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Article

Internal Validation of a Machine Learning Model for Antimicrobial Stewardship: Evaluating Trainability of Data and the Accuracy of Clinical Recommendations within a Clinical Decision Support System

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Abstract: Background: Antimicrobial stewardship programs (ASPs) are essential in combating antimicrobial resistance (AMR), however, limited resources hinder their implementation. Arkstone, a biotechnology company, developed a machine learning (ML)-driven clinical decision support system (CDSS) to guide antimicrobial prescribing. Though widely used, the model had not been previously evaluated. **Methods:** Three components of the ML system were assessed: (1) A prospective observational study tested its ability to distinguish trained from novel data using various validation techniques and BioFire molecular panel inputs; (2) An anonymous retrospective analysis of internal infectious disease lab results evaluated recognition of novel versus trained complex datasets; and (3) A prospective observational validation study reviewed clinical recommendations against standard guidelines by independent clinicians. **Results:** The system achieved 100% accuracy (F1=1) in identifying 111 unique novel data points across 1,110 tests over nine training sessions. It correctly identified all 519 fully trained and 644 novel complex datasets. Among 644 clinician-trained reports, there were no major discrepancies in recommendations, with only 100 showing minor differences. **Conclusion:** This novel ML system demonstrated high accuracy in distinguishing trained from novel data and produced recommendations consistent with clinical guidelines. These results support its value in strengthening CDSS and ASP efforts.

Keywords: machine learning; antimicrobial stewardship; antibiotic resistance; clinical decision support

1. Introduction

Machine learning (ML), a subset of artificial intelligence (AI), has rapidly transformed diverse sectors, including healthcare, by enabling systems to learn from data, identify patterns, and make decisions with minimal human intervention [1,2]. In clinical settings, ML-based clinical decision support systems (CDSS) are increasingly used to aid in diagnosis, treatment planning, and antimicrobial stewardship [3–5]. A critical element in the deployment of these ML models is the rigorous validation process, which ensures the model's reliability, generalizability, and accuracy when presented with new and complex data. This process is important, as models trained on limited data can overfit, capturing noise rather than relevant patterns, leading to poor performance when

exposed to new data [6]. Furthermore, a lack of structured data review processes in some ML systems raises concerns about the accuracy of their recommendations, particularly in healthcare settings where critical variables might not be included in the analysis [7].

The potential of ML-driven CDSS to improve patient care and public health is considerable, yet there are significant barriers to widespread adoption; one of these is the lack of comprehensive validation techniques [8] and confidence in an accurate working model. As previous attempts to implement ML-based systems in healthcare have shown, challenges such as poor data integration, concerns about data privacy, limited clinical applicability, and inaccurate recommendations have led to the discontinuation of various clinical support systems such as IBM Watson for Oncology [9–11] and DeepMind Health's Streams [12–14]. These experiences highlight the need for robust, ethical, and clinically focused validation approaches to ensure the safe and effective integration of ML into clinical practice [15]. In addition, due to the rapid pace of evolving technology, formal validation methods are lacking, with limited data on the ideal validation process and evidence that ML processes are accurate [16]. Lastly, because of the uniqueness of the ML model in this study, to our knowledge, there are no studies evaluating its capabilities and its validation methods. Therefore, it is essential, we examine and test the capabilities of this system [17,18].

Arkstone, a biotechnology company, has developed a unique ML model for real-time, patient-specific infectious disease guidance integrated with laboratory results. This system provides clinicians with actionable recommendations aligned with clinical guidelines, intending to improve antimicrobial stewardship and reduce antibiotic overuse.

This study aimed to internally validate the system by evaluating its performance in training data and the system's ability to recall the trained data and distinguish it from new data accurately. By analyzing the ML model, the integration of these tools into clinical practice can be done confidently. In addition, this study will evaluate the model's robustness, accuracy, and generalizability for clinical use across diverse settings by evaluating the accuracy of the clinical recommendations themselves, which requires human input (human-in-the-loop (HITL) machine learning). This study aims to provide evidence to support the use of similar tools that enhance infectious disease knowledge and antimicrobial stewardship, particularly in resource-limited environments.

2. Materials and Methods

2.1. Model Description

Data entered the ML model occurs via results sent by laboratories. This is typically done in real-time via an HL7 or API interface but can also be done through manual uploads. Data sent includes patient demographics, laboratory findings (organisms, antibiotic susceptibility, and resistance genes), sample sources, diagnostic codes, allergies, and pregnancy status. Within seconds to minutes, a concise, single-page PDF that provides recommendations on the appropriate antimicrobial, if applicable ([Supplement 1](#)).

For scalable and effective processing of data, a unique machine-learning model was developed that incorporates multiple validation techniques simultaneously, including applied K-Fold cross-validation, random subsampling, and holdout validation. The combination of methods, also called Antimicrobial Intelligence, is applied in real-time (prospective validation), leveraging live data streams. A key component of this is the system's inability to suggest new treatment options as well as to provide recommendations to untrained data sets. The system, therefore, relies on HITL processes on multiple levels to ensure the data is trained accurately. This step is critical at every stage of data training, ensuring that expert oversight is consistently applied. In addition to requiring human approval to train data, HITL is required and repeated by different infectious disease experts to ensure consistency and accuracy on how the data is trained, minimizing the risk of human error and bias. Furthermore, once the data is finally trained by multiple infectious disease experts, it does not stay in this status indefinitely. Data that has been rigorously trained previously gets pushed back into a status requiring it to be trained again, ensuring that data is repeatedly and periodically

retrained so that information is up-to-date and error-free. This also allows for updates to medical recommendations that may have changed since initial training. This hybrid model that incorporates both human oversight and set algorithms ensures its adaptability to new and ever-evolving data.

The validation process is divided into three key elements that will be evaluated separately: 1) Evaluation of the system’s ability to distinguish and recall trained from new single data points; 2) Evaluation of the system’s ability to distinguish and recall trained from new complex data sets; (Figure 1) and 3) Evaluation of the accuracy of the clinical recommendations outputted by the system.

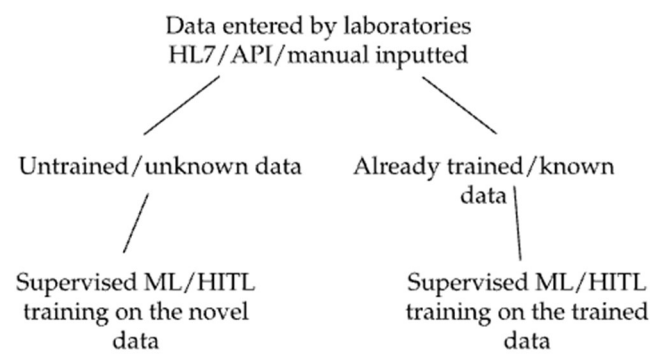


Figure 1. Simplified diagram of processes in Arkstone ML.

2.2. Data Sources and Preparation

Data were obtained from the Arkstone laboratory results database. The data set included positive and negative microbiology results, as well as demographic data such as patient age and sex, ICD-10-CM codes, allergies, pregnancy status, source of specimen, organism, sensitivity information, and resistance gene information. Diagnostic modalities included molecular or standard culture techniques. Prior to analysis, all data were de-identified according to HIPAA (Health Insurance Portability and Accountability Act of 1996) guidelines. Data were pre-processed to ensure consistency in format and coding.

2.3. Study Design

This validation study consisted of two components: a prospective observational phase and a retrospective analysis of real-world data from Arkstone’s database. The study was conducted in three sequential phases, each designed to evaluate different aspects of the machine learning system’s clinical performance. The research team accessed the dataset remotely from their respective locations, without intervening in clinical care or altering the existing workflow of the system. Researchers reviewing reports were not the same individuals involved in formulating the initial clinical recommendations to avoid confirmation bias.

The primary objective of the study was to evaluate the internal validation process of the machine learning (ML) model through three critical components: a) Recognition of novel versus trained data: This component assessed the model’s ability to distinguish between previously unseen (untrained) data and data used during the model’s training process; b) Recognition of complex datasets: This involved evaluating the system’s performance when classifying large and heterogeneous datasets composed of multiple data points from various sources; and c) Human-in-the-Loop (HITL) component: This assessed the accuracy of treatment recommendations generated by the system which requires human input via the HITL process.

2.4. Ethical Considerations and Data Availability

The study protocol was approved by the Institutional Ethics Committee of the Faculty of Health Sciences at the Private University of Tacna (FACSA-CEI/224-12-2024). All procedures complied with the Declaration of Helsinki and HIPAA guidelines. The study involved no risk to patients, and informed consent was not required, as all data were either retrospective, anonymized, or publicly available. The dataset consisted of publicly accessible BioFire panel results and fully de-identified internal laboratory submissions. No protected health information (PHI) was accessed or disclosed (Supplement 2).

2.5. Procedures

2.5.1. Element 1: Evaluation of the System’s Ability to Distinguish and Recall Trained from New Single Data Points

Element 1 involved a retrospective observational approach to evaluate the system’s ability to generate accurate recommendations based on individual data points, using multiple validation techniques. New data, also known as untrained data, refers to information the system has not previously encountered. Recognizing untrained data and subsequently training the system to identify it in future encounters are critical. Data imputed into the system was sourced from FDA-approved molecular diagnostic BioFire panels published on the BioFire website (<https://www.biofire.com/products/the-filmarray-panels/>). Panels do not contain any patient information and contain data regarding the type of panel, source of specimen, organism targets, and resistance gene targets. This was to ensure standardized nomenclature of panel types, organisms, and resistance genes. The new data encompassed six different standardized infectious disease panels (Table 1). The data were selected based on the fact that these panels are among the few FDA-approved comprehensive molecular panels currently available. Additionally, the microbes and resistance markers tested by these panels are considered industry standard. Panel information was uploaded into the system and accessed remotely, from Boca Raton, and repeatedly analyzed on subsequent days.

Table 1. FDA-approved Bio-Fire panels.

Panel Name		Panel Description
Panel 1	Respiratory	Identifies pathogens responsible for respiratory illnesses from sputum or nasopharyngeal samples
Panel 2	Blood	Identifies pathogens and antimicrobial resistance genes from blood samples
Panel 3	Gastrointestinal (GI)	Identifies causes of GI infections from GI specimens
Panel 4	Meningitis	Identifies pathogens that cause meningitis from cerebral spinal fluid (CSF) samples
Panel 5	Pneumonia	Identifies pathogens responsible for pneumonia from sputum, nasopharyngeal samples, or other respiratory samples
Panel 6	Joint	Identifies pathogens that may cause joint infections from synovial fluid samples

To ensure the system recognized the uploaded data as novel and untrained, each data point was enclosed in brackets. The dataset was then collectively input into the system (Table 2), which successfully identified it as entirely new, as bracketed data is never present in the system’s training inputs before. This approach helps prevent overfitting and bias and avoids introducing data already

known to the model. Once the data was established as new and untrained, the data was then categorized according to each panel (Table 2) as designed by the panel manufacturer and entered into the system again. Here, too, the system identified all the variables entered into the system correctly as untrained data. This was expected since training did not occur between training sessions 1 and 2.

Table 2. Summary of processes in element 1.

N data points from Biofire’s six different panels		
N unique data points noted after redundancies across the panels		
Brackets were placed around data points to ensure no overfitting, and they are new to the system.		
Training Session 1: All unique data points were entered as a single set of data. System performance: The System identified all data as new.		
Training Session 2: The data points, then divided into respective panels as per the Biofire manufacturer (respiratory, blood, CNS, joint, etc). System performance: The System identified all data as new within these panels.		
Data was then divided into randomized groups or K-folds.		
Training session 3: Fold 1 was trained and tested against the data in the remaining untrained folds. System performance: Only the data that was form 1 was trained was noted to be trained while the other data remained untrained.		
Training sessions 4 - 7: Training was repeated for 2, 3, 4, and 5 in separate instances. System performance: Only the data that was from a fold that was trained was noted to be trained, while the other data remained untrained.		
Training session 8: Random untrained data was placed within the 5 previously trained folds and tested. System performance: Only the data that was previously trained in the other sessions was noted to be trained. The remaining data was then trained as well.		
Training session 9: All data was entered into the system collectively again, as one set. System performance: The system noted that all data points were trained.		

The panels were then randomized, and the data were split multiple times to form new data sets containing unique random variables (random subsampling). These new data sets were distributed into six groups: five of these groups became the folds for K-fold cross-validation (Table 2), and the sixth data set was used for holdout validation (Table 3). After training on one-fold, the data sets from all six panels were re-entered into the system for analysis (Table 2). Once all the sets were analyzed, all the data were again introduced back into the system in its entirety for evaluation (Table 2).

Table 3. Randomized data set, grouped into sets.

Data Set	# Variables	Description of variables
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K fold-1	21	<i>Staphylococcus aureus</i> , <i>Clostridium perfringens</i> , <i>Cryptosporidium</i> , <i>Varicella zoster virus</i> (VZV), <i>Cryptococcus</i> (<i>C. neoformans</i> / <i>C. gattii</i>), <i>Shigella</i> / <i>Enteroinvasive E. coli</i> (EIEC), <i>Neisseria gonorrhoeae</i> , <i>Vibrio</i> (<i>V. parahaemolyticus</i> / <i>V. vulnificus</i> / <i>V. cholerae</i>), <i>Human metapneumovirus</i> , <i>Klebsiella oxytoca</i> , <i>Enterococcus faecalis</i> , <i>Parainfluenza virus 1</i> , <i>Candida parapsilosis</i> , <i>Klebsiella aerogenes</i> , <i>Enterobacter cloacae</i> complex, <i>Haemophilus influenzae</i> , <i>Adenovirus F40/41</i> , <i>Coronavirus 229E</i> , <i>IMP</i> , <i>mcr-1</i> .
K fold-2	19	<i>Influenza A virus A/H3</i> , <i>Clostridioides</i> (<i>Clostridium</i>) <i>difficile</i> (toxin A/B), <i>Streptococcus agalactiae</i> , <i>Adenovirus</i> , <i>Bordetella pertussis</i> , <i>Candida krusei</i> , <i>Herpes simplex virus 2</i> (HSV-2), <i>Serratia marcescens</i> , <i>Cytomegalovirus</i> (CMV), <i>Parainfluenza virus 2</i> , <i>Moraxella catarrhalis</i> , <i>Staphylococcus lugdunensis</i> , <i>Human herpesvirus 6</i> (HHV-6), <i>Bacteroides fragilis</i> , <i>Campylobacter</i> (<i>C. jejuni</i> / <i>C. coli</i> / <i>C. upsaliensis</i>), <i>Candida albicans</i> , <i>Enterococcus faecalis</i> (<i>EAEC</i>), <i>Coronavirus OC43</i> , <i>OXA-48-like</i>
K fold-3	18	<i>Human rhinovirus/enterovirus</i> , <i>Vibrio cholerae</i> , <i>Mycoplasma pneumoniae</i> , <i>Influenza B virus</i> , <i>Legionella pneumophila</i> , <i>Chlamydia pneumoniae</i> , <i>Candida tropicalis</i> , <i>KPC</i> , <i>Plesiomonas shigelloides</i> , <i>Shiga-like toxin-producing E. coli</i> (STEC) <i>stx1/stx2</i> , <i>Enteropathogenic E. coli</i> (EPEC), <i>Cyclospora cayentanensis</i> , <i>Enterobacteriales</i> , <i>Anaerococcus prevotii/vaginalis</i> , <i>Cutibacterium avidum/granulosum</i> , <i>Parainfluenza virus 3</i> , <i>VIM</i> , <i>NDM</i>
k-fold 4	12	<i>Streptococcus pyogenes</i> , <i>Enterococcus faecium</i> , <i>Influenza A virus A/H1</i> , <i>Rotavirus A</i> , <i>Staphylococcus epidermidis</i> , <i>Human parechovirus</i> (HPeV), <i>Klebsiella pneumoniae</i> group, <i>Neisseria meningitidis</i> , <i>Candida auris</i> , <i>Bordetella parapertussis</i> , <i>Peptostreptococcus anaerobius</i> , <i>Coronavirus NL63</i> .
K fold-5	14	<i>Norovirus GI/GII</i> , <i>Candida glabrata</i> , <i>Escherichia coli</i> , <i>Peptoniphilus</i> , <i>Acinetobacter calcoaceticus-baumannii</i> complex, <i>Streptococcus spp.</i> , <i>Pseudomonas aeruginosa</i> , <i>Escherichia coli</i> K1, <i>Herpes simplex virus 1</i> (HSV-1), <i>E. coli</i> O157, <i>Parainfluenza virus 4</i> , <i>Streptococcus pneumoniae</i> , <i>Coronavirus HKU1</i> , <i>Severe acute respiratory syndrome coronavirus 2</i> (SARS-CoV-2).
Hold-out	27	<i>Influenza A virus A/H1-2009</i> , <i>Influenza A virus</i> , <i>Proteus spp.</i> , <i>Stenotrophomonas maltophilia</i> , <i>Respiratory syncytial virus</i> , <i>Listeria monocytogenes</i> , <i>Staphylococcus spp.</i> , <i>Astrovirus</i> , <i>Sapovirus</i> (I, II, IV, and V), <i>Enterotoxigenic E. coli</i> (ETEC) <i>lt/st</i> , <i>Entamoeba histolytica</i> , <i>Giardia lamblia</i> , <i>Yersinia enterocolitica</i> , <i>Enterovirus</i> (EV), <i>Coronavirus</i> , <i>Citrobacter</i> , <i>Kingella kingae</i> , <i>Morganella morganii</i> , <i>Candida spp.</i> , <i>Finnegoldia magna</i> , <i>Parvimonas micra</i> , <i>CTX-M</i> , <i>mecA/C</i> , <i>vanA/B</i> , <i>ESBL</i> , <i>Klebsiella pneumonia</i> group, <i>Salmonella</i> .

2.5.2. Element 2: Evaluation of the System’s Ability to Distinguish and Recall Trained from New Complex Datasets

An anonymous retrospective analysis was conducted using internal data imputed by various laboratories containing infectious disease results. Data within each lab result includes many variables sent by the lab, such as patient demographics, allergies, organisms, resistance genes, pregnancy status, diagnostic codes, and more. The research team accessed the data remotely from their respective locations. These results were randomly selected by choosing a random 24-hour period (all results from Thursday, August 1, 2024, were used).

Patient-specific information was locked to the researchers and was inaccessible. A team of researchers was tasked with manually reviewing each laboratory result and evaluating the accuracy

of how the system classified it (trained or untrained). The assigned researcher was not involved in the initial training of the data to avoid biases.

Data is considered untrained by the system if either a single variable is new or if a dataset has a new combination of trained data (regardless of whether the individual data points have been trained). Fully trained data sets require not only each data point within the dataset to be trained but also require multiple rounds of HITL training for the entire data set combination. This means that the system must see the same combination of data points multiple times before it is considered trained by the system (Table 4).

Table 4. Definition of status.

	<u>Auto-approve</u>	<u>Auto-match</u>	<u>High confidence</u>	<u>New</u>
	Fully trained data points and data sets	Partially trained data sets with completed trained data points	Untrained data sets with trained data points	Untrained data sets and untrained data points
Required training session	Completed full training of all data points and data sets (at least two data set training sessions)	One data set training session was completed, however at least one more training session is required	≥ 90 percent like previously trained data sets where data points are trained completely	Data sets and points require full training

2.5.3. Element 3: Evaluation of Human in-the-Loop Component and the Accuracy of Clinical Recommendations

A prospective evaluation of recommendations generated by human-enhanced (HITL) models during the training process was conducted. The primary objective was to determine whether errors occurred during the HITL intervention that could have resulted in inaccurate clinical recommendations. To assess this, a team of independent researchers with expertise in infectious diseases reviewed the system’s output. Each recommendation was compared to established clinical guidelines and evaluated using a structured six-question questionnaire:

- Were the microbes being treated as pathogens accurately identified?
- Does the antibiotic recommended in OneChoice have activity against the microbe that is presumed to be the pathogen?
- Was the recommended dose accurate?
- Was the recommended duration of treatment accurate?
- Was the preferred therapy the optimal therapy?
- Were there organisms that should have been addressed but were not?

Based on their responses, reviewers categorized discrepancies in HITL-trained outputs as either major or minor: a) Major discrepancies: Failure to identify a pathogen that required treatment or recommending antibiotics that were ineffective against the identified microbe(s); b) Minor discrepancies: Incorrect antibiotic dosage or treatment duration (outside FDA or guideline-based ranges), a suboptimal choice when a better preferred or alternative therapy was available. This review process aimed to ensure the integrity of HITL-influenced recommendations and identify opportunities for further refinement of the system.

2.6. Data Analysis

Data analysis was performed using the STATA 17 statistical package.

For element 1: evaluating the system’s ability to distinguish between trained vs. untrained data the following metrics were calculated: a) Accuracy: proportion of data points correctly classified as trained or untrained; b) Precision: proportion of instances that were correctly identified as trained data; c) Recall: ability of the trained model to correctly identify previously trained data points; and d) F1 Score: the harmonic meaning of precision and recall

For element 2: evaluating the system’s ability to distinguish between trained vs untrained complex data sets, the following metrics were calculated: a) True Positive Rate (TPR): proportion of fully trained data sets correctly identified; b) True negative rate (TNR): proportion of untrained datasets correctly identified; c) False positive rate (FPR): proportion of untrained datasets incorrectly classified as trained; And d) False negative rate (FNR): proportion of fully trained datasets incorrectly classified as untrained.

For element 3: evaluating the accuracy of HITL in in providing accurate clinical recommendations, the percentage of reports with major and minor discrepancies was calculated. All statistical tests were performed at a significance level of $p < 0.05$.

3. Results

3.1. Element 1: Evaluation of the System’s Ability to Distinguish and Recall Trained from New Single Data Points

From the six panels (Table 1), there were initially 192 data points. However, after removing duplicate variables that spanned across multiple panels, 111 unique variables remained. Each panel was fully input into the system. Since this was novel data, the system correctly identified all these variables as new (Table 2).

The proportion of true positive results (correctly identified as trained) to the total predicted positives (all instances predicted as trained) is a measure of precision. (Table 5). The proportion of true positives to the total actual positives (all actual trained data points) is a measure of recall. This is collectively assessed using the F1 score.

Table 5. Proportion of true positive results to total expected positives.

Folds	True positives (identified trained data)	True negative (Identified new data)	False positive (Identified new data as trained data)	False negatives (Identified trained data as new data)
Fold 1	21	21	0	0
Fold 2	19	19	0	0
Fold 3	18	18	0	0
Fold 4	12	12	0	0
Fold 5	14	14	0	0
Hold- out	27	27	0	0
Total	111	111	0	0

TP: True positives, TN: true negatives, FP: false positives, FN: false negatives.

The proportion of true positive results (i.e., correctly identified as trained) relative to the total number of instances predicted as trained represents the system’s precision (Table 6). Similarly, the ratio of true positives to all actual trained data points reflects the system’s recall. Together, these

metrics demonstrate the system’s flawless performance in accurately distinguishing between trained and untrained data under the evaluated conditions.

Table 6. Performance Metric based on Confusion Matrix Results.

Metric	Formula	Result
Precision	$TP / TP + FP$	$111 / (111 + 0) = 1.00 (100\%)$
Recall (Sensitivity)	$TP / TP + FN$	$111 / (111 + 0) = 1.00 (100\%)$
F1 score	$2 \times \text{precision} \times \text{Recall} / (\text{Precision} + \text{Recall})$	$2 \times 1 \times 1 / (1 + 1) = 1.00 (100\%)$
Positive Predictive Value	$TP / (TP + FP)$	$111 / (111 + 0) = 1.00 (100\%)$
Negative Predictive Value	$TN / (TN + FN)$	$111 / (111 + 0) = 1.00 (100\%)$

TP: true positives, TN: true negatives, FP: false positives, FN: false negatives.

3.2. Element 2: Evaluation of the System’s Ability to Distinguish and Recall Trained from New Complex Data Sets

A total of 1,401 real laboratory results were analyzed from the systems database ([Supplement 2](#)). These results were randomly selected by choosing a 24-hour period at random. The dataset was diverse, encompassing results from 66 laboratories located in 55 different regions across 24 states and one international site. This data included: 176 specified provider locations (12%), 936 unspecified practice locations (62%), 198 specified facility locations (13%), and 200 non specified facility locations (13%). A total of 65% of the clinical samples analyzed were urine specimens, representing the most frequent sample type. This was followed by wound (13%), respiratory (8%), and vaginal samples (5%). Other less frequent sources included rectal (3%), throat (2%), nail (2%), oral (1%), urogenital, epidermal, and unknown sources (all <1%) (Figure 2).

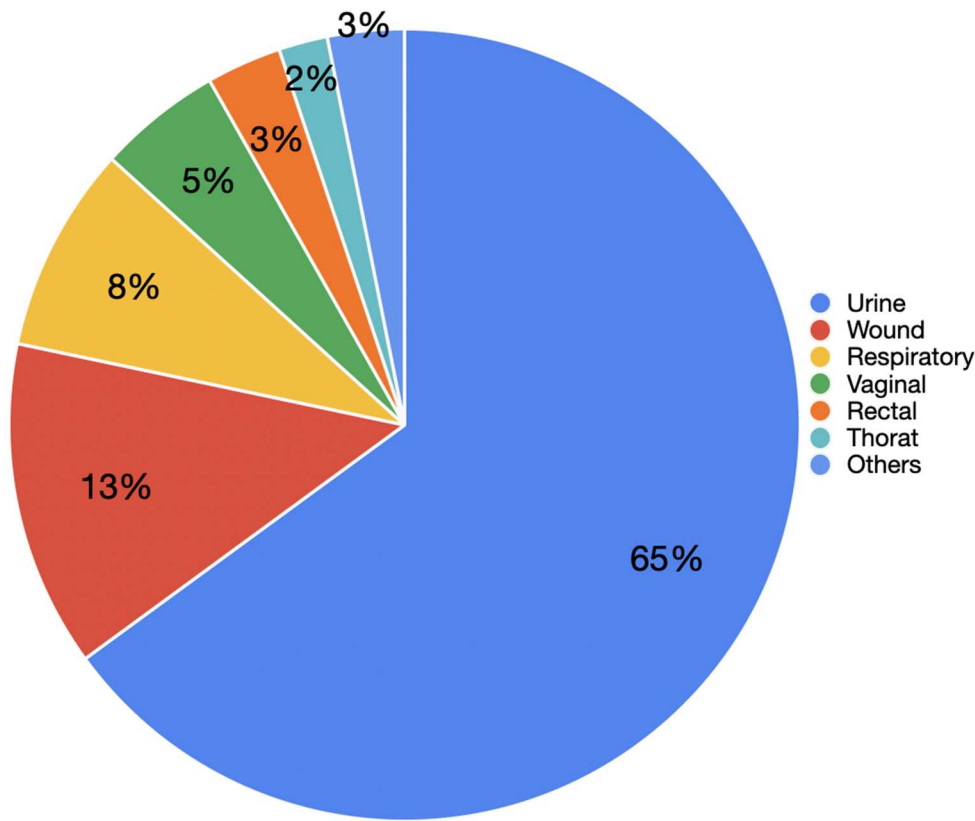


Figure 2. Types of samples submitted for analysis.

Among the 1,401 results analyzed, 238 (16.98%) were confirmed as negative results and 1163 (83.02%) as positive. Of the positive results, 519 (44.62%) were identified by the system as fully trained, while 644 (55.38%) reports required training (untrained). This classification was verified through manual review, verifying the system’s ability to accurately distinguish between trained and untrained datasets. The system’s classification performance was evaluated using the full set of 1,401 reports. Using 238 true negatives and 519 true positives, the F1 Score and related performance metrics were calculated as follows (Table 7).

Table 7. Classification performance of the system on 1401 diagnostic reports.

Classification Outcome	Count	Description
True Positives (TP)	519	Fully trained reports correctly identified as trained
True Negatives (TN)	238	Negative reports correctly identified as trained
False Positives (FP)	0	No untrained reports were incorrectly identified as trained
False Negatives (FN)	0	No trained reports were misclassified as untrained
Untrained (correctly flagged)	644	Reports requiring training correctly identified as untrained
Total reports	1401	

Performance Metrics:

- **Precision:** $TP / (TP + FP) = 519 / (519 + 0) = 1.0$ (100%)
- **Recall:** $TP / (TP + FN) = 519 / (519 + 0) = 1.0$ (100%)

These results indicate that the system achieved perfect accuracy in distinguishing between fully trained and untrained datasets under real-world conditions.

3.3. Element 3. Evaluation of the HITL Component in the Accuracy of Clinical Recommendations

Among the 1,401 results analyzed, 238 (16.98%) were confirmed as negative and 1,163 (83.02%) as positive. Of the positive results, 519 (44.62%) corresponded to fully trained data (Table 11). Additionally, 233 (20.03%) had been trained once but required further reinforcement. Specifically, 164 cases (14.10%) required one additional training session, 61 (5.24%) required two sessions, 7 (0.60%) required three, and 1 case (0.09%) required four sessions. A total of 267 cases (22.95%) were classified as partially untrained, defined as datasets with greater than 90% similarity to data previously seen by the system. Among these, 186 (15.99%) required two training sessions, 41 (3.52%) required three, 19 (1.63%) required four, and five cases each (0.43%) required five or six sessions, respectively. In contrast, 97 cases (8.34%) were classified as completely untrained, having less than 90% similarity to any known dataset. These required more intensive training: 63 cases (5.42%) required two sessions, 15 (1.29%) required three, 12 (1.03%) required four, four (0.34%) required five, two (0.17%) required six, and one case (0.09%) required seven sessions. (Table 8)

Table 8. Number of Steps with Human Intervention Required Before Fully Trained.

n	%	N of Training
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Variable		1	2	3	4	5	6	7
Total	1401							
Negative	238							
Positive	1163							
Complete trained data	519	44.63						
Trained a single time but required additional training	233	20.03	164	61	7	1		
Partially untrained data	267	22.96		186	41	19	5	5
Completely untrained data	97	8.34		97	63	15	12	4

Of the 644 reports that required training, all were reviewed by a clinical team with expertise in infectious disease. According to the consensus of the reviewers, no major discrepancies were identified. Minor discrepancies were observed in 100 (15.52%) of the 644 reports. Specifically, 11 (1.71%) reports involved the system recommending a different antibiotic than what was typically preferred; 36 (5.59%) reports suggested that an alternative antibiotic or combination could have been considered; 34 (5.28%) reports included recommendations where the dose or administration interval lacked formal FDA approval or varied across clinical references; and 20 (3.11%) reports did not address organisms of questionable pathogenicity (Table 9). Overall, these findings indicate a high level of consistency between the system’s recommendations and accepted clinical standards, with only minor variations that reflect the complexity and nuance of clinical decision-making.

Table 9. Frequency of Major Discrepancies and Minor Discrepancies.

Discrepancy	Frequency	%
Mayor discrepancy		
NO discrepancy	644	100
A known pathogen has NOT been addressed	0	0
The recommended antibiotic has NO activity against the microbe detected	0	0
Minor discrepancy		
NO discrepancy	544	84.5
An alternative to OneChoice could have been recommended	11	1.7
Among the alternatives recommendations another antibiotic or a combination of antibiotics could have been recommended	35	5.4
Dosing and length of therapy are not consistent with the FDA guidelines or other literature	34	5.3
Microbes that should have been targeted were NOT addressed	20	3.1
Total	100	15.5

4. Discussion

This study provides a robust assessment of the internal validation of this novel ML system, demonstrating its ability to accurately differentiate between trained and untrained data, both at the level of individual data points and in complex data sets. To our knowledge, this is the only system currently deploying such validation techniques.

It is important to note that once a data point is trained on the system, it remains trained even if it appears in larger data sets. This eliminates the need to retrain the system on the same data point in subsequent data sets. Furthermore, unlike traditional K-fold cross-validation, where training occurs on all but one fold, in this study, the data was trained on a single fold and tested against the other untrained folds. This approach was necessary due to the large volume of data and the nature of the system: once a data point is trained, it cannot be untrained. This method was more efficient and effective in terms of time, allowing for more training sessions within the constraints of the system.

The system shows high accuracy in recognizing new data by distinguishing untrained from trained individual data points using multiple validation techniques (K-fold cross-validation, random subsampling, and holdout validation). As well as accurately classifying complex data sets, with over 1000 real-world laboratory data from diverse sources, which highlights its ability to generalize and avoid overfitting to the training data. Also, the diversity of real-world data with data from 66 laboratories in 55 regions, across multiple states, and one international location ensures the system's adaptability to diverse clinical settings and reduces the possibility of bias [19].

The use of multiple validation techniques (K-fold cross-validation, random subsampling, and holdout validation) in the study mitigates the risks of overfitting and improves the generalizability of the model [7,20]. In the system studied, the HITL component for multi-stage data training enables the participation of clinical experts, prevents error propagation, and ensures compliance with current medical guidelines [4]. This is a key strength, particularly in the context of high-stakes clinical decision-making [21,22].

The critical importance of HITL validation is highlighted by the detection of 100 minor discrepancies out of 644 reviewed reports. This finding reinforces that, although ML algorithms are powerful tools capable of processing large volumes of data and identifying patterns, the nuanced clinical judgment of infectious disease specialists remains essential to ensure patient safety and optimize treatment strategies [3,23]. The nature of these discrepancies is particularly telling: while the automated system excels at data analysis, it lacks the ability to account for complex clinical subtleties and evolving scientific insights. For instance, the system relies on external references, such as FDA guidelines or published literature, for dosing and administration intervals. Although generally accurate, these references may not always reflect the most current evidence or account for patient-specific factors, which clinicians are uniquely positioned to evaluate. Additionally, the system provides generalized recommendations and is not designed to handle rare or atypical clinical scenarios, further emphasizing the necessity of expert oversight. These results suggest that the HITL process successfully mitigated potential errors or suboptimal recommendations, underscoring the vital role of human oversight in AI-supported clinical care [22,24]. This is especially significant in the realm of antimicrobial stewardship, where inappropriate antibiotic prescribing can contribute to the growing threat of antimicrobial resistance [22,25].

The methodology and findings of the study can be contrasted with previous attempts to validate ML-based CDSS; For example, IBM Watson for Oncology, while initially promising, failed due to inaccurate recommendations and integration challenges, ultimately leading to its discontinuation [26,27]. This highlights the crucial need for robust validation processes, as demonstrated in the present study. Mayo Clinic Predictive Analytics: This project faced data complexity and integration issues, which halted its progress [23]. The Pediatric Alert System employed cross-validation techniques, however, this system suffered from limitations due to data sparsity, which impacted the effectiveness of its model [28]. The use of large, diverse datasets and HITL helps overcome this.

This study demonstrates a robust validation process, which emphasizes real-time application and human oversight. It also highlights the advantage of having a system that is not intended to be used for “discovery” of new findings, but instead focuses on adhering to existing guidelines and best practices. The system was carefully trained by infectious disease experts and periodically re-evaluated. This method ensures both the accuracy and up-to-date nature of the database and algorithms. The system was designed to assume that error is possible and, therefore, never allows data to be trained indefinitely.

Limitations of this study include that data curation, particularly for complex data sets, was restricted to a single-day period. Although the sample was geographically diverse, limiting the sample to a single day could introduce some bias into the data sample, possibly related to the capabilities of the quality control team on that particular day. Furthermore, while HITL is essential, the manual review process introduces the potential for bias and variability in data interpretation [7,29]. Automating some components of the HITL process could improve consistency. Lastly, this study does not evaluate the subsequent impact on patient outcomes and antimicrobial stewardship.

The system’s unique approach to antimicrobial guidance is an innovative system, notable for its reliance on established treatment guidelines and avoidance of novel treatment suggestions, thereby prioritizing safety and adherence to proven practices. To meet these goals, this first study demonstrates that the first two steps of the process work robustly, laying the groundwork to demonstrate that the system works efficiently, and if subsequent steps follow suit, will lead to appropriate and useful recommendations in the real world. This is a preliminary study that focuses primarily on the technical performance of the system in identifying trained versus untrained data, and final results and clinical outcomes are not yet included. Therefore, the impact of the system on patient care and antimicrobial stewardship requires further study.

This study provides valuable insights into the design and validation of ML-based CDSS for antimicrobial stewardship. The combination of robust statistical validation techniques with a comprehensive HITL process represents a promising strategy for ensuring the accuracy and clinical relevance of AI-driven recommendations. The findings suggest that such systems can potentially contribute to improved antimicrobial prescribing practices and, ultimately, better patient outcomes.

Future research should focus on external validation and evaluating the performance of the model in diverse clinical settings and patient populations to assess its generalizability and identify potential biases [20,31]. In addition, prospective clinical trials through randomized controlled trials to evaluate the impact of the CDSS on antimicrobial use, patient outcomes, and healthcare costs may be of benefit [4,22]. Lastly, expanding HITL evaluation by collecting more detailed data on the types of discrepancies identified during the HITL process to further refine the model and identify areas where clinician training may be needed. Integrating with Electronic Health Records (EHRs) may facilitate seamless data flow and improve clinician workflow [22,26,30].

By continuing to refine and validate these systems, we can harness the power of AI to support clinicians in making informed decisions about antimicrobial therapy, contributing to the fight against antimicrobial resistance, and improving patient care.

5. Conclusions

This study demonstrates the robust performance of a novel ML model in accurately distinguishing between trained and novel data, achieving 100% accuracy. This result was validated using multiple methodologies, including K-fold cross-validation, random subsampling, and holdout validation. The system also successfully identified complex datasets as either trained or untrained with high reliability. Integrating HITL validation enhanced the model’s adaptability and quality control, reinforcing the value of clinical oversight in AI systems. These findings suggest that advanced ML models can be effectively and consistently trained to support clinical decision support systems (CDSS), helping to bridge key gaps in antimicrobial stewardship. While the system showed excellent performance in identifying data patterns and learning behavior, its clinical decision-making

capabilities also demonstrated strong internal validity. Specifically, it achieved 100% accuracy in avoiding major discrepancies and 84% agreement in minor discrepancies when evaluated against clinical standards. This high level of internal validation represents a promising first step toward broader implementation. Future studies should focus on external validation across diverse healthcare settings and evaluate the impact of these AI-driven recommendations on real-world clinical outcomes and patient care.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. **Supplement 1, Supplement 2.**

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Ethical Considerations and Data Availability: All data used in this study were either publicly available (BioFire panels) or obtained from internal, de-identified laboratory submissions. No protected health information (PHI) was accessed or disclosed. However, the study was submitted to and approved by an Ethics Committee, and all data were managed per HIPAA guidelines. The processed database will be attached to the paper.

Data Availability Statement: The data analyzed in this manuscript, as well as its definitions, can be downloaded at the following link: **Supplement 2.**

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Conflicts of Interest: Ari Frenkel is Co-founder and Chief Science Officer of Arkstone Medical Solutions, the company that produces the OneChoice report evaluated in this study. JC Gómez de la Torre works as the Director of Molecular Informatics at Arkstone Medical Solutions and as Medical Director at Roe Lab in Lima, Perú, while Alicia Rendon and Miguel Hueda Zavaleta serve as Quality Assurance Managers at Arkstone Medical Solutions. These affiliations may be perceived as potential conflicts of interest. However, the design of the study, data collection, analysis, interpretation, manuscript preparation, and the decision to publish the results were conducted independently, with no undue influence from the authors’ affiliations or roles within the company.

Abbreviations

ML	Machine learning
CDSS	Clinical decision support systems
HITL	Human involvement in the loop
AI	Artificial intelligence
ASP	Antimicrobial stewardship program
AMR	Antimicrobial resistance

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