must prioritize:

1. Quantifying f_{u_caseum} and effective diffusion (D_{eff}) of nanocarriers using RED assays.¹⁸
2. Demonstrating intracellular localization and pH-triggered release in primary human macrophages.²²
3. Validating spatial drug penetration gradients in caseous animal models using MALDI-MSI.¹⁷

Conclusion: The persistence of cavitary TB is dictated by the spatial pharmacology of the caseum and the biophysical manipulation of host membranes by Mtb. By integrating AI-optimized, macrophage-targeted lipid nanoparticles with novel bioorthogonal diagnostics, modern medicine can bypass these formidable barriers.¹⁰ This multidisciplinary framework offers a viable path toward shortening treatment regimens, rescuing modern lipophilic drugs from caseum sequestration, and ultimately achieving sterilizing cures in drug-resistant tuberculosis.

References

1. Wikipedia contributors. History of tuberculosis. Wikipedia, The Free Encyclopedia; 2026. Available from: https://en.wikipedia.org/wiki/History_of_tuberculosis
2. Centers for Disease Control and Prevention. History of World TB Day. CDC. Available from: <https://www.cdc.gov/world-tb-day/history/index.html>
3. Frith J. History of Tuberculosis. Part 2 - the Sanatoria and the Discoveries of the Tubercle Bacillus. J Mil Veterans Health. 2014;22(2):36-41. Available from: <https://jmvh.org/article/history-of-tuberculosis-part-2-the-sanatoria-and-the-discoveries-of-the-tubercle-bacillus/>
4. News-Medical.Net. History of Tuberculosis. Available from: <https://www.news-medical.net/health/History-of-Tuberculosis.aspx>
5. Ahmed A, Rodaini S. AI-Optimized, Macrophage-Targeted, pH-Responsive Hybrid Nanocarriers to Improve Penetration of Lipophilic Anti-TB Drugs into Caseous Cavitary Lesions: A Perspective and Experimental Roadmap. [Unpublished manuscript]. 2026.
6. Alsayed SSR, Gunosewoyo H. Tuberculosis: Pathogenesis, Current Treatment Regimens and New Drug Targets. Int J Mol Sci. 2023;24(6):5202. doi: 10.3390/ijms24065202.
7. Sarathy JP, Zuccotto F, Ho H, et al. Prediction of Drug Penetration in Tuberculosis Lesions. ACS Infect Dis. 2016;2(8):552-563. doi: 10.1021/acsinfecdis.6b00051.
8. Sarathy JP, Via LE, Weiner D, et al. Extreme Drug Tolerance of Mycobacterium tuberculosis in Caseum. Antimicrob Agents Chemother. 2018;62(2):e02266-17. doi: 10.1128/AAC.02266-17.
9. Biophysical Society. Researchers Discover How Tuberculosis Bacteria Use a “Stealth” Mechanism to Evade the Immune System. 2026. Available from: <https://www.biophysics.org/news-room/researchers-discover-how-tuberculosis-bacteria-use-a-stealth-mechanism-to-evade-the-immune-system>
10. Koiri D, Mathew L, Panda A, et al. Remote Host Manipulation by Pathogenic Bacterial Extracellular Vesicles. bioRxiv [Preprint]. 2025. doi: 10.64898/2025.12.17.694930.

11. Hayakawa E, Tokumasu F, Nardone GA, et al. A Mycobacterium tuberculosis-Derived Lipid Inhibits Membrane Fusion by Modulating Lipid Membrane Domains. *Biophys J*. 2007;93(11):4018-4030. doi: 10.1529/biophysj.107.108423.
12. Augagneur Y, et al. The conical shape of DIM lipids promotes Mycobacterium tuberculosis infection of macrophages. *bioRxiv* [Preprint]. 2019. doi: 10.1101/649541.
13. Santamaria C, Biegas KJ, Lim PN, et al. Trehalose dimycolate inhibits phagosome maturation and promotes intracellular Mycobacterium tuberculosis growth via noncanonical SNARE interactions. *Proc Natl Acad Sci U S A*. 2025;122(20):e2423292122. doi: 10.1073/pnas.2423292122.
14. Sarathy JP, Ho Liang HP, Weiner D, et al. An In Vitro Caseum Binding Assay that Predicts Drug Penetration in Tuberculosis Lesions. *J Vis Exp*. 2017;(123):55559. doi: 10.3791/55559.
15. Strydom N, et al. Tuberculosis drugs' distribution and emergence of resistance in patient's lung lesions: A mechanistic model and tool for regimen and dose optimization. *PLoS Med*. 2019;16(4):e1002773. doi: 10.1371/journal.pmed.1002773.
16. Sarathy JP, Ho Liang HP, Weiner D, et al. An In Vitro Caseum Binding Assay that Predicts Drug Penetration in Tuberculosis Lesions. *J Vis Exp*. 2017;(123):55559. doi: 10.3791/55559.
17. Zamani FN, Hashim NM, Nair RS, et al. A review on recent advances in lipid-based drug delivery systems for tuberculosis. *Iran J Basic Med Sci*. 2025;28(3):322-333. doi: 10.22038/ijbms.2025.88836.19184.
18. Ma C, Wu M, Ye W, et al. Correction: Inhalable solid lipid nanoparticles for intracellular tuberculosis infection therapy: macrophage-targeting and pH-sensitive properties. *Drug Deliv Transl Res*. 2021;11(3):1236. doi: 10.1007/s13346-021-00958-x.
19. Elbehiry A, Marzouk E, Abalkhail A. Advancing Tuberculosis Treatment with Next-Generation Drugs and Smart Delivery Systems. *Pharmaceutics*. 2026;18(1):60. doi: 10.3390/pharmaceutics18010060.
20. Okafor NI, Nnaji P, Nnolum-Orji NF, Choonara YE. Functionalized Polydopamine Nanoparticles: A Promising Drug Delivery Platform for the Treatment of Tuberculosis. *Drug Dev Res*. 2025;86:e70109. doi: 10.1002/ddr.70109.
21. Li H, et al. Dual pH-Responsive Macrophage-Targeted Isoniazid Glycoparticles for Intracellular Tuberculosis Therapy. *Biomacromolecules*. 2021;22(8):3240-3252. doi: 10.1021/acs.biomac.1c00554.
22. Kapoor DU, Sharma JB, Gandhi SM, et al. AI-driven design and optimization of nanoparticle-based drug delivery systems. *Sci Eng Health Stud*. 2024;18:24010003. doi: 10.69598/sehs.18.24010003.
23. Su K, Qiu J, Xu T, Liu S. Artificial intelligence-guided design of lipid nanoparticles for mRNA delivery. *Acta Pharmaceutica Sinica B*. 2026;16(2):709-727. doi: 10.1016/j.apsb.2025.11.029.
24. Docwire News. AI Finding New Drug Combinations to Fight Tuberculosis. 2020. Available from: <https://www.docwirenews.com/post/using-ai-to-find-new-drug-combinations-to-fight-tuberculosis>
25. Smelyansky SR, Ma CW, Marando VM, et al. Exploiting thioether reactivity to label mycobacterial glycans. *Proc Natl Acad Sci U S A*. 2025;122(19):e2422185122. doi: 10.1073/pnas.2422185122.
26. Smelyansky SR, Ma CW, Marando VM, et al. Exploiting thioether reactivity to label mycobacterial glycans. *Proc Natl Acad Sci U S A*. 2025;122(19):e2422185122. doi: 10.1073/pnas.2422185122.
27. MIT News. New molecular label could lead to simpler, faster tuberculosis tests. 2025. Available from: <https://news.mit.edu/2025/new-molecular-label-could-lead-simpler-faster-tuberculosis-tests-0505>
28. Onnainty R, Granero GE. Advancements in Nanotheranostic Approaches for Tuberculosis: Bridging Diagnosis, Prevention, and Therapy Through Smart Nanoparticles. *J Nanotheranostics*. 2025;6(4):33. doi: 10.3390/jnt6040033.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.