

*Supporting Information*

*for*

**Preparation of Golgi apparatus-targeted polymer dots of high-color-purity near infrared fluorescence for long-term bioimaging**

Yiping Lu,<sup>1</sup> Wei Song,<sup>3</sup> Zhiquan Tang,<sup>1</sup> Wenru Shi,<sup>1</sup> Shumei Gao,<sup>1</sup> Jun Wu,<sup>2,\*</sup>

Yuan Wang,<sup>1</sup> Hu Pan,<sup>1,\*</sup> Yangang Wang,<sup>1</sup> Hong Huang<sup>1,\*</sup>

<sup>1</sup> College of Biological, Chemical Science and Engineering, Jiaxing University,  
Jiaxing 314001, China

<sup>2</sup> College of Advanced Materials Engineering, Jiaxing Nanhu University,  
Jiaxing 314001, China

<sup>3</sup> Institute for Agri-food Standards and Testing Technology, Shanghai Academy  
of Agricultural Science, Shanghai 201403, China

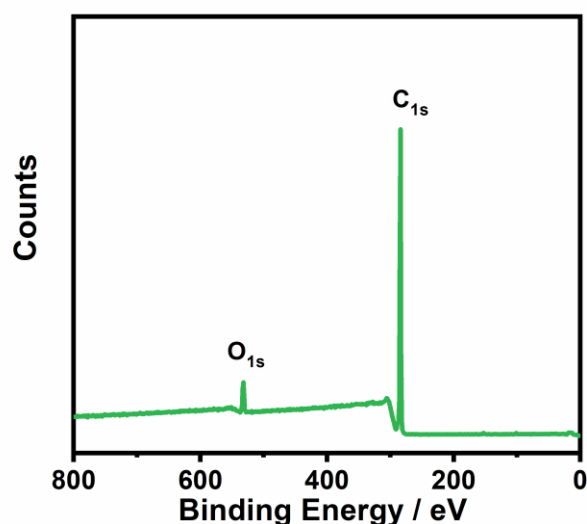
## **Characterizations**

Absorption spectra were recorded on a Shimadzu UV-2550 spectrophotometer. Fluorescence spectra were measured on a F-4600 fluorometer (Hitachi). The fluorescence lifetime and quantum yield were measured by time-correlated single-photon counting on a FLS920 spectrometer. Scanning transmission electron microscopy (STEM) experiments were performed on a Thermo Scientific Talos F200X transmission microscope working at 200 kV. Atomic force microscopic (AFM) characterizations were conducted in the ScanAsyst mode under ambient conditions (Bruker). X-ray photoelectron spectroscopy (XPS) data was collected using a thermoelectron instrument (Thermo Scientific ESCALAB250). Fourier transform infrared spectroscopy (FTIR) spectrum of the samples were acquired by applying a Nicolet iS10 FTIR spectrometer. The contact angles of water on a film of CDs were measured on a JC2000D contact angle goniometer (Powereach). Confocal fluorescence and bright field images (512 × 512 pixels) were acquired with a Leica TCS-SP8 confocal laser scanning microscope. Cells images were captured using a 63× objective lens.

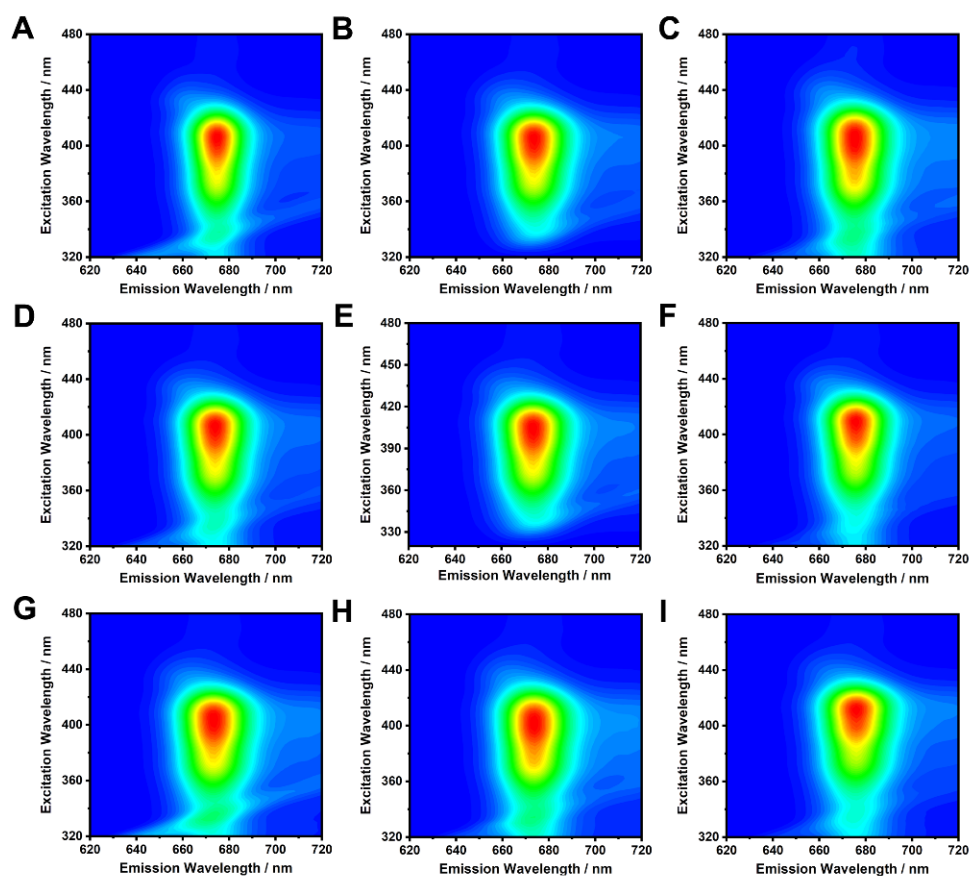
## **Cytotoxicity evaluations**

HeLa cells were fostered at a density of  $\sim 1 \times 10^4$  cells per well in 96-well plates, and grown in DMEM containing 10% fetal bovine serum, 80 U mL<sup>-1</sup> penicillin, and 80 μg mL<sup>-1</sup> streptomycin in a humid incubator with 5% CO<sub>2</sub>/95% air. 12 h later, the culture media were exchanged to fresh one containing Golgi-Pdots of

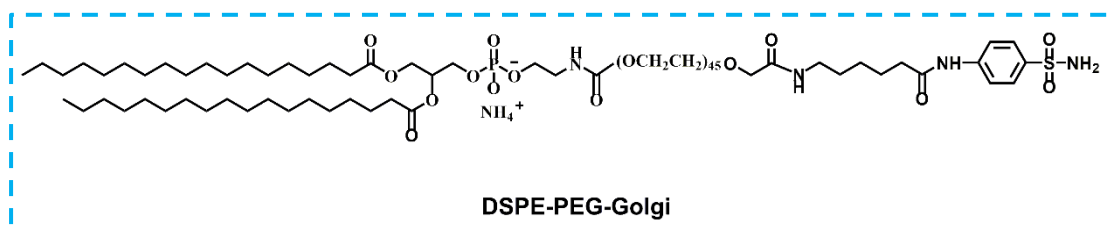
different concentrations (0-100  $\mu\text{g mL}^{-1}$ ) and cultured for 48 h. As to each concentration, five independent experiments were performed. Following, 20  $\mu\text{L}$  of MTT solution (1.0  $\text{mg mL}^{-1}$ ) was introduced into each well, cultivating for 4 h to allow the formation of formazan crystals. Subsequently, 150  $\mu\text{L}$  of DMSO was injected to the wells. Absorbance (A) of the resultant hybrid at 570 nm was measured. Cellular viability values were quantified according to the following equation: cellular viability (%) =  $A_{\text{test}}/A_{\text{control}} \times 100\%$ , where  $A_{\text{control}}$  refers to the absorbance recorded from the control group, and  $A_{\text{test}}$  refers to the absorbance obtained with the existence of Golgi-Pdots.



**Figure S1** XPS survey spectrum of the CDs.



**Figure S2** Excitation-emission contour plots of the CDs in different organic solvents: (A) petroleum ether, (B) DCM, (C) n-butanol, (D) ethyl acetate, (E) dioxane, (F) acetone, (G) acetonitrile, (H) methanol, and (I) DMSO.



**Figure S3** Chemical structure of DSPE-PEG-Golgi.

