

Article

Evaluation of Methods for Collecting Mosquitoes (Culicidae: Diptera) in Canopy and Ground Strata in the Brazilian Savanna

Luis F. Mucci¹, Eduardo S. Bergo¹, Juliana Telles-de-Deus¹, Simone L. Reginato¹, Mariza Pereira¹ and Vera L. F. de Camargo-Neves^{1,*}

¹ Secretaria de Estado da Saúde de São Paulo, Instituto Pasteur, São Paulo 01027-000, SP, Brazil; lfmucci@gmail.com (LFM), edusteber@uol.com.br (ESB), jtelles@pasteur.saude.sp.gov.br (JTD), simone_reginato@yahoo.com.br (SLR), marizap222@gmail.com (MP), vcamargo@pasteur.saude.sp.gov.br (VLFCN)

* Correspondence: lfmucci@gmail.com, vcamargo@pasteur.saude.sp.gov.br

Abstract: The hand-net is the standard method for capturing mosquitoes with sylvatic diurnal activity in disease outbreaks in Brazil. However, occupational risks and biases related to the collectors' abilities and attractiveness are important limitations. In this study, we compared hand-nets with automatic traps (CDC) associated to CO₂ and BG-Lure®, in the Vassununga State Park, a Brazilian Savanna protection area. The collections carried out over 27 days, on the ground and the forest canopy. A total of 1,555 mosquitoes were obtained in 20 taxa. The diversity index ranged between 1.12 and 1.79 and the dominance index, from 0.22 to 0.40. The dominant species in the ground was *Aedes scapularis* (46.0%) and in the canopy, *Hg. janthinomys/capricornii* (31.9%). *Haemagogus leucocelaenus* was rare (n=2). The hand-net resulted in the greatest diversity and abundance of species in both strata, followed by traps associated with CO₂. A low degree of similarity was observed between the hand-net on the ground compared to the other capture methods. The use of BG-Lure® alone resulted in a low number of specimens. In conclusion, the hand-net is still the method of choice for collecting arbovirus vectors in the diurnal period, especially yellow fever vectors.

Keywords: Diurnal mosquitoes; Kairomones; Canopy stratum; Brazilian savanna

1. Introduction

In Brazil, many species of Culicidae (Diptera) are of medical interest [1,2] and much of the knowledge about their bioecology and epidemiological role is the result of research and entomological surveys associated with outbreaks of arboviruses [3–7]. In these studies, the mobile human attraction technique using capture hand-nets was widely used, often in entomological captures in the forest canopy, aimed at increasing the sample of mosquitoes that feed on the blood of birds and non-human primates (NHPs), considered important hosts for several arboviruses [5,8,9]. For this reason, in Brazil, the hand-net was adopted as the standard technique by health surveillance services across the country, with canopy capture recommended as a complementary procedure [10].

However, the use of hand-nets, with humans being the attractant, is subject to variations in individual performance and the attractiveness of each collectors which can imply a bias resulting in sample divergences [11]. Moreover, collections performed by individuals raises concerns about occupational risks [12,13], especially when working at heights in tree canopies, according to the specific safety protocols of each country.

Thus, there is a need for alternative techniques to replace the standard hand-net technique that can ensure representativeness in mosquito sampling. Service (1993) and Santos et al. (2021) point out that the selection of the appropriate method should take into account the specific characteristics of the natural history, biology and ecology of the target species of potential vectors [14,15].

One of the alternative techniques is the use of automatic suction traps with a luminous attractant, commonly used to collect nocturnal species [13,16]. The association of a carbon dioxide source and other types of odor attractants (such as octenol and lactic acid) has been used to increase the sensitivity of this type of trap for daytime use [13,17–21].

In Brazil, most of the studies that use attractants are focused on the urban environment and aimed at capturing *Aedes aegypti* and *Culex quinquefasciatus* [22–24]. With respect to wild mosquitoes, studies are restricted to afternoon and nocturnal periods [21,25–29].

Recently, our group began looking for alternative and/or complementary methods for daytime collections in accordance with forest stratification to carry out routine surveys in different biomes. In a previous study, we conducted a comparative evaluation of in-person capture using hand-nets and automatic traps with carbon dioxide (dry ice) and BG-Lure® as attractants on the ground and in the canopy stratum of an Atlantic Forest environmental reserve [30]. In the present study, we apply the same methodology used in the previous study for a comparative evaluation of these methods in the Cerrado biome.

2. Materials and Methods

2.1. Research Area

The Vassununga state park (Parque Estadual Vassununga - PEV) is a protected conservation area that was created to protect representative areas of the seasonal semi-deciduous forest (inland Atlantic Forest) and different Cerrado (Brazilian savanna) physiognomies. This park, made up of six discontinuous sectors with a total area of 2,071.42 hectares, is located in the municipality of Santa Rita do Passa Quatro, São Paulo, Brazil [31] (Figure 1). The Pé de Gigante sector, chosen for this study, is the largest area in the Park (1,212.92 ha) and contains the three types of Savanna: forested savanna (Cerradão), arborized savanna (Cerrado stricto sensu-ss) and savanna Gramíneo-Lenhosa, according to the classification adopted by Veloso et al. (1991) [32].

According to Köppen climate classification, the climate of the Santa Rita do Passa Quatro region is Cwa; rainy in summer and dry in winter. The average annual temperature is 23.3°C, with an average maximum temperature of approximately 26.0°C in summer (December to February) and an average minimum temperature of approximately 19.0°C in winter (June to August). According to the Park Management Plan [31], the park has an average annual rainfall of 1,365.7 mm and potential evapotranspiration of 1,160.61 mm.

Two sites of the area were selected for the collections: the forest margin and inside the forest (about 3 km apart). In each site, collections were carried out in the forest canopy and at ground level. For the sampling of mosquitoes at the edge of the forest, a platform was installed at the first branch ramification of the tree canopy, seven meters above the ground, accessed using ropes and climbing equipment by a trained capturer. This platform is in an area characterized as arborized savanna (Cerrado ss), a dense savanna with a predominantly arboreal vegetation subtype, with 50 to 70% coverage and an average height of five to eight meters. This physiognomy represents the highest and densest form of savanna. In the second sampling area, inside the forest, we used the platform of a meteorological tower. The capturer worked at a height of eight meters, corresponding to the average canopy height of the surrounding trees. This sector of the park corresponds to an area of dense savanna transitioning into Cerradão, with a predominance of trees in a closed canopy that approaches forest vegetation, with an average tree height of 10 to 12 meters [31].

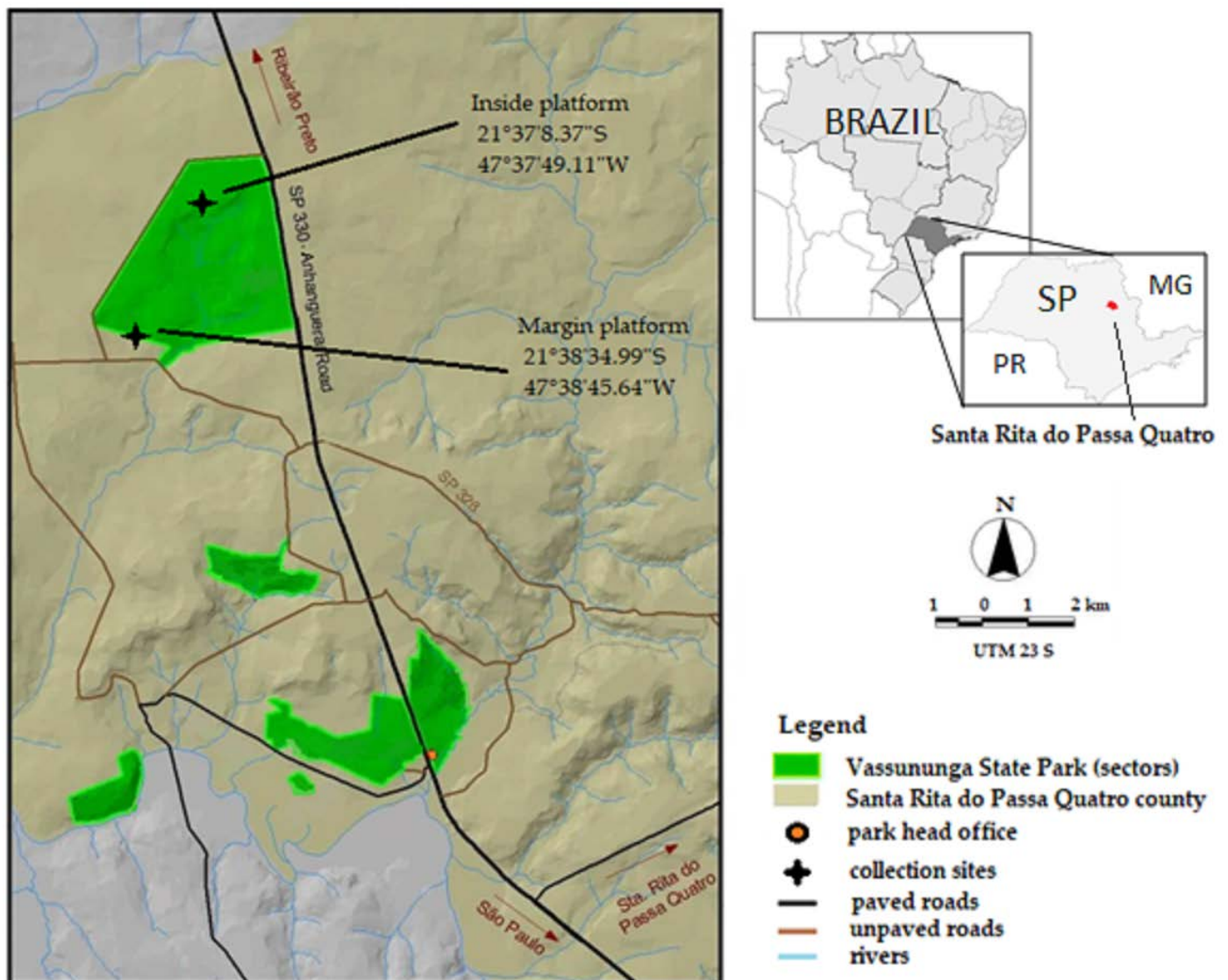


Figure 1. Location of the study area in the Savanna biome, Vassununga State Park. Source: Plano de Manejo PEV, 2009.

2.2. Capture of Specimens

Entomological captures were carried out on three consecutive days per month in February, March, October, November, and December of 2020 and in January, February, April, and May of 2021. The pause in collections was due to COVID-19 pandemic restrictions. Two techniques were used:

2.2.1. Capture by Hand-net

Captures were carried out from 9:00 to 16:00 in both sectors simultaneously. In each sector, the collections were carried out by two capturers, one on the platform and another at ground level, covering approximately 1,500 meters of trails around each platform.

2.2.2. Capture with Automatic Traps

CDC-type electric traps were used [33] with two chemical attractants: carbon dioxide (dry ice) and BG-Lure® (a commercial product), which is a chemical attractant composed of ammonia, L- lactic acid and caproic acid. In each sector (forest margin and interior), four traps were installed close to the ground, between 1 and 1.5 meters in height, and

another four in the tree canopy at a minimum height of 5 meters, depending on the average height of the trees at the site. There were 16 traps in total, with eight positioned at the forest edge and eight inside the forest around each platform, considered the central reference point (Figure 1). The traps were placed at points to the North, South, East, and West, at approximately 250 meters from the platform, and each day the traps were rotated to alternate the position of the exposure of the attractants. One pair of traps (ground-canopy) was used with only CO₂, another pair with only BG-Lure®, and two pairs with the two attractants together (in diametrically opposite positions). The electric traps were exposed in the same collection period as the hand-net method, from 9:00 to 16:00.

Conservation of samples: insects collected with nets and traps were transferred using a mouth aspirator to cryovials and frozen in liquid nitrogen to preserve viral genetic material for future analysis.

2.3. Identification of Material

In the laboratory, the biological material frozen in nitrogen was transferred to a freezer at -80°C until the identification of the genus and species on a cold table at -20°C. Taxonomic keys were used for the morphological identification of Culicidae [1,2,34,35].

2.4. Data analysis

The data were entered into an information system operating on a web server developed especially for the project.

For the estimation of species diversity, species accumulation rate and statistical abundance analysis were performed using the EstimateS version 9 statistics program [36]. Diversity (S) was determined by the number of species collected per month in each environment and stratum and by the capture method. The accumulation curve was calculated using the Coleman rarefaction method. The estimate of the true number of species was performed with the Chao1 estimator and a confidence interval of 95%.

To compare diversity among the different methods (technique-attractant-stratum), the Shannon (H) and Gini-Simpson (1-D) indices were calculated; with Simpson's dominance (D) calculated using EstimateS v.9. For cluster analysis, we used dendrograms based on Bray-Curtis similarity, which considers the abundance of specimens. For the pairwise analysis of the association between methods, we used Spearman's correlation with the PAST version 4.05 statistics program [37].

3. Results

We collected a total of 1,555 Culicidae specimens in 20 taxa, of which 17 species belonged to eight genera. Only 24 specimens were males, corresponding to the genus *Culex* and to the species *Aedes albopictus* and *Psorophora albigena* (Table S1). *Aedes scapularis* was the most abundant species, followed by *Haemagogus janthinomys/capricornii*, *Psorophora albigena*, *Sabethes albiprivus* and *Ae. albopictus* (Table 1). All species, except *Limatus durhamii* and *Sabethes belisarioi*, were more abundant at ground level (n = 1302) than in the canopy (n = 229), including *Hg. janthinomys/capricornii* and *Sa. albiprivus*, which were the dominant species in the canopy. *Haemagogus leucocelaenus* was a rare species at ground level, with only two specimens collected (Table 1).

The hand-net capture technique obtained higher yields than the automatic traps, both in the canopy and on the ground, accounting for 78.6% of the total of specimens collected in the canopy and 72.1% on the ground.

On the ground, the traps using CO₂ + BG-Lure® obtained a higher yield than the traps with only CO₂. Traps using exclusively the BG-Lure® attractant had the lowest yield in both strata (Table 1).

Table 1. Number of Culicidae females collected in the “Pé-do-Gigante” sector of the Vassununga State Park, by forest stratum, technique, and attractant.

TAXA	CANOPY							GROUND							TOTAL	
	BGL	CO2	CO2+ BG1	CO2+ BGL2	NET	TOTAL	%	BGL	CO2	CO2+ BG1	CO2+ BGL2	NET	TOTAL	%	N	%
<i>Aedes albopictus</i>					4	4	1.7	2	9	6	17	115	149	11.4	153	10.0
<i>Aedes scapularis</i>		3	12	1	18	34	14.8	14	65	78	34	408	599	46.0	633	41.3
<i>Aedes serratus</i>												2	2	0.2	2	0.1
<i>Coquillettia juxtamansonia</i>												1	1	0.1	1	0.1
<i>Culex sp</i>				1	7	8	3.5	1	1	4		22	28	2.2	36	2.4
<i>Haemagogus janthinomys/capricornii</i>		1	2	1	69	73	31.9		1	6	5	134	146	11.2	219	14.3
<i>Haemagogus leucocelaenus</i>												2	2	0.2	2	0.1
<i>Limatus durhamii</i>					1	1	0.4								1	0.1
<i>Psorophora albigena</i>		1	11	1	16	29	12.7	8	10	18	9	123	168	12.9	197	12.9
<i>Psorophora discrucians</i>									1				1	0.1	1	0.1
<i>Psorophora ferox</i>									1			3	4	0.3	4	0.3
<i>Psorophora varipes</i>										4		3	7	0.5	7	0.5
<i>Psorophora sp</i>								1				3	4	0.3	4	0.3
<i>Sabethes albiprivus</i>		3		5	35	43	18.8	1	26	7	18	92	144	11.1	187	12.2
<i>Sabethes belisarioi</i>				1	5	6	2.6			1		3	4	0.3	10	0.7
<i>Sabethes glaucodaemon</i>	1		2	3	22	28	12.2		2	3	5	22	32	2.5	60	3.9
<i>Sabethes purpureus</i>					3	3	1.3		2		1	4	7	0.5	10	0.7
<i>Sabethes tarsopus</i>												1	1	0.1	1	0.1
<i>Sabethes sp</i>									1	1			2	0.2	2	0.1
<i>Wyeomyia confusa</i>												1	1	0.1	1	0.1
TOTAL	1	8	27	13	180	229	100.0	27	119	128	89	939	1302	100.0	1531	100.0
%	0.4	3.5	11.8	5.7	78.6	100.0		2.1	9.1	9.8	6.8	72.1	100.0			
				15.0							85.0				100.0	

Techniques-attractants: BGL, CDC with only BG-Lure®; CO2, CDC with only CO2; CO2+BGL1 or CO2+BGL2, CDC with CO2+BG-Lure® and; NET, hand-net.

Table 2. Indicators of abundance, richness, diversity, and dominance of Culicidae specimens collected in the “Pé-do-Gigante” sector of the Vassununga State Park, by forest strata, technique, and attractant.

PARAMETER	CANOPY					GROUND				
	BGL	CO2	CO2+ BG1	CO2+ BGL2	NET	BGL	CO2	CO2+ BG1	CO2+ BGL2	NET
S (Taxa)	1	4	4	7	10	5	10	9	7	16
N (Abundance)	1	8	27	13	180	26	118	127	89	936
D (Dominance)	1	0.31	0.37	0.23	0.22	0.39	0.37	0.41	0.24	0.25
1 - D (Geni-Simpson Index)	0	0.69	0.63	0.77	0.78	0.61	0.63	0.59	0.76	0.75
H (Shannon Index)	0	1.26	1.11	1.69	1.80	1.14	1.37	1.37	1.61	1.70

Techniques-attractants: BGL, CDC with only BG-Lure®; CO2, CDC with only CO2; CO2+BGL1 or CO2+BGL2, CDC with CO2+BG-Lure® and; NET, hand-net.

Regarding the indicators for richness, diversity, and dominance, we observed a qualitative similarity between canopy and ground strata, as the differences between each method in the canopy had the same pattern on the ground. In addition, the values of the same method in the canopy were close to those on the ground. The differences were small between the values for absolute richness and abundance for the different techniques and attractants in each stratum and, in general, with greater richness on the ground compared to the canopy. The biggest difference was in the trap results associated with BG-Lure® in the canopy, where only one specimen was collected and, thus, presented very different indicators of the same method at ground level. Hand-net capture was the technique with greater richness and diversity and less dominance in both strata.

Species accumulation curves show a tendency to anticipate stability for the canopy compared to the ground (Figure 2). This stability in the canopy added to the qualitative and quantitative results, as seen in Table 1.

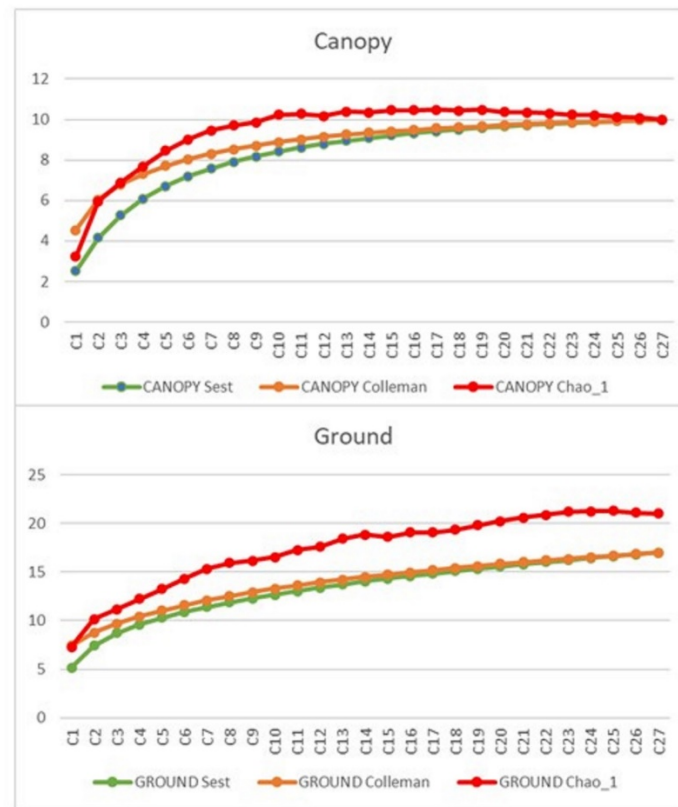


Figure 2. Species accumulation curves for *Culicidae* specimens collected in canopy and ground strata of the Savanna forest in the “Pé-do-Gigante” sector of the Vassununga State Park. Note: C = collection.

Table 3. Spearman's correlation coefficients and statistical significance (p) values from the pair-to-pair comparison between different methods (stratum-technique-attractant) for mosquito collection in the “Pé-do-Gigante” sector of the Vassununga State Park.

STRATA	METHODS	CANOPY					GROUND				
		BGL	CO2	CO2+BG1	CO2+BGL2	NET	BGL	CO2	CO2+BG1	CO2+BGL2	NET
p valor											
CANOPY	BGL		0.622	0.096	0.101	0.184	0.571	0.520	0.853	0.492	0.536
	CO2	-0.12		0.001	0.011	0.001	0.004	0.004	0.001	0.001	0.001
	CO2+BG1	0.39	0.68		0.024	0.002	0.064	0.022	0.008	0.008	0.002
	CO2+BGL2	0.39	0.57	0.52		0.000	0.096	0.006	0.000	0.004	0.040
	NET	0.32	0.69	0.66	0.73		0.011	0.005	0.003	0.002	0.000
GROUND	BGL	-0.14	0.63	0.43	0.39	0.57		0.001	0.001	0.002	0.001
	CO2	0.16	0.62	0.52	0.60	0.62	0.71		0.000	0.000	0.004
	CO2+BG1	0.05	0.69	0.59	0.75	0.65	0.71	0.74		0.000	0.002
	CO2+BGL2	0.17	0.70	0.59	0.63	0.67	0.66	0.91	0.81		0.002
	NET	0.15	0.68	0.66	0.47	0.81	0.69	0.63	0.67	0.65	
p valor											

Techniques-attractants: BGL, CDC with only BG-Lure®; CO2, CDC with only CO2; CO2+BGL1 or CO2+BGL2, CDC with CO2+BG-Lure® and; NET, hand-net.

The Bray-Curtis similarity analysis for the different methods for collecting mosquito species is used to verify clusters of those methods that are most similar in qualitative and quantitative terms of species. Figure 3A shows the similarity dendrogram based on the analysis of all species found. A cluster that stands out includes all the traps at ground level that used CO2 (exclusively or with BG-Lure®) and the hand-net in the canopy, which suggests that one method could replace the other or serve as a replica. At the other extreme, the ground trap methods and traps using only BG-Lure® in the canopy have the

fewest similarities, being isolated from the others. This, however, can be attributed to different reasons, the first being the method presented superior abundance and richness, and the latter, being almost null.

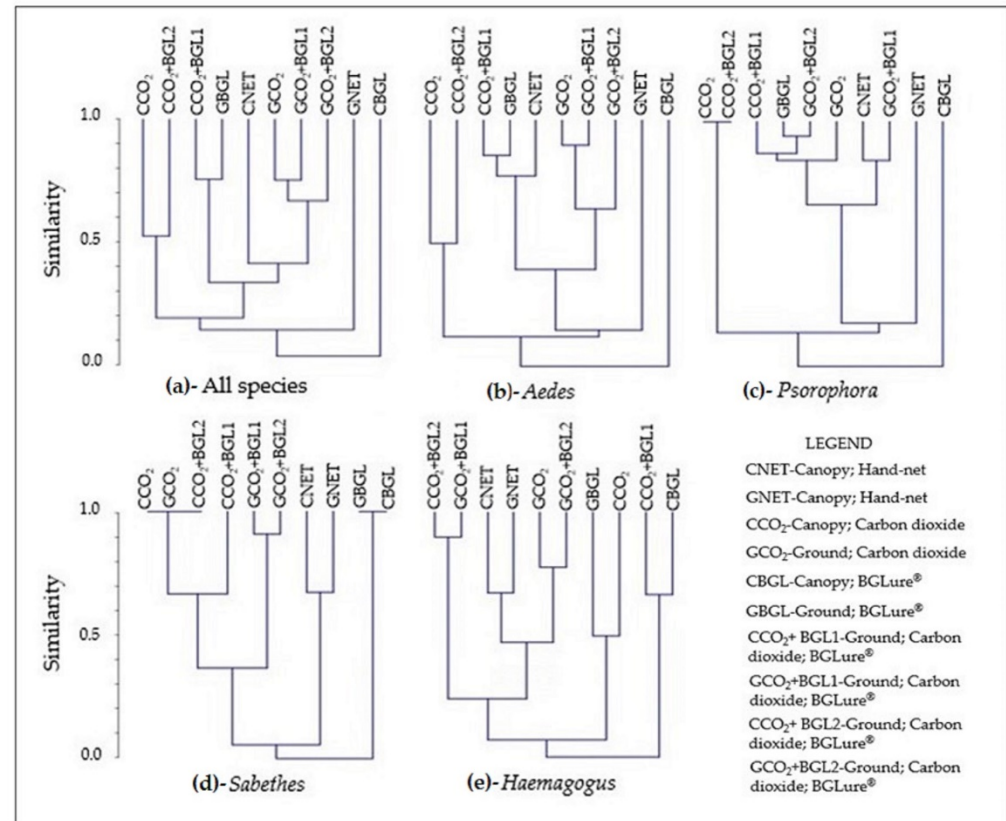


Figure 3. Bray-Curtis similarity dendrogram for different methods (technique-attractant-stratum) for the collection of Culicidae specimens in the “Pé-do-Gigante” sector of the Vassununga State Park, according to all identified species (A) and, according to the four genera with the highest number of specimens: B - *Aedes*; C - *Psorophora*; D - *Sabethes*; E - *Haemagogus*.

The analysis considering each of the four most abundant genera in individualized dendrograms is presented in Figure 3 (B,C,D,E). The genus *Psorophora* presents a very similar large group comprising several methods; ground hand-net methods and traps with only BG-Lure® in the canopy appear in isolation, following a pattern similar to the dendrogram for all species, which also presents the group of traps with CO₂ in the canopy. The genus *Aedes* also shows some variation of this pattern for the central group, with fewer similarities than *Psorophora*. In the case of the *Haemagogus* and *Sabethes* genera, both presented a group with great similarity composed of nets in the canopy and on the ground, suggesting the human attractant as an aggregator. However, *Haemagogus* seems to be the genus that expresses more standardized responses to methods, while *Sabethes* shows more diffuse behavior for technique-attractant-stratum combinations.

4. Discussion

Our study is the most recent survey of diurnal mosquitoes in the Savanna biome, where species richness and abundance were studied according to forest stratification. The main objective was to evaluate alternative capture methods to the hand-net, using the attractants BG-Lure® and carbon dioxide, in order to increase the effectiveness of automatic traps, especially for species of epidemiological interest that frequent the forest canopy. Thus, we sought to carry out collections in a favorable seasonal period, mainly for the species of the Aedini and Sabethini Tribes [38–43].

Based on the results of the species accumulation curves for both strata, we verified that the sampling in the period was efficient, allowing for a good characterization of the diurnal mosquito fauna in the “Pé-do-Gigante” sector of the park. Thus, although we used a shorter collection period (27 days) than in other studies conducted in savanna areas (40-60 days) [3,44,45], we found similar species richness for the genera *Aedes*, *Haemagogus*, *Psorophora* and *Sabethes*, with 14 species found in our study, and 14 to 17 in the others. It is also worth mentioning that these three studies were carried out in central Brazil, between latitude 15.5 and 17.0°S, while our study, at latitude 21.5°S, represents an important record of this fauna in a region close to the southern limit of this biome in Brazil [46].

In addition, due to the methodology of simultaneous collection in the canopy and on the ground, a robust comparison between these two strata was possible. In other systematic studies [18,39,47] or specific entomological surveys associated with epidemic outbreaks [48–50] that also investigate the community of diurnal mosquitoes using the mobile human bait technique with hand-nets, collections were not carried out in the canopy. Thus, all the data on abundance, richness, and dominance of species in the region were related to collections at ground level. Our study provides pioneering results on the stratification of mosquito communities in the northwest region of the state of São Paulo.

Considering the general results from the two strata, we can observe the dominance of *Aedes scapularis* and sub-dominance of *Haemagogus janthinomys/capricornii*, *Psorophora albigena*, *Sabethes albiprivus* and *Ae. albopictus*. In the comparison between strata, *Ae. scapularis* was dominant on the ground, while *Hg. janthinomys/capricornii* dominates in the canopy. Qualitatively, the list of species in the canopy and at ground level was very similar. In quantitative terms, practically all the species found in the canopy had a lower number of specimens compared to collections carried out on the ground, which was also observed in other studies carried out in the savanna [3,44,45].

This suggests that microclimatic conditions in the canopy may be less favorable than at ground level. We should emphasize that the collection points were in the savanna physiognomy, where the shading and size of the trees were less than in other physiognomies found in the park, such as the “Cerradão”, gallery forest, and semi-deciduous seasonal forest [51,52]. In this sense, the canopy was more subject to desiccation, affecting the intensity and maintenance of relative humidity, which is a key environmental factor for the main species found there [39,53,54].

With respect to the indicators for richness, diversity, evenness, and dominance for each technique and attractant, we also observed a qualitative similarity between the canopy and ground, since the differences between each method in the canopy followed the same pattern on the ground. Furthermore, the values of the same method in the canopy were close to those on the ground, but not the parameters of absolute numbers such as species richness and abundance. Spearman's correlation results corroborate this since few significant differences were found between the different methods in the same stratum and between strata. Contrarily, these indicators were different between the strata and methods in a similar study we carried out in the Atlantic Forest biome in Cantareira State Park [30].

However, in both studies, the hand-net was the technique with greater richness and diversity and lower dominance in both strata, while the indicators of traps using only BG-Lure® demonstrated inefficiency, a result also found for wild diurnal mosquitoes in Brazil by other researchers [55,56]. This shows that the exclusive use of BG-Lure® may be inefficient for wild neotropical species. The use of CO₂ as an attractant demonstrated worse performance than its use together with BG-Lure®, a different result from that found in the Atlantic Forest [30] and probably due to the differences in specific environmental conditions in the savanna.

It is worth mentioning that there are few attractants available for commercial use, as can be seen in many studies carried out in Brazil to evaluate the performance of mosquito collection with automatic traps [23–27,30,55–61]. This suggests the need to develop in situ studies using olfactometry to discover different substances that present selective attractiveness for each species of interest, according to the biome where it will be used.

Among the most abundant species of mosquitoes collected in the “Pé-do-Gigante” sector, all of them showed epidemiological importance, especially for the transmission of yellow fever. *Haemagogus janthinomys*, *Hg. leucocelaenus* and *Sa. chloropterus* are considered the main vector species in South America [62], the first two with the greatest range in Brazil [63,64]. In our study, we used the nomenclature *Haemagogus janthinomys/capricornii* because we were unable to determine which of the two species occurred at the site, since they are morphologically differentiated only by the male genitalia, and the investigated site is in the co-occurrence zone [2]. At any rate, whatever the species, it was the most abundant in the canopy compared to the other mosquito species, with a greater number of specimens at ground level than in the canopy. This result differs from several studies in which the species is cited as an acrodendrophile [45,53,55,65,66]. On the other hand, it adds to the results of other studies where variations in this pattern were observed [3,44,56].

The other species with epidemiological importance for yellow fever, which presented the greatest number of specimens, were *Ae. scapularis*, *Ae. albopictus*, *Ps. albigenu*, *Sa. albiprivus* and *Sa. glaucodaemon*, and to a lesser extent *Ae. serratus*, *Ps. ferox*, and *Hg. leucocelaenus*. The rarity of *Hg. leucocelaenus* is noteworthy since it is one of the dominant species in the Cantareira State Park [30] and a vector for the transmission of the yellow fever virus in other areas of the Atlantic Forest [64,67,68]. Perhaps this species has no favorable habitat in the savanna, at least in the *stricto sensu* physiognomy investigated in this study. The assessment of its presence and abundance in enclaves of seasonal semi-deciduous forests, gallery forests, and in the physiognomy of the “Cerradão” deserves to be studied in order to better understand the details of the ecology of the species in this biome since previous studies have not exactly focused on this [18,29,44,45,47,69].

An important result of this research is the assessment of the success of different techniques and attractants in the canopy and on the ground for capturing the genera *Aedes*, *Haemagogus*, *Psorophora*, and *Sabethes*, providing valuable data for arbovirus surveillance services in Brazil. In the similarity cluster analysis, the selectivity of hand-nets for the genera *Haemagogus* and *Sabethes* was evident in the dendrograms, while for the genera *Aedes* and *Psorophora*, the similarity between methods was more similar to the general dendrogram (Figure 3A all species), where the hand-net on the ground and the trap using only BG-Lure® in the canopy appear distant from the other methods, with the former having the highest relative yield and the latter the lowest relative yield.

In investigations of epidemics or epizootic outbreaks, the right place and time are crucial for obtaining good samples [10,70]. Thus, the use of the technique with the highest yield for the target species is very important. However, it is worth considering that the “ideal” collection technique is not always possible for surveillance services. Moreover, the use of automatic traps may be the only way to expand the space-time factor in routine and large-scale surveys [71,72]. Thus, if it is necessary to make use of these alternative techniques, this study, and that of Deus et al. (2022), may represent an important reference for the operational planning of field actions [30].

5. Conclusions

The results of this study highlight the benefits of using hand nets over automatic traps with kairomones to capture diurnal mosquitoes, as they presented higher values of richness and abundance for species of epidemiological interest. Our research suggests that electric traps should be used to collect diurnal mosquitoes in the savanna when used with CO₂ and BG-Lure®, for species richness studies of diurnal Culicidae.

It is clear that there is still much to be explored with respect to attractants and it is important to encourage the development and testing of new kairomones associated with specific hosts, especially when the objective is to increase epidemiological and/or epizootic research based on the infectivity of mosquitoes and the genomic study of the etiologic agent.

Supplementary Materials: Not applicable

Author Contributions: Conceptualization, methodology, investigation, writing—original draft preparation, all the authors participated equally; validation, J.T.D., V.L.C.N., E.S.B., M.P. L.F.M.; formal analysis, V.L.F.C.N.; resources, V.L.F.C.N.; data curation, S.L.R., V.L.F.C.N.; writing—review and editing, J.T.D., L.F.M., V.L.F.C.N.; supervision, J.T.D., V.L.F.C.N.; project administration, V.L.F.C.N.; funding acquisition, V.L.F.C.N. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Fundação de Amparo à Pesquisa do Estado de São Paulo - Fapesp, Brazil grant number: Fapesp - 2017/50345-5 and, the Superintendência de Controle de Endemias – SUCEN, Secretaria de Estado da Saúde de São Paulo, Brazil.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable

Acknowledgments: We thank the teams that carried out the entomological captures and Mr. Agnaldo Nepomuceno, Regional Director of the Metropolitan Region of São Paulo for his support. We are also grateful to Fabrício Pinheiro da Cunha at the Forestry Foundation of Secretaria de Infraestrutura e Meio Ambiente do Estado de São Paulo - SIMA/SP for logistics support and for granting permission to work in the park according to the license SIMA N° 260108 – 009.866/2019 and, Instituto Chico Mendes de Conservação e Biodiversidade, authorization number – SISBIO 72494-1.

Conflicts of Interest: The authors declare no conflict of interest.

Table S1 - Number of male specimens collected by genus and species, according to stratum (canopy and ground) and capture technique (treatment). Santa Rita do Passa Quatro, SP, Brazil.

SPECIE	N	METHOD	STRATUM
<i>Culex</i> sp	8	net	ground /canopy
<i>Aedes albopictus</i>	13	net	ground
<i>Psorophora albigena</i>	3	net/BGL*	ground
TOTAL	24		

* net, hand-net; BGL, CDC with only BG-Lure®

References

1. Consoli, R.A.G.B.; Oliveira, R.L. de *Principais mosquitos de importância sanitária no Brasil*; Editora FIOCRUZ: Rio de Janeiro, RJ, 1994; ISBN 978-85-85676-03-2.
2. Forattini, O.P. *Culicidologia Médica: Identificação, Biologia, Epidemiologia*; Edusp: São Paulo, Brasil, 2002.
3. Pinheiro, F.P.; Rosa, A.P.A.T. da; Moraes, M.A.P. An Epidemic of Yellow Fever in Central Brazil, 1972–1973: II. Ecological Studies. *Am. J. Trop. Med. Hyg.* **1981**, *30*, 204–211, doi:10.4269/ajtmh.1981.30.204.
4. Vasconcelos, P.F.C. Clinical and Ecoepidemiological Situation of Human Arboviruses in Brazilian Amazonia.
5. Iversson, L.B. [Current status of the eco-epidemiological knowledge on arboviruses pathogenic to humans in the Atlantic Forest region of the State of São Paulo]. *Rev. Inst. Med. Trop. Sao Paulo* **1994**, *36*, 343–353, doi:10.1590/s0036-46651994000400007.
6. Vasconcelos, P.F.; Rodrigues, S.G.; Degallier, N.; Moraes, M.A.; da Rosa, J.F.; da Rosa, E.S.; Mondet, B.; Barros, V.L.; da Rosa, A.P. An Epidemic of Sylvatic Yellow Fever in the Southeast Region of Maranhao State, Brazil, 1993-1994: Epidemiologic and Entomologic Findings. *Am. J. Trop. Med. Hyg.* **1997**, *57*, 132–137, doi:10.4269/ajtmh.1997.57.132.
7. Rosa, A.P. de A.T. da; Rosa, A.P. de A.T. da A História Da Arbovirologia No Instituto Evandro Chagas, Belém, Pará, Brasil, de 1954 a 1998. *Rev. Pan-Amaz. Saúde* **2016**, *7*, 61–70, doi:10.5123/s2176-62232016000500007.

8. Dégallier, N.; Rosa, A.P. de A.T. da; Vasconcelos, P.F. da C.; Rosa, E.S.T. da; Rodrigues, S.G.; Sá Filho, G.C.; Rosa, J.F.S.T. da New Entomological and Virological Data on the Vectors of Sylvatic Yellow Fever in Brazil. *Arthropod-Borne Virus Inf Exch* **1993**, *21*–22.
9. Vasconcelos, P.F.; Rosa, A.P.; Rodrigues, S.G.; Rosa, E.S.; Monteiro, H.A.; Cruz, A.C.; Barros, V.L.; Souza, M.R.; Rosa, J.F. Yellow Fever in Pará State, Amazon Region of Brazil, 1998-1999: Entomologic and Epidemiologic Findings. *Emerg. Infect. Dis.* **2001**, *7*, 565–569, doi:10.3201/eid0707.010738.
10. Brasil, M. da S. *Guia de Vigilância de Epizootias Em Primatas Não Humanos e Entomologia Aplicada à Vigilância Da Febre Amarela*; 2nd ed.; Ministério da Saúde, Secretaria de Vigilância em Saúde, Departamento de Vigilância das Doenças Transmissíveis, 2014.
11. Reiter, P.; Gubler, D. *Surveillance and Control of Urban Dengue Vectors*; Dengue and dengue hemorrhagic fever.; Gubler D J, Kuno G CAB International: London, United Kingdom, 1997;
12. Lima, J.B.P.; Rosa-Freitas, M.G.; Rodvalho, C.M.; Santos, F.; Lourenço-de-Oliveira, R. Is There an Efficient Trap or Collection Method for Sampling *Anopheles Darlingi* and Other Malaria Vectors That Can Describe the Essential Parameters Affecting Transmission Dynamics as Effectively as Human Landing Catches? - A Review. *Mem. Inst. Oswaldo Cruz* **2014**, *109*, 685–705, doi:10.1590/0074-0276140134.
13. Achee, N.L.; Youngblood, L.; Bangs, M.J.; Lavery, J.V.; James, S. Considerations for the Use of Human Participants in Vector Biology Research: A Tool for Investigators and Regulators. *Vector Borne Zoonotic Dis.* **2015**, *15*, 89–102, doi:10.1089/vbz.2014.1628.
14. Service, M.W. *Mosquito Ecology: Field Sampling Methods*; Springer Science & Business Media, 1993; ISBN 978-94-011-1868-2.
15. Santos, J.C.; de Almeida, W.R.; Fernandes, G.W. Arthropods: Why It Is So Crucial to Know Their Biodiversity? In *Measuring Arthropod Biodiversity: A Handbook of Sampling Methods*; Santos, J.C., Fernandes, G.W., Eds.; Springer International Publishing: Cham, 2021; pp. 3–11 ISBN 978-3-030-53226-0.
16. Service, M.W. A Critical Review of Procedures for Sampling Populations of Adult Mosquitoes. *Bull. Entomol. Res.* **1977**, *67*, 343–382, doi:10.1017/S0007485300011184.
17. Hocking, B. The Use of Attractants and Repellents in Vector Control. *Bull. World Health Organ.* **1963**, *29*, 121–126.
18. Mucci, L.; Cardoso-Júnior, R.; Paula, M.; Fernandes, A.; Pacchioni, M.; Scandar, S. Potenciais Vetores Da Febre Amarela No Noroeste Do Estado de São Paulo, Brasil: Distribuição Geográfica e Associações Com o Ambiente Florestal. In *Proceedings of the Geotecnologías, Herramientas para la construcción de una nueva visión del cambio global y su transformación para un futuro sostenible: Libro de Actas de XVII Simposio Internacional en Percepción Remota y Sistemas de Información Geográfica*; Walter F. Sione [et al.] EdUnLu: Luján, Argentina, 2016; Vol. 1, pp. 661–672.
19. Watentena, A.; Okoye, I. The Untapped Potential of Mosquito Lures for Malaria Vector Surveillance and Mass Trapping of Mosquitoes: A Review. *Int. J. Mosq. Res.* **2019**, *6*, 132–137.
20. Alencar, J.; Melandri, V.; Silva, J.; Albuquerque, H.G.; Guimarães, A.É. Ecological Characterization of Mosquitoes (Diptera: Culicidae) in Areas of the Mato Grosso Pantanal, Mato Grosso State, Brazil. *Trop. Zool.* **2021**, *34*, doi:10.4081/tz.2021.84.
21. Barrio-Nuevo, K.M.; Cunha, M.S.; Luchs, A.; Fernandes, A.; Rocco, I.M.; Mucci, L.F.; de Souza, R.P.; Medeiros-Sousa, A.R.; Ceretti-Junior, W.; Marrelli, M.T. Detection of Zika and Dengue Viruses in Wild-Caught Mosquitoes Collected during Field Surveillance in an Environmental Protection Area in São Paulo, Brazil. *PLoS One* **2020**, *15*, e0227239.

22. Kröckel, U.; Rose, A.; Eiras, A.E.; Geier, M. New Tools for Surveillance of Adult Yellow Fever Mosquitoes: Comparison of Trap Catches with Human Landing Rates in an Urban Environment. *J. Am. Mosq. Control Assoc.* **2006**, *22*, 229–238.
23. Ázara, T.M.F. de; Degener, C.M.; Roque, R.A.; Ohly, J.J.; Geier, M.; Eiras, Á.E.; Ázara, T.M.F. de; Degener, C.M.; Roque, R.A.; Ohly, J.J.; et al. The Impact of CO₂ on Collection of *Aedes Aegypti* (Linnaeus) and *Culex Quinquefasciatus* Say by BG-Sentinel(r) Traps in Manaus, Brazil. *Mem. Inst. Oswaldo Cruz* **2013**, *108*, 229–232, doi:10.1590/0074-0276108022013016.
24. Degener, C.M.; Ázara, T.M.F. de; Roque, R.A.; Codeço, C.T.; Nobre, A.A.; Ohly, J.J.; Geier, M.; Eiras, Á.E. Temporal Abundance of *Aedes Aegypti* in Manaus, Brazil, Measured by Two Trap Types for Adult Mosquitoes. *Mem. Inst. Oswaldo Cruz* **2014**, *109*, 1030–1040.
25. Gama, R.A.; Silva, I.M. da; Monteiro, H.A. de O.; Eiras, Á.E. Fauna of Culicidae in Rural Areas of Porto Velho and the First Record of *Mansonia (Mansonia) Flaveola* (Coquillett, 1906), for the State of Rondônia, Brazil. *Rev. Soc. Bras. Med. Trop.* **2012**, *45*, 125–127, doi:10.1590/S0037-86822012000100025.
26. Sá, I.L.R. de; Sallum, M.A.M. Comparison of Automatic Traps to Capture Mosquitoes (Diptera: Culicidae) in Rural Areas in the Tropical Atlantic Rainforest. *Mem. Inst. Oswaldo Cruz* **2013**, *108*, 1014–1020, doi:10.1590/0074-0276130474.
27. Chaves, L.S.M.; Laporta, G.Z.; Sallum, M.A.M. Effectiveness of Mosquito Magnet in Preserved Area on the Coastal Atlantic Rainforest: Implication for Entomological Surveillance. *J. Med. Entomol.* **2014**, *51*, 915–924, doi:10.1603/ME14050.
28. Sant’Ana, D.C.; Sá, I.L.R. de; Sallum, M.A.M. Effectiveness of Mosquito Magnet® Trap in Rural Areas in the South-eastern Tropical Atlantic Forest. *Mem. Inst. Oswaldo Cruz* **2014**, *109*, 1021–1029, doi:10.1590/0074-02761400297.
29. Pinheiro, G.G.; Rocha, M.N.; de Oliveira, M.A.; Moreira, L.A.; Andrade Filho, J.D. Detection of Yellow Fever Virus in Sylvatic Mosquitoes during Disease Outbreaks of 2017–2018 in Minas Gerais State, Brazil. *Insects* **2019**, *10*, 136.
30. Deus, J.T. de; Mucci, L.F.; Lucheta Reginatto, S.; Pereira, M.; Bergo, E.S.; de Camargo-Neves, V.L.F. Evaluation of Methods to Collect Diurnal Culicidae (Diptera) at Canopy and Ground Strata, in the Atlantic Forest Biome. *Insects* **2022**, *13*, 202, doi:10.3390/insects13020202.
31. São Paulo, S.; Secretaria do Meio Ambiente do Estado de São Paulo Plano de Manejo PE Vassununga 2009. *Secr. Meio Ambiente Estado São Paulo* **2009**, 328.
32. Veloso, H.P.; Rangel Filho, A.L.R.; Lima, J.C.A. *Classificação da vegetação brasileira, adaptada a um sistema universal*; Ministério da Economia, Fazenda e Planejamento, Fundação Instituto Brasileiro de Geografia e Estatística, Diretoria de Geociências, Departamento de Recursos Naturais e Estudos Ambientais: Rio de Janeiro, 1991; ISBN 978-85-240-0384-4.
33. Sudia, W.D.; Chamberlain, R.W. Battery-Operated Light Trap, an Improved Model. *Mosq. News* **1962**, *22*, 126–129.
34. Reinert, J.E. Revised List of Abbreviations for Genera and Subgenera of Culicidae (Diptera) and Notes on Generic and Subgeneric Changes. *J. Am. Mosq. Control Assoc.* **2001**, *17*, 51–55.
35. Lane, J. *Neotropical Culicidae*; EDUSP: São Paulo, 1953; Vol. 2;.
36. Colwell, R. EstimateS: Statistical Estimation of Species Richness and Shared Species from Samples 2013.
37. Hammer, O.; Harper, D.; Ryan, P. PAST: Paleontological Statistics Software Package for Education and Data Analysis. *Palaeontol. Electron.* **2001**, *4*, 1–9.
38. Guimarães, A.É.; Arlé, M. Mosquitos no Parque Nacional da Serra dos Órgãos, estado do Rio de Janeiro, Brasil: I-distribuição estacional. *Mem. Inst. Oswaldo Cruz* **1984**, *79*, 309–323, doi:10.1590/S0074-02761984000300004.

39. Forattini, O.P.; Gomes, A. de C. Biting Activity of *Aedes Scapularis* (Rondani) and *Haemagogus* Mosquitoes in Southern Brazil (Diptera: Culicidae). *Rev. Saúde Pública* **1988**, *22*, 84–93, doi:10.1590/S0034-89101988000200003.
40. Gomes, A. de C.; Torres, M.A.N.; Paula, M.B. de; Fernandes, A.; Marassá, A.M.; Consales, C.A.; Fonseca, D.F. Ecologia de *Haemagogus* e *Sabethes* (Diptera: Culicidae) Em Áreas Epizooticas Do Vírus Da Febre Amarela, Rio Grande Do Sul, Brasil. *Epidemiol. E Serviços Saúde* **2010**, *19*, 101–113, doi:10.5123/S1679-49742010000200003.
41. Silva, J. dos S.; Pacheco, J.B.; Alencar, J.; Guimarães, A.É. Biodiversity and Influence of Climatic Factors on Mosquitoes (Diptera: Culicidae) around the Peixe Angical Hydroelectric Scheme in the State of Tocantins, Brazil. *Mem. Inst. Oswaldo Cruz* **2010**, *105*, 155–162, doi:10.1590/S0074-02762010000200008.
42. Alencar, J.; Pacheco, J.B.; dos Santos Silva, J.; Silva, S.O.F.; Guimarães, A.É. Influence of Climatic Factors On *Psorophora* (Janthinosoma) *Albigenu* In Pantanal Landscape, Mato Grosso State, Brazil. *J. Am. Mosq. Control Assoc.* **2018**, *34*, 177–181, doi:10.2987/18-6749.1.
43. Couto-Lima, D.; Andrezza, C.S.; Leite, P.J.; Bersot, M.I.L.; Alencar, J.; Lourenço-de-Oliveira, R. Seasonal Population Dynamics of the Primary Yellow Fever Vector *Haemagogus Leucocelaenus* (Dyar & Shannon) (Diptera: Culicidae) Is Mainly Influenced by Temperature in the Atlantic Forest, Southeast Brazil. *Mem. Inst. Oswaldo Cruz* **2020**, *115*, doi:10.1590/0074-02760200218.
44. Carvalho, M.E.S.D.; Naves, H.A.M.; Carneiro, E.; Miranda, M.F. de Distribuição vertical de mosquitos dos gêneros *Haemagogus* e *Sabethes*, em zona urbana de Goiânia-Goiás-Brasil. *Rev. Patol. Trop. J. Trop. Pathol.* **1997**, *26*, doi:10.5216/rpt.v26i1.17371.
45. Lira-Vieira, A.R.; Gurgel-Gonçalves, R.; Moreira, I.M.; Yoshizawa, M.A.C.; Coutinho, M.L.; Prado, P.S.; Souza, J.L. de; Chaib, A.J. de M.; Moreira, J.S.; Castro, C.N. de Ecological Aspects of Mosquitoes (Diptera: Culicidae) in the Gallery Forest of Brasília National Park, Brazil, with an Emphasis on Potential Vectors of Yellow Fever. *Rev. Soc. Bras. Med. Trop.* **2013**, *46*, 566–574, doi:10.1590/0037-8682-0136-2013.
46. Werneck, F.P.; Nogueira, C.; Colli, G.R.; Sites Jr, J.W.; Costa, G.C. Climatic Stability in the Brazilian Cerrado: Implications for Biogeographical Connections of South American Savannas, Species Richness and Conservation in a Biodiversity Hotspot. *J. Biogeogr.* **2012**, *39*, 1695–1706, doi:10.1111/j.1365-2699.2012.02715.x.
47. Camargo-Neves, V.L.F.; Poletto, D.W.; Rodas, L.A.C.; Pacchioli, M.; Cardoso, R.P.; Scandar, S.A.S.; Sampaio, S.M.P.; Koyanagui, P.H.; Botti, M.V.; Mucci, L.F.; Gomes, A. C. I Entomological investigation of a sylvatic yellow fever area in São Paulo State, Brazil. *Cad Saude Públ* **2005**, *21*, 1278–1286. <https://doi.org/10.1590/S0102-311X2005000400031>
48. Moreno, E.S.; Rocco, I.M.; Bergo, E.S.; Brasil, R.A.; Siciliano, M.M.; Suzuki, A.; Silveira, V.R.; Bisordi, I.; Souza, R.P. de Reemergence of Yellow Fever: Detection of Transmission in the State of São Paulo, Brazil, 2008. *Rev. Soc. Bras. Med. Trop.* **2011**, *44*, 290–296.
49. Cunha, M.S.; da Costa, A.C.; de Azevedo Fernandes, N.C.C.; Guerra, J.M.; dos Santos, F.C.P.; Nogueira, J.S.; D'Agostino, L.G.; Komninakis, S.V.; Witkin, S.S.; Ressio, R.A.; et al. Epizootics Due to Yellow Fever Virus in São Paulo State, Brazil: Viral Dissemination to New Areas (2016–2017). *Sci. Rep.* **2019**, *9*, 5474, doi:10.1038/s41598-019-41950-3.
50. Cunha, M.S.; Tubaki, R.M.; de Menezes, R.M.T.; Pereira, M.; Caleiro, G.S.; Coelho, E.; del Castillo Saad, L.; de Azevedo Fernandes, N.C.C.; Guerra, J.M.; Nogueira, J.S. Possible Non-Sylvatic Transmission of Yellow Fever between Non-Human Primates in São Paulo City, Brazil, 2017–2018. *Sci. Rep.* **2020**, *10*, 1–8.
51. Pivello, V.R.; Bitencourt, M.D.; Mantovani, W.; Junior, N. de M.; Batalha, M.A.; Shida, C.N. Proposta de Zoneamento Ecológico para a RESERVA DE Cerrado Pé-de-Gigante (Santa Rita do Passa Quatro, SP). *Braz J Ecol* **1998**, *18*.

52. Fidelis, A.T.; Godoy, S.A.P. de Estrutura de um cerrado strico sensu na Gleba Cerrado Pé-de-Gigante, Santa Rita do Passa Quatro, SP. *Acta Bot. Bras.* **2003**, *17*, 531–539, doi:10.1590/S0102-33062003000400006.
53. Marcondes, C.B.; Alencar, J. Revisão de mosquitos *Haemagogus Williston* (Diptera: Culicidae) do Brasil. **2010**, *21*, 18.
54. Alencar, J.; Dégallier, N.; Hannart, A.; Silva, J. dos S.; Pacheco, J.B.; Guimarães, A.É. Circadian and Seasonal Preferences for Hematophagy among *Haemagogus Capricornii*, *Hg. Janthinomys*, and *Hg. Leucocelaenus* (Diptera: Culicidae) in Different Regions of Brazil. *J. Vector Ecol.* **2008**, *33*, 389–392, doi:10.3376/1081-1710-33.2.389.
55. Hendy, A.; Hernandez-Acosta, E.; Valério, D.; Mendonça, C.; Costa, E.R.; Júnior, J.T.A.; Assunção, F.P.; Scarpassa, V.M.; Gordo, M.; Fé, N.F.; et al. The Vertical Stratification of Potential Bridge Vectors of Mosquito-Borne Viruses in a Central Amazonian Forest Bordering Manaus, Brazil. *Sci. Rep.* **2020**, *10*, 18254, doi:10.1038/s41598-020-75178-3.
56. Hendy, A.; Valério, D.; Fé, N.F.; Hernandez-Acosta, E.; Mendonça, C.; Andrade, E.; Pedrosa, I.; Costa, E.R.; Júnior, J.T.A.; Assunção, F.P.; et al. Microclimate and the Vertical Stratification of Potential Bridge Vectors of Mosquito-borne Viruses Captured by Nets and Ovitrap in a Central Amazonian Forest Bordering Manaus, Brazil. *Sci. Rep.* **2021**, *11*, 21129, doi:10.1038/s41598-021-00514-0.
57. Krockel, U.; Rose, A.; Eiras, A.E.; Geier, M. New Tools for Surveillance of Adult Yellow Fever Mosquitoes: Comparison of Trap Catches with Human Landing Rates in an Urban Environment. *J. Am. Mosq. Control Assoc.* **2006**, *22*, 229–238, doi:10.2987/8756-971X(2006)22[229:NTFSOA]2.0.CO;2.
58. Laporta, G.Z.; Sallum, M.A.M. Effect of CO₂ and 1-Octen-3-Ol Attractants for Estimating Species Richness and the Abundance of Diurnal Mosquitoes in the Southeastern Atlantic Forest, Brazil. *Mem. Inst. Oswaldo Cruz* **2011**, *106*, 279–284.
59. Sant’Ana, D.C.; Sá, I.L.R. de; Sallum, M.A.M. Effectiveness of Mosquito Magnet® Trap in Rural Areas in the Southeastern Tropical Atlantic Forest. *Mem. Inst. Oswaldo Cruz* **2014**, *109*, 1021–1029, doi:10.1590/0074-02761400297.
60. Pinheiro, G.G.; Rocha, M.N.; de Oliveira, M.A.; Moreira, L.A.; Andrade Filho, J.D. Detection of Yellow Fever Virus in Sylvatic Mosquitoes during Disease Outbreaks of 2017–2018 in Minas Gerais State, Brazil. *Insects* **2019**, *10*, 136.
61. Câmara, D.C.P.; Codeço, C.T.; Ayllón, T.; Nobre, A.A.; Azevedo, R.C.; Ferreira, D.F.; da Silva Pinel, C.; Rocha, G.P.; Honório, N.A. Entomological Surveillance of *Aedes* Mosquitoes: Comparison of Different Collection Methods in an Endemic Area in RIO de Janeiro, Brazil. *Trop. Med. Infect. Dis.* **2022**, *7*, 114, doi:10.3390/tropicalmed7070114.
62. Cano, M.E.; Marti, G.A.; Alencar, J.; Silva, S.O.F.; Micieli, M.V. Categorization by Score of Mosquito Species (Diptera: Culicidae) Related to Yellow Fever Epizootics in Argentina. *J. Med. Entomol.* **2022**, *59*, 1766–1777, doi:10.1093/jme/tjac079.
63. Monath, T.P.; Vasconcelos, P.F.C. Yellow Fever. *J. Clin. Virol. Off. Publ. Pan Am. Soc. Clin. Virol.* **2015**, *64*, 160–173, doi:10.1016/j.jcv.2014.08.030.
64. Abreu, F.V.S. de; Ribeiro, I.P.; Ferreira-de-Brito, A.; Santos, A.A.C. dos; Miranda, R.M. de; Bonelly, I. de S.; Neves, M.S.A.S.; Bersot, M.I.; Santos, T.P. dos; Gomes, M.Q.; et al. *Haemagogus Leucocelaenus* and *Haemagogus Janthinomys* Are the Primary Vectors in the Major Yellow Fever Outbreak in Brazil, 2016–2018. *Emerg. Microbes Infect.* **2019**, *8*, 218–231, doi:10.1080/22221751.2019.1568180.
65. Pinto, C.S.; Confalonieri, U.E.; Mascarenhas, B.M. Ecology of *Haemagogus* Sp. and *Sabethes* Sp. (Diptera: Culicidae) in Relation to the Microclimates of the Caxiuanã National Forest, Pará, Brazil. *Mem. Inst. Oswaldo Cruz* **2009**, *104*, 592–598, doi:10.1155/2012/741273.
66. Confalonieri, U.E.C.; Costa Neto, C. Diversity of Mosquito Vectors (Diptera: Culicidae) in Caxiuanã, Pará, Brazil. *Interdiscip. Perspect. Infect. Dis.* **2012**, *2012*, 741273, doi:10.1155/2012/741273.

-
67. Cardoso, J. da C.; De Almeida, M.A.; Dos Santos, E.; Da Fonseca, D.F.; Sallum, M.A.; Noll, C.A.; Monteiro, H.A. de O.; Cruz, A.C.; Carvalho, V.L.; Pinto, E.V. Yellow Fever Virus in *Haemagogus Leucocelaenus* and *Aedes Serratus* Mosquitoes, Southern Brazil, 2008. *Emerg. Infect. Dis.* **2010**, *16*, 1918.
 68. Souza, R.P. de; Petrella, S.; Coimbra, T.L.M.; Maeda, A.Y.; Rocco, I.M.; Bisordi, I.; Silveira, V.R.; Pereira, L.E.; Suzuki, A.; Silva, S.J. dos S.; et al. Isolation of Yellow Fever Virus (YFV) from Naturally Infected *Haemagogus* (Conopostegus) *Leucocelaenus* (Diptera, Cukicudae) in São Paulo State, Brazil, 2009. *Rev. Inst. Med. Trop. São Paulo* **2011**, *53*, 133–139, doi:10.1590/S0036-46652011000300004.
 69. Obara, M.T.; Monteiro, H.; Paula, M.B. de; Gomes, A. de C.; Yoshizawa, M.A.C.; Lira, A.R.; Boffil, M.I.R.; Carvalho, M. do S.L. de Infecção Natural de *Haemagogus Janthinomys* e *Haemagogus Leucocelaenus* Pelo Vírus Da Febre Amarela No Distrito Federal, Brasil, 2007-2008. *Epidemiol. E Serviços Saúde* **2012**, *21*, 457–463, doi:10.5123/S1679-49742012000300011.
 70. Brasil, M. da S. *Guia de Vigilância do Culex quinquefasciatus*; Normas e manuais técnicos; 3rd ed.; Ministério da Saúde: Brasília, D.F, 2011.
 71. Bowman, L.R.; Runge-Ranzinger, S.; McCall, P.J. Assessing the Relationship between Vector Indices and Dengue Transmission: A Systematic Review of the Evidence. *PLoS Negl. Trop. Dis.* **2014**, *8*, e2848, doi:10.1371/journal.pntd.0002848.
 72. Tabbabi, A.; Daaboub, J. First Investigation of Deltamethrin Pyrethroid Susceptibility and Resistance Status of *Anopheles Labbranchiae* (Falleroni, 1926), Potential Malaria Vector in Tunisia. *Asian Pac. J. Trop. Biomed.* **2017**, *7*, 1067–1070, doi:10.1016/j.apjtb.2017.10.007.