Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Valorization of Lignocellulosic Biomass into Biofuels and Value-Added Products for Sustainable Environment: A Comprehensive Review

Surbhi Sharma ^{1, †}, Mei-Ling Tsai ^{2, †}, Vishal Sharma ^{2, 3, †}, Pei-Pei Sun ², Parushi Nargotra ², Bijender Kumar Bajaj ¹, Chiu-Wen Chen ^{3, 4, *}, Cheng-Di Dong ³

- ¹ School of Biotechnology, University of Jammu, India; <u>ssurbhi337@gmail.com</u> (S.S.); <u>bajajbijenderk@gmail.com</u> (B.K.B.)
- ² Department of Seafood Science, National Kaohsiung University of Science and Technology, Kaohsiung City, Taiwan; <u>sudans123@gmail.com</u> (V.S.); <u>mltsai@nkust.edu.tw</u> (M.-L.T.); <u>ppsun@nkust.edu.tw</u> (P.-P.S.); <u>parushi11nargotra@gmail.com</u> (P.N.)
- ³ Department of Marine Environmental Engineering, National Kaohsiung University of Science and Technology, Kaohsiung City, Taiwan
- ⁴ Institute of Aquatic Science and Technology, National Kaohsiung University of Science and Technology, Kaohsiung City, Taiwan
- * Correspondence: cwchen@nkust.edu.tw (C.-W.C.); cddong@nkust.edu.tw (C.-D.D.)
- [†] These authors contributed equally to this work.

Abstract: An upsurge in global population over the years and rapid urbanization have accelerated huge dependence on petroleum-derived fuels and consequent environment concerns owing to greenhouse gas emissions in the atmosphere. An integrated biorefinery uses lignocellulosic feedstock as raw material for the production of renewable biofuels, and other fine chemicals. The sustainable bio-economy and the biorefinery industry would benefit greatly from the effective use of lignocellulosic biomass obtained from agricultural feedstocks to replace petrochemical products. Lignin, cellulose, hemicellulose, and other extractives, which are essential components of lignocellulosic biomass, must be separated or upgraded into useful forms in order to fully realize the potential of biorefinery. The development of low-cost and green pretreatment technologies with effective biomass deconstruction potential is imperative for an efficient bioprocess. The abundance of microorganisms along with their continuous production of various degradative enzymes makes them suited for the environmentally friendly bioconversion of agro-industrial wastes into viable bioproducts. The present review highlights the concept of biorefinery, lignocellulosic biomass and its valorization by green pretreatment strategies into biofuels and other biochemicals. The major barriers and challenges in bioconversion technologies, environmental sustainability of the bioproducts and promising solutions to alleviate those bottlenecks are also summarized.

Keywords: Integrated biorefinery; Circular economy; Lignocellulosic biomass; Pretreatment; Valorization; Enzymes; Biofuels; Biochemicals

1. Introduction

Global population growth and socio-economic expansion have contributed to an enormous rise in pollution and an expedited depletion of energy resources [1,2]. The primary cause of the current global energy crisis and deterioration of the environment is the overconsumption of conventional fossil fuels, which contribute significantly to greenhouse gas emissions, global warming, acid rains and other climatic ill-effects [3,4]. Thus, it is imperative to make enduring technological developments to supply the accelerating energy demand without imperilling the planet's finite resources [5]. The paradigm of "take, make, and dispose off" is currently being replaced by "reuse and recovery" of re-

sources in order to attain 'healthy climate-healthy individual' and 'socio-economic success' [6]. To produce low-carbon, long-lasting, and fossil-free fuels, the development of various non-conventional/renewable/infinite energy resources are being explored. Lignocellulosic biomass (LCB)-based biofuels are getting prominence as a feasible substitute for petroleum-based fuels [3,7]. LCB constituted by agricultural residues/food waste/forest products/municipal and sewage wastes represents the most promising, copious and a low-cost carbon-based substrate, which can be extensively exploited for the production of biofuels and other commercial products in a biorefinery [4,8–10]. The sustainable bioeconomy and the biorefinery industry would benefit greatly from the effective use of lignocellulosic biomass obtained from agricultural feedstocks to replace petrochemical products.

Biorefinery is a strategy which integrates several conversion processes, such as biochemical, thermo-chemical, and microbial growth to generate energy, biofuels, chemicals, and other value-added desired industrial products from LCB [11]. However, the full-scale bioconversion of LCB to bioenergy/biofuels (biodiesel, bioethanol, biobutanol), and biobased commodities (chemicals, feed, food) poses various significant obstacles in terms of energy input and product yield into the overall process [12]. LCB, a hetero-polysaccharide complex, is comprised of a variety of polymers (cellulose, hemicellulose, and lignin), and other polar and non-polar substances [9,13–15]. The structural polysaccharides of LCB are entangled with lignin blocks in a very complicated manner, thereby, imparting high rigidity, and recalcitrance to biomass structure [12]. Hence, a key bottleneck in the valorization of LCB for the production of power, fuels, or other chemicals is its structural recalcitrance, which might be successfully overcomed by implementing an appropriate pretreatment approach [1,16]. Pretreatment aims to disrupt the crystalline structure of biomass by breaking the hydrogen bonds, and cross-linked hydrophobic interactions between cellulose, hemicellulose and lignin, thus, increases the accessibility of biomass to enzyme hydrolysis [17,18]. An effective pretreatment may improve the cellulose's accessibility, pore size, and surface area while lowering its degree of polymerization (DP) and crystallinity [19,20]. The primary pretreatment regimes available for the fractionation of LCB can be categorized as physical, chemical, physicochemical, and biological techniques [6]. However, most of these methods possess several limitations such as high cost, nongreen, generation of inhibitors/toxic substances, and many others. Therefore, in the current times, research is being focused on the development of environmentally-friendly and cost-efficient pretreatment methods to achieve successful LCB-fuel technology [13,15]. The choice of an appropriate pretreatment technique depends upon biomass type, its structural properties, composition, and degree of polymerization. Pretreatment is the principal capital-intensive step in LCB biorefinery, thus, significantly affects the overall economy of a bioprocess [21].

Another crucial step in the chain of LCB-fuel scheme is enzymatic saccharification of suitably pretreated biomass. The enzyme cocktail of cellulases (endo- β -1,4-glucanase, cellobiohydrolase, and β -glucosidase), and hemicellulases (xylanases, glucuronidase, glucomannanase, galactomannanase, β -xylosidase, acetylesterase) work synergistically to rupture the cellulose, and hemicellulose fibres into glucose monomers during enzymatic hydrolysis [19,22]. Currently, lytic polysaccharide monooxygenases (LPMOs), manganese peroxidase, laccases are being employed for enzymatic saccharification; as they hydrolyze the polysaccharides, remove lignin, thus, improve the effectiveness of process by increasing the glucose yield and decreasing the quantity of cellulases [3,22]. However, high sensitivity, high cost, less stability, and high dosage of the enzymes hinder greatly the economic feasibility of the biofuel production process. Thus, novel approaches such as onsite enzyme production, usage of enzyme cocktails, enzyme immobilization, and others are being utilized for achieving the cost-economy of the overall process [13,23]. Besides biofuel, a variety of value-added products are also produced through enzymatic saccharification of pretreated LCB such as xylo-oligosaccharides, xylose, psicose, tagatose, xylene,

toluene, benzene, aliphatic acids, quinines, and lignin monomers including syringols, syringaldehyde, propylphenol eugenol, vanillin, aryl ethers, and many others [24]. For the production of biofuel-ethanol, butanol, lactic acid, acetone, or other end products, the reducing sugars (hexoses and pentoses) obtained are subjected to a biochemical process named 'fermentation' [22]. The fermentation process is intended to be optimized via pretreatment and hydrolysis techniques [25].

Based on the composition of sugary hydrolysate, specific micro-organisms (bacteria, fungi or yeasts) are required for metabolization, and product formation [2]. The commonly used micro-organisms for ethanol fermentation include *Saccharomyces cerevisiae*, *Pichia stipitis*, *Zymomonas mobilis*, *Candida shehatae*, *Candida brassicae*, *Klebsiella oxytoca*, *Mucor indicus*, *Escherichia coli*, *Penicillium*, *Trichoderma*, *Aspergillus* [8,14,17,26]. However, *Saccharomyces cerevisiae* is the principal yeast which has been employed extensively for the production of alcohol, mainly in wine industries and breweries [2]. Thus, valorization of lignocellulosic feedstock into biofuels, and other upgraded products envisage the 'waste to wealth' transformation, and the growth of a biomass-based economy [27]. The current review the article addresses the concept of biorefinery, challenges associated, and the possible solutions which might improve bioprocessing for the development of a sustainable biorefinery. Additionally, a brief description of promising pretreatment strategies for LCB bioconversion and key steps involved in biofuel production processes is outlined, besides discussing the environment sustainability of the fuels and bioproducts.

2. Lignocellulosic Biorefinery concept for circular bioeconomy

In order to boost production of green fuels and adding economic value, the concept of a biorefinery was established. A biorefinery is a fundamental framework developed for the use of raw biomass (LCB, algae, other organic wastes), in which all processing methods are integrated in a systematic manner to produce sustainable bio-based products [2]. The biorefineries are essential to fostering a circular bioeconomy that is knowledge-driven and environmentally sound [2,28]. The idea of circular bioeconomy majorly emphasizes on the recovery of all goods from resources without any waste being discharged, and the lignocelluloisc biomass utilized as a feedstock plays a vital role in the production of both bioproducts and the energy needed to power them. The primary goal of circular bioeconomy is to replace the end-of-life phase through resource recovery, the use of renewable energy, the elimination of harmful chemicals, and the zero discharge concept by altering the design of all relevant systems and business models [11]. Conventional biorefineries implement high levels of process integration and little waste output to transform biomass into a variety of products for different industrial sectors [29]. According to the International Energy Agency (IEA), biorefinery is targets 'the sustainable conversion of various biomasses into bioenergy and diverse bio-products (chemicals, feed, and food), and bioenergy (power, heat, and biofuels)' [25]. Based on the literature survey [2,19,30], biorefinery can be classified into three phases on the basis of biomass, biomass processing technology, and the product:

- a. Phase I biorefinery (one feed, one preset processing technology, and one main product). For example, in European Union, vegetable oil biorefinery involves the production of biodiesel by the transesterification of rapeseed oil.
- b. Phase II biorefinery (one feed, multiple processing technologies, and multiple end products). For example, in Sweden, forestry feedstock is refined to produce cellulose, lignosulphonate and bioethanol.
- c. Phase III biorefinery (multiple feedstocks, multiple processing technologies, and multiple end products). For example, lignocellulosic biomass based biorefinery, two platform biorefinery, green biorefinery, and others.

A successful biorefinery must be the one which produce very little or no waste, while processing. Thus, an integrated strategy which combines conversion strategies (chemical, thermochemical, and biochemical), and potent downstream strategies systematically,

must be used in a zero-waste biorefinery foe achieveing the taget of circula bioeconomy [2,11]. In this way, a sustainable and more economical approach could be developed for an ideal biorefinery, which could potentially provide a plethora of fuels, and value-added industrial products [22]. Figure 1 shows shows how the concept of circular bio-economy can be implemented in a lignocellulosic biorefinery for a sustaiabe clean environment. With the basic layout of a biorefinery process, the unexplored lignocellulosic biomass can serve as potent resource for the production of bio-products. The ability of using various LCBs can provide a consistent supply for biorefining processes throughout the year, thus, boosting the viability of biomass waste valorization with an economic perspective [31]. The accomplishment of LCB based second generation (2G) biorefinery is majorly determined by the type, composition, and properties of biomasses as well as the conversion processes used in the process.

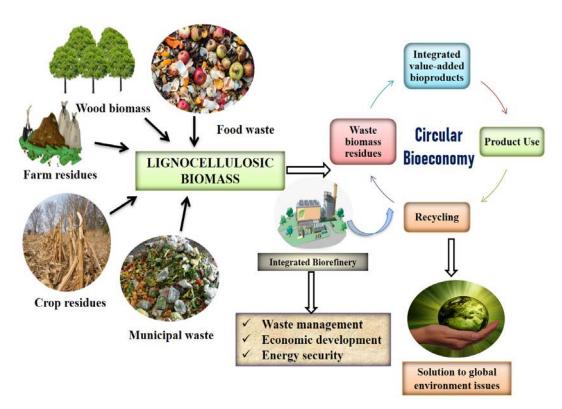


Figure 1. Idea of circular bioeconmy using integrated biorefinery for sustainable environment.

3. Lignocellulose composition and structure

Lignocellulosic biomass is the most copious, low-cost, carbon-neutral, and high-energy density organic polymer present on this earth [32]. The production of renewable fuels from LCB not only boosts the global energy security, but also minimizes the wastes, helps rural economies, and reduces the environmental problems [33]. LCB is a heterogeneous polymeric complex composed of three major components i.e., cellulose (35–55%), hemicellulose (20-40%), and aromatic lignin (10-25%), and minor traces of protein, pectin, ash and other extractives (inorganics, waxes, fats, resin acids, phenolics) [11,34]. Table 1 shows the cellulose, hemicellulose, and lignin content in different types of lignocellulosic feed-stocks.

The percent composition of these polymers may differ in different biomasses due to variations in biomass type (softwood, hardwood, grasses, and farm waste), soil type, climate type, fertilizers type, growing conditions, harvesting technique, and other physical factors [1,8,15].

Table 1. Composition of different lignocellulosic biomass feedstocks.

| S.No. | Lignocellulose feedstocks | Cellulose (%) | Hemicellulose (%) | Lignin (%) | Reference |
|------------|---------------------------|----------------|---|----------------------|-----------|
| 1. | Sugarcane bagasse | 38.6 | 20 | 24.68 | [8] |
| 2. | Sunflower stalks | 27.22 | 11.94 | - | [34] |
| 3. | Oat flakes | 21 | 38% | 27 | [35] |
| 4. | Spruce sawdust | 55.4 | 1.4% arabinose, 4.2% xylose | 28.7 | [36] |
| 5. | Eucalyptus | 41.58 | 15.85 | 29.40 | [37] |
| 6. | Parthenium hysterophorus | 49.98 | 7.61% arabinose, 14.18% xylose | 17.6 | [14] |
| 7. | Saccharum spontaneum | 32.16 | 19.36 | 16.86 | [32] |
| 8. | Birchwood planks | 54.22 | 28.14 | 11.13 | [38] |
| 9. | Oak sawdust | 44.7 | 1.2% arabinose, 14.8% xylose | 26.7 | [36] |
| 10. | Oil palm trunk | 56.1 | 16.15 | 19.11 | [39] |
| 11. | Pine | 36.2 | 23.0 | 32.8 | [40] |
| 12. | Cup plant | 39 | 21 | 21 | [41] |
| 13. | Sunn hemp fibre | 75.6 | 10.05 | 10.32 | [42] |
| 14. | Watermelon rind | 39.67 | 23.21 | 10.6 | [43] |
| | | | 13.2% xylan, 1.5% ga- | 30.2% Klason lignin, | |
| 15. | Peanut shell | 36.9% glucan | lactan, 5.2% arabinan, | 3.9% acid-soluble | [44] |
| | | | 1.0% mannan, | lignin, | |
| 16. | Corn cob | 41 | 22.6 | 14.1 | [45] |
| 17. | Barley straw | 31.1 ± 0.8 | 27.2 ± 0.4 | 18.8 ± 0.2 | [46] |
| 18. | Corn stover | 31.5% glucan | 22.5% xylan, 2.1% galactan, 1.7% arabinan | 18 | [47] |

3.1. Cellulose

Cellulose ($C_6H_{10}O_5$)_x is an unbranched linear polymer of β -D-glucopyranose units, joined together by β -(1,4) glycosidic linkages [48]. It is the primary component of lignocellulosic biomass which accounts for 30-50% of the whole dry mass [2]. The degree of polymerization (DP) of cellulose fibrils is approximately greater than 10,000. The uniform structure of cellulose fibrils is formed by inter- and intramolecular hydrogen bonding as well as covalent interactions which offers great tensile strength to cellulose, and make it resistant for various solvents [33]. Due to the presence of hydrogen bonding between hydroxyl groups in cellulose, different crystalline patterns with varying degrees of crystallinity are produced.

The cellulose polymers are separated from one another and more likely to form hydrogen bonds with other molecules in the more amorphous regions. Due to these two distinct molecular configurations, the polymer has a paracrystalline structure, where both the amorphous and crystalline portions are present [49]. Amorphous cellulose, which is easier to access, can be easily hydrolyzed by cellulase, while the harder-to-access crystalline part cannot be effectively broken down [36]. The important bio-based products obtained from the processing of cellulose include ethanol, succinic acid (SA), levulinic acid (LA), sorbitol, hydroxyacetaldehyde, and many others [2].

3.2. Hemicellulose

Hemicellulose (C5H8O4)m, the most rich carbohydrate polymer after cellulose, constitutes about 15-35% of total biomass. It is structurally a heteropolymer with less polymerization degree, and molecular weight as compared to cellulose [22,26]. It contains both branching and linear polymers that are made from a variety of anhydrous sugars (xylose, arabinose, D-glucoronic acid, L-rhamnose, and others). Since, the hemicellulose components have amorphous nature, and low DP, they do not significantly increase the biomass' recalcitrance [1,48]. Thus, it can be separated rapidly, and utilised to make biofuels like bioethanol, and other value-added products including lactic acid, xylo-oligosaccharides, xylitol, hydroxymethylfurfural (HMF), polyhydroxyalkanoates, and furfural [2]. The most prevalent hemicellulose polysaccharides are known as xylans, which have polymeric

chains made up of 1,4-linked -D-xylose units [50]. Depending on its source, xylan has a different chemical makeup. Hemicellulose also contains mannan components such as glucomannan, galactomannan, glucuronic acid, and galacturonic acid. Mannose residues are connected by β -1,4-bonds, whereas galactomannan is formed when mannose residues use α -1,6-bonds to link the galactose residues.

3.3. Lignin

Lignin [C₉H₁₀O₃(OCH₃)_{0.9-1.7}] n is primarily one of the crucial components of plant cell walls that offers them stiffness and strength while guarding against microbial attack [1,51]. It is an insoluble, stiff, aromatic biopolymer made up of monolignols like sinapyl alcohol, p-coumaryl alcohol, and coniferyl alcohol [19,52]. The presence of these components varies significantly, depending on the type of plant and extraction technique, with softwood and hardwood having higher (25-31%) and lower percentages (16-24%) of lignin, respectively [14,16,34]. With a projected accessibility of more than 3×10¹¹ tonnes, or around 30% of all non-fossil carbon, it is a rich organic non-fossil carbon present on the planet. Due to its abundance availability, lignin macromolecule represents a potential source for the production of fine chemicals, adsorbents, fuels, and polymer processing materials [53]. Lignin generally forms a three-dimensional cross-linked network by integrating with cellulose and hemicellulose components of LCB.

The common interunit linkages in lignin include β -aryl ether (β -O-4'), resinol (β - β '), dibenzodioxocin, phenylcoumaran (β -5'), α -O-4', 4-O-5', β -1 linkages [52]. Ether bonds, which make up roughly 60–75% of all bonding in lignin and are a common type of bond accounting for approximately 45–62% of all connection types, are the main type of bond used to connect the various structural units [1]. Lignin has long been considered a road-block in the lignocellulosic biorefineries due to its non-productive binding with the cellulolytic enzymes [54]. However, the last ten years has seen a paradigm shift in lignin research where it is now considered a bridge for biorefineries and an essential natural polymer for value-addition that can achieve the targets of circular bioeconomy in future.

3.4. Other components

Apart from cellulose, hemicellulose and lignin polymers, LCB is also constituted by proteins, pectins, little amount of extractives like gums, terpenoids, fatty acids, chlorophyll, other phenolic substances, and mineral rich ash (K, Mg,Ca, Si) [33]. Contrary to lignocellulosic content, these components provide colour and smell to the wood, serve as energy storage units, and shield the plant species from microbial invasions [21]. The non-structural parts of lignocellulosic biomass known as extractives can be dissolved in water or neutral organic solvents like hexane and ethanol. Various biopolymers, including terpenes, terpenoids, proteins, fats, waxes, lipids, steroids, and phenolic substances including lignans, stilbenes and tannins are found in extractives [50]. Ash is a solid, inorganic residue that remains after all LCB deposits have been completely burned [55]. It contains elements and minerals such silicates, carbonates, phosphates, calcium, magnesium, sodium, and potassium [56]. In contrast to agricultural wastes, woody biomass often has a very low ash content [50]. For the prediction of deposits forming in the boilers during gasification or incineration, it is crucial to have a thorough understanding of the physical and chemical features of lignocellulosic ashes.

4. Recent advances in pretreatment technologies for lignocellulosic biomass

The highly complex and recalcitrant structure of lignocellulosic biomass is the biggest barrier which comes under 2G biorefineries [57]. The intermeshed matrix of LCB biopolymers (cellulose, hemicellulose, and lignin) provides resistance to various microbial and enzymatic digestions. The other variables which affect the biomass recalcitrance include the biomass porosity, and the presence of proteins, and acetyl groups [58]. Thus,

fractionation of lignocellulosic components becomes essential for the extraction of monomeric sugars to produce biopolymers, biofuels, and bio-chemicals [1,3,32]. Pretreatment of LCB is a very significant step in a biorefinery, which enables the efficient disintegration of biomass, increases the permeability, and surface area availability of polysaccharides towards enzymatic hydrolysis. A desirable pretreatment aims to reduce biomass crystallinity, improve porosity, remove lignin, prevent inhibitors formation, avoid sugar degradation, requires less energy input, and less cost [35,59]. A variety of conventional pretreatment strategies have been developed for the disruption of biomass such as biological, physical, chemical, and physiochemical (Figure 2) [1,50]. Majority of these methods, however, are associated with few limitations such as high operational expense, harsh operational conditions (pressure, pH, and temperature), toxic product formation, and many others [13,14,60]. Hence, novel eco-benign pretreatment techniques are needed to be developed to overcome the constraints of traditional methods.

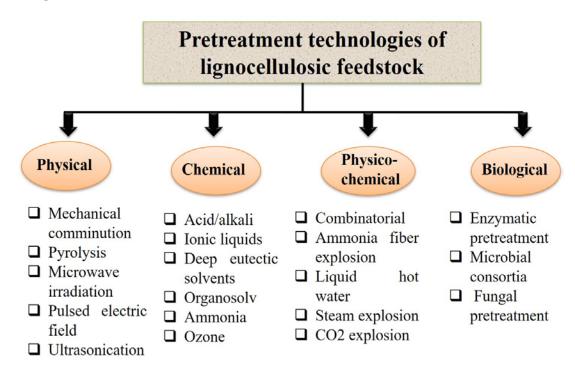


Figure 2. Diferrent pretreatment technoligies for breaking the inherent realcitrant structure of lignocellulosic biomass.

4.1. Physical Pretreatment

Physical pretreatment techniques primarily aim to decrease the particle size of LCB, and increase the surface accessibility of polysaccharides [21,61]. Mechanical comminution (chipping, milling, and/or grinding), freezing, pyrolysis, microwave irradiation, pulsed electric field, ultrasonication, and torrefaction are several pretreatment approaches which are being employed to destruct the lignocellulosic biomass [48,61,62]. However, most of these methods necessitate high energy/power consumption, high-cost processing instruments/equipments, and also produce hazardous chemical compounds. Thus, they are considered less attractive for industrial scale usage [4,21,33,60]. But even so, various methods including ultrasound and microwave irradiations have been developed as very efficient in the decrystallization and solubilization of cellulose and lignin from lignocellulosic feed-stock.

4.1.1. Ultrasound pretreatment

Ultrasound irradiation assistant pretreatment includes application of high frequency (20-500 MHz) mechanical acoustic waves which produce high shear force for proficient

defibrillation, and degradation of lignocellulosic biomass [17]. Ultrasound pretreatment method is based on the basic principle of 'cavitational effect', in which the ultrasonic waves aid in breaking α -O-4 and β -O-4 lignin linkages, which results in the development of cavitation bubbles by fractionating the ligno-polysaccharides complex of biomass [6,8]. When these bubbles reach a threshold size, they rapidly collapse, causing a significant rise in temperature (2000–5000 K) and pressure (1800 atm) [22]. It also offers a number of benefits including less residence duration, high levels of activation energy, and adequate mass transfer for efficient LCB deconstruction [61].

He et al. [63] investigated the structural variations of eucalyptus wood due to ultrasound pretreatment assisted with distilled water, aqueous acetic acid, or aqueous soda solution at 28 KHz frequency and 300W power. It was observed that the proportion of alkali metals was significantly reduced after ultrasound pretreatment. There was an increase in the crystallinity (35.5%) of wood samples after pretreatment which indicated the successful disintegration of wood biomass. Subhedar et al. [64] reported pretreatment of pistachio shells, groundnut shells, and coconut coir using ultrasound irradiations (0.5% w/v biomass loading, 100W power, and 70 min pretreatment time). Compared to alkaline pretreatment, the delignification percentage of LCB was significantly higher (80-100%) in case of ultrasound assisted method. At optimal parameters, the sugar yield obtained through ultrasound assisted digestion was found to be greater for Pistachio shells, groundnut shells, and coconut coir i.e. 18.4, 21.3, and 23.9 g/L, respectively, as compared to alkali hydrolysis i.e. 8.1, 10.2, and 12.1 g/L, respectively in similar biomass order. In another study conducted by Kunaver et al. [65], high frequency ultrasound irradiation approach was employed for the liquefaction of different forest wood waste residues into value-added products. The results demonstrated the high production of low-molecular weight residual products by using short reaction time (decreased by factor 9), and less energy/power consumption. Thus, ultrasound technology (standalone/integrated) has been the top choice for effective LCB pretreatment.

4.1.2. Microwave irradiation

This irradiation method involves non-ionizable, low-frequency electromagnetic radiations called microwaves produced by cycling off of a dipole (two opposite and equal charges separated by a particular distance) [21]. Microwave pretreatment exhibits an excellent potential in the bioprocessing of lignocellulosic biomass as the heat produced by the technique raises the temperature and pressure (thermal effect), throughout the biomass rather than just on its surface, demonstrating greater energy efficiency than traditional heating methods [66]. According to Mankar et al. [61], LCB acts as a good biological conductor due to the presence of non-ionic amorphous, and ionic crystalline cellulose regions. Temperature, initial feedstock components, irradiation time, microwave power, and catalyst loading all had a discernible impact on the microwave-physicochemical processing of biomass [67]. Application of microwave irradiations heats the polar structure of LCB, and leads to the formation of hotspots. Consequently, constant blasts occur inside the biomass structure, which accelerates the reorientation of crystalline cellulose fibrils [66]. In particular, microwave heating has a number of benefits, including short reaction time, fast and homogenous heating, low activation energy, increased product yield, few by-products, environmentally friendly, low-cost, and energy effective operation [67].

Nuchdang et al. [68] investigated the effect of microwave pretreatment for *Brachiara mutica* (paragrass) biomass, treated with acid and alkali. The results showed that use of microwave irradiation at 120°C, 30 min, and 5% w/w alkali to biomass ratio increased the total reducing sugars significantly (137.3%), as compared to native biomass. Kainthola et al. [69] reported highest solubilization of rice straw biomass under the influence of microwave irradiations (for 4 min, at 190°C), resulting in the high specific yield of methane i.e. 325.2318 mL/g/VS. Short reaction time and high heating rate reduced the production of inhibitory products, thus improved the efficacy of the pretreatment process. In another

study, microwave assisted FeCl₃, H₂SO₄, and NaOH pretreatment was carried out for sugarcane bagasse biomass to extract the fermentable sugars. The results demonstrated high lignin removal, high sugar yield including removal of glucose and xylose selectively, at short residence time of 5 to 10 minutes [70]. Despite the amazing advantages of microwave irradiation, the development of industrial microwave heating applications and the incorporation of microwaves into the physicochemical approach for biomass preparation are relatively limited. Additionally, the operation conditions for microwave assisted pretreatment should be tuned to extract the polymeric sugars (cellulose/hemicellulose) as much as possible.

4.2. Chemical pretreatment

This type of pretreatment involves the usage of various chemicals such as acids, alkali, deep eutectic solvents, ionic liquids, ammonia, ozone, and many others for the disruption of recalcitrant lignocellulosic biomass [1,14,32]. Chemical pretreatment majorly aims to decrease the biomass crystallinity, increase its porosity, remove lignin, and thus, enhance the biomass surface availability towards enzymatic hydrolysis [59]. Acids (concentrated and dilute) cleave hemicelluloses fraction, and remove a part of lignin of LCB, thus provides greater cellulose access to hydrolysis by enzymes [71]. But, most of the acids are associated with various limitations such as sugar loss, high cost, corrosive nature, and inhibitors production [22]. Morais et al. [72] carried out pretreatment of sugarcane bagasse with dilute phosphoric acid that under optimal pretreatment conditions (4.95% phosphoric acid, 80°C, 375 min) yielded maximum reducing sugars (98% glucose) with higher saccharification efficiency (99%). Alkaline pretreatment leads to the delignification of biomass by cleaving aryl-ether, ester, and C-C bonds, removes hemicelluloses, and increases the surface area for efficient hydrolysis and fermentation [21,73]. However, long reaction time, and high expense for neutralization of pretreatment slurry are the major limitations of the alkaline pretreatment [61]. A mild-alkali (sodium hydroxide) mediated pretreatment of Cogongrass was performed at 2% w/v NaOH concentration, 90 min time, and 85°C [74]. The results illustrated higher saccharification efficiency (90.8%) after alkali pretreatment, which on fermentation yielded maximum ethanol (76%). Although traditional chemicals (acids/alkalis) results in improved decrystallization and cellulose accessibility in a biorefinery, but they also have a number of drawbacks that prevent their usage on a large scale [75]. These pretreatment techniques' principal downsides include the high expense of recovering the acid/alkali, the requirement for corrosive-resistant equipment, and the production of inhibitory compounds. Therefore, the pursuit of new eco-friendly solvents has been intensified in the recent years.

4.2.1. Ionic liquids (ILs)

Ionic liquids (ILs) have effectively become the most environmentally friendly and reusable organic solvents for LCB processing due to their numerous extraordinary tuneable characteristics [12,17,52]. ILs are organic salts of cationic and anionic species, have lower melting point, non-inflammable, high chemical and thermal stability, and low vapour pressure [58]. ILs mediated pretreatment solubilize the polar and non-polar molecules of ligno-polysaccharides, decrease cellulose crystallinity, increases porosity, eliminate the lignin sheath, and thus provide larger cellulose accessibility to saccharification enzymes (Halder et al., 2019). Several imidazolium based ionic liquids have been used for pretreatment of lignocellulosic biomass.

Alayoubi et al. [36] carried out low temperature (45°C) IL, 1-ethyl-3-methylimidazolium acetate pretreatment of oak sawdust, spruce sawdust, and model cellulose, which on saccharification produced high glucose yield i.e. 59.3%, 49.3%, and 68.2%, respectively. Dotsenko et al. [76] reported an effective delignification, and biotransformation of hardwood (hornbeam wood and spruce), and softwood biomass samples, after pretreatment

with ionic liquid 1-butyl-3-methylimidazolium chloride ([Bmim]Cl). Pretreatment of sugarcane tops biomass using ionic liquid tris (2-hydroxyethyl) methylammonium-methyl-sulfate ([TMA]MeSO₄), followed by saccharification resulted in high sugar yield i.e. 181.18 mg/g biomass [13]. However, expensive synthesis, potential toxicity, recycling issues, and non-biodegradable nature are few limitations of ILs which limit their large-scale usage in industries [8,77].

4.2.2. Deep eutectic solvents (DESs)

Lately, DESs have gained a lot of interest as green chemistry approach for the digestibility of lignocellulosic biomass [1,8]. Deep eutectic solvents primarily consist of a mixture of liquid eutectic materials that include hydrogen-bond donors and acceptors at a particular molar ratio [78]. They exist majorly in liquid form, and possess lower melting point than their single constituents [15]. They exhibit almost similar characteristics with ILs including low vapour pressure, non-flammable, high thermo-chemical stability, high miscibility, high polarity, and thus, can be efficiently used for the fractionation of LCB [61,78]. However, easy synthesis, low-cost, non-toxicity, good recyclability, and increased biodegradability make them preferable solvents over ILs. DESs solubilize lignin effectively from LCB, and transform the crystalline portion of cellulose into amorphous one at moderate process conditions [1]. Deep eutectic solvents' promising properties have forced and encouraged their subsequent use for successful LCB bioconversion, making them superior to diluted acids and ionic liquids.

Jing et al. [79] investigated the production of biohydrogen from corn cob biomass by using deep eutectic solvent (ChCl/ethanolamine) system. The pretreatment resulted in maximum removal of lignin with high percent efficiency (83.12%). Tan et al. [80] carried out pretreatment of oil palm empty fruit bunch (EFB) biomass with choline chloride/formic acid (CC-FA), and choline chloride/lactic acid (CC-LA) solvent system. The results demonstrated improved biomass fractionation with high delignification efficiency of 60 wt%. Thi and Lee [81] investigated the effect of three DESs (choline chloride-urea, ChCl-U; chloride-lactic acid, ChCl-LA; choline chloride-glycerol, ChCl-G) on the pretreatment of oil palm empty fruit bunch biomass. The results illustrated that ChCl-LA pretreatment, followed by saccharification gave highest sugars yield (20.7%), followed by ChCl-G and ChCl-U pretreatment which yield sugar content of approximately 20.0% and 16.9%, respectively. To further improve the effectiveness of the DES pretreatment and maximize the yield of reducing sugars, a thorough investigation of the operating parameters and techno-economic analysis of the process is required.

4.2.3. Organosolv pretreatment

Organosolv pretreatment is one of the promising methods for the effective valorization of lignocellulosic biomass into value-added chemicals that could facilitate the shift towards increased use of renewable biomass substrates. Organosolv pretreatment uses organic solvents (ethanol, methanol, formic acid, acetic acid, phenol, glycerol, or, acetone) or their aqueous solutions to specially extract or remove lignin from lignocellulosic feed-stocks [82]. The organosolv pretreatment provides for highly efficient generation of ethanol, lignin and other biochemicals from biomass when compared to other pretreatment techniques. In this pretreatment, LCB is added to a combination of organic solvent and water that has a solid-to-liquid ratio of between 1:4 and 1:10 (w/w) and a solvent concentration of 35%–70% (w/w) [83]. The rate of pretreatment reaction can also be accelerated by including a catalyst. The pretreatment causes the lignin linkages and lignin-carbohydrate bonds to hydrolyze resulting in a solid phase primarily consisting of cellulose and hemicellulose. To avoid lignin precipitation, this prepared material must be rinsed with an organic solvent. Thereafter, the pretreated biomass is washed in distilled water to remove the organic solvent, and the solid and liquid phases are separated. The pretreated

solids are subsequently enzymatically saccharified fermented to produce value-added biochemicals, whereas the solvent is recovered from the spent liquor using distillation process (Figure 3).

Karnaouri et al. [84] pretreated pine and beechwood biomass with a mild oxidative organosolvs (tetrahydrofuran, acetone and ethanol) that produced cellulose-rich solid fractions for lactic acid production from *Lactobacillus deLCBrueckii* strain. A high lactic acid production of 62 g L $^{-1}$ was obtained from beechwood biomass compared to that obtained from pine biomass (36.4 g L $^{-1}$) showing that organosolv pretreatment is suited for both hardwood and softwood biomass for the production of valuable biochemicals. In another recent study [85], the sorghum biomass was pretreated by modified low-cost organosolv using glycerol solvent and ammonia as catalyst that resulted in a significant improvement in the saccharification (reducing sugars of 421.35 mg/g biomass) and fermentation yields (36 g/L). Tang et al. [86] executed synergetic organosolv and organic amine pretreatment of corn stover biomass to produce salt free high-quality lignin and fermentable sugars. Under optimal process conditions, maximum sugar yield of 83.2%, contributed by 87.1% glucose, 75.4% xylose was obtained with a significant delignification percentage of 83.2% by the application of aqueous ethanol (60%, v/v) as a solvent, and *n*-propylamine (10 mmol/g, biomass) as a catalyst.

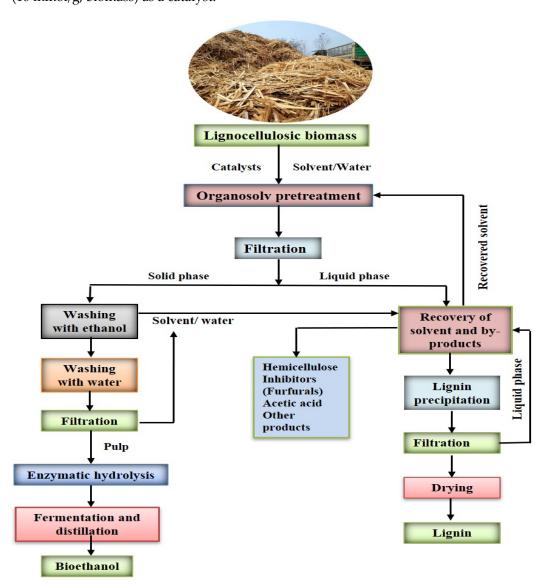


Figure 3. Schematic representation of steps in orgaosolv pretreatment of lignocelluloci feedstock.

The promising results with the organosolv pretreatment has led the establishment of some of the organosolv-based biorefineries in developed countries like United states of America, Brazil, Germany, Australia, France and Canada [83]. The further setting up of organosolv-based biorefineries would need an integration of LCB components into pretreatment processes and the reduction of pretreatment costs. This can be accomplished by lowering the volume of organic solvent used in pretreatment, raising the added value of by-products and optimizing the entire process using statistical approaches.

4.3. Physicochemical pretreatment

The physico-chemical pretreatment technique involves the combination of both chemical and physical methods, and is believed to be more efficient in the disintegration of lignin-polysaccharide complex at varying process conditions i.e. temperature and pressure [87]. Various methods including carbon dioxide explosion, steam explosion, liquid hot water method, ammonia fiber explosion, wet oxidation, and different combinatorial approaches comes under physico-chemical methods [19,88]. Steam explosion is the most common method for LCB pretreatment in which an autohydrolysis reaction is carried out by high pressure saturated steam which decreases the biomass crystallinity, removes lignin, and transforms cellulose and hemicellulose into soluble oligomers [62]. In CO₂ explosion pretreatment, dissolution of LCB is caused due to the application of high-pressure supercritical carbon dioxide at high temperature. Ammonia fiber explosion and CO₂ explosion work at low temperature, thus, are considered more economical [89]. The integration of different techniques such as acids, alkali, DESs, ILs, combined with ultrasound, microwave, or others have been employed by various researchers to improve the effectiveness of the process [8,14,16].

Kuglarz et al. [90] carried out combined pretreatment of industrial hemp biomass with steam at 180°C, and 1% H2SO4, which resulted in the production of maximum glucose yield of 73-74%, and an ethanol content of 75-79%. Nargotra et al. [34] reported an enhanced sugar yield of 63.42 mg/g biomass by the combinatorial pretreatment (ionic liquid, 1-butyl-3-methyl imidazolium chloride+ alkali, NaOH) of sunflower stalk biomass, as compared to IL (79.6 mg/g biomass), and NaOH (97.38 mg/g biomass) in standalone mode. Vaid et al. [91] reported combined application of DES (choline chloride/glycerol) and calcium hydroxide for the pretreatment of Saccharum spontaneum biomass, followed by enzymatic hydrolysis under one pot consolidated bioprocessing. The results demonstrated maximum sugar yield of 372.3 mg/g biomass which was 4.94-fold higher than the sugar yield obtained under unoptimized conditions (75.25 mg/g biomass). In another study conducted by Hou et al. [92], eucalyptus sawdust was pretreated by combined application of microwave and IL [TBA][OH], which on hydrolysis yielded maximum sugar yield of 410.67 mg/g biomass in 48h at optimal pretreatment conditions. Vaid et al. [32] employed a novel pretreatment approach using sodium dodecylsulfate assisted IL tris (2hydroxyethyl) methyl- ammonium methyl sulphate for the deconstruction of Saccharum spontaneum biomass. Hydrolysis with IL stable Bacillus subtilis G2 enzymes yielded maximum reducing sugars of 364.24 mg/g biomass at optimized process conditions, in comparison to unoptimized (154.91 mg/g biomass).

4.4. Biological Pretreatment

Biological pretreatment entails the utilization of microorganisms, mostly fungi and bacteria, which have the ability to produce extracellular lignocellulolytic enzymes for the degradation of lignocellulosic biomass (Sindhu et al., 2016). Brown, white and soft rot fungi such as *Dichmitus squalens, Phanerochaete chrysosporium, Jungua separabilima, Phlebia radiata*, and *Rigidosporus lignosus* are generally used for the digestion of polysaccharides [19,33]. This pretreatment method is regarded as an efficient, affordable, and environmentally friendly process as it does not employ high energy inputs, and produce less harmful substances, compared to other chemical strategies [93]. However, less available surface

area due to biomass structural complexity, slow digestion rate, and requirement of monitored microbial growth conditions restrict the use of biological pretreatment at commercial level [62,94].

5. Enzymatic hydrolysis as an integral step in biorefineries

Enzymatic hydrolysis of pretreated lignocellulosic feedstock is the key step in LCB-biorefinery chain which involves the transformation of polymeric carbohydrates into simple fermentable monomers with the help of saccharification enzymes such as cellulases, hemicellulases, and ligninases [13,32,54,60]. The three major cellulolytic enzymes employed to degrade cellulose are a) endo-1,4- β -D-glucanases (hydrolyze low-crystalline cellulose chains randomly); b) cellobiohydrolases/exoglucanases (hydrolyze cello-oligosaccharides' reducing and non-reducing ends to generate cellobiose units), and c) cellobiases/ β -glucosidases (hydrolyze cellobiose to form D-glucose) [2,3,35]. All the three cellulases work simultaneously and synergistically as a mixture, thus increase the catalytic activity of cellulase enzymes [95].

Besides cellulases, hemicellulases such as β -xylosidase, xylanases, glucomannanase, glucuronidase, galactomannanase, and acetylesterase are employed for the hydrolysis of hemicellulose polymers [22,96,97]. Xylanases solubilize β -1,4-D-xylan fractions of hemicellulose by cleaving β -1,4-D-xylopyranosyl bond, and thus, results in the formation of oligomeric (xylo-oligosaccharides), and monomeric sugars [98,99]. Another class of enzymes named lytic polysaccharide monooxygenases (LPMO) work synergistically with cellulases, and thus, boosts the effectiveness of cellulose degradation [3,22]. LPMOs are copper-dependent enzymes which follows an oxidative mechanism to improve the disintegration of biomass. In the presence of cellobiose dehydrogenase and ascorbic acid (electron donors), copper ions cause reduction of oxygen, thus aid in the cleavage of glycosidic linkages of crystalline cellulose [35,57].

Lignin, an aromatic polymer of LCB acts as a physical barrier in the process of poly-saccharides extraction [1,3,14]. It is degraded by a variety of ligninolytic enzymes such as laccases, versatile peroxidase manganese peroxidase, and lignin peroxidase [100]. Microorganisms (fungi and bacteria) are commonly used for the production of hydrolytic enzymes. Micro-organisms such as *Bacillus velezensis*, *Bacillus paranthracis*, *Aspergillus terreus*, *Aspergillus niger*, *Trametes versicolor*, and *Phanerochaete chrysosporium* are few of the examples which produce enzymes with cellulolytic, hemicellulolytic, and ligninolytic capabilities [2,100]. Various factors such as enzymatic activity of saccharification enzymes, enzyme loading, substrate quantity, process conditions, and inhibitors present in the post-pretreatment sugar hydrolysate [89]. However, addition of various salts/surfactants/metal ions, optimization of process parameters, and utilization of enzyme mixture increase the efficiency of enzymatic hydrolysis [101].

In a study conducted by Cai et al. [102], the addition of polyvinylpyrrolidone (1 g/L) during enzymatic saccharification of dilute acid pretreated eucalyptus biomass increased its digestibility to 73.4% from 28.9%, as compared to other additives (bovine serum aLCBumin, PEG, and Tween). Rocha-Martin et al. [103] reported an improvement in the glucose yield by 32%, 10%, and 7.5% after PEG4000 assisted hydrolysis of pretreated microcrystalline cellulose (Avicel), corn stover, and sugar cane straw biomass, respectively. Addition of PEG4000 increased the endoglucanase and beta-glucosidase catalytic activity by 60% and 20%, respectively and also decreased the liquefaction time (up to 25%).

Although enzymatic hydrolysis is a crucial step in the bio-transformation of LCB at commercial level, application of enzymes still has a number of drawbacks, including high cost, limited availability, less stability, non-recyclability, and non-reusability; thus, limit the effectiveness of a bioprocess [104]. To circumvent these problems, immobilization of microbial enzymes on suitable matrices/support materials provides a promising solution [105]. Nanomaterials (with a size of less than 100 nm) such as nanopartricles, nanofibers, nanoflowers, nanomagnets, and many others have the potential to completely transform

the production of biocatalysts and their application in the bioenergy area [3]. Nanobiocatalysts provides a number of benefits to enzymes including high surface area, increased activity, increased stability, reusability, easy-synthesis, environment friendly, and costeffective nature [106,107]. Furthermore, enzyme engineering or genetic manipulation of genes coding for saccharification enzymes can improve the catalytic activity, and stability of enzymes [3]. The production of competent and resistant micro-organisms, which can survive harsh environments, and development of stable enzymes require the potential application of genetic engineering tools [24].

Karnaouri et al. [108] produced an enzyme called MtEG5A expressed from the endoglucanase gene of Myceliophthora thermophile, a thermophilic fungus, into Pichia pastoris, the methylotrophic yeast. These enzymes have been found to hydrolyze birch, wheat straw, and spruce biomass, producing large quantities of cellobiose and hence increased yield of sugars. In a study, cellulases immobilized on magnetic nanoparticles were utilized for the hydrolysis of Allamanda schottii L flowers pretreated with sodium hydroxide, and highest sugar yield of 25 g/ml was observed in comparison to that with native enzyme i.e. 18 g/ml. Fermentation of the obtained sugar hydrolysates yielded an ethanol content of 252 g/L (immobilized enzyme), and 182 g/L (native enzyme), respectively [109]. Hwangbo et al. [110] investigated the combined pretreatment of NaOH and IL, 1-ethyl-3-methylimidazolium chloride of corn husks, hydrolyzed with five types of saccharifying enzymes which were cross-linked in the presence and absence of magnetic nanoparticles called as magnetite-saccharifying enzyme cross-linked enzyme aggregates, M-SE-CLEAs; and saccharifying enzyme cross-linked enzyme aggregates, SE-CLEAs. The results indicated maximum reducing sugar of 250 mg/g biomass by employing SE-CLEAs. Additionally, reusability of M-SE-CLEAs was successfully achieved up to 3 hydrolysis cycle.

6. Bioconversion of lignocellulosic biomass into biofuels and value-added materials

Carbohydrate rich lignocellulosic biomass obtained from agricultural by-products, forestry residues, municipal wastes, and industrial emissions holds a remarkable potential for the production of green energies i.e. biofuels (bioethanol, biohydrogen, biomethane, biobutanol, biogas), and other value-added products through various aforementioned thermo-chemical and biochemical conversion technologies [1,58,78]. Enzymatic digestion of polysaccharides results in the production of reducing sugars, which on fermentation produces various kinds of biofuels. A biochemical process which catalyzes the conversion of monomeric carbohydrates with the help of micro-organisms under low oxygen environment is known as fermentation [25]. Most widely employed industrial microorganisms for the fermentation process are Schizosaccharomyces, Saccharomyces cerevisiae, Candida shehatae, Zymomonas mobilis, Pichia stipitis and others [89]. For microorganisms to thrive, all the internal conditions (dissolved oxygen, culture medium, and other micronutrients), and necessary parameters (pH, temperature, and sugar content) must be satisfied. They must possess high product yield, and resistance to the toxic inhibitory compounds [111]. The conventional method used for fermentation of biomass hydrolysate has a specific process in which the fermentation is carried out in several types (batch, fed-batch, solid state, consolidated bioprocessing, simultaneous saccharification and fermentation, and separate hydrolysis and fermentation) [22,26]. The processed lignocellulosic biomass after several rounds of steps including pretreatment, enzymatic saccharification act as a suitable substrate for the production number of bioproducts such as biofuels, bioplastics, vanillin, hydroxymethylfurfural, etc. (Figure 4) [1,3,96,112].

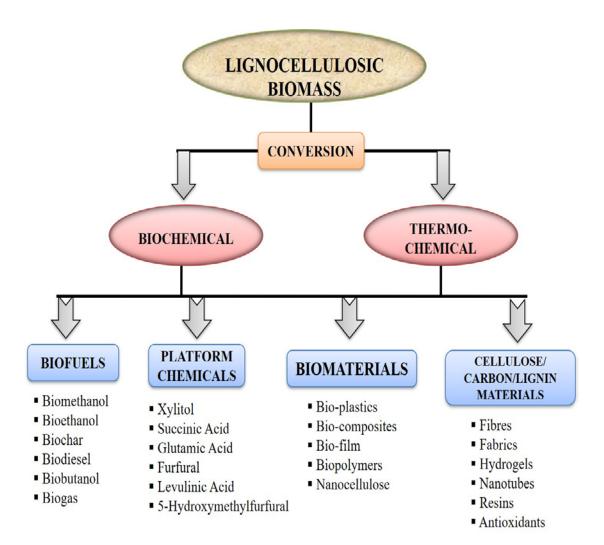


Figure 4. Biochemical and thermochemical conversion of lignocelluloisc substrate into biofuels and value-added bioproducts.

6.1. Bioethanol

Biofuel-ethanol (C2H5OH) is a promising green combustible fuel over gasoline due to various characteristics like higher flammability, high octane number, higher combustion efficiency, high heat of vaporization, shortened burning time, minimum toxicity, and less-polluting than petrol. It is usually mixed with gasoline and used as fuel in the transportation industry [26,113]. India contributes only 2% to the overall global bioethanol production [114]. For the production of bio-ethanol, LCB is first processed into simple sugars (hexoses and pentoses), and then metabolized into ethanol by using fermenting microorganisms such *Saccharomyces cerevisiae*, and *Pichia stipitis* [2]. Jin et al. [115] executed ethanol fermentation of the sugary liquor obtained by hydrolysis of combinatorially (sodium hydroxide and hydroxymethylation reagents) pretreated sugarcane bagasse, and observed the maximum ethanol yield of 10.67 g/L. Vaid et al. [16] reported the highest ethanol concentration of 0.148 g/g pine needle biomass under consolidated bioprocessing of pine needle biomass pretreated with 1-ethyl-3-methylimidazolium methanesulfonate.

6.2. Biodiesel

Another promising alternative for non-renewable fossil fuels is fatty acid methyl ester, called biodiesel due to its sustainability, biodegradability, renewability, and environmentally friendliness, as compared to conventional diesel [116]. It is generally produced from edible and non-edible oils by transesterification, blending, pyrolysis, microemulsion,

or other conversion processes. Biodiesel can replace the fossil derived diesel directly without any modification in the existing engines, and is non-explosive, non-inflammable, and non-toxic [117]. Wang et al. [118] reported two insect (*Tenebrio molitor* L. and *Hermetia illucens* L.) based biorefinery of corn stover biomass, and observed the crude grease of 1.95 g which yielded significant quantity of biofertilizer (111.59 g), protein (6.55 g), biodiesel (1.7g). Bateni et al. [119] carried out alkali pretreatment of castor plant at optimal conditions to improve the bioethanol and biodiesel yield which was obtained to be 27.2 to 71% and $85.0 \pm 1.0\%$, respectively. It was estimated that 149.6 g of biodiesel and approximately 30.1 g of bioethanol was observed from 1 kg of castor plant.

6.3. Biomethane

LCB biopolymers can be digested anaerobically by bacteria to produce an easily storable and flexible biofuel named 'biomethane' or 'biogas' [11]. Firstly, the complex lignopolysaccharides are processed with suitable pretreatment strategy into simpler molecules of amino acids, sugars, and fatty acids. Acidogens (fermentative bacteria) digest these monomers to short chain fatty acids, and then convert into acetate, hydrogen and carbon dioxide. These end products act as substrates to produce biomethane via methanogenesis (Xu et al., 2019). Recently Banu et al. [120] reported maximum biomethane yield of 0.174L/g COD from waste activated sludge pretreated by bacterial pretreatment induced by mild hydrogen peroxide, as compared to methane obtained with bacterial pretreatment i.e. 0.078 L/g COD, and control sludge i.e. 0.02175 L/g COD.

6.4. 5-. Hydroxymethylfurfural (HMF)

5-HMF is generally considered as a multifunctional molecule due to the presence of a furan ring structure joined with an alcohol, and aromatic aldehyde [121]. 5-HMF can produce a variety of derivates including 2,5-diformylfuran (2,5-DFF), levulinic acid, 2,5-dimethylfuran (2,5-DMF), and 2,5-bis(hydroxymethyl) furan (2,5-BHF) [122]. These compounds have been used as precursors in the production of materials like polyesters, polyamides, and polyurethane [123]. [108] Ruby and Schüth [124] reported 5-hydroxymethylfurfural production by fructose dehydration with the application of acid-free and metalfree heterogeneous catalytic system. The results observed 77% of maximum HMF yield. Zhao et al. [125] carried out the as-synthesized solid acid catalyst mediated fructose degradation to produce HMF with maximum yield of 90% under optimal process conditions.

7. Environmental sustainability of biofuels, and value-added biochemicals from lignocellulosics

The continuous emission of greenhouse gases (GHGs) due to combustion of fossil derived fuels lead to an enormous increase in the global temperature causing global warming, and other environmental related problems like pollution, climate change, and oil spills [26]. An exponential rise in global population and depletion of conventional fossil resources cause the upsurge need of energy supply worldwide [126]. Valorization of abundantly available lignocellulosic biomass waste into biofuels and value-added products promises an effective solution. Biofuels, which are made from non-fossilized biomass (plant products) or other organic solid wastes, offer the advantage of lowering net carbon emissions, primarily GHGs, and dependency on oil [114]. Increasing the use of biofuels would benefit several important policy goals such as energy security, fuel quality and environment friendly. Biofuels can easily replace petroleum-based fuels and, in many nations, can serve as an indigenous source of transportation fuel rather than one that must be imported. However, if imported, biodiesel or ethanol will probably originate outside of OPEC (Organisation of Petroleum Exporting Countries), diversifying the supply of transportation fuels on a more global scale.

The advantages of using ethanol to increase fuel octane have caught the attention of refiners and automakers, particularly in cases where MTBE and other possible octane enhancers are restricted or banned. Biofuels are often more environmentally benign than petroleum fuels due to fewer GHG emissions across the whole fuel cycle. Vehicles powered by biofuels release less of various pollutants that exacerbate air quality issues, especially in metropolitan areas, either in their pure form or as blends with conventional petroleum fuels. Combining biofuels with conventional fuels can reduce some air pollutants while increasing others (such as NOx emissions) [127]. The bioconversion of LCB into various value-added chemicals such as formic acid, succinic acid, lactic acid, levulinic acid, 2,5-dimethylfuran (2,5-DMF), 5-hydroxymethylfurfural (5-HMF), and other aromatic compounds can be transformed into a variety of upgraded derivatives which possess remarkable potential in biopolymers, biofuels, and solvent factories [122].

8. Current challenges and future prospects

Second generation biorefinery based on lignocellulosic feedstock is a renewable, sustainable, and eco-benign approach in view of deteriorating fossil fuels, increased energy prices and environmental ill-effects [58]. However, full scale bio-transformation of LCB into desired chemicals is still limited due to high operational and investment costs. This challenge is ascribed to LCB's intrinsically tough structure, which resists microbial invasion and enzyme access [12]. A comprehensive study on the characterization and composition of the lignocellulosic substrate is pre-requisite for the development of a successful biorefinery. The composition, availability, cost, production, collection, storage, and shipping are the factors which are taken into consideration while selecting lignocellulosic feedstock [2]. Bioconversion processes including biomass pretreatment and enzymatic saccharification are the key steps which affect the techno-economic feasibility of a biorefinery. As the green refinery concept is put into practise, it is necessary to maximise the costeffectiveness and quality of fractionated products. Despite improvements in a number of different pretreatment strategies, which produce the high sugar yield with less demerits, there is still a lot of room for enhanced LCB pretreatment and detoxification-based advancements [58]. Thus, adequate and cost-efficient pretreatment methods are needed to be developed which can cause maximum degradation of biomass, less chemical and energy consumption, and boost the subsequent enzymatic hydrolysis. The severity of biomass pretreatment influences the amount of enzyme required for the next step of hydrolysis [26]. Therefore, future research efforts should focus more on developing customized pretreatment techniques while considering various LCB compositions or the targeted products, as well as developing a solid understanding of the reaction mechanism underlying pretreatment.

Another major obstacle which hinders the feasibility of a biorefinery is the high operational expenses of hydrolytic enzymes. Enzymatic hydrolysis step though contributes about 25-30% of the total biomass processing cost, yet have not been fully commercialized for second-generation biofuel production. Therefore, exploration and production of affordable enzyme mixtures are required for the efficient hydrolysis process [89]. Several techniques such as metagenomics, metabolic engineering, and protein engineering can be used to discover the highly stable, and genetically modified hydrolytic enzymes, leading to the production of various desirable products [3,24]. Immobilization of potent enzymes on nanosized materials (organic wastes, nanoparticles, nanofibers, nanotubes, and others) provide a plethora of benefits to enzymes including high stability, recyclability, reusability, high catalytic activity, thus improve the cost effectiveness of 2G biorefinery [110]. Majority of the micro-organisms involved in fermentation can metabolize specific class of sugars, for example, Saccharomyces cerevisiae can ferment only hexoses, while Pichia stipitis can metabolize only five carbon sugars, leaving other residues unused (lignin, unreacted substrates, among others). To boost ethanol yield and productivity, it is crucial to investigate and create genetically modified cellulolytic and fermentative microorganisms and co-culture system. Future commercialization of biorefineries is just the top of the iceberg.

Due to the high cost of renewable energy produced by a single technology, combining various technologies on a large scale for commercial purposes would increase the yield of renewable energy and make it more economical. The integration of biorefineries to scale up the projects to close the gap between the generation and commercialization of biofuels, and value-added products is the subject of extensive research.

9. Conclusions

The concept of lignocellulosic biorefinery is developed by valorizing lignocellulosic biomass and its constituent parts into value-added products and enhanced energy. Introduction of process-efficient and low-cost technologies is the most essential requirement for sustainable and renewable energy production. It not only minimizes the negative environmental impact, but also cuts down on carbon emissions. The review encompasses detailed information on the lignocellulosic biomass, processing technologies, merits, demerits, and challenges associated with a biorefinery. For forest-based biorefineries to be incorporated into industrial supply chains, a sustainable and ecologically friendly solution for the valorisation of lignocellulosic biomass through cost-effective and sustainable processes must be developed.

Author Contributions: Conceptualization, V.S. and P.N.; Validation, V.S., M.-L.T., C.-W.C., P.N., and C.-D.D.; Formal Analysis, V.S., B.K.B.; M.-L.T., P.N., and C.-W.C.; Investigation, V.S., B.K.B. and C.-D.D.; Writing—Original Draft Preparation, S.S. and V.S.; Writing—Review & Editing, P.N.; V.S.; Visualization, V.S., M.-L.T., and C.-D.D.; Resources, M.-L.T.; Super-vision, M.-L.T., P.-P.S., and C.-D.D.; Funding Acquisition, C.-D.D. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by Taiwan MOST for funding support (Ref. No. 109-2222-E-992-002).

Data Availability Statement: Not applicable.

Acknowledgments: Authors V.S. and P.N. gratefully acknowledge National Kaohsiung University of Science and Technology, Kaohsiung City, Taiwan, for providing the post-Doctoral fellowship. Authors M.-L.T., C.-W.C., and C.-D.D. acknowledge Ministry of Science and Technology, Taiwan MOST for funding support (Ref. No. 109-2222-E-992-002).

Conflicts of Interest: The authors declare no conflict of interest.

References:

- Sharma, V.; Tsai, M.-L.; Chen, C.-W.; Sun, P.-P.; Patel, A.K.; Singhania, R.R.; Nargotra, P.; Dong, C.-D. Deep Eutectic Solvents as Promising Pretreatment Agents for Sustainable Lignocellulosic Biorefineries: A Review. *Bioresour. Technol.* 2022, 360, 127631, doi:10.1016/j.biortech.2022.127631.
- 2. Devi, A.; Bajar, S.; Kour, H.; Kothari, R.; Pant, D.; Singh, A. Lignocellulosic Biomass Valorization for Bioethanol Production: A Circular Bioeconomy Approach. *Bioenerg. Res.* **2022**, doi:10.1007/s12155-022-10401-9.
- 3. Sharma, V.; Tsai, M.-L.; Nargotra, P.; Chen, C.-W.; Kuo, C.-H.; Sun, P.-P.; Dong, C.-D. Agro-Industrial Food Waste as a Low-Cost Substrate for Sustainable Production of Industrial Enzymes: A Critical Review. *Catalysts* **2022**, *12*, 1373, doi:10.3390/catal12111373.
- 4. Weng, Z.-H.; Nargotra, P.; Kuo, C.-H.; Liu, Y.-C. Immobilization of Recombinant Endoglucanase (CelA) from Clostridium Thermocellum on Modified Regenerated Cellulose Membrane. *Catalysts* **2022**, *12*, 1356, doi:10.3390/catal12111356.
- 5. Periyasamy, S.; Beula Isabel, J.; Kavitha, S.; Karthik, V.; Mohamed, B.A.; Gizaw, D.G.; Sivashanmugam, P.; Aminabhavi, T.M. Recent Advances in Consolidated Bioprocessing for Conversion of Lignocellulosic Biomass into Bioethanol A Review. *Chem. Eng. J.* **2023**, *453*, 139783, doi:10.1016/j.cej.2022.139783.
- 6. Hassan, S.S.; Williams, G.A.; Jaiswal, A.K. Emerging Technologies for the Pretreatment of Lignocellulosic Biomass. *Bioresour. Technol.* **2018**, *262*, 310–318, doi:10.1016/j.biortech.2018.04.099.
- 7. Razzak, S.A.; Lucky, R.A.; Hossain, M.M.; deLasa, H. Valorization of Microalgae Biomass to Biofuel Production: A Review. *Energy Nexus* **2022**, *7*, 100139, doi:10.1016/j.nexus.2022.100139.

- 8. Sharma, V.; Nargotra, P.; Sharma, S.; Bajaj, B.K. Efficacy and Functional Mechanisms of a Novel Combinatorial Pretreatment Approach Based on Deep Eutectic Solvent and Ultrasonic Waves for Bioconversion of Sugarcane Bagasse. *Renew. Energy* **2021**, *163*, 1910–1922, doi:10.1016/j.renene.2020.10.101.
- 9. Sharma, V.; Nargotra, P.; Sharma, S.; Bajaj, B.K. Efficient Bioconversion of Sugarcane Tops Biomass into Biofuel-Ethanol Using an Optimized Alkali-Ionic Liquid Pretreatment Approach. *Biomass Conv. Bioref.* **2020**, doi:10.1007/s13399-020-01123-z.
- 10. Sharma, V.; Bhat, B.; Gupta, M.; Vaid, S.; Sharma, S.; Nargotra, P.; Singh, S.; Bajaj, B.K. Role of Systematic Biology in Biorefining of Lignocellulosic Residues for Biofuels and Chemicals Production. In *Sustainable Biotechnology-Enzymatic Resources of Renewable Energy*; Singh, O.V., Chandel, A.K., Eds.; Springer International Publishing: Cham, 2018; pp. 5–55 ISBN 978-3-319-95480-6.
- 11. Rajesh Banu, J.; Preethi; Kavitha, S.; Tyagi, V.K.; Gunasekaran, M.; Karthikeyan, O.P.; Kumar, G. Lignocellulosic Biomass Based Biorefinery: A Successful Platform towards Circular Bioeconomy. *Fuel* **2021**, 302, 121086, doi:10.1016/j.fuel.2021.121086.
- 12. Vu, H.P.; Nguyen, L.N.; Vu, M.T.; Johir, M.A.H.; McLaughlan, R.; Nghiem, L.D. A Comprehensive Review on the Framework to Valorise Lignocellulosic Biomass as Biorefinery Feedstocks. *Sci. Total Environ.* **2020**, 743, 140630, doi:10.1016/j.scitotenv.2020.140630.
- 13. Sharma, S.; Nargotra, P.; Sharma, V.; Bangotra, R.; Kaur, M.; Kapoor, N.; Paul, S.; Bajaj, B.K. Nanobiocatalysts for Efficacious Bioconversion of Ionic Liquid Pretreated Sugarcane Tops Biomass to Biofuel. *Bioresour. Technol.* **2021**, 333, 125191, doi:10.1016/j.biortech.2021.125191.
- 14. Nargotra, P.; Sharma, V.; Bajaj, B.K. Consolidated Bioprocessing of Surfactant-Assisted Ionic Liquid-Pretreated *Parthenium hysterophorus* L. Biomass for Bioethanol Production. *Bioresour. Technol.* **2019**, 289, 121611, doi:10.1016/j.biortech.2019.121611.
- 15. Nargotra, P.; Sharma, V.; Sharma, S.; Kapoor, N.; Bajaj, B.K. Development of Consolidated Bioprocess for Biofuel-Ethanol Production from Ultrasound-Assisted Deep Eutectic Solvent Pretreated *Parthenium hysterophorus* Biomass. *Biomass Conv. Bioref.* **2020**, doi:10.1007/s13399-020-01017-0.
- 16. Vaid, S.; Nargotra, P.; Bajaj, B.K. Consolidated Bioprocessing for Biofuel-Ethanol Production from Pine Needle Biomass. *Environ. Prog. Sustain.* **2018**, *37*, 546–552, doi:10.1002/ep.12691.
- 17. Sharma, V.; Nargotra, P.; Bajaj, B.K. Ultrasound and Surfactant Assisted Ionic Liquid Pretreatment of Sugarcane Bagasse for Enhancing Saccharification Using Enzymes from an Ionic Liquid Tolerant *Aspergillus assiutensis* VS34. *Bioresour. Technol.* **2019**, 285, 121319, doi:10.1016/j.biortech.2019.121319.
- 18. Vaid, S.; Bhat, N.; Nargotra, P.; Bajaj, B.K. Combinatorial Application of Ammonium Carbonate and Sulphuric Acid Pretreatment to Achieve Enhanced Sugar Yield from Pine Needle Biomass for Potential Biofuel–Ethanol Production. *Energ. Ecol. Environ.* **2018**, *3*, 126–135, doi:10.1007/s40974-018-0083-1.
- 19. Hazeena, S.H.; Sindhu, R.; Pandey, A.; Binod, P. Lignocellulosic Bio-Refinery Approach for Microbial 2,3-Butanediol Production. *Bioresour. Technol.* **2020**, 302, 122873, doi:10.1016/j.biortech.2020.122873.
- 20. Sharma, H.K.; Xu, C.; Qin, W. Biological Pretreatment of Lignocellulosic Biomass for Biofuels and Bioproducts: An Overview. *Waste Biomass Valor.* **2019**, *10*, 235–251, doi:10.1007/s12649-017-0059-y.
- 21. Kumar, B.; Bhardwaj, N.; Agrawal, K.; Chaturvedi, V.; Verma, P. Current Perspective on Pretreatment Technologies Using Lignocellulosic Biomass: An Emerging Biorefinery Concept. *Fuel Process. Technol.* **2020**, *199*, 106244, doi:10.1016/j.fuproc.2019.106244.
- 22. Haq, I. ul; Qaisar, K.; Nawaz, A.; Akram, F.; Mukhtar, H.; Zohu, X.; Xu, Y.; Mumtaz, M.W.; Rashid, U.; Ghani, W.A.W.A.K.; et al. Advances in Valorization of Lignocellulosic Biomass towards Energy Generation. *Catalysts* **2021**, *11*, 309, doi:10.3390/catal11030309.
- 23. Kumar, V.; Patel, S.K.S.; Gupta, R.K.; Otari, S.V.; Gao, H.; Lee, J.-K.; Zhang, L. Enhanced Saccharification and Fermentation of Rice Straw by Reducing the Concentration of Phenolic Compounds Using an Immobilized Enzyme Cocktail. *Biotechnol. J.* **2019**, *14*, 1800468, doi:10.1002/biot.201800468.
- 24. Manisha; Yadav, S.K. Technological Advances and Applications of Hydrolytic Enzymes for Valorization of Lignocellulosic Biomass. *Bioresour. Technol.* **2017**, 245, 1727–1739, doi:10.1016/j.biortech.2017.05.066.
- 25. Wicker, R.J.; Kumar, G.; Khan, E.; Bhatnagar, A. Emergent Green Technologies for Cost-Effective Valorization of Microalgal Biomass to Renewable Fuel Products under a Biorefinery Scheme. *Chem. Eng. J.* **2021**, *415*, 128932, doi:10.1016/j.cej.2021.128932.

- 26. Toor, M.; Kumar, S.S.; Malyan, S.K.; Bishnoi, N.R.; Mathimani, T.; Rajendran, K.; Pugazhendhi, A. An Overview on Bioethanol Production from Lignocellulosic Feedstocks. *Chemosphere* **2020**, 242, 125080, doi:10.1016/j.chemosphere.2019.125080.
- 27. Elegbede, J.A.; Ajayi, V.A.; Lateef, A. Microbial Valorization of Corncob: Novel Route for Biotechnological Products for Sustainable Bioeconomy. *Environ. Technol. Innov.* **2021**, 24, 102073, doi:10.1016/j.eti.2021.102073.
- 28. Awasthi, M.K.; Sarsaiya, S.; Patel, A.; Juneja, A.; Singh, R.P.; Yan, B.; Awasthi, S.K.; Jain, A.; Liu, T.; Duan, Y.; et al. Refining Biomass Residues for Sustainable Energy and Bio-Products: An Assessment of Technology, Its Importance, and Strategic Applications in Circular Bio-Economy. *Renew. Sust. Energ. Rev.* **2020**, *127*, 109876, doi:10.1016/j.rser.2020.109876.
- 29. Velvizhi, G.; Balakumar, K.; Shetti, N.P.; Ahmad, E.; Kishore Pant, K.; Aminabhavi, T.M. Integrated Biorefinery Processes for Conversion of Lignocellulosic Biomass to Value Added Materials: Paving a Path towards Circular Economy. *Bioresour. Technol.* **2022**, 343, 126151, doi:10.1016/j.biortech.2021.126151.
- 30. A. Linares-Pasten, J.; Andersson, M.; N. Karlsson, E. Thermostable Glycoside Hydrolases in Biorefinery Technologies. *Cur. Biotechnol.* **2014**, *3*, 26–44.
- 31. Jin, Q.; Yang, L.; Poe, N.; Huang, H. Integrated Processing of Plant-Derived Waste to Produce Value-Added Products Based on the Biorefinery Concept. *Trends Food Sci. Technol.* **2018**, 74, 119–131, doi:10.1016/j.tifs.2018.02.014.
- 32. Vaid, S.; Sharma, S.; Bajaj, B.K. Chemo-Enzymatic Approaches for Consolidated Bioconversion of *Saccharum spontaneum* Biomass to Ethanol-Biofuel. *Bioresour. Technol.* **2021**, 329, 124898, doi:10.1016/j.biortech.2021.124898.
- 33. Wagle, A.; Angove, M.J.; Mahara, A.; Wagle, A.; Mainali, B.; Martins, M.; Goldbeck, R.; Raj Paudel, S. Multi-Stage Pre-Treatment of Lignocellulosic Biomass for Multi-Product Biorefinery: A Review. *Sustain. Energy Technol.* **2022**, 49, 101702, doi:10.1016/j.seta.2021.101702.
- 34. Nargotra, P.; Sharma, V.; Gupta, M.; Kour, S.; Bajaj, B.K. Application of Ionic Liquid and Alkali Pretreatment for Enhancing Saccharification of Sunflower Stalk Biomass for Potential Biofuel-Ethanol Production. *Bioresour. Technol.* **2018**, 267, 560–568, doi:10.1016/j.biortech.2018.07.070.
- 35. Houfani, A.A.; Anders, N.; Spiess, A.C.; Baldrian, P.; Benallaoua, S. Insights from Enzymatic Degradation of Cellulose and Hemicellulose to Fermentable Sugars— a Review. *Biomass and Bioenergy* **2020**, *134*, 105481, doi:10.1016/j.biombioe.2020.105481.
- 36. Alayoubi, R.; Mehmood, N.; Husson, E.; Kouzayha, A.; Tabcheh, M.; Chaveriat, L.; Sarazin, C.; Gosselin, I. Low Temperature Ionic Liquid Pretreatment of Lignocellulosic Biomass to Enhance Bioethanol Yield. *Renew. Energy* **2020**, *145*, 1808–1816, doi:10.1016/j.renene.2019.07.091.
- 37. Li, H.-Y.; Chen, X.; Wang, C.-Z.; Sun, S.-N.; Sun, R.-C. Evaluation of the Two-Step Treatment with Ionic Liquids and Alkali for Enhancing Enzymatic Hydrolysis of Eucalyptus: Chemical and Anatomical Changes. *Biotechnol. Biofuels* **2016**, *9*, 166, doi:10.1186/s13068-016-0578-y.
- 38. Kohli, K.; Katuwal, S.; Biswas, A.; Sharma, B.K. Effective Delignification of Lignocellulosic Biomass by Microwave Assisted Deep Eutectic Solvents. *Bioresour. Techno.* **2020**, 303, 122897, doi:10.1016/j.biortech.2020.122897.
- 39. Zulkefli, S.; Abdulmalek, E.; Abdul Rahman, Mohd.B. Pretreatment of Oil Palm Trunk in Deep Eutectic Solvent and Optimization of Enzymatic Hydrolysis of Pretreated Oil Palm Trunk. *Renew. Energy* **2017**, *107*, 36–41, doi:10.1016/j.renene.2017.01.037.
- 40. Haykir, N.I.; Soysal, K.; Yaglikci, S.; Gokce, Y. Assessing the Effect of Protic Ionic Liquid Pretreatment of Pinus Radiata from Different Perspectives Including Solvent-Water Ratio. *J. Wood Chem. Technol.* **2021**, 41, 236–248, doi:10.1080/02773813.2021.1976797.
- 41. Klímek, P.; Meinlschmidt, P.; Wimmer, R.; Plinke, B.; Schirp, A. Using Sunflower (Helianthus Annuus L.), Topinambour (*Helianthus tuberosus* L.) and Cup-Plant (*Silphium perfoliatum* L.) Stalks as Alternative Raw Materials for Particleboards. *Ind Crops Prod* . **2016**, 92, 157–164, doi:10.1016/j.indcrop.2016.08.004.
- 42. Paul, S.K.; Chakraborty, S. Microwave-Assisted Ionic Liquid-Mediated Rapid Catalytic Conversion of Non-Edible Lignocellulosic Sunn Hemp Fibres to Biofuels. *Bioresour. Technol.* **2018**, 253, 85–93, doi:10.1016/j.biortech.2018.01.010.

- 43. Fakayode, O.A.; Akpabli-Tsigbe, N.D.K.; Wahia, H.; Tu, S.; Ren, M.; Zhou, C.; Ma, H. Integrated Bioprocess for Bio-Ethanol Production from Watermelon Rind Biomass: Ultrasound-Assisted Deep Eutectic Solvent Pretreatment, Enzymatic Hydrolysis and Fermentation. *Renew. Energy* **2021**, *180*, 258–270, doi:10.1016/j.renene.2021.08.057.
- 44. Ge, S.; Wu, Y.; Peng, W.; Xia, C.; Mei, C.; Cai, L.; Shi, S.Q.; Sonne, C.; Lam, S.S.; Tsang, Y.F. High-Pressure CO2 Hydrothermal Pretreatment of Peanut Shells for Enzymatic Hydrolysis Conversion into Glucose. *Chem. Eng. J.* **2020**, *385*, 123949, doi:10.1016/j.cej.2019.123949.
- 45. Łukajtis, R.; Rybarczyk, P.; Kucharska, K.; Konopacka-Łyskawa, D.; Słupek, E.; Wychodnik, K.; Kamiński, M. Optimization of Saccharification Conditions of Lignocellulosic Biomass under Alkaline Pre-Treatment and Enzymatic Hydrolysis. *Energies* **2018**, *11*, 886, doi:10.3390/en11040886.
- 46. Oliva, J.M.; Negro, M.J.; Manzanares, P.; Ballesteros, I.; Chamorro, M.Á.; Sáez, F.; Ballesteros, M.; Moreno, A.D. A Sequential Steam Explosion and Reactive Extrusion Pretreatment for Lignocellulosic Biomass Conversion within a Fermentation-Based Biorefinery Perspective. *Fermentation* **2017**, *3*, 15, doi:10.3390/fermentation3020015.
- 47. Park, S.H.; Pham, T.T.H.; Kim, T.H. Effects of Additional Xylanase on Saccharification and Ethanol Fermentation of Ammonia-Pretreated Corn Stover and Rice Straw. *Energies* **2020**, *13*, 4574, doi:10.3390/en13174574.
- 48. Hoang, A.T.; Ong, H.C.; Fattah, I.M.R.; Chong, C.T.; Cheng, C.K.; Sakthivel, R.; Ok, Y.S. Progress on the Lignocellulosic Biomass Pyrolysis for Biofuel Production toward Environmental Sustainability. *Fuel Process. Technol.* **2021**, 223, 106997, doi:10.1016/j.fuproc.2021.106997.
- 49. Siqueira, J.G.W.; Rodrigues, C.; Vandenberghe, L.P. de S.; Woiciechowski, A.L.; Soccol, C.R. Current Advances in On-Site Cellulase Production and Application on Lignocellulosic Biomass Conversion to Biofuels: A Review. *Biomass and Bioenergy* **2020**, *132*, 105419, doi:10.1016/j.biombioe.2019.105419.
- 50. Okolie, J.A.; Nanda, S.; Dalai, A.K.; Kozinski, J.A. Chemistry and Specialty Industrial Applications of Lignocellulosic Biomass. *Waste Biomass Valor.* **2021**, *12*, 2145–2169, doi:10.1007/s12649-020-01123-0.
- 51. Haldar, D.; Dey, P.; Patel, A.K.; Dong, C.-D.; Singhania, R.R. A Critical Review on the Effect of Lignin Redeposition on Biomass in Controlling the Process of Enzymatic Hydrolysis. *Bioenerg. Res.* **2022**, *15*, 863–874, doi:10.1007/s12155-021-10374-1.
- 52. Hasanov, I.; Raud, M.; Kikas, T. The Role of Ionic Liquids in the Lignin Separation from Lignocellulosic Biomass. *Energies* **2020**, *13*, 4864, doi:10.3390/en13184864.
- 53. Lobato-Peralta, D.R.; Duque-Brito, E.; Villafán-Vidales, H.I.; Longoria, A.; Sebastian, P.J.; Cuentas-Gallegos, A.K.; Arancibia-Bulnes, C.A.; Okoye, P.U. A Review on Trends in Lignin Extraction and Valorization of Lignocellulosic Biomass for Energy Applications. *J. Clean. Prod.* **2021**, 293, 126123, doi:10.1016/j.jclepro.2021.126123.
- 54. Singhania, R.R.; Patel, A.K.; Raj, T.; Chen, C.-W.; Ponnusamy, V.K.; Tahir, N.; Kim, S.-H.; Dong, C.-D. Lignin Valorisation via Enzymes: A Sustainable Approach. *Fuel* **2022**, *311*, 122608, doi:10.1016/j.fuel.2021.122608.
- Nanda, S.; Mohanty, P.; Pant, K.K.; Naik, S.; Kozinski, J.A.; Dalai, A.K. Characterization of North American Lignocellulosic Biomass and Biochars in Terms of Their Candidacy for Alternate Renewable Fuels. *Bioenerg. Res.* **2013**, *6*, 663–677, doi:10.1007/s12155-012-9281-4.
- Vassilev, S.V.; Baxter, D.; Andersen, L.K.; Vassileva, C.G.; Morgan, T.J. An Overview of the Organic and Inorganic Phase Composition of Biomass. *Fuel* **2012**, *94*, 1–33, doi:10.1016/j.fuel.2011.09.030.
- 57. Yoo, C.G.; Meng, X.; Pu, Y.; Ragauskas, A.J. The Critical Role of Lignin in Lignocellulosic Biomass Conversion and Recent Pretreatment Strategies: A Comprehensive Review. *Bioresour. Technol.* **2020**, *301*, 122784, doi:10.1016/j.biortech.2020.122784.
- 58. Bhatia, S.K.; Jagtap, S.S.; Bedekar, A.A.; Bhatia, R.K.; Patel, A.K.; Pant, D.; Rajesh Banu, J.; Rao, C.V.; Kim, Y.-G.; Yang, Y.-H. Recent Developments in Pretreatment Technologies on Lignocellulosic Biomass: Effect of Key Parameters, Technological Improvements, and Challenges. *Bioresour. Technol.* **2020**, 300, 122724, doi:10.1016/j.biortech.2019.122724.
- 59. Jatoi, A.S.; Abbasi, S.A.; Hashmi, Z.; Shah, A.K.; Alam, M.S.; Bhatti, Z.A.; Maitlo, G.; Hussain, S.; Khandro, G.A.; Usto, M.A.; et al. Recent Trends and Future Perspectives of Lignocellulose Biomass for Biofuel Production: A Comprehensive Review. *Biomass Conv. Bioref.* **2021**, doi:10.1007/s13399-021-01853-8.
- 60. Nargotra, P.; Vaid, S.; Bajaj, B.K. Cellulase Production from Bacillus Subtilis SV1 and Its Application Potential for Saccharification of Ionic Liquid Pretreated Pine Needle Biomass under One Pot Consolidated Bioprocess. *Fermentation* **2016**, 2, 19, doi:10.3390/fermentation2040019.

- 61. Mankar, A.R.; Pandey, A.; Modak, A.; Pant, K.K. Pretreatment of Lignocellulosic Biomass: A Review on Recent Advances. *Bioresour. Technol.* **2021**, *334*, 125235, doi:10.1016/j.biortech.2021.125235.
- 62. Srivastava, N.; Rawat, R.; Singh Oberoi, H.; Ramteke, P.W. A Review on Fuel Ethanol Production From Lignocellulosic Biomass. Int. J. Green Energy **2015**, *12*, 949–960, doi:10.1080/15435075.2014.890104.
- 63. He, Z.; Wang, Z.; Zhao, Z.; Yi, S.; Mu, J.; Wang, X. Influence of Ultrasound Pretreatment on Wood Physiochemical Structure. *Ultrason. Sonochem.* **2017**, *34*, 136–141, doi:10.1016/j.ultsonch.2016.05.035.
- 64. Subhedar, P.B.; Ray, P.; Gogate, P.R. Intensification of Delignification and Subsequent Hydrolysis for the Fermentable Sugar Production from Lignocellulosic Biomass Using Ultrasonic Irradiation. *Ultrason. Sonochem.* **2018**, 40, 140–150, doi:10.1016/j.ultsonch.2017.01.030.
- 65. Kunaver, M.; Jasiukaitytė, E.; Čuk, N. Ultrasonically Assisted Liquefaction of Lignocellulosic Materials. *Bioresour. Technol.* **2012**, *103*, 360–366, doi:10.1016/j.biortech.2011.09.051.
- 66. Haldar, D.; Purkait, M.K. A Review on the Environment-Friendly Emerging Techniques for Pretreatment of Lignocellulosic Biomass: Mechanistic Insight and Advancements. *Chemosphere* **2021**, 264, 128523, doi:10.1016/j.chemosphere.2020.128523.
- 67. Hoang, A.T.; Nižetić, S.; Ong, H.C.; Mofijur, M.; Ahmed, S.F.; Ashok, B.; Bui, V.T.V.; Chau, M.Q. Insight into the Recent Advances of Microwave Pretreatment Technologies for the Conversion of Lignocellulosic Biomass into Sustainable Biofuel. *Chemosphere* **2021**, *281*, 130878, doi:10.1016/j.chemosphere.2021.130878.
- 68. Nuchdang, S.; Thongtus, V.; Khemkhao, M.; Kirdponpattara, S.; Moore, E.J.; Setiabudi, H.D.B.; Phalakornkule, C. Enhanced Production of Reducing Sugars from Paragrass Using Microwave-Assisted Alkaline Pretreatment. *Biomass Conv. Bioref.* **2021**, *11*, 2471–2483, doi:10.1007/s13399-020-00624-1.
- 69. Kainthola, J.; Shariq, M.; Kalamdhad, A.S.; Goud, V.V. Enhanced Methane Potential of Rice Straw with Microwave Assisted Pretreatment and Its Kinetic Analysis. *J. Environ. Manage.* **2019**, 232, 188–196, doi:10.1016/j.jenvman.2018.11.052.
- 70. Zhu, Z.; Liu, Y.; Yang, X.; McQueen-Mason, S.J.; Gomez, L.D.; Macquarrie, D.J. Comparative Evaluation of Microwave-Assisted Acid, Alkaline, and Inorganic Salt Pretreatments of Sugarcane Bagasse for Sugar Recovery. *Biomass Conv. Bioref.* **2021**, *11*, 2681–2693, doi:10.1007/s13399-020-00680-7.
- 71. Kundu, C.; Samudrala, S.P.; Kibria, M.A.; Bhattacharya, S. One-Step Peracetic Acid Pretreatment of Hardwood and Softwood Biomass for Platform Chemicals Production. *Sci. Rep.* **2021**, *11*, 11183, doi:10.1038/s41598-021-90667-9.
- 72. Morais, W.G.; Pacheco, T.F.; Corrêa, P.S.; Martins, A.A.; Mata, T.M.; Caetano, N.S. Acid Pretreatment of Sugarcane Biomass to Obtain Hemicellulosic Hydrolisate Rich in Fermentable Sugar. *Energy Reports* **2020**, *6*, 18–23, doi:10.1016/j.egyr.2020.10.015.
- 73. Mehrez, I.; Chandrasekhar, K.; Kim, W.; Kim, S.-H.; Kumar, G. Comparison of Alkali and Ionic Liquid Pretreatment Methods on the Biochemical Methane Potential of Date Palm Waste Biomass. *Bioresour. Technol.* **2022**, *360*, 127505, doi:10.1016/j.biortech.2022.127505.
- 74. Goshadrou, A. Bioethanol Production from Cogongrass by Sequential Recycling of Black Liquor and Wastewater in a Mild-Alkali Pretreatment. *Fuel* **2019**, 258, 116141, doi:10.1016/j.fuel.2019.116141.
- 75. Lorenci Woiciechowski, A.; Dalmas Neto, C.J.; Porto de Souza Vandenberghe, L.; de Carvalho Neto, D.P.; Novak Sydney, A.C.; Letti, L.A.J.; Karp, S.G.; Zevallos Torres, L.A.; Soccol, C.R. Lignocellulosic Biomass: Acid and Alkaline Pretreatments and Their Effects on Biomass Recalcitrance Conventional Processing and Recent Advances. *Bioresour. Technol.* **2020**, *304*, 122848, doi:10.1016/j.biortech.2020.122848.
- 76. Dotsenko, A.S.; Dotsenko, G.S.; Senko, O.V.; Stepanov, N.A.; Lyagin, I.V.; Efremenko, E.N.; Gusakov, A.V.; Zorov, I.N.; Rubtsova, E.A. Complex Effect of Lignocellulosic Biomass Pretreatment with 1-Butyl-3-Methylimidazolium Chloride Ionic Liquid on Various Aspects of Ethanol and Fumaric Acid Production by Immobilized Cells within SSF. *Bioresour. Technol.* **2018**, *250*, 429–438, doi:10.1016/j.biortech.2017.11.064.
- 77. Lim, W.-L.; Gunny, A.A.N.; Kasim, F.H.; AlNashef, I.M.; Arbain, D. Alkaline Deep Eutectic Solvent: A Novel Green Solvent for Lignocellulose Pulping. *Cellulose* **2019**, *26*, 4085–4098, doi:10.1007/s10570-019-02346-8.
- 78. Amesho, K.T.T.; Lin, Y.-C.; Mohan, S.V.; Halder, S.; Ponnusamy, V.K.; Jhang, S.-R. Deep Eutectic Solvents in the Transformation of Biomass into Biofuels and Fine Chemicals: A Review. *Environ Chem. Lett.* **2022**, doi:10.1007/s10311-022-01521-x.

- 79. Jing, Y.; Li, F.; Li, Y.; Jiang, D.; Lu, C.; Zhang, Z.; Zhang, Q. Biohydrogen Production by Deep Eutectic Solvent Delignification-Driven Enzymatic Hydrolysis and Photo-Fermentation: Effect of Liquid–Solid Ratio. *Bioresour. Technol.* **2022**, 349, 126867, doi:10.1016/j.biortech.2022.126867.
- 80. Tan, Y.T.; Ngoh, G.C.; Chua, A.S.M. Effect of Functional Groups in Acid Constituent of Deep Eutectic Solvent for Extraction of Reactive Lignin. *Bioresour. Technol.* **2019**, *281*, 359–366, doi:10.1016/j.biortech.2019.02.010.
- 81. Thi, S.; Lee, K.M. Comparison of Deep Eutectic Solvents (DES) on Pretreatment of Oil Palm Empty Fruit Bunch (OPEFB): Cellulose Digestibility, Structural and Morphology Changes. *Bioresour. Technol.* **2019**, *282*, 525–529, doi:10.1016/j.biortech.2019.03.065.
- 82. Vaidya, A.A.; Murton, K.D.; Smith, D.A.; Dedual, G. A Review on Organosolv Pretreatment of Softwood with a Focus on Enzymatic Hydrolysis of Cellulose. *Biomass Conv. Bioref.* **2022**, *12*, 5427–5442, doi:10.1007/s13399-022-02373-9.
- 83. Borand, M.N.; Karaosmanoğlu, F. Effects of Organosolv Pretreatment Conditions for Lignocellulosic Biomass in Biorefinery Applications: A Review. *J. Renew. Sustain. Energy* **2018**, *10*, 033104, doi:10.1063/1.5025876.
- 84. Karnaouri, A.; Asimakopoulou, G.; Kalogiannis, K.G.; Lappas, A.; Topakas, E. Efficient D-Lactic Acid Production by Lactobacillus Delbrueckii Subsp. Bulgaricus through Conversion of Organosolv Pretreated Lignocellulosic Biomass. *Biomass and Bioenergy* **2020**, *140*, 105672, doi:10.1016/j.biombioe.2020.105672.
- 85. Joy, S.P.; Krishnan, C. Modified Organosolv Pretreatment for Improved Cellulosic Ethanol Production from Sorghum Biomass. *Ind. Crops Prod.* **2022**, *177*, 114409, doi:10.1016/j.indcrop.2021.114409.
- 86. Tang, C.; Shan, J.; Chen, Y.; Zhong, L.; Shen, T.; Zhu, C.; Ying, H. Organic Amine Catalytic Organosolv Pretreatment of Corn Stover for Enzymatic Saccharification and High-Quality Lignin. *Bioresour. Technol.* **2017**, 232, 222–228, doi:10.1016/j.biortech.2017.02.041.
- 87. Li, X.; Shi, Y.; Kong, W.; Wei, J.; Song, W.; Wang, S. Improving Enzymatic Hydrolysis of Lignocellulosic Biomass by Bio-Coordinated Physicochemical Pretreatment—A Review. *Energy Reports* **2022**, *8*, 696–709, doi:10.1016/j.egyr.2021.12.015.
- 88. Mockaitis, G.; Bruant, G.; Foresti, E.; Zaiat, M.; Guiot, S.R. Physicochemical Pretreatment Selects Microbial Communities to Produce Alcohols through Metabolism of Volatile Fatty Acids. *Biomass Conv. Bioref.* **2022**, doi:10.1007/s13399-022-02383-7.
- 89. Raud, M.; Kikas, T.; Sippula, O.; Shurpali, N.J. Potentials and Challenges in Lignocellulosic Biofuel Production Technology. *Renew. Sust. Energ. Rev.* **2019**, *111*, 44–56, doi:10.1016/j.rser.2019.05.020.
- 90. Kuglarz, M.; Gunnarsson, I.B.; Svensson, S.-E.; Prade, T.; Johansson, E.; Angelidaki, I. Ethanol Production from Industrial Hemp: Effect of Combined Dilute Acid/Steam Pretreatment and Economic Aspects. *Bioresour. Technol.* **2014**, *163*, 236–243, doi:10.1016/j.biortech.2014.049.
- 91. Vaid, S.; Sharma, S.; Dutt, H.C.; Mahajan, R.; Bajaj, B.K. One Pot Consolidated Bioprocess for Conversion of Saccharum Spontaneum Biomass to Ethanol-Biofuel. *Energy Convers. Manag.* **2021**, 250, 114880, doi:10.1016/j.enconman.2021.114880.
- 92. Hou, X.; Wang, Z.; Sun, J.; Li, M.; Wang, S.; Chen, K.; Gao, Z. A Microwave-Assisted Aqueous Ionic Liquid Pretreatment to Enhance Enzymatic Hydrolysis of Eucalyptus and Its Mechanism. *Bioresour. Technol.* **2019**, 272, 99–104, doi:10.1016/j.biortech.2018.10.003.
- 93. Hosseini Koupaie, E.; Dahadha, S.; Bazyar Lakeh, A.A.; Azizi, A.; Elbeshbishy, E. Enzymatic Pretreatment of Lignocellulosic Biomass for Enhanced Biomethane Production-A Review. *J. Environ. Manage.* **2019**, 233, 774–784, doi:10.1016/j.jenvman.2018.09.106.
- 94. Prasad, R.K.; Chatterjee, S.; Mazumder, P.B.; Gupta, S.K.; Sharma, S.; Vairale, M.G.; Datta, S.; Dwivedi, S.K.; Gupta, D.K. Bioethanol Production from Waste Lignocelluloses: A Review on Microbial Degradation Potential. *Chemosphere* **2019**, 231, 588–606, doi:10.1016/j.chemosphere.2019.05.142.
- 95. Roth, J.C.G.; Hoeltz, M.; Benitez, L.B. Current Approaches and Trends in the Production of Microbial Cellulases Using Residual Lignocellulosic Biomass: A Bibliometric Analysis of the Last 10 Years. *Arch. Microbiol.* **2020**, 202, 935–951, doi:10.1007/s00203-019-01796-9.
- 96. Sharma, S.; Sharma, V.; Nargotra, P.; Bajaj, B.K. Process Desired Functional Attributes of an Endoxylanase of GH10 Family from a New Strain of *Aspergillus terreus* S9. *Int. J. Biol. Macromol.* **2018**, 115, 663–671, doi:10.1016/j.ijbiomac.2018.04.096.

- 97. Sharma, S.; Sharma, V.; Nargotra, P.; Bajaj, B.K. Bioprocess Development for Production of a Process-Apt Xylanase with Multifaceted Application Potential for a Range of Industrial Processes. *SN Appl. Sci.* **2020**, *2*, 739, doi:10.1007/s42452-020-2541-6.
- 98. Madadi, M.; Tu, Y.; Abbas, A. Recent Status on Enzymatic Saccharification of Lignocellulosic Biomass for Bioethanol Production. *Electron. J. Biotechnol.* **2017**, *13*, 135–143.
- 99. Alokika; Singh, B. Enhanced Production of Bacterial Xylanase and Its Utility in Saccharification of Sugarcane Bagasse. *Bioprocess Biosyst. Eng.* **2020**, *43*, 1081–1091, doi:10.1007/s00449-020-02306-8.
- 100. Cui, L.; Wang, Z.; Zeng, Y.; Yang, N.; Liu, M.; Zhao, Y.; Zheng, Y. Lignin Biodegradation and Its Valorization. *Fermentation* **2022**, *8*, 366, doi:10.3390/fermentation8080366.
- 101. Obeng, E.M.; Budiman, C.; Ongkudon, C.M. Identifying Additives for Cellulase Enhancement—A Systematic Approach. *Biocatal. Agric. Biotechnol.* **2017**, *11*, 67–74, doi:10.1016/j.bcab.2017.06.006.
- 102. Cai, C.; Qiu, X.; Zeng, M.; Lin, M.; Lin, X.; Lou, H.; Zhan, X.; Pang, Y.; Huang, J.; Xie, L. Using Polyvinylpyrrolidone to Enhance the Enzymatic Hydrolysis of Lignocelluloses by Reducing the Cellulase Non-Productive Adsorption on Lignin. *Bioresour. Technol.* **2017**, 227, 74–81, doi:10.1016/j.biortech.2016.12.002.
- 103. Rocha-Martín, J.; Martinez-Bernal, C.; Pérez-Cobas, Y.; Reyes-Sosa, F.M.; García, B.D. Additives Enhancing Enzymatic Hydrolysis of Lignocellulosic Biomass. *Bioresour. Technol.* **2017**, 244, 48–56, doi:10.1016/j.biortech.2017.06.132.
- 104. Rai, M.; Ingle, A.P.; Pandit, R.; Paralikar, P.; Biswas, J.K.; da Silva, S.S. Emerging Role of Nanobiocatalysts in Hydrolysis of Lignocellulosic Biomass Leading to Sustainable Bioethanol Production. *Catalysis Reviews* **2019**, *61*, 1–26, doi:10.1080/01614940.2018.1479503.
- 105. Grewal, J.; Ahmad, R.; Khare, S.K. Development of Cellulase-Nanoconjugates with Enhanced Ionic Liquid and Thermal Stability for in Situ Lignocellulose Saccharification. *Bioresour. Technol.* **2017**, 242, 236–243, doi:10.1016/j.biortech.2017.04.007.
- 106. Cheng, G.; Xing, J.; Pi, Z.; Liu, S.; Liu, Z.; Song, F. α-Glucosidase Immobilization on Functionalized Fe3O4 Magnetic Nanoparticles for Screening of Enzyme Inhibitors. *Chin. Chem. Lett.* **2019**, 30, 656–659, doi:10.1016/j.cclet.2018.12.003.
- 107. Hossain, N.; Mahlia, T.M.I.; Saidur, R. Latest Development in Microalgae-Biofuel Production with Nano-Additives. *Biotechnol. Biofuels* **2019**, *12*, 125, doi:10.1186/s13068-019-1465-0.
- 108. Karnaouri, A.; Muraleedharan, M.N.; Dimarogona, M.; Topakas, E.; Rova, U.; Sandgren, M.; Christakopoulos, P. Recombinant Expression of Thermostable Processive MtEG5 Endoglucanase and Its Synergism with MtLPMO from *Myceliophthora thermophila* during the Hydrolysis of Lignocellulosic Substrates. *Biotechnol. Biofuels* **2017**, *10*, 126, doi:10.1186/s13068-017-0813-1.
- 109. Vijayalakshmi, S.; Govindarajan, M.; Al-Mulahim, N.; Ahmed, Z.; Mahboob, S. Cellulase Immobilized Magnetic Nanoparticles for Green Energy Production from *Allamanda schottii* L: Sustainability Research in Waste Recycling. *Saudi Journal of Biological Sciences* **2021**, *28*, 901–910, doi:10.1016/j.sjbs.2020.11.034.
- 110. Hwangbo, M.; Tran, J.L.; Chu, K.-H. Effective One-Step Saccharification of Lignocellulosic Biomass Using Magnetite-Biocatalysts Containing Saccharifying Enzymes. *Sci. Total Environ.* **2019**, *647*, 806–813, doi:10.1016/j.scitotenv.2018.08.066.
- 111. Yan, J. Handbook of Clean Energy Systems, 6 Volume Set; John Wiley & Sons, 2015; ISBN 978-1-118-38858-7.
- 112. Sharma, S.; Sharma, P.; Sharma, V.; Bajaj, B.K. Polyhydroxybutyrate as an Eco-Friendly Alternative of Synthetic Plastics. In *Environmental and Agricultural Microbiology*; John Wiley & Sons, Ltd, 2021; pp. 101–149 ISBN 978-1-119-52589-9.
- 113. Mohd Azhar, S.H.; Abdulla, R.; Jambo, S.A.; Marbawi, H.; Gansau, J.A.; Mohd Faik, A.A.; Rodrigues, K.F. Yeasts in Sustainable Bioethanol Production: A Review. *Biochem. Biophys. Rep.* **2017**, *10*, 52–61, doi:10.1016/j.bbrep.2017.03.003.
- 114. Ramachandra, T.V.; Hebbale, D. Bioethanol from Macroalgae: Prospects and Challenges. *Renew. Sust. Energ. Rev.* **2020**, *117*, 109479, doi:10.1016/j.rser.2019.109479.
- 115. Jin, Y.; Shi, Z.; Xu, G.; Yang, H.; Yang, J. A Stepwise Pretreatment of Sugarcane Bagasse by Alkaline and Hydroxymethyl Reagent for Bioethanol Production. *Ind. Crops Prod.* **2020**, 145, 112136, doi:10.1016/j.indcrop.2020.112136.

- 116. Wang, H.; Peng, X.; Zhang, H.; Yang, S.; Li, H. Microorganisms-Promoted Biodiesel Production from Biomass: A Review. *Energy Convers. Manag.*: X 2021, 12, 100137, doi:10.1016/j.ecmx.2021.100137.
- 117. Yusuf, N.N.A.N.; Kamarudin, S.K.; Yaakub, Z. Overview on the Current Trends in Biodiesel Production. *Energy Convers. Manag.* **2011**, *52*, 2741–2751, doi:10.1016/j.enconman.2010.12.004.
- 118. Wang, H.; Rehman, K. ur; Liu, X.; Yang, Q.; Zheng, L.; Li, W.; Cai, M.; Li, Q.; Zhang, J.; Yu, Z. Insect Biorefinery: A Green Approach for Conversion of Crop Residues into Biodiesel and Protein. *Biotechnol. Biofuels* **2017**, *10*, 304, doi:10.1186/s13068-017-0986-7.
- 119. Bateni, H.; Karimi, K. Biodiesel Production from Castor Plant Integrating Ethanol Production via a Biorefinery Approach. *Chem Eng Res Des*. **2016**, *107*, 4–12, doi:10.1016/j.cherd.2015.08.014.
- 120. Preethi; Rajesh Banu, J.; Kavitha, S.; Yukesh Kannah, R.; Varjani, S.; Gunasekaran, M. Mild Hydrogen Peroxide Interceded Bacterial Disintegration of Waste Activated Sludge for Efficient Biomethane Production. *Sci. Total Environ.* 2022, 817, 152873, doi:10.1016/j.scitotenv.2021.152873.
- 121. Okolie, J.A.; Mukherjee, A.; Nanda, S.; Dalai, A.K.; Kozinski, J.A. Next-Generation Biofuels and Platform Biochemicals from Lignocellulosic Biomass. *Int. J. Energy Res.* **2021**, *45*, 14145–14169, doi:10.1002/er.6697.
- 122. Kohli, K.; Prajapati, R.; Sharma, B.K. Bio-Based Chemicals from Renewable Biomass for Integrated Biorefineries. *Energies* **2019**, *12*, 233, doi:10.3390/en12020233.
- 123. Gandini, A.; Belgacem, M.N. Recent Contributions to the Preparation of Polymers Derived from Renewable Resources. *J. Polym. Environ*. **2002**, *10*, 105–114, doi:10.1023/A:1021172130748.
- 124. Ruby, M.-P.; Schüth, F. Synthesis of N-Alkyl-4-Vinylpyridinium-Based Cross-Linked Polymers and Their Catalytic Performance for the Conversion of Fructose into 5-Hydroxymethylfurfural. *Green Chem.* **2016**, *18*, 3422–3429, doi:10.1039/C5GC02949I.
- 125. Zhao, J.; Zhou, C.; He, C.; Dai, Y.; Jia, X.; Yang, Y. Efficient Dehydration of Fructose to 5-Hydroxymethylfurfural over Sulfonated Carbon Sphere Solid Acid Catalysts. *Catalysis Today* **2016**, 264, 123–130, doi:10.1016/j.cattod.2015.07.005.
- 126. Prabha, S.P.; Nagappan, S.; Rathna, R.; Viveka, R.; Nakkeeran, E. 21 Blue Biotechnology: A Vision for Future Marine Biorefineries. In *Refining Biomass Residues for Sustainable Energy and Bioproducts*; Kumar, R.P., Gnansounou, E., Raman, J.K., Baskar, G., Eds.; Academic Press, 2020; pp. 463–480 ISBN 978-0-12-818996-2.
- 127. Chang, W.-R.; Hwang, J.-J.; Wu, W. Environmental Impact and Sustainability Study on Biofuels for Transportation Applications. *Renew. Sust. Energ. Rev.* **2017**, *67*, 277–288, doi:10.1016/j.rser.2016.09.020.