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Article

Novel Dry Powder of Hyaluronic Acid-Vancomycin Complex for Inhalation Useful on Pulmonary Infections Associated to Cystic Fibrosis

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Abstract: Polyelectrolyte-drug complexes are interesting alternatives to improve unfavourable drugs properties. Vancomycin (VAN) is an antimicrobial used in the treatment of methicillin-resistant *Staphylococcus aureus* pulmonary infections in patients with cystic fibrosis. It is generally administered intravenously with high incidence of adverse side effects, which could be reduced by intrapulmonary administration. Currently, there are no commercially available inhalable formulations containing VAN. Thus, the present work focuses on the preparation and characterization of an ionic complex between hyaluronic acid (HA) and VAN with potential use as inhalable formulations. Particulate-solid HA-VAN₂₅ complex by spray drying from an aqueous dispersion was obtained. FTIR spectroscopy and thermal analysis confirmed the ionic interaction between HA and VAN, while an amorphous diffraction pattern by X-ray was observed. The powder density, geometric size and morphology showed a suitable aerosolization and aerodynamic performance of the powder, indicating the capability of reaching the deep lung. An *in vitro* extended-release profile of VAN from the complex was obtained, exceeding 24 hours. Microbiological assays against methicillin-resistant and -sensitive reference strains of *Staphylococcus aureus*, showed that VAN preserves its antibacterial efficacy. In conclusion, HA-VAN₂₅ exhibited interesting properties for the development of inhalable formulations with potential efficacy and safety advantages over conventional treatment.

Keywords: Vancomycin inhalable; cystic fibrosis; polyelectrolyte-drug complex; hyaluronic acid; pulmonary infection

1. Introduction

Cystic fibrosis (CF) is a disorder that damages the lungs, gastrointestinal tract and other organs. It is a hereditary, potentially fatal disease, caused by a defective gene that affects the cells that produce mucus, sweat and gastrointestinal fluids due to defects in CF transmembrane conductance regulator protein [1,2]. Although it can affect different organs, pulmonary bacterial infections are the most frequent form of clinical presentation and the main cause of morbidity and mortality. In the lungs, viscous airway secretions result in inflammation and chronic infection which vary according to age, in fact, the progressive obstruction and long-term damage to the airways leads progressively to decreased ability to clear secretions, causing increased rates of infections [3,4].

The most frequent respiratory infections associated with CF are caused by *Pseudomonas aeruginosa* and *Staphylococcus aureus*, which are treated systemically with antimicrobials administered by different routes, in most cases intravenous or oral [5]. In particular, infection with methicillin-resistant *Staphylococcus aureus* (MRSA) is associated with significantly worse clinical outcomes. While

intravenous vancomycin (VAN) is standard therapy for MRSA, in the respiratory setting, efficacy is reduced by relatively poor penetration into lung secretions and dose-limiting nephrotoxicity [6,7].

Inhaled antibiotics offer a pharmacotherapeutic alternative for the treatment of chronic respiratory tract infections, for example achieve high concentrations of drugs directly to the target site, mucosa of the lungs, while minimizing the risk of systemic side effects, improving both efficacy and safety of the pharmacotherapy [8]. Nebulizer and dry powder formulations for inhalation are currently available for the treatment of infections caused by *Pseudomonas aeruginosa*, containing tobramycin, aztreonam, levofloxacin or colistin. However, there are no commercially available inhaled antimicrobials that are effective against MRSA, *i.e.*, ciprofloxacin and VAN formulations are in different stages of clinical development, denoting the high potential clinical relevance that could have local optimized treatments against MRSA to increase the survival of patients with CF [8,9]. In this line, off-label use of inhaled VAN by nebulization of the intravenous formulation has been reported for treatment of MRSA in the setting of CF, demonstrating a reduction in colonies but not eradication [10]. Nevertheless, inhaled formulations can present side effects, such as cough, bronchospasm, hoarseness, dysphagia, among others, which are frequently associated with mucosal deposition of crystalline drug particles with slow dissolution rate [11].

Therefore, biocompatible drug delivery carriers are an interesting strategy to generate inhaled pharmaceutical products with improved properties beyond available therapies. Over the past decade, significant progress has been made in the development of new pharmaceutical technology platforms, based on polyelectrolyte-ionizable drugs (PE-D) complexes, both in aqueous dispersions and solid materials forms, with potential drug delivery applications [12–14]. The acid-base interaction between carboxylic groups of a PE and a basic group of a drug, yields a high degree of counterionic condensation. PE-D complexes exhibit several unique and favorable properties to design smart or stimuli-responsive drug delivery systems. Based on this platform, modified drug release systems are designed with emphasis on approaching specific pharmacotherapeutic situations. These systems allow to improve the efficacy, safety and reliability of the drug therapeutic action with recognized clinical impact [15]. Besides, they are an accessible and original alternative to develop sustained release formulations and can be produced by a simple and easy to scale-up process. In this context, hyaluronic acid (HA), a natural, bioadhesive and recognized as safe (GRAS) polysaccharide, is an interesting anionic PE with denoted capacity to act as oppositely charged drugs carrier system [16]. In addition, HA is an important component of the extracellular matrix of the lungs that plays a crucial role in the regulation of fluid balance in the pulmonary interstitium. Due to its protection against bronchoconstriction and its hydrating properties, inhaled HA would increase the fluid volume of the airway surface, resulting in an increase in airway fluid volume [17]. Furthermore, the addition of nebulized HA in patients undergoing continuous hypertonic saline therapy, results in an improved tolerability profile, effectively relieving symptoms such as cough, throat irritation and unpleasant taste associated with this treatment [18].

The aim of this work is to develop a particulate inhaled formulation based on an ionic complex between VAN and HA with optimized aerosolization behavior by spray drying technique. It is expected to achieve a pulmonary VAN delivery system to control locally in the infection site and reduce the severe adverse events produced by systemic administration but also, by the direct deposition of crystalline drugs on lung mucosa.

2. Materials and Methods

2.1. Materials

Vancomycin hydrochloride (VAN) was kindly supplied by Lab. Química Luar SRL (Córdoba, Argentina). Sodium hyaluronate was purchased from Pura Química® (batch n° 313318, Córdoba, Argentina). Cation exchange resin (Amberlite® IR-120 Fluka, Sigma-Aldrich, St. Louis, MO, USA); NaCl, KCl, NaHPO₄ and KH₂PO₄ (Parafarm®, Buenos Aires, Argentina); NaOH and HCl 1N (analytical reagents, Anedra, Córdoba, Argentina), acetonitrile and acetic acid (VWR International, Fontenay-sous-Bois, France), sodium acetate and Span 80 (Merck KGaA, Darmstadt, Germany)

Müeller Hinton broth and agar (Britania, Argentina) were used as provided by suppliers. Ultrapure water (water purification NW-system, Heal Force Group, Shangay, CN) was used for all experiments.

2.2. Preparation of Hyaluronic Acid-Vancomycin Complex

Since not available commercially, the acidic-form of HA was obtained from acid-base neutralization of sodium hyaluronate (NaHA) dispersions using a procedure reported by Battistini *et al.* [12]. Briefly, the purified HA dispersion (pH \leq 2.5) was collected by eluting a 0.50% w/v NaHA dispersion through a column containing an anionic exchange resin and then the solid state was achieved by lyophilization (FreeZone 6, Labconco®, MO, USA, at -40°C and 1×10^{-3} mPa of vacuum).

Prior to use, determination of the ionizable carboxylic groups proportion per mass unit of HA (2.59 mmol/g) was performed by differential scanning potentiometry, according to the following methodology. An appropriate amount of HA powder (approximately 100 mg) was dissolved in ultrapure water containing 10 mL of HCl 0.05N and then titrated with 20 mL of NaOH 0.05N using an automatic acid-base titrator (Titrand® 905, Software Tiamo® 2.0 light, Methrom AG, CH). Another homologue titration containing an identical proportion of strong acid but without adding HA, was used as reference. The results were in agreement with its monomeric structure and with the equivalents of a similar HA previously reported [12].

The next step was the preparation of the complex HA-VAN₂₅ itself, where the subscript 25 refers to the proportion of VAN, expressed in moles, required to neutralize 25% of HA ionizable carboxylate groups. In order to formulate the complex, an aqueous dispersion of HA at 0.3% w/v was elaborated, and the appropriate amount of VAN required to neutralize the 25% of HA was added under constant stirring and left overnight to complete interaction. Then, the pH value of complex dispersion was adjusted, using NaOH 1N solution up to 7.0 ± 0.1 . The particulate complex was obtained by spray-drying the aqueous dispersion with a mini Spray Dryer Büchi-B290 (Büchi® Labortechnik, Flawil, CH) using the following parameters: nozzle: 0.7 mm; inlet temperature: 120°C ; aspiration rate: $35\text{m}^3/\text{h}$, feed rate: $4\text{ mL}\cdot\text{min}^{-1}$ and air flow rate: $600\text{ L}/\text{h}$. The outlet temperature was 70°C .

The powder obtained from the spray drying process was removed from the collection vessel, dried under vacuum for 2 hours and weighed on an analytical balance (Ohaus AR2140, NJ, USA, sensitivity 0.1 mg) to determine the yield. The manufactured powder was stored with a rubber stopper tightly sealed by a metal cup in a glass vial, protected from light and moisture, and stored at 25°C . The yield efficiency (Y) of spray-drying process was calculated using the following equation (1):

$$Y(\%) = \frac{W_s}{W_t} \times 100 \quad (1)$$

where W_s is the weight of the powder obtained from the spray-dryer and W_t is the total weight of the raw materials, HA and VAN.

In parallel, a physical mixture (PM) of VAN and HA was prepared in a mortar with the same composition and proportion as HA-VAN₂₅ which was used as comparative control in the physicochemical characterization.

2.3. Physicochemical Characterization of HA-VAN₂₅ in Solid State

The spray-dried powder of HA-VAN₂₅ complex was characterized at molecular, particulate and bulk level using different analytical and physical methods in order to evaluate the suitability of HA-VAN₂₅ as a potential inhalation powder.

2.3.1. Infrared Spectroscopy

The FTIR spectra of the complex, HA-VAN₂₅, PM and the raw materials were collected in the range from 4000 to 650 cm^{-1} using a FTIR spectrometer (Cary 630, Agilent®, Santa Clara, CA, USA) set to a resolution of 4 cm^{-1} and were scanned 40 times per spectrum, equipped with a specific analysis software OMNIC 8.3 spectrum software (Thermo Scientific).

2.3.2. Thermal Analysis

Differential scanning calorimetry (DSC) and thermogravimetric analysis (TGA), using TA® Discovery series instrument (TA Instruments, New Castle, DE, USA), were used to evaluate the thermal behavior of the complex, the PM and the raw materials. The results were processed and analyzed using the TRIOS® software v4.1.1 (TA Instruments). Approximately, 2 mg of each sample was weighed and sealed in non-hermetic aluminium pans. DSC analysis was performed in a temperature range starting from room temperature (25 °C) until sample decomposition, between 190 and 215 °C using a heating ramp of 10 °C/min under N₂ atmosphere (50 mL/min). TGA analysis was carried out under the same conditions as DSC analysis but with a temperature range from room temperature to 400 °C.

2.3.3. X-ray Diffraction

Powder X-ray diffraction (PXRD) patterns were obtained to evaluate the crystalline or amorphous state of VAN, HA and HA-VAN₂₅. For this purpose, an X-ray powder diffractometer (PW1800, Philips, Amsterdam, the Netherlands) using Cu Ka radiation ($\lambda=1.5418 \text{ \AA}$, tube at 40 kV, 100 mA) was used. Data were collected over an angular range of 5 to 60° 2 θ / θ using a step method, with a step of 0.02 and scan rate of 1 seg per step.

2.4. Powder Characterization

2.4.1. Scanning Electron Microscopy (SEM)

Images were captured using an electron microscope (FE-SEM Sigma, ZEISS Instrument, Oberkochen, Germany) with a magnification of 1,000× for VAN and 1,000× but also 10,000× for HA-VAN₂₅. The samples were prepared by dispersing 1–2 mg of powder directly on the carbon tape, placed on the aluminium stubs. The surface morphology and texture of the particles were investigated in plan-view by using a 1 kV electron beam acceleration voltage.

2.4.2. Density Determination and Flow Properties

The flow properties of pharmaceutical powders are frequently evaluated from the bulk and tap densities determinations, in accordance with United States Pharmacopeia (USP) specifications [19]. To determine bulk density (δ_B), an exactly weighed amount of HA-VAN₂₅, between 0.5-1 g, was gently introduced into a 10 mL calibrated measuring cylinder avoiding compaction. The powder was carefully level and the volume was read to the nearest graduated unit. To measure tap density (δ_T), the powder contained in the cylinder was manually tapped until no further changes in the volume were observed. From these data, Carr's index and Hausner ratio were calculated according to equation 2 and 3, respectively[20]:

$$\text{Carr's index (\%)} = 100 \times \frac{\delta_T - \delta_B}{\delta_T} \quad (2)$$

$$\text{Hausner ratio} = \frac{\delta_T}{\delta_B} \quad (3)$$

where δ_T is the tap density and δ_B is the bulk density. Each sample was tested in triplicate.

2.4.3. Particle Size Distribution by Laser Diffraction

The measurement of particle size distribution (PSD) of dry powder HA-VAN₂₅ was performed using a Mastersizer 3000 (Malvern Instruments Ltd, Worcestershire, UK) laser diffraction. All the samples were dispersed using a dry powder feeder (Aero S, Malvern Panalytical, UK) at a dispersive air pressure of 4.0 bar, 3 mm sample flow control and vibration feed rate was set to 45%. The results of the median volume diameter of the powder were expressed as Dv₉₀, Dv₅₀ and Dv₁₀, namely, cumulative undersize volume diameters at 90%, 50% and 10% of the particle population, respectively.

Another parameter measured was the SPAN value, which is the distribution width and was calculated using the following equation (4):

$$\text{SPAN} = \frac{Dv90 - Dv10}{Dv50} \quad (4)$$

2.5. *In vitro* Biopharmaceutical Performance of HA-VAN₂₅ Complex

2.5.1. Aerodynamic Performance Assessment

The aerodynamic distribution of the powders was assessed using the next generation impactor (NGI) at flow rate of 65 L/min (Copley Scientific Limited, Nottingham, UK). The 30 mg of aerosolized powder contained 10 mg of VAN, according to the formulation, were loaded in HPMC QUALI-V I size 3 capsule (Qualicaps, Madrid, Spain) and insert in RS01® inhaler device (Plastiaple, Lecco, IT). After aerosolization, the powder deposited in the different NGI parts, was collected using distilled water and the VAN was quantified by HPLC (see next section).

Different parameters were calculated according to USP specifications for dry powder inhalers [21]. Between these, the emitted dose (ED) corresponds to the amount of VAN that leaves the inhaler device and enters to the in NGI; the emitted fraction (EF) is the ratio between the ED and the total mass of powder in the inhaler device. From the amount of powder deposited on the impactor and their ratio to ED, the mass median aerodynamic diameter (MMAD, *dae* where 50% of the population is smaller than that value), the fine particle fraction (FPF, fraction of powder with *dae* < 5 µm) and the extra-fine particle fraction (EFPF, fraction of powder with *dae* < 3 µm) were calculated.

2.5.2. Analytical Quantification of Vancomycin

HPLC was performed to determine the VAN content in the spray-dried complex using the Agilent equipment 1200 LC series (Agilent Technologies, Santa Clara, CA, USA) equipped with a UV-Vis detector and the data analyzed by OpenLab CVS Chem Station software (rev.c.01.06 v.A.04.02, Agilent Technologies). A reverse-phase and isocratically method was employed along with a 150×3 mm Luna C18 3 µm (Phenomenex, Torrance, CA, USA) column at 30 °C. VAN was eluted using a flow rate of 0.45 mL/min and an injection volume of 20 µL with a mobile phase formulated with acetonitrile and acetate buffer pH 3.50 8:92 v/v. The detector was set at 240 nm and the retention time was around 1.7 min. The analytical method was validated for linearity, precision and accuracy in the appropriate concentration range of analysis in accordance with ICH Guideline Q2. Calibration curves were constructed using five concentrations of VAN dissolved in deionized water in a range from 12.5 to 1000 µg/mL, by triplicate. Each standard was injected five times and the linearity and precision expressed as relative standard deviation (RSD%) of the assay was calculated. The correlation coefficient (R²) for the calibration curve was 0.9996 with a linearity range of 12.5-1000 µg/ml. The reproducibility was considered acceptable as the RSD% values obtained were for all the concentration tested < 2% (limit defined by the ICH guidelines). The LOQ (lower limit of quantification) was 0.25 µg/mL (RSD% 1.72), and the LOD (limit of detection) was 0.125µg/mL (RSD% 1.22).

2.5.3. Dissolution Study

Dissolution tests of VAN and HA-VAN₂₅, were carried out following USP Method I (SOTAX AT 7 Smart, Westborough, MA, USA) using 500 mL of phosphate buffer solution (PBS) at pH = 7.4 and 37.0 ± 0.5 °C as dissolution medium, with basket at 100 rpm speed. An amount of powder containing 150 mg of VAN as raw material and the equivalent of the same in the complex HA-VAN₂₅ were performed. The latest was also sieved using a 560-630 µm sieve prior to the test. Each sample was incorporated into the dissolution medium at time 0 and 2 mL aliquots were taken at predetermined times (5, 10, 15, 30, 45, 60, 90, and 120 min), these extracted volumes were replaced with a fresh thermostated medium. The aliquots were filtered through a cellulose filter before dilution. The concentration of dissolved VAN was determined by UV-Vis spectrophotometry (Evolution 300,

Thermo Electron Corporation, Waltham, MA, USA) at 280 nm. A calibration curve, by triplicate, was designed with six different concentrations of VAN between 55 and 280 µg/mL dissolved in deionized water, obtaining a R² of 0.9985.

The results were expressed as the mean of three determinations with their SD. The profiles obtained were statistically compared using the difference factor (f_1 , equation 5), two profiles being considered different when the value of f_1 calculated between them was greater than 15.

$$f_1 = \frac{\sum_{i=1}^k |R_i - T_i|}{\sum_{i=1}^k R_i} \quad (5)$$

2.5.4. Franz' Cell Diffusion Study

VAN release from complex dispersions was performed at 37.0 ± 0.1 °C in bicompartamental Franz cells, the donor and the receptor compartment were separated by a semisynthetic cellulose membrane (MW cut-off 12 kDa, Sigma-Aldrich, St. Louis, MO, USA). In the donor compartment of each cell, 1 mL containing 11 mg of VAN or HA-VAN₂₅ in the same proportion of VAN was introduced. Sufficient water or PBS (pH = 7,4), approximately 16 mL, were used as the receptor medium. Samples of 0.8 mL were taken at predetermined time intervals and the extracted volume was replaced with fresh medium heated to 37°C. The concentration of released VAN was analyzed by UV-Vis spectrophotometry at 280 nm. Assays were carried out in triplicate and immersion (sink) conditions were maintained.

The profiles obtained were statistically compared using the difference factor, f_1 , presented previously in equation 5.

In addition, the release data were processed using the empirical power equation proposed by Peppas [22] to evaluate the kinetics and mechanism of drug release:

$$\frac{M_t}{M_\infty} = k \cdot t^n \quad (6)$$

where M_t is the amount of drug permeated at time t ; M_∞ is the initial amount of drug in the donor compartment; k is the kinetic constant and n the diffusion exponent characterizing the release mechanism, $n = 0.5$ (Fickian diffusion), $0.5 < n < 1$ (non-Fickian diffusion) and $n = 1$ (zero order, controlled release). The equation 6 is valid in the range up to 60% of the released drug.

2.6. Antibacterial Activity Assays

The samples tested were HA, VAN and HA-VAN₂₅. Firstly, an agar diffusion assay was performed to determine the sensitivity on *S. aureus* strains, both methicillin-sensitive (MSSA) and -resistant (MRSA). For this purpose, a plate containing Müller Hinton Agar (MHA) was inoculated with three ATCC strains: 29213, 25923 and 43300 of *Staphylococcus aureus* where the last one corresponds to the MRSA strain. Then, wells were made where 40 µL of the samples were placed and incubated at 37 °C for 24 h. The inhibition halo formed by each sample was measured. Secondly, the minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) against MSSA and MRSA strains were determined using the plate dilution method according to CLSI standards [23]. From the stock solutions (SS) of VAN, HA or HA-VAN₂₅ serial dilutions (factor 2) were performed in Müller Hinton Broth (MHB) on a multiwell plate. Samples of 100 µL of standardized bacterial inoculum were added to each well and incubated at 37 °C for 24 h. The MIC corresponds to the lowest concentration that inhibits microbial growth and was determined by verifying the absence of turbidity with the naked eye. After MIC reading, dilutions without bacterial growth were plated on MHA and incubated for 24 h at 37 °C to determine the MBC. The MBC value was considered to be the lowest sample dilution in which no microbial growth was observed and therefore 99.9% of the initial inoculum had been reduced.

3. Results and Discussion

3.1. HA-VAN₂₅ Complex Preparation

The PE-D complex was successfully obtained using VAN and HA. It was important to use HA instead of the sodium salt to favor the interaction with VAN, since it also has a considerably lower viscosity than NaHA which reduces the amount of complex that sticks to the spray-dryer walls, increasing the process yield. The inclusion of HA in this formulation might play a key role in airway diseases with a high inflammatory component, such as CF [17]. In preliminary experiments different proportions of VAN were assayed to neutralize HA in order to obtain a physical stable dispersion without sedimentation of aggregated particles or solid precipitates after adjusting the pH value. From these, it was selected the HA-VAN₂₅ complex, as the more appropriate complex composition that remains as a transparent solution and prevents the formation of agglomerates and solid sedimentation. In this context, HA-VAN₂₅ prepared at 0.3% w/v of HA appeared as a transparent colloidal dispersion, which formed spontaneously after dispersing the raw materials under constant stirring. The addition of NaOH in order to get a more biocompatible pH value did not produce any changes in the appearance of the formulation, like darkening or precipitation, so it could be inferred that the system is physically stable. This dispersion was successfully spray dried obtaining a yield of 69% of microparticulate HA-VAN₂₅ complex in the solid state.

3.2. Physicochemical Characterization of HA-VAN₂₅ in the Solid State

The solid-state integral characterization of HA-VAN₂₅ microparticles was carried out using several characterization techniques including FTIR analysis, PXRD, TGA and DSC.

Figure 1 shows the comparative FT-IR spectra, where the bands that could serve as indicators of acid-base interactions were identified in order to analyze the changes attributable to the ionic association between HA and VAN. Regarding to FT-IR spectrums of the raw materials, HA showed the characteristic bands related to carboxylic groups, such as those at 3357 cm⁻¹ corresponding to the O-H tensile vibration of the COOH group, at 1732 cm⁻¹ attributable to the C=O carbonyl stretching vibration of the COOH group and at 1314 cm⁻¹, corresponding to the C-O bond of the carboxyl group of COOH [12,24]. In the VAN spectrum, a broad band at 3280 cm⁻¹ corresponding to the overlapping O-H and N-H tensile vibrations of the acid and amino groups was observed. In addition, the C=O stretching vibration at 1647 cm⁻¹ overlapped with the N-H bending vibration of the amino groups present, and the signal at 1225 cm⁻¹ was attributed to the C-O bond stretching [25].

Comparing the HA-VAN₂₅ complex with the raw materials and its PM, notable differences were observed. The band at 1732 cm⁻¹, corresponding to the C=O of the COOH group, evident in both HA and the PM, was absent in the complex. Furthermore, a band emerged at 1401 cm⁻¹, attributed to the stretching of the C-O bond in the carboxylate group (COO⁻), which is not present in the PM. These findings suggest a potential ionic interaction between HA and VAN, as only the bands corresponding to the ionized groups are evidenced.

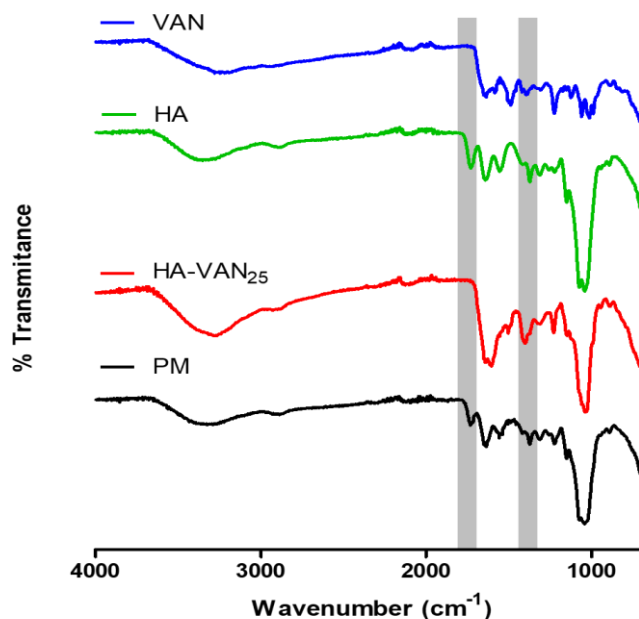


Figure 1. Overlapped Fourier transform Infrared spectroscopy spectra of HA-VAN₂₅ complex; raw materials, vancomycin (VAN) and hyaluronic acid (HA) and the physical mixture (PM).

The DSC and TGA thermograms, depicted in Figure 2, illustrate the thermal characteristics of the raw materials (HA and VAN), the HA-VAN₂₅ complex, and the PM. The TGA analysis reveals two significant weight loss events in VAN. The initial extensive event, starting from room temperature (25 °C) to 100 °C, is attributed to material dehydration resulting from the loss of water adsorbed on the solid particle surfaces, with a weight percent loss of (9.8 ± 0.7) %. This event aligns with the DSC thermogram, represented by a broad and nonspecific endotherm within the same temperature range. The second TGA event likely corresponds to the decomposition temperature, initiating at 208 °C, as evidenced by the sustained weight loss in the TGA curve. Additionally, it is noteworthy that the melting temperature of pure VAN could not be observed under the assay conditions, conducted up to 215 °C. This is consistent with the findings of other authors who investigated the thermal properties of VAN [26].

On the other hand, the DSC thermogram of HA exhibits an initial endothermic peak followed by an exothermic peak at approximately 180 °C, coinciding with mass loss in TGA, indicative of the initiation of its decomposition. The glass transition of pure HA could not be observed at the test temperatures. This aligns with prior findings, as reported by others authors, who similarly demonstrated a multi-stage decomposition process for HA where the initial degradation step involves dehydration from the compound structure, followed by a second stage near 200 °C, consistent with our results, and two additional peaks observed at 320 °C and 415 °C [27].

The thermograms of the HA-VAN₂₅ complex exhibit a weight loss in TGA and an endothermic dehydration peak in DSC between room temperature and 100 °C, mirroring the behavior of the precursors. However, unlike the PM, which decomposes at the same temperature as HA (the precursor that decomposes the first), the TGA spectra of the HA-VAN₂₅ complex reveal a shift in the decomposition temperature towards higher temperatures compared to HA. Consequently, the formation of a complex with VAN appears to shift HA decomposition at temperatures higher than 180 °C, thereby enhancing the polymer's stability. This observation suggests the presence of intermolecular forces between HA and VAN, influencing their thermal behavior, which aligns with the previously presented FT-IR results.

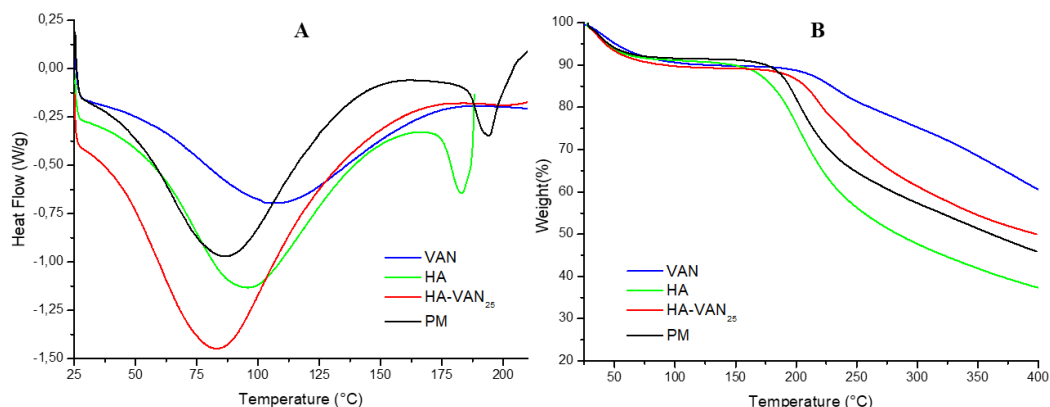


Figure 2. Thermograms of differential scanning calorimetry (A) and thermogravimetric analysis (B) of HA-VAN₂₅ complex, the raw materials (VAN and HA) and the PM.

The p-XRD patterns of VAN, HA and HA-VAN₂₅ were performed. In Figure 3, it can be observed that both precursors, the PM, and the complex are amorphous, evidenced by the absence of diffraction peaks, a consistent observation reported by previous researchers and in alignment with the earlier mentioned DSC spectra [28].

It has been reported that binary PE-D complexes, obtained by mixing a PE with a drug of opposite charge in a convenient medium capable of dissolving one or both components, result in amorphous solid materials where the drug is ionically bound to the polymeric carrier [28].

Generally, amorphous powders are favored over crystalline counterparts for pulmonary drug delivery due to the numerous advantages they offer [29]. For example, the deposition of crystalline particles can induce an inflammatory response, as reported by other authors who studied the inhalation of crystalline rifapentine particles [30]. Currently, TOBI®, an inhalable antibiotic containing tobramycin used in the treatment of *Pseudomonas aeruginosa* infections associated with CF, possesses an amorphous structure, akin to our HA-VAN₂₅ system [31].

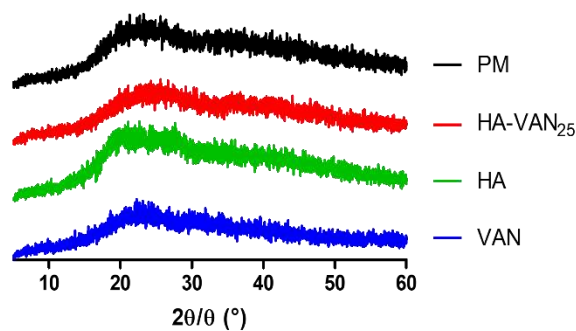


Figure 3. Powder X-ray diffraction patterns of HA-VAN₂₅, the raw materials (VAN and HA) and the PM.

Despite the p-XRD patterns obtained, revealing amorphous structures in both HA-VAN₂₅ and its precursors, HA and VAN, the comprehensive information provided by FT-IR, DSC, and TGA suggests an ionic interaction between the carboxylic groups of HA and the basic groups of VAN.

3.3. Powder Characterization

There are different physical factors that can affect the aerosolization and therefore the deposition in the deep lung of dry powders, such as particle size distribution, flowability of the formulation as well as particle density and shape [32–34].

As it's known, particles having a geometric size range between 1-5 μm are, in principle, suitable to reach the deep lung since particles with sizes higher than 5 μm can be deposited in the upper airways, whereas particles smaller than 1 μm can get exhaled [32].

SEM images were taken after the spray-drying process of the complex as well as the VAN as the raw material at different magnification values. According to the micrographs observed in Figure 4A,B (magnification 1,000 \times), VAN showed a flat and lamellar structure with irregular shape and size of the particles between 20-40 μm , which is considered too large for inhalation purposes. On the contrary, when the HA-VAN₂₅ complex was observed, much smaller particles were created (4B) and in the image with a magnification 10 times higher (4 C, 10,000 \times) it was noticed that most of the particles presented a size between 3-4 μm . Regarding the morphology of HA-VAN₂₅, the particles showed a characteristic hemispherical hollow shape with smooth surface and edges, as was previously described by Martinelli *et al.* [35] and Ceschan *et al.* [36] for similar hyaluronate sodium salts-based spray dried particles. The formation of agglomerates was noticeable, as it was evident also macroscopically, and it could also be seen structures of one-inside-other particles in some cases.

The use of spray-drying as a process for the obtention of the solid can be beneficial in the morphology of the particles as already demonstrated by the spray dried inhalation powder of colistimethate sodium for lung infections in CF [37]. Advantageous properties can be associated with the use of this technique as the shape and diameter uniformity of the particles which can provide good flowability, while the formation of hollow particles allow low-density structures, reducing the aerodynamic diameter and improving the aerosolization performances [38,39].

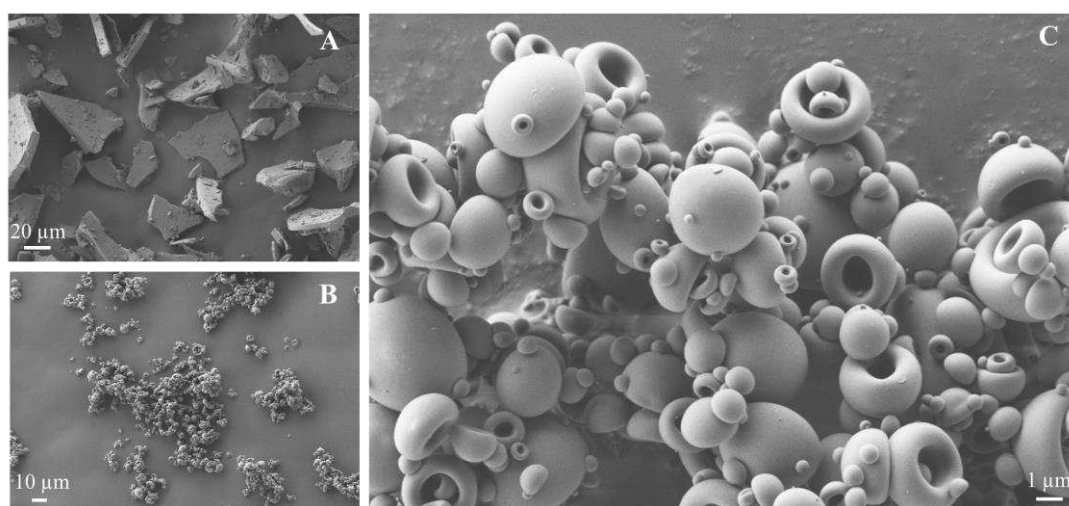


Figure 4. Scanning electron microscopy micrographs of VAN (A), HA-VAN₂₅ (B) with the same magnification (1,000 \times) and HA-VAN₂₅ (C) with a higher magnification (10,000 \times).

In this context, the determination of bulk density and flowability properties of inhalation dry-powders provides important material quality attributes directly related to the aerosolization behavior [40,41]. Taken all into consideration, it was measured the bulk and tapped density of the complex HA-VAN₂₅ showing a value of (0.13 ± 0.02) g/mL and (0.18 ± 0.03) g/mL, respectively. These values are reasonable for inhalable powders since tapped densities below 0.4 g/mL have been reported in many works as cut-off for determining good aerodynamic characteristics [42].

However, since the smaller the size of particles, the higher is the interaction particle-particle related to Van der Waals forces concerning high contact area, these cohesive forces can lead to poor flow properties and formation of agglomerates [40]. Particle agglomerates and poor flowability can impact the aerosolization performance of the powder formulation that ultimately can remain in the inhaler after patient inhalation, resulting in low emitted dose [34]. On the other hand, it is well-established that aerosolization performance in terms of fine particle dose of the emitted powder is optimal with smaller particle sizes, therefore the particle size and density of inhaled powders has to be engineered to balance their properties in order to provide the aerosolization performance as well a proper flowability.

In order to study the flow properties of the complex HA-VAN₂₅, Carr's index (CI) and Hausner ratio (HR) were used according to Equation 2 and 3, obtaining values of 26.9 ± 5.8 and 1.4 ± 0.1 , respectively. The high CI and HR values indicate a powder with poor flow characteristics based on the scale of powder flowability according to USP [20].

As regards to flowability, as less compressible is the powder, less cohesive it will be and for that reason better flow it will have [42,43]. In the case of HA-VAN₂₅, the differences between bulk and tap densities evidenced a high compressibility, leading to a poor flow property, which has been also seen previously for sodium hyaluronate spray-dried particles by Martinelli *et al.* [35].

In addition, in a preliminary determination of the capability of the powder to reach the deep lung, the particle size distribution was measured by laser diffraction, which is correlated with the aerodynamic particle size. HA-VAN₂₅ presented a $Dv_{90} = (6.37 \pm 0.07) \mu\text{m}$, $Dv_{50} = (2.90 \pm 0.02) \mu\text{m}$ and $Dv_{10} = (1.19 \pm 0.02) \mu\text{m}$, meaning that 90 %, 50 % and 10 % of the particle population, respectively, has a particle size below those respective values. Besides, a SPAN value of (1.79 ± 0.03) was measured, which indicates a monodisperse distribution of particle size. These are promising results, where more than half of the population have a geometric size below $5 \mu\text{m}$ within the acceptable range being suitable for deep lung deposition using only the HA-VAN₂₅ complex, without other pharmaceutical excipients.

3.4. In Vitro Biopharmaceutical Performance of HA-VAN₂₅ Complex

The *in vitro* deposition distribution study after the aerosolization of complex powder loaded in rigid HPMC capsules using the NGI equipment was performed. Table 1 summarized the parameters measured, including ED, EF, FPF, Extra-FPF and MMAD, that can be related to the capability of the powder to be transported through the patients' airways. Despite the poor flowability evidenced, the powder provided a good emitted fraction (above 80%), when aerosolized with RS01 device. It appears evident that the agglomerates of HA-VAN₂₅ spray dried microparticles highlighted by SEM analysis and the cohesive forces providing poor flow were disrupted during the capsule spinning, powder extraction and subsequent impaction on the walls and de-agglomeration grid of the RS01 device. The FPF value showed that about 43% of the powder has the potential to reach the deep lung since the particles have an aerodynamic diameter lower than $5 \mu\text{m}$, while the 26% of the particles can go deeper until the alveolus. The MMAD exhibited that 50% of the population have a size lower than $4.29 \mu\text{m}$ which is in good agreement with the PSD obtained by laser diffraction.

Table 1. Aerodynamic parameters of HA-VAN₂₅ spray-dried particles obtained by NGI aerosolization.

ED (mg)	EF (%)	FPF (%)	Extra-FPF (%)	MMAD (μm)
7.9 ± 0.4	84.2 ± 0.1	42.9 ± 0.2	25.9 ± 0.1	4.29 ± 0.03

ED: emitted dose, EF: emitted fraction, FPF: fine particle fraction, Extra-FPF: extra fine particle fraction and MMAD: mass median aerodynamic diameter.

The drug's aerodynamic particle size distribution reported in Figure 5 revealed that approximately 16% of the VAN was not emitted from the capsule and the inhaler device, while the 22% was retained in the IP, which simulates the throat. Those percentages, besides the amount of VAN deposited at stage 1 (9%) correspond to the quantity of drug that will not reach the deep lung. The presence of a high amount of drug from stage 2 onwards is considered desirable for lung deposition [44] since this stage has a cut-off diameter of $4.46 \mu\text{m}$ at the flow rate of 65 L/min, almost half of particles of HA-VAN₂₅ are in these stages. The higher values of VAN at stage 2 and 3 in comparison with the following is consistent with the MMAD value obtained.

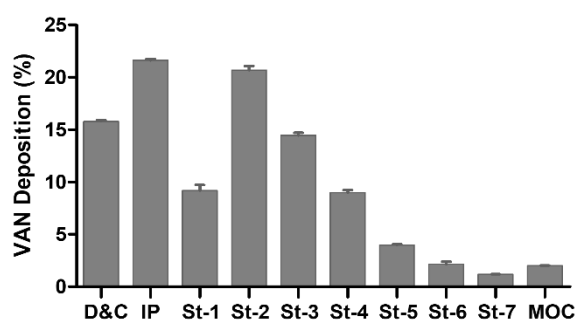


Figure 5. Next Generation Impactor deposition of HA-VAN₂₅ in the different stages of the equipment: device and capsule (D&C), induction port (IP), stages 1 to 7 (St-1 to St-7) and micro-orifice collector (MOC).

Despite the absence of vehicle excipients frequently used in dry powder inhaler formulations, such as lactose or mannitol, the HA-VAN₂₅ powder presented by itself a satisfactory efficiency for pulmonary administration. Moreover, as demonstrated by the optimal value of VAN emitted fraction, the formation of weak agglomerates could result in the easy dis-aggregation either for the inspiratory force generated by the patients and/or the collisions between the particles or particles-wall inside the inhaler device leading to a proper EF [45]. The suitable aerosolization performance observed is consistent with the results earlier informed where the powder showed a low bulk density, the morphology presented a hollow hemispherical shape and the Dv₅₀ value was about 2.9 μm ; and all these together influence the aerodynamic performance which would lead to an adequate pulmonary deposition [46].

In a previous study by Sullivan *et al.* (2015), the *in vitro* and *in vivo* performance of dry powder Vancomycin hydrochloride (VAN) without further processing was investigated in intubated rabbits, revealing promising results. Intubated rabbits administered a 1 mg/kg dose of VAN via inhalation demonstrated a comparable AUC to those receiving the same dose through a single bolus IV infusion. Notably, inhaled VAN exhibited reduced C_{max} and increased T_{max}, indicating a more sustained pulmonary level of the drug. However, the physicochemical and flow properties obtained by these authors were not the most suitable for inhalation therapy, for example, the VAN showed different particle sizes and shapes; presented high bulk and tap densities of (0.35 \pm 0.01) g/cm³ and (0.51 \pm 0.01) g/cm³, respectively; FPF value was no more than 26%, and MMAD value was close of 7 μm , accompanied by a substantial geometric standard deviation [26]. For this reason, our dry powder formulation, based on HA-VAN₂₅ with better physicochemical and flow properties mentioned before, is expected to overcome these results in terms of *in vivo* VAN performance, emphasizing both efficacy and safety.

Following the impaction process, dry powders deposited within the lungs are required to undergo complex drug absorption processes that include wetting, dissolution, and diffusion. Since no established a specific dissolution method for inhalation products is defined by pharmacopoeias and guidelines, it was used the traditional basket apparatus dissolutor (USP Apparatus 1), which has a relatively large volume of dissolution medium where the powder is dispersed. The dissolution profiles of VAN and the HA-VAN₂₅ complex are depicted in Figure 6. It was evident that the complete amount of pure VAN dissolved within 5 min, while the dissolved proportion of VAN from HA-VAN₂₅ was less than 40% at the same time. The comparison between VAN and HA-VAN₂₅ dissolution profiles showed $f_1 = 40.6$ denoting significant differences ($f_1 > 15$). Given the hydrophilic nature of VAN, a prompt drug dissolution was anticipated. Conversely, the formation of an ionic complex with HA leads to a slower dissolution rate. Additionally, it is noteworthy that the complex, when in contact with the dissolution medium, undergoes gelation and slight swelling due to the presence of the polymer, potentially contributing to the observed reduction of the dissolution rate.

Beyond the differences in the dissolution profiles between VAN and HA-VAN₂₅, it is noticeable that more than 85% of the loaded VAN dissolves within the initial 30 min. This aspect is crucial, as inadequate drug dissolution could potentially induce lung irritation, local side effects, and trigger processes like macrophage phagocytosis or mucociliary clearance of solid particles, ultimately leading to a rapid reduction in lung dose [47].

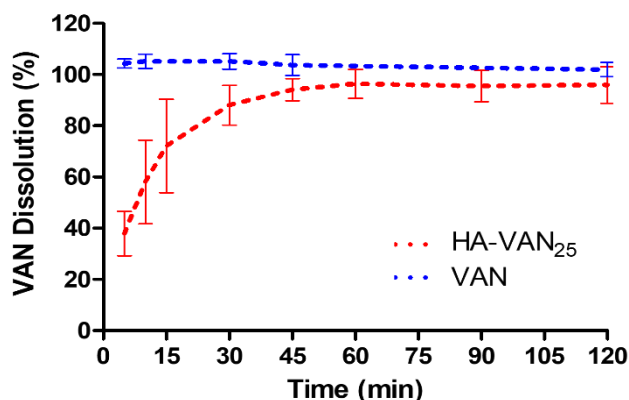


Figure 6. Dissolution profiles of VAN and HA-VAN₂₅ in PBS pH 7.4.

In addition, bicompartamental Franz cells were used in order to study the release behavior of the aqueous HA-VAN₂₅ complex dispersion in comparison with a solution of pure VAN, in two different media: water and PBS pH 7.4.

It can be seen in Figure 7, when water was used as a receptor medium, a fast diffusion of VAN across the dialysis membrane from its solution was observed. However, the release rate of VAN from the aqueous dispersion of its complex was substantially slower. This response can be attributed to the reservoir behavior usually described to PE-D complexes, where the dissociation of ionic complex between HA and VAN is the determining step in the VAN release toward the semipermeable membrane [12,48]. In addition, the electrostatic attraction between the oppositely charged macroions present in the donor compartment, makes the diffusion of ionic species more restricted [14].

When water was replaced by PBS pH 7.4 solution (as a simulated physiological receptor medium), a significant increase of VAN diffusion from the HA-VAN₂₅, in comparison with its release profile in water, was observed, reflected by a f_1 value of 72.6. It is known that the presence of dissolved ions in the receptor medium (from PBS solution) promotes ion exchange from the macromolecular complex microenvironment, increasing the proportion of free ionic species of VAN and consequently significant increase of drug release rates [16].

On the other hand, the similarity of VAN release profiles from pure VAN solution and from the complex, both in PBS medium, ($f_1 = 15$), could be related with the diffusion of ions from the receptor compartment to the donor compartment, promoting the dissociation of VAN HCl into VAN and HCl separately, shifting the acid base equilibrium towards the non-ionized VAN (VAN base) increasing their proportion in solution. The VAN base presents a lower solubility (0.225 mg/mL) than VAN HCl in water (> 100 mg/mL), *i.e.*, the solubility decreases over 400 times [49]. This could be the reason why VAN diffuses faster in water than PBS 7.4, and, at the same time, with a similar release profile than the complex in the saline solution. However, additional experiments assay would be necessary to confirm this observation.

Finally, kinetic analysis of the release profiles showed a strong fit to the power equations (R^2 0.96 and 0.99) above the Higuchi diffusion model. The values of diffusional exponent n between 0.5 and 1, suggest an anomalous or non-Fickian release behavior (Table 2). Consequently, the dissociation of ionic pairs between HA and VAN and the later diffusion of VAN from HA-VAN₂₅ seems to be the principal control mechanisms of drug delivery. The polymer chains reorganize slowly, whereas the diffusion process proceeds very rapidly and leads to anomalous time-dependent effects as it was observed in the majority of PE-D systems [50].

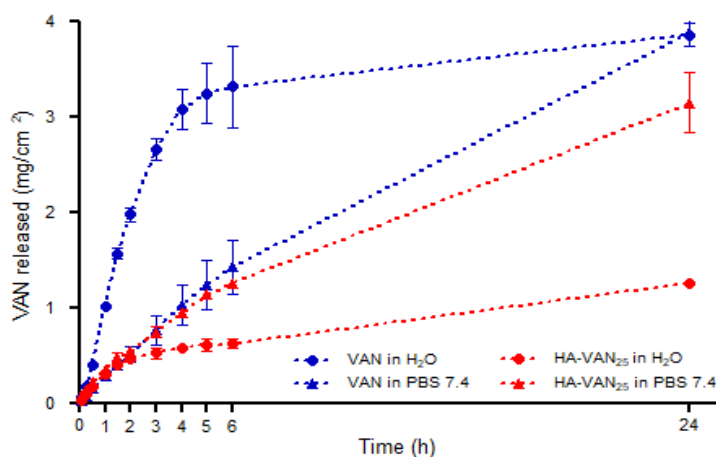


Figure 7. Release profiles of VAN and HA-VAN₂₅ towards water and PBS pH 7.4.

Table 2. Kinetic data obtained from HA-VAN₂₅ towards water and PBS pH=7 using Peppas equation.

	Receptor medium					
	H ₂ O			PBS 7.4		
	<i>k</i>	<i>n</i>	<i>R</i> ²	<i>k</i>	<i>n</i>	<i>R</i> ²
HA-VAN ₂₅	0.5	0.60	0.96	0.4	0.67	0.99

3.5. Antibacterial Activity Tests

The agar diffusion test was carried out to assess the susceptibility against *Staphylococcus aureus* reference strains, specifically strains 29213, 25923, and 43300. The results, presented in Table 3, indicate that HA does not exhibit inhibition, displaying a halo of 8 mm corresponding to the diameter of the original well. HA itself is not considered inherently antimicrobial. In contrast, VAN demonstrates an inhibition halo ranging between 29–31 mm, depending on the strain, and the complex exhibits a similar inhibition pattern. Notably, none of these samples demonstrates reduced inhibition against the MRSA strain.

Table 3. Halo of inhibition obtained for HA, VAN, HA-VAN₂₅ against methicillin-sensitive *Staphylococcus aureus* (MSSA) and methicillin-resistant *Staphylococcus aureus* (MRSA) ATCC strains.

Samples	29213	25923	43300
HA	8 mm±0.0	8 mm±0.0	8 mm±0.0
VAN	29 mm±0.5	31 mm±0.5	30 mm±0.5
HA-VAN ₂₅	30 mm±0.5	31 mm±0.5	31 mm±0.5

The antibacterial efficacy of HA-VAN₂₅ was assessed by examining MIC and MBC against *Staphylococcus aureus*. Table 4 reveals that not only does VAN exhibit antibacterial activity, but so does HA-VAN₂₅ against MRSA and MSSA. Remarkably, HA alone does not demonstrate inhibitory or bactericidal activity in concordance with the absence of halo inhibition previously mentioned. In contrast, VAN exhibits the same MIC and MBC values, consistent with its recognized bactericidal nature and these values are between 4 and 8 µg/mL in agreement with previous findings [51,52].

Notably, HA-VAN₂₅ demonstrates comparable antimicrobial activity with VAN, underscoring that the formation of the complex preserves the well-established antibacterial efficacy of VAN against both MSSA and MRSA strains while simultaneously offering the advantages of being complexed with HA.

Table 4. Minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC) values in $\mu\text{g/mL}$ obtained for HA, VAN and HA-VAN₂₅ against MSSA and MRSA ATCC strains.

Samples	29213		25923		43300	
	MIC	MBC	MIC	MBC	MIC	MBC
HA	> 2500	--	> 2500	--	> 2500	--
VAN	4.88	4.88	4.88	4.88	4.88	4.88
HA-VAN ₂₅	4.88	4.88	4.88	4.88	4.88	4.88

4. Conclusions

A dry powder based on biocompatible HA-VAN₂₅ complex was successfully obtained using a simple and scale-up method without organic solvents. The characterization of the solid state confirmed the ionic interaction between HA and VAN, and the formation of a new entity with properties different to the raw materials. The studies of morphology, size and bulk and tap densities of the powder revealed that it is adequate to reach deep lungs with a spherical shape, decreasing the possibilities to irritate the lungs. The HA-VAN₂₅ complex aerodynamic performance showed that it is suitable for pulmonary administration. The dissolution of the complex was fast and the *in vitro* release showed an extended release of VAN. In addition, the HA-VAN₂₅ complex showed the preservation of the antibacterial efficacy of VAN while simultaneously offering the advantages of being complexed with HA. This unique combination not only maintains the antimicrobial activity of VAN, but also provides additional benefits, such as mitigating irritation and inflammation, and hydrating the airways. These characteristics make the complex highly promising for potential applications in respiratory health, promising a more effective and well-tolerated approach to combat bacterial infections caused by MRSA in patients with CF, improving mainly their quality of life.

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