

Review

Not peer-reviewed version

The Nerve Growth Factor Receptor (NGFR/P75^{ntr}): A Major Player in Alzheimer's Disease

[Francesco Bruno](#)*, [Paolo Abondio](#)*, Alberto Montesanto, [Donata Luiselli](#), [Amalia Cecilia Bruni](#), Raffaele Maletta

Posted Date: 13 January 2023

doi: 10.20944/preprints202301.0239.v1

Keywords: Alzheimer's disease; Nerve growth factor receptor; NGFR; p75NTR; amyloid-beta; expression; signaling pathways; neuropathology; diagnosis; treatment.



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

The Nerve Growth Factor Receptor (NGFR/p75^{NTR}): A Major Player in Alzheimer's Disease

Francesco Bruno ^{1,*}, Paolo Abondio ^{2,3,*}, Alberto Montesanto ⁴, Donata Luiselli ², Amalia C. Bruni ^{1,†} and Raffaele Maletta ¹

¹ Regional Neurogenetic Center (CRN), Department of Primary Care, ASP Catanzaro, 88046 Lamezia Terme, Italy

² Laboratory of Ancient DNA, Department of Cultural Heritage, University of Bologna, Via degli Ariani 1, 48121 Ravenna, Italy

³ Laboratory of Molecular Anthropology and Center for Genome Biology, Department of Biological, Geological and Environmental Sciences, University of Bologna, Via Selmi 3, 40126 Bologna, Italy

⁴ Department of Biology, Ecology and Earth Sciences, University of Calabria, 87036 Rende, Italy

* Correspondence: francescobrunofb@gmail.com (F.B.); paolo.abondio2@unibo.it (P.A.)

† Amalia C. Bruni is former Director of the Regional Neurogenetic Center (CRN).

Abstract: Alzheimer's disease (AD) represents the most prevalent type of dementia in elderly people, primarily characterized by brain accumulation of beta-amyloid (A β) peptides, derived from Amyloid Precursor Protein (APP), in the extracellular space (*amyloid plaques*) and intracellular deposits of the hyperphosphorylated form of the protein tau (p-tau; *tangles or neurofibrillary aggregates*). The Nerve growth factor receptor (NGFR/p75^{NTR}) represents a low-affinity receptor for all known mammals neurotrophins (i.e., pro-NGF, NGF, BDNF, NT-3 e NT-4/5) and it is involved in pathways that determine both survival and death of neurons. Interestingly, also A β peptides can bind to NGFR/p75^{NTR} making it the "ideal" candidate in mediating A β -induced neuropathology. Besides pathogenesis and neuropathology, several data indicated that NGFR/p75^{NTR} could play a key role in AD also from a genetic perspective. Other studies suggested that NGFR/p75^{NTR} could represent a good diagnostic tool, as well as a promising therapeutic target for AD. Here, we comprehensively summarize and review the current experimental evidence on this topic.

Keywords: Alzheimer's disease; Nerve growth factor receptor; NGFR; p75^{NTR}; amyloid-beta; expression; signaling pathways; neuropathology; diagnosis; treatment

1. Introduction

Alzheimer's disease (AD) represents the most prevalent type of dementia in elderly people [1], primarily characterized by brain accumulation of beta-amyloid (A β) peptide in the extracellular space (*amyloid plaques*) and intracellular deposits of paired helical filaments (PHF), mainly formed by the hyperphosphorylated form of the protein tau (p-tau) (*tangles or neurofibrillary aggregates*) [2,3]. The main clinical feature of AD consists of increasing cognitive decline over time, often anticipated, or accompanied by a set of behavioral disturbances [4–6]. Two major forms of the disease exist: sporadic (sAD)-the causes of which are not yet completely understood and can be related to several hypotheses- and familial (fAD)-inherited within families from generation to generation even with a clear autosomal dominant transmission of mutations in Presenilin 1 (*PSEN1*), 2 (*PSEN2*) or Amyloid Precursors Protein (*APP*) genes [7–9]. In both forms, amyloid plaques and neurofibrillary tangles are accompanied by neuronal and synaptic loss [10], microgliosis [11], astrogliosis [12], and neuroinflammation [13].

Amyloid plaques, originate from the APP which is a membrane glycoprotein with a large N-terminal glycosylated domain on the extracellular side and a C-terminal domain on the intracellular side of the cell membrane [14]. Under physiological conditions, APP is processed by two distinct pathways, one non-amyloidogenic (α -secretases pathway, e.g., ADAM10 and ADAM17) and one

amyloidogenic (β -secretase pathway), the latter of which leads to the production of $A\beta$ peptides (e.g., $A\beta_{1-42}$ and $A\beta_{25-35}$) [15]. Specifically, BACE1 is the β -secretase enzyme that cleaves the transmembrane APP and, together with γ -secretase, generates $A\beta$ species that form increasingly large and conformationally complex soluble aggregates [15–18]. Over the years, several hypotheses have been formulated to explain how the neuropathological features of AD are causally related to each other underpinning its pathogenesis. Among these, the most debated in scientific communities is the “amyloid cascade hypothesis” which posed that the progressive accumulation of $A\beta$ in the brain triggers a complex cascade of events that result in the tau hyperphosphorylation, loss of synapses, glial activations, progressive deficiency of neurotransmitters, and neural death [19,20]. However, this hypothesis has been subject to several criticisms or alternative interpretation because of lack of a strong link that can explain how $A\beta$ can cause all these neuropathological phenotypes [21]. Interestingly, an increasing body of evidence indicates that the nerve growth factor receptor (NGFR), also known as p75 neurotrophin receptor (p75^{NTR}), could represent the “missing piece” to elucidate the neuropathological mechanism underpinning both fAD and sAD.

The NGFR/p75^{NTR} protein is encoded by the *NGFR/p75^{NTR}* gene, located on the long arm of chromosome 17 (17q21.33). At least two isoforms of NGFR/p75^{NTR} exist: a full-length isoform and a short isoform (s-NGFR/p75^{NTR}) both of which are expressed in nerve cells [22]. The full-length isoform includes six exons and five introns [23]. The resulting protein comprises an extracellular domain that contains four cysteine-rich motifs (CDR1-CRD4), a transmembrane domain, and an intra-cellular domain formed by a juxtamembrane domain (Chopper) and a death domain (Figure 1) [24,25]. The s-NGFR/p75^{NTR} isoform results from alternative splicing of exon III of the *NGFR/p75^{NTR}* locus which encodes for the CDR 2–4 [26] and consequently lacks the neurotrophin-binding site. For this reason, while the full-length isoform can bind all known neurotrophins, the s-NGFR/p75^{NTR} isoform does not interact with several molecules including Nogo, sortilin and Trk (Figure 1) [27,28]. Sabry and colleagues also reported a human NGFR/p75^{NTR} 36 kD isoform not yet fully characterized [28].

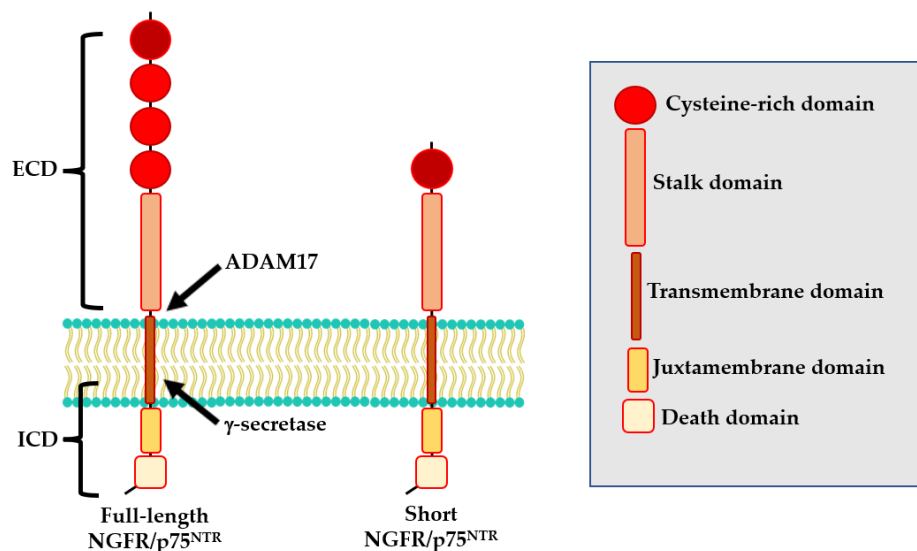


Figure 1. Structure of NGFR/p75^{NTR} isoforms.

NGFR/p75^{NTR} represents a low-affinity receptor for all known mammalian neurotrophins—i.e., nerve growth factor (NGF), brain derived neurotrophic factor (BDNF), NT-3, and NT4/5 [29]—and a higher affinity receptor for their immature forms (i.e., pro-NGF, pro-BDNF) [30,31]. The ligand binding to NGFR/p75^{NTR} mainly causes the activation of apoptosis signaling pathways [9,32]. However, these neurotrophins can also bind other receptors—i.e., NGF to TrkA, BDNF and NT-4/5 to TrkB whereas NT-3 to TrkC—promoting downstream signaling involved in neuronal survival and differentiation [33]. Other findings also showed that NGFR/p75^{NTR} can regulate axonal growth, cell cycle, and synaptic plasticity [34]. The specificity of the signaling pathways to be activated (i.e., death

vs survivor) is reached through the formation of NGFR/p75^{NTR} molecular complexes with different co-receptors, such as Trk family, sortilin and Nogo receptors [35].

Moreover, preliminary data indicated that NGFR/p75^{NTR} can also undergo epigenetic modifications such as methylation and histone acetylation. For example, it has been shown that ibuprofen can increase the mRNA and protein levels of NGFR/p75^{NTR} by inducing its promoter hypomethylation and by increasing N⁶-methyladenosine (m6A)-NGFR/p75^{NTR} expression in cancer cell lines [36].

Interestingly, NGFR/p75^{NTR} could also undergo to a three-step proteolytic cleavage that could modify its functional properties [37,38]. During the first step the extracellular domain (ECD) of NGFR/p75^{NTR} is cleaved by ADAM17, generating a membrane-bound C-terminal fragment (CTF). Then, CTF undergoes to a second cleavage by the PSEN-dependent γ -secretase, releasing the intracellular domain (ICD) into the cytoplasm [29].

Given the multiple functions and implications of NGFR/p75^{NTR} in central nervous system (CNS), this review will explore the genetic and molecular evidence surrounding its relationship with AD, the experimental evidence around NGFR/p75^{NTR}-driven AD neuropathology, as well as the relevance of this protein as a prognostic-diagnostic tool and therapeutic target for AD.

2. Expression and Signaling Pathways of NGFR/p75^{NTR} in AD

NGFR/p75^{NTR} receptors are highly expressed of both neurons and glial cells during the CNS development [39] and after brain injury [40], whereas in the normal adulthood are present only in specific brain regions such as neurons and astrocytes of hippocampal cornus ammon 1 (CA1) [41] and especially in the cholinergic neurons (CNs) of the basal forebrain-of which antibodies against these receptors represent the most used immunohistochemical marker [42–44]. The CNs of the basal forebrain send long projections to several cortical areas, including the entorhinal cortex and visual cortex, as well as the amygdala, resulting in several cholinergic circuits involved in the regulations of working memory, visual discrimination, and attention [45].

It has long been known that the CNs of the basal forebrain play a critical role in AD for different reasons: (i) the AD neurodegeneration could originate in these neurons and only then may affect other brain regions; (ii) CNs represent the type of neurons most affected in AD; (iii) CNs degeneration predicts the cortical spread of AD neuropathology [46]. Interestingly, TrkA expression is reduced in early and late stages of AD [47–49] whereas the NGFR/p75^{NTR} expression is not affected or is increased in AD-damaged CNs of the basal forebrain [49–51]. Besides basal forebrain, AD patients showed higher levels of NGFR/p75^{NTR} receptors also in cortical neurons [52] and hippocampi [53,54]. Since NGFR/p75^{NTR} mediates apoptotic signaling in the absence of TrkA [40,55] the imbalance ratio of NGFR/p75^{NTR} and TrkA may result in an increased programmed cell death. Sáez and colleagues [56] demonstrated that the A β ₂₅₋₃₅ exerts an indirect neurotoxic effect by stimulating the astrocytes synthesis of NGF that in turn causes tau hyperphosphorylation and hippocampal neuron death through NGFR/p75^{NTR} [56]. Therefore, NGFR/p75^{NTR} and TrkA receptors differentially regulate APP processing, promoting the non-amyloidogenic and amyloidogenic pathways, respectively. In particular, the binding of NGF to NGFR/p75^{NTR} leads to the activation of the second messenger ceramide and then to the β -secretase cleavage of APP [57].

Interestingly, an increasing amount of evidence indicates the pivotal role of NGFR/p75^{NTR} in amyloidogenesis and in A β -induced neurotoxicity due to the binding of A β to these receptors. In particular, Devarajan and Sharmila [58] demonstrated that A β ₁₋₄₂ specifically recognizes CRD1 and CRD2 domains of the NGFR/p75^{NTR} receptor and forms a “cap” like structure at the N-terminal that is stabilized by a network of hydrogen bonds [58].

Moreover, the binding of A β ₁₋₄₂ to NGFR/p75^{NTR} increases the extracellular levels of A β ₁₋₄₂ [59], causes neurotoxicity [55] by signaling pathways that involve JNK, NADPH oxidase, and caspases-9/3, as well as by an alternative PLAIDD pathway [60–62], mediates A β -induced tau hyperphosphorylation and neurodegeneration (i.e., synaptic disorder and neuronal loss) through the calpain/CDK5 and AKT/GSK3 β pathways [63] and increases the expression of another pro-apoptotic neurotrophin receptor, sortilin via the RhoA pathway [64] that in turn could interact with

NGFR/p75^{NTR} to induce neuronal apoptosis [65]. Chakravarthy and colleagues suggested that A β accumulation in turn upregulates the expression of membrane-associated NGFR/p75^{NTR} through insulin-like growth factor 1 receptor (IGF-1R) phosphorylation [54,66]. Moreover, Hu and colleagues reported in 2013 that the death receptor 6 (DR6) forms a receptor complex with NGFR/p75^{NTR} that mediates A β -induced neurotoxicity in cortical neurons [67]. Finally, preliminary data also indicated that APP and pro-NGF could directly interact with NGFR/p75^{NTR} increasing apoptosis in AD [68–70] (Figure 2).

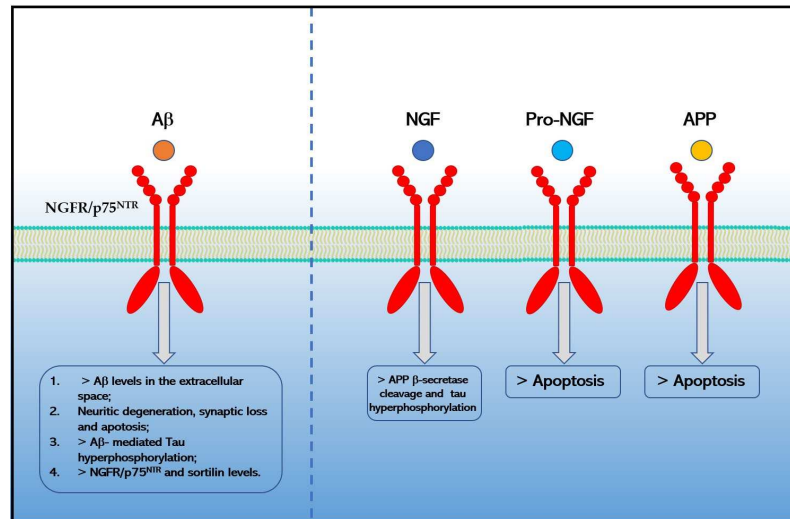


Figure 2. Effects caused by the binding of A β ₁₋₄₂, NGF, pro-NGF and APP to NGFR/p75^{NTR} in Alzheimer's Disease.

Interestingly, several data indicated that NGFR/p75^{NTR} ECD, derived from the proteolytic cleavage by ADAM-17 followed by regulated-intramembranous proteolysis by γ -secretase, represents a protective factor against AD-neuropathology [71]. This effect is probable due to the fact that the NGFR/p75^{NTR} ECD contains the binding sites for A β -i.e., CRD1 and CRD2 domains-and for other ligands that can induce apoptosis through the activation of these receptors [72]. Thus, the impaired NGFR/p75^{NTR} ECD shedding and the imbalanced levels of NGFR/p75^{NTR} and NGFR/p75^{NTR} ECD could play an important role in regulating the production and clearance of A β [73]. Moreover, the *PSEN1* M146V mutation increases γ -secretase cutting of NGFR/p75^{NTR} *in vitro* and potentially exacerbates the pathogenic outcomes observed in fAD [74]. He and colleagues also reported an overproduction of naturally occurring autoantibodies against NGFR/p75^{NTR} ECD in AD patients, strengthening the role of the immune system and immunosenescence in the AD pathogenesis [73].

Further evidence about the involvement of NGFR/p75^{NTR} in AD derives from inactivation and ablation studies. Indeed, the 5xFAD mice, which expresses a signaling-deficient variant of the NGFR/p75^{NTR}, shows a greater neuroprotection from AD neuropathology than animals lacking this receptor [75]. At the same manner, the ablation of NGFR/p75^{NTR} signaling protects hippocampal network against pathophysiological changes observed in AD (e.g., A β -induced degradation of gamma oscillations and gamma-theta interaction) *in vitro* [76].

Thus, the involvement of NGFR/p75^{NTR} in the AD-associated brain alterations appears to be multifactorial and encompasses several ligands, including A β , NGF, pro-NGF and APP, as well as the activity of ADAM17 and γ -secretases, although the complete set of elements needed by the neurons to enter the apoptotic program in response to these interactions remains to be determined.

3. NGFR/p75^{NTR} Genetic Variants and AD

The first evidence of a genetic involvement of NGFR/p75^{NTR} in AD emerged from a study of Cozza and colleagues [77]. The authors analyzed the genetic variability of four functional NGFR/p75^{NTR} polymorphisms (i.e., rs2072445 and rs2072446 located in exon 4, rs741072 located in exon 6, rs734194 located in the 3'-UTR) and the risk of developing AD in an Italian sample consisting

of 151 sAD patients, 100 fAD patients, and 97 healthy subjects. fAD was defined as those patients with at least two first-degree relatives in two generations affected by AD in absence of mutations in *PSEN1*, *PSEN2* and *APP* genes. From the results of this study emerged a significant association between the rs2072446 variant and the risk of fAD. Since the polymorphism is located near the genetic region that codes for the cleavage region recognized by ADAM17, its variability may affect the cleavage efficiency of NGFR/p75^{NTR} and thus the release of NGFR/p75^{NTR} ECD [73]. However, this association did not remain significant after correction for multiple comparisons. In a second study, Cheng and colleagues [78] examined the genetic variability of the four SNPs previously analyzed by Cozza and collaborators [77] together with the rs741073 variant, located on 3'-UTR, and the risk of developing AD. In a Chinese sample consisting of 264 sAD patients and 389 healthy controls the authors found that the rs734194 variant was significantly associated with a decreased risk of AD. However, also in this case, this association did not remain significant after correction for multiple comparisons. Moreover, Matyi and colleagues [79] analyzed the association between the rs2072446 variant and the risk of AD in a population sample of older adults from the Cache County Study on Memory in Aging (CCSMA) of Utah, consisting of 396 AD and 3,272 controls. They found a significant association between the genetic variability of the rs2072446 polymorphism and the risk of developing AD only in females. In particular, they found that compared to male non carriers, female carriers of the minor T allele of rs2072446 showed a 60% higher risk of developing AD in contrast with the finding previously reported by Cozza [77]. Through the analysis of an Australian sample consisting of 258 cases and 247 controls, Vacher and colleagues [80] found a significant association between the variant rs9908234, located in the intron 1 of the *NGFR/p75^{NTR}* gene, and the brain accumulation of A β . More recently, He and collaborators [81] conducted a case-control association study (366 sAD patients and 390 age- and sex-matched controls) in a Chinese Han population. By analyzing the variability of twelve tag-SNPs within the *NGFR/p75^{NTR}* gene located in the promoter region (rs603769 and rs2584665), in intron 1 (rs9908234 and rs3785931), in intron 3 (rs2537706 and rs534561), in exon 4 (rs2072446) and in exon 6 (rs7219709, rs1804011, rs734194, rs741072, and rs741073), they found that the rs2072446-T allele was significantly associated with an increased risk of sAD (OR = 1.79), replicating the finding previously reported by Cozza and colleagues [77] in an Italian population sample. Interestingly, by analyzing another cohort (279 cognitive normal, 480 MCI, and 47 AD) from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database it has also been shown that T allele at the rs2072446 variant was also associated with the heavier A β burden, which further contributed to an increased risk of AD progression in APOE ϵ 4 non-carrier. Probably the heterogeneity of these results might be due to the presence of sex- and population-specific associations that might be also modified by socio-economic and lifestyle factors and epistatic genetic effects that could be more efficiently captured in large population samples with more accurate and detailed clinical data. Thus, other studies are needed to better understand the role of *NGFR/p75^{NTR}* variants in different populations and selecting a set of more informative SNPs to capture most of the genetic variations of the entire gene.

4. NGFR/p75^{NTR} as a Biomarker of AD

The evidence that NGFR/p75^{NTR} could represent a specific biomarker for AD emerged in 2015 from a study of Yao and colleagues [71], who reported a significant reduction of NGFR/p75^{NTR} ECD levels in cerebrospinal fluid (CSF) and in the brains of AD patients and of *APP/PSEN1* double-transgenic mice, due to the A β -induced reduction in the expression and activity of ADAM17. In the same period, Jiao and collaborators [82] reported a distinct and typical NGFR/p75^{NTR} ECD profile in AD patients, characterized by a decreased CSF and an increased serum level, compared to other diseases (i.e., Parkinson disease, stroke) as well to the control group (elderly people without neurological disorders). The serum and CSF levels of NGFR/p75^{NTR} ECD also correlated with the Mini-Mental State Examination (MMSE) scores in AD patients and thus its detection could represent a good tool for the differential diagnosis. In addition, the combination of CSF A β ₁₋₄₂, CSF A β ₄₂₋₄₀, CSF ptau₁₈₁ or CSF ptau₁₈₁/A β ₁₋₄₂ with CSF NGFR/p75^{NTR} ECD improves the diagnostic accuracy. More recently, He and colleagues [73] investigated the presence and alterations of naturally occurring

autoantibodies against NGFR/p75^{NTR} ECD in AD patients, as well as their effects on AD pathology. From the results of this study emerged an increased levels of these naturally occurring antibodies in the CFS and a negative association between these and the CFS levels of NGFR/p75^{NTR} ECD in AD patients. Interestingly, transgenic AD mice actively immunized with NGFR/p75^{NTR} ECD showed a lower level of NGFR/p75^{NTR} ECD and a more severe AD pathology in the brain and worse cognitive functions compared to two different control groups immunized with the reverse sequence of NGFR/p75^{NTR} ECD and phosphate-buffered saline, respectively. Moreover, Crispoltoni and collaborators [83] examined the relationship between the plasma levels of NGF as well as the expression levels of TrkA and NGFR/p75^{NTR} in monocytes of patients with mild cognitive impairment (MCI), mild and severe AD. The authors reported an increased concentration of plasma NGF accompanied by a higher expression of TrkA, but not of NGFR/p75^{NTR}, in monocytes from patients with MCI and mild AD. Conversely, in patients with severe AD it has been reported a decreased NGF plasma concentration whereas in the monocytes, the expression of TrkA and NGFR/p75^{NTR} were decreased and increased, respectively, and were associated with caspase 3-mediated apoptosis. Thus, this study reported a plasmatic-NGF and monocytic TrkA and NGFR/p75^{NTR} variation during the progression from MCI to severe AD. However, as far as we know, these studies did not carry out a genetic screening on AD patients to verify the presence of mutations in the *PS1*, *PS2* and *APP* genes or have included patients with no family history of dementia. Further studies are needed to understand whether these findings can be applied to both sporadic and genetic AD patients, taking into account gender differences as well.

5. NGFR/p75^{NTR} as a Therapeutic Target for AD

Given the results discussed so far regarding the involvement of NGFR/p75^{NTR} in the neuropathology of AD, since the mid-2000s, several research groups have tested *in vivo* and *in vitro* the effect of several molecules capable of blocking or altering the expression of these receptor for the treatment of this currently incurable disease [84]. These include LM11A-31 [85], CATDIKGAEC [86,87] and sulforaphane [88]. Furthermore, other researchers have experimented the efficacy of the administration of NGFR/p75^{NTR} ECD in counteracting AD phenotype [63,89].

LM11A-31 is a small non-peptide NGFR/p75^{NTR} ligand able to selectively activate survival signaling and simultaneously inhibit pro-apoptosis pathways [90]. Several animal data indicated that this molecule is able to inhibit A β - induced neural death and neuritic degeneration, as well as to prevent tau phosphorylation and misfolding, loss of cholinergic neurites, microglia and astrocyte activation, and cognitive decline and to reverse synaptic function [85,91–94]. These encouraging results have prompted the researchers to test its effects in humans as well. Indeed, a phase II clinical trial is currently underway on mild-moderate AD patients (ClinicalTrials.gov accession number NCT03069014; <https://clinicaltrials.gov/ct2/show/NCT03069014?term=LM11A-31-BHS&draw=2&rank=1>).

In 2007, Yaar and colleagues [86,87] tested the efficacy of CATDIKGAEC heterocyclic peptide, homologous to amino acids 28–36 of NGF, in counteracting AD phenotype. Preliminary data showed that also this molecule is able to interfere with Ab₁₋₄₀ signaling and rescue neurons from A β ₁₋₄₀-induced toxicity *in vitro*. In addition, the *in vivo* injections of A β together with CATDIKGAEC into the cerebral cortex of B57BL/6 mice significantly decreased the A β -induced brain inflammation compared to the injection of A β and a control peptide. However, as far as we know, this line of research was no longer pursued. Moreover, in 2017, Zhang and colleagues [88] tested the efficacy of sulforaphane (SFN), a secondary metabolite found in edible cruciferous vegetables, in the treatment of AD. Interestingly, the administration of this molecule to *APP/PSEN1* double-transgenic mice, ameliorated the cognitive dysfunction and protected against the Increment of amyloid plaques. These effects may be associated with an up-regulation of NGFR/p75^{NTR} expression which is mediated, at least partly, by SFN-induced reduction in the expression of histone deacetylases (i.e., HDAC1 and HDAC3) that regulates the NGFR/p75^{NTR} transcription. Other studies are needed to better characterize the effects of this molecule in the attenuation of the AD-phenotype, as well as the underlying molecular mechanisms in which NGFR/p75^{NTR} could be implicated.

Finally, as mentioned above, several data indicated that NGFR/p75^{NTR} ECD could have a protective effect on A β -induced neuropathology and has been proposed as a good candidate AD biomarker [71]. Interestingly, it has also been shown that the restoration of NGFR/p75^{NTR} levels through the intracerebroventricular injection of a soluble NGFR/p75^{NTR} ECD-fusion construct protected Tau P301L mice against pathological tau modifications [63] whereas the intramuscular delivery of an AAV-NGFR/p75^{NTR} ECD vector in *APP/PSEN1* double-transgenic mice ameliorates spatial learning and memory, a reduced A β accumulation in the brain and blood and reduced neurite degeneration, neuronal death, microgliosis, inflammation, and tau phosphorylation [89]. Therefore, also the use of a NGFR/p75^{NTR}-based peripheral approach represents a promising direction to develop novel therapies for AD.

6. Conclusions

The evidence here reported, underline the pivotal role of NGFR/p75^{NTR} in all neuropathological changes observed in AD. In the adult brain, these receptors are highly expressed by the cholinergic neurons CNs of the basal forebrain which represent the type of neurons firstly and most affected in AD. Indeed, the binding of A β to these receptors increases the extracellular levels of A β ₁₋₄₂, causes neurotoxicity, synaptic disorder, and neuronal loss, mediates A β -induced tau hyperphosphorylation and increases the expression of membrane-associated NGFR/p75^{NTR} as well as of its “death” co-receptors sortilin. The presence of reduced NGFR/p75^{NTR} ECD levels in CFS and in the brains of AD patients, which has a protective effect in AD-like pathology since it prevents the binding of A β to NGFR/p75^{NTR}, not only pushes to consider it as an AD powerful biomarker but opens new avenues for understanding why impaired NGFR/p75^{NTR} signaling occurs in this disease. The genome-wide association studies (GWAS) carried out so far indicate that only in part it can be explained by the variability of polymorphisms in the *NGFR/p75^{NTR}* gene, although more extensive and large-scale studies are needed in this topic to detect rare NGFR/p75^{NTR} variants and possible epistatic effects with other genetic variants able to modulate the disease susceptibility. For instance, since NGFR/p75^{NTR} ECD is derived from the proteolytic cleavage by ADAM17, further studies are also needed to investigate the role of this biological and functional interaction in the pathogenesis and neuropathology of AD. Also, since it has been shown that NGFR promoter is subject to epigenetic regulation, it is also important to clarify the role of the gene expression regulation on the AD susceptibility. Nevertheless, the analysis of the epigenetic variability in relation to the AD onset is still poorly investigated. Finally, the use of NGFR/p75^{NTR} antagonist molecules or the administration of NGFR/p75^{NTR} ECD appear to be promising therapeutic approaches for AD; a phase II clinical trial is currently underway on mild-moderate AD patients with the use of the small NGFR/p75^{NTR} ligand LM11A-31.

Author Contributions: F.B. conceptualized the manuscript. F.B., P.A. and A.M. wrote the manuscript. F.B. created the figures. D.L., A.B., and R.M. reviewed and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by funds granted by the Italian Ministry of Health *Ricerca Finalizzata* 2018: SG-2018-12366233.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Breijyeh, Z.; Karaman, R. Comprehensive Review on Alzheimer’s Disease: Causes and Treatment. *Molecules* **2020**, *25*, E5789, doi:10.3390/molecules25245789.
2. Goedert, M.; Spillantini, M.G.; Cairns, N.J.; Crowther, R.A. Tau Proteins of Alzheimer Paired Helical Filaments: Abnormal Phosphorylation of All Six Brain Isoforms. *Neuron* **1992**, *8*, 159–168, doi:10.1016/0896-6273(92)90117-v.
3. Skaper, S.D. Alzheimer’s Disease and Amyloid: Culprit or Coincidence? *Int Rev Neurobiol* **2012**, *102*, 277–316, doi:10.1016/B978-0-12-386986-9.00011-9.

4. Lindeboom, J.; Weinstein, H. Neuropsychology of Cognitive Ageing, Minimal Cognitive Impairment, Alzheimer's Disease, and Vascular Cognitive Impairment. *Eur J Pharmacol* **2004**, *490*, 83–86, doi:10.1016/j.ejphar.2004.02.046.
5. Altomari, N.; Bruno, F.; Laganà, V.; Smirne, N.; Colao, R.; Curcio, S.; Di Lorenzo, R.; Frangipane, F.; Maletta, R.; Puccio, G.; et al. A Comparison of Behavioral and Psychological Symptoms of Dementia (BPSD) and BPSD Sub-Syndromes in Early-Onset and Late-Onset Alzheimer's Disease. *J Alzheimers Dis* **2022**, *85*, 691–699, doi:10.3233/JAD-215061.
6. Laganà, V.; Bruno, F.; Altomari, N.; Bruni, G.; Smirne, N.; Curcio, S.; Mirabelli, M.; Colao, R.; Puccio, G.; Frangipane, F.; et al. Neuropsychiatric or Behavioral and Psychological Symptoms of Dementia (BPSD): Focus on Prevalence and Natural History in Alzheimer's Disease and Frontotemporal Dementia. *Front Neurol* **2022**, *13*, 832199, doi:10.3389/fneur.2022.832199.
7. Abondio, P.; Sarno, S.; Giuliani, C.; Laganà, V.; Maletta, R.; Bernardi, L.; Bruno, F.; Colao, R.; Puccio, G.; Frangipane, F.; et al. Amyloid Precursor Protein A713T Mutation in Calabrian Patients with Alzheimer's Disease: A Population Genomics Approach to Estimate Inheritance from a Common Ancestor. *Biomedicines* **2021**, *10*, 20, doi:10.3390/biomedicines10010020.
8. Bruno, F.; Laganà, V.; Di Lorenzo, R.; Bruni, A.C.; Maletta, R. Calabria as a Genetic Isolate: A Model for the Study of Neurodegenerative Diseases. *Biomedicines* **2022**, *10*, 2288, doi:10.3390/biomedicines10092288.
9. Bruno, F.; Malvaso, A.; Canterini, S.; Bruni, A.C. Antimicrobial Peptides (AMPs) in the Pathogenesis of Alzheimer's Disease: Implications for Diagnosis and Treatment. *Antibiotics (Basel)* **2022**, *11*, 726, doi:10.3390/antibiotics11060726.
10. DeTure, M.A.; Dickson, D.W. The Neuropathological Diagnosis of Alzheimer's Disease. *Mol Neurodegener* **2019**, *14*, 32, doi:10.1186/s13024-019-0333-5.
11. Hansen, D.V.; Hanson, J.E.; Sheng, M. Microglia in Alzheimer's Disease. *J Cell Biol* **2018**, *217*, 459–472, doi:10.1083/jcb.201709069.
12. Jain, P.; Wadhwa, P.K.; Jadhav, H.R. Reactive Astrogliosis: Role in Alzheimer's Disease. *CNS Neurol Disord Drug Targets* **2015**, *14*, 872–879, doi:10.2174/1871527314666150713104738.
13. Heneka, M.T.; Carson, M.J.; Khoury, J.E.; Landreth, G.E.; Brosseron, F.; Feinstein, D.L.; Jacobs, A.H.; Wyss-Coray, T.; Vitorica, J.; Ransohoff, R.M.; et al. Neuroinflammation in Alzheimer's Disease. *The Lancet Neurology* **2015**, *14*, 388–405, doi:10.1016/S1474-4422(15)70016-5.
14. Müller, U.C.; Deller, T.; Korte, M. Not Just Amyloid: Physiological Functions of the Amyloid Precursor Protein Family. *Nat Rev Neurosci* **2017**, *18*, 281–298, doi:10.1038/nrn.2017.29.
15. Kojro, E.; Fahrenholz, F. The Non-Amyloidogenic Pathway: Structure and Function of Alpha-Secretases. *Subcell Biochem* **2005**, *38*, 105–127, doi:10.1007/0-387-23226-5_5.
16. Nalivaeva, N.N.; Turner, A.J. Targeting Amyloid Clearance in Alzheimer's Disease as a Therapeutic Strategy. *Br J Pharmacol* **2019**, *176*, 3447–3463, doi:10.1111/bph.14593.
17. Zhao, J.; Liu, X.; Xia, W.; Zhang, Y.; Wang, C. Targeting Amyloidogenic Processing of APP in Alzheimer's Disease. *Front Mol Neurosci* **2020**, *13*, 137, doi:10.3389/fnmol.2020.00137.
18. Hampel, H.; Hardy, J.; Blennow, K.; Chen, C.; Perry, G.; Kim, S.H.; Vilemagne, V.L.; Aisen, P.; Vendruscolo, M.; Iwatsubo, T.; et al. The Amyloid- β Pathway in Alzheimer's Disease. *Mol Psychiatry* **2021**, *26*, 5481–5503, doi:10.1038/s41380-021-01249-0.
19. Levin, J.; Vöglein, J.; Quiroz, Y.T.; Bateman, R.J.; Ghisays, V.; Lopera, F.; McDade, E.; Reiman, E.; Tariot, P.N.; Morris, J.C. Testing the Amyloid Cascade Hypothesis: Prevention Trials in Autosomal Dominant Alzheimer Disease. *Alzheimers Dement* **2022**, *18*, 2687–2698, doi:10.1002/alz.12624.
20. Wu, T.; Lin, D.; Cheng, Y.; Jiang, S.; Riaz, M.W.; Fu, N.; Mou, C.; Ye, M.; Zheng, Y. Amyloid Cascade Hypothesis for the Treatment of Alzheimer's Disease: Progress and Challenges. *Ageing Dis* **2022**, *13*, 1745–1758, doi:10.14336/AD.2022.0412.
21. Stancu, I.-C.; Vasconcelos, B.; Terwel, D.; Dewachter, I. Models of β -Amyloid Induced Tau-Pathology: The Long and "Folded" Road to Understand the Mechanism. *Mol Neurodegener* **2014**, *9*, 51, doi:10.1186/1750-1326-9-51.
22. von Schack, D.; Casademunt, E.; Schweigreiter, R.; Meyer, M.; Bibel, M.; Dechant, G. Complete Ablation of the Neurotrophin Receptor P75NTR Causes Defects Both in the Nervous and the Vascular System. *Nat Neurosci* **2001**, *4*, 977–978, doi:10.1038/nn730.
23. Wang, Y.-T.; Lu, X.-M.; Shu, Y.-H.; Xiao, L.; Chen, K.-T. Selection of Human P75NTR Tag SNPs and Its Biological Significance for Clinical Association Studies. *Bio-Medical Materials and Engineering* **2014**, *24*, 3833–3839, doi:10.3233/BME-141213.
24. Underwood, C.K.; Coulson, E.J. The P75 Neurotrophin Receptor. *The International Journal of Biochemistry & Cell Biology* **2008**, *40*, 1664–1668, doi:10.1016/j.biochel.2007.06.010.
25. Skeldal, S.; Matusica, D.; Nykjaer, A.; Coulson, E.J. Proteolytic Processing of the P75 Neurotrophin Receptor: A Prerequisite for Signalling?: Neuronal Life, Growth and Death Signalling Are Crucially Regulated by Intra-Membrane Proteolysis and Trafficking of P75(NTR). *Bioessays* **2011**, *33*, 614–625, doi:10.1002/bies.201100036.

26. Casaccia-Bonnel, P.; Gu, C.; Khursigara, G.; Chao, M.V. P75 Neurotrophin Receptor as a Modulator of Survival and Death Decisions. *Microsc. Res. Tech.* **1999**, *45*, 217–224, doi:10.1002/(SICI)1097-0029(19990515/01)45:4/5<217::AID-JEMT5>3.0.CO;2-5.
27. Meeker, R.; Williams, K. The P75 Neurotrophin Receptor: At the Crossroad of Neural Repair and Death. *Neural Regen Res* **2015**, *10*, 721, doi:10.4103/1673-5374.156967.
28. Sabry, M.A.; Fares, M.; Folkesson, R.; Al-Ramadan, M.; Alabkal, J.; Al-Kafaji, G.; Hassan, M. Commentary: Impact of a Deletion of the Full-Length and Short Isoform of P75NTR on Cholinergic Innervation and the Population of Postmitotic Doublecortin Positive Cells in the Dentate Gyrus. *Front Neuroanat* **2016**, *10*, 14, doi:10.3389/fnana.2016.00014.
29. Almeida, R.D.; Duarte, C.B. P75NTR Processing and Signaling: Functional Role. In *Handbook of Neurotoxicity*; Kostrzewa, R.M., Ed.; Springer New York: New York, NY, 2014; pp. 1899–1923 ISBN 978-1-4614-5835-7.
30. Ibáñez, C.F. Jekyll-Hyde Neurotrophins: The Story of ProNGF. *Trends Neurosci* **2002**, *25*, 284–286, doi:10.1016/s0166-2236(02)02169-0.
31. Teng, H.K.; Teng, K.K.; Lee, R.; Wright, S.; Tevar, S.; Almeida, R.D.; Kermani, P.; Torkin, R.; Chen, Z.-Y.; Lee, F.S.; et al. ProBDNF Induces Neuronal Apoptosis via Activation of a Receptor Complex of P75NTR and Sortilin. *J Neurosci* **2005**, *25*, 5455–5463, doi:10.1523/JNEUROSCI.5123-04.2005.
32. Shu, Y.-H.; Lu, X.-M.; Wei, J.-X.; Xiao, L.; Wang, Y.-T. Update on the Role of P75NTR in Neurological Disorders: A Novel Therapeutic Target. *Biomed Pharmacother* **2015**, *76*, 17–23, doi:10.1016/j.biopha.2015.10.010.
33. Lu, B.; Pang, P.T.; Woo, N.H. The Yin and Yang of Neurotrophin Action. *Nat Rev Neurosci* **2005**, *6*, 603–614, doi:10.1038/nrn1726.
34. Ibáñez, C.F.; Simi, A. P75 Neurotrophin Receptor Signaling in Nervous System Injury and Degeneration: Paradox and Opportunity. *Trends Neurosci* **2012**, *35*, 431–440, doi:10.1016/j.tins.2012.03.007.
35. Blöchl, A.; Blöchl, R. A Cell-Biological Model of P75NTR Signaling. *J Neurochem* **2007**, *102*, 289–305, doi:10.1111/j.1471-4159.2007.04496.x.
36. Jin, H.; Wu, Z.; Tan, B.; Liu, Z.; Zu, Z.; Wu, X.; Bi, Y.; Hu, X. Ibuprofen Promotes P75 Neurotrophin Receptor Expression through Modifying Promoter Methylation and N6-Methyladenosine-RNA-Methylation in Human Gastric Cancer Cells. *Bioengineered* **2022**, *13*, 14595–14604, doi:10.1080/21655979.2022.2092674.
37. Vicario, A.; Kisiswa, L.; Tann, J.Y.; Kelly, C.E.; Ibáñez, C.F. Neuron-Type-Specific Signaling by the P75NTR Death Receptor Regulated by Differential Proteolytic Cleavage. *Journal of Cell Science* **2015**, jcs.161745, doi:10.1242/jcs.161745.
38. Bao, X.; Shi, J.; Xie, F.; Liu, Z.; Yu, J.; Chen, W.; Zhang, Z.; Xu, Q. Proteolytic Release of the P75NTR Intracellular Domain by ADAM10 Promotes Metastasis and Resistance to Anoikis. *Cancer Research* **2018**, *78*, 2262–2276, doi:10.1158/0008-5472.CAN-17-2789.
39. Cragolini, A.B.; Friedman, W.J. The Function of P75NTR in Glia. *Trends in Neurosciences* **2008**, *31*, 99–104, doi:10.1016/j.tins.2007.11.005.
40. Nykjaer, A.; Willnow, T.E.; Petersen, C.M. P75NTR--Live or Let Die. *Curr Opin Neurobiol* **2005**, *15*, 49–57, doi:10.1016/j.conb.2005.01.004.
41. Wong, L.-W.; Chong, Y.S.; Lin, W.; Kisiswa, L.; Sim, E.; Ibáñez, C.F.; Sajikumar, S. Age-Related Changes in Hippocampal-Dependent Synaptic Plasticity and Memory Mediated by P75 Neurotrophin Receptor. *Aging Cell* **2021**, *20*, e13305, doi:10.1111/acer.13305.
42. Xuan, A.G.; Long, D.H.; Gu, H.G.; Yang, D.D.; Hong, L.P.; Leng, S.L. BDNF Improves the Effects of Neural Stem Cells on the Rat Model of Alzheimer's Disease with Unilateral Lesion of Fimbria-Fornix. *Neurosci Lett* **2008**, *440*, 331–335, doi:10.1016/j.neulet.2008.05.107.
43. Xuan, A.G.; Luo, M.; Ji, W.D.; Long, D.H. Effects of Engrafted Neural Stem Cells in Alzheimer's Disease Rats. *Neurosci Lett* **2009**, *450*, 167–171, doi:10.1016/j.neulet.2008.12.001.
44. Tiernan, C.T.; Mufson, E.J.; Kanaan, N.M.; Counts, S.E. Tau Oligomer Pathology in Nucleus Basalis Neurons During the Progression of Alzheimer Disease. *J Neuropathol Exp Neurol* **2018**, *77*, 246–259, doi:10.1093/jnen/nlx120.
45. Cade, S.; Zhou, X.-F.; Bobrovskaya, L. The Role of Brain-Derived Neurotrophic Factor and the Neurotrophin Receptor P75NTR in Age-Related Brain Atrophy and the Transition to Alzheimer's Disease. *Rev Neurosci* **2022**, *33*, 515–529, doi:10.1515/revneuro-2021-0111.
46. Schmitz, T.W.; Nathan Spreng, R.; Alzheimer's Disease Neuroimaging Initiative Basal Forebrain Degeneration Precedes and Predicts the Cortical Spread of Alzheimer's Pathology. *Nat Commun* **2016**, *7*, 13249, doi:10.1038/ncomms13249.
47. Boissiere, F.; Faucheux, B.; Ruberg, M.; Agid, Y.; Hirsch, E.C. Decreased TrkA Gene Expression in Cholinergic Neurons of the Striatum and Basal Forebrain of Patients with Alzheimer's Disease. *Exp Neurol* **1997**, *145*, 245–252, doi:10.1006/exnr.1997.6443.

48. Mufson, E.J.; Lavine, N.; Jaffar, S.; Kordower, J.H.; Quirion, R.; Saragovi, H.U. Reduction in P140-TrkA Receptor Protein within the Nucleus Basalis and Cortex in Alzheimer's Disease. *Exp Neurol* **1997**, *146*, 91–103, doi:10.1006/exnr.1997.6504.
49. Ginsberg, S.D.; Che, S.; Wu, J.; Counts, S.E.; Mufson, E.J. Down Regulation of Trk but Not P75NTR Gene Expression in Single Cholinergic Basal Forebrain Neurons Mark the Progression of Alzheimer's Disease. *J Neurochem* **2006**, *97*, 475–487, doi:10.1111/j.1471-4159.2006.03764.x.
50. Goedert, M.; Fine, A.; Dawbarn, D.; Wilcock, G.K.; Chao, M.V. Nerve Growth Factor Receptor mRNA Distribution in Human Brain: Normal Levels in Basal Forebrain in Alzheimer's Disease. *Brain Res Mol Brain Res* **1989**, *5*, 1–7, doi:10.1016/0169-328x(89)90011-9.
51. Ernfors, P.; Lindfors, N.; Chan-Palay, V.; Persson, H. Cholinergic Neurons of the Nucleus Basalis Express Elevated Levels of Nerve Growth Factor Receptor mRNA in Senile Dementia of the Alzheimer Type. *Dement Geriatr Cogn Disord* **1990**, *1*, 138–145, doi:10.1159/000107133.
52. Mufson, E.J.; Kordower, J.H. Cortical Neurons Express Nerve Growth Factor Receptors in Advanced Age and Alzheimer Disease. *Proc Natl Acad Sci U S A* **1992**, *89*, 569–573, doi:10.1073/pnas.89.2.569.
53. Hu, X.-Y.; Zhang, H.-Y.; Qin, S.; Xu, H.; Swaab, D.F.; Zhou, J.-N. Increased P75(NTR) Expression in Hippocampal Neurons Containing Hyperphosphorylated Tau in Alzheimer Patients. *Exp Neurol* **2002**, *178*, 104–111, doi:10.1006/exnr.2002.8018.
54. Chakravarthy, B.; Ménard, M.; Ito, S.; Gaudet, C.; Dal Prà, I.; Armato, U.; Whitfield, J. Hippocampal Membrane-Associated P75NTR Levels Are Increased in Alzheimer's Disease. *J Alzheimers Dis* **2012**, *30*, 675–684, doi:10.3233/JAD-2012-120115.
55. Rabizadeh, S.; Bitler, C.M.; Butcher, L.L.; Bredesen, D.E. Expression of the Low-Affinity Nerve Growth Factor Receptor Enhances Beta-Amyloid Peptide Toxicity. *Proc Natl Acad Sci U S A* **1994**, *91*, 10703–10706, doi:10.1073/pnas.91.22.10703.
56. Sáez, E.T.; Pehar, M.; Vargas, M.R.; Barbeito, L.; Maccioni, R.B. Production of Nerve Growth Factor by Beta-Amyloid-Stimulated Astrocytes Induces P75NTR-Dependent Tau Hyperphosphorylation in Cultured Hippocampal Neurons. *J Neurosci Res* **2006**, *84*, 1098–1106, doi:10.1002/jnr.20996.
57. Costantini, C.; Weindruch, R.; Della Valle, G.; Puglielli, L. A TrkA-to-P75NTR Molecular Switch Activates Amyloid Beta-Peptide Generation during Aging. *Biochem J* **2005**, *391*, 59–67, doi:10.1042/BJ20050700.
58. Devarajan, S.; Sharmila, J.S. Computational Studies of Beta Amyloid (A β 42) with P75NTR Receptor: A Novel Therapeutic Target in Alzheimer's Disease. *Adv Bioinformatics* **2014**, *2014*, 736378, doi:10.1155/2014/736378.
59. Yaar, M.; Zhai, S.; Pilch, P.F.; Doyle, S.M.; Eisenhauer, P.B.; Fine, R.E.; Gilcrest, B.A. Binding of Beta-Amyloid to the P75 Neurotrophin Receptor Induces Apoptosis. A Possible Mechanism for Alzheimer's Disease. *J Clin Invest* **1997**, *100*, 2333–2340, doi:10.1172/JCI119772.
60. Tsukamoto, E.; Hashimoto, Y.; Kanekura, K.; Niikura, T.; Aiso, S.; Nishimoto, I. Characterization of the Toxic Mechanism Triggered by Alzheimer's Amyloid-Beta Peptides via P75 Neurotrophin Receptor in Neuronal Hybrid Cells. *J Neurosci Res* **2003**, *73*, 627–636, doi:10.1002/jnr.10703.
61. Hashimoto, Y.; Kaneko, Y.; Tsukamoto, E.; Frankowski, H.; Kouyama, K.; Kita, Y.; Niikura, T.; Aiso, S.; Bredesen, D.E.; Matsuoka, M.; et al. Molecular Characterization of Neurohybrid Cell Death Induced by Alzheimer's Amyloid-Beta Peptides via P75NTR/PLAIDD. *J Neurochem* **2004**, *90*, 549–558, doi:10.1111/j.1471-4159.2004.02513.x.
62. Sotthibundhu, A.; Sykes, A.M.; Fox, B.; Underwood, C.K.; Thangnipon, W.; Coulson, E.J. Beta-Amyloid(1-42) Induces Neuronal Death through the P75 Neurotrophin Receptor. *J Neurosci* **2008**, *28*, 3941–3946, doi:10.1523/JNEUROSCI.0350-08.2008.
63. Shen, L.-L.; Li, W.-W.; Xu, Y.-L.; Gao, S.-H.; Xu, M.-Y.; Bu, X.-L.; Liu, Y.-H.; Wang, J.; Zhu, J.; Zeng, F.; et al. Neurotrophin Receptor P75 Mediates Amyloid β -Induced Tau Pathology. *Neurobiol Dis* **2019**, *132*, 104567, doi:10.1016/j.nbd.2019.104567.
64. Saadipour, K.; Yang, M.; Lim, Y.; Georgiou, K.; Sun, Y.; Keating, D.; Liu, J.; Wang, Y.-R.; Gai, W.-P.; Zhong, J.-H.; et al. Amyloid Beta₁₋₄₂ (A β 42) up-Regulates the Expression of Sortilin via the P75(NTR)/RhoA Signaling Pathway. *J Neurochem* **2013**, *127*, 152–162, doi:10.1111/jnc.12383.
65. Skeldal, S.; Sykes, A.M.; Glerup, S.; Matusica, D.; Palstra, N.; Autio, H.; Boskovic, Z.; Madsen, P.; Castrén, E.; Nykjaer, A.; et al. Mapping of the Interaction Site between Sortilin and the P75 Neurotrophin Receptor Reveals a Regulatory Role for the Sortilin Intracellular Domain in P75 Neurotrophin Receptor Shedding and Apoptosis. *J Biol Chem* **2012**, *287*, 43798–43809, doi:10.1074/jbc.M112.374710.
66. Chakravarthy, B.; Gaudet, C.; Ménard, M.; Atkinson, T.; Brown, L.; Laferla, F.M.; Armato, U.; Whitfield, J. Amyloid-Beta Peptides Stimulate the Expression of the P75(NTR) Neurotrophin Receptor in SHSY5Y Human Neuroblastoma Cells and AD Transgenic Mice. *J Alzheimers Dis* **2010**, *19*, 915–925, doi:10.3233/JAD-2010-1288.
67. Hu, Y.; Lee, X.; Shao, Z.; Apicco, D.; Huang, G.; Gong, B.J.; Pepinsky, R.B.; Mi, S. A DR6/P75(NTR) Complex Is Responsible for β -Amyloid-Induced Cortical Neuron Death. *Cell Death Dis* **2013**, *4*, e579, doi:10.1038/cddis.2013.110.

68. Pedraza, C.E.; Podlesniy, P.; Vidal, N.; Arévalo, J.C.; Lee, R.; Hempstead, B.; Ferrer, I.; Iglesias, M.; Espinet, C. Pro-NGF Isolated from the Human Brain Affected by Alzheimer's Disease Induces Neuronal Apoptosis Mediated by P75NTR. *Am J Pathol* **2005**, *166*, 533–543, doi:10.1016/S0002-9440(10)62275-4.
69. Podlesniy, P.; Kichev, A.; Pedraza, C.; Saurat, J.; Encinas, M.; Perez, B.; Ferrer, I.; Espinet, C. Pro-NGF from Alzheimer's Disease and Normal Human Brain Displays Distinctive Abilities to Induce Processing and Nuclear Translocation of Intracellular Domain of P75NTR and Apoptosis. *Am J Pathol* **2006**, *169*, 119–131, doi:10.2353/ajpath.2006.050787.
70. Fombonne, J.; Rabizadeh, S.; Banwait, S.; Mehlen, P.; Bredesen, D.E. Selective Vulnerability in Alzheimer's Disease: Amyloid Precursor Protein and P75(NTR) Interaction. *Ann Neurol* **2009**, *65*, 294–303, doi:10.1002/ana.21578.
71. Yao, X.-Q.; Jiao, S.-S.; Saadipour, K.; Zeng, F.; Wang, Q.-H.; Zhu, C.; Shen, L.-L.; Zeng, G.-H.; Liang, C.-R.; Wang, J.; et al. P75NTR Ectodomain Is a Physiological Neuroprotective Molecule against Amyloid-Beta Toxicity in the Brain of Alzheimer's Disease. *Mol Psychiatry* **2015**, *20*, 1301–1310, doi:10.1038/mp.2015.49.
72. Zhou, X.-F.; Wang, Y.-J. The P75NTR Extracellular Domain: A Potential Molecule Regulating the Solubility and Removal of Amyloid- β . *Prion* **2011**, *5*, 161–163, doi:10.4161/pri.5.3.16896.
73. He, C.-Y.; Tian, D.-Y.; Chen, S.-H.; Jin, W.-S.; Cheng, Y.; Xin, J.-Y.; Li, W.-W.; Zeng, G.-H.; Tan, C.-R.; Jian, J.-M.; et al. Elevated Levels of Naturally-Occurring Autoantibodies Against the Extracellular Domain of P75NTR Aggravate the Pathology of Alzheimer's Disease. *Neurosci Bull* **2022**, doi:10.1007/s12264-022-00936-4.
74. Hatchett, C.S.; Tyler, S.; Armstrong, D.; Dawbarn, D.; Allen, S.J. Familial Alzheimer's Disease Presenilin 1 Mutation M146V Increases Gamma Secretase Cutting of P75NTR in Vitro. *Brain Res* **2007**, *1147*, 248–255, doi:10.1016/j.brainres.2007.02.002.
75. Yi, C.; Goh, K.Y.; Wong, L.-W.; Ramanujan, A.; Tanaka, K.; Sajikumar, S.; Ibáñez, C.F. Inactive Variants of Death Receptor P75NTR Reduce Alzheimer's Neuropathology by Interfering with APP Internalization. *EMBO J* **2021**, *40*, e104450, doi:10.15252/embj.2020104450.
76. Andrade-Talavera, Y.; Balleza-Tapia, H.; Dolz-Gaitón, P.; Chen, G.; Johansson, J.; Fisahn, A. Ablation of P75NTR Signaling Strengthens Gamma-Theta Rhythm Interaction and Counteracts A β -Induced Degradation of Neuronal Dynamics in Mouse Hippocampus in Vitro. *Transl Psychiatry* **2021**, *11*, 212, doi:10.1038/s41398-021-01332-8.
77. Cozza, A.; Melissari, E.; Iacopetti, P.; Mariotti, V.; Tedde, A.; Nacmias, B.; Conte, A.; Sorbi, S.; Pellegrini, S. SNPs in Neurotrophin System Genes and Alzheimer's Disease in an Italian Population. *J Alzheimers Dis* **2008**, *15*, 61–70, doi:10.3233/jad-2008-15105.
78. Cheng, H.-C.; Sun, Y.; Lai, L.-C.; Chen, S.-Y.; Lee, W.-C.; Chen, J.-H.; Chen, T.-F.; Chen, H.-H.; Wen, L.-L.; Yip, P.-K.; et al. Genetic Polymorphisms of Nerve Growth Factor Receptor (NGFR) and the Risk of Alzheimer's Disease. *J Negat Results Biomed* **2012**, *11*, 5, doi:10.1186/1477-5751-11-5.
79. Matyi, J.; Tschanz, J.T.; Rattinger, G.B.; Sanders, C.; Vernon, E.K.; Corcoran, C.; Kauwe, J.S.K.; Buhusi, M. Sex Differences in Risk for Alzheimer's Disease Related to Neurotrophin Gene Polymorphisms: The Cache County Memory Study. *J Gerontol A Biol Sci Med Sci* **2017**, *72*, 1607–1613, doi:10.1093/gerona/glx092.
80. Vacher, M.; Porter, T.; Villemagne, V.L.; Milicic, L.; Peretti, M.; Fowler, C.; Martins, R.; Rainey-Smith, S.; Ames, D.; Masters, C.L.; et al. Validation of a Priori Candidate Alzheimer's Disease SNPs with Brain Amyloid-Beta Deposition. *Sci Rep* **2019**, *9*, 17069, doi:10.1038/s41598-019-53604-5.
81. He, C.-Y.; Wang, Z.-T.; Shen, Y.-Y.; Shi, A.-Y.; Li, H.-Y.; Chen, D.-W.; Zeng, G.-H.; Tan, C.-R.; Yu, J.-T.; Zeng, F.; et al. Association of Rs2072446 in the NGFR Gene with the Risk of Alzheimer's Disease and Amyloid- β Deposition in the Brain. *CNS Neurosci Ther* **2022**, *28*, 2218–2229, doi:10.1111/cns.13965.
82. Jiao, S.-S.; Bu, X.-L.; Liu, Y.-H.; Wang, Q.-H.; Liu, C.-H.; Yao, X.-Q.; Zhou, X.-F.; Wang, Y.-J. Differential Levels of P75NTR Ectodomain in CSF and Blood in Patients with Alzheimer's Disease: A Novel Diagnostic Marker. *Transl Psychiatry* **2015**, *5*, e650, doi:10.1038/tp.2015.146.
83. Crispoltoni, L.; Stabile, A.M.; Pistilli, A.; Venturelli, M.; Cerulli, G.; Fonte, C.; Smania, N.; Schena, F.; Rende, M. Changes in Plasma β -NGF and Its Receptors Expression on Peripheral Blood Monocytes During Alzheimer's Disease Progression. *J Alzheimers Dis* **2017**, *55*, 1005–1017, doi:10.3233/JAD-160625.
84. Crooks, A.M.; Meeker, R.B. The New Wave of P75 Neurotrophin Receptor Targeted Therapies. *Neural Regen Res* **2022**, *17*, 95–96, doi:10.4103/1673-5374.314304.
85. Yang, T.; Knowles, J.K.; Lu, Q.; Zhang, H.; Arancio, O.; Moore, L.A.; Chang, T.; Wang, Q.; Andreasson, K.; Rajadas, J.; et al. Small Molecule, Non-Peptide P75 Ligands Inhibit Abeta-Induced Neurodegeneration and Synaptic Impairment. *PLoS One* **2008**, *3*, e3604, doi:10.1371/journal.pone.0003604.
86. Yaar, M.; Zhai, S.; Panova, I.; Fine, R.E.; Eisenhauer, P.B.; Blusztajn, J.K.; Lopez-Coviella, I.; Gilchrest, B.A. A Cyclic Peptide That Binds P75(NTR) Protects Neurones from Beta Amyloid (1-40)-Induced Cell Death. *Neuropathol Appl Neurobiol* **2007**, *33*, 533–543, doi:10.1111/j.1365-2990.2007.00844.x.
87. Yaar, M.; Arble, B.L.; Stewart, K.B.; Qureshi, N.H.; Kowall, N.W.; Gilchrest, B.A. P75NTR Antagonistic Cyclic Peptide Decreases the Size of Beta Amyloid-Induced Brain Inflammation. *Cell Mol Neurobiol* **2008**, *28*, 1027–1031, doi:10.1007/s10571-008-9298-6.

88. Zhang, J.; Zhang, R.; Zhan, Z.; Li, X.; Zhou, F.; Xing, A.; Jiang, C.; Chen, Y.; An, L. Beneficial Effects of Sulforaphane Treatment in Alzheimer's Disease May Be Mediated through Reduced HDAC1/3 and Increased P75NTR Expression. *Front Aging Neurosci* **2017**, *9*, 121, doi:10.3389/fnagi.2017.00121.
89. Wang, Q.-H.; Wang, Y.-R.; Zhang, T.; Jiao, S.-S.; Liu, Y.-H.; Zeng, F.; Li, J.; Yao, X.-Q.; Zhou, H.-D.; Zhou, X.-F.; et al. Intramuscular Delivery of P75NTR Ectodomain by an AAV Vector Attenuates Cognitive Deficits and Alzheimer's Disease-like Pathologies in APP/PS1 Transgenic Mice. *J Neurochem* **2016**, *138*, 163–173, doi:10.1111/jnc.13616.
90. Massa, S.M.; Xie, Y.; Yang, T.; Harrington, A.W.; Kim, M.L.; Yoon, S.O.; Kraemer, R.; Moore, L.A.; Hempstead, B.L.; Longo, F.M. Small, Nonpeptide P75NTR Ligands Induce Survival Signaling and Inhibit ProNGF-Induced Death. *J Neurosci* **2006**, *26*, 5288–5300, doi:10.1523/JNEUROSCI.3547-05.2006.
91. Knowles, J.K.; Simmons, D.A.; Nguyen, T.-V.V.; Vander Griend, L.; Xie, Y.; Zhang, H.; Yang, T.; Pollak, J.; Chang, T.; Arancio, O.; et al. Small Molecule P75NTR Ligand Prevents Cognitive Deficits and Neurite Degeneration in an Alzheimer's Mouse Model. *Neurobiol Aging* **2013**, *34*, 2052–2063, doi:10.1016/j.neurobiolaging.2013.02.015.
92. Simmons, D.A.; Knowles, J.K.; Belichenko, N.P.; Banerjee, G.; Finkle, C.; Massa, S.M.; Longo, F.M. A Small Molecule P75NTR Ligand, LM11A-31, Reverses Cholinergic Neurite Dystrophy in Alzheimer's Disease Mouse Models with Mid- to Late-Stage Disease Progression. *PLoS One* **2014**, *9*, e102136, doi:10.1371/journal.pone.0102136.
93. Yang, T.; Tran, K.C.; Zeng, A.Y.; Massa, S.M.; Longo, F.M. Small Molecule Modulation of the P75 Neurotrophin Receptor Inhibits Multiple Amyloid Beta-Induced Tau Pathologies. *Sci Rep* **2020**, *10*, 20322, doi:10.1038/s41598-020-77210-y.
94. Yang, T.; Liu, H.; Tran, K.C.; Leng, A.; Massa, S.M.; Longo, F.M. Small-Molecule Modulation of the P75 Neurotrophin Receptor Inhibits a Wide Range of Tau Molecular Pathologies and Their Sequelae in P301S Tauopathy Mice. *Acta Neuropathol Commun* **2020**, *8*, 156, doi:10.1186/s40478-020-01034-0.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.