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Article

Interactions of *Opuntia ficus-indica* with *Dactylopius coccus* and *D. opuntiae* (Hemiptera: Dactylopiidae) through the Study of Their Volatile Compounds

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Abstract: *Opuntia ficus-indica* has always interacted with many phytophagous insects at its center of origin; two of them are *Dactylopius coccus* and *D. opuntiae*. *D. coccus*, or true cochineal, is produced to extract carminic acid, and *D. opuntiae* or wild cochineal is a key invasive pest of *O. ficus-indica* in more than 20 countries around the world. Despite the economic and environmental relevance of this cactus and, *D. opuntiae* and *D. coccus*, there are few studies that have explored volatile organic compounds (VOCs) derived from the plant-insect interaction. The aim of this work was to determine VOCs of *D. coccus* and *D. opuntiae* feeding on cultivars suitable for their development and to identify different VOCs in cladodes infested by each *Dactylopius* species. Volatiles were identified through their essential oils, which were obtained by hydrodistillation. A total of 66 VOCs from both *Dactylopius* species were identified, and 125 from the Esmeralda and Rojo Pelón cultivars infested by *D. coccus* and *D. opuntiae*, respectively, were determined. Differential VOCs production due to infestation by each *Dactylopius* species was also found. Some changes in methyl salicylate, terpenes such as linalool or the alcohol *p*-vinylguaiaicol were related to *Dactylopius* feeding on cladodes of their respective cultivars. Changes in these VOCs and their probable role in plant defense mechanisms should receive more attention because this knowledge could improve *D. coccus* rearing, or to its inclusion in breeding programs for *D. opuntiae* control in regions where it is a key pest of *O. ficus-indica*.

Keywords: cactus pear; phytophagous insects; terpenes; methyl salicylate; *p*-vinylguaiaicol

1. Introduction

Dactylopiidae, or cochineals, is a family of scale insects that includes only the genus *Dactylopius* and 11 recognized species [1] that are endemic to North and South America [2,3]. An important characteristic of these insects is that they produce carminic acid, probably as a defense mechanism against predation [4–6]. All the species of the genus are considered obligate parasites of Cactacea with high food specificity, particularly for the genera *Nopalea* Salm-Dyck and *Opuntia* Miller [7].

Because of the high carminic acid concentration (~20–25%) of *Dactylopius coccus* Costa, the true cochineal, it is the only species of commercial interest for production. It is reared on *Opuntia ficus-indica* (L.) Miller, the cactus pear. Carminic acid is recognized as a natural dye with cosmetic, food,

pharmaceutical, textile and plastic applications [8]. In addition, it is currently used in biomedicine [9] and as a photosynthesizing pigment in solar cells [10]. In contrast, *Dactylopius opuntiae* Cockerell, or wild cochineal, whose carminic acid content is less than 5%, is not considered useful for obtaining this substance. Rather, it is considered the key pest of *O. ficus-indica* in commercial plantations in Mexico [11,12], where plant and insect are native [7,13]. Additionally, *D. opuntiae* is an invasive pest in at least 20 countries in America, Europe, Africa, and Asia [14–16] where *O. ficus-indica* was adopted or naturalized and became one of the most important cultivated cactus species in the world because of its economic, environmental, and ecological benefits [13,14,17,18].

From a scientific perspective, most of *D. coccus* research has focused on the basic biology of the species and the quest to understand the mechanisms of carminic acid production and its possible physiological or ecological functions [4,19,20]. On the other hand, research on *D. opuntiae* has focused on control tactics because it is a key pest of *O. ficus-indica* [14,15,21,22]. The different cultivars of *O. ficus-indica* used as hosts of both *Dactylopius* species are likely to have particular physical and chemical characteristics, as well as volatile organic compounds (VOCs) that influence the trophic plant-insect and plant-pest-natural enemy relationship, as has been shown in other models of tritrophic interactions where volatiles cause positive or negative responses in terms of attraction and establishment of insects of the same or different species [23,24].

Volatile organic compounds (VOCs) are synthesized as products of plant metabolism and they are emitted into the environment [25] in response to biotic complexes or abiotic stresses [23,26]. These VOCs and essential oils are released from the leaves, flowers, and fruits into the atmosphere and from the roots into the soil [27,28]. This set of volatiles, essential in the defense mechanisms of plants against herbivores or in interspecific communication [23,24,27] is called volatilome and its analysis, using direct and non-direct approaches based on gas chromatography-mass spectrometry (GC-MS), is generally called volatilomics. A field that is continuously growing with the development of analytical and data processing methods [29]. In this regard, some research has been carried out on VOCs of *O. ficus-indica* emanating from cladodes, flowers, fruits, and the oils of its seeds [30–35]. These studies concluded that VOCs composition is a function of the geographical area, species or cultivar, plant structure, state of development and season, among other factors. However, none of these relatively recent papers included interaction with any of the important *Dactylopius* species, nor did they relate the production of VOCs to insect infestation. To our knowledge, there is only one study that evaluated VOCs in *O. ficus-indica* cladodes uninfested and infested by *D. coccus* [36]. This study reported eight types of compounds in uninfested cladodes and nine in cladodes infested by the insect. Furthermore, no other work is known to have explored VOCs of either *Dactylopius* species.

Because plant VOCs play an important role in interactions between insects and other organisms, e.g., pathogens or predators and parasitoids [23,24,37], as well as in the plant's response to insect attacks [38], the objectives of this work were 1) to determine the VOCs of *D. coccus* and *D. opuntiae* feeding on *O. ficus-indica*, and 2) to establish the changes in the composition and proportion of VOCs in cladodes of *O. ficus-indica* uninfested and infested with *Dactylopius*. This information could contribute to understanding the variation between cultivars of both species of insects and to exploring the potential of the biological functions that these compounds play in interspecies communication.

2. Results

Through essential oils it was possible to recover and identify about 80 % and 90 % of the volatile organic compounds (VOCs) of *D. coccus* and *D. opuntiae*, respectively. The *Dactylopius* species had 20 VOCs in common. In addition, 12 and 34 VOCs were specifically produced by *D. coccus* and *D. opuntiae*, respectively (Figure 1). Thus, the volatilome of each species was 32 or 54 compounds, and the proportion of each compound varied greatly between species (Table 1). The VOCs belonged to eight chemical groups, of which three had the highest relative abundance. Carboxylic acids and their derivatives was the most important group, accounting for 59.28 % and 78.29 % of the VOCs abundance for *D. coccus* and *D. opuntiae*, respectively. The second group was alcohols only for *D. coccus* (12.15 %), and the third group was aldehydes with 5.8 % and 7.68 % of the relative abundance

for *D. coccus* and *D. opuntiae*, respectively. The alkanes recovered were less than 2.5 % for both species. The remaining four groups of recovered compounds (ether, terpenes, ketones, alkenes) were less than 0.55 % of relative abundance per group (Table 1).

As mentioned above, the number and abundance of volatiles in each group of compounds also varied greatly in each *Dactylopius* species. For example, in the carboxylic acids and their derivatives, tetradecanoic acid was the most abundant in both species but decanoic acid, lactic acid and dodecanoic acid presented greater relative abundance in *D. coccus*. On the other hand, for *D. opuntiae*, 2-ethylhexanoic acid and cis-5-dodecenoic acid were detected only in this species in greater relative abundance. Hexadecanoic acid, (Z,Z)-9,12-octadecadienoic acid, (Z)-9-octadecenoic acid and octadecanoic acid occurred in both species, but their abundance differed considerably between species; again, they were more abundant for *D. opuntiae* (Table 1)

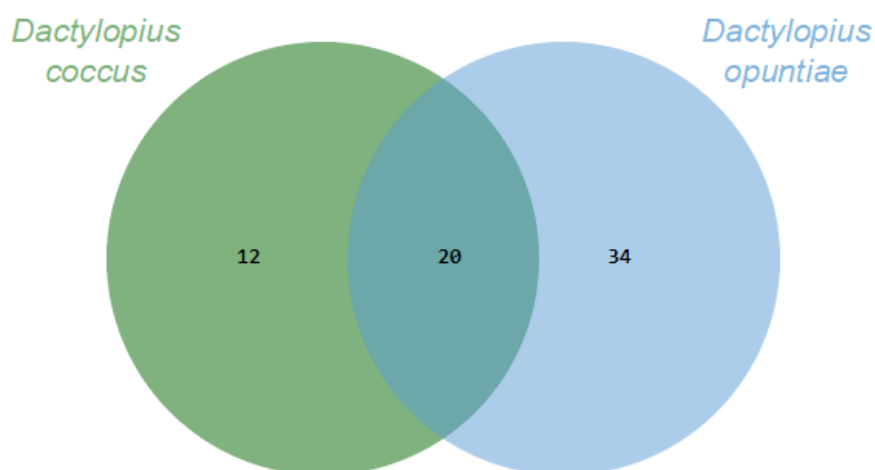


Figure 1. Comparison of the *Dactylopius* volatilomes using Venn diagrams based in the number of VOCs obtained through essential oils for each *Dactylopius* species.

Table 1. Volatile organic compounds (VOCs) obtained through essential oils of each *Dactylopius* species.

No.	Compounds	<i>D. coccus</i>		<i>D. opuntiae</i>		
		RA (%)	KI Exp	RA (%)	KI Exp	KI ref
	<i>Carboxylic acids and derivatives</i>	59.28%		78.29%		
1	Hexanoic acid			1.09±0.08		903
2	2-methylhexanoic acid			0.54±0.01		950
3	Heptanoic acid	0.20±0.05	1021	0.64±0.16	1021	1005
4	2,4-dimethylhexanoic acid			0.37±0.02		1015
5	2-Ethylhexanoic acid			10.90±0.21	1036	1031
6	Lactic acid	5.58±0.58	1061	0.86±0.03	1062	1057
7	Glycolic acid			0.11±0.03	1075	1072
8	2,6-dimethylheptanoic acid	0.61±0.21	1087			
9	Octanoic acid	1.90±0.74	1108	2.20±0.24	1108	1108
10	2,3,4-Trimethylpentanoic acid			0.44±0.02	1127	
11	Ethyl benzoate			0.12±0.01	1153	1141

12	Ethyl octanoate			0.05±1.77	1188	1175
13	Nonanoic acid	2.32±0.64	1214	3.57±0.37	1214	1205
14	2,4-dimethylnonanoic acid	0.31±0.01	1234			
15	Benzoic acid	2.04±0.00	1235	1.04±0.01	1235	1232
16	Ethyl nonanoate	0.26±0.03	1290			1282
17	2-Decenoic acid			0.42±0.02	1310	1290
18	Decanoic acid	8.76±2.53	1316	2.38±0.12	1316	1309
19	Butanedioic acid			0.11±0.03	1318	1314
20	(Z)-4- <i>tert</i> -butylcyclohexyl acetate	0.80±0.02	1358	0.12±0.03	1356	1346
21	Ethyl decanoate			0.18±0.02	1388	1382
22	Undecanoic acid			0.63±0.02	1414	1410
23	<i>cis</i> -5-Dodecenoic acid			3.18±0.04	1504	
24	Dodecanoic acid	4.93±0.90	1512			1509
25	Nonanedioic acid			0.40±0.03	1535	1511
26	Ethyl dodecanoate	1.06±0.07	1580			1566
27	Tridecanoic acid			0.49±0.04	1606	1606
28	<i>p</i> -Hydroxybenzoic acid	0.24±0.00	1615	0.05±0.01	1616	1621
29	Hexyl salicylate	1.05±0.01	1660	0.57±0.02	1658	1684
30	(Z)-9-Tetradecenoic acid	0.42±0.05	1702	0.22±0.09	1707	1691
31	Tetradecanoic acid	21.25±2.90	1717	30.15±1.35	1718	1713
32	Ethyl tetradecanoate			0.40±0.01	1793	1782
33	(Z)-9-Hexadecenoic acid			0.56±0.12	1909	1885
34	Hexadecanoic acid	2.30±1.83	1934	5.89±0.42	1934	1909
35	Ethyl hexadecanoate			0.23±0.01	1974	1968
36	Heptadecanoic acid			1.03±0.06	2039	2009
37	(Z,Z) 9,12-Octadecadienoic acid	0.83±0.73	2105	3.76±0.67	2105	2087
38	(Z) -9-Octadecenoic acid	0.82±1.11	2112	2.44±0.33	2112	2088
39	Octadecanoic acid	1.69±2.32	2139	2.58±0.22	2140	2133
40	Ethyl octadecanoate	0.33±0.03	2208			2181
41	Dehydroabiatic acid	1.58±0.00	2375	0.12±0.02	2376	2385

Aldehydes

5.80

7.68

42	Hexanal			1.79±0.36		964
43	Heptanal			1.24±0.11	1066	1069
44	Octanal			0.23±0.07	1165	1162
45	Nonanal	0.34±0.11	1235	2.57±0.17	1268	1267
46	Decanal			0.28±0.02	1367	1366
47	Dodecanal			0.28±0.10	1663	
48	α -Hexylcinnamaldehyde	5.46±0.08		0.11±0.12	1719	1728

49	Heptadecanal			1.18±0.33	2088	
	<i>Ether</i>			0.09		
50	Benzyl methyl ether			0.09±0.01	966	
	<i>Terpene</i>		0.54	0.08		
51	<i>p</i> -Cymene			0.08±0.04	1018	1025
52	α -Ionone	0.36±0.15	1415			1413
53	β -Ionone	0.18±0.02	1472			1486
	<i>Ketones</i>			0.54		
54	Benzophenone			0.09±0.02	1600	1611
55	2-Nonadecanone			0.45±0.08	2116	2087
	<i>Alcohols</i>		12.15	0.62		
56	Phenol			0.31±0.09	1045	1043
57	2-Ethylhexanol			0.22±0.04	1095	
58	1-Dodecanol	4.31±0.00	1559	0.09±0.01	1559	1575
59	1-Tridecanol	0.42±0.00	1659			1656
60	1-Tetradecanol	3.43±0.00	1765			1770
61	1-Hexadecanol	3.47±0.00	1977			1965
62	1-Octadecanol	0.52±0.00	2175			2159
	<i>Alkene</i>			0.42		
63	1-Tridecene			0.42±0.00	1284	1287
	<i>Alkane</i>		1.80	2.41		
64	Hexadecane	1.80±0.00	1581	1.11±0.29	1581	1600
65	Octadecane			0.25±0.00	1797	1800
66	Heneicosane			1.05±0.21	2309	
	Total		79.57	90.13		

RA Relative Abundance, KI Exp Kovats Index Experimental, KI Ref Kovats Index reference.

The Esmeralda and Rojo Pelón cultivars had VOCs production profiles that differed before and after *Dactylopius* infestation. In both cultivars, 28 VOCs were commonly produced and identified. In addition, 35 specific compounds were identified in Esmeralda and 19 in Rojo Pelón (Figure 2). After infestation by each *Dactylopius* species in the respective *O. ficus-indica* cultivar, a contrasting difference occurred between uninfested and infested cladodes of each cultivar (Table 2). The changes were not only in the number of VOCs, but in their abundance and variation. Sometimes they decreased, sometimes they increased, sometimes some VOCs were no longer detected, and of course there were also some *de novo* compounds (Table 2). After infestation by *D. coccus*, the Esmeralda cultivar increased the number of volatiles from 63 (uninfested) to 87, of which 48 were produced *de novo* and belonged to nine chemical groups. In the case of Rojo Pelón *D. opuntiae*, uninfested cladodes produced 47 VOCs and after infestation they decreased to 38, 13 of which were identified as *de novo*, belonging to seven chemical groups (Table 2, Figure S1 and Figure S2).

Although there was an enormous variation between the number and proportion of VOCs before and after infestation, it was observed that four chemical groups maintained the highest abundance in both infested cultivars. These groups were a) carboxylic acid and derivatives, b) terpenes, c) alcohols, and d) aldehydes and derivatives. Another group, the heterocycles, was only abundant for the uninfested Rojo Pelón cultivar (8.91 %), but after *D. opuntiae* infestation it decreased to less than 1.4 %. The rest of the recovered chemical groups (ethers, ketones, aromatic derivatives and alkanes) were less than 1.16 % of the relative abundance per group in either cultivar infested by the respective *Dactylopius* species. Two of these groups (ethers and aromatic derivatives) were not detected in the infested Rojo Pelón cultivar (Table 2).



Figure 2. Comparison of volatilomes of the *Opuntia ficus-indica* uninfested cladodes of each cultivar using Venn diagram, based on the number VOCs obtained through essential oils.

Table 2. Volatilomes of *Opuntia ficus-indica* (OFI) cultivars before and after infestation by *Dactylopius* species.

No.	Compounds	OFI		KIExp	OFI Rojo Pelón		KIExp	KIRef
		Esmeralda	Esmeralda infested by <i>D. coccus</i>		Rojo Pelón	Pelón infested by <i>D. opuntiae</i>		
		RA (%)	RA (%)		RA (%)	RA (%)		
	Carboxylic acid and derivatives	48.79	44.28		31.78	20.05		
1	Hexanoic acid	0.83±0.63	0.82±0.00	942				904
67	3-Methyl-2-pentenoic acid			959				926
		0.35±0.32						
68	2-Hexenoic acid	0.66±0.01	0.36±0.00	972				939
69	4-Oxopentanoic acid	0.22±0.14		991				956
70	Heptanoic acid	0.59±0.50	0.39±0.05	1022	0.39±0.11	0.25±0.15	1030	1005
5	2-Ethylhexanoic acid				0.11±0.07		1044	1031
71	4-Methylvaleric acid		0.15±0.10	1033				1039
72	2-Methyl-4-pentenoic acid			1062				
			0.11±0.16					
73	Lactic acid	3.22±1.91	1.63±0.16	1065				1057

15	Benzoic acid	1.49±1.77	0.29±0.03	1080	2.06±0.13	0.50±0.09	1080	1084
74	Methyl benzoate	0.41±0.14		1081		0.91±0.34		1084
9	Octanoic acid	1.23±1.52		1112	1.21±0.07	1.34±0.07	1112	1109
11	Ethyl Benzoate	0.05±0.01		1156	0.08±0.04	0.24±0.11	1152	1141
75	Benzeneacetic acid					0.17±0.05	1160	1150
76	Salicylic acid					0.18±0.25	1171	1176
77	Methyl salicylate	1.21±0.37	6.96±0.14	1181		0.32±0.07	1172	1176
78	2-Nonenoic acid		0.86±0.00	1179		0.33±0.08	1184	
13	Nonanoic acid	1.09±0.24	0.76±0.07	1216	1.45±0.12	1.72±0.14	1212	1206
79	Ethyl salicylate		1.03±0.01	1244				1241
18	Decanoic acid	0.77±0.75	0.63±0.00	1318	0.79±0.08		1311	1309
19	Butanedioic acid	0.49±0.19		1320				1314
80	Glyceric acid	0.73±0.02		1346				1342
81	2-Methoxybenzoic acid		0.10±0.01	1331				1362
82	Methyl 2-methoxy benzoate		0.45±0.10	1319				1295
83	Glutaric acid	0.16±0.03		1410				1400
22	Undecanoic acid	0.16±0.02	0.14±0.07	1417	0.08±0.02	0.12±0.08	1408	1410
24	Dodecanoic acid	5.70±6.08	2.77±0.06	1516	7.19±0.50	2.35±0.16	1505	1509
84	2,5-Dimethoxy benzenemethanol acetate				0.13±0.04		1523	
26	Ethyl Dodecanoate	0.26±1.27	0.17±0.09	1582	0.33±0.01		1571	1581
28	<i>p</i> -Hydroxybenzoic acid	0.94±0.45		1620				1621
29	Hexylsalicylate	0.25±3.04	0.46±0.07	1662				1652
85	Methyl tetradecanoate	0.84±0.16		1719				1714
27	Tridecanoic acid		0.06±0.04	1611				1606
86	12-Methyltridecanoic acid		0.07±0.00	1678				1680
31	Tetradecanoic acid	2.98±3.09	1.36±0.06	1720	1.56±0.10	0.19±0.07	1714	1714
87	Methyl benzoate		1.46±0.03	1752				
88	Benzyl Benzoate		2.65±0.51	1741	0.15±0.04		1754	1765
32	Ethyl tetradecanoate		0.10±0.13	1784				1782
89	Nonanedioic acid	0.28±0.01		1808				1788
90	Pentadecanoic acid	0.42±0.47	0.43±0.02	1826				1807
91	Isopropyl tetradecanoate		0.05±0.34	1820				1827
92	Benzyl salicylate		0.31±0.12	1855				1860
34	Hexadecanoic acid	7.19±2.16	4.88±0.16	1935	9.35±0.74	8.25±0.45	1916	1909
93	15-Methylhexadecanoic acid	0.17±0.03		2040				1974
37	(<i>Z,Z</i>)-9,12-Octadecadienoic acid	1.90±0.44	1.03±0.12	2106	0.88±0.14	2.20±1.43	2087	2087
38	(<i>Z</i>)-9-Octadecenoic acid	2.35±1.66	1.94±0.10	2113	2.47±0.98		2090	2100
39	Octadecanoic acid	2.53±0.59	1.76±0.01	2141	2.76±0.15	0.59±0.15	2119	2133

94	Methyl octadecanoate				0.33±0.04		1809	
40	Ethyl octadecanoate		0.11±0.00	2199				2202
41	Dehydroabietic acid	9.32±2.24	9.99±0.04	2376	0.46±0.05	0.39±0.02	2344	2373
<i>Aldehydes and derivatives</i>		2.15	6.25	4.3	4.82			
42	Hexanal	0.44±0.44	0.47±0.00	984				964
43	Heptanal		0.18±0.02	1069				1068
95	Benzaldehyde		0.15±0.07	1094	0.32±0.03	0.55±0.33	1094	1080
96	Diethyl acetal hexanal	0.25±0.14	0.46±0.10	1086				1082
97	5,5-Dimethyl-3-oxo-1-cyclohexene-1-carboxaldehyde		0.15±0.03	1104				
44	Octanal		0.17±0.01	1160	0.40±0.05	0.09±0.05	1165	1167
98	Phenylacetaldehyde	0.62±0.61	0.56±0.06	1198	0.82±0.06	0.81±0.17	1201	1208
45	Nonanal	0.53±0.21	1.03±0.01	1271	1.73±0.12	1.32±0.09	1265	1267
46	Decanal		0.14±0.00	1370	0.14±0.08		1366	1366
99	Nonanaldimethylacetal	0.21±0.10	0.37±0.05	1374				1379
100	3-(4-(tert-butyl)phenyl-2-methylpropanal		0.30±0.04	1497				1500
101	4-Hydroxy-3-methoxybenzaldehyde	0.10±0.04	0.62±0.02	1524	0.89±0.04	2.05±0.01	1511	1544
102	3-Ethoxy-4-hydroxybenzaldehyde		0.11±0.02	1554				1560
48	α-Hexylcinnamaldehyde		1.22±0.74	1725				1726
103	Octadecanal		0.32±0.13	2187				
<i>Heterocycles</i>			0.67		8.91	1.38		
104	2-Isopropyl-3-metoxypirazina		0.25±3.25	1070				1080
105	2-Methoxy-3-isopropylpyrazine				0.30±7.22	1.05±0.33	1083	1089
106	Ethyl 2-(5-methyl-5-vinyltetrahydrofuran-2-yl)propan-2-yl carbonate				8.18±0.10		1064	1090
107	3-Isobutyl-2-methoxypyrazine				0.43±0.03		1164	1170
108	3-Ethyl-4-methyl-1H-pyrrole-2,5-dione		0.35±0.03	1209				1192
109	3-Hydroxy-2-methylpyran-4-one		0.07±0.00	1266				1293

110	2,3-Dihydro-2,2,4,6-tetramethylbenzofuran			0.33±0.01		1410	
	<i>Ethers</i>	0.31	0.16				
50	Benzylmethylether	0.31±0.20			966		966
111	1,2-Dimethoxybenzene		0.16±0.02		1111		1106
	<i>Ketones</i>	1.95	1.16		1.17	0.61	
112	5-Hexen-2-one	0.29±0.01			1007		
113	2,2,6-Trimethylcyclohexanone				0.14±0.09		1031
114	Acetophenone		0.57±0.08		1047	0.45±0.05	1055 1049
115	Isophorone	0.24±0.06	0.01±0.22		1106	0.17±0.03	1038 1094
116	Phenylacetone	0.48±0.05	0.44±0.00		1114	0.24±0.02	0.16±0.07 1110 1116
117	4-Oxoisophorone	0.13±0.02			1131	0.07±0.04	1125 1105
118	2-(1-Hydroxybut-2-enylidene)cyclohexanone					0.14±0.03	1145
119	1-(1-cyclohexen-1-yl)(-1-1-Butenone)	0.70±0.34			1214		
54	Benzophenone	0.11±0.07	0.14±0.01		1607	0.41±0.05	1584 1607
	<i>Terpenes</i>	17.89	13.92		0.8	15.52	
120	Limonene	0.85±0.23			1023		1020
121	Linalool oxide	8.48±0.58	5.06±0.42		1063		1064
122	<i>trans</i> -Linalool oxide		5.70±0.03		1064		1068
123	1,5,5-Trimethyl-3-methylene cyclohexene	0.55±0.63			1071		
124	β-Linalool	5.00±0.58	0.26±0.35		1088		1082
125	α-Terpineol	2.00±0.36			1178		1172
126	Linalool	0.19±0.22			1232	5.61±0.10	1227 1227
127	Geraniol	0.44±0.27	0.33±0.04		1250	1.84±0.06	1357 1238
128	Nerol		0.33±0.06		1232	0.79±0.04	1328 1260
129	β-Damascenone		0.18±0.17		1362	0.55±0.30	1360 1361
52	α-Ionona		0.09±0.26		1404		1413
130	α-Isomethylionone		0.88±0.15		1453		1478
53	β-Ionone		0.40±0.06		1460	0.18±0.04	1458 1486
131	Dihydroactinidiolide	0.38±0.13	0.30±0.02		1537		1532
132	Neophytadiene		0.39±0.08		1832		1842
133	28-Nor-17β(H)-hopane					0.45±0.01	2942
134	β-Sitosterol					0.35±0.03	6.55±0.04 3244 3284

	<i>Alcohols</i>	12.72	9.91	29.37	30.78		
135	1,2-Dihydroxy-4-methylpentane		0.27±0.02	990			
136	Hexanol		0.06±0.01	9994			992
137	(Z)-2-Hexen-1-ol		0.36±0.19	1010	7.65±0.18	4.21±0.08	1025 1001
56	Phenol	0.22±0.09	0.16±1.30	1045			1043
138	Heptanol		0.11±0.23	1067			1092
57	2-Ethylhexanol	0.58±0.23	0.22±0.24	1099	2.13±0.09	1.49±0.05	1103
139	Benzylalcohol	0.27±0.11	0.56±0.22	1143	0.90±0.02	2.08±0.06	1132 1156
140	1-Octanol		0.29±0.14	1158	1.27±0.04	1.51±0.18	1177 1177
141	Guaiacol		0.35±0.03	1209			1192
142	Nonanol		0.07±0.01				
143	Glycerol	0.33±0.52		1290	1.46±0.09		1288 1292
144	<i>p</i> -Vinylguaiacol	10.62±7.24	1.76±0.34	1305	14.67±0.93	17.03±5.14	1294 1282
145	1-Methyl-1(4-methyl-3-cyclohexenyl)ethanol		0.63±0.00	1318			1309
146	Isododecanol		0.09±0.01	1479			
58	1-Dodecanol	0.47±0.09	0.60±0.01	1563	0.76±0.04		1553 1575
60	1-Tetradecanol		0.66±0.00	1756			1768
61	1-Hexadecanol	0.23±0.07	0.98±0.08	1978	0.53±0.03		1960 1965
62	1-Octadecanol		0.92±0.04	2177			2159
147	3,7,11,15-Tetramethyl-2-hexadecenol		1.82±0.15	2198		3.52±0.17	2173 2179
148	Octacosanol					0.94±0.04	3125 3154
	<i>Aromatic derivatives</i>	0.77					
149	1,2-Dihydro-1,1,6-trimethyl naphthalene	0.18±0.07		1338			1332
150	10,18,Bisnorabieta-8,11,13.triene	0.59±0.03		2041			
	<i>Alkanes</i>	0.69	0.55	0.78			
64	Hexadecane	0.69±0.19	0.07±0.01	1585			1600
151	Heptadecane		0.12±0.05	1692			1700
152	Nonadecane		0.36±0.06	1906			1900
153	Eicosane				0.19±2.08		1992 2000
66	Heneicosane				0.23±0.42		2092 2100
154	Docosane				0.36±0.07		2188 2200
Total		85.27	76.9	77.11	73.16		

RA Relative Abundance, KI Exp Kovats Index Experimental, KI Ref Kovats Index reference.

As indicated above, because of *Dactylopius* infestation in each cultivar, there were many changes in the relative abundance of compounds and production of some de novo compounds. The *de novo* compounds were mostly of low relative abundance (equal to or less than 1.0 %), except for some terpenes and alcohols. For example, in the uninfested Rojo Pelón cultivar the relative abundance of terpenes was around 0.8 %, but this relative abundance of terpenes changed to 15.5 % after *D. opuntiae* infestation. On the other hand, the relative abundance of terpenes in the Esmeralda cultivar decreased from 18 to 13.9 % due to *D. coccus* infestation (Table 2). The amount and type of terpenes were different between infested *O. ficus-indica* cultivars, but monoterpenes or their derivatives predominated in both cases (Figure 3).

The terpenes Linalool oxide, trans-Linalool oxide and the alcohol 3,7,11,15-Tetramethyl-2-hexadecenol reached a relative abundance of 5.06 %, 5.7 % and 1.8 % in the Esmeralda cultivar infested by *D. coccus*. On the other hand, the terpenes Linalool, Geraniol and the alcohol 3,7,11,15-Tetramethyl-2-hexadecenol registered 5.6 %, 1.84 % and 3.5 % of the relative abundance in the Rojo Pelón cultivar infested by *D. opuntiae*, respectively. Also, *p*-Vinylguaiacol increased 2.3 % in relative abundance after *D. opuntiae* infestations (Table 2).

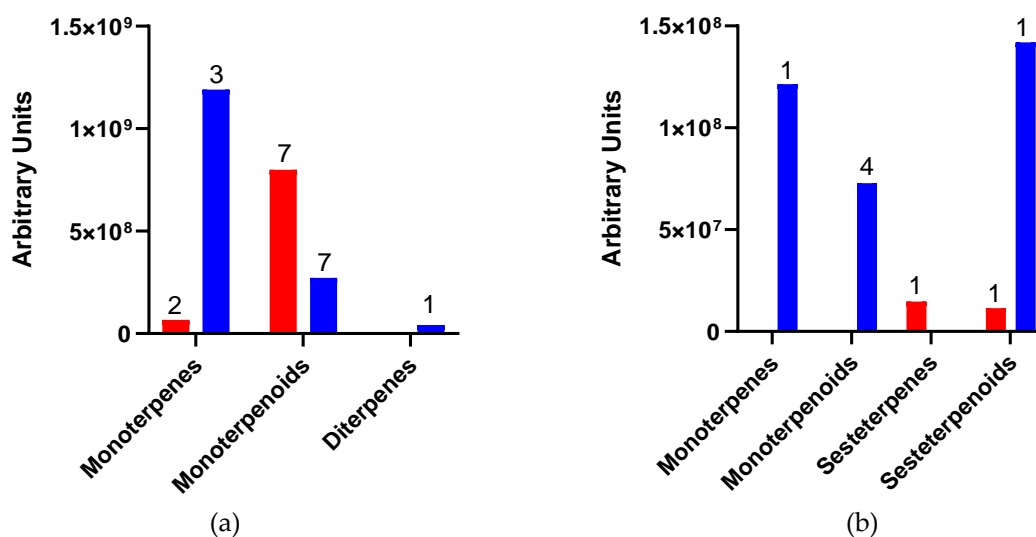


Figure 3. Amount and type of terpenes released by *Opuntia ficus-indica* (OFI) after *Dactylopius* infestation. (a) OFI Esmeralda-*D. coccus*; (b) OFI Rojo Pelón-*D. opuntiae*. The red columns represent uninfested cladodes, and blue columns represent cladodes infested by each *Dactylopius* species. Data are presented as means of the peak area of each terpene (grouped by type and number of compounds).

3. Discussion

Previous assays of *Dactylopius* VOCs extraction such as Headspace (HS-SPME) and extraction by Autosampler Headspace coupled to CG-MS (HS-CG-MS) did not provide the results expected for GC-MS analysis. Thus, to identify the volatiles from *Dactylopius* and its cultivar hosts, we preferred to do so using their essential oils. Essential oils were obtained by the hydrodistillation method (Table S1), which is frequently used to obtain essential oils from plants that contain low vapor pressure compounds, or low volatile compounds. This technique is also used for concentrating compounds with lower concentrations in the essential oil and allows working with a larger sample mass than microextraction techniques, which can potentially improve characterization of insect VOCs [39].

In the volatilome of *D. coccus* and *D. opuntiae*, 32 and 54 VOCs were identified for each species, respectively. To our knowledge, neither of these volatilomes had been reported previously, and this may be the first contribution of this work. By their composition, these VOCs corresponded to eight different chemical groups, but there were three groups of greater abundance. These were a) carboxylic acids and their derivatives, 59.28% and 78.29% abundance for *D. coccus* and *D. opuntiae*, respectively, b) alcohols, which were abundant only for *D. coccus* (12%), and c) aldehydes, 5.8% and

7.68% abundance for *D. coccus* and *D. opuntiae*, respectively (Figure S3 and Figure S4). This composition could be one of the reasons that results were not obtained with the HS-SPME and HS-CG-MS techniques. The VOCs of *Dactylopius* species are mostly fatty acids, some of which may be part of the fat content of the insects, or of the complexity of their waxy coat [40,41]. In fact, each VOC in those groups may have more than one role in structure, function, metabolism, and probably in intra or interspecific communication. For example, *D. coccus* produces a sex pheromone [42], and *D. opuntiae* is suspected to do so as well [43]. Regarding tetradecanoic acid, which is one of the most abundant VOCs for both species of *Dactylopius*, and hexadecanoic acid, relevant to *D. opuntiae*, they have many functions in insect metabolism. One of these is to participate in metabolic pathways of sex pheromones of some *Lepidoptera* such as *Spodoptera littoralis* Boisduval and *Plodia interpunctella* Hubner [44,45], but none of these compounds appear to have relevance in the pheromones of Coccoidea [46], which is the superfamily to which the Dactylopiidae belong. The methodology for identifying insect pheromones begins with live females at a particular moment of maturity and sexual behavior, and so much work remains to be done to decipher the main functions of the VOCs that turned out to be more abundant, and that could lead to novel acids with shorter chains and perhaps more specific for each *Dactylopius* species.

The volatilome of Esmeralda and Rojo Pelón cultivars were different before and after *Dactylopius* infestation. The variation in compound production in cladodes of both cultivars prior to infestation (by *Dactylopius*) may be specific to each cultivar, as variations of other bioactive and volatile compounds have been reported in different cultivars of *O. ficus-indica* [30,34,47]. However, variation in the number and abundance of VOCs within each cultivar after infestation can be attributed to *D. coccus* or *D. opuntiae* feeding on its corresponding cultivar host, as has been demonstrated in other plants where the change in production of VOCs, particularly terpenes and sesquiterpenes, was directly associated with phytophagous insect feeding [23,24,48,49].

In the volatilome of Esmeralda and Rojo Pelón, before or after *Dactylopius* infestation, four chemical groups were identified as the most abundant: a) carboxylic acid and derivatives, b) terpenes, c) alcohols, and d) aldehydes and derivatives (Figure S5 and Figure S6). The structural composition of the host, particularly the quantity of waxes, could be related to the abundance of some of these VOCs in both cultivars [47,50]. This suggestion is related to the anatomical and physiological adaptations of cacti to develop in arid environments, such as a thick and impermeable epidermis covered by a layer of waxy cuticle, a hypodermis with chlorenchyma, plenty of cells with mucilage distributed in the parenchyma, and crassulaceae acid metabolism (CAM) among other characteristics [50]. Of the first and most abundant chemical groups (a), it is probable that we should mention the methyl salicylate, which increased in abundance after infestation by *D. coccus* (about 5 %) in the Esmeralda cultivar. The same compound was identified *de novo* in the Rojo Pelón cultivar infested by *D. opuntiae*, although low in abundance (0.3 %). Methyl salicylate is a phenolic compound that has been reported to be an Herbivore-Induced Plant Volatile (=HIPV) [49,51,52]. Some of these HIPVs can induce direct defense against the phytophagous insect and indirect defense by attracting their natural enemies. It is also useful in communication among plants damaged by phytophagy and others that are not yet damaged. For example, methyl salicylate emitted by plants with phytophagous mite damage was attractive to *Phytoseiulus persimilis* Athias-Henriot (Phytoseiidae) [51,52]. In the same way, it was observed that emission of this compound, after damage by psyllids in pear trees, was attractive to the predatory bug *Anthocoris nemoralis* F. (Hemiptera: Anthocoridae) [53].

In general, a slight decrease in terpene abundance (18 to 14%) was observed after *D. coccus* infestation, but a considerable increase (0.8 to 15%) occurred after *D. opuntiae* infestation. In the Esmeralda cultivar, β -Linalool abundance decreased from 5.0 to 0.3 %, but Linalool oxide and trans-Linalool oxide increased to 5.0 and 5.7 %. On the other hand, in the cultivar Rojo Pelón infested by *D. opuntiae*, five *de novo* terpenes were identified, of which the most abundant was Linalool (5.6%). Terpenes are one of the most studied groups of HIPVs, and it has been shown that some of them have a relevant role in the direct defense system against phytophages, and some volatile terpenes constitute indirect defenses of plants as they attract natural enemies such as predators and parasitoids [23,24,27,48,49].

Linalool is a monoterpene that occurs naturally in flowers and aromatic plants, but it is also produced in response to feeding by phytophagous insects and it is part of the indirect defenses of plants [54]. For example, an increase in linalool production in tobacco plants caused by feeding Lepidoptera larvae increased the rate of egg predation and decreased the oviposition of another Lepidoptera [55]. Linalool also increased due to phytophages feeding on corn, bean, cotton, and potato plants [23], or by a zoophytophagous mirid feeding on pepper plants, and favored the action of natural enemies of their pests [49]. This can suggest that significant changes in the abundance of methyl salicylate from the above group and terpenes, particularly linalool, are probably related to each *Dactylopius* species feeding on its corresponding cultivar host.

The alcohol of greatest abundance and change was *p*-vinylguaiaicol. This compound is common in plants and is part of many essential oils. In addition, it can be found in the gut of some insects probably by the process of lignin degradation [56]. Regarding secondary plant defenses due to damage by phytophagous insects, *p*-vinylguaiaicol stimulated the ovipositional behavior of the natural enemy *Coleomegilla maculata* [57], and it was also a deterrent to the oviposition of the cerambycid *Monochamus alternatus* [56]. Therefore, it is suggested that some changes in *p*-vinylguaiaicol abundance may be a consequence of *Dactylopius* feeding.

In this work, 66 VOCs of both *Dactylopius* species were identified, and 125 of the Esmeralda and Rojo Pelón cladodes infested by *D. coccus* and *D. opuntiae*, respectively. A proportion of VOCs were commonly produced in both insect species or cultivars, but others were specific to each species or cultivar (Figure 4). This is a first approach to the diversity of VOCs produced by *O. ficus-indica*, and the changes that occur due to *D. coccus* and *D. opuntiae* feeding on cultivars suitable for the development of each *Dactylopius* species. More time and work will now be needed to understand the functions performed by the most relevant compounds in these interactions.

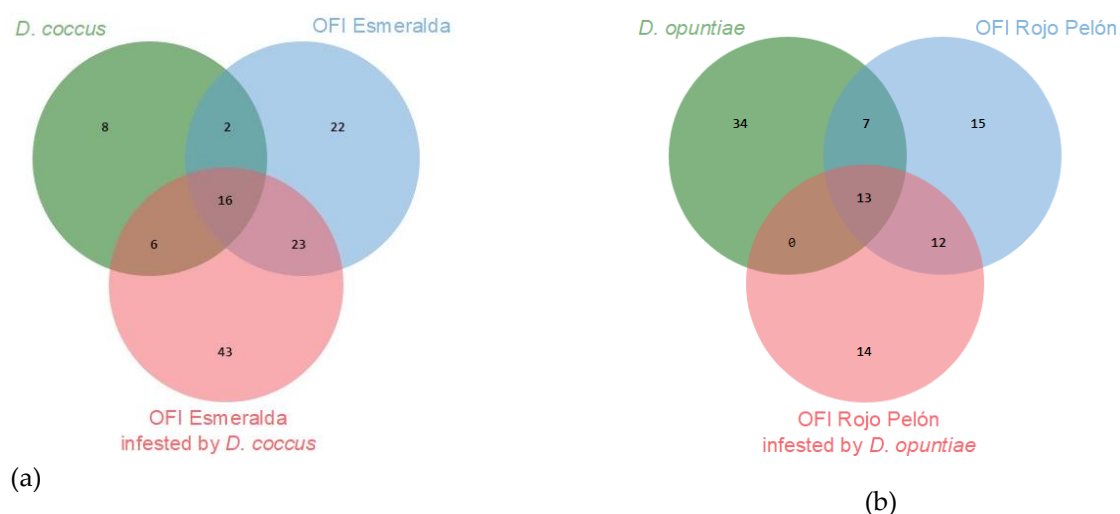


Figure 4. Number of VOCs, obtained through its essential oils, identified as common or de novo compounds between uninfested and infested *O. ficus-indica* (OFI) cultivars by each *Dactylopius* species. Also, identified de novo VOCs from *Dactylopius* species.

If knowledge of the interaction is improved, for example, if it is confirmed that some terpenoids favor the direct or indirect defenses of *O. ficus-indica* against *D. coccus* or *D. opuntiae*, this information could be considered in breeding programs. These programs could be aimed to improve rearing of *D. coccus*, or to induce resistance to *D. opuntiae*. In this regard, breeding programs for *O. ficus-indica* resistant to *D. opuntiae* have already been developed in Brazil and Morocco, and these have focused on physical and biochemical defense mechanisms [15,21,58]. For example, selecting cultivars with high concentrations of calcium oxalates can physically and biochemically limit phytophagous insects [59,60]. However, there are no known breeding programs for *O. ficus-indica* that consider the abundance of terpenes in cultivars, and the response this can trigger in the plant's direct or indirect defenses. This mechanism would be classified as biochemical defense, and measuring terpenes in

different cultivars could improve the direction and understanding of the response. This observation highlights the need to better understand the interaction between *O. ficus-indica* and *Dactylopius*, because it can increase the possibilities of making proposals for sustainable management in the production of *D. coccus*, or in the control of *D. opuntiae*.

4. Materials and Methods

4.1. Chemicals

The reagents used in this study were *N*, *O*-bis(trimethylsilyl) trifluoroacetamide (BSTFA), trimethylsilyl chloride (TMCS), boron trifluoride methanol solution (Sigma-Aldrich, St Louis, MO, USA) ethylic ether (JT Baker, Deventer, Holland).

4.2. Insects and Uninfested and Infested *O. ficus-indica* Cultivars

Dactylopius coccus and *Opuntia ficus-indica* Esmeralda cultivar (infested and uninfested) were originally obtained from a local provider in Jerez, Zacatecas, Mexico. *Dactylopius opuntiae* and *O. ficus-indica* Rojo Pelón cultivar (infested and uninfested) were collected from an experimental field of Colegio de Postgraduados, Campus San Luis Potosí (Salinas, SLP). These cactus pear cultivars were selected with the knowledge that each one is favorable for the development of the respective *Dactylopius* species [58]. The taxonomic identity of *Dactylopius* species was corroborated by S. J. Méndez-Gallegos using De Lotto (1974) [40] and Ferris (1955) keys [61]. To increase material for the samples and analyses, *D. coccus* and *D. opuntiae* colonies were reared on the respective cultivars mentioned under greenhouse conditions (15 ± 2 °C, 22 ± 2 °C and 50 % RH).

4.3. Essential oil of *Dactylopius* Species and Hosts

One hundred grams of adult female *D. coccus* and *D. opuntiae* and 1000 g of infested and uninfested *O. ficus-indica* cladodes were hydrodistilled at the temperature of boiling water. The volatile organic compounds (VOCs) were extracted from the condensed water by liquid-liquid extraction with ethyl ether. The solvent was distilled, and the residual water was removed from organic phase with anhydrous sodium sulfate. The sample was then concentrated (to 1 mL) at 40 °C under vacuum, and the solvent was eliminated at atmospheric pressure at 0 °C.

4.4. Derivatization for Alcohol Detection

Essential oils were diluted to 2 % in 500 μ L heptane and introduced into a 10 mL microwave reaction tube with gasket. Then, 100 μ L of BSTFA/TMCS solution (9:1 v/v) was added to the same tube as a silanizing agent. The mixture was reacted at 90 °C under microwave irradiation (250W microwave power) for 10 min using the Discover System 908,005 (CEM Corporation, NC, USA) with autogenous pressure.

4.5. Derivatization for Aldehydes and Carboxylic Acid Detection

Essential oils were diluted to 2 % in 500 μ L heptane and introduced into a 10 mL microwave reaction tube with gasket. Then, 500 μ L of boron trifluoride 14 % in methanol solution was added to the same tube. The mixture was reacted at 90 °C under microwave irradiation (250 W microwave power) for 10 min using the Discover System 908005 with autogenous pressure.

4.6. Essential Oil GS-MS Analysis

Samples without derivatization were diluted to 2 % in heptane, using 1 μ L of each sample for the analysis, and each sample was analyzed in triplicate. GC-MS analysis was performed using a 7802A Network GC System coupled to a 5977E Network mass selective detector (MSD).

The separation was performed using an HP-5 capillary column (0.25 mm i.d. 30 mm, 0.25 mm film thickness) (J&W, Folsom, CA, USA). The injector was operated in splitless mode at 300 °C, with a flow of 1.0 mL/min, and the oven temperature was programmed to 40 °C for 3 min, and then heated

at 3 °C/min to 300 °C with a holding time of 5 min at the final temperature. The MSD was operated at 70 eV; the ion source was set at 150 °C and the transfer line at 300 °C. VOCs were identified by interpreting their mass spectra fragmentation in the mass range of 15 to 800 atomic mass units. The software MassHunter (Agilent B.07.01.1805) was used for data recording. The compounds were identified by comparing the obtained mass spectra with those of reference compounds from the National Institute of Standards and Technology (NIST11) and Wiley 09. The identities of the compounds were confirmed by the Kovats retention index calculated for each peak with reference to the n-alkane standards (C7–C38) running under the same conditions.

4.7. Statistical Analysis

The relative percentage of each metabolite was calculated considering the peak area obtained by GC-MS of each metabolite in relation to the total area of peaks analyzed. Data represent the mean of the relative percentage of three repeats \pm SD. Metabolites grouped by type for each essential oil were compared with the Mann Whitney U test considering the peak area of each metabolite and a $p \leq 0.05$. The data in the graphics were expressed as median and range of each group. GraphPad Prism 5 was used to perform the analysis. Venn diagrams were constructed using an online tool (<http://jvenn.toulouse.inra.fr/app/example.html> accessed on 23 Nov 2023) [62].

5. Conclusions

This work presents an approach to better understand the interaction between *O. ficus-indica*, *D. coccus* and *D. opuntiae* through identifying volatile compounds in their essential oils. The abundance and proportion of VOCs of *D. coccus* and *D. opuntiae* were determined in the Esmeralda and Rojo Pelón cultivars, viable for the development of each insect species, respectively. Differential VOC production due to infestation by each *Dactylopius* species in each cultivar was also identified. Changes in methyl salicylate, terpenes and *p*-vinylguaiaicol and their likely role in plant defense mechanisms should receive more attention because they could contribute to the development of proposals to improve *D. coccus* rearing, or for the control of *D. opuntiae* in those regions in the world where it is a key pest of *O. ficus-indica*.

Supplementary Materials: The following supporting information can be downloaded at the website of this paper posted on Preprints.org. Table S1 show the yields of essential oils of *Dactylopius* species and *Opuntias* varieties. Figures S1 y S2 show *O. ficus-indica* with *Dactylopius* species relationships by Venn diagrams, Figures S3-S6 show compounds group of *Dactylopius* species and *O. ficus-indica* cultivars.

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Data Availability Statement: The datasets used and analyzed during the current study are available from the corresponding author upon reasonable request.

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Conflicts of Interest: The authors declare no conflicts of interest.

References

1. García-Morales M., Denno B.D., Miller D.R., Miller G.L., Ben-Dov Y., Hardy N.B. ScaleNet: a literature-based model of scale insect biology and systematics. *Database (Oxford)* **2016**, 1-5. DOI: 10.1093/database/bav118

2. Van-Dam A.R., Martínez L.P., Chavez A.J., May B.P. Range wide phylogeography of *Dactylopius coccus* (Hemiptera: Dactylopiidae). *Ann Entomol Soc Am.* **2015**, *108*(3), 299–310. DOI: 10.1093/aesa/sav017.
3. Campana M.G., Robles N.M., Tuross N. America's red gold: Multiple lineages of cultivated cochineal in Mexico. *Ecol Evol.* **2015**, *5*(3), 607–17. DOI: 10.1002/ece3.1398.
4. Barreto-García O.A., Rodríguez-Leyva E., Lomeli-Flores J.R., Vanegas-Rico J.M., Viguera A.L., Portillo L. *Laetilia coccidivora* feeding on two cochineal insect species, Does the prey affect the fitness of the predator? *BioControl* **2020**, *65*(6), 727–36. DOI.org/10.1007/s10526-020-10047-6.
5. Eisner T., Nowicki S., Goetz M., Meinwald J. Red Cochineal Dye (Caminic Acid): Its Role in Nature. *Science* **1980**, *208*(30), 1039–1042. DOI: 10.1126/science.208.4447.1039.
6. Martínez-Martínez S., Rodríguez-Leyva E., Aranda-Ocampo S., Santillán-Galicia M.T., Hernández-López A., Guzmán-Franco A.W. Bacteria associated with carminic acid metabolism in the intestinal tract of three predators of *Dactylopius opuntiae*. *Entomol Exp Appl.* **2023**, *172*, 183-192. DOI: 10.1111/eea.13394.
7. Chávez-Moreno C.K., Tecante A., Casas A. The *Opuntia* (Cactaceae) and *Dactylopius* (Hemiptera: Dactylopiidae) in Mexico: A historical perspective of use, interaction and distribution. *Biodiversity and Conservation* **2009**, *18*, 3337–3355. DOI 10.1007/s10531-009-9647-x.
8. Borges M.E., Tejera R.L., Díaz L., Esparza P., Ibáñez E. Natural dyes extraction from cochineal (*Dactylopius coccus*). New extraction methods. *Food Chemistry* **2012**, *132*, 1855–1860. DOI:10.1016/j.foodchem.2011.12.018.
9. Müller-Maatsch J., Gras C. The “Carminic Problem” and Potential Alternatives. *Handbook on Natural Pigments in Food and Beverages* **2016**, 385–428. DOI.org/10.1016/B978-0-08-100371-8.00018-X
10. Sánchez-García M.A., Bokhimi X., Velázquez S., Jiménez-González A.E. Dye-Sensitized solar cells prepared with Mexican pre-hispanic dyes. *J. Nanotechnol* **2018**, *2018*. DOI.org/10.1155/2018/1236878.
11. Vanegas-Rico J.M., Rodríguez-Leyva E., Lomeli-Flores J.R., González-Hernández H., Pérez-Panduro A., Mora-Aguilera G. Biology and life history of *Hyperaspis trifurcata* feeding on *Dactylopius opuntiae*. *BioControl* **2016** *61*(6), 691–701. DOI 10.1007/s10526-016-9753-0.
12. Vanegas-Rico J.M., Lomeli-Flores J.R., Rodríguez-Leyva E., Mora-Aguilera G., Valdez J.M. Natural enemies of *Dactylopius opuntiae* (Cockerell) on *Opuntia ficus-indica* (L.) Miller in Central Mexico. *Acta Zoológica Mexicana (n.s)* **2010**, *26*(2), 415–33.
13. Griffith M.P. The origins of an important cactus crop, *Opuntia ficus-indica* (Cactaceae): New molecular evidence. *Am. J. Bot.* **2004**, *91*(11), 1915–21. DOI.org/10.3732/ajb.91.11.1915.
14. Mazzeo G., Nucifora S., Russo A., Suma P. *Dactylopius opuntiae*, a new prickly pear cactus pest in the Mediterranean: an overview. *Entomologia Experimentalis et Applicata* **2019**, *167*, 59–72. DOI: 10.1111/eea.12756.
15. Torres J.B., Giorgi J.A. Management of the false carmine cochineal *Dactylopius opuntiae* (Cockerell): perspective from Pernambuco state, Brazil. *Phytoparasitica* **2018**, *46*, 331–40. DOI.org/10.1007/s12600-018-0664-8.
16. Mendel Z., Protasov A., Vanegas-Rico J.M., Lomeli-Flores J.R., Suma P., Rodríguez-Leyva E. Classical and fortuitous biological control of the prickly pear cochineal, *Dactylopius opuntiae*. *Biological Control* **2020**, *142*, 104157. DOI.org/10.1016/j.biocontrol.2019.104157.
17. Silva M.A., Albuquerque T.G., Pereira P., Ramalho R., Vicente F., Oliveira M.B., Costa H. S. *Opuntia ficus-indica* (L.) mill.: A multi-benefit potential to be exploited. *Molecules* **2021**, *26*, 951. DOI.org/10.3390/molecules26040951.
18. Kiesling R. Origen, domesticación y distribución de *Opuntia ficus-indica*. *Journal of the Professional Association for Cactus Development* **1998**, *3*, 50-60. <https://www.researchgate.net/publication/228607157>.
19. Hernandez-Hernandez F.C., García-Gil F., Rojas-Martínez A., Hernández-Martínez S., Lanz-Mendoza H. Carminic acid dye from the homopteran *Dactylopius coccus* hemolymph is consumed during treatment with different microbial elicitors. *Arch Insect Biochem Physiol* **2003**, *54*(1), 37–45. DOI: 10.1002/arch.10099.
20. Bravo-Vinaja A., Méndez-Gallegos J. Emerging trends in scientific research on *Dactylopius coccus* Costa (Hemiptera: Dactylopiidae), carminic acid and its derivatives: a bibliometric analysis. *Agricultura, Sociedad y Desarrollo* **2023**, *20*(2), 139–65. DOI.org/10.22231/asyd.v20i2.1387.
21. Sabbahi R., Hock V. Control of the prickly pear cochineal, *Dactylopius opuntiae* (Cockerell), in Morocco: an overview. *Journal of Plant Diseases and Protection* **2022**, *129*, 1323–30. DOI.org/10.1007/s41348-022-00655-y.
22. García-Pascual E., González-Chávez M.M., Franco A., Rodríguez-Leyva E., Méndez-Gallegos S. J., Morales Rueda J.A., Bravo A. *Dactylopius opuntiae* (Hemiptera: Dactylopiidae) control tactics: a bibliometric analysis. *Investigación Bibliotecológica* **2023**, *38*(98), 13–29. DOI.org/10.22201/iibi.24488321xe.2024.98.58813.
23. Paré P.W., Tumlinson J.H. Update on plant-insect interactions plant volatiles as a defense against insect herbivores. *Plant Physiology* **1999**, *121*, 325-332. DOI.org/10.1104/pp.121.2.325

24. Turlings T.C.J., Erb M. Tritrophic interactions mediated by herbivore-induced plant volatiles: Mechanisms, ecological relevance, and application potential. *Annual Review of Entomology* **2017**, *63*, 433-452; DOI.org/10.1146/annurev-ento-020117-043507.
25. Pophof B., Stange G., Abrell L. Volatile organic compounds as signals in a plant-herbivore system: Electrophysiological responses in olfactory sensilla of the moth *Cactoblastis cactorum*. *Chem Senses* **2005**, *30(1)*, 51–68. DOI.org/10.1093/chemse/bji001.
26. Beck J.J., Baig N., Cook D., Mahoney N.E., Marsico T.D. Semiochemicals from ex situ abiotically stressed cactus tissue: A contributing role of fungal spores? *J. Agric. Food Chem.* **2014**, *62(51)*, 12273–12276. DOI.org/10.1021/jf505735g.
27. Rasmann S., Köllner T., Degenhardt J., Hiltbold I., Toepfer S., Kuhlmann U., Gershenzon J., Turlings T. Recruitment of entomopathogenic nematodes by insect-damaged maize roots. *Nature* **2005**, *434*. www.nature.com/nature.
28. Maffei M.E. Sites of synthesis, biochemistry and functional role of plant volatiles. *South African Journal of Botany* **2010** *76(4)*, 612–31. DOI:10.1016/j.sajb.2010.03.003.
29. Kasote D., Lee J., Sreenivasulu N. Editorial: Volatilomics in plant and agricultural research: recent trends. *Frontiers in Plant Science* **2023**, *14*. DOI 10.3389/fpls.2023.1289998.
30. Flath R.A., Takahashi J.M. Volatile Constituents of Prickly Pear (*Opuntia ficus indica* Mill., de Castilla Variety). *J. Agric. Food Chem.* **1971**, *26(4)*, 835-837. DOI.org/10.1021/jf60218a053.
31. Chahdoura H., Barreira J.C.M., Fernández-Ruiz V., Morales P., Calhella R.C., Flamini G. Soković M., Ferreira I. C. F. R., Achour L. Bioactivity, proximate, mineral and volatile profiles along the flowering stages of *Opuntia microdasys* (Lehm.): Defining potential applications. *Food Funct* **2016**, *7(3)*, 1458–1467. DOI: 10.1039/c5fo01536g.
32. Nounah I., Chbani M., Matthäus B., Charrouf Z., Hajib A., Willenberg I. Profile of volatile aroma-active compounds of cactus seed oil (*opuntia ficus-indica*) from different locations in morocco and their fate during seed roasting. *Foods* **2020**, *9(9)*. DOI:10.3390/foods9091280.
33. Wright C.R., Setzer W.N. Volatile compositions of two cactus species growing in the sonoran desert of Southern Arizona. *American Journal of Essential Oils and Natural Products* **2013**, *1(1)*, 41-47. https://www.researchgate.net/publication/270572666
34. Apablaza E., Sáenz C., Prat L., Ubeda C. Comprehensive characterization and volatile profile of cactus pear fruits from six different species and cultivars. *ACS Food Science and Technology* **2021**, *1(5)*, 928–36. DOI.org/10.1021/acfoodscitech.1c00027.
35. El-Hawary S.S., El-tantawy M.E., Rabeh M.A., Badr W.K. Chemical composition and antimicrobial activity of volatile constituents of cladodes, fruits peel and fruits pulp from *Opuntia ficus indica* (L.) Mill. (Prickly pear) growing in Egypt. *Egypt J. Chem.* **2020**, *64(1)*, 437-444. DOI: 10.21608/EJCHEM.2020.21137.2260.
36. Zhang W.W., Zheng H., Zhang H., Chen X.M., Guo Y.H. Volatile changes of *Opuntia ficus-indica* Mill before and after parasitizing by *Dactylopius coccus* Costa. *Chinese Journal of Applied Entomology* **2012**, *49(4)*, 986-991
37. Stierlin É., Nicolè F., Costes T., Fernandez X., Michel T. Metabolomic study of volatile compounds emitted by lavender grown under open-field conditions: a potential approach to investigate the yellow decline disease. *Metabolomics* **2020**, *16(31)*. DOI.org/10.1007/s11306-020-01654-6.
38. Cui S., Ling P., Zhu H., Keener H.M. Plant pest detection using an artificial nose system: A review. *Sensors* **2018**, *18(2)*, 378. DOI:10.3390/s18020378.
39. Cázares-Samaniego P.J., Castillo C.G., Ramos-López M.A., González-Chávez M.M. Volatilome and essential oil of *Ulomoides dermestoides*: A broad-spectrum medical insect. *Molecules* **2021**, *26(20)*, 6311. DOI.org/10.3390/molecules26206311.
40. De Lotto G. On the status and identity of the cochineal insects (Homoptera: Coccoidea: Dactylopiidae). *J. mi. Soc. sth. Afr.* **1974**, *37(1)*, 167-193. https://hdl.handle.net/10520/AJA00128789_2848.
41. Waku Y., Foldi I. The fine structure of insect glands secreting waxy substances. In: King, R.C., Akai, H. (eds) *Insect Ultrastructure*. Springer, Boston, MA, 1984; pp. 303-322. https://doi.org/10.1007/978-1-4613-2715-8_8.
42. Rodríguez L.C., Faúndez E.H., Niemeyer H.M. Mate searching in the scale insect, *Dactylopius coccus* (Hemiptera: Coccoidea: Dactylopiidae). *Eur. J. Entomol* **2005**, *102*, 305-306
43. Palafox-Luna J.A., Rodríguez-Leyva E., Lomeli-Flores J.R., Viguera-Guzmán A. L., Vanegas-Rico J. M. Ciclo de vida y fecundidad de *Dactylopius opuntiae* (Hemiptera: Dactylopiidae) en *Opuntia ficus-indica* (Caryophyllales: Cactaceae) *Agrociencia* **2018**, *52(1)*, 103-114.
44. Martínez T., Fabriás G., Camps F. Sex pheromone biosynthetic pathway in *Spodoptera littoralis* and its activation by a neurohormone. *The Journal of Biological Chemistry* **1990**, *265(3)*, 1381-1387. https://doi.org/10.1016/S0021-9258(19)40025-2

45. Tsfadia O., Azrielli A., Falach L., Zada A., Roelofs W., Rafaeli A. Pheromone biosynthetic pathways: PBAN-regulated rate-limiting steps and differential expression of desaturase genes in moth species. *Insect Biochem Mol. Biol.* **2008**, *38*(5), 552–67. DOI:10.1016/j.ibmb.2008.01.005.
46. Zou Y., Millar J.G. Chemistry of the pheromones of mealybug and scale insects. *Nat Prod Rep.* **2015**, *32*(7), 1067–1113. DOI: 10.1039/c4np00143e.
47. Rodrigues C., Paula C.D. de, Lahbouki S., Meddich A., Outzourhit A., Rashad M., Pari L., Coelho I., Fernando A.L., Souza V.G.L. *Opuntia* spp.: An overview of the bioactive profile and food applications of this versatile crop adapted to arid lands. *Foods* **2023**, *12*, 1465. DOI.org/10.3390/foods12071465.
48. Holopainen J.K., Blande J.D. Where do herbivore-induced plant volatiles go? *Frontiers in Plant Science* **2013**, *4*. DOI: 10.3389/fpls.2013.00185.
49. Pérez-Hedo M., Bouagga S., Zhang N.X., Moerkens R., Messelink G., Jaques J.A., Flors V., Broufas G., Urbaneja A. Pappas M.L. Induction of plant defenses: the added value of zoophytophagous predators. *Journal of Pest Science* **2022**, *95*, 1501–1517. DOI.org/10.1007/s10340-022-01506-3.
50. Terrazas T, Mauseth J.D. Shoot Anatomy and Morphology', in Park Nobel (ed.), *Cacti: Biology and Uses* (Oakland, CA, 2002; online edn, California Scholarship Online, 22 Mar. 2012) DOI.org/10.1525/california/9780520231573.003.0002.
51. Dicke M., Sabelis M.W., Takabayashi J., Bruin J., Posthumus M.A. Plant strategies of manipulating predator-prey interactions through allelochemicals: prospects for application in pest control. *Journal of Chemical Ecology* **1990**, *16*(11), 3091–3118. DOI.org/10.1007/BF00979614.
52. Ozawa R., Arimura G.I., Takabayashi J., Shimoda T., Nishioka T. Involvement of jasmonate-and salicylate-related signaling pathways for the production of specific herbivore-induced volatiles in plants. *Plant Cell Physiol* **2000**, *41*(4), 381–398. DOI.org/10.1093/pcp/41.4.391.
53. Drukker B., Bruin J., Sabelis M.W. Anthocorid predators learn to associate herbivore-induced plant volatiles with presence or absence of prey. *Physiological Entomology* **2001**, *25*, 260–265. DOI.org/10.1046/j.1365-3032.2000.00190.x.
54. Clavijo-McCormick A., Unsicker S.B., Gershenzon J. The specificity of herbivore-induced plant volatiles in attracting herbivore enemies. *Trends in Plant Science* **2012**, *17*, 303–310. DOI.org/10.1016/j.tplants.2012.03.012.
55. Kessler A., Baldwin I.T. Plant responses to insect herbivory: The emerging molecular analysis. *Annual Review of Plant Biology* **2002**, *53*, 299–328. DOI: 10.1146/annurev.arplant.53.100301.135207
56. Li S.Q., Zhang Z.N. The influence of volatiles from the hindgut of the pine sawyer, *Monochamus alternatus* (Coleoptera: Cerambycidae), on its oviposition behavior. *Zoological Studies* **2007**, *46*(6), 726–733.
57. Smith B.C., Williams R.R. Temperature relations of adult *Coleomegilla maculata lengi* and *C. M. medialis* (Coleoptera: Coccinellidae) and responses to ovipositional stimulants. *The Canadian Entomologist* **1976**, *108*(9), 925–930. DOI:10.4039/Ent108925-9.
58. Berhe Y.K., Portillo L., Viguera A.L. Resistance of *Opuntia ficus-indica* cv 'Rojo Pelon' to *Dactylopius coccus* (Hemiptera: Dactylopiidae) under greenhouse condition. *Journal of the Professional Association for Cactus Development* **2022**, *24*, 293–309. DOI.org/10.56890/jpacd. v24i.509.
59. Nakata P.A. An assessment of engineered calcium oxalate crystal formation on plant growth and development as a step toward evaluating its use to enhance plant defense. *Plos One* **2015**, *10*(10). DOI:10.1371/journal.pone.0141982.
60. War A.R., Buhroo A.A., Hussain B., Ahmad T., Nair R.M., Sharma H.C. Plant defense and insect adaptation with reference to secondary metabolites. *Co-Evolution of Secondary Metabolites* **2020**, 795–822. DOI.org/10.1007/978-3-319-96397-6_60.
61. Ferris G.F. Atlas of the Scale Insects of North America. *Nature* **1943**, *151*, 657. DOI.org/10.1038/151657c0.
62. Bardou P., Mariette J., Escudie F., Djemiel C., Klopp C. Jvenn: an interactive Venn diagram viewer. *BMC Bioinformatics* **2014**, *15*, 293. DOI.org/10.1186/1471-2105-15-293.

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