

Review

Not peer-reviewed version

Exosomal Non-coding RNAs as Potential Biomarkers for Alzheimer's Disease: Advances and Perspectives in Translational Research

<u>Simoneide Souza Titze-de-Almeida</u>*, <u>Clara Luna Marina</u>, Milena Vieira Ramos, Letícia Dias dos Santos Silva, <u>Pedro Renato de Paula Brandão</u>, <u>Diógenes Diego de Carvalho Bispo</u>, <u>Felipe Von Glehn</u>, <u>Ricardo Titze-de-Almeida</u>

Posted Date: 15 April 2025

doi: 10.20944/preprints202504.1201.v1

Keywords: Alzheimer's disease; biomarkers; β-amyloid; Tau protein; miRNAs; circRNAs



Preprints.org is a free multidisciplinary platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This open access article is published under a Creative Commons CC BY 4.0 license, which permit the free download, distribution, and reuse, provided that the author and preprint are cited in any reuse.

Disclaimer/Publisher's Note: The statements, opinions, and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions, or products referred to in the content.

Review

Exosomal Non-Coding RNAs as Potential Biomarkers for Alzheimer's Disease: Advances and Perspectives in Translational Research

Simoneide Souza Titze-de-Almeida ^{1,*}, Clara Luna Marina ¹, Milena Vieira Ramos ², Letícia Dias dos Santos Silva ¹, Pedro Renato de Paula Brandão ^{3,4}, Diógenes Diego de Carvalho Bispo ⁴, Felipe Von Glehn ⁴ and Ricardo Titze-de-Almeida ¹

- ¹ University of Brasília, Research Center for Major Themes Neurodegenerative Disorders Division, Brasília, Federal District, Brazil 1
- ² University of Rio Verde Faculdade de Medicina, Goiás, Brazil
- ³ Sírio-Libanês Hospital, Brasilia, DF, Brazil
- ⁴ University of Brasília, Brasília University Hospital, Federal District, Brazil
- * Correspondence: simoneide.silva@gmail.com

Abstract: Alzheimer's disease (AD) is a progressive neurodegenerative disorder primarily characterized by memory loss and cognitive decline, which significantly impacts patients' quality of life and imposes substantial emotional, practical, and economic burdens on their families. As the most common cause of senile dementia, AD currently affects approximately 50 million people worldwide, with projections indicating a threefold increase by 2050 due to rising life expectancy and an aging global population. Diagnosis of AD remains challenging. Neuroimaging techniques reveal atrophy in critical brain regions, particularly in the cortex, hippocampus, and limbic system, which are essential substrates for memory, personality changes, and other cognitive functions. The hallmark molecular changes associated with AD include the accumulation of β -amyloid plaques and the formation of tau protein tangles. Several underlying mechanisms contribute to neuron loss, such as oxidative stress, neuroinflammation, microbial dysbiosis and insulin resistance. In this context, exosomes - small extracellular vesicles that facilitate cell communication - transport proteins, DNA, mRNA, and non-coding RNA (ncRNA), all of which play a significant role in the neurobiology of AD. Furthermore, emerging research indicates that exosomal ncRNAs may serve as promising biomarkers for AD, offering the possibility of improved diagnostic precision. This review explores the potential of exossomal ncRNAs as non-invasive biomarkers for AD, focusing on recent advances and future directions in translational studies.

Keywords: Alzheimer's disease; biomarkers; β-amyloid; Tau protein; miRNAs; circRNAs

1. Introduction

Dementia has increasingly become a focal point in scientific research and public health initiatives. This attention is driven by the condition's rising prevalence among the aging global population, as well as its profound impact on both affected individuals and their families. Efforts are being made in the realms of prevention, early diagnosis, and clinical intervention to address this growing concern [1]. The increase in life expectancy has contributed to a rise in cases of dementia as well as other neurodegenerative disorders among the elderly. Currently, approximately 50 million individuals worldwide are diagnosed with dementia, a figure projected to triple by 2050 [2]. Alzheimer's disease is the most common form of dementia globally. This chronic neurological disease was first described by Alois Alzheimer in 1906 and is primarily characterized by progressive memory loss and impairment across multiple cognitive functions [3]. Despite considerable progress in

understanding the brain pathology associated with AD, diagnosing and treating this complex disorder remains challenging [1,4].

The incidence of AD is influenced by non-modifiable factors such as advancing age, female sex, lower educational attainment, and genetic predisposition, particularly the presence of the APOE $\epsilon 4$ allele, while $\epsilon 2$ offers some protection. Modifiable factors include cardiovascular conditions (hypertension, diabetes, hypercholesterolemia), lifestyle elements (Mediterranean diet, physical activity, cognitive engagement), and environmental exposures (e.g., air pollution). Depression, obesity, subjective cognitive decline, neurodegenerative markers on imaging, and co-pathologies also contribute to AD risk [5–13].

The pathological features of AD encompass the accumulation of β -amyloid (A β) plaques and neurofibrillary tangles (NFTs) composed of tau protein within the brain. Additional changes of note include an increase in reactive oxygen species (ROS), the proliferation of glial cells, impaired insulin sensitivity, and modifications in the microbiome [14]. These characteristics have been extensively studied due to their critical importance for both diagnostic and potential therapeutic applications [15].

Exosomes are extracellular vesicles (EVs) that have gained significant prominence in AD research in recent years. These small lipid vesicles carry a variety of molecules, including proteins, DNA, mRNA, and non-coding RNA (ncRNA) from their cells of origin. Notably, ncRNAs are regulators of gene expression and can influence multiple signaling pathways [16–18]. AD-related exosomes have been shown to disseminate amyloidogenic peptides, suggesting their involvement in the underlying mechanisms of brain pathology [19].

Research has also investigated blood-derived exosomes as potential biomarkers for AD, focusing on levels of A β , amyloid precursor protein (APP) fragments, tau, and phosphorylated tau (p-tau) [16,17,20]. Furthermore, exosomes isolated from AD patients exhibit aberrant contents of synaptic proteins, inflammatory mediators, growth factors, and lysosomal proteins [20–23]

Exosomal non-coding RNAs (ncRNAs), particularly microRNAs (miRNAs), have emerged as key regulators of APP and tau proteins, both of which are critical in AD pathology [24]. In addition to miRNAs, exosomes transport a variety of other ncRNAs, including long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs), which facilitate cell-to-cell communication and reciprocal influence [25,26]. The expression patterns of these exosomal ncRNAs can vary depending on cell type and disease state, suggesting their potential involvement in AD and other pathologies [27,28]. Recent studies have further clarified the roles of exosomal mRNAs, circRNAs, and miRNAs in AD progression, particularly their regulatory effects on APP and tau proteins [24,29,30]

Regarding the diagnosis of AD, it is crucial to develop simple and effective tests with high specificity to improve diagnostic [15,31,32]. Exosomes represent a promising source of accessible and cost-effective biomarkers for disease monitoring, as their lipid bilayer protects the cargo from enzymatic degradation in the bloodstream [33,34]. This review examines the potential of ncRNAs derived from exosomes as viable biomarkers for AD.

Clinical Diagnosis of AD

Alzheimer's Disease is clinically defined as a major or mild neurocognitive disorder (NCD), as outlined in the Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (DSM-5). The DSM-5 emphasizes a decline in cognitive functioning that is not attributable to delirium or another mental disorder. Key criteria include significant cognitive decline in one or more cognitive domains (e.g., executive function, learning and memory, complex attention, language, social cognition, or perceptual-motor skills) based on concerns from the individual, informants, or clinicians, and preferably documented through standardized neuropsychological testing. For major NCD, these deficits must interfere with independence in daily activities, while for mild NCD, independence remains intact but may require compensatory strategies. A diagnosis of probable AD requires genetic evidence or specific patterns of cognitive decline and progression, whereas possible AD is diagnosed in the absence of such evidence but with clinical support [35–37].

Associated with this clinical presentation, AD is characterized neuropathologically by the presence of amyloid plaques and NFTs, alongside other pathological changes such as inflammatory responses, astrocyte and microglial activation, and synaptic and neuronal loss. Amyloid plaques are extracellular deposits of A β peptides that follow a predictable regional progression, starting in the neocortex and advancing to other brain areas, as outlined in Thal phases [38–42] . NFTs, on the other hand, consist of intracellular aggregates of p-tau, which progress in a stereotyped pattern from the transentorhinal region to the limbic system and eventually the neocortex, as described in Braak stages [43]. The National Institute on Aging-Alzheimer's Association (NIA-AA) guidelines recommend a comprehensive neuropathological assessment using the ABC scoring system, which integrates Thal phases for amyloid plaques, Braak stages for NFTs, and the Consortium to Establish a Registry for Alzheimer's Disease (CERAD) criteria for neuritic plaque density. This approach allows for a detailed classification of AD-related changes, while also accounting for co-pathologies, such as cerebral amyloid angiopathy and other proteinopathies, that may influence clinical presentation and disease progression [38–42].

Historically, the diagnosis of AD relied primarily on clinical assessment, making accurate diagnosis both challenging and time-consuming. Physicians needed to exclude other potential causes of dementia through a rigorous, multi-step process. This typically involved a comprehensive review of medical history, physical examinations, laboratory tests to exclude other conditions (e.g., vitamin B-12, syphilis serology and thyroid hormone levels), imaging studies (e.g., CT scans), and the discontinuation of any medications that might impair cognition. Only after systematically ruling out other causes of cognitive decline could a probable AD diagnosis be established, with confirmation gained by observing disease progression and specific cognitive patterns associated with AD [44]. When clinical findings are inconclusive, re-evaluation is often necessary [45] Moreover, early diagnosis of AD is frequently impeded by social and emotional barriers, including denial, fear of the diagnosis, limited family support, and difficulties in accessing healthcare [46].

Neuropathological studies in individuals diagnosed with AD during their lifetime have significantly advanced our understanding of the disease's complexity and heterogeneity. While the hallmark pathological features—amyloid plaques composed Aβ deposits and NFTs of p-tau proteins—are central to AD diagnosis, not all individuals clinically diagnosed with AD exhibit these findings. Approximately 14% of patients with mild-to-moderate AD lack sufficient neuropathological changes to meet classical diagnostic thresholds, as highlighted in postmortem brain studies. Among clinically diagnosed AD cases, 81% show high or intermediate levels of typical AD pathology, yet only 41% of dementia cases are attributed exclusively to AD pathology. Polymorbidity is common, with frequent co-occurrence of vascular lesions, TDP-43 proteinopathy, and Lewy bodies, which can complicate clinical presentations. This heterogeneity is also reflected in atrophy-based subtypes, such as limbic-predominant and hippocampal-sparing patterns, which correlate with variations in amyloid and tau pathology. The predictable progression of AD pathology described in Thal and Braak staging is often accompanied by comorbidities that may modify disease trajectories and explain variability in treatment outcomes. These findings underscore the importance of integrating neuropathological, clinical, and imaging data to provide a comprehensive understanding of AD and related dementias [41,47–50].

The diagnosis of AD has undergone significant evolution, with recent developments highlighting the integration of biological and clinical criteria. In 2024, the Alzheimer's Association (AA) workgroup, led by Clifford Jack, proposed revised criteria emphasizing the biological definition of AD, centered on the ATN model, which incorporates biomarkers for A β , tau, and neurodegeneration. These criteria distinguish between Core 1 biomarkers - including amyloid PET, CSF measures (specifically CSF A β 42/40, CSF p-tau181/A β 42, CSF t-tau/A β 42), and plasma markers like phosphorylated tau 217 (requiring at least 90% accuracy for diagnostic validity) - and Core 2 biomarkers, comprising tau PET and specific fluid T2 biomarkers such as MTBR-tau243 and other phosphorylated tau forms [51] Core 1 biomarkers define the earliest *in vivo* detectable stage of AD and can identify disease presence in both symptomatic and asymptomatic individuals. In contrast,

Core 2 biomarkers become abnormal later in disease evolution and are more closely linked to symptom onset, serving primarily for biological staging and prognosis rather than diagnosis. Under this framework, an abnormal Core 1 biomarker is sufficient for an AD diagnosis, even in the absence of clinical symptoms.

However, this approach has sparked debate in the scientific community. The conceptualization of AD as a clinical-biological construct has been a cornerstone of the International Working Group's (IWG) approach, emphasizing the necessity of integrating specific clinical phenotypes with pathophysiological biomarkers. According to the IWG, the diagnosis of AD should rest on the presence of distinct clinical syndromes—such as the amnestic variant or other characteristic phenotypes—supported by positive biomarkers, ensuring that these biological findings are interpreted within a clinical context. This approach contrasts with the AA framework, which allows for the diagnosis of AD in cognitively normal individuals based solely on biomarker positivity [52].

A major point of contention lies in the classification of cognitively normal individuals with positive biomarkers. The AA criteria consider these individuals as having AD, while the IWG advocates for alternative terminology, categorizing them as "asymptomatic at risk for AD" or, in specific scenarios with deterministic biomarker profiles, as having "presymptomatic AD." [52,53]. The IWG highlights the potential risks of labeling cognitively normal individuals as having AD based solely on biomarkers, including psychological distress, social consequences, and the possibility of overdiagnosis. Moreover, they emphasize that many biomarker-positive individuals may never develop clinical symptoms, underscoring the importance of distinguishing risk states from active disease to avoid unnecessary interventions and anxiety [52–54].

Both frameworks acknowledge the importance of staging disease progression and recognize factors such as cognitive reserve and comorbidities in shaping clinical outcomes. However, differences persist in how these elements are applied in clinical practice. The growing availability of blood-based biomarkers and new therapeutics targeting the core pathology of AD has heightened the urgency of these discussions, particularly regarding the implications for treatment eligibility and healthcare accessibility. These evolving diagnostic frameworks underscore broader challenges in the field, including the standardization of biomarker testing, the need to account for population diversity, and the ethical considerations surrounding early detection. As AD therapeutics and research continue to advance, these criteria will likely be refined, reflecting the dynamic interplay between biological innovations and clinical application [51].

As an example of the clinical application of this recent knowledge, the Brazilian Academy of Neurology (BAN) underscores a clinically grounded approach to the use of AD biomarkers, tailored to the realities of the healthcare system. According to BAN, biomarker testing should be reserved for symptomatic individuals with objectively verified cognitive deficits and must be conducted under the guidance of qualified specialists in Geriatrics, Neurology, or Psychiatry. The Academy advises against the use of biomarkers in asymptomatic individuals or those with subjective cognitive decline, emphasizing that their main clinical usefulness lies in supporting diagnosis for symptomatic cases [55]. This is also particularly relevant for early-onset dementia, rapidly progressive dementias, atypical clinical presentations, or when disease-modifying treatments are being considered.

BAN supports a structured diagnostic algorithm that prioritizes CSF biomarker analysis over amyloid PET, highlighting the greater accessibility and cost-effectiveness of CSF testing in Brazil. BAN also recognizes the potential of plasma biomarkers, provided they undergo rigorous validation to be effectively integrated into clinical practice. It emphasizes that biomarker results must be interpreted within the broader clinical context, as positive AD biomarker findings do not definitively attribute symptoms to AD pathology alone, given the high prevalence of comorbid neuropathologies in older adults [55] .

Progress in Diagnostic Technologies for AD

Significant advancements in understanding the natural history of AD over the past two decades have been driven by technological breakthroughs, including the development of molecular markers

for neuroimaging, biochemical analyses of biological fluids, and the implementation of bioinformatics and machine learning [56,57].

The most extensively studied biomarkers for AD are A β and tau proteins (Table 1). Key amyloid biomarkers include decreased A β 42/A β 40 ratio in CSF, as increased amyloid deposition in brain parenchyma detected in PET imaging. Additionally, elevated levels of p-tau in CSF and tau deposition in brain parenchyma in PET scans serve as effective indicators for AD pathology[58,59]. While the A β 42/40 ratio is a validated CSF biomarker [60], it poses challenges for blood-based testing due to detection limitations [61,62]. In contrast, recent p-tau blood assays demonstrate higher diagnostic accuracy and disease specificity, making them promising candidates for routine clinical testing [63]. The p-tau217 has been demonstrated to be a superior marker of cognitive progression to dementia in patients with mild cognitive impairment (MCI) compared to p-tau181 [64]. Conversely, p-tau205 showed changes at a later stage and was more strongly associated with tau PET imaging than with amyloid PET imaging. Similar to total tau (t-tau), increased concentrations of neurofilament light chain (NfL) are not specific to AD and can also be elevated in other neurodegenerative disorders, such as amyotrophic lateral sclerosis (ALS) and frontotemporal dementia (FTD).

In a cohort of Chinese participants, changes in CSF biomarker concentrations were observed over the 20 years preceding clinical diagnosis of sporadic AD, with specific timelines as follows: A β 42 levels changed 18 years prior, A β 42/A β 40 ratio 14 years prior, p-tau181 11 years prior, t-tau 10 years prior, and NfL 9 years prior to diagnosis. As cognitive impairment progressed, the alterations in CSF biomarker levels in the AD group initially accelerated before subsequently slowing down [65].

Table 1. Overview of diagnostic methodologies and biomarkers for AD.

Methodology	Biomarker	Advantages	Limitations	Reference
Immunoassay from Biological Fluids				
CSF	Aβ42, t-tau and p-tau, NfL	Widely used in several systematic studies	Requires the presence of cognitive decline that impacts daily activities for the diagnosis of AD	[5]
Plasma	Aβ42 and Aβ40, p-tau181, p-tau217, p-tau231, NfL, GFAP	High diagnostic accuracy for identifying AD compared to standard clinical evaluation	More research is needed	[66,67]
Serum	Aβ, tau, NfL, miRNA	Ability to accurately diagnose early stages of AD and identify individuals at high risk of cognitive decline in older adults	Diagnosis is based on clinical and pathological criteria	[68]
Neuroimaging				
СТ	Brain atrophy and vascular changes	This method also identifies other causes of neurodegeneration	While offering ease of access, there may be less accuracy in the results	[69]
PET	Cerebral metabolism involving glycolysis	Identifies lower glycolysis uptake in medial temporal regions and the cingulum	High cost	[70]

Methodology	Biomarker	Advantages	Limitations	Reference
	or Aβ protein deposition			
MRI	Cerebral atrophy and ventricular dilation	Better visualization of atrophy in the entorhinal cortex, middle temporal lobe, and hippocampus	High cost	[69]
MRS	Measures different metabolites: NAA, mI, Chol, Glu, Gln, and GABA	Provides metabolic information that may aid in understanding AD	Difficult access and high cost of use	[71]
DTI	Description of white matter microstructure through its tensor model	Identifies potential biomarkers in the early stages of AD	Limitations regarding interpretation	[72]
FW	Isolates and quantifies changes in extracellular water	Detects subtle changes in brain tissue that may indicate early stages of DA	Introduces an additional layer of complexity in the analysis and interpretation of data	[73]

Abbreviations: NfL, Neurofilament light chain; GFAP, Glial fibrillary acidic protein; miRNA, MicroRNA; Ccl2, Chemokine; MRI, magnetic resonance imaging; MRS, magnetic resonance spectroscopy; DTI, diffusion tensor imaging; FW, free-water imaging; NAA, N-acetylaspartate; mI, myo-inositol; Chol, cholesterol; Glu, glutamate; Gln, glutamine, and GABA, gamma-aminobutyric acid.

Modern molecular techniques enable the detection of A β 40 and A β 42 in both CSF and blood samples, with blood testing offering a less invasive alternative. Commercially available blood immunoassays for p-tau217 have demonstrated comparable accuracy to CSF biomarkers for identifying AD [67]. In a cohort study involving 786 individuals, Ashton et al. (2024) found that the p-tau217 immunoassay exhibited accuracy similar to that of cerebrospinal fluid biomarkers in identifying A β and tau pathologies. Plasma p-tau217 results from the ALZpath pTau217 assay were compared with magnetic resonance imaging, β -amyloid PET, tau PET, and CSF biomarkers (A β 42/40 and p-tau immunoassays). The method was able to detect the A β protein across three distinct ranges in three patient cohorts. Additionally, changes were observed over an eight-year period as the disease progressed, with the most significant variation in p-tau217 noted in individuals who were positive for both A β and tau.

A study evaluated a two-step diagnostic workflow to screen for A β positivity in patients with MCI using plasma p-tau217 as a biomarker [74]. In the first step, a model combining plasma p-tau217 levels, age, and APOE ϵ 4 status stratified patients into low-, intermediate-, and high-risk categories for A β -PET positivity. Confirmatory CSF testing was conducted only for intermediate-risk individuals. The workflow demonstrated high accuracy (88.2–92.0%) while reducing the need for CSF or PET confirmatory testing by 61.2–85.9%, depending on the thresholds used. The findings suggest that plasma p-tau217 can effectively aid in stratifying patients for A β status, potentially reducing the reliance on more invasive or costly diagnostic methods.

Advances in detection technologies, such as Single-Molecule Array (SIMOA), have enabled precise quantification of low-abundance proteins in biofluids, including A β peptides and p-tau, biomarkers relevant to AD. SIMOA has been employed to measure plasma A β 42/A β 40 ratios, providing a less invasive and more accessible method for assessing amyloid status compared to PET

imaging or CSF sampling [75]. It has also been used for plasma p-tau quantification, supporting its application in early-stage AD detection [76] The method relies on digital bead-based ELISA, where paramagnetic beads coated with capture antibodies isolate single protein molecules. Fluorescence signals are measured in femtoliter-sized wells, allowing for detection of proteins at very low concentrations [77]

Neuroimaging has proven clinically relevant for monitoring AD progression and identifying early-stage indicators. Imaging modalities such as PET, computed tomography (CT), magnetic resonance imaging (MRI), magnetic resonance spectroscopy (MRS), diffusion tensor imaging (DTI), and free-water imaging (FW) enable both structural and functional assessments that aid in predicting AD progression [78–80]. PET imaging is considered the gold standard for non-invasive AD diagnostics, playing a critical role in disease monitoring and differential diagnosis.

The development of PET radiotracers for AD has progressed through distinct phases targeting both amyloid and tau pathologies. The amyloid-targeting timeline began in 2003 with [11C]PiB, the first successful selective radioligand for A β , which led to the development and FDA approval of three ¹⁸F-labeled tracers: [18F]florbetaben (Neuraceq), [18F]florbetapir (Amyvid), and [¹⁸F]flutemetamol (Vizamyl)[81,82]. The second generation of amyloid tracers, including [18F]AZD4694 ([18F]NAV4694), addressed limitations such as white matter retention (Rowe et al., 2013). Parallel developments in tau imaging emerged with first-generation tracers such as [18F]THK5117 and [11C]PBB3, followed by [18F]AV-1451 (flortaucipir) which demonstrated significant binding to neurofibrillary tangles. Second-generation tau tracers, including [18F]MK-6240 and [18F]PI-2620, showed improved selectivity and reduced off-target binding to monoamine oxidases [84,85]. This dual-pathway development of both amyloid and tau radiotracers has enabled in vivo visualization of the two primary pathological hallmarks of AD, facilitating early diagnosis, disease progression monitoring, and therapeutic evaluation [86,87]. The evolution of these imaging agents has particularly enhanced understanding of the temporal relationship between amyloid and tau accumulation in the disease process. These advancements prompted to propose a temporal model of the AD pathological cascade, thereby enhancing our capacity to visualize and quantify AD progression at both the molecular and cellular levels [88].

Additional neuroimaging techniques contribute to a comprehensive assessment in AD diagnostics. While CT imaging is less detailed than MRI, it offers a cost-effective option for initial evaluations. MRI with T1-weighted sequences provides high-resolution images, facilitating an indepth examination of brain structures affected by AD, such as the hippocampus and entorhinal cortex [69]. MRS has demonstrated utility in detecting metabolic changes associated with AD; however, it is used less frequently due to accessibility and cost constraints [89]. DTI, a specialized form of MRI, allows for the examination of white matter microstructure, aiding in the early detection of AD by assessing neural connectivity [90] In parallel, FW offers valuable insights by specifically detecting changes in extracellular water, highlighting early degenerative processes in the brain that conventional imaging techniques might miss [91,92].

Finally, machine learning and regression analysis represent promising tools for early AD diagnosis and predicting disease progression. Techniques such as least squares regression, support vector machines, and regression trees have been utilized to predict neuropsychological scores based on neuroimaging biomarkers [93]. Machine learning models trained on features from multiple imaging modalities have shown potential in accurately predicting cognitive outcomes, offering a compelling approach for precise diagnosis [77,79].

Are exosomes reliable sources of biomarkers for AD diagnosis?

Extracellular vesicles are lipid-coated structures produced by nearly all cell types and released into the extracellular milieu through membrane budding. They carry a variety of intracellular molecules that serve different roles within the organism, including plasma membrane restoration, intercellular communication, and the elimination of cellular waste [94,95] The cargo and function of EVs likely contribute to their heterogeneity, leading to their classification into exosomes (30 to 150 nm) and microvesicles (50 to 1000 nm). Notably, these structures differ in their biogenesis:

microvesicles are formed directly through outward budding, primarily from apoptotic cells, while exosomes are generated during the formation of endosomes (Figure 1). The endocytic pathway also provides the molecular machinery for the autophagy pathway, and both exosome secretion and autophagy play crucial roles in eliminating cellular waste in a coordinated manner [96]

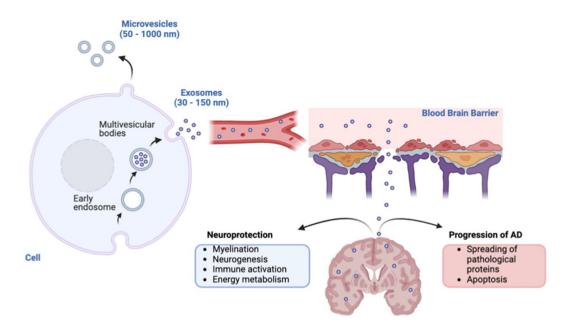


Figure 1. Schematic representation of the origin and function of exosomes in AD. Exosomes (30–150 nm) originate from multivesicular bodies, whereas microvesicles (50–1000 nm) bud directly from the plasma membrane. Once released into circulation, exosomes can cross the blood-brain barrier, as shown. Two main roles of exosomes in AD are depicted: (1) Neuroprotection, contributing to processes such as myelination, neurogenesis, immune activation, and energy metabolism; and (2) Progression of AD, mediating the spread of pathological proteins and apoptosis.

The role of exosomes in AD is complex, as they are implicated in both disease progression and the maintenance of normal brain physiology. Exosomes are known to be neuroprotective, supporting neuronal survival, myelination, neurogenesis, and immune responses following brain injury [97,98] Conversely, exosomes also contribute to AD pathology by facilitating the spread of pathogenic proteins, such as $A\beta$ and p-tau, to the brain and potentially influencing the formation of $A\beta$ plaques (Figure 2) [99,100]

Due to their nanoscale size and lipid composition, exosomes can easily traverse different organs and cells, even crossing the blood-brain barrier to interact with neurons, astrocytes, and microglia. They are delivered to target cells either by fusing with the cell membrane or through endocytosis, releasing their cargo, which may include molecules such as proteins, mRNA, miRNA, and circRNA, into the extracellular milieu. This process can potentially modulate gene expression [101]. The contents of exosomes also reflect the pathophysiological status of the originating cells, resulting in heterogeneity in terms of size, release pathway, content, and function [94]. Non-coding RNAs, such as miRNA and circRNA, present in exosomes are increasingly being explored as potential biomarkers for various neurodegenerative diseases, including AD and chronic cognitive disorders and syndromes.

Exosomes represent a promising source of biomarkers, as they are readily detectable in body fluids, particularly in serum and plasma. Furthermore, they contain specific membrane biomolecules that facilitate the identification of their originating cells. It is now possible to distinguish exosomes from different sources using commercial kits or magnetic beads linked to monoclonal antibodies,

such as those for neuronal-enriched exosomes (L1 Cell Adhesion Molecule - L1CAM) and astrocyte-enriched exosomes (Glutamine aspartate transporter - GLAST) [102,103].

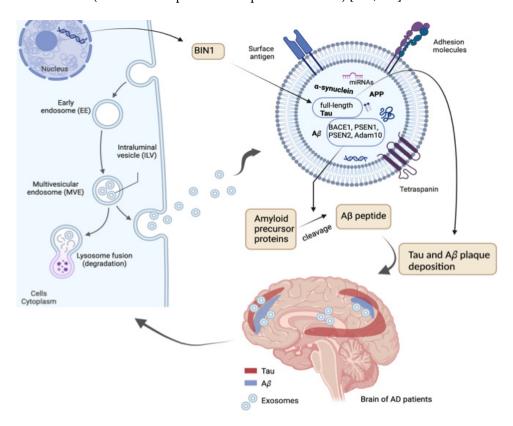


Figure 2. Exosomes and AD neuropathology. The figure depicts the formation of exosomes from multivesicular endosomes (MVEs) in cells and their role in AD. Exosomes carry key molecules, including amyloid precursor protein (APP), $A\beta$ peptides, full-length tau, α-synuclein, BIN1, miRNAs, and tetraspanins, which contribute to AD pathology. These vesicles may transport molecules that participate in the cleavage of APP into $A\beta$ peptides and, also possibly, in the deposition of tau and $A\beta$ plaques in the brain. The bottom section highlights the distribution of tau and $A\beta$ aggregates, alongside exosomes, in the brain of AD patients.

Exosomal Proteins

Previous studies have demonstrated that neuronal exosomes carry a variety of misfolded proteins, including A β , α -synuclein, full-length APP, and p-tau [100] (Figure 2). Notably, the gene locus Bridging Integrator 1 (BIN1), which is linked to late-onset AD, has been shown to facilitate the transport of tau via exosomes both *in vitro* and *in vivo* [104]. The tau protein in exosomes represents a promising strategy for diagnostic purposes, as full-length tau has been found to be more abundant in exosomes than in the free solutions of plasma or cerebrospinal fluid (CSF) of AD patients, whereas healthy individuals do not exhibit full-length tau [105].

Exosomes also carry enzymes such as BACE1, PSEN1, PSEN2, and Adam10 (Figure 2), which are responsible for cleaving APP, generating A β peptides, and consequently contributing to the formation of A β plaques [106]. Interestingly, when peripheral blood exosomes are injected into the hippocampus of an AD mouse model, they diffuse to the cortex and exhibit affinity with microglia, clustering around A β plaques (Figure 2). This suggests that exosomes play a role in amyloid deposition in AD, mediated by microglia ([100]). Consistently, exosomes stimulate the aggregation of A β 1-42 *in vitro*. Furthermore, it has been shown that exosomes from AD patients are more neurotoxic than those from healthy individuals, impairing calcium homeostasis and mitochondrial function [107]. Additionally, the presence of exosomes is enriched in the hippocampus of AD patients compared to those with Parkinson's disease or healthy individuals [108]. The release of exosomes is stimulated by glutamatergic activity in somato-dendritic compartments [109] and by neutral

sphingomyelinase 2 (nSMase2), an enzyme that converts sphingomyelin to ceramide. Notably, inhibiting nSMase2 with GW4869 reduces exosome production, accompanied by a decrease in brain ceramide and A β plaque deposition [110] .Moreover, amyloid peptides induce the secretion of exosomes containing PAR-4 and C18 ceramide, which activate Caspase 3 and consequently lead to astrocyte apoptosis *in vitro* [111].

Exosomal microRNAs

miRNAs are small, double-stranded RNAs that modulate gene expression by targeting mRNA for degradation or inhibiting translation, a process known as RNA interference (RNAi) [112,113] Several miRNAs, including miR-9, miR-29a, miR-107, miR-125b, and miR-135b, regulate various aspects of AD pathology, such as A β metabolism, tau phosphorylation, and neuroinflammation [114]. These miRNAs function as negative regulators of BACE1, thereby reducing A β production and potentially slowing the progression of AD [103].

Exosomal miRNAs are attractive biomarkers due to their stability in bodily fluids and their ability to reflect the origin and function of their originating cells. Studies have shown that miRNAs associated with AD risk genes (e.g., miR-107, miR-135b-5p) are elevated in neuronal exosomes from AD patients, while others, such as miR-320a, are reduced in the CSF of AD patients compared to healthy controls [115].

Exosomal Circular RNAs

CircRNAs, which are abundant in exosomes, play a crucial role in their functionality [111,116]. These stable, single-stranded RNA molecules are formed through back-splicing, resulting in a half-life exceeding 48 hours [117–119]. They are classified into three main types: intronic circRNAs (ciRNAs), exonic circRNAs (EcircRNAs), and exon-intron circRNAs (EIciRNAs) [120,121]. EcircRNAs, which comprise over 80% of circRNAs, are derived from pre-mRNA exons and are found in both the cytoplasm and exosomes, whereas ciRNAs originate from introns and EIciRNAs are primarily located in the nucleus [122,123]

CircRNAs regulate gene expression by binding to miRNA response elements (MREs), effectively acting as sponges that sequester miRNAs from their target mRNAs [116] Additionally, circRNAs interact with proteins, influencing cellular functions and highlighting their complex regulatory roles [124,125]

Studies indicate that circRNAs can circulate through serum and are enriched in exosomes, with their levels influenced by associated miRNA variations, allowing them to transfer biological activity to target cells [116,126–128] A prominent circRNA, CDR1as, exhibits high expression in the normal brain but shows reduced levels in the hippocampus and cortex of individuals with AD. CDR1-as functions as a sponge for miR-7; its decrease impairs the degradation of BACE1 and APP, leading to an abnormal increase in A β levels [129–131].

Exosomal miRNAs as Potential Biomarkers for AD Diagnosis

Recent research has focused on the role of miRNAs in AD pathogenesis associated with EVs such as exosomes (Table 2). Duan et al., 2024, examined serum-derived exosomal miRNAs in AD patients using high-throughput sequencing and RT-qPCR [132]. The authors identified hsa-miR-125b-1-3p, hsa-miR-193a-5p, hsa-miR-378a-3p, hsa-miR-378i, and hsa-miR-451a as differentially expressed miRNAs. Notably, hsa-miR-451a exhibited an area under the curve (AUC) of 0.728. Pathway enrichment analysis linked these miRNAs to neuroactive ligand-receptor interactions, PI3K-Akt signaling, and cytokine-cytokine receptor interactions [132]. Previous studies using EVs isolated from plasma samples also showed that miR-451a was significantly deregulated in AD relative to DLB [133] as well as in prodomal AD patients, highlighting its potential for differential diagnosis.

A study involving 42 AD patients and 19 healthy controls (HC) revealed significant alterations in EV membrane antigens, inflammatory cytokine content, and miRNA expression. Specifically,

myelin oligodendrocyte glycoprotein (MOG) and axonal glycoprotein CD171 correlate with disease severity. Five key miRNAs (let-7g-5p, miR126-3p, miR142-3p, miR146a-5p, and miR223-3p) showed a significant reduction in AD patients, suggesting a link between disease severity and neuroinflammation [134]. Serum exosomal miR-223 levels were significantly downregulated in dementia patients compared to controls. miR-223 levels correlated with Mini-Mental State Examination (MMSE) scores and inflammatory markers (IL-1 β , IL-6, TNF- α). Previous studies showed that its downregulation promotes inflammatory responses in macrophages. ROC curve analysis indicated strong diagnostic potential (AUC = 0.875) [135]. Visconte et al., 2023, found an increased expression of mR-223 in EVs from AD patients, with a positive correlation between miR-223-3p and t-tau levels, suggesting a possible involvement in the cognitive decline and neurodegeneration in AD [136].

The microRNA hsa-miR-126-3p has been identified in two distinct studies on AD. Gámez-Valero et al., 2019, found this miRNA to be significantly reduced in AD patients compared to healthy controls. Aharon et al., 2020, also observed a decrease in this miRNA in patients with severe AD compared to healthy individuals. hsa-miR-126-3p plays crucial roles in neuronal homeostasis and inflammatory response. Its reduced expression may be associated with various pathophysiological mechanisms of AD, such as neuroinflammation and apoptosis [133,134]. miR-126-3p regulates the NF-κB pathway, a key factor in the exacerbated inflammatory response observed in AD [137]. Its downregulation may contribute to a chronic neuroinflammatory environment, leading to sustained microglial activation and progressive neuronal damage.

Furthermore, studies suggest that miR-126-3p plays a fundamental role in maintaining the integrity of the blood-brain barrier (BBB). Its reduced expression may increase BBB permeability, facilitating the infiltration of inflammatory cells and neurotoxic molecules into the central nervous system [138]. The decrease in miR-126-3p may also impact cellular pathways involved in protein degradation, promoting the accumulation of Aβ levels [139]. This alteration is directly linked to the formation of senile plaques and neurofibrillary tangles—hallmark features of AD. Gámez-Valero et al., 2019, suggested that the downregulation of miR-126-3p could help differentiate AD from other dementias, such as DLB. Target gene analysis revealed the regulation of pathways related to phosphorylation enzymes, the proteasome, and apoptosis, reinforcing its involvement in AD pathophysiology. However, further studies with larger cohorts and detailed functional analyses of its molecular interactions are needed to validate the clinical use of miR-126-3p as a biomarker [133].

Cha et al., 2019, observed reduced levels of miR-132 in exosomes derived from the nervous system in the plasma of AD patients compared to controls [140]. miRNA analysis of the brains of AD patients revealed lower miR-132 levels compared to high-risk pathological controls. In plasma, miR-132-3p in exosomes from the nervous system exhibited high sensitivity and specificity for diagnosing AD. miR-132 plays a key role in molecular events leading to A β deposition by regulating A β metabolism, including tau, MAPK, and Sirtuin1 (SIRT1) pathways [141]. Another study demonstrated that miR-132/212 deficiency in an animal model resulted in exacerbated tau pathology and memory deficits, which could be partially reversed by replacing with miR-132 mimics [142]. Furthermore, reduced miR-132 expression in the hippocampus and medial frontal gyrus of AD patients (Braak scores 4 to 6) compared to non-demented controls (Braak scores 0 to 3) correlated with impairments in neuronal differentiation, mediated through the regulation of p250GAP, a Rho family GTPase enriched in the brain ([143].

Recent studies have highlighted the role of miRNAs in the pathogenesis, particularly in relation to gene regulation and disease-modifying factors such as education and depression.

Wang et al., 2022, identified hsa-miR-20a-5p as a key regulator associated with AD risk in a Chinese cohort. The expression of hsa-miR-20a-5p was reduced in AD patients; however, this reduction was less pronounced in individuals with higher levels of education [144]. Notably, miR-20a-5p directly targets several genes implicated in AD, including APP, whose increased expression enhances $A\beta$ production. The study also highlighted hsa-miR-185-5p as another downregulated miRNA in AD patients. Similar to miR-20a-5p, the decrease in hsa-miR-185-5p expression was

partially offset by higher educational attainment. Functional enrichment analysis showed that validated targets of these miRNA are involved in AD-related pathways including Presenilin 1 (PSEN1), an essential component of the γ -secretase complex, which was found to be upregulated in AD patients. Additionally, the downregulation of hsa-miR-185-5p was particularly evident in AD patients with concurrent depressive symptoms. One of its predicted targets, Breast Cancer 1 (BRCA1), plays a role in PSEN1 turnover and was found to be overexpressed in AD. Collectively, these findings suggest that reduced hsa-miR-20a-5p expression contributes to increased APP levels, while the downregulation of hsa-miR-185-5p and hsa-miR-181c-5p leads to upregulation of their respective targets, facilitating A β accumulation and plaque formation in AD pathology [144].

The studies by Duan et al., 2024, and Kumar et al., 2023, highlight the potential role of miR-125b as a biomarker for AD [132]. hsa-miR-125b-1-3p was found to be differentially expressed in serum-derived exosomes of AD patients, showing an AUC of 0.765, with a sensitivity of 82.1%, and a specificity of 67.7%. Functional enrichment analysis indicated its involvement in key neurobiological pathways, including the PI3K-Akt and Hippo signaling pathways—an evolutionarily conserved signaling network that plays a crucial role in regulating numerous biological processes [132]; [145].

Table 2. Exosomal miRNAs in biofluids as prospective biomarkers for AD diagnosis.

EV samples	miRNA's	Biomarkers Characteristics	Reference
Plasma	let-7g; miR126-3p; miR142-3p; miR146a; mir223-3p; mir26b	Low level in severe AD (SAD). KEGG pathway analysis - The analysis revealed 43 KEGG pathways associated with the most significant miRNAs, with a focus on p53, toll-like receptors, MAPK, NF-kappa B, AD, apoptosis, and PI3K-Akt. Increase of endothelial EVs. Elevated levels of EVs expressing the axonal glycoprotein CD171. Increase of inflammatory cytokines and reduction of over 50% growth factors, compared to EVs of HC. SAD score and miRNA expression presented correlation (let-7g r = 0.5223; miR126-3p r = 0.4564; miR142-3p r = 0.4675; miR146a r = 0.5433; mir223-3p r = 0.4779).	[146]
CSF, serum and plasma	miR-193b	,	[147]
Plasma	miR-342-3p miR-185-5p, hsa-miR- 20a-5p, and hsa-miR- 497-5p	Differential expression in AD group and correlated with other miRNAs decreased in AD. Related to AD and education level.	[148]
Plasma	hsa-miR-185-5p, hsa- miR-181c-5p, hsa-miR- 451a, and hsa-miR- 664a-3p	1 1	

		hsa-miR-664a-3p was upregulated in AD patients, which downregulated CREB1 and BDNF expression levels, thereby leading to a cognitive decline in AD patients.
Serum	miR-223	Decreased in patients with dementia. miR-223 level [150] correlated with Mental State Examination (MMSE) scores, Clinical Dementia Rating (CDR) scores, magnetic resonance spectroscopy (MRS) spectral ratios and serum concentrations of IL-1b, IL-6, TNF- α , and CRP. miR-223 downregulated in the dementia group compared to the control group. Differential expression of miR-223 between AD and Vascular Dementia (VaD) groups. Higher miR-223 levels in AD patients under medical care than those at their first clinical visit. Levels of miR-223 in the blood of dementia patients have a positive correlation with the scores on the MMSE and CDR scales (r = 0.365 and 0.4598, respectively). miR-223 levels in patients with dementia present positive correlation with the scores on the MMSE and CDR scales (r = 0.365 and 0.4598, respectively). Levels of IL-1 β , IL-6, TNF- α , and PCR elevated in patients with dementia. Higher in AD compared to VaD. A correlation was found between the levels of miR-223 and the concentrations of IL-1 β , IL-6, TNF- α , and PCR (r = -0.5504, -0.4549, -0.5152, -0.4977, respectively). miR-223 present AUC of 0.875 (95% CI: 0.7779–0.9721).
Plasma	• •	, Validation analysis confirmed significant [151], upregulation of miR-16-5p, miR-25-3p, miR-92a-3p, and miR-451a in prodromal AD patients, suggesting these dysregulated miRNAs are involved in the early progression of AD. Group of AD patients presented positive correlations between A β 42 and miR-30b-5p (r = 0.67) and between h-tau and miR-223-3p (r = 0.62).
Plasma	hsa-miR-451a e hsa- miR-21-5p hsa-miR-23a-3p, hsa- miR-126-3p, hsa-let-7i- 5p e hsa-miR-151a-3p	Down-regulated in AD samples respect to dementia [152] with Lewy bodies (DLB) patients. Decreased in AD respect to controls.
Cortical gray matter	miR-132 and miR-212	Levels of miR-132 separated controls from AD-MCI [140] with an AUC of 0.58 (95% CI: 0.38–0.78) and controls from AD dementia with an AUC of 0.77 (95% CI: 0.61–0.93). miR-212 showed better discrimination than miR-132 between AD-MCI and controls, and AD and controls. miR-212 levels separated controls from AD-MCI with an AUC of 0.68 (95% CI: 0.5–0.86) and controls from AD dementia with an AUC of 0.84 (95% CI: 0.72–0.96).

		TD 010 1: 1 ''' '' (00 00/ (000/ CI	
		miR-212 achieved a sensitivity of 92.2% (95% CI: 68.5–99.6%) and a specificity of 69.0% (95% CI: 50.8–	
Plasma	miR-502-5p	82.7%). AUC is 0.872, sensitivity 79.2 % and specificity 83.3	[153]
	miR-483-5p	%. Area Under the Curve (AUC) is 0.901, sensitivity 79.2 % and specificity 100 %.	
Serum	miR-193a-5p, hsa-miR-	-hsa-miR-125b-1-3p has an AUC of 0.765 in the AD group compared to the healthy group. Sensitivity (82.1) and specificity (67.7%).	[126,154,155]
CSF	miR-455–3p	Elevated levels in AD patients compared to controls (AUC = 0.745).	[156]
Plasma (NCAM/ABCA1 dual- labeled exosomal Aβ42/40)		The AUC of NCAM/ABCA1 dual-labeled exosomal A β 42/40 for diagnosis of SCD was higher than that of A β 42, T-tau, and P-T181-tau; the AUC of NCAM/ABCA1 dual-labeled exosomal miR-384 for diagnosis of SCD was higher than that of A β 42, A β 42/40, T-tau, P-T181-tau, and NfL. miR-384 can downregulate the expression and activity of BACE.	[157]
Plasma (Neurons: EVL1CAM)	miR-29a-5p, miR-125b- 5p, and miR-210-3p miR-210-3p and miR- 132-5p miR-106-5p	MCI, MCI-AD, and AD dementia (AUC = 0.948). MCI (AUC = 0.941). AD dementia (AUC = 1.000).	[158]
	miR-106b-5p	Negative correlation with cortical thickness in regions prone to age-related dementias as imaged in MRI.	ı
Plasma (Astrocytes: sEVGLAST)	miR-107 miR-107 and miR 132-	MCI, MCI-AD, and AD dementia (AUCs = 0.964); AD dementia (AUC = 1.000). Negative correlation with the cortical thickness.	
	5p and miR-210-3p miR-29a-5p and miR- 106-5p	MCI (AUCs = 0.941). Overall cognitive impairment (AUC = 0.925).	
Plasma (Microglia: sEVTMEM119)	miR-29a-5p miR-132-5p and miR- 125b-5p	MCI (AUC = 0.840). AD dementia (AUC = 1.000).	
Plasma	•	Negative correlation with the temporal cortical thickness. AD dementia (AUC = 1.000). Negative correlation	
(Oligodendrocytes: sEVPDGFRα) Plasma (Pericytes: sEVPDGFRα)	miR-9-5p	with temporal cortical thickness. Overall cognitive impairment (AUC = 0.935), MCI (AUC = 0.931), and AD (AUC = 1.000)	
sEVPDGFRβ) Plasma (Endothelial cells: sEVCD31)	miR-132-5p	(AUC = 0.931), and AD (AUC = 1.000). Overall impairment and MCI, and prediction of AD (AUC = 1.000).	
	miR-210-3p	Negative correlation with cortical thickness.	

Plasma (Pericytes: miR-9-5p Overall cognitive impairment (AUC = 1.000).

sEVPDGFRβ and (sEVPDGFRβ) and Endothelial cells: miR-132-5p (sEVCD31)

sEVCD31) miR-132-5p (sEVCD31) MCI and AD.

and miR-135b-5p (sEVPDGFRβ)

Abbreviations: EV, extracellular vesicle; SAD, severe AD; KEGG pathway, Kyoto Encyclopedia of Genes and Genomes; HC, healthy control; CSF: cerebrospinal fluid; MCI, mild cognitive impairment; 3' UTR, 3'-untranslated region; APP, amyloid precursor protein; DAT, dementia of Alzheimer-type; PSEN1, Presenilin 1; GSK3 β , Glycogen synthase kinase-3 β ; IL1, Interleukin-1; CREB1, cAMP-response element binding protein; BDNF, brain-derived neurotrophic factors; MMSE, Mental State Examination scores; CDR, Clinical Dementia Rating; MRS, magnetic resonance spectroscopy; TNF- α , Tumor necrosis factor alpha; VaD ,Vascular Dementia; AUC, area under the curve; DLB, dementia with Lewy bodies; CI, Confidence interval; NCAM/ABCA1, neural cell adhesion molecule (NCAM)/ATP-binding cassette transporter A1 (ABCA1); SCD, subjective cognitive decline; BACE-1, beta-secretase-1.

Concluding Remarks and Prospects

The potential of exosomes and their molecular contents, particularly miRNAs and circRNAs, as biomarkers in the context of AD is an area of significant interest. Exosomes have the capacity to serve as carriers of critical biomarkers linked to disease, and their molecular profiles may provide valuable insights into disease mechanisms and progression.

Future research should prioritize the exploration of exosomal biology to enhance our understanding of how these vesicles and their contents can function as reliable biomarkers for AD diagnosis. Identifying specific exosomal miRNAs and circRNAs associated with AD could lead to the development of novel diagnostic tools. Furthermore, elucidating the mechanisms by which exosomes influence neuronal health may reveal important connections between these biomarkers and the pathophysiology of AD.

In conclusion, exosomal ncRNAs may play a significant role in the diagnosis of AD, alongside well-established biomarkers such as $A\beta$ and tau, thereby helping to refine diagnostic results. The effectiveness of these biomarkers will depend on coordinated changes in the patterns of these molecules, as well as the validation of findings through multicenter studies with independent cohorts.

Author Contributions: Conceptualization, S.S.T.A and R.T.A.; writing—S.S.T.A. original draft preparation, S.S.T.A., R.T.A. C.L.F. M.V.R., R.T.A.; writing—review and editing, S.S.T.A., R.T.A. C.L.F. M.V.R., L.D.S.S., P.R.P.B; D.D.C.B., F.V.G., and R.T.A. All authors have read and agreed to the published version of the manuscript.

Funding: Please add: This research received no external funding.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Knopman DS, Amieva H, Petersen RC, Chételat G, Holtzman DM, Hyman BT, et al. Alzheimer disease. Nat Rev Dis Primers. 2021 May 13;7(1):33.
- 2. Nichols E, Steinmetz JD, Vollset SE, Fukutaki K, Chalek J, Abd-Allah F, et al. Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the Global Burden of Disease Study 2019. Lancet Public Health. 2022 Feb;7(2):e105–25.
- 3. Grøntvedt GR, Schröder TN, Sando SB, White L, Bråthen G, Doeller CF. Alzheimer's disease. Current Biology. 2018 Jun;28(11):R645–9.
- 4. Madnani RS. Alzheimer's disease: a mini-review for the clinician. Front Neurol. 2023 Jun 22;14.

- 5. Dubois B, Villain N, Frisoni GB, Rabinovici GD, Sabbagh M, Cappa S, et al. Clinical diagnosis of Alzheimer's disease: recommendations of the International Working Group. Lancet Neurol. 2021 Jun;20(6):484–96.
- Zhao N, Ren Y, Yamazaki Y, Qiao W, Li F, Felton LM, et al. Alzheimer's Risk Factors Age, APOE Genotype, and Sex Drive Distinct Molecular Pathways. Neuron. 2020 Jun;106(5):727-742.e6.
- Grant WB. A Brief History of the Progress in Our Understanding of Genetics and Lifestyle, Especially Diet, in the Risk of Alzheimer's Disease. Journal of Alzheimer's Disease. 2024 Aug 20;100(s1):S165–78.
- 8. Nordestgaard LT, Christoffersen M, Frikke-Schmidt R. Shared Risk Factors between Dementia and Atherosclerotic Cardiovascular Disease. Int J Mol Sci. 2022 Aug 29;23(17):9777.
- 9. Xu W, Tan L, Wang HF, Jiang T, Tan MS, Tan L, et al. Meta-analysis of modifiable risk factors for Alzheimer's disease. J Neurol Neurosurg Psychiatry. 2015 Aug 20;jnnp-2015-310548.
- Li JQ, Tan L, Wang HF, Tan MS, Tan L, Xu W, et al. Risk factors for predicting progression from mild cognitive impairment to Alzheimer's disease: a systematic review and meta-analysis of cohort studies. J Neurol Neurosurg Psychiatry. 2016 May;87(5):476–84.
- 11. Arenaza-Urquijo EM, Vemuri P. Resistance vs resilience to Alzheimer disease. Neurology. 2018 Apr 10;90(15):695–703.
- 12. Kunkle BW, Grenier-Boley B, Sims R, Bis JC, Damotte V, Naj AC, et al. Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates Aβ, tau, immunity and lipid processing. Nat Genet. 2019 Mar 28;51(3):414–30.
- 13. Monsell SE, Mock C, Fardo DW, Bertelsen S, Cairns NJ, Roe CM, et al. Genetic Comparison of Symptomatic and Asymptomatic Persons With Alzheimer Disease Neuropathology. Alzheimer Dis Assoc Disord. 2017 Jul;31(3):232–8.
- 14. Prpar Mihevc S, Majdič G. Canine Cognitive Dysfunction and Alzheimer's Disease Two Facets of the Same Disease? Front Neurosci. 2019 Jun 12;13.
- 15. Panek WK, Murdoch DM, Gruen ME, Mowat FM, Marek RD, Olby NJ. Plasma Amyloid Beta Concentrations in Aged and Cognitively Impaired Pet Dogs. Mol Neurobiol. 2021 Feb 24;58(2):483–9.
- 16. Fiandaca MS, Kapogiannis D, Mapstone M, Boxer A, Eitan E, Schwartz JB, et al. Identification of preclinical Alzheimer's disease by a profile of pathogenic proteins in neurally derived blood exosomes: A case-control study. Alzheimer's & Dementia. 2015 Jun 14;11(6):600.
- 17. Kapogiannis D, Mustapic M, Shardell MD, Berkowitz ST, Diehl TC, Spangler RD, et al. Association of Extracellular Vesicle Biomarkers With Alzheimer Disease in the Baltimore Longitudinal Study of Aging. JAMA Neurol. 2019 Nov 1;76(11):1340.
- 18. Bhat AA, Younes SN, Raza SS, Zarif L, Nisar S, Ahmed I, et al. Role of non-coding RNA networks in leukemia progression, metastasis and drug resistance. Mol Cancer. 2020 Dec 12;19(1):57.
- 19. Soares Martins T, Trindade D, Vaz M, Campelo I, Almeida M, Trigo G, et al. Diagnostic and therapeutic potential of exosomes in Alzheimer's disease. J Neurochem. 2021 Jan 6;156(2):162–81.
- 20. Goetzl EJ, Kapogiannis D, Schwartz JB, Lobach I V., Goetzl L, Abner EL, et al. Decreased synaptic proteins in neuronal exosomes of frontotemporal dementia and Alzheimer's disease. The FASEB Journal. 2016 Dec 6;30(12):4141–8.
- 21. Goetzl EJ, Schwartz JB, Abner EL, Jicha GA, Kapogiannis D. High complement levels in astrocyte-derived exosomes of Alzheimer disease. Ann Neurol. 2018 Mar 10;83(3):544–52.
- 22. Winston CN, Goetzl EJ, Schwartz JB, Elahi FM, Rissman RA. Complement protein levels in plasma astrocyte-derived exosomes are abnormal in conversion from mild cognitive impairment to Alzheimer's disease dementia. Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring. 2019 Dec 14;11(1):61–6.
- 23. Winston CN, Goetzl EJ, Akers JC, Carter BS, Rockenstein EM, Galasko D, et al. Prediction of conversion from mild cognitive impairment to dementia with neuronally derived blood exosome protein profile. Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring. 2016 Jan 7;3(1):63–72.
- 24. Chen J jiao, Yang G, Yan Q qing, Zhao J, Li S. Exosome-encapsulated microRNAs as promising biomarkers for Alzheimer's disease. Rev Neurosci. 2019 Dec 18;31(1):77–87.

- 25. Huang X, Yuan T, Tschannen M, Sun Z, Jacob H, Du M, et al. Characterization of human plasma-derived exosomal RNAs by deep sequencing. BMC Genomics. 2013;14(1).
- 26. Van Balkom BWM, Eisele AS, Michiel Pegtel D, Bervoets S, Verhaar MC. Quantitative and qualitative analysis of small RNAs in human endothelial cells and exosomes provides insights into localized RNA processing, degradation and sorting. J Extracell Vesicles. 2015;4(2015).
- 27. Mori MA, Ludwig RG, Garcia-Martin R, Brandão BB, Kahn CR. Extracellular miRNAs: From Biomarkers to Mediators of Physiology and Disease. Cell Metab. 2019 Oct;30(4):656–73.
- 28. Zhang J, Li S, Li L, Li M, Guo C, Yao J, et al. Exosome and Exosomal MicroRNA: Trafficking, Sorting, and Function. Genomics Proteomics Bioinformatics. 2015 Feb 1;13(1):17–24.
- 29. Shobeiri P, Alilou S, Jaberinezhad M, Zare F, Karimi N, Maleki S, et al. Circulating long non-coding RNAs as novel diagnostic biomarkers for Alzheimer's disease (AD): A systematic review and meta-analysis. PLoS One. 2023 Mar 22;18(3):e0281784.
- 30. Tang Z, Cheng X, Su X, Wu L, Cai Q, Wu H. *Treponema denticola* Induces Alzheimer-Like Tau Hyperphosphorylation by Activating Hippocampal Neuroinflammation in Mice. J Dent Res. 2022 Jul 22;101(8):992–1001.
- 31. Graff-Radford J, Yong KXX, Apostolova LG, Bouwman FH, Carrillo M, Dickerson BC, et al. New insights into atypical Alzheimer's disease in the era of biomarkers. Lancet Neurol. 2021 Mar;20(3):222–34.
- 32. Phochantachinda S, Chatchaisak D, Temviriyanukul P, Chansawang A, Pitchakarn P, Chantong B. Ethanolic Fruit Extract of Emblica officinalis Suppresses Neuroinflammation in Microglia and Promotes Neurite Outgrowth in Neuro2a Cells. Evidence-Based Complementary and Alternative Medicine. 2021 Sep 7;2021:1–16.
- 33. Armstrong D, Wildman DE. Extracellular Vesicles and the Promise of Continuous Liquid Biopsies. J Pathol Transl Med. 2018 Jan 15;52(1):1–8.
- 34. Kanninen KM, Bister N, Koistinaho J, Malm T. Exosomes as new diagnostic tools in CNS diseases. Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease. 2016 Mar;1862(3):403–10.
- 35. Apostolova L. Early-Stage Alzheimer Primer. J Clin Psychiatry. 2021 May 4;82(3).
- 36. Eramudugolla R, Mortby ME, Sachdev P, Meslin C, Kumar R, Anstey KJ. Evaluation of a research diagnostic algorithm for DSM-5 neurocognitive disorders in a population-based cohort of older adults. Alzheimers Res Ther. 2017 Dec 4;9(1):15.
- 37. Sachdev PS, Blacker D, Blazer DG, Ganguli M, Jeste D V., Paulsen JS, et al. Classifying neurocognitive disorders: the DSM-5 approach. Nat Rev Neurol. 2014 Nov 30;10(11):634–42.
- 38. Thal DR, Capetillo-Zarate E, Del Tredici K, Braak H. The Development of Amyloid β Protein Deposits in the Aged Brain. Science of Aging Knowledge Environment. 2006 Mar 8;2006(6).
- 39. Hyman BT, Phelps CH, Beach TG, Bigio EH, Cairns NJ, Carrillo MC, et al. National Institute on Aging–Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. Alzheimer's & Dementia. 2012 Jan;8(1):1–13.
- 40. Montine TJ, Phelps CH, Beach TG, Bigio EH, Cairns NJ, Dickson DW, et al. National Institute on Aging–Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease: a practical approach. Acta Neuropathol. 2012 Jan 20;123(1):1–11.
- 41. Trejo-Lopez JA, Yachnis AT, Prokop S. Neuropathology of Alzheimer's Disease. Neurotherapeutics. 2022 Jan;19(1):173–85.
- 42. DeTure MA, Dickson DW. The neuropathological diagnosis of Alzheimer's disease. Mol Neurodegener. 2019 Dec 2;14(1):32.
- 43. Braak H, Thal DR, Ghebremedhin E, Del Tredici K. Stages of the pathologic process in alzheimer disease: Age categories from 1 to 100 years. J Neuropathol Exp Neurol. 2011;70(11).
- 44. Schilling LP, Balthazar MLF, Radanovic M, Forlenza OV, Silagi ML, Smid J, et al. Diagnóstico da doença de Alzheimer: recomendações do Departamento Científico de Neurologia Cognitiva e do Envelhecimento da Academia Brasileira de Neurologia. Dement Neuropsychol. 2022 Sep;16(3 suppl 1):25–39.
- 45. Murden RA. The Diagnosis of Alzheimer's Disease. In 1990. p. 59-64.

- 46. Parker TD, Slattery CF, Zhang J, Nicholas JM, Paterson RW, Foulkes AJM, et al. Cortical microstructure in young onset Alzheimer's disease using neurite orientation dispersion and density imaging. Hum Brain Mapp. 2018 Jul 25;39(7):3005–17.
- 47. Serrano-Pozo A, Frosch MP, Masliah E, Hyman BT. Neuropathological Alterations in Alzheimer Disease. Cold Spring Harb Perspect Med. 2011 Sep 1;1(1):a006189–a006189.
- 48. Alafuzoff I, Libard S. Ageing-Related Neurodegeneration and Cognitive Decline. Int J Mol Sci. 2024 Apr 5;25(7):4065.
- 49. Boyle PA, Yu L, Leurgans SE, Wilson RS, Brookmeyer R, Schneider JA, et al. Attributable risk of Alzheimer's dementia attributed to age-related neuropathologies. Ann Neurol. 2019 Jan 19;85(1):114–24.
- 50. White LR, Corrada MM, Kawas CH, Cholerton BA, Edland SE, Flanagan ME, et al. Neuropathologic Changes of Alzheimer's Disease and Related Dementias: Relevance to Future Prevention. Journal of Alzheimer's Disease. 2023 Aug 29;95(1):307–16.
- 51. Jack CR, Andrews JS, Beach TG, Buracchio T, Dunn B, Graf A, et al. Revised criteria for diagnosis and staging of Alzheimer's disease: Alzheimer's Association Workgroup. Alzheimer's & Dementia. 2024 Aug 27;20(8):5143–69.
- 52. Dubois B, Villain N, Schneider L, Fox N, Campbell N, Galasko D, et al. Alzheimer Disease as a Clinical-Biological Construct—An International Working Group Recommendation. JAMA Neurol. 2024 Dec 1;81(12):1304.
- 53. Jack CR, Andrews JS, Beach TG, Buracchio T, Dunn B, Graf A, et al. Revised criteria for diagnosis and staging of Alzheimer's disease: Alzheimer's Association Workgroup. Alzheimer's & Dementia. 2024 Aug 27;20(8):5143–69.
- 54. McKenna MC, O'Philibin L, Foley T, Dolan C, Kennelly S, O'Dowd S, et al. The Use of Biomarkers in Diagnosing Alzheimer's Disease: Recommendations of the Irish Working Group on Biological Approaches to the Diagnosis of Dementia. Ir Med J. 2024 Dec 19;117(10):1062.
- 55. Studart Neto A, Barbosa BJAP, Coutinho AM, Souza LC de, Schilling LP, Silva MNM da, et al. Guidelines for the use and interpretation of Alzheimer's disease biomarkers in clinical practice in Brazil: recommendations from the Scientific Department of Cognitive Neurology and Aging of the Brazilian Academy of Neurology. Dement Neuropsychol. 2024;18.
- 56. Okazawa H, Nogami M, Ishida S, Makino A, Mori T, Kiyono Y, et al. PET/MRI multimodality imaging to evaluate changes in glymphatic system function and biomarkers of Alzheimer's disease. Sci Rep. 2024 May 29;14(1):12310.
- 57. Jack CR. Advances in Alzheimer's disease research over the past two decades. Lancet Neurol. 2022 Oct;21(10):866–9.
- 58. Ashton NJ, Janelidze S, Al Khleifat A, Leuzy A, van der Ende EL, Karikari TK, et al. A multicentre validation study of the diagnostic value of plasma neurofilament light. Nat Commun. 2021 Jun 7;12(1):3400.
- 59. Benedet AL, Milà-Alomà M, Vrillon A, Ashton NJ, Pascoal TA, Lussier F, et al. Differences Between Plasma and Cerebrospinal Fluid Glial Fibrillary Acidic Protein Levels Across the Alzheimer Disease Continuum. JAMA Neurol. 2021 Dec 1;78(12):1471.
- 60. Hampel H, Hardy J, Blennow K, Chen C, Perry G, Kim SH, et al. The Amyloid-β Pathway in Alzheimer's Disease. Mol Psychiatry. 2021 Oct 30;26(10):5481–503.
- 61. Schindler SE, Bollinger JG, Ovod V, Mawuenyega KG, Li Y, Gordon BA, et al. High-precision plasma β-amyloid 42/40 predicts current and future brain amyloidosis. Neurology. 2019 Oct 22;93(17).
- 62. Karikari TK, Ashton NJ, Brinkmalm G, Brum WS, Benedet AL, Montoliu-Gaya L, et al. Blood phospho-tau in Alzheimer disease: analysis, interpretation, and clinical utility. Nat Rev Neurol. 2022 Jul 18;18(7):400–18.
- 63. Ashton NJ, Puig-Pijoan A, Milà-Alomà M, Fernández-Lebrero A, García-Escobar G, González-Ortiz F, et al. Plasma and CSF biomarkers in a memory clinic: Head-to-head comparison of phosphorylated tau immunoassays. Alzheimer's & Dementia. 2023 May 12;19(5):1913–24.
- 64. Mielke MM, Aakre JA, Algeciras-Schimnich A, Proctor NK, Machulda MM, Eichenlaub U, et al. Comparison of CSF phosphorylated tau 181 and 217 for cognitive decline. Alzheimer's & Dementia. 2022 Apr 26;18(4):602–11.

- 65. Jia J, Ning Y, Chen M, Wang S, Yang H, Li F, et al. Biomarker Changes during 20 Years Preceding Alzheimer's Disease. New England Journal of Medicine. 2024 Feb 22;390(8):712–22.
- 66. Palmqvist S, Tideman P, Mattsson-Carlgren N, Schindler SE, Smith R, Ossenkoppele R, et al. Blood Biomarkers to Detect Alzheimer Disease in Primary Care and Secondary Care. JAMA. 2024 Oct 15;332(15):1245.
- 67. Ashton NJ, Brum WS, Di Molfetta G, Benedet AL, Arslan B, Jonaitis E, et al. Diagnostic Accuracy of a Plasma Phosphorylated Tau 217 Immunoassay for Alzheimer Disease Pathology. JAMA Neurol. 2024 Mar 1;81(3):255.
- 68. Farvadi F, Hashemi F, Amini A, Vakilinezhad MA, Raee MJ. Early Diagnosis of Alzheimer's Disease with Blood Test; Tempting but Challenging. Int J Mol Cell Med. 2023;12(2).
- 69. Livingston G, Huntley J, Liu KY, Costafreda SG, Selbæk G, Alladi S, et al. Dementia prevention, intervention, and care: 2024 report of the Lancet standing Commission. The Lancet. 2024 Aug;404(10452):572–628.
- 70. Petrie EC, Cross DJ, Galasko D, Schellenberg GD, Raskind MA, Peskind ER, et al. Preclinical Evidence of Alzheimer Changes. Arch Neurol. 2009 May 1;66(5).
- 71. Sheikh-Bahaei N, Chen M, Pappas I. Magnetic Resonance Spectroscopy (MRS) in Alzheimer's Disease. In 2024. p. 115–42.
- 72. Chen X, Firulyova M, Manis M, Herz J, Smirnov I, Aladyeva E, et al. Microglia-mediated T cell infiltration drives neurodegeneration in tauopathy. Nature. 2023 Mar 23;615(7953):668–77.
- 73. Koji Kamagata, Cristina Andica, Kaito Takabayashi, Yuya Saito, Toshiaki Taoka, Hayato Nozaki, et al. Associação de índices de ressonância magnética do sistema glinfático com deposição amiloide e cognição em comprometimento cognitivo leve e doença de Alzheimer. Neurology. 2022;2648–60.
- 74. Brum WS, Cullen NC, Janelidze S, Ashton NJ, Zimmer ER, Therriault J, et al. A two-step workflow based on plasma p-tau217 to screen for amyloid β positivity with further confirmatory testing only in uncertain cases. Nat Aging. 2023 Aug 31;3(9):1079–90.
- 75. Colmant L, Boyer E, Gerard T, Sleegers K, Lhommel R, Ivanoiu A, et al. Definition of a Threshold for the Plasma Aβ42/Aβ40 Ratio Measured by Single-Molecule Array to Predict the Amyloid Status of Individuals without Dementia. Int J Mol Sci. 2024 Jan 18;25(2):1173.
- 76. Ding XL, Tuo Q zhang, Lei P. An Introduction to Ultrasensitive Assays for Plasma Tau Detection. Journal of Alzheimer's Disease. 2021 Apr 20;80(4):1353–62.
- 77. Hwang YT, Rocchi L, Hammond P, Hardy CJD, Warren JD, Ridha BH, et al. Effect of donepezil on transcranial magnetic stimulation parameters in Alzheimer's disease. Alzheimer's & Dementia: Translational Research & Clinical Interventions. 2018 Jan 2;4(1):103–7.
- 78. Kamagata K, Andica C, Hatano T, Ogawa T, Takeshige-Amano H, Ogaki K, et al. Advanced diffusion magnetic resonance imaging in patients with Alzheimer's and Parkinson's diseases. Neural Regen Res. 2020;15(9):1590.
- 79. Tabarestani S, Eslami M, Cabrerizo M, Curiel RE, Barreto A, Rishe N, et al. A Tensorized Multitask Deep Learning Network for Progression Prediction of Alzheimer's Disease. Front Aging Neurosci. 2022;14:810873.
- 80. Wen J, Thibeau-Sutre E, Diaz-Melo M, Samper-González J, Routier A, Bottani S, et al. Convolutional neural networks for classification of Alzheimer's disease: Overview and reproducible evaluation. Med Image Anal. 2020 Jul;63:101694.
- 81. Bischof GN, Bartenstein P, Barthel H, van Berckel B, Doré V, van Eimeren T, et al. Toward a Universal Readout for ¹⁸ F-Labeled Amyloid Tracers: The CAPTAINs Study. Journal of Nuclear Medicine. 2021 Jul 1;62(7):999–1005.
- 82. Filippi L, Chiaravalloti A, Bagni O, Schillaci O. 18F-labeled radiopharmaceuticals for the molecular neuroimaging of amyloid plaques in Alzheimer's disease. Am J Nucl Med Mol Imaging. 2018;8(4):268–81.
- 83. Rowe CC, Pejoska S, Mulligan RS, Jones G, Chan JG, Svensson S, et al. Head-to-Head Comparison of 11 C-PiB and 18 F-AZD4694 (NAV4694) for β -Amyloid Imaging in Aging and Dementia. Journal of Nuclear Medicine. 2013 Jun;54(6):880–6.

- 84. Murugan NA, Chiotis K, Rodriguez-Vieitez E, Lemoine L, Ågren H, Nordberg A. Cross-interaction of tau PET tracers with monoamine oxidase B: evidence from in silico modelling and in vivo imaging. Eur J Nucl Med Mol Imaging. 2019 Jun 27;46(6):1369–82.
- 85. Groot C, Villeneuve S, Smith R, Hansson O, Ossenkoppele R. Tau PET Imaging in Neurodegenerative Disorders. Journal of Nuclear Medicine. 2022 Jun 1;63(Supplement 1):20S-26S.
- 86. Klunk WE, Engler H, Nordberg A, Wang Y, Blomqvist G, Holt DP, et al. Imaging brain amyloid in Alzheimer's disease with Pittsburgh Compound-B. Ann Neurol. 2004 Mar 21;55(3):306–19.
- 87. Xia C, Arteaga J, Chen G, Gangadharmath U, Gomez LF, Kasi D, et al. [¹⁸ F]T807, a novel tau positron emission tomography imaging agent for Alzheimer's disease. Alzheimer's & Dementia. 2013 Nov 13;9(6):666–76.
- 88. Jack CR, Knopman DS, Jagust WJ, Shaw LM, Aisen PS, Weiner MW, et al. Hypothetical model of dynamic biomarkers of the Alzheimer's pathological cascade. Lancet Neurol. 2010 Jan;9(1):119–28.
- 89. Wittens MMJ, Allemeersch GJ, Sima DM, Naeyaert M, Vanderhasselt T, Vanbinst AM, et al. Inter- and Intra-Scanner Variability of Automated Brain Volumetry on Three Magnetic Resonance Imaging Systems in Alzheimer's Disease and Controls. Front Aging Neurosci. 2021;13.
- 90. Chen X, Firulyova M, Manis M, Herz J, Smirnov I, Aladyeva E, et al. Microglia-mediated T cell infiltration drives neurodegeneration in tauopathy. Nature. 2023 Mar 23;615(7953):668–77.
- 91. Maillard P, Hillmer LJ, Lu H, Arfanakis K, Gold BT, Bauer CE, et al. MRI free water as a biomarker for cognitive performance: Validation in the MarkVCID consortium. Alzheimer's & Dementia: Diagnosis, Assessment & Disease Monitoring. 2022 Jan 13;14(1).
- 92. Sathe A, Yang Y, Schilling KG, Shashikumar N, Moore E, Dumitrescu L, et al. Free-water: A promising structural biomarker for cognitive decline in aging and mild cognitive impairment. Imaging Neuroscience. 2024 Sep 18;2:1–16.
- 93. Moradi E, Hallikainen I, Hänninen T, Tohka J. Rey's Auditory Verbal Learning Test scores can be predicted from whole brain MRI in Alzheimer's disease. Neuroimage Clin. 2017;13:415–27.
- 94. Kalluri R, LeBleu VS. The biology , function , and biomedical applications of exosomes. Science (1979). 2020 Feb 7;367(6478).
- 95. Ciferri MC, Quarto R, Tasso R. Extracellular Vesicles as Biomarkers and Therapeutic Tools: From Pre-Clinical to Clinical Applications. Biology (Basel). 2021 Apr 23;10(5):359.
- 96. Villarroya-Beltri C, Baixauli F, Mittelbrunn M, Fernández-Delgado I, Torralba D, Moreno-Gonzalo O, et al. ISGylation controls exosome secretion by promoting lysosomal degradation of MVB proteins. Nat Commun. 2016 Nov 24;7(1):13588.
- 97. Krämer-Albers E, Bretz N, Tenzer S, Winterstein C, Möbius W, Berger H, et al. Oligodendrocytes secrete exosomes containing major myelin and stress-protective proteins: Trophic support for axons? Proteomics Clin Appl. 2007 Nov 2;1(11):1446–61.
- 98. Frühleis C, Fröhlich D, Kuo WP, Amphornrat J, Thilemann S, Saab AS, et al. Neurotransmitter-Triggered Transfer of Exosomes Mediates Oligodendrocyte–Neuron Communication. PLoS Biol. 2013 Jul 9;11(7):e1001604.
- 99. Drago F, Lombardi M, Prada I, Gabrielli M, Joshi P, Cojoc D, et al. ATP Modifies the Proteome of Extracellular Vesicles Released by Microglia and Influences Their Action on Astrocytes. Front Pharmacol. 2017 Dec 13;8.
- 100. Zhang D, Lee H, Zhu Z, Minhas JK, Jin Y. Enrichment of selective miRNAs in exosomes and delivery of exosomal miRNAs in vitro and in vivo. American Journal of Physiology-Lung Cellular and Molecular Physiology. 2017 Jan 1;312(1):L110–21.
- 101. Migneault F, Dieudé M, Turgeon J, Beillevaire D, Hardy MP, Brodeur A, et al. Apoptotic exosome-like vesicles regulate endothelial gene expression, inflammatory signaling, and function through the NF-κB signaling pathway. Sci Rep. 2020 Jul 28;10(1):12562.
- 102. Martinez-Valbuena I, Visanji NP, Kim A, Lau HHC, So RWL, Alshimemeri S, et al. Alpha-synuclein seeding shows a wide heterogeneity in multiple system atrophy. Transl Neurodegener. 2022 Dec 7;11(1):7.

- 103. Kumar A, Su Y, Sharma M, Singh S, Kim S, Peavey JJ, et al. MicroRNA expression in extracellular vesicles as a novel blood-based biomarker for Alzheimer's disease. Alzheimer's & Dementia. 2023 Nov 18;19(11):4952–66.
- 104. Crotti A, Sait HR, McAvoy KM, Estrada K, Ergun A, Szak S, et al. BIN1 favors the spreading of Tau via extracellular vesicles. Sci Rep. 2019 Jul 1;9(1):9477.
- 105. Guix F, Corbett G, Cha D, Mustapic M, Liu W, Mengel D, et al. Detection of Aggregation-Competent Tau in Neuron-Derived Extracellular Vesicles. Int J Mol Sci. 2018 Feb 27;19(3):663.
- 106. Bellingham SA, Guo BB, Coleman BM, Hill AF. Exosomes: Vehicles for the Transfer of Toxic Proteins Associated with Neurodegenerative Diseases? Front Physiol. 2012;3.
- 107. Eitan E, Hutchison ER, Marosi K, Comotto J, Mustapic M, Nigam SM, et al. Extracellular vesicle-associated $A\beta$ mediates trans-neuronal bioenergetic and Ca2+-handling deficits in Alzheimer's disease models. NPJ Aging Mech Dis. 2016 Sep 22;2(1):16019.
- 108. Rajendran L, Honsho M, Zahn TR, Keller P, Geiger KD, Verkade P, et al. Alzheimer's disease β -amyloid peptides are released in association with exosomes. Proceedings of the National Academy of Sciences. 2006 Jul 25;103(30):11172–7.
- 109. Frühleis C, Fröhlich D, Kuo WP, Amphornrat J, Thilemann S, Saab AS, et al. Neurotransmitter-Triggered Transfer of Exosomes Mediates Oligodendrocyte–Neuron Communication. PLoS Biol. 2013 Jul 9;11(7):e1001604.
- 110. Dinkins MB, Dasgupta S, Wang G, Zhu G, Bieberich E. Exosome reduction in vivo is associated with lower amyloid plaque load in the 5XFAD mouse model of Alzheimer's disease. Neurobiol Aging. 2014 Aug;35(8):1792–800.
- 111. Wang G, Dinkins M, He Q, Zhu G, Poirier C, Campbell A, et al. Astrocytes Secrete Exosomes Enriched with Proapoptotic Ceramide and Prostate Apoptosis Response 4 (PAR-4). Journal of Biological Chemistry. 2012 Jun;287(25):21384–95.
- 112. Fire A, Xu S, Montgomery MK, Kostas SA, Driver SE, Mello CC. Potent and specific genetic interference by double-stranded RNA in Caenorhabditis elegans. Nature. 1998 Feb;391(6669):806–11.
- 113. Titze-de-Almeida R, David C, Titze-de-Almeida SS. The Race of 10 Synthetic RNAi-Based Drugs to the Pharmaceutical Market. Pharm Res. 2017 Jul 7;34(7):1339–63.
- 114. Takousis P, Sadlon A, Schulz J, Wohlers I, Dobricic V, Middleton L, et al. Differential expression of microRNAs in Alzheimer's disease brain, blood, and cerebrospinal fluid. Alzheimer's & Dementia. 2019 Nov 5;15(11):1468–77.
- 115. Tan YJ, Wong BYX, Vaidyanathan R, Sreejith S, Chia SY, Kandiah N, et al. Altered Cerebrospinal Fluid Exosomal microRNA Levels in Young-Onset Alzheimer's Disease and Frontotemporal Dementia. J Alzheimers Dis Rep. 2021 Mar 11;5(1):805–13.
- 116. Li Y, Zheng Q, Bao C, Li S, Guo W, Zhao J, et al. Circular RNA is enriched and stable in exosomes: a promising biomarker for cancer diagnosis. Cell Res. 2015 Aug 3;25(8):981–4.
- 117. Schwanhäusser B, Busse D, Li N, Dittmar G, Schuchhardt J, Wolf J, et al. Global quantification of mammalian gene expression control. Nature. 2011 May 19;473(7347):337–42.
- 118. Jeck WR, Sorrentino JA, Wang K, Slevin MK, Burd CE, Liu J, et al. Circular RNAs are abundant, conserved, and associated with ALU repeats. RNA. 2013 Feb;19(2):141–57.
- 119. Kristensen LS, Andersen MS, Stagsted LVW, Ebbesen KK, Hansen TB, Kjems J. The biogenesis, biology and characterization of circular RNAs. Nat Rev Genet. 2019 Nov 8;20(11):675–91.
- 120. Titze-de-Almeida SS, Titze-de-Almeida R. Progress in circRNA-Targeted Therapy in Experimental Parkinson's Disease. Vol. 15, Pharmaceutics. 2023.
- 121. Isaac R, Reis FCG, Ying W, Olefsky JM. Exosomes as mediators of intercellular crosstalk in metabolism. Vol. 33, Cell Metabolism. 2021.
- 122. Gao Y, Wang J, Zheng Y, Zhang J, Chen S, Zhao F. Comprehensive identification of internal structure and alternative splicing events in circular RNAs. Nat Commun. 2016 Jun 28;7(1):12060.
- 123. Isaac R, Reis FCG, Ying W, Olefsky JM. Exosomes as mediators of intercellular crosstalk in metabolism. Cell Metab. 2021 Sep;33(9):1744–62.

- 124. Ulshöfer CJ, Pfafenrot C, Bindereif A, Schneider T. Methods to study circRNA-protein interactions. Methods. 2021 Dec;196:36–46.
- 125. Aufiero S, van den Hoogenhof MMG, Reckman YJ, Beqqali A, van der Made I, Kluin J, et al. Cardiac circRNAs arise mainly from constitutive exons rather than alternatively spliced exons. RNA. 2018 Jun;24(6):815–27.
- 126. Duan Y, Wang Y, Liu Y, Jin Z, Liu C, Yu X, et al. Circular RNAs in Parkinson's Disease: Reliable Biological Markers and Targets for Rehabilitation. Mol Neurobiol. 2023 Jun 25;60(6):3261–76.
- 127. Preußer C, Hung L, Schneider T, Schreiner S, Hardt M, Moebus A, et al. Selective release of circRNAs in platelet-derived extracellular vesicles. J Extracell Vesicles. 2018 Dec 15;7(1).
- 128. Han QF, Li WJ, Hu KS, Gao J, Zhai WL, Yang JH, et al. Exosome biogenesis: machinery, regulation, and therapeutic implications in cancer. Mol Cancer. 2022 Nov 1;21(1):207.
- 129. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, et al. Natural RNA circles function as efficient microRNA sponges. Nature. 2013 Mar 27;495(7441):384–8.
- 130. Shi L, Yan P, Liang Y, Sun Y, Shen J, Zhou S, et al. Circular RNA expression is suppressed by androgen receptor (AR)-regulated adenosine deaminase that acts on RNA (ADAR1) in human hepatocellular carcinoma. Cell Death Dis. 2017 Nov 16;8(11):e3171–e3171.
- 131. Zhao J, Deng Y, Jiang Z, Qing H. G Protein-Coupled Receptors (GPCRs) in Alzheimer's Disease: A Focus on BACE1 Related GPCRs. Front Aging Neurosci. 2016 Mar 24;8.
- 132. Duan X, Zheng Q, Liang L, Zhou L. Serum Exosomal miRNA-125b and miRNA-451a are Potential Diagnostic Biomarker for Alzheimer's Diseases. Degener Neurol Neuromuscul Dis. 2024;14:21–31.
- 133. Gámez-Valero A, Campdelacreu J, Vilas D, Ispierto L, Reñé R, Álvarez R, et al. Exploratory study on microRNA profiles from plasma-derived extracellular vesicles in Alzheimer's disease and dementia with Lewy bodies. Transl Neurodegener. 2019 Dec 4;8(1):31.
- 134. Aharon A, Spector P, Ahmad RS, Horrany N, Sabbach A, Brenner B, et al. Extracellular Vesicles of Alzheimer's Disease Patients as a Biomarker for Disease Progression. Mol Neurobiol. 2020 Oct 17;57(10):4156–69.
- 135. Han P fei, Wei L, Duan Z qing, Zhang Z liang, Chen T yu, Lu J gong, et al. Contribution of IL-1 β , 6 and TNF- α to the form of post-traumatic osteoarthritis induced by "idealized" anterior cruciate ligament reconstruction in a porcine model. Int Immunopharmacol. 2018 Dec;65:212–20.
- 136. Visconte C, Fenoglio C, Serpente M, Muti P, Sacconi A, Rigoni M, et al. Altered Extracellular Vesicle miRNA Profile in Prodromal Alzheimer's Disease. Int J Mol Sci. 2023 Sep 29;24(19).
- 137. Liang Y, Wang L. Inflamma-MicroRNAs in Alzheimer's Disease: From Disease Pathogenesis to Therapeutic Potentials. Front Cell Neurosci. 2021 Oct 28;15.
- 138. Xi T, Jin F, Zhu Y, Wang J, Tang L, Wang Y, et al. MicroRNA-126-3p attenuates blood-brain barrier disruption, cerebral edema and neuronal injury following intracerebral hemorrhage by regulating PIK3R2 and Akt. Biochem Biophys Res Commun. 2017 Dec;494(1–2):144–51.
- 139. Chum PP, Bishara MA, Solis SR, Behringer EJ. Cerebrovascular miRNAs Track Early Development of Alzheimer's Disease and Target Molecular Markers of Angiogenesis and Blood Flow Regulation. Journal of Alzheimer's Disease. 2024 May 7;99(s2):S187–234.
- 140. Cha DJ, Mengel D, Mustapic M, Liu W, Selkoe DJ, Kapogiannis D, et al. miR-212 and miR-132 Are Downregulated in Neurally Derived Plasma Exosomes of Alzheimer's Patients. Front Neurosci. 2019;13.
- 141. Lugli G, Cohen AM, Bennett DA, Shah RC, Fields CJ, Hernandez AG, et al. Plasma Exosomal miRNAs in Persons with and without Alzheimer Disease: Altered Expression and Prospects for Biomarkers. PLoS One. 2015 Oct 1;10(10):e0139233.
- 142. Smith PY, Hernandez-Rapp J, Jolivette F, Lecours C, Bisht K, Goupil C, et al. miR-132/212 deficiency impairs tau metabolism and promotes pathological aggregation *in vivo*. Hum Mol Genet. 2015 Dec 1;24(23):6721–35.
- 143. Cogswell JP, Ward J, Taylor IA, Waters M, Shi Y, Cannon B, et al. Identification of miRNA Changes in Alzheimer's Disease Brain and CSF Yields Putative Biomarkers and Insights into Disease Pathways. Journal of Alzheimer's Disease. 2008 May 9;14(1):27–41.

- 144. Wang L, Zhen H, Sun Y, Rong S, Li B, Song Z, et al. Plasma Exo-miRNAs Correlated with AD-Related Factors of Chinese Individuals Involved in A β Accumulation and Cognition Decline. Mol Neurobiol. 2022 Nov 30;59(11):6790–804.
- 145. Fu M, Hu Y, Lan T, Guan KL, Luo T, Luo M. The Hippo signalling pathway and its implications in human health and diseases. Vol. 7, Signal Transduction and Targeted Therapy. 2022.
- 146. Aharon A, Spector P, Ahmad RS, Horrany N, Sabbach A, Brenner B, et al. Extracellular Vesicles of Alzheimer's Disease Patients as a Biomarker for Disease Progression. Mol Neurobiol. 2020 Oct 17;57(10):4156–69.
- 147. LIU CG, SONG J, ZHANG YQ, WANG PC. MicroRNA-193b is a regulator of amyloid precursor protein in the blood and cerebrospinal fluid derived exosomal microRNA-193b is a biomarker of Alzheimer's disease. Mol Med Rep. 2014 Nov;10(5):2395–400.
- 148. Lugli G, Cohen AM, Bennett DA, Shah RC, Fields CJ, Hernandez AG, et al. Plasma Exosomal miRNAs in Persons with and without Alzheimer Disease: Altered Expression and Prospects for Biomarkers. PLoS One. 2015 Oct 1;10(10):e0139233.
- 149. Wang L, Zhen H, Sun Y, Rong S, Li B, Song Z, et al. Plasma Exo-miRNAs Correlated with AD-Related Factors of Chinese Individuals Involved in A β Accumulation and Cognition Decline. Mol Neurobiol. 2022 Nov 30;59(11):6790–804.
- 150. Wei H, Xu Y, Xu W, Zhou Q, Chen Q, Yang M, et al. Serum Exosomal miR-223 Serves as a Potential Diagnostic and Prognostic Biomarker for Dementia. Neuroscience. 2018 May;379:167–76.
- 151. Visconte C, Fenoglio C, Serpente M, Muti P, Sacconi A, Rigoni M, et al. Altered Extracellular Vesicle miRNA Profile in Prodromal Alzheimer's Disease. Int J Mol Sci. 2023 Sep 29;24(19):14749.
- 152. Gámez-Valero A, Campdelacreu J, Vilas D, Ispierto L, Reñé R, Álvarez R, et al. Exploratory study on microRNA profiles from plasma-derived extracellular vesicles in Alzheimer's disease and dementia with Lewy bodies. Transl Neurodegener. 2019 Dec 4;8(1):31.
- 153. Liu T, Li N, Pu J, Zhang C, Xu K, Wang W, et al. The plasma derived exosomal miRNA-483-5p/502-5p serve as potential MCI biomarkers in aging. Exp Gerontol. 2024 Feb;186:112355.
- 154. Duan X, Zheng Q, Liang L, Zhou L. Serum Exosomal miRNA-125b and miRNA-451a are Potential Diagnostic Biomarker for Alzheimer's Diseases. Degener Neurol Neuromuscul Dis. 2024 Apr; Volume 14:21–31.
- 155. Cheng L, Doecke JD, Sharples RA, Villemagne VL, Fowler CJ, Rembach A, et al. Prognostic serum miRNA biomarkers associated with Alzheimer's disease shows concordance with neuropsychological and neuroimaging assessment. Mol Psychiatry. 2015 Oct 28;20(10):1188–96.
- 156. Kumar S, Reddy PH. Elevated levels of MicroRNA-455-3p in the cerebrospinal fluid of Alzheimer's patients: A potential biomarker for Alzheimer's disease. Biochimica et Biophysica Acta (BBA) Molecular Basis of Disease. 2021 Apr;1867(4):166052.
- 157. Li Y, Meng S, Di W, Xia M, Dong L, Zhao Y, et al. Amyloid-β protein and MicroRNA-384 in NCAM-Labeled exosomes from peripheral blood are potential diagnostic markers for Alzheimer's disease. CNS Neurosci Ther. 2022 Jul 26;28(7):1093–107.
- 158. Kumar A, Su Y, Sharma M, Singh S, Kim S, Peavey JJ, et al. MicroRNA expression in extracellular vesicles as a novel blood-based biomarker for Alzheimer's disease. Alzheimer's & Dementia. 2023 Nov 18;19(11):4952–66.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.