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Keywords: Colchicine; IL-10; Acute myocardial infarction; Network pharmacology; Reperfusion



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Article

Novel Impact of Colchicine on Interleukin-10 Expression in Acute Myocardial Infarction: An Integrative Approach

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Abstract: Background: Inflammatory plays critical role in myocardial infarction as a critical process in the development of heart failure, involving the development of cardiac fibrosis. Colchicine is a well-established anti-inflammatory drug, but its scientific application in controlling post-acute myocardial infarction remodelling processes has not been established. IL-10, a key cytokine in modulating inflammatory responses, underscoring its potential as a crucial therapeutic target of colchicine. Objectives: To explore the protective role of IL-10 modulated by colchicine in myocardial healing and repair following AMI, particularly cardiac fibrosis. Materials and Methods: The predicted protein of Colchicine was assessed using WAY2DRUG PASS as probability active value. Proteins associated with Colchicine, cardiac fibrosis and acute myocardial infarction were analysed with DisGeNET and Open Target databases. Analysis and visualization of protein-protein interactions were conducted using STRING and Cytoscape. A 3T3 cell line treated with CoCl2 was used to mimic hypoxic. HIF-1 α and IL-10 expression were measured by flow cytometry and analysed using a one-way ANOVA test. This observational clinical trial examined acute myocardial infarction patients undergoing immediate and delayed primary percutaneous coronary interventions. Subjects were randomized into control groups receiving placebo and intervention groups treated with Colchicine. Assessments occurred at 24 hours and five days after the intervention. IL-10 expression in the clinical trial was measured by ELISA assay and analysed using a T-test. Results: Colchicine demonstrates promising bioactivity in treating acute myocardial infarction, with notable probability active values as a tubulin antagonist (0.744), beta-tubulin antagonist (0.673), and NOS2 inhibitor (0.529). Its primary action targets IL-10, with protein-protein interactions analysis indicating interactions between IL-10 and key inflammatory mediators—IL-1β, IFN-γ, CCL2, TNF, and TGFβ1—during acute myocardial infarction and cardiac fibrosis. Hypoxic conditions in the CoCl2-induced 3T3 cell model show significantly elevated HIF-1 α compared to controls (p<0.0001). Acute myocardial infarction patients were predominantly from high-risk populations, with notable smoking prevalence in the early Colchicine group (72%). Hypertension prevalence was significantly different, with 67.31% in the late Colchicine group compared to 38% in the early Colchicine and 39.80% in the early placebo groups. Both early and late Colchicine groups exhibited a higher prevalence of diabetes mellitus. Colchicine use, either early or late, did not significantly alter major adverse cardiovascular events incidence compared to placebo (p>0.05). However, Colchicine significantly increased IL-10 expression in CoCl₂-treated cells (p<0.0001) and in acute myocardial infarction patients within five days (p<0.05). Conclusions: Colchicine may bolster the anti-inflammatory response post-myocardial infarction by activating IL-10 pathways in fibroblasts and in clinical settings,

potentially reducing major adverse cardiovascular events. Further investigation into broader aspects of this pathway, particularly in cardiac fibroblasts, is required.

Keywords: colchicine; IL-10; Acute myocardial infarction; network pharmacology; reperfusion

1. Introduction

Cardiovascular disease, including coronary heart disease and stroke, is a prevalent non-communicable disease worldwide, responsible for approximately 17.8 million deaths in 2017, with more than three-quarters occurring in low to middle-income countries [1]. The incidence of heart and vascular diseases in Indonesia has been steadily rising over the years, making it the leading cause of death in the country. It is estimated that around 15 out of 1,000 people in Indonesia suffer from heart disease [2]. The prevalence of acute myocardial infarction (AMI) approaches three million people worldwide each year [3].

Acute coronary syndrome resulting from coronary artery occlusion leads to ischemic or hypoxic conditions in the heart [4]. Heart disease is characterized by the enlargement of the myocardial infarction site during the acute phase, followed by left ventricular remodelling (VR), including dilation, fibrosis, and heart dysfunction [5]. Ventricular remodelling plays a significant role in the transition from myocardial infarction (MI) to heart failure (HF) mediated by inflammation [6]. The healing and prevention of detrimental VR tissue involve fibroblast proliferation, migration, and phenotypic transformation. Thus, modulating myocardial fibrosis levels after AMI is crucial to delaying or improving VR [7]. Polymorphonuclear cells are the first inflammatory cells recruited to the area of infarction and occurs within a few hours after MI. Polymorphonuclear cells are essentially regulatory early left ventricular remodeling response [8].

Excessive remodelling control to enhance heart function following acute myocardial infarction injury can be observed through several biomarkers, such as galectin-3, type III pro-collagen peptide (PIIINP), and IL-10 [9]. Cytokine therapy for reducing inflammation has been widely used and successful in conditions such as rheumatoid arthritis and is currently an area of investigation for MI and HF treatment. Cardiovascular remodelling is a fluctuating process with observable clinical outcomes, making cytokines a potential therapeutic target [6]. IL-10 can improve left ventricular function, reduce infarction size, and mitigate infarct wall thinning [5].

Primary percutaneous coronary intervention (PCI) is the preferred reperfusion therapy for ST-segment–elevation myocardial infarction (STEMI) [10]. However, the proportion of PCI remains low in some countries and regions [11]. PCI technology may alleviate atherosclerotic vessel narrowing or thrombosis, but exercise tolerance may decrease after revascularization [8].

Colchicine is an alkaloid extracted from plants of the *Colchicum* genus and is known to inhibit the inflammatory process [12]. Colchicine has relatively minor side effects, is readily available and is cost-effective [13]. Colchicine can reduce the expression of TNF- α receptors in macrophages and lower cytokines such as IL-1 β , IFN γ , IL-18, and IL-6. Some studies suggest that Colchicine plays a role in reducing the risk of recurrent MI. Colchicine can lower high-sensitivity C-reactive protein (HsCRP) levels and mean platelet volume (MPV) in AMI [14]. Colchicine triggers apoptosis in liver cells to inhibit hepatic fibrosis [15].

A computational approach to evaluate the pharmacological properties of Colchicine in preventing excessive remodelling in heart disease, in line with experimental advances, employs bioinformatics tools that allow the meta-search of databases to analyse biological interaction signal networks. This approach saves time and resources for laboratory-scale research [16]. To date, no scientific study has revealed the role of Colchicine in postacute myocardial infarction excessive remodelling via IL-10. Therefore, this research aims to analyze the effects of Colchicine on reducing excessive remodelling postmyocardial infarction through IL-10 using both in silico and in vitro studies.

2. Materials and Methods

2.1. In-Silico Study

2.1.1. Prediction of Colchicine Potential Using a Structure-Activity Relationship (SAR) Approach

The Simplified Molecular-Input Line-Entry System (SMILES) profile and the structure of Colchicine were obtained from the PubChem database (https://pubchem.ncbi.nlm.nih.gov/). Colchicine's potential as an anti-inflammatory agent related to cardiac fibrosis and myocardial infarction was then analyzed using WAY2DRUG PASS prediction (http://www.pharmaexpert.ru/passonline/predict.php) by inserting its SMILES structure.

Structure-activity relationship (SAR) analysis was carried out using WAY2DRUG Pass Prediction to compare the input compound with known compounds that exhibit specific potential. The more similar the structures of the compounds are, the higher the prediction value. Compounds with high similarity typically share potential, as depicted by the probability to be active (Pa) value. A Pa value exceeding 0.7 indicates that the compound is predicted to have high potential as an anti-inflammatory agent due to its resemblance to compounds in the database known to be effective therapies. If 0.5 < Pa < 0.7, it suggests a lower likelihood of finding activity in further research, but the compound is not closely related to known pharmaceuticals. When Pa < 0.5, the chances of discovering activity in experiments become even slimmer, but if confirmed, the compound may represent a new chemical entity. Higher Pa values indicate better accuracy. This study used a reference range of 0.5 < Pa < 0.7 [17].

2.1.2. Prediction of Target and Gene Disease Association with Cardiac Fibrosis (CF) and Acute Myocardial Infarction (AMI)

The target analysis of Colchicine was performed using the Comparative Toxicogenomics Database (CTD) (http://ctdbase.org/) and DIGEP-Pred (http://www.way2drug.com/ge/) by entering Colchicine's SMILES structure. Genes and proteins associated with cardiac fibrosis and acute myocardial infarction were obtained from the DisGeNET and Open Target databases, accessed on July 5, 2023 [18]. Targets related to diseases and Colchicine were then mapped using a Venn diagram to identify target intersections. Each identified target was annotated for its function using the Biological Function in the Database for Annotation, Visualization, and Integrated Discovery (DAVID) (http://david.abcc.ncifcrf.gov/home.jsp), accessed on May 29, 2023 [19].

Based on Fisher's exact test, the false discovery rate (FDR) was used as a statistical test method in the DAVID database. To validate the DAVID results, Network Analyst, commonly used for gene expression, functional analysis, and transcription factor analysis, was utilized [20].

2.1.3. Pharmacology Network Analysis

A comprehensive list of target proteins, associated with Cardiac Fibrosis (CF) and Acute Myocardial Infarction (AMI), was compiled from two distinct databases. This collection of proteins was then inputted into the 'multiple protein' feature of the STRING database (https://string-db.org/), a tool used for exploring protein-protein interactions and networks. A compilation of target proteins from both databases was input into the "multiple protein" feature of the STRING database (https://string-db.org/) [21]. The STRING database assesses protein-protein interactions [22]. Additional settings included choosing "Homo sapiens" as the organism, considering network edges as confidence, and setting the minimum needed interaction score to 0.9 for the physical subnetwork. After updating the network based on these additional settings, the visualization results were downloaded in TSV (Tab-separated values) format in the "table/export" menu. The downloaded (.tsv) file was then imported into Cytoscape v.3.10.0 (accessed on July 17, 2023) for further network analysis using the "analyse network" procedure. The analysis network was used to identify the most functional and dominant proteins in the analysed pathway, as depicted using parameters such as degree, betweenness centrality (BC), and closeness centrality (CC). These values help determine significant proteins. A higher BC value indicates a greater role of the protein in a pathway. The degree

3

indicates interactions of the compound with other proteins (represented by nodes). A higher degree value implies more interactions with other nodes. Closeness centrality calculates the role of a protein as a communicator or hub. The higher the closeness centrality value is, the greater the protein's role in information and communication pathways. Degree helps identify the shortest pathways targetable in the path [23].

2.2. In-Vitro Study

2.2.1. Induction of Hypoxia in 3T3 Cell Line

To mimic the model of ischemic in-vitro, we established the hypoxic fibroblast by treating the 3T3 cells with CoCl₂. Cells were cultured in DMEM supplemented with 10% FBS and antibiotics at 37°C in a humidified atmosphere with 5% CO₂ and 95% air. To induce hypoxic conditions, fibroblasts were treated with CoCl₂ (Sigma–Aldrich, St. Louis, MO) at different concentrations of 300 μ M for 24 hours (dose optimization was performed) [24-25]. Colchicine was administered to 3T3 cell lines at a concentration of 10 μ M. The dosage was based on the research protocol conducted by Wang et al. [26].

2.2.2. Measurement of HIF-1 α and IL-10 Expression

The expression of HIF-1 α and IL-10 were measured by using flowcytometry method. Briefly, the 3T3 cell line suspension was centrifuged at 2500 rpm for 5 minutes at a temperature of 10°C. The supernatant was discarded, and the cell pellet was resuspended in 1 mL of PBS, divided into two microtubes (tubes A and B) at 300 μ L each, and centrifuged again. The cell pellet was treated with 100 μ l of BD hotfix/cytoperm fixative solution and incubated for 20 minutes. It was then mixed with 500 μ l of BD Perm/Wash buffer for permeabilization and centrifuged at 2500 rpm at 10°C for 5 minutes. After discarding the supernatant, the cell pellet was stained with FITC-conjugated antihuman HIF-1 α or IL-10 antibodies and incubated for 20 minutes. The cells, now incubated with antibodies, were supplemented with 400 μ l of PBS. The samples were analysed using a FACS CaliburTM flow cytometer [27].

2.3. Clinical Trial Study

2.3.1. Study Design and Subject Recruitment

The investigative framework of this study is anchored in an observational design, integrated within clinical trials targeting patients afflicted with ST-segment elevation acute myocardial infarction (STEMI). These patients were subject to two distinct temporal interventions: primary percutaneous coronary intervention (PCI) executed within 12 hours of symptom onset, and delayed PCI initiated after 12 hours from onset. This expansive, multicenter research was conducted across several esteemed Cardiology Departments: Dr. Saiful Anwar Hospital in Malang City, Dr. Soebandi Hospital in Jember Regency, Binasehat Hospital in Jember Regency, Kaliwates General Hospital in Jember Regency, and Dr. Iskak Hospital in Tulungagung Regency. Data collection spanned from September 2022 to February 2023. In this randomized, double-masked trial, participants were allocated to either a control cohort receiving optimal medical therapy with a placebo or an intervention cohort treated with optimal medical therapy complemented by Colchicine administration. The sample pool included 249 patients, stratified as follows: 98 in the early placebo group, 50 in the early Colchicine group, 49 in the late placebo group, and 52 in the late Colchicine group. Sampling assessments were conducted at 24 hours and 5 days post-intervention. Ethical clearance for this study was granted under the local ethical approval number 400/235/K.3/302/2020, conferred by Saiful Anwar Hospital, Malang, Indonesia. Further to its ethical compliance, this study was registered with the international study registry ISRCTN, bearing the registration number ISRCTN12958502.

2.3.2. Enzyme-Linked Immunosorbent Assay (ELISA) Assay

Sera from Peripheral Blood Mononuclear Cell culturing was subjected to ELISA assay using the Human Interleukin 10, IL-10 ELISA Kit (BT Lab, Bioassay Technology Lab, Zhejiang, China, Cat No: E0102Hu) [28].

2.3.3. Data Analysis

The HIF-1 α and IL-10 data were subjected to One-way ANOVA (p<0.05) and followed by post hoc Tukey's test for comparisons. IL-10 level of sera analysed using T test to determine the difference between control group and intervention. The statistical analysis was generated using GraphPad Prism version 10.0.2 (Software, Inc., San Diego, CA, USA).

3. Results

3.1. In-Silico Study

3.1.1. Prediction of Colchicine Potential Based on SAR

profile **SMILES** The for colchicine is CC(=O)NC1CCC2=CC(=C(C(=C2C3=CC=C(C(=O)C=C13)OC)OC)OC)OC.Α compound is considered to have very high biological activity based on laboratory-scale tests when the Pa (Probability active) value exceeds 0.7 and is greater than Pi (Probability inactive), implying a significant resemblance to drug compounds in terms of similar bioactivity [29]. According to SAR analysis, Colchicine is predicted to act as a tubulin antagonist (0.744), beta-tubulin antagonist (0.673), NOS2 inhibitor (0.529), MAP kinase stimulant (0.44), antimyopathies (0.415), TNF expression inhibitor (0.364), and anti-inflammatory (0.308) (Figure 1) related to cardiac fibrosis and myocardial infarction.

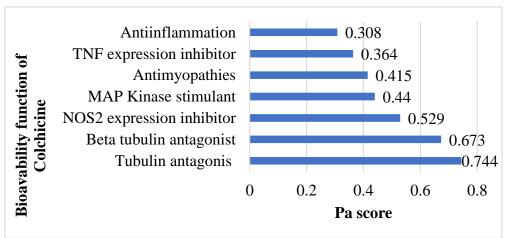


Figure 1. Potential bioavailability of Colchicine. Colchicine actively combats cardiac fibrosis and myocardial infarction by harnessing its anti-inflammatory and preventing excessive tubulin formation.

3.1.2. Prediction of Targets and Genes Related to Colchicine, AMI, and CF

Following the SAR analysis, we performed a modelling analysis to explore possible colchicine targets against CF and AMI. Exploring colchicine targets using CTD & DIGEP-Pred and Gene Disease Association with DisGeNET & Open Target revealed that Colchicine shares 27 targets with CF and AMI. IL-10 interacts with IL-1 β , IFN- γ , CCL2, TNF, and TGF- β 1 in AMI & CF conditions (Figure 2).

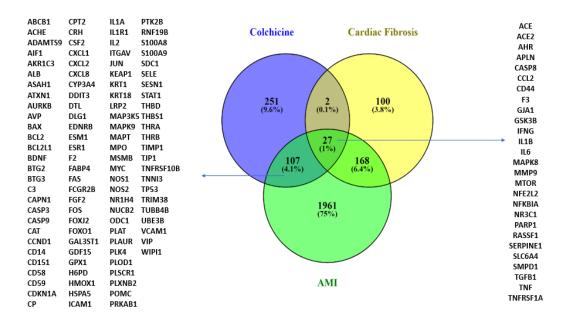


Figure 2. A Venn diagram illustrating colchicine gene targets in CF and AMI.

The targets of Colchicine include NR3C1 and IL-10. IL-10 interacts with IL-1 β , IFN- γ , CCL2, TNF, and TGF- β 1 under conditions associated with AMI and CF.

We annotated each colchicine target through the DAVID web server to identify the functions of these gene targets. As shown in Figure 3. the top 20 targets of colchicine in cardiac fibrosis (CF) and Acute myocardial infarction (AMI) represent their impact on biological responses like hypoxia and inflammation. To verify the reliability of our microarray analysis data, we considered genes with a false discovery rate (FDR) of less than 0.05 as significantly different [30].

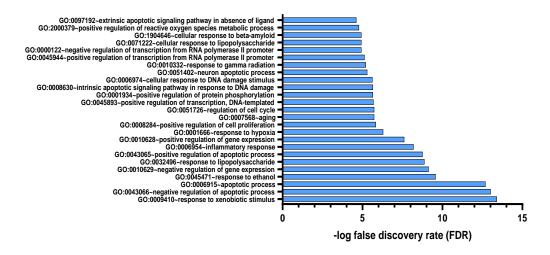


Figure 3. Gene Target Colchicine annotation with FDR < 0.05.

3.1.3. Pharmacology Network Analysis

The prediction scores were obtained based on network analysis using Cytoscape. Based on the entire pathway, the pathways targeted by Colchicine against AMI and cardiac fibrosis were particularly effective in modulating TNF, IL-6, and IL-10 (Supplementary Table 1). When considering laboratory targets, IL-10 interacts most with Colchicine (Figure 4).



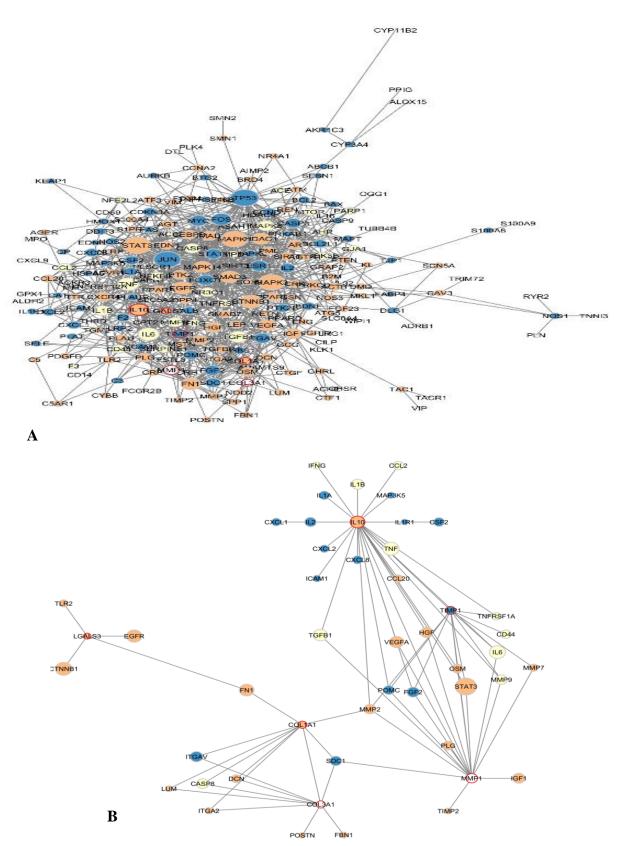


Figure 4. Analysing the Network Target Colchicine and Gene Disease Association in CF and AMI using Cytoscape. Blue (AMI-Colchicine), Orange (AMI-Cardiac Fibrosis), Yellow (AMI-Cardiac Fibrosis-Colchicine), Red (Laboratory Target). A. Protein-protein interaction network of genes related to CF and AMI. B. Functional clusters of protein-protein interactions.

3.2. In-Vitro Study

3.2.1. HIF-1 α Expression

Our research reveals that exposing 3T3 cell lines to 300 μ M CoCl₂ significantly boosts their viability, exceeding 73%. We measured HIF-1 α expression as a successful CoCl₂-mediated hypoxia stimulation marker to confirm the hypoxic model's efficacy in these cells. Our data showed that CoCl₂ treatment significantly induced the percentage of HIF-1 α -positive cells (78.87 ± 1.47%) compared to the control/ normoxia group (35.11 ± 1.21%, p<0.0001, Figure 5A).

Furthermore, colchicine treatment did not affect HIF-1 α expression in normoxic-3T3 cells (p>0.05). On the other hand, CoCl₂ treatment significantly suppressed HIF-1 α expression in hypoxic-3T3 cells (63.43 ± 4.49% vs 78.87 ± 1.47%, p<0.0001, Figure 5B), suggesting that Colchicine has protective role in alleviating hypoxia state in 3T3 cells.

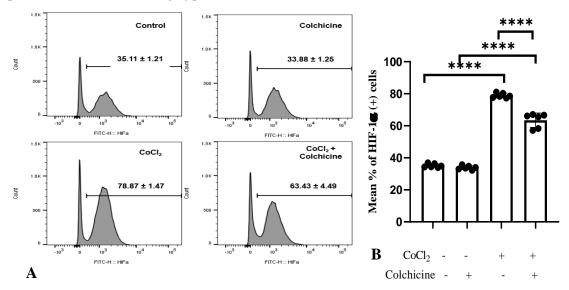


Figure 5. HIF-1 α expression in the treatment groups. (A, B). (1) Control: The cell culture is not exposed to Colchicine or CoCl₂. (2) Administration of 300 μM CoCl₂ for 24 hours significantly increased HIF-1 α expression in 3T3 cells compared to the control. (3) Administration of Colchicine to cells not experiencing hypoxia (without CoCl₂) resulted in a reduction in HIF-1 α compared to the control, but it was not statistically significant. (4) Administering Colchicine for 3 hours post-hypoxia (CoCl₂ 24 hours) reduced HIF-1 α expression compared to that in the CoCl₂ (hypoxia) group. ****p<0.00.

3.2.2. IL-10 Expression

To confirm the association between colchicine and the IL-10 pathway in fibroblasts, we investigated IL-10 expression in 3T3 cells exposed to hypoxia and colchicine. Cells were first treated with CoCl₂ (300 μ M) for 24 hours to induce hypoxia, mimicking the conditions studied in previous experiments. Colchicine (10 μ M) was added, and IL-10 levels were measured 24 hours later using flow cytometry (Figure 6A).

Our data showed that hypoxia stimulation significantly reduced the percentage of IL-10-positive cells compared to normoxic cells ($29.96 \pm 3.45\%$ vs. $48.48 \pm 4.29\%$, p<0.0001, Supplementary Figure 6), suggesting that hypoxia stimulation suppressed the anti-inflammatory properties of 3T3 cells. Interestingly, IL-10 expression in colchicine-treated hypoxic cells was significantly enhanced compared to its corresponding control group ($75.08 \pm 5.79\%$ vs. $48.48 \pm 4.29\%$, p<0.0001, Figure 6B), indicating that colchicine treatment inhibits proinflammatory properties of fibroblast cells.

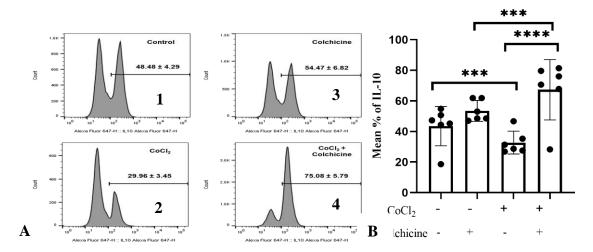


Figure 6. A. IL-10 expression in the treatment groups. A. (1) Control: The cell culture is not exposed to Colchicine or CoCl₂. (2) Administration of 300 μM CoCl₂ for 24 hours significantly reduced IL-10 expression in 3T3 cells compared to the control. (3) Administration of Colchicine to cells not experiencing hypoxia (without CoCl₂) resulted in an increase in IL-10 compared to the control, but it was not statistically significant. (4) Administering Colchicine for 3 hours post-hypoxia (CoCl₂ 24 hours) increased IL-10 expression compared to that in the CoCl₂ (hypoxia) group. Figure 6B. IL-10 expression was assessed using a one-way ANOVA test. The control group did not significantly differ from the colchicine group but showed significant differences when compared to both the CoCl₂ group and the CoCl₂ group. ***p<0.001, ****p<0.0001.

3.3. Clinical Trial Study

3.3.1. Baseline Characteristics of AMI Patients

The baseline patient profile in this heart disease study reflects a diverse demographic and clinical landscape. The average age is in the late fifties, showing slight intergroup variation, with no statistically significant differences in age (p>0.05). The majority of participants are male, with more than three-quarters in each group, and no significant variance in gender distribution (p>0.05). In terms of risk factors, a substantial proportion are former smokers, with the highest percentage in the early placebo group (87.76%) and the lowest in the late Colchicine group (50%), though without statistical significance (p>0.05). The prevalence of hypertension is consistent across all groups, with no meaningful differences (p>0.05). Incidences of Diabetes Mellitus (DM) and dyslipidemia are relatively low, with the highest occurrence of DM in the late Colchicine group (23.08%) and the lowest in the early and late placebo groups at around 10%. Dyslipidemia levels range from 9.62% to 22%, again with no significant differences (p>0.05).

Regarding the infarct-related artery, involvement of the left anterior descending (LAD) artery varies between 29.59% and 51.92%, and non-LAD arteries between 40.82% and 70.41%, with no significant differences between groups (p>0.05).

Table 1. Baseline clinical and echocardiographic AMI patient characteristics.

	Early placebo	Early colchicine	p valu e	Late placebo	Late colchicine	p valu e
	57.34±10.			56.39±10.8		
Age (years) (Mean±SD)	75	58.60±9.13	0.479	5	59.08±9.41	0.457
	84			41	40	
Sex (Male), n (%)	(85.71%)	42 (84%)	0.810	(83.67%)	(76.92%)	0.459

	86			33		
Smoker, n (%)	(87.76%)	43 (86%)	0.800	(67.35%)	26 (50%)	0.106
	34			32	35	
Hypertension, n (%)	(39.80%)	19 (38%)	0.720	(67.35%)	(67.31%)	0.837
			>1.00		12	
Diabetes Melitus, n (%)	9 (9.18%)	5 (10%)	0	6 (12.24%)	(23.08%)	0.197
	14					>1.00
Dyslipidaemia, n (%)	(14.29%)	11 (22%)	0.253	5 (10%)	5 (9.62%)	0
Infarct related artery	29		>1.00	29	27	
(LAD), n (%)	(29.59%)	15 (30%)	0	(59.18%)	(51.92%)	0.670
Infarct related artery (Non	69		>1.00	20	25	
LAD), n (%)	(70.41%)	35 (70%)	0	(40.82%)	(48.08%)	0.670

The table presents the demographic and clinical characteristics of cardiac patients enrolled in a study assessing the effects of colchicine. The data indicates an average age in the late fifties, a predominance of male gender, and a uniform distribution of cardiovascular risk factors—such as smoking history, hypertension, diabetes, and dyslipidemia—across both the intervention and control groups with no significant differences (p>0.05). Furthermore, there was no statistically significant variation in the involvement of infarct-related arteries (both LAD and non-LAD) across all groups (p>0.05), establishing a consistent baseline for subsequent response evaluations to colchicine treatment.

The research findings suggested that administering Colchicine to early STEMI patients for both 24 hours and five days did not result in a significant difference in IL-10 levels between the treatment groups, p > 0.05, Figure 7). However, it is noteworthy that the colchicine-treated group exhibited a tendency towards increased IL-10 levels compared to the non-colchicine group. This trend was observed at both the 24-hour (343.3 \pm 35.28 vs. 307.7 \pm 28.78) and five-day time points (302.2 \pm 40.81 vs. 263.2 \pm 25.82).

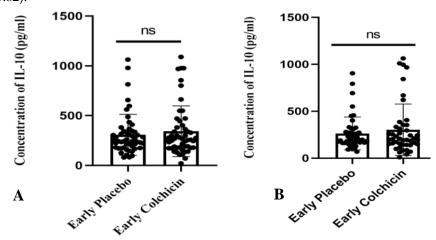


Figure 7. Figure illustrates the administration of both placebo and colchicine at early 24 hours (A) and five days (B) in STEMI patients. It demonstrated no significant difference in IL-10 levels (*P*>0.05). NS: no significance. IL-10 expression was assessed using a T-test.

In contrast, a separate study found that administering colchicine to STEMI patients at late 24 hours did not result in a significant difference in IL-10 levels between the treatment and control groups (p>0.05, as shown in Figure 8A). However, the mean IL-10 level in the colchicine group displayed a trend towards being higher than the control group (413.8 \pm 46.17 vs. 311.5 \pm 31.02), as illustrated in Figure 8. Nevertheless, administration of late five days colchicine significantly differed,

showing higher IL-10 levels compared to the non-colchicine group (p<0.01, as shown in Figure 8B), with average levels of 440.7±50.95 vs 263.8±24.25).

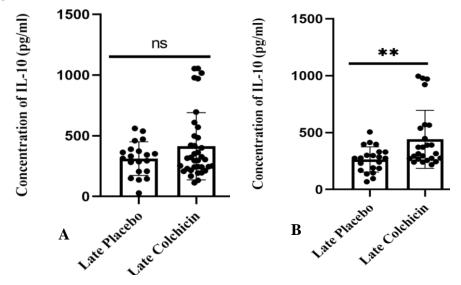


Figure 8. Depicts late colchicine administration at 24 hours (A) showed no significant difference (P>0.05), while at five days (B), there was a significant difference in IL-10 levels. NS: no significance. IL-10 expression was assessed using a T test.

4. Discussion

Age is an independent risk factor for cardiovascular disease (CVD), with a high prevalence in older age groups, highlighting the need for preventive interventions and proactive management [31] . Modifiable risk factors such as hypertension, diabetes mellitus, smoking, and dyslipidemia also significantly contribute to heart disease. Men are more susceptible to coronary heart disease at a younger age due to high testosterone levels, which increase Low Density Lipoprotein (LDL) cholesterol and decrease High Density Lipoprotein (HDL) cholesterol [32]. Hypertension increases workload on the heart and pressure on artery walls, which can lead to atherosclerosis, heart attacks, and strokes [33]. Diabetes mellitus accelerates atherosclerosis through endothelial dysfunction, inflammation, and hypercoagulability. Managing these risk factors is crucial for preventing disease progression and improving patient outcomes [34]. Post-myocardial infarction left ventricular remodelling (LVR) is characterized by a ≥20% increase in left ventricular end-diastolic volume (LVEDV) as measured by echocardiography from day 4 to month 6 following the myocardial infarction event [35] . High LVEDV indicates adverse ventricular remodelling or diastolic dysfunction and is an important predictor for congestive heart failure and mortality [36]. Major Adverse Cardiac Events (MACE) is the most significant and serious primary outcome in CVD, encompassing deaths due to heart disease or related complications MACE measurement in clinical trials or patient management often includes multiple endpoints, such as death, myocardial infarction, stroke, and need for revascularization procedures, which provide a more comprehensive picture of cardiovascular risk and intervention effectiveness [37]. Low MACE events in this study suggest either good safety profiles for colchicine and placebo or low patient population risk for MACE during the study period.

Colchicine is predicted to play a significant role in cardiac fibrosis and myocardial infarction, with a score greater than 0.5 as a tubulin antagonist, beta-tubulin antagonist, and NOS2 inhibitor. Colchicine's mechanism of action involves binding to tubulin, thus hindering microtubule assembly and polymerization. This prediction aligns with the research conducted by McLoughlin and O'Boyle, which states that Colchicine works by blocking β -tubulin and subsequently disrupting microtubule assembly [38]. Microtubules, crucial components of the cytoskeleton, are involved in various cellular processes, including maintaining cell shape, intracellular transport, cytokine and chemokine secretion, cell migration, ion channel regulation, and cell division, and are responsible for the

subcellular arrangement of mitochondria and cardiac muscle cell dynamics [39]. Colchicine inhibits microtubule growth at low concentrations, while higher concentrations induce microtubule depolymerization. Microtubule depolymerization disrupts neutrophil adhesion and recruitment to inflamed tissues [40].

Colchicine targets related to CF and AMI impact biological responses such as hypoxia and inflammation. Hypoxia or ischemia disrupts endothelial cell barrier integrity, increasing blood vessel permeability and initiating leukocyte infiltration. If ischemia persists for an extended period, it can activate programmed cell death and myocardial cell necrosis, including necrosis, apoptosis, and autophagy mechanisms. Subsequent reperfusion exacerbates tissue damage due to sudden reoxygenation of reactive oxygen species (ROS) generation and complement pathway activation [38]. Necrotic cells and stressed/damaged extracellular matrices release danger-associated molecular patterns (DAMPs) and activate a series of inflammatory mediators, including inflammatory cytokines, chemokines (such as monocyte chemoattractant protein-1/chemokine (C-C motives) ligand 2 [CCL2]), and cell adhesion molecules. In the case of MI, this signaling will activate mitogenactivated protein kinases (MAPKs) and nuclear factor (NF)-κB, leading to inflammation [41].

The Colchicine target IL-10 interacts with IL-1 β , IFN- γ , CCL2, TNF- α , and transforming growth factor (TGF- β 1) in conditions such as AMI and CF. Cytokines such as interleukin-1 (IL-1) and monocyte chemoattractant protein-1 (MCP-1) increase during MI (Hartman et al. 2018). Colchicine reduces inflammatory cytokines and TGF- β 1 by inhibiting NF- κ B signalling and activating the NLRP3 inflammasome. IL-10 is produced by T cells and macrophages during acute and chronic inflammation, playing a role in inhibiting apoptosis and TNF- α -induced oxidative stress. Receptormediated IL-10 signaling in the heart controls myocardial hypertrophy in response to excessive pressure stimulation. IL-10 can also reduce proinflammatory cytokines such as TNF- α , IL-1 β , and IL-6, as well as cardiac contractility in heart failure [42].

Hypoxia plays a role in both physiological and pathophysiological conditions. Physiologically, it is involved in processes such as cell differentiation and embryogenesis [43]. Conversely, in pathophysiological scenarios, hypoxia is associated with conditions such as brain and heart ischemia [44]. Myocardial infarction triggers events that lead to the repair and healing of the left ventricle, resulting in scar tissue formation [45]. This process involves critical inflammatory responses necessary for heart repair. However, it also has implications for postinfarction remodelling and heart failure [46].

Cobalt chloride (CoCl₂) induces hypoxia by reducing oxygen levels [43]. It increases the stability of hypoxia-inducible factor (HIF) 1α , linked to changes in cellular oxygen levels [38]. Administration of CoCl₂ led to a significant decrease in IL-10 expression in 3T3 cells compared to the control group [47]. This research aligns with the findings of Tao et al. showed that IL-10 expression decreases at 4, 8, and 16 weeks after myocardial infarction, indicating a correlation between reduced IL-10 and impaired heart function [19].

The administration of Colchicine to nonhypoxic cells did not significantly differ from IL-10 expression in the control group. In the control group, IL-10 expression functions in tissue repair and energy metabolism [45]. IL-10 suppresses the infiltration of inflammatory cells and the expression of proinflammatory cytokines in the myocardium by inhibiting fibrosis through the suppression of p38 mitogen-activated protein kinase (MAPK) and by increasing capillary density through the activation of signal transducer and activator of transcription (STAT3), which promotes myocardial capillary growth and protects the heart from ischemic injury [5]. The administration of Colchicine three hours after hypoxia (after 24 hours of CoCl2exposure) also shows an increase in IL-10 expression compared to the hypoxia group. After MI, leukocytes infiltrate the infarcted area to clear necrotic remnants and respond to myocardial injury by producing proinflammatory cytokines that peak within one-hour post-MI. Between days 3 and 5 post-MI, there is a transition from inflammatory responses to reparative and anti-inflammatory processes, marked by the acceleration of fibroblast activation and proliferation. Prolonged inflammatory responses can disrupt the physiology of the left ventricle (LV), leading to LV dilation and excessive scar tissue formation. Therefore, expediting inflammatory response resolution is critical, and anti-inflammatories such as IL-10 are needed to suppress

inflammation and activate profibrotic processes. IL-10 plays a crucial role in controlling inflammation by regulating the function of macrophages to adopt M2 (anti-inflammatory) polarization [48] and dendritic cells [49]. This research aligns with the findings of Stumpf et al. [50].

The clinical trial results align with the in vitro findings, indicating that administering late Colchicine to STEMI patients can elevate IL-10 levels compared to those not receiving it on the fifth day. CD4 $^{+}$ T cells and Foxp $^{3+}$ regulatory T cells infiltrate the myocardium within days after MI [51]. IL-10 acts as an anti-inflammatory agent by inhibiting antigen presentation by macrophages, suppressing T-cell proliferation, deactivating dendritic cells, or inducing regulatory T-cell function [52], thereby reducing inflammatory products [53] to prevent rupture and left ventricular dilation [51]. Colchicine, as an anti-inflammatory agent in STEMI conditions, aligns with the research conducted by Kaur et al. (2006), demonstrating that the administration of anti-inflammatory drugs such as steroids and dexamethasone for cardiac function improvement can increase IL-10 levels and decrease inflammatory cytokines like TNF- α and NF- κ β transcription factor [54].

In conditions of ischemic heart disorders, macrophages deficient in IL-10 contribute to a shift in macrophage phenotype towards a profibrotic subset, activating fibroblasts. Increased fibroblasts and collagen deposition lead to myocardial relaxation disorders, especially diastolic [55]. Myocardial relaxation disorders occur because inflammatory cytokines in fibroblasts, such as IL-1 β , TNF- α , and IL-6, inhibit proliferation, reduce matrix synthesis, and increase MMP activity [56]. IL-10 in macrophages also functions to reduce ROS, resulting in a decrease in fibroblast numbers [55].

5. Conclusions

Colchicine acts as an anti-inflammatory and antifibrotic agent by increasing IL-10 levels in STEMI patients, supported by in vitro data on 3T3 cell lines and in-silico analysis based on network pharmacology. Colchicine can be used to support the inhibition of heart function disorders. Patients with heart disease who had higher IL-10 levels with colchicine administration in this study experienced protection against progression of increased LVEDV, specifically in the context of managing left ventricular remodelling and preventing heart failure.

In summary, we conclude that the IL-10 pathway is Colchicine's main potential target for ventricular remodelling post-MI, particularly in cardiac fibrosis development. In this study, we showed that a combined approach of network pharmacology along with in vitro experiments to validate the findings is beneficial to unravel the mechanism of Colchicine in this specific pathological entity, age, modifiable risk factors, sex, and LVEDV are important factors in CVD. Risk factor management and MACE measurement are crucial for improving patient outcomes. However, further studies are required to understand how Colchicine affects cardiac fibrosis development in response to MI.

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References

- 1. Kaptoge, S.; Pennells, L.; De Bacquer, D. World Health Organization cardiovascular disease risk charts: revised models to estimate risk in 21 global regions. *The Lancet Global Health* **2019**, 7, pp.e1332–45. Available online: https://doi.org/10.1016/S2214-109X(19)30318-3
- 2. Kementrian Kesehatan RI. Riset Kesehatan Dasar (Riskesdas) 2018.
- 3. Reddy, H.; Hanif, A.M.; Rizkian, T.; Wahyudi. The Role of Galectin-3 in Acute Myocardial Infarction: A Narrative Literature Review. *Bioscmed* **2022**, 6, 2858–2865. Available online: https://doi.org/10.37275/bsm.v6i16.709
- 4. Michiels, C. Physiological and Pathological Responses to Hypoxia. *The American Journal of Pathology* **2004**, 164, pp.1875–82. Available online: https://doi.org/10.1016/S0002-9440(10)63747-9
- 5. Krishnamurthy, P.; Rajasingh, J.; Lambers, E. IL-10 Inhibits Inflammation and Attenuates Left Ventricular Remodeling After Myocardial Infarction via Activation of STAT3 and Suppression of HuR. *Circulation Research* **2009**, 104. Available online: https://doi.org/10.1161/CIRCRESAHA.108.188243
- 6. Hartman, M.H.T.; Groot, H.E.; Leach, I.M. Translational overview of cytokine inhibition in acute myocardial infarction and chronic heart failure. *Trends in Cardiovascular Medicine* **2018**, 28, pp. 369–379. Available online: https://doi.org/10.1016/j.tcm.2018.02.003
- 7. Gourdie, R.G.; Dimmeler, S.; Kohl, P. Novel therapeutic strategies targeting fibroblasts and fibrosis in heart disease. *Nat Rev Drug Discov* **2016**, 15(9), pp. 620-38. Available online: doi: https://doi.org/10.1038/nrd.2016.89.
- 8. Ma, J.; Tai, Y.; Fan, M.; Wang, Z. Cardiac Rehabilitation of Patients with Acute ST-Elevation Myocardial Infarction Undergoing Primary Percutaneous Coronary Intervention in a Han Population in Northern China: A Prospective Cohort Study. *Int J Gen Med* **2021**, 4, pp. 4959–65.
- 9. Suthahar, N.; Meijers, W.C.; Silljé, H.H.W.; De Boer, R.A. From Inflammation to Fibrosis—Molecular and Cellular Mechanisms of Myocardial Tissue Remodelling and Perspectives on Differential Treatment Opportunities. *Curr Heart Fail Rep* **2017**, 14, pp. 235–250. Available online: https://doi.org/10.1007/s11897-017-0343-y
- Chacko, L.P.; Howard, J.; Rajkumar, C. Effects of Percutaneous Coronary Intervention on Death and Myocardial Infarction Stratified by Stable and Unstable Coronary Artery Disease: A Meta-Analysis of Randomized Controlled Trials. Circ Cardiovascular Quality and Outcomes 2020, 13, p. e006363. Available online: https://doi.org/10.1161/CIRCOUTCOMES.119.006363
- 11. Zheng, W.; Yu, C.; Liu, J. Patients with ST-segment elevation of myocardial infarction miss out on early reperfusion: when to undergo delayed revascularization. *J Geriatr Cardiol* **2017**, 14, pp. 524–531
- 12. Ribeiro, S.A.; Lopes, C.; Amaral, R.; Amaral, A. The therapeutic potential of colchicine in the complications of COVID19. Could the immunometabolic properties of an old and cheap drug help? *Metabolism Open* **2020**, 7, p. 100045. Available online: https://doi.org/10.1016/j.metop.2020.100045
- 13. Nawabi, A.Q.; Hassan, W.; Chen, L. Is Colchicine a New Game-Changer in Patients with Acute Coronary Syndrome? *Cureus* **2022**. Available online: https://doi.org/10.7759/cureus.22874
- 14. Diaz-Arocutipa, C.; Benites-Meza, J.K.; Chambergo-Michilot, D. Efficacy and Safety of Colchicine in Postacute Myocardial Infarction Patients: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Front Cardiovasc Med* **2021**, 8, p. 676771. Available online: https://doi.org/10.3389/fcvm.2021.676771
- 15. Zhang, D.; Li, L.; Li, J. Colchicine improves severe acute pancreatitis-induced acute lung injury by suppressing inflammation, apoptosis and oxidative stress in rats. *Biomedicine & Pharmacotherapy* **2022**, 153, p. 113461. Available online: https://doi.org/10.1016/j.biopha.2022.113461
- 16. Liu, H.; Beck, T.N.; Golemis, E.A.; Serebriiskii, I.G. Integrating in silico resources to map a signaling network. *Methods Mol Biol* **2014**, 1101, pp. 197-245. Available online: https://doi.org/10.1007/978-1-62703-721-1 11.
- 17. Basanagouda, M.; Jadhav, V.B.; Kulkarni, M.V.; Rao, R.N. Computer Aided Prediction of Biological Activity Spectra: Study of Correlation between Predicted and Observed Activities for Coumarin-4-Acetic Acids. *Indian J Pharm Sci* **2015**, 73, pp. 88–92
- 18. Mujalli, A.; Banaganapalli, B.; Alrayes, N.M. Myocardial infarction biomarker discovery with integrated gene expression, pathways and biological networks analysis. *Genomics* **2020**, 112, pp. 5072-85. Available online: https://doi.org/10.1016/j.ygeno.2020.09.004
- 19. Tao, Y.; Gao, C.; Qian, D. Regulatory mechanism of fibrosis-related genes in patients with heart failure. *Front Genet* **2022**, 13, p. 1032572. Available online: https://doi.org/10.3389/fgene.2022.1032572

- 20. Shahid, M.; Azfaralariff, A.; Law, D. Comprehensive computational target fishing approach to identify Xanthorrhizol putative targets. *Sci Rep* **2021**, 11, p. 1594. Available online: https://doi.org/10.1038/s41598-021-81026-9
- 21. Wu, Y.; Zhu, Y.; Xie, N. A network pharmacology approach to explore active compounds and pharmacological mechanisms of a patented Chinese herbal medicine in the treatment of endometriosis. *PLoS ONE* **2022**, 17, p. e0263614. Available online: https://doi.org/10.1371/journal.pone.0263614
- 22. Kim, T.H.; Yu, G.R.; Kim, H. Network Pharmacological Analysis of a New Herbal Combination Targeting Hyperlipidemia and Efficacy Validation In Vitro. *CIMB* **2023**, 45, pp. 1314–32. Available online: https://doi.org/10.3390/cimb45020086
- 23. Foutch, D.; Pham, B.; Shen, T. Protein conformational switch discerned via network centrality properties. *Computational and Structural Biotechnology Journal* **2021**, 19, pp. 3599–608. Available online: https://doi.org/10.1016/j.csbj.2021.06.004
- 24. Zhao, C.; Moreno-Nieves, U.; Di Battista, J.A. Chemical Hypoxia Brings to Light Altered Autocrine Sphingosine-1-Phosphate Signalling in Rheumatoid Arthritis Synovial Fibroblasts. *Mediators of Inflammation* **2015**, pp. 1–12. Available online: https://doi.org/10.1155/2015/436525
- 25. Liu, R.; Liu, Q.; Wang, K. Comparative analysis of gene expression profiles in normal hip human cartilage and cartilage from patients with necrosis of the femoral head. *Arthritis Res Ther* **2016**, 18, p. 98. Available online: https://doi.org/10.1186/s13075-016-0991-4
- 26. Wang Y, Zhang Y, Li J, et al. (2023b) Hypoxia Induces M2 Macrophages to Express VSIG4 and Mediate Cardiac Fibrosis After Myocardial Infarction. *Theranostics* **2023**, 13:2192–2209. Available online: https://doi.org/10.7150/thno.78736
- 27. Lestari, S.R.; Rifa'i, M. Regulatory T cells and anti-inflammatory cytokine profile of mice fed a high-fat diet after single-bulb garlic (*Allium sativum L.*) oil treatment. *Trop J Pharm Res* **2019**, 17, p. 2157. Available online: https://doi.org/10.4314/tjpr.v17i11.7
- 28. Al-Masoudi, F.I.; Ali, B.H. Evaluation of Nesfatin 1 and Other Biochemical Markers in diabetic Neuropathy Iraqi patients before and after treatment with tegretol. *Annals of the Romanian Society for Cell Biology* **2021**, pp. 9422–29. Available online: http://annalsofrscb.ro/index.php/journal/article/view/3682
- 29. Emon, N.U.; Alam, S.; Rudra, S. Antidepressant, anxiolytic, antipyretic, and thrombolytic profiling of methanol extract of the aerial part of *Piper nigrum*: In vivo, in vitro, and in silico approaches. *Food Science & Nutrition* **2021**, 9, pp. 833–46. Available online: https://doi.org/10.1002/fsn3.2047
- 30. Liu, Y.; Xu, S.; Zu, T. Reversal of TET-mediated 5-hmC loss in hypoxic fibroblasts by ascorbic acid. *Laboratory Investigation* **2019**, 99, pp. 1193–202. Available online: https://doi.org/10.1038/s41374-019-0235-8
- 31. Qiu, L.; Wang, W.; Sa, R.; Liu, F. Prevalence and Risk Factors of Hypertension, Diabetes, and Dyslipidemia among Adults in Northwest China. *International Journal of Hypertension* **2021**, pp. 1–10. Available online: https://doi.org/10.1155/2021/5528007
- 32. Boden, W.E.; Miller, M.G.; McBride, R. Testosterone concentrations and risk of cardiovascular events in androgen-deficient men with atherosclerotic cardiovascular disease. *American Heart Journal* **2020**, 224, pp. 65–76. Available online: https://doi.org/10.1016/j.ahj.2020.03.016
- 33. Libby, P. The changing landscape of atherosclerosis. *Nature* **2021,** 592, pp. 524–33. Available online: https://doi.org/10.1038/s41586-021-03392-8
- 34. Ambreen, S.; Fatima, S.; Elwakiel, A. Hypercoagulability Impairs Plaque Stability in Diabetes-Induced Atherosclerosis. *Nutrients* **2022**, 14, 1991. Available online: https://doi.org/10.3390/nu14101991
- 35. Logeart D.; Taille, Y.; Derumeaux, G. Patterns of left ventricular remodeling post-myocardial infarction, determinants, and outcome. *Clin Res Cardiol* **2024**. Available online: https://doi.org/10.1007/s00392-023-02331-z
- 36. Lok, D.J.; Lok, S.I.; Bruggink-André, De La Porte, P.W. Galectin-3 is an independent marker for ventricular remodeling and mortality in patients with chronic heart failure. *Clin Res Cardiol* **2013**, 102, pp. 103–10. Available online: https://doi.org/10.1007/s00392-012-0500-y
- 37. Bosco, E.; Hsueh, L.; McConeghy, K.W. Major adverse cardiovascular event definitions used in observational analysis of administrative databases: a systematic review. *BMC Med Res Methodol* **2021**, 21, 241. Available online: https://doi.org/10.1186/s12874-021-01440-5
- 38. McLoughlin, E.C.; O'Boyle, N.M. Colchicine-Binding Site Inhibitors from Chemistry to Clinic: A Review. *Pharmaceuticals (Basel)* **2020**, 13(1), p. 8. doi: 10.3390/ph13010008. Erratum in: Pharmaceuticals (Basel). 2020 Apr 20;13(4):E72. Available online: https://doi.org/10.3390/ph13040072.
- 39. Puurand, M.; Tepp, K.; Timohhina, N. Tubulin βII and βIII Isoforms as the Regulators of VDAC Channel Permeability in Health and Disease. *Cells* **2019**, 8, p. 239. Available online: https://doi.org/10.3390/cells8030239
- 40. Leung, Y.Y.; Hui, L.L.Y.; Kraus, V.B. Colchicine --- update on mechanisms of action and therapeutic uses. *Semin Arthritis Rheum* **2015**, 45, pp. 341–50. Available online: https://doi.org/10.1016/j.semarthrit.2015.06.013

- 41. Prabhu, S.D.; Frangogiannis, N.G. The Biological Basis for Cardiac Repair After Myocardial Infarction: From Inflammation to Fibrosis. *Circ Res* **2016**, 119, pp. 91–112. Available online: https://doi.org/10.1161/CIRCRESAHA.116.303577
- 42. Stafford, N.; Assrafally, F.; Prehar, S. Signaling via the Interleukin-10 Receptor Attenuates Cardiac Hypertrophy in Mice During Pressure Overload, but not Isoproterenol Infusion. *Front Pharmacol* **2020**, 11, p. 559220. Available online: https://doi.org/10.3389/fphar.2020.559220
- 43. Muñoz-Sánchez, J.; Chánez-Cárdenas, M.E. The use of cobalt chloride as a chemical hypoxia model. *J of Applied Toxicology* **2019**, 39, pp. 556–70. Available online: https://doi.org/10.1002/jat.3749
- 44. Michiels, C. Physiological and Pathological Responses to Hypoxia. *The American Journal of Pathology* **2004**, 164, pp. 1875–82. Available online: https://doi.org/10.1016/S0002-9440(10)63747-9
- 45. Jung, M.; Ma, Y.; Iyer, R.P. IL-10 improves cardiac remodeling after myocardial infarction by stimulating M2 macrophage polarization and fibroblast activation. *Basic Res Cardiol* **2017**, 112, 33. Available online: https://doi.org/10.1007/s00395-017-0622-
- 46. Frangogiannis, N.G. The inflammatory response in myocardial injury, repair, and remodelling. *Nat Rev Cardiol* **2014**, 11, pp. 255–65. Available online: https://doi.org/10.1038/nrcardio.2014.28
- 47. Kandilarov, I.; Gardjeva, P.; Georgieva-Kotetarova, M.; Zlatanova, H.; Vilmosh, N.; Kostadinova, I.; Katsarova, M.; Atliev, K.; Dimitrova, S. Effect of Plant Extracts Combinations on TNF-α, IL-6 and IL-10 Levels in Serum of Rats Exposed to Acute and Chronic Stress. *Plants* **2023**, 12, 3049. Available online: https://doi.org/10.3390/plants12173049
- 48. Cabral-Santos, C.; De Lima Jr, E.A.; Fernandes, I.M.D.C. Interleukin-10 responses from acute exercise in healthy subjects: A systematic review. *Journal Cellular Physiology* **2019**, 234, pp. 9956-65. Available online: https://doi.org/10.1002/jcp.27920
- 49. Shoemaker, J.; Saraiva, M.; O'Garra, A. GATA-3 Directly Remodels the *IL-10* Locus Independently of IL-4 in CD4+ T Cells. *The Journal of Immunology* **2006**, 176, pp. 3470–79. Available online: https://doi.org/10.4049/jimmunol.176.6.3470
- 50. Stumpf, C.; Petzi, S.; Seybold, K. Atorvastatin enhances interleukin-10 levels and improves cardiac function in rats after acute myocardial infarction. *Clinical Science* **2009**, 116, pp. 45–52. Available online: https://doi.org/10.1042/CS20080042
- 51. Hofmann, U.; Frantz, S. Role of T-cells in myocardial infarction. *Eur Heart J* **2016**, 37, pp. 873–9. Available online: https://doi.org/10.1093/eurheartj/ehv639
- 52. Meng, X.; Li, J.; Yu, M. Transplantation of mesenchymal stem cells overexpressing IL10 attenuates cardiac impairments in rats with myocardial infarction. *Journal Cellular Physiology* **2018**, 233, pp. 587-95. Available online: https://doi.org/10.1002/jcp.25919
- 53. Lakoski, S.G.; Liu, Y.; Brosnihan, K.B.; Herrington, D.M. Interleukin-10 concentration, and coronary heart disease (CHD) event risk in the estrogen replacement and atherosclerosis (ERA) study. *Atherosclerosis* **2008**, 197, pp. 443–7. Available online: https://doi.org/10.1016/j.atherosclerosis.2007.06.033
- 54. Kaur, K.; Sharma, A.K.; Singal, P.K. Significance of changes in TNF-α and IL-10 levels in the progression of heart failure subsequent to myocardial infarction. *American Journal of Physiology-Heart and Circulatory Physiology* **2006**, 291, pp. H106–13. Available online: https://doi.org/10.1152/ajpheart.01327.2005
- 55. Hulsmans, M.; Sager, H.B.; Roh, J.D. Cardiac macrophages promote diastolic dysfunction. *Journal of Experimental Medicine* **2018**, 215, pp. 423–40. Available online: https://doi.org/10.1084/jem.20171274
- 56. Frieler, R.A.; Mortensen, R.M. Immune Cell and Other Noncardiomyocyte Regulation of Cardiac Hypertrophy and Remodeling. *Circulation* **2015**, 131, pp. 1019–30. Available online: https://doi.org/10.1161/CIRCULATIONAHA.114.008788

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